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[Gut-muscle axis mechanism of](https://www.frontiersin.org/articles/10.3389/fnut.2024.1418778/full) [exercise prevention of sarcopenia](https://www.frontiersin.org/articles/10.3389/fnut.2024.1418778/full)

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Sarcopenia refers to an age-related systemic skeletal muscle disorder, which is characterized by loss of muscle mass and weakening of muscle strength. Gut microbiota can affect skeletal muscle through a variety of mechanisms. Gut microbiota present distinct features among elderly people and sarcopenia patients, including a decrease in microbial diversity, which might be associated with the quality and function of the skeletal muscle. There might be a gut-muscle axis; where gut microbiota and skeletal muscle may affect each other bi-directionally. Skeletal muscle can affect the biodiversity of the gut microbiota, and the latter can, in turn, affect the anabolism of skeletal muscle. This review examines recent studies exploring the relationship between gut microbiota and skeletal muscle, summarizes the effects of exercise on gut microbiota, and discusses the possible mechanisms of the gut-muscle axis.

KEYWORDS

sarcopenia, gut microbiota, gut-muscle axis, exercise, SCFAs

1 Introduction

Sarcopenia is a generalized disease of skeletal muscle consisting of the concurrent combination of reduced muscle mass and muscle strength, the prevalence of which increases with age ([1](#page-6-0)). Sarcopenia can reduce exercise capacity, lead to fall-related injuries such as fractures, reduce the quality of life, and even cause premature death ([2\)](#page-6-1). The etiology of sarcopenia is complex, involving neural activity, hormones, immunity, nutrition, physical activity, age, and other factors. The mechanisms of sarcopenia include inflammation, immune senescence, anabolic resistance, and increased oxidative stress [\(3](#page-6-2)). Exercise intervention and nutritional supplementation, or a combination of both, are currently considered effective interventions for treating and preventing sarcopenia ([4](#page-6-3)). Additionally, the effects of the gut microbiota on sarcopenia has attracted much attention ([5](#page-7-0)). With aging, skeletal muscle degenerates, while gut microbiota also presents specific changes ([6](#page-7-1)). Gut microbiota changes with specific patterns and transition points with age. In fact, with increasing age, the overall abundance of the gut microbiota decreases while some harmful microbial species increase. The differences between the gut microbiota of adults and elderly people are characterized by a decrease in bacterial diversity, a decline in the number of Bifidobacteria, and an increase in the number of *Clostridium* bacteria, lactic acid bacteria, Enterobacteriaceae, and Enterococcus ([7](#page-7-2)). Whether exercise, as one of the most effective ways to improve and delay muscle loss, can establish a cross-talk between sarcopenia and gut microbiota (also known as the gut-muscle axis) still needs more in-depth discussion.

2 Gut microbiota and sarcopenia

2.1 Gut microbiota

Gut microbiota refers to the microbial community of bacteria, archaea, viruses, and fungi that live in symbiotic relationships with their hosts in the gastrointestinal tract. A healthy gut microbiota consists of many species, primarily located in the distal part of the gastrointestinal tract ([8](#page-7-3)).

The host and the microorganisms inhabiting it are called "superorganisms," which can regulate catabolism, synthesize vitamins and amino acids, detoxify foreign substances, regulate immune function, and prevent diseases ([9\)](#page-7-4). It also plays a vital role in regulating mitochondrial activity (10) and is part of the central nervous system ([11](#page-7-6)). Gut microbiota may be a determinant of aging due to its regulatory role in human metabolism and immunity [\(12\)](#page-7-7).

Studies of human gut microbiota have shown that even among healthy people, gut microbiota composition varies significantly between individuals (Human Microbiome Project, 2012). The composition of the gut microbiota is affected by several factors, including diet (13) (13) (13) , host genetics (14) (14) (14) , delivery mode (15) (15) (15) , obesity ([16](#page-7-11)), environment ([17](#page-7-12)), and antibiotic use [\(18\)](#page-7-13).

2.2 Gut microbiota and aging

Gut microbiota have distinct age-related characteristics. Xu quantified the abundance of human gut microbiota through 16S rRNA sequencing and found an aging process in the human gut microbiome, with 35 associated genera proving essential functions through a literature review. Evidence indicated that with age, especially among elderly people older than 90years, the beneficial effects of the gut microbiota decrease while the effects of inflammation and disease increase ([19](#page-7-14)). Zhang performed 16S rRNA sequencing on the gut microbiota of hospitalized elderly people and found that the abundance of the gut microbiota in frail elderly people was low, which harmed intestinal and overall health ([6\)](#page-7-1). Although elderly people have a lower diversity of gut microbiota, different characteristics are found among long-lived elderly people. Tuikhar analyzed the composition of the gut microbiota of centenarians in India and compared it with public data from centenarians in four other countries (India, Italy, Japan, and China). They found that the abundance and biodiversity of the Ruminococcaceae community among centenarians were higher regardless of nationality; the Ruminococcaceae are key symbiotic bacterium of the human gut ecosystem and are a dominant family in healthy ecosystems. The Ruminococcaceae can degrade and convert complex polysaccharides into a variety of nutrients for the host, and their increase may be associated with greater metabolic plasticity and multifunctionality in the gut microbiota of long-lived individuals [\(20\)](#page-7-15). Bianet analyzed the fecal microbiota of more than 1,000 healthy Chinese individuals, including 198 centenarians in good health. They found that the microbiota composition of centenarians was not significantly different from that of adults aged 30–50years. However, the abundance of bacteria producing short-chain fatty acids (SCFAs) was higher in centenarians [\(21\)](#page-7-16).

Among people with physical frailty and sarcopenia, the composition of the gut microbiota has been found to change. It is reported that defects are negatively associated with the abundance and diversity of the gut microbiota in specific categories [\(22\)](#page-7-17). Picca measured gut microbial levels in frail and sarcopenia elderly people and found an increased abundance of Oscillospira and Ruminococcus microbial taxa and a decreased quantity of Barnesiellaceae and Christensenellaceae compared with the healthy group, confirming that frailty is associated with changes in the composition of the gut microbiota [\(23\)](#page-7-18). This is consistent with the previous findings that gut microbiota are associated with weakness and biological aging [\(24\)](#page-7-19).

3 Exercise and gut microbiota

3.1 Effects of exercise on gut microbiota

At present, the changes in gut microbiota induced by exercise have attracted the attention of many researchers ([25](#page-7-20)). It has been reported that exercise can significantly increase the gut microbiota diversity ([26](#page-7-21)). Exercise regulates metabolism in various ways. It helps to increase energy expenditure, promote glycogen and fatty acid breakdown, improve blood glucose and lipid metabolism, and reduce chronic inflammation, thereby promoting overall health. Studies in humans and rodents have extensively validated the effects of exercise on the immune system [\(27,](#page-7-22) [28](#page-7-23)). Exercise also prevents weakness and cognitive dysfunction by altering gut microbiota [\(29\)](#page-7-24).

Animal experiments have shown that exercise can regulate gut microbiota significantly. In a mouse model of obesity induced by a high-fat diet, high-intensity exercise ameliorates obesity-induced dysregulation of gut microbiota and maintains microbial diversity ([30](#page-7-25)). In addition, in obese rats, exercise-induced changes in gut microbiota were different from the effects of a healthy diet, suggesting that exercise has an independent impact on gut microbiota. Subsequent studies have found that exercise exerts stronger, more stable, and more influential roles in promoting intestinal mucosal integrity and metabolic function over time ([31](#page-7-26), [32](#page-7-27)). In mouse models, moderate-intensity exercise can induce significant changes in the composition of gut microbiota, including increased microbial diversity and levels of critical microbial groups that can promote healthy metabolic activity [\(33\)](#page-7-28).

