Check for updates

#### **OPEN ACCESS**

EDITED BY Gislane Lelis Vilela de Oliveira, São Paulo State University, Brazil

REVIEWED BY Claudia Cantoni, Barrow Neurological Institute (BNI), United States Paolo Riccio, University of Basilicata, Italy Leandro Pires Araujo, Columbia University, United States

\*CORRESPONDENCE Javier Ochoa-Repáraz ⊠ jochoareparaz@boisestate.edu

SPECIALTY SECTION This article was submitted to Nutritional Immunology, a section of the journal Frontiers in Nutrition

RECEIVED 17 January 2023 ACCEPTED 14 March 2023 PUBLISHED 29 March 2023

#### CITATION

Hoffman K, Doyle WJ, Schumacher SM and Ochoa-Repáraz J (2023) Gut microbiome-modulated dietary strategies in EAE and multiple sclerosis. *Front. Nutr.* 10:1146748. doi: 10.3389/fnut.2023.1146748

#### COPYRIGHT

© 2023 Hoffman, Doyle, Schumacher and Ochoa-Repáraz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Gut microbiome-modulated dietary strategies in EAE and multiple sclerosis

Kristina Hoffman, William J. Doyle, Sean M. Schumacher and Javier Ochoa-Repáraz\*

Department of Biological Sciences, Boise State University, Boise, ID, United States

Over the last few decades, the incidence of multiple sclerosis has increased as society's dietary habits have switched from a whole foods approach to a high fat, high salt, low dietary fiber, and processed food diet, termed the "Western diet." Environmental factors, such as diet, could play a role in the pathogenesis of multiple sclerosis due to gut microbiota alterations, gut barrier leakage, and subsequent intestinal inflammation that could lead to exacerbated neuroinflammation. This mini-review explores the gut microbiome alterations of various dietary strategies that improve upon the "Western diet" as promising alternatives and targets to current multiple sclerosis treatments. We also provide evidence that gut microbiome modulation through diet can improve or exacerbate clinical symptoms of multiple sclerosis, highlighting the importance of including gut microbiome analyses in future studies of diet and disease.

#### KEYWORDS

diet, gut dysbiosis, multiple sclerosis, EAE, dietary factors, dietary interventions, gut microbiome

# 1. Introduction

Multiple sclerosis (MS) is an autoimmune and neurodegenerative disease that affects the central nervous system (CNS). The estimated number of people with MS in 2020 was 2.8 million worldwide, with a global prevalence of 35.9 per 100,000 people (1). At the initial stages of the disease, T- and B-cell lymphocytes infiltrate into the parenchyma of the CNS, creating inflammatory and demyelination lesions mostly in the white matter but also in the gray matter. As the disease progresses, resident microglia and macrophages play an increasing role in the demyelination process (2). MS symptoms are highly variable and include visual interference, muscle weakness, and fatigue, which ultimately can lead to paralysis. MS disease onset and progression mechanisms are still under investigation due to many confounding factors, including genetics and environmental factors, such as viral infections (specifically Epstein Barr Virus), vitamin D deficiency, and diet. The incidence of MS has increased over the last few decades as people's dietary habits have changed from a whole-food lifestyle to the "Western diet," where people consume more processed foods with low dietary fiber, high fat, and high salt (3). The "Western diet" alteration of the gut microbiome is thought to play a role in the pathogenesis of MS and other autoimmune diseases (3). Intestinal inflammation through gut microbiota alterations could lead to gut barrier leakage and systemic inflammation (4). This intestinal disruption, associated with

10.3389/fnut.2023.1146748

blood-brain barrier disruption, could lead to enhanced neuroinflammation (5, 6). Advancing our understanding of gut microbiome-modulated dietary strategies will contribute to the pathophysiology of MS. In this mini-review, we discuss the gut microbiome alterations with current dietary strategies that are not as widely studied in MS.

# 2. Gut dysbiosis and its role in EAE and MS

Gut microbes are closely co-evolved microbial partners interacting with the host and other microbial commensals. Gut microbes' physiology and metabolism are exposed to environmental changes generated by the host's genetics, immune system, and physiology (7), such as temperature, pH, oxygen, and others, and external modulators, such as diet and drugs. Alteration of this complex system by dietary factors could lead to inflammation and potential disease in gut dysbiosis. In this scenario, dietary factors could lead to dysbiosis, driving intestinal pro-inflammatory responses and subsequently leading to disease or exacerbation of the disease. Gut dysbiosis should then be considered when approaching auto-inflammatory conditions (8). Gut dysbiosis in the context of MS is supported by clinical evidence that indicates significant alterations in the microbiome composition of fecal content isolated from MS patients when compared with healthy individuals, as we recently reviewed (9, 10). Studying the gut microbial communities and their interactions is a complex task, as most taxa are not culturable in the lab. These technical limitations require the use of in vivo experimental models and sample collection studies based on genetic identification. A widely used animal model to study MS is experimental autoimmune encephalomyelitis (EAE). Despite its limitations, EAE is the most commonly used animal model for the study of the disease (11). EAE can be induced in multiple non-rodent and rodent species actively by injecting myelin oligodendrocyte glycoprotein (MOG) or other myelin selfantigens followed by pertussis toxin and passively by the adoptive transfer of encephalitogenic T-cells (12). EAE induction triggers an autoimmune response resulting in the proliferation and activation of pathogenic immune cells that cross the blood-brain barrier (BBB) and their infiltration into the CNS, leading to myelin sheath degradation by mechanisms that still need to be completely elucidated (13). EAE experiments have provided evidence that CNS inflammatory demyelination is linked to the gut microbiome by showing how antibiotic treatments impact disease onset and severity (14-18). Studies in germ-free (GF) mice showed that the lack of microbes results in reduced EAE severity (19, 20) and that the monocolonization of Th17-driving gut bacteria, such as segmented filamentous bacterium, restores the disease's severity (19, 20). In addition, the reconstitution of GF mice with MS patient fecal content exacerbated EAE (21, 22). Creating detailed human microbiota profiles is a necessary step in translating the findings of EAE models (23).

