



# Mechanisms and Future of Non-Small Cell Lung Cancer Metastasis

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Lung cancer, renowned for its fast progression and metastatic potency, is rising to become a leading cause of death globally. It has been long observed that lung cancer is particularly apt in spawning distant metastasis at its early stages, and it can readily colonize virtually any human organ. In recent years, cancer research has shed light on why lung cancer is endowed with its exceptional ability to metastasize. In this review, we will take a comprehensive look at the current research on lung cancer metastasis, including molecular pathways, anatomical features and genetic traits that make lung cancer intrinsically metastatic, as we go from lung cancer's general metastatic potential to the particular metastasis mechanisms in multiple organs. We highly concerned about the advanced discovery and development of lung cancer metastasis, indicating the importance of lung cancer specific gene mutations, heterogeneity or biomarker discovery, and discussing potential opportunities and challenges. We will also introduce some current treatments that targets certain metastatic strategies of non-small cell lung cancer (NSCLC). Advances made in these regards could be critical to our current knowledge base of lung cancer metastasis.

**Keywords:** non-small cell lung cancer, metastasis, treatment, mechanism, systematic literature review

## INTRODUCTION

Lung cancer is notoriously known for its ability to spread readily in its early stages, as well as its potency to spread to a wide range of organs of vastly different anatomy and physiology. Since lung cancer patients are often diagnosed at an advanced stage, multiple metastases would have already developed, making targeted therapy extremely difficult and systemic therapy less effective. In recent years, studies conducted to uncover mechanisms behind lung cancer are rapidly increasing in numbers.

We have learned that lung cancer cells often resist or even thrive under hypoxia, immune cells may be inactivated and manipulated by tumor cells, tumor cells can migrate through various ways, and the different mechanisms underlying lung metastasis of nearly every possible site. All the evidence piece together to reveal how complicated the concept of lung cancer metastasis is due to the numerous pathways involved in the process. In this review, we will discuss the general mechanisms of metastasis including hypoxia, immunocompromise, tumor cell migration.

We shall track non-small-cell lung cancer from its primary tumor to its metastasis in the brain, the bone and the liver, which are the three most frequent sites of lung cancer metastasis. Pathology, molecular pathways and genetic characteristics of lung cancer metastasis to these organs form the bulk of our discussion. While current work of lung cancer metastasis will be reviewed with considerable depth, inferences and connections will also be frequently made to propose possible patterns of lung cancer metastasis. This review shall provide updated progress of lung cancer metastasis research, and give directions about relevant further studies.

## RESULTS

### General Metastatic Potential

Once a cancerous cell mass has established itself, it must address a variety of environmental stress in order to support its rapid expansion as well as initiating processes that can potentially lead to metastasis. Such environmental stress includes a hypoxic stroma, a multitude of immune responses, and antagonistic local cell types that hinders its invasion. Then, tumor cells need to intravasate and survive the circulation to spawn metastasis in various organs. Through manipulation and cooperation, lung cancer cells are able to attain resistance to its hostile surroundings, even converting certain negative influences into signals beneficial to its development, enemy into allies.

### Hypoxia Resilience

Cancer cells, being relatively metabolically active, have a certain oxygen demand. If a tumor tissue is deprived of oxygen for a prolonged period of time, it could undergo necrosis. Unfortunately, due to their low level of vascularization, tumors

are generally less oxygenated compared to their surrounding tissues when they first arise, which invariably leads to intratumor hypoxia. Hypoxia is one of the defining characteristics of lung cancer and indicates poor survival (1). Although low oxygen availability in tumors reduces the effect of radiotherapy, it is also a limiting factor that severely limits tumor growth. It is estimated that tumor cells can only survive within 100 to 150  $\mu\text{M}$  from a blood vessel, given that cells beyond this distance often suffer necrosis (2). To overcome this limitation, lung cancer cells use several regulation mechanisms to avoid lethal hypoxia, while reaping the benefits of low-grade hypoxia which boosts metastasis potential.

An intuitive approach for tumors to obtain adequate oxygen supply is by improving vascularization within the tumor mass through the process of angiogenesis. Several factors that facilitate this process has been identified, some of which are frequently associated with lung cancer. Vascular endothelial growth factors (VEGFs), for instance, can be secreted by tumor cells to stimulate blood vessel formation within the tumor mass. It was observed that hypoxia inducible factor (HIF)  $1\alpha$  expression is positively correlated with VEGF expression, and the upregulation of HIF $1\alpha$  and VEGF often implies poor prognosis in liver cancer (3). Discovered in 1992, HIF has been put under the spotlight as 2019 Nobel Prize in Physiology or Medicine had recently been revealed, and emerges as a promising target for cancer therapy. Among the subtypes of VEGFs, VEGF-A is currently the most extensively studied and well understood. While it can form vasculatures in tumors to compensate for the oxygen demand, newly formed blood vessels are often abnormal in shape and function. Vessels are not neatly divided into arterioles, venules, and capillaries, and are shown to be leaky, even hemorrhagic in all stages of tumor (4). Though the tumor is only getting a suboptimal supply of oxygen and nutrition, this slightly hypoxic environment does yield benefits for tumor cells. Furthermore, since newly formed vasculature is leaky, presumably because of VEGF1-A's participation in vascular permeability regulation, there's an increased likelihood for tumor cells to enter these abnormal vessels early on in tumor progression, especially with increased interstitial pressure accompanied with hemorrhagic leakage. This helps explain why metastasis through blood circulation often occurs at an early stage in lung cancer.

As demonstrated above, angiogenesis can only fulfill a portion of a tumor's oxygen demand. While tumors display significant heterogeneity concerning spatial distribution of hypoxic regions, the vast majority of tumor cells are subsisting under non-lethal hypoxia (5). This characteristic of the tumor microenvironment is shown to facilitate an array of processes contributive to enhanced metastatic potential. It is observed that Epithelial-Mesenchymal Transition (EMT) is profoundly increased in hypoxic tumor tissues, and various mechanisms had been proposed since to explain this phenomenon. Beta-catenin is found to be accumulating in hypoxic tumor cell nucleus and is pinpointed as an agent that increases the expression of EMT-related genes (6). One previous study reveals that beta-catenin enhances HIF-1 mediated transcriptions, promoting tumor survival. This study also confirms that beta-catenin signaling is

**Abbreviations:** AKT, activation of protein kinase; ALI, acute lung injury; APC, antigen-presenting cell; BBB, blood-brain barrier; BMM, bone marrow macrophages; CAA, cancer-associated adipocytes; CAM, cellular adhesion molecules; CLDN5, claudin-5; CTSK, cathepsin k; CYLD, cylindromatosis; DC, dendritic cells; DDR1, discoidin domain receptor 1; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; ERG, ETS-related gene; EMT, epithelial-mesenchymal transition; ETosis, extracellular trap cell death; FASN, fatty acid synthase; HIF, hypoxia inducible factor; HLA-E, human leukocyte antigen e; IFN, interferons; IL, interleukins; IMA, invasive mucinous adenocarcinoma; JAM, junctional adhesion molecules; KP-10, kisspeptin-10; LPS, lipopolysaccharide; LSEC, liver sinusoidal endothelial cells; MF, myofibroblasts; miRNA, Micro-RNAs; MAPK, mitogen-activated protein kinase; MMP, metalloproteinases; MSC, mesenchymal stem cells; MAOA, monoamine oxidase A; NACOS, N-acetylated COS; NSCLC, non-small cell lung cancer; NK, natural killer cells; NO, nitric oxide; OBs, osteoblasts; OC, osteoclasts; PD, polydatin; PDL1, programmed death-ligand 1; PD1, programmed death receptor 1; p-PTEN, phosphorylation of the PTEN C-terminus; PGE2, prostaglandin E2; PTHrP, parathyroid hormone-related protein; RA, reactive astrocytes; ROS, reactive oxygen species; RANKL, RANK ligand; SCFA, short-chain fatty acids; SDF-1, stromal cell-derived factor 1; SNX9, nexin 9; Sfrps, secreted frizzled-related proteins; sRNA, small RNA; TADC, tumor-associated DCs; TAM, tumor-associated macrophages; TGF, tumor growth factors; TJ, tight junctions; TME, tumor microenvironment; TNF, tumor necrosis factors; Tregs, regulatory T cells; TSP, thrombospondin; VCAM-1, vascular cell adhesion molecule-1; VEGF, vascular endothelial growth factors; YY1, yin yang 1.

aberrant under hypoxia (7). Recent studies have attempted to explain beta-catenin's EMT inducing potential, and a link has been drawn between beta-catenin and Wnt pathway, known as Wnt/beta-catenin signaling. Through stabilizing beta-catenin and translocating it into the nucleus, hypoxia in adenocarcinoma cells in lungs eventually results in enhanced Wnt signaling activity. Research had shown that a disruption in regulated Wnt/beta-catenin (8) signaling could induce EMT in tumor cells, and it is proposed that it is caused by the loss of E-cadherin, which is detrimental to the maintenance of epithelial integrity (9). Alternative mechanisms have also been proposed. A study in 2016 concludes that PTEN phosphatase activity can be altered under persistent hypoxia. PTEN phosphatase appears to inhibit EMT in tumor progression, while phosphorylation of the PTEN C-terminus (p-PTEN) leads to diminishing activity of PTEN phosphatase. Lung cancer cells exposed to hypoxia demonstrated sharp decrease in PTEN phosphatase and increase in p-PTEN, which accompanies a high rate of EMT (10). TWIST, recently recognized as another crucial mediator of cancer metastasis, is shown to be influenced by a hypoxic microenvironment. TWIST negatively correlates with E-cadherin expression, while positively correlates with HIF1- $\alpha$  expression. It is considered that HIF-1 $\alpha$  derived from hypoxic environment enhances TWIST expression (11). High-level expression of HIF-1 $\alpha$  and TWIST, therefore, indicate increased EMT occurrence, metastasis likelihood and a poor prognosis (12). EMT grants cancer cells higher mobility and stem-cell properties which both facilitate distant metastasis. Given how fundamental EMT is to most metastasis, hypoxia evidently contributes to the metastasis potential of lung cancer, possibly through multiple signaling pathways.

## Immunocompromise

Aside from having to adapt to unfavorable oxygen status, tumor cells also need to fend off another major threat. Recognizing tumor cells as foreign, a myriad of immune cells will actively attack them upon contact, reducing the tumor's capabilities to expand. In fact, the immune system is so efficient in performing its antitumoral functions that, if a detectable tumor does arise, its cells must have evaded local immunity (13). Tumor cells develop specialized tactics against each component of the local immune system. The following paragraphs will explore the mechanisms tumor cells use to either evade or cooperate with various immune cells.

Given the fact that cancer cells are still bodily cells in nature, they are most prone to attacks launched by members of cell-mediated immunity. One such attack is initiated by cytotoxic T cells (CD8+). CD8+ tumor-infiltrating lymphocytes (CD8+ TILs) can infiltrate into tumors and induce cytotoxic effects through secretion of cytokines, which triggers inflammation and immune responses (14). They can also secrete substances such as granzymes and perforins for immediate cytotoxic effect. Its presence is correlated with a better prognosis, but its function can be suppressed by several factors (15). Programmed death-ligand 1 (PDL1)-Programmed death receptor 1 (PD1) pathway is shown to promote dysfunction in tumor-responding T cells in

lung cancer patients (16). Tumors cells expressed PDL1 can bind to PD1 on the surface of CD8 T cells, reducing its antitumoral effectiveness (17). Immunomodulation therapies targeting this interaction in non-small cell lung cancer (NSCLC) shows promise to enhance therapeutic effects (18).

Apart from directly manipulating CD8 T cell functionality, tumor cells also utilize many indirect mechanisms with the end goal of suppressing CD8 T cell activity. Dendritic cells (DC), a potent antigen-presenting cell (APC) is shown to play a role in immune suppression in lung cancer. DCs isolated from NSCLC display increased secretion of immunosuppressing molecules such as the aforementioned PDL1 (19). DCs are heterogeneous in nature, which can be subdivided into tumor-associated DCs (TADC), which orchestrates T cell antitumoral processes, and regulatory DCs, which accelerates tumor growth by immunosuppression (20). Tumor cells take advantage of DCs' heterogeneity as well as its plasticity to alter the composition of DC pool, impairing the overall antigen-presenting ability of DCs while magnifying its protumorigenic effects (21). Research also suggests that certain regulatory DCs also inhibit helper T cells (CD4+) activity, which disable another antigen-presenting pathway, reducing CD8+ T cell activity further (22). Regulatory T cells (Tregs), which normally function as an immunosuppressing agent through expression of FOXP3, acquire enhanced effectiveness in the tumor microenvironment (TME). Similar to DCs, Tregs exhibit high heterogeneity and plasticity, constituting another pool of immune cells tumor can turn to its favor (23). Tumors seem to possess the ability to chemically attract a subtype of Tregs known as FOXP3(+)/CD25(+)/CD4(+) Treg, which differentiates into a subpopulation that suppresses effector T cells functions (24). This is exemplified by a study involving 92 patients with NSCLC, which discovered increased Treg activity in intratumoral regions, including the overexpression of FOXP3 mRNA (25). Increased level of FOXP3 is shown to enhance the viability and invasiveness of tumors in NSCLCs, thus Treg can be yet another link in immunity which lung cancer might exploit to promote its growth (26). In conclusion, diminishing effect of CD8+ T cells caused by various mechanisms corresponds to greater invasiveness of lung cancer, giving it access to more capillary beds in the lungs, which increases likelihood of blood mediated metastasis.

