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EDITED BY

Le Li,
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REVIEWED BY

Chang Hu,
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China

Georgia Damoraki,
National and Kapodistrian University of
Athens, Greece

Yaya Xu,
Shanghai Jiao Tong University, China

*CORRESPONDENCE

Wenjie Liu
✉ wenjieliu96@163.com
Peng Xie
✉ 2023020221@usc.edu.cn

†These authors have contributed equally to
this work

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Ketogenic diet in treating sepsis-related acquired weakness: is it friend or foe?

Yanmei Miao¹, LeiYu Xie¹, Shaolin Chen², Xiaoming Zhang³,
Wenjie Liu^{4*†} and Peng Xie^{1,5*†}

¹Department of Critical Care Medicine of the Third Affiliated Hospital (The First People's Hospital of Zunyi), Zunyi Medical University, Zunyi, China, ²Department of Nursing of Affiliated Hospital, Zunyi Medical University, Zunyi, China, ³Department of Molecular and Cellular Biology, Baylor College of Medicine, Houston, TX, United States, ⁴Department of Anesthesiology, The Second Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang, China, ⁵Department of Critical Care Medicine, The Second Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang, China

Background: Sepsis is the body's extreme response to an infection leading to organ dysfunction. Sepsis-related acquired weakness (SAW), a critical illness closely related to metabolic disorders, is characterized by generalized sepsis-induced skeletal muscle weakness, mainly manifesting as symmetrical atrophy of respiratory and limb muscles. Muscle accounts for 40% of the body's total mass and is one of the major sites of glucose and energy absorption. Diet affects skeletal muscle metabolism, which further impacts physiology and signaling pathways. The ketogenic diet (KD) is a high-fat, low-carbohydrate diet that has shown benefits in patients with a variety of neuromuscular disorders. Patients with SAW are in a hypermetabolic state and can consume approximately 1% of total body muscle mass in a day. Due to the decreased total body energy expenditure secondary to starvation, skeletal muscles enter a low metabolic state, with reduced gluconeogenesis and protein consumption and elevated levels of ketone bodies. The latest research suggests that KD may be a new strategy for SAW prevention and treatment, but its mechanism is still unclear.

Objective: Our article aims to explore the effect and mechanism of KD on SAW. And we hope that our review will inspire further research on the KD and foster the exploration of novel strategies for combating SAW.

Methods: Search medical databases and related academic websites, using keywords such as "Sepsis-related acquired weakness," "ketogenic diet," and "skeletal muscle," and select representative literature. Using the method of induction and summary, analyze the effect and mechanism of KD on SAW.

Results: Compared with early nutrition, KD has a more protective effect on SAW, but its mechanism is complex. Firstly, KD can alter energy metabolism substrates to affect SAW's energy metabolism; Secondly, KD can directly act as a signaling molecule to improve mitochondrial function in skeletal muscle and stimulate skeletal muscle regeneration signaling molecules; Thirdly, KD can affect the gut microbiota to exert anti-inflammatory effects, enhance immunity, and thus protect SAW.

Conclusion: KD has a protective effect on SAW, which includes improving energy metabolism, stimulating muscle regeneration signals, optimizing gut microbiota composition, and reducing inflammation and oxidative stress.

KEYWORDS

gut microbiota, ICU-acquired weakness, ketone bodies, ketogenic diet, sepsis-related acquired weakness

Introduction

ICU-acquired weakness (ICU-AW) is a neuromuscular dysfunction that has no other reasonable etiology besides critical illness and its treatment (1). ICU-acquired weakness is typically generalized, symmetrical, and affects limb (proximal more than distal) and respiratory muscles, whereas facial and ocular muscles are spared (2). According to statistics, globally, about 13–20 million patients receive life-supporting treatment in ICUs every year, and more than 1 million critically ill patients experience ICU-AW each year (3). Comorbid sepsis, prolonged motor inhibition, the application of mechanical ventilation, and malnutrition are risk factors for ICU-AW in critically ill patients (4), among which sepsis is an independent risk factor for ICU-AW. Up to 60–100% of patients with sepsis develop ICU-AW, which is known as sepsis-related acquired weakness (SAW) (5). SAW is a critical illness with unique metabolisms (6), for which there has been no effective clinical treatment (6). It often results in prolonged mechanical ventilatory support and hospitalization, increasing patient mortality and seriously affecting patients' quality of life after discharge (7). Therefore, there is an urgent need for effective measures to prevent the occurrence and development of SAW, and to improve the survival rate and quality of life of critically ill patients.

Muscle accounts for 40% of the body's total mass and is the primary site of glucose and energy absorption. For organisms that have high energy requirements, sufficient adenosine triphosphate (ATP) production is essential for muscle contraction and the maintenance of muscle function (8). Variations in diets affect not only muscle metabolism (9) but also signaling pathways for growth, survival, and other functions in skeletal muscle cells. Metabolic flexibility in response to a variety of stimuli, including dietary changes, is an important feature of muscle energetics. Changes in dietary metabolism impact muscle tissues not only through energy supply but also through metabolic intermediates. Specific changes in metabolism, such as activation of ketogenic metabolism, may have protective effects against SAW (10).

The ketogenic diet (KD) is formulated with a high percentage of fats, a low percentage of carbohydrates, and adequate proteins and other nutrients. It has been used for more than 100 years since its initial introduction in 1921 by Dr. Wilder at the Mayo Clinic (11). The KD, which was originally used for controlling epilepsy, especially hard-to-control epilepsy (12), is now being used in the management of obesity (13), polycystic ovary syndrome (14), cancer (15), diabetes mellitus (16), and traumatic brain injury (17), as well as amyotrophic lateral sclerosis (18), mitochondrial myopathies (19), primary

sarcopenia (20), and other pathologic conditions (21). Recent studies have demonstrated that the KD may be a novel strategy for the treatment of SAW (22), but the mechanism is still unclear. In addition, KD has great plasticity in the prevention and treatment of muscle atrophy, but a lot of work is needed to clarify the overall situation. Therefore, this review will review the role and mechanism of KD in SAW, providing new methods and avenues for the prevention or treatment of SAW.

The KD and early nutritional support

The KD

The main types of KDs include the classic KD, the medium-chain triglyceride KD, the modified Atkins diet, and the low-blood-sugar KD. The classic KD: Fat accounts for 70–75% of total calories, protein accounts for 20–25%, carbohydrates account for about 5%. Strictly controlling the intake of carbohydrates can promote the body to enter a ketotic state, relying on fat metabolism to produce ketone bodies for energy supply, which can effectively control epileptic seizures, especially for children with refractory epilepsy; The medium-chain triglyceride KD: emphasizes increasing the intake of medium chain triglycerides (MCT), which can be quickly absorbed by the body and converted into ketone bodies. Compared to ordinary fats, MCT is more likely to produce ketone bodies, making it more suitable for people who need to quickly enter a ketogenic state, such as patients with certain neurological diseases; The modified Atkins diet: Compared to the classic ketogenic diet, the restrictions on carbohydrates are relatively relaxed, allowing for a daily intake of 10–20 grams of net carbohydrates. The fat ratio is relatively reduced, and the protein ratio can be appropriately increased. This is suitable for epilepsy patients who are unwilling to accept a strict ketogenic diet and may also have certain benefits in weight loss and improving metabolic indicators; The low-blood-sugar KD: Pay attention to the glycemic index (GI) of food, choose low GI foods with a balanced ratio of fat and protein, which helps stabilize blood sugar levels, reduce blood sugar fluctuations, and improve insulin sensitivity to some extent (23). After metabolizing the KD *in vivo*, the levels of ketone bodies (KBs) (i.e., acetoacetic acid, β -hydroxybutyric acid [β -HB], and acetone) and fatty acids significantly increase. The synthesis of KBs, also known as ketogenesis, predominantly occurs in hepatocytes and less in astrocytes or renal cells (24). KB utilization occurs in the heart, skeletal muscles, and brain (Figure 1) (25). When there is a deficiency or decrease in dietary carbohydrates, the plasma insulin level decreases while the glucagon level increases, thereby promoting hepatic glycogenolysis and gluconeogenesis, as well as lipolysis in adipose tissue, which is mediated by hormone-sensitive lipase. Restricting carbohydrate intake for 4–7 days leads to failed glycogenolysis and increased ketogenesis with elevated levels of free fatty acids (FFA), acetyl coenzyme A (acetyl-CoA), and KBs (26). When glucose levels are low or carbohydrate consumption is insufficient, KBs become the primary source of energy and mediate cell signaling, post-translational

Abbreviations: SAW, Sepsis-related acquired weakness; KD, Ketogenic diet; ICU-AW, ICU-acquired weakness; ATP, Adenosine triphosphate; MCT, Medium chain triglycerides; GI, Glycemic index; KB, Ketone bodies; FFA, Free fatty acid; acetyl-CoA, Acetyl coenzyme A; FAO, Fatty acid oxidation; ROS, Reactive oxygen species; AMPK, Adenosine monophosphate kinase; TCA, Tricarboxylic acid cycle; PDHC, Pyruvate dehydrogenase complex; HDAC, Histone deacetylase; COVID-19, Corona Virus Disease 2019; NF- κ B, Nuclear factor kappa B; TLR4, Toll-like receptor 4; NLRP3, NLR family, pyrin domain containing protein 3; β -HB, β -hydroxybutyric.

