



Reciprocal Interplay Between Fibrillar Collagens and Collagen-Binding Integrins: Implications in Cancer Progression and Metastasis

Isabelle Bourgot[†], Irina Primac[†], Thomas Louis, Agnès Noël and Erik Maquoi*

Laboratory of Tumor and Development Biology, GIGA-Cancer, University of Liège, Liège, Belgium

OPEN ACCESS

Edited by:

Serge Roche,
Institut National de la Santé et de la
Recherche Médicale
(INSERM), France

Reviewed by:

Birgit Leitinger,
Imperial College London,
United Kingdom
Olivier Destaing,
INSERM U1209 Institut pour
l'Avancée des Biosciences
(IAB), France

*Correspondence:

Erik Maquoi
erik.maquoi@uliege.be

[†]These authors share first authorship

Specialty section:

This article was submitted to
Molecular and Cellular Oncology,
a section of the journal
Frontiers in Oncology

Received: 13 February 2020

Accepted: 13 July 2020

Published: 18 August 2020

Citation:

Bourgot I, Primac I, Louis T, Noël A
and Maquoi E (2020) Reciprocal
Interplay Between Fibrillar Collagens
and Collagen-Binding Integrins:
Implications in Cancer Progression
and Metastasis.
Front. Oncol. 10:1488.
doi: 10.3389/fonc.2020.01488

Cancers are complex ecosystems composed of malignant cells embedded in an intricate microenvironment made of different non-transformed cell types and extracellular matrix (ECM) components. The tumor microenvironment is governed by constantly evolving cell-cell and cell-ECM interactions, which are now recognized as key actors in the genesis, progression and treatment of cancer lesions. The ECM is composed of a multitude of fibrous proteins, matricellular-associated proteins, and proteoglycans. This complex structure plays critical roles in cancer progression: it functions as the scaffold for tissues organization and provides biochemical and biomechanical signals that regulate key cancer hallmarks including cell growth, survival, migration, differentiation, angiogenesis, and immune response. Cells sense the biochemical and mechanical properties of the ECM through specialized transmembrane receptors that include integrins, discoidin domain receptors, and syndecans. Advanced stages of several carcinomas are characterized by a desmoplastic reaction characterized by an extensive deposition of fibrillar collagens in the microenvironment. This compact network of fibrillar collagens promotes cancer progression and metastasis, and is associated with low survival rates for cancer patients. In this review, we highlight how fibrillar collagens and their corresponding integrin receptors are modulated during cancer progression. We describe how the deposition and alignment of collagen fibers influence the tumor microenvironment and how fibrillar collagen-binding integrins expressed by cancer and stromal cells critically contribute in cancer hallmarks.

Keywords: fibrillar collagens, extracellular matrix, integrins, cancer, metastasis

INTRODUCTION

Cancer progression is a highly dynamic process implicating distinct features responsible for tumor growth and metastatic dissemination. These features comprise sustained proliferative signals, evading growth suppression, resisting cell death, stimulating angiogenesis, activating invasion and metastasis, deregulating cell metabolism and avoiding immune destruction (1). These hallmarks are obtained through reciprocal interactions between cellular and non-cellular components of tumors, which define the tumor microenvironment (TME). The critical role of the TME in cancer progression has been initially recognized in 1863 by Virchow, who first described that malignancies occurred at sites of chronic inflammation (2). In 1889, the

“seed and soil” hypothesis was proposed by Paget suggesting that the TME was important for tumor progression (3). Solid tumors are very heterogeneous and resemble a complicated organ whose complexity approaches and may even exceed that of normal healthy tissues (4). These tumors contain a complex mixture of non-cancerous cellular elements including blood and lymphatic vessels, pericytes, cancer-associated fibroblasts (CAFs), mesenchymal cells, immune/inflammatory cells and nervous network. Non-soluble or semi-soluble substances, such as the extracellular matrix (ECM), and soluble substances, such as interstitial fluids, cytokines, chemokines, growth factors, and metabolites constitute the acellular components of the TME (5–7). Physico-chemical parameters including interstitial pressures, oxygen level, and pH/redox potential represent additional critical characteristics of the TME. It is important to note that the TME is relatively abundant in comparison to cancer cells, with a proportional ratio nearly always in favor of the TME. In solid tumors, including breast and pancreatic tumors, the TME constitute up to 90% of the tumor mass (8–10). The TME is also characterized by a compositional and spatial heterogeneity, which varies greatly across tumor types, amongst patients with a given cancer type, and across distinct lesions in a given patient.

The ECM, which provides architectural support and anchorage for the cells, is composed of a complex meshwork of highly cross-linked components, including fibrous proteins, glycoproteins, proteoglycans, and polysaccharides (11–13). The biomechanical and biochemical properties of the ECM regulate cell survival, proliferation, differentiation and motility by ligating specific cell surface receptors including integrins, discoidin receptors and syndecans (14, 15). Besides these signaling properties, the ECM also play essential roles in tissue function by providing a structural and mechanical support for tissue integrity. It also influences the availability of cytokines and growth factors and maintains the hydration of the microenvironment. The structure of the ECM is highly dynamic and continuously remodeled through ECM deposition, degradation, or modification (16). Collagens are the most abundant components of the ECM, however, their structure and composition differ across various tissue types (17, 18). For example, the basement membrane, a well-structured matrix underlining epithelial and endothelial cells and separating them from the interstitial stroma, mainly consists of collagens type IV and VIII, while the interstitial stroma is mostly composed of fibrillar collagens type I, II, and III (19, 20).

Abnormal deposition and crosslinking of fibrillar collagens has serious repercussion on tissue homeostasis. Solid tumor's ECM is typically more rigid than normal tissue as a result of the overexpression of several ECM components, including collagens I, II, III, V, IX, XI, and heparan sulfate proteoglycans as well as ECM-crosslinking enzymes such as lysyl oxidases (21, 22). This accumulation generates a stroma characterized by a dense meshwork of fibrillar proteins (**Figure 1**), which progressively causes tissue stiffening, a hallmark of many cancers, such as breast, pancreatic and prostate cancers (23, 24). Stiffened ECM increases integrin-mediated mechanotransduction related signals, thereby promoting cancer cell survival, proliferation, and invasion (25–27). In epithelial cancers, the transition from an

in situ to an invasive carcinoma with associated high mortality is characterized by the focal degradation of the basement membrane (28). The breaching of the basement membrane by malignant cells is significantly influenced by the stiffness of the associated interstitial ECM (29). Tumor cell invasion through the basement membrane exposes malignant cells to a completely different microenvironment mostly dominated by the fibrillar collagens of the interstitial stroma (**Figure 1**). This new microenvironment rewires tumor cells by altering gene expression, cell proliferation, apoptosis, migration and survival, thereby directly affecting the hallmarks of cancer (30–33).

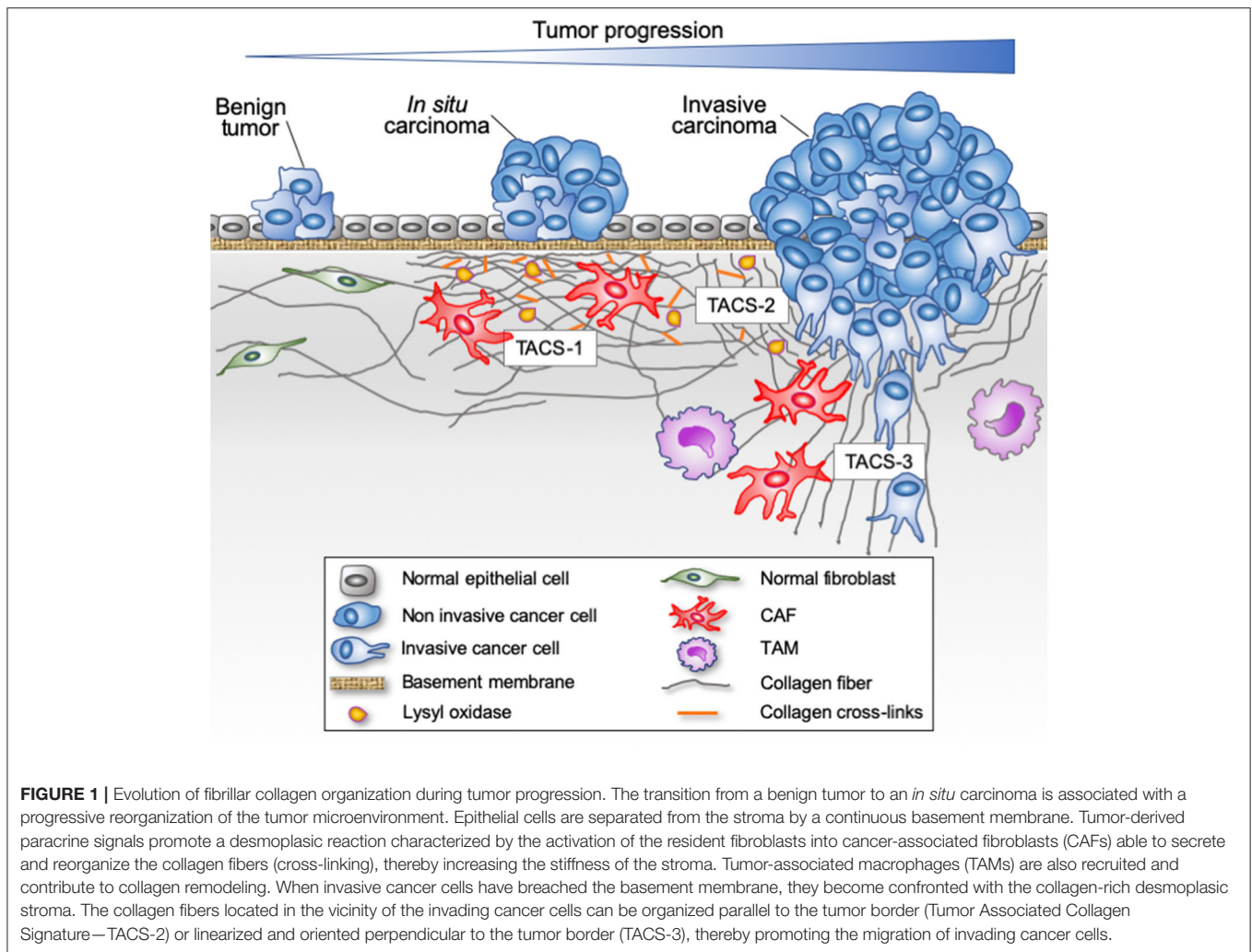
In this review, we describe the different fibrillar collagens and highlight how these proteins and their corresponding integrin receptors are modulated during cancer progression. We describe how the deposition and alignment of collagen fibers influence the TME and how integrin binding fibrillar collagen expressed by cancer and stromal cells represent critical players in cancer hallmarks. A brief overview of the different imaging techniques used to visualize and analyze fibrillar collagens is also provided.

COLLAGENS

Collagens, which constitutes up to 30% of the total protein mass in the human body, represent the most abundant proteins in mammals and are characteristic of the metazoan family (34, 35). In the human genome, 44 collagen genes code for polypeptidic chains and are combined in diverse ways to form 28 collagen types, numbered with roman numerals in vertebrates (I–XXVIII) (36, 37).

The term “collagen” is commonly used to refer to homotrimeric and heterotrimeric proteins formed by three polypeptide chains (α -chains). A characteristic feature of all collagens is the presence of a tight right-handed triple helix composed of three polypeptides α -chains forming a functional collagen molecule (**Figure 2**) (36, 39, 40). The triple helix motif can represent up to 96% of the collagen structure (for collagen I) to <10% (collagen XII) (41). Collagen molecules are made up of a tight right-handed helix composed of three α -chains, each of which contains one or more regions characterized by the repeating amino acid motif (Gly-X-Y)_n, with proline and 4-hydroxyproline amino acids often found at the X and Y positions, respectively (42). The presence of a glycine residue in every third position is required for the assembly into a triple helix. Indeed, the tight packing of the three α -chains near the common axis induces steric constraints on every third amino acid position and only glycine, the smallest amino acid can accommodate without any chain deformation. Consequently, the glycine residues are positioned in the center of the triple helix, where they stabilize the structure (42–44). Some collagen molecules assemble as homotrimers, whereas others assemble as heterotrimers composed of two or three distinct α -chain types. For example, type I collagen contains two identical $\alpha 1$ chains and one $\alpha 2$ chain, [$\alpha 1(I)$]₂ $\alpha 2(I)$. Each α -chain forms an extended left-handed helix with a pitch of 18 amino acid per turn (45).

All members of the collagen family present diverse supramolecular assemblies in the ECM and the capacity to bind



to cell surface receptors or other protein glycosaminoglycans (GAGs) (41). Their size, function and tissue distribution may vary considerably from networks to fibrils. The existence of several α -chains, different supramolecular structures for each collagen type, diverse molecular isoforms as well as the use of alternative promoters and alternative splicing highlight the complexity and diversity of the collagen family (46, 47). Based on this variability, vertebrate collagens have been classified into different families. The most abundant collagen family, with about 90% of the total collagens, is represented by the fibrillar collagens (Table 1). In human, the main fibrillar collagens include types I, II and III (major fibrillar collagens), V and XI (minor fibrillar collagens), and the more recently discovered types XXIV and XXVII (49, 50). Fibrillar collagens are characterized by the presence of uninterrupted triple-helical domains of about 300 nm, forming large extracellular fibrils. Type I collagen represents the archetypal fibrillar collagen due to the presence of a trimeric structure and the absence of imperfection in the triple helix. This molecular organization contrasts with that of other collagen families, which present interruptions in the triple helix or do not assemble into fibrils (Figure 3).

Network-forming collagens and anchoring fibril (ex: type IV and type VII collagens) have extended triple helices (>350 nm) with imperfections in the Gly-X-Y repeat sequences. Types VI, VIII, and X collagen are characterized by the presence of short continuous triple-helical domains. Type VI belongs to beaded filament (51), while types VIII and X form hexagonal networks (40). FACITs (Fibril-Associated Collagens with Interrupted Triple helices) are referred to as fibril-associated molecular bridges. They include collagen IX, XII, XIV, XVI, XIX, XX, XXI, XXII, and XXVI, which are associated to the surface of different collagen fibrils (Figures 2, 3) (40, 52). Collagens type XIII, XVII, XXIII, and XXV represent the MACIT, Membrane-Associated Collagens with Interrupted Triple helices, and function as cell surface molecules with a transmembrane domain. Finally, the multiplexin family (collagens XV and XVIII) is characterized by the presence of multiple triple helix domains interrupted by non-collagenous domains (36).

In this review, we will focus on the fibrillar and fibrillar-associated collagens. For further information on non-fibrillar collagens, the reader is referred to other publications (18, 41, 53, 54).

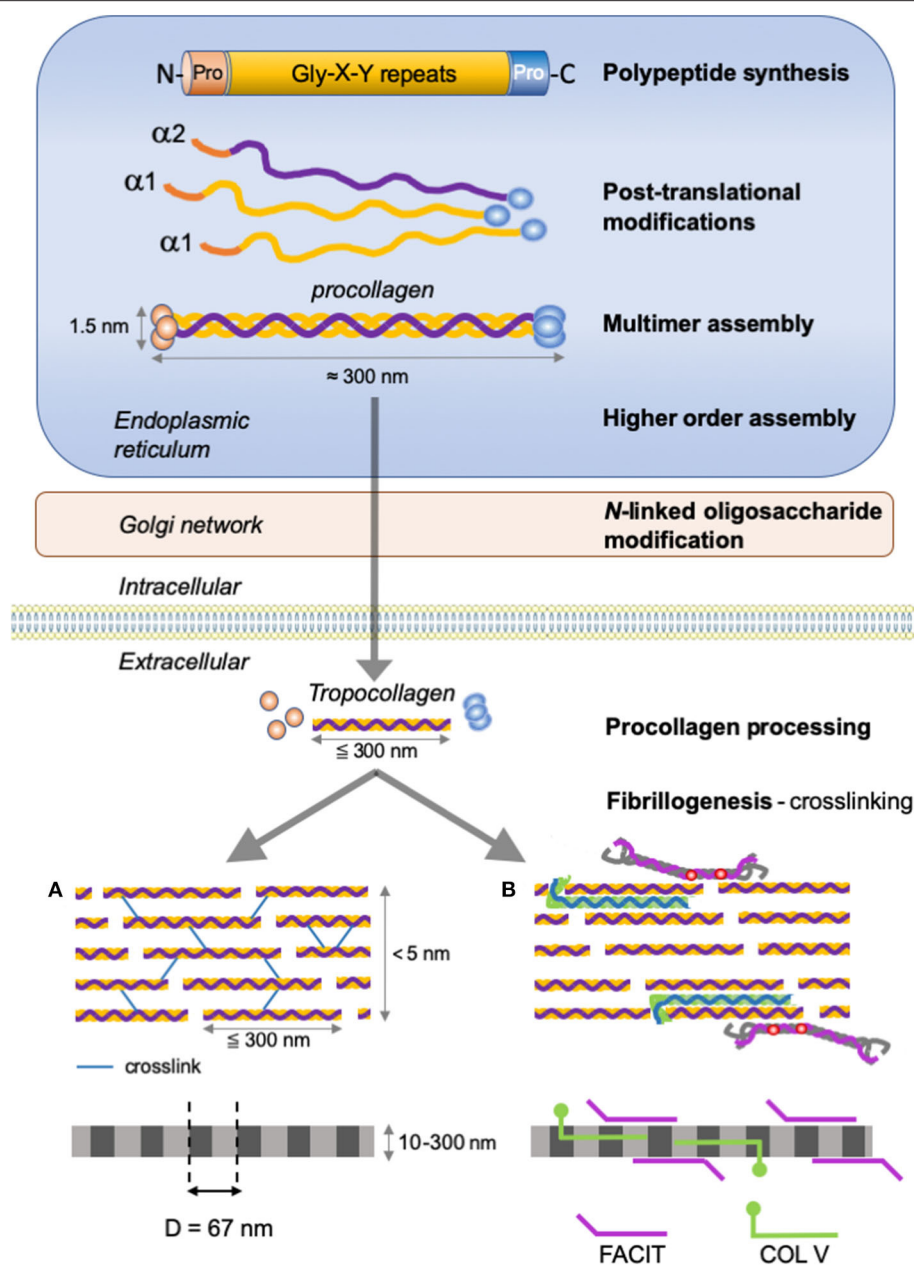


FIGURE 2 | Type I collagen supramolecular assembly pathway. The standard fibrillar collagen molecule is characterized by N- and C-terminal propeptide sequences, which flank a series of Gly-X-Y repeats (where X and Y represent any amino acids but are frequently proline and hydroxyproline). These form the central triple helical structure of procollagen and collagen. Three precursor α -chains (two $\alpha 1$ and one $\alpha 2$) are co-translationally translocated into the endoplasmic reticulum lumen, where specific post-translational modifications occur. Three collagen α -chains associate specifically via their C-terminal domains to form heterotrimers. The helical collagens are trafficked via the Golgi network to the plasma membrane, and secreted into the extracellular space as precursor forms, called procollagens, with N- and C-terminal non-collagenous domains. These domains are removed by the action of specific proteases, and the collagens are assembled into dense fibrils with a characteristic D-periodicity of about 67 nm (A). The fibril is stabilized by covalent lysine- and hydroxylysine-derived crosslinks. In addition to fibrillar collagen, other collagens, such as type V and FACIT collagens, are incorporated into the fibril structure (B). Type V collagen is inserted between strands of the microfibril, and FACIT collagens cling to the surface of the microfibril and work to stabilize higher order structures. Adapted from (38).

FIBRILLAR COLLAGENS

A key characteristic of fibrillar collagens is their ability to assemble and to form highly orientated supramolecular

aggregates. Type I collagen represents 90% of the total collagen. It constitutes the major collagen of the skin, tendons, ligaments, cornea, and other interstitial connective tissues. Type I collagen is mostly incorporated into a composite containing either type

TABLE 1 | Molecular chain compositions of human fibrillar and FACIT collagens.

Classes	Type	Molecular composition	Remarks
Fibrillar collagen	I	$[1\alpha(I)]_2\alpha 2(I)$	Fibrils composed of uninterrupted triple helical domains that are primarily found in skin, bone, tendons and ligaments, cartilage, cornea
	II	$[\alpha 1(II)]_3$	
	III	$[\alpha 1(III)]_3$	
	V	$[\alpha 1(V)]_2, \alpha 2(V); [\alpha 1(V)]_3; \alpha 1(XI)\alpha 2(V)\alpha 3(XI)$	
	XI	$\alpha 1(XI)\alpha 2(XI)\alpha 3(XI)$	
	XXIV	$[\alpha 1(XXIV)]_3$	
	XXVII	$[\alpha 1(XXVII)]_3$	
FACIT collagen	IX	$[\alpha 1(IX), \alpha 2(IX), \alpha 3(IX)]$	Fibril-associated collagens with interrupted triple helices: forming molecular bridges to the surface of collagen fibrils (e.g., IX, XII, XIV, XVI, XIX, XX, XXI)
	XII	$[\alpha 1(XII)]_3$	
	XIV	$[\alpha 1(XIV)]_3$	
	XVI	$[\alpha 1(XVI)]_3$	
	XIX	$[\alpha 1(XIX)]_3$	
	XX	$[\alpha 1(XX)]_3$	
	XXI	$[\alpha 1(XXI)]_3$	
XXII	$[\alpha 1(XXII)]_3$		

Adapted from (41, 48).

III collagen in skin and reticular fibers (55) or type V collagen in bone (56). The biomechanical properties of these composites (e.g., torsional stability, tensile strength, torsional stiffness) ensure the stability and integrity of these tissues (57, 58). The production of fibrillar collagens requires several intracellular and extracellular post-translational steps, which lead to the formation of elongated, cable-like striated fibril structures that are capable of withstanding tensile forces. This illustrates the close relationship between the three-dimensional protein structure and the role of the resultant ECM.

BIOSYNTHESIS AND ARRANGEMENT IN SUPERSTRUCTURE

Collagen biosynthesis is a complex multistep process starting with the synthesis of long α -chains precursors called *procollagens* (Figure 2). The growing peptide chains are co-translationally transported into the rough endoplasmic reticulum where multiple co- and post-translational modifications take place prior to the formation of the triple-helical procollagen.

The fibrillar collagen precursor is synthesized as a multidomain precursor constituted by a long triple helical COL1 domain (about 300 nm in length) flanked by two non-collagenous domains: a specific trimeric C-terminal NC1 domain (C-telopeptide and a C-propeptide) and a N-terminal NC2 domain (a short, non-helical, N-telopeptide and an N-terminal propeptide) (Figure 2). These non-collagenous domains represent important structural components: the C-propeptide plays an essential function in the initiation of triple helix formation and the N-propeptide is implicated in the regulation of primary fibril diameters (38).

Extensive post-translational modifications, including hydroxylation of proline and lysine residues, glycosylation of

lysine and hydroxylysine residues, occur prior to the formation of the triple helix (36). These modifications are stopped by the formation of the triple helix. Hydroxylation of proline and lysine residues are catalyzed by prolyl 3-hydroxylase, collagen prolyl 4-hydroxylase (C-P4Hs), and lysyl hydroxylase, respectively. These proline and hydroxyproline amino acids can comprise up to 20% of the molecule. Prolyl hydroxylation (in Y position) is essential for intramolecular hydrogen bonding and to increase the thermal stability of the triple helix. In fibril forming collagens ~50% of prolines are 4-hydroxylated; the extent of these hydroxylations varies between tissues and collagen types (59). Finally, glucosyl and galactosyl residues are added to the hydroxyl groups of hydroxylysine residues by the hydroxylysyl galactosyltransferase and galactosylhydroxylysyl-glucosyltransferase, respectively. Thus, the three α -chains are maintained together by intramolecular hydrogen bonds (high proline and 4-hydroxyproline content-dependent) and electrostatic interactions involving lysine and aspartate (42, 60–62). Highly ordered hydration networks surround the triple helices allowing a close packaging along the central axis of the molecule. Following hydroxylation and glycosylation processes, C-propeptide domains have an essential function (as nucleus) in the initiation and folding of the triple helix (43, 44, 63, 64). These domains allow the proper selection and alignment of collagen α -chains via intrachain disulfide bonds between the C-terminal propeptides of three procollagens sequences. The central portions of the chains zipper from C- to N-terminus to form the triple helix (65).

