Check for updates

OPEN ACCESS

EDITED BY Senlian Hong, Peking University, China

REVIEWED BY Caiyun Jiang, Fujian Medical University Union Hospital, China Ru Zhou, Shanghai Jiaotong University School of Medicine, China Xiaoke Chen, Shanghai Jiao Tong University, China

*CORRESPONDENCE Tuo Yi, ⊠ yi.tuo@zs-hospital.sh.cn

RECEIVED 20 September 2023 ACCEPTED 14 November 2023 PUBLISHED 22 February 2024

CITATION

Yang K and Yi T (2024), Tumor cell stemness in gastrointestinal cancer: regulation and targeted therapy. *Front. Mol. Biosci.* 10:1297611. doi: 10.3389/fmolb.2023.1297611

COPYRIGHT

© 2024 Yang and Yi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Tumor cell stemness in gastrointestinal cancer: regulation and targeted therapy

Kangqi Yang¹ and Tuo Yi²*

¹School of Basic Medical Sciences, Fudan University, Shanghai, China, ²Department of General Surgery, Zhongshan Hospital, Fudan University, Shanghai, China

The cancer stem cells are a rare group of self-renewable cancer cells capable of the initiation, progression, metastasis and recurrence of tumors, and also a key contributor to the therapeutic resistance. Thus, understanding the molecular mechanism of tumor stemness regulation, especially in the gastrointestinal (GI) cancers, is of great importance for targeting CSC and designing novel therapeutic strategies. This review aims to elucidate current advancements in the understanding of CSC regulation, including CSC biomarkers, signaling pathways, and non-coding RNAs. We will also provide a comprehensive view on how the tumor microenvironment (TME) display an overall tumor-promoting effect, including the recruitment and impact of cancer-associated fibroblasts (CAFs), the establishment of an immunosuppressive milieu, and the induction of angiogenesis and hypoxia. Lastly, this review consolidates mainstream novel therapeutic interventions targeting CSC stemness regulation.

KEYWORDS

gastrointestinal tumor, cancer stem cell, tumor heterogeneity, tumor microenvironment, epithelial-to-mesenchymal transition, cancer-associated fibroblast

1 Introduction

Gastrointestinal (GI) cancer refers to the malignant tumors in the GI tract and its accessory organs, including the esophagus, stomach, small intestine, colon, rectum, anus, liver, gall bladder and pancreas. Cancer statistics issued by American Cancer Society (ACS) estimated 348,840 new cases and 172,010 deaths cause by GI cancer in the United States in 2023 (Siegel et al., 2023). The incidence of GI cancer in the United States between 2015 and 2019 was as high as 50.9 per 100,000, and the mortality rate from 2016 to 2020 was 22.5 per 100,000. Such high morbidity and mortality urges the need to understand the molecular mechanisms underlying the initiation, migration and therapeutic resistance of GI cancer.

Surgical resection presently stands as the standard therapeutic modality for early-stage GI cancer. However, in cases where complete resection is unattainable, the therapeutic focus pivots toward employing adjuvant therapies, particularly chemotherapy and immunotherapy, to extend patient survival. Despite myriad clinical endeavors targeting cancer, sustainable outcomes remain elusive due to the emergence of therapy resistance in cancer cells. With an attempt to uncover the underlying mechanisms, multiple researches have focused on tumor heterogeneity, a salient hallmark of malignancy. Tumor heterogeneity manifests at both intertumoral and intratumoral levels. The former denotes individual variations, while the latter exhibits both as spatial heterogeneity, which refers to a scattered distribution of genetically distinctive cell subpopulation across or within a tumor site, and temporal heterogeneity, described as the temporal variation in the genetic alterations in one individual tumor (Dagogo-Jack and Shaw, 2018).

Focusing on the dynamics in cancer development, the cancer stem cell (CSC) model has gained widespread acceptance. According to this paradigm, CSCs, resembling normal stem cells, are a group of cells characterized by an uncontrolled proliferation potential and an indefinite capacity to drive the growth of new tumors (Shibue and Weinberg, 2017).

Fueled by the notion that tumors are essentially aberrant organs driven by tumor-initiating cells, researches begin to aim for analogies between normal stem cells and tumorigenic cells (Reya et al., 2001). The CSC theory, initially elucidated in hematopoietic malignancy (Lapidot et al., 1994), gains further credence through the evidence from the hierarchical arrangement of phenotypically varied cancer cells (Prasetyanti and Medema, 2017). Termed also as tumor initiating cells, CSC has been shown by mounting evidence to exhibit high tumorigenicity (Huang et al., 2020; Walcher et al., 2020). Extensions of this concept further proposed CSC's role in therapy resistance, ascribing the limited efficacy of conventional therapies to their impotency in targeting CSC (Dean et al., 2005; Shibue and Weinberg, 2017; Dagogo-Jack and Shaw, 2018). Evidence revealing CSC's high resistance in its dormant state (Aguirre-Ghiso, 2007; Talukdar et al., 2019), and the notion that reactivated CSCs continue to act as tumor precursors (Shibue and Weinberg, 2017), prompted extensive investigations on its significance as a trigger of cancer relapse. In summary, CSC stemness is defined as its function in tumor initiation, metastasis, therapeutic resistance and reoccurrence, making it one of the most significant subjects for understanding tumorigenesis.

