Check for updates

OPEN ACCESS

EDITED BY Luis de Lecea, Stanford University, United States

REVIEWED BY Bradley Kerr, University of Alberta, Canada Peter Drummond, Murdoch University, Australia Brooke Schmeichel, East Tennessee State University, United States

*CORRESPONDENCE Alban Latremoliere alatrem1@jhmi.edu Chloe Alexandre calexa41@jhmi.edu

[†]These authors share senior authorship

SPECIALTY SECTION This article was submitted to Sleep and Circadian Rhythms, a section of the journal Frontiers in Neuroscience

RECEIVED 02 August 2022 ACCEPTED 29 November 2022 PUBLISHED 20 December 2022

CITATION

Kourbanova K, Alexandre C and Latremoliere A (2022) Effect of sleep loss on pain—New conceptual and mechanistic avenues. *Front. Neurosci.* 16:1009902. doi: 10.3389/fnins.2022.1009902

COPYRIGHT

© 2022 Kourbanova, Alexandre and Latremoliere. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Effect of sleep loss on pain—New conceptual and mechanistic avenues

Kamila Kourbanova^{1,2}, Chloe Alexandre^{1,2*†} and Alban Latremoliere^{1,2*†}

¹Department of Neurosurgery, Neurosurgery Pain Research Institute, Johns Hopkins School of Medicine, Baltimore, MD, United States, ²Department of Neuroscience, Johns Hopkins School of Medicine, Baltimore, MD, United States

Introduction: Sleep disturbances increase pain sensitivity in clinical and preclinical settings, but the precise mechanisms are unknown. This represents a major public health issue because of the growing sleep deficiency epidemic fueled by modern lifestyle. To understand the neural pathways at the intersection between sleep and pain processes, it is critical to determine the precise nature of the sleep disruptions that increase pain and the specific component of the pain response that is targeted.

Methods: We performed a review of the literature about sleep disturbances and pain sensitivity in humans and rodents by taking into consideration the targeted sleep stage (REMS, non–NREMS, or both), the amount of sleep lost, and the different types of sleep disruptions (partial or total sleep loss, duration, sleep fragmentation or interruptions), and how these differences might affect distinct components of the pain response.

Results: We find that the effects of sleep disturbances on pain are highly conserved among species. The major driver for pain hypersensitivity appears to be the total amount of sleep lost, while REMS loss by itself does not seem to have a direct effect on pain sensitivity. Sleep loss caused by extended wakefulness preferentially increases pain perception, whereas interrupted and limited sleep strongly dysregulates descending controls such as DNIC, especially in women.

Discussion: We discuss the possible mechanisms involved, including an increase in inflammatory processes, a loss of nociceptive inhibitory pathways, and a defect in the cognitive processing of noxious input.

KEYWORDS

sleep deprivation, pain, nucleus accumbens, DNIC, NREMS, REMS, forced awakening, SWS

Introduction

When individuals do not get sufficient sleep or have their sleep curtailed, they fail to maintain normal levels of alertness, which translates into attentional lapses and poor cognitive performances (Van Dongen et al., 2003; Goel et al., 2009; McHill et al., 2018). Insufficient sleep also negatively impacts metabolism

(Spiegel et al., 1999; Tasali et al., 2008), immune and cardiovascular functions (Faraut et al., 2012; Tobaldini et al., 2014; Kohansieh and Makaryus, 2015; Irwin et al., 2016), emotional regulation (Walker, 2009; Simon et al., 2015), and pain (Smith and Haythornthwaite, 2004; National Sleep Foundation, 2015). The overall enhancement in pain sensitivity caused by various sleep disturbances has been widely reported both at the clinical and preclinical levels with consistent effects across species (Lautenbacher et al., 2006; Finan et al., 2013; Schrimpf et al., 2015; Alexandre et al., 2020). In sharp contrast, sleep disturbances do not increase sensitivity to innocuous stimuli of various sensory modalities: thermal discrimination, responsivity to light mechanical stimuli, auditory and visual acuities are not changed by sleep loss (Kundermann et al., 2004; Franzen et al., 2009; Scherer et al., 2013; Alexandre et al., 2017). Altogether, these results indicate that inadequate sleep specifically enhances pain sensitivity, but the precise mechanisms responsible remain largely unknown.

Pain is a complex neural process that involve detecting a potentially harmful stimulus via specialized peripheral sensory neurons called nociceptors, integrating the information at the spinal cord level before sending it to the brain where a pain sensation is generated to incorporate sensory-discriminative, emotional, limbic and even cognitive components (Figure 1A). Virtually all the components of the pain response can be modulated along the nociceptive and pain pathways to either enhance or depress the pain signal (Figure 1A). The ultimate pain sensation generated is the result of the integration of all these pain components, which contributes to the multifactorial and individualized features of pain. Therefore, an overall increase in pain sensitivity can be obtained by various mechanisms, raising the possibility that different sleep disturbances alter pain processing at distinct levels of the nociceptive and pain pathways, and thus could have cumulative effects.

Sleep is also a complex and dynamic neural process that comprises of two major and distinct states, rapid eye movement (REM) sleep and non-rapid eye movement (NREM) sleep. They alternate periodically throughout the sleep cycle and can be distinguished based on their electroencephalogram (EEG) and electromyogram (EMG) features (Aserinsky and Kleitman, 1953). In normal conditions, sleep architecture has a predictable macro-organization with NREMS always preceding REMS (Figure 1B). In humans, three stages of NREMS have been characterized based on their EEG features and to reflect transition into "deeper" sleep (N1, N2, and N3 which is also designated as slow wave sleep SWS). NREMS intensity can be assessed by quantifying the EEG power between 0.5 and 4 Hz, also known as slow-wave activity (SWA) (Borbely, 1982; Borbely and Achermann, 1999). REMS is characterized by an "activated" EEG (high frequency, low amplitude waves), no postural muscular activity on the EMG (atonia), and ocular saccades (that can be quantified with electrooculogram) in both humans and rodents. The distribution and amount of NREMS and REMS are mostly determined by two principal and interacting systems: the endogenous circadian clock and a homeostatic component that regulates the need and intensity of sleep according to the sleep-wake history (i.e., the time previously spent asleep or awake) (Borbely, 1982). Due to these dynamic interrelations between sleep-wake stages, many experimental sleep disruptions are poised to have broader effects on other sleep parameters than originally anticipated, notably NREMS and REMS amount. As a result, the exact effects of various sleep disturbances, especially during chronic settings, on pain processing are not clear.

To understand the neural pathways at the intersection between sleep and pain processes, it is then critical to define precisely which sleep stages (REMS, non–NREMS, or both) and what type of the sleep disruptions (sleep loss, sleep fragmentation or sleep interruptions) affect the pain sensation.

Here, we performed a review of the literature about sleep disturbances and pain sensitivity in humans and rodents by taking into consideration the amount of sleep lost. We find that NREMS loss could be the major driver for pain hypersensitivity, and that the temporal nature of the sleep disruptions could affect different components of the pain response. We then discuss possible neural mechanisms responsible and cellular pathways involved.

Part 1: Total (NREMS+REMS) sleep loss on pain

In humans, a single night of total sleep deprivation caused a decrease in pain thresholds from a heat source (8 studies out of 11), mechanical stimulus (both punctate and pressure; 5 studies out of 6) and cold (5 studies out of 6) when tested immediately at the end of the sleep deprivation (Figure 2). Furthermore, the pain response to a sustained noxious stimulus (ischemia pain, cold pressor pain) was heightened in half of the studies, suggesting a less robust or a ceiling effect of the pain response (Drewes et al., 1997; Onen et al., 2001; Larson and Carter, 2016; Eichhorn et al., 2018; Staffe et al., 2019). Spontaneous pain complaints (general body pain, low back pain, or headache) and migraine crises are more often reported after one night of total sleep deprivation (Busch et al., 2012; Houle et al., 2012; Palma et al., 2013). Both women and men show an hyperalgesic effect of sleep loss, with a stronger trend for women (Eichhorn et al., 2018; Staffe et al., 2019). One night of total sleep deprivation increased temporal summation of pain ratings upon repeated noxious pressure stimulations (Staffe et al., 2019), suggesting a possible spinal nociceptive facilitation (Bosma et al., 2015), although amplification within the brainstem, thalamus,



or cortical areas might also contribute. In contrast, detection of innocuous stimuli (gentle touch, warmth) was not affected by sleep loss (Kundermann et al., 2004, 2008; Lautenbacher et al., 2006; Schuh-Hofer et al., 2013). Other sensory systems such as visual performance and acoustic startle/reactivity, showed a decrease in responsiveness after sleep loss (Franzen et al., 2009; Jung et al., 2011; Liberalesso et al., 2012; Scherer et al., 2013).

In rodents, acute total sleep deprivation is usually performed by variations of the gentle handling paradigm (Tobler et al., 1997; Page et al., 2014; Vanini et al., 2014; Vanini, 2016; Alexandre et al., 2017) with animals tested at the end of the session. Protocols providing novel objects without touching the animals are minimally stressful, allowing for an extension of wakefulness without forced locomotor activity. Nine and twelve hours of sleep deprivation caused an increased sensitivity for noxious heat (hotplate set at 52 C, radiant heat with Hargreaves), pressure (von Frey filaments) and sustained pain (capsaicin), with an overall trend toward higher responses in females (Page et al., 2014; Vanini et al., 2014; Vanini, 2016; Alexandre et al., 2017; Hambrecht-Wiedbusch et al., 2017). Responses to noxious cold (\sim 4C; acetone test) were not changed by sleep loss, in contrast to human data, which might be due to technical differences to generate cold (Harvey et al., 2010; Alexandre et al., 2020). Responses to somatic innocuous stimuli (gentle touch with a brush, warmth (20-37 C) detection though a thermal gradient) were not changed, even after 12 h of sleep deprivation (Alexandre et al., 2017). Startle responses following loud acoustic stimuli were decreased after 9h of sleep loss (Alexandre et al., 2017), similar to humans and presumably reflecting fatigue. Overall, these results indicate that the enhancement of pain responses by total sleep loss is consistent and well preserved in both rodents and humans.

UMANS:				SLEEP			PAIN HYPEF	SENSITIVIT	Υ
	1 st Author	Cohort	Disru	ption technique	Duration	Heat	Mecha	Cold	Sustained pain
Č,	Drewes	10	N	ot described	8h	No	No	n.t.	No
Activities	Onen	9 M	Mess	age every 10min	8h	No	Yes	n.t.	n.t.
	Staffe	25 (16 M, 9	9F) Me	ssage every h	8h	No	Yes	Yes	Yes
	Kundermann	20 (11 M, 9	9 F)	Distraction	8h	Yes	n.t.	No	n.t.
	Eichhorn	36 (18 M, 1	8 F) Distra	ction (actigraphy)	8h	Yes	Yes	Yes	No
	Kundermann	19 (11 M, 8	3 F)	Distraction	8h	Yes	n.t.	n.t.	n.t.
15	Azevedo	28M		Distraction	8h 8h	Yes Yes	n.t. n.t.	n.t. Yes	n.t. n.t.
	Busch	20 (6 M, 14	4F)	Distraction					
Schuh-Hofer		14 (8M, 6	F) Distra	ction (actigraphy)	8h	Yes	Yes	Yes	n.t.
	Larson	27 (14 M, 1	3 F)	Distraction	8h	n.t.	n.t.	Yes	Yes
	Roehrs	7 (1 M, 6 F)		ot described	8h	Yes	n.t.	n.t.	n.t.
	Kamiyama	13 (7 M, 6	F) Distra	ction (actigraphy)	8h	n.t.	Yes	n.t.	n.t.
	Krause	25 (13 M, 1	2 F)	Distraction	8h	Yes	n.t.	n.t.	n.t.
ODENTS:				SLEEF	,		PAIN HYPEF	SENSITIVIT	Υ
ovel object	1 st Author	Species (sex)	Strain	Disruption technique	Duration	Heat	Mecha	Cold	Sustained pain
	Vanini (a)	rat (M)	SD	Gentle Touch	<6h	Yes	n.t.	n.t.	n.t.
5	Hambrecht	rat (M)	SD	Gentle Touch	<6h	Yes	n.t.	n.t.	n.t.
	Page	rat (M)	SD	Gentle Touch	6h	n.t.	No	n.t.	n.t.
total a	Alexandre M	louse (M, F)	C57BL/6J	Novel Object	6h	No	No	No	n.t.
DE	Vanini (b)	rat (M)	SD	Gentle Touch	9h	Yes	n.t.	n.t.	n.t.
	Alexandra			Neval Ohi	9h	Yes	Yes	No	Yes
	Alexandre	iouse (IVI, F)	03/BL/0J	Novel Object	12h	Yes	Yes	No	n t.

Effects of total sleep deprivation in humans and rodents for different pain modalities (heat, mechanical, cold) and sustained pain. M indicates men and F indicates women. n.t., not tested.

Part 2: State-specific sleep deprivation in humans

In humans, SWS (NREMS N3) can be interrupted by monitoring the EEG/EMG signals and applying a short auditory stimulus (80–110 dB) based on the EEG/EMG features observed in real time [for N3 = detection of slow-waves (0.5–3.5 Hz)]. This protocol does not cause a total loss of NREMS or REMS, but it increases the time spent in N1 and N2 (Arima et al., 2001). Three consecutive nights of SWS deprivation did not affect pressure pain responses applied to jaw muscles (Arima et al., 2001), while pain sensitivity to pressure at tender points (commonly used to diagnose fibromyalgia) was reported increased in some studies (Moldofsky et al., 1975; Moldofsky and Scarisbrick, 1976; Lentz et al., 1999) but not in others (Older et al., 1998). All these studies had a relatively low N (ranging from 6 to 19) which could explain the variability and possibly insufficient statistical power. No major differences on pain sensitivity were found between men and women. There is currently no specific SWS-deprivation protocol in rodents.

REMS can be interrupted in humans by a frank awakening (e.g., touching or moving subject) based on EEG/EOG/EMG features observed in real-time. Quantifying the total amount of sleep under these protocols indicates that some REMS deprivation protocols might also cause a significant loss of NREMS (~30%; Roehrs et al., 2006), while others do not (Moldofsky and Scarisbrick, 1976; Azevedo et al., 2011). Interestingly, REMS deprivation protocols that are not associated with total sleep loss do not increase pain even after 4 consecutive days (Azevedo et al., 2011), while a protocol that reduced total sleep amount increased heat pain sensitivity after a single night (Roehrs et al., 2006) (Figure 3). REMS deprivation by administration of clonidine did not change heat pain sensitivity (nor total sleep amount) in either men or women (Chouchou et al., 2015).