TABLE 1 Summary of literature review on early nutrition.

Author(s)	Title	Study aim	Year	Area
Reid CL, et al.	Muscle wasting and energy balance in critical illness	Determine whether muscle ultrasound is suitable for a larger ICU population ($n = 50$) (28)	2004	Britain
Reintam BA, et al.	Early enteral nutrition in critically ill patients: ESICM clinical practice guidelines	To provide evidence-based guidelines for early enteral nutrition (EEN) during critical illness (29)	2017	Switzerland
Heyland DK, et al.	Optimal amount of calories for critically ill patients: depends on how you slice the cake!	To examine the relationship between the amount of calories administered and mortality ($n = 7,872$) (30)	2011	Canada
Harvey SE, et al.	Trial of the route of early nutritional support in critically ill adults	Exploring the most effective ways to provide early nutritional support for critically ill adult patients ($n = 2,400$) (31)	2014	Britain
Rice TW, et al.	Randomized trial of initial trophic versus full-energy enteral nutrition in mechanically ventilated patients with acute respiratory failure	Testing the initial low volume (i.e., nutritional) enteral nutrition can reduce the occurrence of gastrointestinal intolerance/complications and improve the outcome of initial full energy enteral nutrition in patients with acute respiratory failure ($n = 200$) (32)	2011	America
Heidegger CP, et al.	Optimization of energy provision with supplemental parenteral nutrition in critically ill patients: a randomized controlled clinical trial	Evaluating whether using EN plus parenteral nutrition (SPN) to deliver 100% energy target on ICU days 4–8 can optimize clinical outcomes ($n = 153$) (33)	2013	Switzerland
Reintam Blaser A, et al.	How to avoid harm with feeding critically ill patients: a synthesis of viewpoints of a basic scientist, dietitian and intensivist	It is recommend to recommend the best feeding strategy for critically ill patients (34)	2023	Switzerland
Weijs PJ, et al.	Early high protein intake is associated with low mortality and energy overfeeding with high mortality in non-septic mechanically ventilated critically ill patients	Early protein and energy feeding in critically ill patients is heavily debated ($n = 843$) (35)	2014	the Netherlands
Lee ZY, et al.	The effect of higher versus lower protein delivery in critically ill patients: a systematic review and meta-analysis of randomized controlled trials	To compare the effect of higher versus lower protein delivery (with similar energy delivery between groups) on clinical and patient-centered outcomes in critically ill patients ($n = 1,731$) (36)	2021	Canada
Weijs PJ, et al.	Optimal protein and energy nutrition decreases mortality in mechanically ventilated, critically ill patients: a prospective observational cohort study	Exploring mechanical ventilation and optimal nutritional therapy for critically ill patients ($n = 886$) (37)	2012	Austria
Lindner G, et al.	Hypernatremia in the critically ill is an independent risk factor for mortality	Assessing the prevalence of hypernatremia and its impact on mortality and ICU length of stay (LOS) ($n = 981$) (38)	2007	Austria
Gunst J, et al.	Amino acid supplements in critically ill patients	Taking single amino acid supplements during critical illness and administering glutamine may be harmful (39)	2018	Belgium
Li F, et al.	Role of TFEB Mediated Autophagy, Oxidative Stress, Inflammation, and Cell Death in Endotoxin Induced Myocardial Toxicity of Young and Aged Mice	Study the age dependent mechanism behind susceptibility to sepsis (40)	2016	China
Hermans G, et al.	Effect of tolerating macronutrient deficit on the development of intensive-care unit acquired weakness: a subanalysis of the EPaNIC trial	To evaluate whether late PN and early PN have differences in the quality control of autophagy in muscle weakness and muscle fibers ($n = 600$) (41)	2013	Belgium
Zhang J, et al.	Effect of ketogenic diet on exercise tolerance and transcriptome of gastrocnemius in mice	The impact of KD on muscle strength and exercise endurance (42)	2023	China
Weckx R, et al.	Efficacy and safety of ketone ester infusion to prevent muscle weakness in a mouse model of sepsis-induced critical illness	Study on whether ketoester- β -hydroxybutyrate is a safer alternative for treating SAW ($n = 376$) (43)	2022	Belgium
White RH, et al.	Hormonal and metabolic responses to glucose infusion in sepsis studied by the hyperglycemic glucose clamp technique	Initial hormonal and metabolic responses of sepsis patients to glucose under controlled conditions ($n = 23$) (44)	1987	Australia

(Continued)

TABLE 1 (Continued)

Author(s)	Title	Study aim	Year	Area
Giovannini I, et al.	Respiratory quotient and patterns of substrate utilization in human sepsis and trauma	The lower mean respiratory quotient of septic indicates that they depend generally more than nonseptic trauma patients on fat as an energy substrate and confirms a previously obtained evidence of limited hepatic lipogenesis in sepsis ($n=99$) (45)	1983	Italy
Langley RJ, et al.	An integrated clinico-metabolomic model improves prediction of death in sepsis	Different molecular processes between surviving and deceased sepsis patients may allow for the deployment of more appropriate treatment methods (46)	2013	America
Gill SK, et al.	Increased airway glucose increases airway bacterial load in hyperglycemia	Does the increase in airway glucose caused by hyperglycemia lead to an increase in bacterial load 1 (47)	2016	Britain
Darby AM, et al.	High sugar diets can increase susceptibility to bacterial infection in <i>Drosophila melanogaster</i>	Which aspects of host and pathogen physiology are impacted by diet to influence infection dynamics (48)	2024	America
Petronilho F, et al.	Obesity Exacerbates Sepsis-Induced Oxidative Damage in Organs	Assuming obesity exacerbates oxidative damage to peripheral organs in rats receiving sepsis animal models (49)	2016	Brazil
Beylot M, et al.	Regulation of ketone body flux in septic patients	To assess the effect of sepsis on ketone body (KB) kinetics in humans ($n=19$) (50)	1989	France
Rahmel T, et al.	An open-label, randomized controlled trial to assess a ketogenic diet in critically ill patients with sepsis	Assessing whether KD can induce stable ketosis in critically ill sepsis patients ($n=40$) (51)	2024	Germany
Cabo R, et al.	Effects of Intermittent Fasting on Health, Aging, and Disease	Discussing clinical research protocols for testing intermittent fasting regimens in healthy individuals and patients with metabolic disorders (obesity, insulin deficiency) based on the results of preclinical studies and the latest research (52)	2019	America
Dong TA, et al.	Intermittent Fasting: A Heart Healthy Dietary Pattern?	Evaluated existing literature on the potential cardiovascular benefits of intermittent fasting and proposed directions for future research (53)	2020	America
Dulloo AG, et al.	How dieting makes some fatter: from a perspective of human body composition autoregulation	Exploring whether thin individuals have a higher risk of gaining weight due to dieting compared to obese individuals (54)	2012	Switzerland
Daniel NN, et al.	How Does the Ketogenic Diet Work? Four Potential Mechanisms	Discuss the four most likely mechanisms of ketogenic diet in treating epilepsy (55)	2013	America
Goossens C, et al.	Adipose tissue protects against sepsis-induced muscle weakness in mice: from lipolysis to ketones	Exploring whether pre disease obesity prevents ICU acquired weakness due to increased availability of lipids and ketones (22)	2019	Belgium
De Bruyn A, et al.	Impact of withholding early parenteral nutrition in adult critically ill patients on ketogenesis in relation to outcome	Early parenteral nutrition enhances ketogenic activity in critically ill children ($n=110$) (56)	2021	Belgium
Goossens C, et al.	Altered cholesterol homeostasis in critical illness-induced muscle weakness: effect of exogenous 3-hydroxybutyrate	Assuming that altered cholesterol homeostasis is associated with the development of muscle weakness induced by critical illness, and this pathway can be influenced by 3-hydroxybutyrate ($n=600$) (57)	2021	Belgium

It is generally believed that carbohydrates are one of the main sources of energy for the human body and play an important role in the normal function of skeletal muscles. Patients with sepsis manifest a significant shift in overall metabolism from glucose oxidation to fatty acid oxidation (FAO), with critically ill patients having a lower level of glucose oxidation (44) and enhanced lipid metabolism (45). Significant differences in glucose metabolism and fatty acid β -oxidation pathways were found between sepsis

survivors and non-survivors (46). The mortality rate of sepsis patients with high blood sugar is increased (47). Research has found that high carbohydrate nutrition increases the susceptibility of drosophilae to certain bacterial infections. Drosophilae with high carbohydrate nutrition may develop hyperglycemia, and some pathogens may use excess sugar in the host's body to promote growth during infection (48). In addition, obese mice fed high carbohydrate diets can exacerbate skeletal muscle damage in

sepsis (49). The commercially available enteral feeds are often very high in carbohydrates. Many intravenous drugs are administered as glucose-containing solutions. Parenteral nutrition solutions also contain large amounts of glucose. The resulting high carbohydrate load leads to an increased requirement for insulin and the inhibition of FAO and ketogenesis (50). Therefore, alternative dietary management for patients with SAW may include limiting the glycemic load or changing the dietary composition to allow sufficient fasting time to enhance FAO and ketogenesis. A study found that sepsis patients receiving KD improved hyperglycemia, achieved stable ketosis, and reduced immune dysregulation compared to those receiving high carbohydrate nutrition, and were associated with improvements in clinical indicators (51).

