



OPEN ACCESS

EDITED BY

Mora Murri,
University of Malaga, Spain

REVIEWED BY

Siti Wulan,
University of Brawijaya, Indonesia
Amani Alrasheedi,
King Abdulaziz University, Saudi Arabia
Mingjiang Liu,
University of South China, China

*CORRESPONDENCE

Deguang Wang
✉ wangdeguang@ahmu.edu.cn
Bao Zhang
✉ 5962555382@qq.com

RECEIVED 28 March 2024

ACCEPTED 03 June 2024

PUBLISHED 18 June 2024

CITATION

Zu S, Yang ML, Li XD, Wu HH, Li XL, Fan YS,
Wang DG and Zhang B (2024) Flavonoids
intake and weight-adjusted waist index:
insights from a cross-sectional study of
NHANES.

Front. Nutr. 11:1400726.

doi: 10.3389/fnut.2024.1400726

COPYRIGHT

© 2024 Zu, Yang, Li, Wu, Li, Fan, Wang and
Zhang. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Flavonoids intake and weight-adjusted waist index: insights from a cross-sectional study of NHANES

Shuang Zu¹, Meiling Yang², Xiude Li¹, Hanhan Wu¹, Xunliang Li³, Yunshan Fan¹, Deguang Wang^{3*} and Bao Zhang^{1*}

¹Department of Clinical Nutrition, The First Affiliated Hospital of Anhui Medical University, Hefei, Anhui, China, ²Department of Gastroenterology, The First Affiliated Hospital of Anhui Medical University, Hefei, Anhui, China, ³Department of Nephrology, The Second Affiliated Hospital of Anhui Medical University, Hefei, Anhui, China

This study conducted data on 15,446 adults to explore the impact of flavonoids on weight-adjusted waist index (WWI). This was a nationwide cross-sectional study among US adults aged 20 years or older. Dietary intake of flavonoids was assessed through 24-h recall questionnaire. WWI was calculated by dividing waist circumference (WC) by the square root of weight. We utilized weighted generalized linear regression to evaluate the association between flavonoids intake and WWI, and restricted cubic splines (RCS) to explore potential non-linear relationships. Our findings indicated that individuals with lower WWI experienced a notable increase in their consumption of total flavonoids, flavanones, flavones, flavan-3-ols, and anthocyanidins intake (β (95% CI); $-0.05(-0.09, -0.01)$; $-0.07(-0.13, 0.00)$; $-0.07(-0.11, -0.02)$; $-0.06(-0.11, 0.00)$; $-0.13(-0.18, -0.08)$, respectively), with the exception of flavonols and isoflavones. Additionally, consumption of total flavonoids, flavonols, flavanones, isoflavones, and flavan-3-ols had a non-linear relationship with WWI (all P for non-linearity < 0.05). Furthermore, the effect of total flavonoids on WWI varied in race (P for interaction = 0.011), gender (P for interaction = 0.038), and poverty status (P for interaction = 0.002). These findings suggested that increase the intake of flavonoids might prevent abdominal obesity, but further prospective studies are requested before dietary recommendation.

KEYWORDS

flavonoids intake, weight-adjusted waist index, NHANES, abdominal obesity, flavonoids dietary recommendation

1 Introduction

Obesity arises from the disproportionate accumulation of body fat, a consequence of energy intake surpassing energy expenditure. The prevalence of obesity has reached epidemic levels globally, impacting a growing population of people (1, 2). Abdominal obesity, a form of obesity characterized by an accumulation of fat specifically in the abdominal region, is associated with an increased risk of metabolic imbalances, diabetes, high blood pressure, and cardiovascular diseases, beyond overall amount of body fat (3). These elements are poised to create a public health concern, necessitating intervention strategies once the contributing factors have been assessed. Research indicated that people with a high volume

of abdominal fat and a large waist circumference (WC) not only have more intra-abdominal fat but also larger subcutaneous fat cells, accompanied by adipose tissue that is dysfunctional and inflamed (4). Numerous indicators have been utilized to assess obesity, with a particular focus on the detrimental intra-abdominal fat mass. While Body Mass Index (BMI) remains the most prevalent anthropometric measurement for obesity, it has limitations as it does not differentiate between lean and fat mass. WC has been introduced as an alternative for indirectly assessing the increase in visceral fat, serving as a marker for abdominal obesity and aiding in the diagnosis of metabolic syndrome (5). However, similar to BMI, WC on its own is incapable of differentiating between visceral and subcutaneous fat masses. In fact, the accuracy of WC in estimating abdominal fat mass is enhanced when BMI is included as an explanatory variable (6). Weight-adjusted waist index (WWI), which Park et al. (7), is an innovative measure of abdominal obesity that normalizes WC by body weight, leveraging the advantages of WC while reducing its correlation with BMI. The WWI serves not only to differentiate between adipose and muscular tissues but also to address issues of central obesity that are not directly related to overall body weight (8). As reported by Kim et al. (9), there was a positive correlation between both BMI and WC with measurements of fat and muscle. In contrast, the WWI showed a positive relationship with overall and abdominal fat measurements, while it exhibited a negative correlation with the mass of appendicular skeletal muscle (9). In a 40-month longitudinal study involving 1,946 participants, computed tomography (CT) data revealed a rise in measurements of abdominal fat along with a corresponding decline in muscle mass, concurrent with an escalation in the WWI (10). In addition, previous studies had identified a notable association between elevated WWI values and various health issues, including non-alcoholic fatty liver disease (11), diabetic kidney disease (12), bone mineral density (13), and depression (14).

Flavonoid, derived from the Latin term 'flavus' which signifies the color yellow, is omnipresent in the plant kingdom and represent the most prevalent polyphenolic substances in the human diet (15). A vast array of naturally occurring flavonoids, numbering over 5,000, have been identified in a variety of plant species. These compounds feature a 15-carbon (C6–C3–C6) backbone structure, with two benzene rings linked by a straight 3-carbon chain. The classification of flavonoid into multiple subcategories is based on the substitution patterns of the C ring, with further differentiation within each class achieved through the A and B substitutions (16). The main subgroups of flavonoid include flavanols, flavanones, isoflavones, flavones, flavans-3-ols, and anthocyanins. Over the past two decades, flavonoid compounds have been extensively studied for their potential positive effects on human health. Dietary flavonoid intake has been shown to be significantly and negatively associated with hepatic steatosis and fibrosis (17), metabolic syndrome (18), and obesity (19). The anti-obesity effects of flavonoid is believed to operate through a variety of mechanisms, such as inhibiting enzymes, modulating neuro-hormonal responses related to food consumption and satiety, and promoting the generation of mitochondria (20). So, can flavonoids compounds naturally prevent or reduce abdominal obesity? Currently, there is a lack of research regarding the potential of flavonoids consumption in reducing the risk of developing abdominal obesity. Hence, the importance of this study lies in investigating the effects of dietary flavonoids intake on WWI using data from the National Health and Nutrition Examination

Survey (NHANES) database, and uncovering the potential of flavonoids compounds in reducing the risk of abdominal obesity.

