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Neurobiology and changing ecosystems: Toward understanding the impact of anthropogenic influences on neurons and circuits

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Rapid anthropogenic environmental changes, including those due to habitat contamination, degradation, and climate change, have far-reaching effects on biological systems that may outpace animals' adaptive responses. Neurobiological systems mediate interactions between animals and their environments and evolved over millions of years to detect and respond to change. To gain an understanding of the adaptive capacity of nervous systems given an unprecedented pace of environmental change, mechanisms of physiology and behavior at the cellular and biophysical level must be examined. While behavioral changes resulting from anthropogenic activity are becoming increasingly described, identification and examination of the cellular, molecular, and circuit-level processes underlying those changes are profoundly underexplored. Hence, the field of neuroscience lacks predictive frameworks to describe which neurobiological systems may be resilient or vulnerable to rapidly changing ecosystems, or what modes of adaptation are represented in our natural world. In this review, we highlight examples of animal behavior modification and corresponding nervous system adaptation in response to rapid environmental change. The underlying cellular, molecular, and circuit-level component processes underlying these behaviors are not known and emphasize the unmet need for rigorous scientific enquiry into the neurobiology of changing ecosystems.

KEYWORDS

climate change, neural circuits, behavioral neuroscience, adaptation, neural resilience, anthropogenic impacts, neuroethology

Introduction

Nervous systems evolved across hundreds of millions of years to allow organisms to sense and respond to external environmental changes. Being receptive and responsive, yet flexible, are features that lie at the core of the remarkable adaptive capabilities of the brain. Environments and organisms are inseparable. Evolution shapes the nervous system through experience with a dynamic range of environmental possibilities. However, rapid, human-induced environmental change may challenge the range of experience in evolutionary history as a function of the adaptive clock mechanisms imposed by genetics. How resilient are neural systems in the face of unanticipated and potentially prolonged change? What are the rules that govern whether an animal will adapt within its ecological niche, or attempt to find another one? Nervous systems mediate the interactions between organisms and their environments, and thus, understanding their response to anthropogenic environmental change is vital for the ability to predict *what* changes may transpire with rapid environmental perturbation, and *how* neuronal systems may adapt to–or fail to adapt–to those changes (O'Donnell, 2018).

Anthropogenic pressures, including but not limited to, dramatic increases in greenhouse gases and subsequent climate instability, habitat depletion, and pollution have emerged only recently in our planet's history. The most optimistic future climate change scenario, assuming nationally determined contributions are implemented by 2030, predicts warming of an average of 2.4°C by 2,100 (Kemp et al., 2022). Following our current trajectory, warming will increase by an average of 3°C. In more devastating scenarios, positive feedback loops in the carbon cycle may prevent stabilization of climate and trigger sudden and irreversible global warming in unpredictable and catastrophic ways (Steffen et al., 2018). Continuing our current trajectory is predicted to lead to an estimated extinction of one-third of all animal and plant species globally by 2070 (Román-Palacios and Wiens, 2020).

Behavioral responses of individual animals and populations to anthropogenic environmental influence are becoming increasingly well described (Tuomainen and Candolin, 2011; Candolin and Wong, 2012; Goumas et al., 2020; Gunn et al., 2022). Connecting phenomenological observations of anthropogenic influence on animal behavior to causative cellular, molecular, and circuit-level processes, however, is an open area that is ripe for inquiry. For example, certain pollutants reduce the efficiency of escape responses in fish (Weis et al., 1999, 2001; Lürling and Scheffer, 2007). Likewise, the genes, cells, and circuits that mediate escape responses are known, but the connection to the phenomenological observation does not yet exist. How do environmental pollutants modify known behavioral response mechanisms at all these levels of analysis? The advent of tools to target, monitor, and manipulate genes, cells, and circuits has yielded an immense understanding of basic neurobiology that can now be integrated with ethologically inspired approaches and questions.

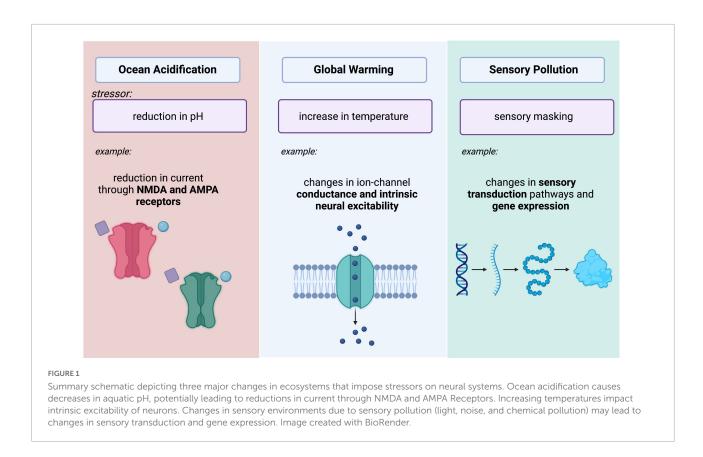
Responses to anthropogenic impacts may originate or affect neural systems at multiple levels, from changes in gene expression and neural plasticity all the way to cognition, each of which likely operate under different timescales and with varying robustness to change. Take for example cases of noise pollution in which machine sounds interfere in the same frequency range that certain species use for communication. If neural circuit activity and behavior are adaptive, frequencies of sounds used to communicate may shift (though this may create new pressures). If neural circuit plasticity does not appropriately occur, perhaps there may instead be selection for different frequency communication schemes spanning across multiple generations. Environmental changes may be too rapid for some species to survive. Species responses will also be variable, ultimately altering ecosystems in potentially irreversible ways.

Unprecedented anthropogenic change also brings forth scientific questions that would not have been imagined decades ago and would not have been approachable without the tools and domain knowledge of the present day. Anthropogenic influence on ecosystems is relatively new; we were unable to observe these phenomena because they simply did not exist or were unmeasurable with the available methods and technologies. Similarly, we did not have the domain knowledge within neuroscience (and we still do not) that would allow us access into the question space. How could we predict how noise pollution impacts auditory system development without an understanding of normal development? As studying disease can provide useful domain knowledge about basic biological processes, studying neural systems under duress may help to explicate both normal and disrupted neural function. Now is the time to combine our understanding of anthropogenic influence on ecosystems with our understanding of neuroscience domain knowledge or utilize current environmental pressures to build additional domain knowledge; now we have an entry point.

Pursuing the mechanistic foundations of some phenomenological observations with the same spatial and temporal specificity that can be achieved in well-controlled lab settings may be unfeasible, these observations have broad implications on animal behavior, sensory plasticity, contemporary evolution, and neurophysiological function. Researchers are, however, addressing questions that can feasibly be approached in the lab or at the very least, inspire new tools and conceptual frameworks. This review describes selected examples of anthropogenic influence on climate with corresponding impacts to animal behavior and explores underlying neural mechanisms. More specifically, we explore stressors imposed by ocean acidification, rises in temperature, and changes in sensory environments and how they may impact neural function (Figure 1). We view these studies as the vanguard in a new and transformative field of neuroscientific research that has the potential to transform our comprehension of the natural world and of nervous system function itself.

Ocean acidification

Rising atmospheric CO_2 from greenhouse gas emissions generates an uptake in carbonic acid within surface ocean levels, resulting in lower pH marine environments. This process,

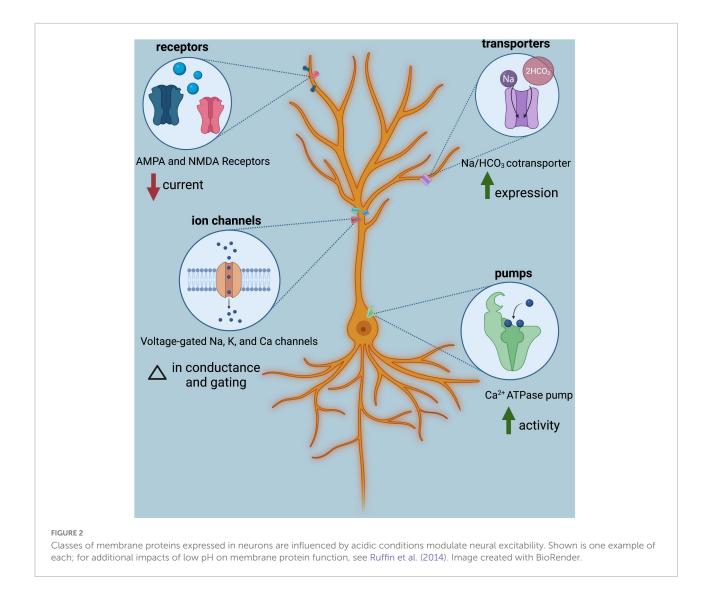


known as ocean acidification (OA) has not been observed at the current rate in the past 300 million years of inferred geologic history and is predicted to result in a global reduction of an estimated 0.7 pH units by the year 2300 (Caldeira and Wickett, 2003). OA has become a major focus of climate change study due to its link to physiological impairments on developmental, metabolic, immune, and respiratory processes (Esbaugh, 2018).