Significant changes in gut microbiome composition have also been observed in both EAE models and MS clinical studies vs. healthy controls (HC) [for a discussion, see our latest reviews (9, 10)]. We recently summarized these studies in a different work (9). For the scope of this mini-review, we will discuss a few notable microbe alterations across EAE and MS studies to provide a base for dietary strategy comparison in the following sections and Table 1. In EAE studies, significant reductions in the relative abundances of microbes associated with antiinflammatory responses were observed, including Lactobacillaceae (24, 25) and Bacteroidaceae (26). Inversely, there were increases in the abundances of Clostridiaceae (24, 25) and Akkermansia (24, 26). In MS vs. healthy control (HC) comparison studies, there were decreases in anti-inflammatory microbes Bacteroidaceae (27-30), and increases in pro-inflammatory microbes Methanobrevibacter (28, 31), Acinetobacter (21, 27, 32), and Akkermansia (21, 22, 30-32). Targeting specific microbes, their metabolites, or the pathways they play a role in for protection with dietary alterations is a potential novel avenue for treatment. For example, current research into the ratio of Firmicutes to Bacteroidetes (F/B) has shown it as a biomarker for dysbiosis and a potential biomarker of disease (33-35). Changes to this ratio are of high interest due to both bacterial phyla composing about 90% of the gut microbiome (35). High F/B has been associated with obesity, while low F/B with inflammatory bowel disease. However, there have been conflicting results, and many confounding factors are not considered, such as age, environmental conditions, and diet (33-35). In the following sections, we will briefly discuss various current dietary strategies and their impact on the gut microbiome in MS and EAE.

## 3. Dietary factors in EAE and MS

In this section, we discuss the dietary factors shown to regulate experimental disease in the EAE model and potential relevance to human disease. Common components of daily dietary habits, such as salt and zinc, are known to regulate EAE severity. The effects of high concentrations of sodium chloride seem to be triggered by the induction of Th17 cell-mediated proinflammation (36–38). Dietary sodium was also shown to exacerbate EAE (39). By contrast, when combined with aspartate, the treatment with zinc and salts reduced disease severity (40). Prophylactic treatment with zinc dissolved in drinking water also reduces EAE clinical scores by modulating Treg/Th17 balances, promoting immunosuppression (41). Below, we summarize the most widely studied dietary factors as modulators of EAE. These and other dietary factors capable of regulating EAE severity were revised extensively by van den Hoogen et al. (42).

### 3.1. Fibers and derived metabolites

The supplementation of diet with fibers is proposed as a supplement strategy for MS patients due to their reduced abundance in the "western diet" (43). Dietary fiber can be either soluble or insoluble and fermentable or non-fermentable complex carbohydrates. Fermentable fiber microbial end products, short chain fatty acids (SCFAs), have evidence supporting protective and therapeutic properties in autoimmune diseases such as MS (3). SCFAs promote a healthy gut and provide systemic effects that could result in beneficial effects against neuroinflammation. As we recently reviewed, an increasing number of studies indicated

Dietary strategies	Alterations of gut microbiome	Form of disease and impact	References**	
No dietary alterations	<ul> <li>EAE*: ↑ Clostridiaceae (24, 25) and Akkermansia (24, 26)</li> <li>↓ Lactobacillaceae (24, 25) and Bacteroidaceae (26)</li> <li>MS*: ↑ Methanobrevibacter (28, 31), Acinetobacter (21, 27, 32), and Akkermansia (21, 22, 30-32)</li> <li>↓ Bacteroidaceae (27-30)</li> </ul>		(22-32)	
Vitamin D	MS: $\uparrow$ Akkermansia, Faecalibacterium, and Coprococcus (30)	MS: conflicting studies on the link of vitamin D and pathogenesis	(30)	
Dietary fiber	EAE: ↑ Ruminococcaceae, Helicobacteraceae and Enterococcaceae (3) and Lactobacillaceae (52) ↓ Sutterellaceae, Lactobacillaceae and Coriobacteriaceae (3)	EAE: normal diet 55% disease incidence vs. cellulose rich diet 23%, and cellulose rich diet delayed neurological symptoms by about 1 week	(3, 52)	
Mediterranean diet	EAE: no gut microbiome analysis, measured bacterial LPS levels MS: ↑ <i>Enterobacteriaceae, Akkermansia</i> and <i>Blautia</i> (80) ↓ <i>Faecalibacterium, Prevotella</i> , and <i>Parabacteroides</i> (80) Lower ratio <i>Firmicutes</i> to <i>Bacteroidetes</i> (80)		(80)	
Isoflavone diet	EAE: ↑ Adlercreutzia and Parabacteroides (3, 82) ↓ Akkermansia (3, 82)	EAE: increased bacteria are deficient in MS and decreased bacteria associated with MS	(3, 82)	
High vegetable/Low protein	MS high vegetable/low protein: ↑ <i>Lachnospiraceae</i> , <i>Coprococcus</i> , <i>Ruminococcus</i> , a sequence of an unclassified <i>Lachnospiraceae</i> strain, <i>Roseburia</i> , and a <i>Hungatella</i> -related unknown <i>Lachnospiraceae</i> (34) MS "Western diet": ↑ <i>Euryarchaeota</i> (83)	MS: samples of patients on different diets were taken after 12 months, no baseline samples for comparison	(34, 83)	
Intermittent fasting	EAE: ↑ Bacteroidaceae, Lactobacillaceae and Prevotellaceae (85) ↓ Bifidobacterium (85) MS: pilot study with similar alterations to EAE (85) EAE: delayed onset and reduced seven MS: intermittent fasting partly linked to all microbiome beneficial effects		(85)	

TABLE 1	Dietary	alterations	of the	gut	microbiome	in	EAE	and	MS.
---------	---------	-------------	--------	-----	------------	----	-----	-----	-----

\*The prevalent microbes across studies shown.

