



# Could the eIF2 $\alpha$ -Independent Translation Be the Achilles Heel of Cancer?

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Eukaryotic initiation factor eIF2 is a key component of the ternary complex whose role is to deliver initiator tRNA into the ribosome. A variety of stimuli, both physiologic and pathophysiologic activate eIF2 kinases that phosphorylate the  $\alpha$  subunit of eIF2, preventing it from forming the ternary complex, thus attenuating cellular protein synthesis. Paradoxically, in cancer cells, the phosphorylation of eIF2 $\alpha$  is associated with activation of survival pathways. This review explores the recently emerged novel mechanism of eIF2 $\alpha$ -independent translation initiation. This mechanism, which appears to be shared by some RNA viruses and Internal Ribosome Entry Site-containing cellular mRNAs and utilizes auxiliary proteins, such as eIF5B, eIF2D, and MCT-1, is responsible for the selective translation of cancer-associated genes and could represent a weak point amenable to specific targeting for the treatment of cancer.

**Keywords:** eIF2 $\alpha$ , selective translation, IRES, stress, oncogene, apoptosis

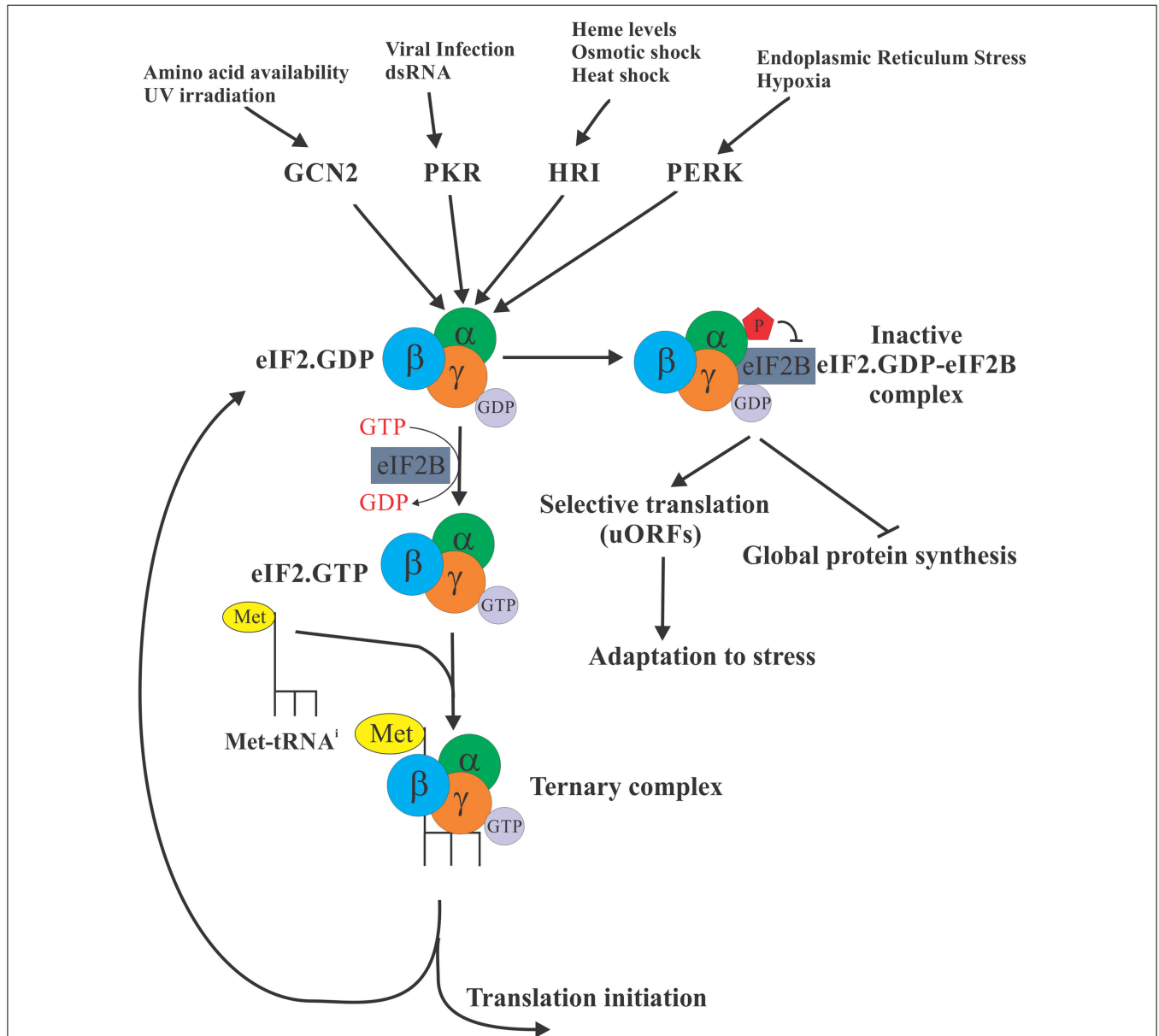
## INTRODUCTION

Regulation of protein synthesis (translation) is a key cellular process, which underpins cellular survival. Traditionally, the transcription and translation of the genome was considered to be a highly correlated phenomenon, this notion has been challenged by the demonstration that mRNA transcriptional outputs correlate with only about 40% of the total protein content in a cell (1, 2). This disconnect is further accentuated upon e.g., epidermal growth factor stimulation where up to 90% of transcripts exhibited uncoupled translation from transcription (2). Interestingly, these studies also showed that highly expressed transcripts are sometimes poorly translated and, *vice versa*, poorly transcribed genes can be translated efficiently. This suggests that transcription and translation are largely independent of each other, further strengthening the notion that translational control has a major impact on regulating the proteome under certain conditions. It is therefore not surprising that many aberrant cellular processes require modification of the translation machinery and translation output (i.e., proteome). This is often the case in various stress responses and diseases and is probably best exemplified in cancer. The link between misregulated translation and cancer has long been suspected, and strong experimental evidence has been obtained by many laboratories in recent years. This review will focus on one specific mechanism of translational dysregulation in oncogenesis, which is the ability of cancer cells to utilize an eIF2 $\alpha$ -independent mode of translation initiation.

