



Efficacy of Disease Modifying Therapies in Progressive MS and How Immune Senescence May Explain Their Failure

Navid Manouchehri¹, Victor H. Salinas¹, Negar Rabi Yeganeh², David Pitt³,
Rehana Z. Hussain¹ and Olaf Stuve^{1,4*}

¹ Department of Neurology, The University of Texas Southwestern Medical Center, Dallas, TX, United States, ² Department of Radiopharmacy, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran, ³ Department of Neurology, Yale University, New Haven, CT, United States, ⁴ Neurology Section, VA North Texas Health Care System, Medical Service Dallas, Veterans Affairs Medical Center, Dallas, TX, United States

OPEN ACCESS

Edited by:

Aurora Zanghi,
Sant'Elia Hospital, Italy

Reviewed by:

Vincent Van Pesch,
Catholic University of
Louvain, Belgium

*Correspondence:

Olaf Stuve
olaf.stuve@utsouthwestern.edu

Specialty section:

This article was submitted to
Multiple Sclerosis and
Neuroimmunology,
a section of the journal
Frontiers in Neurology

Received: 13 January 2022

Accepted: 18 February 2022

Published: 31 March 2022

Citation:

Manouchehri N, Salinas VH, Rabi Yeganeh N, Pitt D, Hussain RZ and Stuve O (2022) Efficacy of Disease Modifying Therapies in Progressive MS and How Immune Senescence May Explain Their Failure. *Front. Neurol.* 13:854390. doi: 10.3389/fneur.2022.854390

The advent of disease modifying therapies (DMT) in the past two decades has been the cornerstone of successful clinical management of multiple sclerosis (MS). Despite the great strides made in reducing the relapse frequency and occurrence of new signal changes on neuroimaging in patients with relapsing remitting MS (RRMS) by approved DMT, it has been challenging to demonstrate their effectiveness in non-active secondary progressive MS (SPMS) and primary progressive MS (PPMS) disease phenotypes. The dichotomy of DMT effectiveness between RRMS and progressive MS informs on distinct pathogeneses of the different MS phenotypes. Conversely, factors that render patients with progressive MS resistant to therapy are not understood. Thus far, age has emerged as the main correlate of the transition from RRMS to SPMS. Whether it is aging and age-related factors or the underlying immune senescence that qualitatively alter immune responses as the disease transitions to SPMS, that diminish DMT effectiveness, or both, is currently not known. Here, we will discuss the role of immune senescence on different arms of the immune system, and how it may explain relative DMT resistance.

Keywords: adaptive immunity, innate immunity, multiple sclerosis, immunosenescence, progressive multiple sclerosis, disease modifying therapies

INTRODUCTION

Multiple sclerosis (MS) is the most prevalent inflammatory disorder of the central nervous system (CNS) with a presumed autoimmune pathogenesis. MS was traditionally viewed as a T cell-mediated inflammatory disorder based on numerous observations made over the span of many decades. Aside from the abundance of lymphocytic infiltrates in MS lesion biopsies, other factors included: (A) the induction of the experimental autoimmune encephalomyelitis (EAE) model of MS in healthy recipient animals by adoptive transfer of myelin-reactive CD4⁺ T helper (Th) cells from previously immunized donor mice (1); (B) the genetic association of MS with human leukocyte antigen (HLA) DRB1*15:01 (2), a major histocompatibility complex (MHC) class II molecule, required for the presentation of linearized peptides to CD4⁺ Th cells; (C) the failed attempt to treat MS patients with an altered peptide ligand of myelin basic protein (MBP) that activated MBP-reactive CD4⁺ Th cells, leading to disease exacerbation instead (3); (D) the initiation

and re-activation of MS with immune checkpoint inhibitors during cancer therapy (4); and (E) the beneficial effects of pharmacotherapies in early relapsing MS that deplete T cells, or sequester them out of the CNS (5, 6). This last aspect has illustrated how in early MS, relapses and new MS brain lesions are triggered and perpetuated by T cells and possibly other bone marrow-derived immune cells (7). The success of B cell depleting therapies in treating active MS, further corroborates the role of bone marrow-derived immune cells outside of the T cell compartment in pathogenesis of the disease (8–10). Changes in the clinical phenotype of MS, including treatment responsiveness will likely be linked to these cells as well.

A clinical course typified by relapses followed by periods of remission defines relapsing-remitting MS (RRMS) (11, 12). Patients with early MS who display clinical and paraclinical magnetic resonance imaging (MRI) disease activity gain a detectable and substantial benefit from receiving disease modifying therapies (DMT); patients without these evidences of disease activity, are defined as progressive MS (PMS); specifically, based on the 2013 Lublin criteria, PMS patients accrue objectively documented neurological disability without intermittent recovery and do not appear to receive any benefit from DMT (12, 13). Thus, the molecular and cellular signature of MS, as the primary therapeutic targets, change with age and disease becomes non-active. PMS at this stage is considered either secondary MS (SPMS) when following a period of RRMS, or primary progressive MS (PPMS) in lieu of relapsing disease activity (11, 12). PPMS patients are ~10 years older upon diagnosis than RRMS patients. A subsection of patients with PMS, do show disease activity as defined above (12). There is no disease biomarker to indicate when the transition from RRMS to SPMS starts or is completed.

Currently, different hypotheses try to explain this transition; to date, age has been the most relevant prognostic factor underlying the transition from active RRMS to non-active SPMS (14–17). In contrast, a meta-analysis of all blinded, randomized clinical trials of DMT for RRMS indicated that DMT efficacy were independent of the recipients' age (18) despite a clear trend toward reduced effectiveness. Unfortunately, individualized data was not made available to the authors, and these results have to be interpreted with some caution.

As a biological correlate to age, immunosenescence has been advocated as a candidate to explain diminished DMT efficacy in PMS (19). Immunosenescence correlates with age relative to overall life expectancy (20). It is often accompanied by a decline in key immune functions such as the capacity for strictly non-self-antigen presentation and breadth of antigen recognition, the formation of long-lasting immune memory, and active immune surveillance (20). Here, we discuss whether immunosenescence contributes to the transition from active to non-active MS and how that correlates to loss of DMT efficacy in the context of PMS.

AGE, IMMUNOSENESCENCE, AND LOSS OF DMT EFFICACY

The transition to SPMS may not be entirely influenced by age. It takes place on average two decades following the

