



OPEN ACCESS

EDITED BY

José Pinela,
Instituto Politécnico de Bragança, Portugal

REVIEWED BY

Zhangsheng Zhu,
South China Agricultural University, China
Jian Jun Lei,
South China Agricultural University, China

*CORRESPONDENCE

Jorge Ariel Torres-Castillo
✉ joatorres@docentes.uat.edu.mx

RECEIVED 22 December 2023

ACCEPTED 27 March 2024

PUBLISHED 15 April 2024

CITATION

Pérez-Ramírez R, Moreno-Ramírez YR,
Ruiz-De-La-Cruz G, Juárez-Aragón MC,
Aguirre-Mancilla CL, Niño-García N and
Torres-Castillo JA (2024) Piquin chili, a wild
spice: natural variation in nutraceutical
contents.

Front. Nutr. 11:1360299.

doi: 10.3389/fnut.2024.1360299

COPYRIGHT

© 2024 Pérez-Ramírez, Moreno-Ramírez,
Ruiz-De-La-Cruz, Juárez-Aragón, Aguirre-
Mancilla, Niño-García and Torres-Castillo.
This is an open-access article distributed
under the terms of the [Creative Commons
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,
distribution or reproduction in other forums is
permitted, provided the original author(s) and
the copyright owner(s) are credited and that
the original publication in this journal is cited,
in accordance with accepted academic
practice. No use, distribution or reproduction
is permitted which does not comply with
these terms.

Piquin chili, a wild spice: natural variation in nutraceutical contents

Rogelio Pérez-Ramírez¹, Yolanda del Rocio Moreno-Ramírez²,
Gilberto Ruiz-De-La-Cruz³, María Cruz Juárez-Aragón¹,
César Leobardo Aguirre-Mancilla⁴, Nohemí Niño-García⁵ and
Jorge Ariel Torres-Castillo^{1,5*}

¹Instituto de Ecología Aplicada, Universidad Autónoma de Tamaulipas, Ciudad Victoria, Tamaulipas, Mexico, ²Facultad de Ingeniería y Ciencias, Universidad Autónoma de Tamaulipas, Ciudad Victoria, Tamaulipas, Mexico, ³Laboratorio de Biotecnología Animal, Centro de Biotecnología Genómica, Instituto Politécnico Nacional, Reynosa, Tamaulipas, Mexico, ⁴Departamento de Posgrado, Instituto Tecnológico Roque, Tecnológico Nacional de México, Celaya, Guanajuato, Mexico, ⁵Unidad Académica Multidisciplinaria Mante Centro, Universidad Autónoma de Tamaulipas, Ciudad Mante, Tamaulipas, Mexico

The piquin chili is a wild spice widely consumed from the South United States to Central America and stands out as a source of flavonoids, essential metabolites with antioxidant properties. The concentrations of flavonoids, carotenoids, and capsaicinoids vary according to regions, maturity stages, and ripening processes. These compounds, which are known for their health benefits and industrial applications, highlight the importance of identifying ideal environmental conditions for collecting fruits with the highest contents. Comprehensive studies of the piquin chili are essential for understanding its properties for the benefit of consumers. This approach fortifies trade, contributes to resource conservation, and advances cultivated chili production.

KEYWORDS

nutraceuticals, bioactive compounds, chili, capsicum, wild plants

1 Introduction

The plants synthesize phytochemical compounds as part of biosynthetic processes to perform ecological and physiological functions. The phytochemical compounds are perceived through aromas, flavors, and colors, which humans could exploit (1). The wide phytochemical diversity with nutraceutical potential from wild plant sources includes phenolics, alkaloids, terpenoids, polysaccharides, glycosides, and compounds of a peptide nature (2–4).

The consumption of nutraceutical sources from vegetal ecosystems is widely practiced across world cultures, showing the importance of conservation and proper management of natural resources (5–7). Nevertheless, consuming products with nutraceutical potential from wild sources must consider the differences in contents and composition compared with cultivated forms, which consequently affect the product quality, organoleptic, and functional characteristics, that are considered as influencing elements of consumer decision (8–11). It is imperative to perform comprehensive studies to maintain optimal production conditions for nutraceuticals, recreating natural processes supported by genetic improvement, analytical techniques, traditional practices, natural resource management, industrial process innovations, specific sustainable forestry production principles, and public policies for their exploitation (12).

The piquin chili, *Capsicum annuum* var. *glabriusculum* (Dunal) Heiser and Pickersgill, is a wild form of *Capsicum*, considered the ancestor of *anunum* species, widely distributed from the southeast of the United States of America to northern South America, with names such as piquin, chile de monte, chiltepin, chiltepe, amashito, timpinchile, amash, chile congo, mosquito chili, kipin, and chilpaya (13–16). This wild resource contains nutraceutical compounds integrated with phenolics, alkaloids, carotenoids, and volatile compounds. Piquin chili fruits are regionally consumed mainly through direct extraction from the ecosystem (17, 18).

The piquin chili represents a food resource that provides nutraceutical compounds to the diet of the rural population since consumption is associated with customs from the chili distribution area, thereby highlighting the interaction of identity and culture (17, 19). As a wild plant, it shows wide genetic variability and plasticity resulting from local adaptations (20). Hence, its nutritional and nutraceutical composition results from genetic influence, the environment in which it is grown, ecological interactions, harvesting practices, harvesting season, and post-harvest storage and processing (21). The research on the phytochemicals and nutraceutical potential of the wild forms of *Capsicum* is limited, compared with improved and commercial varieties of *C. annuum*, *C. frutescens*, and *C. chinense* that have been extensively studied (22–25). Although there are studies about the phytochemistry of piquin chili, management has yet to be archived to ensure the content of bioactive compounds with potential effects. The nutraceutical potential of piquin chili leads is influenced by nature and management conditions, supporting that holistic knowledge can reinforce future initiatives for establishing quality criteria for this spice focused on consumer benefits by means of chemical diversity, variation in contents, and evidence about accumulation patterns in wild and cultivated natural populations.

According to official statistics in Mexico¹, the production of piquin chili is not as high as other varieties; however, the commercial value of a ton of cultivated green piquin chili is close to US\$ 23,800. In comparison, a ton of jalapeño chili is approximately US\$ 500, and a ton of serrano chili is approximately US\$ 640. Regarding its role in the rural economy, it was recently highlighted that the price per kilogram of wild piquin chili is strongly associated with the number of fruits collected per day, which depends on multiple environmental factors and the season during which the plants are produced. For example, in April, in the northeastern region of Mexico, a kilogram of green wild piquin costs up to US\$ 85. The wild piquin chili is considered a social identity resource where the collection is performed by the male gender given the remoteness and isolation of the wild piquin populations; however, the cleaning and selection of the fruits are carried out by women, which shows the relevance of the piquin chili in the territory (26).

