Check for updates

OPEN ACCESS

EDITED BY Sarah M. Temkin, National Institutes of Health (NIH), United States

REVIEWED BY Diego Raimondo, University of Bologna, Italy Komsun Suwannarurk, Thammasat University, Thailand

*CORRESPONDENCE Karl C. Podratz M podratz.karl@mayo.edu

RECEIVED 22 November 2022 ACCEPTED 09 May 2023 PUBLISHED 23 May 2023

CITATION

Gonzalez-Bosquet J, Weroha SJ, Bakkum-Gamez JN, Weaver AL, McGree ME, Dowdy SC, Famuyide AO, Kipp BR, Halling KC, Yadav S, Couch FJ and Podratz KC (2023) Prognostic stratification of endometrial cancers with high microsatellite instability or no specific molecular profile. *Front. Oncol.* 13:1105504. doi: 10.3389/fonc.2023.1105504

COPYRIGHT

© 2023 Gonzalez-Bosquet, Weroha, Bakkum-Gamez, Weaver, McGree, Dowdy, Famuyide, Kipp, Halling, Yadav, Couch and Podratz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Prognostic stratification of endometrial cancers with high microsatellite instability or no specific molecular profile

Jesus Gonzalez-Bosquet¹, S. John Weroha², Jamie N. Bakkum-Gamez³, Amy L. Weaver⁴, Michaela E. McGree⁴, Sean C. Dowdy³, Abimbola O. Famuyide³, Benjamin R. Kipp⁵, Kevin C. Halling⁵, Siddhartha Yadav², Fergus J. Couch^{4,5} and Karl C. Podratz³*

¹Department of Obstetrics and Gynecology, University of Iowa, Iowa City, IA, United States, ²Division of Medical Oncology, Mayo Clinic, Rochester, MN, United States, ³Department of Obstetrics and Gynecology, Mayo Clinic, Rochester, MN, United States, ⁴Department of Quantitative Health Sciences, Mayo Clinic, Rochester, MN, United States, ⁵Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, United States

Objective: To identify high-risk disease in clinicopathologic low-risk endometrial cancer (EC) with high microsatellite instability (MSI-H) or no specific molecular profile (NSMP) and therapeutic insensitivity in clinicopathologic high-risk MSI-H/ NSMP EC.

Methods: We searched The Cancer Genome Atlas for DNA sequencing, RNA expression, and surveillance data regarding MSI-H/NSMP EC. We used a molecular classification system of *E2F1* and *CCNA2* expression and sequence variations in *POLE*, *PPP2R1A*, or *FBXW7* (ECPPF) to prognostically stratify MSI-H/NSMP ECs. Clinical outcomes were annotated after integrating ECPPF and sequence variations in homologous recombination (HR) genes.

Results: Data were available for 239 patients with EC, which included 58 MSI-H and 89 NSMP cases. ECPPF effectively stratified MSI-H/NSMP EC into distinct molecular groups with prognostic implications: molecular low risk (MLR), with low *CCNA2* and *E2F1* expression, and molecular high risk (MHR), with high *CCNA2* and *E2F1* expression and/or *PPP2R1A* and/or *FBXW7* variants. The 3-year disease-free survival (DFS) rate was 43.8% in the MHR group with clinicopathologic low-risk indicators and 93.9% in the MLR group (*P*<.001). In the MHR group, wild-type HR genes were present in 28% of cases but in 81% of documented recurrences. The 3-year DFS rate in patients with MSI-H/NSMP EC with clinicopathologic high-risk indicators was significantly higher in the MLR (94.1%) and MHR/HR variant gene (88.9%) groups than in the MHR/HR wild-type gene group (50.3%, *P*<.001).

Conclusion: ECPPF may resolve prognostic challenges for MSI-H/NSMP EC by identifying occult high-risk disease in EC with clinicopathologic low-risk indicators and therapeutic insensitivity in EC with clinicopathologic high-risk indicators.

KEYWORDS

CCNA2, ECPPF, E2F1, endometrial cancer, FBXW7, MSI, NSMP, PPP2R1A

Introduction

Over the past 4 decades, prognostic stratification of endometrial cancer (EC) has progressively incorporated serous histopathologic findings, lymphatic metastases, lymphovascular space involvement, TP53 variations, ERBB2 expression, mismatch repair and homologous recombination (HR) repair deficiencies, and other variables (1-5). More recently, The Cancer Genome Atlas (TCGA) generated a molecular-based prognostic classification system (6). Despite the evolution of EC risk stratification, corresponding risk-adjusted therapy has minimally affected EC mortality rates. The American Cancer Society 2020 Annual Report concluded that "cancer survival has improved since the mid-1970s for all of the most common cancers except uterine cervix and uterine corpus" and that "stagnant survival rates reflect a lack of major treatment advances" (7). Addressing this lack of major treatment advances requires an enhanced understanding of pivotal tumor-specific molecular drivers to facilitate the discovery of target-specific therapeutics.

TCGA classification of EC identified 4 unique molecular subgroups: 1) POLE ultramutated, characterized by POLE variations; 2) microsatellite instability hypermutated, classified by mismatch repair deficiency and high microsatellite instability (MSI-H); 3) copy-number low, denoted by no specific molecular profile (NSMP); and 4) copy-number high, primarily typified by TP53 variations and serouslike EC (6). Comparative prognoses associated with POLE ultramutated and copy-number high EC are contrasting, in which the prognosis is favorable for POLE ultramutated EC and substantially compromised for copy-number high EC. MSI-H and NSMP ECs have an intermediate prognosis but account for most EC tumors, thereby highly contributing to the annual EC mortality rate (6, 8-10). Therefore, determining the causes of treatment failures in MSI-H and NSMP ECs is paramount. Because MSI-H and NSMP tumors are predominantly low stage, with grade 1 and 2 histopathologic indicators (11), treatment failures may reflect our inability to identify clinicopathologic low-risk cases at high risk for recurrence (i.e., EC harboring occult high-risk disease). Radiotherapy (RT) and/or platinum-based chemotherapy (PbCT) are standard adjuvant treatment modalities for clinicopathologic high-risk MSI-H and NSMP EC. Treatment failures may also result from our inability to predict insensitivity to these standard adjuvant therapeutics (9, 12).