State-specific sleep deprivation in rodents

In rodents, the majority of REMS sleep deprivation has been performed using variations of the "platform-over-water" method (Mendelson et al., 1974) that take advantage of the muscle atonia occurring during REMS (Jouvet et al., 1964). Rats or mice are placed on top of a small platform surrounded by water in which they fall when they enter REMS and lose postural tone, causing them to wake up (Figure 4A). A modification of this paradigm was introduced to circumvent isolation and movement restriction by allowing a socially stable group of animals to be sleep-deprived together in a large tank containing multiple small platforms (Coenen and Van Luijtelaar, 1985). While 6 h of this paradigm does not change heat and mechanical pain sensitivity and 24 h has limited effects (Onen et al., 2000; Wei et al., 2007; Wang et al., 2015; Tomim et al., 2016; Nasehi et al., 2018; Sardi et al., 2018b), two or more consecutive days are systematically associated with an increase in pain responses for mild (44-46 C) (Onen et al., 2000; May et al., 2005; Damasceno et al., 2009; Harvey et al., 2010; Skinner et al., 2011) and intense (52 C) (Nascimento et al., 2007; Damasceno et al., 2009; Araujo et al., 2011; Gurel et al., 2014) heat, pressure (Randall and Selitto) (Ukponmwan et al., 1984; Onen et al., 2000; Tomim et al., 2016) and punctate (von Frey filaments) (Wei et al., 2008, 2010; Damasceno et al., 2009) mechanical

stimuli, and chemical pain (Figure 4A) (Hicks et al., 1979; Onen et al., 2000; Tomim et al., 2016). The development of heat hyperalgesia is stronger in females (Araujo et al., 2011). However, "platform-over-water" protocols significantly reduce NREMS by \sim 30–50%, nearly \sim 4–6 h sleep loss per day (Grahnstedt and Ursin, 1985; Machado et al., 2004), making it difficult to distinguish the relative contribution of REMS or NREMS loss to the development of hyperalgesia. Interestingly, the overall onset of hyperalgesia during "platform-over-water" protocols ranges between 24 and 48 h, (corresponding to \sim 5– 10 h total sleep loss), which is very similar to that of an acute total sleep deprivation (between 6 and 9 h) (Alexandre et al., 2017).

Sleep disruption protocols aiming at fragmenting NREMS can also indirectly prevent the transition from NREMS to REMS, thereby reducing REMS amount without significantly altering NREMS amount in most cases (Figure 4B). These protocols usually involve circular cages equipped with a rotating sweeping bar or cage floor programmed to rotate/move at specific intervals and durations (Figure 4B) (Ringgold et al., 2013; Sutton and Opp, 2014; Alexandre et al., 2017). Under these protocols, loss of REMS (ranging from 50 to 95%) without concomitant loss of total sleep did not alter mechanical or heat pain after one or up to 5 consecutive days in males or females (Sutton and Opp, 2014; Wodarski et al., 2015; Alexandre et al., 2017). In one

"REMS deprivation" Platform(s) over water	1 st Author Wang Nasehi Onen Wei (a) Wei (d) Tomim Sardi Onen May	Species (sex) Rat (M+F Rat (M)	Strain							
Platform(s) over water	Wang Nasehi Onen Wei (a) Wei (d) Tomim Sardi Onen May	Rat (M+F Rat (M)		Disruption technique	Duration (d)	n Daily total sleep loss	Daily REM sleep loss	Heat	Mecha	Other
Platform(s) over water	Nasehi Onen Wei (a) Wei (d) Tomim Sardi Onen May	Rat (M)	SD	Flower Pot; 5.5 d	cm 6h	Yes (<10%)		no (HG)	no (vF)	n.t.
	Onen Wei (a) Wei (d) Tomim Sardi Onen May	Rat (M) Wista	Wistar	Dual Platform; 60	icm 1		yes (~95%)	no (50)	n.t.	n.t.
A CONTRACTOR	Wei (a) Wei (d) Tomim Sardi Onen May	Rat (M)	Wistar	Flower Pot; 5.5 cm	cm 1			n.t.	no (RS)	n.t.
	Wei (d) Tomim Sardi Onen May	Rat (M)	SD & HW	Flower Pot; 7.5 d	cm 1	Yes (~40%)		no (50)	Yes (vF)	n.t.
	Tomim Sardi Onen May	Rat (M)	HW	Flower Pot; 7.5 cm	cm 1			n.t.	Yes (vF)	n.t.
	Sardi Onen May	Rat (M)	Wistar	Flower Pot; 6.5 c	icm 1			n.t.	Yes (RS)	Yes (che
	May	Rat (M)	Wistar	Flower Pot; 6.5 cm	cm 1			n.t.	Yes (RS)	n.t.
	iviay	Rat (M)	Wistar	Flower Pot; 5.5 0	cm 2	2		Yes (46)	Yes (RS)	Yes (ele
	Harvey	Rat (M)	SD	Dual Platform: 7.5	.5 cm 2			Yes (44, 52)	n.t.	nt
	Onen	Rat (IVI)	SD	Elower Pot: 5.5		Ves (~10%)	Vec (~95%)	Yes (45)	No (DO)	nt
	Wei (b)	Rat (M)	VVIStar	Flower Pot: 7.5 c	cm 2	165 (-4076)	yes (~5376)	nt	Yes (RS)	nt
	Wei (c)	Rat (M)		Flower Pot: 7.5 c	cm 2		-	nt	Yes (VF)	nt
	Tomim	Rat (IVI)	Wietor	Flower Pot: 6.5 c	cm 2			nt	Yes (VF)	Vec (che
Atonia	Open (b)	Rat (M)	Wistar	Flower Pot: 5.5 d	cm 3			Voc (A6)	Vec (PS)	nt.
	Damasceno	Rat (M)	Wistar	Flower Pot: 6.5 o	cm 3			Ves (46 52)	n.t.	n.t.
	Araujo	Ms (M+F)	C57BL/6.1	MMP; 3.5 cm	3	Yes (~40%)	ves (~95%)	Yes (50)	n.t.	n.t.
Year	Onen	Rat (M)	Wistar	Flower Pot; 5.5 d	cm 3			n.t.	Yes (RS)	n.t.
2	Hicks (a)	Rat (F)	SD	Flower Pot; 6 ci	:m 3			n.t.	n.t.	Yes (ele
	Nascimento	Rat (M)	Wistar	MMP; 6.5 cm	4			Yes (50)	n.t.	n.t.
	Damasceno	Rat (M)	Wistar	Flower Pot; 6.5 d	cm 4			Yes (46, 52)	n.t.	n.t.
	Skinner Gürel Ukponmwen	Rat (M) Wi Rat (F) S Rat (M) Wi	Wistar	Flower Pot; 6 ci	m 4	4) yes (~95%)	Yes (46)	n.t.	n.t.
Fall in water			SD	MMP; 6 cm	4	Yes (~40%)		Yes (52)	n.t.	n.t.
			Wistar	Flower Pot; 5.5 d	.5 cm 4			n.t.	Yes (RS)	n.t.
	Damasceno	Rat (M)	Wistar	Flower Pot; 6 c	:m 4			n.t.	Yes (vF)	n.t.
	Hicks	Rat (F)	SD	Flower Pot; 6 c	:m 4			n.t.	n.t.	Yes (ele
	1 st Author Wodarski	Species (sex) Rat (M)	Strain Wistar	Disruption technique ting cage; 10s on/60s	Duration (d)	Daily total sleep loss yes (~60%)	Daily REM sleep loss yes (-95%)	Heat yes	Mecha yes	Other
	1 st Author Wodarski Alexandre	(sex) Rat (M) Ms (M+F) (Strain Wistar Rota 10s on 257BL/6J Cag	Disruption technique ting cage; 10s on/60s v/10s off first 17 min er ge tilt; 10 ms mvt / 1 m	Duration (d) off 0.33 ach h 0.33 hin 1	Daily total sleep loss yes (~60%) yes (-30%) no	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%)	Heat yes No No	Mecha yes No No	Other n.t. n.t. No
Rotating bar	1 st Author Wodarski Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M)	Strain Wistar Rota 10s on 257BL/6J Cag 257BL/6J Rot	Disruption technique ting cage; 10s on/60s v/10s off first 17 min er ge tilt; 10 ms mvt / 1 m tating bar; 8s on/30s o	Duration (d) a off 0.33 ach h 0.33 nin 1 off 1	Daily total sleep loss yes (~60%) yes (-30%) no no	Daily REM sleep loss yes (-95%) yes (-50%) yes (-95%)	Heat Ves No n.t.	Mecha yes No No No	Other n.t. n.t. No n.t.
Rotating bar Cage tilt	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) Ms (M+F) (Ms (M)	Strain Wistar Rota 257BL/6J Cag 257BL/6J Cag 257BL/6J Cag 257BL/6J Rot	Disruption technique v10s on/60s v10s off first 17 min er ge tilt, 10 ms mvt /1 m tating bar; 8s on/30s o ge tilt, 10 ms mvt /1 m tating bar; 8s on/30s o	Duration (d) off 0.33 ach h 0.33 nin 1 off 1 off 7	Daily total sleep loss yes (~60%) yes (-30%) no no no no	Daily REM sleep loss yes (-95%) yes (-50%) yes (-95%) yes (-50%) yes (-95%)	Heat No No n.t. No n.t.	Mecha No No No No No	Other n.t. n.t. No n.t. No n.t.
Cage tilt Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) Ms (M+F) (Ms (M+F) (Ms (M+F) (Strain Wistar 557BL/6J Carg 557BL/6J Carg 557BL/6J Rota Carg 577BL/6J Rota	Disruption technique ting cage; 10s on80s v/10s off first 17 min e: ge tilt; 10 ms mv / 1 m tating bar; 8s on/30s o ge tilt; 10 ms mv / 1 m tating bar; 8s on/30s o Species (Sex)	Duration (d) soff 0.33 ach h 0.33 bin 1 off 1 nin 5 off 7	Daily total sleep loss yes (~60%) yes (~30%) no no no no no Disruption	Daily REM sleep loss yes (-55%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) technique	Heat yes No nt. No nt. Heat hyp (%	Mecha No No No No eralgesia bsl)	Other n.t. No n.t. No n.t. Duration (d)
Cage tilt Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Strain Wistar 557BL/6J Cag 57BL/6J Cag 57BL/6J Cag 57BL/6J Rof Alexandre	Disruption technique ting cage; 10s on60s v/10s off first 17 min er ge tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o get tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o Species (sex) Ms (M) Ci	Duration (d) .off 0.33 ach h 0.33 inin 1 .off 7	Daily total sleep loss yes (~60%) yes (~30%) no no no no no Disruption Novel	Daily REM sleep loss yes (-95%) yes (-50%) yes (-95%) yes (-95%) yes (-95%) technique	Heat yes No nt No nt Heat hyp (%) 11	Mecha yes No No No No No eralgesia bsi)	Other n.t. No n.t. No n.t. Duration (d)
Cage tilt Accumulation of heat hypera	1 st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Strain Wistar C57BL/6J Cag C57B	Disruption technique ting cage; 10s on80s v/10s off first 17 min er ge tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o gitt; 10 ms mv/ 1 m tating bar; 8s on/30s o Species (sex) Ms (M) C: Rat (M)	Strain Strain Strain Strain Strain Strain Strain Strain Strain Strain Strain Strain Strain	Daily total sleep loss yes (~60%) yes (~30%) no no no no no Disruption Novel	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-50%) yes (-95%) technique Object gentle handling	Heat yes No No nt. No nt. Heat hype (% 111 g 2 2	Mecha No No No No No So eralgesia bsl)	Other n.t. No n.t. No n.t. Duration (d) 1 1
Cage tilt Accumulation of heat hypera	1 st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M+F) (Ms (M+F) (Ms (M) (Strain Wistar 103 on 575BL/6J 575BL/6J 575BL/6J 1st Author Alexandre Hambrecht Nasehi	Disruption technique ting cage; 10s on/60s V/10s off first 17 min es ge tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o Species (sex) Ms (M) C: Rat (M)	Duration (d) off 0.33 ach h 0.33 in 1 ff 1 in 5 ff 7 Strain Strain Strain Strain	Daily total sleep loss yes (~60%) yes (~30%) no no no no Disruption Novel Novel Object, Dual Platf	Daily REM sleep loss yes (-55%) yes (-50%) yes (-50%) yes (-50%) yes (-50%) yes (-95%) technique Object gentle handling orms; 6 cm	Heat yes No No n.t. Heat hype (% I 11 g 2 2 2 2 2 2	Mecha No No No No No So 4	Other n.t. No n.t. No n.t. Duration (d) 1 1 1
Cage tilt Accumulation of heat hypera	1 st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Strain Wistar 257BL/6J 257BL/6J 257BL/6J 257BL/6J 257BL/6J 267BL/6J 787 787 787 787 787 787 787 78	Disruption technique vi0s off first 17 min ea ge tilt: 10 ms mvt / 1 m talang bar; 8s on/30s o ge tilt: 10 ms mvt / 1 m talting bar; 8s on/30s o ge tilt: 10 ms mvt / 1 m talting bar; 8s on/30s o Species (sex) Ms (M) C: Rat (M) Rat (M)	Duration (d) off 0.33 ach h 0.33 in 1 ff 1 in 5 ff 7 Strain Strain Strain SD ↓	Daily total sleep loss yes (~60%) yes (~30%) no no no Disruption Novel Novel Object, Dual Platf	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) technique l Object gentle handling orms; 6 cm	Heat yes No No n.t. No n.t. Heat hyp (%) (%) 111 9 2 2 2 2 2 2 2 2 2	Mecha yes No No No No No eralgesia bsi) 5 0 4 5	Other n.t. No n.t. No n.t. Duration (d) 1 1 1 2
Cage tilt Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Strain Wistar Rota 105 75/7BL/6J Ca 557BL/6J Ca 557BL/6J Ca 557BL/6J Rot 1st Author 1st Author Alexandre Hambre-tt Naseh May Alexandre	Disruption technique v/tos off frst 17 min e.g ge tilt; 10 ms m/t 1 m tating bar; 8 s on/30 s o ge tilt; 10 ms m/t 1 m tating bar; 8 s on/30 s o Species (sex) Ms (M) C: Rat (M) M Rat (M) C:	Duration (d) off 0.33 ach h 0.33 inin 1 off 7 Strain Stra	Daily total sleep loss yes (~60%) yes (~30%) no no no no Disruption Novel Novel Object, Dual Platf Dual Platf	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) technique Object gentle handling orms; 6 cm Object	Heat yes No No n.t. No n.t. Heat hyp (%) 111 g 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	Mecha No No No No No No So So 4 5 5 6	Other n.t. No n.t. No n.t. Duration (d) 1 1 1 2 2 3
Cage tilt Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Ms (M) (Strain Rota 105 or (578L/6) Cac Cac Cac Cac Cac Cac Cac Cac Cac Cac	Disruption technique ting cage; 10s on80s v/10s off first 17 min e: ge tilt; 10 ms mv / 1 m ge tilt; 10 ms mv / 1 m tating bar; 8s on/30s o ge tilt; 10 ms mv / 1 m tating bar; 10 ms mv / 1 m tating bar; 10 ms mv / 1 m tating bar; 10 ms mv / 1 ms mv / 1 m tating bar; 10 ms mv / 1 ms mv / 1 ms mv / 1 m tating bar; 10 ms mv / 1	Duration (d) 0.33 ach h 0.33 iin 1 off 7	Daily total sleep loss yes (-60%) yes (-30%) no no no no Disruption Novel Novel Object, Dual Platf Dual Platf Novel Flower	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) yes (-95%) technique Object gentle handling orms; 6 cm Object pot, 6 cm	Heat No No n.t. No n.t. Heat hype (% 111 9 2 2 2 2 3 3	Mecha No No No No No No So 4 5 5 6 6	Other n.t. n.t. No n.t. No n.t. Duration (d) 1 1 1 2 2 3 3
Cage tilt Accumulation of heat hypera	1 st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Rota Rota 105 or 105 or 557BL/6J Cag 557BL/6J Cag 557BL/6J Cag 557BL/6J Cag 1st Author Rota Alexandre Hambrecht Nasehi Alexandre Alexandre (a) Alexandre (a) Alexandre (a) Alexandre (a) Alexandre (a) Alexandre (a)	Disruption technique ting cage; 10s on/60s v/10s off first 17 min er ge tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o tilt; 10 ms mv/ 1 m tating bar; 8s on/30s o Species (sex) Ms (M) Ci Rat (M) Rat (M) Ci Rat (M) Ci Rat (M) Ci Rat (M) Ci	Duration (d) off 0.33 ach h 0.33 inin 1 iff 1 off 7	Daily total sleep loss yes (~60%) yes (~30%) no no no no Disruption Novel Novel Object, Dual Platf Dual Platf Novel Flower Flower	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) technique Object gentle handling orms; 6 cm Object pot, 6 cm 3.5 cm	Heat No No nt. No nt. Heat hyp (% 11 g 2 2 2 2 2 2 2 3 3 3	Mecha No No No No No No So (4 5 5 6 1 1 7	Other n.t. No n.t. No n.t. Duration (d) 1 1 1 2 3 3 3 3
Accumulation of heat hypera	1 st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Ms (M) (Strain Rota 10s on 507BL/6J Rota 10s on 507BL/6J 557BL/6J Cag 557BL/6J Rota Cag 557BL/6J Rota Cag 800 1st Author Alexandre May Rota Cag 800 1st Author Alexandre May Rota Cag 800 Alexandre Maraujo Araujo Nascimento Rota 800	Disruption technique vi/0s off first 17 min er ge tilt; 10 ms mv/ 1 m tataling bar; 8s on/30s o ge tilt; 10 ms mv/ 1 m tataling bar; 8s on/30s o ge tilt; 10 ms mv/ 1 m tataling bar; 8s on/30s o Ms (M) Ci Rat (M) M Rat (M) M Rat (M) M Ms (M) Ci Rat (M) M	Duration (d) off 0.33 sach h 0.33 sin 1 off 7 Strain 5	Daily total sleep loss yes (~60%) yes (~30%) no no no no Disruption Novel Novel Object, Dual Platf Dual Platf Novel Flower p	Daily REM sleep loss yes (-50%) yes (-50%) yes (-50%) yes (-50%) yes (-95%) technique Object gentle handling orms; 6 cm Object pot, 6 cm oot, 6.5 cm	Heat yes No No nt No nt Heat hype (%) 11 9 22 2 2 2 3 3 3 3 3	Mecha No No No No No So Co 4 So 6 1 7 7	Other n.t. No n.t. No n.t. Duration (d) 1 1 1 2 3 3 3 3 3
Accumulation of heat hypera	1 st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) ((M) (M) (Ms (M) ((M) (M) (M) (M) (M) (M) (M) (M) (M	Strain Rota 10s on 2057BL/6J Cag 2057BL/6J Cag 2057BL/6J Cag 2057BL/6J Rota 2000 1st Author 1st Author Alexandre Hambre-Ch Nasehi Alexandre Alexandre Hambre-Ch Nasehi Alexandre Alexandre Alexandre Alexandre Alexandre Alexandre May Jaraujo Araujo Araujo	Disruption technique ting cage; 10s on60s v/10s off first 17 min e. ge tilt; 10 ms mk / 1 m tating bar; 8s on/30s o ge tilt; 10 ms mk / 1 m tating bar; 8s on/30s o ge tilt; 10 ms mk / 1 m tating bar; 8s on/30s o (sex) Ms (M) C: Rat (M) M Ms (M) C: Rat (M) M Ms (M) C: Rat (M) M Rat (M) M	Duration (d) orff 0.33 ach h 0.33 inin 1 ff 1 off 7	Daily total sleep loss yes (~60%) yes (~30%) no no no Disruption Novel Disruption Novel Object, Dual Platf Dual Platf Novel Flower p MMP;	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-50%) yes (-95%) technique I Object gentle handling orms; 6 cm Object pot, 6 cm 3.5 cm 6.5 cm	Heat yes No No n.t. No n.t. Heat hype (%) (%) (%) 111 9 2 2 2 3 3 3 3 3 3 3	Mecha No No No No No No So So So So So So So So So So So So So	Other n.t. n.t. No n.t. No nt. Duration (d) 1 2 3 3 3 3 3 3 3 3 3 3 3 3 3
Cage tilt Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (M) (Ms (M) (Ms (M)	Strain Rota 105 or (257BL/6J) Ca Ca Ca C57BL/6J Ca Ca Ca C57BL/6J 1st Author Alexandre Alexandre 1st Author Alexandre Alexandre Mascenco (a) Araco Ca Ca Ca Ca Ca Ca Ca Ca Ca Ca Ca Ca Ca C	Disruption technique v/10s off frst 17 min e. ge tilt; 10 ms m/ 1 m ge tilt; 10 ms m/ 1 m tating bar; 8s on/30s o ge tilt; 10 ms m/ 1 m tating bar; 8s on/30s o (sex) Ms (M) C: Rat (M) M Rat (M) C: Rat (M) C: R	Duration (d) orff 0.33 ach h 0.33 inin 1 iff 1 off 7	Daily total sleep loss yes (~60%) yes (~30%) no no no no Disruption Novel Disruption Novel Object, Dual Platf Dual Platf Novel Flower MMP; Flower	Daily REM sleep loss yes (-50%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) yes (-95%) technique Object gentle handling orms; 6 cm orms; 6 cm Object pot, 6 cm 3.5 cm ot, 6.5 cm Object	Heat yes No No n.t. No n.t. Heat hyp (%6 111 2 2 2 3 3 3 3 3 3 3 3 3 3	Mecha No No No No No No So So 4 4 5 5 6 1 7 7 8 8 9	Other n.t. n.t. No n.t. Duration (d) 1 2 3 3 3 3 3 4
Cage tilt Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (C	Strain Rota 105 or (57BL/6J Cac Cac Cac C57BL/6J Cac Cac Cac C57BL/6J Cac Cac Cac Cac Cac Cac Cac Cac Cac Cac	Disruption technique ting cage; 10s on80s v/10s off first 17 min e: ge tilt; 10 ms mv/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 10 ms m/1 m tating bar; 8s on/30s of ge tilt; 1	Duration (d) orff 0.33 sin 1 offf 7 Strain 7 STBL/6J 5 SD 5 Winstar 57BL/6J STBL/6J 57BL/6J Winstar 57BL/6J STBL/6J 57BL/6J STBL/6J 57BL/6J	Daily total sleep loss yes (-60%) yes (-30%) no no no no Disruption Novel Novel Object, Dual Platf Dual Platf Dual Platf Novel Flower MMP; Flower MMP; Novel	Daily REM sleep loss yes (-95%) yes (-50%) yes (-50%) yes (-95%) yes (-95%) technique Object gentle handling orms; 6 cm orms; 6 cm 0.5 cm 6.5 cm 0.5 cm 0.5 cm	Heat No No nt. No nt. Heat hyp (% (% 111 9 2 2 2 2 2 2 2 3 3 3 3 3 3 3 3 3 3 3 3	Mecha No No No No No No No So So So So So So So So So So So So So	Cother 1 nt. No nt. No nt. No nt. 1 1 1 1 2 3 3 3 3 4 4 5
Accumulation of heat hypera	1st Author Wodarski Alexandre Sutton Alexandre Sutton	Species (sex) Rat (M) Ms (M+F) (Ms (M) (Ms (M+F) (Ms (M) (Ms (M) (Da	Strain Rota 105 or (257BL/6) Cac Cac Cac 575BL/6) Cac Cac 575BL/6) Rof Cac Alexandre Rof Alexandre Nasein (a) Cac May Alexandre (a) Alexandre (a) (a) Araujo Araujo (a)	Disruption technique ting cage; 10s on#0s v/10s off first 17 min experime ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or ge tilt; 10 ms m/ 1 min taing bar; 8s on/30s or Ms (M) C: Rat (M) M Ms (M) C: Rat (M) M Ms (M) C: Rat (M) M Ms (M) C:	Duration (d) orff 0.33 sin 1 offf 7 strain 5	Daily total sleep loss yes (~60%) yes (~30%) no no no no Disruption Novel Novel Object, Dual Platf Dual Platf Novel Flower p MMP; Novel Novel Novel	Daily REM sleep loss yes (-55%) yes (-50%) yes (-50%) yes (-50%) yes (-50%) yes (-50%) yes (-95%) technique Object gentle handling orms; 6 cm orms; 6 cm Object pot, 6 cm 3.5 cm 0.5 cm Object Object 3.5 cm	Heat No No nt. Heat hyp (% 111 g 2 2 2 4 2 2 2 2 3 3 3 3 3 3 3 3 3 3 4	Mecha No No No No No So So 4 5 5 6 1 1 7 7 8 8 9 9 9 1 7	Cther 1 nt. No nt. No nt. No 1 1 1 1 2 3 3 3 3 3 3 4 4 5 3 3

