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SPECIALTY SECTION

This article was submitted to
Cancer Molecular Targets
and Therapeutics,
a section of the journal
Frontiers in Oncology

RECEIVED 21 April 2022

ACCEPTED 27 June 2022

PUBLISHED 15 July 2022

CITATION

Cheng X, Lou K, Ding L, Zou X,
Huang R, Xu G, Zou J and Zhang G
(2022) Clinical potential of the Hippo-
YAP pathway in bladder cancer.
Front. Oncol. 12:925278.
doi: 10.3389/fonc.2022.925278

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Clinical potential of the Hippo-YAP pathway in bladder cancer

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Bladder cancer (BC) is one of the world's most frequent cancers. Surgery coupled with adjuvant platinum-based chemotherapy is the current standard of therapy for BC. However, a high proportion of patients progressed to chemotherapy-resistant or even neoplasm recurrence. Hence, identifying novel treatment targets is critical for clinical treatment. Current studies indicated that the Hippo-YAP pathway plays a crucial role in regulating the survival of cancer stem cells (CSCs), which is related to the progression and reoccurrence of a variety of cancers. In this review, we summarize the evidence that Hippo-YAP mediates the occurrence, progression and chemotherapy resistance in BC, as well as the role of the Hippo-YAP pathway in regulating bladder cancer stem-like cells (BCSCs). Finally, the clinical potential of Hippo-YAP in the treatment of BC was prospected.

KEYWORDS

bladder cancer, Hippo pathway, YAP, chemoresistant, cancer stem cell

Introduction

Worldwide, BC is the 11th most common malignancy, with more than 570,000 new cases and 210,000 deaths in 2020 (1), and the incidence is increasing (2). BC is divided into nonmuscle-invasive bladder cancer (NMIBC) and muscle-invasive bladder cancer (MIBC). NMIBC is less malignant, and the routine treatment is based on TURBT (transurethral resection of bladder tumor) combined with bladder perfusion chemotherapy or immunotherapy (3). MIBC is more aggressive, and the classical treatment is radical cystectomy combined with platinum-based chemotherapeutic (4). The preferred treatment for metastatic MIBC is platinum-based chemotherapy. In cisplatin-ineligible patients, immunotherapy is preferred for PD-L1-positive patients,

and carboplatin is chosen as an alternative therapy for PD-L1-negative patients (4, 5). Unfortunately, even with the tremendous efforts of current research on BC, the mortality rate of BC patients is still high (1). The most important factor affecting the prognosis of BC patients is that a large proportion of patients relapse after the first treatment for BC and are resistant to existing treatment regimens (6), with no effective therapeutic target to date (7, 8). Therefore, it is necessary to further investigate the mechanisms of BC pathogenesis, recurrence and drug resistance, and to screen effective targeted drugs for the treatment of advanced metastatic BC.

The Hippo-YAP signaling pathway plays a key role in stem cells and cancer cells (9, 10). The Hippo pathway, first identified in *Drosophila melanogaster*, has a role in regulating organ size (11) and is conserved in a variety of species, including humans (12). It is an important regulator of organ development, cell proliferation, dynamic balance, and regeneration (10, 13). Extracellular matrix, nutrition, cell density, cell polarity, mechanical transduction, and G protein-coupled receptors are all factors that regulate the Hippo-YAP pathway (14–17). The cytoplasmic kinase cascade and the nuclear transcription module are the two primary components of the Hippo-YAP pathway. The Hippo-kinase cascade is mainly composed of MAP4K, MST1/2, and LATS1/2 (18, 19). The nuclear transcriptional module of the Hippo pathway is a transcriptionally active motif with oncogenic effects composed

of YAP (yes-associate protein), TAZ (transcriptional co-activators with PDZ binding sequences), and TEAD-1 (TEA domain family member 1), which are mainly regulated by the Hippo-kinase cascade (Figure 1). YAP/TAZ has a dominant role in numerous solid tumors (9, 13, 17, 41, 42), and increasing significance of elevated YAP/TAZ activity in BC (43).

In this review, we summarized the evidence that YAP would be a promising therapeutic target, regarding the association of YAP with BC onset, progression, postoperative recurrence, chemoresistance, and metastasis. In addition, we emphasized the role of the Hippo-YAP pathway in regulating BCSCs (bladder cancer stem-like cells), as well as the hitherto unanswered question that how the nuclear transcriptional module of the Hippo pathway is over-activated in BC. At last, the clinical potential and pharmacology direction of Hippo-YAP were discussed in this paper.

Aberrant activation of YAP/TAZ in BC

The role of YAP in BC has received increasing attention, and many studies have shown that YAP is a clinical marker of BC progression (44) and a key molecule contributing to postoperative recurrence and chemotherapy resistance in BC (45). Levels of YAP correlate positively with pathological grade

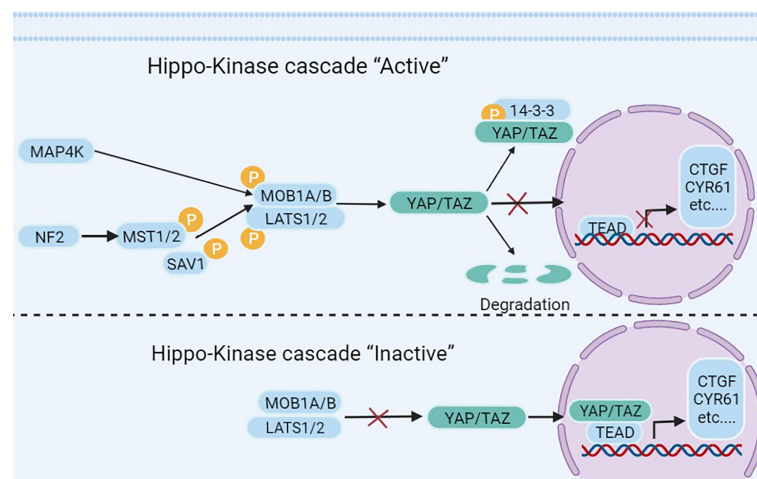


FIGURE 1

The Hippo pathway's upstream serine-threonine kinase cascade regulates YAP/TAZ. MST1/2 and MAP4K families are the main kinases of the Hippo-kinases cascade. When they are phosphorylated, which subsequently inhibits the transcriptional activity of YAP (20) and TAZ (21) through phosphorylating LATS1/2 (22–25). On the contrary, when the Hippo-kinase cascade is "inactive", it leads to YAP dephosphorylation, which translocates to the nucleus and binds to TEAD1–TEAD4, following with the transcription of downstream genes (26–28). Such as multiple anti-apoptotic and proliferative genes, including CTGF (connective tissue growth factor) and CYR61 (cysteine-rich angiogenic factor) (26–28). Other molecules regulating YAP/TAZ phosphorylation have also been reported in the literature, such as NDR1/2 (Nuclear Dbf2-related 1/2) (29), SRC (30–33), NLK (Nemo-like kinase) (34, 35), AMPK (5'-adenosine monophosphate-activated protein kinase) (36–38), and JNK (c-Jun N-terminal kinase) (39) have all been found to directly phosphorylate and hence control YAP/TAZ. Finally, YAP/TAZ is regulated in a kinase-independent manner (18, 19, 40).

of BC (46), and enhanced YAP activity has been shown in the majority of solid tumors (42), including lung, liver, sarcoma, pancreas, and breast (9, 10, 41).

Recent studies have reported that YAP is highly expressed in BC tissues and that knockdown of the YAP gene impaired the proliferation and migratory capacity of BC cells (47). High YAP expression correlates with poor prognosis in patients with BC (48). It is not clear how YAP becomes overactivated and forces BC initiation and progress, but several possible mechanisms have recently been identified (Figure 2 and Table 1).

Mutant GNA13 gene activates YAP/TAZ

Heterotrimeric G-proteins are important signal transduction molecules triggered by a large class of GPCRs (G-protein-coupled receptors) (56). Dysregulation of the GPCRs-G-protein pathway in cancer has been reported to be very common (57–59). G-protein family mutations were related to several malignancies, such as GNAQ or GNA11 ($G_{q/11}$ family) mutations are found in 90% of uveal melanomas (60, 61), 70% of pancreatic ductal carcinomas present GNAS (G_s family) mutations (62, 63), and 24% of epithelial T-cell lymphomas (64) GNAI2 ($G_{i/o}$ family) mutation. *In vitro*, tumorigenic experiments found that the $G_{i/o}$ family, $G_{q/11}$ family, and $G_{12/13}$ (GNA12 and GNA13) family mutation can promote oncogenic transformation (65–70).

Recent research based on bioinformatics analysis has shown that GNA13 mutation may be an oncogene in BC (59, 71, 72) and that the mutated GNA13 gene produces oncogenic effects by

activating YAP/TAZ (51). This was confirmed by research by Dr. Maziarz, who showed that the Arg-200 mutation of GNA13 in BC can significantly increase YAP/TAZ transcriptional activity by upregulating the RhoGEF-Rho GTPase cascade in TCGA database and cellular experiments (51)(Figure 3A). *In vitro*, tumorigenic experiments showed that the GNA13Arg-200 mutant induced cancerization of cells (control group of unmutated cell lines) (51). Dr. Maziarz's findings back up the theory that GNA13 hotspot mutations are a potential cause of BC, and that pharmacological inhibition of the Hippo-YAP pathway might be a feasible treatment option (51). This conclusion should be taken with a grain of salt because Dr. Maziarz's experiment lacks clinical validation in multiple data centers and *in vivo* tumorigenic assays.

