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*CORRESPONDENCE Colleen A. McClung mcclungca@upmc.edu

† These authors have contributed equally to this work and share first authorship

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[Adolescent circadian rhythm](https://www.frontiersin.org/articles/10.3389/fnins.2024.1478508/full) [disruption increases reward and](https://www.frontiersin.org/articles/10.3389/fnins.2024.1478508/full) [risk-taking](https://www.frontiersin.org/articles/10.3389/fnins.2024.1478508/full)

Lauren M. DePoy^{1,2†}, Chelsea A. Vadnie^{1,2,3†}, Kaitlyn A. Petersen^{1,2}, Madeline R. Scott^{1,2}, Wei Zong⁴, RuoFei Yin⁴, Ross C. Matthaei¹, Fernanda Juarez Anaya², Callie I. Kampe³, George C. Tseng⁴ and Colleen A. McClung^{1,2,3*}

1 Department of Psychiatry, Translational Neuroscience Program, University of Pittsburgh School of Medicine, Pittsburgh, PA, United States, 2Center for Neuroscience, University of Pittsburgh, Pittsburgh, PA, United States, ³David O. Robbins Neuroscience Program, Department of Psychology, Ohio Wesleyan University, Delaware, OH, United States, ⁴Department of Biostatistics, University of Pittsburgh, Pittsburgh, PA, United States

Introduction: Circadian rhythm disturbances have long been associated with the development of psychiatric disorders, including mood and substance use disorders. Adolescence is a particularly vulnerable time for the onset of psychiatric disorders and for circadian rhythm and sleep disruptions. Preclinical studies have found that circadian rhythm disruption (CRD) impacts the brain and behavior, but this research is largely focused on adult disruptions. Here, we hypothesized that adolescent CRD would have a greater effect on psychiatricrelated behaviors, relative to adult disruption.

Methods: We determined the long-term behavioral and neurobiological effects of CRD during early adolescence by exposing mice to 12 h shifts in the light/ dark cycle. Adult mice were exposed to the same CRD paradigm. Behavior testing began approximately 4 weeks later for both groups. To identify possible mechanisms, we also measured gene expression in brain regions relevant to circadian rhythms, mood and reward.

Results: CRD during early adolescence, but not adulthood, persistently increased exploratory drive (risk-taking behavior) and cocaine preference when tested later in life. Interestingly, we found sex differences when intravenous cocaine selfadministration was tested. While female mice with a history of adolescent CRD had a greater propensity to self-administer cocaine, as well as increased motivation and cue-induced reinstatement, male adolescent CRD mice had reduced motivation and extinction responding. Importantly, we found that transcripts in the SCN were affected by adolescent CRD and these were largely distinct across sex.

Conclusion: Overall, adolescent CRD in mice caused persistent increases in risky behavior, cocaine reward and cocaine self-administration, which suggests that CRD during adolescence may predispose individuals toward substance use disorders. Future research is required to elucidate how adolescent CRD affects behaviors relevant to mood-and substance use-related disorders across the 24-h day, as well as to identify intervention strategies to alleviate disruption during adolescence and novel therapeutic approaches once symptoms have begun.

KEYWORDS

circadian disruption, adolescence, cocaine, anxiety, risk-taking, reward

1 Introduction

Psychiatric disorders and circadian rhythm disruptions (CRD) commonly co-occur. Variations and disruptions of rhythms (e.g., evening chronotypes, misaligned rhythms, and delayed rhythms) are frequently described in psychiatric disorders [\(Souêtre et al., 1989](#page-15-0); [McClung, 2007;](#page-15-1) [Emens et al., 2009;](#page-14-0) [Merikanto et al., 2015](#page-15-2); [Vadnie and](#page-15-3) [McClung, 2017](#page-15-3)), including substance use disorders (SUDs; [Logan](#page-15-4) [et al., 2014;](#page-15-4) [DePoy et al., 2017\)](#page-14-1). On the other hand, those with disrupted sleep or circadian rhythms commonly exhibit increased symptoms of psychiatric disorders, such as increased risk-taking and drug use [\(Wittmann et al., 2006](#page-15-5); [Gowen et al., 2019;](#page-14-2) [Taylor et al., 2020;](#page-15-6) [Pasch et al., 2012](#page-15-7); [Hasler et al., 2017;](#page-14-3) [Hasler et al., 2016\)](#page-14-4), suggesting a bidirectional relationship between psychiatric disorders and CRD. Supporting this, environmental (e.g., light at night) and genetic insults to circadian rhythms are linked to altered mood [\(Benedetti](#page-13-0) [et al., 2003](#page-13-0), [2008;](#page-13-1) [Easton et al., 2003](#page-14-5); [Johansson et al., 2003](#page-14-6); [Roybal](#page-15-8) [et al., 2007;](#page-15-8) [Mansour et al., 2009;](#page-15-9) [Lavebratt et al., 2010](#page-15-10)) and substance use ([Dong et al., 2011;](#page-14-7) [Forbes et al., 2012](#page-14-8); [Ozburn et al., 2012](#page-15-11), [2013](#page-15-12); [Blomeyer et al., 2013](#page-14-9); [Gamsby et al., 2013](#page-14-10); [Bi et al., 2014;](#page-14-11) [Baranger](#page-13-2) [et al., 2016](#page-13-2)) across species. External or internal disruptions can perturb circadian rhythms by impacting the core molecular clock, a series of interlocking transcriptional-translational feedback loops found in almost every cell in the body [\(Reppert and Weaver, 2002;](#page-15-13) [Ko](#page-14-12) [and Takahashi, 2006;](#page-14-12) [Takahashi, 2017\)](#page-15-14), which controls physiological processes and behavior across the light/dark (LD) cycle. The molecular clock is largely regulated by the CLOCK protein, which dimerizes with ARNTL (BMAL1) to control transcription of many genes including the Period (*Per*) and Cryptochrome (*Cry*) genes. After translation, the PER and CRY proteins enter the nucleus and inhibit the transcriptional activity of CLOCK/BMAL1, closing the negative feedback loop after around 24 hours (h; [Ko and Takahashi, 2006](#page-14-12)).

Additionally, the suprachiasmatic nucleus (SCN), a brain region located in the hypothalamus, is known as the principal pacemaker of the brain and is responsible for the generation and maintenance of circadian rhythms across the brain and periphery. Not only can circadian disruption cause structural and functional changes to the SCN ([Iglesia et al., 2004](#page-14-13); [Yan, 2011](#page-15-15)), but notably, preclinical studies support a role for the SCN in regulating anxiety-like and moodrelated behaviors ([Landgraf et al., 2016;](#page-15-16) [Boehler et al., 2021](#page-14-14); [Whylings](#page-15-17) [et al., 2021;](#page-15-17) [Vadnie et al., 2022](#page-15-18)). Other regions likely to be involved in the effects of CRD on behaviors associated with psychiatric disorders include the prefrontal cortex (PFC) and nucleus accumbens (NAc). In animal models, circadian disruptions via clock gene knockouts and knockdowns have been shown to disrupt dopamine signaling rich areas, such as the NAc, and alter a variety of mood- and rewardrelated behaviors ([Andretic et al., 1999;](#page-13-3) [Falcón and McClung, 2009](#page-14-15); [Ozburn et al., 2012](#page-15-11), [2017;](#page-15-19) [Logan et al., 2014](#page-15-4); [Parekh and McClung,](#page-15-20) [2016;](#page-15-20) [Parekh et al., 2019;](#page-15-21) [DePoy et al., 2020](#page-14-16)). Further, in the PFC, a major site of executive function and cognitive control over behavior, neuronal activity and circadian gene expression are altered following exposure to a disrupted LD cycle ([Karatsoreos et al., 2011;](#page-14-17) [Otsuka](#page-15-22) [et al., 2020;](#page-15-22) [Roberts and Karatsoreos, 2023](#page-15-23)).

Preclinical studies have focused on investigating the effects of disturbing rhythms during adulthood or before weaning, but not during adolescence [\(Karatsoreos, Bhagat et al., 2011](#page-14-17); [LeGates et al.,](#page-15-24) [2012;](#page-15-24) [McGowan and Coogan, 2013](#page-15-25); [Borniger et al., 2014](#page-14-18); [Ben-Hamo](#page-13-4) [et al., 2016](#page-13-4); [Chen et al., 2021b;](#page-14-19) [Ameen et al, 2022](#page-13-5)). However, adolescence is a particularly vulnerable time for both psychiatric disorders and circadian disruptions. As reward-related brain regions mature faster than areas important for top-down cognitive control ([Mills et al., 2014\)](#page-15-26) and individuals seek independence from parents and more social interaction with peers ([Steinberg and Silverberg,](#page-15-27) [1986](#page-15-27)), adolescence is typically a time of increased risk-taking and novelty-seeking behavior ([Spear, 2000\)](#page-15-28). Adolescents are also vulnerable to circadian disruptions due to early morning school, work and societal obligations conflicting with their preferred delayed sleep and wake times, resulting in misaligned rhythms and reductions in sleep which have been shown to negatively impact mental and physical health ([Wahistrom, 2002](#page-15-29); [Owens et al., 2010;](#page-15-30) [Touitou, 2013;](#page-15-31) [Boergers et al., 2014](#page-14-20); [Borisenkov et al., 2022\)](#page-14-21). Notably, circadian rhythms are delayed in adolescents across different countries and cultures [\(Thorleifsdottir et al., 2002](#page-15-32); [Roenneberg et al., 2004](#page-15-33); [Yang](#page-15-34) [et al., 2005](#page-15-34); [Crowley et al., 2014\)](#page-14-22) and phase-shifts are also found in other species, including rodents [\(Golub et al., 1996](#page-14-23); [Hummer et al.,](#page-14-24) [2007](#page-14-24); [Hagenauer et al., 2011](#page-14-25)), suggesting this delay is biologically driven.

In rodents, adolescence is most broadly defined as the period from weaning postnatal day 21, (P21) to sexual maturity (P60), with most adolescent-typical neurobehavioral characteristics being present from P28-42 ([Spear, 2000\)](#page-15-28). During adolescence, there are significant changes occurring with neural circuitry and neurotransmitter systems. A few recent studies suggest that CRD during adolescence can induce lasting behavioral and neurobiological effects, indicating there is a need to further study the impact of circadian disruption during this developmental period when many psychiatric disorders start to arise ([Chen et al., 2021a](#page-14-26); [Cloutier et al., 2022](#page-14-27); [Gonzalez et al., 2023;](#page-14-28) [Bonilla](#page-14-29) [et al., 2024](#page-14-29); [Jameson et al., 2024\)](#page-14-30). Investigating the long-term impact of circadian rhythm disruption on behavior and the neurobiology of relevant brain regions may help us understand how psychiatric disorders develop and how they can be better prevented and treated.

The bidirectional relationship between psychiatric disorders and CRD begins during adolescence, a vulnerable time for engaging in risky behavior, such as initiating substance use, and suffering from CRD. In this study, we sought to determine the long-term behavioral and neurobiological effects of CRD during early adolescence and adulthood by exposing mice to 12 h shifts in the LD cycle. This type of light cycle paradigm is known to reduce exploratory drive and novelty seeking in adult mice [\(McGowan and Coogan, 2013\)](#page-15-25) as well as shift and dampen the amplitude of rhythms ([Kim et al., 2018](#page-14-31)). These types of rhythm disturbances have been implicated in psychiatric disorders in humans ([Souêtre et al., 1989;](#page-15-0) [Lewy, 2010;](#page-15-35) [Hasler et al., 2017\)](#page-14-3). We hypothesized that adolescent CRD would have a more profound effect on psychiatric-related behaviors relative to adult disruption. To identify possible mechanisms by which circadian disruption during adolescence affects behaviors later in life, we also measured gene expression in brain regions relevant to circadian rhythms, mood and reward, the SCN, PFC and NAc, respectively.