study (Wodarski et al., 2015), 8 h of sleep fragmentation that caused a minor total sleep loss (\sim 30%; 2 h 40 min) did not increase heat and mechanical pain responses, while a sleep

fragmentation protocol that also caused ${\sim}60\%$ total sleep loss (${\sim}5\,h$ 30 min) increased pain responses. Such estimation of the amount of total sleep loss required to increase pain

sensitivity is aligned with total sleep deprivation studies (Page et al., 2014; Alexandre et al., 2017).

Overall, these results suggest that traditional "REM sleepspecific" deprivation protocols relying on platforms-over-thewater techniques cause a significant total sleep loss over time, which might be the main driver for pain hypersensitivity, rather than the specific loss of REMS. Indeed, when ranking all studies that used the same pain assay (latency to withdraw on a hotplate set a 50-52 C; to mitigate technical variability) by pain intensity, we found that the main factor associated with the gradual increase in heat pain was the overall amount of sleep loss, irrespective of the sleep disruption protocol or species used (Figure 4B). Furthermore, these compiled studies suggest that when subjected to ~6h total sleep loss per day, animals develop heat hyperalgesia with a relatively linear increase of \sim 7% per day and reach a plateau after 4 d (Figure 4B). There were not enough studies carried out with females to perform this analysis. The overall pain profile from acute and chronic sleep loss experiments suggests females develop stronger hyperalgesia and might reach a plateau sooner compared to males. Altogether, these results fit with the hypothesis that the amount of total sleep loss, and not REMS or sleep continuity, is the major factor associated with the increased pain sensation for heat and mechanical stimuli.

Part 3: Dynamics of sleep loss on pain

While the amount of total sleep loss appears to be a major contributor to the increased pain response, the dynamics of the sleep loss could target different components of pain processing. This can be evidenced when comparing two models of sleep disruption in humans achieving the same sleep deficit after a single night: delayed bedtime and forced awakening (Smith et al., 2007). In the delayed bedtime protocol, subjects stay awake for 4h past their usual bedtime, while under the forced awakening protocol, subjects are awakened for 20 min every hour, and for a full hour at a random time during the night. Both protocols reduce total sleep amount by \sim 45–50% (~4h). While the delayed bedtime protocol initially extends wakefulness then allows for a consolidated sleep (i.e., with increased SWS during the remaining sleep opportunity), the forced awakenings protocol interrupts sleep with transient, short (20 min to 1 h) wake periods (causing more N1 at the expense of SWS) (Figure 5). These different dynamics of sleep loss are associated with preferential changes in pain responses, possibly underlying specific mechanisms. In both men and women, a single 4-h delayed bedtime is sufficient to increase heat and mechanical pain sensitivity when subjects are tested at wake up (Roehrs et al., 2006; Tiede et al., 2010; Faraut et al., 2015; Matre et al., 2015), similar to the effects of a night of total sleep deprivation, while a single night of forced awakenings

does not affect these modalities (Smith et al., 2007; Finan et al., 2017; Iacovides et al., 2017; Letzen et al., 2020). Both delayed bed onset and total sleep deprivation are associated with an extended wake, suggesting that heat and mechanical pain might be caused by a common mechanism sensitive to excessive wakefulness, i.e., a maximum period beyond which normal pain perception and neurobehavioral functioning cannot be maintained (Van Dongen et al., 2003). In support of this, administration of caffeine and modafinil, two drugs that can restore alertness, normalize heat and mechanical pain sensitivity (Vanini, 2016; Alexandre et al., 2017), without removing the sleep debt. Both sleepiness and pain hypersensitivity accumulate during experimental sleep deprivation (Alexandre et al., 2017), and sleepiness ratings in healthy individuals correlate with their pain complaints (Smith et al., 2007). Chemogenetic activation of GABAergic neurons in the anterior cingulate cortex in mice is sufficient to concomitantly increase delta power (which could indicate increased sleep pressure) and mechanical pain (Li et al., 2020). These results suggest that the neural circuits driving sleepiness could also be responsible for the heightened heat and mechanical pain sensitivity after an acute sleep loss.

Another major difference in pain processing between reduced yet consolidated sleep and fragmented sleep concerns diffuse noxious inhibitory controls (DNIC), a type of descending controls that can suppress the nociceptive signal at the spinal cord level. DNIC are triggered when two noxious stimulations occur simultaneously in different dermatomes, and they can be assessed in humans by using "conditioned pain modulation (CPM)" protocols (Yarnitsky, 2010; Yarnitsky et al., 2015). One night of forced awakenings strongly disrupts CPM in women (Smith et al., 2007), whereas a single 4-h delayed bedtime does not (Matre et al., 2016; 14 women, 8 men; Smith et al., 2007; women only; Matre et al., 2017; night shifts: 41 women, 12 men) (Figure 5), indicating that a mild sleep loss with disrupted continuity specifically targets these descending controls. Clinical studies support this assertion as DNICs are strongly altered in fibromyalgia patients who also suffer from mild total sleep loss and sleep disruption by awakenings (lasting more than 3 min; Lautenbacher and Rollman, 1997; Diaz-Piedra et al., 2015; Bjurstrom and Irwin, 2016), as well as patients with temporomandibular joint pain where DNICs were most altered in patients with poor sleep quality (Edwards et al., 2009). Interestingly, experimental sleep disturbances only disrupt DNIC in women (Eichhorn et al., 2018), which could contribute to the strong incidence of fibromyalgia and temporomandibular joint disorder pain in women.

