



Genetic Polymorphisms Involved in Bladder Cancer: A Global Review

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Bladder cancer (BC) has been associated with genetic susceptibility. Single peptide polymorphisms (SNPs) can modulate BC susceptibility. A literature search was performed covering the period between January 2000 and October 2020. Overall, 334 articles were selected, reporting 455 SNPs located in 244 genes. The selected 455 SNPs were further investigated. All SNPs that were associated with smoking and environmental exposure were excluded from this study. A total of 197 genes and 343 SNPs were found to be associated with BC, among which 177 genes and 291 SNPs had congruent results across all available studies. These genes and SNPs were classified into eight different categories according to their function.

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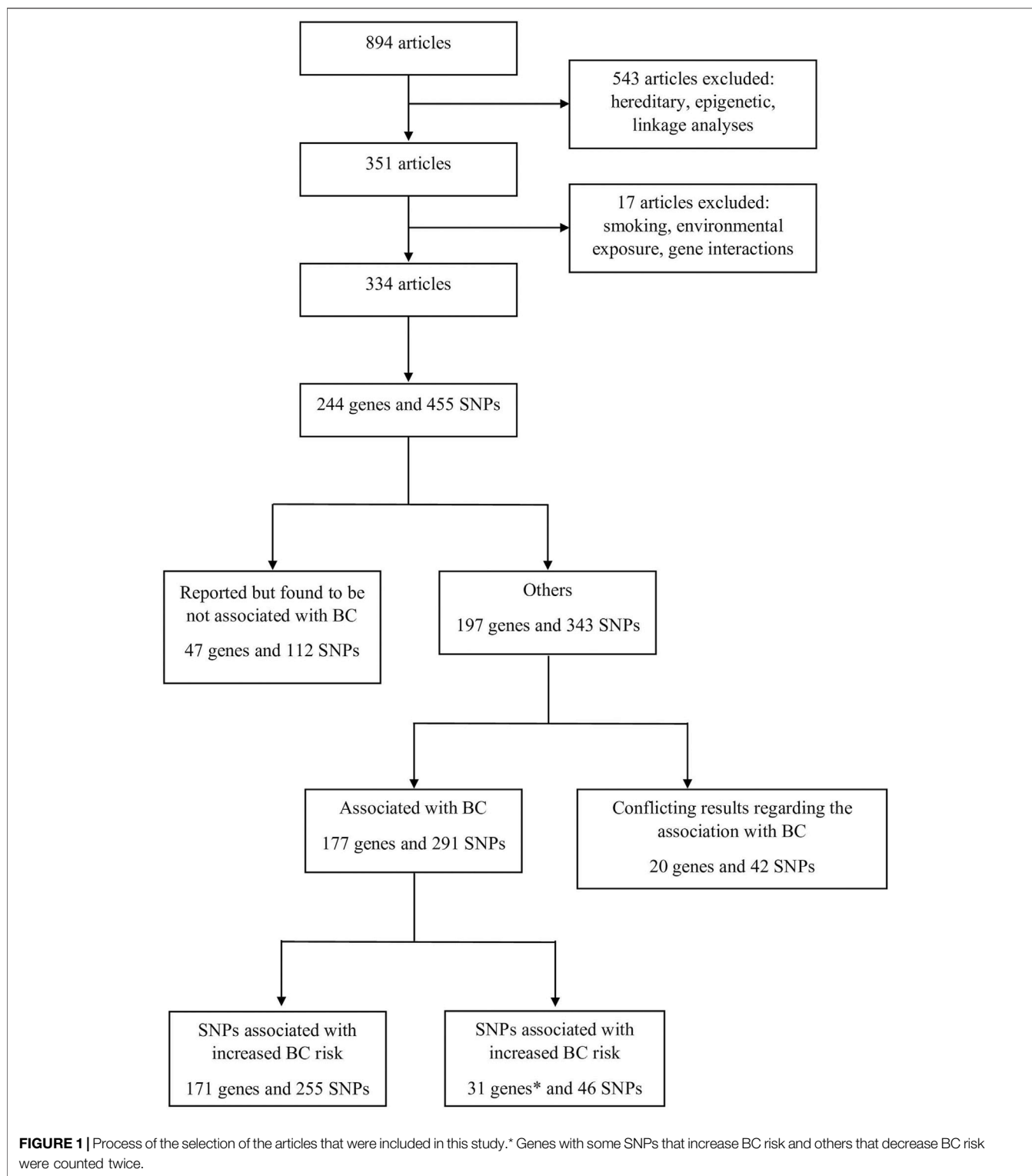
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INTRODUCTION

Bladder cancer (BC), one of the most common cancers worldwide, is particularly prevalent in developed countries [1, 2]. Its global incidence was estimated to be equal to 3%, in 2020, according to GLOBOCAN [1, 2]. It is a complex disease that involves different risk factors, mainly smoking and occupational carcinogen exposure [3], in addition to genetic susceptibility [4]. In fact, the lifetime absolute risk (AR) of developing BC in 50 years-old white non-Hispanic never-smoker men is 1.9%, whereas the AR is 7.1% for current smokers among 50 years-old white non-Hispanic men [5]. Recent advances in DNA sequencing technologies, in particular next-generation sequencing approaches, have opened new horizons, allowing a better understanding of genetic triggers related to BC [6]. Several genomic alterations were found to be linked to this specific cancer, including gene rearrangements, amplifications, deletions, copy number variations, and point variants, including pathogenic and polymorphic variants that are also known as single nucleotide polymorphisms (SNPs) [7]. SNPs, which account for 90% of the human genome's variability, are gaining much interest in the oncogenetics field since many of them have been shown to modulate cancer susceptibility by increasing or decreasing the risk of an individual developing cancer [8]. Among genes associated with cancer, we can cite genes regulating environmental carcinogen metabolism, DNA repair, or cell cycle pathways, all of which are involved in the development and/or progression of any type of cancer, including BC [9, 10].

Many studies have examined the association of SNPs with BC in different populations. This extensive literature review aims to list all SNPs that are significantly associated with BC and discuss their involvement in this type of cancer.



METHODS

In order to gather all available information related to the possible association of various SNPs with BC, an extensive literature search was performed in the PubMed database covering the

period between January 2000 and October 2020. The keywords “bladder cancer” combined with “genetic predisposition” were used with Boolean operators. Overall, 894 articles were selected. The titles and abstracts of these articles were assessed for eligibility before evaluating their contents. Articles that target

TABLE 1 | Summary of genes classification with number of genes in each category.

	Congruent data		Conflicting results	
	Genes	SNPs	Genes	SNPs
Chemical carcinogenesis	13	38	18	53
Signaling	34	54	39	63
Cell death	13	15	14	16
DNA repair	23	32	27	47
Cell cycle	9	16	10	18
Cell architecture	13	14	14	16
Metabolic	20	27	23	34
Others	52	95	52	96
Total	177	291	197	343

Genes and SNPs that showed concordant results between all available studies are designated "associated". Under the category "conflicting results" are classified genes and SNPs that showed discordant results between the available studies. Bold values represent the total number of genes and SNPs in each category.

hereditary cancer, epigenetic studies, and linkage analysis studies were not included. At the end, 334 articles were selected, reporting 455 SNPs located in 244 genes. Selected genes were then classified based on the same method used by [11]. The article selection process is summarized in **Figure 1** as a PRISMA flow diagram.

The selected 455 SNPs were further investigated. All SNPs that were associated with smoking and environmental exposure but did not present a direct association with BC were excluded from this study. However, SNPs associated with BC independently of interfering factors like smoking status, sex, age, and others were included.

RESULTS

In total, 197 genes and 343 SNPs were found to be associated with BC, of which 177 genes and 291 SNPs had congruent results between all available studies, meaning that all studies that assessed a particular SNP had the same results regarding the SNP association with BC. The remaining 20 genes and 42 SNPs were reported with conflicting data, thus representing a huge amount of scattered data that needs a clear classification in order to facilitate their accessibility to researchers. These genes and SNPs were thus thoroughly evaluated and classified into eight different categories according to their function: chemical carcinogenesis, signaling, cell death, DNA repair, cell cycle, cell architecture, metabolic, and other genes that do not correspond to any specific category. Further subclassifications were also enclosed in each category (**Table 1**).

Chemical Carcinogenesis

Genetics and environmental factors impact the quantities of enzymes that participate in the activation and detoxification of chemicals, which can potentially lead to carcinogenesis. Most chemical compounds become carcinogens after being metabolized to chemically reactive electrophiles, which can interact with DNA to generate a carcinogenic response [12]. Genes encoding enzymes modulating chemical carcinogenesis

were found to be linked to BC. In this section, genes were divided according to the different known enzyme families. Genes that encode for phase I and phase II enzymes were subcategorized into four categories: Cytochromes, Glutathione S-Transferase, N-acetyltransferase, and UDP-Glucuronosyltransferase (UGT).

Cytochromes are phase I enzymes whose role is to convert xenobiotics into excretable compounds. They are divided into subfamilies and are primarily located in the liver, but can also be found in the gastrointestinal tract, lungs, and kidneys. Mutations in these genes can affect substrate transformation, which may lead, in some cases, to cancer [13]. For instance, rs2472299 in *CYP1A* is known worldwide as a risk factor for BC [13], while rs4646903 and rs2198843 in the same gene [13, 14] are known to widely increase the risk for the same type of cancer but exclusively in the Asian population, and rs4646421 is specific for BC in Tunisians [15]. Results regarding rs1048943 in *CYP1A1* are contradictory, since it increases the risk for BC in Asian and Brazilian populations but was not associated with BC in many other studies [13, 16–19]. Similarly, rs762551 in *CYP1A2* showed conflicting results [20–22]. Regarding the *CYP1B1* gene: rs2855658 increases BC risks in the European population [23], and rs10012, rs1056827, and rs150799650 showed an increased risk in Indo-European cohorts [24], while rs1056827 was associated with a decreased risk in the Spanish population [25]. Contradictory results were seen for rs1056836, which presents a notable increased risk for BC in the Spanish population [13, 19, 25, 26]. On the other hand, rs4244285 and rs4986893 in *CYP2C19* showed a protective effect against BC in the Chinese population [27]. *CYP2E1* was associated with an increased risk for BC in the Lebanese population [28]. However, rs2031920 in the same gene was widely associated with a decreased risk in Asians [29] with controversial results in Caucasians [29, 30].

Glutathione S-Transferase constitutes a superfamily of phase II multipurpose enzymes that contribute to metabolic detoxification processes that protect macromolecules from environmental carcinogens, reactive oxygen species, and chemotherapeutic agents. The glutathione S-transferase family includes the following enzymes: GSTA, GSTM, GSTP, GSTT, and GSTO. Since these molecules contribute to the metabolism of potential carcinogens, any polymorphism affecting their expression or function may lead to cancer [31]. Most articles show that in most populations, the *GSTM1* null genotype [32–47] and rs1695 in *GSTP1* [16, 19, 33, 40, 42, 48–55] increase the risk for BC. However, a Chinese study showed that the *GSTT1* null genotype was associated with a decreased risk for BC [36]. Rs156697 in *GSTO2* also shows a positive association with BC in the Serbian population [56]. The polymorphisms in *GSTT1* were mostly associated with an increased risk for BC [16, 18, 32–34, 40, 42, 44, 47, 48, 57–63]. Rs3957356, rs3957357, and rs4715332 in *GSTA1* were associated with an increased risk for BC in both Balkan-Ben and Balkan-non Ben [63].

N-acetyltransferase (NAT) enzymes, also known as NAT1 and NAT2, are responsible for acetylating aromatic and heterocyclic amines in the liver, the gastrointestinal tract, and the urinary bladder. Genetic polymorphisms in these enzymes can alter the processing rate of various carcinogenic compounds, thus

increasing the risk for cancer. Hence, slow NAT2 acetylation is associated with increased risk for BC [64]. Many SNPs in *NAT1* are associated with a significantly increased risk for BC in the Lebanese population, including rs15561, rs4986782, and rs1057126 [28, 65, 66]. However, the latter did not show any association with BC in two other meta-analyses [4, 67]. Rs9650592 in *NAT1* was positively associated with BC in the European population [23]. A study conducted on a French Caucasian population demonstrated an increased risk for BC in the presence of rs1208, rs1801280, or rs1041983 in *NAT2* [19]. Rs1041983 presents the same effect in the American population [40]. Rs1799929, rs1799930, and rs1799931 were shown to be associated with BC in French and Bangladeshi populations [19, 68]. However, a Chinese meta-analysis contradicted the French results and found no association between these SNPs and BC [69]. Finally, rs1495741 and rs4646249 increase the risk of BC [11, 37, 70] in the European population [23].

The UDP-Glucuronosyltransferase (UGT) family of enzymes are phase II enzymes that are involved in the glucuronidation of aromatic amines and other carcinogens. They are primarily located in the liver but also in the gastrointestinal tract, lungs, and kidneys. The UGT family is composed of UGT1A, UGT2A, and UGT2B. They present different subtypes that are all, except for UGT2B17, expressed in normal bladder tissue [71]. Conflicting data was documented regarding the association of rs11892031 in *UGT1A10/UGT1A8* with BC [23, 39, 70, 72–75]. Rs17863783 in *UGT1A6* decreases the risk of developing BC [70, 76]. Rs1104892, rs1105880, rs1113193, rs1604144, rs17854828, rs17864684, rs17868322, rs2602374, rs2741042, rs2741044, rs2741045, and rs4148326 in *UGT1A8* are associated with an increased risk in the American population, while rs1113193, rs1604144, rs17854828, rs17864684, rs4148328, rs4233633, and rs7571337 are associated with a decreased risk in the same population [77]. Rs2736520 in *UGT2B4* increases the risk for BC, while Rs3822179 decreases the risk in the American population [77].

CDC-like kinase 3 (CLK3) is a dual specificity kinase that belongs to the serine/threonine type kinase family. Its role in human cancer is still undetermined [78]. However, a Japanese study has established that rs11543198 is associated with an increased risk for BC in the Japanese population [79].

Signaling

Signaling and immune-related genes were found to be associated with BC in the literature. These genes were divided into four categories, including cytokines, toll-like receptors, transcriptional factors, and other genes.