2 The piquin is a wild source of nutraceuticals

The dependence on plant resources to obtain functional benefits lies in the knowledge acquired through generations, traditional

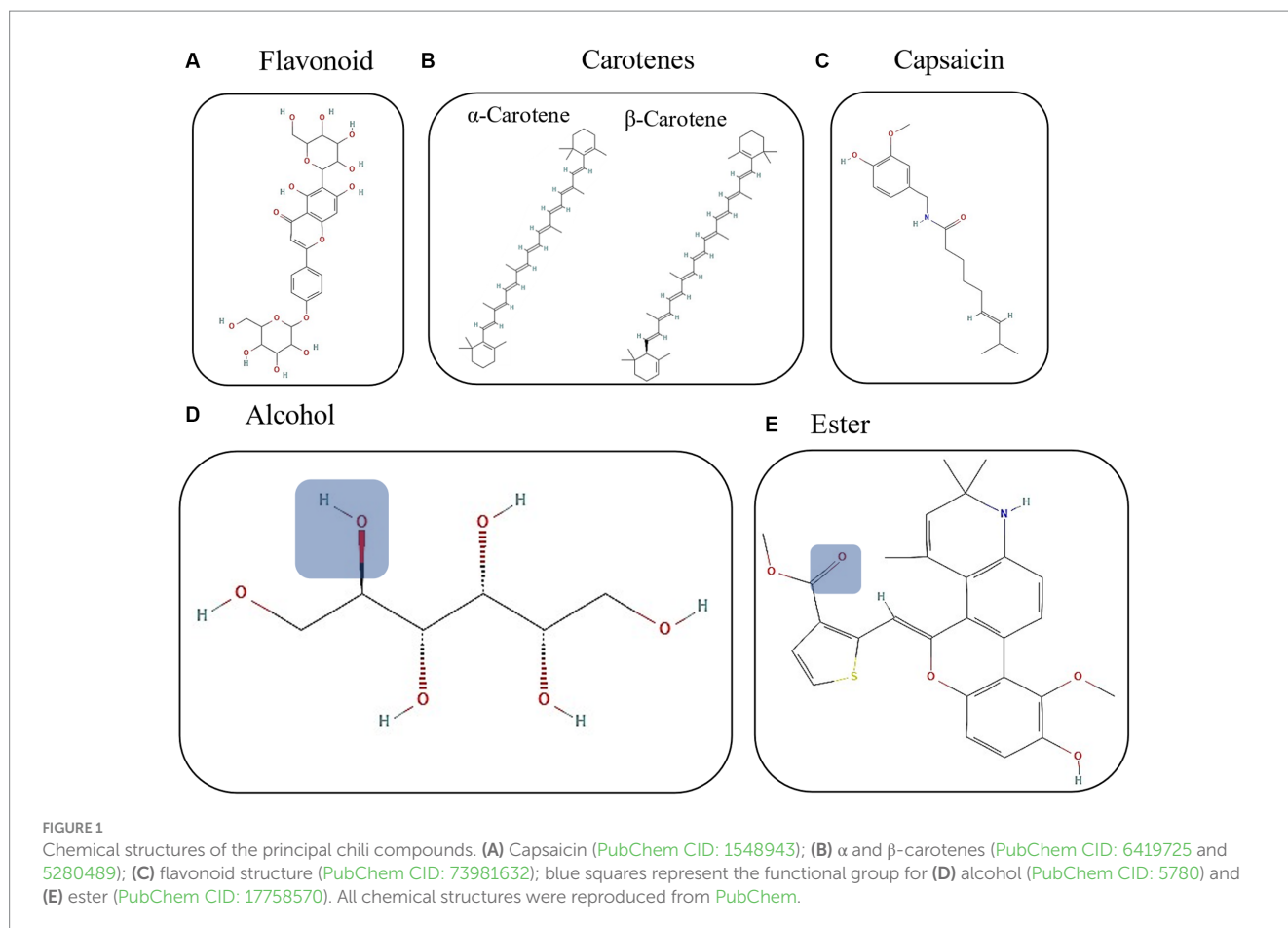
knowledge, and seasonal use related to the productive and phenological cycle of plants in native, indigenous, and rural communities (27, 28). The consumption of wild plants, such as piquin chili, has a close relationship between humans and the ecosystem; for example, this natural resource activates the local economy due to the interest of people to acquire piquin plants, fruits, and derivatives such as spicy oils, sauces, salts, and dry milled chili. All these products originate in rural areas and are sold in rural and cities, where consumers recognize them by their flavors and pungency (29, 30). As a wild species, its organoleptic and phytochemical characteristics vary (Figure 1), causing an impact on the consumer. Although agroforestry or greenhouse production programs exist, few species have achieved the phytochemical contents associated with the characteristics required by the consumer. It is essential to understand the impact of environmental conditions on the growth, organoleptic, and functional characteristics of the piquin chili due to its various compounds with nutraceutical potential that are highly influenced by the environment (Table 1) (20).

3 Flavonoids

Flavonoids are specialized metabolites with a 15-carbon structural core called the flavone skeleton, where different chemical substitutions (Figure 1A) give them biofunctional and antioxidant properties. They are essential in the food, cosmetics, and pharmaceutical industries. Flavonoids can accumulate in specific organs or tissues in significant concentrations, which lead to identifying rich natural sources of these compounds (40). The presence of flavonoids in piquin chili has been repeatedly reported, highlighting its phytochemical potential as a natural source (18, 20).

The piquin chili flavonoid concentrations vary between the evaluated accessions. Samples collected in the areas of Valles Centrales, Sierra Sur, and Oaxaca Isthmus (Southern Mexico) had flavonoid concentration differences according to the immature and mature state, with 0.5 and 0.95 mg per gram of tissue, respectively, showing a difference close to 50% of flavonoid concentration. However, it does not consider the chemical diversity of total flavonoids (41). The mature chili peppers obtained in Sonora (Northwest of Mexico), analyzed through HPLC-DAD, reported piquin chili with 0.065 ± 0.006 mg per gram in dry weight (DW) (42). Mature chili peppers collected in the central, coastal, plain, and southern regions of Tamaulipas (Northeast of Mexico) had contents between 0.24 and 0.36 mg of quercetin equivalents per gram of tissue (20), lower than those reported in the Oaxaca samples (41). Among populations within a region, mature piquin chili fruits have flavonoid contents with variations that reach up to 30%. Even the content of flavonoids is higher than in some chili-cultivated varieties (20, 43, 44). This highlights the precedence of the germplasm and its ripening process to maximize flavonoid contents and unleash the full range of potential benefits associated with consuming these phytochemicals (45). In this regard, the variation in flavonoid contents in several chili species and genotypes has shown dynamic patterns, where the environmental conditions have been remarked as more important in the final contents; however, the genetic interaction, agronomical practices, and the direction of particular flavor by artificial selection according to preferences and uses should not be discarded to reach high yields when the purpose is harvesting chili fruits rich in flavonoids (22, 46).

