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Impact of dietary selenium and blood concentration on liver function: a population-based study

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Background: Evidence on the association between selenium and liver function parameters is limited and controversial.

Methods: Data on dietary selenium intake, blood selenium concentration, and liver function parameters were obtained from the National Health and Nutrition Examination Survey (NHANES) 2017–2020. Associations between selenium (dietary intake and blood concentration) and liver function parameters [alanine aminotransferase (ALT), aspartate aminotransferase (AST), the ALT/ AST ratio, gamma-glutamyl transferase (GGT), and alkaline phosphatase (ALP)] were assessed using multivariate linear regression models. Subgroup analyses and interaction tests were conducted to examine differences in associations according to age, gender, body mass index (BMI), diabetes, and physical activity.

Results: The study included 6,869 participants after screening. The multivariate linear regression model revealed that dietary selenium intake was positively associated with ALT (β = 0.112, 95% CI = 0.041, 0.183) and the ALT/AST ratio (β = 0.002, 95% CI = 0.001, 0.004) after adjustment for covariates. Results of blood selenium concentration also showed that higher blood selenium levels were positively associated with ALT (β = 0.436, 95% CI = 0.308, 0.564), AST (β = 0.112, 95% CI = 0.015, 0.208), and the ALT/AST ratio (β = 0.012, 95% CI = 0.009, 0.015). However, ALP decreased with increasing blood selenium concentration (β = -0.207, 95% CI = -0.414, -0.000). In addition, we found significant differences in the effect of selenium on liver function parameters according to age, gender, and BMI.

Conclusion: Dietary selenium intake and blood concentration affect liver function parameters. These findings suggest that further research is needed to explore these associations to promote liver health and disease prevention.

KEYWORDS

dietary selenium intake, blood selenium concentration, liver function, cross-sectional study, NHANES

1 Introduction

The liver, being the largest metabolic organ in the human body, is essential for metabolism, detoxification, and biochemical synthesis (1). Liver injury can occur due to a variety of factors, including viral infections, excessive alcohol consumption, drug-induced hepatotoxicity, and metabolic disorders (2). Liver function tests (LFTs) are crucial for assessing liver status and the extent of injury. These tests include measurements of alanine aminotransferase (ALT), aspartate aminotransferase (AST), the ALT/AST ratio, gamma-glutamyl transferase (GGT), and alkaline phosphatase (ALP). The liver is not only sensitive to a variety of factors but is also intimately involved in the metabolism of numerous trace elements in the body (3, 4). In the multifaceted study of liver health, the role of trace minerals has received increasing attention (5–8).

Selenium is a vital micronutrient that is absorbed by the human body through the food chain, primarily from seafood, plant sources, and animal products (9). In the human body, selenium is not only a component of several important proteins, such as glutathione peroxidases and selenoprotein P, but also plays a role in antioxidant defense and maintaining cellular redox balance (10). Because of these important functions, selenium intake is closely linked to overall health, impacting cardiovascular disease (11, 12), immune function (13, 14), and thyroid health (15, 16). Blood selenium concentration is a crucial and reliable indicator of an individual's overall selenium nutritional status. However, the relationship between selenium intake and blood concentration is not linear and is influenced by individual absorption, metabolism, and excretion processes (17). Both inadequate and excessive selenium intake can have adverse health effects. Therefore, understanding the relationship between dietary selenium intake and blood selenium concentration and how these factors influence human health, particularly in maintaining liver health, is critical for developing dietary guidelines and preventive strategies.

The role of selenium in liver health has received increasing attention due to its essential functions in enzymatic processes and antioxidant defense mechanisms (18). Selenium contributes significantly to the ability of the liver to mitigate oxidative stress, a known contributor to liver injury (19-21). Oxidative stress in the liver can be caused by several factors, including environmental toxins, metabolic imbalances, and chronic alcohol use, all of which can lead to liver cell damage and subsequent liver disease (22, 23). Research has shown that selenium deficiency may exacerbate liver injury by impairing the liver's antioxidant defense system, thereby increasing susceptibility to oxidative damage (24, 25). However, the relationship between selenium and liver health is complex and dose-dependent, with both selenium deficiency and excess posing risks to liver function. This study aims to elucidate the relationship between dietary selenium intake, blood selenium concentration, and liver damage in US adults using data from the 2017-2020 National Health and Nutrition Examination Survey (NHANES). By examining these associations, this research aims to provide insight into the range of selenium intake and blood selenium concentrations for liver health and to contribute to the development of dietary guidelines and interventions aimed at preventing liver injury.

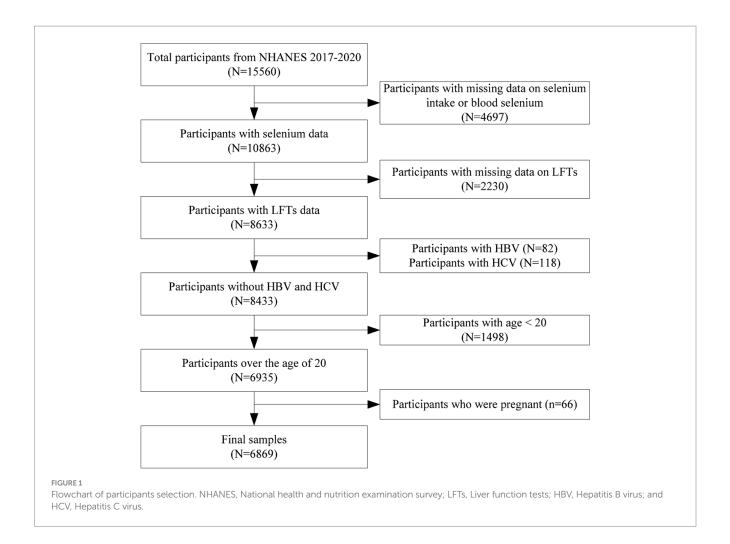
2 Materials and methods

2.1 Study population

The NHANES is an annual, nationally representative survey conducted in the United States by the National Center for Health Statistics (NCHS) of the Centers for Disease Control and Prevention (CDC). NHANES has been utilizing an advanced multistage probability sampling design since 1999 to gather data biennially from a diverse, non-institutionalized section of the United States population. Ethical approval for NHANES was secured from the NCHS Research Ethics Review Board, and each participant willingly provided written informed consent. NHANES data from 2017 to 2020 were used in this study. From the initial pool of 15,560 eligible participants, several exclusions were made to ensure the integrity of the analysis: 4,697 individuals were excluded due to missing data on selenium intake or blood selenium levels, and 2,230 individuals were excluded due to missing LFTs results. The primary objective of this study is to investigate the impact of selenium on liver function in the general population. Including hepatitispositive individuals could complicate the interpretation of results, as their abnormal liver function parameters may be attributable to the disease itself rather than selenium levels. Therefore, we excluded 82 participants who were positive for hepatitis B antibody and 118 participants who were positive for hepatitis C antibody. We excluded 1,498 individuals under the age of 20 because they are in a developmental stage, and their liver function parameters and selenium metabolism may differ significantly from adults, affecting the comparability and interpretability of the results. This exclusion also helps to enhance the homogeneity and data consistency of the study sample, and facilitates the analysis of the impact of educational level on the study results. We also excluded 66 individuals who reported being pregnant to refine the cohort for more accurate and relevant results. The study ultimately encompassed 6,869 participants (Figure 1).