For an effective immune response against cancer cells, a sequence of steps must take place in fighting tumor cells. Each step of this sequence requires the presence of several stimulatory and inhibitory factors [80]. Among these actors, Cluster of Differentiation (CD) 274 [also known as Programmed Death Ligand 1 (PD-L1)], has an inhibitory function toward immune response activation. Rs4143815 in this gene increases the risk for BC in the Chinese population [81]. Cytotoxic T-lymphocyte-associated protein 4 (CTLA4) can also inhibit T cell-mediated immune responses, and rs231775 and rs3087243 in the

corresponding gene have been shown to increase the risk for BC in the Indian population [82]. Intercellular Adhesion Molecule 1 (ICAM1) is responsible for T cell tumor infiltration, and therefore a variation in this gene can increase cancer susceptibility, as shown for rs5498 in the Taiwanese population [83]. Furthermore, chemokines help regulate the immune response and impact cancer progression [84]. Among the genes encoding chemokines, C-C chemokine receptor type 2 (*CCR2*) rs1799864 [85–87] and *CCR5* rs333 [85] were associated with an increased risk for BC in the Turkish population, rs1801157 in C-X-C motif chemokine 12 (*CXCL12*) also has a positive association with BC in Indian [88] and Turkish [85] populations; and rs1126579 in CXC chemokine receptors 2 (*CXCR2*) increases cancer's susceptibility in the Indian population [88] while it shows a protective effect towards BC in the American population [89]. Finally, rs187115 in *CD44* was associated with an increased risk for BC in the Taiwanese population [90]. Tumor-secreted T-helper 17 (Th17) cell-associated interleukins (IL) mobilize immune suppressive cells and promote tumor growth [91]. Rs2275913 in *IL-17A* and rs187238 in *IL-18* increase the risk for BC in the Polish population, with the highest association for *IL-18* [92]. Rs1946518 in *IL-18* also increases the risk of BC in the Indian population [93]. Rs2227485 in *IL-22* is also a risk factor for BC in the Chinese population [94]. Other interleukins that play a role in macrophages and neutrophils' immunosuppressive roles include *IL-10*, *IL-12*, and *IL-23* [95]. Rs1800896, rs1800871, and rs1800872 in *IL-10* [96] are associated with an increased risk for BC in Chinese [97] and Indian [98] populations; rs10889677 in *IL-23R* also increases BC risk in Chinese [99] and Polish [92] populations; and *IL-12* decreases the risk in the Indian population [93]. Other interleukins were also found to play a role in BC risk; for instance, rs2069762 in the *IL-2* gene increases the risk for BC in the Chinese population. Rs4073 in *IL-8/CXCL8* gives high BC susceptibility in the Indian population [100] and rs1800890 in *IL-19* in the Spanish population [101]. Last but not least, rs153109 and rs17855750 in *IL-27* predispose to BC in Spanish and Chinese individuals, respectively [102, 103]. Finally, rs1799964 in *TNF- α* [104] and rs1800470 in *TGFB1* [105] are positively associated with BC in the Indian population.

Toll-like receptors (TLR) play a key role in the initiation of the innate immune response. They are expressed on both immune and tumor cells and regulate the immune responses in tumor progression and as therapeutic targets for cancer [106]. *TLR2* mutations (–196 to –174del) increase the risk of BC in India [107]. Rs11536889 in *TLR4* [108–110] and rs72552316 in *TLR7* [111] constitute separate BC risk factors.

Transcription factors might be linked to BC [112]. For instance, rs2228570 in *VDR* (Vitamin D receptor) was found to increase the risk of developing BC in India [113] and Tunisia [114]. *TP63* can act as a tumor suppressor gene or as an oncogene depending on the cellular setting and pathways where it is implicated, and it can thus regulate the transcription of different genes [115]. Several SNPs have been shown to increase the risk of BC, among them rs4687100 [116] and rs710521 [39, 70, 116]. Of note, the latter has also been proven to present a protective effect in the Indian population

[117], but no association with BC was identified in a Chinese study [118]. Finally, conflicting data were also obtained for rs35592567 since it was shown to have a positive association in one meta-analysis [11] and a negative association in another [119].

Vascular endothelial growth factor (VEGF) regulates angiogenesis and is upregulated and overexpressed in BC [120]. Rs699947 and Rs35569394 in *VEGF* increase and decrease, respectively, the BC risk in the Indian population [121]. Moreover, rs833052, rs25648 [122], rs3025039 [122, 123], and rs699947 [124] in *VEGFA* showed a positive association with BC, while the latter also presented a protective effect in the Tunisian population [125]. Rs3775194, rs1485762, rs6828869, and rs17697515 in *VEGFC*, and rs4557213 in *VEGFR* increase the risk for BC in the American Caucasian population [126] while rs1485766 in *VEGFC* increases the risk in the Taiwanese population [127]. Human Leukocyte Antigen G (HLA-G) expression in tumors promotes the immune suppressive microenvironment, which results in poor treatment response and prognosis [128]. Rs1063320, rs1610696, rs1704, rs1707, rs1710, rs17179101, and rs17179108 in *HLA-G* are associated with increased cancer susceptibility in the Brazilian population [129].

Other SNPs that were also studied in various genes include: rs6593205 and rs7799627 in Epidermal Growth Factor Receptor (*EGFR*), which decrease the risk for BC in the American Caucasian population, while rs11238349 increases the risk in the same population [126] and rs1050171 increases the risk in the Chinese population [130]. Rs696 in *NFKBIA* and Rs11188660 in *BLNK* (B cell linker) increase the risk in the Spanish population [101]. Rs28362491 in *NFKB1* [131–133] and rs7944701 in *MAML2* [134] increase the risk in the Chinese population. A more extensive list is available in **Supplementary Table S1** under the section titled “signaling.”

Cell Death

By evading death, immortal cells can develop into tumors. Mutations in certain pathways promoting cell death can thus lead to tumor formation. Cell death pathways involve multiple genes, including cell-death receptors such as Tumor Necrosis Factor Receptor 1 (*TNFR1*), *FAS*, and TNF-related apoptosis-inducing ligand (*TRAIL*) receptors and effectors such as caspases [135]. Genes and SNPs related to those pathways were reported in BC. Rs2234767 in *FAS*, Rs763110 in *FASLG* [136], and Rs1131580 in *TNFSF10* [137] increase the risk for BC in the Turkish population. In *DR4* (*TRAILR1* or *TNFRSF10A*), rs6557634 increases the risk for BC the Indian population [138] and rs13278062 increases the risk in the Chinese population [139] while rs20575 was found to present a protective effect in the Caucasian population [140]. Moreover, rs2647396 in *BCL10* [101], rs10999426 in *PRF1* [101], and rs42490 in *RIPK2* [141] each individually increase the risk for BC in the Spanish population, while rs401681 in *CLPTMIL* increases the risk widely in all studied populations [39, 73, 74]. Rs4647603 in *CASP3* and rs3181320 and rs507879 in *CASP5* increase the risk for BC in Indians [138] while rs3834129 in *CASP8* [142] and rs4645978 in *CASP9* [143]

decrease the risk for BC in the Chinese and Indian populations, respectively. Finally, rs1045411 in *HMGB1* presents a protective effect in the Chinese population [144].

DNA Repair

The DNA repair genes associated with BC are classified according to the repair mechanism: base excision repair (BER), nucleotide excision repair (NER), homologous recombination (HR), and poly-ADP-ribosylation.

BER is a mechanism of DNA reparation that is not restricted to the repair of single-strand breaks but also of damages resulting from defects in alkylation, oxidation, deamination, and depurination. Given that this mechanism repairs thousands of errors per cell and per day, it plays a major role in cancer prevention by ensuring genome integrity and stability. Therefore, BER guarantees the integrality of apoptosis pathways and prevents mutation accumulation that may initiate tumors [145]. In X-ray repair cross-complementing protein 1 (*XRCC1*), rs1799782 and rs25489 are widely associated with an increased risk for BC, especially in Asian and Indian populations [146–152]. Rs915927 in the same gene increases tumor susceptibility in the Italian population [153] while rs25487 gave contradictory results in different studies [147–149, 154–159]. Rs3218373, rs3218536, and rs6464268 in *XRCC2* present a protective effect towards BC in the Italian population [160]. Rs1805377 in *XRCC4* increases the risk of BC in Spanish [160] and Indian individuals [161]. Rs6869366 increases the risk of BC in Taiwanese people [162] but decreases the risk in the Indian population [161]. Rs828907 in *XRCC5* increases BC risk in the Taiwanese population [163]. Rs3213245 and rs7003908 in *XRCC7* (*PRKDC*) are positively associated with BC, respectively, in the Chinese Han [164] and in the Indians [165]. Other SNPs in BC-associated BER genes include rs1760944 in *APEX1*, which decreases the risk for BC in the Chinese population [146] while rs1130409 in *APE1* showed conflicting data [147, 166, 167]. Rs3136717 in *POLB* [168] and rs1052133 in *hOGG1* [147, 165, 169–171] are associated with a high risk for BC, while rs125701 in *hOGG1* decreases the risk for BC in the Spanish population [168]. Finally, rs2029167 in *SMUG1* and rs3219487 in *MUTYH* increase the risk for BC in the American population [172] and rs11039130 in *DDB2* has the same effect in the Caucasian population [173].

NER is also an important DNA repair pathway consisting of two distinct sub-pathways. The global-genome NER process fixes helix damages over the entire genome, whereas the transcription-coupled NER mechanism acts during transcription to resolve RNA polymerase blocking lesions [174]. Mutations in genes implicated in NER can result in a predisposition to cancer since mutations and chromosomal abnormalities can either activate oncogenes or inactivate tumor suppressor genes [175]. Rs3212961 in *ERCC1* increases BC risk in the Spanish population [176] whereas rs967591, rs735482, and rs2336219 have a protective effect towards BC in the Italian population [153]. Rs13181 [15, 157, 177–182], rs1799793 [166, 170, 177, 178, 183–186] and rs238406 [176, 178, 179] in *ERCC2* (*XPD*) were individually widely associated with an increased BC risk. Rs1047769 [176] and Rs17655 [187] in *ERCC5* increase the

risk of BC, respectively, in Spanish and Chinese people. Rs2228526 and rs2228528 in *ERCC6* increase BC susceptibility in Belarussians [181] with the latter also having the same effect in the Taiwanese population [188]. Rs4150667 in *GTF2H1* is associated with an increased risk for BC in the Caucasian race [173], rs1805335 in *RAD23B* increases the risk in the Spanish population [176] and rs2228000 [189–193] and rs2228001 [155, 187, 193–195] in *XPC* are positively associated with BC in all study populations.

HR is another mechanism that helps maintain the genome's integrity by repairing double-strand breaks. Therefore, HR deficiencies that result from gene mutations make individuals more susceptible to cancer [196]. Rs861539 in *XRCC3* is associated with an increased risk for BC in different populations [160, 197–199]. Rs1799794 and rs861530 in the same gene also increase BC risk in the Chinese population [200]. Rs11571833 in *BRCA2* increases BC risk in those of European descents [201]. Rs1805794 in *NBN* is associated with a high risk for BC [202] while rs8032440 in *FANCI* and rs3739177 in *PNKP* make the American population more susceptible to BC [172].

Poly-ADP-ribosylation is a post-translational modification catalyzed by poly(ADP-ribosyl)ation polymerases (PARPs) as a response to DNA damage [203]. Rs1136410 in *PARP1* is associated with a high BC risk in the Spanish population [168], whereas rs3219123 and rs12568297 in *PARP1*, rs1713413 in *PARP2*, and rs2862907 in *PARP4* increase separately the risk for BC in the American population [172].

Cell Cycle

Genes and their corresponding SNPs related to the cellular cycle were separated into three groups consisting of tumor suppressor genes, inhibitors of apoptosis, and other genes associated with BC.

Tumor suppressor genes work by repairing DNA damage, inhibiting cell division, and, in some cases, triggering apoptosis to stop tumor development. Therefore, inactivation or loss of function resulting from mutations in these genes can lead to cancer [204]. An SNP in intron 3 of the *TP53* gene was associated with a decreased risk for BC in the American population [183]. Moreover, rs1042522 in the same gene is associated with an increased risk for BC in Asia [205–207]. However, this same SNP showed a protective effect towards BC in the Indian [208] and Brazilian [209] populations, and rs17878362 has the same effect in the American population [210]. Rs2839698 in *H19* shows a protective effect towards BC in the Netherlands [211] whereas rs217727 in *H19* [212, 213] and rs760805 in *RUNX3* [214] increase BC risk in the Chinese population. Finally, rs2073636 in *TSC2* is associated with a high risk for BC in the American population [172] and rs17879961 in *CHEK2* with a low risk for BC in European descent [201].

Inhibitors of apoptosis are a family of eight proteins. A mutation activating any of the corresponding genes will become a weapon that will be used by tumors to evade apoptosis [215]. Rs2071214 [216], rs8073069 [216], rs9904341 [216–219], and rs3764383 [219] in *BIRC5* increase

BC risk, while rs17878467 decreases the risk in Asians [216, 219, 220].

Other genes include Cyclin D1 (*CCND1*) which regulates cell cycle progression through the activation of cyclin-dependent kinases 4 and 6 (CDK4 and CDK6) that lead to Rb protein phosphorylation, thus inactivating it and allowing the cell to progress past the G1/S checkpoint and continue its replication. Over-expression of *CCND1* can lead to cancer [221] and the rs9344 in this gene increases the risk for BC [221–225]. Similarly, Cyclin E1 (*CCNE1*) binds to CDK2 and activates it, allowing the cell to progress and enter phase S [226]. Rs8102137 in *CCNE1* widely increases BC development risk [11, 39, 73, 75, 116]. Moreover, the PI3K–AKT–mTOR is implicated in cell growth, tumorigenesis, and cell invasion [227]. Mitochondrial gene *POLG* polymorphism rs3087374 increases BC risk in the American population [172]. Rs2294008 [11, 37, 39, 73, 79, 228–235] and rs2978974 [70] in the prostate stem cell antigen (*PSCA*), an inhibitor of cell proliferation, increase widely BC risk.

Cell Architecture

Caveolin-1 (*CAV1*) is an essential membrane protein expressed in multiple cells. It plays a central role in the formation of caveolae, which are small plasma membrane invaginations involved in signaling and transport. The role of *CAV1* in carcinogenesis has been proven. However, the mechanism is still unknown [236]. Rs3807987 and rs7804372 in *CAV1* increase the risk for BC in the Taiwanese population [237] while rs1049334 widely increases it. *CLTA* and *CLTC*, which encode the light and heavy chains of clathrin, are also associated with BC. Indeed, rs10972786 in *CLTA* and rs7224631 in *CLTC* increase the risk of BC in the European population [23]. Clathrin-mediated vesicle pathways include the *DNM1* and *DNM2* genes. Rs13285411 in *DNM1* and rs4804528 in *DNM2* increase the risk for BC, while rs4804149 in *KANK2* has a significant protective effect towards BC in the European Population [23]. Finally, rs12216499 in *RSPH3* widely increases the risk for BC [70], while rs907611 in *LSP1* increases the risk for BC in the European population [238] but was found not to be associated with BC in Chinese [70].

Rs738141 in Dynein light chain 4 (*DNAL4*) implicated in cell motricity increases the risk for BC in the European population [23].

Rs16260 in Cadherin-1 (*CDH1*) contributes to BC susceptibility in the Chinese population [239].

The extracellular degradation process is mediated by the Matrix Metalloprotease (MMP), which plays a regulatory role in multiple pathways such as apoptosis or angiogenesis. Hence, MMPs participate in carcinogenesis [240]. Conflicting results concerning the association of rs1799750 in *MMP1* with BC were obtained [241–245]. Rs243865 in *MMP2* has shown an increased BC risk in India [246] and in a meta-analysis conducted by [244]. However, another meta-analysis done by L. Tao et al. has shown, for the same SNP, a protective effect towards BC in the Asian population [245]. Contradictory results have also been reported for rs11568818 in *MMP7* [242, 245, 247]. Finally, rs28382575 in *MMP11* increases BC risk in Taiwan [248].

Metabolic

Metabolism-related genes were found to be associated with BC. These genes were divided into alcohol metabolism genes, solute carrier transporters (SLC), folate metabolism enzymes, water-soluble vitamin metabolism genes, and other various metabolism-related genes.

Alcohol metabolism requires multiple enzymes. Rs12529 in *AKR1C3* decreases BC risk in the Turkish population [249] while Rs4680 in *COMT* has shown contradictory results [19, 250, 251].

