Check for updates

OPEN ACCESS

EDITED BY Spiros Vlahopoulos, University of Athens, Greece

REVIEWED BY Sudha Sharma, Howard University, United States

Maria S. Pombo-de-Oliveira, National Cancer Institute (INCA), Brazil

*CORRESPONDENCE Yi Yang ⊠yangyi@tjh.tjmu.edu.cn

[†]These authors have contributed equally to this work and share first authorship

SPECIALTY SECTION

This article was submitted to Hematologic Malignancies, a section of the journal Frontiers in Oncology

RECEIVED 10 October 2022 ACCEPTED 02 March 2023 PUBLISHED 14 March 2023

CITATION

Yuan W, Shang Z, Shen K, Yu Q, Lv Q, Cao Y, Wang J and Yang Y (2023) Case report: Germline RECQL mutation potentially involved in hereditary predisposition to acute leukemia. *Front. Oncol.* 13:1066083. doi: 10.3389/fonc.2023.1066083

COPYRIGHT

© 2023 Yuan, Shang, Shen, Yu, Lv, Cao, Wang and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Case report: Germline RECQL mutation potentially involved in hereditary predisposition to acute leukemia

Wei Yuan^{1†}, Zhen Shang^{2,3†}, Kefeng Shen^{2,3}, Qiuxia Yu^{2,3}, Qiuxia Lv^{2,3}, Yang Cao^{2,3}, Jue Wang^{2,3} and Yi Yang^{4*}

¹Department and Institute of Infectious Disease, Tongji Hospital of Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ²Department of Hematology, Tongji Hospital of Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ³Immunotherapy Research Center for Hematologic Diseases of Hubei Province, Tongji Hospital of Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ⁴Department of geriatrics, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

The pathogenesis of acute leukemia is still complex and vague. Most types of acute leukemia are related to somatic gene mutations, and familial incidence is rare. Here we report a case of familial leukemia. The proband presented to our hospital with vaginal bleeding and disseminated intravascular coagulation at the age of 42 and was diagnosed with acute promyelocytic leukemia with typical PML-RAR α fusion gene caused by t(15;17)(q24;q21) translocation. By taking the history, we found that the patient's second daughter had been diagnosed with B-cell acute leukemia with ETV6-RUNX1 fusion gene at age 6. Then we performed whole exome sequencing in peripheral blood mononuclear cells from these two patients at remission status and identified 8 shared germline gene mutations. Using functional annotation and Sanger sequencing validation, we finally focused on a single nucleotide variant in RecQ like helicase (RECQL), rs146924988, which was negative in the proband's healthy eldest daughter. This gene variant potentially led to a relative lack of RECQL protein, disordered DNA repair and chromatin rearrangement, which may mediate the occurrence of fusion genes, as driving factors for leukemia. This study identified a novel possible leukemia-related germline gene variant and provided a new understanding for the screening and pathogenesis of hereditary predisposition syndromes.

KEYWORDS

case report, acute leukemia, germline mutation, hereditary predisposition, fusion gene

Introduction

In addition to the somatic changes related to diagnosis and prognosis in leukemia cells, many germline mutations have been discovered in hematopoietic malignancies in recent years, namely hereditary predisposition (1, 2). The 'Myeloid Neoplasms with Germline

Predisposition' category has been included in the revised fourth edition of the World Health Organization (WHO) classification of tumors of the hematopoietic and lymphoid tissues (3), which further emphasizes the importance of constitutional mutation in the occurrence and development of hematological neoplasms. Fusion genes are hybrid genes formed by the fusion of two previously separated genes, which are resulting from chromosomal rearrangement, including translocation, inversion, deletion, or tandem duplication (4). Fusion genes are molecular biological characteristics of many leukemia types and have been successfully used as diagnostic markers and therapeutic targets (5). However, its relationship with hereditary predisposition is still unclear.

Here, we report a case of familial acute leukemia with fusion genes and germline RecQ Like Helicase (*RECQL*) mutation. The proband was a 42-year-old female patient with acute promyelocytic leukemia (APL) and *PML-RAR* α fusion gene. She has two daughters, and the younger one developed B-cell precursor acute lymphoblastic leukemia (BCP-ALL) with *ETV6-RUNX1* fusion gene at age 6. As of the date of submission, the eldest daughter is still in good health. Comprehensive genetic testing showed that a single nucleotide variant (SNV) in *RECQL*, rs146924988, was potentially involved in this familial case with acute leukemia.