1 <https://nube.siap.gob.mx/cierreagricola/> (Accessed March 22, 2024).



4 Carotenoids

Carotenoids are lipophilic compounds produced by plants that protect photosynthetic systems from light excess and are precursors of phytohormones. The carotenoids have a 40-carbon skeleton derived from repetitive condensation of isoprene units (Figure 1B) and have been considered of nutraceutical interest against complications associated with oxidative stress (47). For the chili peppers, carotenoids contribute to the coloration of the mature states, which is evident as the chlorophyll degrades (48, 49). In this regard, it is worth highlighting the marked influence of environmental conditions and the genetics of chili peppers regarding the accumulation of carotenoids, which have been regularly identified in various chili genotypes, where specific accumulation patterns have been observed (50–52). As an example, in the piquin chili, the carotenoid content has been reported as contrasting between the immature (green fruit) and mature states (red fruit), in addition to being undetectable in the first state and reaching considerable levels when ripe (33).

However, its accumulation and degradation patterns are highly dynamic at a molecular level, which results in different amounts and a wide chemical diversity of carotenoids and related compounds during the ripening process (51). While it is suggested that the maximum accumulation of some carotenoids is reached when the chilies mature, this process is responsible for the bright and attractive tones of the chilies (53). Mature chili is recommended for use due to its accumulation of some carotenoids and its increased antioxidant property, which is associated with its

industrial uses and consumption to obtain the greatest nutraceutical effects (32). The Cumpas and Sahuaripa chilies, ecotypes of piquin chili, showed levels with significant differences of β -carotene on mature chilies, with 6.03 and 5.70 mg per gram for DW, respectively. Both chilies were from Sonora State (northwest of Mexico) and were grown under greenhouse conditions (34). Two ecotypes of piquin chili from Tabasco State (southeastern Mexico) showed significant differences between carotenoid contents in ripe and unripe fruits. Mature fruits show higher carotenoid content and are subjected to various levels of shading, reaching up to 28.80 mg g⁻¹ of DW in the dry season, in an open sky system for the Garbanzo genotype and 23.71 mg g⁻¹ of DW in the rainy season, in an open sky system for the Amashito genotype (50). The same research shows a difference in carotenoid levels in mature chili peppers, and these differ according to the shading conditions, with higher levels when the plants are exposed to natural light in the open field system and lower levels when shade conditions are increased. The carotenoid levels were different between seasons and humidity. The amount of pigment, associated mainly with carotenoids, present shows a wide heterogeneity between ecotypes of piquin chili from a wide area sampled in Tamaulipas (northeast of Mexico) (20). Thus, the levels of carotenoids present in piquin chili are highly influenced by phenological, genotypic, and environmental conditions, which increase the complexity of the consensus under which this wild plant can have a specific content and stability according to its growth conditions. Although this could be considered a

TABLE 1 The main phytochemicals identified in piquin chili.

Phytochemical	Line, ecotype, and genotype	Content range	Variation source	Reference
Flavonoids	Chiltepin Northwest Mexico	4.24 ± 0.10 mg/g DW	Nr	Hayano-Kanashiro et al. (14)
	Amashito	13.93–20.56 mg/g DW	Phe	De la Cruz-Ricardez et al. (31)
	Garbanzo	6.15–10.32 mg/g DW		
	Cumpas	2.34 ± 0.14–4.14 ± 1.0 mg CE/g DW	Gr, Phe	Vázquez-Flores et al. (32)
	Sahuaripa	1.32 ± 0.17–3.53 ± 1.3 mg CE/g DW		
Carotenoids	Amashito	9.01–11.56 mg QE/g DW	Gn, Na	De la Cruz-Ricardez et al. (33)
	Garbanzo	5.06–9.62 mg QE/g DW		
	IS	0.24, 0.22	Gr	Moreno-Ramírez et al. (20)
	15 Ct, 14 Ct	0.36, 0.70		
	Cumpas	1.60 ± 0.02–6.03 ± 0.08 mgβCE/g DW	Gr, Phe	Vázquez-Flores et al. (34)
Capsaicinoids	Sahuaripa	0.66 ± 0.002–5.70 ± 0.8 mgβCE/g DW		
	Chiltepin Northwest Mexico	12.81 ± 0.7–33.23 ± 0.3	Phe	Hayano-Kanashiro et al. (14)
	Amashito	≈5.0–≈10.0 mg/g DW	Phe	De la Cruz-Ricardez et al. (35)
	Garbanzo	≈2.0–≈15.0 mg/g DW		
	Amashito	5.41–23.71 mg/g DW	Phe, Ag	De la Cruz-Ricardez et al. (33)
Capsaicinoids	Garbanzo	3.71–28.8 mg/g DW		
	III-1, I-3	13.7 ± 0.1, 27.2 ± 0.3 mg/g DW	Na	Moreno-Ramírez et al. (18)
	Pue 01, Nay 02	135 ± 19 μg/ml, 1,379 ± 95 μg/ml	Na	Díaz-Sánchez et al. (36)
	Oax10, Ags01	301 ± 34 μg/ml, 3,719 ± 101 μg/ml	Gn	Díaz-Sánchez et al. (36)
	San Fernando	0.67 ± 0.37–3.13 ± 0.33 mg/g DW	Ag	Valiente-Banuet and Gutiérrez-Ochoa (37)
	Cumpas	1.68 ± 0.1–8.73 ± 0.06 mg/g DW	Gr, Phe	Vázquez-Flores et al. (34)
	Sahuaripa	1.41 ± 0.01–8.59 ± 0.04 mg/g DW		
Capsaicinoids	G8	0.56–1.06 mg/g FW	Phe	Morales-Fernández et al. (38)
	Chiltepin (Querétaro, México)	216.22 ± 0.68–1249.28 ± 12.54 mg/g DW	Phe	Fayos et al. (39)

Phe, phenological state variation; Na, natural variation; Ag, variation due to agronomic management; Gn, genotype variation; Gr, variation for geographic origin; Nr, not reported; DW, dry weight; FW, fresh weight; CE, catechin equivalent; QE, quercetin equivalent; mgβCE, β-carotene equivalent.

disadvantage for intensive or industrial use, it represents an opportunity to strengthen local uses, which take advantage of this heterogeneity to increase added value for local consumers.