## eIF2 IS THE NEXUS THAT CONTROLS TRANSLATION INITIATION EFFICIENCY

The eukaryotic initiation factor 2 (eIF2) is required for the formation of the ternary complex [consisting of eIF2 ( $\alpha$ ,  $\beta$ , and  $\gamma$  subunits), Met-tRNA<sub>i</sub>, and GTP], which in turn brings the initiator Met-tRNA<sub>i</sub> to the P site of the 40S ribosomal subunit, enabling initiation of protein synthesis. eIF2 exists in two distinct configurations – the GDP- or GTP-bound forms. Following the recognition of the start codon by the 43S preinitiation complex,

the GTP bound to eIF2 is hydrolyzed to deliver Met-tRNA<sub>i</sub> and the eIF2. GDP is subsequently released from the 40S subunit to be recycled for the delivery of another Met-tRNA<sub>i</sub> [reviewed in Ref. (3)]. The GDP to GTP exchange is catalyzed by the eIF2B (the guanine exchange factor). This step is necessary for the regeneration of active eIF2, which is the only form of eIF2 that can bind Met-tRNA<sub>i</sub>. Regulation of the GDP–GTP exchange, thus, provides the cell with a control point to match the protein synthesis rates to the physiologic requirements (Figure 1). In response to virtually all stresses, the  $\alpha$  subunit of eIF2 is phosphorylated at serine 51,



**FIGURE 1 | eIF2 $\alpha$  is a control nexus integrating diverse stimuli to regulate translation.** The eukaryotic initiation factor 2 (eIF2) is required for the formation of the ternary complex [consisting of eIF2 ( $\alpha$ ,  $\beta$ , and  $\gamma$  subunits), Met-tRNA<sub>i</sub>, and GTP] whose role is to deliver the initiator Met-tRNA<sub>i</sub> to the P site of the 40S ribosomal subunit, enabling translation initiation. eIF2 exists in two distinct configurations – the GDP bound (inactive) and GTP bound (active), but only the GTP-bound form can bind Met-tRNA<sub>i</sub>. In response to diverse stress stimuli, distinct protein kinases phosphorylate the  $\alpha$  subunit of eIF2 at serine 51, ultimately locking eIF2 in an inactive eIF2 complex with eIF2B. As a result, availability of the ternary complex significantly declines ultimately resulting in the inhibition of translation globally. Selective translation of a subset of mRNAs with uORFs continues, however, allowing cells to express proteins required for adaptation to stress conditions.

which greatly increases its avidity for eIF2B and locks both proteins in an inactive eIF2–eIF2B complex. Since eIF2B is present in limiting concentrations, phosphorylation of even a small fraction of eIF2 $\alpha$  is sufficient to inhibit its recycling. As a result, availability of the ternary complex significantly declines leading to inhibition of 43S formation and attenuation of translation initiation (4).

In mammalian cells, there are four eIF2 $\alpha$  kinases that integrate a variety of distinct stresses to phosphorylate eIF2 $\alpha$ . Thus, haem-regulated inhibitor (HRI) is activated by iron deficiency (5), heavy metals (6), osmotic, or oxidative stress and heat shock (7); protein kinase activated by double-stranded RNA (PKR) is activated by dsRNA during viral infection (8) or interferon-induced apoptosis (9); general control non-derepressible-2 (GCN2) is activated by amino acid starvation (10) and UV irradiation (11); and PKR-like endoplasmic reticulum kinase (PERK) is activated in response to endoplasmic reticulum (ER) stress (12) and UV irradiation (13). Since the ternary complex is absolutely required for the translation of cellular mRNAs, the phosphorylation of eIF2 $\alpha$  is expected to inhibit translation globally. However, for certain mRNAs, phosphorylation of eIF2 $\alpha$  enhances their translation. These mRNAs frequently encode for proteins required for stress adaptation and recovery [reviewed in Ref. (14)]. These mRNAs, best exemplified by the transcription factor ATF4, contain short upstream ORFs (uORFs) in their 5' untranslated regions (UTRs) that engage the ribosome and thus cause the main coding ORF of these mRNAs to be poorly translated when ternary complex is abundant. In contrast, during stress conditions when the abundance of ternary complex is limited, some ribosomes pass through the uORFs without initiating until they acquire a ternary complex upstream of the main coding ORF, resulting in productive translation of the main ORF. It is expected, however, that under conditions of severe stress if no ternary complex is available even translation of these mRNAs would be inhibited.

## EVIDENCE FOR eIF2 $\alpha$ -INDEPENDENT TRANSLATION

In the last decade or so, evidence has begun to emerge pointing to the existence of an eIF2 $\alpha$ -independent initiation of translation. It was observed in several independent studies that phosphorylation of eIF2 $\alpha$  is frequently required to promote cell survival under various stresses, but the genes lying downstream of eIF2 $\alpha$  regulation did not necessarily utilize the uORF-dependent mechanism. For example, following nutritional stress caused by amino acid starvation, induction of amino acid transporter SNAT2, a component of transport system A, which is required to promote recovery of amino acid balance, was blunted in cells unable to phosphorylate eIF2 $\alpha$  (15). Similarly, translation of the inhibitor of apoptosis proteins XIAP and cIAP1 was shown to be resistant to the global attenuation of translation due to eIF2 $\alpha$  phosphorylation during hypoxia (16, 17), glucose deficiency (18), serum deprivation (19–21), or ER stress (22). Other examples of mRNAs whose translation is enhanced by eIF2 $\alpha$  phosphorylation include VEGF and p120 catenin [observed in hypoxic inflammatory breast tumors (23, 24)], Bcl-xL [during hypertonic stress (25)], or c-Src and c-myc [during ER stress (26, 27)]. Interestingly, none of these mRNAs harbor uORFs that would enable enhanced mRNA

translation under conditions of reduced ternary complex availability. Instead, Internal Ribosome Entry Site (IRES) sequences were described in the 5' UTRs of these mRNAs. IRESs are discrete functional RNA motifs that support translation of the downstream reading frame by an alternative mechanism that does not appear to rely on m<sup>7</sup>G cap structure or initiation factors that recognize the cap. This phenomenon, termed internal initiation, was first observed in RNA viruses (in particular *Picornaviridae*) whose RNA is naturally uncapped, and yet, it is efficiently translated by the host translation machinery (28). Instead of cap, IRES precede the protein-coding portion of the viral RNA and directly recruit the 40S ribosome to the vicinity of initiation codon. The recruitment of the 40S can occur either in the complete absence of any other protein factors (dicistrovirus intergenic IRES) or with the aid of various combinations of canonical initiation factors (such as eIF4G and eIF3) and auxiliary proteins in case of the remaining viral IRES [reviewed in Ref. (29)].