Case presentation

Case 1 (mother)

A previously healthy 42-year-old woman presented to the emergency room due to massive vaginal bleeding in November 2021. Blood routine test showed an abnormal increase of white blood cells (WBC) of $32.83 \times 10^9/L$ and monocytes of $20.19 \times 10^9/L$, with hemoglobin of 100.0g/L and decreased platelet count of $43.0 \times 10^9/L$. Coagulation function test indicated a prolonged prothrombin time of 26.8s, decreased fibrinogen level of 0.93g/L and elevated fibrinogen degradation products of $89.7\mu g/mL$ and D-Dimer greater than $21\mu g/mL$ FEU, suggesting a disseminated intravascular coagulation. An intramural uterine fibroid (approximately 1.8cm*1.3cm) on the anterior wall was found by transabdominal pelvic ultrasonography. Peripheral blood cells smear showed that granulocytes and promyelocytes accounted for 92.00% and 88.00% of the total number of nucleated cells, respectively.

The patient was then admitted to the hematology department for further diagnosis and treatment. Morphological examination of bone marrow (BM) aspirates revealed an abnormal myeloid development with an increased percentage of promyelocytes (approximately 92.00%). BM biopsy analysis revealed marked hyperactive hyperplasia and promyelocytic dysplasia and cytosis (about 90%), and the immunohistochemistry stains for MPO and CD117 were both positive. Flow cytometric immunotyping of BM aspirates showed that abnormal myeloblasts expressed CD117, CD45, cMPO, CD64, CD9, CD33bri, CD13 and CD38, accounting for 83.3% of the total nucleated cells. A few abnormal myeloblasts expressed CD34 (approximately 9.9%), CD11c and CD11b. These abnormal cells were large and had side scatter (SSC) values similar to those of granulocytes. Cytogenetic analysis showed an abnormal karyotype of 46, XX, t(15;17)(q24;q21), del (6) (q22;q32). Molecular genetic analysis detected a typical S-type *PML-RAR* α fusion gene transcript caused by t(15;17)(q24;q21) translocation (approximately 60.30%). Based on these results, this patient was diagnosed as APL (7). After being classified as high-risk group, this patient received all-trans retinoic acid (ATRA, 25 mg·m- $^{2} \cdot d^{-1}$, d1-d3⁾ combined with arsenic trioxide (0.16 mg·kg⁻¹·d⁻¹, d1d3) induction chemotherapy, and continued with three courses of IA (idarubicin 8 mg·m⁻²·d⁻¹, d1-d3; cytarabine 100 mg·m⁻²·d⁻¹, d1d5) consolidation regimen (6). Six times of prophylactic intrathecal chemotherapy were administered during this process to prevent central nervous system relapse. And then ATRA combined with compound Huangdai tablet (CHDT) was used as maintenance chemotherapy. To this date, the patient was still undergoing maintenance therapy and staying in complete remission at the cytological and molecular levels both in bone marrow and cerebrospinal fluid, with no signs or symptoms of APL recurrence.

Case 2 (daughter)

During medical history collection of case 1 (mother), it was found that a 6-year-old girl, the younger one of the patient's two daughters, was diagnosed with BCP-ALL (low-risk group) in March 2019, just two years before the occurrence of her mother's APL (Figure 1A). At the diagnosis, the blast cells in bone marrow were CD10, CD19 and terminal deoxynucleotidyl transferase (TdT) positive detected by fluorescence activated cell sorting (FACS), and with ETV6-RUNX1 fusion gene detected by RT-PCR and t (12;21)(p13;q22) translocation by florescence in situ hybridization (FISH). After receiving a high-dose chemotherapy regimen according to the CCCG-ALL-2015 protocol (8), the daughter achieved complete remission. Up to this date, the daughter still underwent oral maintenance therapy with mercaptopurine (6-MP) and methotrexate (MTX) and remained in minimum residual disease (MRD) negative remission at cellular and molecular (ETV6-RUNX1) levels.