2 Materials and methods

2.1 Animals and housing conditions

Male and female C57BL/6 J mice were obtained from The Jackson Laboratory (000664) and then bred in house. Mice were maintained on a 12:12 light/dark (LD) schedule with lights on [Zeitgeber Time (ZT0)] at 0700 except for those exposed to CRD (described below). Behavioral testing occurred during the light phase from ZT2-10. Food and water were provided *ad libitum* unless otherwise indicated. Procedures were approved by the University of Pittsburgh Institutional Animal Care and Use Committee.

2.2 Circadian rhythm disruption

Adolescent CRD began at P28 \pm 1, when light cycles were shifted 12 h to a 12:12 schedule with lights on at 1900. After 3 days, the light cycle was shifted back by 12 h to a 12:12 schedule with lights on at 0700. The 12 h shifts of the light cycle continued every 3 days, resulting in 4 complete reversals of the LD cycle spanning P28-37. Following adolescent CRD, mice were returned to a fixed LD schedule (ZT0 = 0700) and behavioral testing began approximately 4 weeks later, in adulthood. Due to the nature of this shifting paradigm, each experimental group was housed together, and mice were pseudo-randomized to each behavior testing group.

Adult mice experienced CRD as described above from P62-71. Following CRD, mice were returned to a fixed LD schedule $(ZT0 = 0700)$, and behavioral testing began approximately 4 weeks later. Control adolescent and adult mice were physically disturbed at the same ages and times of day with matched cage handling but received no change to their LD schedule.

2.3 Drugs

Cocaine hydrochloride was provided by the National Institute on Drug Abuse. Animals were injected with 2.5, 5 or 10 mg/kg (intraperitoneal, i.p.; volume 10 mL/kg) for conditioned place preference testing and 0–1 mg/kg/infusion for cocaine self-administration.

2.4 Behavioral testing

Mice used for behavior testing were subjected to only 1 of the 3 different groups of behavior testing.

Group 1 mice were used in a battery of tests in the following order to assess locomotor activity, exploratory drive [open field, elevated plus maze (EPM), light/dark (LD) box], spatial working memory (T-maze) and helplessness-like behavior (forced swim test, FST). Behavior testing took place every other day.

Group 2 mice were tested for reward-related behavior. These mice were tested for sucrose preference and cocaine conditioned place preference (CPP).

Group 3 mice were assessed for operant food self-administration and intravenous cocaine self-administration.

2.4.1 Locomotor response to novelty

Mice were placed in clear boxes (field dimensions: 9.5″ x 18.0″) equipped with photobeams (Kinder Scientific Smart Cage Rack System, Poway, CA). Total horizontal distance traveled over 120 minutes (min) was measured for each mouse under standard room lighting.

2.4.2 Open field

Mice were placed in the corner of a large plastic arena (52 \times 52 \times 25 cm) with black walls and a clear bottom. The center was a 24 cm x 24 cm square in the middle of the box that was designated with taped gridlines. Mice were allowed to freely explore the box for 10 min. Behavior was video recorded under dim white light (~20 lux). Center entries and center time were quantified using Ethovision XT (Noldus, Leesburg, VA, United States).

2.4.3 Elevated plus maze

As previously described [\(Vadnie et al., 2022\)](#page-15-18), mice were placed in the center of the maze facing an open arm in dim light $(\sim 20 \text{ lux})$. The EPM consisted of 2 plastic open arms perpendicular to two closed arms (30 \times 5 cm) and was elevated above the ground (81 cm). Behavior was recorded for 10 min and video tracking software (Ethovision XT; Noldus, Leesburg, VA, United States) was used to quantify the time spent in the open arms, as well as the number of entries into each arm.

2.4.4 Light/dark box

Clear boxes (Kinder Scientific Smart Cage Rack System; field dimensions: 9.5″ x 18.0″) were divided into two equally sized chambers, a black opaque chamber kept in the dark with a lid and a brightly lit chamber (~880 lux). An opening was present between the two chambers that was equipped with a door. Mice were first placed in the dark chamber for 2 min and then the door was opened between the two sides. Mice were allowed to explore both chambers for 20 min. Photobeams were used to measure the number of entries into and time spent in the light side.

2.4.5 T-maze

Testing in the T-maze took place under dim white light (~40 lux). The assay consisted of 5 trials made up of 2 runs (a sequestered and a non-sequestered run). For both runs the mouse was placed in the starting arm of the T-maze facing toward the wall, away from the two choice arms. For the first sequestered run of the trial, the mouse was given 2 min to select an arm. Once the mouse had all four paws in an arm, the door was closed so that the mouse was sequestered in the arm for 30 seconds (s). The mouse was then removed from the maze and placed in a clean holding cage for 20 s. Any urine or feces were removed from the T-maze to reduce scent cues. For the non-sequestered run, the mouse was again placed back in the starting arm facing toward the wall, away from the choice arms. The mouse was given 2 min to select an arm. Once the mouse selected an arm and had all 4 paws in the arm, it was placed back in the holding cage for 20 s. A spontaneous alternation was considered when the mouse selected the arm opposite to the one it was sequestered in. % spontaneous alternations were calculated as the number of alternating trials divided by the total completed trials multiplied by 100. If the mouse failed to select an arm, the mouse was placed back in the holding cage for 1 min. Then the mouse was permitted to re-do the trial. If the mouse still failed to select an arm, it was returned to a holding cage for 1 min and then was moved on to the next trial. Testing ended after 5 trials or after the mouse failed to select an arm for 3 consecutive runs. No mouse failed the test completely.

2.4.6 Forced swim test

Mice were individually placed into glass beakers of water (25°C; 18 cm depth) that were separated by dividers to prevent the mice from observing each other during testing. Behavior was video recorded for 6 min. Time spent struggling during the last 4 min of the test and latency to immobility were scored by a blinded, trained observer. Struggling was defined as any movement that was not for the sole purpose of keeping the mouse afloat. Immobility time was calculated by subtracting the time spent struggling from the total time (4 min).

2.4.7 Conditioned place preference

As previously described ([Ozburn et al., 2015](#page-15-36)) mice were first habituated to a testing room for 30 min. On day 1, a pre-conditioning test was conducted, wherein mice were placed in the center of a 3-chamber box. The outer two chambers were distinct with visual and tactile differences. Time in each chamber was recorded over the 20-min session. On the subsequent 4 days, mice were injected with either saline or cocaine and restricted to one of the outer chambers. Saline was injected on days 2 and 4, and cocaine was injected on days 3 and 5 (2.5, 5 or 10 mg/kg, 10 mL/kg, NIDA drug consortium). A biased design was used, since >50% of mice showed a chamber bias during the pre-test, wherein the preferred chamber (>10% preference) was paired with saline or chambers were assigned pseudo-randomly if no side preference was found.

2.4.8 Sucrose preference

Mice were habituated to two bottles containing water or a sucrose solution (0.5% sucrose w/v). After 24 h, bottle position was switched to minimize side preferences. After another 24 h, mice were individually housed and exposed to water and sucrose bottles. Mice were undisturbed overnight, for 18 h, and bottles were weighed to determine consumption of sucrose and water. Control cages were sham handled and fluid loss was recorded. Percent preference for sucrose consumption was calculated with 50% indicating no preference.

2.4.9 Intravenous cocaine self-administration

Mice were first restricted to 85% of their free-feeding weight for food self-administration. Mice were trained to respond for chocolate flavored food pellets (20 mg, grain-based precision pellets, Bio-Serv) in Med-Associates operant conditioning chambers. Responding on one lever was reinforced using a fixed ratio 1 (FR1) schedule. A cue light was illuminated over the active lever for the duration of the experiment. Responses on the inactive lever had no programmed consequences but were recorded. Sessions ended at 60 min or when a maximum of 30 pellets were acquired. Mice were trained for at least five sessions or until they acquired ≥25 pellets for three consecutive sessions.

Mice then underwent jugular catheterization. Mice were anesthetized with isoflurane. As previously described ([DePoy](#page-14-16) [et al., 2020](#page-14-16)), the dorsal and ventral sides were shaved and disinfected. The right jugular vein was exposed by blunt dissection and a sterile polyurethane catheter was placed and secured to the vein. The catheter was exteriorized posterior to the scapulae via a felt mesh mount (Instech). The dorsal and ventral wounds were sutured, and mice were pair housed for the duration of the experiment, unless fighting or uneven sample sizes necessitated single housing. Mice received analgesic treatment (Rimadyl 5 mg/ kg) following surgery and post-operatively for 2 days. Mice recover for 6–7 days (d) before intravenous self-administration training begins. Catheters were maintained by infusing catheters

daily with 0.05 mL gentamicin (0.33 mg/mL) and heparinized saline (30 USP/ml) containing baytril (0.5 mg/kg). Catheter patency was tested approximately once per week using 0.05 mL brevital (3 mg/mL), mice that failed to lose muscle tone were excluded.

After recovery from surgery, mice were trained to respond on an FR1 schedule for cocaine (0.5 mg/kg/infusion, 30 μL over 1.7 s) on the previously inactive lever from food training [\(Ozburn et al., 2012](#page-15-11)). Cocaine was delivered through an armored tether connected to a swivel and syringe pump. Mice were tested 6 d/week with the last day being reserved for patency testing. Drug delivery culminated in extinction of the house light, a compound cue (auditory tone and stimulus light), and a 10-s (s) timeout during which no additional cocaine reinforcers can be delivered. Sessions ended when mice selfadministered 60 infusions or at 120 min. Mice were considered to have acquired the cocaine-reinforced response when mice selfadministered ≥15 reinforcers across three sessions (with ≥2:1 active/ inactive lever press ratio) and a minimum of 10 total acquisition sessions. Drug intake (infusions), discrimination of the active versus inactive and time to reach criteria were measured.

After acquisition, mice were tested on an FR1 schedule with two consecutive sessions of descending unit doses of cocaine (1.0, 0.5, 0.25, 0.125, 0.063, and 0 mg/kg). This dose–response analysis measured the reinforcing properties of cocaine. Next, mice self-administered cocaine at the baseline 0.5 mg/kg unit dose for 2 days before progressive ratio testing. In order to measure motivation to take cocaine in the same mice, three counterbalanced unit doses (1.0, 0.5, and 0.25 mg/kg) were presented for two consecutive sessions under a progressive ratio schedule, wherein each successive infusion requires more lever press responses (1, 2, 4, 6, 9, 12, 16, 20…240). Sessions end after 4 or 1.5 h without acquiring a reinforcer. This test is used to measure motivation, which will be used to describe changes to the breakpoint ratio, or the last ratio obtained for a dose of cocaine. Mice were again returned to their baseline self-administration dose (0.5 mg/kg/infusion) for 2 days before extinction training. Here, all lever presses had no programmed consequences and mice were trained for 10 days or until ≤30% peak active responding at 0.5 mg/kg/infusion was reached. Following extinction, mice were tested for sensitivity to cue-induced reinstatement, a model of relapse where presentations of previously cocaine-associated cues invigorate responding on the previously cocaine-reinforced lever. Mice received one non-contingent cue presentation, followed by cues contingent on responding on the previously active lever.