2.2 Measurement of liver function

In NHANES, fasting blood samples were systematically collected from participants at a mobile examination center, ensuring consistency and standardization in the sample collection process. These samples were then immediately refrigerated and sent to a central laboratory for careful analysis. In the laboratory, serum liver function biomarkers were quantitatively assessed using the advanced Beckman Coulter DxC800 Synchron clinical system. ALT, primarily found in liver cells, is an enzyme involved in amino acid metabolism (26, 27). When liver cells are damaged or die, ALT is released into the blood. Elevated serum ALT levels are often seen as a sign of liver damage. AST, found in liver cells, cardiomyocytes, muscle cells, and other tissues, is not as liverspecific as ALT (26, 27). However, AST levels increase in response to liver injury. The ALT to AST ratio is used to differentiate between various types of liver disease. GGT and ALP are indicators of cholestasis (26, 27). Specifically, in this crosssectional study, ALT, AST, ALT/AST ratio, GGT, and ALP were used to assess liver function impairment.



2.3 Measurement of dietary selenium intake and blood selenium level

The process for gathering dietary selenium data has been carefully designed to ensure accuracy and reliability. Dietary intake data were collected over two non-consecutive days, beginning with a face-to-face interview in a mobile examination center and followed by a telephone interview on a subsequent day. Participants were instructed to provide a detailed account of all food and beverages consumed in the previous 24 h, detailing specific meal contents and quantities. These details were then analyzed using the USDA's Food and Nutrient Database for Dietary Studies, enabling an accurate estimation of selenium intake. To enhance individual dietary data accuracy, this study averaged selenium intake from two recall days, minimizing day-to-day dietary variation. In NHANES 2017–2020, whole blood selenium concentrations were measured using inductively coupled plasma mass spectrometry. Detailed measurements can be found in a previously published study (28).

2.4 Measurement of covariates

Covariates were identified as confounders based on previously published studies (29–31). Demographic characteristics, including gender, education, age, ethnicity, and family income-to-poverty ratio (PIR), were considered. Health-related factors included body mass index (BMI), alcohol consumption (at least 12 alcoholic beverages per year), smoking (at least 100 cigarettes in a lifetime), and physical activity (engaging in moderate or vigorous exercise or recreational activities weekly). Clinical factors included diabetes mellitus, fasting glucose, and total cholesterol (TC).

2.5 Statistical analysis

Dietary selenium intakes and blood selenium concentrations were divided into quartiles, with quartiles 1–4 representing progressively increasing intakes or concentrations. In the sample characteristics, continuous variables were expressed as means±standard deviation, and categorical variables as count (number) and percentage. To compare means between groups, one-way ANOVA was used, while chi-squared tests were employed for percentages. Associations between selenium (dietary intake and blood concentration) and liver function parameters (ALT, AST, ALT/AST, GGT, and ALP) were calculated using multivariate linear regression models. The multivariate analysis utilized three models: crude model with no adjustment; Model 1 adjusted for age, gender, and race; and Model 2 included adjustments for the full range of covariates. Subgroup analyses and interaction tests were conducted to explore variations in the associations by age, gender, race, education, and BMI. p values less than 0.05 were considered statistically significant. The statistical analyses and graphical representations in this study were generated using R (version 4.1.3) and EmpowerStats (version 2.0).

3 Results

3.1 Baseline characteristics

Table 1 presents a comparison of baseline characteristics of participants across four quartiles of dietary selenium intake. A total of 6,869 participants with an average age of 50.788±17.384 were included in this study, of which 48.62% were males and 51.38% were females. Notably, selenium intakes were higher than international recommendations and blood concentrations were also quite high. In addition, liver enzymes as well as cholesterol were within reference ranges. Blood selenium concentrations increased with each quartile of dietary selenium intake (p < 0.05). Significant differences were observed among the quartiles in age, gender, race, education level, family income-to-poverty ratio (PIR), BMI, fasting glucose, drinking habits, physical activity, and various liver function markers (all p < 0.05). Participants in the highest quartile were more likely to be male, consume alcohol, have above high school education, engage in physical activity, and have higher BMI, fasting glucose, ALT, AST, GGT, and ALT/AST levels, but lower ALP levels compared to those in the lowest quartile.

Table 2 compares the baseline characteristics of participants according to four quartiles of blood selenium concentration. With increasing dietary selenium intake, the quartiles of the participants' blood selenium concentrations increased (p < 0.05). Significant differences in age, gender, race, education level, fasting glucose, drinking habits, physical activity, ALT, AST, GGT, ALT/AST, and TC (all p < 0.05). Participants in the highest quartile were more likely to be male, to have above high school education, to be physically active, and to have higher levels of fasting glucose, ALT, AST, GGT, ALT/AST, and TC, compared to participants in the lowest quartile.

3.2 Relationship between dietary selenium intake and liver function parameters

To better observe the relationship between selenium intake and liver function parameters, the unit of selenium intake was converted to $10 \,\mu g$ per day. The results of multivariate linear regression analyses examining the association between dietary selenium intake and liver function parameters are shown in Table 3. The present study found a positive association between dietary selenium intake and ALT and the ALT/AST ratio whatever the model. There was no robust association with other liver function parameters. The effect of selenium intake on liver function parameters was illustrated through a smoothed curve fit (Figure 2). Dietary selenium intake was then categorized into quartile variables to examine the linear relationship (Table 4). In model 2, higher selenium intake was associated with higher effect values for ALT and ALT/AST.

3.3 Relationship between blood selenium concentration and liver function parameters

Similarly, blood selenium concentrations were converted to $10 \mu g/L$. Results from multivariate linear regression analyses examining the association between blood selenium concentration and liver function parameters are presented in Table 5. Consistent with the findings for dietary selenium intake, blood selenium concentrations were positively correlated with ALT and the ALT/AST ratio. Additionally, blood selenium concentrations were positively correlated with ALP in model 2. The effect of blood selenium on liver function parameters was illustrated through a smoothed curve fit (Figure 3). Blood selenium concentration was then categorized into quartile variables to examine the linear relationship (Table 6). In model 2, similar trend test results were observed for ALT and the ALT/AST ratio (*p* for trend <0.001), but not for AST and ALP.

3.4 Subgroup analysis

Based on the above results, we found correlations between dietary selenium and blood concentrations with ALT. Subgroup analyses further explored the effects of age, gender, BMI, diabetes, and physical activity on these associations. The present results showed that correlations between selenium intake and blood concentrations with ALT were not consistent. Significant associations were found between dietary selenium and ALT in participants aged 20-40 years, female, BMI>30, non-diabetic, and with or without physical activity (Figure 4). The results of interaction tests indicated that there were no statistically significant variations in the association between dietary selenium intake and ALT across different stratifications, implying that age, gender, BMI, diabetes, and physical activity did not have a significant impact on this positive correlation. However, interaction tests revealed that the association between selenium blood concentrations and ALT levels is significantly influenced by age, gender, and BMI stratifications (Figure 5). Higher selenium concentrations were associated with a significant increase in ALT levels across all age and gender stratifications, with a more pronounced effect in individuals aged 20-40 and in males. The increase in ALT levels was also significant in obese individuals. In addition, although the interaction test for physical activity and diabetes status was not significant, higher selenium concentrations still significantly increased ALT levels in all stratifications of physical activity and diabetes status.

4 Discussion

In this cross-sectional population-based study, dietary selenium intake was consistently positively associated with ALT and the ALT/ AST ratio before and after adjustment for covariates. In addition, results indicated that high blood selenium was positively associated with ALT, AST, and ALT/AST ratio. However, ALP levels decreased with increasing blood selenium concentrations. In conclusion, this cross-sectional study demonstrated that dietary selenium intake and blood selenium concentration affect liver function parameters. Selenium is an essential trace element, and adequate intake is crucial TABLE 1 Baseline characteristics of participants according to dietary selenium intake.