In human observational studies, only a few have shown exerciseinduced changes in gut microbiota. Bressa found that adult women with an active lifestyle had significantly higher diversity of healthpromoting microbiota in stool samples than their sedentary peers ([34\)](#page-7-29). According to Clarke, compared with a passive control group of the same size, age, and sex, professional football players had a higher diversity of the gut microbiota, including levels of 22 different microbial groups, which are closely related to protein consumption and creatine kinase ([35](#page-7-30)). Barton also found differences in gut microbiota composition between professional football players and the control group. There are more bacteria-producing SCFAs and more bacteria involved in carbohydrate and amino acid metabolism in the feces of professional football players, resulting in higher concentrations of acetic acid, butyric acid, and propionic acid ([36](#page-7-31)). In a human intervention study, Allen found that exerciseinduced changes in the gut microbiota varied based on the obesity status. They conducted 6weeks of endurance exercise training on both obese and lean sedentary individuals. The study showed that

exercise increased fecal short-chain fatty acid concentrations in lean individuals but not in obese individuals. The changes in the gut microbiota were consistent with alterations in genes and bacteria capable of producing SCFAs ([37](#page-7-32)). Moreover, discontinuing exercise training reversed the exercise-induced changes in the gut microbiota, as well as in lean body mass and body fat resulting from the training. Another study by Manuel analyzed the gut microbiota composition of professional cyclists during the Tour de France. It was discovered that the frequency of the consumption of food and sports supplements during the race had a significant impact on the gut microbiota. To enhance athletic performance, future strategies may involve utilizing selective cultures of bacteria that produce SCFAs and targeting specific bacteria [\(38](#page-7-33)). However, the above studies only show the differences in the distribution of the gut microbiota among different athletes. More comparisons of healthy gut microbiota among other athletes are needed; the effects of specific sports programs on healthy gut microbiota still need to be investigated.

3.2 Exercise intensity plays a significant role in modulating the gut microbiome

The human gut microbiota is significantly influenced by the intensity of physical activity. Generally, long-term exercise of low-tomoderate intensity has positive effects on the gut microbiota, specifically enhancing microbial diversity, elevating the abundance of beneficial bacteria, promoting the production of SCFAs, and strengthening gut barrier function and immune regulation ([39](#page-7-34)). One study found that moderate-intensity exercise significantly impacts the gut microbiota of lean individuals, primarily by enhancing microbial diversity and the production of SCFAs [\(37\)](#page-7-32). Another study suggested that acute moderate-intensity exercise has positive effects on the gut microbiota of athletes, including increased microbial diversity, a rise in the abundance of probiotic organisms such as Bifidobacteria and Lactobacilli, a decrease in potential pathogenic bacteria, and promotion of the production of SCFAs, thus enhancing gut barrier function and anti-inflammatory effects [\(40\)](#page-7-35).

In contrast, high-intensity exercise might have different impacts on gut microbiota. During high-intensity workouts, the body's energy is chiefly spent on sustaining muscular activity and other physiological functions, which might reduce the available energy for gut microbiota, thereby affecting their growth and metabolic activities ([41](#page-7-36)). Additionally, high-intensity exercise might induce intestinal and systemic inflammatory responses, thereby increasing the levels of inflammatory markers, such as C-reactive protein (CRP) and tumor necrosis factor-α (TNF-α). These persistent inflammatory responses could disrupt the stability and diversity of the gut microbiota, leading to a decrease in beneficial bacteria and an increase in potential pathogenic bacteria [\(39\)](#page-7-34). Further research has indicated that highintensity exercise could increase intestinal permeability, causing damage to intestinal epithelial cells and heightening the potential risk of leaky gut syndrome [\(42\)](#page-7-37). Some specific changes, e.g., a decrease in beneficial bacteria (such as Lactobacilli and Bifidobacteria) and an increase in potential pathogenic bacteria (such as Clostridia), are common in individuals who engage regularly in high-intensity and long-duration workouts. The gut microbiota characteristics of these athletes show higher counts of bacteria involved in the inflammatory

process ([43](#page-7-38)). Therefore, workout intensity is a crucial factor affecting the state of the gut microbiota.

3.3 Possible mechanisms of how exercise affects gut microbiota

The effects of exercise on human gut microbiota may involve multiple levels, such as regulating immune response ([44](#page-7-39)) and influencing the production of intestinal microbial metabolites, including SCFAs [\(8,](#page-7-3) [45](#page-7-40)) [\(Figure 1](#page-3-0)).

3.3.1 Gut microbiota metabolites of SCFAs

The effect of exercise on gut microbiota mainly manifests as an increase in SCFA concentration. SCFAs, fatty acids of 2–6 carbon atoms, are primarily produced by the fermentation of indigestible sugars, such as dietary fiber and oligosaccharides, by beneficial bacteria, such as *Lactobacillus* and *Bifidobacterium* in the colon. SCFAs can be absorbed by skeletal muscle through systemic circulation. They can regulate metabolic synthesis processes, such as activating AMPK and mTOR signaling pathways and enhancing mitochondrial biogenesis ([46](#page-7-41)), by inhibiting histone deacetylases (HDACs) and activating transcription factors like PPARγ. SCFAs can promote the expression of genes related to fatty acid synthesis and storage, influence the AMPK signaling pathway, and regulate the activity of fatty acid synthase (FAS), balancing the body's fatty acid synthesis and oxidation processes ([47](#page-7-42)). By inducing the translocation of glucose transporter 4 (GLUT4) and regulating the phosphorylation status of insulin receptor substrate (IRS), SCFAs can enhance insulin signal transduction, activate the phosphatidylinositol 3-kinase (PI3K) and protein kinase B (Akt) pathways to promote glucose absorption, and increase insulin sensitivity ([48](#page-7-43)). Through various mechanisms, such as inhibiting the NF-κB signaling pathway, activating G proteincoupled receptors, adjusting the acetylation state of histones, regulating immune cell function, and inhibiting pro-inflammatory metabolic pathways, SCFAs may potentially reduce inflammation ([49](#page-7-44)). SCFAs can alter the permeability of the blood–brain barrier, influencing metabolism and signal transmission within the brain, and improving central nervous system functions [\(50](#page-7-45)). In addition, butyrate among SCFAs can enhance ATP generation and muscle fiber metabolism efficiency ([51\)](#page-7-46).

Physical exercise may stimulate the production of SCFAs. Research by Allen reveals that exercise has the potential to alter the composition and functionality of human gut microbiota. Among lean participants, exercise significantly increases the concentration of SCFAs in fecal matter ([37\)](#page-7-32). Furthermore, exercise-induced intestinal hypoxia involves intricate mechanisms impacting gut microbiota and their metabolic byproducts. The rerouting of blood during physical exercise prioritizes supply to muscles and other active organs, resulting in decreased blood flow and oxygen delivery to the intestines. This oxygen-depleted environment in the gut increases the growth of anaerobic bacteria, whose compositional shifts subsequently influence the formation of metabolic products, such as SCFAs, within the gut. A study by Huang observed that acute exerciseinduced hypoxia increased changes in the gut microbiota of mice. This promoted the proliferation of anaerobic bacteria such as *Akkermansia* and *Bacteroidetes* genera, accompanied by an increase in the production of short-chain fatty acids. These changes ultimately improved the endurance performance of the mice ([52\)](#page-7-47).

The mechanism by which exercise promotes increased SCFA concentration is still being determined. The possible mechanism is that changes in colonic oxygen saturation or pH may encourage the mixing of intestinal contents and the fermentation of dietary fiber through endogenous metabolite input (e.g., lactic acid), thereby increasing the production of SCFAs ([53\)](#page-7-48).