Many chaperone proteins including the heat shock protein 47 (HSP47) may influence and guide the formation of the triple helix during procollagen assembly (66). The binding of HSP47 enables the effective assembly and folding of the procollagen chains and facilitates the stabilization of procollagen triple helix at body temperature.

Once assembled in the endoplasmic reticulum, procollagen molecules are packaged in the Golgi for export into the ECM. Once released in the extracellular space, the procollagen is cleaved to form the mature triple-helical collagen or tropocollagen. Cleavage of N- and C-propeptides allows spontaneous fibrillogenesis in the extracellular environment. During or following exocytosis, extracellular proteinases, the procollagen N-proteinases (identified as the ADAMTS-2, -3, -14) and procollagen C-proteinases (identified as the BMP-1/tolloid proteinases) remove the N- and C-terminal propeptides (67–74). The removal of these propeptides exposes the telopeptides (short non-triple helical extensions of the polypeptide chains), which become binding sites for further covalent crosslinking during fibrillogenesis (75). The complete removal of the propeptides from collagen I, II and III fibrils allows fibrillogenesis in the extracellular space (76). In some cases, propeptides terminal-ends remain attached or partially attached, thereby affecting the solubility of the procollagen in the extracellular space, inhibiting premature fibril assembly or influencing fibril shape and diameter. For example, the uncleaved N-propeptides of collagen type V and XI influence fibril growth by sterically limiting lateral molecule addition (77).

Fibrillar collagens and FACITs fibril

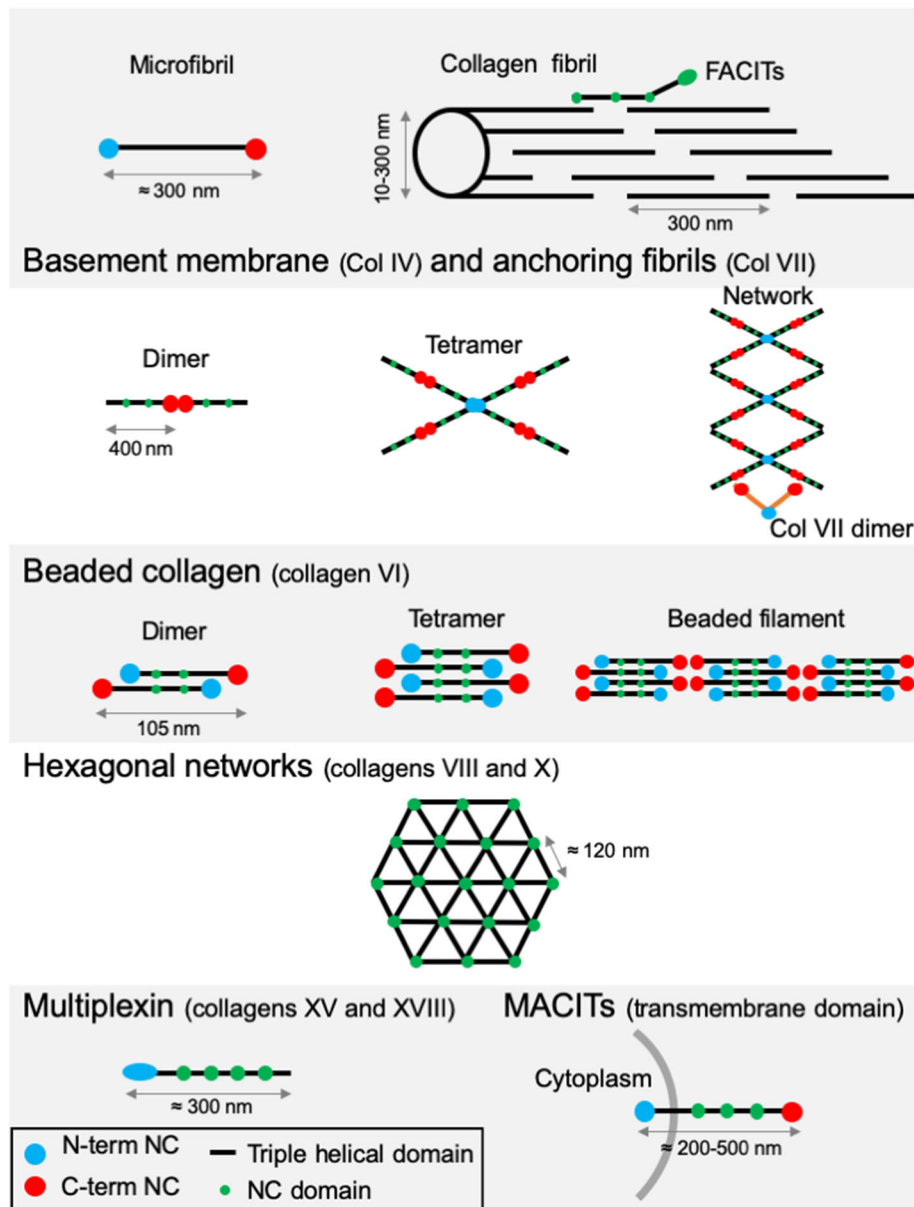


FIGURE 3 | Supramolecular structures formed by some archetypal collagens. *Fibrillar collagens and FACITs fibrils:* the association of mature protomers together leads to the formation of microfibrils which in turn assemble into fibrils. FACITs protomers attach at the surface of fibers with the C-terminal part protruding and regulate fibrillogenesis. *Basement membrane and anchoring fibrils:* formation of type IV collagen dimer occurs by the association of two protomers through their globular NC C-terminal domain. Dimers interact together through their N-terminal domains to constitute tetramers. Networks are the result of the two first steps linked to additional lateral interactions between the molecules. Dimers of type VII collagen interact with the network of type IV collagen. *Beaded collagen:* an association between the type VI collagen dimer and tetramer takes place inside the cells. Connection of tetramers leads to the formation of long filaments called "beaded filaments" according to their appearance in electron microscopy. *Hexagonal networks:* collagens VIII and X form hexagonal networks in Descemet's membrane and in hypertrophic cartilage, respectively. *Multiplexin:* collagens XVIII and XV are found in basement membrane. *MACITs:* transmembrane collagens (XIII, XVII, XXIII, and XXV). The N-terminal NC domain (N-terminal NC) is located inside the cell, whereas the triple helix region is extracellular. NC, non-collagenous domain. Adapted from (41).

After processing, the mature protein (*tropocollagen*), consists almost entirely of a triple-stranded helix and is considered as the building block for higher order fibrils and fibers. The self-assembly of tropocollagen monomers (300 nm long, 1.4 nm

diameter) results in the formation of collagen microfibrils with a quarter-stagger axial D periodicity of 67 nm long repeats to create the characteristic striation observed in collagen-containing tissues (69, 78). The staggered arrangement

optimizes electrostatic and hydrophobic interactions and allows the formation of covalent intermolecular cross-links between lysine/hydroxylysine residues of helical and neighboring non-helical regions thereby stabilizing the collagen fibrils (75, 79, 80). This process is initiated by the oxidative deamination of lysyl and hydroxylsyl residues in the N- and C-terminal telopeptide regions catalyzed by enzymes of the lysyl oxidase family (LOX) (81). The newly generated aldehyde group forms crosslinks with the (hydroxy)lysines in the triple-helical region of neighboring molecules. These links are crucial to confer the mechanical characteristics of collagen-containing tissues (82–84).

The short primary fibrils are unilaterally elongated via a multistage process including nucleation and organization to form intermediate-sized microfibrils. Once assembled, collagen microfibrils grow into fibrils through longitudinal and axial increase (77, 85). The assembly can also be regulated by many collagen binding proteins such as the FACITs (**Figure 2**), which integrate into the fibrils, selectively altering the surface properties of the collagen fibril as well as the interactions with accessory ECM molecules including the small leucine-rich repeat proteoglycans (78, 86).

IMPLICATIONS OF FIBRILLAR COLLAGENS IN CANCER PROGRESSION

In 1986, Dvorak reported an association between the TME and wound healing, suggesting that tumors behave as wounds that do not heal (87). In agreement with this concept, a fibro-inflammatory microenvironment has been shown to play critical roles in supporting tumor progression (88). Indeed, compelling studies suggest that a normal microenvironment prevents premalignant cells from progressing into cancer, whereas an abnormal or wound repair-associated microenvironment can be tumor-promoting (88). Reacting to a disruption in tissue homeostasis, resident fibroblasts are progressively activated into CAFs, which represent the most frequent cell type in the TME of many carcinomas, including pancreas, breast, and hepatic carcinomas (89, 90). During cancer progression, fibroblast activation and expansion is induced (**Figure 1**), such that low-grade premalignant lesions are already surrounded by areas of fibrosis (91). As these CAFs coevolve with the advancing cancer, they take on diverse functions that support tumor progression (92). In that context, CAFs represent key players in tumor fibrosis (also called desmoplasia), which is defined as a fibrotic state characterized by an excessive synthesis, deposition and remodeling of fibrillar collagens in the surroundings of the tumor (22, 93). Increasing evidence suggests heterogeneity among CAFs. Single-cell RNA-seq analyses of different mouse and human tumors highlighted the existence of at least three CAF populations (92, 94, 95). Among these, a population of CAFs has been coined myofibroblastic CAFs or matrix CAFs (mCAF) and abundantly produces a wide variety of ECM components including type I and type III fibrillar collagens, matrix-modifying enzymes such as LOX as well as a contractile phenotype. Consistent with its role as a key regulator of fibrogenic gene expression and the

myofibroblast state, TGF- β signaling promotes the mCAF phenotype (96, 97).

By exerting contraction forces on collagen fibers, CAFs induce the reorientation and alignment of fibrillar proteins. The dense collagen network is also crosslinked, thereby increasing matrix stiffness.

Atomic force microscopy analysis of a murine model of spontaneous breast cancer (MMTV-PyMT mice) revealed that while the elastic modulus of the normal mammary gland was about 400 Pa, it increased to 1,200 and 3,000 Pa in pre-malignant and malignant tumors, respectively (98). In human breast, the stiffness of the normal and non-invasive stroma is around 400 Pa. In sharp contrast, the invasive regions of aggressive tumors were much stiffer (>5 kPa). Interestingly, the tumor invasion front appears stiffer than the tumor core tissue (99).

This desmoplasia-associated collagen remodeling elicits biochemical and biophysical cues which influence stromal and tumor cell properties, providing crucial physical guidance facilitating cell migration, invasion, and metastasis through reciprocal interactions between the cells and the ECM (21, 26, 100–105).

Several key biophysical parameters of type I collagen matrix play a role in cancer cell migration, in particular the mechanical properties, the pore size, the density and orientation of cell adhesion sites presented by the collagen fibers, and the local direction of the fibers (106).

Collagen fiber diameter and pore size play key roles in cell force generation and migration (107, 108), with larger fiber diameter and pore size promoting cell force generation and migration (21).

During migration, cancer cells follow the path of least resistance, they recognize and use open pores within the matrix. A correlation between cell-migration speed and pore size has been shown (107, 109). Cancer-cell migration is also positively correlated with the stiffness of the tumor and associated stromal matrix (110, 111). However, fiber alignment appears as the best predictor of cell speed in collagen matrix.

The mode of cell migration is also dictated by the microstructure of the matrix. At low collagen densities, when the matrix pore size is larger than the nucleus of the migrating cell, matrix cleavage is not critical and cells move rapidly using pseudopodial protrusions in an amoeboid mode of migration (107, 112). Increased deposition of fibrillar collagens may lead to decreased porosity near the tumor. At high collagen densities, when the pore size is significantly smaller than the nucleus of the cell, cell movements rely more extensively on the cleavage of collagen fibers by secreted and cell membrane-associated collagenolytic enzymes including matrix metalloproteinases (MMPs), cathepsins, and serine proteinases. In this case, tumor cells undergo a more mesenchymal mode of migration: they use proteinase-assisted invadopodia to open the pore to the necessary size and move through them by transmitting forces via adhesion. However, some cells are able to migrate through small pores in the absence of matrix remodeling by disrupting and subsequently repairing their nuclei (113).

The proteinase-assisted migration generates channel-like tracks (3–30 μm in diameter and 100–600 μm in length) in the

matrix (114). These migration tracks opened by path-finding cells can subsequently be used by several following cancer cells (109, 115).

CAFs also play an active role in promoting cancer cell infiltration into the tumor stroma by taking the lead and forming tracks in which tumor cells follow (116). The remodeling of fibrillar collagens by CAFs and/or cancer cells may lead to heterogeneities in density and network organization.

The alignment of the fibrillar components of the ECM has a strong influence on the direction and speed of migrating cells (16, 117). Experimental models have demonstrated that cancer cells invade more efficiently through *in vitro* engineered lattices of linear type I collagen than through disorganized lattices (105, 106, 118). *In vivo*, local tumor cell invasion is oriented along radially aligned collagen fibers. These reorganized collagen fibers are called tumor-associated collagen signatures (TACS, **Figure 1**). Three TACS corresponding to different levels of collagen fibers reorganization have been described and represent novel markers to locate and characterize tumors (103, 119–121). In TACS-1 stage, a localized increase in the deposition of collagen without obvious alignment is observed near the tumor. In TACS-2 stage (pre-invasive tissue), the collagen fibers are aligned in parallel to the tumor border. In TACS-3 stage (metastatic stage), the collagen fibers are bundled and aligned perpendicular to the tumor border. Collagen linearization is therefore considered as a key feature of metastatic carcinomas and predictive of poor prognosis in breast carcinoma and *in situ* breast ductal carcinoma (121–123). The aligned collagen fibers provide tracks not only for cancer cells but also for macrophages, thereby promoting their migration into the TME (124). Recently, WISP1, a matricellular protein secreted by cancer cells, was shown to induce collagen linearization by binding directly type I collagen, thereby promoting its linearization, independently of cell-derived mechanical tensions (125). Moreover, fibroblast activation protein (FAP), another important ECM-associated proteinase, promotes the formation *in vitro* of desmoplastic-like aligned matrices (126) and its overexpression has been associated with a poor patient outcome.

The accumulation of cross-linked collagens and the subsequent stiffening of the matrix also lead to elevated interstitial fluid pressure in the desmoplastic TME inducing resistance to treatment by decreasing chemotherapy and immunotherapy drug delivery (127).

In addition to their biomechanical contribution to cancer progression, CAF-derived collagen-rich ECMs also indirectly fuel cancer cells with amino acids. Indeed, collagen uptake and catabolism, with subsequent proline catabolism, has been shown to support cancer cell proliferation (128).

Even if CAFs are usually considered as the key ECM remodelers in the TME, both epithelial and immune cells also contribute to the synthesis and secretion of fibrillar ECM components (129). Indeed, epithelial cells are able to secrete ECM components (including type I collagen) (130–133). In colorectal cancer, tumor associated macrophages (TAMs) could even play a more important role in collagen deposition, cross-linking, and linearization than CAFs (134).

The desmoplastic reaction has been frequently associated with a poor survival of cancer patients (135–140). According to clinical data, tumor collagen content, alignment and distribution have been considered as prognostic factors related to cancer differentiation, invasion and clinical outcome in different cancers (141–143). However, the implications of fibrillar collagens during cancer progression are not restricted to the primary tumors. Despite the successful treatment of a primary tumor, dormant disseminated tumor cells may be reactivated to form actively proliferating metastatic lesions (144, 145). This reactivation is associated with the induction of fibrosis characterized by the deposition of fibrillar collagens in the metastatic microenvironment (146).

Beside the above described functions in desmoplasia, fibrillar collagens are extremely important in major steps of cancer progression. Specific chains of these collagens and procollagens act as effectors and allow modulation of key processes in cancer progression (e.g., proliferation, apoptosis, angiogenesis, invasion, and metastasis). Some studies reveal that these effects can be either pro- or anti-tumorigenic and are collagen type-dependent (147, 148). A non-exhaustive overview of the contribution of each fibrillar collagens in cancer progression is presented in **Table 2**.

In order to complete this overview of the roles of fibrillar collagens in cancer, we explored *in silico*, whether the mRNA level of the different fibrillar collagens were connected with clinical outcome in human cancers. Patients were divided into two groups (low and high expression groups) according to the gene expression level in tumor tissue. Kaplan-Meier log rank analysis showed that a high expression level of a few collagens was associated with a significantly longer overall survival period in some cancers (e.g., COL2A1 in pancreatic ductal adenocarcinoma—**Figure 4A**). However, for most fibrillar collagens, high expression levels were linked with significantly shorter overall survival periods (e.g., COL5A1 in kidney renal papillary cell carcinomas—**Figure 4B**). The complete analysis of the prognostic value of the expression level of 11 fibrillar collagens on the survival of patients suffering from 13 different cancers is summarized in **Figure 4C**.

FIBRILLAR COLLAGENS, AGING AND CANCER: A DANGEROUS TRIO

The incidence of most cancers strongly increases with age and cancer represents the primary cause of death in population aged 60–79 years. The risk of having an invasive cancer in patients over 60 is more than 2-fold that of younger patients (183).

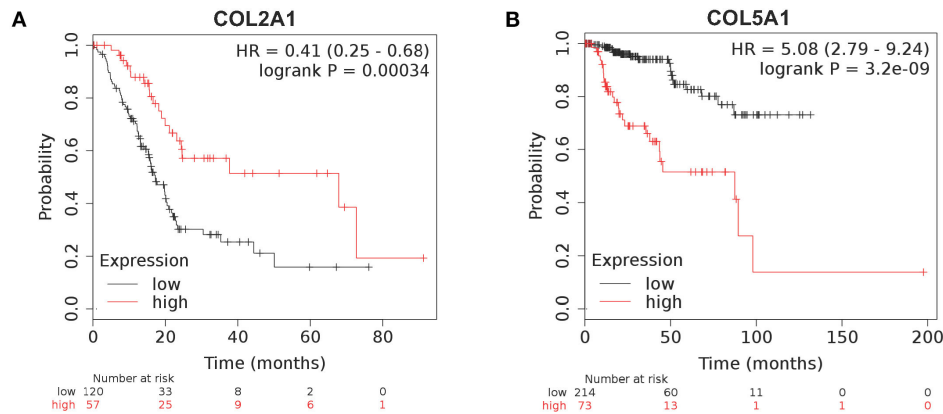
Many studies have shown that aging affects the normal cells of the TME. Among these stromal cells, fibroblasts and immune cells, which represent key actors in tumor progression and metastasis, are notably susceptible to this age-related impact (184). ECM integrity decreases substantially as we age. Age-related alterations in the physical features of the ECM comprise decreases in collagen density (185–187), ECM fiber thickness and area (188, 189).

TABLE 2 | Influence of fibrillar collagen expression on cancer properties.

Type	Subtype	Cancers	Implications	References
COL I	COL1A1	Hepatocellular	Clonogenicity, motility, invasiveness and stemness	(149)
		Bladder	Proliferation, migration, and invasion	(150)
		Malignant astrocytoma	Invasion	(151)
		Breast	Migration	(152)
		Colorectal	Migration	(153)
		Oral squamous cell carcinoma	Proliferation and migration	(154)
		Gastric	Proliferation, migration, and invasion	(155)
COL II	COL2A1	Ovarian	Prognostic value	(156)
		Gastric	Prognostic value	(157)
			Biomarker	(158)
COL III	COL3A1	Ovarian	Lymphovascular metastasis	(159)
			Patient survival	(160)
			Drug resistance	(161)
		Bladder	Patient survival	(162)
		Glioblastoma	Patient survival	(163)
		Colorectal	Patient survival	(164)
			Proliferation	(164)
COL V	COL5A1	Ovarian	Lymphovascular metastasis	(159)
		Breast	Viability and migration	(165)
			Patient survival	(165)
		Lung	Proliferation, apoptosis, invasion	(166)
			Patient survival	(166)
		Renal	Proliferation, apoptosis, migration, invasion	(167)
		Patient survival	(167)	
		Tongue squamous cell carcinoma	Patient survival and metastasis	(168)
	Breast	Prognostic value	(169)	
	COL5A2	Ovarian	Lymphovascular metastasis	(159)
		Tongue squamous cell carcinoma	Patient survival	(168)
Bladder		Patient survival	(170)	
	COL5A3	Breast	Proliferation	(171)
COL XI	COL11A1	Ovarian	Patient survival	(172)
			Invasion	(172)
			Drug resistance	(173)
		Non-small cell lung carcinoma	Proliferation, migration, invasion, drug resistance	(174)
		Pan-cancer	Marker of activated CAF	(175)
		Gastric	Proliferation, migration, invasion	(176)
		Esophageal squamous cell carcinoma	Prognostic value	(177)
		Pancreas	Prognostic value	(178)
		Breast	Prognostic value	(179)
			COL24A1	Head and neck
COL XXIV	COL24A1	Hepatocellular carcinoma	Prognostic value	(181)

Because of their long half-life, fibrillar collagens are subjected to post-translational changes during biological aging. These modifications include mineralization, accumulation of advanced glycation end-products (AGEs) (190), an increase in crosslinks level (191), and the reduction of glycosaminoglycans (GAGs), which impact fiber stability. These biochemical alterations change the structural organization of type I collagen (192–194). Such changes in structural properties of type I collagen affect its susceptibility to matrix metalloproteinases

(MMPs)-mediated degradation (185, 195), and lead to a tissues stiffening. These structural alterations also influence the recognition of the collagen fibers by cell surface receptors such as discoidin domain receptors (DDR), which are sensitive to the structural organization of the collagen. *In vitro* studies have shown that aged type I collagen upregulates the proliferation of fibrosarcoma cells (196) and presents a reduced pro-apoptotic potential toward luminal breast cancer cells (197).