2 Methods

2.1 Study population

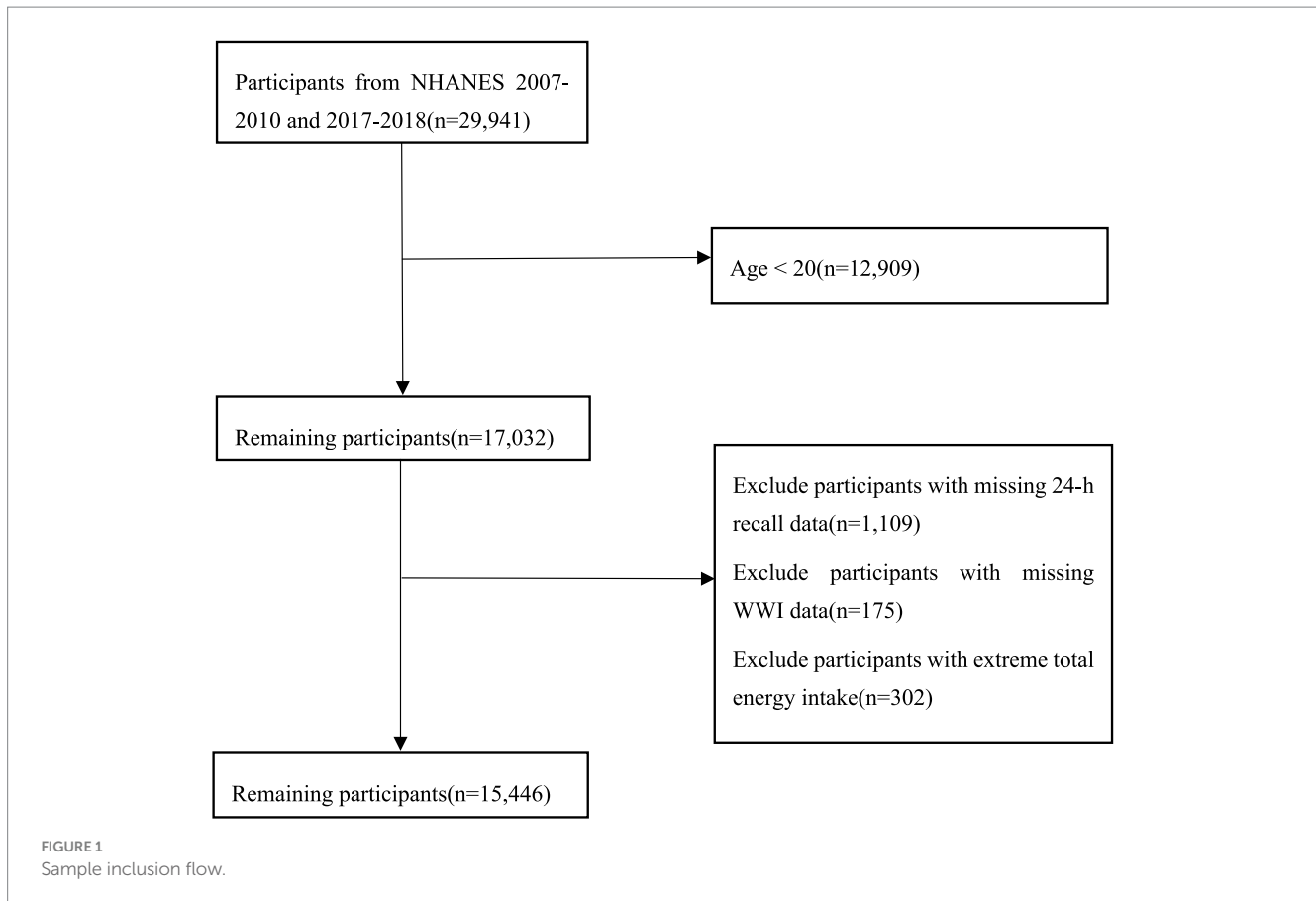
NHANES, overseen by the National Center for Health Statistics (NCHS), is a comprehensive, nationwide survey aimed at assessing the health of the American population. The NCHS Ethics Review Board has endorsed the stratified multistage sampling approach employed in the survey. NHANES data collection involved an initial in-home interview, followed by a health examination at mobile examination center (MEC), and concludes with a follow-up telephone interview. For this study, we utilized data from the survey periods of 2007–2010 and 2017–2018. All participants were required to provide written consent after receiving comprehensive information about the study. Our study included 17,032 participants above the age of 20 and collected information on their flavonoids intake and WWI values. After careful consideration, participants with missing 24-h recall data ($n=1,109$) were removed from the study. Participants who had missing WC and weight data ($n=175$) were omitted, too. Furthermore, a total of 302 participants were excluded as they had either extremely low or high energy intakes: < 500 kcal or $> 5,000$ kcal/day for females and < 500 kcal or $> 8,000$ kcal/day for males. Finally, 15,446 participants were selected for inclusion (Figure 1).

2.2 Dietary assessment of flavonoids

The United States Department of Agriculture (USDA) Food Codes' Flavonoid Database for the years 2007–2010 and 2017–2018 offers detailed information on the flavonoid content of a wide array of foods and beverages, as listed in the USDA and Food and Nutrient Database for Dietary Studies (FNDDS) version 5.0. The database contains detailed information on 29 specific flavonoids, which are divided into six categories: Anthocyanidins, Flavan-3-ols, Flavanones, Flavones, Flavonols, and Isoflavones, aligning with the dietary data provided by NHANES for the years 2007–2010 and 2017–2018. To determine the average daily flavonoids intake during these time-frames, a two-day dietary recall approach was utilized, involving in-person interviews with participants to document their food consumption over 2 days, supplemented by a subsequent telephone interview approximately 1 week later. In this study, flavonoids intake was determined solely from dietary sources, including water, without consideration for the consumption of flavonoid supplements or pharmaceuticals.

2.3 WWI

WWI ($\text{cm}/\sqrt{\text{kg}}$) is calculated by dividing WC (cm) by the square root of the individual's weight (kg). These body measurements were conducted at MEC, with a health technologist responsible for recording and documenting the data. The current anthropometric standards were used to measure weight and WC (21). The participants were asked to wear specific attire for the assessment and then position



themselves in the center of a digital scale, barefooted, with arms resting by their sides and their gaze directed straight ahead. WC was determined using a measuring tape, with MEC health technicians marking a horizontal line above the superior lateral edge of the right ilium. A right midaxillary line was then drawn, and the tape was positioned horizontally at the intersection of these two lines. Technicians, standing on the participant's right side, wrapped the tape around the torso in a horizontal plane.

2.4 Covariates

The survey's covariates encompassed demographic factors such as age, gender, ethnicity, educational attainment, and the family poverty income ratio (PIR), alongside lifestyle factors including alcohol and tobacco consumption, physical activity levels, and dietary intake of calories, carbohydrates, proteins, fats, and fiber. We categorized education levels as follows: below high school (below 11th grade), high school (graduated from high school or obtained a GED equivalent), and beyond high school (completed college or achieved a higher level of education). Poverty status was categorized into two groups based on income level: PIR <130% referred to low income, while PIR ≥130% designated high income (22). Smoking status was categorized into three groups: never smoked (consuming fewer than 100 cigarettes throughout their life), former smokers (having smoked over 100 cigarettes but are currently abstaining), and current smokers (who have smoked over 100 cigarettes and continue to smoke either occasionally or regularly). Alcohol consumption status was categorized

into three groups: lifetime abstainer (who have consumed alcohol less than 12 times in their lifetime), former drinker (who have consumed alcohol more than 12 times in their lifetime but have not consumed it in the past year) and current drinker (who currently consume alcohol more than three times a week). Physical activity was categorized into two levels: low, defined as less than 599 metabolic equivalents (METs) per week, and high, defined as 599 METs or more per week (23).

2.5 Statistical analysis

Taking into account the complex sampling strategy of NHANES, we applied appropriate weights to the data in accordance with NHANES guidelines, which involved using a third of the weights from both the 2007–2010 and 2017–2018 periods. In this study, 15,446 individuals were adjusted for accuracy to represent a population of 218,690,615. Continuous data were presented as the mean with the standard deviation, whereas categorical data were represented as unweighted numbers (weighted percentages). The participant characteristics were analyzed according to tertile of total flavonoids intake, using weighted student's t-tests for continuous variables and weighted chi-squared tests for categorical variables. The levels of flavonoids consumption were divided into three groups (T1, T2 and T3) and employed weighted generalized linear regression models with the low-intake group (T1) as the reference group. The crude model did not take into account any of the variables, whereas model 1 adjusted for potential confounding factors such as age, gender, ethnicity, education levels, and PIR. After considering physical activity levels,

total carbohydrate intake, total protein intake, total fat intake, and total fiber intake, we made additional adjustments in model 2. We selected the above covariates based on professional knowledge, previously identified risk factors for WWI (7, 9, 10), and the observed incomparability of participant characteristics (Table 1). We decided to exclude the other covariates for the reasons that preliminary analyses indicated potential collinearity between them. Furthermore, to maintain model parsimony and avoid overfitting, we aimed to include only above covariates that had the most substantial and theoretically justified impact on the relationship between flavonoid intake and WWI. Restricted cubic splines (RCS) were utilized in Model 2 to explore the possible non-linear relationships between flavonoids intake and WWI. Furthermore, in order to examine the influence of total flavonoids intake on WWI, subgroup analyses and interaction exploration were performed to assess the effect of covariates that exhibited notable variations across different levels of total flavonoids intake. All subgroup variable analyses performed using a fully adjusted Model 2, and generalized linear regression models were additionally applied to assess the interactive effects between these subgroups and WWI. The missing covariate values were imputed by the “mice” package in R. The nationally representative estimates in NHANES were calculated by utilizing “SDMVPSU” and “SDMVSTRA.” The data processing was done in R version 4.3.1. A statistically significant difference was determined by considering a two-sided p -value <0.05 .

3 Results

3.1 Characteristics of participants

The study involved 15,446 participants with an average age of 47.35 ± 16.96 years, of whom 7,533 (47.83%) were male and 7,913 (52.17%) were female. The average WWI across all participants was 10.91 ± 0.90 cm/ $\sqrt{\text{kg}}$. The mean total flavonoids intake for all participants was 227.90 ± 410.82 mg/100 g foods/day, with the values for the different tertiles as follows: T1: 0–36.08 mg; T2: 36.08–160.52 mg; T3: 160.52–8,018.67 mg. Participants in the highest tertile of total flavonoids intake were older, Non-Hispanic White, had a higher level of education, higher PIR, smoked less, and consumed alcohol more frequently. Additionally, higher intakes of total flavonoids were more likely to with higher physical activity, total caloric intake, carbohydrate, protein, fat consumption, as well as fiber intake (Table 1).