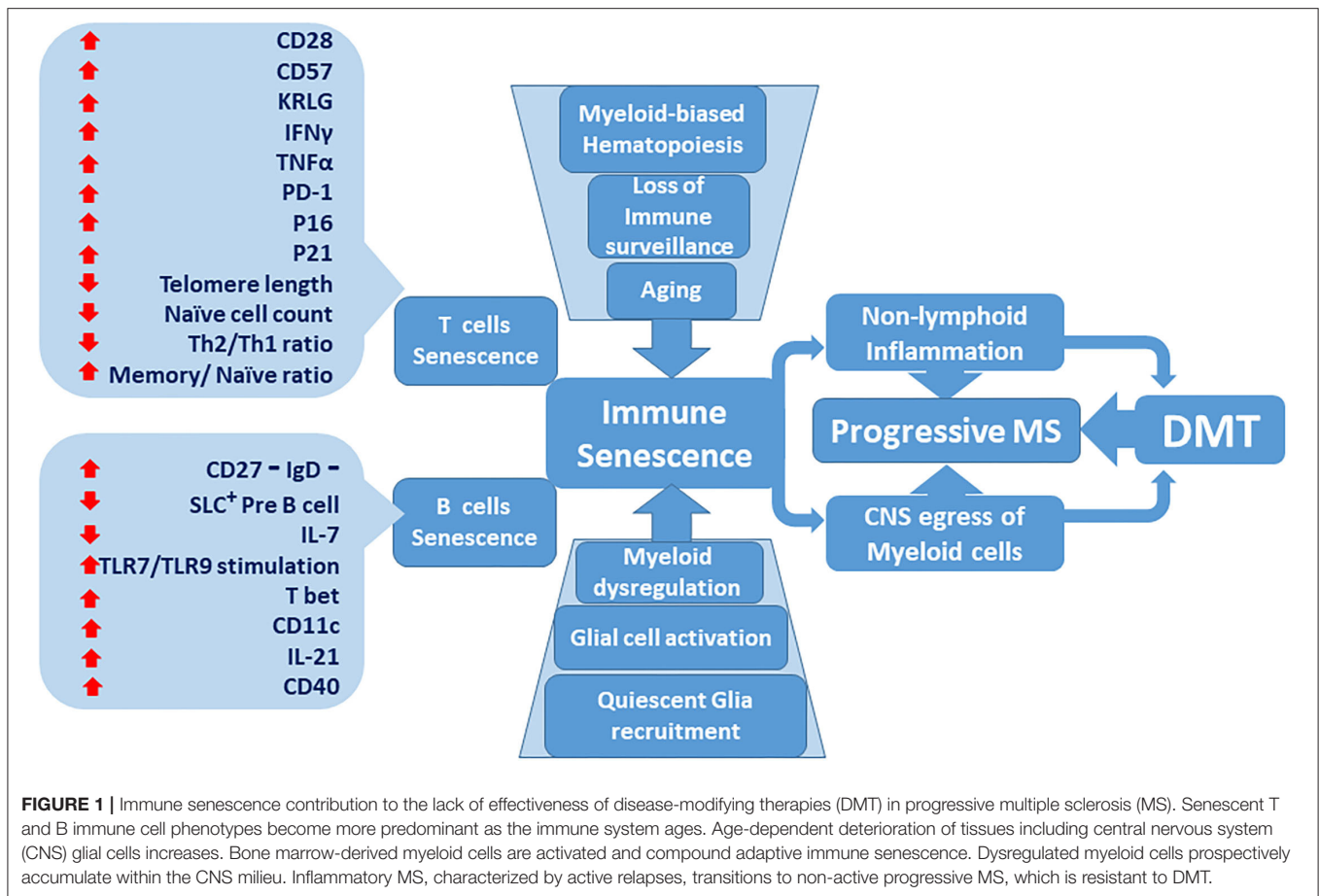
clinical diagnosis of RRMS in adult-onset MS (AOMS) (21). Given that currently approved DMT remain effective even in patients with late-onset MS (LOMS), diagnosed in 50-year-old patients or older, it appears counterintuitive to attribute DMT unresponsiveness to age alone. Although, PPMS is more prevalent among LOMS than AOMS, still, nearly 50% of LOMS cases are RRMS and respond to DMT (18, 22). Immunosenescence as a potential contributor to DMT-resistance may be present in both AOMS and LOMS, and not entirely driven by age.

A correlate of intact adaptive immune function is responsiveness to vaccination with neo-antigens. Expectedly, vaccine efficacy wanes in elder populations (23); however, data on vaccine response among elderly RRMS demonstrate substantial adaptive immune response, despite long term DMT treatment with proven negative effects on vaccine response (24), reiterating how age alone does not define quality of immune response.

Current DMT in MS, including interferons (25–27), copolymers (28) depleting agents against CD20 (8–10) and CD52 (29), nucleoside synthesis antagonists (30–32), sphingosine-1 phosphate receptor modulators (33–36), nuclear 1 factor (erythroid-derived 2)-like 2 modulators (37) or α 4-integrin antagonists (38, 39), aim to either deplete lymphocytes, modulate pro-inflammatory features or inhibit their traffic into the CNS. These DMT classes are approved for use in active MS. The validating trials for the only two FDA-approved DMT for use in SPMS, namely, cladribine (40, 41) and siponimod (34) recruited a mix of active and non-active progressive MS patients, limiting their relevance in pure non-active MS cohorts (13). These trials likely benefited participants with residual active MS (42). Conceivably, immunosenescence, both predates and promotes the transition from DMT-responsive active MS to DMT-resistant non-active MS. However, instigators or accelerators of immunosenescence in the context of MS require further elucidation.

ADAPTIVE IMMUNOSENESCENCE AND DMT-RESISTANT PMS

The adaptive immune system is not fully competent at birth; it becomes fully functional post-puberty and in early adulthood, declining progressively thereafter (43–46). Despite the age-associated decline in thymic epithelial tissue, it has been demonstrated that both the thymic cortex and thymic medulla function throughout life (47–49); however, the inevitable thymic involution is accompanied by the reduction of T cell diversity (49). Intact and functional thymic epithelium continually produces T cells migrating out of thymic medulla to peripheral lymphoid organs (50, 51). T cells generated from thymopoiesis have a full T cell receptor (TCR) repertoire, and are capable of generating responses to neo-antigens. In contrast, expansion of the peripheral T cells, driven by thymic involution may lead to repertoires limited to those of existing memory T cells and reduced capacity of immune response to new antigens (52–55). As mentioned, T cells are critical in initiating and perpetuating inflammation in active MS (7). MS pathogenesis potentiates T cell-antigenic spreading and repeatedly stimulates CNS-specific



T cells (56). Suppressed CD28 expression, mediated by repeated antigenic stimulation is associated with senescent phenotypes in T cells (**Figure 1**) (23, 57–61). A similar phenotype of CD28^{low} T cells is detectable in the pool of circulatory effector memory T cells with senescent attributes in the context of MS. (62, 63). Immunosenescence is not restricted to cellular immunity and similarly carries over to B cell-mediated immune responses (61). Noticeably, non-cellular adaptive immune responses mediated by B cells have been implicated by preclinical models as mediators of CNS autoimmunity (64). The Epstein-Barr virus (EBV), a plausible pathogenic MS trigger, is in fact a B cell tropic infection (65). EBV infection primes polyclonal populations of B cells to avidly present self-antigens to autoreactive CD8⁺ cytotoxic T cells (66). Post-mortem studies that have yet to be reproduced showed abundant EBV infected B cells within actively demyelinating MS lesions (66, 67). Similar to T cells, constant B cell activation may drive premature senescence (68). Interestingly, surface expression of CD40, a correlate of B cells antigen presentation and memory formation, signifies senescence and is elevated in EBV infected B cells (69). In B cell senescence, it is quality rather than quantity of humoral response that declines, resulting in comparable volume of antibodies, albeit less effective. This is evident from diminished antibody specificity for foreign antigens, and decreased predominance of IgG isotypes along with

lowered affinity of antibodies (70). Prematurely senescent B cells provide antigen presentation for expansion and maintenance of T cells in autoimmune disease like MS. Namely, increased signaling via CD80, CD86, CD11c, and CD40 by B cells in MS patients is higher than healthy controls and responsible for promotion of inflammatory T cell responses. Ultimately, the expression of these markers correlates with exhausted or senescent B cell phenotypes (**Figure 1**) (23, 57–61, 71). As discussed earlier, anti-CD20 agents that deplete B cells have become a mainstay of therapy for active MS with proven efficacy (72). However, B cell depletion with the humanized anti-CD20 monoclonal antibody (72) ocrelizumab, which is approved for PPMS, does not deliver significant neuroprotective effects as assessed by serial blood measurements of neurofilament light chain (NFL) (73). CD20⁺ cells are not a single population and range from naïve B cells to fully matured memory cells. Since production of new pro-B cells is directly affected by senescence (74), continued treatment with anti-CD20 therapies likely depletes naïve B cells with diverse B cell receptor (BCR) repertoires. Likewise, continuation of T cell-depleting DMT in non-active PMS will likely result in T cells that lack diverse naïve cells. In this manner, increasing age and DMT therapy compound adaptive immunosenescence in MS. Nevertheless, if adaptive immunity was relevant in perpetuating disease progression in

PMS, DMT effectiveness should have remained constant or even slightly improved since PMS is dominated by senescence. In fact, if DMT unresponsiveness in PMS was driven predominantly by adaptive immunity, decreased potency of senescent adaptive immunity would have led to improvement of clinical outcomes in elder patients. Since DMT optimally address adaptive immune cells in the periphery, DMT resistance in non-active PMS requires explanations beyond adaptive immune components or peripheral compartment.