5 Capsaicinoids

Capsaicinoids (CAPs) are compounds synthesized mainly in the pericarp and placental tissue of chili fruits (*Capsicum*) and are considered within the group of alkaloids. This biosynthesis is highly controlled at a molecular level (54, 55). *Capsicum* plants synthesize them as a defensive mechanism against herbivores and phytopathogens. The CAPs show low polarity and are structurally based on a vanilloid group (an aromatic ring with a hydroxyl and a methyl group), together with a long hydrocarbon chain and the amide group (Figure 1C) (56, 57). The study of CAPs aims to optimize the content of chili fruits, especially the capsaicin responsible for chili's heat and pungency. Effects such as antitumor, antiangiogenic, antineoplastic, thermogenic, and antimicrobial have been related to CAPs (57, 58). It also relates to industrial applications such as functional coatings, clinical applications, food uses, and biotechnological developments (59, 60). Therefore, several investigations address ideal conditions, biosynthesis, and extraction

methods with optimal yields, low prices, high purity, and concentration to meet market demands. In this regard, the accumulation of capsaicinoids in chili fruits depends on the growing conditions, climate, genotype, and agronomic management (61, 62). The biosynthesis of capsaicinoids presents a highly dynamic genetic activity during fruit development, and there is evidence that supports changes in pungency driven during the domestication process in the case of *C. annuum*, being different between wild types and cultivated relatives (63). The accumulation of capsaicinoids in *C. annuum* var. *annuum* has been characterized, indicating that approximately 90 days after anthesis, the fruits reached the highest content. This information highlights the appropriate time for harvesting chili fruits when capsaicinoids are the compounds of interest (31).

For piquin chili, the CAP content evaluations present a wide variation. The content of capsaicin and dihydrocapsaicin under wild conditions is not affected once the fruit has reached its size of commercial interest, being statistically similar between mature and immature fruits (35); however, there is evidence that levels of capsaicinoids could change, the highest level being discovered in mature fruits and when they are cultivated under greenhouse conditions (36). The proportion of capsaicin and dihydrocapsaicin in 16 wild native populations presented similar levels, but there was observed variation in capsaicinoids per population sample (20). Díaz-Sánchez et al. (36)

examined 31 accessions of piquin chili that are grown under greenhouse conditions with similar capsaicin and dihydrocapsaicin ratios. Variations in the total content of capsaicinoids were shown, with the lowest value being 301 ± 34 in the Oax10 accession and the maximum value being $3,719 \pm 101 \mu\text{g/ml}$ in Ags01 (36). During the process of fruit formation and ripening, a high classification of both capsaicinoids has been established, which indicates that these proportions are maintained even when the ripening time varies (39). The number of capsaicinoids could be maintained without variation when the growth conditions are not so drastic or long-lasting as to impact the accumulation of these metabolites in the fruit (64). However, the content of total capsaicinoids has been reported with differences according to the degree of ripening under controlled conditions, with 1.41 and 1.68 mg g^{-1} DW in the immature state and 8.59 – 8.73 mg g^{-1} DW in mature fruits of the Sahuaripa and Cumpas lines, respectively (34). The analysis of ripe fruits of wild piquin chilis from Tamaulipas showed that the contents of total capsaicinoids were statistically different and showed differences among ecotypes in the same municipality (18, 65). The above result suggests that diverse environmental factors may influence the final accumulation of total capsaicinoids more than other factors. Due to this, it is suggested that knowledge of ideal environmental conditions could support the selection of sites with the best characteristics for collecting fruits with the highest CAP contents (65) for diverse purposes, including their biosynthesis, their ecophysiological role, and the promotion of regional or international marketing. Recently, it was demonstrated that the accumulation of CAPs in piquin chili also follows a pattern similar to other chili varieties, which could help in breeding programs to obtain more pungent chilis and to establish times for collecting fruits with optimal contents (66).

All previous studies support the relevance of understanding the impact of growing conditions and genetic components, which influence piquin fruits with specific phytochemical contents. Although some initiatives have been made to stimulate agroforestry and greenhouse production, the yields and qualities are hard to emulate, mainly due to the impact of genetic diversity and multifactorial environmental influences; hence, the more we learn, the closer we get close to improving the management of this species.

6 Volatile compounds

In chili, this category includes a heterogeneous group of chemical compounds (Figures 1D,E) responsible for aroma and partial flavor, which influences the organoleptic perceptions of consumers and criteria for specific uses (67, 68). They are organic, low molecular weight compounds, and volatile at environmental temperatures (69). In *Capsicum* species, several chemical groups have been reported, including aldehydes, organic acids, esters, lipoxygenase cleavage products, nitrogen-containing compounds, hydrocarbons, alcohols, ketones, furans, terpenoids, phenolics, and miscellaneous compounds, highlighting the complexity of volatile compounds in these fruits. However, not all compounds contribute to the aroma, and chemical diversity fluctuates between species, varieties, and phenology (68, 70). Piquin chili is also preferred by consumers due to its aromatic traits, especially in the green stage, with fresh, fruity, and herbal notes, which are associated with its complex chemical array. Piquin chili includes up to 140 compounds, among which esters, alcohols, aldehydes, ketones, terpenes, organic acids, and hydrocarbons were detected (71).

This wild chili shares some volatile compounds responsible for fruity notes with *C. chinense* Jacq. cv. Habanero and *C. frutescens* L. (72, 73). The volatile compounds in piquin chili change during the ripening process, with the high diversity and contents in the green stage decreasing as it matures in the case of esters; however, mature fruit shows slightly higher levels of other compounds. This suggests that flavor and aroma are different, influencing consumer preferences for uses depending on their ripening stages (71).

All this evidence supports the potential of the piquin chili to provide phytochemicals associated with benefits for human health; however, its consumption is driven by cultural issues linked to traditional uses and forms of preparation that influence the intake of these compounds. In this regard, the purpose of the consumption and the traditional use must be considered and linked with all information about the chemical composition in different stages. As mentioned above, the phytochemical groups explored in this study showed great variation as a result of multifactorial influences (genetic background, highly dynamic regulation of biosynthesis, environmental effects, and agronomic issues), which makes it challenging to designate ideal conditions for collecting chili fruits from the wild. Nevertheless, cultivated piquin chili was shown as a better strategy to produce fruits with more stable phytochemical contents to benefit consumers and also with the potential to reduce the extraction pressure in natural populations. Therefore, developing strategies to improve agronomic management to increase the availability of chili with better phytochemical contents derived from optimal growing conditions and versatile genotypes is needed.