3.2 Associations between dietary flavonoids intake and WWI

Table 2 displayed the outcomes of weighted linear regression analyses that evaluate the relationships between total flavonoids and flavonoid subclass intake with WWI. In model 2, A statistically significant negative correlations were observed between total flavonoids, flavanones, flavones, flavan-3-ols, and anthocyanidins ($p=0.025$, $p=0.041$, $p=0.006$, $p=0.0035$, $p<0.001$, respectively) with WWI after adjusting demographic variables and lifestyle variables except flavonols and isoflavones. Notably, when using the first tertile as the reference, the third tertile of total flavonoids ($\beta: -0.05$, 95%CI: $-0.09, -0.01$), flavanones ($\beta: -0.07$, 95%CI: $-0.13, 0.00$), flavones ($\beta: -0.07$, 95%CI: $-0.11, -0.02$), anthocyanidins ($\beta: -0.13$, 95%CI: $-0.18, -0.08$), and the second tertile of

flavan-3-ols ($\beta: -0.06$, 95%CI: $-0.11, 0.00$) had demonstrated the most significant potential for reducing WWI. In all three models, there was a significant negative correlation seen between T3 of total flavonoids (all $p<0.05$), T2 and T3 of flavones ($p<0.01$), T2 of flavan-3-ols ($p<0.05$), T3 of anthocyanidins ($p<0.01$) and WWI. Forest plot was used to show the relationship between the third tertile of flavonoids and linear trend test was conducted by treating each flavonoid as a continuous variable. In model 2, total flavonoids, flavanones, flavones, and anthocyanidins indicated a linear trend with WWI (P -trend=0.033, P -trend=0.014, P -trend=0.047, P -trend=0.002, respectively) (Figure 2).

3.3 The non-linear relationships between flavonoids intake and WWI

We utilized an RCS to examine the non-linear relationships between the intake of flavonoids and WWI, with the median consumption of all flavonoids serving as the baseline for comparison. In Model 2, it was determined that total flavonoids (P for non-linearity <0.001), flavonols (P for non-linearity=0.031), flavanones (P for non-linearity <0.001), isoflavones (P for non-linearity=0.002), and flavan-3-ols (P for non-linearity=0.001) exhibited a statistically significant non-linear association with WWI. Association between total flavonoids, flavonols, flavanones, flavan-3-ols and WWI was J-shaped curve. The WWI showed a consistent decline with the increment in total flavonoids intake, up to a point where the intake reached 108.86 mg/100 g foods/day and the consumption range of total flavonoids in 65.43–217.45 mg/100 g foods/day have the ability to decrease WWI. Beyond this range, WWI started to rise with further increases in total flavonoids consumption. Within the dietary intake range of <8.39 mg/100 g foods/day and 12.12–57.81 mg/100 g foods/day, flavonols exhibited a capacity to decrease WWI and the optimal reduction occurring at a level of 30.77 mg/100 g foods/day. At 2.80 mg/100 g foods/day (flavanones) and 41.85 mg/100 g foods/day (flavan-3-ols) points, WWI values were the lowest, respectively. Additionally, beyond the range of flavanones (1.40–40.63 mg/100 g foods/day) and flavan-3-ols (20.92–104.64 mg/100 g foods/day), WWI started to increase. On the contrary, when isoflavones intake increased, WWI values consistently dropped, and the downward trend then slightly increased (Figure 3).

3.4 Subgroup and interaction analysis

Figure 4 presented the subgroup and interaction analysis between WWI and total flavonoids intake, revealing significant interactions with respect to races, gender, and poverty status (P for interaction = 0.011, P for interaction = 0.038, P for interaction = 0.002, respectively). When compared to Non-Hispanic White, the intake of total dietary flavonoids appeared to have a more pronounced protective effect against obesity in other populations ($p<0.001$). Notably, a higher intake of total flavonoids was linked to a lower WWI among males ($p=0.001$) and individuals in poverty ($p<0.001$).

4 Discussion

Based on a representative sample of 15,446 US adults, the present analyses showed that the average consumption of total flavonoids was

TABLE 1 Basic characteristics of total participants stratified by total flavonoids tertile.

Characteristics	Total (n = 15,446)	Category of Flavonoid Intake, mg/100 g foods/day			p-value
		T1 (n = 5,380)	T2 (n = 5,384)	T3 (n = 4,682)	
Age (year)	47.35 (16.96)	45.02 (17.05)	48.14 (17.22)	48.88 (16,34)	<0.001
Gender, n (%)					0.3
Male	7,533 (47.83%)	2,623 (48.59%)	2,654 (48.31%)	2,257 (46.61%)	
Female	7,913 (52.17%)	2,757 (51.41%)	2,730 (51.69%)	2,425 (53.39%)	
Ethnicity, n (%)					<0.001
Mexican-American	2,551 (8.61%)	979 (10.18%)	1,035 (10.30%)	537 (5.34%)	
Non-Hispanic White	6,847 (67.03%)	2,309 (65.75%)	2,203 (64.14%)	2,334 (71.19%)	
Non-Hispanic Black	3,149 (11.22%)	1,222 (12.99%)	1,050 (11.30%)	877 (9.38%)	
Others	2,901 (13.14%)	870 (11.08%)	1,096 (14.26%)	934 (14.09%)	
Poverty status, n (%)					<0.001
No	9,658 (72.42%)	3,092 (68.28%)	3,383 (73.37%)	3,181 (75.61%)	
Yes	5,790 (27.58%)	2,288 (31.72%)	2,001 (26.63%)	1,501 (24.39%)	
Education levels, n (%)					<0.001
More than high school	7,726 (58.49%)	2,184 (48.93%)	2,831 (62.54%)	2,709 (63.98%)	
High school	3,684 (25.43%)	1,458 (30.70%)	1,184 (21.80%)	1,042 (23.81%)	
Less than high school	4,038 (16.08%)	1,738 (20.37%)	1,369 (15.66%)	931 (12.21%)	
Smoking status, n (%)					<0.001
Never	8,436 (55.21%)	2,597 (48.34%)	3,128 (59.04%)	2,709 (58.24%)	
Former	3,829 (24.65%)	1,311 (23.72%)	1,360 (25.35%)	1,158 (24.89%)	
Current	3,180 (20.14%)	1,472 (27.94%)	895 (15.61%)	813 (16.87%)	
Drinking status, n (%)					0.006
Never	3,241 (16.23%)	1,087 (15.29%)	1,098 (16.29%)	1,055 (17.12%)	
Former	3,062 (16.84%)	1,190 (19.20%)	1,009 (15.30%)	862 (16.01%)	
Current	9,146 (66.93%)	3,103 (65.51%)	3,277 (68.41%)	2,765 (66.87%)	
Physical activity, n (%)					0.001
High	9,137 (65.02%)	2,963(62.51%)	3,275 (65.66%)	2,898 (66.88%)	
Low	6,311 (34.98%)	2,417 (37.49%)	2,109 (34.34%)	1,786 (33.12%)	
BMI, n (%)					<0.001
BMI < 25	4,234 (29.18%)	1,368 (26.33%)	1,497 (29.48%)	1,369 (31.75%)	

(Continued)

TABLE 1 (Continued)

Characteristics	Total (n = 15,446)	Category of Flavonoid Intake, mg/100 g foods/day			p-value
		T1 (n = 5,380)	T2 (n = 5,384)	T3 (n = 4,682)	
BMI ≥ 25	1,1214 (70.82%)	4,012 (73.67%)	3,887 (70.52%)	3,313 (68.25%)	
Weight (kg)	82.86 (21.83)	84.28 (22.44)	82.46 (21.62)	81.83 (21.34)	0.003
WC (cm)	99.13 (16.67)	100.31 (16.93)	98.71 (16.58)	98.37 (16.44)	<0.001
WWI (cm/√kg)	10.91 (0.90)	10.95 (0.90)	10.90 (0.89)	10.89 (0.90)	0.034
Total calories (kcal/d)	2,095.41 (837.56)	1,930.17 (785.30)	2,183.91 (870.05)	2,172.47(830.56)	<0.001
Total carbohydrate (g)	249.87 (108.69)	225.19 (102.75)	259.14 (106.27)	265.31 (112.51)	<0.001
Total protein (g)	81.83 (35.18)	75.82 (33.20)	85.25 (36.73)	84.43 (34.75)	<0.001
Total fat (g)	81.26 (39.58)	77.42 (38.63)	83.14 (41.43)	83.24 (38.32)	<0.001
Total fiber (g)	16.76 (9.03)	12.96 (6.46)	18.61 (8.76)	18.71 (10.21)	<0.001
Total flavonoids (mg)	227.90 (410.82)	17.50 (9.47)	78.79 (33.90)	587.41 (556.19)	<0.001
Flavonols (mg)	18.96 (18.73)	7.83 (5.30)	15.38 (10.11)	33.67 (23.86)	<0.001
Flavanones (mg)	12.60 (26.63)	1.35 (3.80)	18.94 (24.76)	17.53 (36.19)	<0.001
Isoflavones (mg)	2.14 (11.40)	0.54 (2.08)	2.65 (10.20)	3.24 (16.67)	<0.001
Flavones (mg)	0.95 (2.10)	0.49 (0.74)	1.00 (1.44)	1.35 (3.19)	<0.001
Flavan-3-ols (mg)	178.95 (391.91)	5.37 (5.21)	25.32 (27.68)	506.17 (547.01)	<0.001
Anthocyanidins (mg)	14.30 (37.50)	1.92 (3.74)	15.51 (20.84)	25.46 (59.10)	<0.001

T, Tertile; BMI, body mass index; WC, weight circumference; WWI, Weight-adjusted waist index; continuous data were displayed as weighted means (standard deviation) and categorical variables were exhibited as unweighted numbers (weighted percentages).

