Check for updates

OPEN ACCESS

EDITED BY Gaspar Kitange, University of Minnesota Twin Cities, United States

REVIEWED BY

Ganapasam Sudhandiran, University of Madras, India Hongbin Li, Lanzhou University of Technology, China

*CORRESPONDENCE Alesia Ivashkevich alesiai@uow.edu.au

SPECIALTY SECTION

This article was submitted to Radiation Oncology, a section of the journal Frontiers in Oncology

RECEIVED 23 October 2021 ACCEPTED 31 August 2022 PUBLISHED 01 March 2023

CITATION

Ivashkevich A (2023) The role of isoflavones in augmenting the effects of radiotherapy. Front. Oncol. 12:800562. [doi: 10.3389/fonc.2022.800562](https://doi.org/10.3389/fonc.2022.800562)

COPYRIGHT

© 2023 Ivashkevich. This is an openaccess article distributed under the terms of the [Creative Commons](http://creativecommons.org/licenses/by/4.0/) [Attribution License \(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

[The role of iso](https://www.frontiersin.org/articles/10.3389/fonc.2022.800562/full)flavones in [augmenting the effects](https://www.frontiersin.org/articles/10.3389/fonc.2022.800562/full) [of radiotherapy](https://www.frontiersin.org/articles/10.3389/fonc.2022.800562/full)

Alesia Ivashkevich $1.2*$

1 Faculty of Engineering and Information Sciences, University of Wollongong, Wollongong, NSW, Australia, 2Noxopharm, Gordon, NSW, Australia

Cancer is one of the major health problems and the second cause of death worldwide behind heart disease. The traditional soy diet containing isoflavones, consumed by the Asian population in China and Japan has been identified as a protective factor from hormone-related cancers. Over the years the research focus has shifted from emphasizing the preventive effect of isoflavones from cancer initiation and promotion to their efficacy against established tumors along with chemo- and radiopotentiating effects. Studies performed in mouse models and results of clinical trials emphasize that genistein or a mixture of isoflavones, containing in traditional soy diet, could be utilized to both potentiate the response of cancer cells to radiotherapy and reduce radiationinduced toxicity in normal tissues. Currently ongoing clinical research explores a potential of another significant isoflavone, idronoxil, also known as phenoxodiol, as radiation enhancing agent. In the light of the recent clinical findings, this article reviews the accumulated evidence which support the clinically desirable interactions of soy isoflavones with radiation therapy resulting in improved tumor treatment. This review discusses important aspects of the development of isoflavones as anticancer agents, and mechanisms potentially relevant to their activity in combination with radiation therapy of cancer. It gives a critical overview of studies characterizing isoflavone targets such as topoisomerases, ENOX2/PMET, tyrosine kinases and ER receptor signaling, and cellular effects on the cell cycle, DNA damage, cell death, and immune responses.

KEYWORDS

Idronoxil (Veyonda/phenoxodiol), ENOX2, topoisomerase, radiation therapy, apoptosis, prostate cancer, ER receptor, isoflavones

Introduction

Cancer represents a major health problem worldwide and its incidence is predicted to increase in the future [\(1\)](#page-10-0). Radiation therapy is a highly cost-effective cancer treatment received by approximately 50% of all cancer patients as a single modality, or in combination with chemotherapy [\(2\)](#page-10-0).

The occurrence and mortality risk of hormone-related cancers including prostate, breast, endometrium, and ovarian cancers is much higher in the United States and Europe compared to Asian countries such as Japan and China [\(3,](#page-10-0) [4](#page-10-0)). Epidemiologic studies have demonstrated that the observed differences are attributed to the traditional diet rich in soy products [\(3](#page-10-0), [5](#page-10-0)) alongside of a broad spectrum of dietary and lifestyle factors ([6,](#page-10-0) [7\)](#page-10-0). It has been estimated that in Asia soy consumption constitutes around 15-50 mg/day, and in western countries 2 mg/day [\(8,](#page-10-0) [9](#page-10-0)). The isoflavones genistein, daidzein, and glycitein - polyphenolic plant-derived compounds - are mainly found in soybeans and these compounds have been shown to be the principal biologically active anticancer molecules [\(10,](#page-10-0) [11\)](#page-10-0). They have been described as non-steroid phytoestrogens that have both estrogenic and antiestrogenic activity. In plants, isoflavones are mostly present in the inactive form as glycosides (genistin and daidzin). Following consumption, isoflavone glycosides are hydrolyzed in intestines by bacterial beta-glucosidases and converted to the corresponding bioactive aglycones such as genistein and daidzein ([12\)](#page-10-0). They are being absorbed from the intestine to blood and conjugated mainly in the liver to glucuronides which are excreted in the urine. Genistein and daidzein are the major isoflavones detected in the blood and urine of humans ([13](#page-10-0)). Human dietary studies demonstrate that -90% of the total amount of genistein exists in either the glucoronidated or sulphated form ([14](#page-10-0)), while only 10% exists in the free nonconjugated from ([15](#page-10-0)), which have different biologic activity ([16,](#page-10-0) [17\)](#page-10-0). Marked differences between humans, rats, and mice in the profile of major metabolites following phase II metabolism were observed ([18](#page-11-0)).

Anticancer activity of isoflavones in hormone-dependent (prostate, breast, ovary, and endometrium) and hormoneindependent cancers (lung, melanoma) has been extensively studied experimentally in in vivo and in vitro cancer models ([19](#page-11-0)–[21\)](#page-11-0). Dietary supplementation with soy significantly prevented spontaneous and chemically induced prostate carcinogenesis in rat models ([22](#page-11-0)–[25](#page-11-0)) and inhibited growth and metastasis of androgen-sensitive human prostate cancers (PC) in an orthotropic mouse model ([26](#page-11-0)). Growth of PC3 and LNCaP cells have been reported to be inhibited independently of their p53 and AR status ([24](#page-11-0), [27](#page-11-0), [28](#page-11-0)). In women, epidemiological data indicate that dietary isoflavone consumption is inversely related with the risk of breast ([20](#page-11-0), [29](#page-11-0), [30\)](#page-11-0), endometrial [\(21\)](#page-11-0), ovarian ([31](#page-11-0)), and cervical cancers ([32](#page-11-0)). Furthermore, chemopreventive properties of isoflavones were explored in rat ([33](#page-11-0)–[36\)](#page-11-0) and mouse models of mammary [\(37,](#page-11-0) [38\)](#page-11-0) and endometrial [\(39\)](#page-11-0) cancers.

In vitro, genistein inhibited proliferation of hormone independent leukaemia [\(40\)](#page-11-0), lymphoma [\(41,](#page-11-0) [42\)](#page-11-0), melanoma ([19\)](#page-11-0), lung [\(43](#page-11-0)), pancreatic [\(42](#page-11-0), [44\)](#page-11-0), gastric ([44\)](#page-11-0), intestinal, hepatic [\(45](#page-11-0)), urinary [\(46\)](#page-11-0), and head and neck cancer cells [\(47](#page-11-0)).

Although anticancer properties of genistein, an active and predominant dietary isoflavone, have been addressed by large

number of studies, more recent research focuses on anticancer activity of Idronoxil, being currently developed under the Veyonda® trademark. Idronoxil is a synthetic isoflavone metabolite that is a natural intermediate (dehydroequol, 7,40 dihydroxyisoflav-2-ene) in the metabolism of daidzein to equol [\(48](#page-11-0)). Analogously to genistein, however more potently, it exhibits anticancer activity in several tumor types as was demonstrated in head and neck [\(49\)](#page-11-0), ovarian [\(50\)](#page-11-0), renal [\(51\)](#page-11-0) cell lines, and prevents carcinogenesis ([52](#page-11-0)). Several clinical trials have addressed Idronoxil activity as an agent potentiating radiation therapy.

Mechanisms of soy isoflavones as anticancer agents

Scientific studies have mainly addressed the mechanisms of antitumor effectiveness of genistein, daidzein and its metabolites phenoxodiol (idronoxil) and equol. All isoflavones display multifaceted cellular responses resulting from their ability to target several essential cellular processes. Interestingly, despite the same origin and some degree of structural similarity, genistein and daidzein appear to be significantly different in their mechanism of action. Equol, the final product of metabolic conversion of daidzein, has also been found to demonstrate differential activities when compared to daidzein. The chemical structure of equol is substantially similar to its precursor phenoxodiol [\(Figure 1\)](#page-2-0). However, no detailed comparative study of its properties to those of phenoxodiol has been performed to date.

Non-steroidal action of isoflavones on ER receptor linked to antitumor effects

Genistein, daidzein, and the daidzein metabolite equol show structural similarity to estradiol-17 β (E2), and exhibit estrogenic properties. It has been demonstrated that soy phytoestrogens and their metabolites bind to and activate both estrogen receptor (ER) α and β [\(53](#page-11-0), [54](#page-11-0)). They have been described to exhibit very diverse pro- and anti-estrogenic effects on ER related cell function in various cell types including immune cells ([55](#page-11-0)–[58\)](#page-11-0). Comparing to estradiol, these compounds possess relatively limited estrogenic potency. For example, equol shows 10^{-3} - 10^{-5} times the activity of 17β -estradiol ([59](#page-11-0), [60\)](#page-11-0). However, their high levels in humans under certain nutritional conditions or pharmacological delivery could induce significant biological effects. The following sequence of the efficacy of activation of ER α has established [\(53](#page-11-0)), with the EC(50): 17 β -estradiol (0.03 μ M) > equol (3.5 μ M) > genistein (15 μ M) > daidzein (>300

 μ M), and for ER β 17 β -estradiol (0.01 μ M) > genistein (0.03 μ M) $>$ daidzein (0.35 µM) $>$ equol (0.4 µM). The results of another study ([61\)](#page-11-0) suggested that in cells with a predominance of $ER-\alpha$ genistein acts as an agonist to ER- α , and in cells with ER- β alone genistein most likely acts as an anti-estrogen.

Action of isoflavones on ER receptors could result in a direct antitumor activity as $ER-\beta$ appears to play an antiproliferative role in healthy prostate ([62](#page-11-0)-[64\)](#page-12-0). Furthermore, $ER-\beta$ was also found to inhibit epithelial proliferation and diminish breast cancer risk and progression ([65](#page-12-0), [66\)](#page-12-0).

Topoisomerase inhibition, DNA damage, and clastogenic potential of isoflavones

Early studies have established that genistein is a potent inhibitor of both topoisomerases I and II [\(67\)](#page-12-0). Subsequently it has been ascribed properties of non-intercalating topoisomerase poison ([67](#page-12-0)–[69\)](#page-12-0). It was found to have an effect on the cleavage activity of DNA topoisomerase II, which resulted in the induction of protein-linked DNA strand breaks ([70\)](#page-12-0). Topoisomerase II-mediated DNA breakage was demonstrated in colorectal cancer cells in response to genistein ([70\)](#page-12-0). Another study found a significant induction of double strand breaks as detected with yH2AX assay in A549 lung cancer cells at 24 and 48 hrs treatment with 30 μM of soy isoflavones. 40-50% of the mixture of soy isoflavones [SIF, extract from soybeans consisting of 43% genistein, 21% daidzein and 2% glycetein [\(71,](#page-12-0) [72](#page-12-0))] treated cells contained several gH2AX foci as detected 24 hours after the start of the treatment ($p < 0.001$). After 48 hrs treatment with SIFs, a more than twofold increase in the number of gH2AX foci and a greater frequency of cells positive for gH2AX foci (60-70%) was observed when compared with the 24

hrs time point $(p < 0.001)$ ([72](#page-12-0)). However, it would have been helpful for accurate data interpretation to take into the account the accumulation of cells in G2/M phase of the cell cycle, as the G2/M cells are characterized by increased numbers of γ H2AX foci ([73\)](#page-12-0). Consistent with DNA damaging properties as determined with γ H2AX detection, clastogenic activity (induction of micronuclei) was demonstrated for genistein at concentrations of 25 μM and above [\(69\)](#page-12-0).

Catalytic topoisomerase II inhibitor activity, rather than topoisomerase II poison activity, was ascribed to daidzein [\(67\)](#page-12-0). Consistent with properties of a non-inducer of DNA damage, daidzein has not caused any significant clastogenicity in NIH 3T3 cells [\(67](#page-12-0)). Analogously to daidzein, equol was found to be much less efficient in induction of DNA strand breakage than genistein [\(74\)](#page-12-0). A study published by Constantinou and Husband, 2002 ([75\)](#page-12-0) have observed inhibition of the catalytic activity of topoisomerase II and stabilization of the topoisomerase II-mediated cleavable complex by idronoxil in a plasmid based cell free assay, indicating its activity as topoisomerase II poison. However, no cell culture studies have been published so far to confirm the findings of idronoxil activity on topoisomerase II in a cell system. In addition, idronoxil exhibits a significant structural similarity to equol (Figure 1), which has not been ascribed topoisomerase inhibitor properties to. Therefore, additional studies are warranted to clarify the potential activity of idronoxil as topoisomerase II poison.

ENOX2 and PMET inhibition by idronoxil

Multiple studies have established a role of idronoxil as an inhibitor of plasma electron transport (PMET) [\(76](#page-12-0)–[78\)](#page-12-0). Idronoxil has been described as a potent inhibitor of a cancer associated

splice variant of ENOX2 – tNOX, oncofetal protein ([76](#page-12-0)), which is re-expressed by cancer cells upon transformation [\(79,](#page-12-0) [80](#page-12-0)). ENOX is a terminal oxidase of the plasma electron transport chain from cytosolic NAD(P)H via hydroquinons to acceptors at the cell surface ([76](#page-12-0)). Idronoxil has been also demonstrated to inhibit PMET in tumor cells and primary immune cells ([77,](#page-12-0) [78](#page-12-0)). However, its PMET inhibitory activity required much higher concentrations (IC50 of 46 μM in proliferating T cells), compared to that required for promotion of apoptosis (5.4 μM) ([79](#page-12-0)). Therefore, it is more likely that idronoxil-induced apoptosis results from ENOX2 targeting, rather than PMET inhibition. More experimental evidence is needed to describe idronoxil effects on ENOX2 and PMET in more detail.