7 Conclusion

Since piquin chili is a wild spice obtained from natural populations, and in some cases with incipient agronomic management, holistic studies of the piquin chili should be considered to understand the factors that affect the biosynthesis of the compounds responsible for the organoleptic properties and bioactive components to guarantee the ideal contents for the benefit of the consumer, which will strengthen its trade at a local and international level, in addition to promoting the conservation of this resource. This requires genetic knowledge, agronomic knowledge, and technological improvements focused on the integrative management of piquin. Piquin chili is not only considered for its commercial or nutraceutical value but also for its holistic knowledge relevant to being the ancestor of many cultivated varieties of commercial chili peppers, making it a basis for understanding and improving the production of such varieties. On the other hand, piquin domestication focuses on obtaining lines that satisfy consumers or market requirements in specific ways to reduce extraction pressure on natural populations. However, what highlights its significance is the relationship since its recollection and exploitation as a wild resource, linked to customs, regional uses, and biocultural factors.

Author contributions

RP-R: Conceptualization, Validation, Writing – original draft, Writing – review & editing. YM-R: Validation, Writing – original draft, Writing – review & editing. GR-D-L-C: Writing – original draft, Writing – review & editing. MJ-A: Investigation, Writing – original

draft, Writing – review & editing. CA-M: Investigation, Writing – original draft, Writing – review & editing. NN-G: Writing – original draft, Writing – review & editing. JT-C: Conceptualization, Investigation, Supervision, Writing – original draft, Writing – review & editing.

EL Cielo. The authors would also like to thank Moisés Ramírez Meraz for his guidance on the economic aspect of the piquin chili.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This study was supported by the research budget from the Universidad Autónoma de Tamaulipas—Project: UAT/SIP/INV/2023/058.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Acknowledgments

The authors would like to thank the Project UAT/SIP/INV/2023/058-Prospección del aprovechamiento y conservación de ciertos elementos del componente biótico de la Reserva de la Biosfera