To explore the possible hereditary predisposition in this case, we performed whole exome sequencing (WES) in peripheral blood mononuclear cells (PBMCs) from these two patients at remission status with informed consents. The genomic DNA of each sample was sheared with Biorupter (Diagenode, Belgium) and 150 ~ 200bp fragments were obtained. AIExomeV1 Enrichment Kit (iGeneTech, Beijing, China) and Illumina platform (Illumina, San Diego, CA) were applied to capture and sequence the whole exons. Then clean reads were mapped to the reference genome GRCh37 by using Burrow-Wheeler Aligner. After removing low quality reads and duplications, SNV and insertion-deletion (InDel) were called and annotated by using Genome Analysis Toolkit (GATK). Finally, a total of 8 germline gene mutations shared by both patients were identified (Table 1), including 2 InDel and 6 SNV, in which an SNV in RECQL (rs146924988) was noticed by functional annotation. Sanger sequencing of nasopharyngeal swab specimens was used to verify WES results, and to screen the other healthy child of the



A brief summary of medical history and pedigree for the *RECQL* mutation carriers. (A) Timeline of clinical events in 2 patients with acute leukemia. (B) The Pedigree and germline and tumor chromatopherogram confirmed the presence of single nucleotide variation in *RECQL* (rs146924988) in 2 patients with acute leukemia in this family, while it was negative in healthy members. (C) Sanger sequencing of nasopharyngeal swab specimens was used to verify *RECQL* mutation (rs146924988) in the proband and her two daughters. APL, acute promyelocytic leukemia; WES, whole exome sequencing; BCP-ALL, B-cell precursor acute lymphoblastic leukemia; Mut, mutant; WT, wide type.

proband without *RECQL* mutation detected. The pedigree for rs146924988 and leukemia was shown in Figures 1B, C.

Patient perspective

I have an entire family, including a husband and two daughters. I led a busy and happy life, struggling to make ends meet, until my little daughter and I were diagnosed with leukemia in turn. Before I came to Tongji Hospital, I thought my life was coming to an end. Fortunately, after receiving systematic treatment, my daughter and I were cured and there is no sign of recurrence. Our life and work are gradually returning to normal.

Discussion

With the development of genomic sequencing methods, many germline mutations have been found in cancer patients and their immediate relatives. Germline mutations associated with the hereditary predisposition to cancers were highlighted for their clinical relevance and diagnostic values, which were also known as hereditary predisposition syndromes (HPS) (9). For hematological malignancies, hereditary predisposition was first identified in chronic lymphocytic leukemia (CLL) and acute myeloid leukemia (AML), which has led to the identification of several germline mutations, such as *RUNX1*, *CEBPA*, *GATA2*, *ANKRD26*, *DDX41* and *ETV6* mutations. Furthermore, genetic susceptibility was indicated in ALL and

TABLE 1 Summary of shared germline gene mutations at rem	nission status.
--	-----------------

Gene	Vcf_mut	AAChange.HGVS	FRE (case1/case2)	Mut_type	avsnp150
RECQL	chr12:21652503:A/G	c.2T>C:p.0?	45.1%/42.9%	SNP	rs146924988
AK2	chr1:33478892:G/A	c.586C>T:p.(Arg196Trp)	50.5%/49.6%	SNP	rs370429097
C7	chr5:40972681:C/T	c.2059C>T:p.(Arg687Cys)	47.8%/45.9%	SNP	rs117487879
CCDC40	chr17:78069197:G/A	c.2968G>A:p.(Asp990Asn)	50%/50.5%	SNP	rs200958035
NCF4	chr22:37273779:G/T	c.934G>T:p.(Gly312Cys)	46.9%/39.0%	SNP	rs199618052
KCNQ4	chr1:41284190:C/G	c.546C>G:p.(Phe182Leu)	52.7%/41.7%	SNP	rs80358273
GJB2	chr13:20763485:AG/A	c.235del:p.(Leu79Cysfs*3)	47.4%/54.9%	InDel	rs80338943
USH2A	chr1:216595579:G/GA	c.99_100insT:p.(Arg34Serfs*41)	49.0%/41.1%	InDel	rs141672841

Mut, mutation; FRE, frequency; InDel, insertion-deletion; SNP, single nucleotide polymorphism.