C

	COL1 A1	COL1 A2	COL2 A1	COL3 A1	COL5 A1	COL5 A2	COL5 A3	COL11 A1	COL11 A2	COL24 A1	COL27 A1
Bladder carcinoma (n=404)	1.56 <i>0.028</i>	1.57 <i>0.003</i>	1.33 <i>0.085</i>	1.54 <i>0.0072</i>	1.59 <i>0.0018</i>	1.66 <i>0.00079</i>	2.26 <i>7.7x10⁻⁹</i>	1.43 <i>0.022</i>	0.74 <i>0.083</i>	0.8 <i>0.15</i>	0.77 <i>0.11</i>
Breast cancer (n=1089)	1.26 <i>0.16</i>	1.26 <i>0.17</i>	0.75 <i>0.078</i>	1.29 <i>0.013</i>	1.35 <i>0.079</i>	1.38 <i>0.069</i>	1.28 <i>0.016</i>	1.64 <i>0.0027</i>	1.14 <i>0.45</i>	1.22 <i>0.24</i>	0.73 <i>0.048</i>
Cervical squamous cell carcinoma (n=304)	2.19 <i>0.0057</i>	1.61 <i>0.09</i>	1.46 <i>0.12</i>	1.69 <i>0.036</i>	1.86 <i>0.01</i>	1.93 <i>0.0064</i>	1.76 <i>0.019</i>	2.21 <i>0.0006</i>	0.62 <i>0.12</i>	1.31 <i>0.31</i>	0.81 <i>0.39</i>
Esophageal adenocarcinoma (n=78)	1.66 <i>0.12</i>	1.45 <i>0.25</i>	1.91 <i>0.055</i>	1.95 <i>0.011</i>	1.79 <i>0.075</i>	1.59 <i>0.15</i>	1.47 <i>0.25</i>	2.33 <i>0.017</i>	0.78 <i>0.45</i>	1.3 <i>0.45</i>	0.64 <i>0.21</i>
Esophageal squamous cell carcinoma (n=81)	0.59 <i>0.2</i>	0.55 <i>0.25</i>	0.47 <i>0.064</i>	0.57 <i>0.028</i>	0.69 <i>0.37</i>	0.54 <i>0.12</i>	1.67 <i>0.27</i>	0.58 <i>0.23</i>	0.44 <i>0.051</i>	0.63 <i>0.27</i>	0.59 <i>0.2</i>
Head-neck squamous cell carcinoma (n=499)	1.2 <i>0.22</i>	0.85 <i>0.25</i>	1.26 <i>0.11</i>	1.2 <i>0.02</i>	1.27 <i>0.083</i>	1.26 <i>0.11</i>	1.15 <i>0.35</i>	1.327 <i>0.047</i>	0.82 <i>0.16</i>	1.21 <i>0.16</i>	1.13 <i>0.45</i>
Kidney renal clear cell carcinoma (n=530)	1.73 <i>0.0004</i>	1.55 <i>0.0098</i>	1.65 <i>0.0015</i>	1.35 <i>0.061</i>	1.77 <i>0.0002</i>	1.33 <i>0.083</i>	1.61 <i>0.0018</i>	1.92 <i>1.7x10⁻⁵</i>	1.65 <i>0.0023</i>	1.72 <i>0.0003</i>	1.33 <i>0.06</i>
Kidney renal papillary cell carcinoma (n=287)	4.63 <i>2.9x10⁻⁸</i>	4.03 <i>6.5x10⁻⁷</i>	3.39 <i>0.0032</i>	4.95 <i>8.2x10⁻⁹</i>	5.08 <i>8.2x10⁻⁹</i>	4.84 <i>1.2x10⁻⁸</i>	4.35 <i>2.5x10⁻⁷</i>	4.67 <i>3.9x10⁻⁸</i>	2.77 <i>0.00059</i>	3.64 <i>5.6x10⁻⁶</i>	0.47 <i>0.016</i>
Liver hepatocellular carcinoma (n=370)	1.41 <i>0.059</i>	1.51 <i>0.051</i>	1.62 <i>0.022</i>	1.38 <i>0.069</i>	1.4 <i>0.081</i>	1.4 <i>0.067</i>	1.37 <i>0.077</i>	1.62 <i>0.006</i>	0.65 <i>0.017</i>	2.01 <i>8.7x10⁻⁵</i>	0.77 <i>0.14</i>
Lung adenocarcinoma (n=504)	1.63 <i>0.0013</i>	1.38 <i>0.029</i>	0.77 <i>0.094</i>	1.35 <i>0.063</i>	1.46 <i>0.011</i>	1.5 <i>0.019</i>	1.43 <i>0.038</i>	1.91 <i>0.00025</i>	0.88 <i>0.41</i>	0.65 <i>0.0048</i>	0.75 <i>0.063</i>
Lung squamous cell carcinoma (n=495)	1.33 <i>0.038</i>	1.28 <i>0.075</i>	1.26 <i>0.16</i>	1.4 <i>0.022</i>	1.35 <i>0.031</i>	1.36 <i>0.031</i>	1.27 <i>0.079</i>	0.87 <i>0.31</i>	1.36 <i>0.04</i>	1.35 <i>0.077</i>	1.19 <i>0.21</i>
Ovarian cancer (n=373)	1.38 <i>0.019</i>	1.38 <i>0.019</i>	1.41 <i>0.02</i>	1.37 <i>0.026</i>	1.48 <i>0.005</i>	1.3 <i>0.065</i>	1.19 <i>0.21</i>	1.23 <i>0.13</i>	1.31 <i>0.045</i>	1.31 <i>0.052</i>	1.21 <i>0.17</i>
Pancreatic ductal adenocarcinoma (n=177)	1.4 <i>0.11</i>	1.43 <i>0.09</i>	0.41 <i>0.0003</i>	1.6 <i>0.075</i>	1.56 <i>0.09</i>	1.45 <i>0.077</i>	1.62 <i>0.046</i>	1.74 <i>0.011</i>	0.71 <i>0.11</i>	1.55 <i>0.039</i>	0.49 <i>0.0015</i>

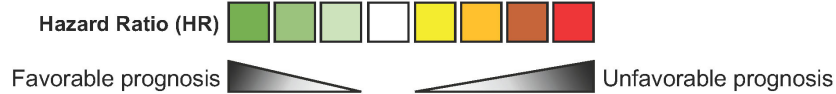


FIGURE 4 | Influence of 11 fibrillar collagen gene expression on patient prognosis outcome in 13 different cancers. Hazard ratio (HR) and log-rank *p*-values were calculated using the pan-cancer RNA-seq Kaplan-Meier plotter (182). **(A)** Kaplan-Meier plot showing that patients with a high COL2A1 gene expression (red lines) have a higher overall survival than those with a low gene expression (black lines). **(B)** Kaplan-Meier plot showing that patients with a high COL5A1 gene expression (red lines) have a lower overall survival than those with a low gene expression (black lines). HR and 95% confidence interval are shown. Log-rank *P* < 0.05 was considered to indicate statistical significance. **(C)** Summary of HR (**bold**) and log-rank *p*-values (*italic*) for 11 fibrillar collagen genes in 13 different cancers. Collagen genes whose mRNA levels were significantly associated with patient's overall survival in a specific cancer were color coded according to the log-rank *p*-values and HR (unfavorable prognosis: yellow to red; favorable prognosis: light to dark green).

IMAGING FIBRILLAR COLLAGENS

Several techniques can be used to image the architecture of collagen networks depending on the experimental conditions and the studied features. Histochemical (van Gieson staining, Picro-Sirius red, etc.) and immunohistochemical techniques are routinely used to stain fibrillar collagens (198–200). These techniques require additional fixation and preparation steps, which can alter the collagen structure of the samples, and hence are limited to *ex vivo* materials. Images of stained samples are obtained using bright-field, polarized and fluorescence microscopy techniques (199, 200). Laser scanning microscopes or multiphoton laser scanning microscopes have also been used to visualize stained tissues in three dimensions (198).

Transmission electron microscopy (TEM) and scanning electron microscopy (SEM) also allow to visualize collagens *ex vivo* and *in vitro*. The analyzed tissues must previously be washed, dehydrated, dried and sputter coated before imaging (103, 201, 202). While SEM and TEM provide highly detailed information on microstructure and nanostructure of collagens, respectively, the preparation steps may greatly alter the samples. SEM can be used to image morphology and arrangement of the collagen fibers but its three-dimensional imaging capability is limited (103). Besides these methods requiring a preparative step, non-disruptive techniques are also available. Among these techniques, confocal reflection microscopy (CRM) is a high resolution technique for imaging specimens which differ in refractive index from their surroundings or which possess a high reflectance (203). It is widely used to visualize polymers and biomaterials. A laser-scanning confocal microscope in reflection is able to detect variations of backscattered light intensity at the collagen-to-media interface for each sequential focal plane, resulting in the reconstruction of a three-dimensional image of the sample (204). CRM is readily applicable to dynamically follow living specimens. This technique can be combined with fluorescence confocal imaging without modifying the microscope hardware (76). While this visualization technique is mostly used *in vitro* to study the interactions between cells and ECM components (76, 194, 201, 203, 204), it also enables the *in vivo* visualization of epidermis and superficial dermis in real time, providing a tool for imaging skin lesions and helping skin cancer diagnosis (205–207). Excitation wavelength is proportional to the minimal resolution but decreasing it reduces the penetration depth (204). Hence, simultaneous excitation at 488 and 567 nm leads to better imaging than individual 488 and 567 nm excitations. In practice, a penetration depth of up to 100 μm is achieved for CRM on 3D collagen scaffolds, while resolution strongly declines after 30 μm .

Second harmonic generation (SHG) microscopy is a highly specific optical method of direct visualization of fibrillar collagens that can be carried out using most two-photon fluorescence microscopes (208). It allows the non-invasive assessment of the abundance and structure of fibrillar collagens with a high resolution and specificity. For those reasons, it represents the most widely used technique for the *in vivo* imaging of fibrillar collagens.

SHG occurs when two photons interact with optically non-linear material and merge to generate a new photon with twice the energy and half the wavelength of the initial photons. Fibrillar collagen non-linear optical response, which results in a strong SHG signal, originates from its non-centrosymmetric triple helical molecular assemblies, which exhibit large hyperpolarizabilities (209, 210). This process is sensitive to the microscopic structure of the scattering material. Therefore, SHG emission directionality is influenced by the diameter of the collagen fibrils that are bundled into fibers, their spacing within the fiber, and the disorder in their packing (209, 211, 212). Among the different collagens, type I collagen has the most ordered structure and, hence, produces the strongest SHG signal (213). Maximum SHG resolution is higher than what is obtained using linear optical microscopy techniques and enables to image collagen fibrils (209, 214). SHG signal intensity is dependent of the fibril/fiber orientation (210). In particular, fibers which are perpendicular to the laser light polarization axis will result in a weak signal. SHG is also very low with fibers perpendicular to the imaging plane (due to the centrosymmetric structure of fiber cross section). It should also be emphasized that, due to resolution limitations, collagen fibrils with a diameter lower than 100 μm will not be detected. The SHG optical sectioning capability enables tissue imaging in three dimensions (215). SHG imaging is therefore a label-free, non-destructive, high resolution, sensitive and specific modality for visualizing the spatial distribution of fibrillar collagens *in vitro*, *ex vivo* (Figure 5), and *in vivo*. SHG images can be subsequently analyzed with specific image analysis tools such as CT-FIRE which can extract individual collagen fibers from images for a quantitative assessment of fiber metrics including fiber angle, fiber length, fiber straightness, and fiber width (216). In the context of cancer, SHG measurement has been used as an independent prognostic indicator of metastatic outcome in patients with estrogen receptor positive, lymph node-negative breast cancer as well as in stage I colon adenocarcinomas (217).

Other techniques can be performed to directly monitor collagen fibers. Small angle X-ray scattering (SAXS) characterizes collagen fibril structure and organization into fibers of whole samples (218, 219). It represents an efficient way for observing and investigating the structural organization of collagen fibrils including their orientation. Collagen fibril orientation can then be used to derive the collagen fiber orientation. When used in combination with Fourier transformed infrared (FTIR) microspectroscopy, which gives chemical information on submolecular bounds and functional groups, and unsupervised hierarchical cluster analysis, it can characterize, classify and map cell clusters in *ex vivo* tissue samples (219). Magnetic resonance imaging (MRI) can be used to study *ex vivo* and *in vivo* the three-dimensional organization and density of collagens (220, 221). Finally, real-time collagen imaging in *in vivo* models can also be achieved by using transgenic lines expressing fluorescent protein-tagged fibrillar collagen (222).

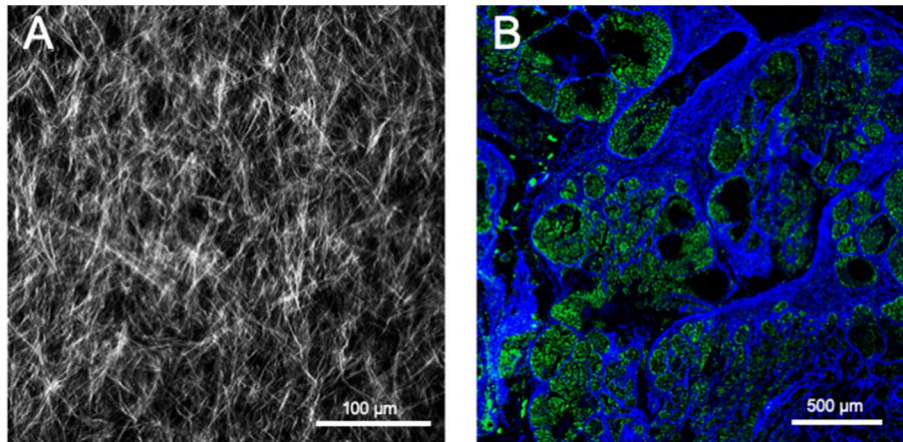


FIGURE 5 | Second harmonic generation (SHG) image of fibrillar collagens. **(A)** SHG image of *in vitro* polymerized collagen I. **(B)** SHG of a mouse mammary PyMT tumor section. Collagen fibers appear in blue, while the cellular structures appear in green due to the autofluorescence of the sample.

STRUCTURAL AND FUNCTIONAL FEATURES OF COLLAGEN-BINDING INTEGRINS

Fibrillar collagen receptors include different classes of molecules such as DDRs, mannose receptors, leucocyte receptor complex, as well as proteolytic enzymes. DDRs (DDR1 and DDR2) are tyrosine kinase receptors that become specifically activated by the native triple helix of fibrillar collagens I–III, followed by tyrosine autophosphorylation, receptor internalization and signaling (223, 224). Leucocyte receptor complex members like OSCAR (osteoclast associated receptor) and GPVI (glycoprotein VI) have been also shown to act as fibrillar collagen receptors and mediate processes such as osteoclastogenesis (225) and platelet activation and aggregation, respectively (226). The uPARAP/Endo180 mannose receptor has been shown to recognize fibrillar collagens such as collagen I, II, and V and acts as an endocytic receptor by modulating the collagen fragment internalization and its lysosomal degradation, playing a role in the fibrillar collagen turnover (227). In this context, proteolytic enzymes such as MMPs and cathepsins also recognize fibrillar collagens and mediate collagen degradation in the extracellular compartment (228). These different non-integrin collagen receptors have been shown to play key roles in specific hallmarks of cancer, including among others proliferation, apoptosis, drug resistance, inflammation, neo-angiogenesis and metastasis (227, 229).

Integrins comprise a large family of matricellular receptors involved in the transduction of the bidirectional signaling between cells and the surrounding ECM. Each integrin consists of a cell surface heterodimer of α and β subunits non-covalently associated and composed by a large ectodomain, a transmembrane domain and a short cytoplasmic tail (Figure 6). The ECM ligand specificity of integrins is ascribed to the large ectodomain, while the cytoplasmic tail is known to bind several intracellular regulatory and cytoskeletal molecules. Integrin α and β subunits heterodimerize in the endoplasmic reticulum,

undergo posttranslational modifications such as glycosylation in Golgi, and are subsequently transported in an inactive form at the cell surface where activated to interact with specific ECM ligands. Recognition of different ECM ligands by specific integrins triggers distinct intracellular signals, enabling cells to regulate their behavior in response to microenvironmental changes.

Among the 24 different integrin heterodimers (18 α subunits and 8 β subunits), collagen binding integrins include four different α subunits bound to a common $\beta 1$ subunit: $\alpha 1\beta 1$, $\alpha 2\beta 1$, $\alpha 10\beta 1$, and $\alpha 11\beta 1$. Other classes of integrins include integrins presenting a Arg-Gly-Asp (RGD) motif, laminin binding integrins and leucocyte receptors. The members of collagen binding integrin subgroup recognize their fibrillar ligands either directly by using an inserted domain in their α subunit (αI domain) (230) or indirectly by a class of non-collagen bridging molecules—COLINBRI” (COLlagen INtegrin BRIDging) (231) (Figure 6).

Specific collagenous motifs are recognized by integrin αI domains, the major functional motif being the GFOGER (O = 4-hydroxyproline) sequence in the triple-helical conformation. The binding of this motif occurs in a metal-dependent manner via a Metal Ion-Dependent Adhesion Site (MIDAS) which coordinates a divalent cation Mg^{2+} and is highly conserved in all four collagen binding integrins (232). Other motifs with a GxOGER sequence (x = hydrophobic residue) include GROGER, GLOGER, GMOGER, and occur at specific loci within the D-periods of fibrillar collagens. A description of all known integrin recognition motifs for fibrillar collagens has been previously addressed by Hamaia and Farnale (233). Previous studies reported that $\alpha 1$ and $\alpha 10$ integrins poorly bind to the fibrillar form of collagens, when compared to their $\alpha 2$ and $\alpha 11$ counterparts, and rather favor the binding to the native monomeric form of other types of collagens such as network-forming (collagen IV and VI) or FACIT collagens (collagen IX) (231). It has been demonstrated that integrin $\alpha 1$ prefers to bind to the monomeric form of collagen I rather than to its mature

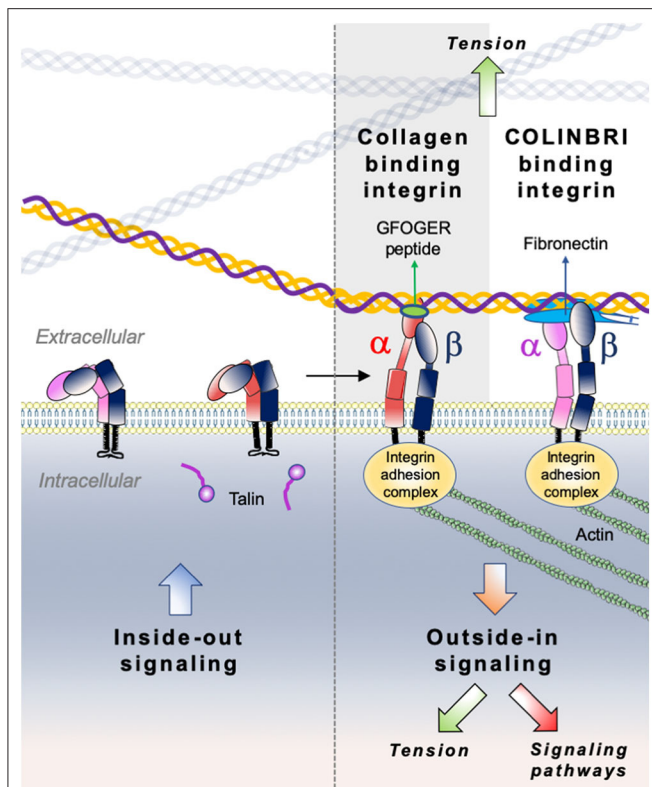


FIGURE 6 | Illustration of direct (collagen-binding integrin-mediated) and indirect (COLINBRI-mediated) integrin heterodimers binding to collagen fibrils. Inactive integrins adopt a compact conformation in which the α - (red/purple) and β -subunit (black) are closely associated. Intracellular signals, culminating in the binding of talin to the β -subunit tail, lead to conformational changes that result in increased affinity for extracellular ligands. The primed integrin binds talin, which represents the end-point of inside-out signaling. The binding of talin and ligand initiate focal contact formation. As the cytoskeleton matures, tension (green arrows) is generated on the integrin receptor across the cell membrane. The force applied to the integrin strengthens receptor-ligand binding and allows the formation of stable focal adhesions and the initiation of intracellular signaling cascades (red arrow), the end-point of outside-in signaling. In the direct cell-binding mechanism, collagen-binding integrins directly interact with the GFOGER sequence of fibrillar collagen to provide cell adhesion. In the indirect way, cell binding involves COLINBRIs like fibronectin represented here in blue. The COLINBRI molecule is anchored to collagen and provides cell attachment by interaction with the COLINBRI-binding integrins.

fibrillar form (234) and also binds with a higher affinity to the collagen IV (a basement membrane collagen) (235). Likewise, integrin $\alpha 10$ displays a higher affinity for network-forming collagen IV and VI when compared to the fibrillar collagens I-III (236). Higher fibrillar collagen specificity was however demonstrated for the $\alpha 2$ and $\alpha 11$ integrin I domains. These integrins prefer the binding of mature fibrillar collagen forms mostly through the GFOGER motif. Along with the binding of collagenous ligands, these integrins show also specificity for other ECM components such as laminins, proteoglycans, and tenascins. A detailed ligand specificity of collagen binding integrins is described in **Table 3**.

Most of the functional motifs in the collagen triple helix contain hydroxylated proline residues. Along with its critical

role in the stabilization of the collagen triple helix and fibril formation, hydroxylation of proline in position Y of the collagen triplet sequence -G-X-Y- has also an important role in the recognition of functional sites by the collagen receptors. It has been reported that $\alpha 2$ integrin binding to the GFPGER motif (P = 4-proline) is weaker than its binding to the consensus GFOGER motif, suggesting a higher avidity of integrin receptors for hydroxyproline-containing motifs of collagens (244). Furthermore, a recent study reported that absence of hydroxyproline residues in the GFOGER motif strongly impairs the avidity of $\alpha 1$ and $\alpha 11$ integrins for the collagen molecule and with a less prominent difference for the $\alpha 2$ integrin (245). This study revealed that absence of proline hydroxylation in collagen can affect integrin binding not only by the structural destabilization of the triple helix, but also by a direct mechanism, in which the residue Arg-218 in the $\alpha 11$ integrin domain directly interacts with the hydroxyproline residue in the integrin-binding motif of collagen.

Indirect binding of fibrillar collagens may involve collagen-binding integrins but also RGD integrins or leucocyte receptors. COLINBRIs have been described as prototypical ECM molecules being able to bridge between integrins and fibrillar collagens. Such molecules include among others fibronectin, vitronectin, periostin, and small leucine-rich proteoglycan/protein (SLRP) family members (231). These proteins show binding sites for specific fibrillar collagens, as well as several integrin-recognizing sites. They also play a role in collagen fibrillogenesis, deposition, and modulation of integrin affinity for collagenous ligands. An example of such a COLINBRI molecule is fibronectin, which displays a binding domain for the collagen I $\alpha 1$ chain in its N-terminal part and several binding sites for RGD-binding integrins ($\alpha 5\beta 1$ or $\alpha v\beta 3$ integrins) and leucocyte receptors ($\alpha 4\beta 1$ or $\alpha 4\beta 7$ integrins). Previous studies have reported the reciprocal mechanoregulation between the two ECM molecules and the active role of fibronectin in the collagen initial fibril-formation and assembly (246). In this context, collagen-binding integrins have also been reported to interact with COLINBRI-like molecules, particularly with SLRPs. Decorin interacts with $\alpha 2\beta 1$ integrin and allosterically modulates its collagen I-binding activity in angiogenic endothelial cells, but not quiescent cells, thus promoting a migratory phenotype (247). Conversely, lumican, another SLRP involved in collagen type I fibrillogenesis, displays an inhibitory effect on melanoma cell migration modulated by the interaction of its proteic core with the activated I domain of the $\alpha 2$ integrin subunit (248). For a detailed comparison of integrin and non-integrin collagen receptor binding sites, we refer the reader to Zeltz et al. (231) and Zhu et al. (249).

MECHANISTIC INSIGHTS INTO COLLAGEN-BINDING INTEGRIN SIGNALING

Collagen-binding integrins, similarly to the other type of integrins operate as bidirectional signaling receptors upon a biochemical or mechanical activation. The two directions

TABLE 3 | Specificity of collagen-binding integrins.

Integrin subunit	Collagen specificity	Motifs	Other ECM ligands	References
$\alpha 1$	Collagen I, II, III, IV, V, VI, IX, XVI and XVIII Highest affinity for collagen IV	GFOGER (collagen I, II, IV) GVOGEA (collagen II) GLOGEN (collagen III)	Laminins, galectins, arresten and semaphorin 7A	(231–233, 237, 238)
$\alpha 2$	Collagen I, II, III, IV, V and XI	GFOGER (collagen I, II, IV) GMOGER	Laminins, Tenascin C, decorin, endorepellin, perlecan, and chondroadherin	(231–233, 239, 240)
$\alpha 10$	Collagen I, II, IV, VI Highest affinity for collagen IV and VI	GFOGER (Collagen I, II, IV)	ND	(233, 236, 241)
$\alpha 11$	Collagen I, IV, V, IX Highest for collagen I	GFOGER (Collagen I, II, IV)	ND	(231, 232, 242, 243)

of integrin signaling have different biological consequences and involve distinct conformational states. In the traditional “outside-in” signaling, integrins act as membrane receptors in transmitting information from the surrounding environment into cells (Figure 6). This signaling refers to multivalent integrin-ligand binding, integrin conformational switch from an inactive low avidity state to a high avidity state, subsequent clustering and association with actin cytoskeleton (250). During “inside-out” signaling, talin, an intracellular activator, binds to the cytoplasmic tail of integrin β -subunit, inducing conformational changes that increase the affinity for extracellular ligands (251). Although conceptually different, the “outside-in” and “inside-out” integrin signaling processes are often complementary and closely linked both leading to regulation of cell polarity and cytoskeleton assembly, of gene expression, cell survival and motility. It is worth noting, that most of the previously described mechanistic studies on integrin signaling were performed on RGD binding integrins, as well as on $\alpha 1\beta 1$ and $\alpha 2\beta 1$ collagen-binding integrins, while integrin $\alpha 10\beta 1$ and $\alpha 11\beta 1$ signaling is still poorly explored. Furthermore, the specificity of integrin signaling relies on multiple factors, including heterodimer subtype, cellular context, ECM organization and ligand recognition. Analogously to other integrin subtypes, the signaling responses of collagen binding integrins are mostly mediated by the common $\beta 1$ subunit and are not specific to the $\alpha\beta$ heterodimer. Only a few studies reported the implication of the α cytoplasmic domain in the cellular signaling of collagen binding integrins. We therefore described below a general integrin signaling pathway applied to most integrins and highlight relevant studies which describe distinct signaling events for collagen binding integrins.