Influences of variable pH on neural excitability have long been studied, though not entirely through the lens of anthropogenic OA. Due to the pH-sensitive nature of membrane proteins expressed in neurons, namely ion channels, neurotransmitter receptors, transporters, and pumps, neural excitability is modified in acidified conditions (Figure 2; for a complete review see Ruffin et al., 2014). For example, acidified conditions change conductance and gating properties of voltage-gated sodium, calcium, and potassium channels (Tombaugh and Somjen, 1996) and also reduce current through AMPA and NMDA receptors (Giffard et al., 1990; McDonald et al., 1998). Activity of Ca²⁺-ATPase pumps increases (Pick and Karlish, 1982; Irwin et al., 1994; OuYang et al., 1994; Wolosker et al., 1997), as does expression of the Na/HCO3 cotransporter, NBCn1 (Park et al., 2010). These pH-sensitive membrane proteins affect neural excitability through a number of processes: regulating the resting membrane potential, setting thresholds for action potential firing, affecting responsiveness to receptor agonists and antagonists, setting the length of refractory periods, and synchronizing neuronal network activity (Ruffin et al., 2014).

The complexity of potential downstream effects of any combination of acidosis on these membrane proteins is difficult to predict, however, one can reasonably speculate that OA may modify neural excitability in aquatic animals and consequently may influence animal behavior. In fact, numerous experimental studies are beginning to link OA to changes specific behaviors such as foraging (Esbaugh, 2018; Jiahuan et al., 2018; Van Colen et al., 2020); reproductive behavior (Milazzo et al., 2016; Nagelkerken et al., 2021); defense behaviors (reviewed in Clements and Comeau, 2019); predatorprey interactions (reviewed in Draper and Weissburg, 2019); and habitat selection (Wang and Wang, 2021). At this time, there is little clarity on which behavioral changes are the result of behavioral adaptations, perturbations on pH-dependent physiological mechanisms, or direct perturbations on cell and circuit neurobiological processes that underlie behavior.

An alternate model suggests that impacts of OA on behavior of aquatic animals are in part mediated by changes in physiological pH which is thought to indirectly decrease inhibitory neurotransmission. In a set of studies, larval clownfish reared in lower pH conditions (higher CO_2) than controls demonstrated altered olfactory discrimination related to predator cues and cues that would allow them to locate suitable habitat settlement sites



(Munday et al., 2009; Dixson et al., 2010). More specifically, fish exposed to high CO₂ were attracted to predator odors, rather than avoidant. Interestingly, the onset timing of these behavioral alterations was dependent on the magnitude of pH reduction but was reversible following 2-day exposure to control seawater (Munday et al., 2010). These results hinted at an unknown CO2-dependent, yet reversible mechanism that altered behavior. Typically, when exposed to high CO2/low pH conditions, known as hypercapnic conditions, fish regulate acid-base relevant ion concentrations, namely bicarbonate [HCO₃⁻] and chloride [Cl⁻], in order to maintain blood and tissue pH (Ishimatsu et al., 2008). This results in an upregulation of active H⁺ excretion and an accumulation of HCO3⁻ in physiological fluids, which is proposed to reduce Cl⁻ in extracellular compartments (Tresguerres and Hamilton, 2017). The authors hypothesized that changes in HCO_3^- and Cl⁻ gradients would reverse net ionic influxes through GABAA channels, switching their function from inhibitory to excitatory.

To test this hypothesis, the experimenters pretreated high CO_2 exposed fish with the $GABA_A$ receptor antagonist, gabazine, and observed a restoration in prey avoidance behavior (Nilsson et al., 2012). These $GABA_A$ receptor antagonist studies have now been replicated in multiple marine and freshwater species (with varying results) and posit that OA conditions result in neuronal hyperexcitability that explains observed changes in behavior (reviewed in Tresguerres and Hamilton, 2017).

The GABA_A model stands as one of the few proposed and experimentally tested basic neurobiological mechanisms downstream of anthropogenic environmental changes. However, it is critical to note that ion concentrations have not been directly measured under hypercapnic conditions that arise from OA and that these results are controversial because animals must be in states of extreme, unnaturally acidic conditions for GABA_A channels to reverse the direction of ion flow. This further highlights the need for targeted approaches to understand how OA may impact membrane protein function in

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neurons as well as how homeostatic mechanisms that regulate physiological pH may impact neural processes. Species-specific neuronal responses to OA are also quite variable (Tresguerres and Hamilton, 2017), suggesting a potential comparative area of study into system variability, adaptation, and resilience.

Sensory ecology and change

Sensory ecology focuses on how and why animals obtain information from their environments. Natural animal behaviors largely rely on the perception of a diversity of biologically important environmental sensory cues, for example temperature fluctuation and length of daylight prompting the migration of monarch butterflies (Guerra, 2020). Anthropogenic environmental change presents an opportunity to re-examine the central questions of sensory ecology; if the sensory environment has changed through anthropogenic activity, what is the impact on behavioral systems? A considerable body of research is focused on sensory ecology related to changing ecosystems, specifically in how noise, light, and chemical and material pollutants impair sensory cue production, transmission, detection, and discrimination, and yet the neurobiological impacts or consequences of change are not well understood (for comprehensive reviews, see Kelley et al., 2018; Draper and Weissburg, 2019).

Anthropogenic activity may modify or mask the native sensory information that animals encounter. Sound, light, and chemical pollution, for example, reduce the quality of auditory, visual, and chemosensory cues required to guide behavior and results in changes to behavioral strategies, as illustrated in the following examples. Specific neurophysiological mechanisms governing these behavioral adaptations, however, are not yet explored.

Auditory systems

Animals may alter their behavior in response to increasing anthropogenic noise from sources such as shipping and boating, automobile, sonar, seismic surveys, deep-sea mining, ocean dredging, wind turbines, and numerous other sources (Tuomainen and Candolin, 2011; Kunc et al., 2016; Harding et al., 2019). Typically, humpback whales rely on high-information content vocal communication with conspecifics, but under high ambient noise conditions shift their communication schemes to instead display more surfacegenerated sounds at the expense of less information-content (Dunlop et al., 2010). When exposed to low-frequency sonar, however, they increase the repetition of phrases in their songs, suggesting that even within the same species, behavioral modulation to different types of noise itself is stimulus-specific and reflects a need to optimize information content fidelity (Miller et al., 2000). Similar findings have been reported in terrestrial species where urbanization generates loud and low-frequency noise (Warren et al., 2006). For example, some bird species compensate for anthropogenic noise by modulating specific components of their song, in a species-specific manner (Lohr et al., 2003; Brumm, 2004; Halfwerk and Slabbekoorn, 2009; Slabbekoorn and Halfwerk, 2009; Zollinger et al., 2011; Ríos-Chelén et al., 2013; Gil et al., 2015).

In addition to potential changes in neurotransmission, ocean acidification also increases ambient noise in the ocean due to the pH dependence of sound absorption in water, specifically within the range of 0.01–10 kHz (Fisher and Simmons, 1977; Hester et al., 2008; Ilyina et al., 2009). This frequency-dependent modification is within the hearing sensitivity range of many invertebrates (Albert and Kozlov, 2016), fishes (Ladich and Fay, 2013) and marine mammals (Weilgart, 2007) and may interfere neuronal circuits that mediate echolocation, navigation, mate choice, habitat selection, and other behaviors that depend on acoustic cues in the marine soundscape.

While impacts of anthropogenic noise on animal behavior are well described in both marine and terrestrial ecosystems, much less, if anything, is known about its influence on neurophysiology and sensory processing within the brain. Several studies monitor changes in auditory sensitivity thresholds using auditory evoked potentials (AEPs) and acoustic brainstem responses (ABRs) and find that anthropogenic noise decreases auditory sensitivity across a range of species including fish (Vasconcelos et al., 2007; Codarin et al., 2009), marine mammals (Nachtigall et al., 2004; Lucke et al., 2009), leatherback turtles (Piniak et al., 2012, 2016), and polar bears (Nachtigall et al., 2007). Damage to auditory systems may be through a hearing organ-such as destruction of hair cells due to high frequency exposure-or through the perturbation of the secondary signal in auditory circuit processing. It remains enigmatic as to how these changes in hearing system function connect to observed behavioral adaptations and how the brain adapts to dynamic changes in signal-to-noise of acoustic signals. What might unraveling these questions teach us about the limits or flexibility of both behavior and sensory processing?

Visual systems

For many animals, vision is a vital sense that drives behaviors, ranging from mate selection to migration to prey capture, and many others (Cronin et al., 2014). Studies of visual ecology related to anthropogenic activity primarily target three sources that effectively mask visual cues: light pollution, changes in water turbidity, and shifts in the ocean depth that animals inhabit (known as bathymetric shifts).