\*\*References are organized in order of dietary strategies.

the potential of triggering SCFA-specific responses in the context of inflammation (9). SCFAs promote anti-inflammatory pathways that regulate the balance between Th17 and Treg responses. Colonic bacteria that produce SCFAs are reduced in fecal samples from MS individuals compared to HC samples (44). Circulating levels of SCFAs appear to be reduced in MS individuals, and they are protective in the EAE model (45), promoting axonal remyelination (46–50). SCFAs produced by colonic bacteria have also been shown to be key to maintaining the integrity of the BBB (51).

Non-fermentable fiber is a common vegetarian diet component and has yet to be as thoroughly investigated due to poor breakdown via gut microbes. In a recent clinical study, non-fermentable fiber dietary cellulose has been shown to increase amounts of long-chain fatty acids and alter gut microbiota's composition, promoting beneficial effects in MS (3). Experimentally, in animals administered with normal diet, the EAE disease incidence was 55% compared to the 23% incidence observed in animals fed with a cellulose-rich diet (3). Neurological symptoms were significantly delayed about 1 week in the cellulose-rich diet EAE group; however, there was no significant difference in disease severity (3). Despite the reduction in SCFA-producing microbes and increase in longchain fatty acids, the cellulose-rich diet demonstrated beneficial effects (3). The fermentable dietary fiber guar gum reported increased SCFAs and significantly delayed EAE (43). A notable mention that might benefit from additional gut microbiome studies is pomegranate peel extract which increased Lactobacillaceae and reduced the severity of EAE (52). Pomegranate peel extract contains dietary fiber, polyphenols, vitamins, and minerals (53). These studies support dietary fibers as potent modulators of EAE and potential protective or therapeutic supplements for MS patients. A more detailed discussion of dietary fibers and their impact on the gut microbiome was recently reviewed by others (54).

Experimental autoimmune encephalomyelitis studies also revealed the protective effects of polyunsaturated fatty acids (FAs) against neuroinflammation, such as omega-6 fatty acid  $\gamma$ -linolenic acid in SJL mice (55), and oil-containing linolenic acids in Lewis rats (56), as well as docosahexaenoic acid (57) and eicosapentaenoic acid (58, 59) in C57BL/6 mice. Interestingly, the protective effects of omega-6 fatty acid  $\gamma$ -linolenic acid in the EAE model contradict previous findings of omega-6 as a pro-inflammatory factor. The notion of omega-6 FAs as precursors of pro- or anti-inflammatory mediators have been discussed by others (60). The form of omega-6 and their interactions with omega-3 FAs might be relevant to the nature of the immunological responses triggered (60). Diet supplementation with omega-3 resulted in moderated beneficial effects in a clinical study (61). These and other dietary factors that regulate EAE severity were already revised (42).

## 3.2. Vitamin supplementation

Vitamin D supplements have been proposed for use combined with disease-modifying therapies against MS. Vitamin D is fatsoluble, naturally present in foods, can be supplemented, and is stimulated from skin exposure to sunlight. Vitamin D plays a role in increasing intestinal absorption and retention of relevant nutrients such as calcium. Vitamin D deficiency is a common worldwide issue, even with the supplementation previously stated. Due to the distribution of MS being more prevalent where there is less sun exposure, vitamin D deficiency is thought to play a role in the pathogenesis of the disease (62). In an exploratory clinical study, MS patients were evaluated for alterations in the gut microbiota with vitamin D. MS patients treated with vitamin D had increased Akkermansia, Faecalibacterium, and Coprococcus vs. untreated MS (30). These microbes tend to be associated with pro-inflammatory properties. In other clinical studies measuring vitamin D levels, there is no consistent link between vitamin D supplementation and reduction or protection of MS (63). The combination of vitamin D and MOG protected against EAE (64). In addition, the administration of vitamin D3 also protected against the experimental disease (65). Another study showed that the protection induced by vitamin D treatment depends on the age of the animals and suggests that early administration of the dietary factor could be preferred (66). Interestingly, the requirement for vitamin D and vitamin D receptors for EAE induction has also been documented (67). A more detailed discussion on the role of vitamin D in regulating neuroinflammation has been recently published (68).

Vitamin A metabolites such as retinoid acid are protective against EAE by regulating the Treg-Th17 axis and Treg function (69–71). The immunomodulatory effects of retinoic acid have been demonstrated in other autoinflammatory models (72, 73).

The effects of vitamin E have also been explored in murine models of EAE through the administration of tocopherol or its derivatives (74, 75). In addition to reductions in EAE clinical scores, vitamin E (and vitamin D3) was also associated with increased remyelination in a toxic rat model of demyelination by injection of ethidium bromide (76).

# 4. Dietary interventions in EAE and MS

In the following paragraphs, we discuss studies that applied dietary interventions in MS patients and supporting pre-clinical evidence using EAE models. For example, the marmoset model EAE was used to address the potential beneficial effects of a diet enriched in fruit juices, carrot juice, yogurt, yeast flakes, vitamin D, among other components, and reduced ground powder. The administration of the diet selected a Bifidobacteria-enriched microbiome and reduced pro-inflammatory responses, inducing neuroprotection (77).

### 4.1. Mediterranean diet

The Mediterranean diet has been proposed to benefit cognitive health (78) and even be neuroprotective (79). The diet is associated with a lower ratio of *Firmicutes* to *Bacteroidetes* and promotes microbial diversity, which is important for gut homeostasis (80). Olive oil is one of the primary ingredients in the Mediterranean diet, in addition to large amounts of fruits, vegetables, whole grains, seafood, nuts, and legumes. Extra-virgin olive oil has previously been shown to have antioxidant and anti-inflammatory properties due to mono-unsaturated fats and phenols (81). Although a gut microbiome analysis was not performed in this extra-virgin olive oil EAE study, it is notable that the investigators detected increased circulating levels of lipopolysaccharide, Gram-negative bacterial endotoxin, in control EAE mice (81). Lipopolysaccharide levels were reduced in animals fed extra-virgin olive oil, suggesting dysbiosis and increased intestinal barrier disruption. Olive oil-fed EAE mice also showed reduced oxidative stress compared to controls (81).