Exercise induces an increase in SCFAs in feces, which may also be due to an increased ability of the gut microbiota to produce SCFAs. In the process of SCFA production, butyryl-CoA is a critical enzyme for butyric acid production [\(54\)](#page-8-0). Exercise may induce the expression of genes for these vital enzymes, which may partially explain the increase in SCFAs in feces.

3.3.2 Systemic inflammation

Another potential mechanism involves changes in systemic inflammation levels. Animal experiments have found that exercise can mediate alterations in metabolism and attenuate colitis in mice [\(55\)](#page-8-1). Gut microbiota plays a role in the intestinal immune response by mediating the transfer of gut microbiota from the intestinal cavity to the blood circulation, a phenomenon known as leaky gut. Exercise can enhance intestinal barrier function and reduce inflammation by regulating the composition of the gut microbiota [\(56\)](#page-8-2). Therefore, exercise may help reduce inflammation.

4 The effects of gut microbiota on skeletal muscle

In recent years, it has been reported that the gut microbiota is involved in skeletal muscle metabolism ([57](#page-8-3)). The composition of the

gut microbiota may affect the quality, structure, and function of skeletal muscle during aging, thus influencing the development of sarcopenia.

4.1 Muscle function

The gut microbiota significantly affects skeletal muscle mass and function in mice. Research has shown that germ-free mice lacking gut microbiota experience skeletal muscle atrophy, reduced expression of insulin-like growth factor 1, and decreased transcription of genes related to muscle growth and mitochondrial function. However, transferring gut microbiota from pathogen-free mice to germ-free mice helps improve these conditions. These findings highlight the crucial role of the gut microbiota in regulating skeletal muscle function ([58](#page-8-4)). In another study, treatment with the antibiotic metronidazole led to dysregulation of the gut flora in mice, resulting in reduced hindlimb muscle weight, smaller fibers in the tibialis anterior muscles, and upregulation of genes associated with neurogenic atrophy of skeletal muscle, including Hdac4, myogenin, MuRF1, and atrogin1, which contribute to skeletal muscle atrophy ([59](#page-8-5)). Additionally, research showed that 6weeks of supplementation with *Lactobacillus plantarum* TWK10 in mice enhanced glucose utilization by increasing the number of gastrocnemius type I muscle fibers, resulting in increased endurance exercise time [\(60\)](#page-8-6).

Human studies have also found that gut microbiota composition is related to skeletal muscle function in elderly people. For instance, when elderly people are transferred to care facilities, their gut microbiota changes, affecting bone and body composition, which can lead to muscle loss, osteoporosis, obesity, and an increased risk of fractures ([61\)](#page-8-7).

4.2 Muscle mass

Animal experiments have shown that changes in gut microbiota can affect skeletal muscle mass. The BaF3 mouse model is a typical model of leukemia that exhibits muscle atrophy, fat loss, anorexia, and inflammatory responses at later stages. Significant changes in the gut microbiota of these models, particularly in the *Lactobacillus* populations, were found in this study. The balance of *Lactobacillus* populations could be restored by supplementing BaF3 mice with *Lactobacillus reuteri* 100-23 and *Lactobacillus gasseri* 311,476, and the expression of muscle atrophy markers (Atrogin-1, MuRF1, LC3, and histone L) was reduced in the gastrocnemius and tibialis muscles. Additionally, the levels of serum factors related to chronic inflammation (e.g., IL-6, MCP-1) were reduced. These findings suggest that the gut microbiota, especially *Lactobacillus* species, can modulate the body's immune-inflammatory response and inhibit muscle atrophy [\(62\)](#page-8-8). Chen has shown that long-term exposure to *L. plantarum* in mice may increase energy absorption and muscle mass [\(60\)](#page-8-6).

Prebiotics may affect skeletal muscle through gut microbiota by promoting the proliferation of beneficial bacteria ([63\)](#page-8-9). Cani found that prebiotic supplements reduced lipopolysaccharide (LPS) levels and inflammation, promoting increased skeletal muscle mass in obese mice [\(64](#page-8-10)). Varian found a significant improvement in muscle mass after treating mice with muscle atrophy using a probiotic mixture containing *L. reuteri* ([65](#page-8-11)). Probiotics and prebiotics may promote skeletal muscle anabolism by improving gut microbiota composition, reducing intestinal endotoxin concentrations, and increasing insulin sensitivity [\(66](#page-8-12)).

4.3 Muscle composition

Increasing age leads to increased infiltration of adipose tissue, altering the composition of skeletal muscle, which associated with a decrease in motor skills ([67](#page-8-13)). Remarkably, resistance training can mitigate this accumulated fat by increasing the size of muscle fibers; in contrast, aerobic exercises contribute to fat burning by boosting the overall metabolic rate ([68\)](#page-8-14). Regular physical activity further enhances metabolic health and helps prevent the accumulation of fat in muscle tissue ([69\)](#page-8-15). Importantly, gut microbiota regulates inflammation and fat metabolism, particularly through the production of substances such as SCFAs. This aids in improving insulin sensitivity, which subsequently helps reduce the accumulation of fat in muscles [\(70](#page-8-16)).

In animal models, changes in gut microbiota also alter the composition of skeletal muscle fibers [\(62\)](#page-8-8). Studies have shown that a high-fat diet induces acute changes in the gut microbiota of 3-month-old rats and significantly affects its overall composition within 28days. This leads to increased circulating inflammatory cytokines and intramuscular fat in skeletal muscle. Furthermore, a high-fat diet alters intestinal barrier function by increasing pro-inflammatory cytokines, such as TNF-α and IL-6, modulating metabolic pathways of SCFAs, affecting insulin signaling pathways, altering intestinal barrier function and skeletal muscle composition and function, and possibly causing shifts in myofiber type. Together, these mechanisms reveal the complex relationship between the gut microbiota and skeletal muscle composition ([71\)](#page-8-17). A study on 10-month-old pigs found that energy metabolism-related gut bacteria were significantly associated with intramuscular fat content through 16S rRNA gene sequencing, validating the critical role of the gut microbiota in regulating muscle composition ([72](#page-8-18)).

5 Possible mechanisms of the gut-muscle axis

Animal models and human studies have confirmed the role of the gut microbiota in regulating skeletal muscle, but its systemic mechanisms still need to be fully understood. Some studies have suggested that there is a "gut-muscle axis" and proposed possible mechanisms ([73](#page-8-19)) by which gut microbiota affects skeletal muscle mass and function, including protein metabolism, systemic chronic inflammation, metabolic resistance (reduced or diminished responsiveness of the organism in metabolic regulation), and mitochondrial dysfunction [\(Figure 2](#page-5-0)).

5.1 Metabolites of the gut microbiota

Metabolites produced by the gut microbiota can be absorbed by the intestinal mucosa and affect skeletal muscle quality ([74\)](#page-8-20). Some of these metabolites are nutrients, such as folic acid ([75](#page-8-21)), riboflavin [\(76\)](#page-8-22), vitamin B12 [\(75\)](#page-8-21), betaine ([77](#page-8-23)), and some amino acids [\(78](#page-8-24)). These nutrients affect skeletal muscle function, from promoting DNA synthesis and repair to mediating insulin growth factor 1, which stimulates anabolism and cell proliferation ([73](#page-8-19)). Healthy gut microbiota can produce these nutrients and improve the bioavailability of amino acids. In contrast, dysregulation of the gut microbiota may lead to reduced production of these nutrients and decreased absorption of amino acids, negatively affecting skeletal muscle anabolism [\(78\)](#page-8-24).