Of all biological programs regulating CSC stemness, the most attributable is epithelial-to-mesenchymal transition (EMT), a process critical for embryonic development, wound healing and tissue regeneration (Shibue and Weinberg, 2017). As the name suggests, the program drives epithelial cells to undergo a reduction in apicobasal polarity and intracellular adhesion (Yilmaz and Christofori, 2009), consequently offering them mesenchymal-like properties including higher invasiveness and motility (Shibue and Weinberg, 2017). Aberrantly triggered EMT has been elucidated to have significant function in CSC stemness activation, inducing cell renewal and therapeutic resistance (Wilson et al., 2020; Zhang et al., 2021). Via signaling pathways, EMT transcription factor ZEB1, TWIST and SNAIL undergoes alterations (De Craene and Berx, 2013), which will suppress epithelial-associated proteins such as E-cadherin, and in turn upregulate genes encoding mesenchymal-associated proteins, including vimentin and N-cadherin (De Craene and Berx, 2013).

The CSC model has become a mainstream concept featured in paramount researches ever since its first release. However, our knowledge about the underlying molecular mechanism of CSC regulation is still limited, due to the lack of connections built between the potential regulators. Meanwhile, based on the spatial heterogeneity of CSC, it is also critical to study the diverse regulatory mechanisms regarding tissue specificity. Thus, a comprehensive analysis of CSC regulators is needed to conclude the complex crosstalk and provide clear insights for therapeutic strategies. In this review, we summarize the current mainstream findings of factors engaged in the regulation of CSC properties in GI tumors, including its biomarkers [Figure 1(1)], signaling pathways [Figure 1(2)] and non-coding RNAs (ncRNAs) [Figure 1(3)]. We will also closely investigate the niche CSCs reside in to assess the complex crosstalk between multiple regulators [Figure 1(4)]. By featuring both organ specificity and cross-tissue comparison, we will provide insights in the plasticity and mutual properties exhibited by CSCs in different GI organs, and propose clues for therapies exclusively targeting GI tumors.

2 CSC surface markers

Identification of CSC, which currently relies heavily on cell surface markers, is crucial for CSC isolation and early detection. Recent literature has selected a list of biomarker candidates highly correlated to elevated CSC phenotypes and worsened prognostic results (Dalerba et al., 2007; Chu et al., 2009; Sun et al., 2013).

2.1 CD44

CD44, one of the most widely identified CSC markers, is a transmembrane glycoprotein originally expressed on epithelial cell surface for regulating intracellular interactions and immunocyte activation (Hassn Mesrati et al., 2021). Extensive literature reports have highlighted an upregulation of CD44 mRNA expression across multiple cancer types, particularly on cells displaying EMT phenotype.

The structural and functional features of CD44 render it particularly well-suited for facilitating CSC metastasis. As a hyaluronic acid receptor, CD44 engages with multiple ligands, adeptly mediating intracellular interactions that activate cancer cell migration and metastasis (Yu and Stamenkovic, 1999). Meanwhile, CD44 participates in metabolic regulation, inducing chemotherapy resistance by suppressing intracellular reactive oxygen species (ROS) in cancer cells (Tamada et al., 2012).

By interacting with downstream pathways, CD44 has been found to influence stemness and migration in various tumor cell types. In a study isolating CD44⁺ cells from human gastric cancer (GC) cells (Sun et al., 2013), Sun et al. conducted a spheroid colony formation assay, revealing heightened stemness and elevated invasiveness in vitro and in vivo. Concordantly, CD44 knockdown resulted in diminished stemness and migration of GC tumor cells, confirming the necessity of CD44 in CSC tumorigenesis (Takaishi et al., 2009). Research comparing colorectal cancer (CRC) xenograft derived from different percentage of EpCAM/CD44 cells discovered that EpCAM^{high}/ CD44+ cell group possesses the highest level of tumorigenicity (Dalerba et al., 2007). Additionally, CRC cell fluorescenceactivated cell sorting (FACS) analysis identified the CD44⁺ cell subgroup as possessing high clonogenicity and in vivo tumorigenicity (Chu et al., 2009). In hepatocellular carcinoma (HCC), CD44 also induces a mesenchymal phenotype by activating TGF β pathway, consequently promoting cancer migration and metastasis (Mima et al., 2012). Subsequent analysis of HCC patient samples revealed high CD44 expression as a poor prognosis indicator, confirming its clinicopathological significance. Furthermore, the separation of pancreatic ductal adenocarcinoma (PDAC) cell lines with different CD44 level uncovered a correlation between high CD44 level and enhanced CSC-like properties, including elevated tumorigenicity, clonogenic

abilities, and therapy resistance *in vivo* (Zhao et al., 2016). Previous investigations into CD44-expressing cells in PDAC also unveiled its greater EMT phenotype and invasiveness *in vitro* (Mani et al., 2008).