Effects of isoflavones on the cell cycle

For genistein, induction of cell cycle arrest in the G2/M phase was observed, which is consistent with its properties as a potent inhibitor of both topoisomerases I and II [\(68](#page-12-0)). The G2/M cell cycle block occurred in a p53-dependent manner and was found to be associated with activation of ATM/P53, p21 and Gadd45 α [\(81](#page-12-0)). Accumulation of cells in the G2/M phase is favorable for enhancing radiosensitization, as G2/M cells are characterized by increased radiosensitivity [\(82\)](#page-12-0). Both equol and daidzein have been shown to induce an accumulation of prostate cancer cells in the G0/G1 phase of the cell cycle, the most radioresistant cell cycle phase ([83](#page-12-0)). Furthermore, similar patterns of cell cycle responses were observed for idronoxil. Its administration leads to G0/G1 arrest [\(49,](#page-11-0) [84](#page-12-0), [85](#page-12-0)) via p53-independent induction of p21WAF1/ CIP1 [\(49\)](#page-11-0). The potential of idronoxil to induce a G0/G1 block is consistent with its properties as ENOX2/PMET inhibitor as well as catalytic topoisomerase II inhibitor. ENOX2 proteins were ascribed an important role in driving cell enlargement ([86](#page-12-0)). When idronoxil-treated cells fail to reach the size sufficient to pass the G1 checkpoint, they fail to divide, and as a result accumulate in the G0/G1 phase of the cell cycle.

Inhibition of tyrosine kinases

Earlier studies [\(87\)](#page-12-0) in extracellular systems and in cultured A431 cells have demonstrated an activity of genistein as a specific and potent inhibitor of tyrosine protein kinases, such as EGF. Interestingly, the study of genistein effects on serum-, E2-, and EGF-stimulated cell growth has revealed that EGF-R was not targeted by genistein at cytostatic concentrations in breast and prostate cancer cells, and only cytotoxic concentrations (more than 50 μg/ml) achieved a noticeable inhibition of EGF-R tyrosine phosphorylation [\(88,](#page-12-0) [89\)](#page-12-0). Those results might indicate that drug is being metabolized or converted upon cellular entry, which leads to reduction of its intracellular concentrations. It also may selectively inhibit single

sites of tyrosine phosphorylation of the EGF-R or engage with other signal transduction mechanisms below assay detection limit. It was also noticed that the daidzein is a much weaker inhibitor of serum-, E2- and EGF-stimulated cell growth.

The structure-activity relationship studies have revealed that removal of a hydroxyl group from the 5' position as in flavone and daidzein [\(Figure 1](#page-2-0)) has drastically decreased their inhibitory activity on tyrosine kinase as compared to genistein (IC50 = 0.7 μg/ml in case of genistein, and >100 μg/ml for daidzein). Furthermore, the findings of tyrosine kinase inhibitory activity were not correlated with IC50 of growth inhibition of RSVtransformed 3Y1 (RSV3Y1) cells (7 μg/ml versus 25 μg/ml), providing additional evidence in support of different mechanisms of action of these compounds ([90\)](#page-12-0).

AKT is an important target and prognostic marker in prostate cancer and signification overexpression of AKT1, AKT2 and mTOR genes was observed in human prostate cancer samples compared with normal prostate gland tissue [\(91,](#page-12-0) [92](#page-12-0)). Genistein-mediated inhibition of AKT kinase activity, as well as inhibition of NF-kB activation was observed by Li and Sarkar, 2002 [\(26\)](#page-11-0). However, the inhibitory effect on AKT and NF-kB was observed after the treatment with genistein for more than 24 hrs, and might simply result from genistein acting on targets upstream of AKT, or accompany the induction of cell death, therefore having less functional significance.

Furthermore, contradicting findings were reported in regards to the effects of idronoxil on AKT. One report has noticed its inhibition of AKT via dephosphorylation in renal cancer cells [\(50\)](#page-11-0). However, in another study, an increase in phospho-AKT was detected after 6-12 hrs of idronoxil treatment. It was followed by rapid decrease just before the onset of apoptosis, which is not functionally significant [\(93\)](#page-12-0). Moreover, activation of the Akt/FOXO3a pathway with another isoflavone, equol S-isoform has been implicated in inhibition of prostate cancer growth in vitro and in vivo ([94](#page-12-0)).

Isoflavone effects of isoflavones on cell adhesion and tumor metastatic potential

It has been demonstrated in rodent models and in vitro studies that genstein, administered at physiologically relevant concentrations of 10 nM inhibits cell detachment, protease production, cell invasion via inhibition of MMP-2 production [\(95](#page-12-0)–[100](#page-12-0)) and human PCa metastasis [\(101](#page-12-0)) as well as B16BL6 murine melanoma pulmonary metastasis [\(102](#page-12-0)). Inhibition of FAK activation which regulates focal adhesion complex formation and turnover, has been demonstrated by several studies [\(103](#page-12-0)–[105](#page-12-0)). Migration of rat prostate cancer cells, murine breast cancer and melanoma cells was found to be inhibited by genistein in a dose–dependent manner over the range of nontoxic concentrations of 1 to 10 μM [\(106](#page-12-0), [107\)](#page-13-0) possibly via inhibition of FAK phosphorylation ([108\)](#page-13-0).

Cytostatic and cytotoxic activity of isoflavones

The activity of genistein in tumor cells has been characterized as being reversible and cytostatic at the lower concentrations of up to IC50 [\(89,](#page-12-0) [109](#page-13-0)) and becoming more cytotoxic with increase in concentration [\(89,](#page-12-0) [110](#page-13-0)). Another isoflavone, daidzein was found to be less effective than genistein at causing cell death. Up to 100 μM of daidzein was required to cause cytotoxicity comparable to that induced by 30 μM genistein as was determined in PC-3 cells ([111\)](#page-13-0).

In prostate and ovarian cancer cells reduction of cell proliferation caused by idronoxil was accompanied by induction of cell death ([112,](#page-13-0) [113](#page-13-0)). It has been determined that idronoxil is at least five to thirty times more potent than genistein at decreasing proliferation of tumor cells [\(48](#page-11-0), [49](#page-11-0), [112\)](#page-13-0). Furthermore, it has been established as a potent inducer of intrinsic and extrinsic apoptotic pathways operating in a caspase-independent fashion [\(113](#page-13-0), [114](#page-13-0)). The evidence available so far has connected idronoxil to early effectors of cellular responses to stress, namely sphingolipid metabolism ([115\)](#page-13-0) and the PI3K/AKT pathway [\(112](#page-13-0)). The apoptotic effect appears to be linked to an increase in ubiquinol occurring as a result of ENOX2 inhibition, which in turn leads to cytosolic accumulation of NADH and decoupling of the sphingosine-1 phosphate (S1P) pro-survival signal transduction cascade [\(115](#page-13-0), [116\)](#page-13-0). It is accompanied by accumulation of ceramide through NADH-mediated activation of plasma membrane sphingomylelinase [\(117](#page-13-0)). Both events (concurrent reduction in S1P and accumulation of ceramide) appear to initiate cell death. Both genistein and idronoxil have been determined to exert antitumor activity in a p53-independent fashion as has been demonstrated in lung, prostate, and breast cancer cells ([44](#page-11-0), [48](#page-11-0), [118\)](#page-13-0).

Idronoxil potential as autophagy inhibitor has been demonstrated in ovarian clear carcinoma KK cells ([117](#page-13-0)). Treatment with 0.5, 1 and 2 μg of idronoxil for 24 hrs caused inhibition of expression of the Atg7, Atg12 and Beclin 1 autophagy proteins, which was especially noticeable at 2 μg of idronoxil, the highest administered concentration.

The final product of daidzein metabolism, equol, has also shown very promising anticancer effects [\(45\)](#page-11-0) and found to be about ten times more potent than daidzein ([83](#page-12-0)).

Selective targeting of tumor cells by isoflavones

Selectivity of isoflavones towards tumor cells was observed for several cell types. Genistein has not caused any significant induction of apoptosis in spontaneously immortalized 'normal' MCF10A and MCF12A breast epithelial cells in vitro as assessed

after 72 hrs of drug exposure. On the contrary, apoptosis has been strongly induced in a p21-dependent fashion by 90 μM of genistein in malignant MDA-MB-231 and MCF10CA1a breast cancer cells ([118](#page-13-0)). However, this study results are inconclusive as apoptosis was measured only at one time point of 72 hrs. This leaves the possibility open that induction of apoptosis in immortalized breast epithelial cells has occurred at a later time, or cells would undergo proliferative cell death leading to a degree of clonogenicity loss equal to that of malignant cells [\(118](#page-13-0)). The authors have also addressed the effects of genistein on the cell cycle. Pronounced induction of G2/M arrest by genistein was observed in all tested cell lines. The same group has also demonstrated that CRL-2221 prostate cells transformed with human papilloma virus 18 (HPV-18) were less sensitive to genistein than PC-3 prostate tumor cells ([27\)](#page-11-0). In comparison, the earlier study ([83](#page-12-0)), which was performed in true primary prostate epithelium cell cultures, has observed the opposite effect. A range of non-transformed prostate epithelial cells was shown to be even more sensitive to both genistein and equol than tumor cells at day 6 of continuous incubation with concentrations attained as a result of dietary soy uptake in Asian men [\(83\)](#page-12-0).

Genistein effects on proliferation and toxicity was also assessed in normal and tumor blood cells ([109,](#page-13-0) [110](#page-13-0)). Normal proliferating lymphocytes were shown to be resistant to 24 hrs exposure to genistein at concentrations as high as 200 μg/ml, whereas 50% inhibition concentration after 24 h of exposure for HL-60 and MOLT-4 cells was 8.5 and 13.0 μg/ml, respectively [\(109](#page-13-0)). Leukaemic CFU-AML cells were much more sensitive to genistein than normal CFU-GM from either marrow, unseparated blood, or CD34+CD45RA– cells derived from blood [\(110](#page-13-0)).

Several studies have provided consistent indications of enhanced sensitivity of tumor cells to idronoxil. In comparison, wild-type mouse embryonic fibroblasts were found to be resistant to idronoxil [\(117](#page-13-0)). It was also noticed that the potential of idronoxil to inhibit proliferation and cause apoptosis in blood cells was to some extent correlated to the proliferation activity of the studied cells ([78\)](#page-12-0). The isoflavone reduced proliferation and caused cell death in primary stimulated lymphocytes, however the resting lymphocytes were more resistant. Idronoxil was also found to be more cytotoxic in acute lymphocytic leukaemia cells and cell lines, than in activated T cells, where acute myeloid leukaemia cells demonstrated a wide range of sensitivities to it ([78](#page-12-0)). The authors hypothesized, that this effect was related to a idronoxil activity as a PMET inhibitor, as rapidly proliferating cells are known to have high demand in energy. Idronoxil was also found to exert antiangiogenic antiproliferative effects on normal endothelial cells, and no significant toxicity was observed on unstimulated endothelial cells [\(119\)](#page-13-0). However, it could as well result from its hypothesized activity as topoisomerase II inhibitor in cells not expressing ENOX2. This is supported by

the available evidence, which demonstrate that resting T- and Blymphocytes are resistant to activity of topoisomerase inhibitors etoposide and camptothecin, and acquire sensitivity during transition from G1 to S phase of the cell cycle ([120\)](#page-13-0).

Concentration dependent effects

As isoflavones undergo extensive metabolism and bioconjugation, it is important to consider that the concentration offree isoflavones such as genistein is in the low nanomolar range in the blood of soy consumers. Therefore, effects induced by low concentrations potentially represent important clinically relevant mechanisms. Relevance of effects low-to-mid micromolar range is less clear as only total genistein (conjugated plus free forms) can reach micromolar concentrations in the blood after high-dose genistein administration.

Concentration-dependent biphasic activity was described for both genistein and idronoxil across different cell types. At a lower concentration range genistein was ascribed an ability to stimulate estrogen-dependent human breast tumor growth ([121](#page-13-0), [122](#page-13-0)). An ER-dependent biphasic effect was observed for genistein in ER-b expressing MCF7 cells. Increase in cell proliferation was noted at 0.1–10 μM of genistein, whereas concentrations of 20–90 μM have led to suppression of cell proliferation and DNA synthesis [\(123](#page-13-0)). Similarly in CHO-K1 cells characterized by high expression of ER-beta over ER-alpha, single treatment of genistein at physiologically achievable low concentration $\langle 2 \mu M \rangle$ induced proliferation, while high concentrations (50 and 100 μM) resulted in suppression of cell proliferation, G2/M arrest and induction of apoptosis consistent with biphasic estrogenic, anti-estrogenic and DNA topoisomerase II targeting effects of genistein ([124\)](#page-13-0). Furthermore, lower concentrations of genistein appeared to stimulate some immune effects, however it acted as immunosuppressant at higher concentrations. When administered at non-physiologic high concentrations (10–100 mmol/L), genistein was found to inhibit cytotoxic T-cell– mediated tumoricidal activity, alter leukocyte adherence, impair T-cell motility, and inhibit the activation of natural killer (NK) cells in response to lipopolysaccharide or fixed bacteria [reviewed in [\(125](#page-13-0))].

Idronoxil at low concentrations (0.05-0.5 μg/ml) enhanced the in vitro cytotoxic responses of PBMC (peripheral blood mononuclear cells) and was demonstrated to specifically activate by several fold the killing capacity of NK cells [\(126](#page-13-0)). However, at concentrations >1 μg/ml (4 μM) it was found to inhibit proliferation and reduced the viability of healthy donorderived PBMC [\(126\)](#page-13-0).

At high concentrations idronoxil was found to stimulate apoptosis in normal trophoblasts [\(127](#page-13-0)). Idronoxil alone had no effect on neurite outgrowth and length in a concentration range

up to 1 μM, however administration of 10 μM idronoxil resulted in neurite toxicity ([128\)](#page-13-0). The concentrations of 30-200 μM of genistein were reported to be efficient at induction of apoptotic cell death in a range of cancer cell lines ([27](#page-11-0), [43](#page-11-0), [46,](#page-11-0) [47](#page-11-0), [71,](#page-12-0) [129](#page-13-0)–[132\)](#page-13-0).

It is likely that the observed biphasic mode of the isoflavone effects on various cell types, including immune cells, results from the concentration-dependent target(s) activation. For example, effects evoked by lower concentrations of genistein result from its engagement of ER receptor associated pathways, whereas high concentrations achieve topoisomerase II targeting, hence growth inhibition, effects on cell cycle and anti-proliferative action can be observed.