CD8+ T cells are not alone in the frontline battle against cancer. Natural Killer (NK) cells, which constitute a major part of the innate immune system, also have antitumorigenic capabilities. Unlike CD8+ T cells, which requires a round of clonal expansion before unleashing its cytotoxic effects, NK cells can directly kill tumor cells, such as through the secretion of TNF $\alpha$ . A study conducted on mice subjects concluded that lung resident NK cells have a dominant role in suppressing metastatic tumor growth in the lungs (27). Numerous factors had been studied to establish pathways leading to NK cell exhaustion. Prostaglandin E2 (PGE2) secreted by lung cancer cells is attributed to diminishing effect of NK cells in one study, which shows that a range of soluble factors including PGE2 can inhibit the release of perforins and granzymes produced by NK cells (28). The loss of interleukin 2 (IL2) is also considered as a

suppressive factor since IL2 is critical to triggering cytolytic activities in NK cells (29). Human leukocyte antigen E (HLA-E), a ligand overexpressed in tumor cells, can bind to inhibitory receptor NKG2A which dampens NK cell response to malignant growth (30). In cisplatin-resistant lung cancer cases, a specific pathway is unmasked. Higher expression of fatty acid synthase (FASN) in cisplatin-resistant tumor cells signals the downstream molecule TGF $\beta$ 1, resulting in elevated PD-L1 expression that suppresses NK cell functions (31). In short, through multiple signaling pathways, lung cancer cells evade the cytolytic effects of NK cells, potentially giving it a greater chance to survive at distant sites surveilled by NK cells.

The last type of immune cells that will be discussed in this section are macrophages. Macrophages make up the majority of tumor-infiltrating immune cells, often triggering local inflammation (32). The dual role of macrophage has been observed in tumor masses. Early on in tumor emergence, macrophages either engulf individual tumor cells or act as APCs to provoke immune response from CD8+ T cells. When CD8+ T cells eventually fail to mount an immune effect on the tumor, tumor-associated macrophages (TAMs) promote tumor growth by secreting growth factors or facilitating angiogenesis (33).

Although alternative classifications have been proposed, a binary system is still widely used, which subdivides macrophages into M1 and M2 types (34). Inhibition of NOTCH-1 is proposed as a possible pathway that enhances M2 differentiation in tumors. When notch-1 is inhibited, enhanced M2 differentiation is observed (35). One study clarifies individual effects M1 and M2 has on lung cancer development. Using lung cancer cell A549, their experiments showed that M2 subtype promotes A549 invasion, while the M1 subtype inhibits angiogenesis in tumors (36). The M2 subtype's protumorigenic function is further proven in an experiment that uses  $\beta$ -elemene to inhibit M2 activity, resulting in suppressed tumor growth (37). M2 macrophages are shown to have angiogenesis effects that M1 macrophages do not possess. FGF signaling for M2a and PlGF signaling for M2c are proposed as possible mechanisms behind the angiogenic effect of M2 macrophages (38). Not only do M2 macrophages stimulate angiogenesis and suppress immune responses, they also prepare target organs for metastasis, as well as promoting intravasation of tumor cells (39). Macrophages are also associated with EMT in tumor cells, since inflammation is shown to be a key factor in inducing EMT. Given that certain macrophages, like M1 macrophages, are proinflammatory, they might increase EMT occurrence in cancerous cells (40). Another 2017 research concludes that M2 macrophages can induce EMT through the TGF- $\beta$ /Smad2 signaling pathway. M2 macrophages secrete large amounts of TGF- $\beta$ , which can induce EMT in lung alveolar epithelial cells. Specific changes in genetic expression include heightened  $\alpha$ -SMA and lowered E-cadherin and CK18, which contributes to cell mobility and the loss of epithelial adhesion (41). Another Chinese team confirms that E-cadherin level does correlate with TAMs density, and reveals that TAMs promote a certain type of EMT in lung adenocarcinoma through FUT4/LeY-mediated fucosylation (42). With angiogenic and

EMT capabilities combined, macrophages are a major candidate for facilitating lung cancer growth and metastasis.

## Cell Motility and Migration

Rather than remaining stationary, many tumor cells turned to a nomadic lifestyle. Tumor cell migration often depends on the crosstalk of different mechanisms. In order to move around in its extracellular matrix (ECM), tumor cells must make changes to themselves and cooperate with other cell types present in the local microenvironment. Better cell motility grants malignant cells enhanced potency to invade normal tissues, as well as a heightened chance of entering vasculature (43).

Tumor cells are not endowed with the ability to move on themselves at an early stage. Therefore, tumor cells must undergo significant changes to enable movement. Cell membrane protrusion is one of the first characteristics of migratory tumor cells acquire. The formation of these protrusions, such as lamellipodia and invadopodia, is driven by the interaction between several proteins and cytoplasmic structures, one of them being the actin cytoskeleton (44). The actin cytoskeleton requires actin-binding proteins to function, and some of these actin-binding proteins are shown to be correlated with cancer progression. A study in 2017 discovered that cortactin, an actin-binding protein, is often overexpressed in tumors. Cortactin promotes tumor cell invasiveness by forming invadopodia and degrading surrounding ECM. It is believed that the overexpression of cortactin is caused by the amplification of a chromosomal band (45). Another study postulated that since actin and actin-binding protein are observed to be accumulating in tumor cell nucleus, they may alter gene expression, which could potentially explain the genetic amplification observed in the previous study (46). Actin has been linked to the formation of multiple membrane protrusions. One research suggests that cortactin and dynamin-2 stabilize F-actin bundles in filopodia in lung cancer cell line H1299 (47). Furthermore, cortactin is also shown to participate in invadopodia formation. Inhibiting cortactin by miR-182 negatively impacted invadopodia formation (48). Cortactin is not the only protein that facilitates the acquisition of cell motility through membrane protrusions. The range of proteins participating in forming membrane protrusions is inconceivably large. Tks5, an adapter protein that contributes to invadopodia formation, is shown to have increased expression in lung carcinoma. The overexpression of Tks5 is correlated with increased metastatic potential and worsened prognosis (49). Another study in 2014 also identifies Tks5 as a critical factor for invadopodia maturation, while pointing to cortactin and Tks4 for invadopodia initiation and function, respectively (50). A study concerning invadopodia pinpointed  $\beta$ 3 integrin as indispensable to invadopodia's formation and its ability to degrade the surrounding ECM. Increased  $\beta$ 3 integrin expression often indicate a greater probability of metastasis in lung cancer (51). A decrease in nexin 9 (SNX9) in invadopodia in NSCLC is correlated with tumor invasiveness. SNX9 normally inhibits invadopodia function by binding to TKS5, therefore the depletion of



SNX9 result in increased degradation of ECM (52). As demonstrated above, lung cancer cells may utilize a range of protein regulation to enhance its motility through the formation of membrane protrusions.

Tumor cells can also steer cells in the ECM into their favor. Myofibroblasts (MF), which normally participate in tissue repair, may cooperate with cancer and promote tumor cell migration (53). TGF- $\beta$  is shown to be involved in the creation of MF. One study established a positive feedback signaling loop formed by TGF- $\beta$  and IL-6/JAK2/STAT3 signaling pathways, which mediates interactions between MF and tumor cells (54). Although TGF- $\beta$  can exist in soluble form, it is also found on surfaces of exosomes produced by tumor cells, which might explain why MF presence is heightened in tumor ECM (55). Exosomal TGF- $\beta$  also has greater potency in signaling myofibroblastic differentiation. When exosomes are eliminated from cancer cell secretome, myofibroblastic differentiation ceases, leading to failure of stroma-assisted tumor growth (56). MF express metalloproteinases (MMPs), which can alter matrix proteins. Studies show that MMPs are highly expressed in lung tumors and are actively involved in ECM degradation (57). A study performed on A549 NSCLC cells demonstrates that upregulation of MMP-9 promotes tumor cell migration and increases invasiveness (58). Moreover, using ECM degradation techniques such as MMPs, lung cancer cells can also degrade basement membrane of capillaries to intravasate. In conclusion, through cooperating with cells like MF, lung cancer cells can attain greater efficiency in degrading its surrounding ECM, allowing it to migrate with increased pace.

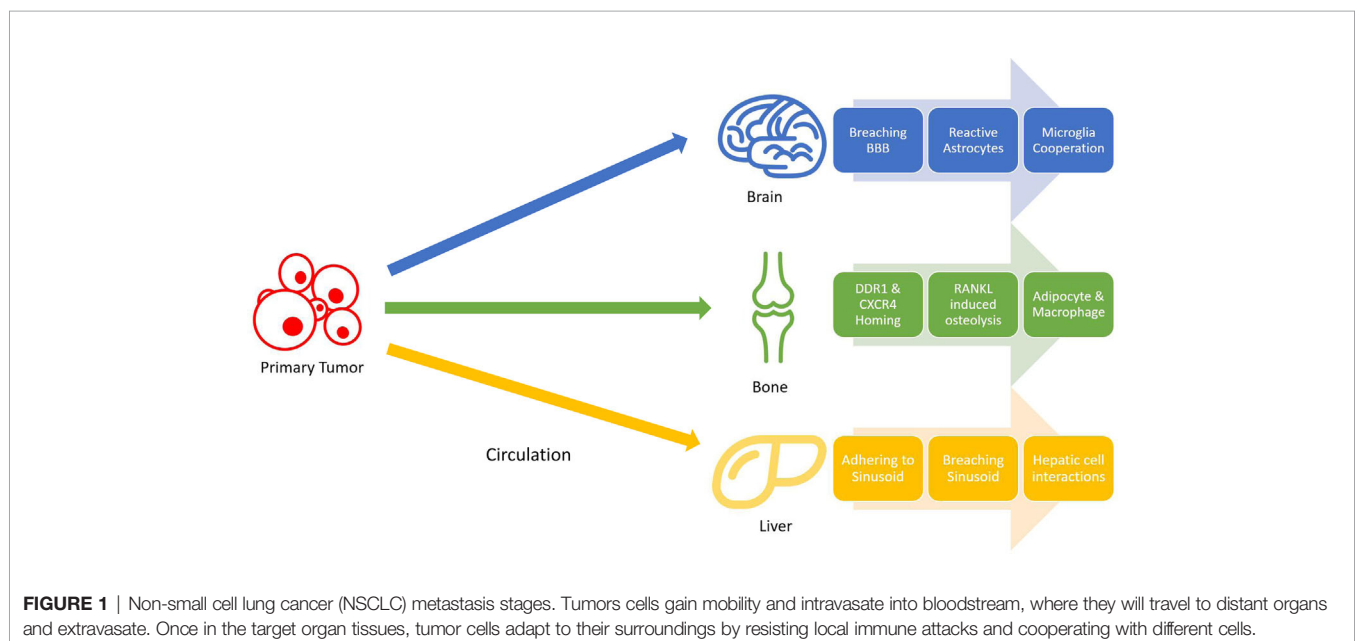
## Going Organ-Specific

Lung cancer is distinctive from many other types of cancer regarding the wide range of organs it could metastasize to. The nervous system, bone, liver, respiratory system, and adrenal gland, ranked by overall incidence rate, are all colonizable by

lung cancer metastasis, and such metastasis all foretell a worsened prognosis (59). Understanding the mechanisms behind such a width of potential metastasis of lung cancer can unlock new therapeutic strategies to prevent the formation of metastasis. The following sections will concern less about why lung cancer metastasize frequently in general, but rather focus more on mechanisms that facilitate metastasis formation in individual organs favored by lung metastasis. Although the colonization of tumor cells in these organs follow a similar pattern, they involve vastly different molecular pathways based on their immediate surroundings (**Figure 1**).

## Brain

Being one of the most protected organs in the body, the brain is surprisingly prone to lung cancer metastasis. There has been some advancement on decrypting the homing mechanisms used by tumor cells. CD15 is a cell adhesion molecule found overexpressed in metastatic NSCLC cells. Its binding counterpart, E-selectin (CD62E), is also found overexpressed in hCMEC/D3 human brain endothelial cells through activation by TGF- $\alpha$ . This increases the chance for metastatic NSCLC cells to bind to the brain endothelium, which opens the gateway for tumor cell intrusion (60). The effect of TGF- $\alpha$  can be dampened by EGFR specific tyrosine kinase inhibitor (61), and is shown to be downregulated by Yin Yang 1 (YY1), a potential therapeutic target (62). CD15-CD62E interaction is exclusively found at adhesion sites of tumor cells and brain endothelium while blocking this interaction significantly reduces cancer cell's ability to attach to the brain endothelium (63). Another study suggests that metastasizing lung cancer cells might secrete substances that damage endothelial glycocalyx. The glycocalyx serves as a protective covering of brain endothelium, which can have various adhesive interactions with tumor cells (64). Its degradation is correlated with enhanced E-selectin-mediated adhesion (65).