Notably, CD44 isoforms exhibit both distributional and functional heterogeneity. CD44 has two major isoforms, CD44 standard (CD44s) and CD44 variants (CD44v). While CD44v exhibit significantly greater tumorigenicity than CD44s in pancreatic carcinoma (Li et al., 2014), the roles are reversed in HCC and gall bladder carcinoma (GBC), where CD44s acts as the predominant CSC-promoter (Mima et al., 2012; Miwa et al., 2017). Due to the heterogeneity of CD44 isoforms, closer investigations are needed before therapeutically targeting CD44 to ensure the treatment's targeting accuracy and organ specificity.

2.2 CD133

CD133 is one of the most well-studied CSC markers. This cell surface marker has been implicated in instigating an upregulation of tumorigenic factors in GC, including EMT-associated SNAIL and N-cadherin, along with the tumor development promoters Oct-4 and c-myc (Soleimani et al., 2022). Similarly, CD133 can mediate tumor stemness and metastatic properties in CRC by exosome secretion (Zhao et al., 2020). Clinically, meta-analysis on GC patients also observed increased lymphatic metastasis (Yiming et al., 2015) and worsened 5-year survival rate among the patients harboring CD133-positive cells (Wen et al., 2013). Similar clinicopathological correlation was recorded in CRC, PDAC and EC patients (Chen et al., 2013b; Wen et al., 2013; Wang et al., 2019b), suggesting the high prognostic value of CD133.

CD133 has been demonstrated to promote the proliferation and CSC-like traits across diverse tumor types, which positions CD133 as a key candidate for tumor identification and therapeutic targeting. However, CD133 has been categorized by some as a less effective marker, as several studies fail to observe a distinguishable tumorigenicity increase in CD133+ cells compared to CD133-groups (Dalerba et al., 2007), possibly due to distributional heterogeneity.

2.3 EpCAM

The epithelial cell adhesion molecule (EpCAM) is found to be expressed on CSC across various cancer types. Based on the notion that EpCAM serves as a marker of normal hepatic stem cells, researchers sought the mechanism of tumorigenicity of EpCAM in the pre-cancerous stage of advanced cirrhosis. Investigations into EpCAM + cells from advanced cirrhosis revealed an elevated expression of the Notch gene, accompanied by enhanced selfrenewal in-vitro and an increased resemblance to HCC regarding genetic products (Khosla et al., 2017). Moreover, Yamashita et al. recorded an increased invasiveness of HCC EpCAM cells in NOD/ SCID mouse model, elucidating the regulatory mechanism through the Wnt/β-catenin signaling pathway (Yamashita et al., 2009). EpCAM is also among the first markers to effectively identify CSCs in PDAC (Ishiwata et al., 2018), CRC (Dalerba et al., 2007) and GC (Dai et al., 2017), making it a promising candidate for the identification of CSCs in GI cancers.

2.4 ALDH

Aldehyde dehydrogenase (ALDH), an enzyme integral to diverse metabolic processes (Toledo-Guzmán et al., 2019), has only recently gained recognition for its involvement in CSC identification, as cumulative evidence suggesting it as a major regulator governing tumorigenic properties (Mele et al., 2018; Toledo-Guzmán et al., 2019). In GC, ALDH^{high} cells were observed to have greater sphere-forming ability *in vitro* and elevated tumorigenicity *in vivo*. The orchestrated regulation of ALDH aligns with the activities of Notch and Sonic Hedgehog (SHH) signaling pathways, emphasizing its critical role in mediating downstream cellular activities (Nishikawa et al., 2013). Furthermore, ALDH has emerged as a promising marker for identification in esophageal cancer (EC) (Islam et al., 2015) and PDAC (Rasheed et al., 2010).

Interestingly, even as the majority of studies concluded ALDH as a key contributor to tumorigenesis and poor clinical outcomes, a few studies posit alternative perspectives. An analysis of 1420 CRC tissue microarrays by Lugli et al. failed to establish a significant correlation between ALDH expression and patient survival rate (Lugli et al., 2010). Meanwhile, a comprehensive analysis of cell surface markers in PDAC documented the ALDH expression abundance in normal pancreatic tissue as similar to that of the cancer tissue, challenging its exclusivity as a CSC marker in PDAC (Ishiwata et al., 2018). Although these reports do not diminish the potential of ALDH as a CSC marker, they underscore the importance of developing a universal CSC identification strategy, and indicate the challenges of CSC identification posed by its spatial heterogeneity.

