Check for updates

OPEN ACCESS

EDITED BY Mahta Moussavi, Prairie View A&M University, United States

REVIEWED BY Ye Peng, Jiangsu University, China Oliver Dean John, Université de Lausanne, Switzerland

*CORRESPONDENCE Jing Yang ⊠ yangjing5152@163.com; ⊠ yangjing5152@nuc.edu.cn

RECEIVED 03 October 2023 ACCEPTED 08 December 2023 PUBLISHED 08 January 2024

CITATION

Wang T, Yang J, Huang Z, Wang F, Liu R, Liu Y and Li X (2024) Integrated 16s RNA sequencing and network pharmacology to explore the effects of polyphenol-rich raspberry leaf extract on weight control. *Front. Nutr.* 10:1306037. doi: 10.3389/fnut.2023.1306037

COPYRIGHT

© 2024 Wang, Yang, Huang, Wang, Liu, Liu and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Integrated 16s RNA sequencing and network pharmacology to explore the effects of polyphenol-rich raspberry leaf extract on weight control

Tao Wang^{1,2}, Jing Yang^{1,2}*, Ziang Huang^{1,2}, Fei Wang³, Ruzi Liu⁴, Yongping Liu¹ and Xiaojun Li¹

¹School of Chemistry and Chemical Engineering, North University of China, Taiyuan, Shanxi, China, ²Dezhou Industrial Technology Research Institute of North University of China, Dezhou, Shandong, China, ³The Hospital of North University of China, Taiyuan, Shanxi, China, ⁴Dezhou Yongshengzhai Braised Chicken Group Co., Ltd., Dezhou, Shangdong, China

Introduction: Obesity is recognized as a chronic low-grade inflammation associated with intestinal flora imbalance, leading to dyslipidemia and inflammation. Modern research has found that polyphenols have anti-obesity effects. However, the mechanism of action of raspberry leaf extract (RLE) with high polyphenols in regulating obesity is still unknown. This study investigated the improvement effect of supplementing RLE on high-fat diet (HFD) induced obesity in mice.

Methods: RLE was used to intervene in HFD induced C57BL/6J male mice during prevention stage (1-16 weeks) and treatment stage (17-20 weeks). Their weight changes and obesity-related biochemical indicators were measured. The changes in intestinal flora were analyzed using 16S rRNA sequencing, and finally the targets and pathways of the 7 typical polyphenols (quercetin-3-O-glucuronide, ellagic acid, kaempferol-3-O-rutinoside, chlorogenic acid, brevifolin carboxylic acid, quercetin-3-O-rutinoside, and quercetin) of RLE in the regulation of obesity were predicted by network pharmacology approach.

Results and discussion: The results showed that RLE effectively prevented and treated weight gain in obese mice induced by HFD, alleviated adipocyte hypertrophy, reduced Interleukin-6 and Tumor Necrosis Factor Alpha levels, and improved intestinal flora, especially *Muriaculaceae*, *Alistipes* and *Alloprevotella*, and decreased the *Firmicutes/Bacteroidota* ratio. Network pharmacology analysis selected 60 common targets for 7 RLE polyphenols and obesity. Combined with protein-protein interaction network, enrichment analysis and experimental results, TNF, IL-6, AKT1, and PPAR were predicted as potential key targets for RLE polyphenols.

Conclusion: The potential mechanism by which polyphenol-rich RLE regulates obesity may be attributed to the specific polyphenols of RLE and their synergistic effects, therefore RLE has a great anti-obesity potential and may be used as a means to alleviate obesity and related diseases.

KEYWORDS

obesity, Rubus idaeus, intestinal flora, network pharmacology, IL-6, TNF- α

Introduction

The World Health Organization (WHO) defines both overweight (a body mass index (BMI) \ge 25 kg/m²) and obesity (BMI \ge 30 kg/m²) as an abnormal or excessive fat accumulation (1). Obesity greatly increases the risk of diseases including type 2 diabetes, fatty liver, hyperglycemia and coronary heart disease, as well as reduces quality of life and life expectancy (2, 3). Since 1980, the frequency of overweight and obese individuals has doubled, hence, obesity has become a growing public health problem (1). Although changing lifestyle, adjusting diet, and increasing physical activity should be the best choices for controlling weight, overweight or obese individuals, even lean individuals may prefer to diet pills. Orlistat, naltrexone/ bupropion and liraglutide are commonly weight-loss pills that have received approval in the USA or European Union (4). Unfortunately, orlistat commonly causes increased bloating, and stomach pain (5), while liraglutide is a glucagon-like peptide-1 agonist, and the most common side effect is nausea, and gastrointestinal disorders such as constipation and diarrhea (6). Therefore, safer and more effective methods should be sought for treating obesity.

Increasing evidence suggests that consuming a diet high in polyphenols may be beneficial for antioxidation and anti-obesity (7). The raspberry (Rubus idaeus L.), or Fu Pen Zi in Chinese, is a Rubus plant in the Rosaceae family (8). Unripe raspberry (Rubus Chingii Hu) is a typical Traditional Chinese Medicine (TCM) (9), rich in ellagic acid, flavanols, flavonols, phenolic acids and so on (10, 11). Among them, chingiitannin A exhibits the best inhibitory activity against α amylase and α - glucosidase (12). In Europe and the United States, raspberry is known as the "Fruit of Life." According to the data from the Food and Agriculture Organization of the United Nations (FAO), it is the third largest high-value small berry in the world, with an annual demand of more than 2 million tons (13). Raspberry leaves are also rich in polyphenols, ellagic acid, quercetin and their derivates (14). They were included in Chinese Mmedicine and Food Homologous List in 2002 (15), the Chinese Cosmetic Raw Materials list in 2015 (16), the Scientific Committee of the British Herbal Medicine Association in 1983 (17), and the European Community Herbal Medicine Monograph published by the European Medicines Agency (EMA) in 2012 (18). In our previous studies, it was shown that the total phenolic content (TPC) of raspberry leaf extract (RLE) reached 540.32 ± 8.96 mg g⁻¹, and RLE supplementation reduced body weight and adipose percentage in mice after ingestion (19). In addition, the 16s RNA sequencing results of co-cultivation of RLE with volunteer feces showed that RLE has potential value in maintaining intestinal health (20).