SLC transporters are membrane proteins whose role is to supply cells with nutrients, neurotransmitters, hormones, and drugs. They are usually upregulated in cancer [252]. Rs17674580 in *SLC14A* [39, 253] and rs1058396 [74], rs10775480 [11, 254], rs10853535 [70, 254], rs17674580 [37, 79, 117], and rs7238033 [39, 70, 254] in *SLC14A1* increase BC susceptibility in different populations. Rs1385129 in *SLC2A1*, also known as Glucose transporter 1 (*GLUT1*), decreases the carcinogenesis risk in the Chinese population [255]. Rs11871756, rs11077654, rs9913017, and rs4969054 in *SLC39A11* increase the risk for BC in different populations [256] and rs2306283 in *SLCO1B1* increases the risk in the Japanese population [257].

Mitochondrial folate metabolic enzymes are associated with cancer [258]. Hence, rs1667627 in *MTHFD2* increases the risk of developing BC in the American population [89]. Other SNPs in genes involved in the folate metabolism are: rs1801131 [259–262] and rs1801133 [260–265] in *MTHFR* that are widely classified as risk factors for BC, whereas rs1476413 in the same gene decreases the risk of cancer occurrence [266], rs1805087 in *MTR* increases the risk for BC in the Tunisian population [262, 267] and rs1801394 in *MTRR* increases the risk for BC only in Saudi Arabia [18].

SNPs in the metabolism of water-soluble vitamin genes that increase BC risk include rs61330082 in *NAMPT* in the Chinese population [268], rs4652795 in *NMNAT2*, rs7636269 in *NMNAT3*, and rs2304191 in *NMRK2* in the European population [23].

Other metabolism genes that are associated with BC are reported in **Supplementary Table S1**.

Others

Finally, unclassified genes that present various actions and are implicated in different pathways were also found to be associated with BC. These genes are reported under the section named “Others” in **Supplementary Table S1**.

Not Associated SNPs

Many other SNPs in different genes were also studied and found not to modulate BC risk. Among these, rs3892097 in *CYP2D6* was not found to be associated with BC in the Indian [62] and Tunisian [269] populations. Moreover, rs1800629 in *TNF- α* [270, 271] and rs833061 in *VEGFA* [122] also showed no association with BC in a meta-analysis. Rs4253211 in *ERCC6* [272], rs1042489 in *BIRC5* [216] and rs4880 in *SOD2* [273] were also not associated with BC. Finally, rs11225395, rs35866072, and rs1940475 in *MMP8* [274], as well as rs3918242 [243, 244, 275],

rs3918241, rs2250889, rs17576, and rs17577 in *MMP9* all showed no association with any risk for carcinogenesis [276]. Other genes and SNPs that strictly showed no association with BC risk are reported in **Supplementary Table S1** under the section “No association.”

DISCUSSION

Several SNPs associated with BC are restricted to specific populations. A generalization on an SNP’s potential role in predisposing or protecting an individual from BC cannot be done without testing this specific SNP in a sufficient number of individuals from different races and origins. Some SNPs were studied in different populations, which enabled us to correlate them to BC risk. On the other hand, many other SNPs that were exclusively found not to be associated with BC were only studied in one or two populations. Therefore, the scarcity of the gathered data related to these SNPs renders the interpretation of their correlation with BC challenging and requires broader studies for the validation of their contribution to this type of cancer. For SNPs with conflicting results between studies, GWAS-specific repositories could be useful.

On the other hand, the risk conferred by these variants cannot alone explain the development of BC [277]. In fact, several environmental factors were found to play a role in the pathogenesis of the disease. These include: gender, age, smoking habits, alcohol consumption, and potential environmental exposure. It is also worth mentioning that some SNPs were found to be associated with more invasiveness and recurrence. For example, *PSCA* rs2294008 was associated with a more invasive disease [234].

Individual SNPs effects on modulating BC risk are minimal. Therefore, studies have shifted towards assessing the polygenic risk score (PRS), as it represents a more accurate representation of an individual’s risk of developing BC. PRS aggregates the effects of multiple SNPs to provide a disease risk prediction [278]. A PRS based on 24 independent GWAS markers showed a fourfold increase in BC risk for both smokers and nonsmokers [5].

Finally, the clinical value of such information has yet to be investigated. In fact, recommendations regarding the clinical management of patients based on their genotypes are still lacking [279].

CONCLUSION

SNPs are genetic variants that are generally population specific [8]. This review shows that several SNPs were associated with BC, depending on the studied cohorts. The generalization of the link between a variant and BC is not always possible, especially in the absence of data related to large cohorts from different ethnicities. For instance, several SNPs associated with BC are private to specific populations. On the other hand, the involvement of many SNPs in BC was

ruled out based on studies focusing on one to two cohorts only. The scarcity of related data urges us to gather all the information we can in order to make it accessible to the scientific community. However, one should consider the complexity of the interpretation of these genomic markers [277], especially that, in many cases, the cumulative effect of several SNPs contributes to modulating the risk for BC [280], in addition to epidemiological or environmental factors such as gender, age, smoking habits, and alcohol consumption. Last but not least, the clinical value of such information has yet to be investigated.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding author.

REFERENCES

- GCO. *900-World-Fact-Sheets.Pdf* (2021). Available From: <https://gco.iarc.fr/today/data/factsheets/populations/900-world-fact-sheets.pdf> (Accessed January 25, 2021).
- GCO. *30-Bladder-Fact-Sheet.Pdf* (2021). Available From: <https://gco.iarc.fr/today/data/factsheets/cancers/30-Bladder-fact-sheet.pdf> (Accessed January 25, 2021).
- Cumberbatch MGK, Jubber I, Black PC, Esperto F, Figueroa JD, Kamat AM, et al. Epidemiology of Bladder Cancer: A Systematic Review and Contemporary Update of Risk Factors in 2018. *Eur Urol* (2018) 74(6):784–95. doi:10.1016/j.eururo.2018.09.001
- Wu K, Wang X, Xie Z, Liu Z, Lu Y. N-Acetyltransferase 1 Polymorphism and Bladder Cancer Susceptibility: A Meta-Analysis of Epidemiological Studies. *J Int Med Res* (2013) 41(1):31–7. doi:10.1177/0300060513476988
- Koutros S, Kiemeny LA, Pal Choudhury P, Milne RL, Lopez de Maturana E, Ye Y, et al. Genome-Wide Association Study of Bladder Cancer Reveals New Biological and Translational Insights. *Eur Urol* (2023) 84(1):127–37. doi:10.1016/j.eururo.2023.04.020
- Audenet F, Attalla K, Sfakianos JP. The Evolution of Bladder Cancer Genomics: What Have We Learned and How Can We Use It? *Urol Oncol Semin Original Invest* (2018) 36(7):313–20. doi:10.1016/j.urolonc.2018.02.017
- Dutt A, Beroukhi R. Single Nucleotide Polymorphism Array Analysis of Cancer. *Curr Opin Oncol* (2007) 19(1):43–9. doi:10.1097/coo.0b013e328011a8c1
- Bernig T, Chanock SJ. Challenges of SNP Genotyping and Genetic Variation: Its Future Role in Diagnosis and Treatment of Cancer. *Expert Rev Mol Diagn* (2006) 6(3):319–31. doi:10.1586/14737159.6.3.319
- Guo CC, Czerniak B. Bladder Cancer in the Genomic Era. *Arch Pathol Lab Med* (2019) 143(6):695–704. doi:10.5858/arpa.2018-0329-ra
- Wu X, Lin X, Dinney CP, Gu J, Grossman HB. Genetic Polymorphism in Bladder Cancer. *Front Biosci* (2007) 12:192–213. doi:10.2741/2058
- de Maturana EL, Rava M, Anumudu C, Sáez O, Alonso D, Malats N. Bladder Cancer Genetic Susceptibility. A Systematic Review. *Bladder Cancer* (2018) 4(2):215–26. doi:10.3233/blc-170159
- Wogan GN, Hecht SS, Felton JS, Conney AH, Loeb LA. Environmental and Chemical Carcinogenesis. *Semin Cancer Biol* (2004) 14(6):473–86. doi:10.1016/j.semcancer.2004.06.010
- Sankhwar M, Sankhwar SN. Variations in CYP Isoforms and Bladder Cancer: A Superfamily Paradigm. *Urol Oncol Semin Original Invest* (2014) 32(1):28.e33–28.e40. doi:10.1016/j.urolonc.2012.10.005
- Wang Y, Kong CZ, Zhang Z, Yang CM, Li J. Relationships Between CYP1A1 Genetic Polymorphisms and Bladder Cancer Risk: A Meta-Analysis. *DNA Cel Biol* (2014) 33(3):171–81. doi:10.1089/dna.2013.2298
- Feki-Tounsi M, Khelif R, Louati I, Fourati M, Mhiri MN, Hamza-Chaffai A, et al. Polymorphisms in XRCC1, ERCC2, and ERCC3 DNA Repair Genes, CYP1A1 Xenobiotic Metabolism Gene, and Tobacco Are Associated With Bladder Cancer Susceptibility in Tunisian Population. *Environ Sci Pollut Res* (2017) 24(28):22476–84. doi:10.1007/s11356-017-9767-x
- Grando JPS, Kuasne H, Losi-Guembarovski R, Sant'ana Rodrigues I, Matsuda HM, Fuganti PE, et al. Association Between Polymorphisms in the Biometabolism Genes CYP1A1, GSTM1, GSTT1 and GSTP1 in Bladder Cancer. *Clin Exp Med* (2009) 9(1):21–8. doi:10.1007/s10238-008-0015-z
- Lu Y, Zhang XL, Xie L, Li TJ, He Y, Peng QL, et al. Lack of Association Between CYP1A1 Polymorphisms and Risk of Bladder Cancer: A Meta-Analysis. *Asian Pac J Cancer Prev* (2014) 15(9):4071–7. doi:10.7314/apjcp.2014.15.9.4071
- Elhawary NA, Nassir A, Saada H, Dannoun A, Qoqandi O, Alsharif A, et al. Combined Genetic Biomarkers Confer Susceptibility to Risk of Urothelial Bladder Carcinoma in a Saudi Population. *Dis Markers* (2017) 2017:1–11. doi:10.1155/2017/1474560
- Fontana L, Delort L, Joumard L, Rabiau N, Bosviel R, Satih S, et al. Genetic Polymorphisms in CYP1A1, CYP1B1, COMT, GSTP1 and NAT2 Genes and Association With Bladder Cancer Risk in a French Cohort. *Anticancer Res* (2009) 29(5):1631–5.
- Sun WX, Chen YH, Liu ZZ, Xie JJ, Wang W, Du YP, et al. Association Between the CYP1A2 Polymorphisms and Risk of Cancer: A Meta-Analysis. *Mol Genet Genomics* (2015) 290(2):709–25. doi:10.1007/s00438-014-0956-8
- Zeng Y, Jiang HY, Wei L, Xu WD, Wang YJ, Wang YD, et al. Association Between the CYP1A2 Rs762551 Polymorphism and Bladder Cancer Susceptibility: A Meta-Analysis Based on Case-Control Studies. *Asian Pac J Cancer Prev* (2015) 16(16):7249–54. doi:10.7314/apjcp.2015.16.16.7249
- Vukovic V, Ianuale C, Leoncini E, Pastorino R, Gualano MR, Amore R, et al. Lack of Association Between Polymorphisms in the CYP1A2 Gene and Risk of Cancer: Evidence From Meta-Analyses. *BMC Cancer* (2016) 16:83. doi:10.1186/s12885-016-2096-5
- Menashe I, Figueroa JD, Garcia-Closas M, Chatterjee N, Malats N, Picornell A, et al. Large-Scale Pathway-Based Analysis of Bladder Cancer Genome-Wide Association Data From Five Studies of European Background. *PLoS One* (2012) 7(1):e29396. doi:10.1371/journal.pone.0029396
- Sankhwar M, Sankhwar SN, Abhishek A, Gupta N, Rajender S. CYP1B1 Gene Polymorphisms Correlate With an Increased Risk of Urinary Bladder Cancer in India. *Urol Oncol Semin Original Invest* (2016) 34(4):167.e1–167.e8. doi:10.1016/j.urolonc.2015.11.010
- Salinas-Sánchez AS, Donate-Moreno MJ, López-Garrido MP, Giménez-Bachs JM, Escribano J. Role of CYP1B1 Gene Polymorphisms in Bladder Cancer Susceptibility. *J Urol* (2012) 187(2):700–6. doi:10.1016/j.juro.2011.10.063

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

CONFLICT OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontierspartnerships.org/articles/10.3389/or.2023.10603/full#supplementary-material>