Further supporting evidence is needed to demonstrate how diet modification can impact the gut microbiome and subsequently have a neuroprotective effect. A clinical study evaluated the impact of a Mediterranean diet on the microbiome of MS patients in a Spanish cohort of MS vs. HC patients (80). The study showed significant differences in the microbiota composition of MS and HC samples. Although the results indicated potential beneficial changes on the microbiota composition when exposed to the Mediterranean diet, the samples obtained from the MS cohort showed increased *Enterobacteriaceae*, a bacterial family that contains several potential pathogens (80).

### 4.2. Isoflavone diet

An isoflavone diet is characterized as a legume-based diet. Dietary legumes are rich in isoflavones and can include leaves, stems, and the fruit or seeds of certain plants, such as beans and peas. Isoflavones are phytoestrogens. Humans do not have the enzymes to break down isoflavones, so we rely on gut microbes for their catabolism and use. Isoflavone-metabolizing bacteria have previously been shown to be reduced in MS and are known for their antioxidant and anti-inflammatory properties for other diseases such as cancer (82). A recent EAE study investigated the effects of diets with and without isoflavones on the composition of the gut microbiome (82). The study found that the isoflavone-rich diet increased the isoflavonemetabolizing microbes Adlercreutzia and Parabacteroides in EAE mice. These bacteria are reduced in MS patients. By contrast, the administration of the diet without isoflavone to EAE mice increased the relative abundances of Akkermansia (82). These results suggest an isoflavone diet may have protective and therapeutic effects in neuroinflammatory diseases.

### 4.3. High vegetable/low protein diet

In line with previous diets, a diet high in vegetables and low in protein has been shown to improve MS clinical symptoms (83). A clinical pilot study was performed with MS patients comparing a high vegetable/low protein diet with the "Western diet" after a minimum of 12 months on the diet. Increased Lachnospiraceae, Coprococcus, Ruminococcus, a sequence of an unclassified Lachnospiraceae strain, Roseburia, and a Hungatellarelated unknown Lachnospiraceae along with reduced disease severity and relapse were found in MS high vegetable/low protein diet patients (83). Lachnospiraceae has been associated with antiinflammatory properties (83). In the MS "Western diet" patients, there was a significant increase in Euryarchaeota (83). For this pilot study, it is important to note they did not take baseline samples before the start of the diets for analysis, so the comparison for alteration of the gut microbiomes is strictly between MS patient diets at the end of a minimum of 12 months (83).

# 4.4. Reduced carbohydrate consumption and intermittent fasting

Ketogenic diets drastically reduce the intake of carbohydrates, inducing ketosis and driving the production of ketone bodies, an alternative energy source for the brain (84). Ketone bodies are believed to promote myelin regeneration, a possible target for therapeutics and protection in MS (84). Intermittent fasting also produces ketone bodies by inducing ketosis (84). Both may activate autophagy pathways that ameliorate disease by renewing and mobilizing non-nuclear and cytoplasmic macromolecules in cells (84). EAE studies have shown the beneficial effects of intermittent fasting; however, clinical studies are limited (84). Bahr et al.'s study is the first clinical study investigating ketogenic and intermittent fasting effects, which they found may modulate MS. However, clinical evidence was lacking. No gut microbiome analysis was performed. A recent EAE intermittent fasting study showed delayed onset and reduced severity of EAE (85). In the intermittent fasting EAE, there was an increase in anti-inflammatory microbes Bacteroidaceae, Lactobacillaceae, and Prevotellaceae, and a decrease in pro-inflammatory microbes Bifidobacterium (85). Cignarella et al. also performed a pilot clinical trial and found gut microbiome alterations resembling protective microbe alterations in their EAE study. They found intermittent fasting to have protective and therapeutic effects linked partly to an altered gut microbiome in EAE and MS.

### 5. Discussion

The studies summarized in this mini-review highlight the importance of future investigation into dietary interventions, specific microbes associated with dietary interventions, their interactions, and the roles they play in biological processes within EAE and MS. This mini-review covered various current and popular dietary strategies; however, the gut microbiota can produce thousands of metabolites and plays a role in many biological processes that need to be further investigated (80). The EAE model has extended our understanding of the multifactorial interactions between the gut microbiota, diet, and host. Less is known about the effects of diet on the MS cohort. The mini-review also highlights the need for more and larger clinical studies evaluating

# References

1. Walton C, King R, Rechtman L, Kaye W, Leray E, Marrie RA, et al. Rising prevalence of multiple sclerosis worldwide: insights from the atlas of MS, third edition. *Mult Scler.* (2020) 26:1816–21. doi: 10.1177/1352458520970841

2. Absinta M, Lassmann H, Trapp BD. Mechanisms underlying progression in multiple sclerosis. *Curr Opin Neurol.* (2020) 33:277-85. doi: 10.1097/WCO. 000000000000818

3. Berer K, Martínez I, Walker A, Kunkel B, Schmitt-Kopplin P, Walter J, et al. Dietary non-fermentable fiber prevents autoimmune neurological disease by changing gut metabolic and immune status. *Sci Rep.* (2018) 8:10431. doi: 10.1038/s41598-018-28839-3

4. Ghezzi L, Cantoni C, Pinget GV, Zhou Y, Piccio L. Targeting the gut to treat multiple sclerosis. J Clin Invest. (2021) 131:e143774. doi: 10.1172/JCI143774

dietary strategies in MS patients, including the impact on the gut microbiome. Nevertheless, the number of studies evaluating the impact of diet on the immune responses and signaling molecules in MS have increased of recent years (42, 86). Future studies that include analyses of gut microbiome modulation *via* dietary strategies are needed to investigate further the mechanisms for use as targets in the protection or treatment of MS.

# Author contributions

KH designed the outline of the manuscript. WD, SS, and JO-R helped with edits and revisions. All authors contributed to the article and approved the submitted version.