MSI-H EC generally harbors a high tumor mutation burden (TMB), and thus patients with MSI-H tumors are considered potential candidates for immunotherapy (6, 13-16). However, the high associated TMB presumably predicts a greater prevalence of HR gene variations, which portend sensitivity to DNA-damaging agents in cases with clinicopathologic high-risk indicators (17-20). Defining specific vulnerabilities in MSI-H tumors is necessary for decreasing the mortality rate in patients with this molecular profile. Furthermore, because of the absence of identifiable molecularly targetable pathways in NSMP, empiric therapy must be used for the largest EC cohort. Therefore, we used our previously described stratification system based on E2F1 and CCNA2 expression and sequence variations in POLE, PPP2R1A, and FBXW7 (termed ECPPF) to molecularly stratify MSI-H and NSMP ECs (21). The primary aim of our study was to use ECPPF to identify MSI-H and NSMP EC cases with clinicopathologic low-risk indicators that are at risk for occult extrauterine disease and recurrence and those with clinicopathologic high-risk indicators that are insensitive to standard therapeutic regimens.

Methods

Study population

The initial report of the ECs curated in the TCGA database consisted of 239 cases with documented DNA sequencing, RNA expression, and sufficient surveillance data (6). According to TCGA molecular classification of these cases, 30 tumors had *POLE* variations, 58 had MSI-H, 62 had *TP53* variations (presumed copy-number high), and an estimated 89 had NSMP (i.e., copy-number low). We previously reported the integration of genomic, transcriptomic, and clinical outcomes data for these cases, which culminated in the generation of the ECPPF stratification system (21). However, the molecular-based stratification of clinical outcomes for these EC cases may have been biased by a relatively high percentage (26%) of tumors with high-risk *TP53* variations;

Abbreviations: DFS, disease-free survival; EC, endometrial cancer; ECPPF, stratification with *E2F1* and *CCNA2* expression and sequence variations in *POLE*, *PPP2R1A*, and *FBXW7*; HR, homologous recombination; LVSI, lymphovascular space involvement; MHR, molecular high risk; MI, myometrial invasion; MLR, molecular low risk; MSI-H, high microsatellite instability; NSMP, no specific molecular profile; PbCT, platinum-based chemotherapy; RT, radiotherapy; TCGA, The Cancer Genome Atlas; TMB, tumor mutation burden.

10.3389/fonc.2023.1105504

therefore, we limited the current study population to only the 147 cases of MSI-H and NSMP EC, which consisted predominantly of early-stage, low-grade tumors. As previously reported, the EC tumors included in TCGA were surgically staged, and adjuvant therapy consisted of chemotherapy and/or RT, with 98% of patients who received chemotherapy receiving PbCT (6).

Data acquisition

TCGA data relevant to EC (6) were downloaded as normalized, formatted, and organized data for integration and analysis as described in previous studies (21, 22). All data collection and processing, including the informed consent process, were performed after approval by the local institutional review board or ethics committee for each contributing institution and in accordance with TCGA Human Subjects Protection and Data Access Policies adopted by the National Cancer Institute and the National Human Genome Research Institute.

Sequence variation analysis

Only validated sequence variations (or TCGA level 3 variations) were used for analysis (6). Sequence variant information was abstracted from exome-sequencing data generated with the Illumina Genome Analyzer GAIIx or HiSeq 2000 sequencing platforms (Illumina, Inc). Silent variations were excluded from the analysis, and only frame-shift insertions and deletions, in-frame insertions or deletions, missense, nonsense, nonstop, and splice-site variations were included in the study. For our analysis, the number of variations for each selected gene and for each patient were recorded.

Gene expression

Normalized and log-transformed gene expression data were downloaded as level 3 RNA-sequencing data. These data were generated with Illumina HiSeq 2000 platforms and annotated with the hg19 version of the human genome. Statistical analyses of RNA expression data were performed with R software (The R Foundation) (23) and Bioconductor packages (Bioconductor) (24).

Molecular stratification of MSI-H and NSMP EC

We previously reported the applicability of ECPPF for stratifying the unabridged EC cohort in TCGA independent of histomorphologic variables (21). Restricting the study population to MSI-H and NSMP EC rendered analysis of *POLE* variations expendable from the ECPPF profiling panel. The log-transformed gene expression equal to or greater than 2.75 for either or both *CCNA2* and *E2F1* was converted to the quantitative expression sum

of CCNA2+E2F1 and categorized as either low expression (sum<4.75) or high expression (sum≥4.75). Assessment of HR gene variations was restricted to the 10 most prevalent genes (*ATR*, *ATM*, *BRCA2*, *BRCA1*, *CDK12*, *BARD1*, *NBN*, *PALB2*, *CHEK1*, and *RAD51*) reportedly linked to EC and MSI-H (3, 25–28).

Statistical analysis

Data were descriptively summarized as frequency and percentage for categorical variables and median (IQR) for continuous variables. Clinicopathologic and molecular risk factors were compared between the MSI-H and NSMP molecular classification groups by using χ^2 or Fisher exact tests for categorical variables. Follow-up duration was calculated from the date of surgical resection to the date of first documented recurrence or latest follow-up. Disease-free survival (DFS) rates were estimated by using the Kaplan-Meier method and compared between groups with log-rank tests. Cox proportional hazards models were fit to evaluate the association between molecular parameters and the risk of recurrence; associations were summarized with hazard ratios and 95% CIs estimated from the models. All calculated P values less than .05 were considered statistically significant. The data were analyzed with SAS, v9.4 (SAS Institute Inc).

Results

Clinicopathologic stratification of MSI-H and NSMP EC

MSI-H (n=58) and NSMP (n=89) tumors had prevalent earlystage, low-grade histopathologic characteristics and 50% or less myometrial invasion (MI) (Table 1). The median (IQR) age of the 147 patients with these tumors was 60 (54-67) years. Recurrences were documented in 26 (18%) cases, and the median (IQR) time to recurrence after surgical resection was 13.9 (10.6-22.9) months. Among the 121 cases without evidence of recurrence, the median (IQR) follow-up period was 28.3 (15.7-44.8) months.