2.5 Transcriptional factors

aforementioned surface markers, Despite the CSC transcriptional factors are also valuable for CSC identification and therapeutic targeting. Sox2, a member of the SRY-related HMG-box (SOX) family, is among the most extensively discussed factors. Employing an *in-vitro* cell culture model, Lundburg et al. substantiated that Sox2-expressing cells exhibit CSC-like features, which is concomitant with a reduced level of CDX2, a state typically observed in late-stage CRC patients (Lugli et al., 2010). These findings are in line with the research conducted by Takeda et al., which also elucidated Sox2's role in the maintenance of stemness and the regulation of differentiation states (Takeda et al., 2018). Similarly, in GC and PDAC, expression of Sox2 correlates with enhanced self-renewal ability, metastatic potential and therapy resistance (Wuebben et al., 2016; Zhang et al., 2016; Sharma et al., 2019).

While the landscape of cell marker research continues to expand, biomarkers are not completely reliable for CSC identification. As previously highlighted, the substantial heterogeneity among tumor subtypes and the phenotypic plasticity inherent in CSCs can compromise the efficacy of surface markers. Due to CSC plasticity, the ablation of biomarkers may not completely diminish the CSC-like traits. A study by Huang et al., in 2013 challenged the effectiveness of biomarkers in distinguishing CSCs. They examined the tumorigenic properties of CSC+ (narrowly defined as tumor cells positive of the well-studied CSC markers) and CSC- cells, and found no significant pathological difference after co-transplanting the 2 cell groups in mouse models (Huang et al., 2013). Conclusions from this research is limited, given its inclusion of only a few markers as "CSC+" standards, while numerous novel biomarkers have been discovered and investigated since its publication. Nonetheless, the findings prompt careful considerations when utilizing surface markers for CSC identification or developing corresponding therapies, regarding the high plasticity and heterogeneity associated with CSCs.

3 Signaling pathways

Multiple pathways have been demonstrated to play critical roles in promoting tumorigenic properties, including the elevation of CSC biomarker expression, the regulation of CSC activity, the augmentation of EMT process and the establishment of an immunosuppressive TME. Moreover, compared to cell biomarkers, signaling pathway components are less influenced by subtype heterogeneity and CSC plasticity, therefore possess higher stability in tumor identification and therapeutic targeting.

3.1 WNT

WNT, a highly complex pathway involved in various biological processes, is crucial for determining cell fate and inducing CSC properties. Aberrant activation of WNT pathway has been reported across multiple cancer types, including CRC (Pan et al., 2017), EC (Chu et al., 2022) and GC (Katoh and Katoh, 2022).

WNT is critical for the proliferation/dormancy state transformation of CSC. This process is of great significance to CSC stemness maintenance and therapeutic resistance, as proliferative CSCs are highly vulnerable to therapies compared to their dormant state (Phan and Croucher, 2020). This transformation is mediated by a dynamic interplay between the two states of WNT, namely, the canonical and noncanonical signaling pathway (Katoh, 2017). The former can directly elevate the transcription level of oncogene MYC and cell cycle activator CCND1 (Katoh and Katoh, 2007), meanwhile activating β -catenin to enhance CSC expansion (Katoh and Katoh, 2017). The noncanonical pathway, on the other hand, contributes to β -catenin degradation and CSC dormancy (Katoh and Katoh, 2017).

Multiple CSC biomarkers rely on WNT pathway to regulate their expression and function. Ishimoto et al. has demonstrated on GC-imitating K19-Wnt1/C2mE mouse model that WNT signaling upregulates CD44⁺ cell residing at gastric gland squamo-columnar junction (SCJ), consequently activating the quiescent tissue to adopt a malignant growth pattern (Ishimoto et al., 2010). Similar regulatory effects have been observed in CRC and PDAC, where WNT promotes tumorigenesis by enhancing CD44 activities (van de Wetering et al., 2002; Wang et al., 2015). WNT can also regulate EpCAM expression and activity in HCC, consequently affecting the stem cell-like features of the EpCAM + subgroup (Yamashita et al., 2009).