TABLE 2 Associations between flavonoids intake and WWI.

	Crude Model β (95% CI)	<i>p</i> -value	Model 1 β (95% CI)	<i>p</i> -value	Model 2 β (95% CI)	<i>p</i> -value
Total flavonoids(mg)						
T1(0–36.08 mg)	Reference	–	Reference	–	Reference	–
T2 (> 36.08, \leq 160.52)	0.05 (–0.10, 0.00)	0.064	–0.10 (–0.14, –0.06)	<0.001	–0.04 (–0.08, 0.00)	0.040
T3 (> 160.52, \leq 8,018.67)	–0.06 (–0.10, –0.01)	0.020	–0.11 (–0.15, –0.07)	<0.001	–0.05 (–0.09, –0.01)	0.025
Flavonols (mg)						
T1 (0–9.31)	Reference	–	Reference	–	Reference	–
T2 (> 9.31, \leq 20.01)	–0.03 (–0.08, 0.02)	0.300	–0.03 (–0.07, 0.02)	0.200	0.01 (–0.03, 0.06)	0.500
T3 (> 20.01, \leq 332.04)	–0.11 (–0.16, –0.05)	<0.001	–0.09 (–0.13, –0.04)	<0.001	–0.02 (–0.06, 0.03)	0.400
Flavanones (mg)						
T1 (0–0.14)	Reference	–	Reference	–	Reference	–
T2 (> 0.14, \leq 4.17)	–0.07 (–0.14, 0.00)	0.053	–0.09 (–0.15, –0.02)	0.009	–0.06 (–0.12, 0.00)	0.054
T3 (> 4.17, \leq 590.62)	–0.07 (–0.14, 0.00)	0.067	–0.12 (–0.18, –0.06)	<0.001	–0.07 (–0.13, 0.00)	0.041
Isoflavones (mg)						
T1 (= 0)	Reference	–	Reference	–	Reference	–
T2 (> 0, \leq 0.01)	0.01 (–0.06, 0.07)	0.031	–0.03 (–0.08, 0.02)	0.200	–0.02 (–0.07, 0.03)	0.700
T3 (> 0.01, \leq 390.60)	–0.07 (–0.12, –0.03)	0.008	–0.06 (–0.10, –0.02)	0.006	–0.01 (–0.06, 0.03)	0.900
Flavones (mg)						
T1 (0–0.28)	Reference	–	Reference	–	Reference	–
T2 (> 0.28, \leq 0.90)	–0.09 (–0.13, –0.04)	<0.001	–0.09 (–0.13, –0.06)	<0.001	–0.06 (–0.10, –0.03)	0.002
T3 (> 0.90, \leq 87.25)	–0.12 (–0.17, –0.07)	<0.001	–0.14 (–0.18, –0.09)	<0.001	–0.07 (–0.11, –0.02)	0.006
Flavan-3-ols (mg)						
T1 (0–8.19)	Reference	–	Reference	–	Reference	–
T2 (> 8.19, \leq 66.11)	–0.08 (–0.14, –0.01)	0.018	–0.11 (–0.16, –0.05)	<0.001	–0.06 (–0.11, 0.00)	0.035
T3 (> 66.11, \leq 7,727.37)	–0.01 (–0.06, 0.05)	0.700	–0.06 (–0.11, –0.01)	0.015	–0.02 (–0.07, 0.03)	0.400
Anthocyanidins (mg)						
T1 (0–0.405)	Reference	–	Reference	–	Reference	–
T2 (> 0.405, \leq 6.92)	–0.01 (–0.06, 0.05)	0.700	–0.08 (–0.13, –0.04)	<0.001	–0.06 (–0.11, –0.02)	0.011
T3 (> 6.92, \leq 1,497.9)	–0.09 (–0.15, –0.03)	0.005	–0.18 (–0.23, –0.14)	<0.001	–0.13 (–0.18, –0.08)	<0.001

T, Tertile; WWI, Weight-adjusted waist index; PIR, family income to poverty. Weighted generalized linear regression models for the association between flavonoids intake and WWI. Model 1: no covariates were adjusted. Model 2: adjusted for age, gender, ethnicity, education levels and PIR. Model 3: adjusted for age, gender, ethnicity, education levels, PIR, physical activity levels, total carbohydrate intake, total protein intake, total fat intake and total fiber intake.

227.90 mg/100 g foods/day mostly from flavan-3-ols (178.95 mg/100 g foods/day) and flavonols (18.96 mg/100 g foods/day). Our findings revealed that WWI decreased with increasing levels of dietary intake of total flavonoids, flavanones, flavones, flavan-3-ols, and anthocyanidins after adjusting demographic variables and lifestyle variables. Additionally, the RCS curves demonstrated that the intake of total flavonoids, flavonols, flavanones, and flavan-3-ols within a certain range can exert the greatest effect in reducing WWI levels. Subgroup and interaction analysis further revealed that other ethnic groups (Mexican American and Others), male, and participants living in poverty had a stronger negative correlation between the total amount of flavonoids and WWI.

Flavonoids, a class of polyphenolic compounds unique to the plant kingdom, have recently sparked significant interest due to their potential health benefits for humans. Notable sources of these dietary components encompass a variety of plant-based foods, such as tea,

berries, citrus fruits, and soy products (24). In the present study, tea emerged as the predominant source for the total flavonoids, flavonols, and flavan-3-ols categories. Meanwhile, a diverse array of foods significantly contributed to the intake of other flavonoid classes, with orange juice being a key source of flavanones, protein powders for isoflavones, sweet peppers for flavones and blueberries for anthocyanins (25). Kim et al. (26) examined the relationship between the regular consumption of various flavonoids categories and obesity in 23,118 adults in Korea found that a higher total intake of flavonoid was associated with a lower prevalence of obesity in women (OR = 0.82; 95%CI: 0.71–0.94) and the consumption of flavonols (OR = 0.88; 95%CI: 0.78–0.99), isoflavones (OR = 0.85; 95%CI: 0.75–0.96) and proanthocyanidins (OR = 0.81; 95%CI: 0.71–0.92) was found to be inversely associated with the risk of abdominal obesity. However, the intake of flavonols (OR = 1.16; 95%CI: 1.02–1.33), flavanones (OR = 1.18; 95%CI: 1.04–1.35), and anthocyanidins