INNATE IMMUNOSENESCENCE AND DMT RESISTANCE IN PMS

Innate immunity is influenced by the immunosenescence. Contrary to adaptive immunity, the innate immune system in mammals is considered functional at birth, and retains most of its function throughout life (61). Cell migration, adhesion and phagocytosis of polymorphonuclear leukocytes (PMN) were believed to stay virtually unaltered by aging (75–78); however studies have shown that certain innate immune cell functions may falter with age (79–81). Immunosenescence pushes innate immune cells toward functional dysregulation that compounds the effects of suboptimal senescent adaptive immunity. In MS, effects of immunosenescence on innate immune are noticeable in four domains; namely, (1) Aged innate immune system manifests with the preponderance of dys-homeostatic phenotypes and *forme pleine* of the ideally self-limiting responses. This is likely to promote chronic and continued tissue destruction, sub-functional remodeling and delayed healing (81–84). (2) Within the CNS, myeloid cells, as antigen presenting cells (APC), re-activate and retain CD4⁺ T cells, and contribute to effective immune surveillance (85). These cells are altered via senescence. There are three compartments within the brain where myeloid cells exert their effect: (A) the parenchyma, (B) cerebral perivascular spaces (CPVS) and (C) meninges. Parenchymal microglia are tissue-intrinsic macrophages of the CNS (86–88). The other relevant compartments for antigen presentation are CPVS and meninges, which are populated by monocyte-derived macrophages and dendritic cells (DC). In CX₃CR₁ GFP⁺ mice, it was demonstrated that microglia and monocyte-derived brain macrophages are distinct entities (89, 90). Meninges have been implicated as an anatomic site in host defense and autoimmunity. Major histocompatibility complex II (MHCII)-positive cells, detectable within all meningeal layers (91), include monocyte-derived macrophages, monocyte-derived DC (mDC) and classical DC (cDC) (92). While some studies reported DC from young and aged humans having similar surface expression of MHCII molecules and elicited equal T cell proliferation, other investigators demonstrated significantly lower MHCII expression by DC in the elderly (93, 94). (3) During hematopoiesis, immunosenescence reduces lymphopoiesis in favor of heightened myelopoiesis, resulting in a net increase in myeloid cell output (59). (4) Inflammaging, defined as chronic, low grade and sterile inflammation, despite the overall diminishment in immune functions, increases with age (81, 95). Bone marrow-derived myeloid cells (BMC), activate

and upregulate surface adhesion molecules in response to inflammation (96). This may allow BMC to penetrate and accrue within target tissues such as CNS (97).

CNS microenvironment in response to the aforementioned events may shift to a dys-homeostatic state adopted by CNS-resident myeloid cells (97). Previous observations postulated that compared to naïve quiescent microglia, activated microglia and infiltrated BMC during CNS inflammation upregulate inflammation-associated signals (92). Myeloid cells within the CPVS are constantly replaced by BMC, and this turnover is accelerated during inflammation (98). During EAE, parenchymal microglia and BMC exhibited mutual activation markers following the onset of clinical disease in mice. Specifically, clinical disease onset was temporally associated with the appearance of BMC in the CNS inflammatory milieu (97). This was not a transient event and the newly present activated BMC were likely retained within the CNS microenvironment and merged with the activated microglial pool (97). It is still unclear to what extent these changes advance functional disarray and homeostatic disturbance; however, current DMT in MS do not primarily target BMC. Certain DMT might modulate the trafficking of BMC into the CNS, as demonstrated with natalizumab therapy (99). However, the ASCEND trial, a phase 3 study on the efficacy of natalizumab therapy in SPMS, failed to show meaningful clinical benefits (100), perhaps suggesting that ongoing migration of BMC in SPMS is no longer highly relevant to disease progression.

“Smoldering” MS lesions are a candidate to explain PMS and chronic destruction of CNS parenchyma without evident inflammation. They are dominated by the presence of perilesional activated myeloid cells without lymphoid inflammation (101, 102). Whether these myeloid cells are CNS intrinsic or bone marrow-derived, or a mixture of both is incompletely understood. Smoldering MS is more prevalent among SPMS patients who are on average older than active MS patients, however, as we discussed before, age likely is not the driving factor. The transition to SPMS takes place faster in LOMS patients; however, accrual of disability for LOMS patients during years living with SPMS is slower in comparison (21). Possibly, later onset and relatively brief inflammatory phase in LOMS, spares them from fully-recognized disease burden. In the context of non-active PMS, antigen-independent and compartmentalized chronic inflammation, shielded from therapeutic efforts, is plausible. The corollary to this hypothesis would be that even outside of common DMT, other anti-proliferative therapies might fail to provide meaningful clinical benefit to PMS (103), a logic that should guide hematopoietic stem cell transplants as well (104). Furthermore, if myeloid cells drive PMS, they probably do so in a stage-specific fashion; (1) BMC likely enter the CNS throughout active and non-active MS and are retained within the CNS; and (2) altered CNS-intrinsic myeloid cells, respond to BMC presence, at sites similar to the border of smoldering lesions. Adaptive immune cells most likely instigate these events at first; during non-active MS, such external cues may no longer be absolutely required and suppression of adaptive immune system *via* DMT, thereafter provide little clinical benefit.

GLIAL SENESENCE AND DMT RESISTANCE IN PMS

Glial cells including astrocytes and microglia are critical in CNS. Immune functions related to astrocytes in the context of MS and its preclinical models have recently attracted renewed interest; however their pathogenic role in MS is currently less clear. For instance, the depletion of astrocytes worsens clinical disease in an acute EAE model but ameliorates progressive EAE (105). A subpopulation of astrocytes expresses complement component 3 (C3) in response to interleukin-1 alpha (IL-1 α) tumor necrosis factor alpha (TNF α) and complement component 1, subcomponent q (C1q), and possesses neurotoxic properties and are upregulated in MS lesions (106). Their development underscores a bi-directional interaction between astrocytes and myeloid cells. Pro-inflammatory signals from Microglia and BMC in CNS are a primary inducer of such astrocytes while activated astrocytes allow further recruitment and entry of pro-inflammatory monocytes to the CNS; this multiplicative effect may have been intercepted by astrocyte depletion leading to the aforementioned amelioration of chronic EAE (106, 107). Furthermore, senescent astrocytes, may contribute to neurodegeneration in an overburdened neural network post demyelination. Astrocytes possess star-shaped appearance, and are intimately associated with the CNS vasculature (108). Together with endothelial cells and pericytes, astrocytes are critical in forming and maintaining the blood brain barrier (BBB). Astrocytes can express MHC class II molecules in defined experimental conditions which endows them to serve as potential APC to CD4⁺ T helper cells (109–111). Cellular changes associated with astrocyte senescence include the increased expression of glial fibrillary acidic protein (GFAP) and vimentin (112, 113). This is at least partly driven by increased signaling of transforming growth factor beta 1 (TGF β 1). TGF β 1 inhibits astrocyte proliferation and induces a senescence-associated secretory phenotype (SASP), which involves an enhanced expression of inflammatory molecules (19, 114).