## Funding

This work was supported by a grant from the National Institutes of Health (R15NS107743). We acknowledge support from the Institutional Development Award (IDeA) from the National Institute of General Medical Sciences of the National Institutes of Health under Grants #P20GM103408 and P20GM109095, and the BSU-Biomolecular Research Center, RRID:SCR\_019174.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

5. Riccio P, Rossano R. Diet, gut microbiota, and vitamins D + A in multiple sclerosis. *Neurotherapeutics.* (2018) 15:75–91. doi: 10.1007/s13311-017-0581-4

6. Camara-Lemarroy CR, Metz L, Meddings JB, Sharkey KA, Wee Yong V. The intestinal barrier in multiple sclerosis: implications for pathophysiology and therapeutics. *Brain.* (2018) 141:1900–16.

7. Ochoa-Repáraz J, Kasper LH. Gut microbiome and the risk factors in central nervous system autoimmunity. *FEBS Lett.* (2014) 588:4214–22. doi: 10.1016/j.febslet. 2014.09.024

8. Belkaid Y, Hand T. Role of the microbiota in immunity and inflammation. *Cell.* (2014) 157:121-41. doi: 10.1016/j.cell.2014.03.011

9. Hoffman, K, Brownell Z, Doyle WJ, Ochoa-Repáraz J. The immunomodulatory roles of the gut microbiome in autoimmune diseases of the central nervous system:

multiple sclerosis as a model. *J Autoimmun*. (2022):102957. doi: 10.1016/j.jaut.2022. 102957 [Epub ahead of print].

10. Ochoa-Reparaz J, Kirby TO, Kasper LH. The gut microbiome and multiple sclerosis. *Cold Spring Harb Perspect Med.* (2018) 8:a029017. doi: 10.1101/cshperspect. a029017

11. Lassmann H. Pathophysiological basis of autoimmune-initiated/mediated neurodegeneration. In: Groppa S, Meuth SG editors. *Translational Methods for Multiple Sclerosis Research*. Totowa, NJ: Humana Press (2021).

12. Karpus WJ. Cytokines and chemokines in the pathogenesis of experimental autoimmune encephalomyelitis. *J Immunol.* (2020) 204:316–26. doi: 10.4049/jimmunol.1900914

13. Van Kaer L, Postoak JL, Wang C, Yang G, Wu L. Innate, innate-like and adaptive lymphocytes in the pathogenesis of MS and EAE. *Cell Mol Immunol.* (2019) 16:531–9. doi: 10.1038/s41423-019-0221-5

14. Yokote H, Miyake S, Croxford JL, Oki S, Mizusawa H, Yamamura T. NKT celldependent amelioration of a mouse model of multiple sclerosis by altering gut flora. *Am J Pathol.* (2008) 173:1714–23. doi: 10.2353/ajpath.2008.080622

15. Ochoa-Repáraz J, Mielcarz DW, Ditrio LE, Burroughs AR, Foureau DM, Haque-Begum S, et al. Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. *J Immunol.* (2009) 183:6041–50. doi: 10.4049/jimmunol.0900747

16. Seifert HA, Benedek G, Nguyen H, Gerstner G, Zhang Y, Kent G, et al. Antibiotics protect against EAE by increasing regulatory and anti-inflammatory cells. *Metab Brain Dis.* (2018) 33:1599–607. doi: 10.1007/s11011-018-0266-7

17. Gödel C, Kunkel B, Kashani A, Lassmann H, Arumugam M, Krishnamoorthy G. Perturbation of gut microbiota decreases susceptibility but does not modulate ongoing autoimmune neurological disease. *J Neuroinflamm*. (2020) 17:79.

18. Rothhammer V, Mascanfroni ID, Bunse L, Takenaka MC, Kenison JE, Mayo L, et al. Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor. *Nat Med.* (2016) 22:586–97. doi: 10.1038/nm.4106

19. Lee YK, Menezes JS, Umesaki Y, Mazmanian SK. Proinflammatory T-cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA*. (2011) 108(Suppl. 1):4615–22. doi: 10.1073/pnas.1000082107

20. Berer K, Mues M, Koutrolos M, Rasbi ZA, Boziki M, Johner C, et al. Commensal microbiota and myelin autoantigen cooperate to trigger autoimmune demyelination. *Nature*. (2011) 479:538–41. doi: 10.1038/nature10554

21. Cekanaviciute E, Yoo BB, Runia TF, Debelius JW, Singh S, Nelson CA, et al. Gut bacteria from multiple sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. *Proc Natl Acad Sci USA*. (2017) 363:201711235.

22. Berer K, Gerdes LA, Cekanaviciute E, Jia X, Xiao L, Xia Z, et al. Gut microbiota from multiple sclerosis patients enables spontaneous autoimmune encephalomyelitis in mice. *Proc Natl Acad Sci USA*. (2017) 145:201711233. doi: 10.1073/pnas.1711233114

23. Boziki MK, Kesidou E, Theotokis P, Mentis AA, Karafoulidou E, Melnikov M, et al. Microbiome in multiple sclerosis: where are we, what we know and do not know. *Brain Sci.* (2020) 10:234. doi: 10.3390/brainsci10040234

24. Colpitts SL, Kasper EJ, Keever A, Liljenberg C, Kirby T, Magori K, et al. A bidirectional association between the gut microbiota and CNS disease in a biphasic murine model of multiple sclerosis. *Gut Microbes.* (2017) 8:561–73. doi: 10.1080/19490976.2017.1353843

25. 2nd DM, Goertz JE, Marin IA, Costello J, Overall CC, Gaultier A. Experimental autoimmune encephalomyelitis is associated with changes of the microbiota composition in the gastrointestinal tract. *Sci Rep.* (2020) 10:15183.

