frontiers | Frontiers in Oncology

CORRECTION
published: 21 June 2022
doi: 10.3389/fonc.2022.927917

Corrigendum: Anti-Tumoral Effect and Action Mechanism of Exosomes Derived From *Toxoplasma gondii*-Infected Dendritic Cells in Mice Colorectal Cancer

Shilan Zhu[1,2], Jinmiao Lu[1,2], Zhibing Lin[3], Asmaa M. I. Abuzeid[1,4], Xiaoyu Chen[1,2], Tingting Zhuang[1], Haiyan Gong[2], Rongsheng Mi[2], Yan Huang[2], Zhaoguo Chen[2]* and Guoqing Li[1]**

[1] Guangdong Provincial Key Laboratory of Zoonosis Prevention and Control, College of Veterinary Medicine, South China Agricultural University, Guangzhou, China, [2] Key Laboratory of Animal Parasitology of Ministry of Agriculture, Laboratory of Quality and Safety Risk Assessment for Animal Products on Biohazards (Shanghai) of Ministry of Agriculture, Shanghai Veterinary Research Institute, Chinese Academy of Agricultural Sciences, Shanghai, China, [3] School of Agriculture and Biology, Shanghai Jiao Tong University, Shanghai, China, [4] Faculty of Veterinary Medicine, Suez Canal University, Ismailia, Egypt

OPEN ACCESS

Edited and reviewed by:
Anil Shanker,
Meharry Medical College,
United States

***Correspondence:**
Guoqing Li
gqli@scau.edu.cn
Zhaoguo Chen
zhaoguochen@shvri.ac.cn
Zhibing Lin
mdzhibing@163.com

Specialty section:
This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Oncology

Received: 25 April 2022
Accepted: 31 May 2022
Published: 21 June 2022

Citation:
Zhu S, Lu J, Lin Z, Abuzeid AMI, Chen X, Zhuang T, Gong H, Mi R, Huang Y, Chen Z and Li G (2022) Corrigendum: Anti-Tumoral Effect and Action Mechanism of Exosomes Derived From Toxoplasma gondii-Infected Dendritic Cells in Mice Colorectal Cancer. Front. Oncol. 12:927917. doi: 10.3389/fonc.2022.927917

Keywords: ***Toxoplasma gondii***, **dendritic cells, exosome, miRNA, macrophage, miR-155-5p**

A Corrigendum on:

Anti-Tumoral Effect and Action Mechanism of Exosomes Derived From Toxoplasma gondii-Infected Dendritic Cells in Mice Colorectal Cancer

By Zhu S, Lu J, Lin Z, Abuzeid AMI, Chen X, Zhuang T, Gong H, Mi R, Huang Y, Chen Z and Li G (2022). Front. Oncol. 12:870528. doi: 10.3389/fonc.2022.870528

In the original article, there was a mistake in **Figure 2** as published. 1. **Figure 2A** (left) missed to indicate the corresponding groups for the panels. The corresponding groups are now correctly indicated. 2. **Figure 2B**: The representative flow cytometry plot was repeated two times (same flow cytometry image shown in the third column). The flow cytometry image are the results of flow cytometry detection of anti-CD86-PC7 in the Me49 group.We replaced the lower flow cytometry image in the third column with a representative flow-cytometry plot of anti-CD206-APC in the Me49 group. The corrected **Figure 2** appears below.

The authors apologize for this error and state that this does not change the scientific conclusions of the article in any way. The original article has been updated.

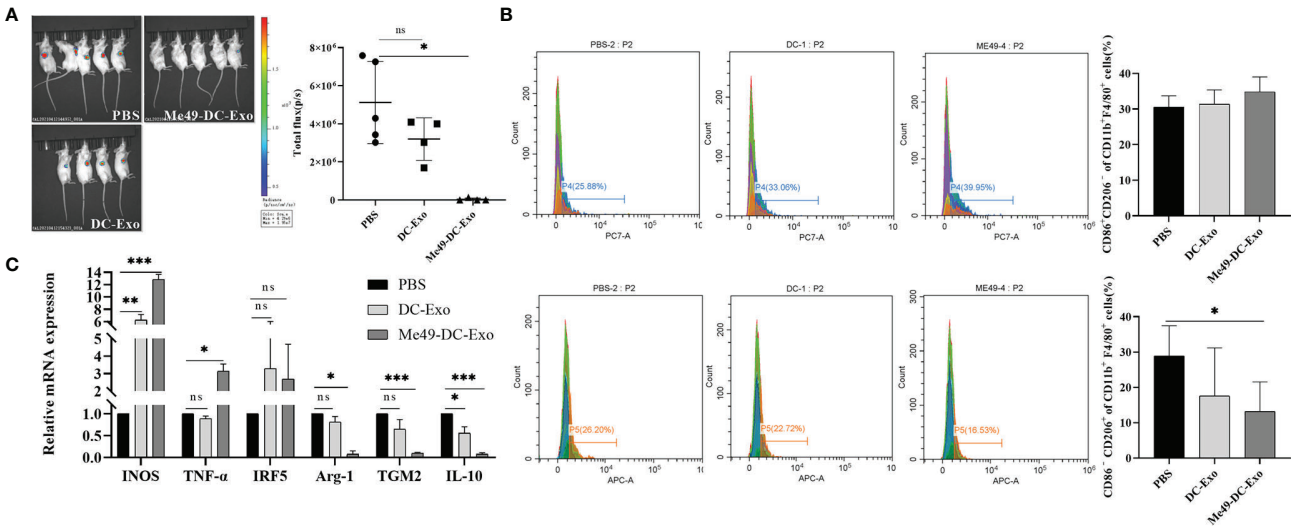

FIGURE 2 | Me49-DC-Exo inhibited tumor growth in mouse and regulated macrophage polarization. **(A)** On day 4 after treatment, the IVIS imager detected bioluminescence images in tumors of mouse and quantified the bioluminescence signal intensity of each tumor in mouse. **(B)** Flow cytometry was used to label CD86 + or CD206 +, and CD45 + CD11b + F4/80 + macrophages in blood of mice injected with DC-Exo and Me49-DC-Exo were stained to detect the percentage of CD86+ CD206 – M1 macrophages and CD86 – CD206 + M2 macrophages. **(C)** mRNA levels of M1 macrophage specific genes (INOS, TNF-a, and IRF5) and M2 macrophage specific genes (TGM2, Arg-1 and IL10) in blood of tumor-bearing mice injected intratumorally with PBS, DC-Exo and Me49-DC-Exo were detected by qRT-PCR. The data were expressed as mean ± standard deviation, and the independent sample t-test was used to compare the statistical differences between two groups. ns ($p \geq 0.05$), *$p < 0.05$, **$p < 0.01$, ***$p < 0.001$.