2.5 Tissue collection

Mice exposed to adolescent CRD, as well as matching controls, were euthanized approximately 4 wks after the CRD or control paradigm ended by cervical dislocation and rapid decapitation at ZT6. Brains were flash frozen on dry ice and stored at −80°C until used for punching.

2.6 RNA-sequencing

The prefrontal cortex (PFC), nucleus accumbens (NAc), and suprachiasmatic nucleus (SCN) were punched and isolated from frozen brain tissue using a cryostat. Individual animals were each used as a sample ($n = 4-8$ per condition). Tissue was homogenized and total RNA was isolated using the RNeasy Plus Micro Kit (Qiagen). RNA quantity and quality were assessed using fluorometry (Qubit RNA High Sensitivity Assay Kit and Fluorometer; Invitrogen) and chromatography (Bioanalyzer and RNA 6000 Pico Kit; Agilent) respectively. Libraries were prepared with 10 ng of RNA from each sample using the Smartseq HT ultra-low input sample preparation kits (Illumina). Paired-end dual-indexed sequencing (75 bp) was performed using the NextSeq 500 platform (Illumina). A total of 30 million reads per sample was targeted. Sequencing was performed at the University of Pittsburgh Health Sciences Sequencing Core at UPMC Children's Hospital of Pittsburgh.

FastQC v0.11.7 was used to assess the quality of the data. Per base sequence quality was high (Quality score generally >30) indicating good data quality. HISAT2 (HISAT2v2.1.0) was used to align reads to the reference (*Mus musculus* Ensembl GRCm38) using default parameters. The resulting bam files were converted to expression count data using HTSeq (HTSeq v0.10.0) with default union mode. Data normalization and gene filtering were performed separately for each region. Genes were retained for analysis if counts per million (CPM) was greater than 1 in 50% or more of the subjects within each group. All Y-chromosome genes were also eliminated from the analysis. A total of 10,386 (NAc), 10,220 (PFC), and 11,339 (SCN) annotated genes retained after filtering were used as the initial input dataset. The median of ratios normalization method in DESeq2 was then used to normalize for sequencing depth and RNA composition followed by a log2 transformation. Log2CPM values were used to calculate z-scores for expression for each transcript. Heatmaps were generated using z-scores to visualize changes in expression across group and brain region.

2.7 Statistical analyses

GraphPad Prism 10 was used. Data are expressed as mean ± SEM with $p \le 0.05$ considered significant and $0.05 < p \le 0.1$ considered trending. Data were analyzed by two- or three-way ANOVAs for effects of CRD, sex and session/lever/dose and repeated measures (RM) were used when appropriate. ANOVAs with significant and trending interactions were followed by post-hoc tests with Bonferroni corrections for multiple comparisons. If a significant/trending interaction or effect of sex was found in a three-way RM ANOVA, data were separated by sex and two-way ANOVAs were performed in order to better examine the effect of CRD and the RM on the dependent variable. If no effect of sex (main or interaction) was found, data were collapsed for visualization and to increase power for subsequent analysis (ANOVA or unpaired two-tailed t-test), without sex as a factor. A Welch's correction was used for unpaired two-tailed t-tests comparing groups that differed in variance, meaning that the F-test to compare variance between the two groups was significant. F-statistics from the higher order ANOVA (main effect of sex and interactions) are shown. Outliers over two standard deviations from the mean were excluded (*a priori*)*. F* or *t* < 1 is noted when appropriate for non-significant findings.

To examine gene expression changes by sex, at the approximate time of our behavioral assessments, a DE analysis was conducted by DESeq2 in each sex and brain region separately at ZT6. Genes were ranked for high DE based on q values and fold change in each brain region and sex. Genes were considered significantly DE with a q-value criteria of *q* < 0.05 to adjust for multiple comparisons. A fold change cut-off of 1.2 was used.

Metascape, a gene annotation and analysis resource, was used to identify enriched biological processes in the top DE genes ([Zhou et al., 2019\)](#page-15-37). To compare between groups, a significance threshold of $q < 0.05$ was used. $p < 0.01$ was used as a significance threshold to identify significantly enriched processes. For the background gene list in Metascape, we used a list of the 12,523 transcripts expressed in any of the 3 brain regions, which is more accurate than using the default full genome background. Further exploratory analysis was done on transcripts that did not pass the false discovery rate, instead using $p < 0.05$ to identify transcripts and identify enriched processes.

3 Results

3.1 Adolescent CRD increases exploratory drive and impairs spatial working memory

Mice were exposed to circadian rhythm disruption (CRD) during adolescence, which consisted of four 12 h shifts in the LD cycle occurring every 3 days, from P28-37 and behavior was tested in adulthood, ~4 weeks later. For most of the behaviors tested, analysis by two-way ANOVA revealed no interactions or effects of sex (*F*s < 1; latency to immobility: interaction $F_{(1,33)} = 1.55$, $p = 0.22$, sex $F_{(1,33)} = 2.20$, $p = 0.15$), so data were collapsed across sex and analyzed by unpaired two-tailed *t*-tests. For the LD box, where we did see a significant interaction, the data are separated by sex.

We first determined if adolescent CRD affected locomotor activity in a novel environment and the exploratory drive of adult mice. Adolescent CRD decreased the distance traveled in a 2 h locomotor test in a novel environment (Figure 1A, $t_{(36)} = 2.07$, $p = 0.046$; [Supplementary Figure 1](#page-13-6)), but increased exploratory drive in the open field, LD box, and EPM in adulthood ([Figures 1B–E](#page-5-0)). Specifically, adolescent CRD increased the amount of time spent in the center of the open field [\(Figure 1B](#page-5-0), $t_{(36)} = 2.18$, $p = 0.036$). There was also a trend for adolescent CRD to increase entries into the center of the open field [\(Figure 1C,](#page-5-0) $t_{(36)} = 1.81$, $p = 0.079$) and the percentage of open arm entries into the EPM ([Figure 1D](#page-5-0), $t_{(36)} = 1.81$, $p = 0.078$). Interestingly, unlike the other measures of exploratory drive, analysis by two-way ANOVA revealed an interaction between the effects of sex and adolescent CRD on time spent on the light side of the LD box ([Figure 1E](#page-5-0), $F_{(1, 33)} = 6.44$, $p = 0.016$). There was also an effect of sex ($F_{(1, 33)} = 9.70$, $p = 0.004$) and an overall effect of CRD ($F_{(1, 133)} = 9.70$, $p = 0.004$) and an overall effect of CRD 33 ₃₃) = 11.82, $p = 0.002$). Specifically for this measure in the LD box, adolescent CRD increased exploratory drive in adult males only. Also, there was a baseline sex difference, with female controls spending more time in the light relative to male controls, which may have caused a ceiling effect. Overall, circadian disruption during adolescence increased exploratory drive or risk-taking behaviors in adult mice, except for the LD box, where only males were affected.

To determine if adolescent CRD may affect cognitive function, we assessed the behavior of adult mice with a history of adolescent

CRD in the T-maze. Adolescent CRD decreased spontaneous alternations in the T-maze suggesting that adolescent CRD impairs spatial working memory ([Figure 1F,](#page-5-0) $t_{(25.37)} = 2.27$, $p = 0.032$).

To assess if adolescent CRD has long-lasting effects on helplessness-like behavior, we measured behavior in the FST and observed no differences in immobility ([Figure 1G,](#page-5-0) *t* < 1) or latency to immobility ([Figure 1H](#page-5-0), *t* < 1) suggesting that adolescent CRD does not alter depressive-like behavior.

3.2 Adult CRD does not affect exploratory drive or spatial working memory

Next, we determined if CRD during adulthood, from P62-71, has similar effects when the same amount of time (~4 wks) was

given between CRD and behavior testing. For most of the behaviors tested, analysis by two-way ANOVA revealed no interactions or effects of sex (*F*s < 1; locomotion: interaction $F_{(1,38)} = 1.50$, $p = 0.23$; center entries open field: sex $F_{(1,38)} = 2.74$, *p* = 0.11; open arm entries EPM: interaction $F_{(1,40)} = 1.45$, *p* = 0.24, sex $F_{(1,40)} = 1.81$, $p = 0.19$), so data were collapsed across sex and analyzed by unpaired two-tailed *t*-tests. For center time in the open field and latency to immobility for the FST, where we did see an effect of sex and interaction respectively, the data are separated by sex.

Adult CRD had no effect on distance traveled in a 2 h locomotor test ([Figure 2A](#page-6-0), *t* < 1; [Supplementary Figure 1\)](#page-13-6). Analysis by two-way ANOVA revealed females overall spent more time in the center of the open field ([Figure 2B](#page-6-0), $F_{(1,39)} = 7.56$, $p = 0.009$). No effects of CRD ($F_{(1,39)} = 1.27$, $p = 0.27$) or interactions between sex and CRD ($F < 1$) were found for

time spent in the center of the open field. Adult CRD also had no effect on the number of center entries in the open field [\(Figure 2C](#page-6-0), $t < 1$). Furthermore, adult CRD had no effect on the percentage of open arm entries in the EPM ([Figure 2D](#page-6-0), $t < 1$) or time spent in the light side in the LD box test [\(Figure 2E,](#page-6-0) *t* < 1). Overall, circadian disruption in adulthood had no effect on locomotor activity or exploratory drive in adult mice that were tested ~4 wks after disruption occurred.

Adult CRD also had no effect on spatial working memory, with no change observed in the number of spontaneous alternations in the T-maze [\(Figure 2F,](#page-6-0) *t* < 1).

To assess if adult CRD has lasting effects on helplessness-like behavior, we assessed behavior in the FST. Adult CRD had no effect on immobility ([Figure 2G,](#page-6-0) *t* < 1). Analysis by two-way ANOVA revealed an interaction between sex and adult CRD on latency to immobility ([Figure 2H](#page-6-0), $F_{(1,39)} = 6.35$, $p = 0.016$). There were no overall effects of sex or adult CRD (*Fs* < 1). Bonferroni test showed a reduction in latency to immobility in adult CRD males, but not females.

3.3 Adolescent CRD, but not adult CRD increases cocaine reward

Since increased exploratory drive or risk-taking-like behavior is associated with increased reward seeking, we then investigated if adolescent CRD specifically affects reward-related behaviors in mice. We first determined if adolescent CRD affects sucrose preference or natural reward. Here we found an interaction for the effects of sex and adolescent CRD on sucrose preference [\(Figure 3A,](#page-7-0) $F_{(1,49)} = 4.87$, $p = 0.032$). Post-hoc analyses revealed a trend for adolescent CRD to decrease sucrose preference in males, but

gray-filled circles and females by open circles. Mean \pm SEM, $n = 20-22$, *p ≤ 0.05

adolescent CRD had no effect on sucrose preference in females. When adult mice experienced CRD and were tested ~4 wks later, sucrose preference was not altered relative to controls ([Figure 3B](#page-7-0)). Analysis by two-way ANOVA revealed no effect of adult CRD or interaction (*F*s < 1), but there was a trend for decreased sucrose preference in all females relative to males $(F_{(1,26)} = 2.99, p = 0.10)$. Overall, CRD during adolescence and adulthood seems to have a minimal effect on natural reward, with only a slight reduction in males following adolescent CRD.