Variables	Dietary selenium intake (µg/day)				
	Q1 (<65.4) <i>N</i> = 1,717	Q2 (65.4–93.3) N = 1,717	Q3 (93.3–127.8) N = 1,715	Q4 (>127.8) <i>N</i> = 1,720	
Age, year, Mean (SD)	49.01 (17.70)	49.45 (17.90)	49.10 (17.16)	45.96 (16.02)	<0.001
Gender, <i>N</i> (%)					<0.001
Male	584 (34.01%)	687 (40.01%)	881 (51.37%)	1,188 (69.07%)	
Female	1,133 (65.99%)	1,030 (59.99%)	834 (48.63%)	532 (30.93%)	
Race, N (%)					<0.001
Mexican American	175 (10.19%)	205 (11.94%)	218 (12.71%)	231 (13.43%)	
Other Hispanic	202 (11.76%)	158 (9.20%)	168 (9.80%)	179 (10.41%)	
Non-Hispanic White	598 (34.83%)	671 (39.08%)	658 (38.37%)	593 (34.48%)	
Non-Hispanic Black	445 (25.92%)	442 (25.74%)	428 (24.96%)	423 (24.59%)	
Other race-including multi-racial	297 (17.30%)	241 (14.04%)	243 (14.17%)	294 (17.09%)	
Education level, N (%)					<0.001
Above high school	873 (50.84%)	986 (57.43%)	1,092 (63.67%)	1,058 (61.51%)	
Completed high school	391 (22.77%)	299 (17.41%)	267 (15.57%)	259 (15.06%)	
Less than high school	453 (26.38%)	432 (25.16%)	356 (20.76%)	403 (23.43%)	
Ratio of family income to poverty, Mean (SD)	2.81 (1.55)	3.10 (1.52)	3.35 (1.53)	3.21 (1.55)	<0.001
BMI, kg/m², Mean (SD)	29.44 (7.36)	29.46 (6.93)	29.88 (6.90)	30.51 (7.69)	<0.001
Diabetes, N (%)					0.948
Yes	287 (16.72%)	260 (15.14%)	265 (15.45%)	239 (13.90%)	
No	1,393 (81.13%)	1,409 (82.06%)	1,389 (80.99%)	1,428 (83.02%)	
Borderline	37 (2.15%)	48 (2.80%)	61 (3.56%)	53 (3.08%)	
Fasting glucose, mg/dL, Mean (SD)	110.96 (20.45)	110.48 (20.27)	112.75 (24.68)	112.63 (25.18)	0.004
Drink, <i>N</i> (%)					<0.001
Yes	647 (37.68%)	755 (43.97%)	839 (48.92%)	896 (52.09%)	
No	1,070 (62.32%)	962 (56.03%)	876 (51.08%)	824 (47.91%)	
Smoke, <i>N</i> (%)					0.105
Yes	693 (40.36%)	702 (40.89%)	710 (41.40%)	772 (44.88%)	
No	1,024 (59.64%)	1,015 (59.11%)	1,005 (58.60%)	948 (55.12%)	
Physical activity, N (%)					<0.001
Yes	723 (42.11%)	793 (46.19%)	864 (50.38%)	913 (53.08%)	
No	994 (57.89%)	924 (53.81%)	851 (49.62%)	807 (46.92%)	
ALT, U/L, Mean (SD)	20.27 (14.59)	20.93 (14.38)	22.98 (16.46)	25.13 (16.64)	<0.001
ALP, U/L, Mean (SD)	77.18 (25.49)	75.31 (24.40)	73.64 (25.44)	75.44 (23.88)	0.016
AST, U/L, Mean (SD)	20.55 (10.12)	21.03 (12.49)	22.18 (12.17)	22.15 (9.20)	<0.001
GGT, U/L, Mean (SD)	27.05 (32.73)	27.39 (31.75)	29.35 (43.50)	31.61 (35.50)	<0.001
ALT/AST, Mean (SD)	0.96 (0.34)	0.98 (0.33)	1.01 (0.35)	1.11 (0.37)	<0.001
TC, mmol/L, Mean (SD)	4.89 (1.06)	4.88 (1.05)	4.86 (1.01)	4.79 (1.05)	0.192
Blood selenium concentration, μg/L, Mean (SD)	187.66 (26.07)	187.52 (28.38)	188.87 (26.45)	190.76 (27.99)	<0.001

Significant results highlighted in bold. BMI, Body mass index; TC, Total cholesterol; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; GGT, Gamma-glutamyl transferase; ALP, Alkaline phosphatase.

TABLE 2 Baseline characteristics of participants according to blood selenium concentration.

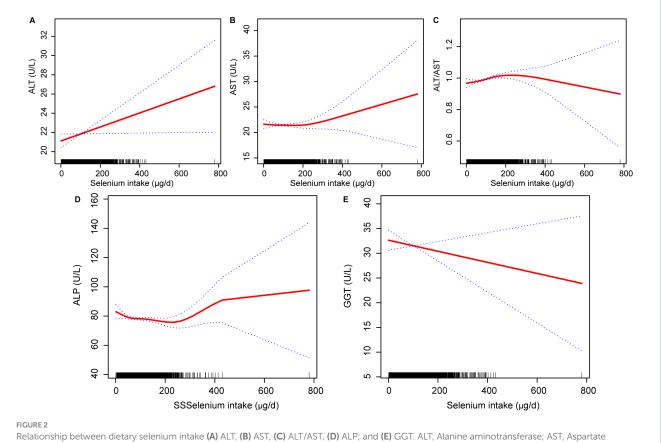
Variables	Blood selenium concentration (µg/L)					
	Q1 (<168.8)	Q2 (168.8– 183.75)	Q3 (183.75– 200.55)	Q4 (>200.55)		
	N = 1,716	N = 1,717	<i>N</i> = 1,718	N = 1,718		
Age, year, Mean (SD)	49.29 (18.08)	47.50 (17.31)	47.91 (17.07)	48.78 (16.63)	0.014	
Gender, N (%)					<0.001	
Male	758 (44.17%)	820 (47.76%)	836 (48.66%)	926 (53.90%)		
Female	958 (55.83%)	897 (52.24%)	882 (51.34%)	792 (46.10%)		
Race, N (%)					<0.001	
Mexican American	157 (9.15%)	212 (12.35%)	242 (14.09%)	218 (12.69%)		
Other Hispanic	189 (11.01%)	185 (10.77%)	159 (9.25%)	174 (10.13%)		
Non-Hispanic White	604 (35.20%)	621 (36.17%)	666 (38.77%)	629 (36.61%)		
Non-Hispanic Black	559 (32.58%)	470 (27.37%)	367 (21.36%)	342 (19.91%)		
Other race-including multi-racial	207 (12.06%)	229 (13.34%)	284 (16.53%)	355 (20.66%)		
Education level, N (%)					<0.001	
Above high school	916 (53.38%)	993 (57.83%)	1,051 (61.18%)	1,049 (61.06%)		
Completed high school	459 (26.75%)	430 (25.04%)	368 (21.42%)	387 (22.53%)		
Less than high school	341 (19.87%)	294 (17.12%)	299 (17.40%)	282 (16.41%)		
Ratio of family income to poverty, Mean (SD)	3.05 (1.60)	3.15 (1.54)	3.16 (1.57)	3.14 (1.50)	0.173	
BMI, kg/m², Mean (SD)	29.65 (7.64)	29.83 (7.28)	29.97 (7.01)	29.87 (7.10)	0.662	
Diabetes, N (%)					0.444	
Yes	266 (15.50%)	247 (14.39%)	249 (14.49%)	289 (16.82%)		
No	1,407 (81.99%)	1,419 (82.64%)	1,418 (82.54%)	1,375 (80.03%)		
Borderline	43 (2.51%)	51 (2.97%)	51 (2.97%)	54 (3.14%)		
Fasting glucose, mg/dL, Mean (SD)	112.00 (27.17)	110.48 (18.44)	110.91 (20.36)	113.50 (25.04)	<0.001	
Drink, <i>N</i> (%)					<0.001	
Yes	728 (42.42%)	798 (46.48%)	838 (48.78%)	773 (44.99%)		
No	988 (57.58%)	919 (53.52%)	880 (51.22%)	945 (55.01%)		
Smoke, <i>N</i> (%)					0.056	
Yes	737 (42.95%)	726 (42.28%)	700 (40.75%)	714 (41.56%)		
No	979 (57.05%)	991 (57.72%)	1,018 (59.25%)	1,004 (58.44%)		
Physical activity, N (%)					0.021	
Yes	741 (43.18%)	825 (48.05%)	868 (50.52%)	859 (50.00%)		
No	975 (56.82%)	892 (51.95%)	850 (49.48%)	859 (50.00%)		
ALT, U/L, Mean (SD)	19.55 (12.01)	21.81 (15.33)	23.27 (15.48)	24.33 (18.20)	<0.001	
ALP, U/L, Mean (SD)	75.93 (28.87)	74.44 (23.69)	76.96 (24.17)	74.07 (22.97)	0.235	
AST, U/L, Mean (SD)	20.80 (10.00)	21.30 (11.02)	21.74 (9.63)	22.06 (13.11)	<0.001	
GGT, U/L, Mean (SD)	28.75 (45.82)	27.06 (31.40)	29.21 (34.06)	30.49 (34.67)	0.044	
ALT/AST, Mean (SD)	0.93 (0.32)	0.99 (0.34)	1.04 (0.36)	1.08 (0.37)	<0.001	
TC, mmol/L, Mean (SD)	4.59 (0.93)	4.83 (1.05)	4.86 (1.02)	5.06 (1.09)	<0.001	
Dietary selenium intake, μg/day, Mean (SD)	157.12 (10.20)	176.47 (4.26)	191.56 (4.86)	220.48 (25.55)	<0.001	