Intermittent fasting is a major mechanism for promoting longevity and mitigating diseases (52). It can effectively activate cytoprotective and cell-repairing pathways, including autophagy, mitochondrial biogenesis, and antioxidant defense (53). However, prolonged fasting ultimately comes at the cost of weight loss (54). Therefore, the beneficial pathways facilitated by fasting can be similarly activated through modified nutritional strategies, such as ketone supplementation (55). The finding that obese patients with sepsis have a lower incidence of SAW than lean patients with sepsis suggests that obesity has a protective effect on SAW, which may be attributed to the presence of KBs (22). Plasma concentrations of β -HB, a type of KB, are increased in pediatric mice with early childhood macronutrient deficiencies in the ICU (56). In a mouse model of sepsis, β -HB treatment was shown to ameliorate SAW (22), suggesting that it has a direct effect on muscle strength. However, the protective effects of the KD or KBs are not solely attributed to their role as energy substrates but also to their function as signaling molecules that impact regenerative pathways (57).

Mechanism of action of the KD on SAW

Energy metabolism

Skeletal muscles require energy for various activities (58), and ATP is required to support muscle contraction. Typically, the majority of ATP is generated through mitochondrial respiration, with anaerobic glycolysis contributing to $\leq 2\%$ of total ATP production. SAW can lead to systemic hypoxia and inflammation, increased reactive oxygen species (ROS), impaired glucose and fatty acid utilization, diminished mitochondrial number and function (59, 60), increased ATP demand, and elevated adenosine monophosphate kinase (AMPK) (60) (Figure 1).

Carbohydrates are the preferred substrate for energy production under healthy conditions. However, in the critically ill population, carbohydrates can cause hyperglycemia, increase mitochondrial oxygen consumption, and increase ROS production, thereby triggering a vicious cycle of mitochondrial damage (61). The impaired translocation of glucose transporter-4 and increased insulin resistance both exacerbate the deleterious effects of hyperglycemia (62). In addition, the metabolic characteristics of SAW patients are impaired ketone production and reduced fatty acid metabolism in the liver and muscles (22). Research has found

that in SAW patients, β -HB is preferentially absorbed by muscles and metabolized into cholesterol precursor mevalonate, rather than TCA metabolites (57). Fat, as an energy substrate, can alter muscle substrate metabolism, from using glycogen to β -HB (63). KD treatment of SAW can increase the release and metabolism of fatty acids into ketone bodies, thereby preventing and treating SAW (22).

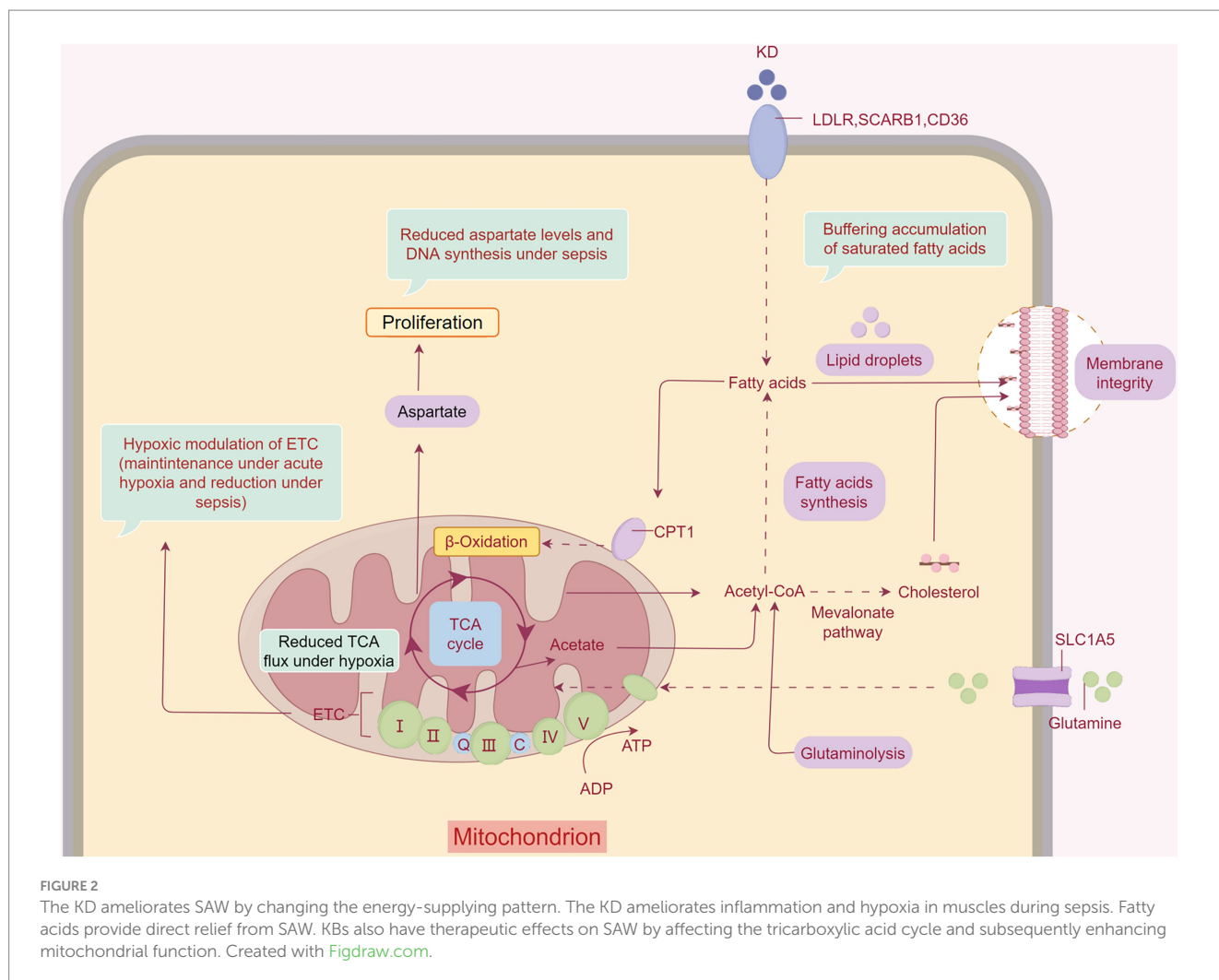
Pyruvate is transferred to mitochondria under aerobic conditions and oxidized to acetyl-CoA via the pyruvate dehydrogenase complex (PDHC) to accelerate aerobic oxidation. The PDHC activity of mononuclear cells in the peripheral blood was significantly lower in patients with sepsis than in healthy controls (64). In addition, rats with sepsis had a 70% reduction in PDHC activity and decreased acetyl-CoA production in skeletal muscle cells, resulting in hypoxia and dysfunction in skeletal muscle cells (Figure 2) (65). KD has been shown to increase pyruvate dehydrogenase levels (66). In mice fed with a KD, pyruvate was oxidatively decarboxylated to acetyl-CoA, catalyzed by pyruvate dehydrogenase (67), and the KD led to increased levels of pyruvate in skeletal muscles (68).

Muscle signaling molecules

Wallace et al. demonstrated the efficacy of the long-term application of the KD in alleviating sarcopenia (20). The KD increases body weight and fat mass (69) but does not change body mass or muscle mass in mice (70). This may be related to its role in providing signaling molecules rather than being solely a metabolic substrate (43). The KD not only promotes the conversion of type IIB skeletal muscle fibers to type IIA fibers and increases the levels of neuromuscular junction remodeling markers, but also facilitates mitochondrial biogenesis, reduces endoplasmic reticulum stress, and enhances protein synthesis and proteasome activity (20). In addition, the KD upregulates genes related to muscle atrophy, such as *Mafbx*, *Murf1*, *Foxo3*, *Lc3b*, and *Klf15*, to reduce muscle mass, fiber cross-sectional area, and grip strength (67).

In humans, a KD can modulate tryptophan metabolism, thereby increasing mitoprotective metabolites. In addition, the anti-inflammatory fatty acids eicosatetraenoic acid and docosahexaenoic acid may be increased. During a KD, there may be increased utilization of carnitine and changes in the tryptophan pathway with decreased quinolinic acid and increased kynurenic acid concentrations. Kynurenic acid is thought to have protective effects on mitochondrial respiration, whereas accumulation of quinolinic acid is associated with mitochondrial dysfunction (71).