NUAK2-LAST-YAP/TAZ positive feedback regulation loop

NUAK2 is a member of the AMPK kinase family, which has been extensively examined for its regulation of the Hippo-YAP pathway by regulating the Hippo kinase cassette (36–38, 73–76). Recent studies have shown that NUAK2 activity is significantly associated with aggressive, high-grade BC. Separate extracts of tumor cells from patients with high-grade and low-grade BC were tested and showed that NUAK2 expression in tumor cells was significantly higher in high-grade patients than in low-grade patients. Knockdown of NUAK2 gene in various cancer cell lines such as BC cell lines (TCCSUP, T24), colon cancer cell lines (SW480) and breast cancer cell lines (MDA-MB231 and MDA-

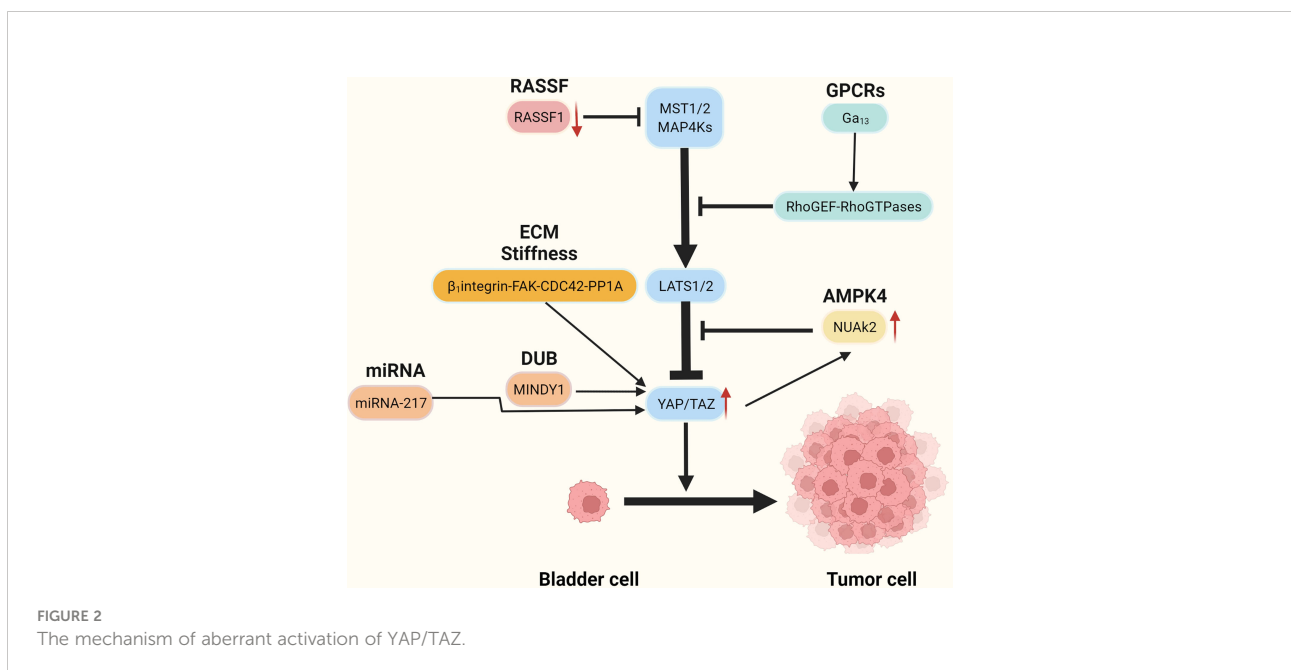


FIGURE 2
The mechanism of aberrant activation of YAP/TAZ.

TABLE 1 Functions of genes associated with the Hippo-YAP pathway.

Gene	Function of Hippo-YAP	Function of BC	Reference
ALDH1	Activated YAP/TAZ	Progression and chemotherapy resistance	(49)
CDC42	Activated YAP/TAZ	Progression and recurrence	(45)
FAK	Activated YAP/TAZ	Progression and recurrence	(45)
FOXM1	Activated YAP/TAZ	Progression and chemotherapy resistance	(50)
GNA13	Activated YAP/TAZ	Progression	(51)
ITGB1	Activated YAP/TAZ	Progression and recurrence	(45)
LATS1/2	Inactivated YAP/TAZ	Inhibition	(18, 19)
MINDY1	Activated YAP/TAZ	Progression	(52)
MST1/2	Inactivated YAP/TAZ	Inhibition	(18, 19)
miRNA-217	Activated YAP/TAZ	Progression	(53)
NUAK2	Inactivated LATS1/2	Progression	(46)
NRF2	Activated YAP/TAZ	Progression and chemotherapy resistance	(50)
RhoA/B/C	Activated YAP/TAZ	Progression	(51)
RASSF1	Inactivated MAST1/2	Progression	(54)
PDGFB	Activated YAP/TAZ	Progression and chemotherapy resistance	(55)
PP1A	Activated YAP/TAZ	Progression and recurrence	(45)

MB468) significantly inhibited the transcriptional activity of YAP/TAZ and the proliferation ability of cancer cells (46). Further experiments revealed that the expression of NUA2 was positively related to YAP/TAZ activity and negatively correlated with LAST activity. The regulatory effect of NUA2 on YAP/TAZ was significantly diminished when LAST was knocked down, and the knockdown of YAP/TAZ decreased the expression of NUA2. The Above research suggests the existence of a NUA2-LAST-YAP/TAZ positive feedback regulatory loop in BCs with high activity of NUA2 (46) (Figure 2).

The ubiquitin-protease system regulates the Hippo-YAP

The ubiquitin proteasomes system (UPS) is a protein degradation pathway that exists in all eukaryotic cells. UPS is the most important regulated protein degradation system, which participates in the cell cycle process, cell survival, apoptosis, DNA repair, and antigen presentation (77). The imbalance of UPS can lead to increased or reduced degradation of key proteins that promote tumorigenesis (78). Recently, it has been reported that several ubiquitin-protein ligases (E3) in UPS, such as PRAJA1, ITC1, SIAH2, FBXW7, and WWP1, play an important role in regulating the expression of YAP. These enzymes can regulate the stability of YAP protein in cancer cells through ubiquitin and proteasome degradation (79, 80). The protein level of LATS kinase is controlled by E3 ubiquitin

ligase-mediated degradation. In addition, LATS has a unique E3 chain, and MST1 also has its unique E3 ligase C-terminal recognition (81). The de-ubiquitin enzyme (DUB) is an enzyme with the opposite function of E3, such as MINDY1, which can increase its stability by removing the K48-linked ubiquitin chain from YAP. When it is exhausted, it can reduce the level of YAP protein and inhibit the YAP-TEAD-1 transcriptional activity, weakening the proliferation and invasiveness of cancer cells (52) (Figure 3B).

ECM stiffness activates YAP

More and more studies have found that the extracellular matrix (ECM) determines the fate and behavior of cancer cells, including differentiation, proliferation, apoptosis, and migration (82). In addition to perlecan, fibrillary collagen, and laminin in ECM, overexpression of agrin leads to increased density of ECM and ECM stiffness (83), leading to abnormal signals activating integrin (mechanosensory receptor) and related pathways (83). It is reported that collagen stiffness in ECM promotes NMIBC to MIBC, which may also be one of the causes of postoperative BC recurrence (84). However, the function and role of the proteins in ECM and the related signal transduction pathways are still opaque. Fortunately, according to the latest research, it has been found that the integrin-FAK-CDC42-PP1A (45) signaling pathway leads to ECM stiffness to promote the progression and recurrence of BC (Figure 3C). In addition to the high expression of β 1-integrin (encoded by ITGB1), FAK, and

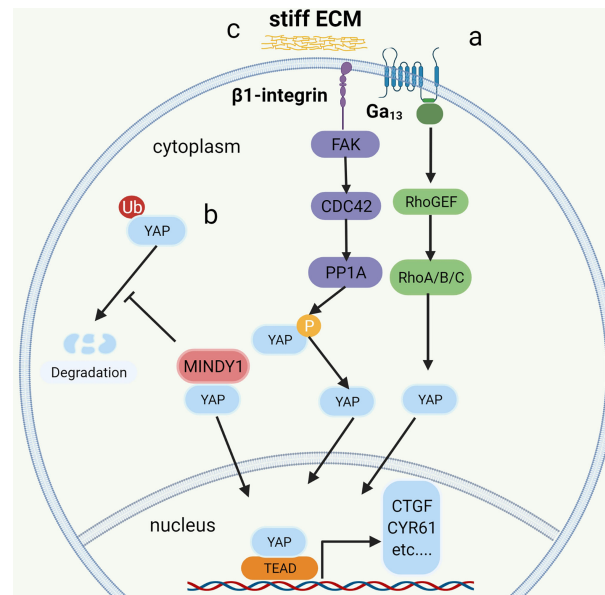


FIGURE 3

The mechanism of YAP regulation in bladder cancer. a: Mutation of $G_{12/13}$ can significantly increase the transcriptional activity of YAP/TAZ by upregulating the RhoGEF-Rho GTPase cascade. b: MINDY1 increases its stability and avoids degradation by removing the ubiquitin chain from YAP. c: ECM stiffness increases the nuclear localization of YAP by activating the integrin-FAK-CDC42-PP1A signaling pathway to dephosphorylate YAP.

CDC42, high ECM stiffness is also associated with increased nuclear localization of YAP (45). Molecular docking data showed that integrin binds to FAK through hydrogen bonding (45). FAK activates CDC42-PP1A kinase and dephosphorylates YAP (85), thus increasing the nuclear localization of YAP (45).

transcription factor YAP and its target proteins CTGF, CYR61, and ANKRD1 (53) (Figure 2).

Role of HIPPO-YAP pathway in BCSCS

Other pathways related to YAP activation

RASSF1 is a tumor suppressor (86). Its inactivation leads to the occurrence and development of many kinds of tumors including BC (87, 88). Low expression of RASSF1 in BC is strongly associated with high expression of YAP, CTGF, and CYR61, in addition to high-risk BC (54). Further studies have found that decreased expression of RASSF1 in BC inactivated MST1/2, which leads to increased activity of the YAP-TEAD-1 and promotes the occurrence and development of BC (54)(Figure 2).

The role of exosomes as novel biological markers in tumorigenesis, progression, diagnosis, and treatment is being increasingly emphasized (89–91). The miRNA-217 is secreted through exosomes by BC mesenchymal cells (53), and miRNA-217 expression is significantly higher in BC cell lines than in normal human bladder cell lines (53). The miRNA-217 affects BC proliferation, migration, and apoptosis by regulating the

Role of BCSCs in BC

BCSCs are a subgroup of BC cells, which have stem-like properties such as high proliferation, self-renewal, and drug resistance (92). Progression, chemotherapy resistance, and heterogeneity of BC are significantly related to cancer stem-like cells (CSCs) (93–95). At present, the markers commonly used to identify BCSCs are CD44, CD133, ALDH1, OV6, BMI1, and ABCG2 (49, 55, 96, 97). Although, the specific mechanism of preserving the stem-like qualities of BCSCs remains unclear, encouragingly, several signaling pathways have recently been reported to regulate the proliferation, tumorigenesis, and chemoresistance of BCSCs, including the Hippo-YAP signaling pathway, Hedgehog signaling pathway, Wnt/ β -catenin pathway, E2F1-EZH2-SUZ12 and KMT1A-GATA3-STAT3 cascade (49, 55, 98–101). A recent single-cell sequencing study showed that variants of GPRC5A, MLL2, and ARID1A drive the proliferation of BCSCs (102). The revelation of the molecular mechanism of

maintaining BCSCs is a very significant breakthrough in the therapeutic target of BC (92, 103).