Beyond its influence on biomarkers, WNT regulates multiple immune-relevant factors to mediate tumor immune evasion. Canonical WNT signaling pathway is shown to directly increase PD-L1 expression (Katoh and Katoh, 2022) and Treg-recruiting C-C motif chemokine ligand (CCL) 28 in GC (Ji et al., 2020). This pathway also facilitates Treg production by limiting IL-12 secretion (Spranger et al., 2017) and downregulating dendritic cell (DC)recruiting chemokines CCL4 and CCL5 (Katoh and Katoh, 2022). Interestingly, previous researches reported that WNT exhibit a positive effect on T cell maturation and activation (Gattinoni et al., 2010), and that WNT downregulates FOXP3 to inhibit Treg immunosuppression (Zhou et al., 2022b). These findings suggest that WNT can display immune-supportive functions under certain conditions. Collective insights into WNT plasticity are needed to underscore the potential for spatiotemporal regulation of WNT signaling to improve tumor immunotherapy.

3.2 JAK/STAT

The Janus kinase/signal transducer and activator of transcription 3 (JAK/STAT3) pathway is crucial in tumorigenesis, exerting regulatory control over tumor initiation, progression and immune responses. Initiated by IL-6, activation of the JAK/STAT axis begins from an induced conformational change of IL-6 receptor, which then activates JAK and initiates the latent transcription factor STAT3 activity (Jones and Jenkins, 2018). The IL-6 family, encompassing IL-11, IL-27, and IL-31, emerges as a set of key regulators influencing immune responses, pathway signaling and CSC expansion (Jones and Jenkins, 2018; Jin, 2020).

By upregulating CD44 ligands and forming a CD44/acetylated STAT3 complex, the IL-6/STAT3 pathway promotes biomarker expression and enhances CSC survival (Lee et al., 2008; Su et al., 2011). Additionally, STAT3 is among the pivotal transcription factors in EMT induction. A study on mouse liver has demonstrated that STAT3 upregulation, triggered by Twist1 expression, activates tumorigenesis by inducing EMT (Uthaya Kumar et al., 2016). Moreover, the JAK/STAT3 pathway mediates therapeutic resistance. As demonstrated by Ma et al., JAK/STAT3-activating IL-11 promotes acquired chemoresistance in GC, followed by a cascading effect that consequently induces antiapoptosis and diminishes therapeutic outcome (Ma et al., 2019a).

Actively engaged in immunocyte crosstalk, the JAK/ STAT3 pathway plays an important role in shaping immunosuppression and indirectly inducing CSC expansion. Members of the IL-6 family, primarily produced under inflammatory states (Tanaka et al., 2016; Yoshida, 2020), contribute to promoting EC and CRC immunosuppression by blocking DC maturation and inducing the M2 polarization of tumor-associated macrophage (TAM) (Wang et al., 2014; Higashino et al., 2019). IL-6 also upregulates cell programmed death 1 (PD-1) expression on CD8 T cells and cell programmed death 1 ligand 1 (PD-L1) on tumor cells, which leads to cytotoxic cell dysfunctionality (Ju et al., 2020). Additionally, the IL-6/STAT3 axis mediates TAM's CSC-promoting property. A coculture of CD44⁺ human HCC cells and TAMs has shown positive correlations between an elevated IL-6 level and the expansion of the CD44⁺ cell group, implicating STAT3's critical role in mediating TAM-CSC crosstalk (Wan et al., 2014).

3.3 TGFβ

Transforming growth factor beta (TGF β) is a cytokine widely expressed in normal tissues, serving key modulatory functions of cell growth, immune responses and intracellular communications (Kubiczkova et al., 2012). In tumor tissues, aberrantly-triggered TGF β is also engaged in activities of many tumor-associated cells including cancer-associated fibroblasts (CAF) and TAM (Trelford et al., 2022).

TGF β release is stimulated by extracellular matrix (ECM) degradation. When ECM degradation reaches a certain rate, CAFs are triggered to release TGF β , inducing the production of pro-oxidant enzymes and ROS, while upregulating the fibrogenesis factors. These effects culminate in desmoplastic aggregation and ECM stiffness promotion (Fuyuhiro et al., 2011; Mortezaee, 2021), which induces the development of CSC-like traits.

TGFB orchestrates the initiation of EMT across various GI malignancies, including GC, CRC and HCC (Calon et al., 2015; Krzysiek-Maczka et al., 2019; Jeng et al., 2023). It directly upregulates the production of multiple EMT transcription factors such as SNAIL, TWIST, SLUG and ZEB1 (Lee and Massague, 2022; Trelford et al., 2022), and also promotes EMT by introducing epigenetic alterations. Via the SMAD signaling pathway, TGFB downregulates DNA demethylation, inducing a hypermethylated state of multiple promoters of genes encoding EMT factors. For example, by inhibiting DNA methyltransferases (DNMT)-induced DNA methylation, TGFβ upregulates CD133 expression (You et al., 2010), enhancing HCC-cell metastasis. Moreover, Malfettone et al. reported a stimulatory effect of TGFβ-induced EMT on CD44 expression in HCC, indicating TGF β 's role in upregulating CSC biomarkers (Malfettone et al., 2017). Tumor cells undergoing TGFβ-induced EMT exhibit increased resistance to therapies including chemotherapy and radiation, establishing a correlation between high TGF β levels and unfavorable prognosis (Zheng et al., 2015).