In addition, gut microbial metabolites represented by SCFAs (such as acetate, propionate, and butyrate) can act as endocrine mediators and influence the metabolism and function of skeletal muscles [\(79\)](#page-8-25). They can improve glucose homeostasis and insulin sensitivity, regulate inflammation and satiety, and stimulate catabolism in adipose tissue and anabolism in skeletal muscle cells. Dysregulation of the gut microbiota is usually characterized by the reduced production of SCFAs, resulting in reduced anabolic or anti-catabolic action of skeletal muscle [\(73\)](#page-8-19).

5.2 Protein anabolism

Gut microbiota are involved in the metabolism and absorption of amino acids. They can improve the bioavailability of amino acids by utilizing several amino acids from nutrient and endogenous proteins, thus affecting the synthesis and decomposition of muscle proteins [\(80\)](#page-8-26).

In addition, gut microbiota can synthesize some essential amino acids, such as tryptophan [\(78](#page-8-24)), which may stimulate the insulin-like growth factor 1/p70S6K/mTOR pathway in muscle cells and promote the expression of muscle fiber synthesis genes [\(81\)](#page-8-27).

The composition of the gut microbiota can also influence muscle mass, anabolism, and host function by producing mediators, such as

SCFAs [\(73](#page-8-19)). They can increase the production of ATP by regulating the protein regulation pathway, affect protein deposition, and stimulate the glucose absorption of skeletal muscle by regulating the balance of synthesis and decomposition throughout the body [\(82](#page-8-28)).

5.3 Inflammation and gut barrier function

The aging process can seriously affect the structure of the human gut microbiota and the balance and inflammatory state of the host immune system. Changes in the gut microbiota can alter the inflammatory state of individuals.

Healthy gut microbiota regulates immune homeostasis, maintains the balance between pro-inflammatory and anti-inflammatory responses, and suppresses chronic inflammation [\(73\)](#page-8-19). Meanwhile, it can induce host reactions in the intestinal mucosa, strengthen the intestinal barrier function, and play an immunomodulatory role inside and outside the intestinal tract ([83](#page-8-29)). SCFAs produced by the gut microbiota can exert anti-inflammatory effects by reducing the secretion of pro-inflammatory cytokines and chemokines as well as the infiltration of local macrophages. Butyric acid, one of the SCFAs can induce the secretion of interleukin-10, retinoic acid, and tumor growth factor-β [\(84\)](#page-8-30), stimulate the production of anti-inflammatory T cells [\(85\)](#page-8-31), and suppress inflammation.

Another mechanism by which gut microbiota regulates inflammation is maintaining intestinal barrier function, regulating LPS, and promoting the absorption of inflammatory bacterial endotoxin ([86\)](#page-8-32). Butyrate also plays a vital role in intestinal barrier

function by strengthening the tight junctions of epithelial cells, thus preventing endotoxin translocation and reducing inflammation [\(87\)](#page-8-33).

5.4 Mitochondrial dysfunction

Mitochondria is the energy factory of the cell. Mitochondrial function, especially ATP production and the efficiency of the respiratory chain, decreases with age. In skeletal muscle, primary aging results in a lack of mitochondrial energy and a loss of muscle mass ([88](#page-8-34)). Downregulation of genes of the mitochondrial energy metabolism pathway (e.g., the tricarboxylic acid cycle and oxidative phosphorylation) causes mitochondrial dysfunction and progressive loss of muscle mass and function, leading to sarcopenia ([89](#page-8-35)). The inducible inflammatory factors in sarcopenia may be the free mtDNA of nucleoid or oxidized cells extruded from the damaged mitochondria, which can activate the innate immune system of the body and induce the production of inflammatory mediators. The release of inflammatory mediators can sustain a vicious cycle within the muscle cells, leading to further mitochondrial damage and, eventually, sarcopenia ([90](#page-8-36)).

There is a regulatory relationship between gut microbiota and mitochondria, and interrupting this relationship may lead to mitochondrial dysfunction [\(91\)](#page-8-37). The gut microbiota can regulate key mitochondrial transcriptional activators, transcription factors, and enzymes such as PGC-1 α , SIRT1, and AMPK genes [\(10\)](#page-7-5).

In addition, SCFAs are recognized as mediators underlying the effects of the gut microbiota on skeletal muscle, and their main targets

are mitochondria of skeletal muscle cells, suggesting a close relationship between gut microbiota and mitochondrial function. SCFAs and secondary bile acids can promote mitochondrial energy production and regulate reactive oxygen species (ROS) by weakening TNF-α-mediated immune responses and inflammatory bodies such as NLRP3. Additionally, SCFAs, such as butyric acid and acetic acid, can directly or indirectly affect mitochondrial energy metabolism through a wide range of transcription factors ([10](#page-7-5)).

5.5 Insulin resistance and metabolism resistance

Elderly people have anabolic resistance and require more protein intake to achieve the same myoprotein synthesis as younger adults ([92](#page-8-38)). Gut microbiota participate directly or indirectly in the hypothesized mechanisms of anabolic resistance in elderly people and may facilitate complex interactions among these hypothesized mechanisms [\(93](#page-8-39)). Thus, a healthy gut microbiota may reduce metabolic resistance.

Disordered gut microbiota and altered intestinal barriers may reduce SCFAs and secondary bile acids but increase the absorption of LPS and branched-chain amino acids (BCAAs). All of these changes can lead to insulin resistance ([85\)](#page-8-31). SCFAs are critical mediators of mitochondrial energy metabolism, acting as ligands for free fatty acid receptors 2 and 3 and regulating glucose and fatty acid metabolism. They can enhance glucose tolerance, promote insulin sensitivity, and play a key role in regulating glucose uptake and metabolism ([73](#page-8-19)). Secondary bile acids can activate the secretion of glucagon-like peptide-1, thereby reducing insulin resistance and maintaining glucose homeostasis [\(94\)](#page-8-40). LPS can activate Toll-like receptor 4 signaling, inducing insulin resistance, inflammation, and obesity [\(85\)](#page-8-31). Circulating BCAA levels may independently increase the risk of insulin resistance and type 2 diabetes ([95](#page-8-41)).

6 Conclusion and limitations

In summary, the impact of exercise on the gut microbiota is multifaceted, leading to notable changes such as increased abundance and the production of SCFAs. Concurrently, the gut microbiota exerts a profound influence on skeletal muscle quality and function through various mechanisms, such as the regulation of protein metabolism, mitigation of insulin resistance, improvement of mitochondrial function, and attenuation of inflammation. As individuals grow older, people may experience sarcopenia. This aging-related process can significantly alter intestinal physiology, particularly the integrity of the intestinal mucosal barrier. Consequently, shifts in gut microbiota composition may arise as a consequence of sarcopenia rather than serving as its primary cause. Moreover, sarcopenia frequently

References

coincides with inadequate nutrition and diminished exercise capacity. Given that both diet and physical activity strongly influence gut microbiota composition, it is plausible that lifestyle factors contribute to sarcopenia rather than solely attributing it to dysbiosis of the gut microbiota.

However, existing research on the relationship between gut microbiota and sarcopenia is limited. Many studies have been conducted on small cohorts, lacking comprehensive assessments of elderly populations. Furthermore, the majority of data concerning the interplay between gut microbiota and muscle mass/function are derived from animal models, with human studies primarily being observational in nature. Therefore, the precise role of the gut microbiota in the development and progression of sarcopenia warrants further investigation and a broader scope of research methodologies.

Author contributions

TL: Investigation, Methodology, Writing – original draft, Writing – review & editing. DY: Methodology, Writing – review & editing, Writing – original draft. RS: Funding acquisition, Methodology, Supervision, Writing – review & editing, Writing – original draft.