26. Moles L, Egimendia A, Osorio-Querejeta I, Iparraguirre L, Alberro A, Suárez J, et al. Gut microbiota changes in experimental autoimmune encephalomyelitis and cuprizone mice models. *ACS Chem Neurosci.* (2021) 12:893–905. doi: 10.1021/acschemneuro.0c00695

27. Tremlett H, Fadrosh DW, Faruqi AA, Zhu F, Hart J, Roalstad S, et al. Gut microbiota in early pediatric multiple sclerosis: a case-control study. *Eur J Neurol.* (2016) 23:1308–21.

28. Tremlett H, Fadrosh D, Faruqi A, Hart J, Roalstad S, Graves J, et al. Associations between the gut microbiota and host immune markers in pediatric multiple sclerosis and controls. *BMC Neurol.* (2016) 16:182. doi: 10.1186/s12883-016-0703-3

29. Miyake S, Kim S, Suda W, Oshima K, Nakamura M, Matsuoka T, et al. Dysbiosis in the gut microbiota of patients with multiple sclerosis, with a striking depletion of species belonging to clostridia XIVa and IV clusters. *PLoS One.* (2015) 10:e0137429. doi: 10.1371/journal.pone.0137429

30. Cantarel BL, Waubant E, Chehoud C, Kuczynski J, DeSantis TZ, Warrington J, et al. Gut microbiota in multiple sclerosis: possible influence of immunomodulators. *J Investig Med.* (2015) 63:729–34.

31. Jangi S, Gandhi R, Cox LM, Li N, Glehn F v, Yan R, et al. Alterations of the human gut microbiome in multiple sclerosis. *Nat Commun.* (2016) 7:12015.

32. Vallino A, Dos Santos A, Mathé CV, Garcia A, Morille J, Dugast E, et al. Gut bacteria Akkermansia elicit a specific IgG response in CSF of patients with MS. *Neurol Neuroimmunol. Neuroinflamm.* (2020) 7:e688. doi: 10.1212/NXI.00000000000688 33. Magne F, Gotteland M, Gauthier L, Zazueta A, Pesoa S, Navarrete P, et al. The Firmicutes/bacteroidetes ratio: a relevant marker of gut dysbiosis in obese patients? *Nutrients.* (2020) 12:1474. doi: 10.3390/nu12051474

34. Mariat D, Firmesse O, Levenez F, Guimaráes V, Sokol H, Dor J, et al. The firmicutes/bacteroidetes ratio of the human microbiota changes with age. *BMC Microbiol.* (2009) 9:123. doi: 10.1186/1471-2180-9-123

35. Nkosi BVZ, Padayachee T, Gront D, Nelson DR, Syed K. Contrasting health effects of bacteroidetes and firmicutes lies in their genomes: analysis of P450s, ferredoxins, and secondary metabolite clusters. *Int J Mol Sci.* (2022) 23:5057. doi: 10.3390/ijms23095057

36. Jörg S, Kissel J, Manzel A, Kleinewietfeld M, Haghikia A, Gold R, et al. High salt drives Th17 responses in experimental autoimmune encephalomyelitis without impacting myeloid dendritic cells. *Exp Neurol.* (2016) 279:212–22. doi: 10.1016/j. expneurol.2016.03.010

37. Wu C, Yosef N, Thalhamer T, Zhu C, Xiao S, Kishi Y, et al. Induction of pathogenic TH17 cells by inducible salt-sensing kinase SGK1. *Nature*. (2013) 496:513–7. doi: 10.1038/nature11984

38. Kleinewietfeld M, Manzel A, Titze J, Kvakan H, Yosef N, Linker RA, et al. Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. *Nature.* (2013) 496:518–22.

39. Krementsov DN, Case LK, Hickey WF, Teuscher C. Exacerbation of autoimmune neuroinflammation by dietary sodium is genetically controlled and sex specific. *FASEB J.* (2015) 29:3446–57. doi: 10.1096/fj.15-272542

40. Schubert C, Guttek K, Grüngreiff K, Thielitz A, Bühling F, Reinhold A, et al. Oral zinc aspartate treats experimental autoimmune encephalomyelitis. *BioMetals.* (2014) 27:1249–62. doi: 10.1007/s10534-014-9786-8

41. Rosenkranz E, Maywald M, Hilgers R, Brieger A, Clarner T, Kipp M, et al. Induction of regulatory T cells in Th1-/Th17-driven experimental autoimmune encephalomyelitis by zinc administration. *J Nutr Biochem*. (2016) 29:116–23. doi: 10.1016/j.jnutbio.2015.11.010

42. van den Hoogen WJ, Laman JD, 't Hart B. Modulation of multiple sclerosis and its animal model experimental autoimmune encephalomyelitis by food and gut microbiota. *Front Immunol.* (2017) 8:1081. doi: 10.3389/fimmu.2017.01081

43. Fettig NM, Robinson HG, Allanach JR, Davis KM, Simister RL, Wang EJ, et al. Inhibition of Th1 activation and differentiation by dietary guar gum ameliorates experimental autoimmune encephalomyelitis. *Cell Rep.* (2022) 40:111328. doi: 10. 1016/j.celrep.2022.111328

44. Ling Z, Cheng Y, Yan X, Shao L, Liu X, Zhou D, et al. Alterations of the fecal microbiota in Chinese patients with multiple sclerosis. *Front Immunol.* (2020) 11:590783. doi: 10.3389/fimmu.2020.590783

45. Chevalier AC, Rosenberger TA. Increasing acetyl-CoA metabolism attenuates injury and alters spinal cord lipid content in mice subjected to experimental autoimmune encephalomyelitis. *J Neurochem*. (2017) 141:721–37. doi: 10.1111/jnc. 14032

46. Chen T, Noto D, Hoshino Y, Mizuno M, Miyake S. Butyrate suppresses demyelination and enhances remyelination. *J Neuroinflamm*. (2019) 16:165. doi: 10. 1186/s12974-019-1552-y

47. Park J, Wang Q, Wu Q, Mao-Draayer Y, Kim CH. Bidirectional regulatory potentials of short-chain fatty acids and their G-protein-coupled receptors in autoimmune neuroinflammation. *Sci Rep.* (2019) 9:8837.