Artificial light has impacts on wildlife as well as human health by altering natural light regimes spatially, temporally, and spectrally (Gaston et al., 2013; Falcón et al., 2020). Light itself is an information source used to regulate metabolic and physiological processes, circadian rhythms and photoperiodism, migratory and orienting behavior, and visual perception. For a review on artificial lighting and LED impacts see Falcón et al. (2020). Take for example, sea turtle hatchlings that use lunar illumination and its reflection on water to navigate to sea. There are several accounts of disoriented hatchlings who travel to erroneous locations owning to light pollution, and even those who do successfully reach the sea sometimes return to the shore when shore-based artificial lighting was present (Truscott et al., 2017).

An emerging body of work is examining plasticity of retinal opsin gene expression in reef fish under differing light regimes (Carleton et al., 2020; Musilova et al., 2021; Fogg et al., 2022). Using transcriptomics and qPCR, experimenters have discovered that artificial light increases expression of cone (but not rod) opsin genes sensitive to a broad range of wavelengths. Shifts in opsin expression under differing light conditions were also reversible and highly plastic. Future work will examine if plastic changes in opsin gene expression are maintained with long-term exposure to artificial light.

Run-off from agricultural and septic sources, dredging, and erosion create murky, high-turbidity environments that mask visual cues in water. In one study, turbidity reduced the reactive distance of predators to their prey, making them less efficient predators (Gregory, 1993). In fact, water turbidity is proposed to shape predator-prey interactions of aquatic species by altering visual sensitivity thresholds (Abrahams and Kattenfeld, 1997; Van de Meutter et al., 2005). Using the optomotor response, researchers have measured reduced visual sensitivity as a function of turbidity (Nieman et al., 2018; Suriyampola et al., 2018). How might the fidelity of visual signals and sensitivity to those signals influence behavioral modifications? In simulated turbid environments, western rainbowfish compensated for turbidity by enhancing the vibrance of their color displays to maintain visual communication with conspecifics (Kelley et al., 2012). It will be fascinating to discover how visual signals are utilized by the brain and integrated with internal and ecological demands to generate behavioral modifications, and at what evolutionary rate.

Warming and ocean acidification have caused marine animals to inhabit deeper waters (known as bathymetric shifts; Perry et al., 2005; Pinsky et al., 2013) where optical conditions are poorer in both illumination and spectral breadth. In these scenarios, marine organisms experience reduced contrast and color-cues that may be used for survival (for a wonderful review, see Caves and Johnsen, 2021). In cases of high turbidity and bathymetric shifts optical properties are similar; both entail reduced illumination and narrowing of spectral breadth. Turbidity, however, can be dynamic and transient while bathymetric conditions are more stable; how might sensory adaptation across these timescales differ? What may be the impact of inhabiting turbid or bathymetric waterscapes on visually-mediated behavioral circuits?

Chemosensory systems

Physiological impacts of environmental pollutants are moderately well studied in terms of toxicology and human health, but less so in the context of the sensory ecology and neuroethology. In humans, multifactorial environmental pollution has been associated with poor health outcomes, including neurological disorders (Manisalidis et al., 2020), in addition to impacts on stress (Thomson et al., 2019) and immunity (Bauer et al., 2012; Thomson et al., 2019). Unsurprisingly, chemical pollutants can impair chemoreception in animals (Troyer and Turner, 2015; Ajmani et al., 2016). This is best described in aquatic species, as opposed to terrestrial species, where the olfactory epithelium is in direct contact with environmental pollutants in water (Whitlock and Palominos, 2022).

Ocean acidification impairs behaviors that rely on chemosensation, such as habitat settlement, predator avoidance, foraging, and others (for a review, see Leduc et al., 2013), though these studies rarely conduct individual sensory manipulations in isolation. High CO₂, and consequently, low pH, are proposed to modify the structure of odorants as well as odorant-receptor affinity, resulting in reduced olfactory sensitivity (Porteus et al., 2021). Brain homogenates of larval sea bream reared in acidified conditions displayed a pH-dependent reduction in GABA and acetylcholine neurotransmitter concentrations as well as changes in expression of genes related to olfactory transduction. Specifically, experimenters observed a downregulation of genes encoding positive gene regulators (the olfaction-specific G protein, Golf, and G-protein signaling 2, RGS2), and opposingly, an upregulation in genes encoding for negative regulators (G protein-coupled receptor kinase, GRK, and arrestin), suggesting that ocean acidification has direct influence on chemosensory transduction pathways, down to the molecular level (Jiahuan et al., 2018).

In addition to pollutants and ocean acidification, nano and micro-plastics also affect animal chemosensation. Breakdown of phytoplankton and marine algae during zooplankton grazing releases dimethyl sulfide. Dimethyl sulfide attracts seabirds to these zooplankton grazing sites in the midst of vast stretches of featureless ocean, effectively serving as an olfactory cue to inform seabirds of zooplankton prey location. Nano-and microplastics accumulate phytoplankton and marine algae on their surfaces, which also release dimethyl sulfide when decomposing; no zooplankton needed. Since dimethyl sulfide is released by decomposing phytoplankton and algae on microplastic surfaces, seabirds mistakenly consume microplastics (Savoca and Nevitt, 2014; Savoca et al., 2016). The same sensory cue seabirds evolved to reliably detect, localize, and depend on for their survival now serves as an anthropogenic sensory trap.

Physiological changes in response to anthropogenic stresses are slowly beginning to be described but underlying genetic or cellular systems driving their change are not well understood, as demonstrated by the ocean acidification example above. In a beautiful set of studies, Thimmegowda et al. (2020) tracked activity of Giant Asian honeybees in distinct locations with varying levels of air pollution across Bangalore, India over the course of 3 years. High-levels of air pollution correlated with changes in honeybee survival, flower visitation, heart rate, and bee morphology. The experimenters found changes in gene expression related to metabolism, stress, and immunity in antennae and heart tissue. Intriguingly, lab-reared Drosophila exposed to the same field sites also underwent similar changes in physiology and gene expression, providing accessibility into the mechanistic consequences underlying human-generated environmental stress. Perhaps, there is an underlying sensory component to olfactory signaling that is impaired by certain pollutants, alongside other physiological processes.

Pollutants can also affect sensory processes outside of chemosensation through neurotoxic effects. A recent study revealed that chronic sublethal exposure to cholinergic pesticides impairs wide-field visual motion detection in honeybees, as assayed through the optomotor response (Parkinson et al., 2022). This impairs the ability for honeybees to fly in straight, directed trajectories when they have gone off-course. At a molecular level, pesticide exposure resulted in altered stress and detoxification gene expression in the brain. Exposure to sulfoxaflor, one of the cholinergic insecticides tested, resulted in increased apoptosis primarily in the optic lobe where visual information is processed. These results suggest that cholinergic pesticides impair encoding of optic flow cues and result in disrupted optomotor behaviors.

Temperature, described in depth in the next section, also produces synergistic effects with pollutants that exacerbates their neurotoxic impacts, potentially by increasing contaminant uptake and/or modifying bioavailability (Sokolova and Lannig, 2008). Polystyrene plastic nanoparticle exposure, for example, exerts changes in specific metabolic pathways that lead to brain tissue degradation and disruption in circadian rhythms in zebrafish. In this study, spectral analyses also detected polystyrene particles in brain homogenates at all experimental temperatures tested but were generally found to be more concentrated at higher temperatures; any level of microplastic exposure is likely detrimental, but even more so at higher temperatures (Sulukan et al., 2022). Sodium cyanide poisoning from illegal, yet still prevalent, cyanide fishing practices also poses a growing threat to reef fish nervous systems due to the synergistic impacts from warming oceans (Madeira et al., 2020). Additional work is needed to understand the effects of combined perturbations on nervous system function.

Other sensory systems

Sensory systems outside of auditory, visual, or chemosensory modalities are also impacted by anthropogenic activity. Magnetic fields created by electric current flow in subsea cables generate electric fields between 0.5 and 100 μ V/m (England and Robert, 2022), which is within the detection range of aquatic electroreceptive organisms (Peters et al., 2007; Hutchison et al., 2020). Experimentally, magnetoreceptive lobsters and electro-sensitive skates exhibit increased exploratory behavior when exposed to electromagnetic fields from simulated underwater cable deployments (Hutchison et al., 2020). Deeper descriptions of electromagnetic sensing are needed in order to understand how these increases in exploratory behavior may functionally affect electro- and magnetoreceptive sensory systems.

It is intriguing to consider how sensory processing may be modified in cases of environmental perturbation, where the cause is indirect or direct; could compensation by alternate sensory systems result in the same efficacy and robustness in behavior? How might the brain know when to produce behavioral or genetic adaptations in response to inadequate sensory cues? Within a neural-circuit context, how might circuit elements compensate for rapid change? How might the *how* and *why* of sensory ecology change? Anthropogenic activity is essentially creating a *new* stimulus set; mechanistic understanding of sensory system biology is tractable in a lab setting if utilizing environmentally informed stimuli.