YAP induces and preserves stem-like qualities of BCSCs

Previous studies have shown that the Hippo-YAP pathway is essential to maintain the stem-like properties of some CSCs (41), such as BC, prostate cancer, breast cancer, lung cancer, and glioblastoma. YAP is a key regulatory protein for CSCs proliferation and carcinogenesis (55, 104–106). YAP is also of great significance in BCSCs. The research of Dr. Wang and Dr. Zhao shows that YAP is necessary for the proliferation and maintenance of stem-like properties of BCSCs and is related to its expressing OV6 and ALDH1 (49, 55).

OV6 is a unique marker of CSCs in epithelial malignant tumors, such as BC, hepatocellular carcinoma, cholangiocarcinoma, and esophageal cancer. CSCs are highly expressed and are associated with poor prognosis (55, 107–110). Dr. Wang et al. have found that BC cells in OV6⁺ have strong characteristics of tumor stem-like cells, which can significantly inhibit its proliferation and chemotherapy resistance when YAP is knocked out. Further experiments showed that YAP maintained the stem-like properties of BC cells of OV6⁺ by activating PDGF β , and the cells lost the characteristics of stem-like when PDGF β was knocked out. The use of YAP or PDGFR inhibitors in a mouse model of BC can block the positive feedback regulatory loop of BCSCs of OV6⁺, thereby overcoming the resistance of advanced BC to cisplatin (55). Dr. Wang's research demonstrated that there is a positive

feedback regulatory pathway in BC cells of OV6⁺. YAP activates PDGF β gene transcription and translation through TEAD-1 to produce PDGF-BB (Platelet-derived growth factor subunit B protein), which in turn prevents YAP from being phosphorylated by LATS1/2, thereby increasing the nuclear localization of YAP (55) (Figure 4).

YAP activity was also found in BCSCs cells of ALDH1⁺. When YAP was inhibited, the expression of ALDH1 decreased, it was more sensitive to chemotherapeutic drugs, and the ability of self-renewal and proliferation decreased significantly (49). In addition, it was also found that Hippo-YAP and COX2/PGE2 pathways co-acted on the proliferation of BCSCs, and their inhibitors successfully blocked the progression of BC (111). Moreover, YAP induces non-CSCs into CSCs (17) and maintains the characteristics of CSCs by inducing autophagy (112). These researches suggest that the Hippo-YAP pathway plays an important role in the proliferation and development of BCSCs and BC.

The HIPPO-YAP in chemotherapy resistance and immunotherapy

Mechanisms of chemotherapy resistance in BC

Drug resistance to chemotherapy and targeted chemotherapy remains a major obstacle to the treatment of various cancers, including BC (4, 113). The causes of chemotherapy resistance are very complex and can be divided into congenital resistance and secondary resistance according to their essential causes.

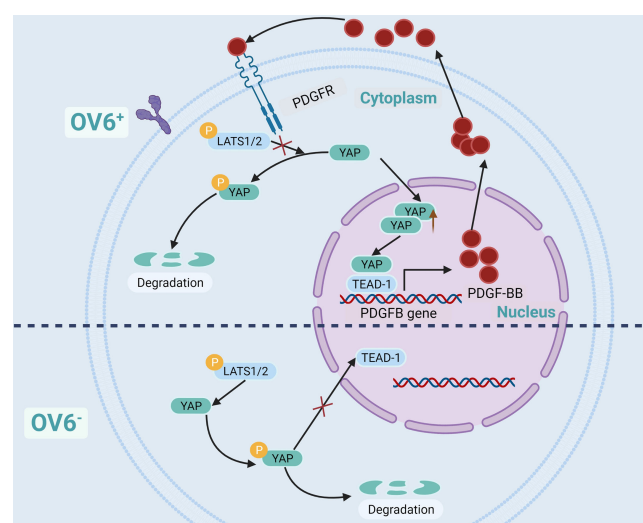


FIGURE 4
YAP/TEAD-1/PDGFB/PDGFR positive feedback regulatory loop in OV6⁺ BCSCs.

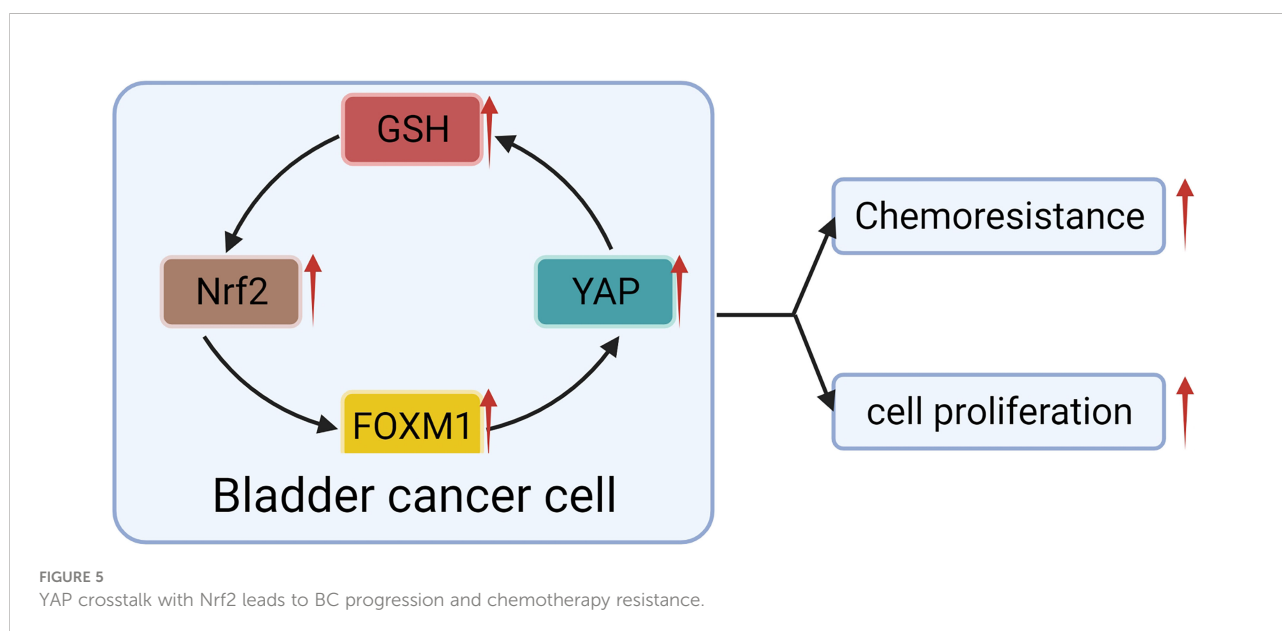
Congenital resistance refers to mutations in the genome or epigenetic mutations that have occurred before treatment. Secondary resistance refers to genomic alterations that occur after treatment with the appropriate drug (113). Several prevalent mechanisms of drug resistance have been reported, such as increased drug efflux, drug target mutations, cell stemming, apoptotic escape, immune escape, and DNA damage repair (114–118). Among them, the role of cell stemness and apoptotic escape in chemotherapy resistance has been emphasized. The active DNA repair capacity and resistance to apoptosis that are characteristic of cell stemness are the main mechanisms of its resistance (119–121). Therefore, further studies targeting the mechanisms that maintain cell stemness are important to improve chemotherapeutic efficacy.

The role of Hippo-YAP in chemotherapy resistance of BC

YAP is reported to be associated with drug resistance, such as cisplatin (122, 123), survivin and erlotinib inhibitors (124), anti-tubulin drugs (125), and radiation therapy (126). The sensitivity of cisplatin was negatively correlated with the expression of YAP in BC (127). Overexpression of YAP in BC was significantly correlated to resistance to cisplatin. Knocking out of the YAP gene not only increased the sensitivity of BC to cisplatin (50, 127) but also increased the sensitivity to other DNA damage drugs (50). YAP was recently reported to mediate chemotherapy resistance by maintaining tumor cell stemness (49, 55). Although there is a lot of evidence that YAP plays an important role in chemotherapy resistance of BC, the specific

mechanism of YAP leading to chemotherapy resistance of BC is limited.

Fortunately, a recent study showed that YAP crosstalk with NRF2, thereby enhancing the antioxidant capacity of tumor cells that mediated BC chemotherapy resistance (50). The escape of apoptosis mediated by antioxidation is recognized as the mechanism of drug resistance in BC (50, 113). NRF2 is a classical regulator of cellular redox response (128, 129). With further research, it has been found that NRF2 has a specific high expression in cancer cells, can promote the progression (129) and metastasis (130) of many kinds of cancer, and make the human body resistant to chemotherapy and radiotherapy (131, 132). The interaction between NRF2 and YAP was found in BC cells. Knocking-out of NRF2 not only inhibited the proliferation and invasion of BC cells but also significantly restrained the expression of YAP (50). When YAP was blocked, the growth, invasion, and NRF2 expression of cancer cells were significantly decreased (50). For example, the chemotherapeutic drug-resistant cell lines were more responsive to Aila (YAP and NRF2 inhibitors) (133). Researchers suggested that NRF2 may interact with YAP through FOXM1 (50). A significant correlation was found among the expression of NRF2, FOXM1, YAP, and GSH in chemotherapy-resistant BC cell lines (50). When NRF2 was knocked out, the expression of YAP, FOXM1 and GSH decreased synchronously, along with decreased proliferation ability of the cell line and increased sensitivity to chemotherapeutic drugs (50). Although the evidence of direct interaction between NRF2 and FOXM1 is not sufficient but combined with the experiments of Dr. Gucci and Professor Eric Ciamporero, we can speculate that there is a vague interaction between NRF2 and YAP in BC, which plays a role in regulating chemotherapy resistance of BC (Figure 5).



Potential role of Hippo-YAP in immunotherapy

Immunotherapy has been widely demonstrated to be effective in BC and is currently a second-line treatment option for metastatic BC and a first-line treatment option for cisplatin-ineligible PD-L1+ patients (4, 5). However, the benefit of immunotherapy for BC patients is limited because of its complex tumor microenvironment-mediated immune escape and the low responsiveness of immunotherapy (5). Although no studies related to the Hippo pathway with immune escape and immunotherapy in BC. However, YAP was found to increase tumor immune escape response by increasing PD-L1 expression in other cancers, such as melanoma (134), and colorectal cancer (135). Interestingly, it was found that in lung cancer, YAP expression increased anti-tumor immune response by decreasing PD-L1 expression (136). Based on the available evidence the Hippo-YAP pathway has a quite complex role in tumor immunity with tissue heterogeneity. Therefore, revealing the role of Hippo-YAP in anti-tumor immunity in bladder cancer may be important for improving the efficacy of immunotherapy in the future.