We integrated tumor stage, grade, and MI data from the MSI-H and NSMP cases to analyze tumors with *clinicopathologic low-risk indicators* (i.e., stage I, grade 1/2, and <75% MI) and *clinicopathologic high-risk indicators* (i.e., stage I, grade 1/2, and \geq 75% MI; stage I, grade 3; or stage II-IV). Although grade 3 histopathologic and clinicopathologic high-risk indicators were more prevalent in MSI-H tumors than in NSMP tumors, the 3year DFS rates for patients with MSI-H and NSMP EC did not significantly differ (P=.34) (Figure 1A). The 3-year DFS rate (95% CI) was similar for patients with MSI-H tumors with clinicopathologic low-risk indicators and those with high-risk indicators (72.3% [53.9%-97.0%] vs 74.2% [59.4%-92.7%], P=.98) (Figure 1B). The 3-year DFS rate (95% CI) also did not significantly differ between patients with NSMP tumors with clinicopathologic low-risk indicators and those with high-risk indicators (79.0% TABLE 1 Clinicopathologic and molecular characteristics of endometrial cancer with MSI-H and NSMP^a.

Characteristic	Total (N=147)	MSI-H (n=58)	NSMP (n=89)	Р
Age, y	60 (54-67)	60 (57-69)	60 (53-65)	.16
Stage				.23
I/II	121 (82.3)	45 (78)	76 (85)	
III/IV	26 (17.7)	13 (22)	13 (15)	
Grade				<.001
1	62 (42.2)	19 (33)	43 (48)	
2	58 (39.5)	19 (33)	39 (44)	
3	27 (18.4)	20 (35)	7 (8)	
MI				.99
≤50%	114 (77.6)	45 (78)	69 (78)	
>50%	33 (22.4)	13 (22)	20 (23)	
Clinicopathologic risk level ^b				.006
Low	91 (61.9)	28 (48)	63 (71)	
High	56 (38.1)	30 (52)	26 (29)	
Gene variations				
HR genes	33 (22.4)	26 (45)	7 (8)	<.001
CTNNB1	49 (33.3)	19 (33)	30 (34)	.91
ARID1A	55 (37.4)	22 (38)	33 (37)	.92
KRAS	30 (20.4)	14 (24)	16 (18)	.37
PIK3CA	82 (55.8)	42 (72)	40 (45)	.001
PTEN	101 (68.7)	37 (64)	64 (72)	.30
TP53	4 (2.7)	4 (7)	0 (0)	.02
ECPPF group				
Molecular low risk	90 (61.2)	27 (47)	63 (71)	.003
Molecular high risk ^c	57 (38.8)	31 (53)	26 (29)	
CCNA2/E2F1≥4.75	40 (27.2)	25 (43)	15 (17)	<.001
PPP2R1A variant	15 (10.2)	4 (7)	11 (12)	.28
FBXW7 variant	9 (6.1)	6 (10)	3 (3)	.16
Surveillance time, mo				.01
To recurrence	13.9 (10.7-22.4)	11.7 (11.1-17.9)	17 (10.6-23.5)	
No recurrence	28.3 (15.7-44.8)	28.5 (15.5-61.6)	28.3 (16.2-40.8)	

ECPPF, stratification according to E2F1 and CCNA2 expression and sequence variations in POLE, PPP2R1A, and FBXW7; HR, homologous recombination; MI, myometrial invasion; MSI-H, high microsatellite instability; NSMP, no specific molecular profile.

^aAge and surveillance time are summarized as median (IQR). All other values are No. (%), and comparisons were made with χ^2 or Fisher exact tests.

^bClinicopathologic low risk: stage I, grade 1/2, <75% MI. Clinicopathologic high risk: stage I, grade 1/2, ≥75% MI; stage I, grade 3; and stage II-IV.

^cCombinations of CCNA2/E2F1≥4.75 and FBWX7 and/or PPP2R1A variant subgroups were present in the MSI-H (n=4 combinations) and NSMP (n=3 combinations) endometrial cancer tumors.

[67.9%-92.0%] vs 87.3% [71.8%-100.0%], P=.42) (Figure 1C). Likewise, the 3-year DFS rate for the combined cohort of patients with MSI-H and NSMP ECs did not differ between those with clinicopathologic low-risk indicators (n=91) and those with high-risk indicators (n=56, P=.79) (Figure 1D).

ECPPF stratification of MSI-H/NSMP EC

ECPPF stratified MSI-H/NSMP cases (N=147) into 2 distinct molecular groups with prognostic implications: 1) molecular low risk (MLR), characterized by low *CCNA2* and/or *E2F1* expression



(i.e., CCNA2+E2F1 quantitative expression sum<4.75), and 2) molecular high risk (MHR). The MHR group was further stratified into 3 subgroups: 1) $CCNA2/E2F1\ge4.75$, characterized by high CCNA2 and/or E2F1 expression (i.e., CCNA2+E2F1 quantitative expression sum ≥4.75); 2) PPP2R1A variant; and 3) FBXW7 variant. The 3-year DFS rate of the MLR group (n=90) was significantly higher than that of the MHR subgroups (P<.001) (Figure 1E). The hazard ratio (95% CI) was 10.48 (3.46-31.77) for the $CCNA2/E2F1\ge4.75$ subgroup, 16.59 (3.67-74.99) for the FBWX7 variant subgroup, and 7.74 (1.93-30.95) for the PPP2R1A variant subgroup (reference: MLR subgroup, P<.001).

ECPPF stratification of MSI-H EC

We assessed clinical outcomes according to the ECPPFstratified groups for MSI-H cases (Figure 2A). The 3-year DFS rate (95% CI) was 95.0% (85.9%-100.0%) for the MLR group, 58.2% (39.4%-85.8%) for the *CCNA2/E2F1* \ge 4.75 subgroup, and 53.6% (25.7%-100.0%) for the combined *PPP2R1A/FBXW7* variant subgroups (*P*=.01).

Integrating ECPPF and HR gene variations in MSI-H EC

Among 58 MSI-H cases, 26 (45%) had at least 1 HR gene variation (Table 1). *ATR*, *ATM*, *BRCA2*, and *BRCA1* had the most sequence variations (Figure 2B). We used ECPPF to compare clinical outcomes of patients with stage I, grade 1/2 (i.e., *low-risk*) MSI-H EC tumors with HR variant genes and those with stage I, grade 3 or stage II-IV (i.e., *at-risk*) MSI-H EC tumors with HR variant genes (generally adjuvant therapy candidates) (Figure 2C). Most of the at-risk MSI-H cases with HR variant genes were stratified by ECPPF as MHR; however, no recurrences were documented in these cases. In contrast, several recurrences were documented in the low-risk MSI-H cases with HR variant genes, and most of these were stratified by ECPPF as MHR (Figure 2C). The 3-year DFS rate was also significantly lower for patients with low-risk MSI-H tumors with HR variant genes than those with at-risk tumors (*P*=.04) (Figure 2D).