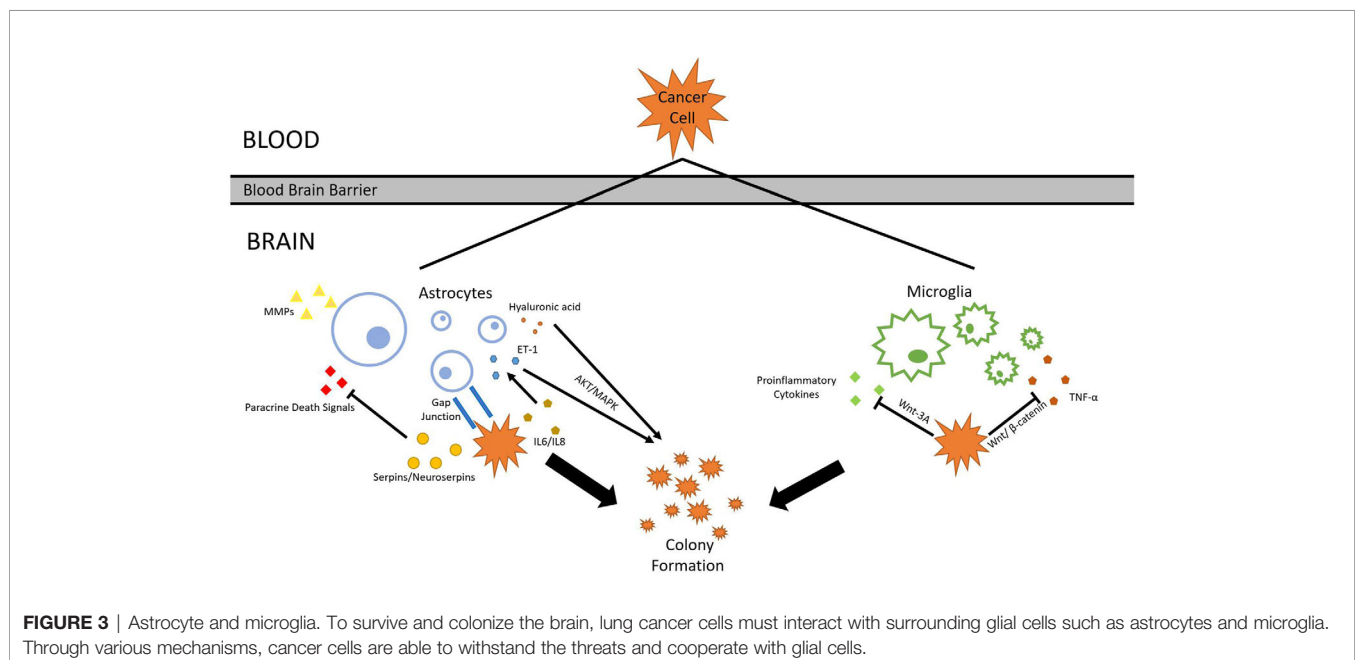


both tumorigenic and cytotoxic effects. Microglia can be broadly divided into pro-inflammatory and anti-inflammatory subtypes. Proinflammatory microglia suppress metastasis formation, while its counterpart increases metastatic tumor burden (85). Lipopolysaccharide (LPS)-activated microglia are cytotoxic to lung cancer cells by inducing apoptosis in them (86). Its activation is also correlated with increased production of proinflammatory cytokines (87). Cancer cells counteract the threat posed by microglia by suppressing its pro-inflammatory properties. Studies show that Wnt pathways are central to the regulation of microglial functions. Some Wnt pathways were once considered proinflammatory and thus disfavored for tumor growth. A 2011 study suggests that Wnt-3A stimulation triggers IL-6, IL-12, and tumor necrosis factor  $\alpha$  production in microglia, which are all strong proinflammatory factors (88). However, multiple subsequent studies reach differing conclusions. Wnt-3A pathway is found to trigger the induction of M2 phenotype in microglia, which are anti-inflammatory and favor tumorigenesis (89). It is recognized as an oncogene, and its activation is correlated with Wnt-catenin pathway. Knocking down Wnt-1 and Wnt-3A suppresses tumor proliferation (90). Other than Wnt-3A, Wnt-5A has also been shown to correlate with increased presence of TAMs, though the effect of WNT-5A seems to be also pro-inflammatory (91). This complication of Wnt-3A and Wnt-5A effects can be potentially explained by a 2013 study. While Wnt-3A and WNT-5A alone upregulate proinflammatory responses, the combination of Wnts and LPS serves anti-inflammatory functions (92). Tumor cells likely utilize this characteristic to favor their own growth by upregulating multiple Wnt pathways, amplifying the anti-inflammatory and tumorigenic effects of microglia. Furthermore, Wnt/ $\beta$ -catenin signaling pathway, which has been discussed earlier in section 1, play a pivotal role in microglia function in brain metastasis. Through Gastrodin

mediation, Wnt/ $\beta$ -catenin signaling pathway can downregulate multiple inflammatory factors such as TNF- $\alpha$  (93). Given that Wnt/ $\beta$ -catenin interaction is highly pronounced in lung cancer cells, lung cancer cells might gain an advantage by using this interaction to cooperate with microglia, resulting in higher metastatic potential in the brain. Secreted frizzled-related proteins (Sfrps) demonstrate differential inhibition of Wnt-3a activity (94). A study in 2011 found that SFRP-1 and the SFRP-like molecule V3Nter can inhibit  $\beta$ -catenin-activated tumor cells growth *in vivo*, though another study argues that sFRPs have dual effects on Wnt/ $\beta$ -catenin signaling depending on various factors (95, 96). With more research currently underway, much more will be uncovered about microglia's role in lung cancer brain metastasis in the future (Figure 3).

## Bone

At 39%, bone metastasis is the most frequent site of metastasis of adenocarcinoma. The presence of bone metastasis also signals worse prognosis compared to other metastasis sites (59). Multiple homing mechanisms have been revealed in lung cancer that metastasizes to the bone. In breast and prostate cancers, stromal cell-derived factor 1 (SDF-1) and its receptor CXCR4 are responsible for attracting tumor cells to the bone marrow (97). This coincides with a study recently published in 2019, which observed increased expression of CXCR4 in lung carcinoma (98). It is not impossible that the overexpression of CXCR4 in lung cancer cells enabled better attachment of tumor cells to the bone. Kisspeptin-10 (KP-10) can inhibit tumor cell invasion and EMT induced by SDF-1 through down-regulating CXCR-4 expression (99). Recent studies also reveal that certain CXCR4-targeting nanobodies inhibit CXCR4-related functions, which may yet unravel another treatment option (100). With striking resemblance to the aforementioned CXCR4/SDF-1 homing mechanism, CXCL10/CXCR3 also emerge as a



**FIGURE 3** | Astrocyte and microglia. To survive and colonize the brain, lung cancer cells must interact with surrounding glial cells such as astrocytes and microglia. Through various mechanisms, cancer cells are able to withstand the threats and cooperate with glial cells.

potential homing mechanism used by lung adenocarcinoma. A 2012 study demonstrates that CXCL10 attracts CXCR3-expressing tumor cells to the bone, though omitting its connection with lung cancer (101). A French team discovered that a CXCL10/CXCR3-A autocrine loop exists in invasive mucinous adenocarcinoma (IMA), in which CXCL10 upregulates CXCR3-A (102). It is thus possible that CXCL10 in the bone stroma may upregulate CXCR3 production in lung cancer cells, which in turn became more attracted to the bone. One confirmed homing mechanism used by lung adenocarcinoma cells is discoidin domain receptor 1 (DDR1). DDR1 is a cell surface receptor that can bind to several types of collagens and enhance cell adhesion. It was observed back in 2010 that DDR1 upregulation is a frequent occurrence in invasive lung adenocarcinoma. Its upregulation is correlated with increased tumor invasiveness and worse prognosis, although the precise mechanisms were elusive (103). The upregulation of DDR1 is found to be inducible by collagen I, which is a major component of osteons in the bone (104). Lung cancer cells may express DDR1 when induced by collagen I in the bone, which completes its homing process. Disrupting DDR1 pathway significantly reduced tumor-bone engagement, lowering cancer cell's potential to metastasize to the bones (105), and multiple means of doing so has been discovered. Tetrahydroisoquinoline-7-carboxamide based DDR1 inhibitor 7ae developed to treat acute lung injury (ALI) bind tightly to DDR1, reducing its kinase activity (106). Other studies reveal that celastrol downregulates E2F1 (107), while E2F1 silencing downregulates the expression of DDR1 (108). However, these studies are primarily concerned with osteosarcomas and hepatocellular carcinoma, therefore their existence in lung cancer bone metastasis awaits further validation.

Maintenance of bone tissues requires cooperation between osteoblasts (OBs) and osteoclasts (OCs). Cancer cells often hijack either OB or OC, forming osteoblastic or osteolytic lesions in bone tissues. Since osteoblastic lesions are infrequent in NSCLC bone metastasis, it will not be discussed in this article. In osteoclastic bone metastasis, which is most common in NSCLC, a "tumor-OC cooperation" is found. Tumor cells secrete pro-osteoclastogenic substances, while osteolysis generates substances that benefit tumor growth (109). One initiator of such a vicious circle is RANK ligand (RANKL). RANKL has been demonstrated to be essential to osteolytic activities induced by NSCLC cells. The inhibition of RANKL reduced osteoclastogenesis stimulated by NSCLC cells, and metastasis progression is slowed, possibly due to fewer growth factors yielded from osteolysis (110). A 2018 study confirms this discovery using mouse models. RANKL expression is found in NSCLC in primary lung cancer, and it is correlated with enhanced tumor growth (111). It is thus possible that NSCLC tumor cells present in the bones express RANKL, which enables osteolytic activities. With such a critical role in bone metastasis formation, RANKL is by far one of the most accurate predictors of bone response in patients with bone metastasis (112). Multiple factors that are associated with RANKL expression in lung tumor cells have also been identified. Thrombospondin (TSP)-2

knockdown is correlated with the inhibition of osteolytic metastasis of lung cancer. It is proposed that TSP-2 enhances osteoclastogenesis through RANKL dependent pathways (113). RANKL can also upregulate Basigin-2, which induces MMPs and VEGF expression, contributing to lung cancer bone metastasis through osteoclastogenesis (114). Extensive research has yielded numerous treatments that target RANKL, most of which are conducted quite recently. Estrogens and androgens are shown to inhibit OB-driven osteoclastogenesis (115). A myriad of drugs, such as Baricitinib, Matrine, and Sciadopitysin all suppress RANKL-mediated osteoclastogenesis (116–118).

With extensive osteolysis underway, a range of tumor trophic substances will be released, one of which being transforming growth factor beta (TGF $\beta$ ). TGF $\beta$  is dysregulated in malignant cells, including lung cancer, and upregulation of TGF $\beta$  in the ECM of lung cancer promotes NSCLC progression and invasion (119). Not only does TGF $\beta$  promote tumor cell proliferation, it also contributes to the vicious cycle by encouraging osteolysis. TGF $\beta$  upregulates Gli2, which in turn promotes osteolysis through increased secretion of osteolytic factors such as parathyroid hormone-related protein (PTHrP) (120). In osteoclastic bone metastasis, micro-RNAs (miRNAs) also play an important role, acting as key regulators in bone metastasis progression. Most miRNAs discovered are found to be inhibitory, and very few are shown to be pro-metastasis. One such miRNA is miR-326, which is found to correlate strongly with tumor burden in lung cancer bone metastasis (121). Serum microRNA-139-5p expression in mesenchymal stem cells (MSCs), progenitors of OBs, is sharply reduced when exposed to lung cancer cells A549 and L9981. The downregulation of microRNA-139-5p is correlated to increased osteolysis (122). miR-33a, which suppresses bone metastasis, is downregulated in lung cancer cells. Decreased miR-33a expression is correlated with increased PTHrP expression and RANKL production, both of which contribute to osteolysis (123). In conclusion, NSCLC cells manipulate OCs through secretion of pro-osteoclastogenesis substances and regulation of various miRNAs, allowing it to form bone metastasis.

Besides osteoclasts, adipocytes and macrophages also participate in bone metastasis formation. Adipocytes store fat, which can be used by lung cancer cells as an energy source. Compared to non-malignant lung cells, lung cancer cell A549 has twenty times more lipid droplets in its cytoplasm, suggesting that it is capable of using lipid as an alternative energy source to glucose (124). When fatty acid production *in vivo* or *in vitro* is inhibited, NSCLC growth is also hindered, suggesting cancer cell's reliance on fatty acid synthesized and stored in adipocytes (125). Cancer-associated adipocytes (CAAs) release fatty acids through lipolysis, while cancer cells can harvest the energy in them using  $\beta$ -oxidation (126). Moreover, CAAs can produce leptin, which has been demonstrated to promote EMT changes in A549 lung cancer cells, contributing to tumor cell mobility and consequently their invasiveness in the bone (127). Macrophages are also correlated with bone metastasis. A reduction in the number of macrophages is accompanied by the inhibition of bone metastasis (128). Rac2, a small GTPase, controls M1 to M2



differentiation in macrophages, which suppresses inflammatory responses (129). IL-13-induced phosphorylation of STAT6 has also been identified as a major pathway leading to macrophage M2 polarization (130). Furthermore, bone marrow macrophages (BMMs) are the major source of cathepsin K (CTSK), which promotes tumor progression in bones (131). Although substantial progress is made regarding the roles of adipocytes and macrophages in bone metastasis, very few researches have explored their interactions with lung cancer cells in particular. That being said, more effort should be invested in clarifying the connections between adipocytes, macrophages, and invading lung cancer cells. In essence, lung cancer bone metastasis involves a unique homing mechanism, a osteoclastic invasion, and diverse tumor-bone interactions.