Dysbiosis of the intestinal flora is a common symptom of obesity and other metabolic diseases (21). It has been found that obese mice have more *Firmicutes* and significantly fewer *Bacteroidetes* than lean mice (22). The ratio of *Firmicutes/Bacteroidetes* (*F/B*) is often used as a marker of obesity, with the ratio being greater in obese animals (23) and in obese subjects (24). In the study of the effects of polyphenols on intestinal flora, supplementation with anthocyanin component and metabolite of Saskatoon berry increased the abundance of *Muribaculaceae* (25), as well as supplementation with polyphenol-rich oolong tea increased the abundance of bacteria (*Bacteroides, Alistipes,* and *Alloprevotella*) that can produce short chain fatty acid (SCFA) (26). These polyphenol supplements show great value in improving the intestinal flora.

Traditional herbs and herbal extracts have complex compositions and usually act on multiple targets to achieve therapeutic effects on

diseases (27). The use of network pharmacology analysis can greatly reduce the initial screening cost and systematically explain the therapeutic mechanism of herbal medicines on diseases (28). For example, Wang et al. (29) reported that the anti-obesity mechanism of baicalein may be related to the up-regulation of SLC2A1 and the down-regulation of TNF by network pharmacology prediction and experimental validation. Li et al. (30) also studied the obesity-relieving mechanism of mulberry (*Morus alba* L.) leaves and screened the potential anti-obesity mechanism using network pharmacology, which was verified through experiments. The results showed that the anti-obesity mechanism of mulberry leaves involved inflammation, lipid metabolism, and PI3K/Akt/Bcl-xl signaling pathway.

The purpose of this study is to further investigate the effects of RLE on obese C57BL/6J male mice induced by high-fat diet (HFD). RLE supplementation was conducted during the prevention and treatment stages of obese mice, respectively. The body weight, blood glucose levels, blood lipids, and inflammatory factors of mice were measured, their fat and liver cells were observed, as well as 16 s RNA intestinal flora were sequencing. Meantime, the targets and pathways of 7 typical polyphenols in RLE in obesity regulation were predicted through network pharmacology. The results will further explore the intrinsic mechanism of RLE and its main compounds in controlling body weight.

Materials and methods

Chemicals

Basic diet (12% of energy from fat in the basal diet SWS9102) was provided by Jiangsu Synergetic Pharmaceutical Bioengineering Co., Ltd. (Jiangsu, China). Interleukin-6 (IL-6) and Tumor Necrosis Factor Alpha (TNF- α) commercial reagent kit were provided by Wuhan Purity Bioengineering Co., Ltd. (Wuhan, China). TIANamp Stool NDA Kit was provided by Tiangen Bioengineering Co., Ltd. (Beijing, China). Paraformaldehyde (4%), hematoxylin, eosin and other reagents were provided by Shanghai yuanye Bioengineering Co., Ltd. (Shanghai, China). Raspberry leaf extract (RLE) with a total phenolic content (TPC) exceeding 50% was prepared in the laboratory according by Yang et al. (19).

Animal experiments

Thirty 4-week-old specific pathogen free C57BL/6J male mice were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd. (Beijing, China). All mice were housed under controlled conditions of 23±2°C and a 12h light-dark cycle. The grouping diagram and experimental process were as follows Figure 1 and Table 1, and the entire experiment period lasted for 20 weeks. After one week of adaptation, the mice were randomly divided into three groups: P-NC (n=6, basal diet SWS9102 with 12% energy), P-HFD (P-HFD group, *n* = 18, HFD: SWS9102 + 19.2% lard with 45% energy), and P-HPH (P-HPH group, n = 6, HFD with 0.2% RLE). After 16 weeks, the remaining mice in the P-HFD group were randomly divided into three groups: T-HFD (T-HFD group, n = 4, HFD), T-HPL (T-HPL group, n=4, HFD with 0.1% RLE), and T-HPH (T-HPH group, n=4, HFD with 0.2% RLE). All animal experimental procedures were in accordance with the requirements of the China National Standard for Laboratory Animals. Mouse body weight was



TABLE 1 Mice grouping and food.

Group	P-NC	P-HFD	P-HPH	T-HFD	T-HPL	T-HPH
Stage		Prevention I (1–16 weeks)	Treatment II (17–20 weeks)		
No.	<i>n</i> = 6	<i>n</i> = 18	<i>n</i> = 6	n = 4	n = 4	n = 4
Food	Normal control diet	High-fat diet				
Fat energy supply ratio	12% (SWS9102)	45% (SWS9102 + 19.2% lard)				
RLE	0	0	0.2%	0	0.1%	0.2%

recorded after fasting for 12 h each week. At the end of each stage, mice were euthanized by cervical dislocation after blood collection from the orbital venous plexus, and white adipose tissue, liver and rectal feces collections for subsequent experiments.

Serum biochemical analysis

After fasting for 12h, blood samples were obtained from mice through the orbital venous plexus. The samples were stored overnight at 4° C and then centrifuged at 3000 rpm for 15 min. The supernatant serum was transferred and stored at -20° C for future use. The contents of total cholesterol (TC), total triglycerides (TG), low-density lipoprotein cholesterol (LDL-C), and high-density lipoprotein cholesterol (HDL-C) in the serum were measured using 7,180 Full-automatic biochemical analyzer (Hitachi High-Tech Diagnostics Co., Ltd., Shanghai, China).

Oral glucose tolerance test (OGTT)

After fasting for 16 h, a glucose solution of 2 g/kg body weight was gavaged to mice. Blood samples were collected by cutting off the tail tip at 0, 30, 60, 90, and 120 min after administration. The blood glucose levels were immediately measured using blood glucose meter (Jiangsu Yuyue Medical Equipment and Supply Co., Ltd., Jiangsu, China). The data of the OGTT were visualized using the area under the curve (AUC).

Histopathological analysis

According to Dai's method (31), fresh mouse liver and white adipose tissue were rapidly separated, fixed in 4% paraformaldehyde solution for 24h. Then they were dehydrated in gradient alcohol, embedded in paraffin, sectioned, and stained with hematoxylin and eosin (H&E staining). After sealing with neutral gum, tissue pathological changes were observed by microscopy at 400 \times magnification. The area of adipocytes was measured using Image J 2.0 software.