MSI-H tumors with wild-type HR genes were equally distributed among low-risk and at-risk cases. ECPPF stratification of these cases showed differential outcomes for the MLR and MHR



the following molecular high-risk (MHR) subgroups: *CCNA2IE2F*124.75 (n=22) and combined *PPP2R1A/FBXW7* variant (n=9). (**B**) Number of documented sequence variations in select homologous recombination (HR) genes in the MSI-H cohort. (**C**) Pattern of stage, grade, recurrences, ECPPF-stratified MLR and MHR groups, and variations in *CTNNB1*, *ARID1A*, *KRAS*, and *TP53* genes in cases of MSI-H EC with HR variant genes (red line delineates between stage I, grade 1/2 and other tumors). (**D**) DFS of patients with MSI-H EC with HR variant genes according to low-risk (n=13) and at-risk (n=13) tumors. (**E**) Pattern of stage, grade, recurrences, ECPPF-stratified MLR and MHR groups, and variations in *CTNNB1*, *ARID1A*, *KRAS*, and *TP53* genes in cases of MSI-H EC with wild-type HR genes (red line delineates between stage I, grade 1/2 and other tumors). (**D**) DFS of patients with MSI-H EC with HR variant genes according to low-risk (n=13) and at-risk (n=13) tumors. (**E**) Pattern of stage, grade, recurrences, ECPPF-stratified MLR and MHR groups, and variations in *CTNNB1*, *ARID1A*, *KRAS*, and *TP53* genes in cases of MSI-H EC with wild-type HR genes (red line delineates between stage I, grade 1/2 and other tumors). (**F**) DFS of patients with MSI-H EC with wild-type HR genes according to ECPPF-stratified MLR (n=16) and MHR (n=16) groups. (**G**) ECPPF stratification of MSI-H EC with clinicopathologic high-risk indicators (CPHR) according to MLR and MHR groups and HR variant gene status. (**H**) DFS of patients with MSI-H EC with clinicopathologic high-risk indicators according to ECPPF-stratified MLR group with wild-type HR genes (n=7), MLR group with HR variant genes (n=14), and MHR group with wild-type HR genes (n=9). Kaplan-Meier curves for DFS were compared with log-rank tests.

groups (Figure 2E). No recurrences were documented in the MSI-H cases with wild-type HR genes that were ECPPF-stratified as MLR, but recurrences were documented in most of the ECPPF-stratified MHR cases. The 3-year DFS rate (95% CI) for patients with MSI-H tumors with wild-type HR genes that were ECPPF-stratified as MHR was 41.0% (21.4%-78.6%), which was significantly lower than that for patients with ECPPF-stratified MLR EC (P=.002) (Figure 2F).

Clinical outcomes were assessed after segregating MSI-H tumors with clinicopathologic high-risk indicators and HR variant genes (n=30). Generally, patients with these tumors are candidates for adjuvant therapy. Clinicopathologic high-risk MSI-H cases were ECPPF-stratified as MLR, HR variant gene, and MHR groups (Figure 2G). The 3-year DFS rate (95% CI) was 100% (100.0%-100.0%) for the MLR group, 91.7% (77.3%-100.0%) for the HR variant gene group, and 33.3% (13.2%-84.0%) for the MHR group (P=.001) (Figure 2H).

ECPPF stratification of NSMP EC

Clinicopathologic low-risk indicators were present more often in NSMP (n=63, 71%) than in MSI-H (n=28, 48%) tumors (*P*=.006) (Table 1) but failed to predict superior DFS (Figure 1C). ECPPF stratification into the MLR group and MHR *CCNA2/E2F1* \geq 4.75 and combined *PPP2R1/FBXW7* variant subgroups resulted in significantly different 3-year DFS rates for patients with NSMP EC (*P*<.001) (Figure 3A). The 3-year DFS rate (95% CI) was 93.6% (86.7%-100.0%) for the MLR group, 36.7% (14.0%-96.0%) for the *CCNA2/E2F1* \geq 4.75 subgroup, and 61.1% (32.4%-100.0%) for the combined *PPP2R1A/FBXW7* variant subgroups. Furthermore, the hazard ratio (95% CI) was 14.29 (3.39-60.19) for the *CCNA2/* $E2F1 \ge 4.75$ subgroup and 9.72 (2.16-43.74) for the combined *PPP2R1A/FBXW7* variant subgroups (reference: MLR subgroup, *P*<.001) (Figure 3B).

ECPPF stratification of early-stage, lowgrade NSMP EC

Because 55 of 89 (62%) NSMP cases were stage I, grade 1/2 with 50% or less MI, which generally confers excellent outcomes without adjuvant chemotherapeutic treatment, we next used ECPPF to



Molecular stratification and clinical outcomes of endometrial cancer (EC) with no specific molecular profile (NSMP). (A) Disease-free survival (DFS) of patients with NSMP EC (n=89) according to the ECPPF-stratified molecular low-risk (MLR) group, characterized by *CCNA2/E2F1*<4.75 (n=63), and the following molecular high-risk (MHR) subgroups: *CCNA2/E2F1* \geq 4.75 (n=12) and combined *PPP2R1A/FBXW7* variant (n=14). DFS (95% CI) at 36 months is indicated for each group. (B) Forest plot of Cox proportional hazard ratios for the 3 ECPPF-stratified groups (reference: *CCNA2/E2F1*<4.75). (C) DFS of patients with stage I, grade 1/2, \leq 50% myometrial invasion (MI) NSMP EC (n=55) according to ECPPF-stratified MLR (n=39) and MHR (n=16) groups. (D) Pattern of recurrences, ECPPF-stratified MLR and MHR groups, and variations in *CTNNB1* and *ARID1A* in cases of NSMP with *CTNNB1* variations (n=21) according to ECPPF-stratified MLR (n=50) groups. (F) DFS of patients with NSMP EC with *CTNNB1* and *ARID1A* variations (n=21) according to ECPPF-stratified MLR (n=39) and MHR (n=15) and MHR (n=13) groups. Kaplan-Meier curves for DFS were compared with log-rank tests.

identify cases of occult high-risk disease in conventionally low-risk EC. ECPPF stratification of these NSMP early-stage, low-grade cases into MLR and MHR groups resulted in significantly different 3-year DFS rates (P<.001) (Figure 3C). The 3-year DFS rate (95% CI) was 93.9% (86.1%-100.0%) for the MLR group and 51.8% (29.7%-90.5%) for the MHR group. The hazard ratio (95% CI) for the MHR group was 12.10 (2.46-59.42) (reference: MLR group, P=.002).