Clinical potential of HIPPO-YAP pathway for BC therapy

The preclinical attempt targeted YAP-TEAD

The aberrant activation of YAP in BC leads to tumor recurrence and chemoresistance, which are major clinical difficulties of BC therapy. Targeting Hippo-YAP possesses the potential in solving this major obstacle. Since YAP exerts transcriptional activity primarily by binding to the transcription factor TEAD-1 (26, 27, 137, 138), inhibition of this interaction makes it the most direct and effective (138). Verteporfin (VP) inhibits the interaction of YAP with TEAD-1 by binding YAP (139). *In vitro* experiments demonstrate that VP inhibits BC growth and the stem-like properties of BCSCs (140–142). Although VP is used to treat macular degeneration, its low metabolic rate and low specificity *in vivo* make it toxic (143, 144), hindering its future use in cancer therapy. VGLL4 (Vestigial like family member 4) binds TEAD-1 competitively with YAP through the TDU (Tondu) structural domain, thereby reducing the transcriptional benefit of YAP (145, 146). Super-TDU (VGLL4-mimetic peptide) has significant anticancer effects in a mouse gastric cancer model induced by *Helicobacter pylori* (145). It has been reported that a YAP analog, namely 17-peptide (147, 148), has now been designed with a super-inhibitory effect on YAP-TEAD-1 and a significant

inhibition of tumor proliferation in an ovarian cancer animal model (149). Unfortunately, even though breaking the YAP-TEAD-1 interaction is the most direct way to target the Hippo-YAP pathway, there are still no relevant drugs approved for clinical treatment of BC use.

Activating Hippo kinase cascade would be a promising attempt

Hippo-kinase cascade, consisting mainly of the MST1/2, LAST1/2, and MAP4K families, whose activation inhibits the transcriptional function of YAP/TAZ (150). Thus, activation of the Hippo-kinase cascade is a viable way to target the Hippo-YAP pathway for cancer treatment. SHAP (STRN3-derived Hippo-activating peptide), a potent activator of MST1/2 enzymes, has better inhibitory effects on YAP than drugs such as VP and super-TDU, in addition to advantages toxicity and physical properties (151). In a mouse model of gastric cancer, SHAP exhibited stronger tumor-suppressive effects than drugs such as VP and super-TDU (151). The RAF (rapidly-accelerated fibrosarcoma) family was shown to inactivate MST1/2 by a mechanism acting upstream of the MST1/2 kinase (152). Therefore, inhibition of RAF leads to activation of MST1/2, which acts as an anticancer agent. Previously, ISIS-1532 oligonucleotide was found to silence the expression of RAF (153, 154). Although ISIS-1532 had a good response in lung cancer (153, 154), however, it performed poorly in phase II clinical trials in people with colon cancer, prostate cancer, and ovarian cancer (154–157). Despite the lack of studies on Hippo-kinase cascade activators in BC, this type of activator holds remarkably positive promise in the treatment of BC (144).

Conclusion and perspective

Overexpression of YAP was verified, and current studies indicated that YAP has a more extensive contribution to the development of BC. YAP plays a key role in BC initiation, progression, chemoresistance, and induction of BCSCs (44, 45, 47, 55). Interestingly, multiple mechanisms are now found to be involved in YAP upregulation in BC. Therefore, the development of inhibitors of YAP is a promising direction. However, current molecular drugs faced a series of challenges, including insufficient clinical trials, uncontrolled side effects, metabolism difficulties, etc. Hence, drug metabolism and toxicology are urgent in the future development of YAP-related drugs. New drug design strategies, like antibody-drug coupling (ADC), should be a promising direction. Moreover, YAP-based chemicals are hard to compare favorably with traditional chemotherapy drugs in killing cancer cells frankly.

However, they would more adjuvant drugs in overcoming the chemotherapy resistance than a single therapeutic target.

Although the mechanisms of Hippo-kinase cascade regulation in BC are poorly understood. However, according to recent findings, targeting the Hippo cascade may be more effective than interfering with the YAP-TEAD combination. In animal models of gastric cancer, SHAP was more efficacious than the conventional direct inhibitors of YAP (151). Therefore, we believe that activation of the Hippo-kinase cascade is a promising direction for the treatment of malignancies. However, developing protein activators is significantly more challenging than protein inhibitors. Therefore, further unraveling the mechanism of Hippo-kinase cascade dysregulation and developing related drugs are important for improving the clinical prognosis and developing individualized treatment plans for BC patients in the future.

Review strategy and methods

The review strategy and inclusion criteria as listed below. The Major review strategy: a total of 41 publications were retrieved from Pubmed with the search terms Hippo/YAP and bladder cancer/urothelial carcinoma/transitional cell carcinomas, including 7 reviews and 34 research articles. The final selection of 22 articles (19 articles and 3 reviews) was based on the inclusion criteria (a. Subjects with bladder cancer or bladder cancer cell lines; b. independent cohort validation with relevant biomarker studies; c. Complete and appropriate controlled experiments). The minor review strategy: 1. Hippo/YAP and CSCs/cancer stem cells/bladder cancer stem cells 178 (58 reviews and 120 papers); 2. Hippo/YAP and chemotherapy resistance/immunotherapy 82(22 reviews,60 papers); 3. Hippo/YAP and therapy 481 (151 reviews and 330 papers).

References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/caac.21660
- Lin S, Gao K, Gu S, You L, Qian S, Tang M, et al. Worldwide trends in cervical cancer incidence and mortality, with predictions for the next 15 years. *Cancer* (2021) 127(21):4030–9. doi: 10.1002/cncr.33795
- Babjuk M, Burger M, Capoun O, Cohen D, Comp erat EM, Dominguez Escrig JL, et al. European Association of urology guidelines on non-muscle-invasive bladder cancer (Ta, T1, and carcinoma *in situ*). *Eur Urol* (2022) 81(1):75–94. doi: 10.1016/j.eururo.2021.08.010
- Witjes JA, Bruins HM, Cathomas R, Comp erat EM, Cowan NC, Gakis G, et al. European Association of urology guidelines on muscle-invasive and metastatic bladder cancer: Summary of the 2020 guidelines. *Eur Urol* (2021) 79(1):82–104. doi: 10.1016/j.eururo.2020.03.055
- Bellmunt J, de Wit R, Vaughn DJ, Fradet Y, Lee JL, Fong L, et al. Pembrolizumab as second-line therapy for advanced urothelial carcinoma. *N Engl J Med* (2017) 376(11):1015–26. doi: 10.1056/NEJMoa1613683
- Knowles MA, Hurst CD. Molecular biology of bladder cancer: New insights into pathogenesis and clinical diversity. *Nat Rev Cancer* (2015) 15(1):25–41. doi: 10.1038/nrc3817
- Chiang CH, Chung JG, Hsu FT. Regorafenib induces extrinsic/intrinsic apoptosis and inhibits Mapk/Nf- b-modulated tumor progression in bladder cancer *in vitro* and *in vivo*. *Environ Toxicol* (2019) 34(6):679–88. doi: 10.1002/tox.22734
- Hua X, Xu J, Deng X, Xu J, Li J, Zhu DQ, et al. New compound chla-f induces autophagy-dependent anti-cancer effect *via* upregulating sestrin-2 in human bladder cancer. *Cancer Lett* (2018) 436:38–51. doi: 10.1016/j.canlet.2018.08.013
- Moroishi T, Hansen CG, Guan KL. The emerging roles of yap and taz in cancer. *Nat Rev Cancer* (2015) 15(2):73–9. doi: 10.1038/nrc3876
- Moya IM, Halder G. Hippo-Yap/Taz signalling in organ regeneration and regenerative medicine. *Nat Rev Mol Cell Biol* (2019) 20(4):211–26. doi: 10.1038/s41580-018-0086-y
- Dong J, Feldmann G, Huang J, Wu S, Zhang N, Comerford SA, et al. Elucidation of a universal size-control mechanism in drosophila and mammals. *Cell* (2007) 130(6):1120–33. doi: 10.1016/j.cell.2007.07.019
- Pan D. The hippo signaling pathway in development and cancer. *Dev Cell* (2010) 19(4):491–505. doi: 10.1016/j.devcel.2010.09.011

Author contributions

GZ, JZ, and XC designed the thesis and outline for the review. LD and GX searched related publications. XC, KL, and RH drafted the manuscript. GZ, JZ, and XZ reviewed the manuscript and polished the grammar. All authors contributed to the manuscript revision and approved the submitted version.

