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*CORRESPONDENCE Xiaoyan Wu Wuxiaoyan2k6@whu.edu.cn Lihua Ni Ni nilihua2016@163.com

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The roles of heme oxygenase-1 in renal disease

Hongfu Zhai¹, Lihua Ni^{1*} and Xiaoyan Wu^{1,2*}

¹Department of Nephrology, Zhongnan Hospital of Wuhan University, Wuhan, China, ²Department of General Practice, Zhongnan Hospital of Wuhan University, Wuhan, China

Heme oxygenase (HO), a heat shock protein containing hemoglobin, is an important enzyme in heme catabolism. It is involved in cell homeostasis and has anti-inflammatory, antioxidant, anti-apoptosis, immunomodulation, and other functions. It is expressed at a modest level in most normal tissues. When the body suffers from ischemia hypoxia, injury, toxins, and other nociceptive stimuli, the expression increases, which can transform the oxidative microenvironment into an antioxidant environment to promote tissue recovery from damage. In recent years, research has continued to verify its value in a variety of human bodily systems. It is also regarded as a key target for the treatment of numerous disorders. With the advancement of studies, its significance in renal disease has gained increasing attention. It is thought to have a significant protective function in preventing acute kidney injury and delaying the progression of chronic renal diseases. Its protective mechanisms include anti-inflammatory, antioxidant, cell cycle regulation, apoptosis inhibition, hemodynamic regulation, and other aspects, which have been demonstrated in diverse animal models. Furthermore, as a protective factor, its potential therapeutic efficacy in renal disease has recently become a hot area of research. Although a large number of preclinical trials have confirmed its therapeutic potential in reducing kidney injury, due to the problems and side effects of HO-1 induction therapy, its efficacy and safety in clinical application need to be further explored. In this review, we summarize the current state of research on the mechanism, location, and treatment of HO and its relationship with various renal diseases.

KEYWORDS

heme oxygenase, renal disease, mechanism, value, therapeutic

1 Overview of HO

HO is an important enzyme in heme degradation and metabolism, and can decompose heme into three metabolites: biliverdin IX, CO, and Fe^{2+} . It includes three distinct active subtypes, namely oxygen stress inducible (HO-1), constitutive heme oxygenase (HO-2) and HO-3, which have not yet been fully described, and was first found by Tenhunen and his colleagues in 1968 (1). Among these subtypes, HO-1 has the most biological activity and is the most frequently studied. It plays a significant role in cell stress and the maintenance of cell

homeostasis, which is linked to numerous clinical disease states (2). Heavy metals, endotoxin, UV, cytokines, growth hormones, and other chemical and physical stimuli can stimulate HO-1 expression, which in turn promotes the generation of antioxidants and anti-apoptosis molecules to protect cells. It also promotes injury repair through immunity, regulation of cell autophagy, inhibition of apoptosis, and the regulation of cell cycle and other pathways. Nath and colleagues first found its kidney-protective effect in a rat model of glycerolinduced hemolytic illness in 1992 (3). They discovered that prior hemoglobin injections promptly increased the expression of HO-1 mRNA and protein in the kidney, delaying the progression of renal failure. Since then, Zager and colleagues have shown that HO-1 has a kidney protective effect in a variety of AKI models, including ischemia/ reperfusion, glycerol-induced rhabdomyolysis, cisplatin nephrotoxicity, and bilateral ureteral obstruction models (4). They also discovered that blood and urine HO-1 levels rise with the severity of acute renal injury, and the degree of kidney impairment is reduced compared to HO-1 deficiency. Later research has gradually clarified its action mechanism and proved its utility in other chronic kidney illnesses such as diabetic nephropathy, hypertensive nephropathy, glomerular nephritis, and renal transplantation.

2 The roles of HO-1

HO-1 expression in renal cells is induced by ischemia, hypoxia, or stress. It has anti-inflammatory and anti-oxidative functions, modulates cell cycle, inhibits apoptosis, regulates autophagy, influences hemodynamics, and regulates immunity, thus promoting the recovery of kidney tissues and cells from injury (Figure 1).

2.1 Anti-inflammatory and anti-oxidative stress

The anti-inflammatory action of HO-1 has been supported in various inflammatory models. On the one hand, HO-1 and its byproducts can inhibit the production of pro-inflammatory cytokines such as IL-1, IL-6, IL-12, IL-1 β , and TNF by activated macrophages, thus playing a role in the macrophage-targeted inflammatory regression stage (5). Up-regulation of HO-1 expression, on the other hand, can limit NF-kB activation and constrain the expression of inflammatory mediators COX-2 and NOS, thus decreasing the generation of COX-2derived prostaglandin E2 and NOS-derived NO to diminish inflammatory mediator production (6–8). HO-1 knockout mice showed leukocyte macrophage increase, hepatosplenomegaly, and renal tubulointerstitial damage after kidney damage, more inflammatory cell infiltration, and fibrosis in comparison to wild mice accentuated HO-1's anti-inflammatory action (9).

HO-1 and its metabolites catalyzing the decomposition of heme can play an anti-oxidative role in many ways (10, 11). On the one hand, Heme induces the production of hydrogen peroxide in renal tubular epithelial cells, causing oxidative stress and the production of a large amount of ROS by amplifying lipid peroxidation stress. Secondly, heme inhibits the action of proteasomes such as glutathione reductase, resulting in cell damage and apoptosis. HO-1 inhibits the deleterious effects of heme. This benefit is confirmed by increased heme content and aggravated kidney injury in HO-1 deficient mice (12). Secondly, HO-1 can also reduce the accumulation of intracellular iron, thus reducing the production of oxygen free radicals and peroxide free radicals in cells to inhibit oxidative stress (13). Heme metabolites such as bilirubin and CO have also been shown to inhibit protein phosphorylation and PKC activity, inhibit oxidation of peroxide radicals caused by chemicals, and reduce the production of lipid peroxidation products in tissues and organs, thereby alleviating oxidative stress (14). Bilirubin, in particular, plays an important antioxidant and cytoprotective role in weakening DNA damage and cell death mediated by Ang II (15). CO also promotes the activation of antioxidants such as superoxide dismutase (SOD), NFkB, and heat shock protein 70 (Hsp70). KD Poss discovered that mice lacking functioning HO-1 were substantially more vulnerable to oxidative stress injury caused by hydrogen peroxide. Anemia resulting from endotoxin injection and the infiltration and fibrosis of renal interstitial inflammatory cells were more visible than in normal mice (10, 16). This also demonstrated that HO-1 upregulation was an adaptive mechanism for protecting cells from stress damage.

Most disease processes are accompanied by inflammation and oxidative stress, so the anti-oxidative stress of HO-1 plays a major role in the protective mechanism of the kidney. Animal studies on renal ischemia-reperfusion injury have shown that the expression of inflammatory mediators such as NF- κ B, MCP-1, TGF- β , and other inflammatory mediators is more obvious in HO-1 deficient mice, and the kidney injury is more serious (17), which also indicated that HO-1 could alleviate inflammatory reaction and improve renal tubular injury caused by stress.

2.2 Regulation of cell cycle

HO-1 regulates cell cycle progression. The cell cycle is divided into four stages: pre-DNA synthesis (G1), DNA synthesis (S), anaphase (G2), and mitotic phase (M). The progression from one phase to the next is tightly controlled by the production of certain cyclins and the cycle-dependent protein kinase (CDK), which ultimately determines whether the cell cycle is arrested or not. P21 is a member of the Cip/Kip family of CDK inhibitors. Nath and his colleagues were the first to identify the direct regulation of HO-1 on P21 expression on kidney (3): HO-1 regulates P21, limiting target protein phosphorylation and blocking cell cycle progression from G1 to S phase. In addition, CO and iron, the by-products of heme metabolism, also play a role in renal tubular injury and repair through P21-mediated cell cycle arrest. The regulatory effect of HO-1 on P21 has also been found in mesangial cells: HO-1 induced p21 expression in mesangial cells, whereas HO-1 inhibitors counteracted this effect (18).