Similarly to the internal initiation in viruses, some cellular mRNAs were shown to be translated by this alternative mode of translation. It was observed that a subset of cellular mRNAs (up to 5%) was efficiently translated in cells infected with poliovirus which cleaves eIF4G and thus inhibits cap-dependent translation (30). Follow-up studies identified functional IRES in a variety of cellular mRNAs, and many cellular IRES elements were shown to be *bona fide* regulatory elements that direct translation of their respective proteins in a cap- and/or eIF4E-independent manner during various cellular stresses or pathological conditions, such as tumorigenesis, in particular. It is, therefore, not surprising that many of the IRES-harboring mRNAs encode proteins that are key regulators of cell proliferation and survival/apoptosis and their misregulated expression is frequently associated with cancer. Thus, we and others have proposed that the selective regulation of IRES-mediated translation is important for the regulation of cell death and survival (14, 31–34). Indeed, the experimental data from many laboratories have now validated this hypothesis in a number of models [reviewed in Ref. (34, 35)]. The *in vivo* evidence supporting this concept, and particularly in the context of cancer formation, is particularly striking. For example, the selective impairment of IRES-mediated translation but not cap-dependent translation is correlated with enhanced apoptosis of hematopoietic progenitors and stem cells, leading to the fatal progressive bone marrow failure syndrome known as *dyskeratosis congenita* and its associated tumorigenesis (36–38). A switch to IRES-dependent translation has been implicated in breast cancer growth and angiogenic potential *in vivo* (39), and in the acquired resistance of cancer cells to radiation-induced apoptosis (40). Similarly, HPV-induced transformation of human keratinocytes is accompanied by a switch from cap-dependent to IRES-dependent translation (41).

## MECHANISMS OF eIF2 $\alpha$ -INDEPENDENT TRANSLATION

The mechanism of how translation initiation can occur without the eIF2 was first described in viruses that utilize IRES mechanism of translation initiation, and whose translation has been reported to be refractory to reduced availability of ternary complex. The

simplest model is the CrPV virus, in which the secondary and tertiary structures of the intergenic region IRES mimic the function of a peptidyl tRNA (Met-tRNA<sub>i</sub>) in the P site, recruit 40S and 60S ribosomal subunits to form 80S, and direct the synthesis of viral polyprotein by inducing ribosomal translocation (42). Hepatitis C virus (HCV) and classical swine fever virus (CSFV) were both shown to use eIF5B instead of eIF2 to deliver initiator tRNA into the P site of the ribosome (43–45). The canonical role of eIF5B is in ribosome subunit joining, and it does not normally promote binding of initiator tRNA to the 40S subunit. However, it appears to “take over” when eIF2 is inactivated due to phosphorylation. In addition to eIF5B, other cellular proteins including eIF2D (formerly called Ligatin) and MCT-1/DENR were all reported to be able to deliver tRNA into the P site of the ribosome during HCV IRES-mediated translation (46, 47). Their role in supporting cellular mRNA translation during stress has not yet been fully elucidated.

Evidence is starting to emerge that a similar eIF5B-dependent mechanism operates on cellular mRNAs as well. We have investigated the molecular mechanism of eIF2 $\alpha$ -independent translation of XIAP mRNA (21). XIAP protein is encoded by two mRNA splice variants that differ only in their 5' UTR regions. The more abundant, shorter transcript produces the majority of XIAP protein under normal growth conditions by cap-dependent translation. However, during cellular stress, the longer transcript, containing the IRES element, directs efficient translation despite attenuation of global, cap-dependent translation (20). During normal proliferative conditions, when the ternary complex is available in abundance, XIAP translation continues in an eIF2 $\alpha$ -dependent mode, similar to other cellular mRNAs. However, upon serum deprivation, the XIAP IRES-dependent translation switches to an alternative, eIF5B-dependent mode to circumvent attenuation due to eIF2 $\alpha$  phosphorylation. The ability of the cell to evade ternary complex requirement suggests that cells have evolved an alternative, eIF2 $\alpha$ -independent mechanism of tRNA delivery to support a “rescue” mechanism of translation of critical survival proteins under conditions when the “normal” mechanism is not available. Interestingly, a limited investigation of other cellular IRES-containing mRNAs (Bcl-xL, cIAP1, Apaf-1, and p97/DAP5) suggests that not all cellular IRESs utilize eIF5B-dependent mode of tRNA delivery during serum deprivation (21). However, the full spectrum of eIF5B-dependent cellular mRNA transcripts still needs to be determined.

Multiple copies in T-cell lymphoma-1 (MCT-1; also called MCST1, malignant T-cell amplified sequence 1) is an oncogene, which is also implicated in the regulation of cellular mRNA translation, although the precise mechanism of this control is not clear (48). MCT-1 recruits DENR (density regulated protein) to aid in the post-termination ribosome recycling (49). In addition, DENR/MCT-1 complex interacts with the cap complex and enhances translation of a subset of mRNAs whose products are involved in the regulation of apoptosis and the cell cycle. Of note, some of these mRNAs, such as cyclin D1, HSP70, and cIAP1, were previously reported to be translated by an IRES-dependent mechanism (22, 50, 51). Recently, a different cohort of specific mRNAs that require DENR/MCT-1 complex for efficient translation was identified by polysome

profiling experiments in DENR-knockdown *Drosophila* cells (48). Interestingly, these mRNAs, enriched for transcriptional regulators and oncogenic kinases, were characterized by the presence of uORFs with strong Kozak sequence. However, further experiments revealed that DENR/MCT-1 promotes reinitiation of translation on these mRNAs, so the mechanism of translation upregulation by DENR/MCT-1 is distinct from the classical uORF-dependent mode that is used by ATF4. Whether or not DENR/MCT-1 can also function in an alternative ternary complex formation in order to deliver initiator tRNA under conditions of cell stress has not been tested.