26. Liu Y, Lin C, Zhang A, Song H, Fan C. The CYP1B1 Leu432Val Polymorphism and Risk of Urinary System Cancers. *Tumor Biol* (2014) 35(5):4719–25. doi:10.1007/s13277-014-1617-6
27. Shi WX, Chen SQ. Frequencies of Poor Metabolizers of Cytochrome P450 2C19 in Esophagus Cancer, Stomach Cancer, Lung Cancer and Bladder Cancer in Chinese Population. *World J Gastroenterol* (2004) 10(13):1961–3. doi:10.3748/wjg.v10.i13.463
28. Basma HA, Kobeissi LH, Jabbour ME, Moussa MA, Dhaini HR. CYP2E1 and NQO1 Genotypes and Bladder Cancer Risk in a Lebanese Population. *Int J Mol Epidemiol Genet* (2013) 4(4):207–17.
29. Yin X, Xiong W, Wang Y, Tang W, Xi W, Qian S, et al. Association of CYP2E1 Gene Polymorphisms With Bladder Cancer Risk: A Systematic Review and Meta-Analysis. *Medicine (Baltimore)* (2018) 97(39):e11910. doi:10.1097/md.00000000000011910
30. Deng XD, Gao Q, Zhang B, Zhang LX, Zhang W, Er ZEM, et al. Functional RsaI/PstI Polymorphism in Cytochrome P450 2E1 Contributes to Bladder Cancer Susceptibility: Evidence From a Meta-Analysis. *Asian Pac J Cancer Prev* (2014) 15(12):4977–82. doi:10.7314/apjcp.2014.15.12.4977
31. Di Pietro G, Magno LAV, Rios-Santos F. Glutathione S-Transferases: An Overview in Cancer Research. *Expert Opin Drug Metab Toxicol* (2010) 6(2):153–70. doi:10.1517/17425250903427980
32. Yu C, Hequn C, Longfei L, Long W, Zhi C, Feng Z, et al. GSTM1 and GSTT1 Polymorphisms Are Associated With Increased Bladder Cancer Risk: Evidence From Updated Meta-Analysis. *Oncotarget* (2017) 8(2):3246–58. doi:10.18632/oncotarget.13702
33. Yu Y, Li X, Liang C, Tang J, Qin Z, Wang C, et al. The Relationship Between GSTA1, GSTM1, GSTP1, and GSTT1 Genetic Polymorphisms and Bladder Cancer Susceptibility: A Meta-Analysis. *Medicine (Baltimore)* (2016) 95(37):e4900. doi:10.1097/md.0000000000004900
34. Ouerhani S, Tebourski F, Slama MRB, Marrakchi R, Rabeh M, Hassine LB, et al. The Role of Glutathione Transferases M1 and T1 in Individual Susceptibility to Bladder Cancer in a Tunisian Population. *Ann Hum Biol* (2006) 33(5–6):529–35. doi:10.1080/03014460600907517
35. Taioli E, Raimondi S. Genetic Susceptibility to Bladder Cancer. *The Lancet* (2005) 366(9486):610–2. doi:10.1016/s0140-6736(05)67115-2
36. Huang WW, Chen DK, Li LJ, Pan QW, Bao WS. Glutathione S-Transferase M1 and T1 Null Genotypes and Bladder Cancer Risk: A Meta-Analysis in a Single Ethnic Group. *J Cancer Res Ther* (2018) 14:S993–7. doi:10.4103/0973-1482.191067
37. Wang M, Chu H, Lv Q, Wang L, Yuan L, Fu G, et al. Cumulative Effect of Genome-Wide Association Study-Identified Genetic Variants for Bladder Cancer. *Int J Cancer* (2014) 135(11):2653–60. doi:10.1002/ijc.28898
38. Kang HW, Song PH, Ha YS, Kim WT, Kim YJ, Yun SJ, et al. Glutathione S-Transferase M1 and T1 Polymorphisms: Susceptibility and Outcomes in Muscle Invasive Bladder Cancer Patients. *Eur J Cancer* (2013) 49(14):3010–9. doi:10.1016/j.ejca.2013.05.019
39. Dudek AM, Grotenhuis AJ, Vermeulen SH, Kiemeny LALM, Verhaegh GW. Urinary Bladder Cancer Susceptibility Markers. What Do We Know About Functional Mechanisms? *Int J Mol Sci* (2013) 14(6):12346–66. doi:10.3390/ijms140612346
40. Yuan JM, Chan KK, Coetzee GA, Castela JE, Watson MA, Bell DA, et al. Genetic Determinants in the Metabolism of Bladder Carcinogens in Relation to Risk of Bladder Cancer. *Carcinogenesis* (2008) 29(7):1386–93. doi:10.1093/carcin/bgn136
41. Schnakenberg E, Breuer R, Werdin R, Dreikorn K, Schloot W. Susceptibility Genes: GSTM1 and GSTM3 as Genetic Risk Factors in Bladder Cancer. *Cytogenet Genome Res* (2000) 91(1–4):234–8. doi:10.1159/000056851
42. Törüner GA, Akyerli C, Uçar A, Akı T, Atsu N, Ozen H, et al. Polymorphisms of Glutathione S-Transferase Genes (GSTM1, GSTP1 and GSTT1) and Bladder Cancer Susceptibility in the Turkish Population. *Arch Toxicol* (2001) 75(8):459–64. doi:10.1007/s002040100268
43. Zhang R, Xu G, Chen W, Zhang W. Genetic Polymorphisms of Glutathione S-Transferase M1 and Bladder Cancer Risk: A Meta-Analysis of 26 Studies. *Mol Biol Rep* (2011) 38(4):2491–7. doi:10.1007/s11033-010-0386-6
44. Zhou T, Li HY, Xie WJ, Zhong Z, Zhong H, Lin ZJ. Association of Glutathione S-Transferase Gene Polymorphism With Bladder Cancer Susceptibility. *BMC Cancer* (2018) 18(1):1088. doi:10.1186/s12885-018-5014-1
45. García-Closas M, Malats N, Silverman D, Dosemeci M, Kogevinas M, Hein DW, et al. NAT2 Slow Acetylation, GSTM1 Null Genotype, and Risk of Bladder Cancer: Results From the Spanish Bladder Cancer Study and Meta-Analyses. *The Lancet* (2005) 366(9486):649–59. doi:10.1016/s0140-6736(05)67137-1
46. Johns LE, Houlston RS. Glutathione S-Transferase Micro1 (GSTM1) Status and Bladder Cancer Risk: A Meta-Analysis. *Mutagenesis* (2000) 15(5):399–404. doi:10.1093/mutage/15.5.399
47. Lee SJ, Cho SH, Park SK, Kim SW, Park MS, Choi HY, et al. Combined Effect of Glutathione S-Transferase M1 and T1 Genotypes on Bladder Cancer Risk. *Cancer Lett* (2002) 177(2):173–9. doi:10.1016/s0304-3835(01)00820-5
48. Safarinejad MR, Safarinejad S, Shafiei N, Safarinejad S. Association of Genetic Polymorphism of Glutathione S-Transferase (GSTM1, GSTT1, GSTP1) With Bladder Cancer Susceptibility. *Urol Oncol Semin Original Invest* (2013) 31(7):1193–203. doi:10.1016/j.urolonc.2011.11.027
49. Zhang R, Xu G, Chen W, Zhang W. Genetic Polymorphisms of Glutathione S-Transferase P1 and Bladder Cancer Susceptibility in a Chinese Population. *Genet Test Mol Biomarkers* (2011) 15(1–2):85–8. doi:10.1089/gtmb.2010.0162
50. Song Y, Chen J, Liu K, Zhou K, Lu Y, Wang X, et al. Glutathione S-Transferase Pi 1 (GSTP1) Gene 313 A/G (Rs1695) Polymorphism Is Associated With the Risk of Urinary Bladder Cancer: Evidence From a Systematic Review and Meta-Analysis Based on 34 Case-Control Studies. *Gene* (2019) 719:144077. doi:10.1016/j.gene.2019.144077
51. Pandith AA, Lateef A, Shah Nawaz S, Hussain A, Malla TM, Azad N, et al. GSTP1 Gene Ile105Val Polymorphism Causes an Elevated Risk for Bladder Carcinogenesis in Smokers. *Asian Pac J Cancer Prev* (2013) 14(11):6375–8. doi:10.7314/apjcp.2013.14.11.6375
52. Wang Z, Xue L, Chong T, Li H, Chen H, Wang Z. Quantitative Assessment of the Association Between Glutathione S-Transferase P1 Ile105Val Polymorphism and Bladder Cancer Risk. *Tumor Biol* (2013) 34(3):1651–7. doi:10.1007/s13277-013-0698-y
53. Wu K, Wang X, Xie Z, Liu Z, Lu Y. Glutathione S-Transferase P1 Gene Polymorphism and Bladder Cancer Susceptibility: An Updated Analysis. *Mol Biol Rep* (2013) 40(1):687–95. doi:10.1007/s11033-012-2109-7
54. Mittal RD, Srivastava DSL, Mandhani A, Mittal B. Genetic Polymorphism of Drug Metabolizing Enzymes (CYP2E1, GSTP1) and Susceptibility to Bladder Cancer in North India. *Asian Pac J Cancer Prev* (2005) 6(1):6–9.
55. Kellen E, Hemelt M, Broberg K, Golka K, Kristensen VN, Hung RJ, et al. Pooled Analysis and Meta-Analysis of the Glutathione S-Transferase P1 Ile 105Val Polymorphism and Bladder Cancer: A HuGE-GSEC Review. *Am J Epidemiol* (2007) 165(11):1221–30. doi:10.1093/aje/kwm003
56. Djukic T, Simic T, Radic T, Matic M, Pljesa-Ercegovac M, Suvakov S, et al. GSTO1*C/GSTO2*G Haplotype Is Associated With Risk of Transitional Cell Carcinoma of Urinary Bladder. *Int Urol Nephrol* (2015) 47(4):625–30. doi:10.1007/s11255-015-0933-0
57. Ma QW, Lin GF, Chen JG, Shen JH. Polymorphism of Glutathione S-Transferase T1, M1 and P1 Genes in a Shanghai population: Patients With Occupational or Non-Occupational Bladder Cancer. *Biomed Environ Sci* (2002) 15(3):253–60.
58. Matic M, Dragicevic B, Pekmezovic T, Suvakov S, Savic-Radojevic A, Pljesa-Ercegovac M, et al. Common Polymorphisms in GSTA1, GSTM1 and GSTT1 Are Associated With Susceptibility to Urinary Bladder Cancer in Individuals From Balkan Endemic Nephropathy Areas of Serbia. *Tohoku J Exp Med* (2016) 240(1):25–30. doi:10.1620/tjem.240.25
59. Goerlitz D, El Daly M, Abdel-Hamid M, Saleh DA, Goldman L, El Kafrawy S, et al. GSTM1, GSTT1 Null Variants, and GPX1 Single Nucleotide Polymorphism Are Not Associated With Bladder Cancer Risk in Egypt. *Cancer Epidemiol Biomarkers Prev* (2011) 20(7):1552–4. doi:10.1158/1055-9965.epi-10-1306
60. Chen YC, Xu L, Guo YLL, Su HJJ, Smith TJ, Ryan LM, et al. Polymorphisms in GSTT1 and P53 and Urinary Transitional Cell Carcinoma in South-Western Taiwan: A Preliminary Study. *Biomarkers* (2004) 9(4–5):386–94. doi:10.1080/13547500400010122
61. Gong M, Dong W, An R. Glutathione S-Transferase T1 Polymorphism Contributes to Bladder Cancer Risk: A Meta-Analysis Involving 50 Studies. *DNA Cel Biol* (2012) 31(7):1187–97. doi:10.1089/dna.2011.1567

62. Sobti RC, Al-Badran AI, Sharma S, Sharma SK, Krishan A, Mohan H. Genetic Polymorphisms of CYP2D6, GSTM1, and GSTT1 Genes and Bladder Cancer Risk in North India. *Cancer Genet Cytogenet* (2005) 156(1):68–73. doi:10.1016/j.cancergencyto.2004.04.001
63. Matic M, Pekmezovic T, Djukic T, Mimic-Oka J, Dragicevic D, Krivic B, et al. GSTA1, GSTM1, GSTP1, and GSTT1 Polymorphisms and Susceptibility to Smoking-Related Bladder Cancer: A Case-Control Study. *Urol Oncol Semin Original Invest* (2013) 31(7):1184–92. doi:10.1016/j.urolonc.2011.08.005
64. Sanderson S, Salanti G, Higgins J. Joint Effects of the N-Acetyltransferase 1 and 2 (NAT1 and NAT2) Genes and Smoking on Bladder Carcinogenesis: A Literature-Based Systematic HuGE Review and Evidence Synthesis. *Am J Epidemiol* (2007) 166(7):741–51. doi:10.1093/aje/kwm167
65. Yassine IA, Kobeissi L, Jabbour ME, Dhaini HR. N-Acetyltransferase 1 (NAT1) Genotype: A Risk Factor for Urinary Bladder Cancer in a Lebanese Population. *J Oncol* (2012) 2012:1–10. doi:10.1155/2012/512976
66. Kobeissi LH, Yassine IA, Jabbour ME, Moussa MA, Dhaini HR. Urinary Bladder Cancer Risk Factors: A Lebanese Case-Control Study. *Asian Pac J Cancer Prev* (2013) 14(5):3205–11. doi:10.7314/apjcp.2013.14.5.3205
67. Xu Z, Li X, Qin Z, Xue J, Wang J, Liu Z, et al. Association of N-Acetyltransferase 1 Polymorphism and Bladder Cancer Risk: An Updated Meta-Analysis and Trial Sequential Analysis. *Int J Biol Markers* (2017) 32(3):e297–304. doi:10.5301/ijbm.5000269
68. Hosen MB, Islam J, Salam MA, Islam MF, Hawlader MZH, Kabir Y. N-Acetyltransferase 2 Gene Polymorphism as a Biomarker for Susceptibility to Bladder Cancer in Bangladeshi Population. *Asia Pac J Clin Oncol* (2015) 11(1):78–84. doi:10.1111/ajco.12291
69. Ma QW, Lin GF, Chen JG, Xiang CQ, Guo WC, Golka K, et al. Polymorphism of N-Acetyltransferase 2 (NAT2) Gene Polymorphism in Shanghai Population: Occupational and Non-Occupational Bladder Cancer Patient Groups. *Biomed Environ Sci : BES* (2004) 17(3):291–8.
70. Selinski S. Urinary Bladder Cancer Risk Variants: Recent Findings and New Challenges of GWAS and Confirmatory Studies. *Arch Toxicol* (2014) 88(7):1469–75. doi:10.1007/s00204-014-1297-4
71. Izumi K, Li Y, Ishiguro H, Zheng Y, Yao JL, Netto GJ, et al. Expression of UDP-Glucuronosyltransferase 1A in Bladder Cancer: Association With Prognosis and Regulation by Estrogen. *Mol Carcinog* (2014) 53(4):314–24. doi:10.1002/mc.21978
72. Selinski S, Lehmann ML, Blaszkewicz M, Ovsiannikov D, Moormann O, Guballa C, et al. Rs11892031[A] on Chromosome 2q37 in an Intronic Region of the UGT1A Locus Is Associated With Urinary Bladder Cancer Risk. *Arch Toxicol* (2012) 86(9):1369–78. doi:10.1007/s00204-012-0854-y
73. Hemminki K, Bermejo JL, Ji J, Kumar R. Familial Bladder Cancer and the Related Genes. *Curr Opin Urol* (2011) 21(5):386–92. doi:10.1097/mou.0b013e32834958ff
74. Zhang Y, Sun Y, Chen T, Hu H, Xie W, Qiao Z, et al. Genetic Variations Rs11892031 and Rs401681 Are Associated With Bladder Cancer Risk in a Chinese Population. *Int J Mol Sci* (2014) 15(11):19330–41. doi:10.3390/ijms151119330
75. Rothman N, Garcia-Closas M, Chatterjee N, Malats N, Wu X, Figueroa JD, et al. A Multi-Stage Genome-Wide Association Study of Bladder Cancer Identifies Multiple Susceptibility Loci. *Nat Genet* (2010) 42(11):978–84. doi:10.1038/ng.687
76. Tang W, Fu YP, Figueroa JD, Malats N, Garcia-Closas M, Chatterjee N, et al. Mapping of the UGT1A Locus Identifies an Uncommon Coding Variant That Affects mRNA Expression and Protects From Bladder Cancer. *Hum Mol Genet* (2012) 21(8):1918–30. doi:10.1093/hmg/ddr619
77. Wang J, Wu X, Kamat A, Barton Grossman H, Dinney CP, Lin J. Fluid Intake, Genetic Variants of UDP-Glucuronosyltransferases, and Bladder Cancer Risk. *Br J Cancer* (2013) 108(11):2372–80. doi:10.1038/bjc.2013.190
78. Zhou Q, Lin M, Feng X, Ma F, Zhu Y, Liu X, et al. Targeting CLK3 Inhibits the Progression of Cholangiocarcinoma by Reprogramming Nucleotide Metabolism. *J Exp Med* (2020) 217(8):e20191779. doi:10.1084/jem.20191779
79. Matsuda K, Takahashi A, Middlebrooks CD, Obara W, Nasu Y, Inoue K, et al. Genome-Wide Association Study Identified SNP on 15q24 Associated With Bladder Cancer Risk in Japanese Population. *Hum Mol Genet* (2015) 24(4):1177–84. doi:10.1093/hmg/ddu512
80. Chen DS, Mellman I. Oncology Meets Immunology: The Cancer-Immunity Cycle. *Immunity* (2013) 39(1):1–10. doi:10.1016/j.immuni.2013.07.012
81. Zou J, Wu D, Li T, Wang X, Liu Y, Tan S. Association of PD-L1 Gene Rs4143815 C>G Polymorphism and Human Cancer Susceptibility: A Systematic Review and Meta-Analysis. *Pathol - Res Pract* (2019) 215(2):229–34. doi:10.1016/j.prp.2018.12.002
82. Jaiswal PK, Singh V, Mittal RD. Cytotoxic T Lymphocyte Antigen 4 (CTLA4) Gene Polymorphism With Bladder Cancer Risk in North Indian Population. *Mol Biol Rep* (2014) 41(2):799–807. doi:10.1007/s11033-013-2919-2
83. Wang SS, Hsieh MJ, Ou YC, Chen CS, Li JR, Hsiao PC, et al. Impacts of ICAM-1 Gene Polymorphisms on Urothelial Cell Carcinoma Susceptibility and Clinicopathologic Characteristics in Taiwan. *Tumor Biol* (2014) 35(8):7483–90. doi:10.1007/s13277-014-1934-9
84. Nagarsheth N, Wicha MS, Zou W. Chemokines in the Cancer Microenvironment and Their Relevance in Cancer Immunotherapy. *Nat Rev Immunol* (2017) 17(9):559–72. doi:10.1038/nri.2017.49
85. Kucukgergin C, Isman FK, Dasdemir S, Cakmakoglu B, Sanli O, Gokkusu C, et al. The Role of Chemokine and Chemokine Receptor Gene Variants on the Susceptibility and Clinicopathological Characteristics of Bladder Cancer. *Gene* (2012) 511(1):7–11. doi:10.1016/j.gene.2012.09.011
86. Huang Y, Chen H, Wang J, Bunjhoo H, Xiong W, Xu Y, et al. Relationship Between CCR2-V64i Polymorphism and Cancer Risk: A Meta-Analysis. *Gene* (2013) 524(1):54–8. doi:10.1016/j.gene.2013.04.011
87. Narter KF, Agachan B, Sozen S, Cincin ZB, Isbir T. CCR2-64I Is a Risk Factor for Development of Bladder Cancer. *Genet Mol Res* (2010) 9(2):685–92. doi:10.4238/vol9-2gmr829
88. Singh V, Jaiswal PK, Kapoor R, Kapoor R, Mittal RD. Impact of Chemokines CCR5Δ32, CXCL12G801A, and CXCR2C1208T on Bladder Cancer Susceptibility in North Indian Population. *Tumor Biol* (2014) 35(5):4765–72. doi:10.1007/s13277-014-1624-7
89. Andrew AS, Gui J, Sanderson AC, Mason RA, Morlock EV, Schned AR, et al. Bladder Cancer SNP Panel Predicts Susceptibility and Survival. *Hum Genet* (2009) 125(5–6):527–39. doi:10.1007/s00439-009-0645-6
90. Weng WC, Huang YH, Yang SF, Wang SS, Kuo WH, Hsueh CW, et al. Effect of CD44 Gene Polymorphisms on Risk of Transitional Cell Carcinoma of the Urinary Bladder in Taiwan. *Tumor Biol* (2016) 37(5):6971–7. doi:10.1007/s13277-015-4566-9
91. Kalvakolanu DV. The ‘Yin-Yang’ of Cytokines in Cancer. *Cytokine* (2019) 118:1–2. doi:10.1016/j.cyto.2018.12.013
92. Krajewski W, Karabon L, Partyka A, Tomkiewicz A, Poletajew S, Tukiendorf A, et al. Polymorphisms of Genes Encoding Cytokines Predict the Risk of High-Grade Bladder Cancer and Outcomes of BCG Immunotherapy. *Cent Eur J Immunol* (2020) 45(1):37–47. doi:10.5114/cej.2020.94674
93. Jaiswal PK, Singh V, Srivastava P, Mittal RD. Association of IL-12, IL-18 Variants and Serum IL-18 With Bladder Cancer Susceptibility in North Indian Population. *Gene* (2013) 519(1):128–34. doi:10.1016/j.gene.2013.01.025
94. Zhao T, Wu X, Liu J. Association Between Interleukin-22 Genetic Polymorphisms and Bladder Cancer Risk. *Clinics* (2015) 70(10):686–90. doi:10.6061/clinics/2015(10)05
95. Galdiero MR, Marone G, Mantovani A. Cancer Inflammation and Cytokines. *Cold Spring Harb Perspect Biol* (2018) 10(8):a028662. doi:10.1101/cshperspect.a028662
96. Shi X, Xie X, Xun X, Jia Y, Li S. Associations of IL-10 Genetic Polymorphisms With the Risk of Urologic Cancer: A Meta-Analysis Based on 18,415 Subjects. *SpringerPlus* (2016) 5(1):2034. doi:10.1186/s40064-016-3705-0
97. Chen Z, Zhou W, Dai M, Wu Z, Jin R. Association Between the Interaction Polymorphisms of Interleukin-10 and Smoking on Patients With Bladder Cancer Risk From a Case-Control Study. *Zhonghua Liu Xing Bing Xue Za Zhi Zhonghua Liuxingbingxue Zazhi* (2013) 34(2):183–6.
98. Ahirwar D, Mandhani A, Mittal RD. Interleukin-10 G-1082A and C-819T Polymorphisms as Possible Molecular Markers of Urothelial Bladder Cancer. *Arch Med Res* (2009) 40(2):97–102. doi:10.1016/j.arcmed.2008.11.006
99. Tang T, Xue H, Cui S, Gong Z, Fei Z, Cheng S, et al. Association of Interleukin-23 Receptor Gene Polymorphisms With Risk of Bladder Cancer in Chinese. *Fam Cancer* (2014) 13(4):619–23. doi:10.1007/s10689-014-9731-6
100. Ahirwar DK, Mandhani A, Mittal RD. IL-8 -251 T > A Polymorphism Is Associated With Bladder Cancer Susceptibility and Outcome After BCG