Immune cell activation as antitumor mechanism of isoflavones

It could be hypothesized that most of the systemic and clinically significant antitumor activities of isoflavones might involve activation of immune cells. Tumor targeting effects of phytoestrogens might be explained not only by direct activity towards tumor cells ([36](#page-11-0), [88,](#page-12-0) [133\)](#page-13-0), but also by their indirect action via activation of the immune system. As a result of genistein administration, increased host resistance to B16F10 tumor cells has been observed in mice and was accompanied by dose related increase of cytotoxic T-cell activity and interleukin (IL)-2 stimulated natural killer (NK) cell activity ([134\)](#page-13-0).

Both genistein and dadzein at nutritionally relevant concentrations, genistein at <0.5 μM/L and genistein and dadzein glucuronides at 0.1-10 μM/L, were found to enhance human NK cell toxicity in vitro ([135\)](#page-13-0). Consistent with biphasic dose response, genistein at concentrations higher than > 0.5 μM/ L has significantly inhibited NK cytotoxicity ($P < 0.05$).

In several studies reviewed by Sakai and Kogiso, 2008 ([136\)](#page-13-0) genistein has been shown to enhance the cytotoxic response mediated by NK and cytotoxic T-cells, and T-cell cytokine production. Furthermore, chemopreventative properties of genistein were shown to be associated with induction of antitumor immunity in adult female mice ([137\)](#page-13-0). At the same time it was found to suppress antigen-specific immune response in vivo and lymphocyte proliferation response in vitro [\(136](#page-13-0)).

Georgaki et al, 2009 ([126\)](#page-13-0) has demonstrated that idronoxil is able to enhance the in vitro cytotoxic responses of PBMCs via activation of the killing capacity of NK cells by several fold. It reduced the tumor burden and prolonged survival in a mouse model of colon cancer ([126\)](#page-13-0).

The mechanisms of the observed isoflavone effects on the immune cells are unclear. Isoflavones might exhibit their antitumor activity indirectly, activating immune cells via ER. Immune cells are known to express sex hormone receptors including ER α and ER β , which has been shown to underlie

sex differences in immune cell numbers and/or functional responses of the innate and the adaptive immune systems ([138](#page-13-0)–[140\)](#page-13-0). Estrogen and ER α signaling has been shown to promote type I IFN synthesis [\(141](#page-13-0)–[144\)](#page-13-0), and influence levels of proinflammatory cytokines (IL-12, IL-6, IL-1b), and IL-10 ([145\)](#page-13-0). Activation of ER receptor mediated immune pathways might improve the antitumor responses.

Another important potential mechanism of engaging beneficial antitumor immune responses includes idronoxil alteration of pyridine nucleotide products of plasma membrane redox, NAD+ and NADH, which stimulate sphingomyelinase (SM) and inhibit sphingosine kinase (SK1) which in turn leads to loss of sphingosine-1-phosphate (S1P) ([93](#page-12-0)). Increased SK1 expression has been observed in several tumor types, and in some cases has been correlated with disease progression and reduced patient survival [\(146](#page-13-0)). Furthermore, S1P and its receptors play a role in migration of immune cells ([147,](#page-13-0) [148](#page-13-0)) and SK1 has been recently shown to act as a key regulator of antitumor immunity ([149\)](#page-13-0). SK1 silencing decreased the expression of immunosuppressive factors in the tumor microenvironment which normally limit infiltration with regulatory T cells. Interestingly, the recent study has demonstrated a potential of autophagy inhibition to promote antitumor immunity, which in turn mediated an improved control of distant non-irradiated lesions via systemic type I IFN signaling [\(150](#page-13-0)). Further studies should establish a potential relevance of autophagy inhibiting activity of idronoxil [\(117](#page-13-0)) to its antitumor efficacy when combined with radiation therapy.

Combination effects

Radiation therapy achieves the best success rate in combination with neoadjuvant or adjuvant chemoradiotherapy. The exploration of anticancer activities and mechanisms of cancer prevention by isoflavones has provided evidence of their potential to enhance anticancer therapies.

The ability to modify the radiation response in tumors and normal tissues by a mixture of soy isoflavones and genistein has been observed and explored by several research groups, whereas no radiosensitizing effect was observed with another isoflavone, daidzein ([151\)](#page-14-0).

The activity of idronoxil in combination with cancer therapy focusing on its chemosensitizer properties ([152](#page-14-0)–[156](#page-14-0)) and its ability to overcome chemoresistance to gemcitabine and doxorubicin and sensitize to 5-FU and oxaliplatin-induced apoptosis ([156\)](#page-14-0) was extensively studied in several tumor types including ovarian and osteosarcoma cancer cells. Furthermore, current clinical trials of Veyonda, the suppository formulation of Idronoxil (current name for idronoxil) in combination with external or targeted radiation hold promise to provide a novel potent therapy of end-stage prostate cancer [\(Table 1](#page-7-0)).

Potentiation of radiation toxicity in in vitro studies

Most of the studies have focused on sensitization of cancer cells to radiation with genistein, whereas not much is known about interaction of other isoflavones with irradiation. Pretreatment with genistein prior to irradiation resulted in radiosensitivity enhancement in tested PC-3 (prostate), Me180 and CaSki (cervical cancer), BR-231, MCF-7, MDA-MB-231 (breast), as well as KCI18 and RC-2 (renal cell carcinoma) and sarcoma cancer cell lines [\(151](#page-14-0), [157](#page-14-0), [158\)](#page-14-0). In PC-3 cells genistein appeared to inhibit radiation-induced NF-kB activation and promoted apoptosis as detected by PARP cleavage ([158](#page-14-0)).

Optimization of the drug administration schedule

The timing of drug administration can be crucial for achieving maximal radiosensitization ([159](#page-14-0)). It has been observed that administration of genistein 24 hours prior to irradiation and its presence in the cell culture medium during the exposure resulted in a better radiosensitization effect than when the drug was administered post-irradiation ([151,](#page-14-0) [158\)](#page-14-0). Furthermore, continuous exposure of cells to genistein was found to be required for maximal effect, as cell growth inhibition induced by genistein in combination with radiation was dependent on its presence in the medium during the 10-day incubation of the colony assay [\(7](#page-10-0)). Interestingly, a significant synergistic effect and radiosensitization was observed in H35 hepatoma cells with 30-90 μM of genistein administered only 30 min prior to irradiation, and present in cell culture medium for further 24 hrs post-IR. No effect of genistein on survival of irradiated cells was found when they were pre-incubated for periods of up to 24 hrs ([160\)](#page-14-0).

Potentiation of the radiation effect with genistein in cervical cancer cells Me180 and CaSki was observed with administration of 10 μM of genistein at 48 hrs prior to irradiation ([151\)](#page-14-0). Radiation enhancement ratios, defined as survival with irradiation and genistein compared to radiation alone, were found to vary from 1.4 to 4.4 over the range of doses. 10 μM of genistein was found to be mildly cytotoxic in cell growth assays. Induction of G2/M arrest by genistein was observed only in Me180 cells, and some accumulation of cells in S phase was observed in CaSki cells. The authors found significant inhibition of Mcl-1 by genistein which correlated with an increase in radiosensitivity in Me180 cells. Inhibition of activated pAKT (Thr 308) with genistein and radiation was noticed in CaSki cells. Using the dosage of $10 \mu M$ genistein, the sensitizer enhancement ratios after exposure to X-rays at a 10% cell survival (IC10) were established to 1.43 for MCF-7 and 1.36 for MDA-MB-231 cells, respectively ([157,](#page-14-0) [158](#page-14-0)).

TABLE 1 Completed and ongoing clinical trials of isoflavones in combination with radiation therapy.

A significant potentiation of radiation cytotoxicity by genistein was observed in HL-60 leukaemia cells ([161\)](#page-14-0). Exposure to 5 Gy alone has caused an increase in sub-G0/G1 population up to 21.2% indicating cell death at 48 hrs postirradiation. Pre-treatment with 20 μM of genistein alone resulted in 27.5%, and combination with irradiation has led to accumulation of about 60% cells in sub-G0/G1 cell population. The authors have also found that genistein exerted a somewhat radioprotective effect on normal lymphocytes ([161\)](#page-14-0). In another study [\(162](#page-14-0)) a significant protection of B-lymphocyte precursors by genistein was observed after administration at 30 μg/ml 30 min prior to irradiation and continuous incubation for 24 hrs.

Effects of isoflavones on repair of radiation-induced DNA damage

A study published by Singh-Gupta et al., 2011 [\(72\)](#page-12-0) demonstrated that SIF potentiated DNA damage induction by irradiation in human A549 non-small cell lung cancer cells. Preincubation with SIF for 24 hrs prior to exposure to 3 Gy has resulted in induction of 21.15 ± 2.38 of γ H2AX foci/cell, which is almost double of foci induced by IR alone at 1 hr post-exposure (13.86 \pm 2.31 of γ H2AX foci per cell), or four times of foci resulted from incubation of cells with SIF for 24 hrs (4.96 ± 1.02) foci per cell). The effects of SIF on the functioning in the base excision repair pathway have also been investigated in this study.

The authors suggest that the observed potentiation of residual DNA damage with SIF occurs via inhibition of the repair activity of apurinic/apyrimidinic endonuclease 1/redox factor-1 (APE1/ Ref-1), the main player in the cellular response to DNA damage and redox regulation against oxidative stress, by analogue with methoxyamine ([72\)](#page-12-0). The mode of interaction between isoflavones and radiation cannot be reliably determined as 3 Gy of irradiation administered alone resulted in 80% reduction in survival, respectively ([72\)](#page-12-0).

Study by Liu et al, 2013 ([157](#page-14-0)) demonstrated that pretreatment of MCF-7 (ER-positive, harboring wild-type p53) and MDA-MB-231 (ER-negative, harboring mutant p53) cells with genistein for 24 hrs followed by 4 Gy of X-rays resulted in increased number of the γ -H2AX foci at 12 h post-irradiation and decreased homologous recombination repair protein Rad51 foci formation. Genistein has significantly potentiated accumulation of cells in radiosensitive G2/M phase of the cell cycle phase and apoptosis.

Taken together, the published results of cell culture experiments and pre-clinical models indicate the potential of soy isoflavones to modify the radiation response of tumor cells. Ideally, the radiosensitization is defined as increased radiation sensitivity of cells in the absence of significant drug-induced cytotoxicity. In the case of genistein, most published studies show describe enhancement of radiation effects by the soy isoflavone administered at weakly cytotoxic concentrations in the range of 5–15 μM. It should also be noted that concentrations of genistein in this range are considered mild and being close to physiological doses achieved as a result of dietary supplementation [\(14](#page-10-0), [163\)](#page-14-0).

Effects of isoflavones on radiationinduced invasion and migration

Recent publication by Liu et al, 2021 ([164\)](#page-14-0) has demonstrated the potential of genistein to inhibit radiation-induced invasion and migration of DNA-PKcs-positive glioblastoma cells. The authors have observed interaction of genistein with DNA-PKcs which resulted in blocking DNA-PKcs/Akt2/Rac1 signaling pathway induced by radiation in DNA-PKcs-positive but not in negative glioblastoma cells and established the exact binding site of genistein to DNA-PKcs. The physiological relevance of these findings remains to be further addressed.

Preclinical models of radiosensitization with isoflavones

Several research groups have explored the aspects of interaction of genistein and soy products with irradiation in pre-clinical mouse models. Potentiation of radiation inhibition of tumor growth by genistein has been observed in the

orthotopic nude mouse model of prostate cancer [\(165](#page-14-0)). When 5 mg/day of genistein was administered orally for 2 days prior to exposure to 5 Gy, and every other day for 4 weeks postirradiation, it led to greater inhibition of primary tumor growth (87%) compared with genistein (30%) or irradiation (73%) alone, and improved survival. No signs of any toxicity which potentially could be associated with genistein treatment were observed ([165\)](#page-14-0). Intriguingly, when genistein was used as a single treatment, it promoted metastasis to regional para-aortic lymph nodes. This metastasis-promoting activity was also observed in a mouse model of renal cancer [\(166](#page-14-0)). Treatment of established kidney tumors with genistein demonstrated a tendency to stimulate the growth of the primary kidney tumor and increase the incidence of metastasis. However, when given in conjunction with radiation therapy, genistein significantly inhibited the growth and progression of established kidney tumors ([166\)](#page-14-0). These results confirm the potentiation of radiotherapy by genistein. Raffoul et al., 2007 have demonstrated that administration of soy isoflavones in the nude mouse model of prostate cancer with implanted PC-3 cells mediated reduction of the prostate weight from 420 mg to 250 mg, and in combination with irradiation from 100 mg to 50 mg [\(167](#page-14-0)). In contrast to the studies where administration of pure genistein promoted metastases ([165,](#page-14-0) [166\)](#page-14-0), no such prometastatic effect was observed with combination of soy isoflavones. Subsequently, it has been determined in the PC-3 prostate tumor model that dadzein is a compound in the SIF mixture capable of negating genistein-induced metastasis ([111\)](#page-13-0). Another study confirmed that daidzein and its derivative equol inhibit the cancer cell invasion in breast cancer cells potentially via down-regulation of matrix metalloproteinase-2, a major matrix degrading enzyme ([168\)](#page-14-0). Interestingly, the angiogenic properties of idronoxil were demonstrated, too and occurred via inhibition of endothelial cell proliferation, migration and capillary tube formation as well as inhibition of expression of the matrix metalloproteinase MMP-2 [\(118](#page-13-0)).

Radioprotective properties of isoflavones explored in pre-clinical models

Numerous studies have described the radioprotective effect of genistein in mouse models of radiation-induced toxicity.

In male mice genistein mitigated radiation-induced testicular dysfunction by an anti-apoptotic effect and recovery of spermatogenesis ([169\)](#page-14-0). Induction of IL-6 and serum granulocyte-colony stimulating factor ([170,](#page-14-0) [171\)](#page-14-0) and increased hematopoiesis [\(172\)](#page-14-0) was found to be associated with genisteinmediated protection of irradiated mice. Davis et al., 2007 have shown a significant radioprotection of the hematopoietic progenitor cell compartment by a single subcutaneous administration of genistein 24 hrs prior to irradiation via induction of quiescence in hematopoietic stem cells [\(173](#page-14-0), [174\)](#page-14-0).