Significant results highlighted in bold. BMI, Body mass index; TC, Total cholesterol; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; GGT, Gamma-glutamyl transferase; and ALP, Alkaline phosphatase.

Exposure	Outcome	β (95%CI), <i>p</i> value				
		Crude	Model 1	Model 2		
Selenium intake	ALT	0.416 (0.345, 0.488) < 0.001	0.163 (0.090, 0.236) < 0.001	0.112 (0.041, 0.183) 0.002		
	AST	0.162 (0.111, 0.213) < 0.001	0.058 (0.005, 0.112) 0.032	0.049 (-0.005, 0.102) 0.075		
	ALT/AST	0.010 (0.009, 0.012) < 0.001	0.004 (0.002, 0.006) < 0.001	0.002 (0.001, 0.004) 0.005		
	ALP	-0.135 (-0.249, -0.020) 0.021	-0.079 (-0.197, 0.039) 0.190	-0.039 (-0.153, 0.076) 0.507		
	GGT	0.266 (0.098, 0.433) 0.002	-0.048 (-0.223, 0.126) 0.588	-0.060 (-0.232, 0.111) 0.492		

TABLE 3 Multiple linear regression associations of dietary selenium intake (continuous) with liver function parameters.

Crude: no covariates were adjusted. Model 1: age, gender, and race were adjusted. Model 2: age, gender, race, education, ratio of family income to poverty, diabetes, BMI, drink, smoke, physical activity, and TC were adjusted. Significant results highlighted in bold. BMI, Body mass index; TC, Total cholesterol, ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; GGT, Gamma-glutamyl transferase; and ALP, Alkaline phosphatase.



aminotransferase; GGT, Gamma-glutamyl transferase; ALP, Alkaline phosphatase.

for maintaining liver health. However, the use of selenium supplements should be approached with caution in light of our findings. While selenium has health benefits, excessive intake can lead to adverse effects, including potential effects on liver function. Therefore, clinicians should consider patients' total selenium intake when recommending supplements to avoid overconsumption.

The role of selenium as a micronutrient essential for various bodily functions has been well-documented over the decades. However, supplementation, especially in the context of the Selenium and Vitamin E Cancer Prevention Trial, has raised concerns due to adverse outcomes like increased incidence of type 2 diabetes and prostate cancer (32, 33). Reports from the 1930s about selenium's potential hepatotoxic effects in overexposure cases set a precedent for careful selenium dosage consideration in supplementation practices (34). In a recent study, Li et al. (35) associated blood selenium with elevated AST in 15,328 samples from NHANES 2011–2018, which is consistent with our findings. However, they did not explore dietary selenium intake, subgroup analyses, or BMI's impact on selenium metabolism. Yang et al.'s cross-sectional study in China with 8,550 participants found significant ALT and AST increases with rising plasma selenium concentrations (36). They observed a linear correlation between plasma selenium and ALT and AST when converting plasma selenium concentrations into a categorical variable. Moreover, our results align with Longnecker et al. (37), who noted a

LFTs	Model		Dietary selenium intake (quartile)				
		Q1	Q2 β (95% CI)	Q3 β (95% CI)	Q4 β (95% CI)		
ALT	Crude	Reference	0.660 (-0.417, 1.738)	2.710 (1.651, 3.769)	4.862 (3.799, 5.924)	<0.001	
	Model 1	Reference	0.046 (-0.995, 1.087)	1.102 (0.069, 2.134)	1.589 (0.524, 2.654)	<0.001	
	Model 2	Reference	-0.064 (-1.073, 0.945)	0.660 (-0.348, 1.668)	0.897 (-0.140, 1.934)	0.039	
AST	Crude	Reference	0.484 (-0.282, 1.250)	1.636 (0.882, 2.389)	1.606 (0.851, 2.361)	<0.001	
	Model 1	Reference	0.198 (-0.562, 0.957)	0.922 (0.169, 1.675)	0.219 (-0.558, 0.996)	0.277	
	Model 2	Reference	0.136 (-0.622, 0.893)	0.809 (0.052, 1.565)	0.113 (-0.666, 0.891)	0.439	
ALT/AST	Crude	Reference	0.021 (-0.003, 0.045)	0.056 (0.032, 0.079)	0.149 (0.126, 0.173)	<0.001	
	Model 1	Reference	0.006 (-0.018, 0.029)	0.016 (-0.007, 0.039)	0.068 (0.045, 0.092)	<0.001	
	Model 2	Reference	0.003 (-0.018, 0.024)	0.001 (-0.021, 0.022)	0.042 (0.021, 0.064)	<0.001	
ALP	Crude	Reference	-1.873 (-3.584, -0.161)	-3.536 (-5.218, -1.853)	-1.734 (-3.421, -0.047)	0.016	
	Model 1	Reference	-1.831 (-3.509, -0.153)	-3.240 (-4.905, -1.575)	-0.735 (-2.452, 0.982)	0.220	
	Model 2	Reference	-0.818 (-2.443, 0.808)	-1.858 (-3.482, -0.234)	-0.050 (-1.721, 1.620)	0.704	
GGT	Crude	Reference	0.337 (-2.174, 2.848)	2.301 (-0.168, 4.770)	4.557 (2.080, 7.033)	<0.001	
	Model 1	Reference	-0.556 (-3.043, 1.931)	0.271 (-2.196, 2.738)	0.716 (-1.828, 3.260)	0.453	
	Model 2	Reference	-0.198 (-2.635, 2.239)	0.405 (-2.029, 2.840)	0.741 (-1.764, 3.246)	0.476	

TABLE 4 Multiple linear regression associations of dietary selenium intake (quartile) with liver function parameters.