We then investigated whether adolescent CRD and adult CRD may differentially affect drug reward by assessing cocaine CPP. Analysis by ANOVA revealed no interactions or effects of sex (*F*s < 1), so data were collapsed across sex and analyzed by a lower order ANOVA. Adolescent mice were conditioned to either 2.5, 5.0, or 10 mg/kg of cocaine. In a two-way ANOVA for effects of dose and adolescent CRD, as expected, higher cocaine doses resulted in greater CPP ([Figure 3C,](#page-7-0) Effect of dose, $F_{(2, 110)} = 5.35$, $p = 0.006$). Interestingly, adolescent CRD overall increased cocaine CPP ($F_{(1)}$, $_{110}$ = 7.76, $p = 0.006$). Adult mice were only conditioned to one dose of cocaine, the middle dose (5 mg/kg), since no behavioral effects had been seen thus far. CRD during adulthood had no effect on cocaine CPP at this dose [\(Figure 3D](#page-7-0); $t_{(34)} = 1.51$, $p = 0.14$). Thus, adolescent CRD, but not adult CRD increased cocaine reward across sex.

3.4 Adolescent CRD has sex-specific effects on cocaine self-administration

Since differences in the sensitivity to cocaine reward after adolescent CRD may affect cocaine use, we then determined the effects of adolescent CRD on intravenous cocaine self-administration. For experiments assessing cocaine administration across RM (sessions, doses, or lever), data were analyzed by three-way ANOVAs for the effects of adolescent CRD, sex, and RM. If sex differences or an interaction with sex was observed, data were separated by sex and analyzed by two-way ANOVAs to better examine effects across our repeated measures (dose/session/lever).

Mice were trained to self-administer food before jugular catheters were placed to investigate whether operant conditioning is affected by adolescent CRD. A three-way ANOVA for the effects of adolescent CRD, sex, and session revealed no interactions or effect of sex (*F*s < 1), therefore sex was pooled. In a two-way ANOVA (adolescent CRD and session), all mice increased their responding for the reinforced lever as shown by an effect of session ([Supplementary Figure 2,](#page-13-6) $F_{(4,376)} = 190.1, p < 0.0001$). We also found a trend for adolescent CRD to increase responding on the food-reinforced lever $(F_{(1,94)} = 3.21,$ $p = 0.077$.

After mice recovered from jugular catheterization, they were trained to self-administer cocaine. In a three-way ANOVA, all mice

FIGURE 3

Effects of adolescent CRD and adult CRD on natural and cocaine reward. Adolescent CRD also altered sucrose preference, decreasing preference compared to controls in male mice, but not female mice (A). There was no effect of adult CRD (B) on sucrose preference. Adolescent CRD (C), but not adult CRD (D) increased cocaine CPP. Individual data points are shown for each sample with males indicated by gray-filled circles and females by open circles. Mean \pm SEM, $n = 12-16$ for sucrose preference and 16-24 for CPP, $* p \le 0.05$

acquired cocaine self-administration as shown by an increase in lever pressing across sessions (effect of session, $F_{(9,522)} = 43.92$, $p < 0.0001$). A trending effect of sex was also found $(F_(1.58) = 2.78, p = 0.10)$, therefore two-way ANOVAs were performed for males and females separately. No change in cocaine self-administration was found in female adolescent CRD mice compared to controls ([Figure 4A,](#page-8-0) effect of adolescent CRD *F* < 1). In males, there was a trend for an interaction between adolescent CRD and session $(F_{(9,261)} = 1.90, p = 0.05)$. While there were no significant post-hoc effects of adolescent CRD on daily cocaine intake, this interaction suggests that male mice with a history of adolescent CRD acquire cocaine self-administration differently than controls. Interestingly, when sessions to reach criteria were quantified, a trend for an interaction was found (two-way ANOVA, adolescent CRD and sex, $F_{(1,54)} = 3.56$, $p = 0.06$). In post-hoc comparisons, female mice that were exposed to adolescent CRD reached criteria for self-administration significantly faster.

Following acquisition, a dose–response curve was used to measure the reinforcing properties of cocaine. Analysis by three-way ANOVA revealed an interaction between the effects of dose, sex and adolescent CRD $(F_{(5,285)} = 2.38, p = 0.039)$. When analyzed separately, two-way ANOVAs did not reveal any significant effects of adolescent CRD within females (*F* < 1) or males (*F*_(1,29) = 1.05, *p* = 0.32) [\(Figure 4B](#page-8-0)). However, the three-way interaction above suggests that male and

female mice have distinct responses to adolescent CRD in the context of a dose–response curve.

Next, we assessed the effects of adolescent CRD on motivation for cocaine by determining the breakpoint ratio for responding during a progressive ratio schedule of reinforcement for three different doses of cocaine. A three-way ANOVA revealed a trending interaction between sex and adolescent CRD $(F_{(1,39)} = 3.10, p = 0.09)$. In females, we found a significant interaction between the effects of cocaine dose and adolescent CRD (two-way ANOVA, Figure 5A, $F_{(2,38)} = 3.44$, $p = 0.042$), however no significant post-hoc comparisons were identified within any individual dose. For males, a trend was observed for an effect of adolescent CRD ($F_{(1,21)} = 2.92$, $p = 0.10$) with a decrease in breakpoint ratio, or motivation, in mice previously exposed to adolescent CRD.

We then assessed extinction and cue-induced reinstatement. In a three-way ANOVA, all mice extinguished responding as shown by a decrease in lever pressing across sessions when cocaine and cocaineassociated cues were withheld ([Figure 5B,](#page-9-0) effect of session, $F_{(9,351)} = 31.57$, $p < 0.0001$). We also found a trending interaction between the effects of session, sex, and adolescent CRD ($F_{(9,351)} = 1.79$, $p = 0.07$). In females, adolescent CRD had no effect on extinction (two-way ANOVA, $F_{(1,19)} = 1.15$, $p = 0.30$), but in males, we observed an interaction between the effects of adolescent CRD and session

adolescent CRD and dose for motivation (breakpoint ratio) to self-administer cocaine in females, but no significant post-hocs were found (left). In males, there was a trend for adolescent CRD to reduce motivation to self-administer cocaine (right). (B) There was no effect of adolescent CRD on responding during extinction in female mice (left), while male mice with a history of CRD had reduced extinction responding (right). (C) For cueinduced reinstatement, responding was increased on the previously active lever in females with adolescent CRD (left), but no effect was found in males (right). Mean $+$ SEM, $n = 9-13$, $* p \le 0.05$.

 $(F_{(9,180)} = 2.30, p = 0.018)$. Males exposed to adolescent CRD responded less on the first session of extinction, compared to controls, indicating a possible decrease in frustrative non-reward or perseverative behavior on the previously active lever.

When we re-introduced the previously cocaine-associated cues, in the continued absence of intravenous cocaine, all mice exhibited cue-induced reinstatement by pressing the previously active lever more than the previously inactive lever (three-way ANOVA, main effect of lever, $F_{(1,37)} = 49.65$, $p < 0.0001$). Importantly, we also found a three-way interaction between lever, adolescent CRD and sex ([Figure 5C,](#page-9-0) $F_{(1,37)} = 5.38$, $p = 0.03$). In subsequent two-way ANOVAs, there was an interaction between the effects of adolescent CRD and

lever in females ($F_{(1,17)}$ = 3.57, p = 0.08) as well as a trend for an overall effect of adolescent CRD ($F_{(1,17)} = 4.12$, $p = 0.06$). Female mice exposed to adolescent CRD showed an increase in cue-induced reinstatement, as shown by increased responding on the active lever, compared to controls. In males, there was no effect of adolescent CRD on cue-induced reinstatement ($F_{(1,20)} = 2.50$, $p = 0.13$).

Although some behavioral differences were small, and we identified many trending effects, these changes taken together suggest phenotypic changes after adolescent CRD. Overall, males with adolescent CRD tended to respond less for intravenous cocaine, with less motivation and reduced responding during extinction, while females with adolescent CRD tended to respond more, and faster, for intravenous cocaine and exhibited an increase in cue-induced reinstatement.

3.5 Adolescent CRD sex-dependently alters gene expression in the SCN

These studies have shown that adolescent CRD alters behavior related to risk-taking, reward, and cocaine self-administration. These adolescent CRD-induced behavioral changes could be caused by lasting changes in gene expression in circadian or reward-related brain regions. Given that most of our behavior assessments were performed in the early- to mid-light phase, we investigated changes in gene expression after adolescent CRD at around that same time of day (ZT6). We used a DE analysis to examine what transcripts were altered in female and male CRD mice. We found many more DE transcripts when females and males were examined separately instead of when they are pooled ([Supplementary Table 1](#page-13-6)), suggesting distinct differences in these populations. Therefore, we did our analysis in each sex individually. In the SCN, 153 and 476 (*q* < 0.05) transcripts were DE in males and females, respectively ([Figure 6A\)](#page-11-0). When we examined these transcripts for enriched processes, we identified pathways in females involved in translation, microtubule-based movement, and membrane potential ([Figure 6B](#page-11-0)). In males, we identified processes regulating circadian rhythms, intracellular signaling, immune responses and axon development as significantly enriched ([Figure 6C\)](#page-11-0). In the NAc and PFC at ZT6, transcripts were not DE at *q* < 0.05 in CRD mice relative to controls. We performed an exploratory analysis ([Supplementary Figures 3–6](#page-13-6)) to identify potential DE transcripts in these regions at looser thresholds of $p < 0.05$ and $p < 0.01$ to investigate in future studies. See the [Supplementary materials](#page-13-6) for a description of this exploratory analysis.

4 Discussion

The goal of our study was to investigate the effects of adolescent CRD on psychiatric-related behaviors and gene expression later in life. Mice exposed to 12 h shifts in the LD schedule during early adolescence (P28-37) exhibited increased exploratory drive and impaired working memory in adulthood. Mice as prey animals typically avoid open, elevated, or brightly lit areas of behavioral apparatuses, thus our findings suggest that adolescent CRD increases risk-taking behavior in mice. The persistent increase in risk-taking in adolescent CRD exposed mice relative to controls does not appear to be driven by differences in activity since disrupted mice moved less in a locomotor assay in a novel environment.

Previous studies in rodents indicate that the behavioral effects of environmentally induced CRD depend on the type of light cycle manipulation, its timing, and duration. In adult rodents, light cycle manipulations like constant light exposure or a fragmented LD schedule with more light overall, have increased exploratory drive/ risk-taking behaviors in adult mice [\(Fonken et al., 2009;](#page-14-32) [Richardson](#page-15-38) [et al., 2021\)](#page-15-38). Interestingly, the effects of exposing adult mice to fragmented LD schedules on risk-taking were not persistent and reverted after 2 weeks. Similarly, we did not observe persistent changes in risk-taking behaviors in adult mice tested ~4 wks after our CRD paradigm. Similar changes have been found when light cycle manipulations occur in juvenile mice. Disrupting circadian rhythms before weaning leads to increased exploratory drive and impaired working memory in adulthood ([Borniger et al., 2014](#page-14-18); [Ameen et al.,](#page-13-5) [2022\)](#page-13-5). Some studies have found that LD cycle manipulations during adolescence alter exploratory drive in adult mice, but there have been few studies. For example, a chronic jet lag paradigm was shown to increase exploratory drive in the EPM, but this finding was specific to male Sandy mice, a line used to study aspects of schizophrenia ([Cloutier et al., 2022](#page-14-27)). On the other hand, in female adolescent mice, exposure to light at night decreased exploratory drive in the open field in adulthood [\(Chen et al., 2021b\)](#page-14-19). Here we found that 12 h shifts in the LD cycle during adolescence resulted in increased exploratory drive or risk-taking behavior in both sexes. The differences in the behavior findings in these studies may be due to differences in how phase shift paradigms and light at night during adolescence affect gene expression in the brain.