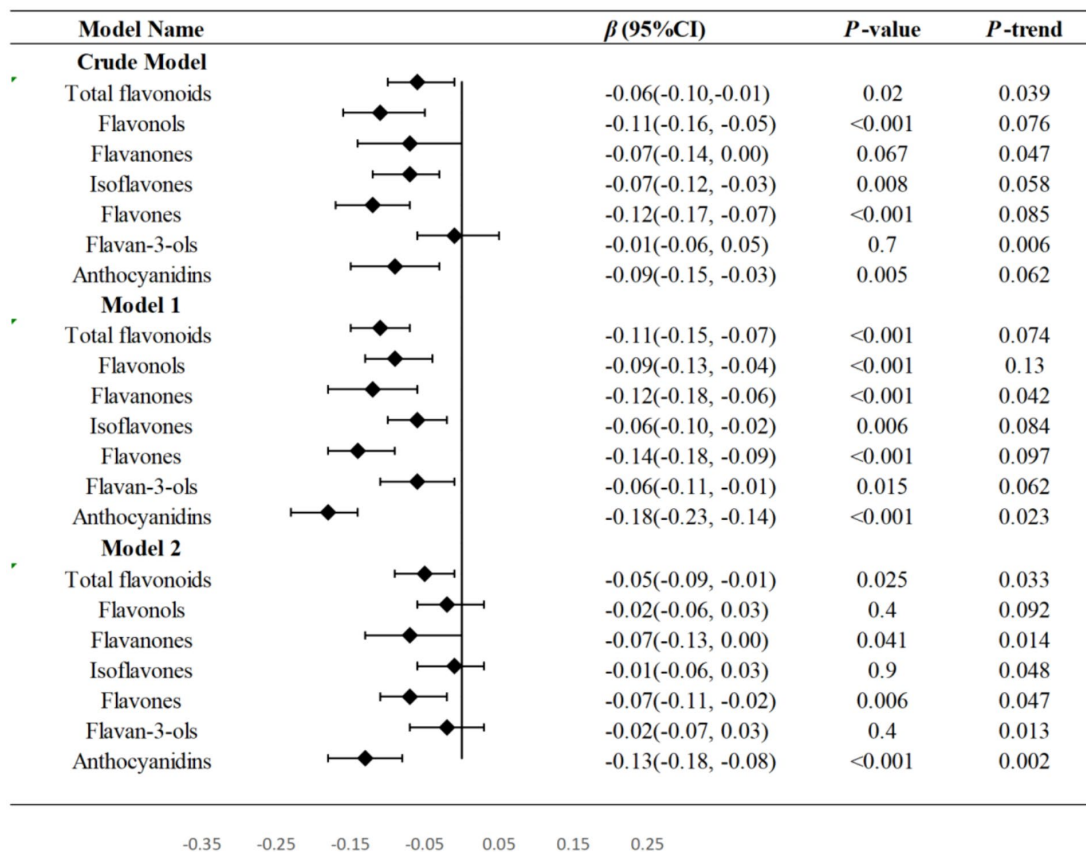
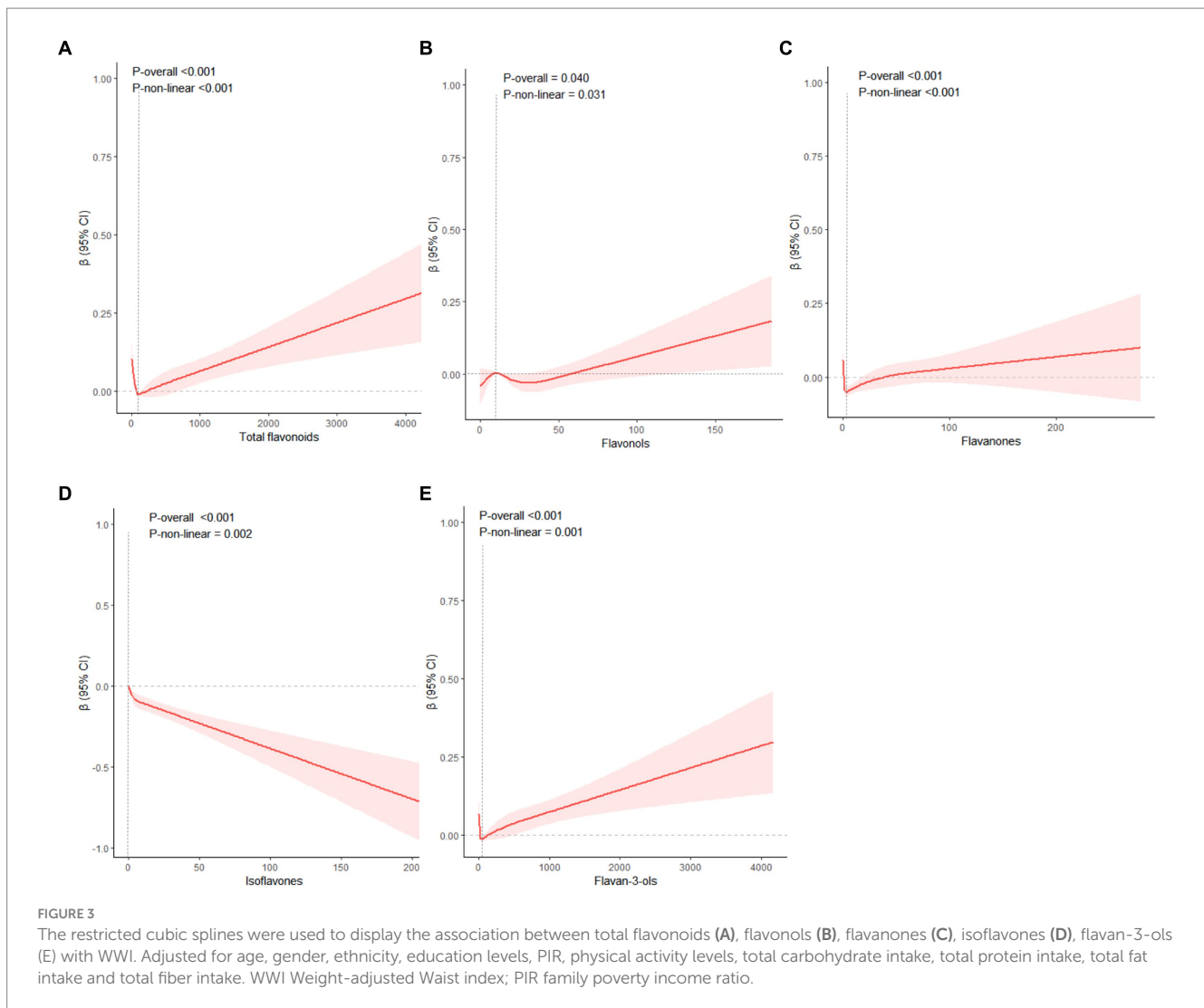


FIGURE 2 Forest plots were used to show the relationship between the third tertile of flavonoids and WWI in three models and linear trend test was conducted by treating each flavonoid as a continuous variable. T Tertile; WWI Weight-adjusted waist index; PIR family income to poverty. Model 1: no covariates were adjusted. Model 2: adjusted for age, gender, ethnicity, education levels and PIR. Model 3: adjusted for age, gender, ethnicity, education levels, PIR, physical activity levels, total carbohydrate intake, total protein intake, total fat intake and total fiber intake.

(OR = 1.27; 95%CI:1.11–1.46) was positively correlated with obesity as determined by BMI in males (26). In our research, we found that flavanones (β : -0.07; 95%CI: -0.13, 0.00) and flavones (β : -0.07; 95%CI: -0.11, -0.02) were more effective in reducing WWI and male participants had a stronger negative correlation between the total amount of flavonoids and WWI. This disparity could be attributed to the fact that the authors of the referenced study utilized the Korean health and nutrition survey, which may have a predisposition towards abdominal obesity and distinct dietary pattern (kimchi was the main source of flavonoid intake) (27, 28). What is more, abdominal obesity was defined by WC in the research of Kim et al. (28), while in the presented study, the WWI was to describe abdominal obesity. Quercetin is the most commonly found individual flavonols in daily dietary intake. The research conducted by Nishimura et al. (29) indicated that quercetin intake was inversely correlated with fat mass, BMI and WC in central obese participants. However, daily quercetin supplement of 60 mg did not alter the overall abdominal fat content in a healthy population (29). Additional research has indicated that the administration of high doses of quercetin (500 or 1,000 mg) did not influence body mass or composition among healthy individual (30). These findings were different from the outcomes reported in our study that the dietary flavonols within a certain range (12.12–57.81 mg/100 g foods/day) had

the ability to attenuated WWI. This could imply that the effects of quercetin on certain parameters might be predominantly observed in individuals with higher body mass or when combined with other flavonols, as these also showed potential effects on obesity-related measures.

In our subgroup analysis, we identified that certain demographic groups may experience greater benefits from the intake of total flavonoids. Given the multi-ethnic composition of the United States, our study revealed that total flavonoids consumption has a particularly significant impact on populations other than Non-Hispanic Whites, notably among Mexican-Americans ($p = 0.007$) and individuals of Others ($p < 0.001$) ethnic backgrounds. This could be attributed to variations in lifestyle, dietary practices, and genetic predispositions across different racial and ethnic groups (31). It is possible that Non-Hispanic Whites have a reduced sensitivity to the effects of flavonoids, although additional research is necessary to confirm this hypothesis. With an increase in total flavonoids intake, we observed a trend of decreasing WWI among male participants. The potential mechanisms behind these associations might be related to sex differences in body composition influenced by dietary consumption. Body fat distribution varies between genders, with men tending to accumulate a greater amount of fat in the abdominal region compared to women. This makes men more prone to developing abdominal



obesity (32). Given the consistent rise in the incidence of abdominal obesity among men in the US (33), the inverse relationship between total flavonoids consumption and abdominal obesity in this demographic suggested that dietary recommendations emphasizing flavonoid-rich foods might be less effective in preventing abdominal obesity in women. Furthermore, a stronger correlation between total flavonoids intake and the WWI had been noted in individuals with PIR below 1.3. This could be attributed to disparities in the primary sources of flavonoids between lower and higher socioeconomic groups, with lower-income families potentially engaging more frequently in home-cooked meals, which may influence their dietary flavonoid intake (34, 35).