multiple myeloma, with constitutional mutations in genes such as IKZF1, SH2B3, PAX5 (ALL) and KDM1A/LSD1 (multiple myeloma). Monogenic genomic alterations have also been found to be involved in inborn errors of the immune system and confer a predisposition to hematological malignancies such as ataxia telangiectasia, Nijmegen breakage syndrome, Bloom's syndrome, xeroderma pigmentosum, constitutional mismatch repair deficiency, Fanconi anemia, and telomere syndromes (2, 10-12). Interestingly, previously identified inherited cancer susceptibility syndromes also increase the possibility to develop hematological malignancies. For example, patients with Diamond-Blackfan anemia, severe congenital neutropenia, or familial thrombocytopenia have a significantly increased incidence of myeloid neoplasms (10). Up to 62% of children with lymphoproliferative diseases have primary immune deficiency disease (PID) (13). Compared with the general population, people with PID have a 1.5 times higher risk of developing cancer (14). At present, HPS-related genes or pathways involve transcription factors, RNA function, DNA methylation, ribosome assembly, histone modification, DNA unwinding and DNA repair (9). Other germline mutations in key transcription and translation processes and signal transduction pathways, such as the RAS pathway, cell proliferation, apoptosis, and tumor suppressor genes (e.g., TP53) were also considered as possible underlying predispositions.

In this case, the proband and her second daughter both suffered from acute leukemia at different times in their lives. Considering the incidence of acute leukemia, we speculated that there might be HPS in this case. Through WES sequencing and bioinformatics analysis, we finally focused on rs146924988, an SNV in *RECQL*, and the mutation frequencies in the proband and her second daughter were 45.1% and 42.9%, respectively. Further Sanger sequencing showed no such mutation of *RECQL* in the healthy child of the proband, which indicated that rs146924988 might play a role in this case of familial leukemia. The proband of this case was an APL patient, carrying a typical *PML-RAR* α fusion gene. The second daughter of the proband was a BCP-ALL patient, carrying the classical *ETV6-RUNX1* fusion gene. Considering the driving role of these two different fusion genes in the occurrences and development of corresponding leukemias (15, 16), it can be inferred that this RECQL gene variant (rs146924988) is potentially associated with the generation of acute leukemia. We evaluated the alteration frequency and expression levels of RECQL in acute leukemia from publicly available datasets using the cBioPortal database (17), including AML datasets from The Cancer Genome Atlas (TCGA) (18) and B-ALL datasets generated by the Therapeutically Applicable Research to Generate Effective Treatments (TARGET, https://ocg.cancer.gov/programs/target) initiative, phs000464. Samples with complete data of gene mutations, copy-number alterations (CNA) and mRNA expressions were retained after screening. The CNA frequency of RECQL was more than 5% both in ALL and AML patients, and the main variants types are deep deletion and shallow deletion that related to low gene expression (Figures 2A, B). Interestingly, the expression of RECQL in AML-M3 patients mostly having PML-RAR fusion gene, was significantly lower than that in other types of AML patients (Figure 2C). These findings highlighted the potential role of RECQL in leukemia.

RECQL gene, as a member of the RecQ helicase family located on chromosome 12p12, encodes a DNA helicase with 649 residues. RECQL protein is a highly conserved protein and plays an important role in DNA repair, replication, recombination and transcription (19). The SNV (rs146924988) detected in this case is an initiator codon variant of RECQL, which may lead to start-lost loss-of-function of RECQL (20, 21) (Figures 3A, B). Prior to this report, the RECQL germline mutation has been found to be associated with hereditary breast cancer and hereditary ovarian cancer syndrome (22, 23), but its role in hematological malignancies has not been reported. RECQL protein is an ATPase and helicase that can bind and unlock the structural intermediate of DNA replication and repair. RECQL unlocks double-stranded DNA and catalyzes the migration of ATP-dependent branches on Holliday junctions and removable D-loop substrates. In addition to cleavage of DNA, RECQL promotes annealing of complementary single-stranded DNA in an ATP-independent manner (24). Moreover, RECQL can also interact with proteins that are involved in DNA replication, repair and mismatch repair, such as FEN1, RPA, PARP1, and MLH1 (25-27). Deletion of RECQL