Integrin signaling requires the assembly of a dynamic multiprotein machinery around their cytoplasmic tails, called “adhesome.” It was reported that integrin adhesome comprises a network of at least 156 components linked via 690 different interactions (252). These components are classified into several functional groups, which comprise 25 adaptor proteins, 24 cytoskeletal molecules, nine actin-binding proteins, 18 tyrosine and serine/threonine kinases, 12 tyrosine and serine/threonine protein phosphatases, seven transmembrane receptors, six adhesion proteins, eight GTPase-activating proteins (GAPs), eight guanine nucleotide exchange factors (GEFs), five GTPases, and 32 other components (252). Integrin connection to

the cytoskeletal machinery is mediated by integrin adhesion complexes (IACs)—macromolecular complexes which transduce biochemical and mechanical signals from the microenvironment into biological responses (253). The formation of several major types of IACs has been reported, including focal complexes, focal adhesions and fibrillar adhesions, which play a central role in cell adhesion and migration (253). Most of the IAC proteomic analyses were performed on complexes isolated from fibronectin-attached cells, while still little is known about the adhesome complexes related to collagen binding integrins. These studies identified around 60 core proteins within IACs which constitute the cell adhesion machinery and link integrins bound to their ECM ligands to the cytoskeleton (254). It is plausible to assume that IACs associated to collagen binding integrins share a great similarity with those derived from fibronectin-attached integrins, as their have a common $\beta 1$ subunit.

Integrin clustering refers to the interaction of integrin $\alpha\beta$ heterodimers to form hetero-oligomers at cell surface. This event is relevant for “outside-in” signaling of integrins, including mechano-transduction processes and integrin recycling (251, 255). Upon clustering, integrins associate with a cytoskeletal signaling complex that promotes the assembly of actin filaments into large stress fibers (251). The resulting intracellular structures formed by integrins and cytoskeletal proteins are known as “focal adhesions” and “focal complexes.” Early steps of integrin signaling involve interactions with tyrosine kinases such as focal adhesion kinase (FAK), Src-kinases, abelson murine leukemia viral oncogene homolog 1 (Abl) and integrin-linked kinase (ILK), scaffold molecules such as p130CRK-associated substrate (p130CAS) and cytoskeletal proteins such as talin and kindlin. In the canonical integrin pathway, the active FAK/Src complex interacts with p130CAS and paxillin that in turn recruit Crk adaptor molecule leading to activation of several downstream molecules, including ras-related C3 botulinum toxin substrate 1 (Rac1), serine/threonine-protein kinase PAK 1, and c-Jun N-terminal kinase (JNK). The FAK/Src complex can also recruit other adaptor molecules leading to activation of major signaling pathways such as PI3K, RhoGTP-ases, p38, Erk, and phospholipase gamma (PLCG) pathways (256). Several studies have reported the implication of collagen binding integrins in multiple pathways mentioned above and mainly for $\alpha 1\beta 1$ and $\alpha 2\beta 1$ integrins. For example, binding of collagen type I by

integrin $\alpha 1\beta 1$ has been shown to sustain mesangial cell spreading via the activation of Erk1/2 pathway (257). Collagen binding of integrin $\alpha 2\beta 1$ through the GFOGER motif has been shown to mediate an “outside-in” signaling and to promote platelet spreading via activation of Src, FAK, and PLCG2 pathways (258). Similarly, integrin $\alpha 2\beta 1$ -mediated adhesion of platelets to type I collagen or GFOGER peptide has been shown to induce a downstream Pyk2 activation and PI3K β and Akt phosphorylation (259). Platelets adhesion to the monomeric collagen type I via $\alpha 2\beta 1$ integrin was also shown to trigger the activation of PLC γ 2 downstream signaling via Src kinases and Rac GTPases (260). Interestingly, both $\alpha 1$ and $\alpha 2$ collagen binding integrins have been shown to be involved in the production of reactive oxygen species (ROS), either by negatively regulating Rac1 activation and collagen synthesis through a crosstalk with the epidermal growth factor receptor (EGFR) (261), or by p38 MAPK phosphorylation and regulation of cell cycle (262). Furthermore, $\alpha 1\beta 1$ integrin has been also shown to be required for an efficient collagen remodeling in wound healing via the non-canonical TGF- β 1-dependent JNK signaling (263).

“Inside-out” signaling of integrins involves interactions with intracellular molecules at the cytoplasmic tails, which regulate integrin conformation. These interactions mainly occur at the cytoplasmic domain of the β integrin subunit. Talins and kindlins play a key role in this process. The binding of adaptor protein talin to the cytoplasmic tail of integrin β -subunit is a critical event in affinity activation of integrins (264). Talin association triggers integrin activation through the disruption of inhibitory interactions between α - and β -subunit cytoplasmic tails with subsequent transition from the bent to the extended conformation and increase in integrin affinity for extracellular ligands (255). Another major regulator of inside-out signaling, kindlin binds to the cytoplasmic tail of integrin β -subunit, supports integrin activation and recruits paxillin to the nascent focal adhesions, which in turn promotes the formation of membrane protrusions and therefore cell migration (265). In this context, talin was shown to be required for $\alpha 2\beta 1$ integrin-mediated platelet adhesion to collagen type I and to promote platelet aggregation (266). Interestingly, another study reported that the interaction between integrin $\alpha 2\beta 1$ in a non-activated conformation and collagen type I results in the activation of FAK in a talin-independent manner and requires the protein kinase C (267). While most of these interactions involve the $\beta 1$ cytoplasmic tail, some studies also reported the role of $\alpha 1$ and $\alpha 2$ subunits in the integrin downstream signaling. Notably, SHARPIN, an inactivator of talin and kindlin recruitment to β subunits, has been found to bind $\alpha 1$ and $\alpha 2$ tails, rather than $\beta 1$ cytoplasmic domain (268). The T-cell protein tyrosine phosphatase (TCPTP) has been also found to bind the cytoplasmic tail of $\alpha 1$ integrin upon cell adhesion to collagen and to negatively regulate EGFR signaling (269). Similarly, it has also been reported that $\alpha 2$ cytoplasmic tail is involved in the downstream activation of p38 signaling and upregulation of collagen gene transcription (270). The distinct signaling events induced by the different collagen binding integrin α subunits is remarkably highlighted by the opposite effect that these subunits have on collagen synthesis. Indeed, while $\alpha 1\beta 1$ integrin is a

known repressor of collagen type I synthesis (271), $\alpha 2\beta 1$ integrin promotes its expression (270). Additionally, several studies have reported a link between the ligand-induced activation of collagen binding integrins and the production of collagen remodeling proteinases such as MMPs (272–274).

Collagen-binding integrins have been also reported to crosstalk with receptor tyrosine kinases (RTKs) and mediate many cellular functions, including signaling, ligand recognition, and RTK endocytosis and trafficking. Multiple studies have reported the spatial coexistence between integrins and RTKs at cell surface with a synergistic signaling between the two receptors mainly related to the “outside-in” signaling of integrins (275). The interaction between the two partners was reported to be ligand-dependent or independent. A detailed description of these interactions was addressed by previous reviews (275–277). In the context of collagen binding integrins, a crosstalk between EGFR and $\alpha 1\beta 1$ or $\alpha 2\beta 1$ integrins has been reported to regulate the downstream signaling and trafficking of this tyrosine kinase receptor (269, 278). The cooperation between HGFR and $\alpha 2\beta 1$ integrin has been shown to regulate the innate immune response upon mast cell adhesion to collagen type I (279). Furthermore, we have recently shown that integrin $\alpha 11$ associates to PDGFR β in cancer associated fibroblasts and activates its JNK downstream signaling (280).

DDRs can also positively and negatively regulate collagen-binding integrin-mediated signal transduction and cell adhesion: DDR-mediated signaling can directly affect the activity of integrins but it can also converge with integrin-triggered pathways to regulate cellular functions, whereby each receptor engages its own downstream pathway (281).

COLLAGEN-BINDING INTEGRINS IN CANCER

Integrin expression patterns undergo major changes during tumor progression and metastasis leading to significant alterations in the phenotype of both cancer and stromal cells. In cancer cells, integrins have a key role in sustaining cell proliferation and invasion, evading tumor suppressors and cell death and promoting the EMT (282). Stromal integrins, on the other hand, are implicated in processes such as tumor angiogenesis, desmoplastic reaction and metastasis by modulating cell adhesion and invasion, and ECM remodeling. In view of their bidirectional signaling, integrins provide spatially restricted communication between cells and the surrounding microenvironment and act as key mechanosensing elements and RTK co-partners to modulate biological processes essential for tumor cell survival. Normal cells rely on integrin-mediated cell adhesion to ECM components to proliferate and survive. Defects in cell ability to attach to ECM are associated with impaired pro-survival integrin-dependent signaling pathways including PI3K/Akt, MAPK, FAK, NF- κ B, leading to anoikis (283). Integrins modulate all steps of the metastatic process, from invasion from the primary tumor and intravasation into the bloodstream, to colonization of the secondary sites. Distinct integrin-ligand binding combinations act as drivers

of organ-specificity colonization of tumor cells and the subsequent cell survival and adaptation to the newly acquired microenvironment. The desmoplastic reaction has been linked to high expression levels of integrins in metastatic cancers (284). Being key modulators of ECM, tumor and stromal integrins regulate matrix composition in order to control cell adhesion and invasion. Along with the aforementioned processes, integrins also contribute to tumor regulation through angiogenesis, metabolic reprogramming, evasion of immune destruction and acquisition of drug resistance.

Several mechanisms have been found to be involved in collagen-binding integrin regulation of tumor growth and metastasis. In the following section, we will discuss each of the four collagen-binding integrins, their expression profile and implication in pathological conditions with a special focus on cancer disease.

$\alpha 1\beta 1$ integrin is widely expressed in normal tissues, particularly in the mesenchyme, vascular and immune system with limited expression in the epithelium (285). As illustrated in **Table 3**, this integrin can bind many types of collagens, including collagen I, II, III, IV, V, VI, IX, XVI, and XVIII, as well as laminins, galectins, arrestin, and semaphorin 7A (232). As previously mentioned, $\alpha 1\beta 1$ integrin is mostly detected in connective tissues and it exhibits a higher affinity for collagen IV when compared to collagen I (235). Integrin $\alpha 1\beta 1$ functions include cell adhesion and survival, collagen synthesis and MMP secretion. This receptor has been associated with pathological conditions such as osteoporosis, renal injury, rheumatoid arthritis, psoriasis, as well as cancer (285). Integrin $\alpha 1$ mutant mice are viable and fertile with no marked phenotype (286). Integrin $\alpha 1$ adult mice exhibit however a mild decrease in body weight, hypocellular dermis (287) and aged animals display different phenotypes, including osteoarthritis (288), and retinal degeneration (289). *In vitro*, integrin $\alpha 1$ -deficient cells display adhesion alteration when cultured on collagen I and IV, as well as failure to recruit and activate Shc adaptor molecule with consequent reduction in cell proliferation and survival (287). When challenged to different pathological conditions, $\alpha 1$ -mutant mice present reduced psoriasis (290), accelerated aging-dependent osteoarthritis (288), diminished bone fracture healing (291), severe hepatic insulin resistance and lower hepatic fat accumulation upon a high fat diet (292), and increased glomerulosclerosis in diabetic mice (293) (**Table 4**).

In the context of cancer, integrin $\alpha 1\beta 1$ is still poorly explored. Only a few studies reported its role in tumor progression and metastasis, mostly in non-small cell lung carcinoma and colorectal cancer. In an orthotopic model of non-small cell lung carcinoma in $\alpha 1$ -null mice with increased MMP9 levels, a decreased number, size and vascularization of primary tumors and metastases was observed, highlighting the proangiogenic features of this integrin (294). Similarly, in another study of a spontaneous non-small cell lung carcinoma mouse model in *KrasLA2*/ $\alpha 1$ -null mice, tumors appeared smaller, less angiogenic, with reduced proliferation and increased apoptosis (295). Overexpression of $\alpha 1\beta 1$ integrin has been reported in 65% of colorectal cancers and correlates for more than 70% with *Myc* oncogene expression (336, 337). Furthermore, in a xenograft

model of colorectal cancers, $\alpha 1$ -deficient tumors displayed extensive necrosis, low mitotic index and reduced angiogenesis (296). Reduced angiogenesis in absence of $\alpha 1\beta 1$ integrin was also reported in an experimental breast cancer model (297). Some melanoma studies also reported a link between high expression of $\alpha 1\beta 1$ integrin and a poor patient outcome (298, 299). Oral squamous cell carcinomas, broncho-alveolar and gastric carcinomas were also reported to display a gain of $\alpha 1\beta 1$ integrin expression (300–302).

$\alpha 2\beta 1$ integrin is mostly expressed by normal epithelial cells, fibroblasts and platelets/megakaryocytes, and depending on the differentiation state also in T lymphocytes and endothelial cells (239). It preferentially binds to fibrillar collagen types I, II, III, V, and XI. It also binds to non-collagenous ligands such as laminins, tenascin C, decorin, endorepellin, and chondroadherin (232). One of the main functions of integrin $\alpha 2\beta 1$ is the adhesion-mediated survival particularly in platelets, where this integrin constitutes the most abundant collagen receptor. It is also a major regulator of cell motility, mainly via p38 MAPK pathway activation (239). Genetic alteration of integrin $\alpha 2\beta 1$ expression has been detected in pathological conditions such as hemostasis, thrombosis, fibrosis, and immune response. The mutant mice for integrin $\alpha 2$ integrin subunit present a mild phenotype in the mammary gland branching morphogenesis defect (305, 306) adhesion defects for platelets, fibroblasts and keratinocytes when tested for collagen I (307), and a reduced age-related bone deterioration (308). Pathologically challenged mice display, reduced wound healing with increased neovascularization (309), defects in age related bone degradation due to the overexpression of collagen I (308), decreased innate immune response to *Listeria* infection and reduced thrombi formation (239), decreased glomerulosclerosis and collagen deposition after renal injury (310), and reduced joint inflammation and cartilage destruction in rheumatoid arthritis (311).

In cancer, $\alpha 2\beta 1$ integrin was extensively studied in the context of angiogenesis. In the MMTV-neu spontaneous mouse model of breast cancer, $\alpha 2\beta 1$ integrin acts as a metastasis suppressor by inhibiting tumor cell intravasation without altering tumor development or growth (312). The same study provided evidence for a correlation between $\alpha 2\beta 1$ integrin expression and estrogen positivity in breast cancer patients and that the decreased $\alpha 2\beta 1$ integrin levels predict metastasis and decreased survival in prostate and breast cancer patients. Conversely, in another recent study of a xenograft model of MCF-7 cells interacting with platelets, $\alpha 2$ downregulation prevented breast cancer metastasis (313). In an experimental model of breast cancer metastasis, the MMP13 expression observed in MDA-MB-231 metastasizing cells to the bones was induced by collagen type I through integrin $\alpha 1\beta 1$ and $\alpha 2\beta 1$ -mediated p38 MAPK pathway (314). The implication of $\alpha 2\beta 1$ integrin, as well as $\alpha 1\beta 1$ integrin was further emphasized by a melanoma xenograft model in a double knockout model, in which absence of both integrins impaired tumor angiogenesis (66). In another melanoma study, $\alpha 2\beta 1$ integrin co-localized with E-cadherin and N-cadherin at tumor cell-cell contacts in both primary and metastatic samples (315). Disturbance of these adhesive networks impaired tumor growth in a xenograft model and N-cadherin downregulation prevented

TABLE 4 | Phenotypical consequences of collagen-binding integrin knockouts in mice.

Integrin subunit	Expression	Knockout phenotype	Pathologically challenged knockout phenotype	Cancer phenotype
$\alpha 1$	Endothelial cells, fibroblasts, pericytes, mesangial cells, bone marrow mesenchymal stem cells, hepatic stellate cells, chondrocytes, neural cells, Schwann cells, immune cells (285, 286)	No pronounced phenotype (286) moderate decrease in body weight, hypocellular dermis (287) aged-related osteoarthritis (288), retinal degeneration (289)	Reduced psoriasis (290) and inflammation (129), accelerated aging-dependent osteoarthritis (288), reduced bone fracture healing (291), increased glomerulosclerosis in diabetic mice (293), hepatic insulin resistance and decreased hepatic fat accumulation on a high fat diet (292)	Reduced tumor number and size, reduced angiogenesis and proliferation, increased apoptosis in models of non-small cell lung carcinoma $\alpha 1$ -null mice (294, 295); overexpression in colorectal cancers and correlation with Myc oncogene; reduced angiogenesis, increased necrosis and low mitotic index in a xenograft model of colorectal cancers (296); reduced angiogenesis in an experimental breast cancer model (297); correlation with a poor patient outcome in melanoma (298, 299); gain of expression in oral squamous cell carcinomas, broncho-alveolar and gastric carcinomas (300–302)
$\alpha 2$	Epithelial cells, platelets/megakaryocytes, endothelial cells fibroblasts (303, 304)	Moderate phenotype in mammary gland morphogenesis (305, 306), adhesion defects for isolated platelets on collagen I (307), reduced age-related osteoporosis (308)	Increased neovascularization in wound healing (309), defects in age-related bone loss (308), decreased immune response to peritoneal <i>Listeria</i> infection and reduced thrombi formation (239), decreased glomerulosclerosis upon renal injury (310), reduced inflammation and cartilage deterioration in rheumatoid arthritis (311), reduced inflammatory bowel disease (65)	Metastasis suppressor in the MMTV-neu mouse breast cancer model and correlation with patient outcome in breast and prostate cancers (312); impaired breast cancer metastasis in a xenograft model of MCF-7 cells interacting with platelets (313); reduced MMP13 expression by metastasizing cells to the bones in an experimental model of MDA-MB-231 breast cancer cells (314); impaired tumor angiogenesis in a melanoma xenograft model in a double knockout model (66); co-localization with E-cadherin and N-cadherin at tumor cell-cell contacts in primary and metastatic samples of melanoma (315); selective metastasis to the bones in prostate cancer (316); implication in doxorubicin-induced drug resistance in leukemia (317, 318); implication in other types of cancers, including pancreatic, colorectal, gastric and lung cancers, oral squamous cell carcinoma and T-cell acute lymphoblastic leukemia (319)
$\alpha 10$	Chondrocytes, mesenchymal stem cells, junctional fibroblasts (241, 320)	Mild chondrodysplasia (321), canine chondrodysplasia and limb dwarfism (322)	ND	Upregulation in malignant melanoma (323); overexpression in glioblastoma (324); high expression correlated with disease-specific death and distant metastasis in high-grade myxofibrosarcoma (325)
$\alpha 11$	Fibroblasts (cancer associated fibroblasts, myofibroblasts), mesenchymal stem cells (326–328)	Dwarfism, altered incisor tooth eruption, increased mortality and decreased IGF-1 serum levels (327, 329, 330)	Reduced wound strength and deposition of granulation tissue in wound healing (263), reduced cardiac fibrosis and left ventricular hypertrophy (331)	Decreased tumorigenicity of cancer cells (328) and matrix stiffness (332) in a challenged xenograft model of lung cancer; correlation with tumor progression and postoperative recurrence in non-small cell lung cancer (333); association with a pro-tumorigenic PDGFR β -positive subset of cancer associated fibroblasts and correlation with a poor clinical outcome in breast cancer (280); correlation with aggressive phenotypes of breast cancer (334); decreased intratumoral interstitial fluid pressure and collagen structure in a challenged mouse tumor model (335)

$\alpha 2\beta 1$ integrin-mediated tumor cell invasion on collagen type I matrix. In prostate cancer, $\alpha 2\beta 1$ integrin contributes to a selective metastasis to the bone, rather than to other sites in patient samples and downregulation of this integrin impairs the adhesion and migration of prostate cancer cells toward

collagen type I within the bone (316). Implication of $\alpha 2\beta 1$ integrin in tumor progression and metastasis was reported in several other types of cancers, including pancreatic, colorectal, gastric and lung cancers, oral squamous cell carcinoma and T-cell acute lymphoblastic leukemia, which were previously reviewed

in (319). Lastly, it has been reported that $\alpha 2\beta 1$ integrin and collagen I participate to doxorubicin-induced drug resistance in leukemia by either protecting leukemia cells from apoptosis through MAPK/ERK pathway activation (317), or by decreasing the DNA damage through the inhibition of Rac1 activation (318).

$\alpha 10\beta 1$ integrin expression is mostly restricted to the cartilaginous tissue and this integrin is considered as a phenotypic marker for chondrocyte differentiation. Integrin $\alpha 10\beta 1$ is a receptor for fibril-forming collagen type II, for network-forming collagen IV and beaded-filaments forming VI (321). It has a lower affinity for other types of fibrillar collagens. Integrin $\alpha 10\beta 1$ is implicated in chondrocyte differentiation potency and is associated to an increase of collagen type II synthesis (338). The major physiological role of $\alpha 10\beta 1$ integrin concerns the skeletal development, as this receptor is mainly expressed in the cartilage. Indeed, $\alpha 10\beta 1$ integrin ablation in mouse models results in mild chondrodysplasia with altered chondrocyte functionality (321). Integrin $\alpha 10$ -deficiency is associated to changes in chondrocyte morphology, increased apoptosis and reduced proliferation, resulting in a mild growth retardation (339). Furthermore, a previous study reported that canine chondrodysplasia is associated with a naturally occurring mutation of $\alpha 10\beta 1$ integrin gene, and the observed limb dwarfism resembled the phenotype detected in the $\alpha 1$ -null mouse model (322). The implication of this integrin in other pathological conditions is poorly documented.

In cancer, the role of $\alpha 10\beta 1$ integrin remains poorly explored. A previous study reported the upregulation of $\alpha 10\beta 1$ integrin in malignant melanoma when compared to primary melanocytes (323). In a recent study, this integrin emerged as a potential therapeutic target for treatment of glioblastoma (324). This study revealed that $\alpha 10\beta 1$ integrin is overexpressed in glioblastoma patient tissues and cells when compared to normal brain tissues, and high $\alpha 10$ expression was associated to increased proliferation and migration. Interestingly, $\alpha 10\beta 1$ integrin was correlated to disease-specific death and distant metastasis in a cohort of 64 primary high-grade myxofibrosarcomas (325). The authors demonstrated that $\alpha 10\beta 1$ integrin promotes tumor cell survival through activation of TRIO-RAC-RICTOR-mTOR signaling, and that this pathway constitutes a promising targeted therapeutic strategy for patients with high-risk myxofibrosarcoma.

$\alpha 11\beta 1$ integrin expression is primarily confined to mesenchymal-like cells during pathological conditions including fibrosis, wound healing and cancer. Integrin $\alpha 11$ is the latest integrin family member to be identified. $\alpha 11\beta 1$ integrin binds with high affinity collagen type I and contains the I domain, which recognizes the triple-helical GFOGER motif sequence (340). The highest $\alpha 11$ transcript levels were detected in embryos and human adult uterus, heart and periodontal ligament. A complete characterization of $\alpha 11$ expression in human adult tissues has not yet been performed. While the other collagen binding integrins $\alpha 1$ and $\alpha 2$ are expressed in a several cell types, $\alpha 11$ expression appears to be restricted to distinct cell subsets of mesenchymal origin. Generally, it is considered that $\alpha 11$ integrin is expressed *in vitro* by mesenchymal derived cells, and *in vivo* by fibroblasts at sites of highly organized collagen structures (230). Integrin $\alpha 11$ expression is promoted by TGF β

(341) and by the mechanical stiffness of the environment in a mechanism involving an autocrine loop of Activin (326). Integrin $\alpha 11$ is a major collagen receptor on fibroblastic cells and *in vitro* studies revealed that this integrin plays a key role in the adhesion and motility of these cells (242). Additionally, $\alpha 11$ integrin plays a critical role in collagen reorganization and remodeling. Several studies reported that *in vitro* integrin $\alpha 11$ -deficiency impacts contraction of collagen matrices (273, 329). Furthermore, integrin $\alpha 11$ was also reported to be also implicated in TGF β -induced myofibroblast differentiation (326). Interestingly, a recent study investigated the contribution of $\alpha 11$ integrin cytoplasmic tail to cell proliferation and invasion via a FAK/ERK-dependent activation (342).

The $\alpha 11$ -deficient mice are viable and fertile but display the following defects: dwarfism, altered teeth, increased mortality, and decreased IGF-1 serum levels (327, 329). Dwarfism of $\alpha 11$ -deficient mice does not seem to be related to structural defects in forming cartilage or bone, but it is rather due to the tooth alteration and IGF-1 serum levels in these mice. Indeed, a strong malnutrition of $\alpha 11$ -deficient mice was observed due to a late incisor eruption and altered tooth shape (327). However, further investigations revealed that $\alpha 11$ -deficient mice are already smaller at birth, before the incisor eruption effect could impact the body size, suggesting that other mechanisms could be involved in the acquired phenotype. Accordingly, IGF1 is a growth factor with a crucial role in growth control and bone mineralization. Indeed, low hepatic IGF1 production was detected in $\alpha 11$ -deficient mice, implying that dwarfism observed in these mice may also be induced by the severely diminished IGF1 serum levels (330).