Another study published by Day, et al. ([175](#page-14-0)) has also demonstrated significant radioprotective properties of genistein in normal tissues. 200 mg/kg genistein delivered subcutaneously protected from acute radiation injury in mice after total body exposure to 7.75 Gy. The authors observed that at 30 days genistein has led to improved survival from 23-53% to 92%, accompanied by reduced lung damage. Mitigation of the radiation induced lung injury for a period of up to 28 weeks was demonstrated by Mahmood et al., 2011 ([176\)](#page-14-0). A study led by Hillman [\(177](#page-14-0)) has further explored the exact mechanisms of radioprotective effects of genistein in normal tissues. It has been established that pre-treatment of mice with SIF for three days prior to lung irradiation, and continuous treatment at various doses for 18 weeks post-exposure resulted in reduction in infiltration and activation of alveolar macrophages and neutrophils in both the bronchoalveolar and lung parenchyma compartments. Soy treatment protected the F4/80+CD11cimmunoregulatory interstitial macrophages, which are normally decreased by irradiation, and led to switch from a pro-inflammatory M1 macrophage to an anti-inflammatory M2 macrophage phenotype [\(177](#page-14-0)).

Using pre-clinical mouse cancer models, studies on the combination of soy isoflavones with tumor irradiation demonstrate a significant anti-cancer effect by these two modalities and indicate the potential and safety of dietary factors to improve conventional radiotherapy for a better control of tumor.

Clinical trials

Combination of isoflavones with radiation therapy yielded good indications of its efficacy and safety in clinical trial in localized prostate cancer [\(178\)](#page-14-0), the most common cancer in men ([179\)](#page-14-0). Despite high level of diagnosis, the relatively successful treatment of localized disease with surgery or radiotherapy results in the 5-year overall survival rate above 90% [\(179](#page-14-0)). However, treatment morbidity leads to worsened quality of life. The results of the pilot clinical study which was published in 2010 by Ahmad et al. ([178\)](#page-14-0), suggested that soy isoflavones taken in conjunction with curative radiation therapy could reduce the urinary, intestinal, and sexual adverse effects in patients with localized prostate cancer. 200 mg soy isoflavone or placebo were administered orally daily in a group of 42 patients for 6 months beginning with the first day of radiation therapy, which was given in 1.8 to 2.5 Gy fractions for a total of 73.8 to 77.5 Gy. At 3 months, soy-treated patients had less urinary incontinence, less urgency and better erectile function compared to the placebo group, and at later time point of 6 months the symptoms in the soy-treated group had further improved. Patients experienced less dripping/leakage of urine (7.7% in Group 1 vs. 28.4% in Group 2), less rectal cramping/ diarrhea (7.7% vs. 21.4%), and less pain with bowel movements

(0% vs. 14.8%) than placebo-treated patients. Another clinical study in pediatric cancer patients has demonstrated that genistein reduced the adverse effects of abdominal radiotherapy. Patients experienced less pain and no diarrhea, which are common side effects of abdominal radiation ([180\)](#page-14-0).

However, there is limited clinical data available on isoflavone antitumor efficacy when combined with radiation therapy [\(Table 1\)](#page-7-0). Several clinical trials have investigated the efficiency of idronoxil in combination with irradiation in late stage metastatic chemotherapy resistant prostate cancer (mCRPC) patients and in head and neck cancer patients ([Table 1\)](#page-7-0) [\(181](#page-14-0)– [185](#page-14-0)). PoC and dose confirmation DARRT study in late-stage prostate cancer patients has addressed safety and potential indications of efficacy of the combination of 1 of 3 doses of idronoxil, given as suppository (NOX66) in dose-escalated fashion (400mg, 800mg and 1200mg) in combination with 20Gy of EBRT given over 5 daily fractions to selected target lesion/s for all cohorts.

The combination of idronoxil with a novel promising treatment for late-stage prostate cancer $[^{177}$ Lu]Lu-PSMAtargeted radionuclide therapy with is also being explored as part of the LuPIN clinical trial [\(182](#page-14-0)–[185\)](#page-14-0). Recently published results ([184,](#page-14-0) [185\)](#page-14-0) of phase I/II study in 56 men with progressive mCRPC with extensive prior treatment which included docetaxel and cabazitaxel in addition to abitraterone and/or enzalutamide indicated safety and tolerability of NOX66 (Idronoxil) delivered at doses up to 1200 mg in suppository form. Suppositories were given on days 1-10 of a 6-week cycle, where 7.5 GBq of $\left[{}^{177}$ Lu $\right]$ Lu-PSMA-617 was delivered on day 1 of a 6-week cycle. 86% of patients have experienced a >50% PSA decline, which was higher than reported in prospective study of [177Lu]Lu-PSMA-617 (56% of PSA decline) ([186](#page-14-0)–[188](#page-14-0)), and the randomized TheraP trial (66%) ([185,](#page-14-0) [190\)](#page-14-0). Pain reduction was observed in more than half of the men. The median overall survival (OS) was found to be 19.7 months (95% CI 9.5– 30 mo), whereas previously reported median OS in study of $[^{177}$ Lu]Lu-PSMA-617 efficacy in mCRPC was found to be 13.3 months (95% CI 10.5-18.7) [\(189](#page-14-0)). These results indicate a potential of NOX66 to improve survival outcome in mCRPC patients. Further research is required to better evaluate the efficacy of NOX66 and LuPSMA-617 combination.

Conclusions

Unique property of soy isoflavones specifically genistein to target tumor cells while sparing normal tissues when combined with radiation have been addressed in a number studies. Showing some structural resemblance, yet causing significantly different biological effects, other isoflavones such as idronoxil are currently being successfully explored to clinical benefit in combination therapies.

Identification and characterization of isoflavone targets has been a continuous endeavor spanning over decades. Activity of isoflavones as protein kinase inhibitors and DNA topoisomerase II inhibitors/poisons has been extensively investigated. However, it might have limited clinical implications as it occurs at supraphysiological concentrations, which are manyfold greater than concentrations that can be obtained through consumption of dietary genistein or even through the use of supplements containing genistein. Limited available evidence do not support kinases as direct targets of idronoxil, and not much is known in regards to its activity on topoisomerases. This leaves ER and ENOX2/PMET as well described potential molecular targets mediating cellular effects of idronoxil in tumor and normal cells. However the activity of idronoxil towards other targets cannot be excluded.

The S1P inhibition properties resulting from ENOX2/PMET targeting in combination with potent apoptosis inducer characteristics of idronoxil hold great promise in indirectly potentiating antitumor therapies via engagement with immune cells. There is also an array of other biological actions of isoflavones including those on cell communications ([191](#page-14-0)) and mitochondrial biogenesis, which need to be better understood.

Completed and ongoing clinical trials are set to explore the clinical utility of phenoxodiol in combination with different forms of radiation therapy, especially targeting patients with unmet need for treatment options. Combination of soy isoflavones with radiotherapy appears to hold the most therapeutical promise, however, the precise mechanisms and molecular targets need to be better understood.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. Cancer J Clin (2018) 68(6):394–424. doi: [10.3322/caac.21492](https://doi.org/10.3322/caac.21492)

2. Delaney G, Jacob S, Featherstone C, Barton M. The role of radiotherapy in cancer treatment: Estimating optimal utilization from a review of evidence-based clinical guidelines. Cancer (2005) 104(6):1129–37. doi: [10.1002/cncr.21324](https://doi.org/10.1002/cncr.21324)

3. Sarkar FH, Li Y. The role of isoflavones in cancer chemoprevention. Front Biosci (2004) 9:2714–24. doi: [10.2741/1430](https://doi.org/10.2741/1430)

4. Giovannucci E. Epidemiologic characteristics of prostate cancer. Cancer (1995) 75(S7):1766–77. doi: [10.1002/1097-0142\(19950401\)75:7+<1766::AID-](https://doi.org/10.1002/1097-0142(19950401)75:7+%3C1766::AID-CNCR2820751604%3E3.0.CO;2-T)[CNCR2820751604>3.0.CO;2-T](https://doi.org/10.1002/1097-0142(19950401)75:7+%3C1766::AID-CNCR2820751604%3E3.0.CO;2-T)

5. Applegate CC, Rowles JL, Ranard KM, Jeon S, Erdman JW. Soy consumption and the risk of prostate cancer: An updated systematic review and meta-analysis. Nutrients (2018) 10(1). doi: [10.3390/nu10010040](https://doi.org/10.3390/nu10010040)

6. Adlercreutz H. Western Diet and Western diseases: Some hormonal and biochemical mechanisms and associations. Scand J Clin Lab Invest Suppl (1990) 201:3–23. doi: [10.1080/00365519009085798](https://doi.org/10.1080/00365519009085798)

7. Severson RK, Nomura AM, Grove JS, Stemmermann GN. A prospective study of demographics, diet, and prostate cancer among men of Japanese ancestry
in Hawaii. *Cancer Res* (1989) 49(7):1857–60.

8. Bedell S, Nachtigall M, Naftolin F. The pros and cons of plant estrogens for menopause. J Steroid Biochem Mol Biol (2014) 139:225–36. doi: [10.1016/](https://doi.org/10.1016/j.jsbmb.2012.12.004) [j.jsbmb.2012.12.004](https://doi.org/10.1016/j.jsbmb.2012.12.004)

9. Rietjens IM, Sotoca AM, Vervoort J, Louisse J. Mechanisms underlying the dualistic mode of action of major soy isoflavones in relation to cell proliferation and cancer risks. Mol Nutr Food Res (2013) 57(1):100–13. doi: [10.1002/mnfr.201200439](https://doi.org/10.1002/mnfr.201200439)

Author contributions

The author confirms being the sole contributor of this work and has approved it for publication.

Funding

This study received funding from Noxopharm Limited. The funder was not involved in the study design, collection, analysis, or interpretation of data. Noxopharm reviewed the article and agreed to publish it. The author declares no other competing interests.

Conflict of interest

AI is/was employed by Noxopharm Limited.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

10. Lampe JW. Isoflavonoid and lignan phytoestrogens as dietary biomarkers. J Nutr (2003) 133 Suppl 3:956s–64s. doi: [10.1093/jn/133.3.956](https://doi.org/10.1093/jn/133.3.956)

11. Izumi T, et al, Piskula MK, Osawa S, Obata A, Tobe K, Saito M. Soy isoflavone aglycones are absorbed faster and in higher amounts than their glucosides in humans. J Nutr (2000) 130(7):1695–9. doi: [10.1093/jn/130.7.1695](https://doi.org/10.1093/jn/130.7.1695)

12. Coldham NG, Darby C, Hows M, King LJ, Zhang AQ, Sauer MJ, et al. Comparative metabolism of genistin by human and rat gut microflora: detection and identification of the end-products of metabolism. Xenobiotica (2002) 32(1):45– 62. doi: [10.1080/00498250110085809](https://doi.org/10.1080/00498250110085809)

13. Kelly GE, Nelson C, Waring MA, Joannou GE, Reeder AY. Metabolites of dietary (soya) isoflavones in human urine. Clin Chim Acta (1993) 223(1-2):9–22. doi: [10.1016/0009-8981\(93\)90058-C](https://doi.org/10.1016/0009-8981(93)90058-C)

14. Adlercreutz H, Markkanen H, Watanabe S. Plasma concentrations of phytooestrogens in Japanese men. Lancet (1993) 342(8881):1209–10. doi: [10.1016/0140-](https://doi.org/10.1016/0140-6736(93)92188-Y) [6736\(93\)92188-Y](https://doi.org/10.1016/0140-6736(93)92188-Y)

15. Pavese JM, Farmer RL, Bergan RC. Inhibition of cancer cell invasion and metastasis by genistein. Cancer Metastasis Rev (2010) 29(3):465–82. doi: [10.1007/](https://doi.org/10.1007/s10555-010-9238-z) [s10555-010-9238-z](https://doi.org/10.1007/s10555-010-9238-z)

16. Murata M, Midorikawa K, Koh M, Umezawa K, Kawanishi S. Genistein and daidzein induce cell proliferation and their metabolites cause oxidative DNA damage in relation to isoflavone-induced cancer of estrogen-sensitive organs. Biochemistry (2004) 43(9):2569–77. doi: [10.1021/bi035613d](https://doi.org/10.1021/bi035613d)

17. Pugazhendhi D, Watson KA, Mills S, Botting N, Pope GS, Darbre PD. Effect of sulphation on the oestrogen agonist activity of the phytoestrogens genistein and daidzein in MCF-7 human breast cancer cells. J Endocrinol (2008) 197(3):503–15. doi: [10.1677/JOE-07-0384](https://doi.org/10.1677/JOE-07-0384)

18. Soukup ST, Helppi J, Mueller DR, Zierau O, Watzl B, Vollmer G. Phase II metabolism of the soy isoflavones genistein and daidzein in humans, rats and mice: a cross-species and sex comparison. Arch Toxicol (2016) 90(6):1335-47. doi: [10.1007/s00204-016-1663-5](https://doi.org/10.1007/s00204-016-1663-5)

19. Yan CH, Ghen XG, Han R. Effects of genistein, a soybean-derived isoflavone, on proliferation and differentiation of B16-BL6 mouse melanoma cells. J Asian Nat Prod Res (1999) 1(4):285–99. doi: [10.1080/10286029908039877](https://doi.org/10.1080/10286029908039877)

20. Adlercreutz CH, Goldin BR, Gorbach SL, Hoeckerstedt KA, Watanabe S, Haemaelaeinen EK, et al. Soybean phytoestrogen intake and cancer risk. *J Nutr*
(1995) 125(3 Suppl):757s–70s. doi: [10.1093/jn/125.3_suppl.757s](https://doi.org/10.1093/jn/125.3_suppl.757s)

21. Horn-Ross PL, John EM, Canchola AJ, Stewart SL, Lee MM. Phytoestrogen intake and endometrial cancer risk. J Natl Cancer Inst (2003) 95(15):1158–64. doi: [10.1093/jnci/djg015](https://doi.org/10.1093/jnci/djg015)

22. Pollard M, Wolter W. Prevention of spontaneous prostate-related cancer in lobund-wistar rats by a soy protein isolate/isoflavone diet. Prostate (2000) 45 (2):101–5. doi: [10.1002/1097-0045\(20001001\)45:2<101::AID-PROS3>3.0.CO;2-P](https://doi.org/10.1002/1097-0045(20001001)45:2%3C101::AID-PROS3%3E3.0.CO;2-P)

23. Wang J, Eltoum IE, Lamartiniere CA. Dietary genistein suppresses chemically induced prostate cancer in lobund-wistar rats. Cancer Lett (2002) 186 (1):11–8. doi: [10.1016/S0304-3835\(01\)00811-4](https://doi.org/10.1016/S0304-3835(01)00811-4)

24. Davis JN, Singh B, Bhuiyan M, Sarkar FH. Genistein-induced upregulation of p21WAF1, downregulation of cyclin b, and induction of apoptosis in prostate cancer cells. Nutr Cancer (1998) 32(3):123–31. doi: [10.1080/01635589809514730](https://doi.org/10.1080/01635589809514730)

25. Onozawa M, Kawamori T, Baba M, Fukuda K. Effects of a soybean isoflavone mixture on carcinogenesis in prostate and seminal vesicles of F344 rats. Jpn J Cancer Res (1999) 90(4):393–8. doi: [10.1111/j.1349-7006.1999.tb00760.x](https://doi.org/10.1111/j.1349-7006.1999.tb00760.x)

26. Zhou JR, Yu L, Zhong Y, Nassr R, Franke A, Gaston S, et al. Inhibition of orthotopic growth and metastasis of androgen-sensitive human prostate tumors in mice by bioactive soybean components. Prostate (2002) 53(2):143–53. doi: [10.1002/](https://doi.org/10.1002/pros.10141) [pros.10141](https://doi.org/10.1002/pros.10141)

27. Li Y, Sarkar FH. Inhibition of nuclear factor kappaB activation in PC3 cells by genistein is mediated via akt signaling pathway. Clin Cancer Res (2002) 8 (7):2369–77.