Inflammation induced phenotypes in microglia also mimic senescence. Specifically, CNS microglia exhibit a phenotypical profile that has been frequently associated with aging in the context of neurodegenerative disorders (115). These attributes, including increased iron storage, production of pro-inflammatory cytokines, lower motility and diminished phagocytic capacity are not strictly age-dependent and are inducible by other insults as well. Single cell transcriptomic studies on EAE models as well as human samples have shown presence of distinct microglia-like cells in CNS inflammation (97, 116). Microglia are maintained by local proliferative self-renewal (90). Within the CNS, functional properties of microglia, as tissue resident myeloid cells, across age groups are likely constant. Interestingly, it was recently shown that microglial density in the brain increases in aged mice (117). However, the authors had utilized markers that correlated with activated microglia, namely Iba-1. Therefore, the observed increase in microglia density with age might point more toward heightened microglial activation and subsequent reduction in the pool of homeostatic quiescent microglia. Senescent microglia

are found within the brains of PMS patients despite pronounced reduction in inactive lesions (118, 119). As discussed, these cells might exert their role at the border of smoldering lesions (102). Phagocytosis of myelin debris supports re-myelination efforts; microglial depletion associates with loss of phagocytic capacity in the microenvironment of MS lesions, likely promoting dysmyelination. Aged human microglia exhibit proclivity to express ferritin, believed to be associated with senescence (120). Increased iron uptake likely follows the destruction of iron-containing oligodendrocytes in MS and is observable in aged microglia. These limitations to CNS re-myelination efforts, possibly lend to PMS phenotype.

Glial cells are not a primary target for current DMT, and their role in relation to why DMT fail remains to be explained. The restricted CNS bioavailability of most DMT, is unlikely to impact glial cells. Further studies are required to elucidate the significance of these observations and their potential for development of novel therapeutics.

TOWARD A THEORY FOR DMT RESISTANCE IN PROGRESSIVE MS

Based on extensive data from clinical trials and post-approval observational studies it is evident that the current therapeutic dogma in MS, namely the depletion of inflammatory adaptive immune cells or their sequestration out of the CNS is effective during active RRMS. Current data gleaned from clinical trials suggest that most DMT have minimal effects on non-active progressive MS without signs of disease activity (13, 34, 73, 121, 122); therefore administration of DMT that target immune components may not be in the best interest of these patients. Alternatively future therapeutic endeavors may benefit from incorporating strategies that cover innate immunity and glial targets within the CNS.

The evidence presented here supports a view of DMT resistance associated with immunosenescence in PMS (**Figure 1**). Possible pharmacological efforts to address immunosenescence may adopt designs that identify senescence-specific factors, amenable to modulation. A desired goal and measure of success could be delayed transition to non-active PMS. Current DMT, as effective as they are in controlling RRMS, are unqualified to address the breadth of ongoing deleterious effects of MS (123–125), including the role of innate immune cells and CNS glial components. Myeloid cells, as potential candidates for targeted novel therapies in non-active PMS, retain phenotypical plasticity. Curtailing the dys-homeostatic signaling in these cells and safeguarding against further disruption of CNS quiescence, is a biological plausibility (126, 127). Given the current availability of analytic tools with single-cell resolution, deep characterization of myeloid sub populations and definition of exclusive phenotypical signatures in pertinent compartments is both feasible and indispensable. Molecular targets, acquired through this process could serve as a map for pre-clinical efforts (128, 129).

In conclusion, immunosenescence as a multivariate phenomenon, is not defined by advancing age alone and its

constellation of immunological effects over time culminates to DMT resistance in PMS. Current DMT mechanisms of action are optimized to mitigate inflammation-induced damage during active MS. They are limited to address the inevitable transition to non-active PMS or remain effective thereafter. Clinical and biological data call for a more targeted approach with an emphasis on myeloid cells, innate immunity components and glial cells in the future.

AUTHOR CONTRIBUTIONS

NM and NR surveyed the literature. NM, VS, and NR drafted the manuscript. RH, DP, and OS provided expert opinion and edited

the final manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

OS was a 2021 recipient of a Grant for Multiple Sclerosis Innovation (GMSI), Merck KGaA, and was funded by a Merit Review grant (federal award document number (FAIN) BX005664-01 from the United States (U.S.) Department of Veterans Affairs, Biomedical Laboratory Research and Development). The funders were not involved in the study design, collection, analysis, interpretation of data, the writing of this article or the decision to submit it for publication.

REFERENCES

- Paterson PY. Transfer of allergic encephalomyelitis in rats by means of lymph node cells. *J Exp Med.* (1960) 111:119–36. doi: 10.1084/jem.111.1.119
- Haines JL, Ter-Minassian M, Bazzyk A, Gusella JF, Kim DJ, Terwedow H, et al. A complete genomic screen for multiple sclerosis underscores a role for the major histocompatibility complex. The Multiple Sclerosis Genetics Group. *Nat Genet.* (1996) 13:469–71. doi: 10.1038/ng0896-469