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Conflict of interest

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^{1.} Kirk B, Cawthon PM, Arai H, Ávila-Funes JA, Barazzoni R, Bhasin S, et al. The conceptual definition of sarcopenia: Delphi consensus from the global leadership initiative in sarcopenia (GLIS). *Age Ageing*. (2024) 53:afae052. doi: [10.1093/ageing/afae052](https://doi.org/10.1093/ageing/afae052)

^{2.} Larsson L, Degens H, Li M, Salviati L, Lee YI, Thompson W, et al. Sarcopenia: aging-related loss of muscle mass and function. *Physiol Rev*. (2019) 99:427–511. doi: [10.1152/physrev.00061.2017](https://doi.org/10.1152/physrev.00061.2017)

^{3.} Narici MV, Maffulli N. Sarcopenia: characteristics, mechanisms and functional significance. *Br Med Bull*. (2010) 95:139–59. doi: [10.1093/bmb/ldq008](https://doi.org/10.1093/bmb/ldq008)

^{4.} Yoshimura Y, Wakabayashi H, Yamada M, Kim H, Harada A, Arai H. Interventions for treating sarcopenia: a systematic review and Meta-analysis of randomized controlled studies. *J Am Med Dir Assoc*. (2017) 18:553.e1–553.e16. doi: [10.1016/j.](https://doi.org/10.1016/j.jamda.2017.03.019) [jamda.2017.03.019](https://doi.org/10.1016/j.jamda.2017.03.019)

6. Zhang L, Liao J, Chen Q, Chen M, Kuang Y, Chen L, et al. Characterization of the gut microbiota in frail elderly patients. *Aging Clin Exp Res*. (2020) 32:2001–11. doi: [10.1007/s40520-019-01385-2](https://doi.org/10.1007/s40520-019-01385-2)

7. Kim S, Jazwinski SM. The gut microbiota and healthy aging: a Mini-review. *Gerontology*. (2018) 64:513–20. doi: [10.1159/000490615](https://doi.org/10.1159/000490615)

8. Schmidt TSB, Raes J, Bork P. The human gut microbiome: from association to modulation. *Cell*. (2018) 172:1198–215. doi: [10.1016/j.cell.2018.02.044](https://doi.org/10.1016/j.cell.2018.02.044)

9. Rinninella E, Raoul P, Cintoni M, Franceschi F, Miggiano GAD, Gasbarrini A, et al. What is the healthy gut microbiota composition? A changing ecosystem across age, environment, diet, and diseases. *Microorganisms*. (2019) 7:14. doi: [10.3390/](https://doi.org/10.3390/microorganisms7010014) [microorganisms7010014](https://doi.org/10.3390/microorganisms7010014)

10. Clark A, Mach N. The crosstalk between the gut microbiota and mitochondria during exercise. *Front Physiol*. (2017) 8:319. doi: [10.3389/fphys.2017.00319](https://doi.org/10.3389/fphys.2017.00319)

11. Dinan TG, Cryan JF. The microbiome-gut-brain Axis in health and disease. *Gastroenterol Clin N Am*. (2017) 46:77–89. doi: [10.1016/j.gtc.2016.09.007](https://doi.org/10.1016/j.gtc.2016.09.007)

12. Candela M, Biagi E, Brigidi P, O'Toole PW, De Vos WM. Maintenance of a healthy trajectory of the intestinal microbiome during aging: a dietary approach. *Mech Ageing Dev*. (2014) 136–137:70–5. doi: [10.1016/j.mad.2013.12.004](https://doi.org/10.1016/j.mad.2013.12.004)

13. Chen J, He X, Huang J. Diet effects in gut microbiome and obesity. *J Food Sci*. (2014) 79:R442–51. doi: [10.1111/1750-3841.12397](https://doi.org/10.1111/1750-3841.12397)

14. Goodrich JK, Waters JL, Poole AC, Sutter JL, Koren O, Blekhman R, et al. Human genetics shape the gut microbiome. *Cell*. (2014) 159:789–99. doi: [10.1016/j.](https://doi.org/10.1016/j.cell.2014.09.053) [cell.2014.09.053](https://doi.org/10.1016/j.cell.2014.09.053)

15. Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, et al. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA*. (2010) 107:11971–5. doi: [10.1073/pnas.1002601107](https://doi.org/10.1073/pnas.1002601107)

16. Cani PD. Gut microbiota and obesity: lessons from the microbiome. *Brief Funct Genomics*. (2013) 12:381–7. doi: [10.1093/bfgp/elt014](https://doi.org/10.1093/bfgp/elt014)

17. Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, et al. Human gut microbiome viewed across age and geography. *Nature*. (2012) 486:222–7. doi: [10.1038/nature11053](https://doi.org/10.1038/nature11053)

18. Bhattacharya S, Bhadra R, Schols A, Sambashivaiah S. Gut microbial dysbiosis as a limiting factor in the management of primary and secondary sarcopenia: an Asian Indian perspective. *Curr Opin Clin Nutr Metab Care*. (2020) 23:404–10. doi: [10.1097/](https://doi.org/10.1097/mco.0000000000000688) [mco.0000000000000688](https://doi.org/10.1097/mco.0000000000000688)

19. Xu C, Zhu H, Qiu P. Aging progression of human gut microbiota. *BMC Microbiol*. (2019) 19:236. doi: [10.1186/s12866-019-1616-2](https://doi.org/10.1186/s12866-019-1616-2)

20. Tuikhar N, Keisam S, Labala RK, Imrat P, Ramakrishnan MC, Arunkumar G, et al. Comparative analysis of the gut microbiota in centenarians and young adults shows a common signature across genotypically non-related populations. *Mech Ageing Dev*. (2019) 179:23–35. doi: [10.1016/j.mad.2019.02.001](https://doi.org/10.1016/j.mad.2019.02.001)

21. Bian G, Gloor GB, Gong A, Jia C, Zhang W, Hu J, et al. The gut microbiota of healthy aged Chinese is similar to that of the healthy young. *mSphere*. (2017) 2:e00327–17. doi: [10.1128/mSphere.00327-17](https://doi.org/10.1128/mSphere.00327-17)

22. Strasser B, Wolters M, Weyh C, Krüger K, Ticinesi A. The effects of lifestyle and diet on gut microbiota composition, inflammation and muscle performance in our aging society. *Nutrients*. (2021) 13:2045. doi: [10.3390/nu13062045](https://doi.org/10.3390/nu13062045)

23. Picca A, Ponziani FR, Calvani R, Marini F, Biancolillo A, Coelho-Junior HJ, et al. Gut microbial, inflammatory and metabolic signatures in older people with physical frailty and sarcopenia: results from the BIOSPHERE study. *Nutrients*. (2019) 12:65. doi: [10.3390/nu12010065](https://doi.org/10.3390/nu12010065)

24. Maffei VJ, Kim S, Blanchard E IV, Luo M, Jazwinski SM, Taylor CM, et al. Welsh: biological aging and the human gut microbiota. *J Gerontol A Biol Sci Med Sci*. (2017) 72:1474–82. doi: [10.1093/gerona/glx042](https://doi.org/10.1093/gerona/glx042)

25. Lavilla-Lerma ML, Aibar-Almazán A, Martı Nez-Amat A, Benomar-El-Bakali N, Abriouel-Hayani H, Hita-Contreras F. The effects of physical activity on the modulation of gut microbiota composition: a systematic review. *Benef Microbes*. (2023) 14:553–64. doi: [10.1163/18762891-20230031](https://doi.org/10.1163/18762891-20230031)

26. Min L, Ablitip A, Wang R, Luciana T, Wei M, Ma X. Effects of exercise on gut microbiota of adults: a systematic review and Meta-analysis. *Nutrients*. (2024) 16:1070. doi: [10.3390/nu16071070](https://doi.org/10.3390/nu16071070)