FIGURE 2

Mutation and expression of *RECQL* potentially involved in acute leukemia. The alteration frequency in *RECQL* in acute leukemia was analyzed by using the cBioPortal database, including data of AML from TCGA database and data of BCP-ALL from TARGET database. Samples with complete data of gene mutation, CNA and mRNA expression (log RNA Seq V2 RSEM) were retained after screening. (A) The alteration frequency in *RECQL* in B-ALL patients (Pediatric Acute Lymphoid Leukemia - Phase II, TARGET). (B) The alteration frequency in *RECQL* in AML patients (TCGA-LAML). (C) Comparison of *RECQL* gene expression in AML patients with different French American British classification. AML, Acute Myeloid Leukemia; TCGA, The Cancer Genome Atlas; B-ALL, acute B lymphoblastic leukemia; CNA, copy-number alterations; RSEM, RNA-Seq by Expectation Maximization; TARGET, therapeutically applicable research to generate effective treatments. Statistically significant differences are indicated as follows: *p < 0.05, **p < 0.01, ***p < 0.01.



FIGURE 3

A theoretical model that *RECQL* mutation (rs146924988) acts as a potential mediator of hereditary leukemia. (A) Variant details of single nucleotide variation in *RECQL* (rs146924988). Dotted box indicates the corresponding DNA sequence of the initiator codon. (B) Hypothetical mechanism that *RECQL* mutation (rs146924988) affecting transcription and translation. The rs146924988 mutation is an initiator codon variant of *RECQL*, which may lead to a start-lost loss-of-function of RECQL. (C) A theoretical model that rs146924988 acts as a mediator of hereditary leukemia. The germline mutation (rs146924988) was proposed to mediate the occurrence of chromosome rearrangement and fusion genes, as well as affect chemotherapy sensitivity. WT, wide type; Mut, mutant; DSB, DNA double-strand break.

resulted in increased DNA damage accumulation, which indicated that RECQL was involved in the solution of replication fork stagnation. RECQL-deficient mice and cells exhibited frequent spontaneous chromosomal breakage and chromatid translocation (28). In a recent in-vitro experiment based on MDA-MB-231 cells, RECQL was considered to be involved in regulating cell stress response (29). It has been found to mediate the recruitment of replication protein A (RPA) in DNA damage sites and to promote the activation of the ATR-Chk1 pathway in response to the DNA damage caused by gemcitabine (29). ATR-Chk1 is a classic pathway of DNA damage response (DDR) induced by DNA double-strand breaks (DSBs), involving cell cycle checkpoint regulation and DNA repair (30). It can promote ubiquitination and proteasome degradation by phosphorylating CDC25 phosphatases (CDC25A, B, C), and induce cell cycle G1-S transition and G2-M transition (31-33). ATR-Chk1 dysfunction is associated with a large number of DNA damage and chromosomal rearrangements in Ataxiatelangiectasia (A-T) patients, especially in lymphocytes, which often leads to the production of fusion genes (34, 35).

Based on these previous findings, we hypothesize that the constitutional mutation in *RECQL* gene (rs146924988) may inhibit the translation of *RECQL* mRNA and lead to a relative lack of RECQL protein, which may further impair the ATR-Chk1 pathway and resulting in functional disorders in DNA replication and DSBs repair. It is conceivable that these functional disorders could ultimately mediate the occurrence of chromosome rearrangement and fusion genes, as driving factors for leukemia (Figure 3C). Admittedly, we have not obtained direct evidence that *RECQL* gene

mutation (rs146924988) will lead to decreased RECQL expression level because this study is a case report in nature, and the biological process of such mutation potentially affecting DSBs repair and mediating the production of fusion genes needs to be verified in in-vitro experiments and larger clinical cohorts in the future. Moreover, there were 7 other shared germline mutations in this case of familial leukemia, which were related to genes AK2, C7, CCDC40, NCF4, KCNQ4, GJB2 and USH2A. These genes were reported to be involved in ADP biosynthesis, complement activation, oxidation-reduction and other biological processes, and their functional annotations were shown in Table S1. By checking the literature and public data repository, there is no direct evidence that these mutations are associated with genomic instability or genetic predisposition to cancer, but their contributions to the onset of leukemia cannot be completely excluded and needs further evaluation. The loss of RECQL function may increase the probability of the somatic events, such as the occurrence of chromosome rearrangements, in positions given by chance and modulated by other factors, resulting in different genetic fusions genes. This partly explained how germline genetic variants in RECQL lead to different types of fusions gene in these 2 cases.