Temperature

A central concern of anthropogenic environmental impacts is temperature fluctuation that arises from elevated greenhouse gases which trap heat in the atmosphere. Climate model simulations predict that the global average temperature will increase between 1.1°C and 5.4°C by the year 2100 (Friedlingstein and Prentice, 2010; Meier et al., 2012), depending on the extent and efficacy of climate action. Temperature itself has profound impacts on a multitude of physiological processes, for example sex determination (Rhen and Schroeder, 2017) and metabolism (Schulte, 2015). Likewise, temperature alters biophysical properties of cells such as membrane fluidity (Fan and Evans, 2015) and ion permeability (Lamas et al., 2019).

In neurons, even slight physiological fluctuations in temperature alter neural physiology and function (Moser et al., 1993; Andersen and Moser, 1995). Minor reductions in temperature have dramatic impacts on synaptic vesicle release processes and presynaptic mechanisms of neural plasticity (Katz and Miledi, 1965; Micheva and Smith, 2005). Elevated temperatures enhance neural excitability by altering intrinsic

membrane properties (Buzatu, 2009; Kim and Connors, 2012). More specifically, ion channels and signaling proteins are temperature sensitive (Tang et al., 2010; Robertson and Money, 2012) such that elevated temperatures result in changes to the resting membrane potential, the generation of action potentials, the velocity of action potential propagation, and action potential duration (Buzatu, 2009). These alterations in membrane excitability are reflected in unstructured neural circuit activity (see examples below) and have been shown to modify a range of neurobiological processes including sensory transduction, motor reflexes, locomotion, learning, and memory (Vornanen, 2017). Mechanisms of thermal acclimation counteract effects of acute elevated temperatures by changing the number of ion channels and through differential expression of temperature-specific isoforms of ion channels (Vornanen, 2017). Biology displays no shortage of temperaturedependent processes. An emerging field of research seeks to understand how temperature change arising from climate change influences neurobiological function and the resilience of neural circuits.

This is perhaps best studied in marine invertebrates where water temperature varies dramatically daily and seasonally. Body temperature of ectothermic organisms track ambient temperatures so they must rely on rapid behavioral and physiological adaptations for survival. In normal conditions ion flow is balanced across neuronal membranes, allowing neural circuits to produce appropriate responses and activity patterns. In the stomatogastric nervous system (STNS) of the crab, for example, ion channel flow is modulated to maintain function of the rhythmic circuit that mediates pyloric filtering of food in the stomach, despite variability in temperature (Tang et al., 2010; Soofi et al., 2014; Kushinsky et al., 2019). Pattern generating neurons in the crab are remarkably robust to changes in temperature up to a point, however above this limit, which is regulated by adaptive homeostatic mechanisms (Tang et al., 2012), temperature causes an imbalance of inhibitory and excitatory conductances leading to membrane leakage and "crashing," or dysfunction of the rhythmic circuit by shunting action potential generation (Stein and Harzsch, 2021). Interestingly, neuromodulatory input enhances the robustness of STNS rhythms (Städele et al., 2015; Haddad and Marder, 2018; DeMaegd and Stein, 2021) which may also contribute to robustness of temperature adaptation.

Using wild-caught crabs, an inadvertently longitudinal study discovered that crash temperatures at which the crab STNS circuit ceased to function varied across years such that higher crash temperatures of STNS neurons reflected increases in ocean temperature (Marder and Rue, 2021). This suggests that the robustness of this circuit relies on the ability to acclimate to ambient temperatures, but begs the question of how robust this adaptive capacity will fare under continually warming oceans; in other words, what are the limits? What features are associated with enhanced resilience and adaptability in individuals and how might this contribute to behavioral adaptation, migration, or attrition?

Learning and cognitive function also suffer temperaturedependent impairment in humans (Taylor et al., 2016; Khan et al., 2021; Park et al., 2021) and in animals (reviewed in Soravia et al., 2021). Zebrafish exposed to temperatures 2°C above their voluntary thermal maximum (the temperature that elicits avoidant behavior; Rey et al., 2015) for a prolonged period, exhibited reduced interest in novel environments and impaired cognitive performance in Y-maze behavioral tests that correlated with a downregulation of proteins associated with synaptic transmission (Toni et al., 2019). Similar impairments and related patterns in gene expression from acute extreme temperatures have been replicated across many species (Soravia et al., 2021). It is proposed that ongoing adaptation of heat shock proteins will determine thermal tolerance of organisms (González et al., 2016; Hupało et al., 2018). In fact, at short timescales, heat shock protein expression instead prevents cellular damage and improves long memory formation in great pond snails (Sunada et al., 2016).

In a fascinating study, Port Jackson sharks raised in predicted end of the century temperatures (3° C increase) out-performed their normal-temperature reared counterparts in a quantity discrimination task, in both time to learning and accuracy at the task (Pouca et al., 2019). While changes in metabolic demands may perhaps be the driver of these observations, there may be other neural correlates or processes associated with cognition. Cognition itself may act as a buffer to counteract detrimental impacts of changing ecosystems (Brown, 2012). More work is needed to understand how temperature-dependent impairments in cognition are preserved or unprotected by compensatory and adaptive mechanisms, and how cognition may play a role in determining species-specific responses to changing ecosystems.

Discussion

Rapid anthropogenic environmental change is occurring and consequently, its influence on animal behavior and corresponding neural substrates is a very large experimentin-progress. While nervous systems evolved to adapt to environmental changes at non-disruptive scales, current change rates may outpace adaptive behavioral responses (Radchuk et al., 2019). The underlying impacts that changing environments have on neural processes are considerably underexplored, providing ample opportunity for scientific research with the potential for transformative discoveries.

Neuroscientists generally draw conclusions about the brain using experimentally generated, well-controlled physiological or environmental perturbations. In the natural world environmental perturbations, however, are variable, as are the physiologies, diets, genetic backgrounds, and other features of natural populations. Neuroscience as a field has gone great lengths to clamp these variables in exchange for reliability, and the subsequent ability to draw conclusions. Experimentally approaching neurobiology and changing ecosystems in such a way that captures true environmental variability while maintaining experimental control will not always be feasible, highlighting the necessity for a coalescence between lab and in-field studies. As a labonly approach, experimenters can implement predictions from global climate models as controlled experimental variables, as demonstrated by multiple studies presented in this perspective piece. Exposure to in-field studies can begin by increased crosstalk and collaboration between ecologists, evolutionary biologists, and neurobiologists. Together, this will not only allow a deeper understanding of limits and capabilities of cell and neural circuit function, but help improve predictive models for climate change impact on species and populations.

Though many natural populations are threatened by anthropogenic activity, some species are thriving. Jellyfish, which have an increasing global ecological and economic significance, are more successful in warmer water, potentially due to longer breeding periods (Brotz et al., 2012). They have also recently been demonstrated as a genetically tractable model organism (Weissbourd et al., 2021). Comparative studies may allow us to understand what parameters enable some organisms to thrive while others fail to adapt.

Changes in seasonality, resource availability, and resource quality are shifting the locations that animals inhabit, leading to new inter-species interactions (Cohen et al., 2018; Kharouba et al., 2018). As new predator-prey interactions emerge, or new forms of inter-species cooperation develop, these imminent phenomenological observations may serve as a basis for fascinating novel studies of animal behavior and adaptive neural circuits. Conceivably, environmental pressures are creating new mechanisms that counteract stress that we have yet to discover.

There is growing advocacy for neuroscientists to take specific actions to mitigate the climate crisis by reducing emissions (e.g., virtual conference attendance, reduced travel, and reductions in waste and energy consumption in lab operations. For useful guides, see Aron et al., 2020; Gardner et al., 2021; Aron, 2022; Duhaime, 2022). Similarly, this emerging area of scientific research may have societal implications that may help to shape climate adaptation, rather than mitigation, responses. In the case of the study of pollutants and pollinators (Thimmegowda et al., 2020), there are direct implications for food security, based on the chemosensory ecology of bees. Potential policy and conservation applications likewise can be derived or informed by neuroscientific studies related to anthropogenic impacts on animals and ecosystems. This new avenue may evolve into applied environmental neuroscience research driven by the underlying question of how nervous systems cope with increasingly variable and extreme changes in ecosystems. Anthropogenic impacts on ecosystems and natural animal populations are inescapable, but they provide a new challenge to discover fundamental properties of cells and neural circuits.

Author contributions

AM conducted the literature review and wrote the manuscript. AB provided the feedback and guidance on the manuscript. Both authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

Abrahams, M. V., and Kattenfeld, M. G. (1997). The role of turbidity as a constraint on predator-prey interactions in aquatic environments. *Behav. Ecol. Sociobiol.* 40, 169–174. doi: 10.1007/s002650050330

Ajmani, G. S., Suh, H. H., and Pinto, J. M. (2016). Effects of ambient air pollution exposure on olfaction: A review. *Environ. Health Perspect.* 124, 1683–1693.