TGF β also plays a critical role in shaping the tumor immune landscape and mediating CSC immune escape. Targeting cytotoxic T cells, TGF_β can directly diminish their aggressiveness by hindering their responses and inducing apoptosis (Busenhart et al., 2022; Chen et al., 2022). Meanwhile, TGFβ can interfere with DC activities, inhibiting its proliferation, antigen presenting function and overall mobilization (Esebanmen and Langridge, 2017; Peng et al., 2022), thereby obstructing the differentiation process of adaptive immune cells. Similarly, TGFβ dampens natural kill (NK) cell stimulation and interferon-gamma (IFNy) production (Hayashi et al., 2003; Peng et al., 2022), attenuating NK cell effects in cytolysis and lymphocyte mediation (Castro et al., 2018). Furthermore, TGFβ generates immunodeficiency by inducing cell differentiation in an immunosuppressive direction. For instance, TGFβ-recruited tumorassociated neutrophils (TANs) generally exhibit decreased cytotoxicity, indicating their differentiation in a tumorigenic fashion (Fridlender et al., 2009; Song et al., 2021).

Notably, a number of TGF β -mediated activities result in a positive feedback loop. For example, while TGF β interacts with

DC to inhibit its normal immune functions, it also upregulates the DC-derived TGF β level, enhancing its effects in DC regulation (Esebanmen and Langridge, 2017). The same mechanism can be observed in TAN and CAF (Esebanmen and Langridge, 2017; Peng et al., 2022), where TGF β interactions can trigger autocrine TGF β loops, magnifying the tumorigenic effects.

3.4 Sonic hedgehog

Throughout mammalian embryogenesis, the SHH signaling pathway emerges as a key mediator orchestrating tissue differentiation and human foregut formation. Consequently, an abundant level of SHH can be found in human gastric mucosa, contributing to epithelial growth, gastric acid production and potential neoplastic transformation (Merchant, 2012; van den Brink et al., 2002). Additionally, SHH-mediated organogenesis is also observed in the maturation of the pancreas, development of the gallbladder, and the repairing of liver tissue (Swiderska-Syn et al., 2016; Jeng et al., 2020).

While SHH activity typically remains dormant in mature adults, aberrations in the SHH pathway can reawaken its influence, promoting cancer initiation and progression (Jeng et al., 2020). For instance, mutated SHH pathways have been discovered in the activation of human pancreatic myofibroblast invasion *in vitro*, underscoring SHH's role in fostering desmoplasia in PDAC and subsequently supporting CSC metastasis (Bailey et al., 2008). Additionally, research by Jeng et al. has demonstrated the expansion of HCC *in vivo* following SHH gene alterations, suggesting its regulatory impact on HCC growth (Jeng et al., 2015). Moreover, by employing xenotransplantation in patient-derived CRC organoids, Regan et al. has elucidated SHH's role in promoting CSC survival via crosstalk with the WNT pathway (Regan et al., 2017).

3.5 Notch

The Notch signaling pathway regulates genes associated with stem cell maintenance, exerting critical influence on cell proliferation, differentiation and tissue maturation (Zhou et al., 2022a). Upon initiation from receptor-ligand binding event, the Notch pathway sets off a series of downstream cascading effects. The ultimate product translocates to the nucleus (Andersson et al., 2011; Zhou et al., 2022a), engaging in transcriptional interactions that lead to the upregulation of genes such as HES, MYC, and CDKN1A, which encode proteins essential for maintaining CSC phenotype, initiating oncogenesis, and inhibiting apoptosis (Clara et al., 2020).

Notably, the Notch pathway exhibits dual effects in tumor progression, acting as an oncogene or a tumor suppressor depending on varied receptor/ligand expressions or upstream regulators (Li et al., 2023c). Aberrant overexpression of Notch, as observed in GC, PDAC, CRC and HCC (Vinson et al., 2016; Hibdon et al., 2019; Tyagi et al., 2020; Chung and Xu, 2023; Jeng et al., 2023), relates the Notch gene to tumor types characterized by increased aggressiveness, promoted drug resistance and poor clinical prognosis. The Notch pathway upregulates CSC activities by interacting with CSC surface markers. Based on an observed increase of Notch1 in ALDH^{high} cells in GC, Nishikawa et al. proposed that Notch pathway regulates CSC maintenance and induces chemotherapy resistance (Nishikawa et al., 2013). Abundant research on Notch-CSC interactions also reveals its contribution in promoting CSC self-renewal, tumor angiogenesis, and metastasis (Miao et al., 2017; Akil et al., 2021). Regarding immune system regulation, Notch is also found to induce immune evasion by promoting TAM activation and inhibiting T cell proliferation (Palaga et al., 2018; Meng et al., 2022).