27. Egan B, Zierath JR. Exercise metabolism and the molecular regulation of skeletal muscle adaptation. *Cell Metab*. (2013) 17:162–84. doi: [10.1016/j.cmet.2012.12.012](https://doi.org/10.1016/j.cmet.2012.12.012)

28. Campbell JP, Turner JE. Debunking the myth of exercise-induced immune suppression: redefining the impact of exercise on immunological health across the lifespan. *Front Immunol*. (2018) 9:648. doi: [10.3389/fimmu.2018.00648](https://doi.org/10.3389/fimmu.2018.00648)

29. Shin HE, Kwak SE, Lee JH, Zhang D, Bae JH, Song W. Exercise, the gut microbiome, and frailty. *Ann Geriatr Med Res*. (2019) 23:105–14. doi: [10.4235/](https://doi.org/10.4235/agmr.19.0014) [agmr.19.0014](https://doi.org/10.4235/agmr.19.0014)

30. Denou E, Marcinko K, Surette MG, Steinberg GR, Schertzer JD. High-intensity exercise training increases the diversity and metabolic capacity of the mouse distal gut microbiota during diet-induced obesity. *Am J Physiol Endocrinol Metab*. (2016) 310:E982–93. doi: [10.1152/ajpendo.00537.2015](https://doi.org/10.1152/ajpendo.00537.2015)

31. Welly RJ, Liu TW, Zidon TM, Rowles JL 3rd, Park YM, Smith TN, et al. Comparison of diet versus exercise on metabolic function and gut microbiota in obese rats. *Med Sci Sports Exerc*. (2016) 48:1688–98. doi: [10.1249/mss.0000000000000964](https://doi.org/10.1249/mss.0000000000000964)

32. Imdad S, So B, Jang J, Park J, Lee SJ, Kim JH, et al. Temporal variations in the gut microbial diversity in response to high-fat diet and exercise. *Sci Rep*. (2024) 14:3282. doi: [10.1038/s41598-024-52852-4](https://doi.org/10.1038/s41598-024-52852-4)

33. Lamoureux EV, Grandy SA, Langille MGI. Moderate exercise has limited but distinguishable effects on the mouse microbiome. *mSystems*. (2017) 2:e00006–17. doi: [10.1128/mSystems.00006-17](https://doi.org/10.1128/mSystems.00006-17)

34. Bressa C, Bailén-Andrino M, Pérez-Santiago J, González-Soltero R, Pérez M, Montalvo-Lominchar MG, et al. Differences in gut microbiota profile between women with active lifestyle and sedentary women. *PLoS One*. (2017) 12:e0171352. doi: [10.1371/](https://doi.org/10.1371/journal.pone.0171352) [journal.pone.0171352](https://doi.org/10.1371/journal.pone.0171352)

35. Clarke SF, Murphy EF, O'Sullivan O, Lucey AJ, Humphreys M, Hogan A, et al. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut*. (2014) 63:1913–20. doi: [10.1136/gutjnl-2013-306541](https://doi.org/10.1136/gutjnl-2013-306541)

36. Barton W, Penney NC, Cronin O, Garcia-Perez I, Molloy MG, Holmes E, et al. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut*. (2017) 67:gutjnl-2016-313627–33. doi: [10.1136/gutjnl-2016-313627](https://doi.org/10.1136/gutjnl-2016-313627)

37. Allen JM, Mailing LJ, Niemiro GM, Moore R, Cook MD, White BA, et al. Exercise alters gut microbiota composition and function in lean and obese humans. *Med Sci Sports Exerc*. (2018) 50:747–57. doi: [10.1249/mss.0000000000001495](https://doi.org/10.1249/mss.0000000000001495)

38. Fernandez-Sanjurjo M, Fernandez J, Martinez-Camblor P, Rodriguez-Alonso M, Ortolano-Rios R, Pinto-Hernandez P, et al. Dynamics of gut microbiota and short-chain fatty acids during a cycling grand tour are related to exercise performance and modulated by dietary intake. *Nutrients*. (2024) 16:661. doi: [10.3390/nu16050661](https://doi.org/10.3390/nu16050661)

39. Clark A, Mach N. Exercise-induced stress behavior, gut-microbiota-brain axis and diet: a systematic review for athletes. *J Int Soc Sports Nutr*. (2016) 13:43. doi: [10.1186/](https://doi.org/10.1186/s12970-016-0155-6) [s12970-016-0155-6](https://doi.org/10.1186/s12970-016-0155-6)

40. Tabone M, Bressa C, García-Merino JA, Moreno-Pérez D, Van EC, Castelli FA, et al. The effect of acute moderate-intensity exercise on the serum and fecal metabolomes and the gut microbiota of cross-country endurance athletes. *Sci Rep*. (2021) 11:3558. doi: [10.1038/s41598-021-82947-1](https://doi.org/10.1038/s41598-021-82947-1)

41. Mach N, Fuster-Botella D. Endurance exercise and gut microbiota: a review. *J Sport Health Sci*. (2017) 6:179–97. doi: [10.1016/j.jshs.2016.05.001](https://doi.org/10.1016/j.jshs.2016.05.001)

42. Ribeiro FM, Petriz B, Marques G, Kamilla LH, Franco OL. Is there an exerciseintensity threshold capable of avoiding the leaky gut? *Front Nutr*. (2021) 8:627289. doi: [10.3389/fnut.2021.627289](https://doi.org/10.3389/fnut.2021.627289)

43. Bonomini-Gnutzmann R, Plaza-Díaz J, Jorquera-Aguilera C, Rodríguez-Rodríguez A, Rodríguez-Rodríguez F. Effect of intensity and duration of exercise on gut microbiota in humans: a systematic review. *Int J Environ Res Public Health*. (2022) 19:9518. doi: [10.3390/ijerph19159518](https://doi.org/10.3390/ijerph19159518)

44. Codella R, Luzi L, Terruzzi I. Exercise has the guts: how physical activity may positively modulate gut microbiota in chronic and immune-based diseases. *Dig Liver Dis*. (2018) 50:331–41. doi: [10.1016/j.dld.2017.11.016](https://doi.org/10.1016/j.dld.2017.11.016)

45. Sales KM, Reimer RA. Unlocking a novel determinant of athletic performance: the role of the gut microbiota, short-chain fatty acids, and "biotics" in exercise. *J Sport Health Sci*. (2023) 12:36–44. doi: [10.1016/j.jshs.2022.09.002](https://doi.org/10.1016/j.jshs.2022.09.002)

46. Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, et al. Gut microbiota composition correlates with diet and health in the elderly. *Nature*. (2012) 488:178–84. doi: [10.1038/nature11319](https://doi.org/10.1038/nature11319)

47. Watterson KR, Hudson BD, Ulven T, Milligan G. Treatment of type 2 diabetes by free fatty acid receptor agonists. *Front Endocrinol (Lausanne)*. (2014) 5:137. doi: [10.3389/](https://doi.org/10.3389/fendo.2014.00137) [fendo.2014.00137](https://doi.org/10.3389/fendo.2014.00137)

48. den Besten G, van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J Lipid Res*. (2013) 54:2325–40. doi: [10.1194/jlr.R036012](https://doi.org/10.1194/jlr.R036012)

49. Kaczmarczyk MM, Miller MJ, Freund GG. The health benefits of dietary fiber: beyond the usual suspects of type 2 diabetes mellitus, cardiovascular disease and colon cancer. *Metabolism*. (2012) 61:1058–66. doi: [10.1016/j.metabol.2012.01.017](https://doi.org/10.1016/j.metabol.2012.01.017)

50. De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchampt A, et al. Microbiota-generated metabolites promote metabolic benefits via gut-brain neural circuits. *Cell*. (2014) 156:84–96. doi: [10.1016/j.cell.2013.12.016](https://doi.org/10.1016/j.cell.2013.12.016)