- Immunotherapy in a Northern Indian Cohort. *Arch Med Res* (2010) 41(2): 97–103. doi:10.1016/j.arcmed.2010.03.005
101. de Maturana EL, Ye Y, Calle ML, Rothman N, Urrea V, Kogevinas M, et al. Application of Multi-SNP Approaches Bayesian LASSO and AUC-RF to Detect Main Effects of Inflammatory-Gene Variants Associated With Bladder Cancer Risk. *PLoS One* (2013) 8(12):e83745. doi:10.1371/journal.pone.0083745
 102. Zhou B, Zhang P, Tang T, Liao H, Zhang K, Pu Y, et al. Polymorphisms and Plasma Levels of IL-27: Impact on Genetic Susceptibility and Clinical Outcome of Bladder Cancer. *BMC Cancer* (2015) 15:433. doi:10.1186/s12885-015-1459-7
 103. Zhang M, Tan X, Huang J, Ke Z, Ge Y, Xiong H, et al. Association of 3 Common Polymorphisms of IL-27 Gene With Susceptibility to Cancer in Chinese: Evidence From an Updated Meta-Analysis of 27 Studies. *Med Sci Monitor* (2015) 21:2505–13. doi:10.12659/msm.895032
 104. AHIRWAR DK, Mandhani A, Dharaskar A, Kesarwani P, Mittal RD. Association of Tumour Necrosis Factor-Alpha Gene (T-1031C, C-863A, and C-857T) Polymorphisms With Bladder Cancer Susceptibility and Outcome After Bacille Calmette-Guérin Immunotherapy. *BJU Int* (2009) 104(6):867–73. doi:10.1111/j.1464-410x.2009.08549.x
 105. Gautam KA, Pooja S, Sankhwar SN, Sankhwar PL, Goel A, Rajender S. c.29C>T Polymorphism in the Transforming Growth Factor-B1 (TGFBI) Gene Correlates With Increased Risk of Urinary Bladder Cancer. *Cytokine* (2015) 75(2):344–8. doi:10.1016/j.cyto.2015.05.017
 106. Ohadian Moghadam S, Nowroozi MR. Toll-Like Receptors: The Role in Bladder Cancer Development, Progression and Immunotherapy. *Scand J Immunol* (2019) 90(6):e12818. doi:10.1111/sji.12818
 107. Singh V, Srivastava N, Kapoor R, Mittal RD. Single-Nucleotide Polymorphisms in Genes Encoding Toll-Like Receptor -2, -3, -4, and -9 in a Case-Control Study With Bladder Cancer Susceptibility in a North Indian Population. *Arch Med Res* (2013) 44(1):54–61. doi:10.1016/j.arcmed.2012.10.008
 108. Shen Y, Liu Y, Liu S, Zhang A. Toll-Like Receptor 4 Gene Polymorphisms and Susceptibility to Bladder Cancer. *Pathol Oncol Res* (2013) 19(2):275–80. doi:10.1007/s12253-012-9579-8
 109. Shen Y, Bu M, Zhang A, Liu Y, Fu B. Toll-Like Receptor 4 Gene Polymorphism Downregulates Gene Expression and Involves in Susceptibility to Bladder Cancer. *Tumor Biol* (2015) 36(4):2779–84. doi:10.1007/s13277-014-2902-0
 110. Wan B, Tan J, Zeng Q, He LY, Gan Y, Dai YB, et al. 729G/C Polymorphism in Toll-Like Receptor 4 Results in Increased Susceptibility to Bladder Cancer. *Genet Mol Res* (2015) 14(4):15482–7. doi:10.4238/2015.november.30.26
 111. Cheng S, Liu J, Zhang Y, Lin Y, Liu Q, Li H, et al. Association Detection Between Genetic Variants in the MicroRNA Binding Sites of Toll-Like Receptors Signaling Pathway Genes and Bladder Cancer Susceptibility. *Int J Clin Exp Pathol* (2014) 7(11):8118–26.
 112. Köstner K, Denzer N, Müller CSL, Klein R, Tilgen W, Reichrath J. The Relevance of Vitamin D Receptor (VDR) Gene Polymorphisms for Cancer: A Review of the Literature. *Anticancer Res* (2009) 29(9):3511–36.
 113. Mittal RD, Manchanda PK, Bhat S, Bid HK. Association of Vitamin-D Receptor (Fok-I) Gene Polymorphism With Bladder Cancer in an Indian Population. *BJU Int* (2007) 99(4):933–7. doi:10.1111/j.1464-410x.2007.06657.x
 114. Ben Fradj MK, Kallel A, Gargouri MM, Chehida MAB, Sallemi A, Ouane Y, et al. Association of FokI Polymorphism of Vitamin D Receptor With Urothelial Bladder Cancer in Tunisians: Role of Tobacco Smoking and Plasma Vitamin D Concentration. *Tumor Biol* (2016) 37(5):6197–203. doi:10.1007/s13277-015-4496-6
 115. Bankhead A, McMaster T, Wang Y, Boonstra PS, Palmboos PL. TP63 Isoform Expression Is Linked With Distinct Clinical Outcomes in Cancer. *EBioMedicine* (2020) 51:102561. doi:10.1016/j.ebiom.2019.11.022
 116. Dudek AM, Vermeulen SH, Kolev D, Grotenhuis AJ, Kiemeny LALM, Verhaegh GW. Identification of an Enhancer Region Within the TP63/LEPRL1 Locus Containing Genetic Variants Associated With Bladder Cancer Risk. *Cell Oncol* (2018) 41(5):555–68. doi:10.1007/s13402-018-0393-5
 117. Singh V, Jaiswal PK, Mittal RD. Replicative Study of GWAS TP63C/T, TERTC/T, and SLC14A1C/T With Susceptibility to Bladder Cancer in North Indians: Equal Contribution. *Urol Oncol Semin Original Invest* (2014) 32(8):1209–14. doi:10.1016/j.urolonc.2014.05.013
 118. Wang M, Wang M, Zhang W, Yuan L, Fu G, Wei Q, et al. Common Genetic Variants on 8q24 Contribute to Susceptibility to Bladder Cancer in a Chinese Population. *Carcinogenesis* (2009) 30(6):991–6. doi:10.1093/carcin/bgp091
 119. Wang M, Du M, Ma L, Chu H, Lv Q, Ye D, et al. A Functional Variant in TP63 at 3q28 Associated With Bladder Cancer Risk by Creating an miR-140-5p Binding Site. *Int J Cancer* (2016) 139(1):65–74. doi:10.1002/ijc.29978
 120. Chen CH, Ho CH, Kuan-Hua Huang S, Shen CH, Wu CC, Wang YH. Association Between VEGF Gene Promoter Polymorphisms and Bladder Cancer: An Updated Meta-Analysis. *Cytokine* (2020) 131:155112. doi:10.1016/j.cyto.2020.155112
 121. Jaiswal PK, Tripathi N, Shukla A, Mittal RD. Association of Single Nucleotide Polymorphisms in Vascular Endothelial Growth Factor Gene With Bladder Cancer Risk. *Med Oncol* (2013) 30(2):509. doi:10.1007/s12032-013-0509-8
 122. Song Y, Yang Y, Liu L, Liu X. Association Between Five Polymorphisms in Vascular Endothelial Growth Factor Gene and Urinary Bladder Cancer Risk: A Systematic Review and Meta-Analysis Involving 6671 Subjects. *Gene* (2019) 698:186–97. doi:10.1016/j.gene.2019.02.070
 123. Zhang LF, Ren KW, Zuo L, Zou JG, Song NH, Mi YY, et al. VEGF Gene rs3025039C/T and rs833052C/A Variants Are Associated With Bladder Cancer Risk in Asian Descendants. *J Cell Biochem* (2019) 120(6):10402–12. doi:10.1002/jcb.28324
 124. Song Y, Hu J, Chen Q, Guo J, Zou Y, Zhang W, et al. Association Between Vascular Endothelial Growth Factor Rs699947 Polymorphism and the Risk of Three Major Urologic Neoplasms (Bladder Cancer, Prostate Cancer, and Renal Cell Carcinoma): A Meta-Analysis Involving 11,204 Subjects. *Gene* (2018) 679:241–52. doi:10.1016/j.gene.2018.09.005
 125. Ben Wafi S, Kallel A, Ben Fradj MK, Sallemi A, Ben Rhouma S, Ben Halima M, et al. Haplotype-Based Association of Vascular Endothelial Growth Factor Gene Polymorphisms With Urothelial Bladder Cancer Risk in Tunisian Population. *J Clin Lab Anal* (2018) 32(9):e22610. doi:10.1002/jcla.22610
 126. Wei H, Kamat AM, Aldousari S, Ye Y, Huang M, Dinney CP, et al. Genetic Variations in the Transforming Growth Factor Beta Pathway as Predictors of Bladder Cancer Risk. *PLoS One* (2012) 7(12):e51758. doi:10.1371/journal.pone.0051758
 127. Tung MC, Hsieh MJ, Wang SS, Yang SF, Chen SS, Wang SW, et al. Associations of VEGF-C Genetic Polymorphisms With Urothelial Cell Carcinoma Susceptibility Differ Between Smokers and Non-Smokers in Taiwan. *PLoS One* (2014) 9(3):e91147. doi:10.1371/journal.pone.0091147
 128. Lin A, Yan WH. Heterogeneity of HLA-G Expression in Cancers: Facing the Challenges. *Front Immunol* (2018) 9:2164. doi:10.3389/fimmu.2018.02164
 129. Castelli EC, Mendes-Junior CT, Viana de Camargo JL, Donadi EA. HLA-G Polymorphism and Transitional Cell Carcinoma of the Bladder in a Brazilian Population. *Tissue Antigens* (2008) 72(2):149–57. doi:10.1111/j.1399-0039.2008.01091.x
 130. Chu H, Wang M, Jin H, Lv Q, Wu D, Tong N, et al. EGFR 3'UTR 774T>C Polymorphism Contributes to Bladder Cancer Risk. *Mutagenesis* (2013) 28(1):49–55. doi:10.1093/mutage/ges051
 131. Li X, Gao Y, Zhou H, Xu W, Li P, Zhou J, et al. The Relationship Between Functional Promoter -94 ins/del ATTG Polymorphism in NF-κ B1 Gene and the Risk of Urinary Cancer. *Cancer Biomarkers* (2016) 16(1):11–7. doi:10.3233/cbm-150536
 132. Li P, Gu J, Yang X, Cai H, Tao J, Yang X, et al. Functional Promoter -94 ins/del ATTG Polymorphism in NFKB1 Gene Is Associated With Bladder Cancer Risk in a Chinese Population. *PLoS One* (2013) 8(8):e71604. doi:10.1371/journal.pone.0071604
 133. Tang T, Cui S, Deng X, Gong Z, Jiang G, Wang P, et al. Insertion/Deletion Polymorphism in the Promoter Region of NFKB1 Gene Increases Susceptibility for Superficial Bladder Cancer in Chinese. *DNA Cel Biol* (2010) 29(1):9–12. doi:10.1089/dna.2009.0937
 134. Shen Y, Lu Q, Ye H, Deng Z, Ma L, Zhang Q, et al. Genetic Variant of MAML2 in the NOTCH Signaling Pathway and the Risk of Bladder Cancer: A STROBE-Compliant Study. *Medicine (Baltimore)* (2020) 99(2):e18725. doi:10.1097/md.00000000000018725
 135. Strasser A, Vaux DL. Cell Death in the Origin and Treatment of Cancer. *Mol Cel* (2020) 78(6):1045–54. doi:10.1016/j.molcel.2020.05.014