Funding

This work was supported by the National Natural Science Foundation of China (No. 81860456 and 81760462); Training plan for the academic and technical leaders of major subjects in Jiangxi Province (No. 20213BCJL22038); Science and Technology Research Project of Jiangxi Provincial Education Department (GJJ211550 and GJJ211523).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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13. Park JH, Shin JE, Park HW. The role of hippo pathway in cancer stem cell biology. *Mol Cells* (2018) 41(2):83–92. doi: 10.14348/molcells.2018.2242
14. Hansen CG, Moroishi T, Guan KL. Yap and taz: A nexus for hippo signaling and beyond. *Trends Cell Biol* (2015) 25(9):499–513. doi: 10.1016/j.tcb.2015.05.002
15. Santinon G, Pocaterra A, Dupont S. Control of Yap/Taz activity by metabolic and nutrient-sensing pathways. *Trends Cell Biol* (2016) 26(4):289–99. doi: 10.1016/j.tcb.2015.11.004
16. Totaro A, Panciera T, Piccolo S. Yap/Taz upstream signals and downstream responses. *Nat Cell Biol* (2018) 20(8):888–99. doi: 10.1038/s41556-018-0142-z
17. Yu FX, Zhao B, Guan KL. Hippo pathway in organ size control, tissue homeostasis, and cancer. *Cell* (2015) 163(4):811–28. doi: 10.1016/j.cell.2015.10.044
18. Dupont S, Morsut L, Aragona M, Enzo E, Giulitti S, Cordenonsi M, et al. Role of Yap/Taz in mechanotransduction. *Nature* (2011) 474(7350):179–83. doi: 10.1038/nature10137
19. Kofler M, Speight P, Little D, Di Ciano-Oliveira C, Szási K, Kapus A. Mediated nuclear import and export of taz and the underlying molecular requirements. *Nat Commun* (2018) 9(1):4966. doi: 10.1038/s41467-018-07450-0
20. Zhao B, Wei X, Li W, Udan RS, Yang Q, Kim J, et al. Inactivation of yap oncoprotein by the hippo pathway is involved in cell contact inhibition and tissue growth control. *Genes Dev* (2007) 21(21):2747–61. doi: 10.1101/gad.1602907
21. Lei QY, Zhang H, Zhao B, Zha ZY, Bai F, Pei XH, et al. Taz promotes cell proliferation and epithelial-mesenchymal transition and is inhibited by the hippo pathway. *Mol Cell Biol* (2008) 28(7):2426–36. doi: 10.1128/mcb.01874-07
22. Chan EH, Nousiainen M, Chalalamalsetty RB, Schäfer A, Nigg EA, Silljé HH. The Ste20-like kinase Mst2 activates the human large tumor suppressor kinase Lats1. *Oncogene* (2005) 24(12):2076–86. doi: 10.1038/sj.onc.1208445
23. Li Q, Li S, Mana-Capelli S, Roth Flach RJ, Danai LV, Amcheslavsky A, et al. The conserved misshapen-warts-yorkie pathway acts in enteroblasts to regulate intestinal stem cells in drosophila. *Dev Cell* (2014) 31(3):291–304. doi: 10.1016/j.devcel.2014.09.012
24. Meng Z, Moroishi T, Mottier-Pavie V, Plouffe SW, Hansen CG, Hong AW, et al. Map4k family kinases act in parallel to Mst1/2 to activate Lats1/2 in the hippo pathway. *Nat Commun* (2015) 6:8357. doi: 10.1038/ncomms9357
25. Zheng Y, Wang W, Liu B, Deng H, Uster E, Pan D. Identification of happyhour/Map4k as alternative Hpo/Mst-like kinases in the hippo kinase cascade. *Dev Cell* (2015) 34(6):642–55. doi: 10.1016/j.devcel.2015.08.014
26. Ota M, Sasaki H. Mammalian tead proteins regulate cell proliferation and contact inhibition as transcriptional mediators of hippo signaling. *Development* (2008) 135(24):4059–69. doi: 10.1242/dev.027151
27. Zhang H, Liu CY, Zha ZY, Zhao B, Yao J, Zhao S, et al. Tead transcription factors mediate the function of taz in cell growth and epithelial-mesenchymal transition. *J Biol Chem* (2009) 284(20):13355–62. doi: 10.1074/jbc.M900843200
28. Zhao B, Ye X, Yu J, Li L, Li W, Li S, et al. Tead mediates yap-dependent gene induction and growth control. *Genes Dev* (2008) 22(14):1962–71. doi: 10.1101/gad.1664408
29. Zhang L, Tang F, Terracciano L, Hynx D, Kohler R, Bichet S, et al. Ndr functions as a physiological Yap1 kinase in the intestinal epithelium. *Curr Biol* (2015) 25(3):296–305. doi: 10.1016/j.cub.2014.11.054
30. Vassilev A, Kaneko KJ, Shu H, Zhao Y, DePamphilis ML. Tead/Tef transcription factors utilize the activation domain of Yap65, a Src/Yes-associated protein localized in the cytoplasm. *Genes Dev* (2001) 15(10):1229–41. doi: 10.1101/gad.888601
31. Zaidi SK, Sullivan AJ, Medina R, Ito Y, van Wijnen AJ, Stein JL, et al. Tyrosine phosphorylation controls Runx2-mediated subnuclear targeting of yap to repress transcription. *EMBO J* (2004) 23(4):790–9. doi: 10.1038/sj.emboj.7600073
32. Vlahov N, Scraze S, Soto MS, Grawenda AM, Bradley L, Pankova D, et al. Alternate Rassf1 transcripts control src activity, e-cadherin contacts, and yap-mediated invasion. *Curr Biol* (2015) 25(23):3019–34. doi: 10.1016/j.cub.2015.09.072
33. Byun MR, Hwang JH, Kim AR, Kim KM, Park JI, Oh HT, et al. Src activates taz for intestinal tumorigenesis and regeneration. *Cancer Lett* (2017) 410:32–40. doi: 10.1016/j.canlet.2017.09.003
34. Moon S, Kim W, Kim S, Kim Y, Song Y, Bilousov O, et al. Phosphorylation by nlk inhibits yap-14-3-3-interactions and induces its nuclear localization. *EMBO Rep* (2017) 18(1):61–71. doi: 10.15252/embr.201642683
35. Hong AW, Meng Z, Yuan HX, Plouffe SW, Moon S, Kim W, et al. Osmotic stress-induced phosphorylation by nlk at Ser128 activates yap. *EMBO Rep* (2017) 18(1):72–86. doi: 10.15252/embr.201642681
36. Mo JS, Meng Z, Kim YC, Park HW, Hansen CG, Kim S, et al. Cellular energy stress induces ampk-mediated regulation of yap and the hippo pathway. *Nat Cell Biol* (2015) 17(4):500–10. doi: 10.1038/ncb3111
37. DeRan M, Yang J, Shen CH, Peters EC, Fitamant J, Chan P, et al. Energy stress regulates hippo-yap signaling involving ampk-mediated regulation of angiomin-like 1 protein. *Cell Rep* (2014) 9(2):495–503. doi: 10.1016/j.celrep.2014.09.036
38. Wang W, Xiao ZD, Li X, Aziz KE, Gan B, Johnson RL, et al. Ampk modulates hippo pathway activity to regulate energy homeostasis. *Nat Cell Biol* (2015) 17(4):490–9. doi: 10.1038/ncb3113
39. Tomlinson V, Gudmundsdottir K, Luong P, Leung KY, Knebel A, Basu S. Jnk phosphorylates yes-associated protein (Yap) to regulate apoptosis. *Cell Death Dis* (2010) 1(2):e29. doi: 10.1038/cddis.2010.7
40. Chan SW, Lim CJ, Chong YF, Pobbati AV, Huang C, Hong W. Hippo pathway-independent restriction of taz and yap by angiomin. *J Biol Chem* (2011) 286(9):7018–26. doi: 10.1074/jbc.C110.212621
41. Zanconato F, Cordenonsi M, Piccolo S. Yap/Taz at the roots of cancer. *Cancer Cell* (2016) 29(6):783–803. doi: 10.1016/j.ccell.2016.05.005
42. Steinhardt AA, Gayyed MF, Klein AP, Dong J, Maitra A, Pan D, et al. Expression of yes-associated protein in common solid tumors. *Hum Pathol* (2008) 39(11):1582–9. doi: 10.1016/j.humpath.2008.04.012
43. Salem O, Hansen CG. The hippo pathway in prostate cancer. *Cells* (2019) 8(4). doi: 10.3390/cells8040370
44. Tran L, Xiao JF, Agarwal N, Duex JE, Theodorescu D. Advances in bladder cancer biology and therapy. *Nat Rev Cancer* (2021) 21(2):104–21. doi: 10.1038/s41568-020-00313-1
45. Ghasemi H, Mousavibahar SH, Hashemnia M, Karimi J, Khodadadi I, Mirzaei F, et al. Tissue stiffness contributes to yap activation in bladder cancer patients undergoing transurethral resection. *Ann N Y Acad Sci* (2020) 1473(1):48–61. doi: 10.1111/nyas.14358
46. Gill MK, Christova T, Zhang YY, Gregorieff A, Zhang L, Narimatsu M, et al. A feed forward loop enforces Yap/Taz signaling during tumorigenesis. *Nat Commun* (2018) 9(1):3510. doi: 10.1038/s41467-018-05939-2
47. Qiu D, Zhu Y, Cong Z. Yap triggers bladder cancer proliferation by affecting the mapk pathway. *Cancer Manag Res* (2020) 12:12205–14. doi: 10.2147/cmar.S273442
48. Poma AM, Torregrossa L, Bruno R, Basolo F, Fontanini G. Hippo pathway affects survival of cancer patients: Extensive analysis of tcga data and review of literature. *Sci Rep* (2018) 8(1):10623. doi: 10.1038/s41598-018-28928-3
49. Zhao AY, Dai YJ, Lian JF, Huang Y, Lin JG, Dai YB, et al. Yap regulates Aldh1a1 expression and stem cell property of bladder cancer cells. *Oncotargets Ther* (2018) 11:6657–63. doi: 10.2147/ott.S170858
50. Ciamporcero E, Daga M, Pizzimenti S, Roetto A, Dianzani C, Compagnone A, et al. Crosstalk between Nrf2 and yap contributes to maintaining the antioxidant potential and chemoresistance in bladder cancer. *Free Radic Biol Med* (2018) 115:447–57. doi: 10.1016/j.freeradbiomed.2017.12.005
51. Maziarz M, Federico A, Zhao J, Dujmusic L, Zhao Z, Monti S, et al. Naturally occurring hotspot cancer mutations in Gα(13) promote oncogenic signaling. *J Biol Chem* (2020) 295(49):16897–904. doi: 10.1074/jbc.AC120.014698
52. Luo Y, Zhou J, Tang J, Zhou F, He Z, Liu T, et al. Mindy1 promotes bladder cancer progression by stabilizing yap. *Cancer Cell Int* (2021) 21(1):395. doi: 10.1186/s12935-021-02095-4
53. Huang ZM, Wang H, Ji ZG. Bladder mesenchymal stromal cell-derived exosomal mirna-217 modulates bladder cancer cell survival through hippo-yap pathway. *Inflammation Res* (2021) 70(9):959–69. doi: 10.1007/s00011-021-01494-7
54. Khandelwal M, Anand V, Appunni S, Seth A, Singh P, Mathur S, et al. Rassf1a-hippo pathway link in patients with urothelial carcinoma of bladder: plausible therapeutic target. *Mol Cell Biochem* (2020) 464(1–2):51–63. doi: 10.1007/s11010-019-03648-y
55. Wang KJ, Wang C, Dai LH, Yang J, Huang H, Ma XJ, et al. Targeting an autocrine regulatory loop in cancer stem-like cells impairs the progression and chemotherapy resistance of bladder cancer. *Clin Cancer Res* (2019) 25(3):1070–86. doi: 10.1158/1078-0432.Ccr-18-0586
56. Oldham WM, Hamm HE. Heterotrimeric G protein activation by G-Protein-Coupled receptors. *Nat Rev Mol Cell Biol* (2008) 9(1):60–71. doi: 10.1038/nrm2299
57. O'Hayre M, Vázquez-Prado J, Kufareva I, Stawiski EW, Handel TM, Seshagiri S, et al. The emerging mutational landscape of G proteins and G-Protein-Coupled receptors in cancer. *Nat Rev Cancer* (2013) 13(6):412–24. doi: 10.1038/nrc3521
58. Kan Z, Jaiswal BS, Stinson J, Janakiraman V, Bhatt D, Stern HM, et al. Diverse somatic mutation patterns and pathway alterations in human cancers. *Nature* (2010) 466(7308):869–73. doi: 10.1038/nature09208
59. Wu V, Yeerna H, Nohata N, Chiou J, Harismendy O, Raimondi F, et al. Illuminating the onco-gpcrome: Novel G protein-coupled receptor-driven oncoendocrine networks and targets for cancer immunotherapy. *J Biol Chem* (2019) 294(29):11062–86. doi: 10.1074/jbc.REV119.005601