- Martin R, Bielekova B, Gran B, McFarland HF. Lessons from studies of antigen-specific T cell responses in Multiple Sclerosis. *J Neural Transm Suppl.* (2000) 2000:361–73. doi: 10.1007/978-3-7091-6301-6_26
- Gerdes LA, Held K, Beltran E, Berking C, Prinz JC, Junker A, et al. CTLA4 as immunological checkpoint in the development of multiple sclerosis. *Ann Neurol.* (2016) 80:294–300. doi: 10.1002/ana.24715
- Zhang Y, Salter A, Cutter G, Stuve O. Clinical trials in multiple sclerosis: milestones. *Ther Adv Neurol Disord.* (2018) 11:1756286418785499. doi: 10.1177/1756286418785499
- Zhang Y, Salter A, Wallstrom E, Cutter G, Stuve O. Evolution of clinical trials in multiple sclerosis. *Ther Adv Neurol Disord.* (2019) 12:1756286419826547. doi: 10.1177/1756286419826547
- Milo R, Korczyn AD, Manouchehri N, Stuve O. The temporal and causal relationship between inflammation and neurodegeneration in multiple sclerosis. *Mult Scler.* (2020) 26:876–86. doi: 10.1177/1352458519886943
- Salzer J, Svenningsson R, Alping P, Novakova L, Bjorck A, Fink K, et al. Rituximab in multiple sclerosis: a retrospective observational study on safety and efficacy. *Neurology.* (2016) 87:2074–81. doi: 10.1212/WNL.0000000000003331
- Hauser SL, Bar-Or A, Comi G, Giovannoni G, Hartung HP, Hemmer B, et al. Ocrelizumab versus interferon beta-1a in relapsing multiple sclerosis. *N Engl J Med.* (2017) 376:221–34. doi: 10.1056/NEJMoa1601277
- Hauser SL, Bar-Or A, Cohen JA, Comi G, Correale J, Coyle PK, et al. Ofatumumab versus teriflunomide in multiple sclerosis. *N Engl J Med.* (2020) 383:546–57. doi: 10.1056/NEJMoa1917246
- Thompson AJ, Banwell BL, Barkhof F, Carroll WM, Coetzee T, Comi G, et al. Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. *Lancet Neurol.* (2018) 17:162–73. doi: 10.1016/S1474-4422(17)30470-2
- Lublin FD, Reingold SC, Cohen JA, Cutter GR, Sorensen PS, Thompson AJ, et al. Defining the clinical course of multiple sclerosis: the 2013 revisions. *Neurology.* (2014) 83:278–86. doi: 10.1212/WNL.0000000000000560
- Manouchehri N, Stuve O. Trials and therapies in secondary progressive MS, simplified. *Nat Rev Neurol.* (2019) 15:431–2. doi: 10.1038/s41582-019-0233-x
- Trapp BD, Nave KA. Multiple sclerosis: an immune or neurodegenerative disorder? *Annu Rev Neurosci.* (2008) 31:247–69. doi: 10.1146/annurev.neuro.30.051606.094313
- Scalfari A, Neuhaus A, Daumer M, Ebers GC, Muraro PA. Age and disability accumulation in multiple sclerosis. *Neurology.* (2011) 77:1246–52. doi: 10.1212/WNL.0b013e318230a17d
- Confavreux C, Vukusic S. The natural history of multiple sclerosis. *Rev Prat.* (2006) 56:1313–20. doi: 10.1016/B978-0-443-07271-0.50006-9
- Kremenchtzky M, Rice GP, Baskerville J, Wingerchuk DM, Ebers GC. The natural history of multiple sclerosis: a geographically based study 9: observations on the progressive phase of the disease. *Brain.* (2006) 129:584–94. doi: 10.1093/brain/awh721
- Zhang Y, Gonzalez Caldito N, Shirani A, Salter A, Cutter G, Culpepper W, et al. Aging and efficacy of disease-modifying therapies in multiple sclerosis: a meta-analysis of clinical trials. *Ther Adv Neurol Disord.* (2020) 13:1756286420969016. doi: 10.1177/1756286420969016
- Papadopoulos D, Magliozzi R, Mitsikostas DD, Gorgoulis VG, Nicholas RS. Aging, cellular senescence, and progressive multiple sclerosis. *Front Cell Neurosci.* (2020) 14:178. doi: 10.3389/fncel.2020.00178
- Ginaldi L, Loreto MF, Corsi MR, Modesti M, De Martinis M. Immunosenescence and infectious diseases. *Microbes Infect.* (2001) 3:851–7. doi: 10.1016/S1286-4579(01)01443-5
- Tremlett H, Yinshan Z, Devonshire V. Natural history of secondary-progressive multiple sclerosis. *Mult Scler.* (2008) 14:314–24. doi: 10.1177/1352458507084264
- Naseri A, Nasiri E, Sahraian MA, Daneshvar S, Talebi M. Clinical features of late-onset multiple sclerosis: a systematic review and meta-analysis. *Mult Scler Relat Disord.* (2021) 50:102816. doi: 10.1016/j.msard.2021.102816
- Goronzy JJ, Weyand CM. Understanding immunosenescence to improve responses to vaccines. *Nat Immunol.* (2013) 14:428–36. doi: 10.1038/ni.2588
- Bar-Or A, Calkwood JC, Chognot C, Evershed J, Fox EJ, Herman A, et al. Effect of ocrelizumab on vaccine responses in patients with multiple sclerosis: the VELOCE study. *Neurology.* (2020) 95:e1999–2008. doi: 10.1212/WNL.0000000000010380
- Paty DW, Li DK. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. II. MRI analysis results of a multicenter, randomized, double-blind, placebo-controlled trial. UBC MS/MRI Study Group and the IFNB Multiple Sclerosis Study Group. *Neurology.* (1993) 43:662–7. doi: 10.1212/WNL.43.4.662
- Jacobs LD, Cookfair DL, Rudick RA, Herndon RM, Richert JR, Salazar AM, et al. Intramuscular interferon beta-1a for disease progression in relapsing multiple sclerosis. The Multiple Sclerosis Collaborative Research Group (MSCRG). *Ann Neurol.* (1996) 39:285–94. doi: 10.1002/ana.410390304
- Calabresi PA, Kieseier BC, Arnold DL, Balcer LJ, Boyko A, Pelletier J, et al. Pegylated interferon beta-1a for relapsing-remitting multiple sclerosis (ADVANCE): a randomised, phase 3, double-blind study. *Lancet Neurol.* (2014) 13:657–65. doi: 10.1016/S1474-4422(14)70068-7
- Johnson KP, Brooks BR, Cohen JA, Ford CC, Goldstein J, Lisak RP, et al. Copolymer 1 reduces relapse rate and improves disability in relapsing-remitting multiple sclerosis: results of a phase III multicenter, double-blind placebo-controlled trial. The Copolymer 1 Multiple Sclerosis Study Group. *Neurology.* (1995) 45:1268–76. doi: 10.1212/WNL.45.7.1268
- Havrdova E, Arnold DL, Cohen JA, Hartung HP, Fox EJ, Giovannoni G, et al. Alemtuzumab CARE-MS I 5-year follow-up: durable efficacy in the absence of continuous MS therapy. *Neurology.* (2017) 89:1107–16. doi: 10.1212/WNL.0000000000004313