We also found that adolescent CRD impaired spatial working memory in adult mice while CRD in adulthood had no subsequent effect on spontaneous alternations in the T-maze. Interestingly, a recent study demonstrated that altering the photoperiod to model social jetlag in adolescent mice impaired novel object recognition and altered neural activity and gene expression in the hippocampus and amygdala ([Bonilla et al., 2024\)](#page-14-29). Similarly, exposing mice to a short-day LD cycle from gestation through P42 impaired memory and social behavior [\(Fang et al., 2021\)](#page-14-33). Furthermore, LD cycle disturbances have been shown to alter cognitive behavior in adult rodents, but it is unclear how long impairments would last once CRD ends ([Fernandez](#page-14-34) [et al., 2014](#page-14-34); [Fuchs et al., 2023\)](#page-14-35). Our results underscore the need for further research identifying the molecular links between CRD and different cognition behaviors across development.

Results from our group and others suggest that CRD at a critical period of development can have long-lasting effects on risky behavior and working memory in mouse models. However, more research is required to study the effects of environmental CRD on drug taking, especially for cocaine and opioids. While a few studies have shown that CRD affects drug intake, these studies have focused on alcohol and methamphetamine. For example, adolescent disruption escalates alcohol intake in adulthood, when animals have had prior exposure to alcohol [\(Gonzalez et al., 2023\)](#page-14-28). CRD has also been shown to increase alcohol intake in adolescents [\(Gamsby et al., 2017](#page-14-36)) and increase methamphetamine consumption in adult rats pre-exposed to methamphetamine ([Doyle et al., 2015\)](#page-14-37). These studies confirm that CRD can significantly affect drug taking, but whether these effects transfer to other drugs of abuse and are exacerbated if disruption occurs before adulthood is not well understood.

We began to answer these questions here. In addition to studying exploratory behaviors, we measured cocaine reward and intake using cocaine CPP and intravenous cocaine self-administration, respectively. We found that adolescent CRD persistently increased cocaine preference in adulthood, and sex-specifically altered intravenous

cocaine self-administration. Female adolescent CRD mice showed a greater propensity to self-administer cocaine, as well as increased motivation and cue-induced reinstatement. On the other hand, male adolescent CRD mice tended to self-administer less cocaine with reduced motivation and extinction responding. These results highlight the importance of studying volitional drug intake. While females showed a persistent increase in risky behavior across measures (exploratory drive, CPP and self-administration), males with a history of adolescent CRD showed a divergent behavioral response. Although unexpected, it is not unusual to find opposing results between conditioned place preference and self-administration ([Larson et al.,](#page-15-39) [2011](#page-15-39)) as in males here. These results suggest that although exploratory behaviors are increased in all mice with adolescent CRD, this unexpectedly only carries over to an increase in cocaine selfadministration in female mice. This might be due to an anhedonic-like response developing in males after adolescent CRD where volitional, reward taking, such as in sucrose preference and cocaine selfadministration, is reduced.

Further, it is possible that results were caused by stress due to the CRD rather than the CRD itself. However, other studies have demonstrated that stress during adolescence has different effects on behavior in adulthood. For example, chronic unpredictable stress during adolescence did not affect locomotor sensitization to amphetamine during adulthood ([Kabbaj et al., 2002\)](#page-14-38). Interestingly, other studies of sensitization following adolescence stress exposure reported similar findings for nicotine in males [\(McCormick and](#page-15-40) [Ibrahim, 2007](#page-15-40); [Cruz et al., 2008\)](#page-14-39). Stress during early adolescence has also been shown to increase anxiety-like behavior or reduce risktaking-like behavior in adulthood [\(Albrecht et al., 2017](#page-13-7)). It is important to note that not everyone that experiences adolescent stress goes on to develop a substance use or other disorder in adulthood. Therefore, it is essential to continue investigating different forms of adverse experiences during development to determine their specific impact.

To identify possible mechanisms by which CRD during adolescence affects behaviors later in life, we measured gene expression in brain regions relevant to circadian rhythms, mood, and reward, including the SCN, PFC, and NAc. Since we found sex-specific effects on drug-related behavior during the early/mid light phase, it is possible that changes at that TOD (ZT6) are driving behavioral changes. Therefore, we measured DE between control and adolescent CRD mice at ZT6 in male and female mice and found that the transcripts affected by adolescent CRD were largely distinct across sex and brain region. The SCN was particularly affected, with more DE transcripts identified at *q* < 0.05. Thus, our observed behavioral changes may be mediated by the SCN or its effects on downstream regions. Interestingly, the identified transcripts altered by CRD were non-overlapping between males and females, suggesting sex-dependent mechanisms. For example, synaptic transmission and microtubule/actin processes are enriched after CRD in females, while immune responses are especially prevalent in males. These differences could then cause further differences in downstream brain regions, including cortical or midbrain regions, possibly occurring via connections in the paraventricular nucleus (PVT).

One possible mechanism for changes downstream in the extra-SCN brain are modulations in hormone signaling, which is also a prevalent method for the SCN to synchronize rhythms in the brain and body. We found a general upregulation in processes related to peptide and hormone secretion and transport in the SCN, suggesting possible changes in important hormones, such as arginine vasopressin and corticosterone, that might affect downstream brain regions. It is important to note that downstream brain regions could also be responding to adolescent CRD independently from the SCN since the retina also transmits light information through the perihabenular nucleus of the thalamus [\(Fernandez et al., 2018\)](#page-14-40). Further studies are needed to better understand how changes in the SCN after adolescent CRD are affecting the rest of the brain, especially the PFC and NAc.

Downstream of the SCN we observed minor changes in the NAc and PFC that did not survive false discovery rate correction. Our exploratory analysis [\(Supplementary material](#page-13-6)) found several interesting results, but future studies with greater sample sizes and more power will be needed to confirm the relevance of these findings. At very minimal statistical cut-offs ($p < 0.05$), we found that in the PFC, transcripts with altered gene expression are associated with the extracellular matrix and kinase signaling in both males and females. These parallel changes may emphasize how critical neural activity and structure in the PFC is during adolescence when significant structural and synaptic maturation are occuring ([Bourgeois et al., 1994;](#page-14-41) [Huttenlocher and Dabholkar, 1997](#page-14-42); [Gourley et al., 2012](#page-14-43); [Koss et al.,](#page-14-44) [2014\)](#page-14-44). We also found many sex-specific changes, which could help explain the sex differences we found in reward-related behavior. These are discussed further in the [Supplementary material.](#page-13-6)

Taken together, the transcripts and enriched processes that were DE at ZT6 provide insight into how adolescent CRD might alter the SCN. We hypothesize that the SCN is directly impacted by light cycle changes and through changes in neuronal activity or neuropeptide/ hormone signaling, communicates those changes to reward-related brain regions, such as the PFC and NAc. However, it is important to note that since the changes found in the PFC and NAc did not reach significance thresholds, the SCN is likely predominately affecting other brain regions, or is affecting many regions, including the PFC and NAc in parallel. Small scale changes across many cortical or midbrain regions could act in concert to change behavior.

In future studies, it will also be critical to investigate how behavior and gene expression change across the 24 h day. Diurnal rhythms in mood- and reward-related behavior are extremely common, for example motivation and drug intake peak during the dark or active phase in nocturnal rodents [\(Baird and Gauvin, 2000](#page-13-8); [Abarca et al.,](#page-13-9) [2002;](#page-13-9) [Roberts et al., 2002;](#page-15-41) [Sleipness et al., 2005](#page-15-42)). Future studies might identify more pronounced behavioral effects after adolescent CRD if behavior is measured at other times.

In addition, while the CRD paradigm used here serves as a valuable model for how multiple, large disruptions to circadian rhythms during adolescence can have differential, long-lasting effects relative to adults, these 12 h LD shifts are not an environmental disruption commonly experienced by humans. Adolescents and adults experience large circadian/sleep disruptions due to school, work, and social obligations for years, not weeks and abnormal LD schedules are typically caused by indoor lighting at night and the use of electronics. In the future, more translationally relevant light paradigms could be used, such as constant light, fragmented light, or social jet lag paradigms, wherein animals are exposed to different schedules during the weekdays and weekends.

Overall, these experiments found that LD cycle disruptions persistently altered reward-related behavior, but only when disruption occurred during adolescence. Furthermore, females showed increased exploratory drive and reward-related behavior, while males showed increases in exploratory drive and cocaine preference, but reductions in volitional reward intake, such as sucrose preference and cocaine self-administration. This was the first study, to our knowledge, to measure cocaine self-administration in mice after environmental CRD. Intravenous self-administration is a highly translational behavioral model of substance use in mice and provides the best evidence of changes in propensity to abuse substances, drug taking and drug seeking in animal models. We also identified changes in gene expression after adolescent CRD in the SCN. These results emphasize how important and long-lasting the effects of CRD during adolescence are. Experiments like these are needed to identify novel intervention strategies to alleviate disruption during adolescence and new molecular targets for drug development.

Data availability statement

Raw and processed RNA-seq data presented in this study are deposited into the NCBI GEO database (accession no. GSE282275).

Ethics statement

The animal study was approved by University of Pittsburgh IACUC. The study was conducted in accordance with the local legislation and institutional requirements.

Author contributions

LD: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Visualization, Writing – original draft, Writing – review & editing. CV: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Visualization, Writing – original draft, Writing – review & editing, Funding acquisition. KP: Investigation, Methodology, Writing – original draft, Writing – review & editing. MS: Data curation, Writing – review & editing. WZ: Formal analysis, Writing – review & editing. RY: Writing – review & editing. RM: Formal analysis, Methodology, Writing – review & editing. FA: Investigation, Methodology, Writing – review & editing. CK: Formal analysis, Writing – review & editing. GT: Formal analysis, Writing – review & editing. CM: Conceptualization, Funding acquisition, Supervision, Writing – review & editing.