2.3 Inhibiting apoptosis

Heme inhibits the proteasome and causes mitochondrial malfunction, which results in cell damage and apoptosis. HO-1 counteracts this impact by decreasing heme aggregation. HO-1 also inhibits the expression of pro-apoptosis genes like Bak while

increasing the expression of anti-apoptosis genes like bcl-2 (19). CO can also prevent apoptosis in fibroblasts and endothelial cells through modulating the MAPK pathway and activating NF-KB in vascular smooth muscle cells, according to recent research (20, 21).

2.4 Regulating autophagy

Autophagy is a complex process in which damaged cell structures are transported to the lysosome for destruction and recycling. Autophagy can develop stress adaption and maintain intracellular homeostasis under stress situations such as acute injury and ischemia. Improper autophagy, on the other hand, may contribute to tissue injury. A previous study has shown that HO-1 is an efficient autophagy regulator (22). On the one hand, HO-1 inhibits the production of beclin, a crucial protein involved in autophagy initiation. HO-1 deficient mice showed enhanced autophagy in the proximal tubules, as shown by increased autophagic vesicles and beclin expression in the proximal tubules. On the other hand, a delayed autophagy response may be attributed to the decrease of ROS and heme.

2.5 Affect hemodynamics

HO-1 inhibits the level of vasoconstrictor molecules in a tissuespecific manner, regulates the expression of anti-angiogenic CXCL-

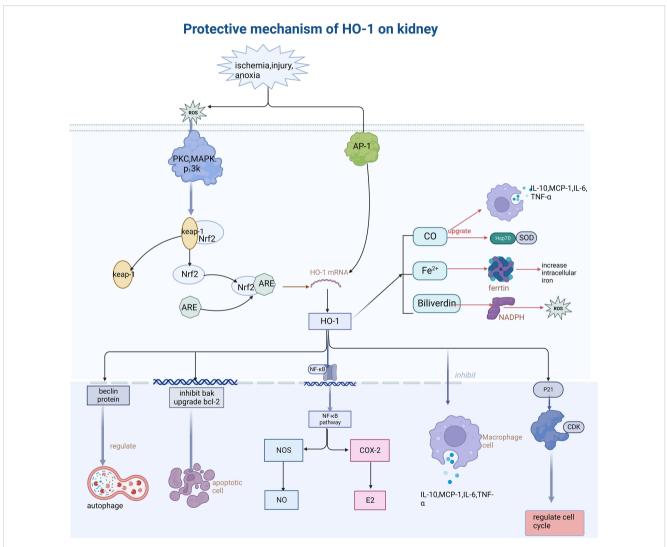


FIGURE 1

Describes how oxidative stress injury induced HO-1 expression and its function mechanism. Stimulation such as ischemia, anoxic, and injury leads to the generation of ROS, further activates PKC and MAPK protein kinases, dissociates Nrf2 from keap-1, and binds the activated Nrf2 to ARE protein, thus inducing the increase of HO-1 expression. HO-1 promotes the body to recover from injury through various mechanisms such as anti-inflammation, antioxidative stress, cell cycle regulation, anti-apoptosis, etc. (This article introduces its role and mechanism in detail). Secondly, its heme-decomposing products CO, Fe^{2+} , and biliverdin also play a role in cell protection by regulating antioxidants, inhibiting iron accumulation in cells, and regulating the secretion of inflammatory mediators. PKC, protein kinase C, MAPK, mitogen- activated protein-1; NF - kB, nuclear factor-k-gene binding; CDK, Cyclin dependent kinase; ROS, reactive oxygen species; NOS, nitric oxide synthase; COX-2, cyclooxygenase-2.

10, and may alter the key balance between angiogenesis and antiangiogenesis factors (23). CO also functions as a physiological regulator of vascular tension *via* cGMP-mediated artery response, activating calcium channels and expanding smaller renal arteries. Further study speculated that CO not only relaxes vascular smooth muscles, but also regulates the proliferation and death of smooth muscle cells and may play a role in blood pressure regulation of acute hypertension, which is critical for maintaining the renal microvascular system following damage.

In conclusion, HO-1 alleviates kidney injury after stress injury through the above mechanisms. Recent investigations on kidney transplantation have proven its immune-regulatory effects: HO-1 inhibits the proliferation of T cells and promotes the differentiation of T cells into $CD^{4+} CD^{25+}$ regulatory T cells. It may also participate in the changes in the proportion of immune-related cytokines, and is a key suppressor of B cells and NK cells, which, in conjunction with the foregoing methods, play a role in alleviating kidney damage (24).

3 HO-1 in different renal cells

HO-1 is expressed differentially in different regions of the kidney, and its functions are outlined in Table 1. The mechanism in different kidney cells can be seen in Figure 2.

Renal tubular epithelial cells, especially proximal tubules, are particularly sensitive to ischemia-reperfusion injury and heme due to the presence of TLR4 (a heme receptor), and the ability to express HO-1 is also the strongest (25). HO-1 regulates constitutive expression of TLR4 in the proximal tubule. It can inhibit autophagy and reduce reactive oxygen species' accumulation and ferroptosis in renal tubular epithelial cells. Therefore HO-1 can be employed as a key marker to indicate renal tubular damage when the body is severely harmed beyond its adaptive capacity. This is supported by the observation that the renal tubular epithelium is most severely damaged in the HO-1 deficient mouse model of intravascular hemolysis and rhabdomyolysis.

The second is the monocyte-macrophage system, which includes macrophages, dendritic cells, and monocytes (26, 27). Agarwal studied Lewis rats with kidney transplantation and found a significant increase in HO mRNA and protein expression in renal allografts on day 5 after transplantation (28). And the expression of HO protein was most pronounced in the graft's monocytes, of

which more than 80% were macrophages. Their study confirmed the ability of macrophages in the kidney to actively express HO-1 under inflammatory conditions, especially in acute allograft rejection.

Because of their filtering function, renal endothelial cells are the first to be affected by various stressors. Up-regulation of HO-1 in endothelial cells can not only regulate complement activation and thrombomodulin expression, but also inhibit heme-mediated proinflammatory and prothrombotic effects, thereby reducing endothelial injury (29, 30). Endothelial cells in different parts of the kidney, however, differ in their ability to induce HO-1 expression: current evidence suggests that macrovascular endothelial cells are more likely to induce HO-1 upregulation than glomerular endothelial cells, while peritubular capillaries have received little attention (35). Yachie showed that HO-1 is critical for endothelial cell survival by studying the first human example of HO-1 deficiency (31). They found a complete absence of HO-1 production in the tissues of the patient. Glomerular electron microscopy showed glomerular endothelial cell detachment, a large amount of dense material deposited under the endothelium, and severe and persistent endothelial damage in the kidney.

Podocytes are also an important part of the glomerular filtration barrier, which has a filtration function and is particularly sensitive to various damage. The role of HO-1 in podocytes was demonstrated in studies of diabetic rats (32). Vimentin, desmin, and antioxidant proteins are elevated in podocytes during oxidative stress injury, and further upregulates Nrf2 to increase HO-1 production. Heme also stimulated the expression of HO-1 in podocytes in a Nrf2-dependent manner (33). This effect was specifically enhanced in diabetic rat podocytes, and the same results were observed in high-glucose-treated podocytes, implying that this enhancement was associated with the increase of AGEs (34). In glomeruli isolated from diabetic rats, it was observed that the content and activity of HO-1 were significantly increased, the ratio of Bax/Bcl-2 was also increased, and the number of apoptotic cells was obviously increased. Such changes were not observed in Nrf2-deficient rats.