Only a limited number of studies have investigated the function of integrin $\alpha 11$ in pathological conditions. In wound healing, it has been reported that upregulated integrin $\alpha 11$ promotes wound strength and deposition of granulation tissue through a TGF β -dependent JNK signaling (263). Overexpression of integrin $\alpha 11$ has been also linked to left ventricular hypertrophy and cardiac fibrosis, as a result of soluble factor secretion and increase collagen deposition in the heart (331). Recent studies have extensively investigated integrin $\alpha 11$ in the context of cancer associated fibroblasts. In lung cancer, stromal integrin $\alpha 11$ increases the tumorigenicity of cancer cells in xenograft models by modulating both IGF-2 production (328) and matrix stiffness (332). Likewise, another study reported integrin $\alpha 11$ association with tumor progression and postoperative recurrence in non-small cell lung cancer (333). We have recently demonstrated that integrin $\alpha 11$ identifies a PDGFR β -positive subset of cancer associated fibroblasts displaying pro-tumorigenic features in breast cancer. Our study revealed that high stromal integrin $\alpha 11$ /PDGFR β expression is correlated with a poorer clinical outcome in breast cancer patients and that this integrin harnesses the PDGFR β /JNK signaling pathway to promote the invasion of cancer cells (280). The implication of stromal $\alpha 11\beta 1$ integrin in breast cancer was further investigated in another recent study, which emphasized the correlation of this integrin expression with aggressive phenotypes of breast cancer (334). Furthermore, an additional report highlighted the contribution of integrin $\alpha 11$ to breast

cancer progression by regulating the intratumoral interstitial fluid pressure and collagen structure (335). Interestingly, a recent study analyzed the expression of $\alpha 11\beta 1$ integrin in a panel of 14 different types of cancers and identified that this integrin is expressed by subset of non-pericyte-derived cancer associated fibroblasts and constitutes an important receptor for collagen remodeling. Integrin $\alpha 11$ contribution to other neoplastic malignancies and metastatic dissemination has not been yet documented. Its restricted expression, confined to a distinct mesenchymal cell subset, makes integrin $\alpha 11$ a good candidate for targeting the stromal compartment in cancer.

CONCLUSIONS AND PERSPECTIVES

Even if the hallmarks of cancer are driven by oncogenic mutations, most of them are modulated by the biochemical and biomechanical properties of the ECM that surrounds the tumor. By using different cell surface receptors, including integrins, cancer, and stromal cells are able to both impact and be impacted by ECM components in general and, more specifically, fibrillar collagens. In agreement with this concept, collagen fibers should no longer be viewed as passive scaffolds but as complex and constantly evolving signaling hubs that modulate a multitude of cellular functions.

It is now well-accepted that cancer should be viewed as a disease affecting the entire tissue rather than single cells. Indeed, tumors evolve in complex, dynamic, and functionally diverse TMEs. The diversity of TMEs depends on several factors including among others the tissue wherein the tumor develops, the stage of tumor evolution, the age of the patient. In contrast to cancer cells, the different cell populations which compose the TME are genetically stable, making them attractive targets for the development of innovative therapies with a low risk of development of treatment resistance. Furthermore, the efficacy of several standard chemotherapies and targeted agents is modulated by the TME, supporting the relevance of combining cancer cell-targeting agents with TME-directed therapies. However, issues related to cancer cell heterogeneity hold true for the TME. In that respect, a high degree of variability has been observed in ECM deposition and stiffness in a single tumor (343). Heterogeneity of the ECM could unravel the lack of success of the clinical trials of therapeutics targeting this

feature of tumor development (343). We should also keep in mind that specific ECM components might have opposing roles during tumor progression in different cancers, suggesting that the impact of the ECM on the hallmarks of cancer cannot be broadly extrapolated to all cancer types.

It is worth noting that fibrillar collagens-derived proteolytic fragments are released in the blood stream where they represent easily accessible biomarkers to monitor cancer progression (140, 344–346).

While many questions related to how the biochemical and biomechanical properties of fibrillar collagens determine tumor progression remain unanswered, the potential for these ECM components to be effective markers and/or targets in treating cancer patients remains very promising.

AUTHOR CONTRIBUTIONS

IB, IP, TL, AN, and EM drafted the first version of the manuscript and IB, IP, and EM the figures and tables. All authors have critically reviewed and approved the manuscript.

FUNDING

This work was supported by: grants from the Fonds de la Recherche Scientifique - FNRS (F.R.S.-FNRS, Belgium), the Fondation contre le Cancer (foundation of public interest, Belgium), the Fonds spéciaux de la Recherche (University of Liège), the Fondation Hospitalo Universitaire Léon Fredericq (FHULF, University of Liège), the Direction Générale Opérationnelle de l'Economie, de l'Emploi et de la Recherche from the Service Public de Wallonie (SPW, Belgium), the Walloon Region through the FRFS-WELBIO strategic research programme, the Wallonia-Brussels Federation (grant for Concerted Research Actions). TL was the recipient of a FNRS-Télévie grant. EM was a Research Associate from the Fund for Scientific Research—FNRS (Belgium).

ACKNOWLEDGMENTS

We apologize to authors whose work could not be discussed here because of space limitations.

REFERENCES

- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013
- Balkwill F, Mantovani A. Inflammation and cancer: back to virchow? *Lancet*. (2001) 357:539–45. doi: 10.1016/S0140-6736(00)04046-0
- Paget S. The distribution of secondary growths in cancer of the breast. *Lancet*. (1889) 133:571–3. doi: 10.1016/S0140-6736(00)49915-0
- Egeblad M, Nakasone ES, Werb Z. Tumors as organs: complex tissues that interface with the entire organism. *Dev Cell*. (2010) 18:884–901. doi: 10.1016/j.devcel.2010.05.012
- Chen F, Zhuang X, Lin L, Yu P, Wang Y, Shi Y, et al. New horizons in tumor microenvironment biology: challenges and opportunities. *BMC Med*. (2015) 13:14–45. doi: 10.1186/s12916-015-0278-7
- Patel H, Nilendu P, Jahagirdar D, Pal JK, Sharma NK. Modulating secreted components of tumor microenvironment: a masterstroke in tumor therapeutics. *Cancer Biol Ther*. (2018) 19:3–12. doi: 10.1080/15384047.2017.1394538
- Denton AE, Roberts EW, Fearon DT. Stromal cells in the tumor microenvironment BT - stromal immunology. In: Owens B, Lakin M, editors. *Stromal Immunology*. Cham: Springer (2018). p. 99–114.
- Neesse A, Bauer CA, Öhlund D, Lauth M, Buchholz M, Michl P, et al. Stromal biology and therapy in pancreatic cancer: ready for clinical translation? *Gut*. (2018) 68:159–71. doi: 10.1136/gutjnl-2018-316451
- Polyak K, Kalluri R. The role of the microenvironment in mammary gland development and cancer. *Cold Spring Harb Perspect Biol*. (2010) 2:a003244. doi: 10.1101/cshperspect.a003244

10. Conklin MW, Keely PJ. Why the stroma matters in breast cancer. *Cell Adhes Migr.* (2012) 6:249–60. doi: 10.4161/cam.20567
11. Hynes RO. The extracellular matrix: not just pretty fibrils. *Science.* (2009) 326:1216–19. doi: 10.1126/science.1176009
12. Hynes RO, Naba A. Overview of the matrisome—an inventory of extracellular matrix constituents and functions. *Cold Spring Harb Perspect Biol.* (2012) 4:a004903. doi: 10.1101/cshperspect.a004903
13. Oskarsson T. Extracellular matrix components in breast cancer progression and metastasis. *Breast.* (2013) 22(Suppl. 2):S66–72. doi: 10.1016/j.breast.2013.07.012
14. Leitinger B, Hohenester E. Mammalian collagen receptors. *Matrix Biol.* (2007) 26:146–55. doi: 10.1016/j.matbio.2006.10.007
15. Xian X, Gopal S, Couchman JR. Syndecans as receptors and organizers of the extracellular matrix. *Cell Tissue Res.* (2010) 339:31–46. doi: 10.1007/s00441-009-0829-3
16. Walker C, Mojares E, del Río Hernández A. Role of extracellular matrix in development and cancer progression. *Int J Mol Sci.* (2018) 19:3028–31. doi: 10.3390/ijms19103028
17. Yue B. Biology of the extracellular matrix: an overview. *J Glaucoma.* (2014) 23:S20–3. doi: 10.1097/IJG.0000000000000108
18. Mouw JK, Ou G, Weaver VM. Extracellular matrix assembly: a multiscale deconstruction. *Nat Publ Gr.* (2014) 15:771–85. doi: 10.1038/nrm3902
19. Theocharis AD, Manou D, Karamanos NK. The extracellular matrix as a multitasking player in disease. *FEBS J.* (2019) 286:2830–69. doi: 10.1111/febs.14818
20. Frantz C, Stewart KM, Weaver VM. The extracellular matrix at a glance. *J Cell Sci.* (2010) 123:4195–200. doi: 10.1242/jcs.023820
21. Levental KR, Yu H, Kass L, Lakins JN, Egeblad M, Erler JT, et al. Matrix crosslinking forces tumor progression by enhancing integrin signaling. *Cell.* (2009) 139:891–906. doi: 10.1016/j.cell.2009.10.027
22. Venning FA, Wullkopf L, Erler JT. Targeting ECM disrupts cancer progression. *Front Oncol.* (2015) 5:224. doi: 10.3389/fonc.2015.00224
23. Gkretsi V, Stylianopoulos T. Cell adhesion and matrix stiffness: coordinating cancer cell invasion and metastasis. *Front Oncol.* (2018) 8:145. doi: 10.3389/fonc.2018.00145
24. Jinka R, Kapoor R, Sistla PG, Raj TA, Pande G. Alterations in cell-extracellular matrix interactions during progression of cancers. *Int J Cell Biol.* (2012) 2012:219196–8. doi: 10.1155/2012/219196
25. Paszek MJ, Zahir N, Johnson KR, Lakins JN, Rozenberg GI, Gefen A, et al. Tensional homeostasis and the malignant phenotype. *Cancer Cell.* (2005) 8:241–54. doi: 10.1016/j.ccr.2005.08.010
26. Butcher DT, Alliston T, Weaver VM. A tense situation: forcing tumour progression. *Nat Rev Cancer.* (2009) 9:108–22. doi: 10.1038/nrc2544
27. Sulzmaier FJ, Jean C, Schlaepfer DD. FAK in cancer: mechanistic findings and clinical applications. *Nat Rev Cancer.* (2014) 14:598–610. doi: 10.1038/nrc3792
28. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell.* (2000) 100:57–70. doi: 10.1016/S0092-8674(00)81683-9
29. Chang TT, Thakar D, Weaver VM. Force-dependent breaching of the basement membrane. *Matrix Biol.* (2017) 57–58:178–89. doi: 10.1016/j.matbio.2016.12.005
30. Pickup MW, Mouw JK, Weaver VM. The extracellular matrix modulates the hallmarks of cancer. *EMBO Rep.* (2014) 15:1243–53. doi: 10.15252/embr.201439246
31. Assent D, Bourgot I, Hennuy B, Geurts P, Noël A, Foidart J-M, et al. A membrane-type-1 matrix metalloproteinase (MT1-MMP) - discoidin domain receptor 1 axis regulates collagen-induced apoptosis in breast cancer cells. *PLoS ONE.* (2015) 10:e0116006. doi: 10.1371/journal.pone.0116006
32. Maquoi E, Assent D, Detilleux J, Pequeur C, Foidart J-M, Noël A. MT1-MMP protects breast carcinoma cells against type I collagen-induced apoptosis. *Oncogene.* (2012) 31:480–93. doi: 10.1038/onc.2011.249
33. Saby C, Collin G, Sinane M, Buache E, Van Gulick L, Saltel F, et al. DDR1 and MT1-MMP expression levels are determinant for triggering BIK-mediated apoptosis by 3D type I collagen matrix in invasive basal-like breast carcinoma cells. *Front Pharmacol.* (2019) 10:462. doi: 10.3389/fphar.2019.00462
34. Exposito J-Y, Valcourt U, Cluzel C, Lethias C. The fibrillar collagen family. *Int J Mol Sci.* (2010) 11:407–26. doi: 10.3390/ijms11020407
35. Exposito J-Y, Cluzel C, Garrone R, Lethias C. Evolution of collagens. *Anat Rec.* (2002) 268:302–16. doi: 10.1002/ar.10162
36. Myllyharju J, Kivirikko KI. Collagens, modifying enzymes and their mutations in humans, flies and worms. *Trends Genet.* (2004) 20:33–43. doi: 10.1016/j.tig.2003.11.004
37. Ricard-Blum S, Ruggiero F. The collagen superfamily: from the extracellular matrix to the cell membrane. *Pathol Biol.* (2005) 53:430–42. doi: 10.1016/j.patbio.2004.12.024
38. Bateman JF, Boot-Handford RP, Lamandé SR. Genetic diseases of connective tissues: cellular and extracellular effects of ECM mutations. *Nat Rev Genet.* (2009) 10:173–83. doi: 10.1038/nrg2520
39. Mayne R, Burgeson RE. *Structure and function of collagen types.* Orlando: Academic Press (1987).
40. Van der Rest M, Garrone R. Collagen family of proteins. *FASEB J.* (1991) 5:2814–23. doi: 10.1096/fasebj.5.13.1916105
41. Ricard-Blum S. The collagen family. *Cold Spring Harb Perspect Biol.* (2011) 3:a004978. doi: 10.1101/cshperspect.a004978
42. Brodsky B, Persikov AV. Molecular structure of the collagen triple helix. *Adv Protein Chem.* (2005) 70:301–39. doi: 10.1016/S0065-3233(05)70009-7
43. Hulmes DJS. Collagen diversity, synthesis and assembly BT – collagen. In: Fratzl P, editor. *Collagen.* Boston, MA: Springer (2008). p. 15–47.
44. Lees JF, Tasab M, Bulleid NJ. Identification of the molecular recognition sequence which determines the type-specific assembly of procollagen. *EMBO J.* (1997) 65:908–16. doi: 10.1093/emboj/16.5.908
45. Hofmann H, Fietzek PP, Kuhn K. The role of polar and hydrophobic interactions for the molecular packing of type I collagen: a three-dimensional evaluation of the amino acid sequence. *J Mol Biol.* (1978) 125:137–65. doi: 10.1016/0022-2836(78)90342-X
46. Hoffman GG, Branam AM, Huang G, Pelegri F, Cole WG, Wenstrup RM, et al. Characterization of the six zebrafish clade B fibrillar procollagen genes, with evidence for evolutionarily conserved alternative splicing within the pro- α 1(V) C-propeptide. *Matrix Biol.* (2010) 29:261–75. doi: 10.1016/j.matbio.2010.01.006
47. Fang M, Jacob R, McDougal O, Oxford JT. Minor fibrillar collagens, variable regions alternative splicing, intrinsic disorder, and tyrosine sulfation. *Protein Cell.* (2012) 3:419–33. doi: 10.1007/s13238-012-2917-5
48. Eyre DR, Weis MA, Wu J-J. Articular cartilage collagen: an irreplaceable framework? *Eur Cells Mater.* (2006) 12:57–63. doi: 10.22203/eCM.v012a07
49. Boot-Handford RP, Tuckwell DS, Plumb DA, Rock CF, Poulsom R. A novel and highly conserved collagen (pro(α)1(XXVII)) with a unique expression pattern and unusual molecular characteristics establishes a new clade within the vertebrate fibrillar collagen family. *J Biol Chem.* (2003) 278:31067–77. doi: 10.1074/jbc.M212889200
50. Koch M, Schulze J, Hansen U, Ashwodt T, Keene DR, Brunken WJ, et al. A novel marker of tissue junctions, collagen XXII. *J Biol Chem.* (2004) 279:22514–21. doi: 10.1074/jbc.M400536200
51. von der Mark H, Aumailley M, Wick G, Fleischmajer R, Timpl R. Immunochemistry, genuine size tissue localization of collagen VI. *Eur J Biochem.* (1984) 142:493–502. doi: 10.1111/j.1432-1033.1984.tb08313.x
52. Shaw LM, Olsen BR. FACIT collagens: diverse molecular bridges in extracellular matrices. *Trends Biochem Sci.* (1991) 16:191–4. doi: 10.1016/0968-0004(91)90074-6
53. Kadler KE, Baldock C, Bella J, Boot-Handford RP. Collagens at a glance. *J Cell Sci.* (2007) 120:1955–8. doi: 10.1242/jcs.03453
54. Gelse K. Collagens—structure, function, and biosynthesis. *Adv Drug Deliv Rev.* (2003) 55:1531–46. doi: 10.1016/j.addr.2003.08.002
55. Fleischmajer R, Perlsh JS, Burgeson RE, Bahai FS, Timpl R. Type I and Type III collagen interactions during fibrillogenesis. *Ann N Y Acad Sci.* (1990) 580:161–75. doi: 10.1111/j.1749-6632.1990.tb17927.x
56. Niyibizi C, Eyre DR. Bone type V collagen: chain composition and location of a trypsin cleavage site. *Connect Tissue Res.* (2009) 20:247–50. doi: 10.3109/03008208909023894
57. Mayne R. Cartilage collagens. what is their function, and are they involved in articular disease? *Arthritis Rheum.* (1989) 32:241–6. doi: 10.1002/anr.1780320302
58. von der Mark K, Schöber S, Goodman SL. Integrins in cell migration BT – integrin Protocols. In: Howlett A, editor. *Integrin Protocols.* NJ: Humana Press (1999). p. 219–230.

59. Kivirikko KI, Kishida Y, Sakakibara S, Prockop DJ. Hydroxylation of (X-Pro-Gly)_n by procollagen proline hydroxylase effect of chain length, helical conformation and amino acid sequence in the substrate. *Biochim Biophys Acta Protein Struct.* (1972) 271:347–56. doi: 10.1016/0005-2795(72)90209-7
60. Persikov AV, Ramshaw JAM, Kirkpatrick A, Brodsky B. Electrostatic interactions involving lysine make major contributions to collagen triple-helix stability. *Biochemistry.* (2005) 44:1414–22. doi: 10.1021/bi048216r
61. Fallas JA, Gauba V, Hartgerink JD. Solution structure of an ABC collagen heterotrimer reveals a single-register helix stabilized by electrostatic interactions. *J Biol Chem.* (2009) 284:26851–9. doi: 10.2210/pdb2klw/pdb
62. Bella J, Eaton M, Brodsky B, Berman HM. Crystal and molecular structure of a collagen-like peptide at 1.9 Å resolution. *Science.* (1994) 266:75–81. doi: 10.1126/science.7695699
63. Bourhis J-M, Mariano N, Zhao Y, Harlos K, Exposito J-Y, Jones EY, et al. Structural basis of fibrillar collagen trimerization and related genetic disorders. *Nat Struct Mol Biol.* (2012) 19:1031–6. doi: 10.1038/nsmb.2389
64. McLaughlin SH, Bulleid NJ. Molecular recognition in procollagen chain assembly. *Matrix Biol.* (1998) 16:369–77. doi: 10.1016/S0945-053X(98)90010-5
65. Gillberg L, Berg S, de Verdier PJ, Lindbom L, Werr J, Hellstrom PM. Effective treatment of mouse experimental colitis by alpha 2 integrin antibody: comparison with alpha 4 antibody and conventional therapy. *Acta Physiol.* (2013) 207:326–36. doi: 10.1111/apha.12017
66. Ghatak S, Niland S, Schulz JN, Wang F, Eble JA, Leitges M, et al. Role of integrins $\alpha 1\beta 1$ and $\alpha 2\beta 1$ in wound and tumor angiogenesis in mice. *Am J Pathol.* (2016) 186:3011–27. doi: 10.1016/j.ajpath.2016.06.021
67. Bekhouche M, Colige A. The procollagen N-proteinases ADAMTS2, 3 and 14 in pathophysiology. *Matrix Biol.* (2015) 44–46:46–53. doi: 10.1016/j.matbio.2015.04.001
68. Vadon-Le Goff S, Hulmes DJS, Moali C. BMP-1/tolloid-like proteinases synchronize matrix assembly with growth factor activation to promote morphogenesis and tissue remodeling. *Matrix Biol.* (2015) 44–46:14–23. doi: 10.1016/j.matbio.2015.02.006
69. Hulmes DJS. Building collagen molecules, fibrils, and suprafibrillar structures. *J Struct Biol.* (2002) 137:2–10. doi: 10.1006/j.sbi.2002.4450
70. Porter S, Clark IM, Kevorkian L, Edwards DR. The ADAMTS metalloproteinases. *Biochem J.* (2005) 386:15–27. doi: 10.1042/BJ20040424
71. Hopkins DR, Keles S, Greenspan DS. The bone morphogenetic protein 1/Tolloid-like metalloproteinases. *Matrix Biol.* (2007) 26:508–23. doi: 10.1016/j.matbio.2007.05.004
72. Colige A, Ruggiero F, Vandenberghe I, Dubail J, Kesteloot F, Van Beeumen J, et al. Domains and maturation processes that regulate the activity of ADAMTS-2, a metalloproteinase cleaving the aminopropeptide of fibrillar procollagens types I–III and V. *J Biol Chem.* (2005) 280:34397–408. doi: 10.1074/jbc.M506458200
73. Le Goff C, Somerville RPT, Kesteloot F, Powell K, Birk DE, Colige AC, et al. Regulation of procollagen amino-propeptide processing during mouse embryogenesis by specialization of homologous ADAMTS proteases: insights on collagen biosynthesis and dermatosparaxis. *Development.* (2006) 133:1587–96. doi: 10.1242/dev.02308
74. Prockop DJ, Sieron AL, Li S-W. Procollagen N-proteinase and procollagen C-proteinase. Two unusual metalloproteinases that are essential for procollagen processing probably have important roles in development and cell signaling. *Matrix Biol.* (1998) 16:399–408. doi: 10.1016/S0945-053X(98)90013-0
75. Prockop DJ, Fertala A. Inhibition of the self-assembly of collagen I into fibrils with synthetic peptides. Demonstration that assembly is driven by specific binding sites on the monomers. *J Biol Chem.* (1998) 273:15598–604. doi: 10.1074/jbc.273.25.15598
76. Artym VV, Matsumoto K. Imaging cells in three-dimensional collagen matrix. *Curr Protoc Cell Biol.* (2010) 48:10.18.1–10.18.20. doi: 10.1002/0471143030.cb1018s48
77. Wenstrup RJ, Florer JB, Brunskill EW, Bell SM, Chervoneva I, Birk DE. Type V collagen controls the initiation of collagen fibril assembly. *J Biol Chem.* (2004) 279:53331–7. doi: 10.1074/jbc.M409622200
78. Kadler KE, Holmes DF, Trotter JA, Chapman JA. Collagen fibril formation. *Biochem J.* (1996) 316:1–11. doi: 10.1042/bj3160001
79. Yamauchi M, Sricholpech M. Lysine post-translational modifications of collagen. *Essays Biochem.* (2012) 52:113–33. doi: 10.1042/bse0520113
80. Trackman PC. Enzymatic and non-enzymatic functions of the lysyl oxidase family in bone. *Matrix Biol.* (2016) 52–54:7–18. doi: 10.1016/j.matbio.2016.01.001
81. Mäki JM. Lysyl oxidases in mammalian development and certain pathological conditions. *Histol Histopathol.* (2009) 24:651–60. doi: 10.14670/HH-24.651
82. Lucero HA, Kagan HM. Lysyl oxidase: an oxidative enzyme and effector of cell function. *Cell Mol Life Sci.* (2006) 63:2304–16. doi: 10.1007/s00018-006-6149-9
83. Rucker RB, Murray J. Cross-linking amino acids in collagen and elastin. *Am J Clin Nutr.* (1978) 31:1221–36. doi: 10.1093/ajcn/31.7.1221
84. Orgel JP, Wess TJ, Miller A. The *in situ* conformation and axial location of the intermolecular cross-linked non-helical telopeptides of type I collagen. *Structure.* (2000) 8:137–42. doi: 10.1016/S0969-2126(00)00089-7
85. Bruckner P. Suprastructures of extracellular matrices: paradigms of functions controlled by aggregates rather than molecules. *Cell Tissue Res.* (2009) 339:7–18. doi: 10.1007/s00441-009-0864-0
86. Molnar J, Fong KSK, He QP, Hayashi K, Kim Y, Fong SFT, et al. Structural and functional diversity of lysyl oxidase and the LOX-like proteins. *Biochim Biophys Acta Proteins Proteomics.* (2003) 1647:220–24. doi: 10.1016/S1570-9639(03)00053-0
87. Dvorak HF. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N Engl J Med.* (1986) 315:1650–9. doi: 10.1056/NEJM198612253152606
88. Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell.* (2012) 21:309–22. doi: 10.1016/j.ccr.2012.02.022
89. Bissell MJ, Hines WC. Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. *Nat Med.* (2011) 17:320–29. doi: 10.1038/nm.2328
90. Pietras K, Östman A. Hallmarks of cancer: interactions with the tumor stroma. *Exp Cell Res.* (2010) 316:1324–31. doi: 10.1016/j.yexcr.2010.02.045
91. Collins MA, Bednar F, Zhang Y, Brisset J-C, Galbán S, Galbán CJ, et al. Oncogenic Kras is required for both the initiation and maintenance of pancreatic cancer in mice. *J Clin Invest.* (2012) 122:639–53. doi: 10.1172/JCI59227
92. Helms E, Onate MK, Sherman MH. Fibroblast heterogeneity in the pancreatic tumor microenvironment. *Cancer Discov.* (2020) 10:648–56. doi: 10.1158/2159-8290.CD-19-1353
93. Pankova D, Chen Y, Terajima M, Schliekelman MJ, Baird BN, Fahrenholtz M, et al. Cancer-associated fibroblasts induce a collagen cross-link switch in tumor stroma. *Mol Cancer Res.* (2016) 14:287–95. doi: 10.1158/1541-7786.MCR-15-0307
94. Costa A, Kieffer Y, Scholer-Dahirel A, Pelon F, Bourachot B, Cardon M, et al. Fibroblast heterogeneity and immunosuppressive environment in human breast cancer. *Cancer Cell.* (2018) 33:463–79.e10. doi: 10.1016/j.ccell.2018.01.011
95. Bartoschek M, Oskolkov N, Bocci M, Lötvrot J, Larsson C, Sommarin M, et al. Spatially and functionally distinct subclasses of breast cancer-associated fibroblasts revealed by single cell RNA sequencing. *Nat Commun.* (2018) 9:5150. doi: 10.1038/s41467-018-07582-3
96. Evans RA, Tian YC, Steadman R, Phillips AO. TGF- $\beta 1$ -mediated fibroblast-myofibroblast terminal differentiation—the role of smad proteins. *Exp Cell Res.* (2003) 282:90–100. doi: 10.1016/S0014-4827(02)00015-0
97. Vaughan MB, Howard EW, Tomasek JJ. Transforming growth factor- $\beta 1$ promotes the morphological and functional differentiation of the myofibroblast. *Exp Cell Res.* (2000) 257:180–89. doi: 10.1006/excr.2000.4869
98. Lopez JI, Kang I, You W-K, McDonald DM, Weaver VM. *In situ* force mapping of mammary gland transformation. *Integr Biol.* (2011) 3:910–21. doi: 10.1039/c1ib00043h
99. Acerbi I, Cassereau L, Dean I, Shi Q, Au A, Park C, et al. Human breast cancer invasion and aggression correlates with ECM stiffening and immune cell infiltration. *Integr Biol.* (2015) 7:1120–34. doi: 10.1039/c5ib00040h
100. Cox TR, Erler JT. Remodeling and homeostasis of the extracellular matrix: implications for fibrotic diseases and cancer. *Dis Model Mech.* (2011) 4:165–78. doi: 10.1242/dmm.004077