## Liver

Compared to NSCLC metastasis in the brain and the bone, metastasis in the liver is less frequent, but still at a high occurrence. It is estimated that 16.7% of metastasized NSCLC patients develop liver metastasis (132). With an average median survival time of 4 months, liver metastasis in NSCLC confers a worse prognosis than lung, brain, and bone metastasis according to a recent study (133). NSCLC liver metastasis often predicts poor progression-free survival with treatments underway, including Nivolumab (134) and erlotinib targeted therapy (135).

When tumor cells enter the liver through the hepatic artery or portal vein, they will encounter the sinusoid, which endothelium is composed of liver sinusoidal endothelial cells (LSECs). LSECs have dual roles in metastasis formation. When tumor cells obstruct the sinusoid, inflammatory response will be triggered by LSECs, leading to production of various substances inhibiting metastasis (136). Among these substances is TNF- $\alpha$ , capable of inducing apoptosis in tumor cells. Lung cancer cells, however, often express phenotypes that resist the effect of TNF- $\alpha$ . A study dating back to 2000 suggests that NF- $\kappa$ B activation increases lung cancer cells' resistance to TNF- $\alpha$  (137). Cyldromatosis (CYLD) is considered to positively correlate with the apoptotic effect of TNF- $\alpha$ . Its expression in lung cancer cells is low, suggesting that lung cancer cells may downregulate CYLD in order to prevent TNF- $\alpha$  induced apoptosis (138). The effect of TNF- $\alpha$  can only be realized when it binds to TNF- $\alpha$  receptor (TNFR1), which contains TNFR1 promoter -223/-29 in lung cancer cells. The downregulation of TNFR1 is associated with decreased effectiveness of TNF- $\alpha$ , pointing to a potential mechanism lung cancer cells may use to evade TNF- $\alpha$  (139). Tumor cells may even benefit from exposure to TNF- $\alpha$ . One study concluded that through the upregulation of MMP-13, TNF- $\alpha$  promotes tumor growth in lung cancer cells (140). Significant progress is made to inhibit cancer growth through manipulating TNF- $\alpha$  expression. Highly N-acetylated COS (NACOS) inhibits TNF- $\alpha$ -induced E-selectin expression in ECs *via* the JNK/NF- $\kappa$ B pathways, potentially attenuates tumor cell adhesion with ECs (141). A research conducted in 2013 proposed that Acacetin can also inhibit TNF- $\alpha$ -induced E-selectin expression, as well as the activation of NF- $\kappa$ B by TNF- $\alpha$  (142). ZLJ-6 inhibits a range of adhesion molecules including

E-selectin, ICAM-1, and VCAM-1, and, although it was once believed that the COX/5-LOX pathway underlies its effect, was found to be independent of this pathway and is rather mediated by NF- $\kappa$ B (143).

Other than TNF- $\alpha$ , LSECs also produce IFN- $\gamma$ . IFN- $\gamma$  induces extracellular trap cell death (ETosis) in A549 lung cancer cells. Reactive oxygen species (ROS), another substance LSECs secrete, regulates IFN- $\gamma$ -induced mimic ETosis in lung cancer cells (144). This mechanism, however, has been found to be defective in many lung cancer cells. PC14PE6/AS2 human lung cancer cells were shown to be unsusceptible to IFN- $\gamma$ -induced autophagy (145). PC14PE6/AS2 human lung adenocarcinoma cells are presented with reduced responsiveness in IFN- $\gamma$  signaling, possibly due to natural PTEN loss. PTEN silenced A549 cells also demonstrate reduced susceptibility to IFN- $\gamma$  through ROS/SHP2 signaling (146). It is also suggested that IFN- $\gamma$  alone may not be able to inhibit lung cancer proliferation, as only PD-L1+ lung carcinomas are affected by IFN- $\gamma$ . Thus PDL1- lung cancer cells may be immune to IFN- $\gamma$  induced ETosis (147). Possible mechanisms have been proposed. IL-10 and IL-10R are found to be overexpressed in cells surrounding NSCLC cells in the lungs, and confer resistance to IFN- $\gamma$  through regulation of the PD1/PD-L1 pathway, although such phenomenon has not been studied in the liver (148). LSECs contribute to the formation of liver metastasis by expressing various adhesion molecules. Early studies have shown that the presence of tumor cells in hepatic circulation can trigger a rapid induction of E-selectin expression in LSECs, which could serve as adhesive molecules that facilitate tumor cell colonization (149). Blocking E-selectin expression causes a 97% reduction in liver metastasis compared to the control group (150). Tumor growth is also suppressed when E-selectin expression in LSECs is inhibited through the prevention of angiogenesis, suggesting E-selectin's importance in metastasis progression in the liver (151). Vascular cell adhesion molecule-1 (VCAM-1) is also found expressed in LSECs, which participates in tumor-stromal interactions (152). Downregulation of VCAM-1 is correlated with suppressed lung cancer cell growth (153), while VCAM-1 knockdown reduces A549 cells' ability to migrate (154).

Furthermore, since LSECs are fenestrated, tumor cells can directly interact with the basement membrane underneath (155). One type of the receptors present on the surface of basement membrane matrix is integrin receptors (156). Integrins  $\alpha$ v,  $\alpha$ 5,  $\beta$ 1,  $\beta$ 3, and  $\beta$ 5 are shown to promote survival and metastasis of lung cancer cells through either providing mechanical attachment to ECM, or the generation of various cell signals stimulating metastasis formation (157). Suppressing metastasis-related integrins such as integrin  $\alpha$ v and integrin  $\beta$ 3 in lung cancer inhibits motility of tumor cells (158). Interestingly, nitric oxide (NO), which is released by LSECs, is demonstrated to increase the expression of integrin  $\alpha$ v and  $\beta$ 1 in three types of NSCLC cell lines. Focal adhesion kinase, active RhoA, and active cell division control 42, all of which are migration associated proteins, are downstream targets of these two integrins (159). Integrin  $\beta$ 4 is also found overexpressed in NSCLC cells, which

increases venous invasion and correlates with a poor prognosis (160). Inhibiting integrin expression may serve as a new therapeutic strategy to suppress liver metastasis. E3 ubiquitin ligase Smurf1, for example, was demonstrated to be an effective inhibitor of integrin expression (161).

After tumor cells extravasate from sinusoid, they will encounter hepatocytes, the main cells of the liver. There has been evidence pointing to the presence of tumor cell-hepatocytes interactions. Claudin-2 level is elevated in breast cancer liver metastasis, and it is responsible for adhering breast cancer cells to hepatocytes (162). Although such interaction is not yet studied between lung adenocarcinoma cells and hepatocytes, there are clues that suggest the existence of this interaction. Claudin-2 is highly expressed in lung adenocarcinoma and is linked to increased tumor cell proliferation (163), and another study in the same year also concluded that decreased claudin-2 hinders lung carcinoma development (164). Therefore, it is not impossible that lung cancer cells may employ a similar technique to better adhere to hepatocytes. Hepatocytes also alter E-cadherin expression in cancer cells. E-cadherin expression is critical in EMT and EMrT, which are essential in metastatic processes. p0071/E-cadherin interaction in lung cancer cells is shown to increase their metastatic potential (165). The downregulation of E-cadherin in A549 cells induces cancer stem cell properties, which are necessary for metastasis. The upregulation of E-cadherin, in contrast, causes cancer stem cells to diminish (166). Therefore, when lung cancer stem cells reach liver tissues, they must upregulate E-cadherin levels to perform EMrT, which trades their stem cell features with epithelial features, invasiveness. Hepatocytes have the ability to upregulate E-cadherin expression in tumor cells in prostate cancer liver metastasis, which seemed to be regulated by lowering epidermal growth factor receptor (EGFR) signaling (167). Since EGFR mutation is relatively common in lung cancer, this mechanism found between prostate cancer cells and hepatocyte may also be used by lung cancer cells, although currently it is not thoroughly studied.

## New Treatment Strategy and Molecular Mechanisms of NSCLC Metastasis

In recent years, many new drugs have been put into clinical trials for the treatment of metastatic NSCLC to verify their toxicity and efficacy to enrich the treatment first-line options for patients. Several carcinogenic drivers including EGFR have been identified and studied. The development of new therapies, including targeted therapy and immunotherapy, has shown encouraging results in prolonging the survival of patients with NSCLC. For example, inhibitors targeting the PD-1/PD-L1 immune checkpoint have been developed as an effective immunotherapy for metastatic NSCLC. In NSCLC cells, the binding of PD-1 and PD-L1 promotes T-cell tolerance and escape from host immunity. Pembrolizumab, nivolumab, and cemiplimab are anti-PD-1 inhibitors, and atezolizumab, durvalumab, and avelumab are anti-PD-L1 inhibitors (168). PD-1 inhibitors offer enhanced survival benefits and fewer adverse events than PD-L1 inhibitors (169). As mentioned

above, any malignant transformation of EGFR may lead to the spread of lung cancer (170), so EGFR as a target has become the main research scheme to inhibit the metastasis of NSCLC. For example, from 2003 to 2019, first-generation drugs (erlotinib, gefitinib), second-generation drugs (afatinib, dacomitinib), third-generation drugs (osimertinib, rociletinib), combination therapy (docetaxel+pemetrexed), and fourth-generation drugs (EAI001 and EAI045) were successively developed (171). Although many drugs have been proved to be effective in clinical trials, some patients will have drug resistance and complex toxicity after a period of treatment. Therefore, it is still necessary to further study the molecular mechanism of NSCLC metastasis and explore potential biomarkers for the development of new therapeutic strategies

HER2 overexpression has been observed in 3%–38% of NSCLC, while strong HER2 protein overexpression is found in 2.5% of NSCLC (172). Leeza Shrestha et al., designed a peptidomimetic (compound 18) that binds to domain IV of the HER2 receptor, disrupts the homo/heterodimerization of HER2, and inhibits downstream signaling for cellular proliferation and growth, hence, can be used as a potential therapeutic agent for the treatment of NSCLC (173). A study from China suggests that traditional Chinese herbal medicine may play a role in antimetastasis of malignant tumors. Chinese herbal medicine Wenxia Changfu formula reverses cell adhesion-mediated drug resistance in lung cancer. Xiaoi Jiedu recipe can inactivate p38 MAPK signaling pathway in NSCLC cells to inhibit its proliferation and metastasis (174). Recent studies have found that CERS6 has important roles in lung cancer migration and metastasis, which proves that the miR-101-CERS6 pathway can be targeted, with potential benefits provided for affected patients (175). High expression of monoamine oxidase A (MAOA) in NSCLC is related to EMT and the development of clinicopathological features of NSCLC (176). The potential MAOA inhibitor G11 may inhibit paclitaxel-resistant NSCLC metastasis and growth by impacting on p-AKT, VEGF, HIF1 $\alpha$ , and MMP2 or MMP9. These findings have established MAOA as a promising therapeutic target in drug-resistant NSCLC and provide a feasible method of MAOA-targeted combined therapeutics (177). The combination of chemical and mechanical signals should also be considered in the new direction of drug development. Baicalein inhibited the formation of protrusive structures and leader cells by the combined effects that decreased both ezrin S-nitrosylation (chemical signaling) and ezrin tension transduction (mechanical signaling), and hampered NSCLC invasion and metastasis. Future study and improved clinical prospects for patients with NSCLC will depend on continued focus on combination of basic discovery and clinical translational research.

## DISCUSSION AND PERSPECTIVE

As showcased in this review, a substantial amount of effort has already been devoted to unraveling mechanisms behind NSCLC's high metastasis potency as well as the wide range of

potential organs for colonization that make NSCLC stand out from the majority of cancers. The chronological distribution of the references indicates that discoveries in this field have been made at an ever-increasing pace. Through studying the numerous molecular pathways involved in NSCLC metastasis, we are starting to find patterns that may one day fit into the puzzle of NSCLC metastasis containment and management. Many novel pathways identified to be essential to NSCLC metastasis had yielded new treatment options, and many of these targeted therapies were shown to be effective in restricting NSCLC metastasis. Still, for every question clarified, dozens of confusions arise. As demonstrated by this extensive review, lung cancer metastasis is an extremely complicated event and is further complicated by the type of lung cancer, metastatic organ, and more. Here, we list out some inquiries and topics that may be worth further investigation, including the limitations and unsolved enigmas of current research. Advances made in these regards could be critical to our current knowledge base of NSCLC metastasis.

- In many organs studied for cancer metastasis, such as the bone, very few researches studied lung cancer metastasis in these organs. As a result, extrapolation might be frequently used in an attempt to apply progress in other cancer types to NSCLC. These inferences are usually made by finding similarities between the discussed cancer types and lung cancer, and then suggest that lung cancer could utilize a similar mechanism. These inferences, being extrapolations, have the factor of conjecture. In order to clarify whether these suggestions are valid in NSCLC, we encourage more studies to be done to either validate or dismiss the existence of these mechanisms.
- Cancer progression can be viewed as a failure of the immune system to identify and eradicate cancer cells. Thus, many immune cells are associated with cancer, some actively involved in cancer invasion and metastasis. Each part of the human body has a unique blend of different immune cells, such as BMM in the bone and microglia in the brain. Most of the works currently available explain how cancer cells evade a certain immune cell's immune response. Few works, however, studied whether one mechanism cancer cell used to fend off a certain type of immune cell might be involved in another interaction between cancer cells and a different type of immune cell. Since there are plenty of overlaps between the molecules secreted by or the signal pathways involved in

different immune cells, it may be worth the effort to identify these overlaps to discover treatment options that can simultaneously attack multiple tumor cell-immune cell interactions, yielding a better prognosis.