Animal studies have found that the administration of β -HB in combination with parenteral nutrition reduces muscle weakness (43). Moreover, β -HB was shown to increase the levels of muscle regeneration markers and decrease the expression of histone deacetylase (HDAC) 4 and 5, which are inhibitors of the muscle regeneration pathways (43). β -OHB injections in healthy adults reduce leucine oxidation and enhance the incorporation of leucine into the skeletal muscles (72). In addition, supplementing SAW mice with β -HB increased markers of early muscle regeneration and reduced the expression of class IIA histone deacetylases (22), which are known factors that inhibit the regeneration pathway by suppressing myocyte enhancer factor 2 (73).



Immunity and the gut microbiota

Immunometabolism plays a critical role in host defense (74). The metabolic determinants of the host response are complex and specific to the types of infection and immune cells involved. A patient's nutritional status alters their gut microbiota and intestinal integrity, affecting inflammation and their host response to sepsis (75). Therefore, it is necessary to consider the balance of these potentially conflicting metabolic demands when designing nutritional interventions.

Sepsis changes the gut microbiota and metabolites. Specifically, the beneficial microorganisms are replaced by the pathogenic ones (76). The gut microbiota is closely intertwined with the host immune system (77) and serves an essential function in host metabolism and resistance against pathogen colonization (78). The disturbance of the gut microbiota occurs in a variety of diseases, including infections and sepsis (79). The gut microbiota may potentially link the immune system and sepsis (80). Recent preclinical evidence has suggested a complex crosstalk between the gut microbial environment and skeletal muscles (81). Lack of gut microbiota found in animal studies can lead to muscle mass loss (82). An increase in the Rikenellaceae of elderly mice was found to be associated with muscle atrophy due to the presence of Riederia bacteria (83). In addition, compared with healthy adult rats,

muscle atrophy rats have a higher ratio of Sutterella to Barnesiella, reduced size of gastrocnemius and triceps, and altered immune function (84). Luckily, the KD can maintain gut microbiota homeostasis. For instance, it increases the beneficial gut microbiota, such as *Akkermansia muciniphila* and *Lactobacillus* (85). Another study in mice and humans found that a KD led to a reduction in *Bifidobacterium* and a decrease in pro-inflammatory Th17 cells in the gut and visceral fat (86). In addition, a KD maintains immune cell homeostasis and promotes cell survival after bacterial infections (86). For instance, it promotes macrophage polarization (87), resulting in a 50% increase in M2-type macrophages and a 50% decrease in M1-type macrophages (88).

Inflammation and gut microbiota

Persistent inflammation is one of the main mechanisms leading to the loss of skeletal muscle mass and function. In the clinical-translational setting, the fundamental importance of ketogenic metabolism for human T cells has already been demonstrated in the context of critically ill COVID-19 patients and, in particular, the improvement of T cell immune metabolism by ketone bodies has been demonstrated in compromised T cells of intensive care patients (89). Due to the profound change in macronutrient composition, a

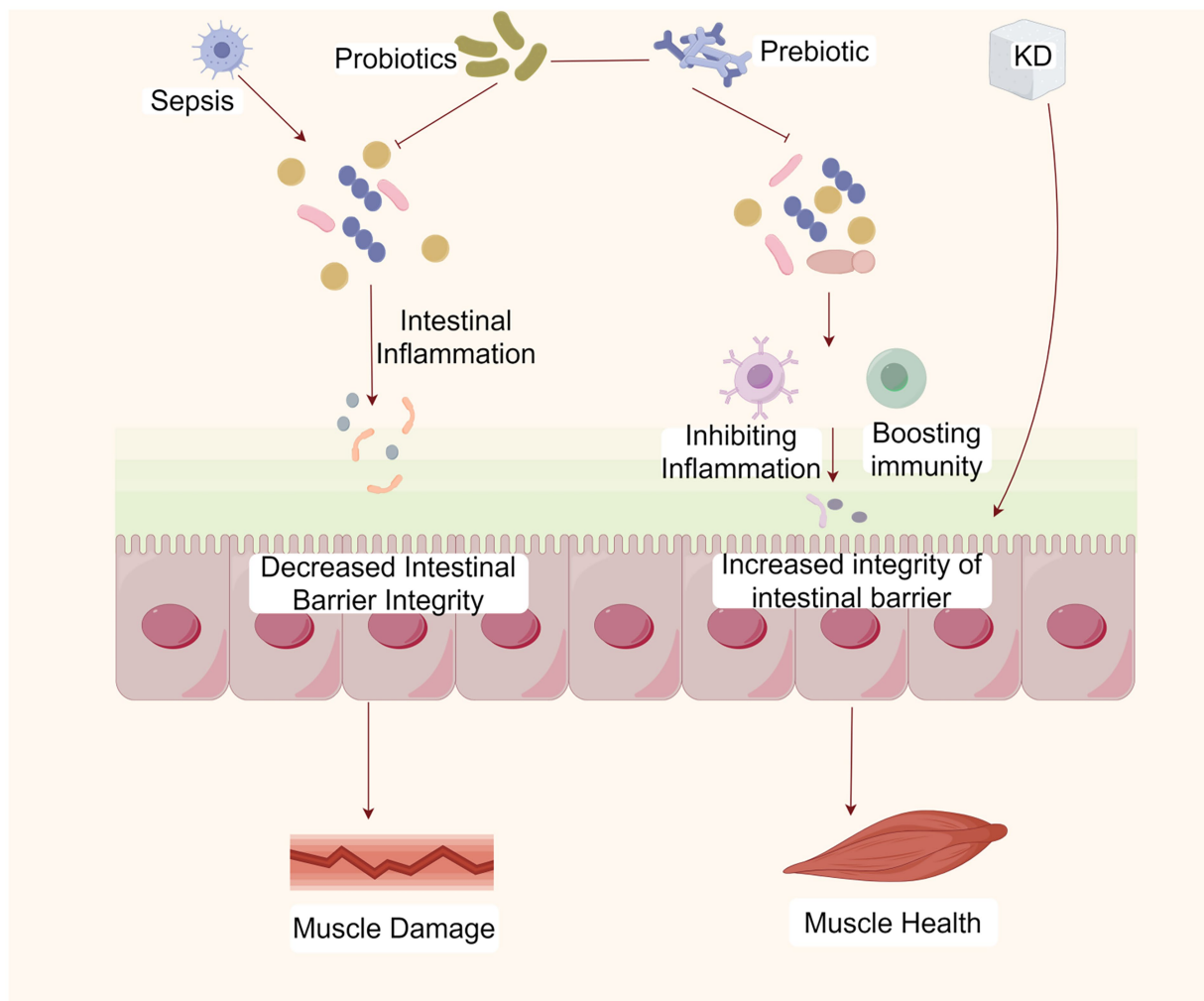


FIGURE 3

The KD ameliorates SAW by maintaining gut microbiota homeostasis. The KD can maintain the homeostasis of the gut microbiota, the integrity of the intestinal barrier, and immune homeostasis, thereby inhibiting inflammation and ameliorating SAW. Created with [Figdraw.com](https://figdraw.com).

ketogenic diet also has significant effects on the cerebrospinal fluid metabolome, which indirectly exerts additional modulating influences on immune cell populations (90).

Intestinal disorders and the loss of gut microbiota diversity impair the integrity of the intestinal barrier and can allow harmful microbial products, such as lipopolysaccharides, to enter the bloodstream, leading to systemic inflammation and metabolic disorders, which can weaken muscle function and reduce muscle mass (91) (Figure 3). The gut microbiota promotes metabolic homeostasis and immune function by strengthening the intestinal barrier. The loss of gut microbiota diseases and diversity can damage the integrity of the intestinal barrier, allowing harmful microbial products such as lipopolysaccharides (LPS) to enter the bloodstream. These harmful substances can cause systemic inflammation, leading to metabolic disorders and decreased muscle function and quality (91). When the intestinal epithelial barrier function is impaired, under lipopolysaccharide stimulation, nuclear factor kappa B (NF- κ B) translocates from the cytoplasm to the nucleus, and stimulates dendritic cells and macrophages to produce pro-inflammatory cytokines and mediators, such as cyclooxygenase-2, TNF- α , inducible nitric oxide synthase, and IL-6, which regulate

intestinal and systemic inflammation (92). And TNF- α regulates the activation of NF- κ B signaling pathway by expressing atrophy related genes, and promotes protein degradation through transcription of ubiquitin proteasome E3 ligase (93). It was reported that the expression level of SOCS3, the target gene of IL-6, was elevated in the skeletal muscles of patients with SAW and that IL-6 mediates sepsis-induced muscle atrophy through the gp130/JAK2/STAT3 pathway (94). Research has found that KD not only alleviates TNF α -induced apoptosis and inflammation of intestinal cells (95), but also reduces the increase in IL-6 levels caused by gut microbiota disorders (96).