DNICs are triggered specifically by the activation of lamina V spinal neurons (Villanueva and Le Bars, 1995) that project to the subnucleus reticularis dorsalis (SRD, also known as dorsal reticular nucleus DRt, and called medullary reticular nucleus in mice; Villanueva and Le Bars, 1995; Le Bars, 2002). Some SRD neurons project directly back into the deep dorsal



horn lamina (V-VII; Villanueva et al., 1995) while others activate several medullar pathways (Bouhassira et al., 1992b) that will also eventually project back to the spinal cord to inhibit projecting neurons (Leite-Almeida et al., 2006). It is important to note that DNICs are physiologically distinct from "traditional" descending inhibitory controls that originate from the rostroventral medulla (RVM), notably the locus coeruleus (LC) and the median raphe nucleus (MRN) (Basbaum and Fields, 1978, 1984; Bouhassira et al., 1992a; Le Bars et al., 1992; Kucharczyk et al., 2022), and the effects of forced awakenings on those are not known. Nonetheless, because delayed bed onset does not disrupt DNICs, this raises the possibility that a mild sleep loss coupled with sleep interruptions caused by the forced awakening protocol might affect more strongly lamina V pain pathways.

In contrast to heat and mechanical pain, cold pain responses are not changed by delayed bed protocol (Faraut et al., 2015), but they are increased after forced awakenings (Rosseland et al., 2018) (Figure 5).

Part 4: Chronic sleep disruptions and recovery sleep

The similar increase in acute pain responses to heat and mechanical stimuli observed after a single night of total sleep deprivation or at wake time after a delayed bedtime indicates that a short sleep opportunity is not sufficient to restore normal pain sensitivity. Indeed, the increase in heat and mechanical pain caused by sleep loss persists in humans (Roehrs et al., 2006; Tiede et al., 2010; Faraut et al., 2015; Matre et al., 2015, 2016; Odegard et al., 2015; Staffe et al., 2019) and even aggravates in rodents (Page et al., 2014; Alexandre et al., 2017; Sardi et al., 2018a). Cumulative deficits after sleep loss have also been reported for diverse biological processes, such as cognitive performance (Van Dongen et al., 2003; Banks et al., 2010), energetic metabolism (Spiegel et al., 1999; Van Cauter et al., 2008; Everson and Szabo, 2011), and immune system functions (Bryant et al., 2004). This is particularly relevant, because chronic insufficient



DEVELOPMENT OF HEAT PAIN HYPERSENSITIVITY DURING CHRONIC SLEEP DEPRIVATION:

sleep is becoming more frequent in the general population (up to 30%) and can be caused by poor sleep hygiene (TV, screens, etc.), work conditions (schedules/shiftwork, multiple jobs) or diseases (e.g., obesity-related sleep disorders; Holingue et al., 2018; Freire et al., 2022).

We quantified the cumulative wake amount over 5 consecutive days of experimental sleep restriction (6h of sleep deprivation daily, starting at light onset) in mice and confirmed that despite a normal homeostatic sleep response (increased in NREMS SWA and NREMS amount) occurring each day during the sleep opportunity period, there was an accumulation of sleep loss/deficit over time (Alexandre et al., 2017). We compared the degree of heat hyperalgesia caused by an acute, extended wakefulness of 6, 9, and 12 h with the effects of the repeated sleep restriction protocol (6 h of extended wake per day) and found they both gradually increased, with a plateau after 12 h acute sleep deprivation or 4 d of sleep restriction (Figure 6A). Interestingly, we found that the development of heat hyperalgesia fits best when incorporating the carryover amount of wake (i.e., the sleep debt at the beginning of the active period, in blue) with the extended wake (in red, Figure 6B), rather than the relative cumulative sleep loss measured at the time of testing (i.e., at the end of the sleep deprivation session; in pink; Figure 6C).

Overall, the cumulative effects of sleep debt on pain could explain why heat and mechanical pain hypersensitivity takes several days to develop in complex chronic settings that are associated with a moderate total sleep loss in humans. For example, the forced awakenings protocol, which causes a 4h total sleep loss per day, is insufficient to increase heat or mechanical (pinprick) pain after a single night (see Figure 5), but increases pain responses to both modalities after two consecutive nights (Smith et al., 2007; Iacovides et al., 2017; Irwin et al., 2022). As the individual has to maintain wakefulness throughout the day despite the homeostatic sleep pressure caused by the first night of forced awakenings (4h sleep loss and broken sleep continuity), this maintained wakefulness could represent an additional extended wake period. After the second night of forced awakenings, the net result is accumulation of sleep debt coupled with an episode of extended wakefulness (similar to an acute night of delayed sleep onset), which causes heat and mechanical hypersensitivity. Similarly, platform-over-the-water protocols occur over several days, causing a buildup of sleep debt that increases heat pain hypersensitivity over time. Finally, these dynamic processes could explain how longitudinal protocols where individuals go through stage-specific sleep deprivations followed by total sleep deprivation can lead to complex and sometimes misleading interpretations.

In conclusion, both clinical and preclinical studies confirm a strong effect of sleep disturbances on pain sensitivity in a highly conserved manner. However, different sleep disturbances affect preferentially specific components of the pain response. REMS deprivation is not associated with a strong increase in pain responses, and the major driver for hyperalgesia appears to be the total amount of sleep lost. Further, the dynamics of sleep loss reveal potential mechanistic insights: sleep loss caused by extended wake is strongly associated with heat and mechanical pain hypersensitivity; whereas a similar loss achieved by disrupting sleep continuity causes a loss of diffuse noxious inhibitory controls. Finally, the sleep debt and its deleterious effects on pain sensitivity are cumulative over days, despite a preserved homeostatic sleep rebound, which over time could contribute to the complex pain responses profile observed in chronic sleep disturbances settings.

Part 5: Possible sites for pain modulation by sleep loss

Nociceptive signals can be amplified at many levels along the pain pathways (see Figure 1A). Below we describe where the nociceptive input might be enhanced by acute and chronic sleep loss, and the potential mechanisms.

Nociceptors

Nociceptors can lower their activation threshold, notably when exposed to pro-inflammatory cytokines/chemokines, trophic factors or even foreign bodies (Chiu et al., 2013), a process known as peripheral sensitization. A major source of proinflammatory cytokines are immune cells. Immunity is composed of the innate and the adaptive immune systems and both are heavily regulated by circadian and sleep-wake cycles (Imeri and Opp, 2009; Lange et al., 2010; Besedovsky et al., 2012; Dimitrov et al., 2015; Irwin and Opp, 2017; Liu et al., 2021) and strongly affected by sleep loss (Bryant et al., 2004). An acute total sleep loss of 3-4 h is sufficient to alter the innate immune system that consists of monocytes (blood-circulating precursors of macrophages), macrophages, eosinophils, neutrophils, basophils, mast cells, and natural killer cells (NK cells). In particular, the number of circulating neutrophils and monocytes increase after sleep loss in humans (Irwin et al., 2008; Christoffersson et al., 2014; Faraut et al., 2015; Lasselin et al., 2015), rats (Ibarra-Coronado et al., 2015) and mice (Guariniello et al., 2012). In these cells, sleep loss enhances the transcriptional activity of the proinflammatory nuclear factor NFkB, which in turn promotes the production and release of TNF α , IL-1 β , IL-6, IL-8, and C-reactive protein (Hu et al., 2003; Haack et al., 2007; Irwin et al., 2008, 2015, 2022; Yehuda et al., 2009; Aho et al., 2013; Carroll et al., 2015; Dimitrov et al., 2015). These cytokines can bind to and sensitize nociceptors directly (Brenn et al., 2007; Binshtok et al., 2008; Kawasaki et al., 2008; Stemkowski et al., 2015). In addition, IL-1ß triggers the induction of cyclooxygenase 2 (COX2) in macrophages (Samad et al., 2001) which leads to the production of the prostaglandin PGE2, a potent sensitizer of nociceptors (Chapman and Dickenson, 1992; Chen et al., 1999). PGE2 levels correlate with migraine and headache complaints after sleep deprivation (Haack et al., 2007, 2009). The overall shift into a pro-inflammatory profile while factors that can terminate the immune response (i.e., anti-inflammatory cytokines and chemokines), could extend the duration of inflammation.

T cells and B cells (adaptive immune system) are further classified into helper (Th), regulator (Treg), killer (cytotoxic) and memory T cells, and they produce a highly targeted cellular response that relies on the prior exposure to the foreign bodies. Sleep loss durations lasting from 3 to 21 days affect the



adaptive immune system, with an overall downregulation of the "cellular immune response" carried out by Th-1 lymphocytes in favor of Th-2 and Th-17 lymphocytes-mediated responses in humans and rodents (Van Leeuwen et al., 2009; Axelsson et al., 2013; Nunes et al., 2018). This promotion of Th-2-mediated responses could be further strengthened by PGE2 produced by monocytes/macrophages (Harris et al., 2002). IL-17 directly causes pain (Pinto et al., 2010) by sensitizing nociceptors (Segond von Banchet et al., 2013), while the pro-nociceptive effects of IL-22 are possibly indirectly mediated by neutrophil recruitment (Pinto et al., 2015). IL-22 can also promote the differentiation into Th-2 lymphocyte profile, further amplifying

this shift (Lou et al., 2017). Finally, the dysregulation of B cell function after chronic sleep loss impairs the efficiency of immunization (Zielinski and Krueger, 2011; Prather et al., 2012).

Overall, these results indicate that sleep loss initially promotes the proinflammatory component of the innate immune response (monocytes/macrophages, neutrophils) and then a shift toward allergic reactions and autoimmunity at the adaptive immune system level (Th2/Th17). Because these changes are associated with an increase in pronociceptive signaling (IL-6, IL-1 β , TNF α) and a reduction of antinociception (IL2), they could contribute to pain hypersensitivity or a delayed recovery after tissue injury. However, administration of

10.3389/fnins.2022.1009902

the non-specific COX1/2 inhibitor ibuprofen does not reduce acute or chronic sleep loss-induced heat or mechanical evoked pain hypersensitivity (Wodarski et al., 2015; Alexandre et al., 2017), suggesting a limited role of inflammatory processes in these symptoms. In contrast, spontaneous pain complaints or migraine attacks, whose frequency increases after repeated nights of insufficient sleep (Haack and Mullington, 2005; Haack et al., 2007; Odegard et al., 2011) might be mediated by the sleep loss-induced increase in prostaglandin levels (Haack et al., 2009). Finally, sleep loss-induced increase in IL-6 or TNF α levels might aggravate the defects in emotional status (Hunt et al., 2021), possibly by crossing the blood brain barrier and reaching the mesolimbic system (Hunt et al., 2022) (Figure 7A).

Spinal cord

The spinal cord is a major site for nociceptive signal modulation. Only a small fraction of spinal neurons project to brain structures and they are mostly located in the lamina I and V of the dorsal horns (Todd, 2010). These neurons are under strong modulatory controls, both excitatory and inhibitory that originate both locally ("segmental controls") and at the supra-spinal level (mostly located within the rostroventral medulla and the pons); (Alexandre et al., 2020). Changes in the excitatory/inhibitory balance in the dorsal horns of the spinal cord will affect the nociceptive signal sent to the brain. One potent form of spinal plasticity is called central sensitization, which corresponds to a heightened functional state of the spinal nociceptive neurons where their activation threshold is reduced, they produce stronger and longer responses upon activation by noxious stimuli (a symptom referred to as hyperalgesia), and they can be activated by innocuous stimulations (allodynia) (Latremoliere and Woolf, 2009). Sleep loss could enhance spinal excitability by modulating both segmental and supraspinal controls.

Proinflammatory cytokines and PGE2 produced at the periphery after sleep loss can penetrate the spinal cord and promote nociceptive transmission (Baba et al., 2001; Samad et al., 2001; Latremoliere and Woolf, 2009). In addition, sleep loss increases sensitivity of NMDAR and mGluR in the dorsal horns (Wei et al., 2007, 2010), and this is associated with an increase in mechanical pain in rats. Capsaicin (active ingredient of red-hot chili peppers that activate TRPV1 channels) can be used to trigger central sensitization and measure associated behavioral changes caused by spinal plasticity. Four nights of platform over water paradigm (rats) increased c-fos immunoreactivity in the trigeminal nucleus upon supradural capsaicin infusion, suggesting increased spinal neuronal excitability, or longer time to return to normal sensitivity at the first relay of nociceptive transmission (Kim et al., 2019). After two nights of forced awakenings (humans), women displayed increased windup pain (but not men), while men developed mechanical secondary hyperalgesia (but not women) (Smith et al., 2019). These results indicate a possible spinal amplification of nociceptive signaling with probable sex difference. However, the lack of development of secondary allodynia in sleep-deprived subjects suggests that forced awakenings and moderate sleep loss do not trigger a state of central sensitization (where innocuous stimuli activate spinal nociceptive neurons and cause pain), but rather specifically amplify the nociceptive/pain signal. This amplification could involve a spinal facilitation, suggested by the increase in cfos spinal staining and windup pain readout (Figure 7B). The increased pain ratings during repeated nociceptive stimulations could also involve supraspinal changes (in brainstem, thalamus, cortex) or an increasing discomfort or anxiety with repeated painful stimulations.

Several lines of evidence suggest sleep loss could also dysregulate descending controls into the spinal cord. Traditional descending inhibitory controls are mostly recruited by the spino-periaqueductal gray (PAG) pathway, which originates from lamina I spinal projection neurons (Gauriau and Bernard, 2002). The PAG is critical for the analgesic effects of morphine (Fields, 2004; Ossipov et al., 2014). Under normal conditions, morphine binds to inhibitory PAG interneurons, which release their brake on glutamatergic PAG neurons that project onto the pain-inhibiting RVM neurons ("OFF-cells"). This disinhibition of RVM "OFF cells" allows the recruitment of inhibitory descending controls. After a single night of sleep deprivation, there is a loss of morphine analgesic efficacity in humans and rodents (Ukponmwan et al., 1984; Nascimento et al., 2007; Steinmiller et al., 2010; Skinner et al., 2011; Alexandre et al., 2017), which could be due in part to an hyperexcitability of GABAergic PAG interneurons preventing the recruitment of inhibitory descending pathways from the RVM (Tomim et al., 2016).

Administration of amitriptyline, a non-selective 5-HT/NA reuptake inhibitor (which increases levels of these monoamines in the spinal cord), improves heat pain hypersensitivity in rats after sleep loss (but not mechanical pain hypersensitivity), thus supporting the hypothesis of a partial loss of noradrenergic and serotoninergic descending controls (Wei et al., 2008; Wodarski et al., 2015). Amitriptyline has also been shown to be an agonist of Kv7.2/7.3 channels (Punke and Friederich, 2007), and selective activation of these channels phenocopied amitriptyline in sleep-deprived animals (Wodarski et al., 2015), which could suggest Kv7.2/7.3 are involved in sleep loss-induced heat hyperalgesia. However, one caveat is that activation of Kv7.2/7.3 channels causes analgesia in well-rested animals (Teng et al., 2016) and are not restricted to the RVM-spinal projections. Finally, in parallel to this possible loss of inhibitory descending controls, sleep loss increases RVM cholecystokininergic transmission, which promotes pronociceptive descending controls ("ON-cells"; Tomim et al., 2016).