60. Van Raamsdonk CD, Bezrookove V, Green G, Bauer J, Gaugler L, O'Brien JM, et al. Frequent somatic mutations of gnaq in uveal melanoma and blue naevi. *Nature* (2009) 457(7229):599–602. doi: 10.1038/nature07586
61. Van Raamsdonk CD, Griewank KG, Crosby MB, Garrido MC, Vemula S, Wiesner T, et al. Mutations in Gna11 in uveal melanoma. *N Engl J Med* (2010) 363(23):2191–9. doi: 10.1056/NEJMoa1000584
62. Ideno N, Yamaguchi H, Ghosh B, Gupta S, Okumura T, Steffen DJ, et al. Gnas(R201c) induces pancreatic cystic neoplasms in mice that express activated kras by inhibiting Yap1 signaling. *Gastroenterology* (2018) 155(5):1593–607.e12. doi: 10.1053/j.gastro.2018.08.006
63. Wu J, Matthaei H, Maitra A, Dal Molin M, Wood LD, Eshleman JR, et al. Recurrent gnas mutations define an unexpected pathway for pancreatic cyst development. *Sci Transl Med* (2011) 3(92):92ra66. doi: 10.1126/scitranslmed.3002543
64. Nairismägi ML, Tan J, Lim JQ, Nagarajan S, Ng CC, Rajasegaran V, et al. Jak-stat and G-Protein-Coupled receptor signaling pathways are frequently altered in epitheliotropic intestinal T-cell lymphoma. *Leukemia* (2016) 30(6):1311–9. doi: 10.1038/leu.2016.13
65. Xu N, Bradley L, Ambudkar I, Gutkind JS. A mutant alpha subunit of G12 potentiates the eicosanoid pathway and is highly oncogenic in nih 3t3 cells. *Proc Natl Acad Sci U.S.A.* (1993) 90(14):6741–5. doi: 10.1073/pnas.90.14.6741
66. Voyno-Yasenetskaya TA, Pace AM, Bourne HR. Mutant alpha subunits of G12 and G13 proteins induce neoplastic transformation of rat-1 fibroblasts. *Oncogene* (1994) 9(9):2559–65.
67. Wong YH, Chan JS, Yung LY, Bourne HR. Mutant alpha subunit of g2 transforms swiss 3t3 cells. *Oncogene* (1995) 10(10):1927–33.
68. Pace AM, Wong YH, Bourne HR. A mutant alpha subunit of Gi2 induces neoplastic transformation of rat-1 cells. *Proc Natl Acad Sci U.S.A.* (1991) 88(16):7031–5. doi: 10.1073/pnas.88.16.7031
69. Ram PT, Horvath CM, Iyengar R. Stat3-mediated transformation of nih-3t3 cells by the constitutively active Q205L galphao protein. *Science* (2000) 287(5450):142–4. doi: 10.1126/science.287.5450.142
70. Gupta SK, Gallego C, Lowndes JM, Pleiman CM, Sable C, Eisfelder BJ, et al. Analysis of the fibroblast transformation potential of gtpase-deficient G12 oncogenes. *Mol Cell Biol* (1992) 12(1):190–7. doi: 10.1128/mcb.12.1.190-197.1992
71. Robertson AG, Kim J, Al-Ahmadie H, Bellmunt J, Guo G, Cherniack AD, et al. Comprehensive molecular characterization of muscle-invasive bladder cancer. *Cell* (2017) 171(3):540–56.e25. doi: 10.1016/j.cell.2017.09.007
72. Bailey MH, Tokheim C, Porta-Pardo E, Sengupta S, Bertrand D, Weerasinghe A, et al. Comprehensive characterization of cancer driver genes and mutations. *Cell* (2018) 174(4):1034–5. doi: 10.1016/j.cell.2018.07.034
73. Mohseni M, Sun J, Lau A, Curtis S, Goldsmith J, Fox VL, et al. A genetic screen identifies an Lkb1-mark signalling axis controlling the hippo-yap pathway. *Nat Cell Biol* (2014) 16(1):108–17. doi: 10.1038/ncb2884
74. Heidary Arash E, Shiban A, Song S, Attisano L. Mark4 inhibits hippo signaling to promote proliferation and migration of breast cancer cells. *EMBO Rep* (2017) 18(3):420–36. doi: 10.15252/embr.201642455
75. Kwan J, Sczaniecka A, Heidary Arash E, Nguyen L, Chen CC, Ratkovic S, et al. Dlg5 connects cell polarity and hippo signaling protein networks by linking par-1 with Mst1/2. *Genes Dev* (2016) 30(24):2696–709. doi: 10.1101/gad.284539.116
76. Wehr MC, Holder MV, Gailite I, Saunders RE, Maile TM, Ciirdaeva E, et al. Salt-inducible kinases regulate growth through the hippo signalling pathway in drosophila. *Nat Cell Biol* (2013) 15(1):61–71. doi: 10.1038/ncb2658
77. Manasanch EE, Orlowski RZ. Proteasome inhibitors in cancer therapy. *Nat Rev Clin Oncol* (2017) 14(7):417–33. doi: 10.1038/nrclinonc.2016.206
78. Orlowski RZ, Dees EC. The role of the ubiquitination-proteasome pathway in breast cancer: applying drugs that affect the ubiquitin-proteasome pathway to the therapy of breast cancer. *Breast Cancer Res* (2003) 5(1):1–7. doi: 10.1186/bcr460
79. Tu K, Yang W, Li C, Zheng X, Lu Z, Guo C, et al. Fbxw7 is an independent prognostic marker and induces apoptosis and growth arrest by regulating yap abundance in hepatocellular carcinoma. *Mol Cancer* (2014) 13:110. doi: 10.1186/1476-4598-13-110
80. Ma B, Chen Y, Chen L, Cheng H, Mu C, Li J, et al. Hypoxia regulates hippo signalling through the Siah2 ubiquitin E3 ligase. *Nat Cell Biol* (2015) 17(1):95–103. doi: 10.1038/ncb3073
81. He M, Zhou Z, Shah AA, Hong Y, Chen Q, Wan Y. New insights into posttranslational modifications of hippo pathway in carcinogenesis and therapeutics. *Cell Div* (2016) 11:4. doi: 10.1186/s13008-016-0013-6
82. Chin L, Xia Y, Discher DE, Janney PA. Mechanotransduction in cancer. *Curr Opin Chem Eng* (2016) 11:77–84. doi: 10.1016/j.coche.2016.01.011
83. Chakraborty S, Hong W. Linking extracellular matrix agrin to the hippo pathway in liver cancer and beyond. *Cancers (Basel)* (2018) 10(2). doi: 10.3390/cancers10020045
84. Zhu H, Chen H, Wang J, Zhou L, Liu S. Collagen stiffness promoted non-muscle-invasive bladder cancer progression to muscle-invasive bladder cancer. *Onco Targets Ther* (2019) 12:3441–57. doi: 10.2147/ott.s194568
85. Hicks-Berthet J, Varelas X. Integrin-Fak-Cdc42-Pp1a signaling gnaws at Yap/Taz activity to control incisor stem cells. *Bioessays* (2017) 39(10). doi: 10.1002/bies.201700116
86. Amin KS, Banerjee PP. The cellular functions of Rassf1a and its inactivation in prostate cancer. *J Carcinog* (2012) 11. doi: 10.4103/1477-3163.93000
87. Hesson LB, Cooper WN, Latif F. The role of rassf1a methylation in cancer. *Dis Markers* (2007) 23(1-2):73–87. doi: 10.1155/2007/291538
88. Pfeifer G, Dammann R. Methylation of the tumor suppressor gene Rassf1a in human tumors. *Biochem (Moscow)* (2005) 70(5):576–83. doi: 10.1007/s10541-005-0151-y
89. Cheng L, Hill AF. Therapeutically harnessing extracellular vesicles. *Nat Rev Drug Discovery* (2022) 21(5):379–99. doi: 10.1038/s41573-022-00410-w
90. Hu W, Liu C, Bi ZY, Zhou Q, Zhang H, Li LL, et al. Comprehensive landscape of extracellular vesicle-derived rnas in cancer initiation, progression, metastasis and cancer immunology. *Mol Cancer* (2020) 19(1):102. doi: 10.1186/s12943-020-01199-1
91. Huang D, Chen J, Hu D, Xie F, Yang T, Li Z, et al. Advances in biological function and clinical application of small extracellular vesicle membrane proteins. *Front Oncol* (2021) 11:675940. doi: 10.3389/fonc.2021.675940
92. Lytle NK, Barber AG, Reya T. Stem cell fate in cancer growth, progression and therapy resistance. *Nat Rev Cancer* (2018) 18(11):669–80. doi: 10.1038/s41568-018-0056-x
93. Park S, Shim J, Park H, Eum D, Park M, Mi Yi J, et al. Macroh2a1 downregulation enhances the stem-like properties of bladder cancer cells by transactivation of Lin28b. *Oncogene* (2016) 35(10):1292–301. doi: 10.1038/onc.2015.187
94. Garcia-Heredia JM, Lucena-Cacace A, Verdugo-Sivianes EM, Pérez M, Carnero A. The cargo protein Map17 (Pdzk1p1) regulates the cancer stem cell pool activating the notch pathway by abducting numb. *Clin Cancer Res* (2017) 23(14):3871–83. doi: 10.1158/1078-0432.CCR-16-2358
95. Park NI, Guilhamon P, Desai K, McAdam RF, Langille E, O'Connor M, et al. Ascl1 reorganizes chromatin to direct neuronal fate and suppress tumorigenicity of glioblastoma stem cells. *Cell Stem Cell* (2017) 21(2):209–24. e7. doi: 10.1016/j.stem.2017.06.004
96. Fang D, Kitamura H. Cancer stem cells and epithelial-mesenchymal transition in urothelial carcinoma: Possible pathways and potential therapeutic approaches. *Int J Urol* (2018) 25(1):7–17. doi: 10.1111/iju.13404
97. Chen D, Wu M, Li Y, Chang I, Yuan Q, Ekimyan-Salvo M, et al. Targeting Bmi1(+) cancer stem cells overcomes chemoresistance and inhibits metastases in squamous cell carcinoma. *Cell Stem Cell* (2017) 20(5):621–34.e6. doi: 10.1016/j.stem.2017.02.003
98. Li C, Du Y, Yang Z, He L, Wang Y, Hao L, et al. Galnt1-mediated glycosylation and activation of sonic hedgehog signaling maintains the self-renewal and tumor-initiating capacity of bladder cancer stem cells. *Cancer Res* (2016) 76(5):1273–83. doi: 10.1158/0008-5472.CAN-15-2309
99. Jia D, Li L, Andrew S, Allan D, Li X, Lee J, et al. An autocrine inflammatory forward-feedback loop after chemotherapy withdrawal facilitates the repopulation of drug-resistant breast cancer cells. *Cell Death Dis* (2017) 8(7):e2932–e. doi: 10.1038/cddis.2017.319
100. Yang Z, He L, Lin K, Zhang Y, Deng A, Liang Y, et al. The Kmt1a-Gata3-Stat3 circuit is a novel self-renewal signaling of human bladder cancer stem cells. *Clin Cancer Res* (2017) 23(21):6673–85. doi: 10.1158/1078-0432.CCR-17-0882
101. Lee S-R, Roh Y-G, Kim S-K, Lee J-S, Seol S-Y, Lee H-H, et al. Activation of Ezh2 and Suz12 regulated by E2f1 predicts the disease progression and aggressive characteristics of bladder cancer. *Clin Cancer Res* (2015) 21(23):5391–403. doi: 10.1158/1078-0432.CCR-14-2680
102. Yang Z, Li C, Fan Z, Liu H, Zhang X, Cai Z, et al. Single-cell sequencing reveals variants in Arid1a, Gprc5a and Mll2 driving self-renewal of human bladder cancer stem cells. *Eur Urol* (2017) 71(1):8–12. doi: 10.1016/j.eururo.2016.06.025
103. Kripnerova M, Parmar HS, Pesta M, Kohoutova M, Kuncova J, Drbal K, et al. Urothelial cancer stem cell heterogeneity. *Adv Exp Med Biol* (2019) 1139:127–51. doi: 10.1007/978-3-030-14366-4_8
104. Guo Y, Cui J, Ji Z, Cheng C, Zhang K, Zhang C, et al. Mir-302/367/Lats2/Yap pathway is essential for prostate tumor-propagating cells and promotes the development of castration resistance. *Oncogene* (2017) 36(45):6336–47. doi: 10.1038/onc.2017.240