Crude: no covariates were adjusted. Model 1: age, gender, and race were adjusted. Model 2: age, gender, race, education, ratio of family income to poverty, diabetes, BMI, drink, smoke, physical activity, and TC were adjusted. Significant results highlighted in bold. BMI, Body mass index; TC, Total cholesterol; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; GGT, Gamma-glutamyl transferase; and ALP, Alkaline phosphatase.

TABLE 5 Multiple linear regression associations of blood selenium concentration (continuous) with liver function parameters.

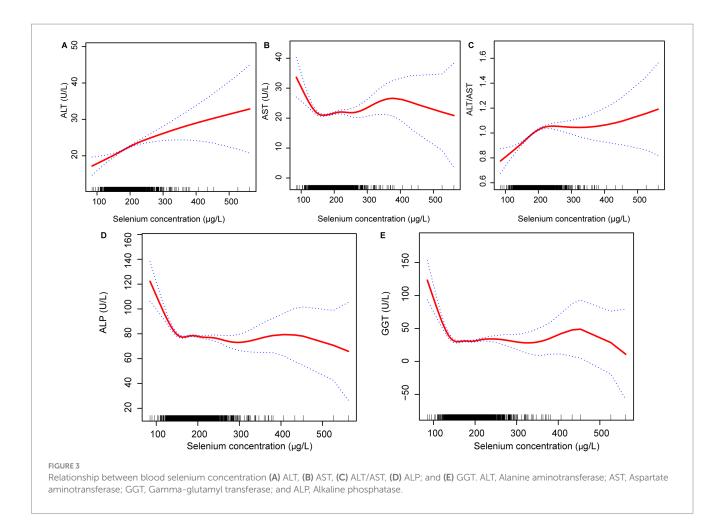
Exposure	Outcome	β (95%CI), <i>P</i> value			
		Crude	Model 1	Model 2	
Blood selenium	ALT	0.644 (0.509, 0.780) < 0.001	0.527 (0.397, 0.658) < 0.001	0.436 (0.308, 0.564) <0.001	
	AST	0.205 (0.109, 0.301) < 0.001	0.154 (0.058, 0.250) 0.002	0.112 (0.015, 0.208) 0.023	
	ALT/AST	0.017 (0.014, 0.020) < 0.001	0.014 (0.011, 0.017) < 0.001	0.012 (0.009, 0.015) < 0.001	
	ALP	-0.228 (-0.443, -0.013) 0.038	-0.211 (-0.423, 0.000) 0.051	-0.207 (-0.414, -0.000) 0.049	
	GGT	0.215 (-0.101, 0.530) 0.182	0.099 (-0.214, 0.413) 0.535	-0.109 (-0.420, 0.201) 0.489	

Crude: no covariates were adjusted. Model 1: age, gender, and race were adjusted. Model 2: age, gender, race, education, ratio of family income to poverty, diabetes, BMI, drink, smoke, physical activity, and TC were adjusted. Significant results highlighted in bold. BMI, Body mass index; TC, Total cholesterol; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; GGT, Gamma-glutamyl transferase; and ALP, Alkaline phosphatase.

positive correlation between selenium intake and ALT levels in United States selenium-enriched regions. Such corroborative findings reinforce the theoretical link between selenium and liver health.

The major antioxidant effects of selenium are mediated by selenium-dependent glutathione peroxidase (GPx) and selenoproteins (38–40). Maximum selenoprotein levels in the blood were in the range of 70–90 μ g/L (41). The increase in blood selenium above these levels does not reflect an increase in selenoprotein levels but primarily reflects non-specific binding of selenomethionine to albumin instead of methionine (41). The mean serum selenium level of the participants was 186.11 μ g/L, and the increase in selenomethionine. A previous report also showed that the dietary selenium intake of United States adults far exceeds the amount of selenium needed to achieve maximal blood selenoprotein levels (42). Adequate selenium intake regulates the activity of selenoproteins, such as thioredoxin reductase (TrxR), thereby enhancing the body's antioxidant capacity and mitigating the adverse effects of oxidative stress (43). TrxR and GPx, as core

components of the thioredoxin (Trx) and glutathione (GSH) systems, constitute key thiol/selenol-dependent antioxidant systems. These systems maintain redox balance within the body and protect cells from oxidative damage (44-46). Selenium deficiency leads to decreased TrxR activity (47, 48). Compared to low selenium levels, selenium supplementation increases TrxR activity but stabilizes at normal levels. Notably, excessive selenium does not further increase TrxR activity. Conversely, in an acetaminophen (APAP)-induced liver injury model, selenium-enriched mice exhibit more severe liver damage than mice with normal selenium levels (49). Typical selenium compounds, such as sodium selenite, have been widely used at high concentrations to inhibit tumor cell growth (50). In mammals, selenite is reduced by TrxR to produce hydrogen selenide, which is highly reactive with oxygen, thereby generating reactive oxygen species (ROS). Selenomethionine, an organic selenium compound, is one of the predominant forms of selenium in nature. It is metabolized through various pathways within cells. Compared to selenite, higher selenium doses generates hydrogen selenide as a metabolic product,



leading to oxidative stress (51). In summary, excessive selenium significantly increases oxidative stress levels within hepatocytes, causing cellular damage, which may be the underlying biological mechanism for the observed elevation in ALT levels induced by excessive selenium.

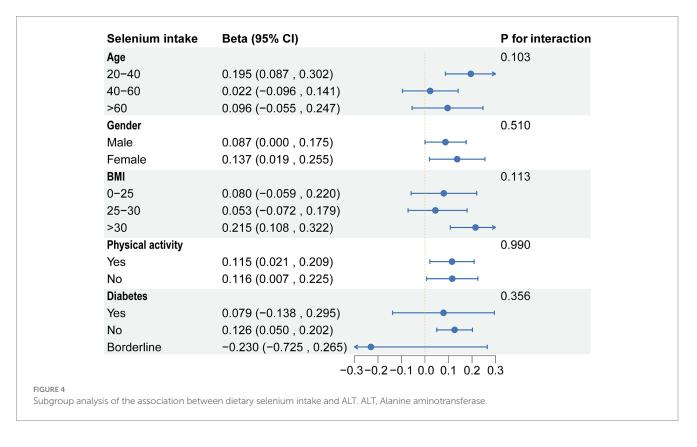
In subgroup analyses, significant associations were found between dietary selenium and ALT in participants aged 20-40 years, female, BMI >30, and non-diabetes. The results of interaction tests indicated that there were no statistically significant variations in the association between dietary selenium intake and ALT across different stratifications. However, interaction tests revealed that the association between selenium blood concentrations and ALT levels was significantly influenced by age, gender, and BMI stratifications. Higher selenium concentrations were associated with a significant increase in ALT levels across all age and gender stratifications, with a more pronounced effect in individuals aged 20-40 and in males. The increase in ALT levels was also significant in obese individuals. Adults between the ages of 20 and 40 are in a period of high metabolic activity, which may make them more sensitive to selenium intake, especially if the intake exceeds the body's needs, which may have adverse effects on liver function (52). Individuals with higher body weight, especially those who are obese, may have altered selenium distribution and action owing to metabolic activities in adipose tissue, which could amplify adverse effects on liver function (53). These findings underscore the need to consider individual variability in research on the relationship between selenium intake and liver health. Future strategies and guidelines should be personalized, accounting for the unique needs and risks of diverse populations to ensure beneficial selenium intake levels for liver health.