48. Duscha A, Gisevius B, Hirschberg S, Yissachar N, Stangl G, Eilers E, et al. Propionic acid shapes the multiple sclerosis disease course by an immunomodulatory mechanism. *Cell.* (2020) 180:1067–80.e16. doi: 10.1016/j.cell.2020.02.035

49. Luu M, Pautz S, Kohl V, Singh R, Romero R, Lucas S, et al. The short-chain fatty acid pentanoate suppresses autoimmunity by modulating the metabolic-epigenetic crosstalk in lymphocytes. *Nat Commun.* (2019) 10:760. doi: 10.1038/s41467-019-08711-2

50. Mizuno M, Noto D, Kaga N, Chiba A, Miyake S. The dual role of short fatty acid chains in the pathogenesis of autoimmune disease models. *PLoS One.* (2017) 12:e0173032. doi: 10.1371/journal.pone.0173032

51. Braniste V, Al-Asmakh M, Kowal C, Anuar F, Abbaspour A, Tóth M, et al. The gut microbiota influences blood-brain barrier permeability in mice. *Sci Transl Med.* (2014) 6:263ra158.

52. Lu X, Han B, Deng X, Deng S, Zhang Y, Shen P, et al. Pomegranate peel extract ameliorates the severity of experimental autoimmune encephalomyelitis via modulation of gut microbiota. *Gut Microbes.* (2020) 12:1857515. doi: 10.1080/19490976.2020.1857515

53. Mo Y, Ma J, Gao W, Zhang L, Li J, Li J, et al. Pomegranate peel as a source of bioactive compounds: a mini review on their physiological functions. *Front Nutr.* (2022) 9:887113. doi: 10.3389/fnut.2022.887113

54. Puhlmann M-L, de Vos WM. Intrinsic dietary fibers and the gut microbiome: rediscovering the benefits of the plant cell matrix for human health. *Front Immunol.* (2022) 13:954845. doi: 10.3389/fimmu.2022.954845

55. Harbige LS, Layward L, Morris-Downes MM, Dumonde DC, Amor S. The protective effects of omega-6 fatty acids in experimental autoimmune encephalomyelitis (EAE) in relation to transforming growth factor-beta 1 (TGF- $\beta$  1)

up-regulation and increased prostaglandin E2 (PGE2) production. *Clin Exp Immunol.* (2001) 122:445–52. doi: 10.1046/j.1365-2249.2000.01399.x

56. Harbige LS, Yeatman N, Amor S, Crawford MA. Prevention of experimental autoimmune encephalomyelitis in Lewis rats by a novel fungal source of  $\gamma$ -linolenic acid. *Br J Nutr*. (1995) 74:701–15. doi: 10.1079/bjn19950173

57. Kong W, Yen J-H, Ganea D. Docosahexaenoic acid prevents dendritic cell maturation, inhibits antigen-specific Th1/Th17 differentiation and suppresses experimental autoimmune encephalomyelitis. *Brain Behav Immun.* (2011) 25:872–82. doi: 10.1016/j.bbi.2010.09.012

58. Unoda K, Doi Y, Nakajima H, Yamane K, Hosokawa T, Ishida S, et al. Eicosapentaenoic acid (EPA) induces peroxisome proliferator-activated receptors and ameliorates experimental autoimmune encephalomyelitis. *J Neuroimmunol.* (2013) 256:7–12. doi: 10.1016/j.jneuroim.2012.12.003

59. Salvati S, Biase AD, Attorri L, Benedetto RD, Sanchez M, Lorenzini L, et al. Ethyl-eicosapentaenoic acid ameliorates the clinical course of experimental allergic encephalomyelitis induced in dark agouti rats. *J Nutr Biochem.* (2013) 24:1645–54. doi: 10.1016/j.jnutbio.2013.02.005

60. Innes JK, Calder PC. Omega-6 fatty acids and inflammation. *Prostaglandins Leukot Essent Fatty Acids*. (2018) 132:41–8.

61. Weinstock-Guttman B, Baier M, Park Y, Feichter J, Lee-Kwen P, Gallagher E, et al. Low fat dietary intervention with  $\omega$ -3 fatty acid supplementation in multiple sclerosis patients. *Prostaglandins Leukot Essent Fatty Acids.* (2005) 73:397–404. doi: 10.1016/j.plefa.2005.05.024

62. Jagannath VA, Filippini G, Pietrantonj CD, Asokan GV, Robak EW, Whamond L, et al. Vitamin D for the management of multiple sclerosis. *Cochrane Database Syst Rev.* (2018) 9:CD008422. doi: 10.1002/14651858.CD008422.pub3

63. Marrie RA, Beck CA. Preventing multiple sclerosis: to (take) vitamin D or not to (take) vitamin D? *Neurology*. (2017) 89:1538–9. doi: 10.1212/WNL.000000000004506

64. Chiuso-Minicucci F, Ishikawa LL, Mimura LA, Fraga-Silva TF, França TG, Zorzella-Pezavento SF, et al. Treatment with vitamin D/MOG association suppresses experimental autoimmune encephalomyelitis. *PLoS One.* (2015) 10:e0125836. doi: 10. 1371/journal.pone.0125836

65. Mimura LA, Fraga-Silva TF, Oliveira LR, Ishikawa LL, Borim PA, Machado C d, et al. Preclinical therapy with vitamin D3 in experimental encephalomyelitis: efficacy and comparison with paricalcitol. *Int J Mol Sci.* (2021) 22:1914. doi: 10.3390/ ijms22041914

66. Adzemovic MZ, Zeitelhofer M, Hochmeister S, Gustafsson SA, Jagodic M. Efficacy of vitamin D in treating multiple sclerosis-like neuroinflammation depends on developmental stage. *Exp Neurol.* (2013) 249:39–48. doi: 10.1016/j.expneurol.2013. 08.002