16s RNA sequencing

The intestinal microbial DNA of each group of mice was extracted using TIANamp Stool NDA Kit. A pair of specific primers, 341F 5'-CTACGGGNGGCWGCAG-3' and 805R 5'-GACTACHVGGG TATCTAATCC-3', were used to amplify the V3-V4 region of the bacterial 16S rRNA gene through PCR. The raw data was sequenced on the Illumina NovaSeq PE250 platform by Shanghai Weihuan Bioengineering Co., Ltd. (Beijing, China). Trimmmomatic PE was used to trim low-quality bases from the original data (32). The analysis process of Qiime 2 (33) was used for filtering, deduplication, base correction, and removal of chimeric sequences. DADA2 (34) was employed to denoise the original data, which was equivalent to clustering at 100% similarity, and only removing and correcting low-quality sequences, identifying chimeras using an algorithm, and directly removing redundant sequences. The obtained ASV/OUT sequences were annotated to obtain corresponding species information and species-based abundance distribution. Visualization and analysis of data using R (Version 3.6.1).

Collection of seven polyphenolic targets

The Traditional Chinese Medicine Systems Pharmacology Database (TCMSP)¹ (35) and the Swiss Target Prediction database²

¹ https://old.tcmsp-e.com/tcmsp.php

² http://swisstargetprediction.ch/

(36) were used to predict potential targets of 7 polyphenols (chlorogenic acid, brevifolin carboxylic acid, ellagic acid, quercetin-3-O-rutinoside, quercetin-3-O-glucuronide (Q3G), kaempferol-3-Orutinoside (K3R), and quercetin). The target obtained from TCMSP were converted to *Homo sapiens* gene symbols using the UniProt database³ (37). The Swiss Target Prediction database was combined with the PubChem database⁴ (38). Canonical SMILES for 7 polyphenols were obtained from PubChem, and then they were imported into Swiss Target Prediction. The species selected "*Homo sapiens*," and a screening condition of "probability">0 was used to screen for prediction of component action targets.

Prediction of targets for obesity

Using "obesity" as a keyword, "Obesity"-associated target genes were predicted through the following 3 databases: GeneCards⁵ (39), Online Mendelian Inheritance in Man (OMIM)⁶ (40), and Therapeutic Target Database (TTD)⁷ (41). The overlapping targets between 7 polyphenol and obesity were considered as potential targets for obesity treatment. A Venn diagram was constructed using Jvenn⁸ (42) for visualization analysis.

Protein-protein interaction analysis

The overlapping target were imported into the STRING database9 (43) to construct a protein-protein interaction (PPI) network. The network and enrichment facilities in STRING comprehensively characterized user gene lists and functional genomics datasets, and created and shared highly customized and enhanced protein-protein association networks (44). The species selected "Homo sapiens" and a minimum interaction score was set to the highest confidence (> 0.400). The PPI network data were imported into Cytoscape 3.9.0 software for visualization, and then the Cytoscape plugin centiscape 2.2 was used to calculate three specific centrality parameters of the PPI network topology structure. The parameters were key indicators for the nodes: "betweenness centrality (BC)," "closeness centrality (CC)," and "degree centrality (DC)" (45). Subsequently, using each parameter as a filtering condition, seven polyphenolic substances regulating obesity were selected as key targets. Finally, each parameter as a filtering condition screened key targets of 7 polyphenols for regulating obesity.

Go and KEGG pathway enrichment analysis

The Metascape database¹⁰ (46) was used molecular function (MF) of Gene Ontology (GO), the enrichment of biological process (BP),

4 https://pubchem.ncbi.nlm.nih.gov/

and cellular component (CC), as well as the pathway analysis of Kyoto Encyclopedia of Genes and Genomes (KEGG). Then, the bubble charts and bar charts for data analysis and visualization were generated by online platform.¹¹

Statistical analysis

The data was represented as mean \pm standard deviation (SD). Statistical analysis was conducted using SPSS 22.0 software. The data was evaluated through one-way analysis of variance (ANOVA) and then followed by Duncan's multiple range test. *p* < 0.05 was defined as statistically significant.

Results

Effects of RLE supplementation on body weight in mice

To investigate the effect of RLE on the body weight of mice fed HFD, the mice were divided into a prevention group (1-16 weeks) and a treatment group (17-20 weeks) at the 16-week mark (Figure 1; Table 1). As shown in Figure 2A, two weeks later, compared to the P-NC group, P-HFD group showed a significant increase in body weight (p < 0.05). Six weeks later, the weight of the P-HPH mice was significantly lower than that of the P-HFD group (p < 0.05). After 16 weeks, the weights of the P-NC, P-HFD, and P-HPH groups were 25.79 ± 0.69 g, 32.01 ± 0.80 g, and 28.97 ± 1.37 g, respectively. Compared with the P-NC group, the P-HPH group only increased by 12% in weight, while the P-HFD group increased by 24%. Subsequently, 12 mice with uniform body weight from P-HFD group were randomly divided into three groups (T-HFD, T-HPL, and T-HPH) for further treatment experiment. Their average weight was 31.82 ± 0.50 g, which was 23% heavier than the P-NC group. After 4 weeks, the body weights in the T-HPL group (28.11±0.55g) and the T-HPH group $(29.48 \pm 0.67 \text{ g})$ were significantly lower than those in the T-HFD group $(33.07 \pm 0.61 \text{ g})$ (p < 0.05) (Figure 2B). Meanwhile, there was no significant difference in food intake between RLE supplementation group and corresponding HFD group (Figures 2C,D). These results indicated that RLE supplementation could significantly reduce the weight gain induced by HFD in the preventive stage, and effectively reduced the body weight of obese mice in the treatment. These effects were not related to dietary intake.