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Abha MK, Thriveni V, Shukla G, Chakravarty S. Nutraceutical potential of tropical wild edible plants of India In: A Kumar, P Singh, S Singh and B Singh, editors. *Wild food plants for zero hunger and resilient agriculture. Plant life and environment dynamics*. Singapore: Springer Nature Singapore (2023). 237–57.
- Bhatt ID, Rawat S, Badhani A, Rawal RS. Nutraceutical potential of selected wild edible fruits of the Indian Himalayan region. *Food Chem.* (2017) 215:84–91. doi: 10.1016/j.foodchem.2016.07.143
- Tran TD, Ogbourne SM, Brooks PR, Sánchez-Cruz N, Medina-Franco JL, Quinn RJ. Lessons from exploring chemical space and chemical diversity of propolis components. *Int J Mol Sci.* (2020) 21:4988. doi: 10.3390/ijms21144988
- Ravivarman K, Mathiventhan U. Proximate and phytochemical analysis of selected wild edible green leafy vegetables in Batticaloa. *J Sci.* (2022) 13:40–50. doi: 10.4038/jsc.v13i2.47
- Pawera L, Khomsan A, Zuhud EA, Hunter D, Ickowitz A, Polesny Z. Wild food plants and trends in their use: from knowledge and perceptions to drivers of change in West Sumatra. *Indonesia Foods.* (2020) 9:1240. doi: 10.3390/foods9091240
- Ray A, Ray R, Sreevidya EA. How many wild edible plants do we eat—their diversity, use, and implications for sustainable food system: an exploratory analysis in India. *Front Sustain Food Syst.* (2020) 4:56. doi: 10.3389/fsufs.2020.00056
- Yeşil Y, Çelik M, Yılmaz B. Wild edible plants in Yeşilli (Mardin-Turkey), a multicultural area. *J Ethnobiol Ethnomed.* (2019) 15:1–19. doi: 10.1186/s13002-019-0327-y
- Nemzer B, Al-Taher F, Abshiru N. Phytochemical composition and nutritional value of different plant parts in two cultivated and wild purslane (*Portulaca oleracea* L.) genotypes. *Food Chem.* (2020) 320:126621. doi: 10.1016/j.foodchem.2020.126621
- Delporte C, Noret N, Vanhaverbeke C, Hardy OJ, Martin JF, Tremblay-Franco M, et al. Does the phytochemical diversity of wild plants like the *Erythrophleum* genus correlate with geographical origin? *Molecules.* (2021) 26:668. doi: 10.3390/molecules26061668
- Prakash OM, Chauhan AS, Kudachikar VB. Traditional uses, nutrition, phytochemistry and various pharmacological properties of Indian wild pear. *Int J Funct Nutr.* (2021) 2:1–13. doi: 10.3892/ijfn.2021.19
- Zeroual A, Sakar EH, Mahjoubi F, Chaouch M, Chaqroune A, Taleb M. Effects of extraction technique and solvent on phytochemicals, antioxidant, and antimicrobial activities of cultivated and wild rosemary (*Rosmarinus officinalis* L.) from Taounate region (northern Morocco). *Biointerface Res Appl Chem.* (2022) 12:8441–52. doi: 10.33263/BRIAC126.84418452
- Karjalainen E, Sarjala T, Raitio H. Promoting human health through forests: overview and major challenges. *Environ Health Prev Med.* (2010) 15:1–8. doi: 10.1007/s12199-008-0069-2
- Krafi KH, Luna-Ruiz JJ, Gepts P. A new collection of wild populations of *Capsicum* in Mexico and the southern United States. *Genet Resour Crop Evol.* (2013) 60:225–32. doi: 10.1007/s10722-012-9827-5
- Hayano-Kanashiro C, Gámez-Meza N, Medina-Juárez LÁ. Wild Pepper *Capsicum annuum* L. var. *glabriusculum*: taxonomy, plant morphology, distribution, genetic diversity, genome sequencing, and phytochemical compounds. *Crop Sci.* (2016) 56:1–11. doi: 10.2135/cropsci2014.11.0789
- Chacón-Hernández JC, Ordaz-Silva S, Mireles-Rodríguez E, Rocandio-Rodríguez M, López-Sánchez IV, Heinz-Castro RTQ, et al. Resistance of wild chili (*Capsicum annuum* L. var. *glabriusculum*) to *Tetranychus merganser* Boudreaux. *Southwest Entomol.* (2020) 45:89–98. doi: 10.3958/059.045.0110
- Torres-Morán MI, Rodríguez-Guzmán E, Velasco-Ramírez AP, Escoto-Delgado M, Rijoas-López ME, Durán-Puga N, et al. Estudio preliminar de identificación a nivel molecular, de ecotipos de Chile piquín. *eCUCBA.* (2022) 9:192–7. doi: 10.32870/ecucba.vi18.254
- Rodríguez-Del Bosque LA. Preferencia del consumidor por el Chile piquín en comparación con otros chiles en el noreste de México. *Rev Chapingo Ser Hortic.* (2005) XI:279–81. doi: 10.5154/r.rchsh.2004.06.029
- Moreno-Ramírez YDR, Martínez-Ávila GC, González-Hernández VA, Castro-López C, Torres-Castillo JA. Free radical-scavenging capacities, phenolics and capsaicinoids in wild piquin chili (*Capsicum annuum* var. *glabriusculum*). *Molecules.* (2018) 23:2655. doi: 10.3390/molecules23102655
- Castellón-Martínez É, Chávez-Servia JL, Carrillo-Rodríguez JC, Vera-Guzmán AM. Preferencias de consumo de chiles (*Capsicum annuum* L.) nativos en los valles centrales de Oaxaca, México. *Rev Fitotec Mex.* (2012) 35:27–35. doi: 10.35196/rfm.2012.Especial_5.27
- Moreno-Ramírez YDR, Hernández-Bautista A, López PA, Vanoye-Eligio V, Torres-Rodríguez ML, Torres-Castillo JA. Variability in the phytochemical contents and free radical-scavenging capacity of *Capsicum annuum* var. *glabriusculum* (wild Piquin chili). *Chem Biodivers.* (2019) 16:e1900381. doi: 10.1002/cbdv.201900381
- Mares-Quiñones MD, Valiente-Banuet JL. Horticultural aspects for the cultivated production of piquin peppers (*Capsicum annuum* L. var. *glabriusculum*)—a review. *HortScience Horts.* (2019) 54:70–5. doi: 10.21273/HORTSCI113451-18
- Meckelmann SW, Riegel DW, van Zonneveld M, Ríos L, Peña K, Mueller-Seitz E, et al. Capsaicinoids, flavonoids, tocopherols, antioxidant capacity and color attributes in 23 native Peruvian chili peppers (*Capsicum* spp.) grown in three different locations. *Eur Food Res Technol.* (2015) 240:273–83. doi: 10.1007/s00217-014-2325-6
- Kantar MB, Anderson JE, Lucht SA, Mercer K, Bernau V, Case KA, et al. Vitamin variation in *Capsicum* spp. provides opportunities to improve nutritional value of human diets. *PLoS One.* (2016) 11:e0161464. doi: 10.1371/journal.pone.0161464
- Sosa-Moguel O, Pino JA, Ayora-Talavera G, Sauri-Duch E, Cuevas-Glory L. Biological activities of volatile extracts from two varieties of habanero pepper (*Capsicum chinense* Jacq.). *Int J Food Prop.* (2017) 20:S3042–51. doi: 10.1080/10942912.2017.1397694
- Treto-Alemán KM, Torres-Castillo JA, Contreras-Toledo AR, Moreno-Ramírez YDR. Enriquecimiento del aceite comestible por compuestos fenólicos y antioxidantes de Chile piquín (*Capsicum annuum* var. *glabriusculum*). *CienciaUAT.* (2021) 15:156–68. doi: 10.29059/cienciauat.v15i2.1459
- Villalón-Mendoza H, Manzanera-Miranda N, Ramírez-Meraz M, Mejorado-Martínez YM, Soto-Ramos JM. Origin and cultural impact of wild Chilili pepper (*Capsicum annuum* L. var. *glabriusculum*) in northeastern Mexico In: F Garza-Oceñas, editor. *Sustainable management of natural resources: diversity, ecology, taxonomy and sociology*. Cham: Springer International Publishing (2023). 143–59.
- Bhaskarachary K, Vemula SR, Gavaravarapu SRM, Joshi AKR. Traditional foods, functional foods and nutraceuticals. *Proc Indian Natn Sci Acad.* (2016) 82:1565–77. doi: 10.16943/ptinsa/2016/48888