67. Wang Y, Marling SJ, Zhu JG, Severson KS, DeLuca HF. Development of experimental autoimmune encephalomyelitis (EAE) in mice requires vitamin D and the vitamin D receptor. *Proc Natl Acad Sci USA*. (2012) 109:8501–4. doi: 10.1073/pnas. 1206054109

68. Miclea A, Bagnoud M, Chan A, Hoepner R. A brief review of the effects of vitamin D on multiple sclerosis. *Front Immunol.* (2020) 11:781. doi: 10.3389/fimmu. 2020.00781

69. Racke MK, Burnett D, Pak SH, Albert PS, Cannella B, Raine CS, et al. Retinoid treatment of experimental allergic encephalomyelitis. IL-4 production correlates with improved disease course. *J Immunol.* (1995) 154:450–8. doi: 10.4049/jimmunol.154. 1.450

70. Zhan X, Liu Y, Yang J, Wang G, Mu L, Zhang T, et al. All- trans -retinoic acid ameliorates experimental allergic encephalomyelitis by affecting dendritic cell and monocyte development. *Immunology.* (2013) 138:333–45. doi: 10.1111/imm. 12040

71. Xiao S, Jin H, Korn T, Liu SM, Oukka M, Lim B, et al. Retinoic acid increases Foxp3+ regulatory T cells and inhibits development of Th17 cells by enhancing TGF- $\beta$ -driven Smad3 signaling and inhibiting IL-6 and IL-23 receptor expression. *J Immunol.* (2008) 181:2277–84. doi: 10.4049/jimmunol.181.4.2277

72. Pino-Lagos K, Guo Y, Brown C, Alexander MP, Elgueta R, Bennett KA, et al. A retinoic acid–dependent checkpoint in the development of CD4+ T cell–mediated immunity. *J Exp Med.* (2011) 208:1767–75. doi: 10.1084/jem.20102358

73. Oliveira L, Teixeira F, Sato M. Impact of retinoic acid on immune cells and inflammatory diseases. *Mediat Inflamm*. (2018) 2018:1–17.

74. Blanchard B, Heurtaux T, Garcia C, Moll NM, Caillava C, Grandbarbe L, et al. Tocopherol derivative TFA-12 promotes myelin repair in experimental models of multiple sclerosis. *J Neurosci.* (2013) 33:11633–42.

75. Xue H, Ren H, Zhang L, Sun X, Wang W, Zhang S, et al. Alpha-tocopherol ameliorates experimental autoimmune encephalomyelitis through the regulation of Th1 cells. *Iran J Basic Med Sci.* (2016) 19:561–6.

76. Goudarzvand M, Javan M, Mirnajafi-Zadeh J, Mozafari S, Tiraihi T. Vitamins E and D3 attenuate demyelination and potentiate remyelination processes of hippocampal formation of rats following local injection of ethidium bromide. *Cell Mol Neurobiol.* (2010) 30:289–99. doi: 10.1007/s10571-009-9451-x

77. Kap YS, Bus-Spoor C, Driel N v, Dubbelaar ML, Grit C, Kooistra SM, et al. Targeted diet modification reduces multiple sclerosis-like disease in adult marmoset monkeys from an outbred colony. *J Immunol.* (2018) 1950:ji1800822. doi: 10.4049/jimmunol.1800822

78. Moustafa B, Trifan G, Isasi CR, Lipton RB, Sotres-Alvarez D, Cai J, et al. Association of mediterranean diet with cognitive decline among diverse Hispanic or Latino adults from the Hispanic community health study/study of Latinos. *JAMA Netw Open.* (2022) 5:e2221982.

79. Angeloni C, Malaguti M, Barbalace M, Hrelia S. Bioactivity of olive oil phenols in neuroprotection. *Int J Mol Sci.* (2017) 18:2230.

80. Moles L, Delgado S, Gorostidi-Aicua M, Sepúlveda L, Alberro A, Iparraguirre L, et al. Microbial dysbiosis and lack of SCFA production in a Spanish cohort of patients with multiple sclerosis. *Front Immunol.* (2022) 13:960761. doi: 10.3389/fimmu.2022. 960761

 Conde C, Escribano BM, Luque E, Feijóo M, Caballero-Villarraso J, Valdelvira ME, et al. Extra-virgin olive oil modifies the changes induced in non-nervous organs and tissues by experimental autoimmune encephalomyelitis models. *Nutrients*. (2019) 11:2448

82. Jensen SN, Cady NM, Shahi SK, Peterson SR, Gupta A, Gibson-Corley KN, et al. Isoflavone diet ameliorates experimental autoimmune encephalomyelitis through modulation of gut bacteria depleted in patients with multiple sclerosis. *Sci Adv.* (2021) 7:eabd4595. doi: 10.1126/sciadv.abd4595

83. Saresella M, Mendozzi L, Rossi V, Mazzali F, Piancone F, LaRosa F, et al. Immunological and clinical effect of diet modulation of the gut microbiome in multiple sclerosis patients: a pilot study. *Front Immunol.* (2017) 8:1391. doi: 10.3389/fimmu. 2017.01391

84. Bahr LS, Bock M, Liebscher D, Bellmann-Strobl J, Franz L, Prüß A, et al. Ketogenic diet and fasting diet as nutritional approaches in multiple sclerosis (NAMS): protocol of a randomized controlled study. *Trials*. (2020) 21:3. doi: 10.1186/s13063-019-3928-9

85. Cignarella F, Cantoni C, Ghezzi L, Salter A, Dorsett Y, Chen L, et al. Intermittent fasting confers protection in CNS autoimmunity by altering the gut microbiota. *Cell Metab.* (2018) 27:1222–35.e6. doi: 10.1016/j.cmet.2018.05.006

86. Montgomery TL, Eckstrom K, Lile KH, Caldwell S, Heney ER, Lahue KG, et al. *Lactobacillus reuteri* tryptophan metabolism promotes host susceptibility to CNS autoimmunity. *Microbiome*. (2022) 10:198. doi: 10.1186/s40168-022-01408-7