105. Noto A, De Vitis C, Pisanu ME, Roscilli G, Ricci G, Catizone A, et al. Stearoyl-Coa-Desaturase 1 regulates lung cancer stemness via stabilization and nuclear localization of Yap/Taz. *Oncogene* (2017) 36(32):4573–84. doi: 10.1038/onc.2017.75
106. Escoll M, Gargini R, Cuadrado A, Anton I, Wandosell F. Mutant P53 oncogenic functions in cancer stem cells are regulated by wip through Yap/Taz. *Oncogene* (2017) 36(25):3515–27. doi: 10.1038/onc.2016.518
107. Van Den Heuvel MC, Slooff MJ, Visser L, Muller M, De Jong KP, Poppema S, et al. Expression of anti-Ov6 antibody and anti-n-cam antibody along the biliary line of normal and diseased human livers. *Hepatology* (2001) 33(6):1387–93. doi: 10.1053/jhep.2001.24453
108. Wang C, Yang W, Yan HX, Luo T, Zhang J, Tang L, et al. (Hbx) induces tumorigenicity of hepatic progenitor cells in 3, 5-Diethoxycarbonyl-1, 4-Dihydrocollidine-Treated hbx transgenic mice. *Hepatology* (2012) 55(1):108–20. doi: 10.1002/hep.24675
109. Wang C, Yan F-h, Zhang J-j, Huang H, Cui Q-s, Dong W, et al. Ov6+ cancer stem cells drive esophageal squamous cell carcinoma progression through Atg7-dependent B-catenin stabilization. *Cancer Lett* (2017) 391:100–13. doi: 10.1016/j.canlet.2017.01.026
110. Yang W, Wang C, Lin Y, Liu Q, Yu L-X, Tang L, et al. Ov6+ tumor-initiating cells contribute to tumor progression and invasion in human hepatocellular carcinoma. *J Hepatol* (2012) 57(3):613–20. doi: 10.1016/j.jhep.2012.04.024
111. Ooki A, Pena MDCR, Marchionni L, Dinalankara W, Begum A, Hahn NM, et al. Yap1 and Cox2 coordinately regulate urothelial cancer stem-like cells. *Cancer Res* (2018) 78(1):168–81. doi: 10.1158/0008-5472.CAN-17-0836
112. García-Prat L, Martínez-Vicente M, Perdiguero E, Ortet L, Rodríguez-Ubrea J, Rebollo E, et al. Autophagy maintains stemness by preventing senescence. *Nature* (2016) 529(7584):37–42. doi: 10.1038/nature16187
113. Rebutti M, Michiels C. Molecular aspects of cancer cell resistance to chemotherapy. *Biochem Pharmacol* (2013) 85(9):1219–26. doi: 10.1016/j.bcp.2013.02.017
114. Saxena M, Stephens MA, Pathak H, Rangarajan A. Transcription factors that mediate epithelial-mesenchymal transition lead to multidrug resistance by upregulating abc transporters. *Cell Death Dis* (2011) 2(7):e179. doi: 10.1038/cddis.2011.61
115. Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. *Nat Rev Cancer* (2005) 5(4):275–84. doi: 10.1038/nrc1590
116. Tonnessen-Murray CA, Frey WD, Rao SG, Shahbandi A, Ungerleider NA, Olayiwola JO, et al. Chemotherapy-induced senescent cancer cells engulf other cells to enhance their survival. *J Cell Biol* (2019) 218(11):3827–44. doi: 10.1083/jcb.201904051
117. Li C, Du L, Ren Y, Liu X, Jiao Q, Cui D, et al. Skp2 promotes breast cancer tumorigenesis and radiation tolerance through pcd4 ubiquitination. *J Exp Clin Cancer Res* (2019) 38(1):76. doi: 10.1186/s13046-019-1069-3
118. Schulz A, Meyer F, Dubrovskaya A, Borgmann K. Cancer stem cells and radioresistance: DNA repair and beyond. *Cancers (Basel)* (2019) 11(6). doi: 10.3390/cancers11060862
119. Roesch A, Fukunaga-Kalabis M, Schmidt EC, Zabierowski SE, Brafford PA, Vultur A, et al. A temporarily distinct subpopulation of slow-cycling melanoma cells is required for continuous tumor growth. *Cell* (2010) 141(4):583–94. doi: 10.1016/j.cell.2010.04.020
120. Zhou BB, Zhang H, Damelin M, Geles KG, Grindley JC, Dirks PB. Tumour-initiating cells: Challenges and opportunities for anticancer drug discovery. *Nat Rev Drug Discovery* (2009) 8(10):806–23. doi: 10.1038/nrd2137
121. Quintana E, Shackleton M, Sabel MS, Fullen DR, Johnson TM, Morrison SJ. Efficient tumour formation by single human melanoma cells. *Nature* (2008) 456(7222):593–8. doi: 10.1038/nature07567
122. Hall CA, Wang R, Miao J, Oliva E, Shen X, Wheeler T, et al. Hippo pathway effector yap is an ovarian cancer oncogene. *Cancer Res* (2010) 70(21):8517–25. doi: 10.1158/0008-5472.Can-10-1242
123. Zhang X, George J, Deb S, Degoutin J, Takano E, Fox S, et al. The hippo pathway transcriptional co-activator, yap, is an ovarian cancer oncogene. *Oncogene* (2011) 30(25):2810–22. doi: 10.1038/onc.2011.8
124. Huang JM, Nagatomo I, Suzuki E, Mizuno T, Kumagai T, Berezov A, et al. Yap modifies cancer cell sensitivity to egfr and survivin inhibitors and is negatively regulated by the non-receptor type protein tyrosine phosphatase 14. *Oncogene* (2013) 32(17):2220–9. doi: 10.1038/onc.2012.231
125. Zhao Y, Khanal P, Savage P, She YM, Cyr TD, Yang X. Yap-induced resistance of cancer cells to antitubulin drugs is modulated by a hippo-independent pathway. *Cancer Res* (2014) 74(16):4493–503. doi: 10.1158/0008-5472.Can-13-2712
126. Fernandez LA, Squatrito M, Northcott P, Awan A, Holland EC, Taylor MD, et al. Oncogenic yap promotes radioresistance and genomic instability in medulloblastoma through Igf2-mediated akt activation. *Oncogene* (2012) 31(15):1923–37. doi: 10.1038/onc.2011.379
127. Ciamporcero E, Shen H, Ramakrishnan S, Ku SY, Chintala S, Shen L, et al. Yap activation protects urothelial cell carcinoma from treatment-induced DNA damage. *Oncogene* (2016) 35(12):1541–53. doi: 10.1038/onc.2015.219
128. de la Vega MR, Chapman E, Zhang DD. Nrf2 and the hallmarks of cancer. *Cancer Cell* (2018) 34(1):21–43. doi: 10.1016/j.ccell.2018.03.022
129. Satoh H, Moriguchi T, Takai J, Ebina M, Yamamoto M. Nrf2 prevents initiation but accelerates progression through the kras signaling pathway during lung carcinogenesis. *Cancer Res* (2013) 73(13):4158–68. doi: 10.1158/0008-5472.Can-12-4499
130. Wang H, Liu X, Long M, Huang Y, Zhang L, Zhang R, et al. Nrf2 activation by antioxidant antidiabetic agents accelerates tumor metastasis. *Sci Transl Med* (2016) 8(334):334ra51–ra51. doi: 10.1126/scitranslmed.aad6095
131. Singh A, Misra V, Thimmulappa RK, Lee H, Ames S, Hoque MO, et al. Dysfunction Keap1–Nrf2 interaction in non-small-cell lung cancer. *PLoS Med* (2006) 3(10):e420. doi: 10.1371/journal.pmed.0030420
132. Padmanabhan B, Tong KI, Ohta T, Nakamura Y, Scharlock M, Ohtsujii M, et al. Structural basis for defects of Keap1 activity provoked by its point mutations in lung cancer. *Mol Cell* (2006) 21(5):689–700. doi: 10.1016/j.molcel.2006.01.013
133. Cucci MA, Grattarola M, Dianzani C, Damia G, Ricci F, Roetto A, et al. Ailanthone increases oxidative stress in cddp-resistant ovarian and bladder cancer cells by inhibiting of Nrf2 and yap expression through a post-translational mechanism. *Free Radic Biol Med* (2020) 150:125–35. doi: 10.1016/j.freeradbiomed.2020.02.021
134. Kim MH, Kim CG, Kim SK, Shin SJ, Choe EA, Park SH, et al. Yap-induced pd-L1 expression drives immune evasion in brca1-resistant melanoma. *Cancer Immunol Res* (2018) 6(3):255–66. doi: 10.1158/2326-6066.Cir-17-0320
135. Ni W, Mo H, Liu Y, Xu Y, Qin C, Zhou Y, et al. Targeting cholesterol biosynthesis promotes anti-tumor immunity by inhibiting long noncoding rna Snhg29-mediated yap activation. *Mol Ther* (2021) 29(10):2995–3010. doi: 10.1016/j.ymthe.2021.05.012
136. Wu A, Wu Q, Deng Y, Liu Y, Lu J, Liu L, et al. Loss of Vgll4 suppresses tumor pd-L1 expression and immune evasion. *EMBO J* (2019) 38(1). doi: 10.15252/emboj.201899506
137. Stein C, Bardet AF, Roma G, Bergling S, Clay I, Ruchti A, et al. Yap1 exerts its transcriptional control via tead-mediated activation of enhancers. *PLoS Genet* (2015) 11(8):e1005465. doi: 10.1371/journal.pgen.1005465
138. Holden JK, Cunningham CN. Targeting the hippo pathway and cancer through the tead family of transcription factors. *Cancers (Basel)* (2018) 10(3). doi: 10.3390/cancers10030081
139. Liu-Chittenden Y, Huang B, Shim JS, Chen Q, Lee SJ, Anders RA, et al. Genetic and pharmacological disruption of the tead-yap complex suppresses the oncogenic activity of yap. *Genes Dev* (2012) 26(12):1300–5. doi: 10.1101/gad.192856.112
140. Dong L, Lin F, Wu W, Liu Y, Huang W. Verteporfin inhibits yap-induced bladder cancer cell growth and invasion via hippo signaling pathway. *Int J Med Sci* (2018) 15(6):645–52. doi: 10.7150/ijms.23460
141. Elbadawy M, Sato Y, Mori T, Goto Y, Hayashi K, Yamanaka M, et al. Anti-tumor effect of trametinib in bladder cancer organoid and the underlying mechanism. *Cancer Biol Ther* (2021) 22(5-6):357–71. doi: 10.1080/15384047.2021.1919004
142. Shibata M, Hoque MO. Targeting cancer stem cells: A strategy for effective eradication of cancer. *Cancers (Basel)* (2019) 11(5). doi: 10.3390/cancers11050732
143. Dasari VR, Mazack V, Feng W, Nash J, Carey DJ, Gogoi R. Verteporfin exhibits yap-independent anti-proliferative and cytotoxic effects in endometrial cancer cells. *Oncotarget* (2017) 8(17):28628–40. doi: 10.18632/oncotarget.15614
144. Lin KC, Park HW, Guan K-L. Deregulation and therapeutic potential of the hippo pathway in cancer. *Annu Rev Cancer Biol* (2018) 2:59–79. doi: 10.1146/annurev-cancerbio-030617-050202
145. Zhang W, Gao Y, Li P, Shi Z, Guo T, Li F, et al. Vgll4 functions as a new tumor suppressor in lung cancer by negatively regulating the yap-tead transcriptional complex. *Cell Res* (2014) 24(3):331–43. doi: 10.1038/cr.2014.10
146. Guo T, Lu Y, Li P, Yin MX, Lv D, Zhang W, et al. A novel partner of scalloped regulates hippo signaling via antagonizing scalloped-yorkie activity. *Cell Res* (2013) 23(10):1201–14. doi: 10.1038/cr.2013.120
147. Zhang Z, Lin Z, Zhou Z, Shen HC, Yan SF, Mayweg AV, et al. Structure-based design and synthesis of potent cyclic peptides inhibiting the yap-tead protein-protein interaction. *ACS Med Chem Lett* (2014) 5(9):993–8. doi: 10.1021/ml500160m
148. Zhou Z, Hu T, Xu Z, Lin Z, Zhang Z, Feng T, et al. Targeting hippo pathway by specific interruption of yap-tead interaction using cyclic yap-like peptides. *FASEB J* (2015) 29(2):724–32. doi: 10.1096/fj.14-262980