- When tumor cells arrive at a distant organ, it can instantaneously spawn a new metastasis, but it can also enter a period of dormancy and become activated at a later time. The latter have been first discovered in the bone marrow, but it is now evident that many organs including the liver can house dormant tumor cells. These dormant cells can survive chemotherapy since it is not hyper-proliferating nor over metabolically active, thus they may spawn new metastasis even though all visible tumors have been eradicated. This mechanism has been extensively studied in the bone, but fewer studies are devoted to other organs such as the liver, and even few of them use NSCLC as their cancer type. A better understanding of this mechanism can be promising to the invention of therapies that prevent cancer recurrence.

Looking at only the tip of an iceberg of NSCLC metastasis, there is obviously still a quite formidable amount of knowledge to be discovered. However, with the progress made with high pace fueled by new generations of biotechnology, preventing NSCLC metastasis, or even curing NSCLC, may become reality in the future not far away.

## AUTHOR CONTRIBUTIONS

TZhu, XB, MC, RL, JZ, TZhe, and KX wrote the original draft. TZhu, XB, WZ, and SZ wrote, reviewed, and edited the manuscript. All authors contributed to the article and approved the submitted version.

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## REFERENCES

1. Salem A, Asselin MC, Reymen B, Jackson A, Lambin P, West CML, et al. Targeting Hypoxia to Improve Non-Small Cell Lung Cancer Outcome. *J Natl Cancer Inst* (2018) 110(1):14–30. doi: 10.1093/jnci/djx160
2. Brown JM. Tumor hypoxia in cancer therapy. *Methods Enzymol* (2007) 435:297–321. doi: 10.1016/S0076-6879(07)35015-5
3. Guo LY, Zhu P, Jin XP. Association between the expression of HIF-1alpha and VEGF and prognostic implications in primary liver cancer. *Genet Mol Res* (2016) 15(2):gmr8107. doi: 10.4238/gmr.15028107
4. Carmeliet P. VEGF as a key mediator of angiogenesis in cancer. *Oncology* (2005) 69 Suppl 3:4–10. doi: 10.1159/000088478
5. Vaupel P, Mayer A. Tumor Hypoxia: Causative Mechanisms, Microregional Heterogeneities, and the Role of Tissue-Based Hypoxia Markers. *Adv Exp Med Biol* (2016) 923:77–86. doi: 10.1007/978-3-319-38810-6\_11
6. Liu KH, Tsai YT, Chin SY, Lee WR, Chen YC, Shen SC. Hypoxia Stimulates the Epithelial-to-Mesenchymal Transition in Lung Cancer Cells Through Accumulation of Nuclear beta-Catenin. *Anticancer Res* (2018) 38(11):6299–308. doi: 10.21873/anticancer.12986



7. Kaidi A, Williams AC, Paraskeva C. Interaction between beta-catenin and HIF-1 promotes cellular adaptation to hypoxia. *Nat Cell Biol* (2007) 9(2):210–7. doi: 10.1038/ncb1534
8. Hong CF, Chen WY, Wu CW. Upregulation of Wnt signaling under hypoxia promotes lung cancer progression. *Oncol Rep* (2017) 38(3):1706–14. doi: 10.3892/or.2017.5807
9. Ghahhari NM, Babashah S. Interplay between microRNAs and WNT/beta-catenin signalling pathway regulates epithelial-mesenchymal transition in cancer. *Eur J Cancer* (2015) 51(12):1638–49. doi: 10.1016/j.ejca.2015.04.021
10. Kohnoh T, Hashimoto N, Ando A, Sakamoto K, Miyazaki S, Aoyama D, et al. Hypoxia-induced modulation of PTEN activity and EMT phenotypes in lung cancers. *Cancer Cell Int* (2016) 16:33. doi: 10.1186/s12935-016-0308-3
11. Wei L, Sun JJ, Cui YC, Jiang SL, Wang XW, Lv LY, et al. Twist may be associated with invasion and metastasis of hypoxic NSCLC cells. *Tumour Biol* (2016) 37(7):9979–87. doi: 10.1007/s13277-016-4896-2
12. Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, et al. Direct regulation of TWIST by HIF-1alpha promotes metastasis. *Nat Cell Biol* (2008) 10(3):295–305. doi: 10.1038/ncb1691
13. Gajewski TF, Schreiber H, Fu YX. Innate and adaptive immune cells in the tumor microenvironment. *Nat Immunol* (2013) 14(10):1014–22. doi: 10.1038/ni.2703
14. Ribas A. Adaptive Immune Resistance: How Cancer Protects from Immune Attack. *Cancer Discov* (2015) 5(9):915–9. doi: 10.1158/2159-8290.CD-15-0563
15. Rashed HE, Abdelrahman AE, Abdelgawad M, Balata S, Shabrawy ME. Prognostic Significance of Programmed Cell Death Ligand 1 (PD-L1), CD8+ Tumor-Infiltrating Lymphocytes and p53 in Non-Small Cell Lung Cancer: An Immunohistochemical Study. *Turk Patoloji Derg* (2017) 1(1):211–22. doi: 10.5146/tjpath.2017.01398
16. Prado-Garcia H, Romero-Garcia S, Puerto-Aquino A, Rumbo-Nava U. The PD-L1/PD-1 pathway promotes dysfunction, but not “exhaustion”, in tumor-responding T cells from pleural effusions in lung cancer patients. *Cancer Immunol Immunother* (2017) 66(6):765–76. doi: 10.1007/s00262-017-1979-x
17. Nowicki TS, Akiyama R, Huang RR, Shintaku IP, Wang X, Tumeh PC, et al. Infiltration of CD8 T Cells and Expression of PD-1 and PD-L1 in Synovial Sarcoma. *Cancer Immunol Res* (2017) 5(2):118–26. doi: 10.1158/2326-6066.CIR-16-0148
18. Guilleminault L, Carmier D, Heuze-Vourc'h N, Diot P, Pichon E. [Immunotherapy in non-small cell lung cancer: inhibition of PD1/PDL1 pathway]. *Rev Pneumol Clin* (2015) 71(1):44–56. doi: 10.1016/j.pneumo.2014.11.004
19. Upham JW, Xi Y. Dendritic Cells in Human Lung Disease: Recent Advances. *Chest* (2017) 151(3):668–73. doi: 10.1016/j.chest.2016.09.030
20. Keirse J, Van Damme H, Van Ginderachter JA, Laoui D. Exploiting tumor-associated dendritic cell heterogeneity for novel cancer therapies. *J Leukoc Biol* (2017) 102(2):317–24. doi: 10.1189/jlb.4MR1116-466R
21. Tang M, Diao J, Catral MS. Molecular mechanisms involved in dendritic cell dysfunction in cancer. *Cell Mol Life Sci* (2017) 74(5):761–76. doi: 10.1007/s00018-016-2317-8
22. Liu Q, Zhang C, Sun A, Zheng Y, Wang L, Cao X. Tumor-educated CD11bhighalow regulatory dendritic cells suppress T cell response through arginase I. *J Immunol* (2009) 182(10):6207–16. doi: 10.4049/jimmunol.0803926
23. Whiteside TL. FOXP3+ Treg as a therapeutic target for promoting anti-tumor immunity. *Expert Opin Ther Targets* (2018) 22(4):353–63. doi: 10.1080/14728222.2018.1451514
24. Nishikawa H, Sakaguchi S. Regulatory T cells in cancer immunotherapy. *Curr Opin Immunol* (2014) 27:1–7. doi: 10.1016/j.coi.2013.12.005
25. Akimova T, Zhang T, Negorev D, Singhal S, Stadanlick J, Rao A, et al. Human lung tumor FOXP3+ Tregs upregulate four “Treg-locking” transcription factors. *JCI Insight* (2017) 2(16):e94075. doi: 10.1172/jci.insight.94075
26. Peng J, Yu Z, Xue L, Wang J, Li J, Liu D, et al. The effect of foxp3-overexpressing Treg cells on non-small cell lung cancer cells. *Mol Med Rep* (2018) 17(4):5860–8. doi: 10.3892/mmr.2018.8606
27. Yamamoto Y, Miyazato K, Takahashi K, Yoshimura N, Tahara H, Hayakawa Y. Lung-resident natural killer cells control pulmonary tumor growth in mice. *Cancer Sci* (2018) 109(9):2670–6. doi: 10.1111/cas.13703
28. Hodge G, Barnawi J, Jurisevic C, Moffat D, Holmes M, Reynolds PN, et al. Lung cancer is associated with decreased expression of perforin, granzyme B and interferon (IFN)-gamma by infiltrating lung tissue T cells, natural killer (NK) T-like and NK cells. *Clin Exp Immunol* (2014) 178(1):79–85. doi: 10.1111/cei.12392
29. Balkhi MY, Ma Q, Ahmad S, Junghans RP. T cell exhaustion and Interleukin 2 downregulation. *Cytokine* (2015) 71(2):339–47. doi: 10.1016/j.cyt.2014.11.024
30. Lo Monaco E, Tremante E, Cerboni C, Melucci E, Sibilio L, Zingoni A, et al. Human leukocyte antigen E contributes to protect tumor cells from lysis by natural killer cells. *Neoplasia* (2011) 13(9):822–30. doi: 10.1593/neo.101684
31. Shen M, Tsai Y, Zhu R, Keng PC, Chen Y, Chen Y, et al. FASN-TGF-beta1-PD-L1 axis contributes to the development of resistance to NK cell cytotoxicity of cisplatin-resistant lung cancer cells. *Biochim Biophys Acta Mol Cell Biol Lipids* (2018) 1863(3):313–22. doi: 10.1016/j.bbalip.2017.12.012
32. Conway EM, Pikor LA, Kung SH, Hamilton MJ, Lam S, Lam WL, et al. Macrophages, Inflammation, and Lung Cancer. *Am J Respir Crit Care Med* (2016) 193(2):116–30. doi: 10.1164/rccm.201508-1545CI
33. Lamagna C, Aurrand-Lions M, Imhof BA. Dual role of macrophages in tumor growth and angiogenesis. *J Leukoc Biol* (2006) 80(4):705–13. doi: 10.1189/jlb.1105656
34. Aras S, Zaidi MR. TAMEless traitors: macrophages in cancer progression and metastasis. *Br J Cancer* (2017) 117(11):1583–91. doi: 10.1038/bjc.2017.356
35. Singla DK, Wang J, Singla R. Primary human monocytes differentiate into M2 macrophages and involve Notch-1 pathway. *Can J Physiol Pharmacol* (2017) 95(3):288–94. doi: 10.1139/cjpp-2016-0319
36. Yuan A, Hsiao YJ, Chen HY, Chen HW, Ho CC, Chen YY, et al. Opposite Effects of M1 and M2 Macrophage Subtypes on Lung Cancer Progression. *Sci Rep* (2015) 5:14273. doi: 10.1038/srep14273
37. Yu X, Xu M, Li N, Li Z, Li H, Shao S, et al. beta-elemene inhibits tumor-promoting effect of M2 macrophages in lung cancer. *Biochem Biophys Res Commun* (2017) 490(2):514–20. doi: 10.1016/j.bbrc.2017.06.071
38. Jetten N, Verbruggen S, Gijbels MJ, Post MJ, De Winther MP, Donners MM. Anti-inflammatory M2, but not pro-inflammatory M1 macrophages promote angiogenesis in vivo. *Angiogenesis* (2014) 17(1):109–18. doi: 10.1007/s10456-013-9381-6
39. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* (2010) 141(1):39–51. doi: 10.1016/j.cell.2010.03.014
40. Suarez-Carmona M, Lesage J, Cataldo D, Gilles C. EMT and inflammation: inseparable actors of cancer progression. *Mol Oncol* (2017) 11(7):805–23. doi: 10.1002/1878-0261.12095
41. Zhu L, Fu X, Chen X, Han X, Dong P. M2 macrophages induce EMT through the TGF-beta/Smad2 signaling pathway. *Cell Biol Int* (2017) 41(9):960–8. doi: 10.1002/cbin.10788
42. Wang A, Lu C, Ning Z, Gao W, Xie Y, Zhang N, et al. Tumor-associated macrophages promote Ezrin phosphorylation-mediated epithelial-mesenchymal transition in lung adenocarcinoma through FUT4/LeY up-regulation. *Oncotarget* (2017) 8(17):28247–59. doi: 10.18632/oncotarget.16001
43. Yamaguchi H, Wyckoff J, Condeelis J. Cell migration in tumors. *Curr Opin Cell Biol* (2005) 17(5):559–64. doi: 10.1016/j.ccb.2005.08.002
44. Yamaguchi H, Condeelis J. Regulation of the actin cytoskeleton in cancer cell migration and invasion. *Biochim Biophys Acta* (2007) 1773(5):642–52. doi: 10.1016/j.bbamcr.2006.07.001
45. Yin M, Ma W, An L. Cortactin in cancer cell migration and invasion. *Oncotarget* (2017) 8(50):88232–43. doi: 10.18632/oncotarget.21088
46. Izdebska M, Zielinska W, Grzanka D, Gagat M. The Role of Actin Dynamics and Actin-Binding Proteins Expression in Epithelial-to-Mesenchymal Transition and Its Association with Cancer Progression and Evaluation of Possible Therapeutic Targets. *BioMed Res Int* (2018) 2018:4578373. doi: 10.1155/2018/4578373
47. Yamada H, Takeda T, Michiue H, Abe T, Takei K. Actin bundling by dynamin 2 and cortactin is implicated in cell migration by stabilizing filopodia in human non-small cell lung carcinoma cells. *Int J Oncol* (2016) 49(3):877–86. doi: 10.3892/ijo.2016.3592
48. Li Y, Zhang H, Gong H, Yuan Y, Li Y, Wang C, et al. Liu H et al: miR-182 suppresses invadopodia formation and metastasis in non-small cell lung