28. Davis JN, Kucuk O, Sarkar FH. Genistein inhibits NF-kappa b activation in prostate cancer cells. Nutr Cancer (1999) 35(2):167–74. doi: [10.1207/](https://doi.org/10.1207/S15327914NC352_11) [S15327914NC352_11](https://doi.org/10.1207/S15327914NC352_11)

29. Deapen D, Liu L, Perkins C, Bernstein L, Ross R. Rapidly rising breast cancer incidence rates among Asian-American women. Int J Cancer (2002) 99(5):747–50. doi: [10.1002/ijc.10415](https://doi.org/10.1002/ijc.10415)

30. Ziegler RG, Hoover RN, Pike MC, Hildersheim A, Nomura AM, West DW, et al. Migration patterns and breast cancer risk in Asian-American women. J Natl Cancer Inst (1993) 85(22):1819–27. doi: [10.1093/jnci/85.22.1819](https://doi.org/10.1093/jnci/85.22.1819)

31. Lee JY, Kim HS, Song YS. Genistein as a potential anticancer agent against ovarian cancer. J Tradit Complement Med (2012) 2(2):96–104. doi: [10.1016/S2225-](https://doi.org/10.1016/S2225-4110(16)30082-7) [4110\(16\)30082-7](https://doi.org/10.1016/S2225-4110(16)30082-7)

32. Paul P, Koh W-P, Jin A, Michel A, Waterboer T, Pawlita M, Wang R, et al, et al. Soy and tea intake on cervical cancer risk: the Singapore Chinese health study. Cancer Causes Control (2019) 30(8):847–57. doi: [10.1007/s10552-019-01173-3](https://doi.org/10.1007/s10552-019-01173-3)

33. Barnes S. The chemopreventive properties of soy isoflavonoids in animal models of breast cancer. Breast Cancer Res Treat (1997) 46(2-3):169–79. doi: [10.1023/A:1005956326155](https://doi.org/10.1023/A:1005956326155)

34. Hawrylewicz EJ, Zapata JJ, Blair WH. Soy and experimental cancer: animal studies. J Nutr (1995) 125(3 Suppl):698s–708s. doi: [10.3390/ijms160511728](https://doi.org/10.3390/ijms160511728)

35. Gotoh T, Yamada K, Yin H, Ito A. Chemoprevention of n-nitroso-Nmethylurea-induced rat mammary carcinogenesis by soy foods or biochanin a. Jpn J Cancer Res (1998) 89(2):137–42. doi: [10.1111/j.1349-7006.1998.tb00541.x](https://doi.org/10.1111/j.1349-7006.1998.tb00541.x)

36. Constantinou AI, Mehta RG, Vaughan A. Inhibition of n-methyl-Nnitrosourea-induced mammary tumors in rats by the soybean isoflavones. Anticancer Res (1996) 16(6a):3293–8.

37. Mizunuma H, Kanazawa K, Ogura S, Otsuka S, Nagai H, et al. Anticarcinogenic effects of isoflavones may be mediated by genistein in mouse mammary tumor virus-induced breast cancer. Oncology (2002) 62(1):78-84. doi: [10.1159/000048250](https://doi.org/10.1159/000048250)

38. Constantinou AI, Krygier AE, Mehta RR. Genistein induces maturation of cultured human breast cancer cells and prevents tumor growth in nude mice. Am J
Clin Nutr (1998) 68(6 Suppl):1426s–30s. doi: [10.1093/ajcn/68.6.1426S](https://doi.org/10.1093/ajcn/68.6.1426S)

39. Lian Z, Niwa K, Tagami K, Hashimoto M, Gao J, Yokoyama Y, et al. Preventive effects of isoflavones, genistein and daidzein, on estradiol-17beta-related endometrial carcinogenesis in mice. Jpn J Cancer Res (2001) 92(7):726-34. doi: [10.1111/j.1349-7006.2001.tb01154.x](https://doi.org/10.1111/j.1349-7006.2001.tb01154.x)

40. Uckun FM, Evans WE, Forsyth CJ, Waddick KG, et al. Biotherapy of b-cell precursor leukemia by targeting genistein to CD19-associated tyrosine kinases.
S*cience* (1995) 267(5199):886–91. doi: [10.1126/science.7531365](https://doi.org/10.1126/science.7531365)

41. Buckley AR, Buckey DJ, Gout PW, Liang H, Rao YP, Blake MJ. Inhibition by genistein of prolactin-induced Nb2 lymphoma cell mitogenesis. Mol Cell Endocrinol (1993) 98(1):17–25. doi: [10.1016/0303-7207\(93\)90231-8](https://doi.org/10.1016/0303-7207(93)90231-8)

42. Buchler P, Gukovskaya AS, Mouria M, Buechler MC, Buechler MW, Friess H, et al. Prevention of metastatic pancreatic cancer growth in vivo by induction of apoptosis with genistein, a naturally occurring isoflavonoid. Pancreas (2003) 26 (3):264–73. doi: [10.1097/00006676-200304000-00010](https://doi.org/10.1097/00006676-200304000-00010)

43. Lian F, Li Y, Bhuiyan M, Sarkar FH. p53-independent apoptosis induced by genistein in lung cancer cells. *Nutr Cancer* (1999) 33(2):125–31. doi: [10.1207/](https://doi.org/10.1207/S15327914NC330202)
[S15327914NC330202](https://doi.org/10.1207/S15327914NC330202)

44. Yanagihara K, Ito A, Toge T, Numoto M. Antiproliferative effects of isoflavones on human cancer cell lines established from the gastrointestinal tract. Cancer Res (1993) 53(23):5815–21.

45. Lei B, Roncaglia V, Vigano R, Cremonini C, De Maria N, Del Buono MG, et al. Phytoestrogens and liver disease. Mol Cell Endocrinol (2002) 193(1-2):81–4. doi: [10.1016/S0303-7207\(02\)00099-0](https://doi.org/10.1016/S0303-7207(02)00099-0)

46. Su SJ, Yeh TM, Lei HY, Chow NH. The potential of soybeanfoods as a chemoprevention approach for human urinary tract cancer. Clin Cancer Res (2000) 6(1):230–6.

47. Alhasan SA, Pietrasczkiwicz H, Alonso MD, Ensley J, Sarkar FH. Genisteininduced cell cycle arrest and apoptosis in a head and neck squamous cell carcinoma cell line. Nutr Cancer (1999) 34(1):12–9. doi: [10.1207/S15327914NC340102](https://doi.org/10.1207/S15327914NC340102)

48. Joannou GE, Kelly GE, Reeder AY, Waring M, Nelson C. A urinary profile study of dietary phytoestrogens. the identification and mode of metabolism of new isoflavonoids. J Steroid Biochem Mol Biol (1995) 54(3-4):167–84. doi: [10.1016/](https://doi.org/10.1016/0960-0760(95)00131-I) [0960-0760\(95\)00131-I](https://doi.org/10.1016/0960-0760(95)00131-I)

49. Aguero MF, Facchinetti MM, Sheleg Z, Senderowicz AM. Phenoxodiol, a novel isoflavone, induces G1 arrest by specific loss in cyclin-dependent kinase 2 activity by p53-independent induction of p21WAF1/CIP1. Cancer Res (2005) 65 (8):3364–73. doi: [10.1158/0008-5472.CAN-04-2429](https://doi.org/10.1158/0008-5472.CAN-04-2429)

50. Mor G, Fu HH, Alvero AB. Phenoxodiol, a novel approach for the treatment of ovarian cancer. Curr Opin Investig Drugs (2006) 7(6):542–8.

51. Isono M, Sato A, Asano T, Okubo K, Asano T. Evaluation of therapeutic potential of phenoxodiol, a novel isoflavone analog, in renal cancer cells. Anticancer Res (2018) 38(10):5709–16. doi: [10.21873/anticanres.12908](https://doi.org/10.21873/anticanres.12908)

52. Constantinou AI, Mehta R, Husband A. Phenoxodiol, a novel isoflavone derivative, inhibits dimethylbenz[a]anthracene (DMBA)-induced mammary carcinogenesis in female sprague-dawley rats. Eur J Cancer (2003) 39(7):1012–8. doi: [10.1016/S0959-8049\(03\)00124-2](https://doi.org/10.1016/S0959-8049(03)00124-2)

53. Kostelac D, Rechkemmer G, Briviba K. Phytoestrogens modulate binding response of estrogen receptors alpha and beta to the estrogen response element. J Agric Food Chem (2003) 51(26):7632–5. doi: [10.1021/jf034427b](https://doi.org/10.1021/jf034427b)

54. Morito K, Hirose T, Kinjo J, Hirakawa T, Okawa M, Nohara T, et al. Interaction of phytoestrogens with estrogen receptors alpha and beta. Biol Pharm Bull (2001) 24(4):351–6. doi: [10.1248/bpb.24.351](https://doi.org/10.1248/bpb.24.351)

55. Richter DU, Mylonas J, Toth B, Scholz C, Briese V, Friese K, et al. Effects of phytoestrogens genistein and daidzein on progesterone and estrogen (estradiol) roduction of human term trophoblast cells in vitro. Gynecol Endocrinol (2009) 25 (1):32–8. doi: [10.1080/09513590802485020](https://doi.org/10.1080/09513590802485020)

56. Cooke PS, Selvaraj V, Yellayi S. Genistein, estrogen receptors, and the acquired immune response. J Nutr (2006) 136(3):704–8. doi: [10.1093/jn/136.3.704](https://doi.org/10.1093/jn/136.3.704)

57. Sathyamoorthy N, Wang TT. Differential effects of dietary phyto-oestrogens daidzein and equol on human breast cancer MCF-7 cells. Eur J Cancer (1997) 33 (14):2384–9. doi: [10.1016/S0959-8049\(97\)00303-1](https://doi.org/10.1016/S0959-8049(97)00303-1)

58. Tang BY, Adams NR. Effect of equol on oestrogen receptors and on synthesis of DNA and protein in the immature rat uterus. *J Endocrinol* (1980) 85(2):291–7. doi: [10.1677/joe.0.0850291](https://doi.org/10.1677/joe.0.0850291)

59. Axelson M, Kirk DN, Farrant RD, Cooley G, Lawson AM, Setchell KD, et al. The identification of the weak oestrogen equol [7-hydroxy-3-(4'-hydroxyphenyl) chroman] in human urine. Biochem J (1982) 201(2):353–7. doi: [10.1042/bj2010353](https://doi.org/10.1042/bj2010353)

60. Shutt DA, Cox RI. Steroid and phyto-oestrogen binding to sheep uterine receptors in vitro. J Endocrinol (1972) 52(2):299–310. doi: [10.1677/joe.0.0520299](https://doi.org/10.1677/joe.0.0520299)

61. Rajah TT, Dum N, Drews N, Cohn R. Genistein in the presence of 17betaestradiol inhibits proliferation of ERbeta breast cancer cells. Pharmacology (2009) 84(2):68–73. doi: [10.1159/000226123](https://doi.org/10.1159/000226123)

62. Stettner M, Kaulfuss S, Burfeind P, Schweyer S, Strauss A, Ringert R-H. The relevance of estrogen receptor-beta expression to the antiproliferative effects observed with histone deacetylase inhibitors and phytoestrogens in prostate cancer treatment. Mol Cancer Ther (2007) 6(10):2626–33. doi: [10.1158/1535-](https://doi.org/10.1158/1535-7163.MCT-07-0197) [7163.MCT-07-0197](https://doi.org/10.1158/1535-7163.MCT-07-0197)

63. Chaurasiya S, Widmann S. Estrogen receptor β exerts tumor suppressive effects in prostate cancer through repression of androgen receptor activity. PloS One (2020) 15(5):e0226057. doi: [10.1371/journal.pone.0226057](https://doi.org/10.1371/journal.pone.0226057)

64. Piccolella M, Crippa V, Messi E, Tetel MJ, Poletti A. Modulators of estrogen receptor inhibit proliferation and migration of prostate cancer cells. Pharmacol Res (2014) 79:13–20. doi: [10.1016/j.phrs.2013.10.002](https://doi.org/10.1016/j.phrs.2013.10.002)

65. Lazennec G, Bresson D, Lucas A, Chauveau C, Vignon F. ER beta inhibits proliferation and invasion of breast cancer cells. Endocrinology (2001) 142 (9):4120–30. doi: [10.1210/endo.142.9.8395](https://doi.org/10.1210/endo.142.9.8395)

66. Paruthiyil S, Parmar H, Kerekatte V, Cunha GR, Firestone leitman GL DC. Estrogen receptor beta inhibits human breast cancer cell proliferation and tumor formation by causing a G2 cell cycle arrest. Cancer Res (2004) 64(1):423–8. doi: [10.1158/0008-5472.CAN-03-2446](https://doi.org/10.1158/0008-5472.CAN-03-2446)

67. Okura A, Arakawa H, Oka H, Yoshinari T, Monden Y. Effect of genistein on topoisomerase activity and on the growth of [Val 12]Ha-ras-transformed NIH 3T3 cells. Biochem Biophys Res Commun (1988) 157(1):183–9. doi: [10.1016/S0006-](https://doi.org/10.1016/S0006-291X(88)80030-5) [291X\(88\)80030-5](https://doi.org/10.1016/S0006-291X(88)80030-5)

68. Markovits J, Linassier C, Fosse P, Couprie J, Pierre J, Jacquemin-Sablon A. Inhibitory effects of the tyrosine kinase inhibitor genistein on mammalian DNA topoisomerase II. Cancer Res (1989) 49(18):5111–7.