References

Abarca, C., Albrecht, U., and Spanagel, R. (2002). Cocaine sensitization and reward are under the influence of circadian genes and rhythm. *Proc. Natl. Acad. Sci.* 99, 9026–9030. doi: [10.1073/pnas.142039099](https://doi.org/10.1073/pnas.142039099)

Albrecht, A., Müller, I., Ardi, Z., Çalışkan, G., Gruber, D., Ivens, S., et al. (2017). Neurobiological consequences of juvenile stress: A GABAergic perspective on risk and resilience. Neurosci. Biobehav. Rev. 74, 21-43. doi: 10.1016/j. and resilience. *Neurosci. Biobehav. Rev.* 74, 21–43. doi: [10.1016/j.](https://doi.org/10.1016/j.neubiorev.2017.01.005) [neubiorev.2017.01.005](https://doi.org/10.1016/j.neubiorev.2017.01.005)

Ameen, R. W., Warshawski, A., Fu, L., and Antle, M. C. (2022). Early life circadian rhythm disruption in mice alters brain and behavior in adulthood. *Sci. Rep.* 12:7366. doi: [10.1038/s41598-022-11335-0](https://doi.org/10.1038/s41598-022-11335-0)

Andretic, R., Chaney, S., and Hirsh, J. (1999). Requirement of circadian genes for cocaine sensitization in Drosophila. *Science* 285, 1066–1068. doi: [10.1126/science.285.5430.1066](https://doi.org/10.1126/science.285.5430.1066)

Baird, T. J., and Gauvin, D. V. (2000). Characterization of cocaine Self-administration and pharmacokinetics as a function of time of day in the rat. *Pharmacol. Biochem. Behav.* 65, 289–299. doi: [10.1016/S0091-3057\(99\)00207-5](https://doi.org/10.1016/S0091-3057(99)00207-5)

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fnins.2024.1478508/](https://www.frontiersin.org/articles/10.3389/fnins.2024.1478508/full#supplementary-material) [full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fnins.2024.1478508/full#supplementary-material)

Baranger, D. A. A., Ifrah, C., Prather, A. A., Carey, C. E., Corral-Frías, N. S., Drabant Conley, E., et al. (2016). PER1 rs3027172 genotype interacts with early life stress to predict problematic alcohol use, but not reward-related ventral striatum activity. *Front. Psychol.* 7:464. doi: [10.3389/fpsyg.2016.00464](https://doi.org/10.3389/fpsyg.2016.00464)

Benedetti, F., Dallaspezia, S., Colombo, C., Pirovano, A., Marino, E., and Smeraldi, E. (2008). A length polymorphism in the circadian clock gene Per3 influences age at onset of bipolar disorder. *Neurosci. Lett.* 445, 184–187. doi: [10.1016/j.neulet.2008.09.002](https://doi.org/10.1016/j.neulet.2008.09.002)

Benedetti, F., Serretti, A., Colombo, C., Barbini, B., Lorenzi, C., Campori, E., et al. (2003). Influence of CLOCK gene polymorphism on circadian mood fluctuation and illness recurrence in bipolar depression. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 123B, 23–26. doi: [10.1002/ajmg.b.20038](https://doi.org/10.1002/ajmg.b.20038)

Ben-Hamo, M., Larson, T. A., Duge, L. S., Sikkema, C., Wilkinson, C. W., de la Iglesia, H. O., et al. (2016). Circadian forced Desynchrony of the master clock leads to phenotypic manifestation of depression in rats. *eNeuro* 3. doi: [10.1523/](https://doi.org/10.1523/ENEURO.0237-16.2016) [ENEURO.0237-16.2016](https://doi.org/10.1523/ENEURO.0237-16.2016)

Bi, J., Gelernter, J., Sun, J., and Kranzler, H. R. (2014). Comparing the utility of homogeneous subtypes of cocaine use and related behaviors with DSM-IV cocaine dependence as traits for genetic association analysis. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 165, 148–156. doi: [10.1002/ajmg.b.32216](https://doi.org/10.1002/ajmg.b.32216)

Blomeyer, D., Buchmann, A. F., Lascorz, J., Zimmermann, U. S., Esser, G., Desrivieres, S., et al. (2013). Association of PER2 genotype and stressful life events with alcohol drinking in young adults. *PLoS One* 8:e59136. d

Boehler, N. A., Fung, S. W., Hegazi, S., Cheng, A. H., and Cheng, H.-Y. M. (2021). Sox2 ablation in the suprachiasmatic nucleus perturbs anxiety- and depressive-like behaviors. *Neurol. Int.* 13, 541–554. doi: [10.3390/neurolint13040054](https://doi.org/10.3390/neurolint13040054)

Boergers, J., Gable, C. J., and Owens, J. A. (2014). Later school start time is associated with improved sleep and daytime functioning in adolescents. *J. Dev. Behav. Pediatr.* 35, 11–17. doi: [10.1097/DBP.0000000000000018](https://doi.org/10.1097/DBP.0000000000000018)

Bonilla, P., Shanks, A., Nerella, Y., and Porcu, A. (2024). Effects of chronic light cycle disruption during adolescence on circadian clock, neuronal activity rhythms, and behavior in mice. *Front. Neurosci.* 18:1418694. doi: [10.3389/fnins.2024.1418694](https://doi.org/10.3389/fnins.2024.1418694)

Borisenkov, M. F., Popov, S. V., Smirnov, V. V., Dorogina, O. I., Pechеrkina, A. A., and Symaniuk, E. E. (2022). Later school start time is associated with better academic performance, sleep-wake rhythm characteristics, and eating behavior. *Chronobiol. Int.* 39, 1444–1453. doi: [10.1080/07420528.2022.2117050](https://doi.org/10.1080/07420528.2022.2117050)

Borniger, J. C., McHenry, Z. D., Abi Salloum, B. A., and Nelson, R. J. (2014). Exposure to dim light at night during early development increases adult anxiety-like responses. *Physiol. Behav.* 133, 99–106. doi: [10.1016/j.physbeh.2014.05.012](https://doi.org/10.1016/j.physbeh.2014.05.012)

Bourgeois, J.-P., Goldman-Rakic, P. S., and Rakic, P. (1994). Synaptogenesis in the prefrontal cortex of Rhesus monkeys. *Cereb. Cortex* 4, 78–96. doi: [10.1093/cercor/4.1.78](https://doi.org/10.1093/cercor/4.1.78)

Chen, R., Weitzner, A. S., McKennon, L. A., and Fonken, L. K. (2021a). Chronic circadian phase advance in male mice induces depressive-like responses and suppresses neuroimmune activation. *Brain Behav. Immun. - Health* 17:100337. doi: [10.1016/j.](https://doi.org/10.1016/j.bbih.2021.100337) [bbih.2021.100337](https://doi.org/10.1016/j.bbih.2021.100337)

Chen, R., Weitzner, A. S., McKennon, L. A., and Fonken, L. K. (2021b). Light at night during development in mice has modest effects on adulthood behavior and neuroimmune activation. *Behav. Brain Res.* 405:113171. doi: [10.1016/j.bbr.2021.113171](https://doi.org/10.1016/j.bbr.2021.113171)

Cloutier, M.-È., Srivastava, L. K., and Cermakian, N. (2022). Exposure to circadian disruption during adolescence interacts with a genetic risk factor to modify schizophrenia-relevant behaviors in a sex-dependent manner. *J. Biol. Rhythm.* 37, 655–672. doi: [10.1177/07487304221125363](https://doi.org/10.1177/07487304221125363)

Crowley, S. J., Van Reen, E., LeBourgeois, M. K., Acebo, C., Tarokh, L., Seifer, R., et al. (2014). A longitudinal assessment of sleep timing, circadian phase, and phase angle of entrainment across human adolescence. *PLoS One* 9:e112199. doi: [10.1371/journal.](https://doi.org/10.1371/journal.pone.0112199) [pone.0112199](https://doi.org/10.1371/journal.pone.0112199)

Cruz, F. C., DeLucia, R., and Planeta, C. S. (2008). PRECLINICAL STUDY: effects of chronic stress on nicotine-induced locomotor activity and corticosterone release in adult and adolescent rats. *Addict. Biol.* 13, 63–69. doi: [10.1111/j.1369-1600.2007.00080.x](https://doi.org/10.1111/j.1369-1600.2007.00080.x)

DePoy, L. M., Becker-Krail, D. D., Zong, W., Petersen, K., Shah, N. M., Brandon, J. H., et al. (2020). Circadian- and sex-dependent increases in intravenous cocaine selfadministration in Npas2 mutant mice. *J. Neurosci.* 41, 1046–1058. doi: [10.1523/JNEUROSCI.1830-20.2020](https://doi.org/10.1523/JNEUROSCI.1830-20.2020)

DePoy, L. M., McClung, C. A., and Logan, R. W. (2017). Neural mechanisms of circadian regulation of natural and drug reward. *Neural Plast.* 2017, 1–14. doi: [10.1155/2017/5720842](https://doi.org/10.1155/2017/5720842)

Dong, L., Bilbao, A., Laucht, M., Henriksson, R., Yakovleva, T., Ridinger, M., et al. (2011). Effects of the circadian rhythm gene period 1 (Per1) on psychosocial stress-induced alcohol drinking. *Am. J. Psychiatry* 168, 1090–1098. doi: [10.1176/appi.ajp.2011.10111579](https://doi.org/10.1176/appi.ajp.2011.10111579)

Doyle, S. E., Feng, H., Garber, G., Menaker, M., and Lynch, W. J. (2015). Effects of circadian disruption on methamphetamine consumption in methamphetamine-exposed rats. *Psychopharmacology* 232, 2169–2179. doi: [10.1007/s00213-014-3845-2](https://doi.org/10.1007/s00213-014-3845-2)

Easton, A., Arbuzova, J., and Turek, F. W. (2003). The circadian clock mutation increases exploratory activity and escape-seeking behavior. *Genes Brain Behav.* 2, 11–19. doi: [10.1034/j.1601-183X.2003.00002.x](https://doi.org/10.1034/j.1601-183X.2003.00002.x)

Emens, J., Lewy, A., Kinzie, J. M., Arntz, D., and Rough, J. (2009). Circadian misalignment in major depressive disorder. *Psychiatry Res.* 168, 259–261. doi: [10.1016/j.](https://doi.org/10.1016/j.psychres.2009.04.009) [psychres.2009.04.009](https://doi.org/10.1016/j.psychres.2009.04.009)

Falcón, E., and McClung, C. A. (2009). A role for the circadian genes in drug addiction. *Neuropharmacology* 56, 91–96. doi: [10.1016/j.neuropharm.2008.06.054](https://doi.org/10.1016/j.neuropharm.2008.06.054)

Fang, K., Liu, D., Pathak, S. S., Yang, B., Li, J., Karthikeyan, R., et al. (2021). Disruption of circadian rhythms by ambient light during neurodevelopment leads to autistic-like molecular and behavioral alterations in adult mice. *Cells* 10:3314. doi: [10.3390/](https://doi.org/10.3390/cells10123314) [cells10123314](https://doi.org/10.3390/cells10123314)

Fernandez, D. C., Fogerson, P. M., Lazzerini Ospri, L., Thomsen, M. B., Layne, R. M., Severin, D., et al. (2018). Light affects mood and learning through distinct retina-brain pathways. *Cell* 175:e18, 71–84.e18. doi: [10.1016/j.](https://doi.org/10.1016/j.cell.2018.08.004) [cell.2018.08.004](https://doi.org/10.1016/j.cell.2018.08.004)

Fernandez, F., Lu, D., Ha, P., Costacurta, P., Chavez, R., Heller, H. C., et al. (2014). Dysrhythmia in the suprachiasmatic nucleus inhibits memory processing. *Science* 346, 854–857. doi: [10.1126/science.1259652](https://doi.org/10.1126/science.1259652)

Fonken, L. K., Finy, M. S., Walton, J. C., Weil, Z. M., Workman, J. L., Ross, J., et al. (2009). Influence of light at night on murine anxiety- and depressive-like responses. *Behav. Brain Res.* 205, 349–354. doi: [10.1016/j.bbr.2009.07.001](https://doi.org/10.1016/j.bbr.2009.07.001)

Forbes, E. E., Dahl, R. E., Almeida, J. R. C., Ferrell, R. E., Nimgaonkar, V. L., Mansour, H., et al. (2012). *PER2 rs2304672* polymorphism moderates circadian-relevant reward circuitry activity in adolescents. *Biol. Psychiatry* 71, 451–457. doi: [10.1016/j.](https://doi.org/10.1016/j.biopsych.2011.10.012) [biopsych.2011.10.012](https://doi.org/10.1016/j.biopsych.2011.10.012)