Effects of RLE on glucose tolerance and serum indexes

To investigate the effects of RLE on blood glucose regulation in obese mice, the OGTT test was implemented. As shown in Figure 2E, glucose absorption in the P-HFD group peaked at 30 min and then rapidly declined, while the P-HPH group showed slower absorption, with a similar overall trend to the P-NC group. At the AUC level

³ https://www.uniprot.org/

⁵ https://www.genecards.org/

⁶ https://www.omim.org/

⁷ https://db.idrblab.net/ttd/

⁸ http://jvenn.toulouse.inra.fr/app/index.html

⁹ https://cn.string-db.org/

¹⁰ https://metascape.org

¹¹ https://www.bioinformatics.com.cn/



(Figure 2F), a significant decrease was observed in the P-HPH group compared with the P-HFD group, suggesting that RLE ameliorated the glucose intolerance induced by HFD.

The ameliorative effect of RLE on lipid accumulation induced by HFD was determined by measuring biochemical parameters, TC, TG, HDL-C, and LDL-C levels (Figures 3A–D). In the prevention stage, RLE reduced the HFD-induced increased in TC, TG, and LDL-C levels (Figures 4A–D). In addition, supplementing with RLE reduced the increase in IL-6 and TNF- α levels caused by HFD (Figures 3E,F). In the treatment stage, low-dose RLE significantly decreased TC and LDL-C levels, while high-dose RLE significantly decreased TG and TC levels. Both doses of RLE reduced IL-6 and TNF- α levels during the treatment stage (Figures 4E,F).

Effects of RLE white adipose tissue and liver tissue

In this study, white adipocytes were observed by H&E staining, and their sizes between groups were quantified using ImageJ software at the same scale. As shown in Figure 5A and Supplementary Table S5, the size of the P-HFD group significantly increased by about twice compared with the P-NC group, and the hypertrophic symptom was significantly alleviated in the P-HPH, T-HPL, and T-HPH groups with RLE supplementation. No significant liver fat accumulation was observed in all groups during the preventive stage, while during the treatment stage, T-HFD group continued to be fed HFD and showed significant differences in hepatocyte morphology and size, as well as noticeable lipid accumulation (Figure 5B).

Effect of RLE supplementation on intestinal flora

As shown in Table 2, there was no significant difference in abundance and diversity between the P-HFD group and the P-NC group at the prevention stage (16 weeks) (p > 0.05), while the diversity in the P-HPH group was higher than both P-HFD and P-NC groups (p < 0.05). Similarly, the abundance and diversity indices of the T-HPL group significantly increased compared with the T-HFD group and T-HPH group (p < 0.05). Overall, P-HPH, T-HPL and T-HPH groups showed increases in diversity.

The composition and abundance of the intestinal microbiota in prevention and treatment groups were showed in Figure 6. *Bacteroidota* and *Firmicutes* were the most abundant phylum among 11 bacterial phyla displayed in each group. Compared with the P-NC group, the abundance of *Firmicutes* increased and the abundance of *Bacteroidota* decreased in P-HFD and T-HFD groups, indicating a higher *F/B* ratio. However, RLE supplementation increased the abundance of *Bacteroidota* and decreased the abundance of *Verrucomicrobiota*.

At the genus level, among the top 20 species, *Muribaculaceae*, *Akkermansia*, *Lachnospiraceae_NK4A136_group*, *Clostridia_UCG-014*, *Alistipes*, *Alloprevotella* and *Turicibacter* were the most abundant species. Compared with the corresponding HFD groups, the relative abundance of *Muriaculaceae*, *Alistipes* and *Alloprevotella* increased in all three groups supplemented with RLE. *Muriaculaceae* had the highest relative abundance (P-HPH: 38.6%, T-HPL: 38.7%, and T-HPH: 44.9%), while the relative abundance of *Akkermansia* and *Clostridia_UCG-014* decreased.



FIGURE 3

Effect of RLE in the prevention group on lipid and inflammatory factor levels in mice fed a high-fat diet. (A–F) Serum levels of TC, TG, LDL-C, HDL-C, HDL-G, and TNF- α in mice. #p < 0.05, P-HFD compared with P-LFD group. *p < 0.05, P-HFD compared with P-HPH group.



Effect of RLE in the treatment group on lipid and inflammatory factor levels in mice fed a high-fat diet. (A–F) Serum levels of TC, TG, LDL-C, HDL-C, IL-6, and TNF- α in mice. *p < 0.05, T-HFD compared with T-HPL group. *p < 0.05, T-HFD compared with T-HPL group.

Targets of polyphenols and genes by network pharmacology

Based on multiple studies and our results, polyphenols play a critical role in alleviating obesity from different perspectives.

Therefore, weight control mechanisms of 7 typical polyphenols (chlorogenic acid, brevifolin carboxylic acid, ellagic acid, quercetin-3-O-rutinoside, Q3G, K3R, and quercetin) in RLE were explored through network pharmacology, especially the correlation between the weight control effect of RLE and these polyphenols. For



Histological observation of white adipose tissue and H&E staining of liver tissue in the prevention and treatment groups (400× magnification). (A) White adipose tissue. (B) Liver tissue.

Groups	ACE	Chao	Shannon	Simpson
P-NC	$555.85 \pm 36.19^{\text{b}}$	$555.59 \pm 36.29^{\mathrm{b}}$	$6.09\pm0.10^{\rm d}$	$0.019 \pm 0.000^{\rm e}$
P-HFD	575.70 ± 19.73^{ab}	575.56 ± 19.71^{ab}	$6.19\pm0.08^{\rm d}$	$0.017 \pm 0.000^{\rm e}$
P-HPH	$503.21 \pm 8.70^{\circ}$	$503.08 \pm 8.73^{\circ}$	$6.89\pm0.04^{\rm b}$	$0.088 \pm 0.004^{\rm c}$
T-HFD	$591.59 \pm 19.48^{\rm ab}$	591.34 ± 19.50^{ab}	$6.68\pm0.08^\circ$	$0.029 \pm 0.002^{\rm d}$
T-HPL	604.73 ± 29.87^{a}	$604.72 \pm 29.90^{\rm a}$	$7.66\pm0.02^{\rm a}$	$0.156 \pm 0.007^{\rm a}$
Т-НРН	$513.24 \pm 10.28^{\circ}$	$513.02 \pm 10.18^{\circ}$	$6.93\pm0.01^{\rm b}$	$0.106 \pm 0.003^{\rm b}$

TABLE 2 Supplementary RLE effect on alpha diversity.