Overall, several studies indicate a probable change in spinal-midbrain pathways' excitability after sleep loss. Sleep disturbances do not trigger a frank state of central sensitization, but rather a relative gain in pronociceptive signaling, in part mediated by changes in RVM spinal projections (Figure 7C). Whether this plasticity also involves intrinsic changes within the spinal nociceptive circuits remains unclear.

Brain: Mesolimbic system

While the spino-thalamo-cortical pathway plays a key role for the sensory-discriminative component of pain and the spino-parabrachial pathway generates its emotional (escape and aversive memory) and autonomic (heart rate, pupil dilatation, sweating) components (Gauriau and Bernard, 2002; Tracey et al., 2019; Chiang et al., 2020), several brain areas associated with cognitive and emotional processes can also modulate the final pain response based on its context (e.g., attention to pain, distraction analgesia, pain tolerance, etc.). The mesolimbic system could play a major role in this contextualization of the pain sensation, notably by determining the salience of the stimulus (Knutson et al., 2001; Roitman et al., 2005; Cooper and Knutson, 2008; Al-Hasani et al., 2015) and by modulating the reward system during and after a pain stimulation (Lammel et al., 2014; Navratilova et al., 2015; Yang et al., 2021). Several lines of evidence have recently pointed to a critical role of the mesolimbic system, notably the nucleus accumbens (NAc) on the proalgesic effects of sleep loss on pain (Finan et al., 2013; Sardi et al., 2018b; Alexandre et al., 2020). The NAc is subdivided in three functionally distinct areas: the NAc core, the medial part of the NAc shell, and the lateral part of the NAc shell (Lammel et al., 2014), which all likely play a distinct role in the overall pain experience. The NAc core is critical to determine the salience of stimuli (Chen and Bruchas, 2021), the medial NAc shell is involved in the aversive-encoding prediction of an unpleasant stimulus (including pain) (de Jong et al., 2019); the lateral NAc shell is a key structure for the reward system (Lammel et al., 2014; de Jong et al., 2019). NAc activity is strongly modulated by dopamine released from the ventral tegmental area (VTA) via mostly segregated pathways: the medial VTA projects to the NAc core and the medial NAc shell, while the lateral VTA almost exclusively projects onto the lateral NAc shell (Lammel et al., 2014). A painful stimulation causes an activation of DA neurons located in the most ventral part of the medial VTA (Brischoux et al., 2009; Lammel et al., 2014; de Jong et al., 2019), presumably reinforcing aversion (de Jong et al., 2019), while it mostly inhibits dopamine release in the lateral NAc shell (de Jong et al., 2019), likely blocking pleasant/reward pathways during ongoing pain. This inhibition of lateral VTA neurons is notably mediated by GABAergic SNR neurons, which are directly contacted by lateral parabrachial nucleus neurons (Yang et al., 2021). At the cessation of the pain stimulation, there is a rebound of dopamine release onto neurons of the lateral NAc shell, a phenomenon known as "pain cessation reward" (Navratilova et al., 2015).

Overall, these studies indicate that the dynamics of dopamine release onto different regions of NAc are extremely important for an adequate pain response. Functional magnetic resonance imaging (fMRI) studies found that sleep loss strongly alters these activation dynamics of NAc upon noxious stimulation (Krause et al., 2019; Seminowicz et al., 2019), notably through a defect in D2R/D3R transmission (Volkow et al., 2008, 2012). In rodents, excitotoxic lesions, local silencing of the NAc (Sardi et al., 2018a,b) or administration of modafinil (Alexandre et al., 2017), which enhances DAergic tone (Boutrel and Koob, 2004), normalize pain sensitivity without any analgesic activity on their own. This dysregulation of VTA-NAc function upon painful stimulation after sleep loss could affect how a nociceptive stimulus is evaluated (salience), how pain (or its cessation) is going to be anticipated, or dampen the positive sensation associated with the termination of a painful stimulus. VTA neurons are strongly implicated in sleep-wake behaviors and both DAergic and GABAergic neurons of the VTA are active during wakefulness and quiet during NREMS (Eban-Rothschild et al., 2016). Their sustained activation during extended wake coupled with a limited recovery during shorter sleep opportunity could contribute to the accumulation of sleep loss-induced hyperalgesia. In addition, an increased input from the spinalparabrachial pathway after sleep loss (originating from lamina I nociceptive neurons) could in turn overactivate SNR GABAergic neurons projecting to the lateral VTA, and thereby contribute to the reduced DAergic transmission onto NAc neurons and disrupt normal activation dynamics of this nucleus upon painful stimulation (Figure 7D).

The majority of NAc neurons expressing D2R express the excitatory adenosine A2A receptors (Valjent et al., 2009) (A2AR). Blockade of A2AR, notably by caffeine, normalizes pain in sleep-deprived animals (Alexandre et al., 2017; Sardi et al., 2018a) while local administration of A2AR agonists extends sleep loss-induced hyperalgesia in rats (Sardi et al., 2018b). Adenosine accumulates with wakefulness, which would contribute to the overexcitability of NAc neurons during extended wake.

Altogether, these data suggest that sleep loss might lead to a state of hyperexcitability of D2R/A2AR NAc neurons, which would be caused by a combination of reduced function of D2R (internalization, or decoupling of G-protein), but also an accumulation of adenosine during wakefulness activating A2AR (Gq-coupled). Restoring an inhibitory brake on these neurons (by promoting D2R or blocking A2AR) could restore normal pain sensitivity by maintaining the normal excitatory/inhibitory balance required for the different subnuclei of the NAc to function (Figure 7D). Finally, it is worth mentioning that in humans, laser-evoked potential signal was reduced after sleep loss, despite an increase in pain ratings (Tiede et al., 2010; Azevedo et al., 2011), which would suggest that the nociceptive input might not be increased in higher brain structures, but rather by promoting how the signal is perceived and contextualized—two major features of the mesolimbic system.

Conclusions

Despite significant differences in sleep-wake patterns between humans and rodents and a great heterogeneity in protocols used to disturb sleep, the effects of sleep disturbances on pain sensitivity are remarkably conserved across species. In addition, in all species assessed, females developed more pain hypersensitivity than males, and this might involve distinct neural mechanisms. We propose that the major driver for pain hypersensitivity is the total amount of sleep lost, while REMS loss by itself does not seem to have a direct effect on pain. REMS deprivation however increases the febrile response and might contribute to impaired recovery upon injury (Sutton and Opp, 2014; Wang et al., 2015; Vanini, 2016; Dai et al., 2022). The dynamics of sleep loss might affect different components of the pain response: sleep loss caused by extended wakefulness preferentially increases pain perception, whereas interrupted and limited sleep strongly dysregulates descending controls such as DNIC, especially in women.

Sleep disturbances affect pain pathways at several anatomical levels. Insufficient sleep causes a shift in the immune system toward a proinflammatory profile. While the increase of these proinflammatory factors (notably prostaglandins and cytokines) at the periphery is not sufficient to trigger evoked pain hypersensitivity, these agents might promote migraine as well as the risk of developing abnormal pain after injury (and inflammation). Sleep disturbances might enhance pain transmission at the spinal level, without however causing a state of full central sensitization (i.e., no development of allodynia). This increased spinal excitability has a strong sexual dimorphism, which could be caused by sex-dependent spinal mechanisms or changes from supraspinal structures. For example, sleep disturbances affect DNICs only in women, while men are mostly spared. Sleep loss causes an overexcitability of PAG neurons, which reduces morphine analgesic efficacy and could also alter "traditional" descending controls. Finally, sleep loss caused by an extended wake, similar to that experienced by a large body of the general population, might preferentially affect the mesolimbic system. This system plays a key role in the emotional and cognitive component of the pain response. Sleep loss-induced dysfunctions of the mesolimbic system could increase pain by promoting the attention to pain, amplifying catastrophizing (expectation of when a painful stimulus will stop), and reducing distraction analgesia.

Because the deleterious effects of sleep loss on pain sensitivity are cumulative, they likely contribute to the complex sleep and pain profiles observed in chronic clinical settings. Understanding how specific pain components are dysregulated by different sleep disturbances will help decipher how they interact over time, and the best strategies to promote sleep features that normalize pain sensitivity.

Methodology

We searched for articles studying the effects of sleep disturbances on pain on PubMed and Google Scholar. Key words included "sleep deprivation"; "sleep loss"; "chronic sleep deprivation"; "chronic sleep loss"; "chronic sleep restriction"; "REMS deprivation"; "total sleep deprivation"; "pain"; "nociception" and their combination. For this review, we selected studies that measured evoked pain sensitivity in laboratory settings (no diaries). Only experiments in healthy subjects were used. For animal studies, we only used data from healthy animals (in some cases, non-injured animals that acted as controls to injury models). Determination of pain hypersensitivity (or normal pain sensitivity) was analyzed for all time points reported within the studies. In studies using multiple sleep disruption protocols, we only analyzed the pain results from the first disruption used. If studies not obtained by the PubMed or Google Scholar searches were identified by reading manuscripts, they were added to this study.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Author contributions

KK, CA, and AL analyzed the data and wrote the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by NIH grant R01NS112266 (AL) and the Blaustein Pain Research Fund Grants (to AL and CA).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Aho, V., Ollila, H. M., Rantanen, V., Kronholm, E., Surakka, I., Van Leeuwen, W. M., et al. (2013). Partial sleep restriction activates immune response-related gene expression pathways: experimental and epidemiological studies in humans. *PLoS ONE* 8, e77184. doi: 10.1371/journal.pone.0077184

Alexandre, C., Latremoliere, A., Ferreira, A., Miracca, G., Yamamoto, M., Scammell, T. E., et al. (2017). Decreased alertness due to sleep loss increases pain sensitivity in mice. *Nat. Med.* 23, 768–774. doi: 10.1038/nm.4329

Alexandre, C., Latremoliere, A., and Finan, P. H. (2020). "Effect of sleep loss on pain," in *The Oxford Handbook of the Neurobiology of Pain* (Oxford University Press), 557–608.

Al-Hasani, R., Mccall, J. G., Shin, G., Gomez, A. M., Schmitz, G. P., Bernardi, J. M., et al. (2015). Distinct subpopulations of nucleus accumbens dynorphin neurons drive aversion and reward. *Neuron* 87, 1063–1077. doi: 10.1016/j.neuron.2015.08.019

Araujo, P., Mazaro-Costa, R., Tufik, S., and Andersen, M. L. (2011). Impact of sex on hyperalgesia induced by sleep loss. *Horm. Behav.* 59, 174–179. doi: 10.1016/j.yhbeh.2010.11.003

Arima, T., Svensson, P., Rasmussen, C., Nielsen, K. D., Drewes, A. M., and Arendt-Nielsen, L. (2001). The relationship between selective sleep deprivation, nocturnal jaw-muscle activity and pain in healthy men. *J. Oral Rehabil.* 28, 140–148. doi: 10.1046/j.1365-2842.2001.00687.x

Aserinsky, E., and Kleitman, N. (1953). Regularly occurring periods of eye motility, and concomitant phenomena, during sleep. *Science* 118, 273–274. doi: 10.1126/science.118.3062.273

Axelsson, J., Rehman, J.-U., Akerstedt, T., Ekman, R., Miller, G. E., Höglund, C. O., et al. (2013). Effects of sustained sleep restriction on mitogen-stimulated cytokines, chemokines and T helper 1/T helper 2 balance in humans. *PLoS ONE* 8, e82291. doi: 10.1371/journal.pone.0082291

Azevedo, E., Manzano, G. M., Silva, A., Martins, R., Andersen, M. L., and Tufik, S. (2011). The effects of total and REM sleep deprivation on laser-evoked potential threshold and pain perception. *Pain* 152, 2052–2058. doi:10.1016/j.pain.2011.04.032

Baba, H., Kohno, T., Moore, K. A., and Woolf, C. J. (2001). Direct activation of rat spinal dorsal horn neurons by prostaglandin E2. *J. Neurosci.* 21, 1750–1756. doi: 10.1523/JNEUROSCI.21-05-01750.2001

Banks, S., Van Dongen, H. P., Maislin, G., and Dinges, D. F. (2010). Neurobehavioral dynamics following chronic sleep restriction: dose-response effects of one night for recovery. *Sleep* 33, 1013–1026. doi: 10.1093/sleep/33.8.1013

Basbaum, A. I., and Fields, H. L. (1978). Endogenous pain control mechanisms: review and hypothesis. *Ann. Neurol.* 4, 451–462. doi: 10.1002/ana.410040511

Basbaum, A. I., and Fields, H. L. (1984). Endogenous pain control systems: brainstem spinal pathways and endorphin circuitry. *Annu. Rev. Neurosci.* 7, 309–338. doi: 10.1146/annurev.ne.07.030184.001521

Besedovsky, L., Lange, T., and Born, J. (2012). Sleep and immune function. *Pflugers Arch.* 463, 121-137. doi: 10.1007/s00424-011-1044-0

Binshtok, A. M., Wang, H., Zimmermann, K., Amaya, F., Vardeh, D., Shi, L., et al. (2008). Nociceptors are interleukin-1beta sensors. *J. Neurosci.* 28, 14062–14073. doi: 10.1523/JNEUROSCI.3795-08.2008

Bjurstrom, M. F., and Irwin, M. R. (2016). Polysomnographic characteristics in nonmalignant chronic pain populations: A review of controlled studies. *Sleep Med. Rev.* 26, 74–86. doi: 10.1016/j.smrv.2015.03.004

Borbely, A. A. (1982). A two process model of sleep regulation. *Hum. Neurobiol.* 1, 195–204.