Fuchs, F., Robin-Choteau, L., Schneider, A., Hugueny, L., Ciocca, D., Serchov, T., et al. (2023). Delaying circadian sleep phase under ultradian light cycle causes time-of-daydependent alteration of cognition and mood. *Sci. Rep.* 13:20313. doi: [10.1038/](https://doi.org/10.1038/s41598-023-44931-9) [s41598-023-44931-9](https://doi.org/10.1038/s41598-023-44931-9)

Gamsby, J. J., Pribish, A. M., Stevanovic, K. D., Yunus, A., and Gulick, D. (2017). Alcohol intake increases in adolescent C57BL/6J mice during intermittent cycles of phase-delayed, Long-light conditions. *Front. Behav. Neurosci.* 11:152. doi: [10.3389/](https://doi.org/10.3389/fnbeh.2017.00152) [fnbeh.2017.00152](https://doi.org/10.3389/fnbeh.2017.00152)

Gamsby, J. J., Templeton, E. L., Bonvini, L. A., Wang, W., Loros, J. J., Dunlap, J. C., et al. (2013). The circadian *Per1* and *Per2* genes influence alcohol intake, reinforcement, and blood alcohol levels. *Behav. Brain Res.* 249, 15–21. doi: [10.1016/j.bbr.2013.04.016](https://doi.org/10.1016/j.bbr.2013.04.016)

Golub, M. S., Takeuchi, P. T., Keen, C. L., Hendrickx, A. G., and Gershwin, M. E. (1996). Activity and attention in zinc-deprived adolescent monkeys. *Am. J. Clin. Nutr.* 64, 908–915. doi: [10.1093/ajcn/64.6.908](https://doi.org/10.1093/ajcn/64.6.908)

Gonzalez, D., Justin, H., Reiss, S., Faulkner, J., Mahoney, H., Yunus, A., et al. (2023). Circadian rhythm shifts and alcohol access in adolescence synergistically increase alcohol preference and intake in adulthood in male C57BL/6 mice. *Behav. Brain Res.* 438:114216. doi: [10.1016/j.bbr.2022.114216](https://doi.org/10.1016/j.bbr.2022.114216)

Gourley, S. L., Olevska, A., Warren, M. S., Taylor, J. R., and Koleske, A. J. (2012). Arg kinase regulates prefrontal dendritic spine refinement and cocaine-induced plasticity. *J. Neurosci.* 32, 2314–2323. doi: [10.1523/JNEUROSCI.2730-11.2012](https://doi.org/10.1523/JNEUROSCI.2730-11.2012)

Gowen, R., Filipowicz, A., and Ingram, K. K. (2019). Chronotype mediates gender differences in risk propensity and risk-taking. *PLoS One* 14:e0216619. doi: [10.1371/](https://doi.org/10.1371/journal.pone.0216619) [journal.pone.0216619](https://doi.org/10.1371/journal.pone.0216619)

Hagenauer, M. H., King, A. F., Possidente, B., McGinnis, M. Y., Lumia, A. R., Peckham, E. M., et al. (2011). Changes in circadian rhythms during puberty in *Rattus norvegicus*: developmental time course and gonadal dependency. *Horm. Behav.* 60, 46–57. doi: [10.1016/j.yhbeh.2011.03.001](https://doi.org/10.1016/j.yhbeh.2011.03.001)

Hasler, B. P., Kirisci, L., and Clark, D. B. (2016). Restless sleep and variable sleep timing during late childhood accelerate the onset of alcohol and other drug involvement. *Journal of studies on alcohol and drugs*, 77, 649–655. doi: [10.15288/jsad.2016.77.649](https://doi.org/10.15288/jsad.2016.77.649)

Hasler, B. P., Casement, M. D., Sitnick, S. L., Shaw, D. S., and Forbes, E. E. (2017). Eveningness among late adolescent males predicts neural reactivity to reward and alcohol dependence 2 years later. *Behav. Brain Res.* 327, 112–120. doi: [10.1016/j.](https://doi.org/10.1016/j.bbr.2017.02.024) [bbr.2017.02.024](https://doi.org/10.1016/j.bbr.2017.02.024)

Hummer, D. L., Jechura, T. J., Mahoney, M. M., and Lee, T. M. (2007). Gonadal hormone effects on entrained and free-running circadian activity rhythms in the developing diurnal rodent *Octodon degus*. *Am. J. Physiol.-Regul. Integr. Comp. Physiol.* 292, R586–R597. doi: [10.1152/ajpregu.00043.2006](https://doi.org/10.1152/ajpregu.00043.2006)

Huttenlocher, P. R., and Dabholkar, A. S. (1997). Regional differences in synaptogenesis in human cerebral cortex. *J. Comp. Neurol.* 387, 167–178. doi: [10.1002/\(SICI\)1096-9861](https://doi.org/10.1002/(SICI)1096-9861(19971020)387:2<167::AID-CNE1>3.0.CO;2-Z) [\(19971020\)387:2<167::AID-CNE1>3.0.CO;2-Z](https://doi.org/10.1002/(SICI)1096-9861(19971020)387:2<167::AID-CNE1>3.0.CO;2-Z)

Iglesia, H. O., La, D., Cambras, T., and Schwartz, W. J. (2004). Forced desynchronization of dual circadian oscillators within the rat suprachiasmatic nucleus. *Curr. Biol.* 14, 796–800. doi: [10.1016/j.cub.2004.04.034](https://doi.org/10.1016/j.cub.2004.04.034)

Jameson, A. N., Siemann, J. K., Grueter, C. A., Grueter, B. A., and McMahon, D. G. (2024). Effects of age and sex on photoperiod modulation of nucleus accumbens monoamine content and release in adolescence and adulthood. Neurobiol. Sleep Circadian Rhythms 16:100103. doi: [10.1016/j.nbscr.2024.100103](https://doi.org/10.1016/j.nbscr.2024.100103)

Johansson, C., Willeit, M., Smedh, C., Ekholm, J., Paunio, T., Kieseppä, T., et al. (2003). Circadian clock-related polymorphisms in seasonal affective disorder and their relevance to diurnal preference. *Neuropsychopharmacology* 28, 734–739. doi: [10.1038/](https://doi.org/10.1038/sj.npp.1300121) si.npp.1300121

Kabbaj, M., Isgor, C., Watson, S. J., and Akil, H. (2002). Stress during adolescence alters behavioral sensitization to amphetamine. *Neuroscience* 113, 395–400. doi: [10.1016/](https://doi.org/10.1016/S0306-4522(02)00188-4) [S0306-4522\(02\)00188-4](https://doi.org/10.1016/S0306-4522(02)00188-4)

Karatsoreos, I. N., Bhagat, S., Bloss, E. B., Morrison, J. H., and McEwen, B. S. (2011). Disruption of circadian clocks has ramifications for metabolism, brain, and behavior. *Proc. Natl. Acad. Sci.* 108, 1657–1662. doi: [10.1073/pnas.1018375108](https://doi.org/10.1073/pnas.1018375108)

Kim, S.-M., Neuendorff, N., Alaniz, R. C., Sun, Y., Chapkin, R. S., and Earnest, D. J. (2018). Shift work cycle-induced alterations of circadian rhythms potentiate the effects of high-fat diet on inflammation and metabolism. *FASEB J.* 32, 3085–3095. doi: [10.1096/](https://doi.org/10.1096/fj.201700784R) [fj.201700784R](https://doi.org/10.1096/fj.201700784R)

Ko, C. H., and Takahashi, J. S. (2006). Molecular components of the mammalian circadian clock. *Hum. Mol. Genet.* 15, R271–R277. doi: [10.1093/hmg/ddl207](https://doi.org/10.1093/hmg/ddl207)

Koss, W. A., Belden, C. E., Hristov, A. D., and Juraska, J. M. (2014). Dendritic remodeling in the adolescent medial prefrontal cortex and the basolateral amygdala of male and female rats. *Synapse* 68, 61–72. doi: [10.1002/syn.21716](https://doi.org/10.1002/syn.21716)

Landgraf, D., Long, J. E., and Welsh, D. K. (2016). Depression-like behaviour in mice is associated with disrupted circadian rhythms in nucleus accumbens and periaqueductal grey. *Eur. J. Neurosci.* 43, 1309–1320. doi: [10.1111/ejn.13085](https://doi.org/10.1111/ejn.13085)

Larson, E. B., Graham, D. L., Arzaga, R. R., Buzin, N., Webb, J., Green, T. A., et al. (2011). Overexpression of CREB in the nucleus Accumbens Shell increases cocaine reinforcement in Self-administering rats. *J. Neurosci.* 31, 16447–16457. doi: [10.1523/](https://doi.org/10.1523/JNEUROSCI.3070-11.2011) [JNEUROSCI.3070-11.2011](https://doi.org/10.1523/JNEUROSCI.3070-11.2011)

Lavebratt, C., Sjöholm, L. K., Soronen, P., Paunio, T., Vawter, M. P., Bunney, W. E., et al. (2010). CRY2 is associated with depression. *PLoS One* 5:e9407. doi: [10.1371/journal.](https://doi.org/10.1371/journal.pone.0009407) [pone.0009407](https://doi.org/10.1371/journal.pone.0009407)

LeGates, T. A., Altimus, C. M., Wang, H., Lee, H.-K., Yang, S., Zhao, H., et al. (2012). Aberrant light directly impairs mood and learning through melanopsin-expressing neurons. *Nature* 491, 594–598. doi: [10.1038/nature11673](https://doi.org/10.1038/nature11673)

Lewy, A. J. (2010). Depressive disorders may more commonly be related to circadian phase delays rather than advances: time will tell. *Sleep Med.* 11, 117–118. doi: [10.1016/j.](https://doi.org/10.1016/j.sleep.2009.09.002) [sleep.2009.09.002](https://doi.org/10.1016/j.sleep.2009.09.002)

Logan, R. W., Williams, W. P. III, and McClung, C. A. (2014). Circadian rhythms and addiction: mechanistic insights and future directions. *Behav. Neurosci.* 128, 387–412. doi: [10.1037/a0036268](https://doi.org/10.1037/a0036268)

Mansour, H. A., Talkowski, M. E., Wood, J., Chowdari, K. V., McClain, L., Prasad, K., et al. (2009). Association study of 21 circadian genes with bipolar I disorder, schizoaffective disorder, and schizophrenia. *Bipolar Disord.* 11, 701–710. doi: [10.1111/j.](https://doi.org/10.1111/j.1399-5618.2009.00756.x) [1399-5618.2009.00756.x](https://doi.org/10.1111/j.1399-5618.2009.00756.x)

McClung, C. A. (2007). Circadian genes, rhythms and the biology of mood disorders. *Pharmacol. Ther.* 114, 222–232. doi: [10.1016/j.pharmthera.2007.02.003](https://doi.org/10.1016/j.pharmthera.2007.02.003)

McCormick, C. M., and Ibrahim, F. N. (2007). Locomotor activity to nicotine and Fos immunoreactivity in the paraventricular nucleus of the hypothalamus in adolescent sociallystressed rats. *Pharmacol. Biochem. Behav.* 86, 92–102. doi: [10.1016/j.pbb.2006.12.012](https://doi.org/10.1016/j.pbb.2006.12.012)