- cancer by targeting cortactin gene. *J Exp Clin Cancer Res* (2018) 37(1):141. doi: 10.1186/s13046-018-0824-1
49. Stylli SS, Luwor RB, Kaye AH, I ST, Hovens CM, Lock P. Expression of the adaptor protein Tks5 in human cancer: prognostic potential. *Oncol Rep* (2014) 32(3):989–1002. doi: 10.3892/or.2014.3310
  50. Leong HS, Robertson AE, Stoletov K, Leith SJ, Chin CA, Chien AE, et al. Invadopodia are required for cancer cell extravasation and are a therapeutic target for metastasis. *Cell Rep* (2014) 8(5):1558–70. doi: 10.1016/j.celrep.2014.07.050
  51. Pelaez R, Morales X, Salvo E, Garasa S, Ortiz de Solorzano C, Martinez A, et al. beta3 integrin expression is required for invadopodia-mediated ECM degradation in lung carcinoma cells. *PLoS One* (2017) 12(8):e0181579. doi: 10.1371/journal.pone.0181579
  52. Bendris N, Stearns CJ, Reis CR, Rodriguez-Canales J, Liu H, Witkiewicz AW, et al. Sorting nexin 9 negatively regulates invadopodia formation and function in cancer cells. *J Cell Sci* (2016) 129(14):2804–16. doi: 10.1242/jcs.188045
  53. Darby IA, Zakuan N, Billet F, Desmouliere A. The myofibroblast, a key cell in normal and pathological tissue repair. *Cell Mol Life Sci* (2016) 73(6):1145–57. doi: 10.1007/s00018-015-2110-0
  54. Shi J, Feng J, Xie J, Mei Z, Shi T, Wang S, et al. Cheng X et al: Targeted blockade of TGF-beta and IL-6/JAK2/STAT3 pathways inhibits lung cancer growth promoted by bone marrow-derived myofibroblasts. *Sci Rep* (2017) 7(1):8660. doi: 10.1038/s41598-017-09020-8
  55. Webber J, Steadman R, Mason MD, Tabi Z, Clayton A. Cancer exosomes trigger fibroblast to myofibroblast differentiation. *Cancer Res* (2010) 70(23):9621–30. doi: 10.1158/0008-5472.CAN-10-1722
  56. Webber JP, Spary LK, Sanders AJ, Chowdhury R, Jiang WG, Steadman R, et al. Differentiation of tumour-promoting stromal myofibroblasts by cancer exosomes. *Oncogene* (2015) 34(3):290–302. doi: 10.1038/onc.2013.560
  57. Merchant N, Nagaraju GP, Rajitha B, Lammata S, Jella KK, Buchwald ZS, et al. Matrix metalloproteinases: their functional role in lung cancer. *Carcinogenesis* (2017) 38(8):766–80. doi: 10.1093/carcin/bgx063
  58. An J, Xue Y, Long M, Zhang G, Zhang J, Su H. TargetingCCR2 with its antagonist suppresses viability, motility and invasion by downregulating MMP-9 expression in non-small cell lung cancer cells. *Oncotarget* (2017) 8(24):39230–40. doi: 10.18632/oncotarget.16837
  59. Riihimaki M, Hemminki A, Fallah M, Thomsen H, Sundquist K, Sundquist J, et al. Metastatic sites and survival in lung cancer. *Lung Cancer* (2014) 86(1):78–84. doi: 10.1016/j.lungcan.2014.07.020
  60. Jassam SA, Maheraly Z, Smith JR, Ashkan K, Roncaroli F, Fillmore HL, et al. TNF-alpha enhancement of CD62E mediates adhesion of non-small cell lung cancer cells to brain endothelium via CD15 in lung-brain metastasis. *Neuro Oncol* (2016) 18(5):679–90. doi: 10.1093/neuonc/nov248
  61. Karki P, Johnson J Jr., Son DS, Aschner M, Lee E. Transcriptional Regulation of Human Transforming Growth Factor-alpha in Astrocytes. *Mol Neurobiol* (2017) 54(2):964–76. doi: 10.1007/s12035-016-9705-9
  62. El-Obeid A, Hesselager G, Westermarck B, Nister M. TGF-alpha-driven tumor growth is inhibited by an EGF receptor tyrosine kinase inhibitor. *Biochem Biophys Res Commun* (2002) 290(1):349–58. doi: 10.1006/bbrc.2001.6210
  63. Jassam SA, Maheraly Z, Smith JR, Ashkan K, Roncaroli F, Fillmore HL, et al. CD15/CD62E Interaction Mediates the Adhesion of Non-Small Cell Lung Cancer Cells on Brain Endothelial Cells: Implications for Cerebral Metastasis. *Int J Mol Sci* (2017) 18(7):1474. doi: 10.3390/ijms18071474
  64. Kutuzov N, Flyvbjerg H, Lauritzen M. Contributions of the glycocalyx, endothelium, and extravascular compartment to the blood-brain barrier. *Proc Natl Acad Sci USA* (2018) 115(40):E9429–38. doi: 10.1073/pnas.1802155115
  65. Rai S, Nejadhamzeeiliani Z, Gutowski NJ, Whatmore JL. Loss of the endothelial glycocalyx is associated with increased E-selectin mediated adhesion of lung tumour cells to the brain microvascular endothelium. *J Exp Clin Cancer Res* (2015) 34:105. doi: 10.1186/s13046-015-0223-9
  66. Soto MS, Serres S, Anthony DC, Sibson NR. Functional role of endothelial adhesion molecules in the early stages of brain metastasis. *Neuro Oncol* (2014) 16(4):540–51. doi: 10.1093/neuonc/not222
  67. Jia W, Lu R, Martin TA, Jiang WG. The role of claudin-5 in blood-brain barrier (BBB) and brain metastases (review). *Mol Med Rep* (2014) 9(3):779–85. doi: 10.3892/mmr.2013.1875
  68. Ma SC, Li Q, Peng JY, Zhouwen JL, Diao JF, Niu JX, et al. Claudin-5 regulates blood-brain barrier permeability by modifying brain microvascular endothelial cell proliferation, migration, and adhesion to prevent lung cancer metastasis. *CNS Neurosci Ther* (2017) 23(12):947–60. doi: 10.1111/cns.12764
  69. Yuan L, Le Bras A, Sacharidou A, Itagaki K, Zhan Y, Kondo M, et al. ETS-related gene (ERG) controls endothelial cell permeability via transcriptional regulation of the claudin 5 (CLDN5) gene. *J Biol Chem* (2012) 287(9):6582–91. doi: 10.1074/jbc.M111.300236
  70. Aslam M, Ahmad N, Srivastava R, Hemmer B. TNF-alpha induced NFkappaB signaling and p65 (RelA) overexpression repress Cldn5 promoter in mouse brain endothelial cells. *Cytokine* (2012) 57(2):269–75. doi: 10.1016/j.cyto.2011.10.016
  71. Verma PK, Bala M, Kumar N, Singh B. Therapeutic potential of natural products from terrestrial plants as TNF-alpha antagonist. *Curr Top Med Chem* (2012) 12(13):1422–35. doi: 10.2174/156802612801784425
  72. Fong LY, Ng CT, Zakaria ZA, Baharuldin MT, Arifah AK, Hakim MN, et al. Asiaticoside Inhibits TNF-alpha-Induced Endothelial Hyperpermeability of Human Aortic Endothelial Cells. *Phytother Res* (2015) 29(10):1501–8. doi: 10.1002/ptr.5404
  73. Valiente M, Obenaus AC, Jin X, Chen Q, Zhang XH, Lee DJ, et al. Brogi E et al: Serpins promote cancer cell survival and vascular co-option in brain metastasis. *Cell* (2014) 156(5):1002–16. doi: 10.1016/j.cell.2014.01.040
  74. Wasilewski D, Priego N, Fustero-Torre C, Valiente M. Reactive Astrocytes in Brain Metastasis. *Front Oncol* (2017) 7:298. doi: 10.3389/fonc.2017.00298
  75. Stevens LE, Cheung WKC, Adua SJ, Arnal-Estape A, Zhao M, Liu Z, et al. Extracellular Matrix Receptor Expression in Subtypes of Lung Adenocarcinoma Potentiates Outgrowth of Micrometastases. *Cancer Res* (2017) 77(8):1905–17. doi: 10.1158/0008-5472.CAN-16-1978
  76. Kim SW, Choi HJ, Lee HJ, He J, Wu Q, Langley RR, et al. Role of the endothelin axis in astrocyte- and endothelial cell-mediated chemoprotection of cancer cells. *Neuro Oncol* (2014) 16(12):1585–98. doi: 10.1093/neuonc/nou128
  77. Ye J, Piao H, Jiang J, Jin G, Zheng M, Yang J, et al. Li L et al: Polydatin inhibits mast cell-mediated allergic inflammation by targeting PI3K/Akt, MAPK, NF-kappaB and Nrf2/HO-1 pathways. *Sci Rep* (2017) 7(1):11895. doi: 10.1038/s41598-017-12252-3
  78. Ren J, Yuan L, Wang Y, Chen G, Hu K. Benzyl sulforaphane is superior to sulforaphane in inhibiting the Akt/MAPK and activating the Nrf2/ARE signalling pathways in HepG2 cells. *J Pharm Pharmacol* (2018) 70(12):1643–53. doi: 10.1111/jphp.13015
  79. Li L, Sapkota M, Gao M, Choi H, Soh Y. Macrolactin F inhibits RANKL-mediated osteoclastogenesis by suppressing Akt, MAPK and NFATc1 pathways and promotes osteoblastogenesis through a BMP-2/smad/Akt/Runx2 signaling pathway. *Eur J Pharmacol* (2017) 815:202–9. doi: 10.1016/j.ejphar.2017.09.015
  80. Seike T, Fujita K, Yamakawa Y, Kido MA, Takiguchi S, Teramoto N, et al. Interaction between lung cancer cells and astrocytes via specific inflammatory cytokines in the microenvironment of brain metastasis. *Clin Exp Metastasis* (2011) 28(1):13–25. doi: 10.1007/s10585-010-9354-8
  81. Chen Q, Boire A, Jin X, Valiente M, Er EE, Lopez-Soto A, et al. Carcinoma-astrocyte gap junctions promote brain metastasis by cGAMP transfer. *Nature* (2016) 533(7604):493–8. doi: 10.1038/nature18268
  82. Wang L, Cossette SM, Rarick KR, Gershan J, Dwinell MB, Harder DR, et al. Astrocytes directly influence tumor cell invasion and metastasis in vivo. *PLoS One* (2013) 8(12):e80933. doi: 10.1371/journal.pone.0080933
  83. Kim SJ, Kim JS, Park ES, Lee JS, Lin Q, Langley RR, et al. Weihua Z et al: Astrocytes upregulate survival genes in tumor cells and induce protection from chemotherapy. *Neoplasia* (2011) 13(3):286–98. doi: 10.1593/neo.11112
  84. Menachem A, Makovski V, Bodner O, Pasmanik-Chor M, Stein R, Shomron N, et al. Intercellular transfer of small RNAs from astrocytes to lung tumor cells induces resistance to chemotherapy. *Oncotarget* (2016) 7(11):12489–504. doi: 10.18632/oncotarget.7273
  85. Andreou KE, Soto MS, Allen D, Economopoulos V, de Bernardi A, Larkin JR, et al. Anti-inflammatory Microglia/Macrophages As a Potential Therapeutic Target in Brain Metastasis. *Front Oncol* (2017) 7:251. doi: 10.3389/fonc.2017.00251
  86. He BP, Wang JJ, Zhang X, Wu Y, Wang M, Bay BH, et al. Differential reactions of microglia to brain metastasis of lung cancer. *Mol Med* (2006) 12(7-8):161–70. doi: 10.2119/2006-00033.He