Borbely, A. A., and Achermann, P. (1999). Sleep homeostasis and models of sleep regulation. J. Biol. Rhythms 14, 557–568. doi: 10.1177/074873099129000894

Bosma, R. L., Ameli Mojarad, E., Leung, L., Pukall, C., Staud, R., and Stroman, P. W. (2015). Neural correlates of temporal summation of second pain

in the human brainstem and spinal cord. Hum. Brain Mapp. 36, 5038-5050. doi: 10.1002/hbm.22993

Bouhassira, D., Bing, Z., and Le Bars, D. (1992a). Effects of lesions of locus coeruleus/subcoeruleus on diffuse noxious inhibitory controls in the rat. *Brain Res.* 571, 140–144. doi: 10.1016/0006-8993(92)90520-J

Bouhassira, D., Villanueva, L., Bing, Z., and Le Bars, D. (1992b). Involvement of the subnucleus reticularis dorsalis in diffuse noxious inhibitory controls in the rat. *Brain Res.* 595, 353–357. doi: 10.1016/0006-8993(92)91071-L

Boutrel, B., and Koob, G. F. (2004). What keeps us awake: the neuropharmacology of stimulants and wakefulness-promoting medications. *Sleep* 27, 1181–1194. doi: 10.1093/sleep/27.6.1181

Brenn, D., Richter, F., and Schaible, H. G. (2007). Sensitization of unmyelinated sensory fibers of the joint nerve to mechanical stimuli by interleukin-6 in the rat: an inflammatory mechanism of joint pain. *Arthritis Rheum.* 56, 351–359. doi: 10.1002/art.22282

Brischoux, F., Chakraborty, S., Brierley, D. I., and Ungless, M. A. (2009). Phasic excitation of dopamine neurons in ventral VTA by noxious stimuli. *Proc. Natl. Acad. Sci. U. S. A.* 106, 4894–4899. doi: 10.1073/pnas.0811507106

Bryant, P. A., Trinder, J., and Curtis, N. (2004). Sick and tired: does sleep have a vital role in the immune system? *Nat. Rev. Immunol.* 4, 457-467. doi: 10.1038/nri1369

Busch, V., Haas, J., Cronlein, T., Pieh, C., Geisler, P., Hajak, G., et al. (2012). Sleep deprivation in chronic somatoform pain-effects on mood and pain regulation. *Psychiatry Res.* 195, 134–143. doi: 10.1016/j.psychres.2011.07.021

Carroll, J. E., Carrillo, C., Olmstead, R., Witarama, T., Breen, E. C., Yokomizo, M., et al. (2015). Sleep deprivation and divergent toll-like receptor-4 activation of cellular inflammation in aging. *Sleep* 38, 205–211. doi: 10.5665/sleep.4398

Chapman, V., and Dickenson, A. H. (1992). The spinal and peripheral roles of bradykinin and prostaglandins in nociceptive processing in the rat. *Eur. J. Pharmacol.* 219, 427–433. doi: 10.1016/0014-2999(92)90484-L

Chen, J., and Bruchas, M. (2021). Neuromodulation: a model for dopamine in salience encoding. *Curr. Biol.* 31, R1426–R1429. doi: 10.1016/j.cub.2021.09.038

Chen, X., Tanner, K., and Levine, J. D. (1999). Mechanical sensitization of cutaneous C-fiber nociceptors by prostaglandin E2 in the rat. *Neurosci. Lett.* 267, 105–108. doi: 10.1016/S0304-3940(99)00345-6

Chiang, M. C., Nguyen, E. K., Canto-Bustos, M., Papale, A. E., Oswald, A. M., and Ross, S. E. (2020). Divergent neural pathways emanating from the lateral parabrachial nucleus mediate distinct components of the pain response. *Neuron* 106, 927–939.e5. doi: 10.1016/j.neuron.2020.03.014

Chiu, I. M., Heesters, B. A., Ghasemlou, N., Von Hehn, C. A., Zhao, F., Tran, J., et al. (2013). Bacteria activate sensory neurons that modulate pain and inflammation. *Nature* 501, 52–57. doi: 10.1038/nature12479

Chouchou, F., Chauny, J. M., Rainville, P., and Lavigne, G. J. (2015). Selective REM sleep deprivation improves expectation-related placebo analgesia. *PLoS ONE* 10, e0144992. doi: 10.1371/journal.pone.0144992

Christoffersson, G., Vagesjo, E., Pettersson, U. S., Massena, S., Nilsson, E. K., Broman, J. E., et al. (2014). Acute sleep deprivation in healthy young men: impact on population diversity and function of circulating neutrophils. *Brain Behav. Immun.* 41, 162–172. doi: 10.1016/j.bbi.2014.05.010

Coenen, A., and Van Luijtelaar, E. (1985). Stress induced by three procedures of deprivation of paradoxical sleep. *Physiol. Behav.* 35, 501–504. doi: 10.1016/0031-9384(85)90130-1

Cooper, J. C., and Knutson, B. (2008). Valence and salience contribute to nucleus accumbens activation. *Neuroimage* 39, 538–547. doi: 10.1016/j.neuroimage.2007.08.009

Dai, Y., Liu, S., Chen, J., Liu, L., Zhou, C., and Zuo, Y. (2022). Microglial responses and pain behaviors are exacerbated by chronic sleep deprivation in

rats with chronic pain via neuroinflammatory pathways. *Neuroscience* 503, 83–94. doi: 10.1016/j.neuroscience.2022.09.004

Damasceno, F., Skinner, G. O., Gomes, A., Araujo, P. C., and De Almeida, O. M. (2009). Systemic amitriptyline administration does not prevent the increased thermal response induced by paradoxical sleep deprivation. *Pharmacol. Biochem. Behav.* 94, 51–55. doi: 10.1016/j.pbb.2009.07.005

de Jong, J. W., Afjei, S. A., Pollak Dorocic, I., Peck, J. R., Liu, C., Kim, C. K., et al. (2019). A neural circuit mechanism for encoding aversive stimuli in the mesolimbic dopamine system. *Neuron* 101, 133–151.e7. doi: 10.1016/j.neuron.2018.11.005

Diaz-Piedra, C., Catena, A., Sanchez, A. I., Miro, E., Martinez, M. P., and Buela-Casal, G. (2015). Sleep disturbances in fibromyalgia syndrome: the role of clinical and polysomnographic variables explaining poor sleep quality in patients. *Sleep Med.* 16, 917–925. doi: 10.1016/j.sleep.2015.03.011

Dimitrov, S., Besedovsky, L., Born, J., and Lange, T. (2015). Differential acute effects of sleep on spontaneous and stimulated production of tumor necrosis factor in men. *Brain Behav. Immun.* 47, 201–210. doi: 10.1016/j.bbi.2014.11.017

Drewes, A. M., Rossel, P., Arendt-Nielsen, L., Nielsen, K. D., Hansen, L. M., Birket-Smith, L., et al. (1997). Sleepiness does not modulate experimental joint pain in healthy volunteers. *Scand. J. Rheumatol.* 26, 399–400. doi: 10.3109/03009749709065709

Eban-Rothschild, A., Rothschild, G., Giardino, W. J., Jones, J. R., and De Lecea, L. (2016). VTA dopaminergic neurons regulate ethologically relevant sleep-wake behaviors. *Nat. Neurosci.* 19, 1356–1366. doi: 10.1038/nn.4377

Edwards, R. R., Grace, E., Peterson, S., Klick, B., Haythornthwaite, J. A., and Smith, M. T. (2009). Sleep continuity and architecture: associations with pain-inhibitory processes in patients with temporomandibular joint disorder. *Eur. J. Pain* 13, 1043–1047. doi: 10.1016/j.ejpain.2008.12.007

Eichhorn, N., Treede, R. D., and Schuh-Hofer, S. (2018). The role of sex in sleep deprivation related changes of nociception and conditioned pain modulation. *Neuroscience* 387, 191–200. doi: 10.1016/j.neuroscience.2017.09.044

Everson, C. A., and Szabo, A. (2011). Repeated exposure to severely limited sleep results in distinctive and persistent physiological imbalances in rats. *PLoS ONE* 6, e22987. doi: 10.1371/journal.pone.0022987

Faraut, B., Boudjeltia, K. Z., Vanhamme, L., and Kerkhofs, M. (2012). Immune, inflammatory and cardiovascular consequences of sleep restriction and recovery. *Sleep Med. Rev.* 16, 137–149. doi: 10.1016/j.smrv.2011.05.001

Faraut, B., Leger, D., Medkour, T., Dubois, A., Bayon, V., Chennaoui, M., et al. (2015). Napping reverses increased pain sensitivity due to sleep restriction. *PLoS ONE* 10, e0117425. doi: 10.1371/journal.pone.0117425

Fields, H. (2004). State-dependent opioid control of pain. Nat. Rev. Neurosci. 5, 565. doi: 10.1038/nrn1431

Finan, P. H., Goodin, B. R., and Smith, M. T. (2013). The association of sleep and pain: an update and a path forward. *J. Pain* 14, 1539–1552. doi: 10.1016/j.jpain.2013.08.007

Finan, P. H., Quartana, P. J., Remeniuk, B., Garland, E. L., Rhudy, J. L., Hand, M., et al. (2017). Partial sleep deprivation attenuates the positive affective system: effects across multiple measurement modalities. *Sleep* 40, zsw017. doi: 10.1093/sleep/zsw017

Franzen, P. L., Buysse, D. J., Dahl, R. E., Thompson, W., and Siegle, G. J. (2009). Sleep deprivation alters pupillary reactivity to emotional stimuli in healthy young adults. *Biol. Psychol.* 80, 300–305. doi: 10.1016/j.biopsycho.2008.10.010

Freire, C., Sennes, L. U., and Polotsky, V. Y. (2022). Opioids and obstructive sleep apnea. J. Clin. Sleep Med. 18, 647–652. doi: 10.5664/jcsm.9730

Gauriau, C., and Bernard, J. F. (2002). Pain pathways and parabrachial circuits in the rat. *Exp. Physiol.* 87, 251–258. doi: 10.1113/eph8702357

Goel, N., Rao, H., Durmer, J. S., and Dinges, D. F. (2009). Neurocognitive consequences of sleep deprivation. *Semin. Neurol.* 29, 320–339. doi: 10.1055/s-0029-1237117

Grahnstedt, S., and Ursin, R. (1985). Platform sleep deprivation affects deep slow wave sleep in addition to REM sleep. *Behav. Brain Res.* 18, 233–239. doi: 10.1016/0166-4328(85)90031-2

Guariniello, L. D., Vicari, P., Lee, K. S., De Oliveira, A. C., and Tufik, S. (2012). Bone marrow and peripheral white blood cells number is affected by sleep deprivation in a murine experimental model. *J. Cell. Physiol.* 227, 361–366. doi: 10.1002/jcp.22743

Gurel, E. E., Ural, K., Ozturk, G., and Ozturk, L. (2014). Flurbiprofen in rapid eye movement sleep deprivation induced hyperalgesia. *Physiol. Behav.* 128, 155–158. doi: 10.1016/j.physbeh.2014.02.003

Haack, M., Lee, E., Cohen, D. A., and Mullington, J. M. (2009). Activation of the prostaglandin system in response to sleep loss in healthy humans:

potential mediator of increased spontaneous pain. Pain 145, 136-141. doi: 10.1016/j.pain.2009.05.029

Haack, M., and Mullington, J. M. (2005). Sustained sleep restriction reduces emotional and physical well-being. *Pain* 119, 56–64. doi: 10.1016/j.pain.2005.09.011

Haack, M., Sanchez, E., and Mullington, J. M. (2007). Elevated inflammatory markers in response to prolonged sleep restriction are associated with increased pain experience in healthy volunteers. *Sleep* 30, 1145–1152. doi: 10.1093/sleep/30.9.1145

Hambrecht-Wiedbusch, V. S., Gabel, M., Liu, L. J., Imperial, J. P., Colmenero, A. V., and Vanini, G. (2017). Preemptive caffeine administration blocks the increase in postoperative pain caused by previous sleep loss in the rat: a potential role for preoptic adenosine A2A receptors in sleep-pain interactions. *Sleep* 40. doi: 10.1093/sleep/zsx116

Harris, S. G., Padilla, J., Koumas, L., Ray, D., and Phipps, R. P. (2002). Prostaglandins as modulators of immunity. *Trends Immunol.* 23, 144–150. doi: 10.1016/S1471-4906(01)02154-8

Harvey, M. T., Kline, R. H. T., May, M. E., Roberts, A. C., Valdovinos, M. G., Wiley, R. G., et al. (2010). Parametric analysis of thermal preference following sleep deprivation in the rat. *Neurosci. Lett.* 485, 98–101. doi: 10.1016/j.neulet.2010.08.070

Hicks, R. A., Coleman, D. D., Ferrante, F., Sahatjian, M., and Hawkins, J. (1979). Pain thresholds in rats during recovery from REM sleep deprivation. *Percept. Mot. Skills* 48, 687–690. doi: 10.2466/pms.1979.48.3.687

Holingue, C., Wennberg, A., Berger, S., Polotsky, V. Y., and Spira, A. P. (2018). Disturbed sleep and diabetes: A potential nexus of dementia risk. *Metab. Clin. Exp.* 84, 85–93. doi: 10.1016/j.metabol.2018.01.021

Houle, T. T., Butschek, R. A., Turner, D. P., Smitherman, T. A., Rains, J. C., and Penzien, D. B. (2012). Stress and sleep duration predict headache severity in chronic headache sufferers. *Pain* 153, 2432–2440. doi: 10.1016/j.pain.2012.08.014

Hu, J., Chen, Z., Gorczynski, C., Gorczynski, L., Kai, Y., Lee, L., et al. (2003). Sleep-deprived mice show altered cytokine production manifest by perturbations in serum IL-1ra, TNFa, and IL-6 levels. *Brain Behav. Immun.* 17, 498–504. doi: 10.1016/j.bbi.2003.03.001

Hunt, C. A., Mun, C. J., Owens, M. A., Lerman, S. F., Kunatharaju, S., Tennen, H. A., et al. (2022). Sleep, positive affect, and circulating interleukin-6 in women with temporomandibular joint disorder. *Psychosom. Med.* 84, 383–392. doi: 10.1097/PSY.000000000001047

Hunt, C. A., Smith, M. T., Mun, C. J., Irwin, M. R., and Finan, P. H. (2021). Trait positive affect buffers the association between experimental sleep disruption and inflammation. *Psychoneuroendocrinology* 129, 105240. doi:10.1016/j.psyneuen.2021.105240

Iacovides, S., George, K., Kamerman, P., and Baker, F. C. (2017). Sleep fragmentation hypersensitizes healthy young women to deep and superficial experimental pain. *J Pain* 18, 844–854. doi: 10.1016/j.jpain.2017. 02.436

Ibarra-Coronado, E. G., Velazquéz-Moctezuma, J., Diaz, D., Becerril-Villanueva, L. E., Pavón, L., and Morales-Montor, J. (2015). Sleep deprivation induces changes in immunity in *Trichinella spiralis*-infected rats. *Int. J. Biol. Sci.* 11, 901. doi: 10.7150/ijbs.11907

Imeri, L., and Opp, M. R. (2009). How (and why) the immune system makes us sleep. *Nat. Rev. Neurosci.* 10, 199–210. doi: 10.1038/nrn2576

Irwin, M. R., Olmstead, R., Bjurstrom, M. F., Finan, P. H., and Smith, M. T. (2022). Sleep disruption and activation of cellular inflammation mediate heightened pain sensitivity: a randomized clinical trial. *Pain.* doi: 10.1097/j.pain.00000000002811

Irwin, M. R., Olmstead, R., and Carroll, J. E. (2016). Sleep disturbance, sleep duration, and inflammation: a systematic review and meta-analysis of cohort studies and experimental sleep deprivation. *Biol. Psychiatry* 80, 40–52. doi: 10.1016/j.biopsych.2015.05.014

Irwin, M. R., and Opp, M. R. (2017). Sleep health: reciprocal regulation of sleep and innate immunity. *Neuropsychopharmacology* 42, 129–155. doi: 10.1038/npp.2016.148

Irwin, M. R., Wang, M., Ribeiro, D., Cho, H. J., Olmstead, R., Breen, E. C., et al. (2008). Sleep loss activates cellular inflammatory signaling. *Biol. Psychiatry* 64, 538–540. doi: 10.1016/j.biopsych.2008.05.004

Irwin, M. R., Witarama, T., Caudill, M., Olmstead, R., and Breen, E. C. (2015). Sleep loss activates cellular inflammation and signal transducer and activator of transcription (STAT) family proteins in humans. *Brain Behav. Immun.* 47, 86–92. doi: 10.1016/j.bbi.2014.09.017