It is known that the pathogenesis of abdominal obesity was different from generalized obesity (36). Human omental preadipocytes cultured preadipocytes undergo lower adipogenic transcription factor expression and more TNF- α -induced apoptosis than human abdominal subcutaneous. Additionally, there are preadipocytes within visceral fat that have limited ability to proliferate and differentiate compared to subcutaneous fat (37). Omental adipocytes have been observed to display comparable or reduced levels of basal lipolysis and exhibit decreased sensitivity to

the insulin-induced antilipolytic effects *in vivo* (38). In fact, there is a higher level of glucose absorption in visceral fat in comparison to subcutaneous fat (39). Furthermore, the omental fat demonstrates a higher capacity to absorb plasma free fatty acids compared to the abdominal subcutaneous fat, which suggests that this specific process might play a role in the selective buildup of visceral fat (40). For our research, we observed a strong inverse relationship between the consumption of total flavonoids, flavanones, flavones, flavan-3-ols, and anthocyanidins with WWI. This finding indicated that a sufficient intake of above flavonoids could potentially be a mildly effective intervention for combating abdominal obesity. Naringenin (4',5,7-trihydroxyflavanone) is a flavanone commonly found in a variety of vegetables, fruits, herbs, and nuts that are widely consumed by humans (41, 42). Research conducted by Atsuyoshi et al. had shed light on the anti-diabetic properties of naringenin. Their findings indicated that naringenin can increase the secretion of adiponectin and reduce the size of 3T3-L1 cells in a concentration-dependent manner during the process of adipocyte differentiation (43). Adiponectin is significant for its role in enhancing glucose uptake without the need for insulin receptor stimulation, improving fatty acid metabolism, and reducing fatty acid levels, which in turn

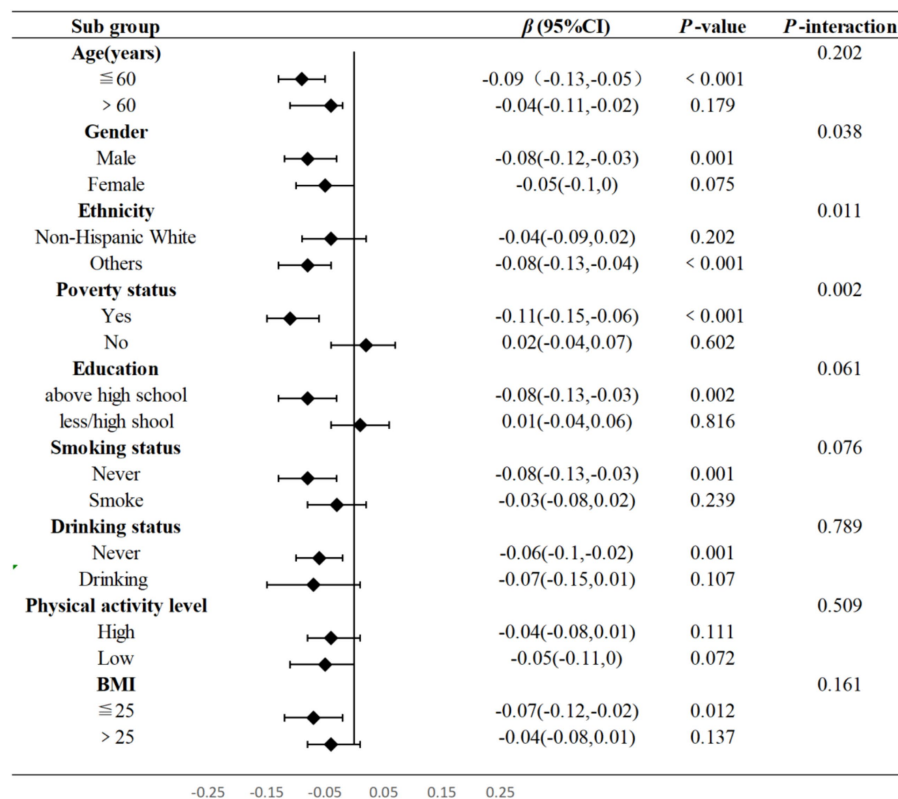


FIGURE 4

Subgroup and interaction analysis between total flavonoids and WWI. Adjusted for age, gender, ethnicity, education levels, PIR, physical activity levels, total carbohydrate intake, total protein intake, total fat intake and total fiber intake. T, Tertile, BMI, Body Mass Index, WWI, Weight-adjusted Waist, PIR, family poverty income ratio.

increases the sensitivity of the insulin receptor. Additionally, it promotes insulin sensitivity through the activation of liver AMP-activated protein kinase (AMPK), prevents arteriosclerosis, exhibits anti-inflammatory effects, and suppresses myocardial hypertrophy (44). Baicalein, which root constitute flavones baicalein (45), has been demonstrated in multiple studies to effectively alleviate insulin resistance and ameliorate complications associated with obesity (46, 47). Additionally, baicalein had been shown to exert protective effects on the functionality of INS382/13 cells and human pancreatic islets, thereby enhancing their secretory capacity (48). Luteolin, a widely available flavone in the diet, had also been identified for its potential anti-obesity characteristics (49). According to research by Lin et al. (50), luteolin was capable of diminishing fat storage in *Caenorhabditis elegans* by stimulating central serotonin signaling, which triggers fat loss. Treatment with luteolin led to an increase in serotonin synthesis within ADF neurons and the subsequent activation of serotonin-linked receptors MOD-1 and SER-6. This activation sequence ultimately results in an upregulation of lipolysis and fatty acid β -oxidation in the nematode model, *Caenorhabditis elegans* (50). Flavan-3-ols, encompassing the catechin group of compounds, are characterized by a 2-phenyl-3,4-dihydro-2H-chromen-3-ol framework. Green tea is particularly abundant in catechins, including epigallocatechin 3-gallate (EGCG) and catechin 3-gallate, which are both classified under flavan-3-ols (51). The impact of catechins and EGCG on abdominal obesity and metabolic syndrome has been a focus of extensive research,

particularly in animal models. For instance, a study involving mice on a high-fat diet, which constituted 60% of their caloric intake, demonstrated that a dietary regimen incorporating EGCG at a concentration of 0.32% for 16 weeks led to significant reductions in body weight gain, overall body fat, and visceral fat weight when compared to the control group without EGCG supplementation (52). Zang et al. (53) conducted research on the effects of green tea extract in juvenile and adult zebrafish models of obesity. Their findings indicated that green tea extract not only significantly reduced the accumulation of visceral adipose tissue in juveniles but also improved visceral adiposity and plasma triglyceride levels in adult zebrafish. The proposed molecular mechanism behind the anti-obesity effects of green tea extract involves the activation of the Wnt/ β -catenin and AMPK signaling pathways, which may help to ameliorate obesity-related phenotypes (53). Multiple studies indicated that the transition of preadipocytes into adipocytes is governed by a tightly regulated sequence of transcriptional events, with proliferator-activated receptor gamma (PPAR γ) and CCAT/enhancer-binding protein α (C/EBP α) being pivotal mediators in this process (54, 55). PPAR γ functions as a regulator in the lipid synthesis pathway, and its overexpression leads to an enlargement and multiplication of lipid molecules within 3T3-L1 preadipocytes undergoing differentiation (56). The black soybean contains various anthocyanins like cyanidin-3-O-glucoside, and delphinidin-3-O-glucoside, which have significant implications. They have been studied to increase preadipocyte differentiation through

downregulated the expression of PPAR γ and C/EBP α in a concentration-dependent manner (57). Additionally, anthocyanins can stimulate lipolysis in mature adipocytes. Oral administration of cyanidin had been shown to decrease hepatic and plasma triglyceride levels, reduce adiposity, and enhance glucose tolerance in mice on a high-fat diet. Metabolomic analysis of the liver indicated that cyanidin influenced the metabolic profile towards increased fatty acid oxidation and ketogenesis (58).

The robustness and broad applicability of our research findings were bolstered by the large and diverse sample size, enabling us to detect subtle association between dietary flavonoids consumption and abdominal obesity within multiple subgroups. Our study suggested that dietary flavonoids could pave the way for creating functional foods to combat abdominal obesity. However, it is crucial to recognize and address the limitations inherent in our study. Initially, our research was confined to examining the link between flavonoids consumption and the WWI, and further investigation is required to confirm the correlation between flavonoid intake and other metabolic health markers. Additionally, different dietary sources (foods/drinks) affect flavonoids bioavailability, the intake data may overestimated the actual amount of flavonoids intake that exhibit physiological health benefits. Lastly, the exclusion of supplement data may limit the generalizability of our findings, as it does not account for all possible sources of flavonoids that participants may have consumed. Future studies should consider including data on the use of flavonoids supplements to provide a more comprehensive evaluation of total flavonoids intake.