Albert, J. T., and Kozlov, A. S. (2016). Comparative aspects of hearing in vertebrates and insects with antennal ears. *Curr. Biol.* 26, R1050–R1061. doi: 10.1016/j.cub.2016.09.017

Andersen, P., and Moser, E. I. (1995). Brain temperature and hippocampal function. *Hippocampus* 5, 491-498. doi: 10.1002/hipo.450050602

Aron, A. R. (2022). A neurosurgeon's climate fight minding the climate: How neuroscience can help solve our environmental crisis *Ann-Christine Duhaime* Harvard University Press, 2022. 336 pp. *Science* 378, 255–255. doi: 10.1126/science.ade5672

Aron, A. R., Ivry, R. B., Jeffery, K. J., Poldrack, R. A., Schmidt, R., Summerfield, C., et al. (2020). How can neuroscientists respond to the climate emergency? *Neuron* 106, 17–20. doi: 10.1016/j.neuron.2020.02.019

Bauer, R. N., Diaz-Sanchez, D., and Jaspers, I. (2012). Effects of air pollutants on innate immunity: The role of toll-like receptors and nucleotide-binding oligomerization domain-like receptors. *J. Allergy Clin. Immunol.* 129, 14–24. doi: 10.1016/j.jaci.2011.11.004

Brotz, L., Cheung, W. W. L., Kleisner, K., Pakhomov, E., and Pauly, D. (2012). Increasing jellyfish populations: Trends in Large Marine Ecosystems. *Jellyfish Blooms IV* 220, 3–20. doi: 10.1007/978-94-007-5316-7_2

Brown, C. (2012). "Experience and learning in changing environments," in *Behavioural responses to a changing world*, eds U. Candolin and B. B. M. Wong (Oxford: Oxford University Press), doi: 10.1093/acprof:osobl/9780199602568.003. 0004

Brumm, H. (2004). Causes and consequences of song amplitude adjustment in a territorial bird: A case study in nightingales. *An. Acad. Bras. Cienc.* 76, 289–295. doi: 10.1590/s0001-37652004000200017

Buzatu, S. (2009). The temperature-induced changes in membrane potential. *Riv. Biol.* 102, 199–217.

Caldeira, K., and Wickett, M. E. (2003). Anthropogenic carbon and ocean pH. *Nature* 425, 365–365. doi: 10.1038/425365a

Candolin, U., and Wong, B. B. M. (2012). Behavioural responses to a changing world: Mechanisms and consequences. Oxford: OUP Oxford.

Carleton, K. L., Escobar-Camacho, D., Stieb, S. M., Cortesi, F., and Marshall, N. J. (2020). Seeing the rainbow: Mechanisms underlying spectral sensitivity in teleost fishes. *J. Exp. Biol.* 223;jeb193334. doi: 10.1242/jeb.193334

Caves, E. M., and Johnsen, S. (2021). The sensory impacts of climate change: Bathymetric shifts and visually mediated interactions in aquatic species. *Proc. Biol. Sci.* 288:20210396. doi: 10.1098/rspb.2021.0396

Clements, J. C., and Comeau, L. A. (2019). Behavioral defenses of shellfish prey under ocean acidification. J. Shellfish Res. 38:725. doi: 10.2983/035.038. 0324

Codarin, A., Wysocki, L. E., Ladich, F., and Picciulin, M. (2009). Effects of ambient and boat noise on hearing and communication in three fish species living in a marine protected area (Miramare, Italy). *Mar. Pollut. Bull.* 58, 1880–1887. doi: 10.1016/j.marpolbul.2009.07.011

Cohen, J. M., Lajeunesse, M. J., and Rohr, J. R. (2018). A global synthesis of animal phenological responses to climate change. *Nat. Clim. Change* 8, 224–228. doi: 10.1038/s41558-018-0067-3

Cronin, T. W., Johnsen, S., Justin Marshall, N., and Warrant, E. J. (2014). Visual ecology. Princeton, NJ: Princeton University Press. doi: 10.23943/princeton/9780691151847.001.0001

DeMaegd, M. L., and Stein, W. (2021). Neuropeptide modulation increases dendritic electrical spread to restore neuronal activity disrupted by temperature. *J. Neurosci.* 41, 7607–7622. doi: 10.1523/JNEUROSCI.0101-21.2021

Dixson, D. L., Munday, P. L., and Jones, G. P. (2010). Ocean acidification disrupts the innate ability of fish to detect predator olfactory cues. *Ecol. Lett.* 13, 68–75. doi: 10.1111/j.1461-0248.2009.01400.x

Draper, A. M., and Weissburg, M. J. (2019). Impacts of global warming and elevated CO2 on sensory behavior in predator-prey interactions: A review and synthesis. *Front. Ecol. Evol.* 7:72. doi: 10.3389/fevo.2019.00072

Duhaime, A.-C. (2022). *Minding the climate: How neuroscience can help solve our environmental crisis.* Cambridge MA: Harvard University Press.

Dunlop, R. A., Cato, D. H., and Noad, M. J. (2010). Your attention please: Increasing ambient noise levels elicits a change in communication behaviour in humpback whales (*Megaptera novaeangliae*). *Proc. Biol. Sci.* 277, 2521–2529. doi: 10.1098/rspb.2009.2319

England, S. J., and Robert, D. (2022). The ecology of electricity and electroreception. *Biol. Rev.* 97, 383-413. doi: 10.1111/brv.12804

Esbaugh, A. J. (2018). Physiological implications of ocean acidification for marine fish: Emerging patterns and new insights. *J. Comp. Physiol. B* 188, 1–13. doi: 10.1007/s00360-017-1105-6

Falcón, J., Torriglia, A., Attia, D., Viénot, F., Gronfier, C., Behar-Cohen, F., et al. (2020). Exposure to artificial light at night and the consequences for flora, fauna, and ecosystems. *Front. Neurosci.* 14:602796. doi: 10.3389/fnins.2020.602796

Fan, W., and Evans, R. M. (2015). Turning up the heat on membrane fluidity. *Cell* 161, 962–963. doi: 10.1016/j.cell.2015.04.046

Fisher, F. H., and Simmons, V. P. (1977). Sound absorption in sea water. J. Acoust. Soc. Am. 62, 558–564. doi: 10.1121/1.381574

Fogg, L. G., Cortesi, F., Gache, C., Lecchini, D., Justin Marshall, N., and de Busserolles, F. (2022). Developing and adult reef fish show rapid, reversible lightinduced plasticity in their visual system. *bioRxiv* [Preprint]. doi: 10.1101/2022.06. 02.494615

Friedlingstein, P., and Prentice, I. C. (2010). Carbon-climate feedbacks: A review of model and observation based estimates. *Curr. Opin. Environ. Sustain.* 2, 251–257. doi: 10.1016/j.cosust.2010.06.002

Gardner, C. J., Thierry, A., Rowlandson, W., and Steinberger, J. K. (2021). From publications to public actions: The role of universities in facilitating academic advocacy and activism in the climate and ecological emergency. *Front. Sustain.* 2:679019. doi:10.3389/frsus.2021.679019

Gaston, K. J., Bennie, J., Davies, T. W., and Hopkins, J. (2013). The ecological impacts of nighttime light pollution: A mechanistic appraisal. *Biol. Rev. Camb. Philos. Soc.* 88, 912–927. doi: 10.1111/brv.12036

Giffard, R. G., Monyer, H., Christine, C. W., and Choi, D. W. (1990). Acidosis reduces NMDA receptor activation, glutamate neurotoxicity, and oxygen-glucose deprivation neuronal injury in cortical cultures. *Brain Res.* 506, 339–342. doi: 10.1016/0006-8993/90)91276-m

Gil, D., Honarmand, M., Pascual, J., Pérez-Mena, E., and Garcia, C. M. (2015). Birds living near airports advance their dawn chorus and reduce overlap with aircraft noise. *Behav. Ecol.* 26, 435–443. doi: 10.1093/beheco/aru207

González, K., Gaitán-Espitia, J., Font, A., Cárdenas, C. A., and González-Aravena, M. (2016). Expression pattern of heat shock proteins during acute thermal stress in the Antarctic sea urchin, *Sterechinus neumayeri. Rev. Chil. Hist. Nat.* 89:2. doi: 10.1186/s40693-016-0052-z

Goumas, M., Lee, V. E., Boogert, N. J., Kelley, L. A., and Thornton, A. (2020). The role of animal cognition in human-wildlife interactions. *Front. Psychol.* 11:589978. doi: 10.3389/fpsyg.2020.589978

Gregory, R. S. (1993). Effect of turbidity on the predator avoidance behaviour of juvenile chinook salmon (*Oncorhynchus tshawytscha*). *Can. J. Fish. Aquatic Sci.* 50, 241–246. doi: 10.1139/f93-027

Guerra, P. A. (2020). The monarch butterfly as a model for understanding the role of environmental sensory cues in long-distance migratory phenomena. *Front. Behav. Neurosci.* 14:600737. doi: 10.3389/fnbeh.2020.600737

Gunn, R. L., Hartley, I. R., Algar, A. C., Niemelä, P. T., and Keith, S. A. (2022). Understanding behavioural responses to human-induced rapid environmental change: A meta-analysis. *Oikos* 2022:e08366. doi: 10.1111/oik. 08366

Haddad, S. A., and Marder, E. (2018). Circuit robustness to temperature perturbation is altered by neuromodulators. *Neuron* 100, 609.e-623.e.