136. Verim L, Timirci-Kahraman O, Akbulut H, Akbas A, Ozturk T, Turan S, et al. Functional Genetic Variants in Apoptosis-Associated FAS and FASL Genes and Risk of Bladder Cancer in a Turkish Population. In vivo (*Athens, Greece*) (2014) 28(3):397–402.
137. Timirci-Kahraman O, Ozkan NE, Turan S, Farooqi AA, Verim L, Ozturk T, et al. Genetic Variants in the Tumor Necrosis Factor-Related Apoptosis-Inducing Ligand and Death Receptor Genes Contribute to Susceptibility to Bladder Cancer. *Genet Test Mol Biomarkers* (2015) 19(6):309–15. doi:10.1089/gtmb.2015.0050
138. Mittal RD, Srivastava P, Mittal T, Verma A, Jaiswal PK, Singh V, et al. Association of Death Receptor 4, Caspase 3 and 5 Gene Polymorphism With Increased Risk to Bladder Cancer in North Indians. *Eur J Surg Oncol (Ejso)* (2011) 37(8):727–33. doi:10.1016/j.ejso.2011.05.013
139. Wang M, Wang M, Cheng G, Zhang Z, Fu G, Zhang Z. Genetic Variants in the Death Receptor 4 Gene Contribute to Susceptibility to Bladder Cancer. *Mutat Research/Fundamental Mol Mech Mutagenesis* (2009) 661(1–2):85–92. doi:10.1016/j.mrfmmm.2008.11.009
140. Hazra A, Chamberlain RM, Grossman HB, Zhu Y, Spitz MR, Wu X. Death Receptor 4 and Bladder Cancer Risk. *Cancer Res* (2003) 63(6):1157–9.
141. Guirado M, Gil H, Saenz-Lopez P, Reinboth J, Garrido F, Cozar JM, et al. Association Between C13ORF31, NOD2, RIPK2 and TLR10 Polymorphisms and Urothelial Bladder Cancer. *Hum Immunol* (2012) 73(6):668–72. doi:10.1016/j.humimm.2012.03.006
142. Wang M, Zhang Z, Tian Y, Shao J, Zhang Z. A Six-Nucleotide Insertion-Deletion Polymorphism in the CASP8 Promoter Associated With Risk and Progression of Bladder Cancer. *Clin Cancer Res* (2009) 15(7):2567–72. doi:10.1158/1078-0432.ccr-08-2829
143. Gangwar R, Mandhani A, Mittal RD. Caspase 9 and Caspase 8 Gene Polymorphisms and Susceptibility to Bladder Cancer in North Indian Population. *Ann Surg Oncol* (2009) 16(7):2028–34. doi:10.1245/s10434-009-0488-3
144. Hung SC, Wang SS, Li JR, Chen CS, Yang CK, Chiu KY, et al. Effect of HMGB1 Polymorphisms on Urothelial Cell Carcinoma Susceptibility and Clinicopathological Characteristics. *Int J Med Sci* (2018) 15(14):1731–6. doi:10.7150/ijms.27901
145. Wallace SS. Base Excision Repair: A Critical Player in Many Games. *DNA Repair* (2014) 19:14–26. doi:10.1016/j.dnarep.2014.03.030
146. Wang M, Qin C, Zhu J, Yuan L, Fu G, Zhang Z, et al. Genetic Variants of XRCC1, APE1, and ADPRT Genes and Risk of Bladder Cancer. *DNA Cel Biol* (2010) 29(6):303–11. doi:10.1089/dna.2009.0969
147. Mittal RD, Mandal RK, Gangwar R. Base Excision Repair Pathway Genes Polymorphism in Prostate and Bladder Cancer Risk in North Indian Population. *Mech Ageing Dev* (2012) 133(4):127–32. doi:10.1016/j.mad.2011.10.002
148. Mao Y, Xu X, Lin Y, Chen H, Wu J, Hu Z, et al. Quantitative Assessment of the Associations Between XRCC1 Polymorphisms and Bladder Cancer Risk. *World J Surg Oncol* (2013) 11:58. doi:10.1186/1477-7819-11-58
149. Zhang F, Wu JH, Zhao W, Liu HT. XRCC1 Polymorphisms Increase Bladder Cancer Risk in Asians: A Meta-Analysis. *Tumor Biol* (2013) 34(5):2659–64. doi:10.1007/s13277-013-0816-x
150. Fang Z, Chen F, Wang X, Yi S, Chen W, Ye G. XRCC1 Arg194Trp and Arg280His Polymorphisms Increase Bladder Cancer Risk in Asian Population: Evidence From a Meta-Analysis. *PLoS One* (2013) 8(5):e64001. doi:10.1371/journal.pone.0064001
151. Li S, Peng Q, Chen Y, You J, Chen Z, Deng Y, et al. DNA Repair Gene XRCC1 Polymorphisms, Smoking, and Bladder Cancer Risk: A Meta-Analysis. *PLoS One* (2013) 8(9):e73448. doi:10.1371/journal.pone.0073448
152. Liu C, Yin Q, Li L, Jiao G, Wang M, Wang Y. XRCC1 Arg194Trp and Arg280His Polymorphisms in Bladder Cancer Susceptibility: A Meta-Analysis. *Crit Rev Eukaryot Gene Expr* (2013) 23(4):339–54. doi:10.1615/critrevukaryotgeneexpr.2013007781
153. Ricceri F, Guarnera S, Sacerdote C, Polidoro S, Allione A, Fontana D, et al. ERCC1 Haplotypes Modify Bladder Cancer Risk: A Case-Control Study. *DNA Repair* (2010) 9(2):191–200. doi:10.1016/j.dnarep.2009.12.002
154. Yang D, Liu C, Shi J, Wang N, Du X, Yin Q, et al. Association of XRCC1 Arg399Gln Polymorphism With Bladder Cancer Susceptibility: A Meta-Analysis. *Gene* (2014) 534(1):17–23. doi:10.1016/j.gene.2013.10.038
155. Fontana L, Bosviel R, Delort L, Guy L, Chalabi N, Kwiatkowski F, et al. DNA Repair Gene ERCC2, XPC, XRCC1, XRCC3 Polymorphisms and Associations With Bladder Cancer Risk in a French Cohort. *Anticancer Res* (2008) 28(3B):1853–6.
156. Wang C, Sun Y, Han R. XRCC1 Genetic Polymorphisms and Bladder Cancer Susceptibility: A Meta-Analysis. *Urology* (2008) 72(4):869–72. doi:10.1016/j.urology.2007.12.059
157. Gao W, Romkes M, Zhong S, Nukui T, Persad RA, Smith PJB, et al. Genetic Polymorphisms in the DNA Repair Genes XPD and XRCC1, P53 Gene Mutations and Bladder Cancer Risk. *Oncol Rep* (2010) 24(1):257–62. doi:10.3892/or_00000854
158. Zhuo W, Zhang L, Cai L, Zhu B, Chen Z. XRCC1 Arg399Gln Polymorphism and Bladder Cancer Risk: Updated Meta-Analyses Based on 5767 Cases and 6919 Controls. *Exp Biol Med* (2013) 238(1):66–76. doi:10.1258/ebm.2012.012209
159. Dong LM, Zhang XY, Teng H, Li MS, Wang P. Meta-Analysis Demonstrates No Association Between XRCC1 Arg399Gln Polymorphism and Bladder Cancer Risk. *Genet Mol Res* (2014) 13(4):9976–85. doi:10.4238/2014.november.28.2
160. Figueroa JD, Malats N, Rothman N, Real FX, Silverman D, Kogevinas M, et al. Evaluation of Genetic Variation in the Double-Strand Break Repair Pathway and Bladder Cancer Risk. *Carcinogenesis* (2007) 28(8):1788–93. doi:10.1093/carcin/bgm132
161. Mittal RD, Gangwar R, Mandal RK, Srivastava P, Ahirwar DK. Gene Variants of XRCC4 and XRCC3 and Their Association With Risk for Urothelial Bladder Cancer. *Mol Biol Rep* (2012) 39(2):1667–75. doi:10.1007/s11033-011-0906-z
162. Chang CH, Chang CL, Tsai CW, Wu HC, Chiu CF, Wang RF, et al. Significant Association of an XRCC4 Single Nucleotide Polymorphism With Bladder Cancer Susceptibility in Taiwan. *Anticancer Res* (2009) 29(5):1777–82.
163. Chang CH, Chiu CF, Liang SY, Wu HC, Chang CL, Tsai CW, et al. Significant Association of Ku80 Single Nucleotide Polymorphisms With Bladder Cancer Susceptibility in Taiwan. *Anticancer Res* (2009) 29(4):1275–9.
164. Zhi Y, Yu J, Liu Y, Wei Q, Yuan F, Zhou X, et al. Interaction Between Polymorphisms of DNA Repair Genes Significantly Modulated Bladder Cancer Risk. *Int J Med Sci* (2012) 9(6):498–505. doi:10.7150/ijms.4799
165. Gangwar R, Ahirwar D, Mandhani A, Mittal RD. Do DNA Repair Genes OGG1, XRCC3 and XRCC7 Have an Impact on Susceptibility to Bladder Cancer in the North Indian Population? *Mutat Research/Genetic Toxicol Environ Mutagenesis* (2009) 680(1–2):56–63. doi:10.1016/j.mrgentox.2009.09.008
166. Gangwar R, Ahirwar D, Mandhani A, Mittal RD. Influence of XPD and APE1 DNA Repair Gene Polymorphism on Bladder Cancer Susceptibility in North India. *Urology* (2009) 73(3):675–80. doi:10.1016/j.urology.2008.09.043
167. Liu C, Yin Q, Li L, Zhuang YZ, Zu X, Wang Y. APE1 Asp148Glu Gene Polymorphism and Bladder Cancer Risk: A Meta-Analysis. *Mol Biol Rep* (2013) 40(1):171–6. doi:10.1007/s11033-012-2046-5
168. Figueroa JD, Malats N, Real FX, Silverman D, Kogevinas M, Chanock S, et al. Genetic Variation in the Base Excision Repair Pathway and Bladder Cancer Risk. *Hum Genet* (2007) 121(2):233–42. doi:10.1007/s00439-006-0294-y
169. Ma L, Chu H, Wang M, Shi D, Zhong D, Li P, et al. HOGG1 Ser326Cys Polymorphism Is Associated With Risk of Bladder Cancer in a Chinese Population: A Case-Control Study. *Cancer Sci* (2012) 103(7):1215–20. doi:10.1111/j.1349-7006.2012.02290.x
170. Ramaniuk VP, Nikitchenko NV, Savina NV, Kuzhir TD, Rolevich AI, Krasny SA, et al. Polymorphism of DNA Repair Genes OGG1, XRCC1, XPD and ERCC6 in Bladder Cancer in Belarus. *Biomarkers* (2014) 19(6):509–16. doi:10.3109/1354750x.2014.943291
171. Karahalil B, Kocabas NA, Ozçelik T. DNA Repair Gene Polymorphisms and Bladder Cancer Susceptibility in a Turkish Population. *Anticancer Res* (2006) 26(6C):4955–8.
172. Xie H, Gong Y, Dai J, Wu X, Gu J. Genetic Variations in Base Excision Repair Pathway and Risk of Bladder Cancer: A Case-Control Study in the United States. *Mol Carcinogenesis* (2015) 54(1):50–7. doi:10.1002/mc.22073
173. Xing J, Dinney CP, Shete S, Huang M, Hildebrandt MA, Chen Z, et al. Comprehensive Pathway-Based Interrogation of Genetic Variations in the