However, the ability of the glomerulus to induce HO-1 is very limited, and infiltrating macrophages rather than glomerular cells are the principal producers of HO-1 in the glomerulus. Mosley investigated heterologous (HNTN) and accelerated (ANTN) nephrotoxic nephritis in Lewis rats, and discovered that HO-1 was not detected by immunohistochemistry in glomerular cells of

TABLE 1 The	roles	of	HO-1	in	renal	cells.
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Cells	Function	Model	References
Renal tubular epithelial cells	Inhibits autophagy and apoptosis, regulates cell cycle	intravascular hemolysis mice	(25)
The macrophage system	Anti-inflammatory and anti-oxidative stress	Rat allogeneic kidney transplant, diabetic nephropathy mice	(26–28)
Endothelium	Anti-oxidative stress, reduces endothelial injury	Human cases lacking HO-1 HO-1 knockout mice	(29–31)
Podocytes	Adjusts Bax/Bcl - 2 protein ratio, inhibits apoptosis	Diabetic nephropathy rats High glucose podocytes	(32-34)
Glomerular cells	Affects hemodynamics	Glomerulonephritis rats	(35)

damaged tissue but was present in cells with macrophage morphology (36).

In short, the expression of HO-1 in the kidneys is highly variable. Renal tubular epithelial cells have the strongest ability to express HO-1 in AKI, and the monocyte-macrophage system cannot be ignored. HO in podocytes is significantly increased in diabetic nephropathy, while its expression in glomeruli is less. However, the reason for its differential expression is still unclear, and it is speculated that it may be related to the site of kidney injury, which needs further exploration.

4 Study on the relationship between HO-1 and kidney diseases

The role of HO-1 in kidney disease was first discovered in hemolytic diseases and rhabdomyolysis-associated kidney injury, which was thought to be related to the large release of free hemoglobin. Subsequent studies gradually confirmed its beneficial value in other non-blood-mediated kidney disease (37, 38).

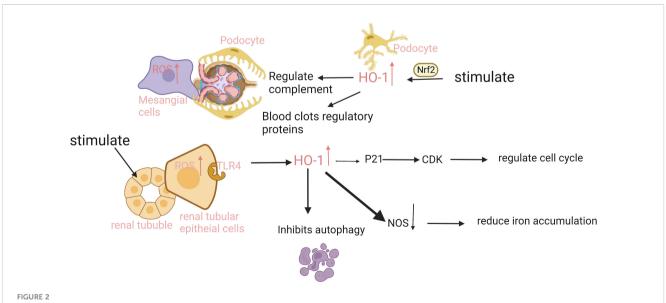
4.1 Acute kidney injury

The role of HO-1 in AKI was first discovered by Nath et al. through a glycerol-induced AKI model; they found that a single prior injection of the inducer rapidly induced HO-1 mRNA and protein expression in the kidney, preventing the progression to ARF (3). Follow-up studies were performed in a variety of different models of AKI, such as ischemia-reperfusion injury, hemolytic disease, rhabdomyolysis, cisplatin, and lipopolysaccharide (LPS). The results showed that various stress factors could induce HO-1 RNA and protein expression, which repeatedly confirmed the protective effect of HO-1 on AKI (39).

M J Tracz established a renal IRI model by clamping the renal artery. In this model, it was found that the expression of HO-1 in the kidney, especially in the renal tubules, was significantly increased after renal ischemia-reperfusion injury (23). The renal blood flow of HO-1 knockout mice (-/-) was much lower than that of wild mice at 4 hours after ischemia, and the level of IL-6 was also significantly increased. Subsequent studies further clarified the mechanisms of the protective effects including regulating NO synthesis and hemodynamics, inhibiting apoptosis, antiinflammation and antioxidant stress factors, etc (40). Shimizu et al. also found a significant increase in HO-1expression in bilateral renal artery ligation-reperfusion rat, and this change was earlier than the increase of serum creatinine concentration (41). However, pretreatment with Sn-MP, a HO-specific competitive inhibitor, aggravated the damage of HO-1 renal tubular epithelial cells in rats.

Rubio-Navarro established a mouse kidney injury model of intravascular hemolytic by injecting the hemolytic drug PHZ. It was found that KIM-1 and NGAL, markers related to renal tubular injury, and HO-1 expression were significantly increased after hemolysis. While Nrf2 deficiency mice did not display an increase in HO-1 expression, KIM-1 and NGAL were higher, and there was extensive proximal tubular necrosis (42). This indicates that HO-1 expression was up-regulated in renal tubules during hemolytic injury and this up-regulation was mediated by Nrf2.

Acute kidney injury caused by trauma-induced rhabdomyolysis, especially crush syndrome, causes a large number of deaths in the ICU. The mechanism of injury includes renal vasoconstriction,



The expression and function of HO-1 is different in different parts of the kidney. The renal tubular epithelial cells have the strongest ability to express HO-1. When injury occurs, ROS in cells increases, and heme binds to the TLR4 receptor on tubule epithelium to promote HO-1 expression. HO-1 upregulates CDK inhibitor P21, reduces tubular cell apoptosis, and inhibits proximal tubular autophagy. In addition, injury can also induce HO-1 expression in podocytes, mesangial cells, and endothelial cells in an Nrf2-dependent manner, regulate complement activation and thrombomodulin, mediate inflammation and thrombosis, and protect cells. ROS, reactive oxygen species; TLR4, Toll-like receptor 4; CDK, Cyclin dependent kinase; NOS, nitric oxide synthase. oxidative stress, myoglobin participation, and so on. Zager and Johnson found that the contents of TNF- α , MCP-1, and TGF- β 1 mRNA increased progressively after glycerol was induced in a mouse model of glycerol-induced rhabdomyolysis. HO-1 increased significantly and reached the maximum after 48 hours of injection (4). The mortality of the mice whose expression of HO-1 was enhanced by a single infusion of hemoglobin was significantly lower than that of the control group. Mice with HO-1 deficiency, meanwhile, were prone to fulminant irreversible AKI, and the mortality rate was almost 100%. The increase of HO-1 expression after rhabdomyolysis-induced AKI can improve renal microcirculation, regulate oxidative stress, protect the kidney, and reduce the risk of death (43).

4.2 Sepsis-related renal injury

Sepsis-associated kidney injury is a common renal complication with a high mortality rate in severe patients in the ICU. The beneficial role of HO-1 in sepsis-associated AKI has been gradually clarified in recent years (44, 45). Chung SW et al. (45) established a model of polymicrobial sepsis in HO-1 gene deficient mice and wild-type mice, and found that HO-1 expression increased after sepsis. However, it does not inhibit the generation and accumulation of inflammatory cells, but enhances bacterial clearance by increasing bacterial phagocytosis and the body's antibacterial response. However, HO-1 gene-deficient mice showed more reduced regional renal blood perfusion and showed higher mortality. Injection of CO-releasing molecules into these mice can increase phagocytosis and alleviate kidney injury in HO-1 deficient mice, which may provide new ideas for further treatment.

4.3 Diabetic kidney disease

Diabetic kidney disease (DKD) is one of the major causes of chronic kidney disease and end-stage renal diseases. Previous studies have confirmed the role of HO-1 in high glucose environments (46, 47). A high glucose environment increases the expression of HO-1 in podocytes and up-regulates extracellular SOD and NOS, thus leading to endothelial relaxation and reduction of ROS to affect renal hemodynamics. Inhibition of HO-1 by HO-1 inhibitors such as ZnPP showed an increase in the number of apoptotic cells, which also indicated the protective effect of HO-1. Ahmed et al., using Streptozotocininduced diabetes as a model in spontaneously hypertensive rats (SHR), found that glomerular albumin permeability was higher and apoptosis was increased in diabetic SHR compared with the control group (48). After induction of HO-1 overexpression by COPP, the permeability of albumin and apoptosis decreased. These results indicated that HO-1 reduces glomerular inflammation and oxidative stress induced by diabetes. HO-1 also has an effect on insulin sensitivity (49); induction of HO-1 expression increased insulin sensitivity, reduced adipose tissue volume, caused adipose tissue remodeling, and improved hyperinsulinism of insulin resistance in diabetes.