ECPPF stratification of CTNNB1 variations in NSMP EC

Among 30 *CTNNB1*-variant NSMP cases, all were grade 1/2, and 22 were ECPPF-stratified as MLR (15 stage I with \leq 50% MI, 4 with >50% MI, and 3 stage II/III), and 8 were ECPPF-stratified as MHR (all stage I, and 2 with >50% MI). All documented *CTNNB1*variant-associated recurrences occurred in the MHR group (Figure 3D). The 3-year DFS rate for patients with early-stage, low-grade NSMP tumors with *CTNNB1* variations significantly differed between the MLR and MHR groups (*P*<.001) (Figure 3E). ECPPF stratification of NSMP tumors with *ARID1A* and/or *CTNNB1* variations into MLR (n=39) and MHR (n=13) groups showed that all documented recurrences occurred in the MHR group. Consequently, the 3-year DFS rate (95% CI) for the MHR group was 32.1% (12.9%-80.3%), which was significantly lower than that for the MLR group (*P*<.001) (Figure 3F).

ECPPF stratification predicts recurrence in MSI-H/NSMP EC with clinicopathologic low-risk indicators

ECPPF stratification of MSI-H/NSMP EC with clinicopathologic low-risk indicators (n=91) into the MLR and MHR groups (Figure 4A) yielded distinctly diverse clinical outcomes. The 3-year DFS rate (95% CI) was 93.9% (87.4%-100.0%) for the MLR group (n=61) and 43.8% (26.1%-73.6%) for the MHR group (n=30) (P<.001, Figure 4B). The hazard ratio (95% CI) for the MHR group was 12.37 (3.54-43.30) (reference: MLR group, P<.001).

ECPPF stratification and HR variant gene status predict treatment sensitivity in MSI-H/NSMP EC with clinicopathologic highrisk indicators

We next assessed treatment sensitivity according to ECPPF stratification and HR variant gene status (Figure 4C). The 3-year DFS rate (95% CI) was significantly higher for the ECPPF-stratified MLR group (94.1% [83.6%-100.0%]) and the MHR group with HR variant genes (88.9% [70.6%-100.0%]) than that for the ECPPF-stratified MHR group with wild-type HR genes (50.3% [29.8%-84.6%]; P<.001) (Figure 4D).

ECPPF stratification, clinicopathologic indicators, and wild-type HR gene status predict treatment sensitivity in MSI-H/ NSMP EC

Segregating MSI-H and NSMP cases according to clinicopathologic low-risk and high-risk indicators first and then according to ECPPF-stratified MLR, MHR with HR variant gene, and MHR with wild-type HR gene groups may provide a molecularbased strategy for managing EC. Of 26 total documented recurrences, 21 (81%) occurred in the 41 (51%) ECPPF-stratified MHR group tumors with wild-type HR genes, including 14 of 25 (56%) of these tumors with clinicopathologic low-risk indicators.

Integrating ECPPF molecular stratification and treatment algorithms for MSI-H/NSMP EC in a hypothetical but similar population would most likely entail the following molecular-based therapy recommendations: observation for 41% (67% with clinicopathologic low-risk indicators) of patients, RT and/or PbCT for 31% of patients (including 5% with clinicopathologic low-risk indicators), and innovative clinical trials for 28% of patients (including 27% with clinicopathologic low-risk indicators) (Figure 4E).

Discussion

Endometrioid histopathologic characteristics are present in more than 80% of EC tumors, with most classified as MSI-H or NSMP, low stage, and low grade (6, 11, 29). The observation that MSI-H/NSMP EC with clinicopathologic low-risk indicators accounted for most of the MSI-H/NSMP recurrences denotes major limitations in conventional risk assignment strategies. Improving clinical outcomes requires the identification of molecular vulnerabilities to permit target-specific risk designation and choice of therapeutics. Consequently, we assessed our previously described ECPPF molecular stratification system for its ability to resolve MSI-H/NSMP EC prognoses (21).

Because sensitivity to RT and chemotherapeutics are in part predicated on attenuated DNA-damage repair pathway elements (18, 30, 31), we integrated HR variant gene status into our stratification system. Our findings show that ECPPF prognostically stratifies MSI-H and NSMP ECs and that integration of HR variant gene status may facilitate development of new molecular-based therapeutic strategies. The ECPPF-stratified MHR group identified cases at high risk for recurrence in those conventionally classified as clinicopathologic low risk. ECPPF stratification also identified NSMP EC with variant *CTNNB1* as a subgroup with a high risk of recurrence and the MHR group with wild-type HR genes as insensitive to standard adjuvant PbCT and/or RT. In contrast, the ECPPF-stratified MLR group predicted sensitivity to standard therapies (PbCT and/or RT) in EC with clinicopathologic high-risk indicators and/or HR variant genes.

The high TMB associated with MSI-H EC has understandably led to embracing immunotherapy in persistent or recurrent disease (13–16). High TMB also predicts an increased prevalence of HR variant genes and most likely sensitivity to RT and DNA-damaging agents (17). Consistent with this precept, recent studies reported



improved clinical outcomes with RT in patients with MSI-H EC (32, 33). In the current study, ECPPF effectively stratified MSI-H cases, in which patients in the MHR group with early-stage, low-grade tumors, who are rarely candidates for adjuvant therapy, had a poor DFS rate. Moreover, the ECPPF-stratified MLR and MHR groups with HR variant genes and clinicopathologic high-risk indicators had higher DFS rates than did the MHR group with wild-type HR genes and clinicopathologic high-risk indicators.

These observations advocate for strategizing treatment options for MSI-H and NSMP EC tumors with clinicopathologic low-risk indicators according to ECPPF-stratified MLR and MHR groups. Thus, pending HR gene status, conservative surveillance is recommended for the MLR group, and early therapeutic intervention (innovative clinical trials) is recommended for the MHR group. Furthermore, a therapeutic strategy for MSI-H and NSMP EC with clinicopathologic high-risk indicators would consist of treatment with contemporary PbCT and/or RT for patients with ECPPF-stratified MLR tumors and MHR tumors with HR variant genes but innovative clinical trials for those with ECPPF-stratified MHR tumors with wild-type HR genes.