Different letters were significantly different at the level of p < 0.05.

polyphenol-related targets, a total of 160 and 269 potential targets were screened out from the TCMSP and Swiss Target Predictiondatabases, respectively. After combining the two databases and removing duplicate target, 270 potential targets for 7 RLE polyphenols were obtained (Supplementary Table S1). For obesityrelated targets, a total of 600 potential targets were identified after removing duplicates from the GeneCards, OMIM, and TTD databases, among which those with eligible relevance scores >5 were recognized as potential targets in the GeneCards database (Supplementary Table S2). Then, the overlapping targets of the seven polyphenols in RLE and obesity were visualized by Veen diagram (Figure 7A). Finally, a total of 60 overlapping targets were obtained as potential therapeutic targets of 7 polyphenols in RLE for the treatment of obesity.

PPI networks and key target genes

The common 60 targets of 7 polyphenols and obesity were imported into the STRING database to construct a PPI network (Figure 7B), which had 60 nodes and 607 edges. The larger the degree value, the larger size and darker red color of the nodes, indicating a higher probability of being a target of polyphenols in RLE for obesity treatment. The PPI network was downloaded and uploaded to Cytoscape 3.9.0 software, the three parameters "BC," "CC," and "DC" were calculated, and the middle values of the three parameters were used as the filtering condition. The three thresholds were "BC" > 51.933, "CC" > 0.009, and "DC" > 20.233, respectively. Nodes that meet these three thresholds were considered as key target. A total of 12 potential key targets for treating obesity with RLE polyphenols



FIGURE 6

Effect of RLE on the composition of intestinal flora in HFD-induced obese mice. (A) Histogram of relative abundance at phylum level. (B) Histogram of relative abundance at genus level.



were AKT1, TP53, TNF, IL-6, VEGFA, PPARG, HIF1A, ESR1, ERBB2, CRP, MAPK1, and PPARA.

results showed that the major pathways included cancer pathways, fluid shear stress and atherosclerosis, and prostate cancer.

Go and KEGG enrichment analysis

The sixty overlapping targets were uploaded to the Metascape database using *Homo sapiens* (species) for GO and KEGG enrichment analysis. A total of 53 items were obtained from the GO enrichment, all with p < 0.05 (Supplementary Table S3). BP, CC, and MF were 20, 14 and 19 items, respectively, and then BP, CC and MF genes were visualized (Figure 8B). The results showed that hormone to response, concave and signaling receptor activator activity were the most significant. Moreover, KEGG enrichment analysis yielded 18 signaling pathways with p < 0.05 (Supplementary Table S4), and they were plotted by bubble maps for visualization and analysis (Figure 8A). The

Discussion

Obesity is one of the major public health problems worldwide, and long-term intake of HFD can lead to fat accumulation, metabolic disorders, and an increased risk of obesity (3). Compared to drugs that can produce certain side effects, dietary intervention with natural diets rich in polyphenols can be a promising means of treating obesity. These bioactive compounds in dietary diets can act as antioxidants and anti-inflammatory agents, which is beneficial for weight loss and alleviation of metabolic disorders (7). Raspberry leaf, a by-products with huge yield (> 1 million tons/per year) has high health care and medicinal values in our previous studies (13, 14). It has been reported



that raspberry leaf containing quercetin, ellagic acid, chlorogenic acid, quercetin-3-O-rutinoside, and Q3G polyphenols exhibit good antioxidant activity (19, 47). In this study, the TPC of the RLE was more than 50% and the most important phenolic compounds of RLE were Q3G, K3R and ellagic acid. RLE significantly reduced the body weight of HFD-induced obese mice in both prevention and treatment phases, but the obesity-relieving effects of RLE and possible underlying mechanisms remain limited.

Obesity also is recognized as a chronic low-grade inflammation. Higher levels of IL-1β and IL-6 are characteristics of chronic low-grade inflammation (26). Studies have shown that polyphenols and extracts rich in polyphenols have beneficial effects on obesity-related inflammation. This is not only related to high polyphenol content, but also involves the synergistic effects between polyphenol molecules in regulating obesity and related complications (7). For example, Q3G inhibits the production of IL-6 and TNF- α (48), as well as quercetin inhibits the inflammatory factors IL-1 β and IL-6, and stimulates the secretion of the anti-inflammatory factor (49). Quercetin-3-Orutinoside reduces glucose uptake by modulating the PPAR-y signaling pathway (50). Chlorogenic acid also weakens disordered lipid metabolism through PPAR- α (51). In the study of obesityrelated signaling pathways, AKT regulated energy homeostasis by maintaining the level of ATP in cells and initiated lipolysis (52), while PPAR was found to be an adipose gene sensor with fat-burning, lipid metabolism and inflammatory response effects (53). This suggests that the significant reduction of IL-6 and TNF- α inflammation levels in HFD-induced obese mice by RLE may be due to its abundance of polyphenolic substances. In this study, although the interaction analysis of the 7 main phenolic compounds in RLE and obesity through network pharmacology had predicted 12 potential targets, based on experimental results and literature reports, it was speculated that IL-6, TNF, AKT1, and PPAR may be key targets for RLE polyphenols in regulating obesity.

The composition and function of the intestinal flora are thought to be associated with obesity, which can be alleviated by regulating the intestinal flora (54). In this study, diversity is increased and increase the F/B ratio in both RLE-supplemented groups compared with the corresponding HFD group. This indicate that RLE supplementation has improved the diversity of intestinal flora in obese mice during prevention and treatment stages. At phylum level, Verrucomicrobiota abundance decreased in all three groups supplemented with RLE. Study shows that the intestinal flora of patients with inflammatory bowel disease (IBD) reveals high levels of Verrucomicrobiota in the feces of IBD patients (55). At the genus level, RLE supplementation can up-regulate Muriaculaceae, Alistipes and Alloprevotella abundance, while the relative abundance of Akkermansia and Clostridia_UCG-014 decreased. Genomic analysis reveal that Muribaculaceae is functionally diverse in the degradation of complex carbohydrates, and this species has also been implicated in the regulation of obesity, with low abundance in obesity models (25, 56), this is consistent with our findings. It has been shown that Muriaculaceae, Alistipes and Alloprevotella are associated with colitis and are negatively correlated with inflammation (57). Lachnospiracea_NK4A136_Group, Alistipes, and Alloprevotella have been reported to be producers of SCFAs, which plays an active role in the prevention and treatment of obesity (58, 59). It is worth mentioning that Akkermansia muciniphila is generally considered a beneficial bacterium and it can reduce fat storage and alleviate lipid metabolism disorders (60). In animal models, administration of polyphenols, fructo oligosaccharides, oat bran, and whole grain barley increased the abundance of Akkermansia, which proved to be lower in mice fed HFD and all had obesity or type II diabetes-like symptoms. However, opposite results were found in some rat studies, an increase in the abundance of Akkermansia was observed in HFD rats supplemented with barley, while a decrease in the abundance of Akkermansia was observed in NC diet supplemented with barley. These were consistent with our experimental results. The different result may be the result of environmental and genetic factors (61). In addition, Clostridia_UCG-014 abundance was higher in the obesity model. Overall, RLE can alleviate HFD-induced obesity by improving intestinal flora.