30. O'Connor P, Wolinsky JS, Confavreux C, Comi G, Kappos L, Olsson TP, et al. Randomized trial of oral teriflunomide for relapsing multiple sclerosis. *N Engl J Med.* (2011) 365:1293–303. doi: 10.1056/NEJMoa1014656
31. Giovannoni G, Comi G, Cook S, Rammohan K, Rieckmann P, Soelberg Sorensen P, et al. A placebo-controlled trial of oral cladribine for relapsing multiple sclerosis. *N Engl J Med.* (2010) 362:416–26. doi: 10.1056/NEJMoa0902533
32. Warnke C, Wiendl H, Hartung HP, Stuve O, Kieseier BC. Identification of targets and new developments in the treatment of multiple sclerosis—focus on cladribine. *Drug Des Devel Ther.* (2010) 4:117–26. doi: 10.2147/DDDT.S6627
33. Calabresi PA, Radue EW, Goodin D, Jeffery D, Rammohan KW, Reder AT, et al. Safety and efficacy of fingolimod in patients with relapsing-remitting multiple sclerosis (FREEDOMS II): a double-blind, randomised, placebo-controlled, phase 3 trial. *Lancet Neurol.* (2014) 13:545–56. doi: 10.1016/S1474-4422(14)70049-3
34. Kappos L, Bar-Or A, Cree BAC, Fox RJ, Giovannoni G, Gold R, et al. Siponimod versus placebo in secondary progressive multiple sclerosis (EXPAND): a double-blind, randomised, phase 3 study. *Lancet.* (2018) 391:1263–73. doi: 10.1016/S0140-6736(18)30475-6
35. Comi G, Kappos L, Selmaj KW, Bar-Or A, Arnold DL, Steinman L, et al. Safety and efficacy of ozanimod versus interferon beta-1a in relapsing multiple sclerosis (SUNBEAM): a multicentre, randomised, minimum 12-month, phase 3 trial. *Lancet Neurol.* (2019) 18:1009–20. doi: 10.1016/S1474-4422(19)30239-X
36. Kappos L, Fox RJ, Burcklen M, Freedman MS, Havrdova EK, Hennessy B, et al. Ponesimod compared with teriflunomide in patients with relapsing multiple sclerosis in the active-comparator phase 3 OPTIMUM study: a randomized clinical trial. *JAMA Neurol.* (2021) 78:558–67. doi: 10.1001/jamaneurol.2021.0405
37. Dubey D, Kieseier BC, Hartung HP, Hemmer B, Warnke C, Menge T, et al. Dimethyl fumarate in relapsing-remitting multiple sclerosis: rationale, mechanisms of action, pharmacokinetics, efficacy and safety. *Expert Rev Neurother.* (2015) 15:339–46. doi: 10.1586/14737175.2015.1025755
38. Shirani A, Stuve O. Natalizumab for multiple sclerosis: a case in point for the impact of translational neuroimmunology. *J Immunol.* (2017) 198:1381–6. doi: 10.4049/jimmunol.1601358
39. Shirani A, Stuve O. Natalizumab: perspectives from the bench to bedside. *Cold Spring Harb Perspect Med.* (2018) 8:a029066. doi: 10.1101/cshperspect.a029066
40. Rice GP, Filippi M, Comi G. Cladribine and progressive MS: clinical and MRI outcomes of a multicenter controlled trial. Cladribine MRI Study Group. *Neurology.* (2000) 54:1145–55. doi: 10.1212/WNL.54.5.1145
41. Montalban X, Leist TP, Cohen BA, Moses H, Campbell J, Hicking C, et al. Cladribine tablets added to IFN-beta in active relapsing MS: the ONWARD study. *Neural Neuroimmunol Neuroinflamm.* (2018) 5:e477. doi: 10.1212/NXI.0000000000000477
42. Naegelien Y, Naegelien P, von Felten S, Lorscheider J, Sonder J, Uitdehaag BMJ, et al. Association of rituximab treatment with disability progression among patients with secondary progressive multiple sclerosis. *JAMA Neurol.* (2019) 76:274–81. doi: 10.1001/jamaneurol.2018.4239
43. Makinodan T. Studies on the influence of age on immune response to understand the biology of immunosenescence. *Exp Gerontol.* (1998) 33:27–38. doi: 10.1016/S0531-5565(97)00058-2
44. Miller RA. The aging immune system: primer and prospectus. *Science.* (1996) 273:70–4. doi: 10.1126/science.273.5271.70
45. Pawelec G, Solana R. Immunosenescence. *Immunol Today.* (1997) 18:514–6. doi: 10.1016/S0167-5699(97)01145-6
46. Burns EA, Goodwin JS. Immunodeficiency of aging. *Drugs Aging.* (1997) 11:374–97. doi: 10.2165/00002512-199711050-00005
47. Smith SM, Ossa-Gomez LJ. A quantitative histologic comparison of the thymus in 100 healthy and diseased adults. *Am J Clin Pathol.* (1981) 76:657–65. doi: 10.1093/ajcp/76.5.657
48. Bertho JM, Demarquay C, Mouliau N, Van Der Meer A, Berrih-Aknin S, Gourmelon P. Phenotypic and immunohistological analyses of the human adult thymus: evidence for an active thymus during adult life. *Cell Immunol.* (1997) 179:30–40. doi: 10.1006/cimm.1997.1148
49. Palmer DB. The effect of age on thymic function. *Front Immunol.* (2013) 4:316. doi: 10.3389/fimmu.2013.00316
50. Scollay R, Wilson A, Shortman K. Thymus cell migration: analysis of thymus emigrants with markers that distinguish medullary thymocytes from peripheral T cells. *J Immunol.* (1984) 132:1089–94.
51. Penit C, Vasseur F. Expansion of mature thymocyte subsets before emigration to the periphery. *J Immunol.* (1997) 159:4848–56.
52. Mackall CL, Bare CV, Granger LA, Sharrow SO, Titus JA, Gress RE. Thymic-independent T cell regeneration occurs via antigen-driven expansion of peripheral T cells resulting in a repertoire that is limited in diversity and prone to skewing. *J Immunol.* (1996) 156:4609–16.
53. Mackall CL, Fleisher TA, Brown MR, Andrich MP, Chen CC, Feuerstein IM, et al. Distinctions between CD8+ and CD4+ T-cell regenerative pathways result in prolonged T-cell subset imbalance after intensive chemotherapy. *Blood.* (1997) 89:3700–7. doi: 10.1182/blood.V89.10.3700
54. Mackall CL, Hakim FT, Gress RE. T-cell regeneration: all repertoires are not created equal. *Immunol Today.* (1997) 18:245–51. doi: 10.1016/S0167-5699(97)81664-7
55. Mackall CL, Granger L, Sheard MA, Cepeda R, Gress RE. T-cell regeneration after bone marrow transplantation: differential CD45 isoform expression on thymic-derived versus thymic-independent progeny. *Blood.* (1993) 82:2585–94. doi: 10.1182/blood.V82.8.2585.2585
56. Davies S, Nicholson T, Laura M, Giovannoni G, Altmann DM. Spread of T lymphocyte immune responses to myelin epitopes with duration of multiple sclerosis. *J Neuropathol Exp Neurol.* (2005) 64:371–7. doi: 10.1093/jnen/64.5.371
57. Rodriguez IJ, Lalinde Ruiz N, Llano Leon M, Martinez Enriquez L, Montilla Velasquez MDP, Ortiz Aguirre JP, et al. Immunosenescence study of T cells: a systematic review. *Front Immunol.* (2020) 11:604591. doi: 10.3389/fimmu.2020.604591
58. Fukushima Y, Minato N, Hattori M. The impact of senescence-associated T cells on immunosenescence and age-related disorders. *Inflamm Regen.* (2018) 38:24. doi: 10.1186/s41232-018-0082-9
59. Ma S, Wang C, Mao X, Hao Y. B cell dysfunction associated with aging and autoimmune diseases. *Front Immunol.* (2019) 10:318. doi: 10.3389/fimmu.2019.00318
60. Swain SL, Blomberg BB. Immune senescence: new insights into defects but continued mystery of root causes. *Curr Opin Immunol.* (2013) 25:495–7. doi: 10.1016/j.coi.2013.08.001
61. Linton P, Thoman ML. T cell senescence. *Front Biosci.* (2001) 6:D248–61. doi: 10.2741/Linton
62. Tomas-Ojer P, Puthenparampil M, Cruciani C, Docampo MJ, Martin R, Sospedra M. Characterization of antigen-induced CD4+ T-cell senescence in multiple sclerosis. *Front Neurol.* (2022) 13:790884. doi: 10.3389/fneur.2022.790884
63. Broux B, Pannemans K, Zhang X, Markovic-Plese S, Broekmans T, Eijnde BO, et al. CXCR1 drives cytotoxic CD4(+)CD28(-) T cells into the brain of multiple sclerosis patients. *J Autoimmun.* (2012) 38:10–9. doi: 10.1016/j.jaut.2011.11.006
64. Negron A, Robinson RR, Stuve O, Forsthuber TG. The role of B cells in multiple sclerosis: current and future therapies. *Cell Immunol.* (2019) 339:10–23. doi: 10.1016/j.cellimm.2018.10.006
65. Guan Y, Jakimovski D, Ramanathan M, Weinstock-Guttman B, Zivadinov R. The role of Epstein-Barr virus in multiple sclerosis: from molecular pathophysiology to in vivo imaging. *Neural Regen Res.* (2019) 14:373–86. doi: 10.4103/1673-5374.245462
66. Morandi E, Jagessar SA, Hart BA, Gran B. EBV infection empowers human B cells for autoimmunity: role of autophagy and relevance to multiple sclerosis. *J Immunol.* (2017) 199:435–48. doi: 10.4049/jimmunol.1700178
67. Serafini B, Scorsi E, Rosicarelli B, Rigau V, Thouvenot E, Aloisi F. Massive intracerebral Epstein-Barr virus reactivation in lethal multiple sclerosis relapse after natalizumab withdrawal. *J Neuroimmunol.* (2017) 307:14–7. doi: 10.1016/j.jneuroim.2017.03.013
68. Palma P, Rinaldi S, Cotugno N, Santilli V, Pahwa S, Rossi P, et al. Premature B-cell senescence as a consequence of chronic immune activation. *Hum Vaccin Immunother.* (2014) 10:2083–8. doi: 10.4161/hv.28698