Although NSMP EC is primarily early stage and low grade, it is associated with an intermediate prognosis (6, 8-10). This paradox

suggests that its high recurrence rate is linked to unrecognized highrisk disease in conventionally classified low-risk EC. ECPPF appeared to effectively stratify NSMP EC cases, particularly for early-stage, lowrisk tumors. The MLR group had a distinctly favorable DFS rate over that of the MHR group with occult high-risk disease. *CTNNB1* variations in low-grade NSMP EC are reportedly associated with poor patient outcomes (18, 34, 35). ECPPF efficiently stratified NSMP tumors with variant *CTNNB1*, and all documented recurrences occurred in the MHR group. ECPPF appears to provide a molecular profile for TCGA-designated copy-number low tumors by identifying occult high-risk disease within conventional low-risk cases and high-risk disease that is insensitive to standard adjuvant PbCT and/or RT.

The recently published version of the National Comprehensive Cancer Network guidelines encourages molecular assessment of EC and states that probing for *POLE* hotspot variations and performing immunohistochemical analysis of mismatch repair deficiency and wild-type *TP53* expression may identify MSI-H and NSMP, respectively (36). However, without specific profiling of adverse molecular drivers, targetable vulnerabilities remain unrecognized. Therefore, ECPPF classification provides new profiling insights to refine prognoses and improve therapeutic efficacy.

The predominant challenges for treating MSI-H and NSMP ECs are distinguishing occult high-risk disease in conventionally characterized histomorphologic low-risk EC and predicting insensitivity of EC with clinicopathologic high-risk indicators to contemporary therapies (12). The 43.8% 3-year DFS rate of the ECPPF-stratified MHR group with clinicopathologic low-risk indicators is the first evidence that molecular stratification can distinguish high-risk disease in conventionally classified low-risk MSI-H/NSMP EC. Importantly, 81% of documented recurrences occurred in the ECPPF-stratified MHR group with wild-type HR genes which accounted for only 28% of all TCGA MSI-H/NSMP EC cases. Furthermore, our analysis of MSI-H/NSMP EC with clinicopathologic high-risk indicators showed that the 3-year DFS rate of the MLR group and MHR group with HR variant genes was considerably disparate from that of the MHR group with wild-type HR genes. This finding is consistent with treatment sensitivity to standard therapeutic regimens in the MLR group and MHR group with HR variant genes and treatment insensitivity in the MHR group with wild-type HR genes. These observations introduce a potential molecular-based paradigm shift for precision therapeutics in MSI-H and NSMP ECs. On the basis of our previously reported molecular schematic of disrupted EC signaling pathways (21), reasonable therapeutic options may include modulators of the CCNA2-E2F1-CIP2A axis or PP2A activity and FBXW7 substrates alone or in combination with PbCT or PARP inhibitors (27, 36-40).

Strengths of our analyses include the size of the patient population, centralized pathology review, and availability of robust genomic and transcriptomic databases. Assessing the primary clinical outcome of DFS rather than overall survival is an additional strength of the study that afforded analysis of limitations in contemporary staging, prognostication, and therapeutic efficacy. Limitations of this study included the variability and length of follow-up periods. Because the upper quartile of time to recurrence exceeded the lower quartile of follow-up time, the extent of recurrences in our study population may be underestimated. Recent meta-analyses of prognostic indicators independent of the TCGA signature suggest that lymphovascular space involvement (LVSI) has prognostic value for EC and that deep MI may be a predictor of recurrence (41, 42). LVSI information was not accessible in the TCGA study population and warrants inclusion in subsequent ECPPF stratification studies. In previously reported multivariate analyses incorporating the ECPPF elements with numerous traditional risk factors, deep MI was not an independently significant risk factor (21, 43). Moreover, definitive information in TCGA regarding treatment strategies for the patients was not available. However, because specimens were submitted chiefly from comprehensive cancer centers, it is reasonable to assume that the therapies were consistent with contemporary established guidelines. Such therapies include surgical staging and adjuvant RT and/or chemotherapy in patients with clinically identified at-risk disease; as such, 98% of patients receiving chemotherapy received PbCT (6). Knowledge of the sites of recurrence (e.g., local, regional, or distant) would have also been preferable for this study, particularly for tumors with predominantly clinicopathologic low-risk indicators.

In summary, ECPPF identifies occult high-risk disease in MSI-H/ NSMP EC with clinicopathologic low-risk indicators and appears to efficiently triage treatment sensitivity of MSI-H/NSMP EC with clinicopathologic high-risk indicators to contemporary adjuvant therapies. Consequently, ECPPF stratification confers a new therapeutic paradigm for MSI-H/NSMP EC: 1) observation only for MLR cases with clinicopathologic low-risk indicators; 2) PbCT and/or RT for MHR cases with HR variant genes and clinicopathologic lowrisk indicators, MLR cases with clinicopathologic high-risk indicators, and MHR cases with HR variant genes; and 3) innovative clinical trials for MHR cases with wild-type HR genes and clinicopathologic low-risk or high-risk indicators.

Expanded gene symbols

CCNA2, cyclin A2; CTNNB1, catenin beta 1; E2F1, E2F transcription factor 1; FBXW7, F-box and WD repeat domaincontaining protein 7; POLE, DNA polymerase epsilon, catalytic subunit; PPP2R1A, protein phosphatase 2 scaffold subunit alpha.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article.

Author contributions

Conceptualization: JG-B and KP; Data curation: JG-B and KP; Formal analysis: JG-B, JB-G, BK, KH, and KP; Funding acquisition: SD, AF, and KP; Investigation: JG-B, SW, JB-G, AW, MM, SD, AF, BK, KH, SY, FC, and KP; Methodology: JG-B, BK, KH, and KP; Project administration: KP; Resources: JG-B and KP; Software: JG-B, AW, and MM; Supervision: KP; Pathology: BK, KH, and FC; Writing – original draft: JG-B, JB-G, AW, MM, BK, KH, SY, and KP; Writing – review and editing: JG-B, SW, JB-G, AW, MM, SD, AF, BK, KH, SY, FC, and KP. All authors contributed to the article and approved the submitted version.