Conclusion

In this study, the effects of RLE supplementation on HFD-induced obese mice carefully evaluated. During the prevention and treatment stages, RLE alleviated weight gain, fat accumulation, lipid disorders, inflammatory levels, and improved intestinal flora in HFD-induced obese mice. The potential mechanism may be attributed to the specific polyphenols of RLE and their synergistic effects on IL-6, TNF, AKT1 and PPAR pathways. RLE has a great anti-obesity potential and may be used as a means to alleviate obesity and related diseases. It may also promote the application of raspberry leaves or raspberry leaf extracts, and the development of related products.

Data availability statement

The datasets presented in this study can be found in online repositories. This data can be found here: Sequence Read Archive (SRA) database is BioProject ID: PRJNA1025257 (https://www.ncbi. nlm.nih.gov/bioproject/?term=PRJNA1025257).

Ethics statement

The animal study was approved by Committee on Biological and Medical Ethics, North University of China. The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

TW: Data curation, Formal analysis, Investigation, Software, Visualization, Writing – original draft. JY: Funding acquisition, Methodology, Supervision, Validation, Visualization, Writing – review & editing. ZH: Data curation, Formal analysis, Validation, Writing – original draft. FW: Data curation, Visualization, Writing – original draft. RL: Data curation, Investigation, Writing – original draft. YL: Data curation, Formal analysis, Writing – review & editing. XL: Data curation, Writing – review & editing.

References

1. Chooi YC, Ding C, Magkos F. The epidemiology of obesity. *Metabolism.* (2019) 92:6–10. doi: 10.1016/j.metabol.2018.09.005

2. Blüher M. Obesity: global epidemiology and pathogenesis. *Nat Rev Endocrinol.* (2019) 15:288–98. doi: 10.1038/s41574-019-0176-8

3. Chu H, Du C, Yang Y, Feng X, Zhu L, Chen J, et al. MC-LR aggravates liver lipid metabolism disorders in obese mice fed a high-fat diet via PI3K/AKT/mTOR/SREBP1 signaling pathway. *Toxins*. (2022) 14:833. doi: 10.3390/toxins14120833

4. Bray GA, Frühbeck G, Ryan DH, Wilding JP. Management of obesity. Lancet. (2016) 387:1947–56. doi: 10.1016/S0140-6736(16)00271-3

5. Drew BS, Dixon AF, Dixon JB. Obesity management: update on orlistat. Vasc Health Risk Manag. (2007) 3:817–21. doi: 10.2147/vhrm.s12187420

6. Alruwaili H, Dehestani B, le Roux CW. Clinical impact of liraglutide as a treatment of obesity. *Clin Pharmacol Adv Appl.* (2021) 13:53–60. doi: 10.2147/CPAA. S276085

7. Boccellino M, D'Angelo S. Anti-obesity effects of polyphenol intake: current status and future possibilities. *Int J Mol Sci.* (2020) 21:5642. doi: 10.3390/ijms21165642

8. Editorial Committee of Flora of China. *Flora Republicae Popularis Sinicae*, vol. 37. Beijing: Science Press (1985). 55 p.

9. Chinese Pharmacopoeia Commission. *Pharmacopoeia of the People's Republic of China 2020*. Beijing: China Medical Science Press (2020).

10. He B, Dai L, Jin L, Liu Y, Li X, Luo M, et al. Bioactive components, pharmacological effects, and drug development of traditional herbal medicine *Rubus chingii* Hu (Fu-pen-Zi). *Front Nutr.* (2023) 9:1052504. doi: 10.3389/fnut.2022.1052504

11. Yu G, Luo Z, Wang W, Li Y, Zhou Y, Shi Y. *Rubus chingii* Hu: a review of the phytochemistry and pharmacology. *Front Pharmacol.* (2019) 10:799. doi: 10.3389/ fphar.2019.00799

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This research was supported by Natural Science Foundation of Shanxi Province (202203021221109), Natural Science Foundation of Shandong Province (ZR2022MC118), and Research Project Supported by Shanxi Scholarship Council of China (2020-101).

Conflict of interest

RL was employed by Dezhou Yongshengzhai Braised Chicken Group Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2023.1306037/ full#supplementary-material

12. Chen Y, Chen Z, Guo Q, Gao X, Ma Q, Xue Z, et al. Identification of ellagitannins in the unripe fruit of *Rubus Chingii* Hu and evaluation of its potential antidiabetic activity. *J Agric Food Chem.* (2019) 67:7025–39. doi: 10.1021/acs.jafc.9b02293

13. FAOSTAT. (2022). Crops and livestock products. Available at: http://www.fao.org/faostat/en/?#data/QC (Accessed December 20, 2015).

14. Wu L, Yang J, Wang C, Li N, Liu Y, Duan A, et al. Chemical compositions of raspberry leaves influenced by growth season, cultivars and leaf position. *Sci Hortic*. (2022) 304:111349. doi: 10.1016/j.scienta.2022.111349

15. National Health Commission of the People's Republic of China. (2002). Notice from the Ministry of Health on further standardizing the management of health food raw materials. Available at: http://www.nhc.gov.cn/sps/s3593/200810/ bc239ea3d226449b86379f645dfd881d.shtml (Accessed March 11, 2002, No. 51).