However, Notch signaling also exhibits anti-tumor properties by influencing NK cell proliferation and DC cell maturation (Felices et al., 2014; Meng et al., 2016). Understanding the precise influence and regulators of Notch's dual effects in tumors, particularly GI tumors, is still in its early stages. Further investigations are needed to elucidate the complex mechanisms underlying the Notch signaling pathway and to gain a comprehensive understanding of tumor heterogeneity.

Interplay among diverse pathways contributes to coordinated regulation of tumorigenesis. For instance, the IL-6/JAK/ STAT3 pathway can induce a cascading effect, resulting in the upregulation of TGF β and the collaborative orchestration of EMT in HCC (Wang et al., 2018a). Conversely, a TGF β -induced CAF has been identified as a source of IL-11, which can activate STAT3 and further enhance metastasis in CRC (Calon et al., 2012). Therefore, a holistic perspective is imperative for the comprehensive exploration of the regulatory roles of signaling pathways in tumorigenesis.

4 Non-coding RNA

While more than 90% of genomes undergo a transcription process, only 2% are determined to be protein-encoding (Wilusz et al., 2009). With accumulating evidence revealing their cell-specific expression and distribution, the blank spaces left by the non-coding RNAs (ncRNAs) has recently drawn increasing attention. ncRNA includes microRNA (miRNA), long non-coding RNA (lncRNA) and circular RNA (circRNA), all proven to be critical in physiological and pathological processes.

4.1 MiRNA

MiRNAs, a group of small ncRNA with approximately 22 nucleotides (Bartel, 2004), block gene expression by binding to the 3'-UTR of target mRNA (Pan et al., 2021). Their impacts are highly heterogenous across different tumors, some promoting cancer stemness and drug resistance, while others demonstrating overall tumor suppressive effects.

MiRNAs modulate the expression of CSC biomarkers to exert control over cancer progression. For instance, miR-328 acts as an effective inhibitor of CD44 (Xue et al., 2016). Its reduced levels are associated with higher GC recurrence rates and enriched CD44⁺ cells, establishing a negative correlation between miR-328 and cancer development (Ishimoto et al., 2014). Targeting CD44, miR-145 also exerts tumor-suppressive effects, attenuating EMT responses and reducing chemotherapy resistance (Zeng et al., 2017; Zhu et al., 2018; Mozammel et al., 2023). Similar effects were observed in miR-17 and miR-1185, which inhibits the expression of SOX2 and CD24, respectively (Li et al., 2015a; Wang et al., 2020a).

Multiple pathway signaling reactions are also regulated by miRNA. MiR-1247-5p is found to be downregulated in HCC patients, suggesting its negative correlation with tumor progression. Further investigation unveiled its target as Wnt3, the inhibition of which induces CSC apoptosis in vitro and suppresses cancer cell growth in vivo (Chu et al., 2017). Similarly, miR-4319 emerges as a potent anticancer factor. Targeting multiple genes associated with GI tumors, it inhibits cell proliferation, reduces CSC stemness and suppresses EMT in HCC (Han et al., 2019), CRC (Huang et al., 2019a) and EC (Hu et al., 2020). Additionally, miR-30a and miR-9 downregulates TM4SF1 (Park et al., 2016; Park et al., 2017), a key oncogenic gene known for its role in activating the Wnt/ β-catenin pathway and enhancing EMT (Tang et al., 2020). MiRNAs also exert indirect regulatory effects via responding to TME alterations. MiR-215 is upregulated by hypoxia, a tumorinitiating condition in TME. However, as miR-215 operates in a negative feedback loop, the elevation of which curtails CSC biomarker LGR5 and suppresses tumor initiation (Ullmann et al., 2019).

Though the majority of CSC-related miRNAs demonstrate tumor-suppressive effects (Pan et al., 2021), a few exhibit CSCpromoting functions. Aberrant miR-5188 expression promotes HCC stemness by mediating β -catenin translocation (Lin et al., 2019), while miR-577, a highly heterogenous factor, shows an upregulation in GC (Luo et al., 2019), reflecting the complex and heterogeneous nature of these regulatory molecules. Further investigations are needed to clarify the mechanisms underlying the dysfunctional expression of CSC-associated miRNAs and explore therapeutic strategies employing tumor suppressor miRNAs or inhibiting oncomiRNAs.

4.2 LncRNA

LncRNAs are noncoding transcripts that contain more than 200 nucleotides (McCabe and Rasmussen, 2021). By initiating chromatin modification on protein-coding genes, lncRNA can exert alterations in transcription, pro-transcription and translation (Castro-Oropeza et al., 2018; McCabe and Rasmussen, 2021).