Jouvet, D., Vimont, P., and Delorme, F. (1964). Étude de la privation sélective de la phase paradoxale du sommeil chez le chat. *J. Physiol.*

Jung, C. M., Ronda, J. M., Czeisler, C. A., and Wright, K. P. Jr. (2011). Comparison of sustained attention assessed by auditory and visual psychomotor vigilance tasks prior to and during sleep deprivation. *J. Sleep Res.* 20, 348–355. doi: 10.1111/j.1365-2869.2010.00877.x

Kawasaki, Y., Zhang, L., Cheng, J. K., and Ji, R. R. (2008). Cytokine mechanisms of central sensitization: distinct and overlapping role of interleukinlbeta, interleukin-6, and tumor necrosis factor-alpha in regulating synaptic and neuronal activity in the superficial spinal cond. *J. Neurosci.* 28, 5189–5194. doi: 10.1523/JNEUROSCI.3338-07.2008

Kim, S. H., Park, J. Y., Shin, H. E., Lee, S. B., Ryu, D. W., Kim, T. W., et al. (2019). The influence of rapid eye movement sleep deprivation on nociceptive transmission and the duration of facial allodynia in rats: a behavioral and Fos immunohistochemical study. *J. Headache Pain* 20, 21. doi:10.1186/s10194-019-0977-0

Knutson, B., Adams, C. M., Fong, G. W., and Hommer, D. (2001). Anticipation of increasing monetary reward selectively recruits nucleus accumbens. *J. Neurosci.* 21, RC159. doi: 10.1523/JNEUROSCI.21-16-j0002.2001

Kohansieh, M., and Makaryus, A. N. (2015). Sleep deficiency and deprivation leading to cardiovascular disease. *Int. J. Hypertens.* 2015, 5. doi: 10.1155/2015/615681

Krause, A. J., Prather, A. A., Wager, T. D., Lindquist, M. A., and Walker, M. P. (2019). The pain of sleep loss: a brain characterization in humans. *J. Neurosci.* 39, 2291–2300. doi: 10.1523/JNEUROSCI.2408-18.2018

Kucharczyk, M. W., Di Domenico, F., and Bannister, K. (2022). Distinct brainstem to spinal cord noradrenergic pathways inversely regulate spinal neuronal activity. *Brain*. 145:2293–2300. doi: 10.1093/brain/awac085

Kundermann, B., Hemmeter-Spernal, J., Huber, M. T., Krieg, J. C., and Lautenbacher, S. (2008). Effects of total sleep deprivation in major depression: overnight improvement of mood is accompanied by increased pain sensitivity and augmented pain complaints. *Psychosom. Med.* 70, 92–101. doi:10.1097/PSY.0b013e31815c1b5d

Kundermann, B., Spernal, J., Huber, M. T., Krieg, J. C., and Lautenbacher, S. (2004). Sleep deprivation affects thermal pain thresholds but not somatosensory thresholds in healthy volunteers. *Psychosom. Med.* 66, 932–937. doi: 10.1097/01.psy.0000145912. 24553.c0

Lammel, S., Lim, B. K., and Malenka, R. C. (2014). Reward and aversion in a heterogeneous midbrain dopamine system. *Neuropharmacology* 76, 351–359. doi: 10.1016/j.neuropharm.2013.03.019

Lange, T., Dimitrov, S., and Born, J. (2010). Effects of sleep and circadian rhythm on the human immune system. *Ann. N. Y. Acad. Sci.* 1193, 48–59. doi: 10.1111/j.1749-6632.2009.05300.x

Larson, R. A., and Carter, J. R. (2016). Total sleep deprivation and pain perception during cold noxious stimuli in humans. *Scand. J. Pain* 13, 12–16. doi: 10.1016/j.sjpain.2016.05.037

Lasselin, J., Rehman, J. U., Akerstedt, T., Lekander, M., and Axelsson, J. (2015). Effect of long-term sleep restriction and subsequent recovery sleep on the diurnal rhythms of white blood cell subpopulations. *Brain Behav. Immun.* 47, 93–99. doi: 10.1016/j.bbi.2014.10.004

Latremoliere, A., and Woolf, C. J. (2009). Central sensitization: a generator of pain hypersensitivity by central neural plasticity. *J. Pain* 10, 895–926. doi: 10.1016/j.jpain.2009.06.012

Lautenbacher, S., Kundermann, B., and Krieg, J. C. (2006). Sleep deprivation and pain perception. *Sleep Med. Rev.* 10, 357–369. doi: 10.1016/j.smrv.2005.08.001

Lautenbacher, S., and Rollman, G. B. (1997). Possible deficiencies of pain modulation in fibromyalgia. *Clin. J. Pain* 13, 189–196. doi: 10.1097/00002508-199709000-00003

Le Bars, D. (2002). The whole body receptive field of dorsal horn multireceptive neurones. *Brain Res. Rev.* 40, 29–44. doi: 10.1016/S0165-0173(02)00186-8

Le Bars, D., Villanueva, L., Bouhassira, D., and Willer, J. C. (1992). Diffuse noxious inhibitory controls (DNIC) in animals and in man. *Patol. Fiziol. Eksp. Ter.* 55-65.

Leite-Almeida, H., Valle-Fernandes, A., and Almeida, A. (2006). Brain projections from the medullary dorsal reticular nucleus: an anterograde and retrograde tracing study in the rat. *Neuroscience* 140, 577–595. doi: 10.1016/j.neuroscience.2006.02.022

Lentz, M. J., Landis, C. A., Rothermel, J., and Shaver, J. L. (1999). Effects of selective slow wave sleep disruption on musculoskeletal pain and fatigue in middle aged women. *J. Rheumatol.* 26, 1586–1592.

Letzen, J. E., Remeniuk, B., Smith, M. T., Irwin, M. R., Finan, P. H., and Seminowicz, D. A. (2020). Individual differences in pain sensitivity are associated

with cognitive network functional connectivity following one night of experimental sleep disruption. *Hum. Brain Mapp.* 41, 581–593. doi: 10.1002/hbm.24824

Li, Y. D., Ge, J., Luo, Y. J., Xu, W., Wang, J., Lazarus, M., et al. (2020). High cortical delta power correlates with aggravated allodynia by activating anterior cingulate cortex GABAergic neurons in neuropathic pain mice. *Pain* 161, 288–299. doi: 10.1097/j.pain.00000000001725

Liberalesso, P. B., D'andrea, K. F., Cordeiro, M. L., Zeigelboim, B. S., Marques, J. M., and Jurkiewicz, A. L. (2012). Effects of sleep deprivation on central auditory processing. *BMC Neurosci.* 13, 83. doi: 10.1186/1471-2202-13-83

Liu, P. Y., Irwin, M. R., Krueger, J. M., Gaddameedhi, S., and Van Dongen, H. P. A. (2021). Night shift schedule alters endogenous regulation of circulating cytokines. *Neurobiol. Sleep Circadian Rhythms* 10, 100063. doi: 10.1016/j.nbscr.2021.100063

Lou, H., Lu, J., Choi, E. B., Oh, M. H., Jeong, M., Barmettler, S., et al. (2017). Expression of IL-22 in the skin causes Th2-biased immunity, epidermal barrier dysfunction, and pruritus via stimulating epithelial Th2 cytokines and the GRP pathway. *J. Immunol.* 198, 2543–2555. doi: 10.4049/jimmunol.1600126

Machado, R. B., Hipolide, D. C., Benedito-Silva, A. A., and Tufik, S. (2004). Sleep deprivation induced by the modified multiple platform technique: quantification of sleep loss and recovery. *Brain Res.* 1004, 45–51. doi: 10.1016/j.brainres.2004.01.019

Matre, D., Andersen, M. R., Knardahl, S., and Nilsen, K. B. (2016). Conditioned pain modulation is not decreased after partial sleep restriction. *Eur. J. Pain* 20, 408–416. doi: 10.1002/ejp.741

Matre, D., Hu, L., Viken, L. A., Hjelle, I. B., Wigemyr, M., Knardahl, S., et al. (2015). Experimental sleep restriction facilitates pain and electrically induced cortical responses. *Sleep* 38, 1607–1617. doi: 10.5665/sleep.5058

Matre, D., Knardahl, S., and Nilsen, K. B. (2017). Night-shift work is associated with increased pain perception. *Scand. J. Work Environ. Health* 43, 260–268. doi: 10.5271/sjweh.3627

May, M. E., Harvey, M. T., Valdovinos, M. G., Kline, R. H. T., Wiley, R. G., and Kennedy, C. H. (2005). Nociceptor and age specific effects of REM sleep deprivation induced hyperalgesia. *Behav. Brain Res.* 159, 89–94. doi: 10.1016/j.bbr.2004.10.005

McHill, A. W., Hull, J. T., Wang, W., Czeisler, C. A., and Klerman, E. B. (2018). Chronic sleep curtailment, even without extended (>16-h) wakefulness, degrades human vigilance performance. *Proc. Nat. Acad. Sci.* 115, 6070–6075. doi: 10.1073/pnas.1706694115

Mendelson, W. B., Guthrie, R. D., Frederick, G., and Wyatt, R. J. (1974). The flower pot technique of rapid eye movement (REM) sleep deprivation. *Pharmacol. Biochem. Behav.* 2, 553–556. doi: 10.1016/0091-3057(74)90018-5

Moldofsky, H., and Scarisbrick, P. (1976). Induction of neurasthenic musculoskeletal pain syndrome by selective sleep stage deprivation. *Psychosom. Med.* 38:35–44. doi: 10.1097/00006842-197601000-00006

Moldofsky, H., Scarisbrick, P., England, R., and Smythe, H. (1975). Musculosketal symptoms and non-REM sleep disturbance in patients with "fibrositis syndrome" and healthy subjects. *Psychosom. Med.* 37, 341-351. doi: 10.1097/00006842-197507000-00008

Nascimento, D. C., Andersen, M. L., Hipolide, D. C., Nobrega, J. N., and Tufik, S. (2007). Pain hypersensitivity induced by paradoxical sleep deprivation is not due to altered binding to brain mu-opioid receptors. *Behav. Brain Res.* 178, 216–220. doi: 10.1016/j.bbr.2006.12.016

Nasehi, M., Mosavi-Nezhad, S. M., Khakpai, F., and Zarrindast, M. R. (2018). The role of omega-3 on modulation of cognitive deficiency induced by REM sleep deprivation in rats. *Behav. Brain Res.* 351, 152–160. doi: 10.1016/j.bbr.2018.06.002

National Sleep Foundation (2015). 2015 sleep in America poll -pain and sleep. Sleep Health 1, e14-e375. doi: 10.1016/j.sleh.2015.02.005

Navratilova, E., Atcherley, C. W., and Porreca, F. (2015). Brain circuits encoding reward from pain relief. *Trends Neurosci.* 38, 741–750. doi: 10.1016/j.tins.2015.09.003

Nunes, J. O., De Souza Apostolico, J., Andrade, D. A., Ruiz, F. S., Fernandes, E. R., Andersen, M. L., et al. (2018). Sleep deprivation predisposes allergic mice to neutrophilic lung inflammation. *J. Allergy Clin. Immunol.* 141, 1018–1027.e4. doi: 10.1016/j.jaci.2017.06.025

Odegard, S. S., Omland, P. M., Nilsen, K. B., Stjern, M., Gravdahl, G. B., and Sand, T. (2015). The effect of sleep restriction on laser evoked potentials, thermal sensory and pain thresholds and suprathreshold pain in healthy subjects. *Clin. Neurophysiol.* 126, 1979–1987. doi: 10.1016/j.clinph.2014.12.011

Odegard, S. S., Sand, T., Engstrom, M., Stovner, L. J., Zwart, J. A., and Hagen, K. (2011). The long-term effect of insomnia on primary headaches: a prospective population-based cohort study (HUNT-2 and HUNT-3). *Headache* 51, 570–580. doi: 10.1111/j.1526-4610.2011.01859.x

Older, S. A., Battafarano, D. F., Danning, C. L., Ward, J. A., Grady, E. P., Derman, S., et al. (1998). The effects of delta wave sleep interruption on pain thresholds and fibromyalgia-like symptoms in healthy subjects; correlations with insulin-like growth factor I. J. Rheumatol. 25, 1180–1186.

Onen, S. H., Alloui, A., Eschalier, A., and Dubray, C. (2000). Vocalization thresholds related to noxious paw pressure are decreased by paradoxical sleep deprivation and increased after sleep recovery in rat. *Neurosci. Lett.* 291, 25–28. doi: 10.1016/S0304-3940(00)01383-5

Onen, S. H., Alloui, A., Gross, A., Eschallier, A., and Dubray, C. (2001). The effects of total sleep deprivation, selective sleep interruption and sleep recovery on pain tolerance thresholds in healthy subjects. *J. Sleep Res.* 10, 35–42. doi: 10.1046/j.1365-2869.2001.00240.x

Ossipov, M. H., Morimura, K., and Porreca, F. (2014). Descending pain modulation and chronification of pain. *Curr. Opin. Support. Palliat. Care* 8, 143. doi: 10.1097/SPC.00000000000055

Page, G. G., Opp, M. R., and Kozachik, S. L. (2014). Reduced sleep, stress responsivity, and female sex contribute to persistent inflammation-induced mechanical hypersensitivity in rats. *Brain Behav. Immun.* 40, 244–251. doi: 10.1016/j.bbi.2014.02.013

Palma, J.-A., Urrestarazu, E., and Iriarte, J. (2013). Sleep loss as risk factor for neurologic disorders: a review. *Sleep Med.* 14, 229–236. doi: 10.1016/j.sleep.2012.11.019

Pinto, L. G., Cunha, T. M., Vieira, S. M., Lemos, H. P., Verri, W. A. Jr., Cunha, F. Q., et al. (2010). IL-17 mediates articular hypernociception in antigen-induced arthritis in mice. *Pain* 148, 247–256. doi: 10.1016/j.pain.2009.11.006

Pinto, L. G., Talbot, J., Peres, R. S., Franca, R. F., Ferreira, S. H., Ryffel, B., et al. (2015). Joint production of IL-22 participates in the initial phase of antigen-induced arthritis through IL-1beta production. *Arthritis Res. Ther.* 17, 235. doi: 10.1186/s13075-015-0759-2

Prather, A. A., Hall, M., Fury, J. M., Ross, D. C., Muldoon, M. F., Cohen, S., et al. (2012). Sleep and antibody response to hepatitis B vaccination. *Sleep* 35, 1063–1069. doi: 10.5665/sleep.1990

Punke, M. A., and Friederich, P. (2007). Amitriptyline is a potent blocker of human Kv1.1 and Kv7.2/7.3 channels. Anesth. Analg. 104, 1256–1264. doi: 10.1213/01.ane.0000260310.63117.a2

Ringgold, K. M., Barf, R. P., George, A., Sutton, B. C., and Opp, M. R. (2013). Prolonged sleep fragmentation of mice exacerbates febrile responses to lipopolysaccharide. *J. Neurosci. Methods* 219, 104–112. doi: 10.1016/j.jneumeth.2013.07.008

Roehrs, T., Hyde, M., Blaisdell, B., Greenwald, M., and Roth, T. (2006). Sleep loss and REM sleep loss are hyperalgesic. *Sleep* 29, 145–151. doi: 10.1093/sleep/29.2.145