69. Mathias A, Perriard G, Canales M, Soneson C, Delorenzi M, Schlupe M, et al. Increased *ex vivo* antigen presentation profile of B cells in multiple sclerosis. *Mult Scler.* (2017) 23:802–9. doi: 10.1177/1352458516664210
70. LeMaout J, Szabo P, Weksler ME. Effect of age on humoral immunity, selection of the B-cell repertoire and B-cell development. *Immunol Rev.* (1997) 160:115–26. doi: 10.1111/j.1600-065X.1997.tb01032.x
71. Fraussen J, Claes N, Van Wijmeersch B, van Horsen J, Stinissen P, Hupperts R, et al. B cells of multiple sclerosis patients induce autoreactive proinflammatory T cell responses. *Clin Immunol.* (2016) 173:124–32. doi: 10.1016/j.clim.2016.10.001
72. Forsthuber TG, Cimbara DM, Ratchford JN, Katz E, Stuve O. B cell-based therapies in CNS autoimmunity: differentiating CD19 and CD20 as therapeutic targets. *Ther Adv Neurol Disord.* (2018) 11:1756286418761697. doi: 10.1177/1756286418761697
73. Manouchehri N, Stüve O. Should ocrelizumab be used in non-active primary progressive multiple sclerosis? Time for a re-assessment. *Ther Adv Neurol Disord.* (2021) 14:1756286421990500. doi: 10.1177/1756286421990500
74. Cancro MP, Hao Y, Scholz JL, Riley RL, Frasca D, Dunn-Walters DK, et al. B cells and aging: molecules and mechanisms. *Trends Immunol.* (2009) 30:313–8. doi: 10.1016/j.it.2009.04.005
75. Fulop T, Foris G, Leovey A. Age-related-changes in camp and Cgmp levels during phagocytosis in human polymorphonuclear leukocytes. *Mech Ageing Dev.* (1984) 27:233–7. doi: 10.1016/0047-6374(84)90048-4
76. Niwa Y, Kasama T, Miyachi Y, Kanoh T. Neutrophil chemotaxis, phagocytosis and parameters of reactive oxygen species in human aging - cross-sectional and longitudinal-studies. *Life Sci.* (1989) 44:1655–64. doi: 10.1016/0024-3205(89)90482-7
77. Damte B, Spagnuolo PJ, Goldsmith GGH, Marino JA. Neutrophil adhesion in the elderly - inhibitory effects of plasma from elderly patients. *Clin Immunol Immunop.* (1990) 54:247–55. doi: 10.1016/0090-1229(90)90086-6
78. Biasi D, Carletto A, Dellagnola C, Caramaschi P, Montesanti F, Zaveri G, et al. Neutrophil migration, oxidative metabolism, and adhesion in elderly and young subjects. *Inflammation.* (1996) 20:673–81. doi: 10.1007/BF01488803
79. Hazeldine J, Harris P, Chapple IL, Grant M, Greenwood H, Livesey A, et al. Impaired neutrophil extracellular trap formation: a novel defect in the innate immune system of aged individuals. *Aging Cell.* (2014) 13:690–8. doi: 10.1111/accel.12222
80. Butcher SK, Chahal H, Nayak L, Sinclair A, Henriquez NV, Sapey E, et al. Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans. *J Leukoc Biol.* (2001) 70:881–6.
81. Shaw AC, Joshi S, Greenwood H, Panda A, Lord JM. Aging of the innate immune system. *Curr Opin Immunol.* (2010) 22:507–13. doi: 10.1016/j.coi.2010.05.003
82. Nomellini V, Faunce DE, Gomez CR, Kovacs EJ. An age-associated increase in pulmonary inflammation after burn injury is abrogated by CXCR2 inhibition. *J Leukocyte Biol.* (2008) 83:1493–501. doi: 10.1189/jlb.1007672
83. Franceschi C, Capri M, Monti D, Giunta S, Olivieri F, Sevini F, et al. Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans. *Mech Ageing Dev.* (2007) 128:92–105. doi: 10.1016/j.mad.2006.11.016
84. Harper JM, Wilkinson JE, Miller RA. Macrophage migration inhibitory factor-knockout mice are long lived and respond to caloric restriction. *Faseb Journal.* (2010) 24:2436–42. doi: 10.1096/fj.09-152223
85. Greter M, Heppner FL, Lemos MP, Odermatt BM, Goebels N, Laufer T, et al. Dendritic cells permit immune invasion of the CNS in an animal model of multiple sclerosis. *Nat Med.* (2005) 11:328–34. doi: 10.1038/nm1197
86. Kreutzberg GW. Microglia: a sensor for pathological events in the CNS. *Trends Neurosci.* (1996) 19:312–8. doi: 10.1016/0166-2236(96)10049-7
87. Ransohoff RM, Perry VH. Microglial physiology: unique stimuli, specialized responses. *Annu Rev Immunol.* (2009) 27:119–45. doi: 10.1146/annurev.immunol.021908.132528
88. Kettenmann H, Hanisch UK, Noda M, Verkhratsky A. Physiology of microglia. *Physiol Rev.* (2011) 91:461–553. doi: 10.1152/physrev.00011.2010
89. Mildner A, Schmidt H, Nitsche M, Mekler D, Hanisch UK, Mack M, et al. Microglia in the adult brain arise from Ly-6Chi monocytes only under defined host conditions. *Multiple Sclerosis.* (2007) 13:S255. doi: 10.1038/nn2015
90. Ajami B, Bennett JL, Krieger C, McNagny KM, Rossi FM. Infiltrating monocytes trigger EAE progression, but do not contribute to the resident microglia pool. *Nat Neurosci.* (2011) 14:1142–9. doi: 10.1038/nn.2887
91. Braun JS, Kaissling B, Le Hir M, Zenker W. Cellular components of the immune barrier in the spinal meninges and dorsal root ganglia of the normal rat: immunohistochemical (MHC class II) and electron-microscopic observations. *Cell Tissue Res.* (1993) 273:209–17. doi: 10.1007/BF00312822
92. Giles DA, Duncker PC, Wilkinson NM, Washnock-Schmid JM, Segal BM. CNS-resident classical DCs play a critical role in CNS autoimmune disease. *J Clin Invest.* (2018) 128:5322–34. doi: 10.1172/JCI123708
93. Steger MM, Maczek C, Grubeck-Loebenstien B. Morphologically and functionally intact dendritic cells can be derived from the peripheral blood of aged individuals. *Clin Exp Immunol.* (1996) 105:544–50. doi: 10.1046/j.1365-2249.1996.d01-790.x
94. Lung TL, Saurwein-Teissl M, Parson W, Schonitzer D, Grubeck-Loebenstien B. Unimpaired dendritic cells can be derived from monocytes in old age and can mobilize residual function in senescent T cells. *Vaccine.* (2000) 18:1606–12. doi: 10.1016/S0264-410X(99)00494-6
95. Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A. Inflammaging: a new immune-metabolic viewpoint for age-related diseases. *Nat Rev Endocrinol.* (2018) 14:576–90. doi: 10.1038/s41574-018-0059-4
96. Yoo H, Park SH, Ye SK, Kim M. IFN-gamma-induced BST2 mediates monocyte adhesion to human endothelial cells. *Cell Immunol.* (2011) 267:23–9. doi: 10.1016/j.cellimm.2010.10.011
97. Manouchehri N, Hussain RZ, Cravens PD, Esaurova E, Artyomov MN, Edelson BT, et al. CD11c(+)CD88(+)CD317(+) myeloid cells are critical mediators of persistent CNS autoimmunity. *Proc Natl Acad Sci USA.* (2021) 118:e2014492118. doi: 10.1073/pnas.2014492118
98. Yona S, Kim KW, Wolf Y, Mildner A, Varol D, Breker M, et al. Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasis. *Immunity.* (2013) 38:79–91. doi: 10.1016/j.immuni.2012.12.001
99. del Pilar Martin M, Cravens PD, Winger R, Frohman EM, Racke MK, Eagar TN, et al. Decrease in the numbers of dendritic cells and CD4+ T cells in cerebral perivascular spaces due to natalizumab. *Arch Neurol.* (2008) 65:1596–603. doi: 10.1001/archneur.65.12.noc80051
100. Kapoor R, Ho PR, Campbell N, Chang I, Deykin A, Forrestal F, et al. Effect of natalizumab on disease progression in secondary progressive multiple sclerosis (ASCEND): a phase 3, randomised, double-blind, placebo-controlled trial with an open-label extension. *Lancet Neurol.* (2018) 17:405–15. doi: 10.1016/S1474-4422(18)30069-3
101. Absinta M, Sati P, Schindler M, Leibovitch EC, Ohayon J, Wu T, et al. Persistent 7-tesla phase rim predicts poor outcome in new multiple sclerosis patient lesions. *J Clin Invest.* (2016) 126:2597–609. doi: 10.1172/JCI86198
102. Kuhlmann T, Ludwin S, Prat A, Antel J, Bruck W, Lassmann H. An updated histological classification system for multiple sclerosis lesions. *Acta Neuropathol.* (2017) 133:13–24. doi: 10.1007/s00401-016-1653-y
103. Fakhri R, Matiello M, Chitnis T, Stankiewicz JM. Efficacy and safety of mycophenolate mofetil in progressive multiple sclerosis patients. *J Neurol.* (2018) 265:2688–94. doi: 10.1007/s00415-018-9050-1
104. Smith JA, Nicaise AM, Ionescu RB, Hamel R, Peruzzotti-Jametti L, Pluchino S. Stem cell therapies for progressive multiple sclerosis. *Front Cell Dev Biol.* (2021) 9:696434. doi: 10.3389/fcell.2021.696434
105. Mayo L, Trauger SA, Blain M, Nadeau M, Patel B, Alvarez JI, et al. Regulation of astrocyte activation by glycolipids drives chronic CNS inflammation. *Nat Med.* (2014) 20:1147–56. doi: 10.1038/nm.3681
106. Liddel SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, et al. Neurotoxic reactive astrocytes are induced by activated microglia. *Nature.* (2017) 541:481–7. doi: 10.1038/nature21029
107. Ambrosini E, Columba-Cabezas S, Serafini B, Muscella A, Aloisi F. Astrocytes are the major intracerebral source of macrophage inflammatory protein-3alpha/CCL20 in relapsing experimental autoimmune encephalomyelitis and in vitro. *Glia.* (2003) 41:290–300. doi: 10.1002/glia.10193
108. Stuve O, Yousef S, Slavin AJ, King CL, Patarroyo JC, Hirschberg DL, et al. The role of the MHC class II transactivator in class II expression and antigen presentation by astrocytes and in susceptibility to central