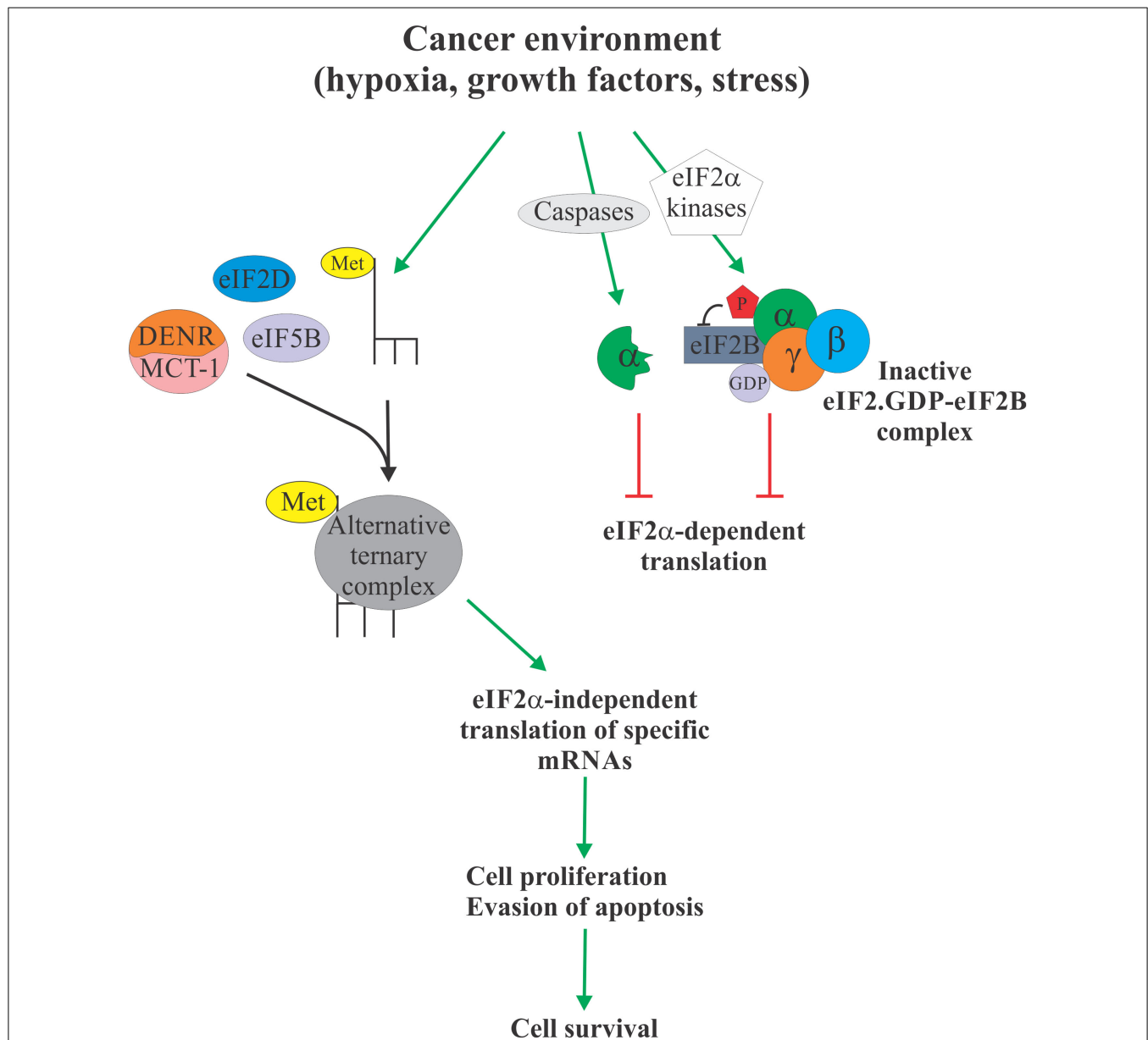
## eIF2 $\alpha$ -INDEPENDENT TRANSLATION AND CANCER

Phosphorylation of eIF2 $\alpha$  serves to integrate diverse stress signals, both intra- and extracellular, and for both physiologic and pathophysiologic conditions. The role of eIF2 $\alpha$  in tumorigenesis has been recently summarized (3). Studies in mouse models and human tumors suggest that targeted disruption of eIF2 $\alpha$  phosphorylation could be exploited to increase the efficacy of therapeutic intervention. In addition, initiation of apoptosis (the main mechanism of action of chemotherapy drugs) results in the caspase-mediated cleavage of eIF2 $\alpha$  (52–54), suggesting that inhibition of the ternary complex formation may be part of the translational reprogramming accompanying apoptosis.

It is, therefore, important to explore the possibility that tumor cells could gain a selective growth advantage and evade apoptosis by utilizing an eIF2 $\alpha$ -independent mode of translation initiation of cancer-related mRNAs, such as XIAP, cIAP1, cyclin D1, and others. As noted above, MCT-1 is an oncogene involved in the regulation of cell cycle and apoptosis. Expression of MCT-1 transforms NIH3T3 and MCF-10A cells (55, 56) and promotes aggressive growth, angiogenesis, tissue invasiveness, and inhibition of apoptosis in breast (57) and lymphoma (58) tumor models. Furthermore, high expression of MCT-1 was reported in 83.9% of human lung carcinomas (59). MCT-1 acts in conjunction with DENR; however, there is no information available on the expression and role of DENR in tumorigenesis yet. A quick survey of the antibody staining data in The Human Protein Atlas<sup>1</sup> showed that DENR is detected in 74% of the cancers, but the discordantly high expression relative to normal tissue is most noticeable in gliomas, lymphomas, melanoma, skin, and urothelial cancers. Of note, expression of MCT-1 is also particularly high in glioma, melanoma, and urothelial cancers (see text footnote 1), suggesting that the concomitant expression of DENR and MCT-1 in these particular cancers may lead to translational activation of specific mRNAs. This notion, however, awaits experimental validation.