- Nucleotide Excision DNA Repair Pathway and Risk of Bladder Cancer. *Cancer* (2012) 118(1):205–15. doi:10.1002/cncr.26224
174. Jager M, Blokzijl F, Kuijk E, Bertl J, Vougioukalaki M, Janssen R, et al. Deficiency of Nucleotide Excision Repair Is Associated With Mutational Signature Observed in Cancer. *Genome Res* (2019) 29(7):1067–77. doi:10.1101/gr.246223.118
 175. Martein J, Lans H, Vermeulen W, Hoeijmakers JHJ. Understanding Nucleotide Excision Repair and Its Roles in Cancer and Ageing. *Nat Rev Mol Cell Biol*. (2014) 15(7):465–81. doi:10.1038/nrm3822
 176. Garcia-Closas M, Malats N, Real FX, Welch R, Kogevinas M, Chatterjee N, et al. Genetic Variation in the Nucleotide Excision Repair Pathway and Bladder Cancer Risk. *Cancer Epidemiol Biomarkers Prev* (2006) 15(3):536–42. doi:10.1158/1055-9965.epi-05-0749
 177. Li SX, Dai QS, Chen SX, Zhang SD, Liao XY, Deng X, et al. Xeroderma Pigmentosum Complementation Group D (XPD) Gene Polymorphisms Contribute to Bladder Cancer Risk: A Meta-Analysis. *Tumor Biol* (2014) 35(4):3905–15. doi:10.1007/s13277-013-1519-z
 178. Wu Y, Yang Y. Complex Association Between ERCC2 Gene Polymorphisms, Gender, Smoking and the Susceptibility to Bladder Cancer: A Meta-Analysis. *Tumor Biol* (2014) 35(6):5245–57. doi:10.1007/s13277-014-1682-x
 179. Shao J, Gu M, Xu Z, Hu Q, Qian L. Polymorphisms of the DNA Gene XPD and Risk of Bladder Cancer in a Southeastern Chinese Population. *Cancer Genet Cytogenet* (2007) 177(1):30–6. doi:10.1016/j.cancergencyto.2007.05.005
 180. Sobti RC, Kaur S, Sharma VL, Singh SK, Hosseini SA, Kler R. Susceptibility of XPD and RAD51 Genetic Variants to Carcinoma of Urinary Bladder in North Indian Population. *DNA Cell Biol* (2012) 31(2):199–210. doi:10.1089/dna.2011.1283
 181. Savina NV, Nikitchenko NV, Kuzhir TD, Rolevich AI, Krasny SA, Goncharova RI. The Involvement of ERCC2/XPD and ERCC6/CSB Wild Type Alleles in Protection Against Aging and Cancer. *Curr Aging Sci* (2018) 11(1):45–54. doi:10.2174/1874609810666170707101548
 182. Li C, Jiang Z, Liu X. XPD Lys(751)Gln and Asp(312)Asn Polymorphisms and Bladder Cancer Risk: A Meta-Analysis. *Mol Biol Rep* (2010) 37(1):301–9. doi:10.1007/s11033-009-9693-1
 183. Wu X, Gu J, Grossman HB, Amos CI, Etzel C, Huang M, et al. Bladder Cancer Predisposition: A Multigenic Approach to DNA-Repair and Cell-Cycle-Control Genes. *Am J Hum Genet* (2006) 78(3):464–79. doi:10.1086/500848
 184. Wang M, Gu D, Zhang Z, Zhou J, Zhang Z. XPD Polymorphisms, Cigarette Smoking, and Bladder Cancer Risk: A Meta-Analysis. *J Toxicol Environ Health A* (2009) 72(11–12):698–705. doi:10.1080/15287390902841029
 185. Li C, Jiang Z, Liu X. XPD Lys(751)Gln and Asp(312)Asn Polymorphisms and Bladder Cancer Risk: A Meta-Analysis. *Mol Biol Rep* (2010) 37(1):301–9. doi:10.1007/s11033-009-9693-1
 186. Chang CH, Wang RF, Tsai RY, Wu HC, Wang CH, Tsai CW, et al. Significant Association of XPD Codon 312 Single Nucleotide Polymorphism With Bladder Cancer Susceptibility in Taiwan. *Anticancer Res* (2009) 29(10):3903–7.
 187. Wen H, Ding Q, Fang Z, Xia G, Fang J. Population Study of Genetic Polymorphisms and Superficial Bladder Cancer Risk in Han-Chinese Smokers in Shanghai. *Int Urol Nephrol* (2009) 41(4):855–64. doi:10.1007/s11255-009-9560-y
 188. Chang CH, Chiu CF, Wang HC, Wu HC, Tsai RY, Tsai CW, et al. Significant Association of ERCC6 Single Nucleotide Polymorphisms With Bladder Cancer Susceptibility in Taiwan. *Anticancer Res* (2009) 29(12):5121–4.
 189. Zhang D, Chen C, Fu X, Gu S, Mao Y, Xie Y, et al. A Meta-Analysis of DNA Repair Gene XPC Polymorphisms and Cancer Risk. *J Hum Genet* (2008) 53(1):18–33. doi:10.1007/s10038-007-0215-5
 190. Sankhwar M, Sankhwar SN, Bansal SK, Gupta G, Rajender S. Polymorphisms in the XPC Gene Affect Urinary Bladder Cancer Risk: A Case-Control Study, Meta-Analyses and Trial Sequential Analyses. *Sci Rep* (2016) 6:27018. doi:10.1038/srep27018
 191. Qiu L, Wang Z, Shi X, Wang Z. Associations Between XPC Polymorphisms and Risk of Cancers: A Meta-Analysis. *Eur J Cancer* (2008) 44(15):2241–53. doi:10.1016/j.ejca.2008.06.024
 192. Wang Y, Li Z, Liu N, Zhang G. Association Between CCND1 and XPC Polymorphisms and Bladder Cancer Risk: A Meta-Analysis Based on 15 Case-Control Studies. *Tumor Biol* (2014) 35(4):3155–65. doi:10.1007/s13277-013-1412-9
 193. Dai QS, Hua RX, Zeng RF, Long JT, Peng ZW. XPC Gene Polymorphisms Contribute to Bladder Cancer Susceptibility: A Meta-Analysis. *Tumor Biol* (2014) 35(1):447–53. doi:10.1007/s13277-013-1062-y
 194. Zhang Y, Wang X, Zhang W, Gong S. An Association Between XPC Lys939Gln Polymorphism and the Risk of Bladder Cancer: A Meta-Analysis. *Tumor Biol* (2013) 34(2):973–82. doi:10.1007/s13277-012-0633-7
 195. Dou K, Xu Q, Han X. The Association Between XPC Lys939Gln Gene Polymorphism and Urinary Bladder Cancer Susceptibility: A Systematic Review and Meta-Analysis. *Diagn Pathol* (2013) 8:112. doi:10.1186/1746-1596-8-112
 196. Prakash R, Zhang Y, Feng W, Jasin M. Homologous Recombination and Human Health: The Roles of BRCA1, BRCA2, and Associated Proteins. *Cold Spring Harb Perspect Biol* (2015) 7(4):a016600. doi:10.1101/cshperspect.a016600
 197. Ma Q, Zhao Y, Wang S, Zhang X, Zhang J, Du M, et al. Genetic Polymorphisms of XRCC3 Thr241Met (C18067T, Rs861539) and Bladder Cancer Risk: A Meta-Analysis of 18 Research Studies. *Tumor Biol* (2014) 35(2):1473–80. doi:10.1007/s13277-013-1203-3
 198. He XF, Wei W, Li JL, Shen XL, Ding Dp, Wang SL, et al. Association Between the XRCC3 T241M Polymorphism and Risk of Cancer: Evidence From 157 Case-Control Studies. *Gene* (2013) 523(1):10–9. doi:10.1016/j.gene.2013.03.071
 199. Zhu X, Zhong Z, Zhang X, Zhao X, Xu R, Ren W, et al. DNA Repair Gene XRCC3 T241M Polymorphism and Bladder Cancer Risk in a Chinese Population. *Genet Test Mol Biomarkers* (2012) 16(6):640–3. doi:10.1089/gtmb.2011.0334
 200. Zhu G, Su H, Lu L, Guo H, Chen Z, Sun Z, et al. Association of Nineteen Polymorphisms From Seven DNA Repair Genes and the Risk for Bladder Cancer in Gansu Province of China. *Oncotarget* (2016) 7(21):31372–83. doi:10.18632/oncotarget.9146
 201. Ge Y, Wang Y, Shao W, Jin J, Du M, Ma G, et al. Rare Variants in BRCA2 and CHEK2 Are Associated With the Risk of Urinary Tract Cancers. *Sci Rep* (2016) 6:33542. doi:10.1038/srep33542
 202. Zhang Y, Huang YS, Lin WQ, Zhang SD, Li QW, Hu YZ, et al. NBS1 Glu185Gln Polymorphism and Susceptibility to Urinary System Cancer: A Meta-Analysis. *Tumor Biol* (2014) 35(11):10723–9. doi:10.1007/s13277-014-2346-6
 203. Hou WH, Chen SH, Yu X. Poly-ADP Ribosylation in DNA Damage Response and Cancer Therapy. *Mutat Research/Reviews Mutat Res* (2019) 780:82–91. doi:10.1016/j.mrrev.2017.09.004
 204. Wang LH, Wu CF, Rajasekaran N, Shin YK. Loss of Tumor Suppressor Gene Function in Human Cancer: An Overview. *Cell Physiol Biochem* (2018) 51(6):2647–93. doi:10.1159/000495956
 205. Liu ZH, Bao ED. Quantitative Assessment of the Association Between TP53 Arg72Pro Polymorphism and Bladder Cancer Risk. *Mol Biol Rep* (2013) 40(3):2389–95. doi:10.1007/s11033-012-2319-z
 206. Lin HY, Huang CH, Yu TJ, Wu WJ, Yang MC, Lung FW. p53 Codon 72 Polymorphism Was Associated With Vulnerability, Progression, But Not Prognosis of Bladder Cancer in a Taiwanese Population: An Implication of Structural Equation Modeling to Manage the Risks of Bladder Cancer. *Urol Int* (2011) 86(3):355–60. doi:10.1159/000323599
 207. Hosen MB, Salam MA, Islam MF, Hossain A, Hawlader MZH, Kabir Y. Association of TP53 Gene Polymorphisms With Susceptibility of Bladder Cancer in Bangladeshi Population. *Tumor Biol* (2015) 36(8):6369–74. doi:10.1007/s13277-015-3324-3
 208. Srivastava P, Jaiswal PK, Singh V, Mittal RD. Role of P53 Gene Polymorphism and Bladder Cancer Predisposition in Northern India. *Cancer Biomarkers* (2011) 8(1):21–8. doi:10.3233/dma-2011-0816
 209. Murgel de Castro Santos LE, Trindade Guilhen AC, Alves de Andrade R, Garcia Sumi L, Ward LS. The Role of TP53 PRO47SER and ARG72PRO Single Nucleotide Polymorphisms in the Susceptibility to Bladder Cancer. *Urol Oncol Semin Original Invest* (2011) 29(3):291–4. doi:10.1016/j.urolonc.2009.03.026
 210. Ye Y, Yang H, Grossman HB, Dinney C, Wu X, Gu J. Genetic Variants in Cell Cycle Control Pathway Confer Susceptibility to Bladder Cancer. *Cancer* (2008) 112(11):2467–74. doi:10.1002/cncr.23472

211. Verhaegh GW, Verkleij L, Vermeulen SHHM, den Heijer M, Witjes JA, Kiemeny LA. Polymorphisms in the H19 Gene and the Risk of Bladder Cancer. *Eur Urol* (2008) 54(5):1118–26. doi:10.1016/j.eururo.2008.01.060
212. Li Z, Niu Y. Association Between lncRNA H19 (Rs217727, Rs2735971 and Rs3024270) Polymorphisms and the Risk of Bladder Cancer in Chinese Population. *Minerva Urologica e Nefrologica* (2019) 71(2):161–7. doi:10.23736/s0393-2249.18.03004-7
213. Hua Q, Lv X, Gu X, Chen Y, Chu H, Du M, et al. Genetic Variants in lncRNA H19 Are Associated With the Risk of Bladder Cancer in a Chinese Population. *Mutagenesis* (2016) 31(5):531–8. doi:10.1093/mutage/gew018
214. Zhang Z, Wang S, Wang M, Tong N, Fu G, Zhang Z. Genetic Variants in RUNX3 and Risk of Bladder Cancer: A Haplotype-Based Analysis. *Carcinogenesis* (2008) 29(10):1973–8. doi:10.1093/carcin/bgn183
215. Frazzi R. BIRC3 and BIRC5: Multi-Faceted Inhibitors in Cancer. *Cell Biosci* (2021) 11(1):8. doi:10.1186/s13578-020-00521-0
216. Zhu Y, Li Y, Zhu S, Tang R, Liu Y, Li J. Association of Survivin Polymorphisms With Tumor Susceptibility: A Meta-Analysis. *PLoS One* (2013) 8(9):e74778. doi:10.1371/journal.pone.0074778
217. Mazoochi T, Karimian M, Ehteram H, Karimian A. Survivin c.-31G>C (Rs9904341) Gene Transversion and Urinary System Cancers Risk: A Systematic Review and a Meta-Analysis. *Personalized Med* (2019) 16(1):67–78. doi:10.2217/pme-2018-0053
218. Kawata N, Tsuchiya N, Horikawa Y, Inoue T, Tsuruta H, Maita S, et al. Two Survivin Polymorphisms Are Cooperatively Associated With Bladder Cancer Susceptibility. *Int J Cancer* (2011) 129(8):1872–80. doi:10.1002/ijc.25850
219. Jaiswal PK, Goel A, Mandhani A, Mittal RD. Functional Polymorphisms in Promoter Survivin Gene and Its Association With Susceptibility to Bladder Cancer in North Indian Cohort. *Mol Biol Rep* (2012) 39(5):5615–21. doi:10.1007/s11033-011-1366-1
220. Fan S, Meng J, Zhang L, Zhang X, Liang C. CAV1 Polymorphisms Rs1049334, Rs1049337, Rs7804372 Might Be the Potential Risk in Tumorigenicity of Urinary Cancer: A Systematic Review and Meta-Analysis. *Pathol - Res Pract* (2019) 215(1):151–8. doi:10.1016/j.prp.2018.11.009
221. Cortessis VK, Siegmund K, Xue S, Ross RK, Yu MC. A Case-Control Study of Cyclin D1 CCND1 870A-->G Polymorphism and Bladder Cancer. *Carcinogenesis* (2003) 24(10):1645–50. doi:10.1093/carcin/bgg128
222. Wang L, Habuchi T, Takahashi T, Mitsumori K, Kamoto T, Kakehi Y, et al. Cyclin D1 Gene Polymorphism Is Associated With an Increased Risk of Urinary Bladder Cancer. *Carcinogenesis* (2002) 23(2):257–64. doi:10.1093/carcin/23.2.257
223. Li J, Luo F, Zhang H, Li L, Xu Y. The CCND1 G870A Polymorphism and Susceptibility to Bladder Cancer. *Tumor Biol* (2014) 35(1):171–7. doi:10.1007/s13277-013-1021-7
224. Ito M, Habuchi T, Watanabe J, Higashi S, Nishiyama H, Wang L, et al. Polymorphism Within the Cyclin D1 Gene Is Associated With an Increased Risk of Carcinoma *In Situ* in Patients With Superficial Bladder Cancer. *Urology* (2004) 64(1):74–8. doi:10.1016/j.urol.2004.03.001
225. Lin HH, Ke HL, Hsiao KH, Tsai CW, Wu WJ, Bau DT, et al. Potential Role of CCND1 G870A Genotype as a Predictor for Urothelial Carcinoma Susceptibility and Muscle-Invasiveness in Taiwan. *Chin J Physiol* (2011) 54(3):196–202. doi:10.4077/cjp.2011.amm123
226. Teixeira LK, Reed SI, Cyclin E. Cyclin E Deregulation and Genomic Instability. *Adv Exp Med Biol* (2017) 1042:527–47. doi:10.1007/978-981-10-6955-0_22
227. Chen M, Gu J, Delclos GL, Killary AM, Fan Z, Hildebrandt MAT, et al. Genetic Variations of the PI3K-AKT-mTOR Pathway and Clinical Outcome in Muscle Invasive and Metastatic Bladder Cancer Patients. *Carcinogenesis* (2010) 31(8):1387–91. doi:10.1093/carcin/bgq110
228. Wu X, Ye Y, Kiemeny LA, Sulem P, Rafnar T, Matullo G, et al. Genetic Variation in the Prostate Stem Cell Antigen Gene PSCA Confers Susceptibility to Urinary Bladder Cancer. *Nat Genet* (2009) 41(9):991–5. doi:10.1038/ng.421
229. Wang S, Tang J, Wang M, Yuan L, Zhang Z. Genetic Variation in PSCA and Bladder Cancer Susceptibility in a Chinese Population. *Carcinogenesis* (2010) 31(4):621–4. doi:10.1093/carcin/bgp323
230. Fu YP, Kohaar I, Rothman N, Earl J, Figueroa JD, Ye Y, et al. Common Genetic Variants in the PSCA Gene Influence Gene Expression and Bladder Cancer Risk. *Proc Natl Acad Sci U S A* (2012) 109(13):4974–9. doi:10.1073/pnas.1202189109
231. Wang P, Ye D, Guo J, Liu F, Jiang H, Gong J, et al. Genetic Score of Multiple Risk-Associated Single Nucleotide Polymorphisms Is a Marker for Genetic Susceptibility to Bladder Cancer. *Genes Chromosomes Cancer* (2014) 53(1):98–105. doi:10.1002/gcc.22121
232. Zhao Y, Gui ZL, Liao S, Gao F, Ge YZ, Jia RP. Prostate Stem Cell Antigen Rs2294008 (C>T) Polymorphism and Bladder Cancer Risk: A Meta-Analysis Based on Cases and Controls. *Genet Mol Res* (2014) 13(3):5534–40. doi:10.4238/2014.july.25.7
233. Lee JH, Song HR, Kim HN, Kweon SS, Yun YW, Choi JS, et al. Genetic Variation in PSCA Is Associated With Bladder Cancer Susceptibility in a Korean Population. *Asian Pac J Cancer Prev* (2014) 15(20):8901–4. doi:10.7314/apjcp.2014.15.20.8901
234. Yang J, Li W, Zhang Z, Shen J, Zhang N, Yang M, et al. PSCARs2294008 T Polymorphism Increases the Risk of Bladder Cancer in Bai, Dai, and Han Ethnicity in China and a Potential Mechanism. *Genes Genomics* (2018) 40(5):531–41. doi:10.1007/s13258-018-0653-9
235. Deng S, Ren ZJ, Jin T, Yang B, Dong Q. Contribution of Prostate Stem Cell Antigen Variation Rs2294008 to the Risk of Bladder Cancer. *Medicine (Baltimore)* (2019) 98(16):e15179. doi:10.1097/md.00000000000015179
236. Ketteler J, Klein D. Caveolin-1, Cancer and Therapy Resistance. *Int J Cancer* (2018) 143(9):2092–104. doi:10.1002/ijc.31369
237. Bau DT, Chang CH, Tsai RY, Wang HC, Wang RF, Tsai CW, et al. Significant Association of Caveolin-1 Genotypes With Bladder Cancer Susceptibility in Taiwan. *Chin J Physiol* (2011) 54(3):153–60. doi:10.4077/cjp.2011.amm009
238. Figueroa JD, Ye Y, Siddiq A, Garcia-Closas M, Chatterjee N, Prokunina-Olsson L, et al. Genome-Wide Association Study Identifies Multiple Loci Associated With Bladder Cancer Risk. *Hum Mol Genet* (2014) 23(5):1387–98. doi:10.1093/hmg/ddt519
239. Ma X, Xu H, Zheng T, Li HZ, Shi TP, Wang BJ, et al. DNA Polymorphisms in Exon 1 and Promoter of the CDH1 Gene and Relevant Risk of Transitional Cell Carcinoma of the Urinary Bladder. *BJU Int* (2008) 102(5):633–6. doi:10.1111/j.1464-410x.2008.07634.x
240. Gobin E, Bagwell K, Wagner J, Mysona D, Sandirasegarane S, Smith N, et al. A Pan-Cancer Perspective of Matrix Metalloproteinases (MMP) Gene Expression Profile and Their Diagnostic/Prognostic Potential. *BMC Cancer* (2019) 19(1):581. doi:10.1186/s12885-019-5768-0
241. Tasci AI, Tugcu V, Ozbek E, Ozbay B, Simsek A, Koksak V. A Single-Nucleotide Polymorphism in the Matrix Metalloproteinase-1 Promoter Enhances Bladder Cancer Susceptibility. *BJU Int* (2008) 101(4):503–7. doi:10.1111/j.1464-410x.2007.07315.x
242. Srivastava P, Gangwar R, Kapoor R, Mittal RD. Bladder Cancer Risk Associated With Genotypic Polymorphism of the Matrix Metalloproteinase-1 and 7 in North Indian Population. *Dis Markers* (2010) 29(1):37–46. doi:10.1155/2010/149651
243. Wieczorek E, Reszka E, Jablonowski Z, Jablonska E, Beata Krol M, Grzegorzczak A, et al. Genetic Polymorphisms in Matrix Metalloproteinases (MMPs) and Tissue Inhibitors of MMPs (TIMPs), and Bladder Cancer Susceptibility. *BJU Int* (2013) 112(8):1207–14. doi:10.1111/bju.12230
244. Yan Y, Liang H, Li T, Li M, Li R, Qin X, et al. The MMP-1, MMP-2, and MMP-9 Gene Polymorphisms and Susceptibility to Bladder Cancer: A Meta-Analysis. *Tumor Biol* (2014) 35(4):3047–52. doi:10.1007/s13277-013-1395-6
245. Tao L, Li Z, Lin L, Lei Y, Hongyuan Y, Hongwei J, et al. MMP1, 2, 3, 7, and 9 Gene Polymorphisms and Urinary Cancer Risk: A Meta-Analysis. *Genet Test Mol Biomarkers* (2015) 19(10):548–55. doi:10.1089/gtmb.2015.0123
246. Srivastava P, Kapoor R, Mittal RD. Association of Single Nucleotide Polymorphisms in Promoter of Matrix Metalloproteinase-2, 8 Genes With Bladder Cancer Risk in Northern India. *Urol Oncol Semin Original Invest* (2013) 31(2):247–54. doi:10.1016/j.urolonc.2011.01.001
247. Liao CH, Chang WS, Tsai CW, Hu PS, Wu HC, Hsu SW, et al. Association of Matrix Metalloproteinase-7 Genotypes With the Risk of Bladder Cancer. *In Vivo* (2018) 32(5):1045–50. doi:10.21873/in vivo.11345
248. Li CC, Hsieh MJ, Wang SS, Hung SC, Lin CY, Kuo CW, et al. Impact of Matrix Metalloproteinases 11 Gene Variants on Urothelial Cell Carcinoma Development and Clinical Characteristics. *Int J Environ Res Public Health* (2020) 17(2):475. doi:10.3390/ijerph17020475