Funding

The study was internally funded by Mayo Clinic, which had no involvement in the study design; collection, analysis, and interpretation of data; writing of the report; or decision to submit the article for publication.

Acknowledgments

Nisha Badders, PhD, ELS, Mayo Clinic, substantively edited the manuscript. The Scientific Publications staff at Mayo Clinic

References

1. Bokhman JV. Two pathogenetic types of endometrial carcinoma. *Gynecol Oncol* (1983) 15:10–7. doi: 10.1016/0090-8258(83)90111-7

2. Hamilton CA, Pothuri B, Arend RC, Backes FJ, Gehrig PA, Soliman PT, et al. Endometrial cancer: a society of gynecologic oncology evidence-based review and recommendations. *Gynecol Oncol* (2021) 160:817–26. doi: 10.1016/j.ygyno.2020.12.021

3. Deshpande M, Romanski PA, Rosenwaks Z, Gerhardt J. Gynecological cancers caused by deficient mismatch repair and microsatellite instability. *Cancers (Basel).* (2020) 12. doi: 10.3390/cancers12113319

4. Ashley CW, Da Cruz Paula A, Kumar R, Mandelker D, Pei X, Riaz N, et al. Analysis of mutational signatures in primary and metastatic endometrial cancer reveals distinct patterns of DNA repair defects and shifts during tumor progression. *Gynecol Oncol* (2019) 152:11–9. doi: 10.1016/j.ygyno.2018.10.032

5. Wallbillich JJ, Morris RT, Ali-Fehmi R. Comparing mutation frequencies for homologous recombination genes in uterine serous and high-grade serous ovarian carcinomas: a case for homologous recombination deficiency testing in uterine serous carcinoma. *Gynecol Oncol* (2020) 159:381–6. doi: 10.1016/j.ygyno.2020.08.012

6. Cancer Genome Atlas Research Network, Kandoth C, Schultz N, Cherniack AD, Akbani R, Liu Y, et al. Integrated genomic characterization of endometrial carcinoma. *Nature*. (2013) 497:67–73. doi: 10.1038/nature12113

7. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin (2020) 70:7-30. doi: 10.3322/caac.21590

8. Stelloo E, Nout RA, Osse EM, Jurgenliemk-Schulz IJ, Jobsen JJ, Lutgens LC, et al. Improved risk assessment by integrating molecular and clinicopathological factors in early-stage endometrial cancer-combined analysis of the PORTEC cohorts. *Clin Cancer Res* (2016) 22:4215–24. doi: 10.1158/1078-0432.CCR-15-2878

9. Leon-Castillo A, de Boer SM, Powell ME, Mileshkin LR, Mackay HJ, Leary A, et al. Molecular classification of the PORTEC-3 trial for high-risk endometrial cancer: impact on prognosis and benefit from adjuvant therapy. *J Clin Oncol* (2020) 38:3388–97. doi: 10.1200/JCO.20.00549

10. Pasanen A, Loukovaara M, Ahvenainen T, Vahteristo P, Butzow R. Differential impact of clinicopathological risk factors within the 2 largest ProMisE molecular subgroups of endometrial carcinoma. *PloS One* (2021) 16:e0253472. doi: 10.1371/journal.pone.0253472

11. Kolehmainen A, Pasanen A, Tuomi T, Koivisto-Korander R, Butzow R, Loukovaara M. Clinical factors as prognostic variables among molecular subgroups of endometrial cancer. *PloS One* (2020) 15:e0242733. doi: 10.1371/journal.pone.0242733

12. Jamieson A, Bosse T, McAlpine JN. The emerging role of molecular pathology in directing the systemic treatment of endometrial cancer. *Ther Adv Med Oncol* (2021) 13. 17588359211035959. doi: 10.1177/17588359211035959

13. Eggink FA, Van Gool IC, Leary A, Pollock PM, Crosbie EJ, Mileshkin L, et al. Immunological profiling of molecularly classified high-risk endometrial cancers identifies POLE-mutant and microsatellite unstable carcinomas as candidates for provided proofreading and administrative and clerical support. Portions of the Methods section were previously published in PLoS One. 2021 Jan;16(1):e0245664 and Gynecol Oncol. 2021 Jul;162(1):182-9.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

checkpoint inhibition. Oncoimmunology. (2017) 6:e1264565. doi: 10.1080/2162402X.2016.1264565

14. Marabelle A, Le DT, Ascierto PA, Di Giacomo AM, De Jesus-Acosta A, Delord JP, et al. Efficacy of pembrolizumab in patients with noncolorectal high microsatellite Instability/Mismatch repair-deficient cancer: results from the phase II KEYNOTE-158 study. *J Clin Oncol* (2020) 38:1–10. doi: 10.1200/JCO.19.02105

15. Makker V, Taylor MH, Aghajanian C, Oaknin A, Mier J, Cohn AL, et al. Lenvatinib plus pembrolizumab in patients with advanced endometrial cancer. *J Clin Oncol* (2020) 38:2981–92. doi: 10.1200/JCO.19.02627

16. Makker V, Colombo N, Herráez AC, Santin A, Colomba E, Miller D, et al. A multicenter, open-label, randomized, phase III study to compare the efficacy and safety of lenvatinib in combination with pembrolizumab versus treatment of physician's choice in patients with advanced endometrial cancer. *Gynecologic Oncol* (2021) 162:S4. doi: 10.1016/S0090-8258(21)00657-0

17. Zhao H, Thienpont B, Yesilyurt BT, Moisse M, Reumers J, Coenegrachts L, et al. Mismatch repair deficiency endows tumors with a unique mutation signature and sensitivity to DNA double-strand breaks. *Elife.* (2014) 3:e02725. doi: 10.7554/eLife.02725.041

18. Damia G, Broggini M. Platinum resistance in ovarian cancer: role of DNA repair. *Cancers (Basel).* (2019) 11. doi: 10.3390/cancers11010119

19. Garutti M, Pelizzari G, Bartoletti M, Malfatti MC, Gerratana L, Tell G, et al. Platinum salts in patients with breast cancer: a focus on predictive factors. *Int J Mol Sci* (2019) 20:3390. doi: 10.3390/ijms20143390

20. da Costa A, Baiocchi G. Genomic profiling of platinum-resistant ovarian cancer: the road into druggable targets. *Semin Cancer Biol* (2021) 77:29–41. doi: 10.1016/j.semcancer.2020.10.016

21. Bosquet JG, Zhang Q, Cliby WA, Bakkum-Gamez JN, Cen L, Dowdy SC, et al. Association of a novel endometrial cancer biomarker panel with prognostic risk, platinum insensitivity, and targetable therapeutic options. *PloS One* (2021) 16: e0245664. doi: 10.1371/journal.pone.0245664

22. Gonzalez-Bosquet J, Bakkum-Gamez JN, Weaver AL, McGree ME, Dowdy SC, Famuyide AO, et al. PP2A and E3 ubiquitin ligase deficiencies: seminal biological drivers in endometrial cancer. *Gynecol Oncol* (2021) 162:182–9. doi: 10.1016/j.ygyno.2021.04.008

23. Core Team R. *R: a language and environment for statistical computing* (2013). Available at: https://www.R-project.org.