The protective role of HO-1 in DKD has also been demonstrated in relevant clinical studies. One study found that HO-1 can be used as an early biomarker of DKD (48). Urinary HO-1 levels were significantly higher in the diabetic nephropathy group than in the non-diabetic nephropathy group, and the increase in urinary HO-1 levels in these patients preceded significant proteinuria. The level of urinary HO-1 is synchronized with the abnormalities of traditional renal function indicators such as creatinine and urea nitrogen. Heme oxygenase and creatinine are jointly involved in the occurrence and development of diabetic nephropathy.

4.4 Hypertensive nephropathy

Hypertensive nephropathy is defined as renal insufficiency caused by essential hypertension arteriosclerosis, which is one of the major causes of ESRD. HO-1 is not involved in the maintenance of hypertension under physiological conditions, but plays a key role in the reduction of pathological hypertension, especially Ang-2dependent hypertension (50). Philip Wenzel found that, in the aorta of Ang-2-induced hypertensive rats, infusion of Ang-2 increased HO-1 expression in aortic adventitia and endothelial cells (51). HO-1-deficient rats were more likely to develop hypertension and cardiac hypertrophy than wild mice. The more extensive and severe the ischemic injury at the cortical-medullary junction, the more obvious the inflammatory mediators and inflammatory cell infiltration in the endothelium. This suggests the expression of HO-1 may decrease the effect of Ang on elevated blood pressure. Sylvia and colleagues established several models of HO-1 specific overexpression, which also confirmed that HO-1 has a specific role in protecting TALH cells from Ang-2 invasion (52). It prevents the increase in Ang2-mediated peroxidase production, significantly decreases prostaglandin E2 levels, decreases blood pressure, and reduces the sensitivity of medullary sodium potassium chloride cotransporter 2 (NKCC2) to furosemide, thereby regulating urinary sodium and attenuating the development of Ang II-dependent hypertensive nephropathy.

4.5 Chronic kidney disease

Chronic kidney disease is mainly characterized by renal interstitial fibrosis and glomerulosclerosis. HO-1 inhibits the activation of apoptosis pathway and inhibits macrophagemediated inflammation and fibrosis progression, thereby inhibiting the loss of periosteal capillaries, inhibiting the activation and proliferation of myofibroblasts, and reducing tubulointerstitial infiltration to delay renal fibrosis (53). The mouse model of ureteral obstruction is a rapid and standard model for studying renal interstitial fibrosis. Correa-Costa demonstrated the ability of HO-1 to prevent and reverse fibrosis formation in CKD through the study of a UUO mouse model (54). They observed the mice treated with ureteral obstruction and found that their glomerular and renal interstitial HO-1 was continuously up-regulated from an early stage. This change occurred earlier than the changes of serum creatinine and urine protein. In HO-1 knockout mice, the fibrosis-related growth factors (such as TGF- β 1 and CTGF) and inflammatory factors (TNF- α , IL-1 β , IL-6, and IL-10) were significantly higher than those in wild mice. These results suggest that HO-1 can alleviate renal interstitial fibrosis after UUO, inhibit the activation and proliferation of myofibroblasts, and reduce the infiltration of inflammatory cytokines, thereby alleviating renal injury and fibrosis.

4.6 Kidney transplantation

Kidney transplantation is currently the best treatment for patients with ESRD, and its technology has been relatively mature. However, transplantation-related ischemia-reperfusion injury, rejection, delayed graft function (DGF), and immunosuppression are still important limiting factors affecting the success rate of transplantation in patients. Reducing renal injury and rejection after transplantation and improving graft function and recipient survival rate are still the research hotspots. As a protective molecule, HO-1 was found to be upregulated in many processes of renal transplantation, including ischemia-reperfusion, DGF, acute and chronic rejection, and xenotransplantation (55, 56). HO-1 improves long-term outcomes after renal transplantation by anti-oxidative stress and inhibiting immune responses. Tullius et al. pretreated donor rats with COPP (a HO-1 inducer) and found that the expression of several inflammatory mediators such as IL-10, IFN- γ , and TNF- α decreased, the polymorphonuclear cell infiltration reduced, and the renal survival rate was significantly improved after transplantation compared with that of unpretreated donor kidneys (57). This effect was observed even if the ischemia time was prolonged to 44h. HO-1 improved the microcirculation of the graft and improved the renal blood perfusion.

HO-1 also plays a vital role in cell proliferation and immune regulation (58): in COPP-treated mice, observed overexpression of HO-1 in the kidney, spleen, and liver were observed. T cell-mediated cellular damage and NK cell-mediated cytotoxicity in the spleen were severely inhibited, and lymphoproliferative hypersensitivity decreased. This indicated that overexpression of HO-1 can lead to inhibition of immune effector cells, which play a vital beneficial role in improving host rejection of grafts during acute rejection. But other studies also showed that HO-1 plays a crucial role in the proliferation and differentiation of B cells: comparing the B lymphocyte of HO-1 knockout mice with those of wild mice, it was found that the B cell counts of HO-1 knockout mice were significantly lower, with growth and development delayed (59). Their results suggested that HO-1 attenuates B cell cycle arrest and apoptosis and stimulates B cell activation and differentiation, which in turn increases the risk of humoral immunity during acute rejection.

In general, many preclinical studies on renal transplantation have shown that induction of HO-1 expression can alleviate graft rejection and attenuate renal injury by inhibiting immune response and oxidative stress. Early HO-1 induction therapy before kidney transplantation is a promising approach to improve renal perfusion and increase graft survival. HO-1 plays a beneficial role in various acute and chronic kidney diseases and can be used as an early biomarker of AKI and some CKD. Clinical studies have found that the level of HO-1 in the blood and urine of patients with AKI is significantly higher than that of control groups, and it is detectable earlier than the change of serum creatinine and urinary albumin, which can be used as an early marker of AKI (60). A cross-sectional study found that serum HO-1 levels were significantly elevated in sepsis patients with kidney damage, and were positively correlated with sepsis-induced AKI (61). Changes in the level of HO-1 are also seen in CKD such as diabetic nephropathy, which can reflect kidney damage (48).

5 Therapeutic potential of HO-1 in renal diseases

A growing body of preclinical evidence has shown that HO-1 plays a beneficial role in multiple kidney diseases through antiinflammatory, anti-oxidative, anti-fibrosis, and anti-apoptosis mechanisms (62, 63), which has triggered research on the therapeutic value of HO-1 and its inducers in kidney diseases. Whether the HO-1 cell protection pathway can be a promising target for clinical intervention to alleviate kidney injury is an important research field.

A variety of drugs have shown beneficial effects in inducing HO-1 expression and promoting renal protection in preclinical experiments, including heme and its derivatives, metal inducers, pharmaceutical compounds, natural inducers, and HO enzyme byproducts. The therapeutic effects are summarized in Table 2.

Heme and its analogues are one of the most potent inducers of HO-1 (36). Pretreatment with a low dose of heme before ischemia can induce HO-1, which leads to a favorable biological response by inhibiting the production of proinflammatory cytokines and producing cytopathic metabolites, thus alleviating renal IRI injury (64). The beneficial effects of arginine heme and hemin on reversing renal oxidative stress injury by upregulating HO-1 in the kidney have also been confirmed in animal experiments, among which arginine heme has been approved for the treatment of acute intermittent purpura in clinic. However, excess heme has prooxidative and lipid peroxidation effects and a risk of thrombophlebitis.

CoCl₂, SnCl₂, ZnCl₂, COPP, and other strong inducers containing metal ions are effective and specific inducers to induce HO-1 expression in the kidney (66, 67). Salom et al. injected CoCl₂ into IRI rats in advance and found that HO-1 was up-regulated, which further inhibited the outbreak of inflammatory factors, reduced the concentration of basal NO, improved the blood perfusion after ischemia, and relieved renal tubular damage. In the rat ischemia-reperfusion model, similar results were observed when SnCl₂ was injected intraperitoneally and subcutaneously into mice. In addition, some natural inducers have also shown effects in related studies (68).