87. Awada R, Saulnier-Blache JS, Gres S, Bourdon E, Rondeau P, Parimisetty A, et al. Autotaxin downregulates LPS-induced microglia activation and pro-inflammatory cytokines production. *J Cell Biochem* (2014) 115(12):2123–32. doi: 10.1002/jcb.24889
88. Halleskog C, Mulder J, Dahlstrom J, Mackie K, Hortobagyi T, Tanila H, et al. WNT signaling in activated microglia is proinflammatory. *Glia* (2011) 59(1):119–31. doi: 10.1002/glia.21081
89. Matias D, Dubois LG, Pontes B, Rosario L, Ferrer VP, Balca-Silva J, et al. GBM-Derived Wnt3a Induces M2-Like Phenotype in Microglial Cells Through Wnt/beta-Catenin Signaling. *Mol Neurobiol* (2019) 56(2):1517–30. doi: 10.1007/s12035-018-1150-5
90. Kaur N, Chettiar S, Rathod S, Rath P, Muzumdar D, Shaikh ML, et al. Wnt3a mediated activation of Wnt/beta-catenin signaling promotes tumor progression in glioblastoma. *Mol Cell Neurosci* (2013) 54:44–57. doi: 10.1016/j.mcn.2013.01.001
91. Dijksterhuis JP, Arthofer E, Marinescu VD, Nelander S, Uhlen M, Ponten F, et al. High levels of WNT-5A in human glioma correlate with increased presence of tumor-associated microglia/monocytes. *Exp Cell Res* (2015) 339(2):280–8. doi: 10.1016/j.yexcr.2015.10.022
92. Halleskog C, Schulte G. WNT-3A and WNT-5A counteract lipopolysaccharide-induced pro-inflammatory changes in mouse primary microglia. *J Neurochem* (2013) 125(6):803–8. doi: 10.1111/jnc.12250
93. Yao YY, Bian LG, Yang P, Sui Y, Li R, Chen YL, et al. Gastrin attenuates proliferation and inflammatory responses in activated microglia through Wnt/beta-catenin signaling pathway. *Brain Res* (2019) 1717:190–203. doi: 10.1016/j.brainres.2019.04.025
94. Galli LM, Barnes T, Cheng T, Acosta L, Anglade A, Willert K, et al. Differential inhibition of Wnt-3a by Sfrp-1, Sfrp-2, and Sfrp-3. *Dev Dyn* (2006) 235(3):681–90. doi: 10.1002/dvdy.20681
95. Lavergne E, Hendaoui I, Coulouarn C, Ribault C, Leseur J, Eliat PA, et al. Blocking Wnt signaling by SFRP-like molecules inhibits in vivo cell proliferation and tumor growth in cells carrying active beta-catenin. *Oncogene* (2011) 30(4):423–33. doi: 10.1038/nc.2010.432
96. Xavier CP, Melikova M, Chuman Y, Uren A, Baljinyam B, Rubin JS. Secreted Frizzled-related protein potentiation versus inhibition of Wnt3a/beta-catenin signaling. *Cell Signal* (2014) 26(1):94–101. doi: 10.1016/j.cellsig.2013.09.016
97. Hirbe AC, Morgan EA, Weibaecher KN. The CXCR4/SDF-1 chemokine axis: a potential therapeutic target for bone metastases? *Curr Pharm Des* (2010) 16(11):1284–90. doi: 10.2174/138161210791034012
98. Liang T, Wang B, Li J, Liu Y. LINC00922 Accelerates the Proliferation, Migration and Invasion of Lung Cancer Via the miRNA-204/CXCR4 Axis. *Med Sci Monit* (2019) 25:5075–86. doi: 10.12659/MSM.916327
99. Grundker C, Bauerschmitz G, Knapp J, Schmidt E, Olbrich T, Emons G. Inhibition of SDF-1/CXCR4-induced epithelial-mesenchymal transition by kisspeptin-10. *Breast Cancer Res Treat* (2015) 152(1):41–50. doi: 10.1007/s10549-015-3463-7
100. Van Hout A, Klarenbeek A, Bobkov V, Doijen J, Arimont M, Zhao C, et al. CXCR4-targeting nanobodies differentially inhibit CXCR4 function and HIV entry. *Biochem Pharmacol* (2018) 158:402–12. doi: 10.1016/j.bcp.2018.10.015
101. Lee JH, Kim HN, Kim KO, Jin WJ, Lee S, Kim HH, et al. CXCL10 promotes osteolytic bone metastasis by enhancing cancer outgrowth and osteoclastogenesis. *Cancer Res* (2012) 72(13):3175–86. doi: 10.1158/0008-5472.CAN-12-0481
102. Duruisseau X, Rabbe N, Antoine M, Vieira T, Poulot V, Cadranet J, et al. Pro-tumoural CXCL10/CXCR3-A autocrine loop in invasive mucinous lung adenocarcinoma. *ERJ Open Res* (2017) 3(1):00047–2016. doi: 10.1183/23120541.00047-2016
103. Yang SH, Baek HA, Lee HJ, Park HS, Jang KY, Kang MJ, et al. Discoidin domain receptor 1 is associated with poor prognosis of non-small cell lung carcinomas. *Oncol Rep* (2010) 24(2):311–9. doi: 10.3892/or\_00000861
104. Miao L, Zhu S, Wang Y, Li Y, Ding J, Dai J, et al. Discoidin domain receptor 1 is associated with poor prognosis of non-small cell lung cancer and promotes cell invasion via epithelial-to-mesenchymal transition. *Med Oncol* (2013) 30(3):626. doi: 10.1007/s12032-013-0626-4
105. Valencia K, Ormazabal C, Zanduetta C, Luis-Ravelo D, Anton I, Pajares MJ, et al. Inhibition of collagen receptor discoidin domain receptor-1 (DDR1) reduces cell survival, homing, and colonization in lung cancer bone metastasis. *Clin Cancer Res* (2012) 18(4):969–80. doi: 10.1158/1078-0432.CCR-11-1686
106. Wang Z, Zhang Y, Bartual SG, Luo J, Xu T, Du W, et al. Tetrahydroisoquinoline-7-carboxamide Derivatives as New Selective Discoidin Domain Receptor 1 (DDR1) Inhibitors. *ACS Med Chem Lett* (2017) 8(3):327–32. doi: 10.1021/acsmchemlett.6b00497
107. Ma L, Peng L, Fang S, He B, Liu Z. Celastrol downregulates E2F1 to induce growth inhibitory effects in hepatocellular carcinoma HepG2 cells. *Oncol Rep* (2017) 38(5):2951–8. doi: 10.3892/or.2017.5971
108. Wang Z, Sun X, Bao Y, Mo J, Du H, Hu J, et al. E2F1 silencing inhibits migration and invasion of osteosarcoma cells via regulating DDR1 expression. *Int J Oncol* (2017) 51(6):1639–50. doi: 10.3892/ijo.2017.4165
109. Guise TA. The vicious cycle of bone metastases. *J Musculoskelet Neuronal Interact* (2002) 2(6):570–2.
110. Miller RE, Jones JC, Tometsko M, Blake ML, Dougall WC. RANKL inhibition blocks osteolytic lesions and reduces skeletal tumor burden in models of non-small-cell lung cancer bone metastases. *J Thorac Oncol* (2014) 9(3):345–54. doi: 10.1097/JTO.000000000000070
111. Faget J, Contat C, Zangger N, Peters S, Meylan E. RANKL Signaling Sustains Primary Tumor Growth in Genetically Engineered Mouse Models of Lung Adenocarcinoma. *J Thorac Oncol* (2018) 13(3):387–98. doi: 10.1016/j.jtho.2017.11.121
112. Ibrahim T, Ricci M, Scarpi E, Bongiovanni A, Ricci R, Riva N, et al. RANKL: A promising circulating marker for bone metastasis response. *Oncol Lett* (2016) 12(4):2970–5. doi: 10.3892/ol.2016.4977
113. Wang M, Chao CC, Chen PC, Liu PI, Yang YC, Su CM, et al. Thrombospondin enhances RANKL-dependent osteoclastogenesis and facilitates lung cancer bone metastasis. *Biochem Pharmacol* (2019) 166:23–32. doi: 10.1016/j.bcp.2019.05.005
114. Liao CG, Yao L, Xie W, Liu L, Wu SD, Lu N, et al. Basigin-2 upregulated by receptor activator of NF-kappaB ligand enhances lung cancer-induced osteolytic lesions. *Cancer Cell Int* (2016) 16:28. doi: 10.1186/s12935-016-0302-9
115. Martin A, Yu J, Xiong J, Khalid AB, Katzenellenbogen B, Kim SH, et al. Estrogens and androgens inhibit association of RANKL with the pre-osteoblast membrane through post-translational mechanisms. *J Cell Physiol* (2017) 232(12):3798–807. doi: 10.1002/jcp.25862
116. Murakami K, Kobayashi Y, Uehara S, Suzuki T, Koide M, Yamashita T, et al. A Jak1/2 inhibitor, baricitinib, inhibits osteoclastogenesis by suppressing RANKL expression in osteoblasts in vitro. *PLoS One* (2017) 12(7):e0181126. doi: 10.1371/journal.pone.0181126
117. Chen X, Zhi X, Pan P, Cui J, Cao L, Weng W, et al. Matrine prevents bone loss in ovariectomized mice by inhibiting RANKL-induced osteoclastogenesis. *FASEB J* (2017) 31(11):4855–65. doi: 10.1096/fj.201700316R
118. Cao J, Lu Q, Liu N, Zhang YX, Wang J, Zhang M, et al. Sciadopitysin suppresses RANKL-mediated osteoclastogenesis and prevents bone loss in LPS-treated mice. *Int Immunopharmacol* (2017) 49:109–17. doi: 10.1016/j.intimp.2017.05.029
119. Eser PO, Janne PA. TGFbeta pathway inhibition in the treatment of non-small cell lung cancer. *Pharmacol Ther* (2018) 184:112–30. doi: 10.1016/j.pharmthera.2017.11.004
120. Johnson RW, Nguyen MP, Padalecki SS, Grubbs BG, Merkel AR, Oyajobi BO, et al. TGF-beta promotion of Gli2-induced expression of parathyroid hormone-related protein, an important osteolytic factor in bone metastasis, is independent of canonical Hedgehog signaling. *Cancer Res* (2011) 71(3):822–31. doi: 10.1158/0008-5472.CAN-10-2993
121. Valencia K, Martin-Fernandez M, Zanduetta C, Ormazabal C, Martinez-Canarias S, Bandres E, et al. miR-326 associates with biochemical markers of bone turnover in lung cancer bone metastasis. *Bone* (2013) 52(1):532–9. doi: 10.1016/j.bone.2012.10.033
122. Xu S, Yang F, Liu R, Li X, Fan H, Liu J, et al. Serum microRNA-139-5p is downregulated in lung cancer patients with lytic bone metastasis. *Oncol Rep* (2018) 39(5):2376–84. doi: 10.3892/or.2018.6316
123. Kuo PL, Liao SH, Hung JY, Huang MS, Hsu YL. MicroRNA-33a functions as a bone metastasis suppressor in lung cancer by targeting parathyroid hormone related protein. *Biochim Biophys Acta* (2013) 1830(6):3756–66. doi: 10.1016/j.bbagen.2013.02.022