In this cross-sectional study, the effects of dietary selenium intake on ALT, ALT/AST, and GGT were consistent with the results for blood selenium concentration. However, the findings regarding AST and ALP were inconsistent. Methodologically, the differences might be due to the nature of the data collection. Dietary selenium intake data is typically based on 24-h dietary recalls, which can be affected by recall bias and reporting accuracy. Blood selenium levels, on the other hand, reflect the actual storage and utilization of selenium in the body, which may have a time lag and individual variability. Additionally, the bioavailability of dietary selenium can vary depending on the type of food and an individual's absorption capacity, leading to different impacts of dietary selenium intake and blood selenium levels on AST and ALP. Biologically, the metabolism and function of selenium are complex and varied. Selenium primarily exerts its antioxidant effects through GPx and selenoprotein P, which have different expression and activity levels in various tissues, possibly resulting in differential impacts on liver enzymes. ALT is predominantly found in hepatocytes, while AST is expressed in multiple tissues including the liver, heart, and muscles, which might explain the different mechanisms of selenium's effects on ALT and AST. Furthermore, selenium might influence ALP levels through its role in regulating apoptosis and inflammatory responses, which can vary depending on an individual's genetic background and health status.

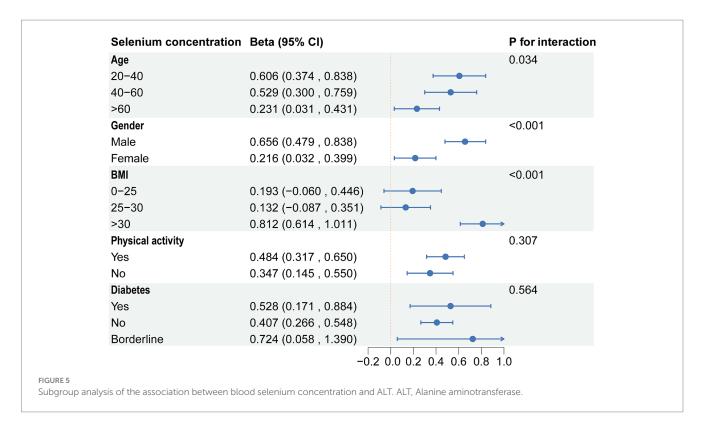
LFTs	Model		Blood selenium concentration (quartile)				
		Q1	Q2 β (95% CI)	Q3 β (95% CI)	Q4 β (95% CI)		
ALT	Crude	Reference	2.263 (1.161, 3.364)	3.723 (2.643, 4.803)	4.779 (3.711, 5.848)	<0.001	
	Model 1	Reference	1.947 (0.889, 3.004)	3.191 (2.153, 4.230)	3.772 (2.742, 4.802)	<0.001	
	Model 2	Reference	1.468 (0.441, 2.495)	2.591 (1.579, 3.602)	2.783 (1.771, 3.795)	<0.001	
AST	Crude	Reference	0.501 (-0.282, 1.284)	0.942 (0.174, 1.710)	1.256 (0.496, 2.016)	<0.001	
	Model 1	Reference	0.432 (-0.341, 1.206)	0.784 (0.024, 1.544)	0.862 (0.108, 1.616)	0.018	
	Model 2	Reference	0.217 (-0.556, 0.990)	0.504 (-0.257, 1.266)	0.432 (-0.330, 1.194)	0.209	
ALT/AST	Crude	Reference	0.064 (0.039, 0.089)	0.107 (0.082, 0.131)	0.145 (0.121, 0.169)	<0.001	
	Model 1	Reference	0.054 (0.031, 0.078)	0.091 (0.068, 0.114)	0.119 (0.096, 0.141)	<0.001	
	Model 2	Reference	0.044 (0.023, 0.066)	0.078 (0.057, 0.099)	0.097 (0.075, 0.118)	<0.001	
ALP	Crude	Reference	-1.495 (-3.242, 0.253)	1.022 (-0.692, 2.735)	-1.866 (-3.561, -0.170)	0.235	
	Model 1	Reference	-1.008 (-2.718, 0.703)	1.622 (-0.058, 3.303)	-1.448 (-3.115, 0.218)	0.455	
	Model 2	Reference	-0.952 (-2.608, 0.704)	1.990 (0.359, 3.621)	-1.841 (-3.473, -0.209)	0.259	
GGT	Crude	Reference	-1.694 (-4.260, 0.872)	0.456 (-2.060, 2.972)	1.740 (-0.749, 4.230)	0.044	
	Model 1	Reference	-1.550 (-4.083, 0.984)	0.509 (-1.980, 2.997)	0.978 (-1.490, 3.447)	0.167	
	Model 2	Reference	-2.733 (-5.219, -0.248)	-0.823 (-3.271, 1.625)	-1.525 (-3.976, 0.925)	0.590	

TABLE 6 Multiple linear regression associations of blood selenium concentration (quartile) with liver function parameters.

Crude: no covariates were adjusted. Model 1: age, gender, and race were adjusted. Model 2: age, gender, race, education, ratio of family income to poverty, diabetes, BMI, drink, smoke, physical activity, and TC were adjusted. Significant results highlighted in bold. BMI, Body mass index; TC, Total cholesterol; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; GGT, Gamma-glutamyl transferase; and ALP, Alkaline phosphatase.



While our study has revealed associations between blood selenium levels, dietary selenium intake, and liver function parameters, it is important to further investigate the different forms of selenium in the blood and their varying effects on liver function. Selenium exists in multiple forms in the blood, including inorganic selenium (such as selenite and selenate) and organic selenium (such as selenomethionine



and selenocysteine). These different forms of selenium have distinct biological activities and metabolic pathways, which may impact liver function in various ways. For instance, organic forms of selenium like selenomethionine can be non-specifically incorporated into body proteins as a long-term selenium storage, whereas inorganic selenium is mainly involved in the synthesis of selenoproteins like GPx. These different forms of selenium may differentially affect antioxidant capacity, cellular protection, and inflammatory responses, thus exhibiting varying effects on liver function parameters. Therefore, future studies should consider selenium speciation analysis to differentiate between these various forms of selenium and their respective impacts on liver function. This will provide a more comprehensive and detailed understanding of the biological roles of selenium and its mechanisms in liver function regulation, thereby offering a scientific basis for developing more precise public health strategies.

This study assessed the impact of dietary selenium intake and blood selenium concentration on liver function parameters among US adults using the latest NHANES dataset. A key strength of this study is its large sample size, coupled with a detailed characterization of the study population. Additionally, the adjustment for covariates enhances the reliability of results. Finally, we performed subgroup analyses and interaction tests to explore differences in these associations by age, gender, race, education level, and BMI.

Although this study provided a comprehensive analysis of dietary selenium intake and blood selenium concentration's effects on liver function, it has several limitations. First, blood selenium concentration, a direct biomarker of exposure, can be influenced by short-term dietary changes, individual metabolic differences, and physiological conditions, potentially not reflecting long-term intake accurately. Additionally, the relationship between dietary intake and blood concentration can be complicated by individual differences in absorption and metabolism. Second, despite adjusting for numerous potential confounders, the study cannot eradicate all confounding biases. For instance, dietary records and questionnaires depend on self-reporting, susceptible to recall or reporting bias. Furthermore, this observational study identified associations between dietary selenium intake, blood concentrations, and liver function, but it cannot establish causality. Future research should confirm these associations via prospective or interventional studies. Third, despite this study exploring the association between selenium levels and various liver function indicators (such as ALT, AST, ALP, and GGT), we did not include bilirubin in our analysis. Bilirubin is an important indicator of liver metabolic capacity and biliary system patency, and its levels are influenced by multiple factors including hemolysis, liver metabolic function, and biliary tract patency. While ALT, AST, ALP, and GGT are also affected by various factors, the metabolic pathway of bilirubin is more complex and involves additional potential confounders. Therefore, we chose not to include bilirubin in the analysis for this study. We acknowledge that this decision introduces certain limitations, making the interpretation of selenium's impact on overall liver function less comprehensive. Future research should incorporate bilirubin into the analysis to more accurately assess the multifaceted effects of selenium on liver function. Lastly, given that the NHANES database does not provide the exact dates of dietary recall interviews and blood sample collections, we cannot directly analyze the temporal relationship between these two variables. This hinders our ability to assess the time-dependent effects of dietary selenium intake and blood selenium concentrations on liver function parameters. Future studies also should consider the temporal relationship between dietary intake and biospecimen collection in their design to improve the accuracy and reliability of the findings.