149. Wei X, Jia Y, Lou H, Ma J, Huang Q, Meng Y, et al. Targeting yap suppresses ovarian cancer progression through regulation of the Pi3k/Akt/Mtor pathway. *Oncol Rep* (2019) 42(6):2768–76. doi: 10.3892/or.2019.7370
150. Piccolo S, Dupont S, Cordenonsi M. The biology of Yap/Taz: Hippo signaling and beyond. *Physiol Rev* (2014) 94(4):1287–312. doi: 10.1152/physrev.00005.2014
151. Tang Y, Fang G, Guo F, Zhang H, Chen X, An L, et al. Selective inhibition of Strn3-containing Pp2a phosphatase restores hippo tumor-suppressor activity in gastric cancer. *Cancer Cell* (2020) 38(1):115–28.e9. doi: 10.1016/j.ccell.2020.05.019
152. O'Neill E, Rushworth L, Baccharini M, Kolch W. Role of the kinase Mst2 in suppression of apoptosis by the proto-oncogene product raf-1. *Science* (2004) 306(5705):2267–70. doi: 10.1126/science.1103233
153. Monia BP, Sasmor H, Johnston JF, Freier SM, Lesnik EA, Muller M, et al. Sequence-specific antitumor activity of a phosphorothioate oligodeoxyribonucleotide targeted to human c-raf kinase supports an antisense mechanism of action *in vivo*. *Proc Natl Acad Sci U.S.A.* (1996) 93(26):15481–4. doi: 10.1073/pnas.93.26.15481
154. Khazak V, Astsaturov I, Serebriiskii IG, Golemis EA. Selective raf inhibition in cancer therapy. *Expert Opin Ther Targets* (2007) 11(12):1587–609. doi: 10.1517/14728222.11.12.1587
155. Tolcher AW, Reyno L, Venner PM, Ernst SD, Moore M, Geary RS, et al. A randomized phase ii and pharmacokinetic study of the antisense oligonucleotides Isis 3521 and Isis 5132 in patients with hormone-refractory prostate cancer. *Clin Cancer Res* (2002) 8(8):2530–5.
156. Cripps MC, Figueredo AT, Oza AM, Taylor MJ, Fields AL, Holmlund JT, et al. Phase ii randomized study of Isis 3521 and Isis 5132 in patients with locally advanced or metastatic colorectal cancer: A national cancer institute of canada clinical trials group study. *Clin Cancer Res* (2002) 8(7):2188–92.
157. Oza AM, Elit L, Swenerton K, Faught W, Ghatage P, Carey M, et al. Phase ii study of cgp 69846a (Isis 5132) in recurrent epithelial ovarian cancer: An nci clinical trials group study (Nci Ind.116). *Gynecol Oncol* (2003) 89(1):129–33. doi: 10.1016/s0090-8258(02)00144-0

Glossary

ABCG2	ATP-binding cassette super-family G member 2	ITCH	itchy E3 ubiquitin protein ligase
ADC	antibody-drug coupling	LATS1	Large tumor suppressor kinase 1
ALDH1	Aldehyde dehydrogenase 1 family, member A1	LATS2	Large tumor suppressor kinase 2
AMPK 5'	AMP-activated protein kinase	MAP4K	Mitogen-activated protein kinase kinase kinase kinase
ANKRD1	Ankyrin repeat domain-containing protein 1	MIBC	muscle-invasive bladder cancer
ARID1A	AT-rich interactive domain-containing protein 1A	MINDY1	MINDY lysine 48 deubiquitinase 1
BC	Bladder cancer	MLL2	Histone-lysine N-methyltransferase 2D
BCSCs	bladder cancer stem-like cells	MST1	macrophage-stimulating 1
BMI1	Polycomb complex protein BMI-1	MST2	Serine/threonine-protein kinase 3
CD133	antigen	NMIBC	nonmuscle-invasive bladder cancer
CD44	CD44 antigen	NRF2	Nuclear factor erythroid 2-related factor 2
COX2	prostaglandin-endoperoxide synthase 2	NUAK2	NUAK family SNF1-like kinase 2
CSCs	cancer stem cells	OV6	Ov6 protein
CTGF	connective tissue growth factor	PDGFB	Platelet-derived growth factor subunit B
CYR61	Cysteine-rich angiogenic inducer 61	PDGF-BB	Platelet-derived growth factor subunit B protein
DUB	de-ubiquitin enzyme	PD-L1	Programmed cell death 1 ligand 1
E3	ubiquitin-protein ligases	PGE2	Prostaglandin E2
FBXW7	F-box/WD repeat-containing protein 7	PRAJA1	E3 ubiquitin-protein ligase Praja1
FOXM1	Forkhead box protein M1	RAF	rapidly-accelerated fibrosarcoma
GNA11	Guanine nucleotide-binding protein subunit alpha-11	RASSF1	Ras association domain-containing protein 1
GNA12	Guanine nucleotide-binding protein subunit alpha-12	Rho	GTPase Rho family of GTPases
GNA13	Guanine nucleotide-binding protein subunit alpha-13	RhoGEF	RhoGEF domain
GNAI2	Guanine nucleotide-binding protein G(i), alpha-2 subunit	RhoA	Ras homolog family member A
GNAQ	Guanine nucleotide-binding protein G(q) subunit alpha	RhoB	Ras homolog family member B
GNAS	Heterotrimeric G-protein alpha subunit Gs- α	RhoC	Ras homolog family member C
GPCRs	G-protein-coupled receptors	SHAP	STRN3-derived Hippo-activating peptide
GPRC5A	Retinoic acid-induced protein 3	SIAH2	siah E3 ubiquitin protein ligase 2
GSH	Glutathione	Super-TDU	VGLL4-mimetic peptide
		TAZ	Tafazzin
		TEAD-1	TEA domain family member 1
		TURBT	transurethral resection of bladder tumor
		UPS	ubiquitin proteasomes system
		VGLL4	Vestigial like family member 4
		VP	Verteporfin

WWP1	WW domain containing E3 ubiquitin protein ligase 1	17-peptide	YAP-like peptide
YAP	Yes-associated protein 1		