There is no available literature on the possible involvement of eIF2D in cancer with the exception of eIF2D being identified as a hepatocellular carcinoma (HCC) – associated antigen in a study of four HCC patients (60). A survey of The Human Protein Atlas shows increased expression in some head and neck, lymphoma, melanoma, ovarian, pancreatic, stomach, and urothelial cancers

<sup>1</sup><http://www.proteinatlas.org>



**FIGURE 2 | eIF2 $\alpha$ -independent translation in cancer.** Cancer environment, such as hypoxia, growth factor and cytokine signaling, and various forms of stress, leads to enhanced phosphorylation or proteolytic degradation of eIF2 $\alpha$  which causes attenuation of protein synthesis due to unavailability of the ternary complex (right). However, this allows Met-tRNA<sub>i</sub> to form an alternative ternary complex with proteins like eIF5B, eIF2D, and DENR/MCT-1 (left). Alternative ternary complex supports translation of select mRNAs, including those regulating apoptosis and cell cycle progression, in an eIF2 $\alpha$ -independent manner, ultimately providing the cancer cell with a survival advantage. Specific targeting of the eIF2 $\alpha$ -independent translation mechanism therefore offers a possible therapeutic target to treat cancer.

(see text footnote 1). Interestingly, eIF2D belongs to a family of PUA (pseudouridine synthase and archaeosine transglycosylase) domain containing proteins, along with DENR and MCT-1 (61). Another notable member of this family is dyskerin (DKC1), which has been shown to play a key role in enabling translation of cancer-related IRES-containing mRNAs, such as Bcl-xL, XIAP (36), p27 (38), and p53 (37). DKC1 deficiency results in the decrease in modification of ribosomal RNA uridines, which consequently renders the ribosome unable to efficiently engage select mRNAs for translation (62). An intriguing possibility, which ought to be experimentally investigated is whether, like DENR/MCT-1

or eIF2D, DKC1 can also participate in the formation of an alternative ternary complex for these mRNAs.

eIF5B was shown to control cell cycle transition and developmental stages during oocytes maturation, by inhibiting cell cycle arrest (63). Interestingly, this study also demonstrated that both the levels of eIF5B and the association of eIF5B with Met-tRNA<sub>i</sub> were increased during serum starvation of oocytes, ES cells, or in proliferating T cells. This was accompanied by increased phosphorylation of eIF2 $\alpha$ , suggesting that under these conditions eIF5B may preferentially enhance translation of specific mRNAs. In addition, depletion of eIF5B impaired translation in mature,

but not proliferating cells, further supporting a stage-specific role for eIF5B. Given the essential role of eIF5B in general translation, it is expressed in all normal tissues; however, there is an enhanced expression in some melanoma and ovarian cancers (see text footnote 1). Of note, the majority of the surveyed cancers show reduced eIF5B staining, when compared to the expression of eIF5B in normal tissues, suggesting that a relationship between eIF5B and tumorigenesis is likely not straightforward.

## FUTURE PERSPECTIVES

Over the past two decades, eIF2 $\alpha$  has emerged as a critical nexus integrating diverse cellular and extracellular signals to control protein synthesis. More recently, the evidence is mounting to link eIF2 $\alpha$  phosphorylation to tumorigenesis [reviewed in Ref. (3)]. In particular, elevated levels of eIF2 $\alpha$  phosphorylation were shown to correlate with cancer cell survival. These observations are somewhat paradoxical, since cancer cells typically display elevated protein synthesis levels (64), which would not be compatible with elevated levels of eIF2 $\alpha$  phosphorylation. Phosphorylation of eIF2 $\alpha$ , therefore, likely serves to activate specific survival pathways, some of which may rely on the formation of an alternative, eIF2 $\alpha$ -independent ternary complexes to promote translation of cancer-specific mRNAs (Figure 2). These alternative pathways and their components, such as eIF5B, eIF2D, and DENR/MCT-1, could, therefore, represent as of yet unexplored Achilles heels of cancer. For example, we have shown that eIF5B promotes IRES-dependent translation of an inhibitor of apoptosis XIAP (21). Chemoresistance in several types of cancer cells has been

associated with increased expression of XIAP (65–69). Importantly, XIAP has been validated as a potent therapeutic target, and various anti-XIAP strategies for cancer treatment are currently being tested in clinical trials (70). As with many other tumor-specific targets, the specificity of the therapeutics intervention for tumor cells is desirable, since XIAP deficiency in non-cancerous cells can lead to disorders, such as X-linked lymphoproliferative disease (71, 72). Specific targeting of the eIF2 $\alpha$ -independent mode of translation of XIAP and other cancer-related mRNAs may provide this needed specificity.

## AUTHOR CONTRIBUTIONS

MH conceived and wrote the manuscript.