16. National Medical Products Administration. (2015). Notice of the State Food and Drug Administration on the release of the catalog of names of used cosmetic raw materials (No. 105). Available at: https://www.nmpa.gov.cn/hzhp/hzhpfgwj/hzhpgzwj/20151223120001361.html (Accessed December 23, 2015).

17. Scientific Committee of the British Herbal Medicine Association. (1983). British Herbal Medicine Association: Bournemouth. Available at: https://bhma.info/product/british-herbal-pharmacopoeia-1983/ (2015, ebook version).

18. Committee on Herbal Medicinal Products (HMPC). Community herbal monograph on Rubus idaeus L., folium. London: European Medicines Agency (2012).

19. Yang J, Cui J, Han H, Chen J, Yao J, Liu Y. Determination of active compounds in raspberry leaf extracts and the effects of extract intake on mice. *Food Sci Technol.* (2019) 40:124–31. doi: 10.1590/fst.35518

20. Yang J, Hao Y, Li N, Wang C, Liu Y. Metabolic and microbial modulation of phenolic compounds from raspberry leaf extract under *in vitro* digestion and fermentation. *Int J Food Sci Technol.* (2021) 56:5168–77. doi: 10.1111/ijfs.15083

21. Aron-Wisnewsky J, Clément K, Nieuwdorp M. Fecal microbiota transplantation: a future therapeutic option for obesity/diabetes? *Curr Diabetes Rep.* (2019) 19:51–9. doi: 10.1007/s11892-019-1180-z

22. Kang Y, Cai Y. Gut microbiota and obesity: implications for fecal microbiota transplantation therapy. *Hormones.* (2017) 16:223–34. doi: 10.1007/BF03401517

23. Magne F, Gotteland M, Gauthier L, Zazueta A, Pesoa S, Navarrete P, et al. The *firmicutes/bacteroidetes* ratio: a relevant marker of gut dysbiosis in obese patients? *Nutrients*. (2020) 12:1474. doi: 10.3390/nu12051474

24. Stojanov S, Berlec A, Štrukelj B. The influence of probiotics on the *firmicutes/bacteroidetes* ratio in the treatment of obesity and inflammatory bowel disease. *Microorganisms*. (2020) 8:1715. doi: 10.3390/microorganisms8111715

25. Zhao R, Shen GX. Impact of anthocyanin component and metabolite of Saskatoon berry on gut microbiome and relationship with fecal short chain fatty acids in diet-induced insulin resistant mice. *J Nutr Biochem.* (2023) 111:109201. doi: 10.1016/j. jnutbio.2022.109201

26. Li A, Wang J, Kou R, Chen M, Zhang B, Zhang Y, et al. Polyphenol-rich oolong tea alleviates obesity and modulates gut microbiota in high-fat diet-fed mice. *Front Nutr.* (2022) 9:937279. doi: 10.3389/fnut.2022.937279

27. Mao T, Zhang J, Qiao Y, Liu B, Zhang S. Uncovering synergistic mechanism of Chinese herbal medicine in the treatment of atrial fibrillation with obstructive sleep apnea hypopnea syndrome by network pharmacology. *Evid Based Complement Altern Med.* (2019) 2019:1–13. doi: 10.1155/2019/8691608

28. Jiao X, Jin X, Ma Y, Yang Y, Li J, Liang L, et al. A comprehensive application: molecular docking and network pharmacology for the prediction of bioactive constituents and elucidation of mechanisms of action in component-based Chinese medicine. *Comput Biol Chem.* (2021) 90:107402. doi: 10.1016/j. compbiolchem.2020.107402

29. Wang Z, Jiang Z, Xiao P, Jiang Y, Liu W, Liu E. The mechanisms of baicalin ameliorate obesity and hyperlipidemia through a network pharmacology approach. *Eur J Pharmacol.* (2020) 878:173103. doi: 10.1016/j.ejphar.2020.173103

30. Li R, Wang C, Chen Y, Li N, Wang Q, Zhang M, et al. A combined network pharmacology and molecular biology approach to investigate the active ingredients and potential mechanisms of mulberry (*Morus alba* L.) leaf on obesity. *Phytomedicine*. (2021) 92:153714. doi: 10.1016/j.phymed.2021.153714

31. Dai K, Song Y, Zhang D, Wei Y, Jiang S, Xu F, et al. Thinned peach polyphenols alleviate obesity in high fat mice by affecting gut microbiota. *Food Res Int.* (2022) 157:111255. doi: 10.1016/j.foodres.2022.111255

32. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics*. (2014) 30:2114–20. doi: 10.1093/bioinformatics/btu170

33. Bolyen E, Rideout JR, Dillon MR, Bokulich NA, Abnet CC, Al-Ghalith GA, et al. Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. *Nat Biotechnol.* (2019) 37:852–7. doi: 10.1038/s41587-019-0209-9

34. Callahan B, McMurdie P, Rosen M, Han A, Johnson A, Holmes S. DADA2: high-resolution sample inference from Illumina amplicon data. *Nat Methods*. (2016) 13:581–3. doi: 10.1038/nmeth.3869

35. Ru J, Li P, Wang J, Zhou W, Li B, Huang C, et al. TCMSP: a database of systems pharmacology for drug discovery from herbal medicines. *J Cheminformatics*. (2014) 6:1–6. doi: 10.1186/1758-2946-6-13

36. Daina A, Michielin O, Zoete V. SwissTargetPrediction: updated data and new features for efficient prediction of protein targets of small molecules. *Nucleic Acids Res.* (2019) 47:W357-64. doi: 10.1093/nar/gkz382

37. UniProt Consortium. UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res. (2019) 47:D506–15. doi: 10.1093/nar/gky1049

38. Kim S, Chen J, Cheng T, Gindulyte A, He J, He S, et al. PubChem 2019 update: improved access to chemical data. *Nucleic Acids Res.* (2019) 47:D1102–9. doi: 10.1093/nar/gky1033

39. Safran M, Dalah I, Alexander J, Rosen N, Iny Stein T, Shmoish M, et al. GeneCards version 3: the human gene integrator. *Database*. (2010) 2010:20. doi: 10.1093/database/baq020