69. Snyder RD, Gillies PJ. Evaluation of the clastogenic, DNA intercalative, and topoisomerase II-interactive properties of bioflavonoids in Chinese hamster V79 cells. Environ Mol Mutagen (2002) 40(4):266–76. doi: [10.1002/em.10121](https://doi.org/10.1002/em.10121)

70. Salti GI, Grewal S, Mehta RR, Das Gupta TK, Boddie AW Jr, Constantinou AI. Genistein induces apoptosis and topoisomerase II-mediated DNA breakage in colon cancer cells. Eur J Cancer (2000) 36(6):796–802. doi: [10.1016/S0959-8049](https://doi.org/10.1016/S0959-8049(00)00017-4) [\(00\)00017-4](https://doi.org/10.1016/S0959-8049(00)00017-4)

71. Hillman GG. Soy isoflavones protect normal tissues while enhancing radiation responses. Semin Radiat Oncol (2019) 29(1):62–71. doi: [10.1016/](https://doi.org/10.1016/j.semradonc.2018.10.002) [j.semradonc.2018.10.002](https://doi.org/10.1016/j.semradonc.2018.10.002)

72. Singh-Gupta V, Joiner MC, Runyan L, Yunker CK, Sarkar FH, Miller S, et al. Soy isoflavones augment radiation effect by inhibiting APE1/Ref-1 DNA repair activity in non-small cell lung cancer. J Thorac Oncol (2011) 6(4):688–98. doi: [10.1097/JTO.0b013e31821034ae](https://doi.org/10.1097/JTO.0b013e31821034ae)

73. Martin OA, Ivashkevich A, Choo S, Woodbine L, Jeggo PA, Martin RF, et al. Statistical analysis of kinetics, distribution and co-localisation of DNA repair foci in irradiated cells: cell cycle effect and implications for prediction of radiosensitivity. DNA Repair (Amst) (2013) 12(10):844–55. doi: [10.1016/j.dnarep.2013.07.002](https://doi.org/10.1016/j.dnarep.2013.07.002)

74. Mitchell JH, Duthie SJ, Collins AR. Effects of phytoestrogens on growth and DNA integrity in human prostate tumor cell lines: PC-3 and LNCaP. Nutr Cancer (2000) 38(2):223–8. doi: [10.1207/S15327914NC382_12](https://doi.org/10.1207/S15327914NC382_12)

75. Constantinou AI, Husband A. Phenoxodiol (2H-1-benzopyran-7-0,1,3-(4 hydroxyphenyl)), a novel isoflavone derivative, inhibits DNA topoisomerase II by stabilizing the cleavable complex. Anticancer Res (2002) 22(5):2581-5.

76. Cho N, Morre DJ. Early developmental expression of a normally tumorassociated and drug-inhibited cell surface-located NADH oxidase (ENOX2) in non-cancer cells. Cancer Immunol Immunother (2009) 58(4):547–52. doi: [10.1007/](https://doi.org/10.1007/s00262-008-0577-3) [s00262-008-0577-3](https://doi.org/10.1007/s00262-008-0577-3)

77. Herst PM, Davis JE, Neeson P, Berridge MV, Ritchie DS. The anti-cancer drug, phenoxodiol, kills primary myeloid and lymphoid leukemic blasts and
rapidly proliferating T cells. *Haematologica* (2009) 94(7):928–34. doi: [10.3324/](https://doi.org/10.3324/haematol.2008.003996) [haematol.2008.003996](https://doi.org/10.3324/haematol.2008.003996)

78. Herst PM, Petersen T, Jerram P, Baty J, Berridge MV. The antiproliferative effects of phenoxodiol are associated with inhibition of plasma membrane electron transport in tumour cell lines and primary immune cells. Biochem Pharmacol (2007) 74(11):1587–95. doi: [10.1016/j.bcp.2007.08.019](https://doi.org/10.1016/j.bcp.2007.08.019)

79. Tang X, Morre DJ, Morre DM. Antisense experiments demonstrate an exon 4 minus splice variant mRNA as the basis for expression of tNOX, a cancer-specific cell surface protein. Oncol Res (2007) 16(12):557–67. doi: [10.3727/](https://doi.org/10.3727/000000007783630006) [000000007783630006](https://doi.org/10.3727/000000007783630006)

80. Tang X, Tian Z, Chueh P-J, Chen S, Morre DM, Morre DM. Alternative splicing as the basis for specific localization of tNOX, a unique hydroquinone (NADH) oxidase, to the cancer cell surface. Biochemistry (2007) 46(43):12337–46. doi: [10.1021/bi700973k](https://doi.org/10.1021/bi700973k)

81. Zhang Z, Wang C-Z, Du G-J, Qi L-W, Calway T, He T-C, et al. Genistein induces G2/M cell cycle arrest and apoptosis via ATM/p53-dependent pathway in human colon cancer cells. Int J Oncol (2013) 43(1):289–96. doi: [10.3892/](https://doi.org/10.3892/ijo.2013.1946) [ijo.2013.1946](https://doi.org/10.3892/ijo.2013.1946)

82. Pawlik TM, Keyomarsi K. Role of cell cycle in mediating sensitivity to radiotherapy. Int J Radiat Oncol Biol Phys (2004) 59(4):928–42. doi: [10.1016/](https://doi.org/10.1016/j.ijrobp.2004.03.005) [j.ijrobp.2004.03.005](https://doi.org/10.1016/j.ijrobp.2004.03.005)

83. Hedlund TE, Johannes WU, Miller GJ. Soy isoflavonoid equol modulates the growth of benign and malignant prostatic epithelial cells in vitro. Prostate (2003) 54 (1):68–78. doi: [10.1002/pros.10137](https://doi.org/10.1002/pros.10137)

84. Mahoney S, Arfuso F, Millward M, Dharmarajan A. The effects of phenoxodiol on the cell cycle of prostate cancer cell lines. Cancer Cell Int (2014) 14(1):110. doi: [10.1186/s12935-014-0110-z](https://doi.org/10.1186/s12935-014-0110-z)

85. Aguero MF, Venero M, Brown DM, Smulson ME, Espinoza LA. Phenoxodiol inhibits growth of metastatic prostate cancer cells. Prostate (2010) 70(11):1211–21. doi: [10.1002/pros.21156](https://doi.org/10.1002/pros.21156)

86. Morre DJ, Morre DM. Cell surface NADH oxidases (ECTO-NOX proteins) with roles in cancer, cellular time-keeping, growth, aging and neurodegenerative diseases. Free Radic Res (2003) 37(8):795–808. doi: [10.1080/1071576031000083107](https://doi.org/10.1080/1071576031000083107)

87. Akiyama T, Ishida J, Nakagawa S, Ogawara H, Watanabe S, Itoh N, et al. Genistein, a specific inhibitor of tyrosine-specific protein kinases. J Biol Chem (1987) 262(12):5592–5. doi: [10.1016/S0021-9258\(18\)45614-1](https://doi.org/10.1016/S0021-9258(18)45614-1)

88. Peterson G, Barnes S. Genistein and biochanin a inhibit the growth of human prostate cancer cells but not epidermal growth factor receptor tyrosine autophosphorylation. Prostate (1993) 22(4):335–45. doi: [10.1002/pros.2990220408](https://doi.org/10.1002/pros.2990220408)

89. Peterson G, Barnes S. Genistein inhibits both estrogen and growth factorstimulated proliferation of human breast cancer cells. Cell Growth Differ (1996) 7 (10):1345–51.

90. Ogawara H, Akiyama T, Watanabe S, Ito N, Kobori M, Seoda Y. Inhibition of tyrosine protein kinase activity by synthetic isoflavones and flavones. J Antibiot (Tokyo) (1989) 42(2):340–3. doi: [10.7164/antibiotics.42.340](https://doi.org/10.7164/antibiotics.42.340)

91. Eke I, Makinde AY, Aryankalayil MJ, Sandfort V, Palayoor ST, Rath BH, et al. Exploiting radiation-induced signaling to increase the susceptibility of resistant cancer cells to targeted drugs: AKT and mTOR inhibitors as an example. Mol Cancer Ther (2018) 17(2):355–67. doi: [10.1158/1535-7163.MCT-](https://doi.org/10.1158/1535-7163.MCT-17-0262)[17-0262](https://doi.org/10.1158/1535-7163.MCT-17-0262)

92. Vivanco I, Sawyers CL. The phosphatidylinositol 3-kinase AKT pathway in human cancer. Nat Rev Cancer (2002) 2(7):489–501. doi: [10.1038/nrc839](https://doi.org/10.1038/nrc839)

93. De Luca T, Bosneaga E, Morre DM, Morre DJ. Downstream targets of altered sphingolipid metabolism in response to inhibition of ENOX2 by phenoxodiol. Biofactors (2008) 34(3):253–60. doi: [10.3233/BIO-2009-1079](https://doi.org/10.3233/BIO-2009-1079)

94. Lu Z, Zhou R, Kong Y, Wang J, Xia W, Guo J, et al. S-equol, a secondary metabolite of natural anticancer isoflavone daidzein, inhibits prostate cancer growth in vitro and in vivo, though activating the Akt/FOXO3a pathway. Curr Cancer Drug Targets (2016) 16(5):455–65. doi: [10.2174/1568009616666151207105720](https://doi.org/10.2174/1568009616666151207105720)

95. Huang X, Chen S, Xu L, Liu Y, Deb DK, Platanias LC, et al. Genistein inhibits p38 map kinase activation, matrix metalloproteinase type 2, and cell invasion in human prostate epithelial cells. Cancer Res (2005) 65(8):3470–8. doi: [10.1158/0008-5472.CAN-04-2807](https://doi.org/10.1158/0008-5472.CAN-04-2807)

96. Xu L, Bergan RC. Genistein inhibits matrix metalloproteinase type 2 activation and prostate cancer cell invasion by blocking the transforming growth factor beta-mediated activation of mitogen-activated protein kinase-activated protein kinase 2-27-kDa heat shock protein pathway. Mol Pharmacol (2006) 70 (3):869–77. doi: [10.1124/mol.106.023861](https://doi.org/10.1124/mol.106.023861)

97. Xu L, Ding Y, Catalona WJ, Yang XJ, Anderson WF, Jovanovic B, et al. MEK4 function, genistein treatment, and invasion of human prostate cancer cells. J Natl Cancer Inst (2009) 101(16):1141–55. doi: [10.1093/jnci/djp227](https://doi.org/10.1093/jnci/djp227)

98. Xu L, Chen S, Bergan RC. MAPKAPK2 and HSP27 are downstream effectors of p38 MAP kinase-mediated matrix metalloproteinase type 2 activation and cell invasion in human prostate cancer. Oncogene (2006) 25(21):2987–98. doi: [10.1038/sj.onc.1209337](https://doi.org/10.1038/sj.onc.1209337)

99. El Touny LH, Banerjee PP. Identification of a biphasic role for genistein in the regulation of prostate cancer growth and metastasis. Cancer Res (2009) 69 (8):3695–703. doi: [10.1158/0008-5472.CAN-08-2958](https://doi.org/10.1158/0008-5472.CAN-08-2958)

100. El Touny LH, Banerjee PP. Genistein induces the metastasis suppressor kangai-1 which mediates its anti-invasive effects in TRAMP cancer cells. Biochem Biophys Res Commun (2007) 361(1):169–75. doi: [10.1016/j.bbrc.2007.07.010](https://doi.org/10.1016/j.bbrc.2007.07.010)

101. Pavese JM, Krishna SN, Bergan RC. Genistein inhibits human prostate cancer cell detachment, invasion, and metastasis. Am J Clin Nutr (2014) 100 Suppl 1(1):431s–6s. doi: [10.3945/ajcn.113.071290](https://doi.org/10.3945/ajcn.113.071290)

102. Li D, Yee JA, McGuire MH, Murphy PA, Yan L. Soybean isoflavones reduce experimental metastasis in mice. J Nutr (1999) 129(5):1075–8. doi: [10.1093/jn/](https://doi.org/10.1093/jn/129.5.1075) [129.5.1075](https://doi.org/10.1093/jn/129.5.1075)

103. Bergan R, Kyle E, Nguyen P, Trepel J, Ingui C, Neckers L. Genisteinstimulated adherence of prostate cancer cells is associated with the binding of focal adhesion kinase to beta-1-integrin. Clin Exp Metastasis (1996) 14(4):389–98. doi: [10.1007/BF00123398](https://doi.org/10.1007/BF00123398)

104. Gu Y, Zhu C-F, Dai Y-L, Zhong Q, Sun B. Inhibitory effects of genistein on metastasis of human hepatocellular carcinoma. World J Gastroenterol (2009) 15 (39):4952–7. doi: [10.3748/wjg.15.4952](https://doi.org/10.3748/wjg.15.4952)

105. Sawai H, Okada Y, Funahashi H, Matsuo Y, Takahashi H, Takeyama H, et al. Activation of focal adhesion kinase enhances the adhesion and invasion of pancreatic cancer cells via extracellular signal-regulated kinase-1/2 signaling pathway activation. Mol Cancer (2005) 4:37. doi: [10.1186/1476-4598-4-37](https://doi.org/10.1186/1476-4598-4-37)

106. Miękus K, Madeja Z. Genistein inhibits the contact-stimulated migration of prostate cancer cells. Cell Mol Biol Lett (2007) 12(3):348–61. doi: [10.2478/](https://doi.org/10.2478/s11658-007-0007-0) [s11658-007-0007-0](https://doi.org/10.2478/s11658-007-0007-0)

107. Farina HG, Pomies M, Alonso DF, Gomez DE. Antitumor and antiangiogenic activity of soy isoflavone genistein in mouse models of melanoma and breast cancer. Oncol Rep (2006) 16(4):885–91. doi: [10.3892/or.16.4.885](https://doi.org/10.3892/or.16.4.885)

108. Kyle E, Neckers L, Takimoto C, Curt G, Bergan R. Genistein-induced apoptosis of prostate cancer cells is preceded by a specific decrease in focal adhesion kinase activity. Mol Pharmacol (1997) 51(2):193–200. doi: [10.1124/](https://doi.org/10.1124/mol.51.2.193) [mol.51.2.193](https://doi.org/10.1124/mol.51.2.193)

109. Traganos F, Ardelt B, Halko N, Bruno S, Darzynkiewicz Z. Effects of genistein on the growth and cell cycle progression of normal human lymphocytes
and human leukemic MOLT-4 and HL-60 cells. *Cancer Res* (1992) 52(22):6200–8.