Some pharmaceutical compounds such as statins, 5aminosalicylic acid (5-ASA), paricalcitol, and celastrol can also induce HO-1 expression and reduce tissue damage (30, 69–71):

Tzong-Shyuan Lee et al. conducted in vitro experiments on human and rat aortic smooth muscle cells and found that the level of HO-1 was significantly increased in statin-treated cells, with a decrease in inflammatory factors and an increase in the expression of anti-proliferative factors such as p21. The opposite results were detected when treated with HO-1 inhibitors. The effect of 5-ASA on HO-1 induction was found in colitis animal models (72). Palicalcitol mediated HO-1 expression through Nrf2 signal transduction pathway to reduce kidney inflammation and injury (79). Celastrol has previously attracted attention for its antiinflammatory and anti-cancer effects. Later studies found that it increased the expression of HO-1 in CKD, and dose-dependently inhibited inflammation, vascular calcification, and renal fibrosis (73, 74). This demonstrates its important role in delaying the progression of kidney disease. HO-1 inhibitor could offset its anti-inflammatory and anti-vascular calcification effects.

As an ROS scavenger, the bilirubin and its protective effects have been confirmed. High bilirubin concentration is significantly associated with enhanced endothelial function and reduction of oxidative stress damage (78). As a toxic gas, it was found that a small dose of CO inhalation $(1.3 \pm 0.2\text{ml/min})$ before IRI could reveal an increase in HO-1 levels. HO-1 further alleviated renal injury through anti-inflammatory and anti-apoptosis effects, and effectively reduced the chronic fibrosis of kidney and delayed the progression of renal disease (75, 76). Studies in the pig cardiopulmonary bypass model (CPB) also showed that CO inhalation before CPB can improve renal function and reduce tissue damage (77).

However, although preclinical studies on HO-1 and its inducers in the treatment of acute and chronic kidney diseases are abundant and have shown beneficial effects, their side effects are also evident (80): First, in recent years, more and more evidence supports that the excessive upregulation of HO-1 is related to the poor prognosis of certain malignant tumors (81), especially breast cancer, renal cell carcinoma, and liver cancer. It can promote tumor growth, metastasis, and angiogenesis, thus promoting cancer progression. However, it also exhibits the opposite effect in some cancer cells, and its effect on tumors is still controversial. Second, induction of HO-1 improves AKI after IRI. However, most of the relevant preclinical trials are induced by prophylactic administration of HO-1 before the occurrence of ischemia, and the experimental effect is related to the dose and time of administration (82). Its effective therapeutic window is narrow. Some experiments have also found that the application of heme after the occurrence of ischemia can actually promote the progression of injury. Furthermore, the increased expression of HO-1 leads to the accumulation of its by-products such as iron and CO, which further leads to iron-mediated ROS release and CO-mediated vascular effects, aggravating the injury. It may also lead to changes in plasma lipoprotein levels, thereby affecting the susceptibility of the body to atherosclerosis (16, 83). Moreover, some metals and natural inducers have non-specific effects and toxicity, and their safety remains to be confirmed.

The dual role of HO-1 induction therapy and the complexity of the disease hinders the progress from preclinical research to clinical trials. There are still only a few relevant clinical studies. Bharucha et al. confirmed for the first time that the activity and content of HO-1 can be safely induced in humans by infusing hemin into healthy volunteers (65). Small doses of oral Dglyceric acid (DGA) have also been found to upregulate HO-1 and bilirubin in the blood and reduce inflammatory damage in the body. This is especially beneficial to chronic diseases mediated by inflammation (84). However, further clinical

Drug		Disease models	Main mechanisms	References
Heme and its	Heme	Rat renal IR	Inhibit pro-inflammatory cytokines, participate in cell metabolism	(36, 64, 65)
analogs	Arginine heme Hemin			
Inducers	CoCl2, SnCl2, ZnCl2,	Mouse IR model	Inhibit inflammatory outbreak, reduce basal NO, improve perfusion	(66, 67)
COPP	СОРР	Mouse kidney transplant model,	Inhibit inflammation, inhibit apoptosis, participate in metabolism	(55)
	Natural inducers	mouse model	HO-1 expression	(68)
Drug compounds	Statins	Rat renal IR,	Induce HO-1 via the Nrf2 pathway	(30, 69–71)
		aortic smooth muscle cells		
-	5-ASA	Mouse colitis model	Induce HO-1 via the Nrf2 pathway	(72)
	Celastrol	UUO mouse model	Increase HO-1, anti-inflammatory, inhibit vascular calcification, inhibit fibrosis	(73, 74)
Metabolites of HO	СО	IR mouse model, sepsis mouse model	Anti-inflammatory, regulating P21, inhibiting apoptosis	(75–77)
	Bilirubin	Endotoxemia rats	Inhibit NADPH,ROS	(78)

TABLE 2 The therapeutic potential of HO-1 inductors in renal diseases.

research is needed to determine the safety and efficacy of HO-1 induction therapy in humans. Whether long-term HO-1 induction therapy can safely and effectively slow the progression of kidney disease and improve the prognosis needs to be further studied in the future.

6 Summary and prospect

Due to the attention and in-depth research on HO-1, its role in kidney diseases has been gradually confirmed. It not only plays a role in heme-mediated pathological diseases, but also participates in various acute and chronic kidney diseases. It is differentially induced in glomeruli, renal tubules, and renal blood vessels to respond to injury and alleviate renal injury through anti-inflammatory and anti-oxidation, immune regulation, apoptosis inhibition, autophagy regulation, and hemodynamic effects. HO-1 induction therapy has shown beneficial effects in preclinical trials, but there are side effects, and relevant clinical research evidence is still insufficient. Several points have been made clear: The expression level of HO-1 affects the progress of kidney disease, and the lack or inhibition of HO-1 will worsen the renal structure and function after injury, while the increased expression has a protective effect; Plasma and urine levels of HO-1 can be used as early biomarkers of certain kidney diseases, earlier than changes in proteinuria and serum creatinine; and HO-1 and its effective inducers can alleviate acute and chronic kidney injury and delay the progression of kidney disease by inducing the expression of HO-1. But HO-1 induction therapy also has many limitations. Inappropriate induction may produce side effects

References

1. Grunenwald A, Roumenina LT, Frimat M. Heme oxygenase 1: a defensive mediator in kidney diseases. Int J Mol Sci (2021) 22(4):2009. doi: 10.3390/ijms22042009

2. Drummond GS, Baum J, Greenberg M, Lewis D, Abraham NG. HO-1 overexpression and underexpression: clinical implications. *Arch Biochem Biophys* (2019) 673:108073. doi: 10.1016/j.abb.2019.108073

3. Nath M, Agarwal A. New insights into the role of heme oxygenase-1 in acute kidney injury[J]. *Kidney Res Clin Practice Korean Soc Nephrol* (2020) 39(4):387-401. doi: 10.23876/j.krcp.20.091

4. Zager RA, Johnson ACM. Progressive histone alterations and proinflammatory gene activation: consequences of heme protein/iron-mediated proximal tubule injury. *Am J Physiol-Renal Physiol Am Physiol Soc* (2010) 298(3):F827–37. doi: 10.1152/ajprenal.00683.2009

5. Facchinetti MM. Heme-Oxygenase-1. Antioxidants Redox Signaling (2020) 32 (17):1239-42. doi: 10.1089/ars.2020.8065

6. Gáll T, Balla G, Balla J. Heme, heme oxygenase, and endoplasmic reticulum stress-a new insight into the pathophysiology of vascular diseases. *Int J Mol Sci* (2019) 20(15):E3675. doi: 10.3390/ijms20153675

7. Han SJ, Kim M, D'Agati VD, Lee HT. 6-shogaol protects against ischemic acute kidney injury by modulating NF- κ B and heme oxygenase-1 pathways. *Am J Physiol Renal Physiol* (2019) 317(3):F743–56. doi: 10.1152/ajprenal.00182.2019

8. Salom MG, Nieto Cerón S, Rodriguez F, Lopez B, Hernández I, Martínez JG, et al. Heme oxygenase-1 induction improves ischemic renal failure: role of nitric oxide and peroxynitrite. *Am J Physiol-Heart Circulatory Physiol Am Physiol Soc* (2007) 293(6): H3542–9. doi: 10.1152/ajpheart.00977.2007

9. Kovtunovych G, Eckhaus MA, Ghosh MC. Dysfunction of the heme recycling system in heme oxygenase 1–deficient mice: effects on macrophage viability and tissue iron distribution. *Blood* (2010) 116(26):6054–62. doi: 10.1182/blood-2010-03-272138

such as toxicity, promotion of cancer, promotion of atherosclerosis, and aggravation of kidney damage. Therefore, the HO-1 enzyme system, as one of the important targets for the treatment of various kidney diseases, provides a new idea for the targeted therapy of kidney diseases. However, it is still in the exploratory stage, and a lot of further research is needed to explore its effectiveness and safety in human applications.