5 Conclusion

This population-based, cross-sectional study demonstrated that dietary selenium intake and blood selenium concentration impact liver function parameters in a population with high selenium intakes and blood concentrations and without major liver disturbances. Public health strategies should consider promoting moderate rather than excessive selenium intake as part of liver disease prevention efforts. Further research should also focus on the relationship between dietary selenium intake and blood selenium concentration with liver function in diverse populations to develop more precise prevention strategies.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material; further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving humans were approved by the National Center for Health Statistics Research Ethics Review Board. The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and institutional requirements.

Author contributions

QL: Conceptualization, Writing - original draft. RH: Funding acquisition, Investigation, Writing - review & editing. ZP:

References

1. Trefts E, Gannon M, Wasserman DH. The liver. Curr Biol. (2017) 27:R1147-51. doi: 10.1016/j.cub.2017.09.019

2. Palmer M, Kleiner DE, Goodman Z, Brunt E, Avigan MI, Regev A, et al. Liver biopsy for assessment of suspected drug-induced liver injury in metabolic dysfunctionassociated steatohepatitis clinical trials: expert consensus from the liver forum. *Aliment Pharmacol Ther.* (2024) 59:201–16. doi: 10.1111/apt.17762

3. Himoto T, Fujita K, Mimura S, Tani J, Morishita A, Kubota S, et al. Involvement of essential trace elements in the pathogenesis of hepatitis C virus-related chronic liver disease and nonalcoholic steatohepatitis. *Exp Ther Med.* (2024) 27:19. doi: 10.3892/etm.2023.12307

4. Nakahata Y, Hanai T, Miwa T, Maeda T, Imai K, Suetsugu A, et al. Effect of selenium deficiency on the development of overt hepatic encephalopathy in patients with chronic liver disease. *J Clin Med.* (2023) 12:2829. doi: 10.3390/jcm12082869

5. Xu L, Lu Y, Wang N, Feng Y. The role and mechanisms of selenium supplementation on fatty liver-associated disorder. *Antioxidants.* (2022) 11:922. doi: 10.3390/antiox11050922

6. Tinkov AA, Korobeinikova TV, Morozova GD, Aschner M, Mak DV, Santamaria A, et al. Association between serum trace element, mineral, and amino acid levels with non-alcoholic fatty liver disease (Nafld) in adult women. *J Trace Elem Med Biol.* (2024) 83:127397. doi: 10.1016/j.jtemb.2024.127397

7. Zhang D, Wu S, Lan Y, Chen S, Wang Y, Sun Y, et al. Blood manganese and nonalcoholic fatty liver disease: a cohort-based case-control study. *Chemosphere*. (2022) 287:132316. doi: 10.1016/j.chemosphere.2021.132457 Conceptualization, Supervision, Writing – review & editing. MZ: Conceptualization, Funding acquisition, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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8. Chung SM, Moon JS, Yoon JS, Won KC, Lee HW. The sex-specific effects of blood Lead, mercury, and cadmium levels on hepatic steatosis and fibrosis: Korean Nationwide cross-sectional study. J Trace Elem Med Biol. (2020) 62:126601. doi: 10.1016/j.jtemb.2020.126601

9. Rayman MP. Food-chain selenium and human health: emphasis on intake. Br J Nutr. (2008) 100:254–68. doi: 10.1017/S0007114508939830

10. Rayman MP. Selenium and human health. Lancet. (2012) 379:1256-68. doi: 10.1016/S0140-6736(11)61452-9

11. Rose AH, Hoffmann PR. Selenoproteins and cardiovascular stress. *Thromb Haemost.* (2015) 113:494–504. doi: 10.1160/th14-07-0603

12. Shimada BK, Alfulaij N, Seale LA. The impact of selenium deficiency on cardiovascular function. Int J Mol Sci. (2021) 22:10713. doi: 10.3390/ijms221910713

13. Ye D, Sun X, Guo Y, Shao K, Qian Y, Huang H, et al. Genetically determined selenium concentrations and risk for autoimmune diseases. *Nutrition*. (2021) 91-92:111391. doi: 10.1016/j.nut.2021.111391

14. Schomburg L. Selenium deficiency due to diet, pregnancy, severe illness, or Covid-19-a preventable trigger for autoimmune disease. *Int J Mol Sci.* (2021) 22:8532. doi: 10.3390/ijms22168532

15. Wang F, Li C, Li S, Cui L, Zhao J, Liao L. Selenium and thyroid diseases. Front Endocrinol. (2023) 14:1133000. doi: 10.3389/fendo.2023.1133000

16. Liu F, Wang K, Nie J, Feng Q, Li X, Yang Y, et al. Relationship between dietary selenium intake and serum thyroid function measures in U.S. adults: data from Nhanes 2007-2012. *Front Nutr.* (2022) 9:1002489. doi: 10.3389/fnut.2022.1002489

17. Schomburg L. Selenoprotein P-selenium transport protein, enzyme and biomarker of selenium status. *Free Radic Biol Med.* (2022) 191:150–63. doi: 10.1016/j. freeradbiomed.2022.08.022

18. Shen Y, Huang H, Wang Y, Yang R, Ke X. Antioxidant effects of se-glutathione peroxidase in alcoholic liver disease. *J Trace Elem Med Biol.* (2022) 74:127048. doi: 10.1016/j.jtemb.2022.127048

19. Zhang T, Sun S, Gavrilović A, Li D, Tang R. Selenium alleviates cadmium-induced oxidative stress, endoplasmic reticulum stress, and apoptosis in L8824 cells. *Ecotoxicol Environ Saf.* (2023) 262:115337. doi: 10.1016/j.ecoenv.2023.115337

20. Abdallah AAM, Abdelrahman MM, Attia H, Hafez A, Anwar Rashed S, Amin YA, et al. (2022). Decreased serum zinc, selenium, and vitamin E as possible risk factors of hepatic fibrosis in non-alcoholic fatty liver disease. Nutr Health:2601060221103032. doi: 10.1177/02601060221103032 (Epub ahead of print).