Halfwerk, W., and Slabbekoorn, H. (2009). A behavioural mechanism explaining noise-dependent frequency use in urban birdsong. *Anim. Behav.* 78, 1301–1307. doi: 10.1016/j.anbehav.2009.09.015

Harding, H. R., Gordon, T. A. C., Eastcott, E., Simpson, S. D., and Radford, A. N. (2019). Causes and consequences of intraspecific variation in animal responses to anthropogenic noise. *Behav. Ecol.* 30, 1501–1511. doi: 10.1093/beheco/arz114

Hester, K. C., Peltzer, E. T., Kirkwood, W. J., and Brewer, P. G. (2008). Unanticipated consequences of ocean acidification: A noisier ocean at lower pH. *Geophys. Res. Lett.* 35:L1960135. doi: 10.1029/2008gl034913

Hupało, K., Wolfgang Riss, H., Grabowski, M., Thiel, J., Bącela-Spychalska, K., and Meyer, E. I. (2018). Climate change as a possible driver of invasion and differential in HSP70 expression in two genetically distinct populations of the

invasive killer shrimp. Dikerogammarus villosus. Biol. Invasion. 20, 2047–2059. doi: 10.1007/s10530-018-1679-2

Hutchison, Z. L., Gill, A. B., Sigray, P., He, H., and King, J. W. (2020). Anthropogenic electromagnetic fields (EMF) influence the behaviour of bottomdwelling marine species. *Sci. Rep.* 10:4219. doi: 10.1038/s41598-020-60793-x

Ilyina, T., Zeebe, R., and Brewer, P. (2009). Changes in underwater sound propagation caused by ocean acidification. *IOP Conf. Series Earth Environ. Sci.* 6:462007. doi: 10.1088/1755-1307/6/46/462007

Irwin, R. P., Lin, S. Z., Long, R. T., and Paul, S. M. (1994). N-methyl-Daspartate induces a rapid, reversible, and calcium-dependent intracellular acidosis in cultured fetal rat hippocampal neurons. *J. Neurosci.* 14, 1352–1357. doi: 10. 1523/JNEUROSCI.14-03-01352.1994

Ishimatsu, A., Kojima, Y., and Hayashi, M. (2008). "Effects of CO2 ocean sequestration on deep-sea animals," in *Proceedings of the OCEANS 2008 – MTS/IEEE kobe techno-ocean*, Kobe, 1–4. doi: 10.1109/oceanskobe.2008.4531034

Jiahuan, R., Wenhao, S., Xiaofan, G., Wei, S., Shanjie, Z., Maolong, H., et al. (2018). Ocean acidification impairs foraging behavior by interfering with olfactory neural signal transduction in black sea bream. *Front. Physiol.* 9:1592. doi: 10.3389/fphys.2018.01592

Katz, B., and Miledi, R. (1965). The effect of temperature on the synaptic delay at the neuromuscular junction. *J. Physiol.* 181, 656–670.

Kelley, J. L., Chapuis, L., Davies, W. I. L., and Collin, S. P. (2018). Sensory system responses to human-induced environmental change. *Front. Ecol. Evol.* 6:95. doi: 10.3389/fevo.2018.00095

Kelley, J. L., Phillips, B., Cummins, G. H., and Shand, J. (2012). Changes in the visual environment affect colour signal brightness and shoaling behaviour in a freshwater fish. *Anim. Behav.* 83, 783–791.

Kemp, L., Xu, C., Depledge, J., Ebi, K. L., Gibbins, G., Kohler, T. A., et al. (2022). Climate endgame: Exploring catastrophic climate change scenarios. *Proc. Natl. Acad. Sci. U.S.A.* 119:e2108146119. doi: 10.1073/pnas.2108146119

Khan, A. M., Finlay, J. M., Clarke, P., Sol, K., Melendez, R., Judd, S., et al. (2021). Association between temperature exposure and cognition: A cross-sectional analysis of 20,687 aging adults in the United States. *BMC Public Health* 21:1484. doi: 10.1186/s12889-021-11533-x

Kharouba, H. M., Ehrlén, J., Gelman, A., Bolmgren, K., Allen, J. M., Travers, S. E., et al. (2018). Global shifts in the phenological synchrony of species interactions over recent decades. *Proc. Natl. Acad. Sci. U.S.A.* 115, 5211–5216. doi: 10.1073/pnas.1714511115

Kim, J. A., and Connors, B. W. (2012). High temperatures alter physiological properties of pyramidal cells and inhibitory interneurons in hippocampus. *Front. Cell. Neurosci.* 6:27. doi: 10.3389/fncel.2012.00027

Kunc, H. P., McLaughlin, K. E., and Schmidt, R. (2016). Aquatic noise pollution: Implications for individuals, populations, and ecosystems. *Proc. Biol. Sci.* 283:8. doi: 10.1098/rspb.2016.0839

Kushinsky, D., Morozova, E. O., and Marder, E. (2019). In vivo effects of temperature on the heart and pyloric rhythms in the crab, *Cancer borealis. J. Exp. Biol.* 222(Pt 5):jeb199190. doi: 10.1242/jeb.199190

Ladich, F., and Fay, R. R. (2013). Auditory evoked potential audiometry in fish. *Rev. Fish Biol. Fish.* 23, 317-364.

Lamas, J. A., Rueda-Ruzafa, L., and Herrera-Pérez, S. (2019). Ion channels and thermosensitivity: TRP, TREK, or Both. *Int. J. Mol. Sci.* 20:2371. doi: 10.3390/ ijms20102371

Leduc, A. O. H. C., Leduc, A. O. H., Munday, P. L., Brown, G. E., and Ferrari, M. C. O. (2013). Effects of acidification on olfactory-mediated behaviour in freshwater and marine ecosystems: A synthesis. *Philos. Trans. R. Soc. B Biol. Sci.* 368:20120447. doi: 10.1098/rstb.2012.0447

Lohr, B., Wright, T. F., and Dooling, R. J. (2003). Detection and discrimination of natural calls in masking noise by birds: Estimating the active space of a signal. *Anim. Behav.* 65, 763–777. doi: 10.1006/anbe.2003.2093

Lucke, K., Siebert, U., Lepper, P. A., and Blanchet, M.-A. (2009). Temporary shift in masked hearing thresholds in a harbor porpoise (*Phocoena phocoena*) after exposure to seismic airgun stimuli. *J. Acoust. Soc. Am.* 125, 4060–4070. doi: 10.1121/1.3117443

Lürling, M., and Scheffer, M. (2007). Info-disruption: Pollution and the transfer of chemical information between organisms. *Trends Ecol. Evol.* 22, 374–379. doi: 10.1016/j.tree.2007.04.002

Madeira, D., Andrade, J., Leal, M. C., Ferreira, V., Rocha, R. J. M., Rosa, R., et al. (2020). Synergistic effects of ocean warming and cyanide poisoning in an ornamental tropical reef fish. *Front. Mar. Sci.* 7:246. doi: 10.3389/fmars.2020.00246

Manisalidis, I., Stavropoulou, E., Stavropoulos, A., and Bezirtzoglou, E. (2020). Environmental and health impacts of air pollution: A review. *Front. Public Health* 8:14. doi: 10.3389/fpubh.2020.00014

Marder, E., and Rue, M. C. P. (2021). From the neuroscience of individual variability to climate change. J. Neurosci. 41, 10213–10221.

Meier, H. E. M., Markus Meier, H. E., Müller-Karulis, B., Andersson, H. C., Dieterich, C., Eilola, K., et al. (2012). Impact of climate change on ecological quality indicators and biogeochemical fluxes in the baltic sea: A multi-model ensemble study. *AMBIO* 41, 558–573. doi: 10.1007/s13280-012-0320

McDonald, J. W., Bhattacharyya, T., Sensi, S. L., Lobner, D., Ying, H. S., Canzoniero, L. M., et al. (1998). Extracellular acidity potentiates AMPA receptormediated cortical neuronal death. *J. Neurosci.* 18, 6290–6299.