Author contributions

XW and LN conceived and designed the study. HZ and LN wrote the review article. All authors read and approved the final manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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10. Poss KD, Tonegawa S. Reduced stress defense in heme oxygenase 1-deficient cells. *Proc Natl Acad Sci* (1997) 94(20):10925-30. doi: 10.1073/pnas.94.20.10925

11. Li X, Yu J, Gong L, Zhang Y, Dong S, Shi J, et al. Heme oxygenase-1(HO-1) regulates golgi stress and attenuates endotoxin-induced acute lung injury through hypoxia inducible factor-1 α (HIF-1 α)/HO-1 signaling pathway. *Free Radical Biol Med* (2021) 165:243–53. doi: 10.1016/j.freeradbiomed.2021.01.028

12. Nath KA, Haggard JJ, Croatt AJ, Grande JP, Poss KD, Alam J. The indispensability of heme oxygenase-1 in protecting against acute heme protein-induced toxicity *in vivo*. *Am J Pathol* (2000) 156(5):1527–35. doi: 10.1016/S0002-9440(10)65024-9

13. Adedoyin O, Boddu R, Traylor A, Lever JM, Bolisetty S, George JF, et al. Heme oxygenase-1 mitigates ferroptosis in renal proximal tubule cells. *Am J Physiol Renal Physiol* (2018) 314(5):F702–14. doi: 10.1152/ajprenal.00044.2017

14. Consoli V, Sorrenti V, Grosso S, Vanella L. Heme oxygenase-1 signaling and redox homeostasis in physiopathological Conditions. *Biomolecules* (2021) 11(4):589. doi: 10.3390/biom11040589

15. Lanone S, Bloc S, Foresti R, Almolki A, Taillé C, Callebert J, et al. Bilirubin decreases nos2 expression via inhibition of NAD(P)H oxidase: implications for protection against endotoxic shock in rats. FASEB journal: Off Publ Fed Am Societies Exp Biol (2005) 19(13):1890–2. doi: 10.1096/fj.04-2368fje

16. Ca P, Ms C, Hb S. Carbon monoxide, oxidative stress, and mitochondrial permeability pore transition. *Free Radic Biol Med* (2006) 40(8):1332–9. doi: 10.1016/j.freeradbiomed.2005.11.020

17. Atsaves V, Detsika MG, Poulaki E, Gakiopoulou H, Lianos EA. Phenotypic characterization of a novel HO-1 depletion model in the rat[J]. *Transgenic Res* (2017) 26(1):51–64. doi: 10.1007/s11248-016-9986-9

18. Kumar D, Bhaskaran M, Alagappan L, Tori D, Yadav I, Konkimalla S, et al. Heme oxygenase-1 modulates mesangial cell proliferation by P21Waf1 upregulation. *Renal Failure Taylor Francis* (2010) 32(2):254–8. doi: 10.3109/08860220903491240

19. Nesovic Ostojic J, Ivanov M, Mihailovic-Stanojevic N, Karanovic D, Kovacevic S, Brkic P, et al. Hyperbaric oxygen preconditioning upregulates heme OxyGenase-1 and anti-apoptotic bcl-2 protein expression in spontaneously hypertensive rats with induced postischemic acute kidney injury. *Int J Mol Sci* (2021) 22(3):1382. doi: 10.3390/ ijms22031382

20. Maines MD. Heme oxygenase: function, multiplicity, regulatory mechanisms, and clinical applications. FASEB journal: Off Publ Fed Am Societies Exp Biol (1988) 2 (10):2557-68. doi: 10.1096/fasebj.2.10.3290025

21. Wagner M, Cadetg P, Ruf R, Mazzucchelli L, Ferrari P, Redaelli CA. Heme oxygenase-1 attenuates ischemia/reperfusion-induced apoptosis and improves survival in rat renal allografts. *Kidney Int* (2003) 63(4):1564–73. doi: 10.1046/j.1523-1755.2003.00897.x

22. Ryter SW. Heme oxgenase-1, a cardinal modulator of regulated cell death and inflammation. *Cells* (2021) 10(3):515. doi: 10.3390/cells10030515

23. Tracz MJ, Juncos JP, Croatt AJ, Ackerman AW, Grande JP, Knutson KL, et al. Deficiency of heme oxygenase-1 impairs renal hemodynamics and exaggerates systemic inflammatory responses to renal ischemia. *Kidney Int* (2007) 72(9):1073–80. doi: 10.1038/sj.ki.5002471

24. Athanassiadou V, Plavoukou S, Grapsa E, Detsika MG. The role of heme oxygenase-1 as an immunomodulator in kidney disease. *Antioxidants* (2022) 11 (12):2454. doi: 10.3390/antiox11122454

25. Jackson KE, Jackson DW, Quadri S, Reitzell MJ, Navar LG. Inhibition of heme oxygenase augments tubular sodium reabsorption. *Am J Physiol-Renal Physiol Am Physiol Soc* (2011) 300(4):F941–6. doi: 10.1152/ajprenal.00024.2010

26. Kovtunovych G, Ghosh MC, Ollivierre W, Weitzel RP, Eckhaus MA, Tisdale JF, et al. Wild-type macrophages reverse disease in heme oxygenase 1-deficient mice. *Blood* (2014) 124(9):1522–30. doi: 10.1182/blood-2014-02-554162

27. Landis RC, Quimby KR, Greenidge AR. M1/M2 macrophages in diabetic nephropathy: Nrf2/HO-1 as therapeutic targets. *Curr Pharm Design* (2018) 24 (20):2241-9. doi: 10.2174/1381612824666180716163845

28. Agarwal A, Kim Y, Matas AJ, Alam J, Nath KA. GAS-GENERATING SYSTEMS IN ACUTE RENAL ALLOGRAFT REJECTION IN THE RAT: Co-induction of heme oxygenase and nitric oxide synthase: 1:: 2. *Transplantation* (1996) 61(1):93. doi: 10.1097/00007890-199601150-00019

29. Jourde-Chiche N, Fakhouri F, Dou L, Bellien J, Burtey S, Frimat M, et al. Endothelium structure and function in kidney health and disease. *Nat Rev Nephrol* (2019) 15(2):87–108. doi: 10.1038/s41581-018-0098-z

30. Grosser N, Hemmerle A, Berndt G, Erdmann K, Hinkelmann U, Schürger S, et al. The antioxidant defense protein heme oxygenase 1 is a novel target for statins in endothelial cells. *Free Radical Biol Med* (2004) 37(12):2064–71. doi: 10.1016/j.freeradbiomed.2004.09.009

31. Yachie A, Niida Y, Wada T, Igarashi N, Kaneda H, Toma T, et al. Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. J Clin Investigation Am Soc Clin Invest (1999) 103(1):129–35. doi: 10.1172/JCI4165

32. Lee SC, Han SH, Li JJ, Lee SH, Jung D-S, Kwak S-J, et al. Induction of heme oxygenase-1 protects against podocyte apoptosis under diabetic conditions. *Kidney Int* (2009) 76(8):838–48. doi: 10.1038/ki.2009.286

33. Rubio-Navarro A, Sanchez-Niño MD, Guerrero-Hue M, García-Caballero C, Gutiérrez E, Yuste C, et al. Podocytes are new cellular targets of haemoglobin-mediated renal damage. *J Pathol* (2018) 244(3):296–310. doi: 10.1002/path.5011