124. Chowdhury R, Amin MA, Bhattacharyya K. Intermittent Fluorescence Oscillations in Lipid Droplets in a Live Normal and Lung Cancer Cell: Time-Resolved Confocal Microscopy. *J Phys Chem B* (2015) 119(34):10868–75. doi: 10.1021/jp5120042
125. Svensson RU, Parker SJ, Eichner LJ, Kolar MJ, Wallace M, Brun SN, et al. Inhibition of acetyl-CoA carboxylase suppresses fatty acid synthesis and tumor growth of non-small-cell lung cancer in preclinical models. *Nat Med* (2016) 22(10):1108–19. doi: 10.1038/nm.4181
126. Nieman KM, Romero IL, Van Houten B, Lengyel E. Adipose tissue and adipocytes support tumorigenesis and metastasis. *Biochim Biophys Acta* (2013) 1831(10):1533–41. doi: 10.1016/j.bbali.2013.02.010
127. Feng H, Liu Q, Zhang N, Zheng L, Sang M, Feng J, et al. Leptin promotes metastasis by inducing an epithelial-mesenchymal transition in A549 lung cancer cells. *Oncol Res* (2013) 21(3):165–71. doi: 10.3727/096504014X13887748696662
128. Hiraoka K, Zenmyo M, Watari K, Iguchi H, Fotovati A, Kimura YN, et al. Inhibition of bone and muscle metastases of lung cancer cells by a decrease in the number of monocytes/macrophages. *Cancer Sci* (2008) 99(8):1595–602. doi: 10.1111/j.1349-7006.2008.00880.x
129. Joshi S, Singh AR, Zulcic M, Bao L, Messer K, Ideker T, et al. Rac2 controls tumor growth, metastasis and M1-M2 macrophage differentiation in vivo. *PLoS One* (2014) 9(4):e95893. doi: 10.1371/journal.pone.0095893
130. Tariq M, Zhang JQ, Liang GK, He QJ, Ding L, Yang B. Gefitinib inhibits M2-like polarization of tumor-associated macrophages in Lewis lung cancer by targeting the STAT6 signaling pathway. *Acta Pharmacol Sin* (2017) 38(11):1501–11. doi: 10.1038/aps.2017.124
131. Herroon MK, Rajagurubandara E, Rudy DL, Chalasani A, Hardaway AL, Podgorski I. Macrophage cathepsin K promotes prostate tumor progression in bone. *Oncogene* (2013) 32(12):1580–93. doi: 10.1038/onc.2012.166
132. Tamura T, Kurishima K, Nakazawa K, Kagohashi K, Ishikawa H, Satoh H, et al. Specific organ metastases and survival in metastatic non-small-cell lung cancer. *Mol Clin Oncol* (2015) 3(1):217–21. doi: 10.3892/mco.2014.410
133. Wu B, Wei S, Tian J, Song X, Hu P, Cui Y. [Comparison of the Survival Time in the Non-small Cell Lung Cancer Patients with Different Organ Metastasis]. *Zhongguo Fei Ai Za Zhi* (2019) 22(2):105–10. doi: 10.3779/j.issn.1009-3419.2019.02.05
134. Funazo T, Nomizo T, Kim YH. Liver Metastasis Is Associated with Poor Progression-Free Survival in Patients with Non-Small Cell Lung Cancer Treated with Nivolumab. *J Thorac Oncol* (2017) 12(9):e140–1. doi: 10.1016/j.jtho.2017.04.027
135. He Y, Wang Y, Zhang S, Ren S, Li J, Zhou C. Hepatic metastasis is a poor predictive marker for erlotinib in lung adenocarcinoma. *Med Hypotheses* (2016) 94:20–2. doi: 10.1016/j.mehy.2016.06.009
136. Clark AM, Ma B, Taylor DL, Griffith L, Wells A. Liver metastases: Microenvironments and ex-vivo models. *Exp Biol Med (Maywood)* (2016) 241(15):1639–52. doi: 10.1177/1535370216658144
137. Kim JY, Lee S, Hwangbo B, Lee CT, Kim YW, Han SK, et al. NF-kappaB activation is related to the resistance of lung cancer cells to TNF-alpha-induced apoptosis. *Biochem Biophys Res Commun* (2000) 273(1):140–6. doi: 10.1006/bbrc.2000.2909
138. Lin X, Chen Q, Huang C, Xu X. CYLD Promotes TNF-alpha-Induced Cell Necrosis Mediated by RIP-1 in Human Lung Cancer Cells. *Mediators Inflammation* (2016) 2016:1542786. doi: 10.1155/2016/1542786
139. You BR, Han BR, Park WH. Suberoylanilide hydroxamic acid increases anti-cancer effect of tumor necrosis factor-alpha through up-regulation of TNF receptor 1 in lung cancer cells. *Oncotarget* (2017) 8(11):17726–37. doi: 10.18632/oncotarget.14628
140. Yan HQ, Zhang D, Shi YY, You X, Shi L, Li Q, et al. Ataxia-telangiectasia mutated activation mediates tumor necrosis factor-alpha induced MMP-13 up-regulation and metastasis in lung cancer cells. *Oncotarget* (2016) 7(38):62070–83. doi: 10.18632/oncotarget.11386
141. Lin CW, Chen LJ, Lee PL, Lee CI, Lin JC, Chiu JJ. The inhibition of TNF-alpha-induced E-selectin expression in endothelial cells via the JNK/NF-kappaB pathways by highly N-acetylated chitoooligosaccharides. *Biomaterials* (2007) 28(7):1355–66. doi: 10.1016/j.biomaterials.2006.11.006
142. Tanigawa N, Hagiwara M, Tada H, Komatsu T, Sugiura S, Kobayashi K, et al. Acacetin inhibits expression of E-selectin on endothelial cells through regulation of the MAP kinase signaling pathway and activation of NF-kappaB. *Immunopharmacol Immunotoxicol* (2013) 35(4):471–7. doi: 10.3109/08923973.2013.811596
143. Chen L, Zhao Q, Wang XL, You R, Zhang YH, Ji H, et al. LJ-6, a novel COX/5-LOX inhibitor, attenuates TNF-alpha-induced endothelial E-selectin, ICAM-1 and VCAM-1 expression and monocyte-endothelial interactions via a COX/5-LOX-independent mechanism. *Vascul Pharmacol* (2011) 55(5-6):135–42. doi: 10.1016/j.vph.2011.07.003
144. Lin CF, Chen CL, Chien SY, Tseng PC, Wang YC, Tsai TT. Oxidative Stress Facilitates IFN-gamma-Induced Mimic Extracellular Trap Cell Death in A549 Lung Epithelial Cancer Cells. *PLoS One* (2016) 11(8):e0162157. doi: 10.1371/journal.pone.0162157
145. Lin CF, Chien SY, Chen CL, Hsieh CY, Tseng PC, Wang YC. IFN-gamma Induces Mimic Extracellular Trap Cell Death in Lung Epithelial Cells Through Autophagy-Regulated DNA Damage. *J Interferon Cytokine Res* (2016) 36(2):100–12. doi: 10.1089/jir.2015.0011
146. Chen CL, Chiang TH, Tseng PC, Wang YC, Lin CF. Loss of PTEN causes SHP2 activation, making lung cancer cells unresponsive to IFN-gamma. *Biochem Biophys Res Commun* (2015) 466(3):578–84. doi: 10.1016/j.bbrc.2015.09.085
147. Gao Y, Yang J, Cai Y, Fu S, Zhang N, Fu X, et al. IFN-gamma-mediated inhibition of lung cancer correlates with PD-L1 expression and is regulated by PI3K-AKT signaling. *Int J Cancer* (2018) 143(4):931–43. doi: 10.1002/ijc.31357
148. Vahl JM, Friedrich J, Mittler S, Trump S, Heim L, Kachler K, et al. Interleukin-10-regulated tumour tolerance in non-small cell lung cancer. *Br J Cancer* (2017) 117(11):1644–55. doi: 10.1038/bjc.2017.336
149. Khatib AM, Kontogianna M, Fallavollita L, Jamison B, Meterissian S, Brodt P. Rapid induction of cytokine and E-selectin expression in the liver in response to metastatic tumor cells. *Cancer Res* (1999) 59(6):1356–61.
150. Brodt P, Fallavollita L, Bresalier RS, Meterissian S, Norton CR, Wolitzky BA. Liver endothelial E-selectin mediates carcinoma cell adhesion and promotes liver metastasis. *Int J Cancer* (1997) 71(4):612–9. doi: 10.1002/(SICI)1097-0215(19970516)71:4<612::AID-IJC17>3.0.CO;2-D
151. Borentain P, Carmona S, Mathieu S, Jouve E, El-Battari A, Gerolami R. Inhibition of E-selectin expression on the surface of endothelial cells inhibits hepatocellular carcinoma growth by preventing tumor angiogenesis. *Cancer Chemother Pharmacol* (2016) 77(4):847–56. doi: 10.1007/s00280-016-3006-x
152. Chen Q, Massague J. Molecular pathways: VCAM-1 as a potential therapeutic target in metastasis. *Clin Cancer Res* (2012) 18(20):5520–5. doi: 10.1158/1078-0432.CCR-11-2904
153. Chen M, Peng W, Hu S, Deng J. miR-126/VCAM-1 regulation by naringin suppresses cell growth of human non-small cell lung cancer. *Oncol Lett* (2018) 16(4):4754–60. doi: 10.3892/ol.2018.9204
154. Kim MR, Jang JH, Park CS, Kim TK, Kim YJ, Chung J, et al. A Human Antibody That Binds to the Sixth Ig-Like Domain of VCAM-1 Blocks Lung Cancer Cell Migration In Vitro. *Int J Mol Sci* (2017) 18(3):566. doi: 10.3390/ijms18030566
155. Enomoto K, Nishikawa Y, Omori Y, Tokairin T, Yoshida M, Ohn N, et al. Cell biology and pathology of liver sinusoidal endothelial cells. *Med Electron Microsc* (2004) 37(4):208–15. doi: 10.1007/s00795-004-0261-4
156. Pasco S, Ramont L, Maquart FX, Monboisse JC. Control of melanoma progression by various matrikines from basement membrane macromolecules. *Crit Rev Oncol Hematol* (2004) 49(3):221–33. doi: 10.1016/j.critrevonc.2003.09.006
157. Aksorn N, Chanvorachote P. Integrin as a Molecular Target for Anti-cancer Approaches in Lung Cancer. *Anticancer Res* (2019) 39(2):541–8. doi: 10.21873/anticancer.13146
158. Petpiroon N, Sritularak B, Chanvorachote P. Phoyunnanin E inhibits migration of non-small cell lung cancer cells via suppression of epithelial-to-mesenchymal transition and integrin alphav and integrin beta3. *BMC Complement Altern Med* (2017) 17(1):553. doi: 10.1186/s12906-017-2059-7
159. Saisongkorh V, Maiuthed A, Chanvorachote P. Nitric oxide increases the migratory activity of non-small cell lung cancer cells via AKT-mediated integrin alphav and beta1 upregulation. *Cell Oncol (Dordr)* (2016) 39(5):449–62. doi: 10.1007/s13402-016-0287-3
160. Stewart RL, West D, Wang C, Weiss HL, Gal T, Durbin EB, et al. Elevated integrin alpha6beta4 expression is associated with venous invasion and decreased overall survival in non-small cell lung cancer. *Hum Pathol* (2016) 54:174–83. doi: 10.1016/j.humpath.2016.04.003
161. Wei X, Wang X, Zhan J, Chen Y, Fang W, Zhang L, et al. Smurf1 inhibits integrin activation by controlling Kiflin-2 ubiquitination and degradation. *J Cell Biol* (2017) 216(5):1455–71. doi: 10.1083/jcb.201609073

162. Tabaries S, Dupuy F, Dong Z, Monast A, Annis MG, Spicer J, et al. Claudin-2 promotes breast cancer liver metastasis by facilitating tumor cell interactions with hepatocytes. *Mol Cell Biol* (2012) 32(15):2979–91. doi: 10.1128/MCB.00299-12
163. Hichino A, Okamoto M, Taga S, Akizuki R, Endo S, Matsunaga T, et al. Down-regulation of Claudin-2 Expression and Proliferation by Epigenetic Inhibitors in Human Lung Adenocarcinoma A549 Cells. *J Biol Chem* (2017) 292(6):2411–21. doi: 10.1074/jbc.M116.762807
164. Sonoki H, Tanimae A, Endo S, Matsunaga T, Furuta T, Ichihara K, et al. Kaempferol and Luteolin Decrease Claudin-2 Expression Mediated by Inhibition of STAT3 in Lung Adenocarcinoma A549 Cells. *Nutrients* (2017) 9(6):597. doi: 10.3390/nu9060597
165. Zhao H, Zhang D, Yang L, Wang E. p0071 interacts with E-cadherin in the cytoplasm so as to promote the invasion and metastasis of non-small cell lung cancer. *Mol Carcinog* (2018) 57(1):89–96. doi: 10.1002/mc.22734
166. Farmakovskaya M, Khromova N, Rybko V, Dugina V, Kopnin B, Kopnin P. E-Cadherin repression increases amount of cancer stem cells in human A549 lung adenocarcinoma and stimulates tumor growth. *Cell Cycle* (2016) 15(8):1084–92. doi: 10.1080/15384101.2016.1156268
167. Yates CC, Shepard CR, Stolz DB, Wells A. Co-culturing human prostate carcinoma cells with hepatocytes leads to increased expression of E-cadherin. *Br J Cancer* (2007) 96(8):1246–52. doi: 10.1038/sj.bjc.6603700
168. Hsu PC, Jablons DM, Yang CT, You L. Epidermal Growth Factor Receptor (EGFR) Pathway, Yes-Associated Protein (YAP) and the Regulation of Programmed Death-Ligand 1 (PD-L1) in Non-Small Cell Lung Cancer (NSCLC). *Int J Mol Sci* (2019) 20(15):3821. doi: 10.3390/ijms20153821
169. Li ZQ, Yan HC, Gu JJ, Yang YL, Fang XJ, Zhang MK. Comparative efficacy and safety of PD-1/PD-L1 Inhibitors versus platinum-based chemotherapy for the first-line treatment of advanced non-small cell lung cancer: a meta analysis of randomized controlled trials. *Pharmacol Res* (2020) 160:105194. doi: 10.1016/j.phrs.2020.105194
170. Carcereny E, Moran T, Capdevila L, Cros S, Vila L, de Los Llanos Gil M, et al. The epidermal growth factor receptor (EGFR) in lung cancer. *Transl Respir Med* (2015) 3:1. doi: 10.1186/s40247-015-0013-z
171. Maity S, Pai KSR, Nayak Y. Advances in targeting EGFR allosteric site as anti-NSCLC therapy to overcome the drug resistance. *Pharmacol Rep* (2020) 72(4):799–813. doi: 10.1007/s43440-020-00131-0
172. Oh DY, Bang YJ. HER2-targeted therapies - a role beyond breast cancer. *Nat Rev Clin Oncol* (2020) 17(1):33–48. doi: 10.1038/s41571-019-0268-3
173. Shrestha L, Singh SS, Parajuli P, Dahal A, Mattheolabakis G, Meyer S, et al. In vivo studies of a peptidomimetic that targets EGFR dimerization in NSCLC. *J Cancer* (2020) 11(20):5982–99. doi: 10.7150/jca.46320
174. Wang Y, Xu C, Xu B, Li L, Li W, Wang W, et al. Xiaohai Jiedu Recipe Inhibits Proliferation and Metastasis of Non-Small Cell Lung Cancer Cells by Blocking the P38 Mitogen-Activated Protein Kinase (MAPK) Pathway. *Med Sci Monit* (2019) 25:7538–46. doi: 10.12659/MSM.917115
175. Suzuki M, Cao K, Kato S, Mizutani N, Tanaka K, Arima C, et al. CERS6 required for cell migration and metastasis in lung cancer. *J Cell Mol Med* (2020) 24(20):11949–59. doi: 10.1111/jcmm.15817
176. Shih JC. Monoamine oxidase isoenzymes: genes, functions and targets for behavior and cancer therapy. *J Neural Transm (Vienna)* (2018) 125(11):1553–66. doi: 10.1007/s00702-018-1927-8
177. Yang X, Zhao D, Li Y, Li Y, Cui W, Li Y, et al. Potential monoamine oxidase A inhibitor suppressing paclitaxel-resistant non-small cell lung cancer metastasis and growth. *Thorac Cancer* (2020) 11(10):2858–66. doi: 10.1111/1759-7714.13617

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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