101. Friedl P, Wolf K. Tube travel: the role of proteases in individual and collective cancer cell invasion. *Cancer Res.* (2008) 68:7247–9. doi: 10.1158/0008-5472.CAN-08-0784
102. Gritsenko PG, Ilina O, Friedl P. Interstitial guidance of cancer invasion. *J Pathol.* (2012) 226:185–99. doi: 10.1002/path.3031
103. Provenzano PP, Eliceiri KW, Campbell JM, Inman DR, White JG, Keely PJ. Collagen reorganization at the tumor-stromal interface facilitates local invasion. *BMC Med.* (2006) 4:215–23. doi: 10.1186/1741-7015-4-38
104. Condeelis J, Segall JE. Intravital imaging of cell movement in tumours. *Nat Rev Cancer.* (2003) 3:921–30. doi: 10.1038/nrc1231
105. Kraning-Rush CM, Carey SP, Lampi MC, Reinhart-King CA. Microfabricated collagen tracks facilitate single cell metastatic invasion in 3D. *Integr Biol.* (2013) 5:606–16. doi: 10.1039/c3ib20196a
106. Riching KM, Cox BL, Salick MR, Pehlke C, Riching AS, Ponik SM, et al. 3D collagen alignment limits protrusions to enhance breast cancer cell persistence. *Biophys J.* (2014) 107:2546–58. doi: 10.1016/j.bpj.2014.10.035
107. Wolf K, te Lindert M, Krause M, Alexander S, te Riet J, Willis AL, et al. Physical limits of cell migration: control by ECM space and nuclear deformation and tuning by proteolysis and traction force. *J Cell Biol.* (2013) 201:1069–84. doi: 10.1083/jcb.201210152
108. Pathak A, Kumar S. Independent regulation of tumor cell migration by matrix stiffness and confinement. *Proc Natl Acad Sci USA.* (2012) 109:10334–9. doi: 10.1073/pnas.1118073109
109. Doyle AD, Petrie RJ, Kutys ML, Yamada KM. Dimensions in cell migration. *Curr Opin Cell Biol.* (2013) 25:642–9. doi: 10.1016/j.ccb.2013.06.004
110. Fenner J, Stacer AC, Winterroth F, Johnson TD, Luker KE, Luker GD. Macroscopic stiffness of breast tumors predicts metastasis. *Sci Rep.* (2014) 4:5512. doi: 10.1038/srep05512
111. Reid SE, Kay EJ, Neilson LJ, Henze AT, Serneels J, McGhee EJ, et al. Tumor matrix stiffness promotes metastatic cancer cell interaction with the endothelium. *EMBO J.* (2017) 36:2373–89. doi: 10.15252/emj.201694912
112. Harada T, Swift J, Irianto J, Shin J-W, Spinler KR, Athirasala A, et al. Nuclear lamin stiffness is a barrier to 3D migration, but softness can limit survival. *J Cell Biol.* (2014) 204:669–82. doi: 10.1083/jcb.201308029
113. Denais CM, Gilbert RM, Isermann P, McGregor AL, te Lindert M, Weigel B, et al. Nuclear envelope rupture and repair during cancer cell migration. *Science.* (2016) 352:353–8. doi: 10.1126/science.aad7297
114. Paul CD, Mistriotis P, Konstantopoulos K. Cancer cell motility: lessons from migration in confined spaces. *Nat Rev Cancer.* (2017) 17:131–40. doi: 10.1038/nrc.2016.123
115. Petrie RJ, Yamada KM. Multiple mechanisms of 3D migration: the origins of plasticity. *Curr Opin Cell Biol.* (2016) 42:7–12. doi: 10.1016/j.ccb.2016.03.025
116. Gaggioli C, Hooper S, Hidalgo-Carcedo C, Grosse R, Marshall JF, Harrington K, et al. Fibroblast-led collective invasion of carcinoma cells with differing roles for RhoGTPases in leading and following cells. *Nat Cell Biol.* (2007) 9:1392–400. doi: 10.1038/ncb1658
117. Sharma P, Ng C, Jana A, Padhi A, Szymanski P, Lee JSH, et al. Aligned fibers direct collective cell migration to engineer closing and nonclosing wound gaps. *Mol Biol Cell.* (2017) 28:2579–88. doi: 10.1091/mbc.e17-05-0305
118. Ray A, Slama ZM, Morford RK, Madden SA, Provenzano PP. Enhanced directional migration of cancer stem cells in 3D aligned collagen matrices. *Biophys J.* (2017) 112:1023–36. doi: 10.1016/j.bpj.2017.01.007
119. Malik R, Lelkes PI, Cukierman E. Biomechanical and biochemical remodeling of stromal extracellular matrix in cancer. *Trends Biotechnol.* (2015) 33:230–6. doi: 10.1016/j.tibtech.2015.01.004
120. Beacham DA, Cukierman E. Stromagenesis: the changing face of fibroblastic microenvironments during tumor progression. *Semin Cancer Biol.* (2005) 15:329–41. doi: 10.1016/j.semcancer.2005.05.003
121. Conklin MW, Eickhoff JC, Riching KM, Pehlke CA, Eliceiri KW, Provenzano PP, et al. Aligned collagen is a prognostic signature for survival in human breast carcinoma. *AJPA.* (2011) 178:1221–32. doi: 10.1016/j.ajpath.2010.11.076
122. Conklin MW, Gangnon RE, Sprague BL, Van Gemert L, Hampton JM, Eliceiri KW, et al. Collagen alignment as a predictor of recurrence after ductal carcinoma *in situ*. *Cancer Epidemiol Prev Biomarkers.* (2018) 27:138–45. doi: 10.1158/1055-9965.EPI-17-0720
123. Colpaert C, Vermeulen P, Jeuris W, van Beest P, Goovaerts G, Weyler J, et al. Early distant relapse in ‘node-negative’ breast cancer patients is not predicted by occult axillary lymph node metastases, but by the features of the primary tumour. *J Pathol.* (2001) 193:442–9. doi: 10.1002/path.829
124. Ford AJ, Orbach SM, Rajagopalan P. Fibroblasts stimulate macrophage migration in interconnected extracellular matrices through tunnel formation and fiber alignment. *Biomaterials.* (2019) 209:88–102. doi: 10.1016/j.biomaterials.2019.03.044
125. Jia H, Janjanam J, Wu SC, Wang R, Pano G, Celestine M, et al. The tumor cell-secreted matricellular protein WISP1 drives pro-metastatic collagen linearization. *EMBO J.* (2019) 38:315–55. doi: 10.15252/emj.2018101302
126. Lee H-O, Mullins SR, Franco-Barraza J, Valianou M, Cukierman E, Cheng JD. FAP-overexpressing fibroblasts produce an extracellular matrix that enhances invasive velocity and directionality of pancreatic cancer cells. *BMC Cancer.* (2011) 11:245. doi: 10.1186/1471-2407-11-245
127. Heldin C-H, Rubin K, Pietras K, Östman A. High interstitial fluid pressure — an obstacle in cancer therapy. *Nat Rev Cancer.* (2004) 4:806–13. doi: 10.1038/nrc1456
128. Olivares O, Mayers JR, Gouirand V, Torrence ME, Gicquel T, Borge L, et al. Collagen-derived proline promotes pancreatic ductal adenocarcinoma cell survival under nutrient limited conditions. *Nat Commun.* (2017) 8:16014–31. doi: 10.1038/ncomms16031
129. de Fougères AR, Sprague AG, Nickerson-Nutter CL, Chi-Rosso G, Rennett PD, Gardner H, et al. Regulation of inflammation by collagen-binding integrins alpha1beta1 and alpha2beta1 in models of hypersensitivity and arthritis. *J Clin Invest.* (2000) 105:721–9. doi: 10.1172/JCI7911
130. Naba A, Clauser KR, Hoersch S, Liu H, Carr SA, Hynes RO. The matrisome: *in silico* definition and *in vivo* characterization by proteomics of normal and tumor extracellular matrices. *Mol Cell Proteomics.* (2012) 11:M1111.014647. doi: 10.1074/mcp.M1111.014647
131. Naba A, Clauser KR, Lamar JM, Carr SA, Hynes RO. Extracellular matrix signatures of human mammary carcinoma identify novel metastasis promoters. *Elife.* (2014) 3:e01308. doi: 10.7554/eLife.01308
132. Zahir N, Weaver VM. Death in the third dimension: apoptosis regulation and tissue architecture. *Curr Opin Genet Dev.* (2004) 14:71–80. doi: 10.1016/j.gde.2003.12.005
133. Jenkins MH, Croteau W, Mullins DW, Brinckerhoff CE. The BRAFV600E inhibitor, PLX4032, increases type I collagen synthesis in melanoma cells. *Matrix Biol.* (2015) 48:66–77. doi: 10.1016/j.matbio.2015.05.007
134. Afik R, Zigmund E, Vugman M, Klepfish M, Shimshoni E, Pasmanik-Chor M, et al. Tumor macrophages are pivotal constructors of tumor collagenous matrix. *J Exp Med.* (2016) 213:2315–31. doi: 10.1084/jem.20151193
135. Drifka CR, Loeffler AG, Mathewson K, Keikhosravi A, Eickhoff JC, Liu Y, et al. Highly aligned stromal collagen is a negative prognostic factor following pancreatic ductal adenocarcinoma resection. *Oncotarget.* (2016) 7:76197–213. doi: 10.18632/oncotarget.12772
136. Hanley CJ, Noble F, Ward M, Bullock M, Drifka C, Mellone M, et al. A subset of myofibroblastic cancer-associated fibroblasts regulate collagen fiber elongation, which is prognostic in multiple cancers. *Oncotarget.* (2016) 7:6159–74. doi: 10.18632/oncotarget.6740
137. Kehlet SN, Sanz-Pamplona R, Brix S, Leeming DJ, Karsdal MA, Moreno V. Excessive collagen turnover products are released during colorectal cancer progression and elevated in serum from metastatic colorectal cancer patients. *Sci Rep.* (2016) 6:30599. doi: 10.1038/srep30599
138. Willumsen N, Bager CL, Leeming DJ, Smith V, Christiansen C, Karsdal MA, et al. Serum biomarkers reflecting specific tumor tissue remodeling processes are valuable diagnostic tools for lung cancer. *Cancer Med.* (2014) 3:1136–45. doi: 10.1002/cam4.303
139. Lu P, Weaver VM, Werb Z. The extracellular matrix: a dynamic niche in cancer progression. *J Cell Biol.* (2012) 196:395–406. doi: 10.1083/jcb.201102147
140. Bager CL, Willumsen N, Leeming DJ, Smith V, Karsdal MA, Dornan D, et al. Collagen degradation products measured in serum can separate ovarian and breast cancer patients from healthy controls: a preliminary study. *Cancer Biomark.* (2015) 15:783–8. doi: 10.3233/CBM-150520
141. Mammoto T, Jiang A, Jiang E, Panigrahy D, Kieran MW, Mammoto A. Role of collagen matrix in tumor angiogenesis and glioblastoma multiforme progression. *Am J Pathol.* (2013) 183:1293–305. doi: 10.1016/j.ajpath.2013.06.026

142. Thangavelu PU, Krenács T, Dray E, Duijf PHG. In epithelial cancers, aberrant COL17A1 promoter methylation predicts its misexpression and increased invasion. *Clin Epigenetics*. (2016) 8:120. doi: 10.1186/s13148-016-0290-6
143. Matte BF, Kumar A, Placone JK, Zanella VG, Martins MD, Engler AJ, et al. Matrix stiffness mechanically conditions EMT and migratory behavior of oral squamous cell carcinoma. *J Cell Sci*. (2019) 132:jcs224360. doi: 10.1242/jcs.224360
144. Townson JL, Chambers AF. Dormancy of solitary metastatic cells. *Cell Cycle*. (2006) 5:1744–50. doi: 10.4161/cc.5.16.2864
145. Wikman H, Vessella R, Pantel K. Cancer micrometastasis and tumour dormancy. *APMIS*. (2008) 116:754–70. doi: 10.1111/j.1600-0463.2008.01033.x
146. Barkan D, El Touny LH, Michalowski AM, Smith JA, Chu I, Davis AS, et al. Metastatic growth from dormant cells induced by a col-I-enriched fibrotic environment. *Cancer Res*. (2010) 70:5706–16. doi: 10.1158/0008-5472.CAN-09-2356
147. Brisson BK, Mauldin EA, Lei W, Vogel LK, Power AM, Lo A, et al. Type III collagen directs stromal organization and limits metastasis in a murine model of breast cancer. *Am J Pathol*. (2015) 185:1471–86. doi: 10.1016/j.ajpath.2015.01.029
148. Chintala SK, Sawaya R, Gokaslan ZL, Rao JS. The effect of type III collagen on migration and invasion of human glioblastoma cell lines *in vitro*. *Cancer Lett*. (1996) 102:57–63. doi: 10.1016/0304-3835(96)04163-8
149. Ma H-P, Chang H-L, Bamodu OA, Yadav VK, Huang T-Y, Wu ATH, et al. Collagen 1A1 (COL1A1) is a reliable biomarker and putative therapeutic target for hepatocellular carcinogenesis and metastasis. *Cancers*. (2019) 11:786. doi: 10.3390/cancers11060786
150. Zhu H, Chen H, Wang J, Zhou L, Liu S. Collagen stiffness promoted non-muscle-invasive bladder cancer progression to muscle-invasive bladder cancer. *Onco Targets Ther*. (2019) 12:3441–57. doi: 10.2147/OTT.S194568
151. Sun S, Wang Y, Wu Y, Gao Y, Li Q, Abdulrahman AA, et al. Identification of COL1A1 as an invasion-related gene in malignant astrocytoma. *Int J Oncol*. (2018) 53:2542–54. doi: 10.3892/ijo.2018.4568
152. Liu J, Shen J-X, Wu H-T, Li X-L, Wen X-F, Du C-W, et al. Collagen 1A1 (COL1A1) promotes metastasis of breast cancer and is a potential therapeutic target. *Discov Med*. (2018) 25:211–23.
153. Zhang Z, Wang Y, Zhang J, Zhong J, Yang R. COL1A1 promotes metastasis in colorectal cancer by regulating the WNT/PCP pathway. *Mol Med Rep*. (2018) 17:5037–42. doi: 10.3892/mmr.2018.8533
154. He B, Lin X, Tian F, Yu W, Qiao B. MiR-133a-3p inhibits oral squamous cell carcinoma (OSCC) proliferation and invasion by suppressing COL1A1. *J Cell Biochem*. (2018) 119:338–46. doi: 10.1002/jcb.26182
155. Wang Q, Yu J. MiR-129-5p suppresses gastric cancer cell invasion and proliferation by inhibiting COL1A1. *Biochem Cell Biol*. (2018) 96:19–25. doi: 10.1139/bcb-2016-0254
156. Ji J, Zhao L, Budhu A, Forgues M, Jia H-L, Qin L-X, et al. Let-7g targets collagen type I alpha2 and inhibits cell migration in hepatocellular carcinoma. *J Hepatol*. (2010) 52:690–7. doi: 10.1016/j.jhep.2009.12.025
157. Wang J, Gao P, Song Y, Sun J, Chen X, Yu H, et al. Prognostic value of gastric cancer-associated gene signatures: evidence based on a meta-analysis using integrated bioinformatics methods. *J Cell Mol Med*. (2018) 22:5743–7. doi: 10.1111/jcmm.13823
158. Yan Z, Xu W, Xiong Y, Cheng Y, Xu H, Wang Z, et al. Highly accurate two-gene signature for gastric cancer. *Med Oncol*. (2013) 30:584. doi: 10.1007/s12032-013-0584-x
159. Yue H, Wang J, Chen R, Hou X, Li J, Lu X. Gene signature characteristic of elevated stromal infiltration and activation is associated with increased risk of hematogenous and lymphatic metastasis in serous ovarian cancer. *BMC Cancer*. (2019) 19:1266. doi: 10.1186/s12885-019-6470-y
160. Engqvist H, Parris TZ, Kovács A, Nemes S, Werner Rönnerman E, De Lara S, et al. Immunohistochemical validation of COL3A1, GPR158 and PITHD1 as prognostic biomarkers in early-stage ovarian carcinomas. *BMC Cancer*. (2019) 19:912–28. doi: 10.1186/s12885-019-6084-4
161. Januchowski R, Swierczewska M, Sterzynska K, Wojtowicz K, Nowicki M, Zabel M. Increased expression of several collagen genes is associated with drug resistance in ovarian cancer cell lines. *J Cancer*. (2016) 7:1295–310. doi: 10.7150/jca.15371
162. Yuan L, Shu B, Chen L, Qian K, Wang Y, Qian G, et al. Overexpression of COL3A1 confers a poor prognosis in human bladder cancer identified by co-expression analysis. *Oncotarget*. (2017) 8:70508–20. doi: 10.18632/oncotarget.19733
163. Gao Y-F, Mao X-Y, Zhu T, Mao C-X, Liu Z-X, Wang Z-B, et al. COL3A1 and SNAP91: novel glioblastoma markers with diagnostic and prognostic value. *Oncotarget*. (2016) 7:70494–503. doi: 10.18632/oncotarget.12038
164. Wang X-Q, Tang Z-X, Yu D, Cui S-J, Jiang Y-H, Zhang Q, et al. Epithelial but not stromal expression of collagen alpha-1(III) is a diagnostic and prognostic indicator of colorectal carcinoma. *Oncotarget*. (2016) 7:8823–38. doi: 10.18632/oncotarget.6815
165. Ren W, Zhang Y, Zhang L, Lin Q, Zhang J, Xu G. Overexpression of collagen type V $\alpha 1$ chain in human breast invasive ductal carcinoma is mediated by TGF- $\beta 1$. *Int J Oncol*. (2018) 52:1694–704. doi: 10.3892/ijo.2018.4317
166. Liu W, Wei H, Gao Z, Chen G, Liu Y, Gao X, et al. COL5A1 may contribute the metastasis of lung adenocarcinoma. *Gene*. (2018) 665:57–66. doi: 10.1016/j.gene.2018.04.066
167. Feng G, Ma H-M, Huang H-B, Li Y-W, Zhang P, Huang J-J, et al. Overexpression of COL5A1 promotes tumor progression and metastasis and correlates with poor survival of patients with clear cell renal cell carcinoma. *Cancer Manag Res*. (2019) 11:1263–74. doi: 10.2147/CMAR.S188216
168. Chen H-C, Tseng Y-K, Shu C-W, Weng T-J, Liou H-H, Yen L-M, et al. Differential clinical significance of COL5A1 and COL5A2 in tongue squamous cell carcinoma. *J Oral Pathol Med*. (2019) 48:468–76. doi: 10.1111/jop.12861
169. Wu M, Sun Q, Mo C-H, Pang J-S, Hou J-Y, Pang L-L, et al. Prospective molecular mechanism of COL5A1 in breast cancer based on a microarray, RNA sequencing and immunohistochemistry. *Oncol Rep*. (2019) 42:151–75. doi: 10.3892/or.2019.7147
170. Zeng X-T, Liu X-P, Liu T-Z, Wang X-H. The clinical significance of COL5A2 in patients with bladder cancer: a retrospective analysis of bladder cancer gene expression data. *Medicine*. (2018) 97:e0091. doi: 10.1097/MD.00000000000010091
171. Huang G, Ge G, Izzi V, Greenspan DS. $\alpha 3$ chains of type V collagen regulate breast tumour growth via glypican-1. *Nat Commun*. (2017) 8:14351. doi: 10.1038/ncomms14351
172. Wu YH, Chang TH, Huang YF, Huang HD, Chou C-Y. COL11A1 promotes tumor progression and predicts poor clinical outcome in ovarian cancer. *Oncogene*. (2014) 33:3432–40. doi: 10.1038/onc.2013.307
173. Wu Y-H, Chang T-H, Huang Y-F, Chen C-C, Chou C-Y. COL11A1 confers chemoresistance on ovarian cancer cells through the activation of Akt/c/EBP β pathway and PDK1 stabilization. *Oncotarget*. (2015) 6:23748–63. doi: 10.18632/oncotarget.4250
174. Shen L, Yang M, Lin Q, Zhang Z, Zhu B, Miao C. COL11A1 is overexpressed in recurrent non-small cell lung cancer and promotes cell proliferation, migration, invasion and drug resistance. *Oncol Rep*. (2016) 36:877–85. doi: 10.3892/or.2016.4869
175. Jia D, Liu Z, Deng N, Tan TZ, Huang RY-J, Taylor-Harding B, et al. A COL11A1-correlated pan-cancer gene signature of activated fibroblasts for the prioritization of therapeutic targets. *Cancer Lett*. (2016) 382:203–14. doi: 10.1016/j.canlet.2016.09.001
176. Li A, Li J, Lin J, Zhuo W, Si J. COL11A1 is overexpressed in gastric cancer tissues and regulates proliferation, migration and invasion of HGC-27 gastric cancer cells *in vitro*. *Oncol Rep*. (2017) 37:333–40. doi: 10.3892/or.2016.5276
177. Zhang B, Zhang C, Yang X, Chen Y, Zhang H, Liu J, et al. Cytoplasmic collagen XI α I as a prognostic biomarker in esophageal squamous cell carcinoma. *Cancer Biol Ther*. (2018) 19:364–72. doi: 10.1080/15384047.2018.1423915
178. Sun D, Jin H, Zhang J, Tan X. Integrated whole genome microarray analysis and immunohistochemical assay identifies COL11A1, GJB2 and CTRL as predictive biomarkers for pancreatic cancer. *Cancer Cell Int*. (2018) 18:174–11. doi: 10.1186/s12935-018-0669-x
179. Toss MS, Miligy IM, Gorringer KL, Aleskandarany MA, Alkawas A, Mittal K, et al. Collagen (XI) alpha-1 chain is an independent prognostic factor in breast ductal carcinoma *in situ*. *Mod Pathol*. (2019) 32:1460–72. doi: 10.1038/s41379-019-0286-9
180. Misawa K, Kanazawa T, Imai A, Endo S, Mochizuki D, Fukushima H, et al. Prognostic value of type XXII and XXIV collagen mRNA