34. Poulaki E, Detsika MG, Fourtziala E, Lianos EA, Gakiopoulou H. Podocytetargeted heme oxygenase (HO)-1 overexpression exacerbates age-related pathology in the rat kidney. *Sci Rep* (2020) 10(1):5719. doi: 10.1038/s41598-020-62016-9

35. May O, Merle NS, Grunenwald A, Gnemmi V, Leon J, Payet C, et al. Heme drives susceptibility of glomerular endothelium to complement overactivation due to inefficient upregulation of heme oxygenase-1. *Front Immunol* (2018) 9. doi: 10.3389/fimmu.2018.03008

36. Mosley K, Wembridge DE, Cattell V, Cook HT. Heme oxygenase is induced in nephrotoxic nephritis and hemin, a stimulator of heme oxygenase synthesis, ameliorates disease. *Kidney Int* (1998) 53(3):672-8. doi: 10.1046/j.1523-1755.1998.00798.x

37. Kanwar YS. Heme oxygenase-1 in renal injury: conclusions of studies in humans and animal models. *Kidney Int* (2001) 59(1):378–9. doi: 10.1046/j.1523-1755.2001.00505.x

38. Lever JM, Boddu R, George JF, Agarwal A. Heme oxygenase-1 in kidney health and disease. *Antioxidants Redox Signaling* (2016) 25(3):165-83. doi: 10.1089/ars.2016.6659

39. Rossi M, Delbauve S, Roumeguère T, Wespes E, Leo O, Flamand V, et al. HO-1 mitigates acute kidney injury and subsequent kidney-lung cross-talk. *Free Radical Res* (2019) 53(9–10):1035–43. doi: 10.1080/10715762.2019.1668936

40. Guo S, Guo L, Fang Q, Yu M, Zhang L, You C, et al. Astaxanthin protects against early acute kidney injury in severely burned rats by inactivating the TLR4/MyD88/NFκB axis and upregulating heme oxygenase-1. *Sci Rep* (2021) 11(1):6679. doi: 10.1038/ 341598-021-86146-w 41. Shimizu H, Takahashi T, Suzuki T, Yamasaki A, Fujiwara T, Odaka Y, et al. Protective effect of heme oxygenase induction in ischemic acute renal failure. *Crit Care Med* (2000) 28(3):809. doi: 10.1097/00003246-200003000-00033

42. Rubio-Navarro A, Vázquez-Carballo C, Guerrero-Hue M, García-Caballero C, Herencia C, Gutiérrez E, et al. Nrf2 plays a protective role against intravascular hemolysis-mediated acute kidney injury. *Front Pharmacol* (2019) 10. doi: 10.3389/ fphar.2019.00740

43. Nath KA, Balla G, Vercellotti GM, Balla J, Jacob HS, Levitt MD, et al. Induction of heme oxygenase is a rapid, protective response in rhabdomyolysis in the rat. *J Clin Investigation Am Soc Clin Invest* (1992) 90(1):267–70. doi: 10.1172/JCl115847

44. Kang K, Nan C, Fei D, Meng X, Liu W, Zhang W, et al. Heme oxygenase 1 modulates thrombomodulin and endothelial protein c receptor levels to attenuate septic kidney injury. *Shock* (2013) 40(2):136–43. doi: 10.1097/SHK.0b013e31829d23f5

45. Chung SW, Liu X, Macias AA, Baron RM, Perrella MA. Heme oxygenase-1derived carbon monoxide enhances the host defense response to microbial sepsis in mice. J Clin Investigation Am Soc Clin Invest (2008) 118(1):239–47. doi: 10.1172/ JCI32730

46. Ptilovanciv EO, Fernandes GS, Teixeira LC, Reis LA, Pessoa EA, Convento MB, et al. Heme oxygenase 1 improves glucoses metabolism and kidney histological alterations in diabetic rats. *Diabetol Metab Syndrome* (2013) 5(1):3. doi: 10.1186/1758-5996-5-3

47. da Fonseca CD, Watanabe M, Couto SMF, Santos AACD, Borges FT, Vattimoet MFF. The renoprotective effects of heme oxygenase-1 during contrast-induced acute kidney injury in preclinical diabetic models. *Clinics* (2021) 76:e3002. doi: 10.6061/ clinics/2021/e3002

48. Ahmed F, Abo el leel H, Hebah HA, Magdy A. P1035urinary heme oxygenase 1 as a marker for early diagnosis of diabetic nephropathy. *Nephrol Dialysis Transplant* (2020) 35(Supplement_3):gfaa142.P1035. doi: 10.1093/ndt/gfaa142.P1035

49. Nicolai A, Li M, Kim DH, Peterson SJ, Vanella L, Positano V, et al. Heme oxygenase-1 induction remodels adipose tissue and improves insulin sensitivity in obesity-induced diabetic rats. *Hypertension Am Heart Assoc* (2009) 53(3):508–15. doi: 10.1161/HYPERTENSIONAHA.108.124701

50. Elmarakby AA, Faulkner J, Baban B, Saleh MA, Sullivan JC. Induction of hemeoxygenase-1 reduces glomerular injury and apoptosis in diabetic spontaneously hypertensive rats. *Am J Physiol-Renal Physiol Am Physiol Soc* (2012) 302(7):F791–800. doi: 10.1152/ajprenal.00472.2011

 Wenzel P, Rossmann H, Müller C, Kossmann S, Oelze M, Schulz A, et al. Heme oxygenase-1 suppresses a pro-inflammatory phenotype in monocytes and determines endothelial function and arterial hypertension in mice and humans. *Eur Heart J* (2015) 36(48):3437–46. doi: 10.1093/eurhearti/ehv544

52. Quan S, Yang L, Shnouda S, Schwartzman ML, Nasjletti A, Goodman AI, et al. Expression of human heme oxygenase-1 in the thick ascending limb attenuates angiotensin II-mediated increase in oxidative injury11See editorial by Chang and perrella, p. 1968. *Kidney Int* (2004) 65(5):1628–39. doi: 10.1111/j.1523-1755.2004.00562.x

53. Kie J-H, Kapturczak MH, Traylor A, Agarwal A, Hill-Kapturczak N. Heme oxygenase-1 deficiency promotes epithelial-mesenchymal transition and renal fibrosis. *J Am Soc Nephrol : JASN* (2008) 19(9):1681. doi: 10.1681/ASN.2007101099

54. Correa-Costa M, Semedo P, Monteiro APFS, Silva RS, Pereira RL, Gonçalves GM, et al. Induction of heme oxygenase-1 can halt and even reverse renal tubule-interstitial fibrosis. *PloS One* (2010) 5(12):e14298. doi: 10.1371/journal.pone.0014298

55. Corona D, Ekser B, Gioco R, Caruso M, Schipa C, Veroux P, et al. Hemeoxygenase and kidney transplantation: a potential for target therapy? *Biomolecules* (2020) 10(6):E840. doi: 10.3390/biom10060840

56. Hölzen JP, August C, Bahde R, Minin E, Lang D, Heidenreich S, et al. Influence of heme oxygenase-1 on microcirculation after kidney transplantation. *J Surg Res* (2008) 148(2):126–35. doi: 10.1016/j.jss.2007.10.007

57. Tullius SG, Nieminen-Kelhä M, Bachmann U, Reutzel-Selke A, Jonas S, Pratschke J, et al. Induction of heme-oxygenase-1 prevents ischemia/reperfusion injury and improves long-term graft outcome in rat renal allografts. *Transplant Proc* (2001) 33(1):1286–7. doi: 10.1016/S0041-1345(00)02480-5

58. Martins PNA, Reuzel-Selke A, Jurisch A, Atrott K, Pascher A, Pratschke J, et al. Induction of carbon monoxide in the donor reduces graft immunogenicity and chronic graft deterioration. *Transplant Proc* (2005) 37(1):379–81. doi: 10.1016/ j.transproceed.2004.11.079