24. Huber W, Carey VJ, Gentleman R, Anders S, Carlson M, Carvalho BS, et al. Orchestrating high-throughput genomic analysis with bioconductor. *Nat Methods* (2015) 12:115–21. doi: 10.1038/nmeth.3252

25. Bilbao C, Ramirez R, Rodriguez G, Falcon O, Leon L, Diaz-Chico N, et al. Double strand break repair components are frequent targets of microsatellite instability in endometrial cancer. *Eur J Cancer.* (2010) 46:2821–7. doi: 10.1016/j.ejca.2010.06.116

26. Lord CJ, Ashworth A. BRCAness revisited. Nat Rev Cancer. (2016) 16:110–20. doi: 10.1038/nrc.2015.21

27. Heeke AL, Pishvaian MJ, Lynce F, Xiu J, Brody JR, Chen WJ, et al. Prevalence of homologous recombination-related gene mutations across multiple cancer types. *JCO Precis Oncol* (2018) 2018. doi: 10.1200/PO.17.00286

28. de Jonge MM, Auguste A, van Wijk LM, Schouten PC, Meijers M, Ter Haar NT, et al. Frequent homologous recombination deficiency in high-grade endometrial carcinomas. *Clin Cancer Res* (2019) 25:1087–97. doi: 10.1158/1078-0432.CCR-18-1443

29. Bell DW, Ellenson LH. Molecular genetics of endometrial carcinoma. *Annu Rev Pathol* (2019) 14:339–67. doi: 10.1146/annurev-pathol-020117-043609

30. Goldstein M, Kastan MB. The DNA damage response: implications for tumor responses to radiation and chemotherapy. *Annu Rev Med* (2015) 66:129–43. doi: 10.1146/annurev-med-081313-121208

31. Brasseur K, Gevry N, Asselin E. Chemoresistance and targeted therapies in ovarian and endometrial cancers. *Oncotarget.* (2017) 8:4008–42. doi: 10.18632/ oncotarget.14021

32. Reijnen C, Kusters-Vandevelde HVN, Prinsen CF, Massuger L, Snijders M, Kommoss S, et al. Mismatch repair deficiency as a predictive marker for response to adjuvant radiotherapy in endometrial cancer. *Gynecol Oncol* (2019) 154:124–30. doi: 10.1016/j.ygyno.2019.03.097

33. McEachron J, Zhou N, Spencer C, Chatterton C, Shanahan L, Katz J, et al. Adjuvant chemoradiation associated with improved outcomes in patients with microsatellite instability-high advanced endometrial carcinoma. *Int J Gynecol Cancer*. (2021) 31:203–8. doi: 10.1136/ijgc-2020-001709

34. Kurnit KC, Kim GN, Fellman BM, Urbauer DL, Mills GB, Zhang W, et al. CTNNB1 (beta-catenin) mutation identifies low grade, early stage endometrial cancer patients at increased risk of recurrence. *Mod Pathol* (2017) 30:1032–41. doi: 10.1038/modpathol.2017.15

35. De Leo A, de Biase D, Lenzi J, Barbero G, Turchetti D, Grillini M, et al. ARID1A and CTNNB1/beta-catenin molecular status affects the clinicopathologic features and prognosis of endometrial carcinoma: implications for an improved surrogate molecular classification. *Cancers (Basel)* (2021) 13:950. doi: 10.3390/cancers13050950

36. Abu-Rustum N, Yashar C, Arend R, Barber E, Bradley K, Brooks R, et al. Uterine neoplasms, version 1.2023, NCCN clinical practice guidelines in oncology. J Natl Compr Canc Netw (2023) 21:181–209. doi: 10.6004/jnccn.2023.0006

37. Westermarck J. Targeted therapies don't work for a reason; the neglected tumor suppressor phosphatase PP2A strikes back. *FEBS J* (2018) 285:4139–45. doi: 10.1111/ febs.14617

38. Remmerie M, Janssens V. PP2A: a promising biomarker and therapeutic target in endometrial cancer. *Front Oncol* (2019) 9:462. doi: 10.3389/fonc.2019.00462

39. Kim H, Ronai ZA. PRMT5 function and targeting in cancer. Cell Stress. (2020) 4:199–215. doi: 10.15698/cst2020.08.228

40. Lodovichi S, Cervelli T, Pellicioli A, Galli A. Inhibition of DNA repair in cancer therapy: toward a multi-target approach. *Int J Mol Sci* (2020) 21:6684. doi: 10.3390/ ijms21186684

41. Raffone A, Travaglino A, Raimondo D, Neola D, Renzulli F, Santoro A, et al. Prognostic value of myometrial invasion and TCGA groups of endometrial carcinoma. *Gynecol Oncol* (2021) 162:401–6. doi: 10.1016/j.ygyno.2021.05.029

42. Raffone A, Travaglino A, Raimondo D, Neola D, Maletta M, Santoro A, et al. Lymphovascular space invasion in endometrial carcinoma: a prognostic factor independent from molecular signature. *Gynecol Oncol* (2022) 165:192–7. doi: 10.1016/j.ygyno.2022.01.013

43. Gonzalez-Bosquet J, Weroha SJ, Bakkum-Gamez JN, Weaver AL, McGree ME, Dowdy SC, et al. ECPPF (E2F1, CCNA2, POLE, PPP2R1A, FBXW7) stratification: profiling high-risk subtypes of histomorphologically low-risk and treatment-insensitive endometrial cancer. *PloS One* (2022) 17:e0278408. doi: 10.1371/journal.pone.0278408