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## REFERENCES

- Schwanhauser B, Busse D, Li N, Dittmar G, Schuchhardt J, Wolf J, et al. Global quantification of mammalian gene expression control. *Nature* (2011) 473:337–42. doi:10.1038/nature10098
- Tebaldi T, Re A, Viero G, Pegoretti I, Passerini A, Blanzieri E, et al. Widespread uncoupling between transcriptome and translational variations after a stimulus in mammalian cells. *BMC Genomics* (2012) 13:220. doi:10.1186/1471-2164-13-220
- Koromilas AE. Roles of the translation initiation factor eIF2 $\alpha$  serine 51 phosphorylation in cancer formation and treatment. *Biochim Biophys Acta* (2015) 1849:871–80. doi:10.1016/j.bbagr.2014.12.007
- Ron D, Harding HP. *eIF2 $\alpha$  Phosphorylation in Cellular Stress Responses and Disease*. New York, NY: Cold Spring Harbor Laboratory Press (2007).
- Ranu RS, London IM. Regulation of protein synthesis in rabbit reticulocyte lysates: additional initiation factor required for formation of ternary complex (eIF-2.GTP.Met-tRNA<sup>f</sup>) and demonstration of inhibitory effect of heme-regulated protein kinase. *Proc Natl Acad Sci U S A* (1979) 76:1079–83. doi:10.1073/pnas.76.3.1079
- Matts RL, Schatz JR, Hurst R, Kagen R. Toxic heavy metal ions activate the heme-regulated eukaryotic initiation factor-2 alpha kinase by inhibiting the capacity of heme-supplemented reticulocyte lysates to reduce disulfide bonds. *J Biol Chem* (1991) 266:12695–702.
- Lu L, Han AP, Chen JJ. Translation initiation control by heme-regulated eukaryotic initiation factor 2 $\alpha$  kinase in erythroid cells under cytoplasmic stresses. *Mol Cell Biol* (2001) 21:7971–80. doi:10.1128/MCB.21.23.7971-7980.2001
- Taylor DR, Lee SB, Romano PR, Marshak DR, Hinnebusch AG, Esteban M, et al. Autophosphorylation sites participate in the activation of the double-stranded-RNA-activated protein kinase PKR. *Mol Cell Biol* (1996) 16:6295–302. doi:10.1128/MCB.16.11.6295
- Samuel CE. Mechanism of interferon action: phosphorylation of protein synthesis initiation factor eIF-2 in interferon-treated human cells by a ribosome-associated kinase processing site specificity similar to heme-regulated rabbit reticulocyte kinase. *Proc Natl Acad Sci U S A* (1979) 76:600–4. doi:10.1073/pnas.76.2.600
- Roussou I, Thireos G, Hauge BM. Transcriptional-translational regulatory circuit in *Saccharomyces cerevisiae* which involves the GCN4 transcriptional activator and the GCN2 protein kinase. *Mol Cell Biol* (1988) 8:2132–9. doi:10.1128/MCB.8.5.2132
- Jiang HY, Wek RC. GCN2 phosphorylation of eIF2 $\alpha$  activates NF-kappaB in response to UV irradiation. *Biochem J* (2005) 385:371–80. doi:10.1042/BJ20041164
- Harding HP, Zhang Y, Ron D. Protein translation and folding are coupled by an endoplasmic-reticulum-resident kinase. *Nature* (1999) 397:271–4. doi:10.1038/16729
- Wu S, Hu Y, Wang J-L, Chatterjee M, Shi Y, Kaufman RJ. Ultraviolet light inhibits translation through activation of the unfolded protein response kinase PERK in the lumen of the endoplasmic reticulum. *J Biol Chem* (2002) 277:18077–83. doi:10.1074/jbc.M110164200
- Holcik M, Sonenberg N. Translational control in stress and apoptosis. *Nat Rev Mol Cell Biol* (2005) 6:318–27. doi:10.1038/nrm1618
- Gaccioli F, Huang CC, Wang C, Bevilacqua E, Franchi-Gazzola R, Gazzola GC, et al. Amino acid starvation induces the SNAT2 neutral amino acid transporter by a mechanism that involves eukaryotic initiation factor 2 $\alpha$  phosphorylation and cap-independent translation. *J Biol Chem* (2006) 281:17929–40. doi:10.1074/jbc.M600341200
- Aird KM, Ding X, Baras A, Wei J, Morse MA, Clay T, et al. Trastuzumab signaling in ErbB2-overexpressing inflammatory breast cancer correlates with X-linked inhibitor of apoptosis protein expression. *Mol Cancer Ther* (2008) 7:38–47. doi:10.1158/1535-7163.mct-07-0370
- Aird KM, Ghanayem RB, Peplinski S, Lyerly HK, Devi GR. X-linked inhibitor of apoptosis protein inhibits apoptosis in inflammatory breast cancer cells with acquired resistance to an ErbB1/2 tyrosine kinase inhibitor. *Mol Cancer Ther* (2010) 9:1432–42. doi:10.1158/1535-7163.mct-10-0160