59. Zhou Z, Ma D, Liu P, Wang P, Wei D, Yu K, et al. Deletion of HO-1 blocks development of b lymphocytes in mice. *Cell Signalling* (2019) 63:109378. doi: 10.1016/j.cellsig.2019.109378

60. Zager RA, Johnson ACM, Becker K. Plasma and urinary heme oxygenase-1 in AKI. J Am Soc Nephrol (2012) 23(6):1048. doi: 10.1681/ASN.2011121147

61. Xia S, Zhang M, Liu H, Dong H, Wu N, Wiedermann CJ, et al. Heme oxygenase-1 as a predictor of sepsis-induced acute kidney injury: a cross-sectional study. *Ann Trans Med* (2022) 10(21):1177-7. doi: 10.21037/atm-22-4793

62. Bolisetty S, Zarjou A, Agarwal A. Heme oxygenase 1 as a therapeutic target in acute kidney injury. *Am J Kidney Dis* (2017) 69(4):531-45. doi: 10.1053/j.ajkd.2016.10.037

63. Uddin MJ, Kim EH, Hannan MA, HA, H. Pharmacotherapy against oxidative stress in chronic kidney disease: promising small molecule natural products targeting Nrf2-HO-1 signaling. *Antioxidants* (2021) 10(2):258. doi: 10.3390/antiox10020258

64. Guo S, Yu M, Fang Q, Zhang L, You C, Wang X, et al. Heme oxygenase-1 induction mitigates burn-associated early acute kidney injury *via* the TLR4 signaling pathway. *Burns* (2022) 48(1):156–67. doi: 10.1016/j.burns.2021.04.013

65. Bharucha AE, Kulkarni A, Choi KM, Lempke M, Brunn GJ, Gibbons SJ, et al. First-in-Human study demonstrating pharmacological activation of heme oxygenase-1 in humans. *Clin Pharmacol Ther* (2010) 87(2):187–90. doi: 10.1038/clpt.2009.221

66. Li Volti G, Sorrenti V, Murabito P, Galvano F, Veroux M, Gullo A, et al. Pharmacological induction of heme oxygenase-1 inhibits iNOS and oxidative stress in renal ischemia-reperfusion injury. *Transplant Proc* (2007) 39(10):2986–91. doi: 10.1016/j.transproceed.2007.09.047

67. Toda N, Takahashi T, Mizobuchi S, Fujii H, Nakahira K, Takahashi S, et al. Tin chloride pretreatment prevents renal injury in rats with ischemic acute renal failure. *Crit Care Med* (2002) 30(7):1512. doi: 10.1097/00003246-200207000-00020

68. Funes SC, Rios M, Fernández-Fierro A, Covián C, Bueno SM, Riedel CA, et al. Naturally derived heme-oxygenase 1 inducers and their therapeutic application to immune-mediated diseases. *Front Immunol* (2020) 11:1467. doi: 10.3389/fimmu.2020.01467

69. Mansouri A, Reiner Ž, Ruscica M, Tedeschi-Reiner E, Radbakhsh S, Ekta MB, et al. Antioxidant effects of statins by modulating Nrf2 and Nrf2/HO-1 signaling in different diseases. *J Clin Med* (2022) 11(5):1313. doi: 10.3390/jcm11051313

70. Gueler F, Park J-K, Rong S, Kirsch T, Lindschau C, Zheng W, et al. Statins attenuate ischemia-reperfusion injury by inducing heme oxygenase-1 in infiltrating macrophages. *Am J Pathol* (2007) 170(4):1192. doi: 10.2353/ajpath.2007.060782

71. Lee T-S, Chang C-C, Zhu Y, Shyy JY-J. Simvastatin induces heme oxygenase-1. *Circulation Am Heart Assoc* (2004) 110(10):1296-302. doi: 10.1161/ 01.CIR.0000140694.67251.9C

72. Horváth K, Varga C, Berkó A, Pósa A, László F, Whittle BJR. The involvement of heme oxygenase-1 activity in the therapeutic actions of 5-aminosalicylic acid in rat colitis. *Eur J Pharmacol* (2008) 581(3):315–23. doi: 10.1016/j.ejphar.2007.12.004

73. Tang M, Cao X, Zhang K, Li Y, Zheng Q, Li G, et al. Celastrol alleviates renal fibrosis by upregulating cannabinoid receptor 2 expression. *Cell Death Disease Nat Publishing Group* (2018) 9(6):1–12. doi: 10.1038/s41419-018-0666-y

74. Yang X, Chen A, Liang Q, Dong Q, Fu M, Liu X, et al. Up-regulation of heme oxygenase-1 by celastrol alleviates oxidative stress and vascular calcification in chronic kidney disease. *Free Radical Biol Med* (2021) 172:530–40. doi: 10.1016/j.freeradbiomed.2021.06.020

75. Nakao A, Neto JS, Kanno S, Stolz DB, Kimizuka K, Liu F, et al. Protection against Ischemia/Reperfusion injury in cardiac and renal transplantation with carbon monoxide, biliverdin and both. *Am J Transplant* (2005) 5(2):282–91. doi: 10.1111/j.1600-6143.2004.00695.x

76. Neto JS, Nakao A, Toyokawa H, Nalesnik MA, Romanosky AJ, Kimizuka K, et al. Low-dose carbon monoxide inhalation prevents development of chronic allograft nephropathy. *Am J Physiol-Renal Physiol* (2006) 290(2):F324–34. doi: 10.1152/ajprenal.00026.2005

77. Goebel U, Siepe M, Schwer CI, Schibilsky D, Foerster K, Neumann J, et al. Inhaled carbon monoxide prevents acute kidney injury in pigs after cardiopulmonary bypass by inducing a heat shock response. *Anesth Analgesia* (2010) 111(1):29. doi: 10.1213/ANE.0b013e3181e0cca4

78. Maruhashi T, Soga J, Fujimura N, Idei N, Mikami S, Iwamoto Y, et al. Hyperbilirubinemia, augmentation of endothelial function, and decrease in oxidative stress in Gilbert syndrome. *Circulation Am Heart Assoc* (2012) 126(5):598–603. doi: 10.1161/CIRCULATIONAHA.112.105775

79. Wang S, Huang S, Liu X, He Y, Liu Y, et al. Paricalcitol ameliorates acute kidney injury in mice by suppressing oxidative stress and inflammation *via* Nrf2/HO-1 signaling. *Int J Mol Sci* (2023) 24(2):969. doi: 10.3390/ijms24020969

80. Rossi M, Delbauve S, Wespes E, Roumeguère T, Leo O, Flamand V, et al. Dual effect of hemin on renal ischemia-reperfusion injury. *Biochem Biophys Res Commun* (2018) 503(4):2820–5. doi: 10.1016/j.bbrc.2018.08.046

81. Nitti M, Piras S, Marinari UM, Moretta L, Pronzato MA, Furfaro AL, et al. HO-1 induction in cancer progression: a matter of cell adaptation. *Antioxidants* (2017) 6 (2):29. doi: 10.3390/antiox6020029

82. Zou L, Sato N, Attuwaybi B, Kone BC. Delayed administration of alphamelanocyte-stimulating hormone or combined therapy with BAY 11-7085 protects against gut ischemia-reperfusion injury. *Shock (Augusta Ga.) Shock* (2003) 20(5):469– 75. doi: 10.1097/01.shk.0000091205.08003.fd

83. Freigang S, Ampenberger F, Spohn G, Heer S, Shamshiev AT, Kisielow J, et al. Nrf2 is essential for cholesterol crystal-induced inflammasome activation and exacerbation of atherosclerosis. *Eur J Immunol* (2011) 41(7):2040–51. doi: 10.1002/eji.201041316

84. Hirvonen OP, Lehti M, Kyröläinen H, Kainulainen H. Heme oxygenase-1 and blood bilirubin are gradually activated by oral d-glyceric acid. *Antioxidants* (2022) 11 (12):2319. doi: 10.3390/antiox11122319