Insulin resistance and reduction in skeletal muscles are associated with increased Toll-like receptor 4 (TLR4) expression and signaling in an aging model, which may be due to the development of secondary endotoxemia (97). Activation of the TLR4 signaling pathway results in significant increases in the NF- κ B protein level and the phosphorylation level of c-Jun N-terminal kinase (97). The TLR4 signaling pathway can also induce a systemic inflammatory response by upregulating pro-inflammatory cytokine (IL-6 and TNF- α) levels through a cascade reaction (98). These inflammatory cytokines are involved in regulating muscle atrophy. Inhibition of the production of

TABLE 2 Effects of ketogenic diet on immunity.

Disease	Intestinal flora	Inflammation	References
Cognitive impairment (CI)	Changed the gut microbiota	Th1 cells ↓	Olson et al. (104)
Neuroinflammation	Firmicutes and Proteobacteria ↑ Bacteroidetes ↓	TNF- α , IL-1 β , and IL-6 mRNA ↓	Li et al. (105)
Parkinson's disease (PD)	Bifidobacterium ↓	Th17 cells ↓	Ang et al. (106)
Parkinson's disease (PD)	Citrobacter, Desulfovibrio, Lactobacillus, and Ruminococcus ↓	TNF- α , IL-1 β , and IL-6 ↓	Jiang et al. (107)
Herpes simplex virus type 1 (HSV-1), Infection-associated herpes simplex encephalitis (HSE)	Lactobacillus and <i>Akkermansia muciniphila</i> ↑	TNF α , IL-6, and NOS2 ↓	Shan et al. (108)
Alzheimer's disease (AD)	Proteobacteria, Enterobacteriales ↑	TNF- α and IL-1 β mRNA ↓	Park et al. (109)
Drug-resistant epilepsy	Actinobacteria, Bifidobacteria ↑, and Proteobacteria ↓	Anti-inflammatory	Rohwer et al. (110)
Acute pancreatitis	Enterobacteriales ↑	IL-1 α , IFN- γ ↓	Xia et al. (111)
Drug-resistant epilepsy	Bifidobacteria ↓	IL-17A, IL-17C, TNE, IL-12B, IL-18R1, and GDNF ↓	Dahlin et al. (112)
Sepsis	Intestinal epithelial cells ↓	IL-1 β , IL-6, and TNF- α ↑	Quan et al. (113)
Critical illnesses	Bacilli, Lactobacillales ↑	Immune ↓	Xu et al. (114)
Sarcopenia	Gut dysbiosis	Immune response ↓ and promoting inflammation	Nardone et al. (115)

these inflammatory cytokines by modulating the gut microbiota can alleviate muscle atrophy (99). Many studies have now confirmed that a KD improves the inflammatory environment by regulating the gut microbiota (Table 2). A KD upregulates antioxidant and anti-inflammatory pathways (100). For example, Ketones are anti-inflammatory and can suppress chronic low-grade inflammation by inhibiting the NLRP3 inflammasome (101). At the same time, however, the antiviral immune response of gamma/delta T cells is enhanced by a ketogenic diet, and the use of ketone bodies has been postulated as an antiviral therapy option (102). And β -HB inhibits the formation of NLRP3 inflammatory vesicles and prevents the release of pro-inflammatory cytokines (103).

Conclusion and outlook

SAW involves unique metabolic entities compared to other critical illnesses. Therefore, optimal metabolic and nutritional management strategies may differ between critically ill patients with SAW and non-SAW. Increasing evidence suggests that transitioning from a carbohydrate-centric metabolism to one that prioritizes lipid metabolism may offer protective effects against SAW. This involves mechanisms like maintaining mitochondrial homeostasis, producing anti-inflammatory effects, and regulating immune homeostasis and gut microbiota. However, the benefits of a KD on SAW remain controversial and need validation with further clinical and basic studies. In addition, this article is a narrative review and lacks a systematic retrieval strategy in the literature collection process, requiring more systematic evidence summary and analysis.

Author contributions

YM: Conceptualization, Data curation, Investigation, Methodology, Project administration, Software, Supervision,