- expression in head and neck cancer patients. *Mol Clin Oncol.* (2014) 2:285–91. doi: 10.3892/mco.2013.233
181. Wang F, Dong J, Zhang Y, Yue S, Guo H, Liang P, et al. Identification of novel diagnostic and prognostic biomarkers for hepatocellular carcinoma. *Oncol Rep.* (2019) 43:87–98. doi: 10.3892/or.2019.7415
 182. Nagy Á, Lániczky A, Menyhart O, Gyorffy B. Validation of miRNA prognostic power in hepatocellular carcinoma using expression data of independent datasets. *Sci Rep.* (2018) 8:9227–9. doi: 10.1038/s41598-018-29514-3
 183. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. *CA Cancer J Clin.* (2018) 68:7–30. doi: 10.3322/caac.21442
 184. Fane M, Weeraratna AT. How the ageing microenvironment influences tumour progression. *Nat Rev Cancer.* (2019) 20:89–106. doi: 10.1038/s41568-019-0222-9
 185. Panwar P, Lamour G, Mackenzie NCW, Yang H, Ko F, Li H, et al. Changes in structural-mechanical properties and degradability of collagen during aging-associated modifications. *J Biol Chem.* (2015) 290:23291–306. doi: 10.1074/jbc.M115.644310
 186. Diridollou S, Vabre V, Berson M, Vaillant L, Black D, Lagarde JM, et al. Skin ageing: changes of physical properties of human skin *in vivo*. *Int J Cosmet Sci.* (2001) 23:353–62. doi: 10.1046/j.0412-5463.2001.00105.x
 187. Fisher GJ, Kang S, Varani J, Bata-Csorgo Z, Wan Y, Datta S, et al. Mechanisms of photoaging and chronological skin aging. *Arch Dermatol.* (2002) 138:1462–70. doi: 10.1001/archderm.138.11.1462
 188. Oh J-H, Kim YK, Jung J-Y, Shin J, Kim KH, Cho KH, et al. Intrinsic aging- and photoaging-dependent level changes of glycosaminoglycans and their correlation with water content in human skin. *J Dermatol Sci.* (2011) 62:192–201. doi: 10.1016/j.jdermsci.2011.02.007
 189. Marcos-Garcés V, Molina Aguilar P, Bea Serrano C, García Bustos V, Benavent Seguí J, Ferrández Izquierdo A, et al. Age-related dermal collagen changes during development, maturation and ageing - a morphometric and comparative study. *J Anat.* (2014) 225:98–108. doi: 10.1111/joa.12186
 190. Simm A, Müller B, Nass N, Hofmann B, Bushnaq H, Silber R-E, et al. Protein glycation - between tissue aging and protection. *Exp Gerontol.* (2015) 68:71–75. doi: 10.1016/j.exger.2014.12.013
 191. Monnier VM, Mustata GT, Biemel KL, Reihl O, Lederer MO, Zhenyu D, et al. Cross-linking of the extracellular matrix by the maillard reaction in aging and diabetes: an update on 'a puzzle nearing resolution'. *Ann N Y Acad Sci.* (2005) 1043:533–44. doi: 10.1196/annals.1333.061
 192. Ait-Belkacem D, Guilbert M, Roche M, Duboisset J, Ferrand P, Sockalingum G, et al. Microscopic structural study of collagen aging in isolated fibrils using polarized second harmonic generation. *J Biomed Opt.* (2012) 17:80501–6. doi: 10.1117/1.JBO.17.8.080506
 193. Wilson SL, Guilbert M, Sulé-Suso J, Torbet J, Jeannesson P, Sockalingum GD, et al. A microscopic and macroscopic study of aging collagen on its molecular structure, mechanical properties, and cellular response. *FASEB J.* (2014) 28:14–25. doi: 10.1096/fj.13-227579
 194. Guilbert M, Roig B, Terryn C, Garnotel R, Jeannesson P, Sockalingum GD, et al. Highlighting the impact of aging on type I collagen: label-free investigation using confocal reflectance microscopy and diffuse reflectance spectroscopy in 3D matrix model. *Oncotarget.* (2016) 7:8546–55. doi: 10.18632/oncotarget.7385
 195. Panwar P, Butler GS, Jamroz A, Azizi P, Overall CM, Brömme D. Aging-associated modifications of collagen affect its degradation by matrix metalloproteinases. *Matrix Biol.* (2018) 65:30–44. doi: 10.1016/j.matbio.2017.06.004
 196. Saby C, Buache E, Brassart-Pasco S, El Btaouri H, Courageot M-P, Van Gulick L, et al. Type I collagen aging impairs discoidin domain receptor 2-mediated tumor cell growth suppression. *Oncotarget.* (2016) 7:24908–27. doi: 10.18632/oncotarget.8795
 197. Saby C, Rammal H, Magnien K, Buache E, Brassart-Pasco S, Van-Gulick L, et al. Age-related modifications of type I collagen impair DDR1-induced apoptosis in non-invasive breast carcinoma cells. *Cell Adhes Migr.* (2018) 12:335–47. doi: 10.1080/19336918.2018.1472182
 198. Alcaraz J, Carrasco JL, Millares L, Luis I-C, Fernández-Porras FJ, Martínez-Romero A, et al. Stromal markers of activated tumor associated fibroblasts predict poor survival and are associated with necrosis in non-small cell lung cancer. *Lung Cancer.* (2019) 135:151–60. doi: 10.1016/j.lungcan.2019.07.020
 199. Mayorca-Guiliani AE, Willacy O, Madsen CD, Rafaeva M, Heumüller SE, Bock F, et al. Decellularization and antibody staining of mouse tissues to map native extracellular matrix structures in 3D. *Nat Protoc.* (2019) 14:3395–425. doi: 10.1038/s41596-019-0225-8
 200. Rich L, Whittaker P. Collagen and picrosirius red staining: a polarized light assessment of fibrillar hue and spatial distribution. *Braz J Morphol Sci.* (2005) 22:97–104.
 201. Friedl P, Maaser K, Klein CE, Niggemann B, Krohne G, Zänker KS. Migration of highly aggressive MV3 melanoma cells in 3-dimensional collagen lattices results in local matrix reorganization and shedding of alpha2 and beta1 integrins and CD44. *Cancer Res.* (1997) 57:2061–70.
 202. Choi S, Friedrichs J, Song YH, Werner C, Estroff LA, Fischbach C. Intrafibrillar, bone-mimetic collagen mineralization regulates breast cancer cell adhesion and migration. *Biomaterials.* (2019) 198:95–106. doi: 10.1016/j.biomaterials.2018.05.002
 203. Brightman AO, Rajwa BP, Sturgis JE, McCallister ME, Robinson JP, Harbin SLV. Time-lapse confocal reflection microscopy of collagen fibrillogenesis and extracellular matrix assembly *in vitro*. *Biopolymers.* (2000) 54:222–34. doi: 10.1002/1097-0282(200009)54:3<222::AID-BIP80>3.0.CO;2-K
 204. Friedl P. Dynamic imaging of cellular interactions with extracellular matrix. *Histochem Cell Biol.* (2004) 122:183–90. doi: 10.1007/s00418-004-0682-0
 205. Wielowieyska-Szybinska D, Białek-Galas K, Podolec K, Wojas-Pelc A. The use of reflectance confocal microscopy for examination of benign and malignant skin tumors. *Postepy Dermatol Alergol.* (2014) 31:380–7. doi: 10.5114/pdia.2014.40961
 206. Bozkurt A, Kose K, Alessi-Fox C, Dy JG, Brooks DH, Rajadhyaksha M. Unsupervised delineation of stratum corneum using reflectance confocal microscopy and spectral clustering. *Skin Res Technol.* (2017) 23:176–85. doi: 10.1111/srt.12316
 207. Ahlgrimm-Siess V, Laimer M, Rabinovitz HS, Oliviero M, Hofmann-Wellenhof R, Marghoob AA, et al. Confocal microscopy in skin cancer. *Curr Dermatol Rep.* (2018) 7:105–18. doi: 10.1007/s13671-018-0218-9
 208. Abraham T, Hogg J. Lung alveolar wall disruption in three-dimensional space identified using second-harmonic generation multiphoton excitation fluorescence In: Vo-Dinh T, Grundfest WS, Mahadevan-Jansen A, editors. *Advanced Biomedical Clinical Diagnostic Systems VIII*. San Francisco, CA: International Society for Optics and Photonics (2010). 75550X.
 209. Williams RM, Zipfel WR, Webb WW. Interpreting second-harmonic generation images of collagen I fibrils. *Biophys J.* (2005) 88:1377–86. doi: 10.1529/biophysj.104.047308
 210. Mostaçõ-Guidolin L, Rosin NL, Hackett T-L. Imaging collagen in scar tissue: developments in second harmonic generation microscopy for biomedical applications. *Int J Mol Sci.* (2017) 18:1772. doi: 10.3390/ijms18081772
 211. Han X, Burke RM, Zettel ML, Tang P, Brown EB. Second harmonic properties of tumor collagen: determining the structural relationship between reactive stroma and healthy stroma. *Opt Express.* (2008) 16:1846–59. doi: 10.1364/OE.16.001846
 212. Lacombe R, Nadiarykh O, Townsend SS, Campagnola PJ. Phase matching considerations in second harmonic generation from tissues: effects on emission directionality, conversion efficiency and observed morphology. *Opt Commun.* (2008) 281:1823–32. doi: 10.1016/j.optcom.2007.10.040
 213. Roth S, Freund I. Optical second-harmonic scattering in rat-tail tendon. *Biopolymers.* (1981) 20:1271–90. doi: 10.1002/bip.1981.360200613
 214. Zipfel WR, Williams RM, Webb WW. Nonlinear magic: multiphoton microscopy in the biosciences. *Nat Biotechnol.* (2003) 21:1369–77. doi: 10.1038/nbt899
 215. Gailhouste L, Le Grand Y, Odin C, Guyader D, Turlin B, Ezan F, et al. Fibrillar collagen scoring by second harmonic microscopy: a new tool in the assessment of liver fibrosis. *J Hepatol.* (2010) 52:398–406. doi: 10.1016/j.jhep.2009.12.009
 216. Bredfeldt JS, Liu Y, Pehlke CA, Conklin MW, Szulzewski JM, Inman DR, et al. Computational segmentation of collagen fibers from second-harmonic generation images of breast cancer. *J Biomed Opt.* (2014) 19:16007. doi: 10.1117/1.JBO.19.1.016007
 217. Burke K. Using second harmonic generation to predict patient outcome in solid tumors. *BMC Cancer.* (2015) 15:929. doi: 10.1186/s12885-015-1911-8
 218. Suhonen H, Fernández M, Serimaa R, Suortti P. Simulation of small-angle x-ray scattering from collagen fibrils and

- comparison with experimental patterns. *Phys Med Biol.* (2005) 50:5401–16. doi: 10.1088/0031-9155/50/22/012
219. Ooi GJ, Fox J, Siu K, Lewis R, Bamberg KR, McNaughton D, et al. Fourier transform infrared imaging and small angle x-ray scattering as a combined biomolecular approach to diagnosis of breast cancer. *Med Phys.* (2008) 35:2151–61. doi: 10.1118/1.2890391
220. Nissi MJ, Töyräs J, Laasanen MS, Rieppo J, Saarakkala S, Lappalainen R, et al. Proteoglycan and collagen sensitive MRI evaluation of normal and degenerated articular cartilage. *J Orthop Res.* (2004) 22:557–64. doi: 10.1016/j.orthres.2003.09.008
221. Egnell L, Vidić I, Jerome NP, Bofin AM, Bathen TF, Goa PE. Stromal collagen content in breast tumors correlates with *in vivo* diffusion-weighted imaging: a comparison of multi B-value DWI with histologic specimen from benign and malignant breast lesions. *J Magn Reson Imaging.* (2019) 51:1868–87. doi: 10.1002/jmri.27018
222. Morris JL, Cross SJ, Lu Y, Kadler KE, Lu Y, Dallas SL, et al. Live imaging of collagen deposition during skin development and repair in a collagen I – GFP fusion transgenic zebrafish line. *Dev Biol.* (2018) 441:4–11. doi: 10.1016/j.ydbio.2018.06.001
223. Vogel W, Gish GD, Alves F, Pawson T. The discoidin domain receptor tyrosine kinases are activated by collagen. *Mol Cell.* (1997) 1:13–23. doi: 10.1016/S1097-2765(00)80003-9
224. Fu HL, Valiathan RR, Arkwright R, Sohail A, Mihai C, Kumarasiri M, et al. Discoidin domain receptors: unique receptor tyrosine kinases in collagen-mediated signaling. *J Biol Chem.* (2013) 288:7430–37. doi: 10.1074/jbc.R112.444158
225. Barrow AD, Raynal N, Andersen TL, Slatter DA, Bihan D, Pugh N, et al. OSCAR is a collagen receptor that costimulates osteoclastogenesis in DAP12-deficient humans and mice. *J Clin Invest.* (2011) 121:3505–16. doi: 10.1172/JCI45913
226. Nieswandt B, Watson SP. Platelet-collagen interaction: is GPVI the central receptor? *Blood.* (2003) 102:449–61. doi: 10.1182/blood-2002-12-3882
227. Melander MC, Jurgensen HJ, Madsen DH, Engelholm LH, Behrendt N. The collagen receptor uPARAP/Endo180 in tissue degradation and cancer. *Int J Oncol.* (2015) 47:1177–88. doi: 10.3892/ijo.2015.3120
228. Sprangers S, Everts V. Molecular pathways of cell-mediated degradation of fibrillar collagen. *Matrix Biol.* (2019) 75–76:190–200. doi: 10.1016/j.matbio.2017.11.008
229. Henriët E, Sala M, Abou Hammoud A, Tuarihionoa A, Di Martino J, Ros M, et al. Multitasking discoidin domain receptors are involved in several and specific hallmarks of cancer. *Cell Adhes Migr.* (2018) 12:363–77. doi: 10.1080/19336918.2018.1465156
230. Gullberg D. *I Domain Integrins*. Dordrecht: Springer (2014).
231. Zeltz C, Orgel J, Gullberg D. Molecular composition and function of integrin-based collagen glues-introducing COLINBRIs. *Biochim Biophys Acta.* (2014) 1840:2533–48. doi: 10.1016/j.bbagen.2013.12.022
232. Heino J. Cellular signaling by collagen-binding integrins. *Adv Exp Med Biol.* (2014) 819:143–55. doi: 10.1007/978-94-017-9153-3_10
233. Hamaia S, Farnsdale RW. Integrin recognition motifs in the human collagens. *Adv Exp Med Biol.* (2014) 819:127–42. doi: 10.1007/978-94-017-9153-3_9
234. Jokinen J, Dadu E, Nykvist P, Käpylä J, White DJ, Ivaska J, et al. Integrin-mediated cell adhesion to type I collagen fibrils. *J Biol Chem.* (2004) 279:31956–63. doi: 10.1074/jbc.M401409200
235. Eble JA, Golbik R, Mann K, Kuhn K. The alpha 1 beta 1 integrin recognition site of the basement membrane collagen molecule [alpha 1(IV)]2 alpha 2(IV). *EMBO J.* (1993) 12:4795–802. doi: 10.1002/j.1460-2075.1993.tb06168.x
236. Tulla M, Pentikainen OT, Viitasalo T, Kapyla J, Impola U, Nykvist P, et al. Selective binding of collagen subtypes by integrin alpha 1I, alpha 2I, and alpha 10I domains. *J Biol Chem.* (2001) 276:48206–12. doi: 10.1074/jbc.M104058200
237. Hamaia SW, Pugh N, Raynal N, Nemoz B, Stone R, Gullberg D, et al. Mapping of potent and specific binding motifs, GLOGEN and GVOGEA, for integrin alpha1beta1 using collagen toolkits II and III. *J Biol Chem.* (2012) 287:26019–28. doi: 10.1074/jbc.M112.353144
238. Kapyla J, Jaalinoja J, Tulla M, Ylostalo J, Nissinen L, Viitasalo T, et al. The fibril-associated collagen IX provides a novel mechanism for cell adhesion to cartilaginous matrix. *J Biol Chem.* (2004) 279:51677–87. doi: 10.1074/jbc.M409412200
239. Madamanchi A, Santoro SA, Zutter MM. Alpha2beta1 integrin. *Adv Exp Med Biol.* (2014) 819:41–60. doi: 10.1007/978-94-017-9153-3_3
240. Carafoli F, Hamaia SW, Bihan D, Hohenester E, Farnsdale RW. An activating mutation reveals a second binding mode of the integrin alpha2 I domain to the GFOGER motif in collagens. *PLoS ONE.* (2013) 8:e69833. doi: 10.1371/journal.pone.0069833
241. Camper L, Hellman U, Lundgren-Akerlund E. Isolation, cloning, and sequence analysis of the integrin subunit alpha10, a beta1-associated collagen binding integrin expressed on chondrocytes. *J Biol Chem.* (1998) 273:20383–9. doi: 10.1074/jbc.273.32.20383
242. Tiger CF, Fougerousse F, Grundstrom G, Velling T, Gullberg D. alpha110beta1 integrin is a receptor for interstitial collagens involved in cell migration and collagen reorganization on mesenchymal nonmuscle cells. *Dev Biol.* (2001) 237:116–29. doi: 10.1006/dbio.2001.0363
243. Zhang WM, Kapyla J, Puranen JS, Knight CG, Tiger CF, Pentikainen OT, et al. alpha 11beta 1 integrin recognizes the GFOGER sequence in interstitial collagens. *J Biol Chem.* (2003) 278:7270–7. doi: 10.1074/jbc.M210313200
244. Knight CG, Morton LF, Peachey AR, Tuckwell DS, Farnsdale RW, Barnes MJ. The collagen-binding A-domains of integrins alpha(1)beta(1) and alpha(2)beta(1) recognize the same specific amino acid sequence, GFOGER, in native (triple-helical) collagens. *J Biol Chem.* (2000) 275:35–40. doi: 10.1074/jbc.275.1.35
245. Sipilä KH, Drushin K, Rappu P, Jokinen J, Salminen TA, Salo AM, et al. Proline hydroxylation in collagen supports integrin binding by two distinct mechanisms. *J Biol Chem.* 293:7645–58. doi: 10.1074/jbc.RA118.002200
246. Kubow KE, Vukmirovic R, Zhe L, Klotzsch E, Smith ML, Gourdon D, et al. Mechanical forces regulate the interactions of fibronectin and collagen I in extracellular matrix. *Nat Commun.* (2015) 6:8026. doi: 10.1038/ncomms9026
247. Fiedler LR, Schonherr E, Waddington R, Niland S, Seidler DG, Aeschlimann D, et al. Decorin regulates endothelial cell motility on collagen I through activation of insulin-like growth factor I receptor and modulation of alpha2beta1 integrin activity. *J Biol Chem.* (2008) 283:17406–15. doi: 10.1074/jbc.M710025200
248. Zeltz C, Brezillon S, Kapyla J, Eble JA, Bobichon H, Terryn C, et al. Lumican inhibits cell migration through alpha2beta1 integrin. *Exp Cell Res.* (2010) 316:2922–31. doi: 10.1016/j.yexcr.2010.08.002
249. Zhu J, Madhurapantula RS, Kalyanasundaram A, Sabharwal T, Antipova O, Bishnoi SW, et al. Ultrastructural location and interactions of the immunoglobulin receptor binding sequence within fibrillar type I collagen. *Int J Mol Sci.* (2020) 21:4166. doi: 10.3390/ijms21114166
250. Seguin L, Desgrosellier JS, Weis SM, Cheresh DA. Integrins and cancer: regulators of cancer stemness, metastasis, and drug resistance. *Trends Cell Biol.* (2015) 25:234–40. doi: 10.1016/j.tcb.2014.12.006
251. Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. *Nat Rev Cancer.* (2018) 18:533–48. doi: 10.1038/s41568-018-0038-z
252. Zaidel-Bar R, Itzkovitz S, Ma'ayan A, Iyengar R, Geiger B. Functional atlas of the integrin adhesome. *Nat Cell Biol.* (2007) 9:858–67. doi: 10.1038/ncb0807-858
253. Humphries JD, Chastney MR, Askari JA, Humphries MJ. Signal transduction via integrin adhesion complexes. *Curr Opin Cell Biol.* (2019) 56:14–21. doi: 10.1016/j.ccb.2018.08.004
254. Horton ER, Byron A, Askari JA, Ng DHJ, Millon-Fremillon A, Robertson J, et al. Definition of a consensus integrin adhesome and its dynamics during adhesion complex assembly and disassembly. *Nat Cell Biol.* (2015) 17:1577–87. doi: 10.1038/ncb3257
255. Shattil SJ, Kim C, Ginsberg MH. The final steps of integrin activation: the end game. *Nat Rev Mol Cell Biol.* (2010) 11:288–300. doi: 10.1038/nrm2871
256. Kechagia JZ, Ivaska J, Roca-Cusachs P. Integrins as biomechanical sensors of the microenvironment. *Nat Rev Mol Cell Biol.* 20:1–17. doi: 10.1038/s41580-019-0134-2
257. Kagami S, Urushihara M, Kondo S, Loster K, Reutter W, Tamaki T, et al. Requirement for tyrosine kinase-ERK1/2 signaling in alpha 1 beta 1 integrin-mediated collagen matrix remodeling by rat mesangial cells. *Exp Cell Res.* (2001) 268:274–83. doi: 10.1006/excr.2001.5279