Early recognition of the hereditary predisposition of hematological malignancies is critical for timely diagnosis and individualized treatment. In this study, a potential HPS-related gene mutation was identified by WES sequencing, which highlights the clinical application of WES, especially in patients with earlyonset leukemia or familial leukemia. In addition to diagnostic value, screening of the HPS-related gene mutations may provide prognosis evaluation reference for an optimal intervention approach. For example, previous studies have shown that *RECQL* gene defects not only cause genomic instability and chromatin recombination, but also make cells more sensitive to toxic genomic stress and cytotoxic therapy (29). Particularly, polymorphic variants in *RECQL* was found to be related to the overall survival rate of pancreatic cancer patients treated with gemcitabine (36). RECQL helicase has also been found to protect multiple myeloma cells from melphalan and bortezomib cytotoxicity (37). Therefore, the significance of *RECQL* mutation in the treatment of acute leukemia needs to be evaluated in future studies.

In this study, we performed WES sequencing in a familial leukemia case, and for the first time to our knowledge, found a germline *RECQL* mutation potentially involved in hereditary predisposition to acute leukemia. The hypothetical biological process in which *RECQL* gene mutation (rs146924988) affect DSBs repair and mediate the generation of fusion genes provides a new understanding of the pathogenesis of leukemia, and highlights the necessity for next-generation sequencing-based screening of genes involved in this process in potential HPS patients.

Data availability statement

The datasets presented in this article are not readily available because of ethical/privacy restrictions. Requests to access the datasets should be directed to the corresponding author.

Ethics statement

This study was approved by the Medical Ethics Committee of the Department of Hematology, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

Author contributions

All authors designed the study, interpreted the findings and revised the manuscript. WY, ZS, KS, QY, and QL carried out data management and statistical analysis and drafted the manuscript. YC

References

1. Pui CH, Nichols KE, Yang JJ. Somatic and germline genomics in paediatric acute lymphoblastic leukaemia. *Nat Rev Clin Oncol* (2019) 16(4):227–40. doi: 10.1038/ s41571-018-0136-6

2. Bochtler T, Haag GM, Schott S, Kloor M, Krämer A, Müller-Tidow C. Hematological malignancies in adults with a family predisposition. *Dtsch Arztebl Int* (2018) 115(50):848–54. doi: 10.3238/arztebl.2018.0848

3. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, et al. The 2016 revision to the world health organization classification of myeloid neoplasms and acute leukemia. *Blood* (2016) 127(20):2391–405. doi: 10.1182/blood-2016-03-643544

4. Mertens F, Johansson B, Fioretos T, Mitelman F. The emerging complexity of gene fusions in cancer. *Nat Rev Cancer*. (2015) 15(6):371-81. doi: 10.1038/nrc3947

and JW helped with English language editing and data management. YY performed project administration. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by the National Natural Science Foundation of China No.82171575 and the National Key Research and Development Program of China 2021YFA1101504.

Acknowledgments

The authors would like to thank the patients and their families for agreeing to report this case, and all the faculty and staff in our clinical and laboratory unit for clinical and technical support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2023.1066083/ full#supplementary-material

5. Papaemmanuil E, Gerstung M, Bullinger L, Gaidzik VI, Paschka P, Roberts ND, et al. Genomic classification and prognosis in acute myeloid leukemia. *N Engl J Med* (2016) 374(23):2209–21. doi: 10.1056/NEJMoa1516192

6. Chinese Society of Hematology, Chinese Medical Doctor Association, Chinese Medical Association and Chinese Medical Doctor Association. Chinese Guidelines for diagnosis and treatment of acute promyelocytic leukemia. *Zhonghua Xue Ye Xue Za Zhi* (2018) 39(3):179–83. doi: 10.3760/cma.j.issn.0253-2727.2018.03.002