110. Carlo-Stella C, Regazzi E, Garau D, Mangoni L, Rizzo MT, et al. Effect of the protein tyrosine kinase inhibitor genistein on normal and leukaemic haemopoietic progenitor cells. Br J Haematol (1996) 93(3):551–7. doi: [10.1046/](https://doi.org/10.1046/j.1365-2141.1996.d01-1694.x) [j.1365-2141.1996.d01-1694.x](https://doi.org/10.1046/j.1365-2141.1996.d01-1694.x)

111. Singh-Gupta V, Zhang H, Yunker CK, Ahmad Z, Zwier D, Sarkar FH, et al. Daidzein effect on hormone refractory prostate cancer in vitro and in vivo compared to genistein and soy extract: potentiation of radiotherapy. Pharm Res (2010) 27(6):1115–27. doi: [10.1007/s11095-010-0107-9](https://doi.org/10.1007/s11095-010-0107-9)

112. Kamsteeg M, Rutherford T, Sapi E, Hanczaruk B, Shahabi S, Flick M, et al. Phenoxodiol–an isoflavone analog–induces apoptosis in chemoresistant ovarian cancer cells. Oncogene (2003) 22(17):2611–20. doi: [10.1038/sj.onc.1206422](https://doi.org/10.1038/sj.onc.1206422)

113. Mahoney S, Arfuso F, Rogers Pl Hisheh S, Brown D, Millward M, et al. Cytotoxic effects of the novel isoflavone, phenoxodiol, on prostate cancer cell lines. J Biosci (2012) 37(1):73–84. doi: [10.1007/s12038-011-9170-6](https://doi.org/10.1007/s12038-011-9170-6)

114. Alvero AB, O'Malley D, Brown D, Kelly G, Garg M, Chen W, et al. Molecular mechanism of phenoxodiol-induced apoptosis in ovarian carcinoma cells. Cancer (2006) 106(3):599–608. doi: [10.1002/cncr.21633](https://doi.org/10.1002/cncr.21633)

115. Morre DJ, Chueh PJ, Yagiz K, Balicki A, Kim C, Morre DM. ECTO-NOX target for the anticancer isoflavene phenoxodiol. Oncol Res (2007) 16(7):299–312. doi: [10.3727/000000006783980973](https://doi.org/10.3727/000000006783980973)

116. De Luca T, Morre DM, Zhao H, Morre DJ. NAD+/NADH and/or CoQ/ CoQH2 ratios from plasma membrane electron transport may determine ceramide and sphingosine-1-phosphate levels accompanying G1 arrest and apoptosis. Biofactors (2005) 25(1-4):43–60. doi: [10.1002/biof.5520250106](https://doi.org/10.1002/biof.5520250106)

117. Miyamoto M, Bhuiyan M, Sarkar FH. Phenoxodiol increases cisplatin sensitivity in ovarian clear cancer cells through XIAP down-regulation and autophagy inhibition. Anticancer Res (2018) 38(1):301–6. doi: [10.21873/](https://doi.org/10.21873/anticanres.12222) [anticanres.12222](https://doi.org/10.21873/anticanres.12222)

118. Upadhyay S, Neburi M, Chinni SR, Alhasan S, Miller F, Sakar FH, et al. Differential sensitivity of normal and malignant breast epithelial cells to genistein is partly mediated by p21(WAF1). Clin Cancer Res (2001) 7(6):1782–9.

119. Gamble JR, Xia P, Hahn CN, Drew JJ, Drogemuller CJ, Brown D. Phenoxodiol, an experimental anticancer drug, shows potent antiangiogenic
properties in addition to its antitumour effects. Int J Cancer (2006) 118 (10):2412–20. doi: [10.1002/ijc.21682](https://doi.org/10.1002/ijc.21682)

120. Ferraro C, Quemeneur L, Fournel S, Prigent AF, Revillard JP, Bonnefoy-Berard N. The topoisomerase inhibitors camptothecin and etoposide induce a CD95-independent apoptosis of activated peripheral lymphocytes. Cell Death Differ (2000) 7(2):197–206. doi: [10.1038/sj.cdd.4400595](https://doi.org/10.1038/sj.cdd.4400595)

121. Ju YH, Allred CD, Allred KF, Karko KL, Doerge DR, Helferich WG. Physiological concentrations of dietary genistein dose-dependently stimulate growth of estrogen-dependent human breast cancer (MCF-7) tumors implanted in athymic nude mice. J Nutr (2001) 131(11):2957–62. doi: [10.1093/jn/131.11.2957](https://doi.org/10.1093/jn/131.11.2957)

122. Allred CD, Allred KF, Ju YH, Clausen LM, Doerge DR, Schantz SL, et al. Dietary genistein results in larger MNU-induced, estrogen-dependent mammary tumors following ovariectomy of sprague-dawley rats. Carcinogenesis (2004) 25 $(2):211-8.$

123. Wang C, Kurzer MS. Phytoestrogen concentration determines effects on DNA synthesis in human breast cancer cells. Nutr Cancer (1997) 28(3):236–47. doi: [10.1080/01635589709514582](https://doi.org/10.1080/01635589709514582)

124. Jenie RI, Amalina ND, Ilmawati GPN, Utomo RY, Ikawati M, Khumaira A, et al. Cell cycle modulation of CHO-K1 cells under genistein treatment correlates with cells senescence, apoptosis and ROS level but in a dose-dependent manner. Adv Pharm Bull (2019) 9(3):453–61. doi: [10.15171/apb.2019.054](https://doi.org/10.15171/apb.2019.054)

125. Polkowski K, Mazurek AP. Biological properties of genistein. a review of in vitro and in vivo data. Acta Pol Pharm (2000) 57(2):135–55.

126. Georgaki S, Skopeliti M, Tsiatas M, Nicolaou KA, Ioannou K, Husband A, et al. Phenoxodiol, an anticancer isoflavene, induces immunomodulatory effects in vitro and in vivo. J Cell Mol Med (2009) 13(9b):3929–38. doi: [10.1111/j.1582-](https://doi.org/10.1111/j.1582-4934.2009.00695.x) [4934.2009.00695.x](https://doi.org/10.1111/j.1582-4934.2009.00695.x)

127. Straszewski-Chavez SL, et al. X-Linked inhibitor of apoptosis (XIAP) confers human trophoblast cell resistance to fas-mediated apoptosis. Mol Hum Reprod (2004) 10(1):33–41. doi: [10.1093/molehr/gah001](https://doi.org/10.1093/molehr/gah001)

128. Klein R, Brown D, Turnley AM. Phenoxodiol protects against cisplatin induced neurite toxicity in a PC-12 cell model. BMC Neurosci (2007) 8:61. doi: [10.1186/1471-2202-8-61](https://doi.org/10.1186/1471-2202-8-61)

129. Li Y, et al. Induction of apoptosis in breast cancer cells MDA-MB-231 by genistein. Oncogene (1999) 18(20):3166–72. doi: [10.1038/sj.onc.1202650](https://doi.org/10.1038/sj.onc.1202650)

130. Xu J, Loo G. Different effects of genistein on molecular markers related to apoptosis in two phenotypically dissimilar breast cancer cell lines. J Cell Biochem (2001) 82(1):78–88. doi: [10.1002/jcb.1147](https://doi.org/10.1002/jcb.1147)

131. Onozawa M, Fukuda K, Ohtani M, Akaza H, Sugimura T, Wakabayashi K, et al. Effects of soybean isoflavones on cell growth and apoptosis of the human prostatic cancer cell line LNCaP. Jpn J Clin Oncol (1998) 28(6):360–3. doi: [10.1093/](https://doi.org/10.1093/jjco/28.6.360) $\frac{1}{10}$ ico/28.6.360

132. Zhang XM, Chen J, Xia Y-G, Xu Q. Apoptosis of murine melanoma B16- BL6 cells induced by quercetin targeting mitochondria, inhibiting expression of PKC-alpha and translocating PKC-delta. Cancer Chemother Pharmacol (2005) 55 (3):251–62. doi: [10.1007/s00280-004-0863-5](https://doi.org/10.1007/s00280-004-0863-5)

133. Peterson G. Evaluation of the biochemical targets of genistein in tumor cells. J Nutr (1995) 125(3 Suppl):784s–9s.

134. Guo TL, McCay JA, Zhang LX, Brown RD, You L, Karrow NA, et al. Genistein modulates immune responses and increases host resistance to B16F10 tumor in adult female B6C3F1 mice. J Nutr (2001) 131(12):3251–8. doi: [10.1093/jn/](https://doi.org/10.1093/jn/131.12.3251) [131.12.3251](https://doi.org/10.1093/jn/131.12.3251)

135. Zhang Y, Song TT, Cunnick JE, Murphy PA, Hendrich S. Daidzein and genistein glucuronides in vitro are weakly estrogenic and activate human natural killer cells at nutritionally relevant concentrations. J Nutr (1999) 129(2):399–405. doi: [10.1093/jn/129.2.399](https://doi.org/10.1093/jn/129.2.399)

136. Sakai T, Kogiso M. Soy isoflavones and immunity. J Med Invest (2008) 55 (3-4):167–73. doi: [10.2152/jmi.55.167](https://doi.org/10.2152/jmi.55.167)

137. Guo TL, Chi RP, Hernandez DM, Auttachoat W, Zheng JF, et al. Decreased 7,12-dimethylbenz[a]anthracene-induced carcinogenesis coincides with the induction of antitumor immunities in adult female B6C3F1 mice pretreated with genistein. Carcinogenesis (2007) 28(12):2560–6. doi: [10.1093/carcin/bgm223](https://doi.org/10.1093/carcin/bgm223)

138. Kadel S, Kovats S. Sex hormones regulate innate immune cells and promote sex differences in respiratory virus infection. Front Immunol (2018) 9:1653. doi: 10.3389/fi[mmu.2018.01653](https://doi.org/10.3389/fimmu.2018.01653)

139. Kovats S. Estrogen receptors regulate an inflammatory pathway of dendritic cell differentiation: mechanisms and implications for immunity. Horm Behav (2012) 62(3):254–62. doi: [10.1016/j.yhbeh.2012.04.011](https://doi.org/10.1016/j.yhbeh.2012.04.011)

140. Kovats S. Estrogen receptors regulate innate immune cells and signaling pathways. Cell Immunol (2015) 294(2):63–9. doi: [10.1016/j.cellimm.2015.01.018](https://doi.org/10.1016/j.cellimm.2015.01.018)

141. Lélu K, laffont S, Delphy L, Paulet P-E, Perinat T, Tschanz SA, et al. Estrogen receptor α signaling in T lymphocytes is required for estradiol-mediated
inhibition of Th1 and Th17 cell differentiation and protection against experimental inhibition of Th1 and Th17 cell differentiation and protection against experimental
autoimmune encephalomyelitis. *J Immunol* (2011) 187(5):2386–93. doi: [10.4049/](https://doi.org/10.4049/jimmunol.1101578) [jimmunol.1101578](https://doi.org/10.4049/jimmunol.1101578)

142. Matalka KZ. The effect of estradiol, but not progesterone, on the production of cytokines in stimulated whole blood, is concentration-dependent. Neuro Endocrinol Lett (2003) 24(3-4):185–91.

143. Ascenzi P, Bocedi A, Marino M. Structure-function relationship of estrogen receptor alpha and beta: impact on human health. *Mol Aspects Med* (2006) 27(4):299–402. doi: [10.1016/j.mam.2006.07.001](https://doi.org/10.1016/j.mam.2006.07.001)

144. Ansar Ahmed S, Penhale WJ, Talal N. Sex hormones, immune responses, and autoimmune diseases. mechanisms of sex hormone action. Am J Pathol (1985) 121(3):531–51.

145. Cuzzocrea S, Genovese T, Mazzon E, Esposito E, Di Paola R, Muia C, et al. Effect of 17beta-estradiol on signal transduction pathways and secondary damage in experimental spinal cord trauma. Shock (2008) 29(3):362–71. doi: [10.1097/](https://doi.org/10.1097/SHK.0b013e31814545dc) [SHK.0b013e31814545dc](https://doi.org/10.1097/SHK.0b013e31814545dc)

146. Pyne NJ, El Buri A, Adams DR, Pyne S. Sphingosine 1-phosphate and cancer. Adv Biol Regul (2018) 68:97–106. doi: [10.1016/j.jbior.2017.09.006](https://doi.org/10.1016/j.jbior.2017.09.006)

147. Kumar A, Saba JD. Regulation of immune cell migration by sphingosine-1- Phosphate. Cell Mol Biol (OMICS) (2015) 61(2).

148. Rivera J, Proia RL, Olivera A. The alliance of sphingosine-1-phosphate and its receptors in immunity. Nat Rev Immunol (2008) 8(10):753–63. doi: [10.1038/](https://doi.org/10.1038/nri2400) [nri2400](https://doi.org/10.1038/nri2400)

149. Imbert C, Montfort A, Fraisse M, Marcheteau E, Gillhodes J, Martin E, et al. Resistance of melanoma to immune checkpoint inhibitors is overcome by targeting the sphingosine kinase-1. Nat Commun (2020) 11(1):437. doi: [10.1038/](https://doi.org/10.1038/s41467-019-14218-7) [s41467-019-14218-7](https://doi.org/10.1038/s41467-019-14218-7)

150. Yamazaki T, Kirchmair A. Mitochondrial DNA drives abscopal responses to radiation that are inhibited by autophagy. Nat Immunol (2020) $21(10):1160-71$. doi: [10.1038/s41590-020-0751-0](https://doi.org/10.1038/s41590-020-0751-0)

151. Yashar CM, Spanos WJ, Taylor DD, Gercel-Taylor C. Potentiation of the radiation effect with genistein in cervical cancer cells. Gynecol Oncol (2005) 99 (1):199–205. doi: [10.1016/j.ygyno.2005.07.002](https://doi.org/10.1016/j.ygyno.2005.07.002)

152. Sapi E, Alvero Chen AB W, O'Malley D, Hao X-Y, Dwipoyono B, et al. Resistance of ovarian carcinoma cells to docetaxel is XIAP dependent and reversible by phenoxodiol. Oncol Res (2004) 14(11-12):567–78. doi: [10.3727/](https://doi.org/10.3727/0965040042707943) [0965040042707943](https://doi.org/10.3727/0965040042707943)

153. Kluger HM, McCarthy MM, Alvero AB, Sznol M, Ariyan S, Camp RL, et al. The X-linked inhibitor of apoptosis protein (XIAP) is up-regulated in metastatic melanoma, and XIAP cleavage by phenoxodiol is associated with carboplatin sensitization. J Transl Med (2007) 5:6. doi: [10.1186/1479-5876-5-6](https://doi.org/10.1186/1479-5876-5-6)

154. McPherson RA, Galettis PT, de Souza PL. Enhancement of the activity of phenoxodiol by cisplatin in prostate cancer cells. Br J Cancer (2009) 100(4):649–55. doi: [10.1038/sj.bjc.6604920](https://doi.org/10.1038/sj.bjc.6604920)

155. Alvero AB, Brown Montagna D M, Matthews M, Mor G. Phenoxodioltopotecan co-administration exhibit significant anti-tumor activity without major adverse side effects. Cancer Biol Ther (2007) 6(4):612–7. doi: [10.4161/cbt.6.4.3891](https://doi.org/10.4161/cbt.6.4.3891)

156. Yaylaci E, Onen HI, Yar Saglam AS. Phenoxodiol sensitizes metastatic colorectal cancer cells to 5-fluorouracil- and oxaliplatin-induced apoptosis through intrinsic pathway. EXCLI J (2020) 19:936–49.