51. den Besten G, Gerding A, van Dijk TH, Ciapaite J, Bleeker A, van Eunen K, et al. Protection against the metabolic syndrome by guar gum-derived short-chain fatty acids depends on peroxisome proliferator-activated receptor γ and glucagon-like Peptide-1. *PLoS One*. (2015) 10:e0136364. doi: [10.1371/journal.pone.0136364](https://doi.org/10.1371/journal.pone.0136364)

52. Huang L, Li T, Zhou M, Deng M, Zhang L, Yi L, et al. Hypoxia improves endurance performance by enhancing short chain fatty acids production via gut microbiota remodeling. *Front Microbiol*. (2021) 12:820691. doi: [10.3389/fmicb.2021.](https://doi.org/10.3389/fmicb.2021.820691) [820691](https://doi.org/10.3389/fmicb.2021.820691)

53. Turnbaugh PJ, Gordon JI. The core gut microbiome, energy balance and obesity. *J Physiol*. (2009) 587:4153–8. doi: [10.1113/jphysiol.2009.174136](https://doi.org/10.1113/jphysiol.2009.174136)

54. Louis P, McCrae SI, Charrier C, Flint HJ. Organization of butyrate synthetic genes in human colonic bacteria: phylogenetic conservation and horizontal gene transfer. *FEMS Microbiol Lett*. (2007) 269:240–7. doi: [10.1111/j.1574-6968.2006.](https://doi.org/10.1111/j.1574-6968.2006.00629.x) [00629.x](https://doi.org/10.1111/j.1574-6968.2006.00629.x)

55. Allen JM, Mailing LJ, Cohrs J, Salmonson C, Fryer JD, Nehra V, et al. Exercise training-induced modification of the gut microbiota persists after microbiota colonization and attenuates the response to chemically-induced colitis in gnotobiotic mice. *Gut Microbes*. (2018) 9:115–30. doi: [10.1080/19490976.2017.1372077](https://doi.org/10.1080/19490976.2017.1372077)

56. Pasini E, Corsetti G, Assanelli D, Testa C, Romano C, Dioguardi FS, et al. Effects of chronic exercise on gut microbiota and intestinal barrier in human with type 2 diabetes. *Minerva Med*. (2019) 110:3–11. doi: [10.23736/s0026-4806.18.05589-1](https://doi.org/10.23736/s0026-4806.18.05589-1)

57. Wen C, Wang Q, Gu S, Jin J, Yang N. Emerging perspectives in the gut-muscle axis: the gut microbiota and its metabolites as important modulators of meat quality. *Microb Biotechnol*. (2024) 17:e14361. doi: [10.1111/1751-7915.14361](https://doi.org/10.1111/1751-7915.14361)

58. Lahiri S, Kim H, Garcia-Perez I, Reza MM, Martin KA, Kundu P, et al. The gut microbiota influences skeletal muscle mass and function in mice. *Sci Transl Med*. (2019) 11:eaan5662. doi: [10.1126/scitranslmed.aan5662](https://doi.org/10.1126/scitranslmed.aan5662)

59. Manickam R, Oh HYP, Tan CK, Paramalingam E, Wahli W. Metronidazole causes skeletal muscle atrophy and modulates muscle Chronometabolism. *Int J Mol Sci*. (2018) 19:2418. doi: [10.3390/ijms19082418](https://doi.org/10.3390/ijms19082418)

60. Chen YM, Wei L, Chiu YS, Hsu YJ, Tsai TY, Wang MF, et al. *Lactobacillus plantarum* TWK10 supplementation improves exercise performance and increases muscle mass in mice. *Nutrients*. (2016) 8:205. doi: [10.3390/nu8040205](https://doi.org/10.3390/nu8040205)

61. Inglis JE, Ilich JZ. The microbiome and Osteosarcopenic obesity in older individuals in long-term care facilities. *Curr Osteoporos Rep*. (2015) 13:358–62. doi: [10.1007/s11914-015-0287-7](https://doi.org/10.1007/s11914-015-0287-7)

62. Bindels LB, Beck R, Schakman O, Martin JC, De Backer F, Sohet FM, et al. Restoring specific lactobacilli levels decreases inflammation and muscle atrophy markers in an acute leukemia mouse model. *PLoS One*. (2012) 7:e37971. doi: [10.1371/journal.](https://doi.org/10.1371/journal.pone.0037971) [pone.0037971](https://doi.org/10.1371/journal.pone.0037971)

63. Vallianou N, Stratigou T, Christodoulatos GS, Tsigalou C, Dalamaga M. Probiotics, prebiotics, Synbiotics, Postbiotics, and obesity: current evidence, controversies, and perspectives. *Curr Obes Rep*. (2020) 9:179–92. doi: [10.1007/s13679-020-00379-w](https://doi.org/10.1007/s13679-020-00379-w)

64. Everard A, Lazarevic V, Derrien M, Girard M, Muccioli GG, Neyrinck AM, et al. Responses of gut microbiota and glucose and lipid metabolism to prebiotics in genetic obese and diet-induced leptin-resistant mice. *Diabetes*. (2011) 60:2775–86. doi: [10.2337/](https://doi.org/10.2337/db11-0227) [db11-0227](https://doi.org/10.2337/db11-0227)

65. Varian BJ, Gourishetti S, Poutahidis T, Lakritz JR, Levkovich T, Kwok C, et al. Beneficial bacteria inhibit cachexia. *Oncotarget*. (2016) 7:11803–16. doi: [10.18632/](https://doi.org/10.18632/oncotarget.7730) [oncotarget.7730](https://doi.org/10.18632/oncotarget.7730)

66. Prokopidis K, Giannos P, Kirwan R, Ispoglou T, Galli F, Witard OC, et al. Impact of probiotics on muscle mass, muscle strength and lean mass: a systematic review and meta-analysis of randomized controlled trials. *J Cachexia Sarcopenia Muscle*. (2023) 14:30–44. doi: [10.1002/jcsm.13132](https://doi.org/10.1002/jcsm.13132)

67. Hawley JA. Microbiota and muscle highway – two way traffic. *Nat Rev Endocrinol*. (2020) 16:71–2. doi: [10.1038/s41574-019-0291-6](https://doi.org/10.1038/s41574-019-0291-6)

68. Hintikka JE, Ahtiainen JP, Permi P, Jalkanen S, Lehtonen M, Pekkala S. Aerobic exercise training and gut microbiome-associated metabolic shifts in women with overweight: a multi-omic study. *Sci Rep*. (2023) 13:11228. doi: [10.1038/](https://doi.org/10.1038/s41598-023-38357-6) [s41598-023-38357-6](https://doi.org/10.1038/s41598-023-38357-6)

69. Chew W, Lim YP, Lim WS, Chambers ES, Frost G, Wong SH, et al. Gut-muscle crosstalk. A perspective on influence of microbes on muscle function. *Front Med (Lausanne)*. (2022) 9:1065365. doi: [10.3389/fmed.2022.1065365](https://doi.org/10.3389/fmed.2022.1065365)

70. Ni Lochlainn M, Bowyer RCE, Moll JM, García MP, Wadge S, Baleanu AF, et al. Steves: effect of gut microbiome modulation on muscle function and cognition: the PROMOTe randomised controlled trial. *Nat Commun*. (2024) 15:1859. doi: [10.1038/](https://doi.org/10.1038/s41467-024-46116-y) [s41467-024-46116-y](https://doi.org/10.1038/s41467-024-46116-y)

71. Collins KH, Paul HA, Hart DA, Reimer RA, Smith IC, Rios JL, et al. A high-fat high-sucrose diet rapidly alters muscle integrity, inflammation and gut microbiota in male rats. *Sci Rep*. (2016) 6:37278. doi: [10.1038/srep37278](https://doi.org/10.1038/srep37278)