7. Sanz MA, Fenaux P, Tallman MS, Estey EH, Löwenberg B, Naoe T, et al. Management of acute promyelocytic leukemia: updated recommendations from an expert panel of the European LeukemiaNet. *Blood* (2019) 133(15):1630–43. doi: 10.1182/blood-2019-01-894980 8. Yang W, Cai J, Shen S, Gao J, Yu J, Hu S, et al. Pulse therapy with vincristine and dexamethasone for childhood acute lymphoblastic leukaemia (CCCG-ALL-2015): an open-label, multicentre, randomised, phase 3, non-inferiority trial. *Lancet Oncol* (2021) 22(9):1322–32. doi: 10.1016/S1470-2045(21)00328-4

9. Garber JE, Offit K. Hereditary cancer predisposition syndromes. J Clin Oncol (2005) 23(2):276–92. doi: 10.1200/JCO.2005.10.042

10. Mangaonkar AA, Patnaik MM. Hereditary predisposition to hematopoietic neoplasms: When bloodline matters for blood cancers. *Mayo Clin Proc* (2020) 95 (7):1482–98. doi: 10.1016/j.mayocp.2019.12.013

11. Bakry D, Aronson M, Durno C, Rimawi H, Farah R, Alharbi QK, et al. Genetic and clinical determinants of constitutional mismatch repair deficiency syndrome: report from the constitutional mismatch repair deficiency consortium. *Eur J Cancer.* (2014) 50(5):987–96. doi: 10.1016/j.ejca.2013.12.005

12. Oetjen KA, Levoska MA, Tamura D, Ito S, Douglas D, Khan SG, et al. Predisposition to hematologic malignancies in patients with xeroderma pigmentosum. *Haematologica* (2020) 105(4):e144-6. doi: 10.3324/haematol. 2019.223370

13. Forbes LR, Eckstein OS, Gulati N, Peckham-Gregory EC, Ozuah NW, Lubega J, et al. Genetic errors of immunity distinguish pediatric nonmalignant lymphoproliferative disorders. *J Allergy Clin Immunol* (2022) 149(2):758-66. doi: 10.1016/j.jaci.2021.07.015

14. Ballow M, Sánchez-Ramón S, Walter JE. Secondary immune deficiency and primary immune deficiency crossovers: Hematological malignancies and autoimmune diseases. *Front Immunol* (2022) 13:928062. doi: 10.3389/fimmu.2022.928062

15. Tan Y, Wang X, Song H, Zhang Y, Zhang R, Li S, et al. A PML/RARα direct target atlas redefines transcriptional deregulation in acute promyelocytic leukemia. *Blood* (2021) 137(11):1503–16. doi: 10.1182/blood.2020005698

16. Nishii R, Baskin-Doerfler R, Yang W, Oak N, Zhao X, Yang W, et al. Molecular basis of ETV6-mediated predisposition to childhood acute lymphoblastic leukemia. *Blood* (2021) 137(3):364–73. doi: 10.1182/blood.2020006164

17. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal* (2013) 6(269):11. doi: 10.1126/scisignal.2004088

18. Cancer Genome Atlas Research Network, Ley TJ, Miller C, Ding L, Raphael BJ, Mungall AJ, et al. Genomic and epigenomic landscapes of adult *de novo* acute myeloid leukemia. *N Engl J Med* (2013) 368(22):2059–74. doi: 10.1056/NEJMoa1301689

19. Veith S, Mangerich A. RecQ helicases and PARP1 team up in maintaining genome integrity. Ageing Res Rev (2015) 23(Pt A):12–28. doi: 10.1016/j.arr.2014.12.006

20. Wang X, Liu Z, Li G, Dang L, Huang S, He L, et al. Efficient gene silencing by adenine base Editor-mediated start codon mutation. *Mol Ther* (2020) 28(2):431–40. doi: 10.1016/j.ymthe.2019.11.022

21. Mengel D, Traschütz A, Reich S, Leyva-Gutiérrez A, Bender F, Hauser S, et al. A *de novo* STUB1 variant associated with an early adult-onset multisystemic ataxia phenotype. *J Neurol* (2021) 268(10):3845–51. doi: 10.1007/s00415-021-10524-7

22. Cybulski C, Carrot-Zhang J, Kluźniak W, Rivera B, Kashyap A, Wokołorczyk D, et al. Germline RECQL mutations are associated with breast cancer susceptibility. *Nat Genet* (2015) 47(6):643–6. doi: 10.1038/ng.3284