McGowan, N. M., and Coogan, A. N. (2013). Circadian and behavioural responses to shift work-like schedules of light/dark in the mouse. *J. Mol. Psychiatry* 1:7. doi: [10.1186/2049-9256-1-7](https://doi.org/10.1186/2049-9256-1-7)

Merikanto, I., Kronholm, E., Peltonen, M., Laatikainen, T., Vartiainen, E., and Partonen, T. (2015). Circadian preference links to depression in general adult population. *J. Affect. Disord.* 188, 143–148. doi: [10.1016/j.jad.2015.08.061](https://doi.org/10.1016/j.jad.2015.08.061)

Mills, K. L., Goddings, A.-L., Clasen, L. S., Giedd, J. N., and Blakemore, S.-J. (2014). The developmental mismatch in structural brain maturation during adolescence. *Dev. Neurosci.* 36, 147–160. doi: [10.1159/000362328](https://doi.org/10.1159/000362328)

Otsuka, T., Thi Le, H., Kohsaka, A., Sato, F., Ihara, H., Nakao, T., et al. (2020). Adverse effects of circadian disorganization on mood and molecular rhythms in the prefrontal cortex of mice. *Neuroscience* 432, 44–54. doi: [10.1016/j.neuroscience.2020.02.013](https://doi.org/10.1016/j.neuroscience.2020.02.013)

Owens, J. A., Belon, K., and Moss, P. (2010). Impact of delaying school start time on adolescent sleep, mood, and behavior. *Arch. Pediatr. Adolesc. Med.* 164, 608–614. doi: [10.1001/archpediatrics.2010.96](https://doi.org/10.1001/archpediatrics.2010.96)

Ozburn, A. R., Falcon, E., Mukherjee, S., Gillman, A., Arey, R., Spencer, S., et al. (2013). The role of clock in ethanol-related behaviors. *Neuropsychopharmacology* 38, 2393–2400. doi: [10.1038/npp.2013.138](https://doi.org/10.1038/npp.2013.138)

Ozburn, A. R., Falcon, E., Twaddle, A., Nugent, A. L., Gillman, A. G., Spencer, S. M., et al. (2015). Direct regulation of diurnal Drd3 expression and cocaine reward by NPAS2. *Biol. Psychiatry* 77, 425–433. doi: [10.1016/j.biopsych.2014.07.030](https://doi.org/10.1016/j.biopsych.2014.07.030)

Ozburn, A. R., Kern, J., Parekh, P. K., Logan, R. W., Liu, Z., Falcon, E., et al. (2017). NPAS2 regulation of anxiety-like behavior and GABAA receptors. *Front. Mol. Neurosci.* 10:360. doi: [10.3389/fnmol.2017.00360](https://doi.org/10.3389/fnmol.2017.00360)

Ozburn, A. R., Larson, E. B., Self, D. W., and McClung, C. A. (2012). Cocaine selfadministration behaviors in ClockΔ19 mice. *Psychopharmacology* 223, 169–177. doi: [10.1007/s00213-012-2704-2](https://doi.org/10.1007/s00213-012-2704-2)

Parekh, P. K., Logan, R. W., Ketchesin, K. D., Becker-Krail, D., Shelton, M. A., Hildebrand, M. A., et al. (2019). Cell-type-specific regulation of nucleus Accumbens synaptic plasticity and cocaine reward sensitivity by the circadian protein, NPAS2. *J. Neurosci.* 39, 4657–4667. doi: [10.1523/JNEUROSCI.2233-18.2019](https://doi.org/10.1523/JNEUROSCI.2233-18.2019)

Parekh, P. K., and McClung, C. A. (2016). Circadian mechanisms underlying rewardrelated neurophysiology and synaptic plasticity. *Front. Psychol.* 6:187. doi: [10.3389/](https://doi.org/10.3389/fpsyt.2015.00187) [fpsyt.2015.00187](https://doi.org/10.3389/fpsyt.2015.00187)

Pasch, K. E., Latimer, L. A., Cance, J. D., Moe, S. G., and Lytle, L. A. (2012). Longitudinal bi-directional relationships between sleep and youth substance use. *Journal of youth and adolescence*, 41, 1184–1196. doi: [10.1007/s10964-012-9784-5](https://doi.org/10.1007/s10964-012-9784-5)

Reppert, S. M., and Weaver, D. R. (2002). Coordination of circadian timing in mammals. *Nature* 418, 935–941. doi: [10.1038/nature00965](https://doi.org/10.1038/nature00965)

Richardson, M. E. S., Brown, D., Honore, D., and Labossiere, A. (2021). Fragmented day-night cycle induces period lengthening, lowered anxiety, and anhedonia in male mice. *Behav. Brain Res.* 413:113453. doi: [10.1016/j.](https://doi.org/10.1016/j.bbr.2021.113453) [bbr.2021.113453](https://doi.org/10.1016/j.bbr.2021.113453)

Roberts, D. C. S., Brebner, K., Vincler, M., and Lynch, W. J. (2002). Patterns of cocaine self-administration in rats produced by various access conditions under a discrete trials procedure. *Drug Alcohol Depend.* 67, 291–299. doi: [10.1016/](https://doi.org/10.1016/S0376-8716(02)00083-2) [S0376-8716\(02\)00083-2](https://doi.org/10.1016/S0376-8716(02)00083-2)

Roberts, B. L., and Karatsoreos, I. N. (2023). Circadian desynchronization disrupts physiological rhythms of prefrontal cortex pyramidal neurons in mice. *Sci. Rep.* 13:9181. doi: [10.1038/s41598-023-35898-8](https://doi.org/10.1038/s41598-023-35898-8)

Roenneberg, T., Kuehnle, T., Pramstaller, P. P., Ricken, J., Havel, M., Guth, A., et al. (2004). A marker for the end of adolescence. *Curr. Biol.* 14, R1038–R1039. doi: [10.1016/j.](https://doi.org/10.1016/j.cub.2004.11.039) [cub.2004.11.039](https://doi.org/10.1016/j.cub.2004.11.039)

Roybal, K., Theobold, D., Graham, A., DiNieri, J. A., Russo, S. J., Krishnan, V., et al. (2007). Mania-like behavior induced by disruption of CLOCK. *Proc. Natl. Acad. Sci.* 104, 6406–6411. doi: [10.1073/pnas.0609625104](https://doi.org/10.1073/pnas.0609625104)

Sleipness, E. P., Sorg, B. A., and Jansen, H. T. (2005). Time of day alters long-term sensitization to cocaine in rats. *Brain Res.* 1065, 132–137. doi: [10.1016/j.](https://doi.org/10.1016/j.brainres.2005.10.017) [brainres.2005.10.017](https://doi.org/10.1016/j.brainres.2005.10.017)

Souêtre, E., Salvati, E., Belugou, J.-L., Pringuey, D., Candito, M., Krebs, B., et al. (1989). Circadian rhythms in depression and recovery: evidence for blunted amplitude as the main chronobiological abnormality. *Psychiatry Res.* 28, 263–278. doi: [10.1016/0165-1781\(89\)90207-2](https://doi.org/10.1016/0165-1781(89)90207-2)

Spear, L. P. (2000). The adolescent brain and age-related behavioral manifestations. *Neurosci. Biobehav. Rev.* 24, 417–463. doi: [10.1016/s0149-7634\(00\)00014-2](https://doi.org/10.1016/s0149-7634(00)00014-2)

Steinberg, L., and Silverberg, S. B. (1986). The vicissitudes of autonomy in early adolescence. *Child Dev.* 57, 841–851. doi: [10.2307/1130361](https://doi.org/10.2307/1130361)

Takahashi, J. S. (2017). Transcriptional architecture of the mammalian circadian clock. *Nat. Rev. Genet.* 18, 164–179. doi: [10.1038/nrg.2016.150](https://doi.org/10.1038/nrg.2016.150)

Taylor, B. J., Bowman, M. A., Brindle, A., Hasler, B. P., Roecklein, K. A., Krafty, R. T., et al. (2020). Evening chronotype, alcohol use disorder severity, and emotion regulation in college students. *Chronobiol. Int.* 37, 1725–1735. doi: [10.1080/07420528.2020.1800028](https://doi.org/10.1080/07420528.2020.1800028)

Thorleifsdottir, B., Björnsson, J. K., Benediktsdottir, B., Gislason, T. H., and Kristbjarnarson, H. (2002). Sleep and sleep habits from childhood to young adulthood over a 10-year period. *J. Psychosom. Res.* 53, 529–537. doi: [10.1016/S0022-3999\(02\)00444-0](https://doi.org/10.1016/S0022-3999(02)00444-0)

Touitou, Y. (2013). Adolescent sleep misalignment: a chronic jet lag and a matter of public health. *J. Physiol.-Paris* 107, 323–326. doi: [10.1016/j.jphysparis.2013.03.008](https://doi.org/10.1016/j.jphysparis.2013.03.008)

Vadnie, C. A., and McClung, C. A. (2017). Circadian rhythm disturbances in mood disorders: insights into the role of the suprachiasmatic nucleus. *Neural Plast.* 2017:e1504507, 1–28. doi: [10.1155/2017/1504507](https://doi.org/10.1155/2017/1504507)

Vadnie, C. A., Petersen, K. A., Eberhardt, L. A., Hildebrand, M. A., Cerwensky, A. J., Zhang, H., et al. (2022). The suprachiasmatic nucleus regulates anxiety-like behavior in mice. *Front. Neurosci.* 15:765850. doi: [10.3389/fnins.2021.765850](https://doi.org/10.3389/fnins.2021.765850)

Wahistrom, K. (2002). Changing times: findings from the first longitudinal study of later high school start times. *NASSP Bull.* 86, 3–21. doi: [10.1177/019263650208663302](https://doi.org/10.1177/019263650208663302)

Whylings, J., Rigney, N., de Vries, G. J., and Petrulis, A. (2021). Reduction in vasopressin cells in the suprachiasmatic nucleus in mice increases anxiety and alters fluid intake. *Horm. Behav.* 133:104997. doi: [10.1016/j.yhbeh.2021.104997](https://doi.org/10.1016/j.yhbeh.2021.104997)

Wittmann, M., Dinich, J., Merrow, M., and Roenneberg, T. (2006). Social jetlag: misalignment of biological and social time. *Chronobiol. Int.* 23, 497–509. doi: [10.1080/07420520500545979](https://doi.org/10.1080/07420520500545979)

Yan, L. (2011). Structural and functional changes in the suprachiasmatic nucleus following chronic circadian rhythm perturbation. *Neuroscience* 183, 99–107. doi: [10.1016/j.neuroscience.2011.03.041](https://doi.org/10.1016/j.neuroscience.2011.03.041)

Yang, C.-K., Kim, J. K., Patel, S. R., and Lee, J.-H. (2005). Age-related changes in sleep/ wake patterns among Korean teenagers. *Pediatrics* 115, 250–256. doi: [10.1542/](https://doi.org/10.1542/peds.2004-0815G) [peds.2004-0815G](https://doi.org/10.1542/peds.2004-0815G)

Zhou, Y., Zhou, B., Pache, L., Chang, M., Khodabakhshi, A. H., Tanaseichuk, O., et al. (2019). Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat. Commun.* 10:1523. doi: [10.1038/s41467-019-09234-6](https://doi.org/10.1038/s41467-019-09234-6)