Micheva, K. D., and Smith, S. J. (2005). Strong effects of subphysiological temperature on the function and plasticity of mammalian presynaptic terminals. *J. Neurosci.* 25, 7481–7488. doi: 10.1523/JNEUROSCI.1801-05.2005

Milazzo, M., Cattano, C., Alonzo, S. H., Foggo, A., Gristina, M., Rodolfo-Metalpa, R., et al. (2016). Ocean acidification affects fish spawning but not paternity at CO2 seeps. *Proc. Biol. Sci.* 283:20161021. doi: 10.1098/rspb.2016.1021

Miller, P. J., Biassoni, N., Samuels, A., and Tyack, P. L. (2000). Whale songs lengthen in response to sonar. *Nature* 405:903. doi: 10.1038/35016148

Moser, E., Mathiesen, I., and Andersen, P. (1993). Association between brain temperature and dentate field potentials in exploring and swimming rats. *Science* 259, 1324–1326. doi: 10.1126/science.8446900

Munday, P. L., Dixson, D. L., Donelson, J. M., Jones, G. P., Pratchett, M. S., Devitsina, G. V., et al. (2009). Ocean acidification impairs olfactory discrimination and homing ability of a marine fish. *Proc. Natl. Acad. Sci. U.S.A.* 106, 1848–1852. doi: 10.1073/pnas.0809996106

Munday, P. L., Dixson, D. L., McCormick, M. I., Meekan, M., Ferrari, M. C. O., and Chivers, D. P. (2010). Replenishment of fish populations is threatened by ocean acidification. *Proc. Natl. Acad. Sci. U.S.A.* 107, 12930–12934.

Musilova, Z., Salzburger, W., and Cortesi, F. (2021). The visual opsin gene repertoires of teleost fishes: Evolution. Ecology, and Function. *Annu. Rev. Cell Dev. Biol.* 37, 441–468.

Nachtigall, P. E., Supin, A. Y., Amundin, M., Röken, B., Møller, T., Mooney, T. A., et al. (2007). Polar bear *Ursus maritimus* hearing measured with auditory evoked potentials. *J. Exp. Biol.* 210, 1116–1122. doi: 10.1242/jeb.02734

Nachtigall, P. E., Supin, A. Y., Pawloski, J., and Au, W. W. L. (2004). Temporary threshold shifts after noise exposure in the bottlenose dolphin (*Tursiops truncatus*) measured using evoked auditory potentials. *Mar. Mamm. Sci.* 20, 673–687. doi: 10.1111/j.1748-7692.2004.tb01187.x

Nagelkerken, I., Alemany, T., Anquetin, J. M., Ferreira, C. M., Ludwig, K. E., Sasaki, M., et al. (2021). Ocean acidification boosts reproduction in fish via indirect effects. *PLoS Biol.* 19:e3001033. doi: 10.1371/journal.pbio.3001033

Nieman, C. L., Oppliger, A. L., McElwain, C. C., and Gray, S. M. (2018). Visual detection thresholds in two trophically distinct fishes are compromised in algal compared to sedimentary turbidity. *Conserv. Physiol.* 6:coy044. doi: 10.1093/ conphys/coy044

Nilsson, G. E., Dixson, D. L., Domenici, P., McCormick, M. I., Sørensen, C., Watson, S.-A., et al. (2012). Near-future carbon dioxide levels alter fish behaviour by interfering with neurotransmitter function. *Nat. Clim. Change* 2, 201–204. doi: 10.1038/nclimate1352

O'Donnell, S. (2018). The neurobiology of climate change. Sci. Nat. 105:11. doi: 10.1007/s00114-017-1538-5

OuYang, Y. B., Mellergård, P., Kristián, T., Kristiánova, V., and Siesjö, B. K. (1994). Influence of acid-base changes on the intracellular calcium concentration of neurons in primary culture. *Exp. Brain Res.* 101, 265–271. doi: 10.1007/BF00228746

Park, H. J., Rajbhandari, I., Yang, H. S., Lee, S., Cucoranu, D., Cooper, D. S., et al. (2010). Neuronal expression of sodium/bicarbonate cotransporter NBCn1 (SLC4A7) and its response to chronic metabolic acidosis. *Am. J. Physiol. Cell Physiol.* 298, C1018–C1028. doi: 10.1152/ajpcell.00492.2009

Park, R. J., Behrer, A. P., and Goodman, J. (2021). Learning is inhibited by heat exposure, both internationally and within the United States. *Nat. Hum. Behav.* 5, 19–27.

Parkinson, R. H., Fecher, C., and Gray, J. R. (2022). Chronic exposure to insecticides impairs honeybee optomotor behaviour. *Front. Insect Sci.* 2:936826. doi: 10.3389/finsc.2022.936826

Perry, A. L., Low, P. J., Ellis, J. R., and Reynolds, J. D. (2005). Climate change and distribution shifts in marine fishes. *Science* 308, 1912–1915. Peters, R. C., Eeuwes, L. B. M., and Bretschneider, F. (2007). On the electrodetection threshold of aquatic vertebrates with ampullary or mucous gland electroreceptor organs. *Biol. Rev. Camb. Philos. Soc.* 82, 361–373.

Pick, U., and Karlish, S. J. (1982). Regulation of the conformation transition in the Ca-ATPase from sarcoplasmic reticulum by pH, temperature, and calcium ions. *J. Biol. Chem.* 257, 6120–6126.

Piniak, W. E. D., Dow Piniak, W. E., Mann, D. A., Eckert, S. A., and Harms, C. A. (2012). Amphibious hearing in sea turtles. *Adv. Exp. Med. Biol.* 730, 83–87. doi: 10.1007/978-1-4419-7311-5_18

Piniak, W. E. D., Mann, D. A., Harms, C. A., Todd Jones, T., and Eckert, S. A. (2016). Hearing in the juvenile green sea turtle (*Chelonia mydas*): A comparison of underwater and aerial hearing using auditory evoked potentials. *PLoS One* 11:e0159711. doi: 10.1371/journal.pone.0159711

Pinsky, M. L., Worm, B., Fogarty, M. J., Sarmiento, J. L., and Levin, S. A. (2013). Marine taxa track local climate velocities. *Science* 341, 1239–1242. doi: 10.1126/science.1239352

Porteus, C. S., Roggatz, C. C., Velez, Z., Hardege, J. D., and Hubbard, P. C. (2021). Acidification can directly affect olfaction in marine organisms. *J. Exp. Biol.* 224;jeb237941. doi: 10.1242/jeb.237941

Pouca, C. V., Gervais, C., Reed, J., Michard, J., and Brown, C. (2019). Quantity discrimination in Port Jackson sharks incubated under elevated temperatures. *Behav. Ecol. Sociobiol.* 73:93. doi: 10.1007/s00265-019-2706-8

Radchuk, V., Reed, T., Teplitsky, C., van de Pol, M., Charmantier, A., Hassall, C., et al. (2019). Adaptive responses of animals to climate change are most likely insufficient. *Nat. Commun.* 10:3109.

Rey, S., Huntingford, F. A., Boltaña, S., Vargas, R., Knowles, T. G., and Mackenzie, S. (2015). Fish can show emotional fever: Stress-induced hyperthermia in zebrafish. *Proc. Biol. Sci.* 282:20152266. doi: 10.1098/rspb.2015.2266

Rhen, T., and Schroeder, A. L. (2017). The genetics of thermosensitive sex determination. *Temperature (Austin)* 4, 109-111.

Ríos-Chelén, A. A., Quirós-Guerrero, E., Gil, D., and Garcia, C. M. (2013). Dealing with urban noise: Vermilion flycatchers sing longer songs in noisier territories. *Behav. Ecol. Sociobiol.* 67, 145–152. doi: 10.1007/s00265-012-1434-0

Robertson, R. M., and Money, T. G. A. (2012). Temperature and neuronal circuit function: Compensation, tuning and tolerance. *Curr. Opin. Neurobiol.* 22, 724–734. doi: 10.1016/j.conb.2012.01.008

Román-Palacios, C., and Wiens, J. J. (2020). Recent responses to climate change reveal the drivers of species extinction and survival. *Proc. Natl. Acad. Sci. U.S.A.* 117, 4211–4217. doi: 10.1073/pnas.1913007117

Ruffin, V. A., Salameh, A. I., Boron, W. F., and Parker, M. D. (2014). Intracellular pH regulation by acid-base transporters in mammalian neurons. *Front. Physiol.* 5:43. doi: 10.3389/fphys.2014.00043

Savoca, M. S., and Nevitt, G. A. (2014). Evidence that dimethyl sulfide facilitates a tritrophic mutualism between marine primary producers and top predators. *Proc. Natl. Acad. Sci. U.S.A.* 111, 4157–4161. doi: 10.1073/pnas.1317120111

Savoca, M. S., Wohlfeil, M. E., Ebeler, S. E., and Nevitt, G. A. (2016). Marine plastic debris emits a keystone infochemical for olfactory foraging seabirds. *Sci. Adv.* 2:e1600395.