23. Kaneyasu T, Mori S, Yamauchi H, Ohsumi S, Ohno S, Aoki D, et al. Prevalence of disease-causing genes in Japanese patients with BRCA1/2-wildtype hereditary breast and ovarian cancer syndrome. *NPJ Breast Cancer* (2020) 6:25. doi: 10.1038/s41523-020-0163-1

24. Sharma S, Sommers JA, Choudhary S, Faulkner JK, Cui S, Andreoli L, et al. Biochemical analysis of the DNA unwinding and strand annealing activities catalyzed by human RECQ1. J Biol Chem (2005) 280(30):28072–84. doi: 10.1074/jbc.M500264200

25. Sami F, Lu X, Parvathaneni S, Roy R, Gary RK, Sharma S. RECQ1 interacts with FEN-1 and promotes binding of FEN-1 to telomeric chromatin. *Biochem J* (2015) 468 (2):227–44. doi: 10.1042/BJ20141021

26. Doherty KM, Sharma S, Uzdilla LA, Wilson TM, Cui S, Vindigni A, et al. RECQ1 helicase interacts with human mismatch repair factors that regulate genetic recombination. *J Biol Chem* (2005) 280(30):28085–94. doi: 10.1074/jbc.M500265200

27. Sharma S, Phatak P, Stortchevoi A, Jasin M, Larocque JR. RECQ1 plays a distinct role in cellular response to oxidative DNA damage. *DNA Repair (Amst)* (2012) 11 (6):537–49. doi: 10.1016/j.dnarep.2012.04.003

28. Sharma S, Stumpo DJ, Balajee AS, Bock CB, Lansdorp PM, Brosh RM, et al. RECQL, a member of the RecQ family of DNA helicases, suppresses chromosomal instability. *Mol Cell Biol* (2007) 27(5):1784–94. doi: 10.1128/MCB.01620-06

29. Parvathaneni S, Sharma S. The DNA repair helicase RECQ1 has a checkpointdependent role in mediating DNA damage responses induced by gemcitabine. *J Biol Chem* (2019) 294(42):15330–45. doi: 10.1074/jbc.RA119.008420

30. Sørensen CS, Syljuåsen RG. Safeguarding genome integrity: the checkpoint kinases ATR, CHK1 and WEE1 restrain CDK activity during normal DNA replication. *Nucleic Acids Res* (2012) 40(2):477–86. doi: 10.1093/nar/gkr697

31. Kastan MB, Bartek J. Cell-cycle checkpoints and cancer. *Nature* (2004) 432 (7015):316–23. doi: 10.1038/nature03097

32. Matthews HK, Bertoli C, de Bruin RAM. Cell cycle control in cancer. Nat Rev Mol Cell Biol (2022) 23(1):74–88. doi: 10.1038/s41580-021-00404-3

33. Karlsson-Rosenthal C, Millar JBA. Cdc25: mechanisms of checkpoint inhibition and recovery. *Trends Cell Biol* (2006) 16(6):285–92. doi: 10.1016/j.tcb.2006.04.002

34. Taylor AM, Metcalfe JA, Thick J, Mak YF. Leukemia and lymphoma in ataxia telangiectasia. Blood (1996) 87(2):423–38. doi: 10.1182/blood.V87.2.423.bloodjournal872423

35. Yamauchi M. Mechanisms underlying the suppression of chromosome rearrangements by ataxia-telangiectasia mutated. *Genes* (2021) 12(8):1232. doi: 10.3390/genes12081232

36. Li D, Frazier M, Evans DB, Hess KR, Crane CH, Jiao L, et al. Single nucleotide polymorphisms of RecQ1, RAD54L, and ATM genes are associated with reduced survival of pancreatic cancer. *J Clin Oncol* (2006) 24(11):1720–8. doi: 10.1200/JCO.2005.04.4206

37. Viziteu E, Klein B, Basbous J, Lin YL, Hirtz C, Gourzones C, et al. RECQ1 helicase is involved in replication stress survival and drug resistance in multiple myeloma. *Leukemia* (2017) 31(10):2104–13. doi: 10.1038/leu.2017.54