28. Gokhale JS, Lele SS, Ananthanarayan L. Indian traditional foods and diets: combining traditional wisdom with modern science of nutraceuticals and functional foods In: SIS Rattan and G Kraur, editors. *Nutrition, food and diet in ageing and longevity. Healthy ageing and longevity*. Cham: Springer (2021). 357–92.
29. Villalón-Mendoza H, Soto-Ramos JM, Ramírez-Meráz M, Medina-Martínez T, Carrillo-Parra A, Garza-Ocañas F, et al. Estudio de procedencias de Chile silvestre “piquín”. *Sociedad Mexicana Recursos Forestales AC*. (2009):103–10.
30. Boesi A. Traditional knowledge of wild food plants in a few Tibetan communities. *J Ethnobiol Ethnomed*. (2014) 10:1–19. doi: 10.1186/1746-4269-10-75
31. Barbero GF, de Aguiar AC, Carrera C, Olachea Á, Ferreira-González M, Martínez J, et al. Evolution of capsaicinoids in Peter pepper (*Capsicum annuum* var. *annuum*) during fruit ripening. *Chem Biodivers*. (2016) 13:1068–75. doi: 10.1002/cbdv.201500503
32. Pola W, Sugaya S, Photchanachai S. Influence of postharvest temperatures on carotenoid biosynthesis and phytochemicals in mature green chili (*Capsicum annuum* L.). *Antioxidants*. (2020) 9:203. doi: 10.3390/antiox9030203
33. De la Cruz-Ricardez D, Lagunes-Espinoza LDC, Ortiz-García CF, Hernández-Nataren E, Soto-Hernández RM, Acosta-Pech RG. Phenology, yield, and phytochemicals of *Capsicum* spp. in response to shading. *Bot Sci*. (2023) 101:865–82. doi: 10.17129/botsci.3234
34. Vazquez-Flores AA, Góngora-Pérez O, Olivas-Orduña I, Muñoz-Bernal OA, Osuna-Avila P, Rodrigo-García J, et al. Phytochemical profile and antioxidant activity of chiltepin chili (*Capsicum annuum* var. *glabriusculum*). *Sonora, Mexico J Food Bioact*. (2020) 11:57–65. doi: 10.31665/JFB.2020.11237
35. De la Cruz-Ricardez DD, Ortiz-García CF, Lagunes-Espinoza LDC, Torres-de la Cruz M, Hernández-Nataren E. Compuestos fenólicos, carotenoides y capsaicinoides en frutos de *Capsicum* spp. de Tabasco, México. *Agrociencia*. (2020) 54:505–19. doi: 10.47163/agrociencia.v54i4.2047
36. Díaz-Sánchez DD, López-Sánchez H, Silva-Rojas HV, Gardea-Béjar AA, Cruz-Huerta N, Ramírez-Ramírez I, et al. Pungency and fruit quality in Mexican landraces of piquín pepper (*Capsicum annuum* var. *glabriusculum*) as affected by plant growth environment and postharvest handling. *Chil J Agric Res*. (2021) 81:546–56. doi: 10.4067/S0718-58392021000400546
37. Valiente-Banuet JJ, Gutiérrez-Ochoa A. Effect of irrigation frequency and shade levels on vegetative growth, yield, and fruit quality of piquín pepper (*Capsicum annuum* L. var. *glabriusculum*). *HortScience*. (2016) 51:573–9. doi: 10.21273/HORTSCI.51.5.573
38. Morales-Fernández SD, Moreno-Velázquez D, Jesús TD, Vázquez-Cruz F, Ibáñez-Martínez A, Tobar-Reyes JR. Phenology and content of capsaicinoids in chili fruits produced under greenhouse conditions. *Rev Mex Cienc Agríc*. (2021) 11:663–75. doi: 10.29312/remexca.v11i3.2159
39. Fayos-Abellán OF, Martínez O, Alejo NO, Savirón M, Orduna J, Claver AG. Evolución del contenido de capsinoides y capsaicinoides durante la maduración de los frutos de “Chiltepin” *C. annuum* L. var. *glabriusculum*. *IX Congreso Mejora Genét Plantas*. (2018) 1:140–3.
40. Dias MC, Pinto DC, Silva AM. Plant flavonoids: chemical characteristics and biological activity. *Molecules*. (2021) 26:5377. doi: 10.3390/molecules26175377
41. Vera-Guzmán AM, Chávez-Servia JL, Carrillo-Rodríguez JC, López MG. Phytochemical evaluation of wild and cultivated pepper (*Capsicum annuum* L. and *C. pubescens* Ruiz & Pav.) from Oaxaca, Mexico. *Chil J Agric Res*. (2011) 71:578–85. doi: 10.4067/S0718-58392011000400013
42. Ovando-Martínez M, Láscara-Meza N, Molina-Domínguez CC, Hayano-Kanashiro C, Medina-Juárez LA. Simulated gastrointestinal digestion, bioaccessibility and antioxidant capacity of polyphenols from red chiltepin (*Capsicum annuum* L. var. *glabriusculum*) grown in Northwest Mexico. *Plant Foods Hum Nutr*. (2018) 73:116–21. doi: 10.1007/s11130-018-0669-y
43. Vera-Guzmán AM, Aquino-Bolaños EN, Heredia-García E, Carrillo-Rodríguez JC, Hernández-Delgado S, Chávez-Servia JL. *Flavonoid and capsaicinoid contents and consumption of Mexican chili pepper (Capsicum annuum L.) landraces. Flavonoids-from biosynthesis to human health*. InTechOpen: London (2017) 405–437
44. De la Cruz-Ricardez D, del Carmen L-EL, Soto-Hernández RM, Hernández-Nataren E, Ortiz-García CF, Acosta-Pech RG. Phytochemical profile of *Capsicum* spp. fruits related to ripeness level, shading and harvest season in the southeast of Mexico. *Chil J Agr Res*. (2024) 84:211–24. doi: 10.4067/s0718-58392024000200211
45. Jucá MM, Cysne-Filho FMS, de Almeida JC, Mesquita DDS, Barriga JRDM, Dias KCF, et al. Flavonoids: biological activities and therapeutic potential. *Nat Prod Res*. (2020) 34:692–705. doi: 10.1080/14786419.2018.1493588
46. Mi S, Zhang X, Wang Y, Zheng M, Zhao J, Gong H, et al. Effect of different genotypes on the fruit volatile profiles, flavonoid composition and antioxidant activities of chili peppers. *Food Chem*. (2022) 374:131751. doi: 10.1016/j.foodchem.2021.131751
47. Saini RK, Prasad P, Lokesh V, Shang X, Shin J, Keum YS, et al. Carotenoids: dietary sources, extraction, encapsulation, bioavailability, and health benefits—a review of recent advancements. *Antioxidants*. (2022) 11:795. doi: 10.3390/antiox11040795
48. Navarro JM, Flores P, Garrido C, Martínez V. Changes in the contents of antioxidant compounds in pepper fruits at different ripening stages, as affected by salinity. *Food Chem*. (2006) 96:66–73. doi: 10.1016/j.foodchem.2005.01.057
49. Bhandari SR, Jung BD, Baek HY, Lee YS. Ripening-dependent changes in phytonutrients and antioxidant activity of red pepper (*Capsicum annuum* L.) fruits cultivated under open-field conditions. *HortScience*. (2013) 48:1275–82. doi: 10.21273/HORTSCI.48.10.1275
50. Kim EH, Lee KM, Lee SY, Kil M, Kwon OH, Lee SG, et al. Influence of genetic and environmental factors on the contents of carotenoids and phenolic acids in red pepper fruits (*Capsicum annuum* L.). *Appl Biol Chem*. (2021) 64:1–11. doi: 10.1186/s13765-021-00657-8