Writing – original draft, Writing – review & editing. LX: Conceptualization, Methodology, Project administration. SC: Project administration, Supervision, Writing – review & editing. XZ: Investigation, Writing – original draft. WL: Data curation, Resources, Writing – review & editing. PX: Formal analysis, Funding acquisition, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Stevens RD, Marshall SA, Cornblath DR, Hoke A, Needham DM, de Jonghe B, et al. A framework for diagnosing and classifying intensive care unit-acquired weakness. *Crit Care Med.* (2009) 37:S299–308. doi: 10.1097/CCM.0b013e3181b6e6f7
- Piva S, Fagoni N, Latronico N. Intensive care unit-acquired weakness: unanswered questions and targets for future research. *F1000Res.* (2019):F1000 Faculty Rev-508:8. doi: 10.12688/f1000research.17376.1
- Fan E, Cheek F, Chlan L, Gosselink R, Hart N, Herridge MS, et al. An official American Thoracic Society clinical practice guideline: the diagnosis of intensive care unit-acquired weakness in adults. *Am J Respir Crit Care Med.* (2014) 190:1437–46. doi: 10.1164/rccm.201411-2011ST
- Ozdemir M, Bomkamp MP, Hyatt HW, Smuder AJ, Powers SK. Intensive care unit acquired weakness is associated with rapid changes to skeletal muscle Proteostasis. *Cells (Basel).* (2022) 11:4005. doi: 10.3390/cells11244005
- Mitobe Y, Morishita S, Ohashi K, Sakai S, Uchiyama M, Abeywickrama H, et al. Skeletal muscle index at intensive care unit admission is a predictor of intensive care unit-acquired weakness in patients with Sepsis. *J Clin Med Res.* (2019) 11:834–41. doi: 10.14740/jocmr4027
- Koutroulis I, Batabyal R, McNamara B, Ledda M, Hoptay C, Freishtat RJ. Sepsis Immunometabolism: from defining Sepsis to understanding how energy production affects immune response. *Crit Care Explor.* (2019) 1:e61:e0061. doi: 10.1097/CCE.0000000000000061
- Tortuyaux R, Davion JB, Jourdain M. Intensive care unit-acquired weakness: questions the clinician should ask. *Rev Neurol (Paris).* (2022) 178:84–92. doi: 10.1016/j.neurol.2021.12.007
- Chadt A, Al-Hasani H. Glucose transporters in adipose tissue, liver, and skeletal muscle in metabolic health and disease. *Pflugers Arch.* (2020) 472:1273–98. doi: 10.1007/s00424-020-02417-x
- Yakupova EI, Bocharnikov AD, Plotnikov EY. Effects of ketogenic diet on muscle metabolism in health and disease. *Nutrients.* (2022) 14:14. doi: 10.3390/nu14183842
- Zhu H, Bi D, Zhang Y, Kong C, Du J, Wu X, et al. Ketogenic diet for human diseases: the underlying mechanisms and potential for clinical implementations. *Signal Transduct Target Ther.* (2022) 7:11. doi: 10.1038/s41392-021-00831-w
- McGaugh E, Barthel B. A review of ketogenic diet and lifestyle. *Mo Med.* (2022) 119:84–8.
- Wells J, Swaminathan A, Paseka J, Hanson C. Efficacy and safety of a ketogenic diet in children and adolescents with refractory epilepsy—a review. *Nutrients.* (2020) 12:1809. doi: 10.3390/nu12061809
- Bruci A, Tuccinardi D, Tozzi R, Balena A, Santucci S, Frontani R, et al. Very low-calorie ketogenic diet: a safe and effective tool for weight loss in patients with obesity and mild kidney failure. *Nutrients.* (2020) 12:12. doi: 10.3390/nu12020333
- Mavropoulos JC, Yancy WS, Hepburn J, Westman EC. The effects of a low-carbohydrate, ketogenic diet on the polycystic ovary syndrome: a pilot study. *Nutr Metab (Lond).* (2005) 2:35. doi: 10.1186/1743-7075-2-35
- Yang L, TeSlaa T, Ng S, Nofal M, Wang L, Lan T, et al. Ketogenic diet and chemotherapy combine to disrupt pancreatic cancer metabolism and growth. *Med.* (2022) 3:119–136.e8. doi: 10.1016/j.medj.2021.12.008
- Accurso A, Bernstein RK, Dahlqvist A, Draznin B, Feinman RD, Fine EJ, et al. Dietary carbohydrate restriction in type 2 diabetes mellitus and metabolic syndrome: time for a critical appraisal. *Nutr Metab (Lond).* (2008) 5:9. doi: 10.1186/1743-7075-5-9
- Wood TR, Stubbs BJ, Juul SE. Exogenous ketone bodies as promising neuroprotective agents for developmental brain injury. *Dev Neurosci (Basel).* (2019) 40:451–62. doi: 10.1159/000499563
- Capluiere Llopis J, Peralta Chamba T, Carrera Juliá S, Cuedra Ballester M, Drehmer Rieger E, López Rodríguez MM, et al. Therapeutic alternative of the ketogenic Mediterranean diet to improve mitochondrial activity in amyotrophic lateral sclerosis (ALS): a comprehensive review. *Food Sci Nutr.* (2019) 8:23–35. doi: 10.1002/fsn3.1324
- Ahola-Erkkilä S, Carroll CJ, Peltola-Mjösund K, Tulkki V, Mattila I, Seppänen-Laakso T, et al. Ketogenic diet slows down mitochondrial myopathy progression in mice. *Hum Mol Genet.* (2010) 19:1974–84. doi: 10.1093/hmg/ddq076
- Wallace MA, Aguirre NW, Marcotte GR, Marshall AG, Baehr LM, Hughes DC, et al. The ketogenic diet preserves skeletal muscle with aging in mice. *Aging Cell.* (2021) 20:e13322. doi: 10.1111/acel.13322
- Paoli A, Grimaldi K, Toniolo L, Canato M, Bianco A, Fratter A. Nutrition and acne: therapeutic potential of ketogenic diets. *Skin Pharmacol Phys.* (2012) 25:111–7. doi: 10.1159/000336404
- Goossens C, Weckx R, Derde S, Dufour T, Vander Perre S, Pauwels L, et al. Adipose tissue protects against sepsis-induced muscle weakness in mice: from lipolysis to ketones. *Crit Care.* (2019) 23:236. doi: 10.1186/s13054-019-2506-6
- Barzegar M, Afghan M, Tarmahi V, Behtari M, Rahimi Khamaneh S, Raeisi S. Ketogenic diet: overview, types, and possible anti-seizure mechanisms. *Nutr Neurosci.* (2021) 24:307–16. doi: 10.1080/1028415X.2019.1627769
- Puchalska P, Crawford PA. Multi-dimensional roles of ketone bodies in fuel metabolism, signaling, and therapeutics. *Cell Metab.* (2017) 25:262–84. doi: 10.1016/j.cmet.2016.12.022
- Bentourkia M, Tremblay S, Pifferi F, Rousseau J, Lecomte R, Cunnane S. PET study of 11C-acetoacetate kinetics in rat brain during dietary treatments affecting ketosis. *Am J Physiol Endocrinol Metab.* (2009) 296:E796–801. doi: 10.1152/ajpendo.90644.2008
- Paoli A. Ketogenic diet for obesity: friend or foe? *Int J Environ Res Public Health.* (2014) 11:2092–107. doi: 10.3390/ijerph110202092
- Barry D, Ellul S, Watters L, Lee D, Haluska RJ, White R. The ketogenic diet in disease and development. *Int J Dev Neurosci.* (2018) 68:53–8. doi: 10.1016/j.ijdevneu.2018.04.005
- Reid CL, Campbell IT, Little RA. Muscle wasting and energy balance in critical illness. *Clin Nutr.* (2004) 23:273–80. doi: 10.1016/S0261-5614(03)00129-8
- Reintam BA, Starkopf J, Alhazzani W, Berger MM, Casaer MP, Deane AM, et al. Early enteral nutrition in critically ill patients: ESICM clinical practice guidelines. *Intensive Care Med.* (2017) 43:380–98. doi: 10.1007/s00134-016-4665-0
- Heyland DK, Cahill N, Day AG. Optimal amount of calories for critically ill patients: depends on how you slice the cake! *Crit Care Med.* (2011) 39:2619–26. doi: 10.1097/CCM.0b013e318226641d
- Harvey SE, Parrott F, Harrison DA, Bear DE, Segaran E, Beale R, et al. Trial of the route of early nutritional support in critically ill adults. *N Engl J Med.* (2014) 371:1673–84. doi: 10.1056/NEJMoa1409860
- Rice TW, Mogan S, Hays MA, Bernard GR, Jensen GL, Wheeler AP. Randomized trial of initial trophic versus full-energy enteral nutrition in mechanically ventilated patients with acute respiratory failure. *Crit Care Med.* (2011) 39:967–74. doi: 10.1097/CCM.0b013e31820a905a
- Heidegger CP, Berger MM, Graf S, Zingg W, Darmon P, Costanza MC, et al. Optimisation of energy provision with supplemental parenteral nutrition in critically ill patients: a randomised controlled clinical trial. *Lancet.* (2013) 381:385–93. doi: 10.1016/S0140-6736(12)61351-8
- Reintam Blaser A, Rooyackers O, Bear DE. How to avoid harm with feeding critically ill patients: a synthesis of viewpoints of a basic scientist, dietician and intensivist. *Crit Care.* (2023) 27:258. doi: 10.1186/s13054-023-04543-1
- Weijs PJ, Looijaard WG, Beishuizen A, Girbes AR, Oudemans-van SH. Early high protein intake is associated with low mortality and energy overfeeding with high mortality in non-septic mechanically ventilated critically ill patients. *Crit Care.* (2014) 18:701. doi: 10.1186/s13054-014-0701-z
- Lee ZY, Yap C, Hasan MS, Engkasan JP, Barakatun-Nisak MY, Day AG, et al. The effect of higher versus lower protein delivery in critically ill patients: a systematic review and meta-analysis of randomized controlled trials. *Crit Care.* (2021) 25:260. doi: 10.1186/s13054-021-03693-4
- Weijs PJ, Stapel SN, de Groot SD, Driessen RH, de Jong E, Girbes AR, et al. Optimal protein and energy nutrition decreases mortality in mechanically ventilated, critically ill patients: a prospective observational cohort study. *JPEN J Parenter Enteral Nutr.* (2012) 36:60–8. doi: 10.1177/0148607111415109
- Lindner G, Funk GC, Schwarz C, Kneidinger N, Kaider A, Schneeweiss B, et al. Hypernatremia in the critically ill is an independent risk factor for mortality. *Am J Kidney Dis.* (2007) 50:952–7. doi: 10.1053/j.ajkd.2007.08.016
- Gunst J, Vanhorebeek I, Thiessen SE, Van den Bergh G. Amino acid supplements in critically ill patients. *Pharmacol Res.* (2018) 130:127–31. doi: 10.1016/j.phrs.2017.12.007
- Li F, Lang F, Zhang H, Xu L, Wang Y, Hao E. Role of TFEB mediated autophagy, oxidative stress, inflammation, and cell death in endotoxin induced myocardial toxicity of young and aged mice. *Oxidative Med Cell Longev.* (2016) 2016:5380319. doi: 10.1155/2016/5380319
- Hermans G, Casaer MP, Clerckx B, Guiza F, Vanhullebusch T, Derde S, et al. Effect of tolerating macronutrient deficit on the development of intensive-care unit acquired weakness: a subanalysis of the EPaNIC trial. *Lancet Respir Med.* (2013) 1:621–9. doi: 10.1016/S2213-2600(13)70183-8