18. Muaddi H, Majumder M, Peidis P, Papadakis AI, Holcik M, Scheuner D, et al. Phosphorylation of eIF2 $\alpha$  at serine 51 is an important determinant of cell survival and adaptation to glucose deficiency. *Mol Biol Cell* (2010) **21**:3220–31. doi:10.1091/mbc.E10-01-0023
19. Holcik M, Lefebvre C, Yeh C, Chow T, Korneluk RG. A new internal-ribosome-entry-site motif potentiates XIAP-mediated cytoprotection. *Nat Cell Biol* (1999) **1**:190–2. doi:10.1038/11109
20. Riley A, Jordan LE, Holcik M. Distinct 5' UTRs regulate XIAP expression under normal growth conditions and during cellular stress. *Nucleic Acids Res* (2010) **38**:4665–74. doi:10.1093/nar/gkq241
21. Thakor N, Holcik M. IRES-mediated translation of cellular messenger RNA operates in eIF2 $\alpha$ -independent manner during stress. *Nucleic Acids Res* (2012) **40**:541–52. doi:10.1093/nar/gkr701
22. Warnakulasuriyarachchi D, Cerquozzi S, Cheung HH, Holcik M. Translational induction of the inhibitor of apoptosis protein HIAP2 during endoplasmic reticulum stress attenuates cell death and is mediated via an inducible internal ribosome entry site element. *J Biol Chem* (2004) **279**:17148–57. doi:10.1074/jbc.M308737200
23. Silvera D, Arju R, Darvishian F, Levine PH, Zolfaghari L, Goldberg J, et al. Essential role for eIF4G1 overexpression in the pathogenesis of inflammatory breast cancer. *Nat Cell Biol* (2009) **11**:903–8. doi:10.1038/ncb1900
24. Silvera D, Schneider RJ. Inflammatory breast cancer cells are constitutively adapted to hypoxia. *Cell Cycle* (2009) **8**:3091–6. doi:10.4161/cc.8.19.9637
25. Bevilacqua E, Wang X, Majumder M, Gaccioli F, Yuan CL, Wang C, et al. eIF2 $\alpha$  phosphorylation tips the balance to apoptosis during osmotic stress. *J Biol Chem* (2010) **285**:17098–111. doi:10.1074/jbc.M110.109439
26. Allam H, Ali N. Initiation factor eIF2-independent mode of c-Src mRNA translation occurs via an internal ribosome entry site. *J Biol Chem* (2010) **285**:5713–25. doi:10.1074/jbc.M109.029462
27. Shi Y, Yang Y, Hoang B, Bardeleben C, Holmes B, Gera J, et al. Therapeutic potential of targeting IRES-dependent c-myc translation in multiple myeloma cells during ER stress. *Oncogene* (2015) **11**:156. doi:10.1038/onc.2015.156
28. Pelletier J, Sonenberg N. Internal initiation of translation of eukaryotic mRNA directed by a sequence derived from poliovirus RNA. *Nature* (1988) **334**:320–5. doi:10.1038/334320a0
29. Jackson RJ. The current status of vertebrate cellular mRNA IRESs. *Cold Spring Harb Perspect Biol* (2013) **5**:a011569. doi:10.1101/cshperspect.a011569
30. Johannes G, Carter MS, Eisen MB, Brown PO, Sarnow P. Identification of eukaryotic mRNAs that are translated at reduced cap binding complex eIF4F concentrations using a cDNA microarray. *Proc Natl Acad Sci U S A* (1999) **96**:13118–23. doi:10.1073/pnas.96.23.13118
31. Sachs AB, Sarnow P, Hentze MW. Starting at the beginning, middle, and end: translation initiation in eukaryotes. *Cell* (1997) **89**:831–8. doi:10.1016/S0092-8674(00)80268-8
32. Holcik M, Sonenberg N, Korneluk RG. Internal ribosome initiation of translation and the control of cell death. *Trends Genet* (2000) **16**:469–73. doi:10.1016/S0168-9525(00)02106-5
33. Lewis SM, Holcik M. IRES in distress: translational regulation of the inhibitor of apoptosis proteins XIAP and HIAP2 during cell stress. *Cell Death Differ* (2005) **12**:547–53. doi:10.1038/sj.cdd.4401602
34. Silvera D, Formenti SC, Schneider RJ. Translational control in cancer. *Nat Rev Cancer* (2010) **10**:254–66. doi:10.1038/nrc2824
35. Elroy-Stein O, Merrick W. Translation initiation via cellular internal ribosome entry sites. In: Mathews MB, Sonenberg N, Hershey J, editors. *Translational Control in Biology and Medicine*. Cold Spring Harbor: Cold Spring Harbor Laboratory Press (2007). p. 155–72.
36. Yoon A, Peng G, Brandenburg Y, Zollo O, Xu W, Rego E, et al. Impaired control of IRES-mediated translation in X-linked dyskeratosis congenita. *Science* (2006) **312**:902–6. doi:10.1126/science.1123835
37. Bellodi C, Kopmar N, Ruggero D. Dereglulation of oncogene-induced senescence and p53 translational control in X-linked dyskeratosis congenita. *EMBO J* (2010) **29**:1865–76. doi:10.1038/emboj.2010.83
38. Bellodi C, Krasnykh O, Haynes N, Theodoropoulou M, Peng G, Montanaro L, et al. Loss of function of the tumor suppressor DKC1 perturbs p27 translation control and contributes to pituitary tumorigenesis. *Cancer Res* (2010) **70**:6026–35. doi:10.1158/0008-5472.CAN-09-4730
39. Braunstein S, Karpisheva K, Pola C, Goldberg J, Hochman T, Yee H, et al. A hypoxia-controlled cap-dependent to cap-independent translation switch in breast cancer. *Mol Cell* (2007) **28**:501–12. doi:10.1016/j.molcel.2007.10.019
40. Gu L, Zhu N, Zhang H, Durden DL, Feng Y, Zhou M. Regulation of XIAP translation and induction by MDM2 following irradiation. *Cancer Cell* (2009) **15**:363–75. doi:10.1016/j.ccr.2009.03.002
41. Mizrachi-Schwartz S, Kravchenko-Balasha N, Ben-Bassat H, Klein S, Levitzki A. Optimization of energy-consuming pathways towards rapid growth in HPV-transformed cells. *PLoS One* (2007) **2**:e628. doi:10.1371/journal.pone.0000628
42. Jan E, Kinzy TG, Sarnow P. Divergent tRNA-like element supports initiation, elongation, and termination of protein biosynthesis. *Proc Natl Acad Sci U S A* (2003) **100**(26):15410–5. doi:10.1073/pnas.2535183100
43. Pestova TV, de Breynne S, Pisarev AV, Abaeva IS, Hellen CU. eIF2-dependent and eIF2-independent modes of initiation on the CSFV IRES: a common role of domain II. *EMBO J* (2008) **27**:1060–72. doi:10.1038/emboj.2008.49
44. Terenin IM, Dmitriev SE, Andreev DE, Shatsky IN. Eukaryotic translation initiation machinery can operate in a bacterial-like mode without eIF2. *Nat Struct Mol Biol* (2008) **6**:6. doi:10.1038/nsmb.1445
45. Yamamoto H, Unbehaun A, Loerke J, Behrmann E, Collier M, Burger J, et al. Structure of the mammalian 80S initiation complex with initiation factor 5B on HCV-IRES RNA. *Nat Struct Mol Biol* (2014) **21**:721–7. doi:10.1038/nsmb.2859
46. Dmitriev SE, Terenin IM, Andreev DE, Ivanov PA, Dunaevsky JE, Merrick WC, et al. GTP-independent tRNA delivery to the ribosomal P-site by a novel eukaryotic translation factor. *J Biol Chem* (2010) **285**:26779–87. doi:10.1074/jbc.M110.119693
47. Skabkin MA, Skabkina OV, Dhote V, Komar AA, Hellen CU, Pestova TV. Activities of ligatin and MCT-1/DENR in eukaryotic translation initiation and ribosomal recycling. *Genes Dev* (2010) **24**:1787–801. doi:10.1101/gad.1957510
48. Schleich S, Strassburger K, Janiesch PC, Koledachkina T, Miller KK, Haneke K, et al. DENR-MCT-1 promotes translation re-initiation downstream of uORFs to control tissue growth. *Nature* (2014) **512**:208–12. doi:10.1038/nature13401
49. Skabkin MA, Skabkina OV, Hellen CU, Pestova TV. Reinitiation and other unconventional posttermination events during eukaryotic translation. *Mol Cell* (2013) **51**:249–64. doi:10.1016/j.molcel.2013.05.026
50. Rubtsova MP, Sizova DV, Dmitriev SE, Ivanov DS, Prassolov VS, Shatsky IN. Distinctive properties of the 5'-untranslated region of human hsp70 mRNA. *J Biol Chem* (2003) **278**:22350–6. doi:10.1074/jbc.M303213200
51. Shi Y, Sharma A, Wu H, Lichtenstein A, Gera J. Cyclin D1 and c-myc internal ribosome entry site (IRES)-dependent translation is regulated by AKT activity and enhanced by rapamycin through a p38 MAPK- and ERK-dependent pathway. *J Biol Chem* (2005) **280**:10964–73. doi:10.1074/jbc.M407874200
52. Bushell M, Wood W, Clemens MJ, Morley SJ. Changes in integrity and association of eukaryotic protein synthesis initiation factors during apoptosis. *Eur J Biochem* (2000) **267**:1083–91. doi:10.1046/j.1432-1327.2000.01101.x
53. Clemens MJ, Bushell M, Jeffrey IW, Pain VM, Morley SJ. Translation initiation factor modifications and the regulation of protein synthesis in apoptotic cells. *Cell Death Differ* (2000) **7**:603–15. doi:10.1038/sj.cdd.4400695
54. Jeffrey IW, Bushell M, Tilleray VJ, Morley S, Clemens MJ. Inhibition of protein synthesis in apoptosis: differential requirements by the tumor necrosis factor alpha family and a DNA-damaging agent for caspases and the double-stranded RNA-dependent protein kinase. *Cancer Res* (2002) **62**:2272–80.
55. Prosniak M, Dierov J, Okami K, Tilton B, Jameson B, Sawaya BE, et al. A novel candidate oncogene, MCT-1, is involved in cell cycle progression. *Cancer Res* (1998) **58**:4233–7.
56. Nandi S, Reinert LS, Hachem A, Mazan-Mamczarz K, Hagner P, He H, et al. Phosphorylation of MCT-1 by p44/42 MAPK is required for its stabilization in response to DNA damage. *Oncogene* (2007) **26**:2283–9. doi:10.1038/sj.onc.1210447
57. Mazan-Mamczarz K, Hagner PR, Corl S, Srikantan S, Wood WH, Becker KG, et al. Post-transcriptional gene regulation by HuR promotes a more tumorigenic phenotype. *Oncogene* (2008) **27**:6151–63. doi:10.1038/onc.2008.215
58. Mazan-Mamczarz K, Hagner P, Dai B, Corl S, Liu Z, Gartenhaus RB. Targeted suppression of MCT-1 attenuates the malignant phenotype through a translational mechanism. *Leuk Res* (2009) **33**:474–82. doi:10.1016/j.leukres.2008.08.012
59. Shih HJ, Chen HH, Chen YA, Wu MH, Liou GG, Chang WW, et al. Targeting MCT-1 oncogene inhibits Shc pathway and xenograft tumorigenicity. *Oncotarget* (2012) **3**:1401–15. doi:10.18632/oncotarget.688
60. Wang Y, Han KJ, Pang XW, Vaughan HA, Qu W, Dong XY, et al. Large scale identification of human hepatocellular carcinoma-associated antigens