Schulte, P. M. (2015). The effects of temperature on aerobic metabolism: Towards a mechanistic understanding of the responses of ectotherms to a changing environment. J. Exp. Biol. 218, 1856–1866. doi: 10.1242/jeb.118851

Slabbekoorn, H., and Halfwerk, W. (2009). Behavioural ecology: Noise annoys at community level. *Curr. Biol.* 19, R693–R695. doi: 10.1016/j.cub.2009.0 7.002

Sokolova, I. M., and Lannig, G. (2008). Interactive effects of metal pollution and temperature on metabolism in aquatic ectotherms: Implications of global climate change. *Clim. Res.* 37, 181–201. doi: 10.3354/cr00764

Soofi, W., Goeritz, M. L., Kispersky, T. J., Prinz, A. A., Marder, E., and Stein, W. (2014). Phase maintenance in a rhythmic motor pattern during temperature changes in vivo. *J. Neurophysiol.* 111, 2603–2613. doi: 10.1152/jn.00906. 2013

Soravia, C., Ashton, B. J., Thornton, A., and Ridley, A. R. (2021). The impacts of heat stress on animal cognition: Implications for adaptation to a changing climate. *WIREs Clim. Change* 12:e713. doi: 10.1002/wcc.713

Städele, C., Heigele, S., and Stein, W. (2015). Neuromodulation to the rescue: Compensation of temperature-induced breakdown of rhythmic motor patterns via extrinsic neuromodulatory input. *PLoS Biol.* 13:e1002265. doi: 10.1371/journal. pbio.1002265 Steffen, W., Rockström, J., Richardson, K., Lenton, T. M., Folke, C., Liverman, D., et al. (2018). Trajectories of the earth system in the anthropocene. *Proc. Natl. Acad. Sci. U.S.A.* 115, 8252–8259.

Stein, W., and Harzsch, S. (2021). The neurobiology of ocean change - insights from decapod crustaceans. *Zoology* 144:125887. doi: 10.1016/j.zool.2020.125887

Sulukan, E., Baran, A., Şenol, O., Yildirim, S., Mavi, A., Ceyhun, H. A., et al. (2022). The synergic toxicity of temperature increases and nanopolystrene on zebrafish brain implies that global warming may worsen the current risk based on plastic debris. *Sci. Total Environ.* 808:152092. doi: 10.1016/j.scitotenv.2021.152092

Sunada, H., Riaz, H., de Freitas, E., Lukowiak, K., Swinton, C., Swinton, E., et al. (2016). Heat stress enhances LTM formation in *Lymnaea*: Role of HSPs and DNA methylation. *J. Exp. Biol.* 219, 1337–1345. doi: 10.1242/jeb.134296

Suriyampola, P. S., Cacéres, J., and Martins, E. P. (2018). Effects of short-term turbidity on sensory preference and behaviour of adult fish. *Anim. Behav.* 146, 105–111. doi: 10.1016/j.anbehav.2018.10.014

Tang, L. S., Goeritz, M. L., Caplan, J. S., Taylor, A. L., Fisek, M., and Marder, E. (2010). Precise temperature compensation of phase in a rhythmic motor pattern. *PLoS Biol.* 8:e1000469. doi: 10.1371/journal.pbio.1000469

Tang, L. S., Taylor, A. L., Rinberg, A., and Marder, E. (2012). Robustness of a rhythmic circuit to short- and long-term temperature changes. *J. Neurosci.* 32, 10075–10085. doi: 10.1523/JNEUROSCI.1443-12.2012

Taylor, L., Watkins, S. L., Marshall, H., Dascombe, B. J., and Foster, J. (2016). The impact of different environmental conditions on cognitive function: A focused review. *Front. Physiol.* 6:372. doi: 10.3389/fphys.2015.00372

Thimmegowda, G. G., Mullen, S., Sottilare, K., Sharma, A., Mohanta, S. S., Brockmann, A., et al. (2020). A field-based quantitative analysis of sublethal effects of air pollution on pollinators. *Proc. Natl. Acad. Sci. U.S.A.* 117, 20653–20661. doi: 10.1073/pnas.2009074117

Thomson, E. M., Filiatreault, A., and Guénette, J. (2019). Stress hormones as potential mediators of air pollutant effects on the brain: Rapid induction of glucocorticoid-responsive genes. *Environ. Res.* 178:108717. doi: 10.1016/j.envres. 2019.108717

Tombaugh, G. C., and Somjen, G. G. (1996). Effects of extracellular pH on voltage-gated Na+. K+ and Ca2+ currents in isolated rat CA1 neurons. *J. Physiol.* 493(Pt 3), 719–732. doi: 10.1113/jphysiol.1996.sp021417

Toni, M., Angiulli, E., Miccoli, G., Cioni, C., Alleva, E., Frabetti, F., et al. (2019). Environmental temperature variation affects brain protein expression and cognitive abilities in adult zebrafish (*Danio rerio*): A proteomic and behavioural study. *J. Proteom.* 204:103396. doi: 10.1016/j.jprot.2019.103396

Tresguerres, M., and Hamilton, T. J. (2017). Acid-base physiology, neurobiology and behaviour in relation to CO-induced ocean acidification. *J. Exp. Biol.* 220, 2136–2148. doi: 10.1242/jeb.144113

Troyer, R. R., and Turner, A. M. (2015). Chemosensory perception of predators by larval amphibians depends on water quality. *PLoS One* 10:e0131516. doi: 10. 1371/journal.pone.0131516

Truscott, Z., Booth, D. T., and Limpus, C. J. (2017). The effect of on-shore light pollution on sea-turtle hatchlings commencing their off-shore swim. *Wildlife Res.* 44:127. doi: 10.1071/wr16143

Tuomainen, U., and Candolin, U. (2011). Behavioural responses to humaninduced environmental change. *Biol. Rev. Camb. Philos. Soc.* 86, 640–657.

Van Colen, C., Vanhove, B., Diem, A., and Moens, T. (2020). Does microplastic ingestion by zooplankton affect predator-prey interactions? An experimental study on larviphagy. *Environ. Pollut.* 256:113479. doi: 10.1016/j.envpol.2019. 113479

Van de Meutter, F., De Meester, L., and Stoks, R. (2005). Water turbidity affects predator-prey interactions in a fish-damselfly system. *Oecologia* 144, 327–336. doi: 10.1007/s00442-005-0050-3

Vasconcelos, R. O., Amorim, M. C. P., and Ladich, F. (2007). Effects of ship noise on the detectability of communication signals in the Lusitanian toadfish. *J. Exp. Biol.* 210, 2104–2112. doi: 10.1242/jeb.004317

Vornanen, M. (2017). Electrical excitability of the fish heart and its autonomic regulation. *Fish Physiol.* 36, 99–153. doi: 10.1016/bs.fp.2017.04.002

Wang, Y., and Wang, T. (2021). "Behavioral impacts of ocean acidification on marine animals," in *Ocean acidification and marine wildlife*, ed. G. Liu (Amsterdam: Academic Press), 109–153. doi: 10.1016/b978-0-12-822330-7. 00002-2

Warren, P. S., Katti, M., Ermann, M., and Brazel, A. (2006). Urban bioacoustics: It's not just noise. *Animal Behaviour* 71, 491–502. doi: 10.1016/j.anbehav.2005. 07.014 Weilgart, L. S. (2007). The impacts of anthropogenic ocean noise on cetaceans and implications for management. *Can. J. Zool.* 85, 1091–1116. doi: 10.1139/z07-101

Weis, J. S., Samson, J., Zhou, T., Skurnick, J., and Weis, P. (2001). Prey capture ability of mummichogs (*Fundulus heteroclitus*) as a behavioral biomarker for contaminants in estuarine systems. *Can. J. Fish. Aqua. Sci.* 58, 1442–1452. doi: 10.1139/f01-086

Weis, J. S., Smith, G. M., and Zhou, T. (1999). Altered predator/prey behavior in polluted environments: Implications for fish conservation. *Environ. Biol. Fish* 55, 43–51. doi: 10.1023/a:1007496528012

Weissbourd, B., Momose, T., Nair, A., Kennedy, A., Hunt, B., and Anderson, D. J. (2021). A genetically tractable jellyfish model for systems

and evolutionary neuroscience. Cell 184, 5854.e-5868.e. doi: 10.1016/j.cell.2021. 10.021

Whitlock, K. E., and Palominos, M. F. (2022). The olfactory tract: Basis for future evolution in response to rapidly changing ecological niches. *Front. Neuroanat.* 16:831602. doi: 10.3389/fnana.2022.83 1602

Wolosker, H., Rocha, J. B., Engelender, S., Panizzutti, R., De Miranda, J., and de Meis, L. (1997). Sarco/endoplasmic reticulum Ca2+-ATPase isoforms: Diverse responses to acidosis. *Biochem. J.* 321:545.

Zollinger, S. A., Goller, F., and Brumm, H. (2011). Metabolic and respiratory costs of increasing song amplitude in zebra finches. *PLoS One* 6:e23198. doi: 10.1371/journal.pone.0023198