72. Fang S, Xiong X, Su Y, Huang L, Chen C. 16S rRNA gene-based association study identified microbial taxa associated with pork intramuscular fat content in feces and cecum lumen. *BMC Microbiol*. (2017) 17:162. doi: [10.1186/s12866-017-](https://doi.org/10.1186/s12866-017-1055-x) [1055-x](https://doi.org/10.1186/s12866-017-1055-x)

73. Ticinesi A, Lauretani F, Milani C, Nouvenne A, Tana C, Del Rio D, et al. Aging gut microbiota at the cross-road between nutrition, physical frailty, and sarcopenia: is there a gut-muscle Axis? *Nutrients*. (2017) 9:1303. doi: [10.3390/nu9121303](https://doi.org/10.3390/nu9121303)

74. Thye AY, Law JW, Tan LT, Thurairajasingam S, Chan KG, Letchumanan V, et al. Exploring the gut microbiome in myasthenia gravis. *Nutrients*. (2022) 14:1647. doi: [10.3390/nu14081647](https://doi.org/10.3390/nu14081647)

75. LeBlanc JG, Milani C, de Giori GS, Sesma F, van Sinderen D, Ventura M. Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol*. (2013) 24:160–8. doi: [10.1016/j.copbio.2012.08.005](https://doi.org/10.1016/j.copbio.2012.08.005)

76. Magnúsdóttir S, Ravcheev D, de Crécy-Lagard V, Thiele I. Systematic genome assessment of B-vitamin biosynthesis suggests co-operation among gut microbes. *Front Genet*. (2015) 6:148. doi: [10.3389/fgene.2015.00148](https://doi.org/10.3389/fgene.2015.00148)

77. Zeisel SH, da Costa KA. Choline: an essential nutrient for public health. *Nutr Rev*. (2009) 67:615–23. doi: [10.1111/j.1753-4887.2009.00246.x](https://doi.org/10.1111/j.1753-4887.2009.00246.x)

78. Lin R, Liu W, Piao M, Zhu H. A review of the relationship between the gut microbiota and amino acid metabolism. *Amino Acids*. (2017) 49:2083–90. doi: [10.1007/](https://doi.org/10.1007/s00726-017-2493-3) [s00726-017-2493-3](https://doi.org/10.1007/s00726-017-2493-3)

79. Liu H, Xi Q, Tan S, Qu Y, Meng Q, Zhang Y, et al. The metabolite butyrate produced by gut microbiota inhibits cachexia-associated skeletal muscle atrophy by regulating intestinal barrier function and macrophage polarization. *Int Immunopharmacol*. (2023) 124:111001. doi: [10.1016/j.intimp.2023.111001](https://doi.org/10.1016/j.intimp.2023.111001)

80. Neis EP, Dejong CH, Rensen SS. The role of microbial amino acid metabolism in host metabolism. *Nutrients*. (2015) 7:2930–46. doi: [10.3390/nu7042930](https://doi.org/10.3390/nu7042930)

81. Dukes A, Davis C, El Refaey M, Upadhyay S, Mork S, Arounleut P, et al. The aromatic amino acid tryptophan stimulates skeletal muscle IGF1/p70s6k/mTor signaling in vivo and the expression of myogenic genes in vitro. *Nutrition*. (2015) 31:1018–24. doi: [10.1016/j.nut.2015.02.011](https://doi.org/10.1016/j.nut.2015.02.011)

82. Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nat Rev Endocrinol*. (2015) 11:577–91. doi: [10.1038/](https://doi.org/10.1038/nrendo.2015.128) [nrendo.2015.128](https://doi.org/10.1038/nrendo.2015.128)

83. Lin L, Zhang J. Role of intestinal microbiota and metabolites on gut homeostasis and human diseases. *BMC Immunol*. (2017) 18:2. doi: [10.1186/s12865-016-0187-3](https://doi.org/10.1186/s12865-016-0187-3)

84. Shapiro H, Thaiss CA, Levy M, Elinav E. The cross talk between microbiota and the immune system: metabolites take center stage. *Curr Opin Immunol*. (2014) 30:54–62. doi: [10.1016/j.coi.2014.07.003](https://doi.org/10.1016/j.coi.2014.07.003)

85. Saad MJ, Santos A, Prada PO. Linking gut microbiota and inflammation to obesity and insulin resistance. *Physiology (Bethesda)*. (2016) 31:283–93. doi: [10.1152/](https://doi.org/10.1152/physiol.00041.2015) [physiol.00041.2015](https://doi.org/10.1152/physiol.00041.2015)

86. Stecher B, Hardt WD. The role of microbiota in infectious disease. *Trends Microbiol*. (2008) 16:107–14. doi: [10.1016/j.tim.2007.12.008](https://doi.org/10.1016/j.tim.2007.12.008)

87. Peng L, Li ZR, Green RS, Holzman IR, Lin J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. *J Nutr*. (2009) 139:1619–25. doi: [10.3945/jn.109.104638](https://doi.org/10.3945/jn.109.104638)

88. Cartee GD, Hepple RT, Bamman MM, Zierath JR. Exercise promotes healthy aging of skeletal muscle. *Cell Metab*. (2016) 23:1034–47. doi: [10.1016/j.cmet.2016.05.007](https://doi.org/10.1016/j.cmet.2016.05.007)

89. Ibebunjo C, Chick JM, Kendall T, Eash JK, Li C, Zhang Y, et al. Genomic and proteomic profiling reveals reduced mitochondrial function and disruption of the neuromuscular junction driving rat sarcopenia. *Mol Cell Biol*. (2013) 33:194–212. doi: [10.1128/mcb.01036-12](https://doi.org/10.1128/mcb.01036-12)

90. Picca A, Lezza AMS, Leeuwenburgh C, Pesce V, Calvani R, Bossola M, et al. Circulating mitochondrial DNA at the crossroads of mitochondrial dysfunction and inflammation during aging and muscle wasting disorders. *Rejuvenation Res*. (2018) 21:350–9. doi: [10.1089/rej.2017.1989](https://doi.org/10.1089/rej.2017.1989)

91. Picca A, Fanelli F, Calvani R, Mulè G, Pesce V, Sisto A, et al. Gut Dysbiosis and muscle aging: searching for novel targets against sarcopenia. *Mediat Inflamm*. (2018) 2018:7026198–15. doi: [10.1155/2018/7026198](https://doi.org/10.1155/2018/7026198)

92. Mitchell CJ, Milan AM, Mitchell SM, Zeng N, Ramzan F, Sharma P, et al. The effects of dietary protein intake on appendicular lean mass and muscle function in elderly men: a 10-wk randomized controlled trial. *Am J Clin Nutr*. (2017) 106:1375–83. doi: [10.3945/ajcn.117.160325](https://doi.org/10.3945/ajcn.117.160325)

93. Ni Lochlainn M, Bowyer RCE, Steves CJ. Steves: dietary protein and muscle in aging people: the potential role of the gut microbiome. *Nutrients*. (2018) 10:929. doi: [10.3390/nu10070929](https://doi.org/10.3390/nu10070929)

94. Pols TW, Noriega LG, Nomura M, Auwerx J, Schoonjans K. The bile acid membrane receptor TGR5: a valuable metabolic target. *Dig Dis*. (2011) 29:37–44. doi: [10.1159/000324126](https://doi.org/10.1159/000324126)

95. Wang-Sattler R, Yu Z, Herder C, Messias AC, Floegel A, He Y, et al. Novel biomarkers for pre-diabetes identified by metabolomics. *Mol Syst Biol*. (2012) 8:615. doi: [10.1038/msb.2012.43](https://doi.org/10.1038/msb.2012.43)