40. Hamosh A, Scott AF, Amberger JS, Bocchini CA, McKusick VA. Online mendelian inheritance in man (OMIM), a knowledgebase of human genes and genetic disorders. *Nucleic Acids Res.* (2005) 33:D514–7. doi: 10.1093/nar/gki033

41. Chen X, Ji ZL, Chen YZ. TTD: Therapeutic target database. *Nucleic Acids Res.* (2002) 30:412–5. doi: 10.1093/nar/30.1.412

42. Bardou P, Mariette J, Escudié F, Djemiel C, Klopp C. jvenn: An interactive Venn diagram viewer. *BMC Bioinformatics*. (2014) 15:1–7. doi: 10.1186/1471-2105-15-293

43. Mering CV, Huynen M, Jaeggi D, Schmidt S, Bork P, Snel B. STRING: a database of predicted functional associations between proteins. *Nucleic Acids Res.* (2003) 31:258–61. doi: 10.1093/nar/gkg034

44. Szklarczyk D, Gable AL, Nastou KC, Lyon D, Kirsch R, Pyysalo S, et al. The STRING database in 2021: customizable protein–protein networks, and functional characterization of user-uploaded gene/measurement sets. *Nucleic Acids Res.* (2021) 49:D605–12. doi: 10.1093/nar/gkaa1074

45. Scardoni G, Petterlini M, Laudanna C. Analyzing biological network parameters with CentiScaPe. *Bioinformatics*. (2009) 25:2857–9. doi: 10.1093/bioinformatics/btp517

46. Zhou Y, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, et al. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat Commun.* (2019) 10:1523. doi: 10.1038/s41467-019-09234-6

47. Lopez-Corona AV, Valencia-Espinosa I, González-Sánchez FA, Sánchez-López AL, Garcia-Amezquita LE, Garcia-Varela R. Antioxidant, anti-inflammatory and cytotoxic activity of phenolic compound family extracted from raspberries (*Rubus idaeus*): a general review. *Antioxidants.* (2022) 11:1192. doi: 10.3390/antiox11061192

48. Guo X, Zhang D, Gao X, Parry J, Liu K, Liu B, et al. Quercetin and quercetin-3-Oglucuronide are equally effective in ameliorating endothelial insulin resistance through inhibition of reactive oxygen species-associated inflammation. *Mol Nutr Food Res.* (2013) 57:1037–45. doi: 10.1002/mnfr.201200569

49. Seo M, Lee Y, Hwang J, Kim K, Lee B. The inhibitory effects of quercetin on obesity and obesity-induced inflammation by regulation of MAPK signaling. *J Nutr Biochem.* (2015) 26:1308–16. doi: 10.1016/j.jnutbio.2015.06.005

50. Ganjayi MS, Karunakaran RS, Gandham S, Meriga B. Quercetin-3-O-rutinoside from *moringa oleifera* downregulates adipogenesis and lipid accumulation and improves glucose uptake by activation of AMPK/Glut-4 in 3T3-L1 cells. *Rev Bras.* (2023) 33:334–43. doi: 10.1007/s43450-022-00352-9

51. Jin S, Chang C, Zhang L, Liu Y, Huang X, Chen Z. Chlorogenic acid improves late diabetes through adiponectin receptor signaling pathways in *db/db* mice. *PLoS One.* (2015) 10:e0120842. doi: 10.1371/journal.pone.0120842

52. Faes S, Dormond O. PI3K and AKT: unfaithful partners in cancer. *Int J Mol Sci.* (2015) 16:21138–52. doi: 10.3390/ijms160921138

53. Evans RM, Barish GD, Wang Y-X. PPARs and the complex journey to obesity. *Nat Med.* (2004) 10:355–61. doi: 10.1038/nm1025

54. Abenavoli L, Scarpellini E, Colica C, Boccuto L, Salehi B, Sharifi-Rad J, et al. Gut microbiota and obesity: a role for probiotics. *Nutrients*. (2019) 11:2690. doi: 10.3390/ nu11112690

55. Bamola VD, Ghosh A, Kapardar RK, Lal B, Cheema S, Sarma P, et al. Gut microbial diversity in health and disease: experience of healthy Indian subjects, and colon carcinoma and inflammatory bowel disease patients. *Microb Ecol Health Dis.* (2017) 28:1322447. doi: 10.1080/16512235.2017.1322447

56. Lagkouvardos I, Lesker TR, Hitch TC, Gálvez EJ, Smit N, Neuhaus K, et al. Sequence and cultivation study of *Muribaculaceae* reveals novel species, host preference, and functional potential of this yet undescribed family. *Microbiome*. (2019) 7:28–15. doi: 10.1186/s40168-019-0637-2

57. Wang H, Huang J, Ding Y, Zhou J, Gao G, Han H, et al. Nanoparticles isolated from porcine bone soup ameliorated dextran sulfate sodium-induced colitis and regulated gut microbiota in mice. *Front Nutr.* (2022) 9:821404. doi: 10.3389/fnut.2022.821404

58. He G, Chen T, Huang L, Zhang Y, Feng Y, Qu S, et al. *Tremella fuciformis* polysaccharide reduces obesity in high-fat diet-fed mice by modulation of gut microbiota. *Front Microbiol.* (2022) 13:13. doi: 10.3389/fmicb.2022.1073350

59. Li AL, Ni WW, Zhang QM, Li Y, Zhang X, Wu HY, et al. Effect of cinnamon essential oil on gut microbiota in the mouse model of dextran sodium sulfate-induced colitis. *Microbiol Immunol.* (2020) 64:23–32. doi: 10.1111/1348-0421.12749

60. Zheng X, Huang W, Li Q, Chen Y, Wu L, Dong Y, et al. Membrane protein Amuc_1100 derived from *Akkermansia muciniphila* facilitates lipolysis and Browning via activating the AC3/PKA/HSL pathway. *Microbiol Spectr.* (2023) 11:e04323–2. doi: 10.1128/spectrum.04323-22

61. Derrien M, Belzer C, de Vos WM. Akkermansia muciniphila and its role in regulating host functions. *Microb Pathog.* (2017) 106:171–81. doi: 10.1016/j. micpath.2016.02.005