42. Zhang J, Chen B, Zou K. Effect of ketogenic diet on exercise tolerance and transcriptome of gastrocnemius in mice. *Open Life Sci.* (2023) 18:20220570. doi: 10.1515/biol-2022-0570
43. Weckx R, Goossens C, Derde S, Pauwels L, Vander PS, Van den Berghe G, et al. Efficacy and safety of ketone ester infusion to prevent muscle weakness in a mouse model of sepsis-induced critical illness. *Sci Rep.* (2022) 12:10591. doi: 10.1038/s41598-022-14961-w
44. White RH, Frayn KN, Little RA, Threlfall CJ, Stoner HB, Irving MH. Hormonal and metabolic responses to glucose infusion in sepsis studied by the hyperglycemic glucose clamp technique. *JPEN J Parenter Enteral Nutr.* (1987) 11:345–53. doi: 10.1177/0148607187011004345
45. Giovannini I, Boldrini G, Castagneto M, Sganga G, Nanni G, Pittiruti M, et al. Respiratory quotient and patterns of substrate utilization in human sepsis and trauma. *JPEN J Parenter Enteral Nutr.* (1983) 7:226–30. doi: 10.1177/0148607183007003226
46. Langley RJ, Tsalik EL, van Velkinburgh JC, Glickman SW, Rice BJ, Wang C, et al. An integrated clinico-metabolomic model improves prediction of death in sepsis. *Sci Transl Med.* (2013) 5:195ra95. doi: 10.1126/scitranslmed.3005893
47. Gill SK, Hui K, Farne H, Garnett JP, Baines DL, Moore LS, et al. Increased airway glucose increases airway bacterial load in hyperglycaemia. *Sci Rep.* (2016) 6:27636. doi: 10.1038/srep27636
48. Darby AM, Okoro DO, Aredas S, Frank AM, Pearson WH, Dionne MS, et al. High sugar diets can increase susceptibility to bacterial infection in *Drosophila melanogaster*. *PLoS Pathog.* (2024) 20:e1012447. doi: 10.1371/journal.ppat.1012447
49. Petronilho F, Giustina AD, Nascimento DZ, Zarbato GF, Vieira AA, Florentino D, et al. Obesity exacerbates Sepsis-induced oxidative damage in organs. *Inflammation.* (2016) 39:2062–71. doi: 10.1007/s10753-016-0444-x
50. Beylot M, Guiraud M, Grau G, Bouletreau P. Regulation of ketone body flux in septic patients. *Am J Phys.* (1989) 257:E665–74. doi: 10.1152/ajpendo.1989.257.5.E665
51. Rahmel T, Effinger D, Bracht T, Griep L, Koos B, Sitek B, et al. An open-label, randomized controlled trial to assess a ketogenic diet in critically ill patients with sepsis. *Sci Transl Med.* (2024) 16:n9285. doi: 10.1126/scitranslmed.adn9285
52. de Cabo R, Mattson MP. Effects of intermittent fasting on health, aging, and disease. *N Engl J Med.* (2019) 381:2541–51. doi: 10.1056/NEJMr1905136
53. Dong TA, Sandesara PB, Dhindsa DS, Mehta A, Arneson LC, Dollar AL, et al. Intermittent fasting: a heart healthy dietary pattern? *Am J Med.* (2020) 133:901–7. doi: 10.1016/j.amjmed.2020.03.030
54. Dullloo AG, Jacquet J, Montani J. How dieting makes some fatter: from a perspective of human body composition autoregulation. *Proc Nutr Soc.* (2012) 71:379–89. doi: 10.1017/S0029665112000225
55. Danial NN, Hartman AL, Stafstrom CE, Thio LL. How does the ketogenic diet work? Four potential mechanisms. *J Child Neurol.* (2013) 28:1027–33. doi: 10.1177/0883073813487598
56. De Bruyn A, Langouche L, Vander PS, Gunst J, Van den Berghe G. Impact of withholding early parenteral nutrition in adult critically ill patients on ketogenesis in relation to outcome. *Crit Care.* (2021) 25:102. doi: 10.1186/s13054-021-03519-3
57. Goossens C, Weckx R, Derde S, Vander PS, Derese I, Van Veldhoven PP, et al. Altered cholesterol homeostasis in critical illness-induced muscle weakness: effect of exogenous 3-hydroxybutyrate. *Crit Care.* (2021) 25:252. doi: 10.1186/s13054-021-03688-1
58. Westerblad H, Bruton JD, Katz A. Skeletal muscle: energy metabolism, fiber types, fatigue and adaptability. *Exp Cell Res.* (2010) 316:3093–9. doi: 10.1016/j.yexcr.2010.05.019
59. Puthuchery ZA, Rawal J, McPhail M, Connolly B, Ratnayake G, Chan P, et al. Acute skeletal muscle wasting in critical illness. *JAMA.* (2013) 310:1591. doi: 10.1001/jama.2013.278481
60. Friedrich O, Reid MB, Van den Berghe G, Vanhorebeek I, Hermans G, Rich MM, et al. The sick and the weak: neuropathies/myopathies in the critically ill. *Physiol Rev.* (2015) 95:1025–109. doi: 10.1152/physrev.00028.2014
61. McClave SA, Wischmeyer PE, Miller KR, van Zanten A. Mitochondrial dysfunction in critical illness: implications for nutritional therapy. *Curr Nutr Rep.* (2019) 8:363–73. doi: 10.1007/s13668-019-00296-y
62. Marik PE, Bellomo R. Stress hyperglycemia: an essential survival response! *Crit Care.* (2013) 17:305. doi: 10.1186/cc12514
63. Cox PJ, Kirk T, Ashmore T, Willerton K, Evans R, Smith A, et al. Nutritional ketosis alters fuel preference and thereby endurance performance in athletes. *Cell Metab.* (2016) 24:256–68. doi: 10.1016/j.cmet.2016.07.010
64. Nuzzo E, Berg KM, Andersen LW, Balkema J, Montissol S, Cocchi MN, et al. Pyruvate dehydrogenase activity is decreased in the peripheral blood mononuclear cells of patients with Sepsis. A prospective observational trial. *Ann Am Thorac Soc.* (2015) 12:1662–6. doi: 10.1513/AnnalsATS.201505-267BC
65. Vary TC. Sepsis-induced alterations in pyruvate dehydrogenase complex activity in rat skeletal muscle: effects on plasma lactate. *Shock.* (1996) 6:89–94. doi: 10.1097/00024382-199608000-00002
66. Inui T, Wada Y, Shibuya M, Arai-Ichinoi N, Okubo Y, Endo W, et al. Intravenous ketogenic diet therapy for neonatal-onset pyruvate dehydrogenase complex deficiency. *Brain and Development.* (2022) 44:244–8. doi: 10.1016/j.braindev.2021.11.005
67. Nakao R, Abe T, Yamamoto S, Oishi K. Ketogenic diet induces skeletal muscle atrophy via reducing muscle protein synthesis and possibly activating proteolysis in mice. *Sci Rep.* (2019) 9:19652. doi: 10.1038/s41598-019-56166-8
68. Spriet LL, Tunstall RJ, Watt MJ, Mehan KA, Hargreaves M, Cameron-Smith D. Pyruvate dehydrogenase activation and kinase expression in human skeletal muscle during fasting. *J Appl Physiol.* (1985). (2004) 96:2082–7. doi: 10.1152/jappphysiol.01318.2003
69. Parker BA, Walton CM, Carr ST, Andrus JL, Cheung E, Duplisa MJ, et al. Beta-Hydroxybutyrate elicits favorable mitochondrial changes in skeletal muscle. *Int J Mol Sci.* (2018) 19:19. doi: 10.3390/ijms19082247
70. Saito H, Wada N, Iida K. Isonitrogenous low-carbohydrate diet elicits specific changes in metabolic gene expression in the skeletal muscle of exercise-trained mice. *PLoS One.* (2022) 17:e262875:e0262875. doi: 10.1371/journal.pone.0262875
71. Effinger D, Hirschberger S, Yoncheva P, Schmid A, Heine T, Newels P, et al. A ketogenic diet substantially reshapes the human metabolome. *Clin Nutr.* (2023) 42:1202–12. doi: 10.1016/j.clnu.2023.04.027
72. Nair KS, Welle SL, Halliday D, Campbell RG. Effect of beta-hydroxybutyrate on whole-body leucine kinetics and fractional mixed skeletal muscle protein synthesis in humans. *J Clin Invest.* (1988) 82:198–205. doi: 10.1172/JCI113570
73. Berkes CA, Tapscott SJ. MyoD and the transcriptional control of myogenesis. *Semin Cell Dev Biol.* (2005) 16:585–95. doi: 10.1016/j.semcdb.2005.07.006
74. Lercher A, Baazim H, Berghthaler A. Systemic Immunometabolism: challenges and opportunities. *Immunity.* (2020) 53:496–509. doi: 10.1016/j.immuni.2020.08.012
75. Otani S, Coopersmith CM. Gut integrity in critical illness. *J Intensive Care.* (2019) 7:17. doi: 10.1186/s40560-019-0372-6
76. Zhou Y, Luo Y, Wang X, Luan F, Peng Y, Li Y, et al. Early gut microbiological changes and metabolomic changes in patients with sepsis: a preliminary study. *Int Microbiol.* (2023) 26:1131–42. doi: 10.1007/s10123-023-00363-z
77. Kau AL, Ahern PP, Griffin NW, Goodman AL, Gordon JI. Human nutrition, the gut microbiome, and immune system: envisioning the future. *Nature (London).* (2011) 474:327–36. doi: 10.1038/nature10213
78. Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. *Science.* (2012) 336:1268–73. doi: 10.1126/science.1223490
79. Zeng MY, Inohara N, Nunez G. Mechanisms of inflammation-driven bacterial dysbiosis in the gut. *Mucosal Immunol.* (2017) 10:18–26. doi: 10.1038/mi.2016.75
80. Karlsson FH, Fak F, Nookaew I, Tremaroli V, Fagerberg B, Petranovic D, et al. Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat Commun.* (2012) 3:1245. doi: 10.1038/ncomms2266
81. Ticinesi A, Nouvenne A, Cerundolo N, Catania P, Prati B, Tana C, et al. Gut microbiota, muscle mass and function in aging: a focus on physical frailty and sarcopenia. *Nutrients.* (2019) 11:11. doi: 10.3390/nu11071633
82. Blanton LV, Charbonneau MR, Salih T, Barratt MJ, Venkatesh S, Ilkaveya O, et al. Gut bacteria that prevent growth impairments transmitted by microbiota from malnourished children. *Science.* (2016) 351:351. doi: 10.1126/science.aad3311
83. Langille MG, Meehan CJ, Koenig JE, Dhanani AS, Rose RA, Howlett SE, et al. Microbial shifts in the aging mouse gut. *Microbiome.* (2014) 2:50. doi: 10.1186/s40168-014-0050-9
84. Siddharth J, Chakrabarti A, Pannerec A, Karaz S, Morin-Rivron D, Masoodi M, et al. Aging and sarcopenia associate with specific interactions between gut microbes, serum biomarkers and host physiology in rats. *Aging (Albany NY).* (2017) 9:1698–720. doi: 10.18632/aging.101262
85. Ma D, Wang AC, Parikh I, Green SJ, Hoffman JD, Chlipala G, et al. Ketogenic diet enhances neurovascular function with altered gut microbiome in young healthy mice. *Sci Rep.* (2018) 8:6670. doi: 10.1038/s41598-018-25190-5
86. Rahmel T, Hübner M, Koos B, Wolf A, Willemsen K, Strauß G, et al. Impact of carbohydrate-reduced nutrition in septic patients on ICU: study protocol for a prospective randomised controlled trial. *BMJ Open.* (2020) 10:e38532:e038532. doi: 10.1136/bmjopen-2020-038532
87. Lin J, Huang Z, Liu J, Huang Z, Liu Y, Liu Q, et al. Neuroprotective effect of ketone metabolism on inhibiting inflammatory response by regulating macrophage polarization after acute cervical spinal cord injury in rats. *Front Neurosci.* (2020) 14:583611. doi: 10.3389/fnins.2020.583611
88. Kesarwani P, Kant S, Zhao Y, Miller CR, Chinnaiyan P. The influence of the ketogenic diet on the immune tolerant microenvironment in glioblastoma. *Cancers.* (2022) 14:5550. doi: 10.3390/cancers14225550
89. Hirschberger S, Gellert L, Effinger D, Muenchhoff M, Herrmann M, Briegel JM, et al. Ketone bodies improve human CD8(+) cytotoxic T-cell immune response during COVID-19 infection. *Front Med (Lausanne).* (2022) 9:923502. doi: 10.3389/fmed.2022.923502
90. Masino SA, Ruskin DN, Freedgood NR, Lindefeldt M, Dahlin M. Differential ketogenic diet-induced shift in CSF lipid/carbohydrate metabolome of pediatric epilepsy patients with optimal vs. no anticonvulsant response: a pilot study. *Nutr Metab (Lond).* (2021) 18:23. doi: 10.1186/s12986-020-00524-1
91. Grosicki GJ, Fielding RA, Lustgarten MS. Gut microbiota contribute to age-related changes in skeletal muscle size, composition, and function: biological basis for a gut-muscle Axis. *Calcif Tissue Int.* (2018) 102:433–42. doi: 10.1007/s00223-017-0345-5