258. Inoue O, Suzuki-Inoue K, Dean WL, Frampton J, Watson SP. Integrin alpha2beta1 mediates outside-in regulation of platelet spreading on collagen through activation of Src kinases and PLCgamma2. *J Cell Biol.* (2003) 160:769–80. doi: 10.1083/jcb.200208043
259. Consonni A, Cipolla L, Guidetti G, Canobbio I, Ciralo E, Hirsch E, et al. Role and regulation of phosphatidylinositol 3-kinase β in platelet integrin $\alpha 2\beta 1$ signaling. *Blood.* (2012) 119:847–56. doi: 10.1182/blood-2011-07-364992
260. Guidetti GF, Bernardi B, Consonni A, Rizzo P, Gruppi C, Balduini C, et al. Integrin alpha2beta1 induces phosphorylation-dependent and phosphorylation-independent activation of phospholipase Cgamma2 in platelets: role of Src kinase and Rac GTPase. *J Thromb Haemost.* (2009) 7:1200–6. doi: 10.1111/j.1538-7836.2009.03444.x
261. Chen X, Abair TD, Ibanez MR, Su Y, Frey MR, Dise RS, et al. Integrin alpha1beta1 controls reactive oxygen species synthesis by negatively regulating epidermal growth factor receptor-mediated Rac activation. *Mol Cell Biol.* (2007) 27:3313–26. doi: 10.1128/MCB.01476-06
262. Honoré S, Kovacic H, Pichard V, Briand C, Rognoni J-B. Alpha2beta1-integrin signaling by itself controls G1/S transition in a human adenocarcinoma cell line (Caco2): implication of NADPH oxidase-dependent production of ROS. *Exp Cell Res.* (2003) 285:59–71. doi: 10.1016/S0014-4827(02)00038-1
263. Schulz JN, Zeltz C, Sorensen IW, Barczyk M, Carracedo S, Hallinger R, et al. Reduced granulation tissue and wound strength in the absence of alpha1beta1 integrin. *J Invest Dermatol.* (2015) 135:1435–44. doi: 10.1038/jid.2015.24
264. Harburger DS, Calderwood DA. Integrin signalling at a glance. *J Cell Sci.* (2009) 122:159–63. doi: 10.1242/jcs.018093
265. Bottcher RT, Veelders M, Rombaut P, Faix J, Theodosiou M, Stradal TE, et al. Kindlin-2 recruits paxillin and Arp2/3 to promote membrane protrusions during initial cell spreading. *J Cell Biol.* (2017) 216:3785–98. doi: 10.1083/jcb.201701176
266. Petrich BG, Marchese P, Ruggeri ZM, Spiess S, Weichert RA, Ye F, et al. Talin is required for integrin-mediated platelet function in hemostasis and thrombosis. *J Exp Med.* (2007) 204:3103–11. doi: 10.1084/jem.20071800
267. Salmela M, Jokinen J, Tiitta S, Rappu P, Cheng RH, Heino J. Integrin alpha2beta1 in nonactivated conformation can induce focal adhesion kinase signaling. *Sci Rep.* (2017) 7:3414. doi: 10.1038/s41598-017-03640-w
268. Rantala JK, Pouwels J, Pellinen T, Veltel S, Laasola P, Mattila E, et al. SHARPIN is an endogenous inhibitor of $\beta 1$ -integrin activation. *Nat Cell Biol.* (2011) 13:1315–24. doi: 10.1038/ncb2340
269. Mattila E, Pellinen T, Nevo J, Vuoriluoto K, Arjonen A, Ivaska J. Negative regulation of EGFR signalling through integrin-alpha1beta1-mediated activation of protein tyrosine phosphatase TCPTP. *Nat Cell Biol.* (2005) 7:78–85. doi: 10.1038/ncb1209
270. Ivaska J, Reunanen H, Westermarck J, Koivisto L, Kähäri VM, Heino J. Integrin alpha2beta1 mediates isoform-specific activation of p38 and upregulation of collagen gene transcription by a mechanism involving the alpha2 cytoplasmic tail. *J Cell Biol.* (1999) 147:401–16. doi: 10.1083/jcb.147.2.401
271. Langholz O, Röckel D, Mauch C, Kozłowska E, Bank I, Krieg T, et al. Collagen and collagenase gene expression in three-dimensional collagen lattices are differentially regulated by alpha 1 beta 1 and alpha 2 beta 1 integrins. *J Cell Biol.* (1995) 131:1903–15. doi: 10.1083/jcb.131.6.1903
272. Ravanti L, Heino J, López-Otin C, Kähäri VM. Induction of collagenase-3 (MMP-13) expression in human skin fibroblasts by three-dimensional collagen is mediated by p38 mitogen-activated protein kinase. *J Biol Chem.* (1999) 274:2446–55. doi: 10.1074/jbc.274.4.2446
273. Barczyk MM, Lu N, Popova SN, Bolstad AI, Gullberg D. alpha1beta1 integrin-mediated MMP-13-dependent collagen lattice contraction by fibroblasts: evidence for integrin-coordinated collagen proteolysis. *J Cell Physiol.* (2013) 228:1108–19. doi: 10.1002/jcp.24261
274. Lochter A, Navre M, Werb Z, Bissell MJ. alpha1 and alpha2 integrins mediate invasive activity of mouse mammary carcinoma cells through regulation of stromelysin-1 expression. *Mol Biol Cell.* (1999) 10:271–82. doi: 10.1091/mbc.10.2.271
275. Yamada KM, Even-Ram S. Integrin regulation of growth factor receptors. *Nat Cell Biol.* (2002) 4:E75–6. doi: 10.1038/ncb0402-e75
276. Ivaska J, Heino J. Cooperation between integrins and growth factor receptors in signaling and endocytosis. *Annu Rev Cell Dev Biol.* (2011) 27:291–320. doi: 10.1146/annurev-cellbio-092910-154017
277. Schnittert J, Bansal R, Storm G, Prakash J. Integrins in wound healing, fibrosis and tumor stroma: high potential targets for therapeutics and drug delivery. *Adv Drug Deliv Rev.* (2018) 129:37–53. doi: 10.1016/j.addr.2018.01.020
278. Guha D, Saha T, Bose S, Chakraborty S, Dhar S, Khan P, et al. Integrin-EGFR interaction regulates anoikis resistance in colon cancer cells. *Apoptosis.* (2019) 24:958–71. doi: 10.1007/s10495-019-01573-5
279. McCall-Culbreath KD, Li Z, Zutter MM. Crosstalk between the alpha2beta1 integrin and c-met/HGF-R regulates innate immunity. *Blood.* (2008) 111:3562–70. doi: 10.1182/blood-2007-08-107664
280. Primac I, Maquoi E, Blacher S, Heljasvaara R, Van Deun J, Smeland HY, et al. Stromal integrin alpha11 regulates PDGFR-beta signaling and promotes breast cancer progression. *J Clin Invest.* (2019) 130:4609–28. doi: 10.1172/JCI125890
281. Leitinger B. Discoidin domain receptor functions in physiological and pathological conditions. *Int Rev Cell Mol Biol.* (2014) 310:39–87. doi: 10.1016/B978-0-12-800180-6.00002-5
282. Danen EHJ. Integrin signaling as a cancer drug target. *ISRN Cell Biol.* (2013) 201:14. doi: 10.1155/2013/135164
283. Vachon PH. Integrin signaling, cell survival, and anoikis: distinctions, differences, and differentiation. *J Signal Transduct.* (2011) 2011:738137. doi: 10.1155/2011/738137
284. Conti JA, Kendall TJ, Bateman A, Armstrong TA, Papa-Adams A, Xu Q, et al. The desmoplastic reaction surrounding hepatic colorectal adenocarcinoma metastases aids tumor growth and survival via alphav integrin ligation. *Clin Cancer Res.* (2008) 14:6405–13. doi: 10.1158/1078-0432.CCR-08-0816
285. Gardner H. Integrin alpha1beta1. *Adv Exp Med Biol.* (2014) 819:21–39. doi: 10.1007/978-94-017-9153-3_2
286. Gardner H, Kreidberg J, Koteliensky V, Jaenisch R. Deletion of integrin alpha 1 by homologous recombination permits normal murine development but gives rise to a specific deficit in cell adhesion. *Dev Biol.* (1996) 175:301–13. doi: 10.1006/dbio.1996.0116
287. Pozzi A, Wary KK, Giancotti FG, Gardner HA. Integrin alpha1beta1 mediates a unique collagen-dependent proliferation pathway *in vivo*. *J Cell Biol.* (1998) 142:587–94. doi: 10.1083/jcb.142.2.587
288. Zemmyo M, Meharrar EJ, Kuhn K, Creighton-Achermann L, Lotz M. Accelerated, aging-dependent development of osteoarthritis in alpha1 integrin-deficient mice. *Arthritis Rheum.* (2003) 48:2873–80. doi: 10.1002/art.11246
289. Peng YW, Zalloccchi M, Meehan DT, Delimont D, Chang B, Hawes N, et al. Progressive morphological and functional defects in retinas from alpha1 integrin-null mice. *Invest Ophthalmol Vis Sci.* (2008) 49:4647–54. doi: 10.1167/iovs.08-2011
290. Conrad C, Boyman O, Tonel G, Tun-Kyi A, Lagner U, de Fougères L, et al. Alpha1beta1 integrin is crucial for accumulation of epidermal T cells and the development of psoriasis. *Nat Med.* (2007) 13:836–42. doi: 10.1038/nm1605
291. Ekholm E, Hankenson KD, Uusitalo H, Hiltunen A, Gardner H, Heino J, et al. Diminished callus size and cartilage synthesis in alpha 1 beta 1 integrin-deficient mice during bone fracture healing. *AJPA.* (2002) 160:1779–85. doi: 10.1016/S0002-9440(10)61124-8
292. Williams AS, Kang L, Zheng J, Grueter C, Bracy DP, James FD, et al. Integrin alpha1-null mice exhibit improved fatty liver when fed a high fat diet despite severe hepatic insulin resistance. *J Biol Chem.* (2015) 290:6546–57. doi: 10.1074/jbc.M114.615716
293. Zent R, Yan X, Su Y, Hudson BG, Borza DB, Moeckel GW, et al. Glomerular injury is exacerbated in diabetic integrin alpha1-null mice. *Kidney Int.* (2006) 70:460–70. doi: 10.1038/sj.ki.5000359
294. Chen X, Su Y, Fingleton B, Acuff H, Matrisian LM, Zent R, et al. An orthotopic model of lung cancer to analyze primary and metastatic NSCLC growth in integrin alpha1-null mice. *Clin Exp Metastasis.* (2005) 22:185–93. doi: 10.1007/s10585-005-7453-8
295. Macias-Perez I, Borza C, Chen X, Yan X, Ibanez R, Mernaugh G, et al. Loss of integrin alpha1beta1 ameliorates Kras-induced lung cancer. *Cancer Res.* (2008) 68:6127–35. doi: 10.1158/0008-5472.CAN-08-1395

296. Boudjadi S, Bernatchez G, Senicourt B, Beausejour M, Vachon PH, Carrier JC, et al. Involvement of the integrin $\alpha 1 \beta 1$ in the progression of colorectal cancer. *Cancers*. (2017) 9:96. doi: 10.3390/cancers9080096
297. Pozzi A, Moberg PE, Miles LA, Wagner S, Soloway P, Gardner HA. Elevated matrix metalloprotease and angiotatin levels in integrin $\alpha 1$ knockout mice cause reduced tumor vascularization. *Proc Natl Acad Sci USA*. (2000) 97:2202–7. doi: 10.1073/pnas.040378497
298. Schadendorf D, Fichtner I, Makki A, Alijagic S, Kupper M, Mrowietz U, et al. Metastatic potential of human melanoma cells in nude mice—characterisation of phenotype, cytokine secretion and tumour-associated antigens. *Br J Cancer*. (1996) 74:194–9. doi: 10.1038/bjc.1996.337
299. Schadendorf D, Gawlik C, Haney U, Ostmeier H, Suter L, Czarnetzki BM. Tumour progression and metastatic behaviour *in vivo* correlates with integrin expression on melanocytic tumours. *J Pathol*. (1993) 170:429–34. doi: 10.1002/path.1711700405
300. Ratzinger S, Grassel S, Dowejko A, Reichert TE, Bauer RJ. Induction of type XVI collagen expression facilitates proliferation of oral cancer cells. *Matrix Biol*. (2011) 30:118–25. doi: 10.1016/j.matbio.2011.01.001
301. Koukoulis GK, Warren WH, Virtanen I, Gould VE. Immunolocalization of integrins in the normal lung and in pulmonary carcinomas. *Hum Pathol*. (1997) 28:1018–25. doi: 10.1016/S0046-8177(97)90054-X
302. Fukuda K, Saikawa Y, Yagi H, Wada N, Takahashi T, Kitagawa Y. Role of integrin $\alpha 1$ subunits in gastric cancer patients with peritoneal dissemination. *Mol Med Rep*. (2012) 5:336–40. doi: 10.3892/mmr.2011.642
303. Wu JE, Santoro SA. Complex patterns of expression suggest extensive roles for the $\alpha 2 \beta 1$ integrin in murine development. *Dev Dyn*. (1994) 199:292–314. doi: 10.1002/aja.1001990405
304. Zutter MM, Santoro SA. Widespread histologic distribution of the $\alpha 2 \beta 1$ integrin cell-surface collagen receptor. *AJPA*. (1990) 137:113–20.
305. Holtkotter O, Nieswandt B, Smyth N, Muller W, Hafner M, Schulte V, et al. Integrin $\alpha 2$ -deficient mice develop normally, are fertile, but display partially defective platelet interaction with collagen. *J Biol Chem*. (2002) 277:10789–94. doi: 10.1074/jbc.M112307200
306. Chen J, Diacovo TG, Grenache DG, Santoro SA, Zutter MM. The $\alpha 2$ integrin subunit-deficient mouse: a multifaceted phenotype including defects of branching morphogenesis and hemostasis. *AJPA*. (2002) 161:337–44. doi: 10.1016/S0002-9440(10)64185-5
307. Barczyk M, Carracedo S, Gullberg D. Integrins. *Cell Tissue Res*. (2010) 339:269–80. doi: 10.1007/s00441-009-0834-6
308. Stange R, Kronenberg D, Timmen M, Everding J, Hidding H, Eckes B, et al. Age-related bone deterioration is diminished by disrupted collagen sensing in integrin $\alpha 2 \beta 1$ deficient mice. *Bone*. (2013) 56:48–54. doi: 10.1016/j.bone.2013.05.003
309. Zweers MC, Davidson JM, Pozzi A, Hallinger R, Janz K, Quondamatteo F, et al. Integrin $\alpha 2 \beta 1$ is required for regulation of murine wound angiogenesis but is dispensable for reepithelialization. *J Invest Dermatol*. (2007) 127:467–78. doi: 10.1038/sj.jid.5700546
310. Borza CM, Su Y, Chen X, Yu L, Mont S, Chetyrkin S, et al. Inhibition of integrin $\alpha 2 \beta 1$ ameliorates glomerular injury. *J Am Soc Nephrol*. (2012) 23:1027–38. doi: 10.1681/ASN.2011040367
311. Peters MA, Wendholt D, Strietholt S, Frank S, Pundt N, Korb-Pap A, et al. The loss of $\alpha 2 \beta 1$ integrin suppresses joint inflammation and cartilage destruction in mouse models of rheumatoid arthritis. *Arthritis Rheum*. (2012) 64:1359–68. doi: 10.1002/art.33487
312. Ramirez NE, Zhang Z, Madamanchi A, Boyd KL, O'Rear LD, Nashabi A, et al. The $\alpha 2 \beta 1$ integrin is a metastasis suppressor in mouse models and human cancer. *J Clin Invest*. (2011) 121:226–37. doi: 10.1172/JCI42328
313. Zuo XX, Yang Y, Zhang Y, Zhang ZG, Wang XF, Shi YG. Platelets promote breast cancer cell MCF-7 metastasis by direct interaction: surface integrin $\alpha 2 \beta 1$ -contacting-mediated activation of Wnt-beta-catenin pathway. *Cell Commun Signal*. (2019) 17:142. doi: 10.1186/s12964-019-0464-x
314. Ibaragi S, Shimo T, Hassan NM, Isowa S, Kurio N, Mandai H, et al. Induction of MMP-13 expression in bone-metastasizing cancer cells by type I collagen through integrin $\alpha 1 \beta 1$ and $\alpha 2 \beta 1$ -p38 MAPK signaling. *Anticancer Res*. (2011) 31:1307–13.
315. Siret C, Terciolo C, Dobric A, Habib MC, Germain S, Bonnier R, et al. Interplay between cadherins and $\alpha 2 \beta 1$ integrin differentially regulates melanoma cell invasion. *Br J Cancer*. (2015) 113:1445–53. doi: 10.1038/bjc.2015.358
316. Sottnik JL, Daignault-Newton S, Zhang X, Morrissey C, Hussain MH, Keller ET, et al. Integrin $\alpha 2 \beta 1$ ($\alpha 2 \beta 1$) promotes prostate cancer skeletal metastasis. *Clin Exp Metastasis*. (2013) 30:569–78. doi: 10.1007/s10585-012-9561-6
317. Naci D, El Azreq MA, Chetoui N, Lauden L, Sigaux F, Charron D, et al. $\alpha 2 \beta 1$ integrin promotes chemoresistance against doxorubicin in cancer cells through extracellular signal-regulated kinase (ERK). *J Biol Chem*. (2012) 287:17065–76. doi: 10.1074/jbc.M112.349365
318. Naci D, Berrazouane S, Barabe F, Aoudjit F. Cell adhesion to collagen promotes leukemia resistance to doxorubicin by reducing DNA damage through the inhibition of Rac1 activation. *Sci Rep*. (2019) 9:19455. doi: 10.1038/s41598-019-55934-w
319. Naci D, Vuori K, Aoudjit F. $\alpha 2 \beta 1$ integrin in cancer development and chemoresistance. *Semin Cancer Biol*. (2015) 35:145–53. doi: 10.1016/j.semcancer.2015.08.004
320. Varas L, Ohlsson LB, Honeth G, Olsson A, Bengtsson T, Wiberg C, et al. $\alpha 10$ integrin expression is up-regulated on fibroblast growth factor-2-treated mesenchymal stem cells with improved chondrogenic differentiation potential. *Stem Cells Dev*. (2007) 16:965–78. doi: 10.1089/scd.2007.0049
321. Lundgren-Akerlund E, Aszodi A. Integrin $\alpha 10 \beta 1$: a collagen receptor critical in skeletal development. *Adv Exp Med Biol*. (2014) 819:61–71. doi: 10.1007/978-94-017-9153-3_4
322. Kyostila K, Lappalainen AK, Lohi H. Canine chondrodysplasia caused by a truncating mutation in collagen-binding integrin α subunit 10. *PLoS ONE*. (2013) 8:e75621. doi: 10.1371/journal.pone.0075621
323. Wenke AK, Kjellman C, Lundgren-Akerlund E, Uhlmann C, Haass NK, Herlyn M, et al. Expression of integrin $\alpha 10$ is induced in malignant melanoma. *Cell Oncol*. (2007) 29:373–86. doi: 10.1097/00008390-200609001-00192
324. Munksgaard Thoren M, Chmielarska Masoumi K, Krona C, Huang X, Kundu S, Schmidt L, et al. Integrin $\alpha 10$, a novel therapeutic target in glioblastoma, regulates cell migration, proliferation, and survival. *Cancers*. (2019) 11:587. doi: 10.3390/cancers11040587
325. Okada T, Lee AY, Qin LX, Agaram N, Mimae T, Shen Y, et al. Integrin- $\alpha 10$ dependency identifies RAC and RICTOR as therapeutic targets in high-grade myxofibrosarcoma. *Cancer Discov*. (2016) 6:1148–65. doi: 10.1158/2159-8290.CD-15-1481
326. Carracedo S, Lu N, Popova SN, Jonsson R, Eckes B, Gullberg D. The fibroblast integrin $\alpha 11 \beta 1$ is induced in a mechanosensitive manner involving actin A and regulates myofibroblast differentiation. *J Biol Chem*. (2010) 285:10434–45. doi: 10.1074/jbc.M109.078766
327. Popova SN, Rodriguez-Sanchez B, Liden A, Betsholtz C, Van Den Bos T, Gullberg D. The mesenchymal $\alpha 11 \beta 1$ integrin attenuates PDGF-BB-stimulated chemotaxis of embryonic fibroblasts on collagens. *Dev Biol*. (2004) 270:427–42. doi: 10.1016/j.ydbio.2004.03.006
328. Zhu C-Q, Popova SN, Brown ER, Barsyte-Lovejoy D, Navab R, Shih W, et al. Integrin $\alpha 11$ regulates IGF2 expression in fibroblasts to enhance tumorigenicity of human non-small-cell lung cancer cells. *Proc Natl Acad Sci USA*. (2007) 104:11754–9. doi: 10.1073/pnas.0703040104
329. Popova SN, Barczyk M, Tiger CF, Beertsen W, Zigrino P, Aszodi A, et al. $\alpha 11 \beta 1$ integrin-dependent regulation of periodontal ligament function in the erupting mouse incisor. *Mol Cell Biol*. (2007) 27:4306–16. doi: 10.1128/MCB.00041-07
330. Blumbach K, Niehoff A, Belgardt BF, Ehlen HW, Schmitz M, Hallinger R, et al. Dwarfism in mice lacking collagen-binding integrins $\alpha 2 \beta 1$ and $\alpha 11 \beta 1$ is caused by severely diminished IGF-1 levels. *J Biol Chem*. (2012) 287:6431–40. doi: 10.1074/jbc.M111.283119
331. Romaine A, Sorensen IW, Zeltz C, Lu N, Erusappan PM, Melleby AO, et al. Overexpression of integrin $\alpha 11$ induces cardiac fibrosis in mice. *Acta Physiol*. (2018) 222. doi: 10.1111/apha.12932
332. Navab R, Strumpf D, To C, Pasko E, Kim KS, Park CJ, et al. Integrin $\alpha 11 \beta 1$ regulates cancer stromal stiffness and promotes tumorigenicity and metastasis in non-small cell lung cancer. *Oncogene*. (2016) 35:1899–908. doi: 10.1038/ncr.2015.254
333. Ando T, Kage H, Matsumoto Y, Zokumasu K, Yotsumoto T, Maemura K, et al. Integrin $\alpha 11$ in non-small cell lung cancer is associated

- with tumor progression and postoperative recurrence. *Cancer Sci.* (2020) 111:200–8. doi: 10.1111/cas.14257
334. Smeland HY, Askeland C, Wik E, Knutsvik G, Molven A, Edelman RJ, et al. Integrin alpha11beta1 is expressed in breast cancer stroma and associates with aggressive tumor phenotypes. *J Pathol Clin Res.* (2019) 6:69–82. doi: 10.1002/cjp2.148
335. Smeland HY, Lu N, Karlens T V, Salvesen G, Reed RK, Stuhr L. Stromal integrin alpha11-deficiency reduces interstitial fluid pressure and perturbs collagen structure in triple-negative breast xenograft tumors. *BMC Cancer.* (2019) 19:234. doi: 10.1186/s12885-019-5449-z
336. Boudjadi S, Carrier JC, Beaulieu JF. Integrin alpha1 subunit is up-regulated in colorectal cancer. *Biomark Res.* (2013) 1:16. doi: 10.1186/2050-7771-1-16
337. Boudjadi S, Carrier JC, Groulx JF, Beaulieu JF. Integrin alpha1beta1 expression is controlled by c-MYC in colorectal cancer cells. *Oncogene.* (2016) 35:1671–8. doi: 10.1038/onc.2015.231
338. Benya PD, Padilla SR. Modulation of the rabbit chondrocyte phenotype by retinoic acid terminates type II collagen synthesis without inducing type I collagen: the modulated phenotype differs from that produced by subculture. *Dev Biol.* (1986) 118:296–305. doi: 10.1016/0012-1606(86)90096-5
339. Bengtsson T, Aszodi A, Nicolae C, Hunziker EB, Lundgren-Akerlund E, Fassler R. Loss of alpha10beta1 integrin expression leads to moderate dysfunction of growth plate chondrocytes. *J Cell Sci.* (2005) 118:929–36. doi: 10.1242/jcs.01678
340. Velling T, Kusche-Gullberg M, Sejersen T, Gullberg D. cDNA cloning and chromosomal localization of human alpha(11) integrin. A collagen-binding, I domain-containing, beta(1)-associated integrin alpha-chain present in muscle tissues. *J Biol Chem.* (1999) 274:25735–42. doi: 10.1074/jbc.274.36.25735
341. Lu N, Carracedo S, Ranta J, Heuchel R, Soiminen R, Gullberg D. The human alpha11 integrin promoter drives fibroblast-restricted expression *in vivo* and is regulated by TGF-beta1 in a Smad- and Sp1-dependent manner. *Matrix Biol.* (2010) 29:166–76. doi: 10.1016/j.matbio.2009.11.003
342. Erusappan P, Alam J, Lu N, Zeltz C, Gullberg D. Integrin alpha11 cytoplasmic tail is required for FAK activation to initiate 3D cell invasion and ERK-mediated cell proliferation. *Sci Rep.* (2019) 9:15283. doi: 10.1038/s41598-019-51689-6
343. Rombouts K, Carloni V. The fibrotic microenvironment as a heterogeneity facet of hepatocellular carcinoma. *Fibrogenes Tissue Repair.* (2013) 6:17. doi: 10.1186/1755-1536-6-17
344. Banys-Paluchowski M, Loibl S, Witzel I, Mundhenke C, Lederer B, Solbach C, et al. Clinical relevance of collagen protein degradation markers C3M and C4M in the serum of breast cancer patients treated with neoadjuvant therapy in the geparquinto trial. *Cancers.* (2019) 11:1186. doi: 10.1055/s-0039-1687939
345. Willumsen N, Ali SM, Leitzel K, Drabick JJ, Yee N, Polimera HV, et al. Collagen fragments quantified in serum as measures of desmoplasia associate with survival outcome in patients with advanced pancreatic cancer. *Sci Rep.* (2019) 9:19761. doi: 10.1038/s41598-019-56268-3
346. Lipton A, Leitzel K, Ali SM, Polimera H V., Nagabhairu V, Marks E, et al. High turnover of extracellular matrix reflected by specific protein fragments measured in serum is associated with poor outcomes in two metastatic breast cancer cohorts. *Int J Cancer.* (2018) 143:3027–34. doi: 10.1002/ijc.31627

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Bourgot, Primac, Louis, Noël and Maquoi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.