21. Staneviciene I, Sulinskiene J, Sadauskiene I, Liekis A, Ruzgaite A, Naginiene R, et al. Effect of selenium on the Iron homeostasis and oxidative damage in brain and liver of mice. *Antioxidants*. (2022) 11:1216. doi: 10.3390/antiox11071216

22. Villanueva-Paz M, Morán L, López-Alcántara N, Freixo C, Andrade RJ, Lucena MI, et al. Oxidative stress in drug-induced liver injury (Dili): from mechanisms to biomarkers for use in clinical practice. *Antioxidants*. (2021) 10:390. doi: 10.3390/antiox10030390

23. Zhou S, Tan Q, Wen B, Bai Y, Che Q, Cao H, et al. Galacto-oligosaccharide alleviates alcohol-induced liver injury by inhibiting oxidative stress and inflammation. *Meta*. (2022) 12:867. doi: 10.3390/metabo12090867

24. Ramírez-Acosta S, Uhlírová R, Navarro F, Gómez-Ariza JL, García-Barrera T. Antagonistic interaction of selenium and cadmium in human hepatic cells through Selenoproteins. *Front Chem.* (2022) 10:891933. doi: 10.3389/fchem.2022.891933

25. Tang C, Li S, Zhang K, Li J, Han Y, Zhan T, et al. Selenium deficiency-induced redox imbalance leads to metabolic reprogramming and inflammation in the liver. *Redox Biol.* (2020) 36:101519. doi: 10.1016/j.redox.2020.101519

26. Nobes J, Leith D, Handjiev S, Dillon JF, Dow E. Intelligent liver function testing (11ft): an intelligent laboratory approach to identifying chronic liver disease. *Diagnostics*. (2024) 14:960. doi: 10.3390/diagnostics14090960

27. Rajesh R, Sangameshwar A, Tan YY, Teh Kim Jun K, Tham TY, Cheah Chang Chuen M. Approach to abnormal liver biochemistries in the primary care setting. *Cureus*. (2024) 16:e56541. doi: 10.7759/cureus.56541

 Laclaustra M, Stranges S, Navas-Acien A, Ordovas JM, Guallar E. Serum selenium and serum lipids in us adults: National Health and nutrition examination survey (Nhanes) 2003-2004. Atherosclerosis. (2010) 210:643–8. doi: 10.1016/j. atherosclerosis.2010.01.005

29. Attanasio R. Sex differences in the association between Perfluoroalkyl acids and liver function in us adolescents: analyses of Nhanes 2013-2016. *Environ Pollut.* (2019) 254:113061. doi: 10.1016/j.envpol.2019.113061

30. Malin AJ, Lesseur C, Busgang SA, Curtin P, Wright RO, Sanders AP. Fluoride exposure and kidney and liver function among adolescents in the United States: Nhanes, 2013-2016. *Environ Int.* (2019) 132:105012. doi: 10.1016/j.envint.2019.105012

31. Xiang S, Dong J, Li X, Li C. Urine phthalate levels and liver function in us adolescents: analyses of Nhanes 2007-2016. *Front Public Health*. (2022) 10:843971. doi: 10.3389/fpubh.2022.843971

32. Lippman SM, Klein EA, Goodman PJ, Lucia MS, Thompson IM, Ford LG, et al. Effect of selenium and vitamin E on risk of prostate Cancer and other cancers: the selenium and vitamin E Cancer prevention trial (select). *JAMA*. (2009) 301:39–51. doi: 10.1001/jama.2008.864

33. Kristal AR, Darke AK, Morris JS, Tangen CM, Goodman PJ, Thompson IM, et al. Baseline selenium status and effects of selenium and vitamin E supplementation on prostate Cancer risk. *J Natl Cancer Inst.* (2014) 106:djt 456. doi: 10.1093/jnci/djt456

34. Smith MI, Westfall BB. Further field studies on the selenium problem in relation to public health. *Public Health Rep.* (1937) 1937:1375–84. doi: 10.2307/4582321

35. Li W, Li X, Su J, Chen H, Zhao P, Qian H, et al. Associations of blood metals with liver function: analysis of Nhanes from 2011 to 2018. *Chemosphere*. (2023) 317:137854. doi: 10.1016/j.chemosphere.2023.137854

36. Yang Z, Yan C, Liu G, Niu Y, Zhang W, Lu S, et al. Plasma selenium levels and nonalcoholic fatty liver disease in Chinese adults: a cross-sectional analysis. *Sci Rep.* (2016) 6:37288. doi: 10.1038/srep37288

37. Longnecker MP, Taylor PR, Levander OA, Howe M, Veillon C, McAdam PA, et al. Selenium in diet, blood, and toenails in relation to human health in a Seleniferous area. *Am J Clin Nutr.* (1991) 53:1288–94. doi: 10.1093/ajcn/53.5.1288

38. Avery JC, Hoffmann PR. Selenium, Selenoproteins, and immunity. *Nutrients*. (2018) 10:1203. doi: 10.3390/nu10091203

39. Díaz-Castro J, Ojeda ML, Alférez MJ, López-Aliaga I, Nestares T, Campos MS. Se bioavailability and glutathione peroxidase activity in Iron deficient rats. *J Trace Elem Med Biol.* (2011) 25:42–6. doi: 10.1016/j.jtemb.2010.12.005

40. Weaver K, Skouta R. The Selenoprotein glutathione peroxidase 4: from molecular mechanisms to novel therapeutic opportunities. *Biomedicine*. (2022) 10:891. doi: 10.3390/biomedicines10040891

41. Combs GF Jr. Biomarkers of selenium status. Nutrients. (2015) 7:2209-36. doi: 10.3390/nu7042209

42. Fairweather-Tait SJ, Bao Y, Broadley MR, Collings R, Ford D, Hesketh JE, et al. Selenium in human health and disease. *Antioxid Redox Signal.* (2011) 14:1337–83. doi: 10.1089/ars.2010.3275

43. Shang N, Wang X, Shu Q, Wang H, Zhao L. The functions of selenium and Selenoproteins relating to the liver diseases. *J Nanosci Nanotechnol.* (2019) 19:1875–88. doi: 10.1166/jnn.2019.16287

44. Lu J, Holmgren A. The Thioredoxin antioxidant system. Free Radic Biol Med. (2014) 66:75–87. doi: 10.1016/j.freeradbiomed.2013.07.036

45. Trovato A, Siracusa R, Di Paola R, Scuto M, Fronte V, Koverech G, et al. Redox modulation of cellular stress response and Lipoxin A4 expression by Coriolus versicolor in rat brain: relevance to Alzheimer's disease pathogenesis. *Neurotoxicology*. (2016) 53:350–8. doi: 10.1016/j.neuro.2015.09.012

46. Trovato A, Siracusa R, Di Paola R, Scuto M, Ontario ML, Bua O, et al. Redox modulation of cellular stress response and Lipoxin A4 expression by Hericium Erinaceus in rat brain: relevance to Alzheimer's disease pathogenesis. *Immun Ageing*. (2016) 13:23. doi: 10.1186/s12979-016-0078-8

47. Hill KE, McCollum GW, Boeglin ME, Burk RF. Thioredoxin reductase activity is decreased by selenium deficiency. *Biochem Biophys Res Commun.* (1997) 234:293–5. doi: 10.1006/bbrc.1997.6618

48. Sunde RA. Selenium regulation of Selenoprotein enzyme activity and transcripts in a pilot study with founder strains from the collaborative cross. *PloS One.* (2018) 13:e0191449. doi: 10.1371/journal.pone.0191449

49. Li J, Cheng P, Li S, Zhao P, Han B, Ren X, et al. Selenium status in diet affects acetaminophen-induced hepatotoxicity via interruption of redox environment. *Antioxid Redox Signal.* (2021) 34:1355–67. doi: 10.1089/ars.2019.7909

50. Brozmanová J, Mániková D, Vlčková V, Chovanec M. Selenium: a double-edged sword for defense and offence in Cancer. *Arch Toxicol.* (2010) 84:919–38. doi: 10.1007/s00204-010-0595-8

51. Lu J, Berndt C, Holmgren A. Metabolism of selenium compounds catalyzed by the mammalian Selenoprotein Thioredoxin reductase. *Biochim Biophys Acta.* (2009) 1790:1513–9. doi: 10.1016/j.bbagen.2009.04.013

52. McAdam PA, Smith DK, Feldman EB, Hames C. Effect of age, sex, and race on selenium status of healthy residents of Augusta. *Georg Biol Trace Element Res.* (1984) 6:3–9. doi: 10.1007/BF02918316

53. Combs GF Jr, Watts JC, Jackson MI, Johnson LK, Zeng H, Scheett AJ, et al. Determinants of selenium status in healthy adults. *Nutr J.* (2011) 10:75. doi: 10.1186/1475-2891-10-75