51. Song S, Song SY, Nian P, Lv D, Jing Y, Lu S, et al. Transcriptomic analysis suggests a coordinated regulation of carotenoid metabolism in ripening chili pepper (*Capsicum annuum* var. *conoides*) fruits. *Antioxidants*. (2022) 11:2245. doi: 10.3390/antiox11112245
52. Giuffrida D, Dugo P, Torre G, Bignardi C, Cavazza A, Corradini C, et al. Characterization of 12 *Capsicum* varieties by evaluation of their carotenoid profile and pungency determination. *Food Chem*. (2013) 140:794–802. doi: 10.1016/j.foodchem.2012.09.060
53. Ayob O, Hussain PR, Suradkar P, Naqash F, Rather SA, Joshi S, et al. Evaluation of chemical composition and antioxidant activity of Himalayan red chilli varieties. *LWT*. (2021) 146:111413. doi: 10.1016/j.lwt.2021.111413
54. Zhu Z, Sun B, Cai W, Zhou X, Mao Y, Chen C, et al. Natural variations in the MYB transcription factor MYB31 determine the evolution of extremely pungent peppers. *New Phytol*. (2019) 223:922–38. doi: 10.1111/nph.15853
55. Sun B, Chen C, Song J, Zheng P, Wang J, Wei J, et al. The *Capsicum* MYB31 regulates capsaicinoid biosynthesis in the pepper pericarp. *Plant Physiol Bioch*. (2022) 176:21–30. doi: 10.1016/j.plaphy.2022.02.014
56. Friedman JR, Richbart SD, Merritt JC, Brown KC, Denning KL, Tirona MT, et al. Capsaicinoids: multiple effects on angiogenesis, invasion and metastasis in human cancers. *Biomed Pharmacother*. (2019) 118:109317. doi: 10.1016/j.biopha.2019.109317
57. Batiha GES, Alqahtani A, Ojo OA, Shaheen HM, Wasef L, Elzeiny M, et al. Biological properties, bioactive constituents, and pharmacokinetics of some *Capsicum* spp. and capsaicinoids. *Int J Mol Sci*. (2020) 21:5179. doi: 10.3390/ijms21155179
58. Irandoost P, Yagin NL, Namazi N, Keshtkar A, Farsi F, Alamdari NM, et al. The effect of Capsaicinoids or Capsinoids in red pepper on thermogenesis in healthy adults: a systematic review and meta-analysis. *Phytother Res*. (2021) 35:1358–77. doi: 10.1002/ptr.6897
59. Baenas N, Belović M, Ilic N, Moreno DA, García-Viguera C. Industrial use of pepper (*Capsicum annuum* L.) derived products: technological benefits and biological advantages. *Food Chem*. (2019) 274:872–85. doi: 10.1016/j.foodchem.2018.09.047
60. Rezaazadeh A, Hamishehkar H, Ehsani A, Ghasempour Z, Moghaddas EK. Applications of capsaicin in food industry: functionality, utilization and stabilization. *Crit Rev Food Sci*. (2023) 63:4009–25. doi: 10.1080/10408398.2021.1997904
61. Kim EH, Lee SY, Baek DY, Park SY, Lee SG, Ryu TH, et al. A comparison of the nutrient composition and statistical profile in red pepper fruits (*Capsicum annuum* L.) based on genetic and environmental factors. *Appl Biol Chem*. (2019) 62:1–13. doi: 10.1186/s13765-019-0456-y
62. Tripodi P, Lo Scalzo R, Ficcadenti N. Dissection of heterotic, genotypic and environmental factors influencing the variation of yield components and health-related compounds in chili pepper (*Capsicum annuum*). *Euphytica*. (2020) 216:112. doi: 10.1007/s10681-020-02648-0
63. Zhang B, Hu F, Cai X, Cheng J, Zhang Y, Lin H, et al. Integrative analysis of the metabolome and transcriptome of a cultivated pepper and its wild progenitor Chiltepin (*Capsicum annuum* L. var. *glabriusculum*) revealed the loss of pungency during *Capsicum* domestication. *Front Plant Sci*. (2022) 12:783496. doi: 10.3389/fpls.2021.783496
64. Ricardez-Miranda LE, Lagunes-Espinoza LC, Hernández-Nataren E, Palma-López DJ, Conde-Martínez FV. Water restriction during the vegetative and reproductive stages of *Capsicum annuum* var. *glabriusculum*, and its effect on growth, secondary metabolites and fruit yield. *Sci Hortic-Amsterdam*. (2021) 285:110129. doi: 10.1016/j.scienta.2021.110129
65. Aguirre-Hernández E, San Miguel-Chávez R, Tenango MP, González-Trujano ME, De La Rosa-Manzano E, Sánchez-Ramos G, et al. Capsaicinoids concentration in *Capsicum annuum* var. *glabriusculum* collected in Tamaulipas, Mexico. *Phyton*. (2017) 86:46–52. doi: 10.32604/phyton.2017.86.046
66. Fayos O, Ochoa-Alejo N, De La Vega OM, Savirón M, Orduna J, Mallor C, et al. Assessment of capsaicinoid and capsinoid accumulation patterns during fruit development in three chili pepper genotypes (*Capsicum* spp.) carrying Pun1 and pAMT alleles related to pungency. *J Agric Food Chem*. (2019) 67:12219–27. doi: 10.1021/acs.jafc.9b05332
67. Cirlini M, Luzzini G, Morini E, Folloni S, Ranieri R, Dall’Asta C, et al. Evaluation of the volatile fraction, pungency and extractable color of different Italian *Capsicum annuum* cultivars designed for food industry. *Eur Food Res Technol*. (2019) 245:2669–78. doi: 10.1007/s00217-019-03378-x
68. Zhang J, Wang C, Wang J, Yang Y, Han K, Bakpa EP, et al. Comprehensive fruit quality assessment and identification of aroma-active compounds in green pepper (*Capsicum annuum* L.). *Front Nutr*. (2023) 9:1027605. doi: 10.3389/fnut.2022.1027605
69. Cicolella A. Volatile organic compounds (VOC): definition, classification and properties. *Rev Mal Respir*. (2008) 25:155–63. doi: 10.1016/S0761-8425(08)71513-4

70. Taiti C, Costa C, Migliori CA, Comparini D, Figorilli S, Mancuso S. Correlation between volatile compounds and spiciness in domesticated and wild fresh chili peppers. *Food Bioprocess Technol.* (2019) 12:1366–80. doi: 10.1007/s11947-019-02297-9

71. Forero MD, Quijano CE, Pino JA. Volatile compounds of Chile pepper (*Capsicum annuum* L. var. *glabriusculum*) at two ripening stages. *Flavour Frag J.* (2009) 24:25–30. doi: 10.1002/ffj.1913

72. Pino J, Fuentes V, Barrios O. Volatile constituents of cachucha peppers (*Capsicum chinense* Jacq.) grown in Cuba. *Food Chem.* (2011) 125:860–4. doi: 10.1016/j.foodchem.2010.08.073

73. Bogusz Junior S, Marchi Tavares A, Teixeira Filho J, Alcaraz Zini C, Teixeira Godoy H. Analysis of the volatile compounds of Brazilian chilli peppers (*Capsicum* spp.) at two stages of maturity by solid phase micro-extraction and gas chromatography-mass spectrometry. *Food Res Int.* (2012) 48:98–107. doi: 10.1016/j.foodres.2012.02.005