5 Conclusion

Our research contributed novel insights into the intricate relationship between flavonoids consumption and abdominal obesity, with our findings indicating a significant inverse correlation between flavonoids intake and the WWI among adults in the US. This underscored the potential significance of flavonoids intake in the management of abdominal obesity and laid a foundation for the development of dietary guidelines for flavonoid consumption. Nonetheless, the observed associations between flavonoids and WWI should not be interpreted as indicative of a causal relationship, and additional prospective cohort studies or randomized controlled trials are necessary to confirm the causal relationship between flavonoids intake and abdominal obesity, as well as to determine the safe and effective dosages of flavonoids for various populations.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

References

1. Yanovski JA. Trends in underweight and obesity — scale of the problem. *Nat Rev Endocrinol.* (2018) 14:5–6. doi: 10.1038/nrendo.2017.157
2. Abarca-Gómez L, Abdeen ZA, Hamid ZA, Abu-Rmeileh NM, Acosta-Cazares B, Acuin C, et al. Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: a pooled analysis of 2416 population-based measurement

Ethics statement

The studies involving humans were approved by The National Centre for Health Statistics' research ethics review board. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

SZ: Writing – original draft. MY: Writing – original draft. XiL: Writing – original draft. HW: Writing – original draft. XuL: Writing – original draft. YF: Writing – original draft. DW: Funding acquisition, Writing – review & editing. BZ: Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This work was supported by the Anhui Provincial Natural Science Foundation (2008085MH244) and grants from The National Key Research and Development Program of China (2020YFC2006502). The funding agency had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgments

This study used data from the NHANES. We thank the NCHS and the US Centers for Disease Control and Prevention for their financial support for data collection and analysis. We also thank the study participants and staff.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

studies in 128.9 million children, adolescents, and adults. *Lancet.* (2017) 390:2627–42. doi: 10.1016/S0140-6736(17)32129-3

3. Pischon T, Boeing H, Hoffmann K, Bergmann M, Schulze MB, Overvad K, et al. General and abdominal adiposity and risk of death in Europe. *N Engl J Med.* (2008) 359:2105–20. doi: 10.1056/NEJMoa0801891