- nervous system autoimmune disease. *J Immunol.* (2002) 169:6720–32. doi: 10.4049/jimmunol.169.12.6720
109. Traugott U, Scheinberg LC, Raine CS. On the presence of Ia-positive endothelial cells and astrocytes in multiple sclerosis lesions and its relevance to antigen presentation. *J Neuroimmunol.* (1985) 8:1–14. doi: 10.1016/S0165-5728(85)80043-6
 110. Sakai K, Tabira T, Endoh M, Steinman L. Ia expression in chronic relapsing experimental allergic encephalomyelitis induced by long-term cultured T cell lines in mice. *Lab Invest.* (1986) 54:345–52.
 111. Traugott U. Detailed analysis of early immunopathologic events during lesion formation in acute experimental autoimmune encephalomyelitis. *Cell Immunol.* (1989) 119:114–29. doi: 10.1016/0008-8749(89)90228-1
 112. Porchet R, Probst A, Bouras C, Draberova E, Draber P, Riederer BM. Analysis of glial acidic fibrillary protein in the human entorhinal cortex during aging and in Alzheimer's disease. *Proteomics.* (2003) 3:1476–85. doi: 10.1002/pmic.200300456
 113. Sloane JA, Hollander W, Rosene DL, Moss MB, Kemper T, Abraham CR. Astrocytic hypertrophy and altered GFAP degradation with age in subcortical white matter of the rhesus monkey. *Brain Res.* (2000) 862:1–10. doi: 10.1016/S0006-8993(00)02059-X
 114. Coppe JB, Desprez PY, Krtolica A, Campisi J. The senescence-associated secretory phenotype: the dark side of tumor suppression. *Annu Rev Pathol.* (2010) 5:99–118. doi: 10.1146/annurev-pathol-121808-102144
 115. Angelova DM, Brown DR. Microglia and the aging brain: are senescent microglia the key to neurodegeneration? *J Neurochem.* (2019) 151:676–88. doi: 10.1111/jnc.14860
 116. Esaulova E, Cantoni C, Shchukina I, Zaitsev K, Bucelli RC, Wu GF, et al. Single-cell RNA-seq analysis of human CSF microglia and myeloid cells in neuroinflammation. *Neuro Neuroimmunol Neuroinflamm.* (2020) 7:e732. doi: 10.1212/NXI.0000000000000732
 117. Poliani PL, Wang YM, Fontana E, Robinette ML, Yamanish Y, Gilfillan S, et al. TREM2 sustains microglial expansion during aging and response to demyelination. *J Clin Invest.* (2015) 125:2161–70. doi: 10.1172/JCI77983
 118. Hametner S, Wimmer I, Haider L, Pfeifenbring S, Bruck W, Lassmann H. Iron and neurodegeneration in the multiple sclerosis brain. *Ann Neurol.* (2013) 74:848–61. doi: 10.1002/ana.23974
 119. Zrzavy T, Hametner S, Wimmer I, Butovsky O, Weiner HL, Lassmann H. Loss of 'homeostatic' microglia and patterns of their activation in active multiple sclerosis. *Brain.* (2017) 140:1900–13. doi: 10.1093/brain/awx113
 120. Lopes KO, Sparks DL, Streit WJ. Microglial dystrophy in the aged and Alzheimer's disease brain is associated with ferritin immunoreactivity. *Glia.* (2008) 56:1048–60. doi: 10.1002/glia.20678
 121. Montalban X, Hauser SL, Kappos L, Arnold DL, Bar-Or A, Comi G, et al. Ocrelizumab versus placebo in primary progressive multiple sclerosis. *New Engl J Med.* (2016) 376:209–20. doi: 10.1056/NEJMoa1606468
 122. Hawker K, O'Connor P, Freedman MS, Calabresi PA, Antel J, Simon J, et al. Rituximab in patients with primary progressive multiple sclerosis: results of a randomized double-blind placebo-controlled multicenter trial. *Ann Neurol.* (2009) 66:460–71. doi: 10.1002/ana.21867
 123. Machado-Santos J, Saji E, Troscher AR, Paunovic M, Liblau R, Gabriely G, et al. The compartmentalized inflammatory response in the multiple sclerosis brain is composed of tissue-resident CD8+ T lymphocytes and B cells. *Brain.* (2018) 141:2066–82. doi: 10.1093/brain/awy151
 124. Pardo G, Jones DE. The sequence of disease-modifying therapies in relapsing multiple sclerosis: safety and immunologic considerations. *J Neurol.* (2017) 264:2351–74. doi: 10.1007/s00415-017-8594-9
 125. Smith AL, Cohen JA, Hua LH. Therapeutic targets for multiple sclerosis: current treatment goals and future directions. *Neurotherapeutics.* (2017) 14:952–60. doi: 10.1007/s13311-017-0548-5
 126. Acosta JC, Banito A, Wuestefeld T, Georgilis A, Janich P, Morton JP, et al. A complex secretory program orchestrated by the inflammasome controls paracrine senescence. *Nat Cell Biol.* (2013) 15:978–90. doi: 10.1038/ncb2784
 127. Kuilman T, Peeper DS. Senescence-messaging secretome: SMS-ing cellular stress. *Nat Rev Cancer.* (2009) 9:81–94. doi: 10.1038/nrc2560
 128. Acosta JC, Snijders AP, Gil J. Unbiased characterization of the senescence-associated secretome using SILAC-based quantitative proteomics. *Methods Mol Biol.* (2013) 965:175–84. doi: 10.1007/978-1-62703-239-1_11
 129. Purcell M, Kruger A, Tainsky MA. Gene expression profiling of replicative and induced senescence. *Cell Cycle.* (2014) 13:3927–37. doi: 10.4161/15384101.2014.973327

Conflict of Interest: OS serves on the editorial boards of Therapeutic Advances in Neurological Disorders, has served on data monitoring committees for Genentech-Roche, Pfizer, Novartis, and TG Therapeutics without monetary compensation, has advised EMD Serono, Celgene, Genentech, Genzyme, TG Therapeutics, and VYNE, receives grant support from EMD Serono and Exalys, was a 2021 recipient of a Grant for Multiple Sclerosis Innovation (GMSI), Merck KGaA, and was funded by a Merit Review grant (federal award document number (FAIN) BX005664-01 from the United States (U.S.) Department of Veterans Affairs, Biomedical Laboratory Research and Development).

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Manouchehri, Salinas, Rabi Yeganeh, Pitt, Hussain and Stuve. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.