Roitman, M. F., Wheeler, R. A., and Carelli, R. M. (2005). Nucleus accumbens neurons are innately tuned for rewarding and aversive taste stimuli, encode their predictors, and are linked to motor output. *Neuron* 45, 587–597. doi: 10.1016/j.neuron.2004.12.055

Rosseland, R., Pallesen, S., Nordhus, I. H., Matre, D., and Blagestad, T. (2018). Effects of sleep fragmentation and induced mood on pain tolerance and pain sensitivity in young healthy adults. *Front. Psychol.* 9, 2089. doi: 10.3389/fpsyg.2018.02089

Samad, T. A., Moore, K. A., Sapirstein, A., Billet, S., Allchorne, A., Poole, S., et al. (2001). Interleukin-1beta-mediated induction of Cox-2 in the CNS contributes to inflammatory pain hypersensitivity. *Nature* 410, 471–475. doi: 10.1038/35068566

Sardi, N. F., Lazzarim, M. K., Guilhen, V. A., Marcilio, R. S., Natume, P. S., Watanabe, T. C., et al. (2018a). Chronic sleep restriction increases pain sensitivity over time in a periaqueductal gray and nucleus accumbens dependent manner. *Neuropharmacology* 139, 52–60. doi: 10.1016/j.neuropharm.2018.06.022

Sardi, N. F., Tobaldini, G., Morais, R. N., and Fischer, L. (2018b). Nucleus accumbens mediates the pronociceptive effect of sleep deprivation: the role of adenosine A2A and dopamine D2 receptors. *Pain* 159, 75–84. doi: 10.1097/j.pain.00000000001066

Scherer, M. R., Claro, P. J., and Heaton, K. J. (2013). Sleep deprivation has no effect on dynamic visual acuity in military service members who are healthy. *Phys. Ther.* 93, 1185–1196. doi: 10.2522/ptj.20120144

Schrimpf, M., Liegl, G., Boeckle, M., Leitner, A., Geisler, P., and Pieh, C. (2015). The effect of sleep deprivation on pain perception in healthy subjects: a meta-analysis. *Sleep Med.* 16, 1313–1320. doi: 10.1016/j.sleep.2015.07.022

Schuh-Hofer, S., Wodarski, R., Pfau, D. B., Caspani, O., Magerl, W., Kennedy, J. D., et al. (2013). One night of total sleep deprivation promotes a state of generalized hyperalgesia: a surrogate pain model to study the relationship of insomnia and pain. *Pain* 154, 1613–1621. doi: 10.1016/j.pain.2013. 04.046

Segond von Banchet, G., Boettger, M. K., Konig, C., Iwakura, Y., Brauer, R., and Schaible, H. G. (2013). Neuronal IL-17 receptor upregulates TRPV4 but not TRPV1 receptors in DRG neurons and mediates mechanical but not thermal hyperalgesia. *Mol. Cell. Neurosci.* 52, 152–160. doi: 10.1016/j.mcn.2012.11.006

Seminowicz, D. A., Remeniuk, B., Krimmel, S. R., Smith, M. T., Barrett, F. S., Wulff, A. B., et al. (2019). Pain-related nucleus accumbens function: modulation by reward and sleep disruption. *Pain* 160, 1196–1207. doi: 10.1097/j.pain.00000000001498

Simon, E. B., Oren, N., Sharon, H., Kirschner, A., Goldway, N., Okon-Singer, H., et al. (2015). Losing neutrality: the neural basis of impaired emotional control without sleep. *J. Neurosci.* 35, 13194–13205. doi: 10.1523/JNEUROSCI.1314-15.2015

Skinner, G. O., Damasceno, F., Gomes, A., and De Almeida, O. M. (2011). Increased pain perception and attenuated opioid antinociception in paradoxical sleep-deprived rats are associated with reduced tyrosine hydroxylase staining in the periaqueductal gray matter and are reversed by L-dopa. *Pharmacol. Biochem. Behav.* 99, 94–99. doi: 10.1016/j.pbb.2011.04.009

Smith, M., Remeniuk, B., Finan, P., Speed, T., Tompkins, D., Robinson, M., et al. (2019). Sex differences in measures of central sensitization and pain sensitivity to experimental sleep disruption: implications for sex differences in chronic pain. *Sleep* 42, zsy209. doi: 10.1093/sleep/zsy209

Smith, M. T., Edwards, R. R., Mccann, U. D., and Haythornthwaite, J. A. (2007). The effects of sleep deprivation on pain inhibition and spontaneous pain in women. *Sleep* 30, 494–505. doi: 10.1093/sleep/30.4.494

Smith, M. T., and Haythornthwaite, J. A. (2004). How do sleep disturbance and chronic pain inter-relate? Insights from the longitudinal and cognitive-behavioral clinical trials literature. *Sleep Med. Rev.* 8, 119–132. doi: 10.1016/S1087-0792(03)00044-3

Spiegel, K., Leproult, R., and Van Cauter, E. (1999). Impact of sleep debt on metabolic and endocrine function. *Lancet* 354, 1435–1439. doi: 10.1016/S0140-6736(99)01376-8

Staffe, A. T., Bech, M. W., Clemmensen, S. L. K., Nielsen, H. T., Larsen, D. B., and Petersen, K. K. (2019). Total sleep deprivation increases pain sensitivity, impairs conditioned pain modulation and facilitates temporal summation of pain in healthy participants. *PLoS ONE* 14, e0225849. doi: 10.1371/journal.pone.0225849

Steinmiller, C. L., Roehrs, T. A., Harris, E., Hyde, M., Greenwald, M. K., and Roth, T. (2010). Differential effect of codeine on thermal nociceptive sensitivity in sleepy versus nonsleepy healthy subjects. *Exp. Clin. Psychopharmacol.* 18, 277. doi: 10.1037/a0018899

Stemkowski, P. L., Noh, M. C., Chen, Y., and Smith, P. A. (2015). Increased excitability of medium-sized dorsal root ganglion neurons by prolonged interleukin-1beta exposure is K(+) channel dependent and reversible. *J. Physiol.* 593, 3739–3755. doi: 10.1113/JP270905

Sutton, B. C., and Opp, M. R. (2014). Sleep fragmentation exacerbates mechanical hypersensitivity and alters subsequent sleep-wake behavior in a mouse model of musculoskeletal sensitization. *Sleep* 37, 515–524. doi: 10.5665/sleep.3488

Tasali, E., Leproult, R., Ehrmann, D. A., and Van Cauter, E. (2008). Slowwave sleep and the risk of type 2 diabetes in humans. *Proc. Nat. Acad. Sci.* 105, 1044–1049. doi: 10.1073/pnas.0706446105

Teng, B. C., Song, Y., Zhang, F., Ma, T. Y., Qi, J. L., Zhang, H. L., et al. (2016). Activation of neuronal Kv7/KCNQ/M-channels by the opener QO58-lysine and its anti-nociceptive effects on inflammatory pain in rodents. *Acta Pharmacol. Sin.* 37, 1054–1062. doi: 10.1038/aps.2016.33

Tiede, W., Magerl, W., Baumgartner, U., Durrer, B., Ehlert, U., and Treede, R. D. (2010). Sleep restriction attenuates amplitudes and attentional modulation of pain-related evoked potentials, but augments pain ratings in healthy volunteers. *Pain* 148, 36–42. doi: 10.1016/j.pain.2009.08.029

Tobaldini, E., Pecis, M., and Montano, N. (2014). Effects of acute and chronic sleep deprivation on cardiovascular regulation. *Arch. Ital. Biol.* 152, 103–110.

Tobler, I., Deboer, T., and Fischer, M. (1997). Sleep and sleep regulation in normal and prion protein-deficient mice. *J. Neurosci.* 17, 1869–1879. doi: 10.1523/JNEUROSCI.17-05-01869.1997

Todd, A. J. (2010). Neuronal circuitry for pain processing in the dorsal horn. *Nat. Rev. Neurosci.* 11, 823–836. doi: 10.1038/nrn2947

Tomim, D. H., Pontarolla, F. M., Bertolini, J. F., Arase, M., Tobaldini, G., Lima, M. M. S., et al. (2016). The pronociceptive effect of paradoxical sleep deprivation in rats: evidence for a role of descending pain modulation mechanisms. *Mol. Neurobiol.* 53, 1706–1717. doi: 10.1007/s12035-014-9059-0

Tracey, I., Woolf, C. J., and Andrews, N. A. (2019). Composite pain biomarker signatures for objective assessment and effective treatment. *Neuron* 101, 783–800. doi: 10.1016/j.neuron.2019.02.019

Ukponmwan, O. E., Rupreht, J., and Dzoljic, M. R. (1984). REM sleep deprivation decreases the antinociceptive property of enkephalinase-inhibition, morphine and cold-water-swim. *Gen. Pharmacol.* 15, 255–258. doi: 10.1016/0306-3623(84)90170-8

Valjent, E., Bertran-Gonzalez, J., Herve, D., Fisone, G., and Girault, J. A. (2009). Looking BAC at striatal signaling: cell-specific analysis in new transgenic mice. *Trends Neurosci.* 32, 538–547. doi: 10.1016/j.tins.2009.06.005

Van Cauter, E., Spiegel, K., Tasali, E., and Leproult, R. (2008). Metabolic consequences of sleep and sleep loss. *Sleep Med.* 9, S23–S28. doi: 10.1016/S1389-9457(08)70013-3

Van Dongen, H. P., Maislin, G., Mullington, J. M., and Dinges, D. F. (2003). The cumulative cost of additional wakefulness: dose-response effects on neurobehavioral functions and sleep physiology from chronic sleep restriction and total sleep deprivation. *Sleep* 26, 117–126. doi: 10.1093/sleep/26.2.117

Van Leeuwen, W. M., Lehto, M., Karisola, P., Lindholm, H., Luukkonen, R., Sallinen, M., et al. (2009). Sleep restriction increases the risk of developing cardiovascular diseases by augmenting proinflammatory responses through IL-17 and CRP. *PLoS ONE* 4, e4589. doi: 10.1371/journal.pone.0004589

Vanini, G. (2016). Sleep deprivation and recovery sleep prior to a noxious inflammatory insult influence characteristics and duration of pain. *Sleep* 39, 133-142. doi: 10.5665/sleep.5334

Vanini, G., Nemanis, K., Baghdoyan, H. A., and Lydic, R. (2014). GABAergic transmission in rat pontine reticular formation regulates the induction phase of anesthesia and modulates hyperalgesia caused by sleep deprivation. *Eur. J. Neurosci.* 40, 2264–2273. doi: 10.1111/ejn.12571

Villanueva, L., Bernard, J. F., and Le Bars, D. (1995). Distribution of spinal cord projections from the medullary subnucleus reticularis dorsalis and the adjacent cuneate nucleus: a *Phaseolus vulgaris*-leucoagglutinin study in the rat. *J. Comp. Neurol.* 352, 11–32. doi: 10.1002/cne.903520103

Villanueva, L., and Le Bars, D. (1995). The activation of bulbo-spinal controls by peripheral nociceptive inputs: diffuse noxious inhibitory controls. *Biol. Res.* 28, 113–125.

Volkow, N. D., Tomasi, D., Wang, G. J., Telang, F., Fowler, J. S., Logan, J., et al. (2012). Evidence that sleep deprivation downregulates dopamine D2R in ventral striatum in the human brain. *J. Neurosci.* 32, 6711–6717. doi: 10.1523/JNEUROSCI.0045-12.2012

Volkow, N. D., Wang, G. J., Telang, F., Fowler, J. S., Logan, J., Wong, C., et al. (2008). Sleep deprivation decreases binding of [11C]raclopride to dopamine D2/D3 receptors in the human brain. *J. Neurosci.* 28, 8454–8461. doi: 10.1523/JNEUROSCI.1443-08.2008

Walker, M. P. (2009). The role of sleep in cognition and emotion. Ann. N. Y. Acad. Sci. 1156, 168–197. doi: 10.1111/j.1749-6632.2009.04416.x

Wang, P. K., Cao, J., Wang, H., Liang, L., Zhang, J., Lutz, B. M., et al. (2015). Short-term sleep disturbance-induced stress does not affect basal pain perception, but does delay postsurgical pain recovery. *J. Pain* 16, 1186–1199. doi: 10.1016/j.jpain.2015.07.006pag

Wei, H., Hao, B., Huang, J. L., Ma, A. N., Li, X. Y., Wang, Y. X., et al. (2010). Intrathecal administration of a gap junction decoupler, an inhibitor of Na(+)-K(+)-2Cl(-) cotransporter 1, or a GABA(A) receptor agonist attenuates mechanical pain hypersensitivity induced by REM sleep deprivation in the rat. *Pharmacol. Biochem. Behav.* 97, 377–383. doi: 10.1016/j.pbb.2010. 09.007

Wei, H., Ma, A., Wang, Y. X., and Pertovaara, A. (2008). Role of spinal 5-HT receptors in cutaneous hypersensitivity induced by REM sleep deprivation. *Pharmacol. Res.* 57, 469–475. doi: 10.1016/j.phrs.2008. 05.007

Wei, H., Zhao, W., Wang, Y. X., and Pertovaara, A. (2007). Pain-related behavior following REM sleep deprivation in the rat: influence of peripheral nerve injury, spinal glutamatergic receptors and nitric oxide. *Brain Res.* 1148, 105–112. doi: 10.1016/j.brainres.2007.02.040

Wodarski, R., Schuh-Hofer, S., Yurek, D., Wafford, K., Gilmour, G., Treede, R. D., et al. (2015). Development and pharmacological characterization of a model of sleep disruption-induced hypersensitivity in the rat. *Eur. J. Pain* 19, 554–566. doi: 10.1002/ejp.580

Yang, H., De Jong, J. W., Cerniauskas, I., Peck, J. R., Lim, B. K., Gong, H., et al. (2021). Pain modulates dopamine neurons via a spinal-parabrachial-mesencephalic circuit. *Nat. Neurosci.* 24, 1402–1413. doi: 10.1038/s41593-021-00903-8

Yarnitsky, D. (2010). Conditioned pain modulation (the diffuse noxious inhibitory control-like effect): its relevance for acute and chronic pain states. *Curr. Opin. Anaesthesiol.* 23, 611–615. doi: 10.1097/ACO.0b013e32833c348b

Yarnitsky, D., Bouhassira, D., Drewes, A. M., Fillingim, R. B., Granot, M., Hansson, P., et al. (2015). Recommendations on practice of conditioned pain modulation (CPM) testing. *Eur. J. Pain* 19, 805–806. doi: 10.1002/ejp.605

Yehuda, S., Sredni, B., Carasso, R. L., and Kenigsbuch-Sredni, D. (2009). REM sleep deprivation in rats results in inflammation and interleukin-17 elevation. *J. Interferon Cytokine Res.* 29, 393–398. doi: 10.1089/jir.2008.0080

Zielinski, M. R., and Krueger, J. M. (2011). Sleep and innate immunity. Front. Biosci. 3, 632-642. doi: 10.2741/s176