4. Klötting N, Fasshauer M, Dietrich A, Kovacs P, Schön MR, Kern M, et al. Insulin-sensitive obesity. *Am. J. Physiol. Endocrinol. Metab.* (2010) 299:E506–15. doi: 10.1152/ajpendo.00586.2009
5. Kawada T, Andou T, Fukumitsu M. Waist circumference, visceral abdominal fat thickness and three components of metabolic syndrome. *Diabetes Metab Syndr.* (2016) 10:4–6. doi: 10.1016/j.dsx.2015.08.013
6. Berentzen TL, Ångquist L, Kotronen A, Borra R, Yki-Järvinen H, Iozzo P, et al. Waist circumference adjusted for body mass index and intra-abdominal fat mass. *PLoS One.* (2012) 7:e32213. doi: 10.1371/journal.pone.0032213
7. Park Y, Kim NH, Kwon TY, Kim SG. A novel adiposity index as an integrated predictor of cardiometabolic disease morbidity and mortality. *Sci Rep.* (2018) 8:16753. doi: 10.1038/s41598-018-35073-4
8. Qin Z, Chang K, Yang Q, Yu Q, Liao R, Su B. The association between weight-adjusted-waist index and increased urinary albumin excretion in adults: a population-based study. *Front Nutr.* (2022) 9:941926. doi: 10.3389/fnut.2022.941926
9. Kim NH, Park Y, Kim NH, Kim SG. Weight-adjusted waist index reflects fat and muscle mass in the opposite direction in older adults. *Age Ageing.* (2021) 50:780–6. doi: 10.1093/ageing/afaa208
10. Kim JY, Choi J, Vella CA, Criqui MH, Allison MA, Kim NH. Associations between weight-adjusted waist index and abdominal fat and muscle mass: multi-ethnic study of atherosclerosis. *Diabetes Metab J.* (2022) 46:747–55. doi: 10.4093/dmj.2021.0294
11. Hu Q, Han K, Shen J, Sun W, Gao L, Gao Y. Association of weight-adjusted-waist index with non-alcoholic fatty liver disease and liver fibrosis: a cross-sectional study based on NHANES. *Eur J Med Res.* (2023) 28:263. doi: 10.1186/s40001-023-01205-4
12. Wang Z, Shao X, Xu W, Xue B, Zhong S, Yang Q. The relationship between weight-adjusted-waist index and diabetic kidney disease in patients with type 2 diabetes mellitus. *Front Endocrinol.* (2024) 15:1345411. doi: 10.3389/fendo.2024.1345411
13. Zhang Y, Wu H, Li C, Liu C, Liu M, Liu X, et al. Associations between weight-adjusted waist index and bone mineral density: results of a nationwide survey. *BMC Endocr Disord.* (2023) 23:162. doi: 10.1186/s12902-023-01418-y
14. Li M, Yu X, Zhang W, Yin J, Zhang L, Luo G, et al. The association between weight-adjusted-waist index and depression: results from NHANES 2005–2018. *J Affect Disord.* (2024) 347:299–305. doi: 10.1016/j.jad.2023.11.073
15. Prasad S, Phromnoi K, Yadav VR, Chaturvedi MM, Aggarwal BB. Targeting inflammatory pathways by flavonoids for prevention and treatment of cancer. *Planta Med.* (2010) 76:1044–63. doi: 10.1055/s-0030-1250111
16. Havsteen BH. The biochemistry and medical significance of the flavonoids. *Pharmacol Ther.* (2002) 96:67–202. doi: 10.1016/s0163-7258(02)00298-x
17. Xie R, Zhang Y. Associations between dietary flavonoid intake with hepatic steatosis and fibrosis quantified by VCTE: evidence from NHANES and FNDSS. *Nutr Metab Cardiovasc Dis.* (2023) 33:1179–89. doi: 10.1016/j.numecd.2023.03.005
18. Zhao Z, Gao W, Ding X, Xu X, Xiao C, Mao G, et al. The association between dietary intake of flavonoids and its subclasses and the risk of metabolic syndrome. *Front Nutr.* (2023) 10:1195107. doi: 10.3389/fnut.2023.1195107
19. Vernarelli JA, Lambert JD. Flavonoid intake is inversely associated with obesity and C-reactive protein, a marker for inflammation, in US adults. *Nutr Diabetes.* (2017) 7:e276. doi: 10.1038/nutd.2017.22
20. Aloo S-O, Ofosu FK, Kim N-H, Kilonzi SM, Oh D-H. Insights on dietary polyphenols as agents against metabolic disorders: obesity as a target disease. *Antioxidants.* (2023) 12:416. doi: 10.3390/antiox12020416
21. Ogden CL, Carroll MD, Lawman HG, Fryar CD, Kruszon-Moran D, Kit BK, et al. Trends in obesity prevalence among children and adolescents in the United States, 1988–1994 through 2013–2014. *JAMA.* (2016) 315:2292–9. doi: 10.1001/jama.2016.6361
22. Ali MK, Bullard KM, Beckles GL, Stevens MR, Barker L, Venkat Narayan KM, et al. Household income and cardiovascular disease risks in U.S. children and young adults. *Diabetes Care.* (2011) 34:1998–2004. doi: 10.2337/dc11-0792
23. Castro-Barquero S, Ruiz-León AM, Sierra-Pérez M, Estruch R, Casas R. Dietary strategies for metabolic syndrome: a comprehensive review. *Nutrients.* (2020) 12:2983. doi: 10.3390/nu12102983
24. Beecher GR. Overview of dietary flavonoids: nomenclature, occurrence and intake. *J Nutr.* (2003) 133:3248S–54S. doi: 10.1093/jn/133.10.3248S
25. Sebastian RS, Fanelli Kuczmariski M, Wilkinson Enns C, Goldman JD, Murayi T, Moshfegh AJ, et al. Application of the database of flavonoid values for USDA food codes 2007–2010 in assessing intake differences between the healthy aging in neighborhoods of diversity across the life span (HANDLS) study and what we eat in America (WWEIA), NHANES. *J Food Compos Anal.* (2021) 104:104124. doi: 10.1016/j.jfca.2021.104124
26. Kim S-A, Kim J, Jun S, Wie G-A, Shin S, Joung H. Association between dietary flavonoid intake and obesity among adults in Korea. *Appl Physiol Nutr Metab.* (2020) 45:203–12. doi: 10.1139/apnm-2019-0211
27. Lim U, Ernst T, Buchthal SD, Latch M, Albright CL, Wilkens LR, et al. Asian women have greater abdominal and visceral adiposity than Caucasian women with similar body mass index. *Nutr Diabetes.* (2011) 1:e6. doi: 10.1038/nutd.2011.2
28. Kim YJ, Park MY, Chang N, Kwon O. Intake and major sources of dietary flavonoid in Korean adults: Korean National Health and nutrition examination survey 2010–2012. *Asia Pac J Clin Nutr.* (2015) 24:456–63. doi: 10.6133/apjcn.2015.24.3.04
29. Nishimura M, Muro T, Kobori M, Nishihira J. Effect of daily ingestion of quercetin-rich onion powder for 12 weeks on visceral fat: a randomised, double-blind, placebo-controlled, parallel-group study. *Nutrients.* (2019) 12:91. doi: 10.3390/nu12010091
30. Knab AM, Shanely RA, Jin F, Austin MD, Sha W, Nieman DC. Quercetin with vitamin C and niacin does not affect body mass or composition. *Appl Physiol Nutr Metab.* (2011) 36:331–8. doi: 10.1139/h11-015
31. Byrd AS, Toth AT, Stanford FC. Racial disparities in obesity treatment. *Curr Obes Rep.* (2018) 7:130–8. doi: 10.1007/s13679-018-0301-3
32. Power ML, Schulkin J. Sex differences in fat storage, fat metabolism, and the health risks from obesity: possible evolutionary origins. *Br J Nutr.* (2008) 99:931–40. doi: 10.1017/S0007114507853347
33. Sun J-Y, Huang W-J, Hua Y, Qu Q, Cheng C, Liu H-L, et al. Trends in general and abdominal obesity in US adults: evidence from the National Health and nutrition examination survey (2001–2018). *Front Public Health.* (2022) 10:925293. doi: 10.3389/fpubh.2022.925293
34. Fanelli Kuczmariski M, Sebastian RS, Goldman JD, Murayi T, Steinfeldt LC, Eosso JR, et al. Dietary flavonoid intakes are associated with race but not income in an urban population. *Nutrients.* (2018) 10:1749. doi: 10.3390/nu10111749
35. Monsivais P, Aggarwal A, Drewnowski A. Time spent on home food preparation and indicators of healthy eating. *Am J Prev Med.* (2014) 47:796–802. doi: 10.1016/j.amepre.2014.07.033
36. Breitfeld J, Kehr S, Müller L, Stadler PF, Böttcher Y, Blüher M, et al. Developmentally driven changes in Adipogenesis in different fat depots are related to obesity. *Front Endocrinol.* (2020) 11:138. doi: 10.3389/fendo.2020.00138
37. Tchkonja T, Tchoukalova YD, Giorgadze N, Pirtskhalava T, Karagiannides I, Forse RA, et al. Abundance of two human preadipocyte subtypes with distinct capacities for replication, adipogenesis, and apoptosis varies among fat depots. *Am J Physiol Endocrinol Metab.* (2005) 288:E267–77. doi: 10.1152/ajpendo.00265.2004
38. Boivin A, Brochu G, Marceau S, Marceau P, Houllé F-S, Tchernof A. Regional differences in adipose tissue metabolism in obese men. *Metabolism.* (2007) 56:533–40. doi: 10.1016/j.metabol.2006.11.015
39. Veilleux A, Blouin K, Rhéaume C, Daris M, Marette A, Tchernof A. Glucose transporter 4 and insulin receptor substrate-1 messenger RNA expression in omental and subcutaneous adipose tissue in women. *Metabolism.* (2009) 58:624–31. doi: 10.1016/j.metabol.2008.12.007
40. Koutsari C, Ali AH, Mundi MS, Jensen MD. Storage of circulating free fatty acid in adipose tissue of postabsorptive humans: quantitative measures and implications for body fat distribution. *Diabetes.* (2011) 60:2032–40. doi: 10.2337/db11-0154
41. Lin L-Z, Mukhopadhyay S, Robbins RJ, Harnly JM. Identification and quantification of flavonoids of Mexican oregano (*Lippia graveolens*) by LC-DAD-ESI/MS analysis. *J Food Compos Anal.* (2007) 20:361–9. doi: 10.1016/j.jfca.2006.09.005
42. Bugianesi R, Catasta G, Spigno P, D’Uva A, Maiani G. Naringenin from cooked tomato paste is bioavailable in men. *J Nutr.* (2002) 132:3349–52. doi: 10.1093/jn/132.11.3349
43. Nishina A, Sato D, Yamamoto J, Kobayashi-Hattori K, Hirai Y, Kimura H. Antidiabetic-like effects of Naringenin-7-O-glucoside from edible *Chrysanthemum* ‘Kotobuki’ and Naringenin by activation of the PI3K/Akt pathway and PPAR γ . *Chem Biodivers.* (2019) 16:e1800434. doi: 10.1002/cbdv.201800434
44. Lazra Y, Falach A, Frenkel L, Rozenberg K, Sampson S, Rosenzweig T. Autocrine/paracrine function of globular adiponectin: inhibition of lipid metabolism and inflammatory response in 3T3-L1 adipocytes. *J Cell Biochem.* (2015) 116:754–66. doi: 10.1002/jcb.25031
45. Gao Y, Snyder SA, Smith JN, Chen YC. Anticancer properties of baicalein: a review. *Med Chem Res.* (2016) 25:1515–23. doi: 10.1007/s00044-016-1607-x
46. Xing Y, Ren X, Li X, Sui L, Shi X, Sun Y, et al. Baicalein enhances the effect of Acarbose on the improvement of nonalcoholic fatty liver disease associated with prediabetes via the inhibition of *De novo* lipogenesis. *J Agric Food Chem.* (2021) 69:9822–36. doi: 10.1021/acs.jafc.1c04194
47. Guo S, Zhang L, Wu S, Liu H. Research progress of typical flavonoids in improving insulin resistance. *Arch Med Sci Atheroscler Dis.* (2020) 5:335–e342. doi: 10.5114/amsad.2020.103472
48. Liu Y, Zhao G, Zhang S. Effects and action mechanism of baicalein on glucolipid metabolism and renal function of the rats with type 2 diabetes mellitus. *Western J Traditional Chin Med.* (2016) 29:23–6.
49. Xu N, Zhang L, Dong J, Zhang X, Chen Y-G, Bao B, et al. Low-dose diet supplement of a natural flavonoid, luteolin, ameliorates diet-induced obesity and insulin resistance in mice. *Mol Nutr Food Res.* (2014) 58:1258–68. doi: 10.1002/mnfr.201300830
50. Lin Y, Yang N, Bao B, Wang L, Chen J, Liu J. Luteolin reduces fat storage in *Caenorhabditis elegans* by promoting the central serotonin pathway. *Food Funct.* (2020) 11:730–40. doi: 10.1039/C9FO02095K
51. Yang YJ, Kim YJ, Yang YK, Kim JY, Kwon O. Dietary flavan-3-ols intake and metabolic syndrome risk in Korean adults. *Nutr Res Pract.* (2012) 6:68–77. doi: 10.4162/nrp.2012.6.1.68
52. Bose M, Lambert JD, Ju J, Reuhl KR, Shapses SA, Yang CS. The major green tea polyphenol, (–)-Epigallocatechin-3-Gallate, inhibits obesity, metabolic syndrome, and fatty liver disease in high-fat-fed mice. *J Nutr.* (2008) 138:1677–83. doi: 10.1093/jn/138.9.1677

53. Zang L, Shimada Y, Nakayama H, Kim Y, Chu D-C, Juneja LR, et al. RNA-seq based transcriptome analysis of the anti-obesity effect of green tea extract using zebrafish obesity models. *Molecules*. (2019) 24:3256. doi: 10.3390/molecules24183256
54. Assi M, Kenawi M, Ropars M, Rébillard A. Interleukin-6, C/EBP- β and PPAR- γ expression correlates with intramuscular liposarcoma growth in mice: the impact of voluntary physical activity levels. *Biochem Biophys Res Commun*. (2017) 490:1026–32. doi: 10.1016/j.bbrc.2017.06.158
55. Kim E-J, Kang M-J, Seo YB, Nam S-W, Kim G-D. Acer okamotoanum Nakai leaf extract inhibits Adipogenesis via suppressing expression of PPAR γ and C/EBP α in 3T3-L1 cells. *J Microbiol Biotechnol*. (2018) 28:1645–53. doi: 10.4014/jmb.1802.01065
56. Guo L, Li X, Tang Q-Q. Transcriptional regulation of adipocyte differentiation: a central role for CCAAT/enhancer-binding protein (C/EBP) β . *J Biol Chem*. (2015) 290:755–61. doi: 10.1074/jbc.R114.619957
57. Park M, Sharma A, Lee H-J. Anti-Adipogenic effects of Delphinidin-3-O- β -glucoside in 3T3-L1 Preadipocytes and primary white adipocytes. *Molecules*. (2019) 24:1848. doi: 10.3390/molecules24101848
58. Jia Y, Wu C, Kim Y-S, Yang SO, Kim Y, Kim J-S, et al. A dietary anthocyanin cyanidin-3-O-glucoside binds to PPARs to regulate glucose metabolism and insulin sensitivity in mice. *Commun Biol*. (2020) 3:514. doi: 10.1038/s42003-020-01231-6