249. Tiryakioglu NO, Tunali NE. Association of AKR1C3 Polymorphisms With Bladder Cancer. *Urol J* (2016) 13(2):2615–21.
250. Wolpert BJ, Amr S, Saleh DA, Ezzat S, Gouda I, Loay I, et al. Associations Differ by Sex for Catechol-O-Methyltransferase Genotypes and Bladder Cancer Risk in South Egypt. *Urol Oncol Semin Original Invest* (2012) 30(6):841–7. doi:10.1016/j.urolonc.2010.09.007
251. Chen Y, Yu X, Li T, Yan H, Mo Z. Significant Association of Catechol-O-Methyltransferase Val158Met Polymorphism With Bladder Cancer Instead of Prostate and Kidney Cancer. *Int J Biol Markers* (2016) 31(2):e110–117. doi:10.5301/ijbm.5000204
252. Zhang Y, Wang J. Targeting Uptake Transporters for Cancer Imaging and Treatment. *Acta Pharmaceutica Sinica B* (2020) 10(1):79–90. doi:10.1016/j.apsb.2019.12.005
253. Rafnar T, Vermeulen SH, Sulem P, Thorleifsson G, Aben KK, Witjes JA, et al. European Genome-Wide Association Study Identifies SLC14A1 as a New Urinary Bladder Cancer Susceptibility Gene. *Hum Mol Genet* (2011) 20(21):4268–81. doi:10.1093/hmg/ddr303
254. Garcia-Closas M, Ye Y, Rothman N, Figueroa JD, Malats N, Dinney CP, et al. A Genome-Wide Association Study of Bladder Cancer Identifies a New Susceptibility Locus Within SLC14A1, a Urea Transporter Gene on Chromosome 18q12.3. *Hum Mol Genet* (2011) 20(21):4282–9. doi:10.1093/hmg/ddr342
255. Xu C, Yang X, Wang Y, Ding N, Han R, Sun Y, et al. An Analysis of the Polymorphisms of the GLUT1 Gene in Urothelial Cell Carcinomas of the Bladder and Its Correlation With P53, Ki67 and GLUT1 Expressions. *Cancer Gene Ther* (2017) 24(7):297–303. doi:10.1038/cgt.2017.17
256. Wu L, Chaffee KG, Parker AS, Sicotte H, Petersen GM. Zinc Transporter Genes and Urological Cancers: Integrated Analysis Suggests a Role for ZIP11 in Bladder Cancer. *Tumor Biol* (2015) 36(10):7431–7. doi:10.1007/s13277-015-3459-2
257. Bui HTT, Fujimoto N, Kubo T, Inatomi H, Matsumoto T. SLCO1B1, SLCO2B1, and SLCO1B3 Polymorphisms and Susceptibility to Bladder Cancer Risk. *Cancer Invest* (2014) 32(6):256–61. doi:10.3109/07357907.2014.907421
258. Miyo M, Konno M, Colvin H, Nishida N, Koseki J, Kawamoto K, et al. The Importance of Mitochondrial Folate Enzymes in Human Colorectal Cancer. *Oncol Rep* (2017) 37(1):417–25. doi:10.3892/or.2016.5264
259. Safarinejad MR, Shafiei N, Safarinejad S. Genetic Susceptibility of Methylenetetrahydrofolate Reductase (MTHFR) Gene C677T, A1298C, and G1793A Polymorphisms With Risk for Bladder Transitional Cell Carcinoma in Men. *Med Oncol* (2011) 28(1):S398–412. doi:10.1007/s12032-010-9723-9
260. Kouidhi S, Rouissi K, Khedhiri S, Ouerhani S, Cherif M, Benammar-Elgaaied A. MTHFR Gene Polymorphisms and Bladder Cancer Susceptibility: A Meta-Analysis Including Race, Smoking Status and Tumour Stage. *Asian Pac J Cancer Prev : APJCP* (2011) 12(9):2227–32.
261. You W, Li Z, Jing C, Qian-Wei X, Yu-Ping Z, Weng-Guang L, et al. MTHFR C677T and A1298C Polymorphisms Were Associated With Bladder Cancer Risk and Disease Progression: A Meta-Analysis. *DNA Cel Biol* (2013) 32(5):260–7. doi:10.1089/dna.2012.1931
262. Ouerhani S, Oliveira E, Marrakchi R, Ben Slama MR, Sfaxi M, Ayed M, et al. Methylenetetrahydrofolate Reductase and Methionine Synthase Polymorphisms and Risk of Bladder Cancer in a Tunisian Population. *Cancer Genet Cytogenet* (2007) 176(1):48–53. doi:10.1016/j.cancergencyto.2007.03.007
263. Izmirli M, Inandikloglu N, Abat D, Alptekin D, Demirhan O, Tansuz Z, et al. MTHFR Gene Polymorphisms in Bladder Cancer in the Turkish Population. *Asian Pac J Cancer Prev : APJCP* (2011) 12(7):1833–5.
264. Cai DW, Liu XF, Bu RG, Chen XN, Ning L, Cheng Y, et al. Genetic Polymorphisms of MTHFR and Aberrant Promoter Hypermethylation of the RASSF1A Gene in Bladder Cancer Risk in a Chinese Population. *J Int Med Res* (2009) 37(6):1882–9. doi:10.1177/147323000903700625
265. Lin J, Spitz MR, Wang Y, Schabath MB, Gorlov IP, Hernandez LM, et al. Polymorphisms of Folate Metabolic Genes and Susceptibility to Bladder Cancer: A Case-Control Study. *Carcinogenesis* (2004) 25(9):1639–47. doi:10.1093/carcin/bgh175
266. Beebe-Dimmer JL, Iyer PT, Nriagu JO, Keele GR, Mehta S, Meliker JR, et al. Genetic Variation in Glutathione S-Transferase Omega-1, Arsenic Methyltransferase and Methylenetetrahydrofolate Reductase, Arsenic Exposure and Bladder Cancer: A Case-Control Study. *Environ Health* (2012) 11:43. doi:10.1186/1476-069x-11-43
267. Rouissi K, Ouerhani S, Oliveira E, Marrakchi R, Cherni L, Othman FB, et al. Polymorphisms in One-Carbon Metabolism Pathway Genes and Risk for Bladder Cancer in a Tunisian Population. *Cancer Genet Cytogenet* (2009) 195(1):43–53. doi:10.1016/j.cancergencyto.2009.06.007
268. Zhang K, Zhou B, Zhang P, Zhang Z, Chen P, Pu Y, et al. Genetic Variants in NAMPT Predict Bladder Cancer Risk and Prognosis in Individuals From Southwest Chinese Han Group. *Tumor Biol* (2014) 35(5):4031–40. doi:10.1007/s13277-013-1527-z
269. Ouerhani S, Marrakchi R, Bouhaha R, Ben Slama MR, Sfaxi M, Ayed M, et al. The Role of CYP2D6*4 Variant in Bladder Cancer Susceptibility in Tunisian Patients. *Bull Cancer (Paris)* (2008) 95(2):E1–4. doi:10.1684/bdc.2008.0583
270. Yang Z, Lv Y, Lv Y, Wang Y. Meta-Analysis Shows Strong Positive Association of the TNF- α Gene With Tumor Stage in Bladder Cancer. *Urol Int* (2012) 89(3):337–41. doi:10.1159/000341701
271. Cai J, Yang MY, Hou N, Li X. Association of Tumor Necrosis Factor- α 308G/A Polymorphism With Urogenital Cancer Risk: A Systematic Review and Meta-Analysis. *Genet Mol Res* (2015) 14(4):16102–12. doi:10.4238/2015. december.7.22
272. Hong Z, Wu J, Li Q, Zhang S, Shi Z. Meta-Analysis Reveals No Significant Association Between ERCC6 Polymorphisms and Bladder Cancer Risk. *Int J Biol Markers* (2017) 32(1):e113–7. doi:10.5301/ijbm.5000236
273. Cao M, Mu X, Jiang C, Yang G, Chen H, Xue W. Single-Nucleotide Polymorphisms of GPX1 and MnSOD and Susceptibility to Bladder Cancer: A Systematic Review and Meta-Analysis. *Tumor Biol* (2014) 35(1):759–64. doi:10.1007/s13277-013-1103-6
274. Zhang LF, Zhu LJ, Zhang W, Yuan W, Song NH, Zuo L, et al. MMP-8 C-799 T, Lys460Thr, and Lys87Glu Variants Are Not Related to Risk of Cancer. *BMC Med Genet* (2019) 20(1):162. doi:10.1186/s12881-019-0890-z
275. Pençe S, İ-zbek E, Ozan Tiryakioglu N, Ersoy Tunali N, Pençe HH, Tunali H. rs3918242 Variant Genotype Frequency and Increased TIMP-2 and MMP-9 Expression Are Positively Correlated With Cancer Invasion in Urinary Bladder Cancer. *Cell Mol Biol* (2017) 63(9):46–52. doi:10.14715/cmb/2017.63.9.9
276. Meng J, Wang S, Shen X, Bai Z, Niu Q, Ma D, et al. Polymorphism of MMP-9 Gene Is Not Associated With the Risk of Urinary Cancers: Evidence From an Updated Meta-Analysis. *Pathol - Res Pract* (2018) 214(12):1966–73. doi:10.1016/j.prp.2018.09.011
277. Mavaddat N, Pharoah PDP, Michailidou K, Tyrer J, Brook MN, Bolla MK, et al. Prediction of Breast Cancer Risk Based on Profiling With Common Genetic Variants. *J Natl Cancer Inst* (2015) 107(5):djv036. doi:10.1093/jnci/djv036
278. Marees AT, de Kluiver H, Stringer S, Vorspan F, Curis E, Marie-Claire C, et al. A Tutorial on Conducting Genome-Wide Association Studies: Quality Control and Statistical Analysis. *Int J Methods Psychiatr Res* (2018) 27(2):e1608. doi:10.1002/mpr.1608
279. Tempfer CB, Hefler LA, Schneeberger C, Huber JC. How Valid Is Single Nucleotide Polymorphism (SNP) Diagnosis for the Individual Risk Assessment of Breast Cancer? *Gynecol Endocrinol* (2006) 22(3):155–9. doi:10.1080/09513590600629175
280. Vadva Z, Larsen CE, Propp BE, Trautwein MR, Alford DR, Alper CA. A New Pedigree-Based SNP Haplotype Method for Genomic Polymorphism and Genetic Studies. *Cells* (2019) 8(8):835. doi:10.3390/cells8080835

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