- by autoantibodies. *J Immunol* (2002) **169**:1102–9. doi:10.4049/jimmunol.169.2.1102
61. Perez-Arellano I, Gallego J, Cervera J. The PUA domain – a structural and functional overview. *FEBS J* (2007) **274**:4972–84. doi:10.1111/j.1742-4658.2007.06031.x
  62. Jack K, Bellodi C, Landry DM, Niederer RO, Meskauskas A, Musalgaonkar S, et al. rRNA pseudouridylation defects affect ribosomal ligand binding and translational fidelity from yeast to human cells. *Mol Cell* (2011) **44**:660–6. doi:10.1016/j.molcel.2011.09.017
  63. Lee S, Truesdell SS, Bukhari SI, Lee JH, LeTonqueze O, Vasudevan S. Upregulation of eIF5B controls cell-cycle arrest and specific developmental stages. *Proc Natl Acad Sci U S A* (2014) **111**:E4315–22. doi:10.1073/pnas.1320477111
  64. Pelletier J, Graff J, Ruggero D, Sonenberg N. Targeting the eIF4F translation initiation complex: a critical nexus for cancer development. *Cancer Res* (2015) **75**:250–63. doi:10.1158/0008-5472.CAN-14-2789
  65. Tamm I, Kornblau SM, Segall H, Krajewski S, Welsh K, Kitada S, et al. Expression and prognostic significance of IAP-family genes in human cancers and myeloid leukemias. *Clin Cancer Res* (2000) **6**:1796–803.
  66. Pardo OE, Lesay A, Arcaro A, Lopes R, Ng BL, Warne PH, et al. Fibroblast growth factor 2-mediated translational control of IAPs blocks mitochondrial release of Smac/DIABLO and Apoptosis in small cell lung cancer cells. *Mol Cell Biol* (2003) **23**:7600–10. doi:10.1128/MCB.23.21.7600-7610.2003
  67. Schimmer AD, Welsh K, Pinilla C, Wang Z, Krajewska M, Bonneau MJ, et al. Small-molecule antagonists of apoptosis suppressor XIAP exhibit broad antitumor activity. *Cancer Cell* (2004) **5**:25–35. doi:10.1016/S1535-6108(03)00332-5
  68. Carter BZ, Gronda M, Wang Z, Welsh K, Pinilla C, Andreeff M, et al. Small-molecule XIAP inhibitors derepress downstream effector caspases and induce apoptosis of acute myeloid leukemia cells. *Blood* (2005) **105**:4043–50. doi:10.1182/blood-2004-08-3168
  69. LaCasse EC, Mahoney DJ, Cheung HH, Plenchette S, Baird S, Korneluk RG. IAP-targeted therapies for cancer. *Oncogene* (2008) **27**:6252–75. doi:10.1038/onc.2008.302
  70. Flygare JA, Fairbrother WJ. Small-molecule pan-IAP antagonists: a patent review. *Expert Opin Ther Pat* (2010) **20**:251–67. doi:10.1517/13543770903567077
  71. Rigaud S, Fondaneche MC, Lambert N, Pasquier B, Mateo V, Soulas P, et al. XIAP deficiency in humans causes an X-linked lymphoproliferative syndrome. *Nature* (2006) **444**:110–4. doi:10.1038/nature05257
  72. Marsh RA, Madden L, Kitchen BJ, Mody R, McClimon B, Jordan MB, et al. XIAP deficiency: a unique primary immunodeficiency best classified as X-linked familial hemophagocytic lymphohistiocytosis and not as X-linked lymphoproliferative disease. *Blood* (2010) **116**:1079–82. doi:10.1182/blood-2010-01-256099
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