92. Wu XX, Huang XL, Chen RR, Li T, Ye HJ, Xie W, et al. Paeoniflorin prevents intestinal barrier disruption and inhibits lipopolysaccharide (LPS)-induced inflammation in Caco-2 cell monolayers. *Inflammation*. (2019) 42:2215–25. doi: 10.1007/s10753-019-01085-z
93. Cohen S, Nathan JA, Goldberg AL. Muscle wasting in disease: molecular mechanisms and promising therapies. *Nat Rev Drug Discov*. (2015) 14:58–74. doi: 10.1038/nrd4467
94. Zanders L, Kny M, Hahn A, Schmidt S, Wundersitz S, Todiras M, et al. Sepsis induces interleukin 6, gp130/JAK2/STAT3, and muscle wasting. *J Cachexia Sarcopenia Muscle*. (2022) 13:713–27. doi: 10.1002/jcsm.12867
95. Kim JT, Napier DL, Kim J, Li C, Lee EY, Weiss HL, et al. Ketogenesis alleviates TNF α -induced apoptosis and inflammatory responses in intestinal cells. *Free Radic Biol Med*. (2021) 172:90–100. doi: 10.1016/j.freeradbiomed.2021.05.032
96. Alsharairi NA. The therapeutic role of short-chain fatty acids mediated very low-calorie ketogenic diet-gut microbiota relationships in paediatric inflammatory bowel diseases. *Nutrients*. (2022) 14:14. doi: 10.3390/nu14194113
97. Ghosh S, Lertwattanarak R, Garduno JJ, Galeana JJ, Li J, Zamarripa F, et al. Elevated muscle TLR4 expression and metabolic endotoxemia in human aging. *J Gerontol A Biol Sci Med Sci*. (2015) 70:232–46. doi: 10.1093/gerona/glu067
98. Thevaranjan N, Puchta A, Schulz C, Naidoo A, Szamosi JC, Verschoor CP, et al. Age-associated microbial Dysbiosis promotes intestinal permeability, systemic inflammation, and macrophage dysfunction. *Cell Host Microbe*. (2017) 21:455–466.e4. doi: 10.1016/j.chom.2017.03.002
99. Sartori R, Romanello V, Sandri M. Mechanisms of muscle atrophy and hypertrophy: implications in health and disease. *Nat Commun*. (2021) 12:330. doi: 10.1038/s41467-020-20123-1
100. Manolis AS, Manolis TA, Manolis AA. Ketone bodies and cardiovascular disease: an alternate fuel source to the rescue. *Int J Mol Sci*. (2023) 24:24. doi: 10.3390/ijms24043534
101. Goldberg EL, Asher JL, Molony RD, Shaw AC, Zeiss CJ, Wang C, et al. beta-Hydroxybutyrate deactivates neutrophil NLRP3 Inflammasome to relieve gout flares. *Cell Rep*. (2017) 18:2077–87. doi: 10.1016/j.celrep.2017.02.004
102. Stubbs BJ, Koutnik AP, Goldberg EL, Upadhyay V, Turnbaugh PJ, Verdin E, et al. Investigating ketone bodies as Immunometabolic countermeasures against respiratory viral infections. *Med*. (2020) 1:43–65. doi: 10.1016/j.medj.2020.06.008
103. Choi IY, Piccio L, Childress P, Bollman B, Ghosh A, Brandhorst S, et al. A diet mimicking fasting promotes regeneration and reduces autoimmunity and multiple sclerosis symptoms. *Cell Rep*. (2016) 15:2136–46. doi: 10.1016/j.celrep.2016.05.009
104. Olson CA, Iniguez AJ, Yang GE, Fang P, Pronovost GN, Jameson KG, et al. Alterations in the gut microbiota contribute to cognitive impairment induced by the ketogenic diet and hypoxia. *Cell Host Microbe*. (2021) 29:1378–1392.e6. doi: 10.1016/j.chom.2021.07.004
105. Li C, Pan J, Sun P, Wang S, Wang S, Feng W, et al. Ketogenic diet alleviates hypoglycemia-induced Neuroinflammation via modulation the gut microbiota in mice. *Mol Nutr Food Res*. (2023) 67:e2200711. doi: 10.1002/mnfr.202200711
106. Ang QY, Alexander M, Newman JC, Tian Y, Cai J, Upadhyay V, et al. Ketogenic diets Alter the gut microbiome resulting in decreased intestinal Th17 cells. *Cell*. (2020) 181:1263–1275.e16. doi: 10.1016/j.cell.2020.04.027
107. Jiang Z, Wang X, Zhang H, Yin J, Zhao P, Yin Q, et al. Ketogenic diet protects MPTP-induced mouse model of Parkinson's disease via altering gut microbiota and metabolites. *MedComm*. (2023) 4:e268. doi: 10.1002/mco2.268
108. Shan T, Huang Y, Zhao Z, Li F, Wang Y, Ye C, et al. Ketogenic diet restrains herpes simplex encephalitis via gut microbes. *Microbes Infect*. (2023) 25:105061. doi: 10.1016/j.micinf.2022.105061
109. Park S, Zhang T, Wu X, Yi QJ. Ketone production by ketogenic diet and by intermittent fasting has different effects on the gut microbiota and disease progression in an Alzheimer's disease rat model. *J Clin Biochem Nutr*. (2020) 67:188–98. doi: 10.3164/jcbs.19-87
110. Rohwer N, El HR, Smyl C, Ocvirk S, Goris T, Grune T, et al. Ketogenic diet has moderate effects on the fecal microbiota of wild-type mice. *Nutrients*. (2023) 15:15. doi: 10.3390/nu15214629
111. Xia H, Guo J, Shen J, Jiang S, Han S, Li L. Butyrate ameliorated the intestinal barrier dysfunction and attenuated acute pancreatitis in mice fed with ketogenic diet. *Life Sci*. (2023) 334:122188. doi: 10.1016/j.lfs.2023.122188
112. Dahlin M, Singleton SS, David JA, Basuchoudhary A, Wickstrom R, Mazumder R, et al. Higher levels of Bifidobacteria and tumor necrosis factor in children with drug-resistant epilepsy are associated with anti-seizure response to the ketogenic diet. *EBioMedicine*. (2022) 80:104061. doi: 10.1016/j.ebiom.2022.104061
113. Quan R, Chen C, Yan W, Zhang Y, Zhao X, Fu Y. BAFF blockade attenuates inflammatory responses and intestinal barrier dysfunction in a murine Endotoxemia model. *Front Immunol*. (2020) 11:570920. doi: 10.3389/fimmu.2020.570920
114. Xu J, Kong X, Li J, Mao H, Zhu Y, Zhu X, et al. Pediatric intensive care unit treatment alters the diversity and composition of the gut microbiota and antimicrobial resistance gene expression in critically ill children. *Front Microbiol*. (2023) 14:1237993. doi: 10.3389/fmicb.2023.1237993
115. Nardone OM, de Sire R, Petito V, Testa A, Villani G, Scalfaferrì F, et al. Inflammatory bowel diseases and sarcopenia: the role of inflammation and gut microbiota in the development of muscle failure. *Front Immunol*. (2021) 12:694217. doi: 10.3389/fimmu.2021.694217