157. Liu X, Sun C, Jin X, Li P, Ye F, Zhao T, et al. Genistein enhances the radiosensitivity of breast cancer cells via G2/M cell cycle arrest and apoptosis. Molecules (2013) 18:13200–17. doi: [10.3390/molecules181113200](https://doi.org/10.3390/molecules181113200)

158. Raffoul JJ, Wang Y, Kucuk O, Forman JD, Sarkar FH, Hillman GG. Genistein inhibits radiation-induced activation of NF-kappaB in prostate cancer cells promoting apoptosis and G2/M cell cycle arrest. BMC Cancer (2006) 6:107. doi: [10.1186/1471-2407-6-107](https://doi.org/10.1186/1471-2407-6-107)

159. Andersson U, Johansson D, Behnam-Motlagh P, Johansson M, Malmer M, et al. Treatment schedule is of importance when gefitinib is combined with irradiation of glioma and endothelial cells in vitro. Acta Oncol (2007) 46(7):951– 60. doi: [10.1080/02841860701253045](https://doi.org/10.1080/02841860701253045)

160. van Rijn J, van den Berg J. Flavonoids as enhancers of x-ray-induced cell damage in hepatoma cells. Clin Cancer Res (1997) 3(10):1775–9.

161. Kim IG, Kim JS, Lee JH, Cho EW. Genistein decreases cellular redox potential, partially suppresses cell growth in HL60 leukemia cells and sensitizes cells to gammaradiationinduced cell death. Mol Med Rep (2014) 10(6):2786–92. doi: [10.3892/mmr.2014.2611](https://doi.org/10.3892/mmr.2014.2611)

162. Uckun FM, Tuel-Ahlgren L, Song CW, Waddick K, Myers DE, Kirihara J, et al. Ionizing radiation stimulates unidentified tyrosine-specific protein kinases in human b-lymphocyte precursors, triggering apoptosis and clonogenic cell death. Proc Natl Acad Sci USA (1992) 89(19):9005–9. doi: [10.1073/pnas.89.19.9005](https://doi.org/10.1073/pnas.89.19.9005)

163. Xu X, Harris KS, Wang HJ, Murphy PA, Hendrich S. Bioavailability of soybean isoflavones depends upon gut microflora in women. J Nutr (1995) 125 (9):2307–15. doi: [10.1093/jn/125.9.2307](https://doi.org/10.1093/jn/125.9.2307)

164. Liu X, Li P, Hirayama R, Niu Y, Liu X, Chen W, et al. Genistein sensitizes glioblastoma cells to carbon ions via inhibiting DNA-PKcs phosphorylation and subsequently repressing NHEJ and delaying HR repair pathways. Radiother Oncol (2018) 129(1):84–94. doi: [10.1016/j.radonc.2018.04.005](https://doi.org/10.1016/j.radonc.2018.04.005)

165. Hillman GG, Wang Y, Kucuk O, Che M, Doerge DR, Yudelev M, et al. Genistein potentiates inhibition of tumor growth by radiation in a prostate cancer orthotopic model. Mol Cancer Ther (2004) 3(10):1271–9. doi: [10.1158/1535-](https://doi.org/10.1158/1535-7163.1271.3.10) [7163.1271.3.10](https://doi.org/10.1158/1535-7163.1271.3.10)

166. Hillman GG, Wang Y, Che M, Raffoul JJ, Yudelev M, Kucuk O, et al. Progression of renal cell carcinoma is inhibited by genistein and radiation in an orthotopic model. BMC Cancer (2007) 7:4. doi: [10.1186/1471-2407-7-4](https://doi.org/10.1186/1471-2407-7-4)

167. Raffoul JJ, Banerjee S, Che M, Knoll ZE, Doerge DR, Abrams J, et al. Soy isoflavones enhance radiotherapy in a metastatic prostate cancer model. Int J Cancer (2007) 120(11):2491–8. doi: [10.1002/ijc.22548](https://doi.org/10.1002/ijc.22548)

168. Magee PJ, Allsopp P, Samaletdin A, Rowland IR. Daidzein, r-(+)equol and s-(-)equol inhibit the invasion of MDA-MB-231 breast cancer cells potentially via the down-regulation of matrix metalloproteinase-2. Eur J Nutr (2014) 53(1):345-50. doi: [10.1007/s00394-013-0520-z](https://doi.org/10.1007/s00394-013-0520-z)

169. Kim JS, Heo K, Yi J-M, Gong EJ, Yang K, Moon C, et al. Genistein mitigates radiation-induced testicular injury. Phytother Res (2012) 26(8):1119–25. doi: [10.1002/ptr.3689](https://doi.org/10.1002/ptr.3689)

170. Singh VK, Grace MB, Parekh VI, Whitnall MH, Landauer MR. Effects of genistein administration on cytokine induction in whole-body gamma irradiated mice. Int Immunopharmacol (2009) 9(12):1401–10. doi: [10.1016/j.intimp.2009.08.012](https://doi.org/10.1016/j.intimp.2009.08.012)

171. Landauer MR, Srinivasan V, Seed TM. Genistein treatment protects mice from ionizing radiation injury. J Appl Toxicol (2003) 23(6):379–85. doi: [10.1002/jat.904](https://doi.org/10.1002/jat.904)

172. Zhou Y, Mi MT. Genistein stimulates hematopoiesis and increases survival in irradiated mice. J Radiat Res (2005) 46(4):425–33. doi: [10.1269/jrr.46.425](https://doi.org/10.1269/jrr.46.425)

173. Davis TA, Clarke TK, Mog SR, Landauer MR. Subcutaneous administration of genistein prior to lethal irradiation supports multilineage,

hematopoietic progenitor cell recovery and survival. Int J Radiat Biol (2007) 83 (3):141–51. doi: [10.1080/09553000601132642](https://doi.org/10.1080/09553000601132642)

174. Davis TA, Mungunsukh O, Zins S, Day RM, Landauer MR. Genistein induces radioprotection by hematopoietic stem cell quiescence. Int J Radiat Biol (2008) 84(9):713–26. doi: [10.1080/09553000802317778](https://doi.org/10.1080/09553000802317778)

175. Day RM, Barshishat-Kupper M, Mog SR, McCart EA, Prasanna PGS, Davis TA, et al. Genistein protects against biomarkers of delayed lung sequelae in mice surviving high-dose total body irradiation. J Radiat Res (2008) 49(4):361–72. doi: [10.1269/jrr.07121](https://doi.org/10.1269/jrr.07121)

176. Mahmood J, Jelveh S, Calveley V, Zaidi A, Doctrow SR, Hill RP. Mitigation of radiation-induced lung injury by genistein and EUK-207. Int J Radiat Biol (2011) 87(8):889–901. doi: [10.3109/09553002.2011.583315](https://doi.org/10.3109/09553002.2011.583315)

177. Abernathy LM, Fountain MD, Rothstein SE, David JM, Yunker CK, Rakowski J, et al. Soy isoflavones promote radioprotection of normal lung tissue by inhibition of radiation-induced activation of macrophages and neutrophils. J Thorac Oncol (2015) 10(12):1703–12. doi: [10.1097/JTO.0000000000000677](https://doi.org/10.1097/JTO.0000000000000677)

178. Ahmad IU, Forman JD, Sarkar FH, Hillman GG, Heath E, Vaishampayan U, et al. Soy isoflavones in conjunction with radiation therapy in patients with prostate cancer. Nutr Cancer (2010) 62(7):996-1000. doi: [10.1080/](https://doi.org/10.1080/01635581.2010.509839) [01635581.2010.509839](https://doi.org/10.1080/01635581.2010.509839)

179. Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ, et al. Cancer statistics, 2009. CA Cancer J Clin (2009) 59(4):225–49. doi: [10.3322/caac.20006](https://doi.org/10.3322/caac.20006)

180. Tacyildiz N, Ozyoruk D, Yavuz G, Unal E, Dincaslan H, Dogu F, et al. Soy isoflavones ameliorate the adverse effects of chemotherapy in children. Nutr Cancer (2010) 62(7):1001–5. doi: [10.1080/01635581.2010.509841](https://doi.org/10.1080/01635581.2010.509841)

181. Souza P, Capp AL, Chikhladze N, Mezvrishvilli Z, Messina M, Mautner G, et al. Phase I study of a novel S1P inhibitor, NOX66, in combination with radiotherapy in patients with metastatic castration-resistant prostate cancer. J Clin Oncol (2020) 38(Suppl 15):5533–3. doi: [10.1200/JCO.2020.](https://doi.org/10.1200/JCO.2020.38.15_suppl.5533) [38.15_suppl.5533](https://doi.org/10.1200/JCO.2020.38.15_suppl.5533)

182. Emmett L, Crumbaker M, Nguyen A, Ho B, Yin C, et al. Interim results of a PhaseI/II prospective dose escalation trial evaluating safety and efficacy of combination 177Lu PSMA 617 and NOX66 in men with mCRPC post androgen signalling inhibition and 2 lines of taxane chemotherapy (LuPIN trial). *I Nucl Med* (2019) 60:465.

183. Crumbaker M, Pathmanandavel S, Yam AO, Nguyen A, Ho B, et al. Phase I/II trial of the combination of 177Lutetium prostate specific membrane antigen 617 and idronoxil (NOX66) in men with end-stage metastatic castration-resistant prostate cancer (LuPIN). Eur Urol Oncol (2021) 4(6):963–70. doi: [10.1016/](https://doi.org/10.1016/j.euo.2020.07.002) [j.euo.2020.07.002](https://doi.org/10.1016/j.euo.2020.07.002)

184. Emmett L, Pathmanandavel S, Crumbaker M, Rofe C, Yam AW, Ho B, et al. Updated results of a phase I/II prospective dose escalation trial evaluating safety and efficacy of combination 177Lu PSMA 617 and idronoxil in men with mCRPC post androgen signalling inhibition and taxane chemotherapy (LuPIN trial). J Clin Oncol (2020) 38(Suppl 15):5557–7. doi: [10.1200/JCO.2020.](https://doi.org/10.1200/JCO.2020.38.15_suppl.5557) [38.15_suppl.5557](https://doi.org/10.1200/JCO.2020.38.15_suppl.5557)

185. Pathmanadavel S, Crumbaker M, Yam AO, Nguyen A, Rofe C, Hovey E, et al. 177Lu-PSMA-617 and idronoxil in men with end-stage metastatic castrationresistant prostate cancer (LuPIN): patient outcomes and predictors of treatment response in a phase I/II trial. J Nucl Med (2022) 63(4):560–6.

186. Li Y, Bhuiyan M, Sarkar FH. Induction of apoptosis and inhibition of cerbB-2 in MDA-MB-435 cells by genistein. Int J Oncol (1999) 15(3):525-33. doi: [10.3892/ijo.15.3.525](https://doi.org/10.3892/ijo.15.3.525)

187. Hillman GG, Forman JD, Kucuk O, Yudelev M, Maughan RL, Rubio J, et al. Genistein potentiates the radiation effect on prostate carcinoma cells. Clin Cancer Res (2001) 7(2):382-90.

188. Hofman MS, Violet J, Hicks RJ, Ferdinandus J, Thang SP, Akhurst T, et al. [177Lu]Lu-PSMA-617 radionuclide treatment in patients with metastatic castration-resistant prostate cancer (LuPSMA trial): a single-centre, single-arm, phase 2 study. Lancet Oncol (2018) 19(6):825–33. doi: [10.1016/S1470-2045\(18\)](https://doi.org/10.1016/S1470-2045(18)30198-0) [30198-0](https://doi.org/10.1016/S1470-2045(18)30198-0)

189. Violet J, Sandhu S, Iravani A, Ferdinandus J, Thang S-P, Kong G, et al. Long-term follow-up and outcomes of retreatment in an expanded 50-patient single-center phase II prospective trial of $[1^{177}Lu]Lu$ -PSMA-617 theranostics in metastatic castration-resistant prostate cancer. J Nucl Med (2020) 61(6):857–65. doi: [10.2967/jnumed.119.236414](https://doi.org/10.2967/jnumed.119.236414)

190. Hofman MS, Emmett L, Sandhu SK, Iravani A, Joshua AM, Goh JC, et al. TheraP: A randomised phase II trial of $[177 \text{Lu}]\text{Lu-PSMA-617 (LuPSMA)}$ theranostic versus cabazitaxel in metastatic castration resistant prostate cancer (mCRPC) progressing after docetaxel: Initial results (ANZUP protocol 1603). J Clin Oncol (2020) 38(15_suppl):5500–0. doi: [10.1200/](https://doi.org/10.1200/JCO.2020.38.15_suppl.5500) [JCO.2020.38.15_suppl.5500](https://doi.org/10.1200/JCO.2020.38.15_suppl.5500)

191. Deng XL, Wang Y, Xiao GS. Effects of equol on multiple k+ channels stably expressed in HEK 293 cells. PloS One (2017) 12(8):e0183708. doi: [10.1371/](https://doi.org/10.1371/journal.pone.0183708) [journal.pone.0183708](https://doi.org/10.1371/journal.pone.0183708)