LncRNAs play an active role in determining cellular fate and engaging directly with oncogenic regulatory factors. Among them, Hox transcript antisense intergenic RNA (HOTAIR) has been identified in CRC, GC, PDAC and HCC (Castro-Oropeza et al., 2018). It regulates EMT-associated gene expression by recruiting chromaffin-mediating enzymes and inducing the activation or silencing of target genes. Through transcriptional alterations, HOTAIR upregulates factors such as ZEB1, TWIST and SNAIL, meanwhile stimulates TGF\$1 pathway (Pádua Alves et al., 2013), instigating EMT and CSC clonogenicity, thereby promoting tumor metastasis and growth (Castro-Oropeza et al., 2018). Similarly, metastasis-associated lung adenocarcinoma transcript 1 (MALAT-1) was found to promote tumorigenesis in CSC derived from PDAC, CRC and GC (Hu et al., 2016; Chen et al., 2017; Wu et al., 2018). Same as most oncogenic lncRNAs, MALAT-1 elevates the expression of EMT-associated factors (McCabe and Rasmussen,

2021), maintains CSC renewal and induces angiogenesis by stabilizing SOX2 mRNA (Jiao et al., 2015).

Moreover, lncRNAs can function as miRNA sponges with specific effects. CAF-derived lncRNA H19 amplifies CRC stemness by acting as miRNA-41 sponge, whose enrichment activates the β -catenin pathway, elevating CSC stemness and chemoresistance *in vitro* and *in vivo* (Ren et al., 2018). Similarly, lncRNA NALT1 correlates with increased mortality rate in GC and CRC patients via sponging miRNA574-5p, which mediates the elevation of PEG10, a gene responsible for CSC proliferation and invasion (Ye et al., 2022).

4.3 CircRNA

CircRNAs are a class of single-stranded nucleotides characterized by their covalently closed loop structure. Each circRNA undergoes back-slicing, in which the absent 5' and 3' of pre-mRNA bind together to form a structure with enhanced stability (Liu and Chen, 2022).

With their abundant expression levels, circRNA exhibits tumorpromoting properties by functioning as regulators of transcription and splicing. CRC-derived circCTIC1 was found to correlate to worse prognosis of CRC patient. Research uncovered its interaction with a protein complex in the nucleus, directly triggering the expression of c-Myc, a significant tumor-promoting gene (Zhan et al., 2019). Another circRNA, circFNDC3B, has been associated with EMT levels in GC, contributing to the downregulation of E-cadherin expression and interacting with CD44 mRNA to bolster aggressive invasion (Hong et al., 2019). Another mechanism proposes that, the generation of circRNAs through the back-splicing mechanism disrupts alternative linear splicing of pre-mRNA, potentially influencing gene expression levels (Samavarchi Tehrani et al., 2023).

Similar to lncRNA, circRNA can also be employed as miRNA sponges. In a genome-wide sequencing study focused on CRC-associated RNAs, Rengganaten et al. elucidated a circRNA-miRNA-mRNA axis mainly mediated by circRNA hsa_circ_0066631 and hsa_circ_0082096. These circRNAs take effects via the downregulation of target miRNAs, thus inhibiting the degradation of mRNAs encoding CSC-promoting proteins, including activators of WNT/ β -catenin and TGF β (Rengganaten et al., 2020). Notably, though circRNA is mostly investigated as ncRNA, emerging evidence suggests their mRNA-resembling potential in protein encoding (Pamudurti et al., 2017), opening a promising field for further research.

5 Tumor microenvironment

Tumor microenvironment (TME) encompasses a diverse array of non-tumorigenic cells, including immune cells, endothelial cells, mesenchymal stroma-like cells (MSC) and CAF, all embedded in a matrix of structured proteins forming ECM (Osipov et al., 2019). TME form specialized compartments called niches, which reside CSCs and regulate cellular activities by intracellular contacts or secreted factors including cytokines, growth factors and hormones (Plaks et al., 2015). Current discussions are now investigating how niches in GI tumors contribute to the survival, growth, differentiation, migration and therapeutic resistance of CSC.

5.1 Immune cell moderations

TME is characterized by a chronic inflammatory state (Plaks et al., 2015). A growing body of evidence have demonstrated a negative correlation between cellular stemness and immune response within the niche (Miranda et al., 2019; Zheng et al., 2022). CSC-derived immunosuppressive factors, including various anti-apoptotic proteins, are its direct means of self-protection (Olver et al., 2006; Abdullah and Chow, 2013). Mediated by CSC-tumor system interactions, TME is exhibited as a milieu for immune detection avoidance (Clara et al., 2020). In CRC, GC and PDAC, studies have consistently highlighted a link between diminished populations of cytotoxic cells and lower survival rates, as well as unfavorable clinical outcomes (Peng et al., 2017; Yu et al., 2018; Zheng et al., 2022). Therefore, it is of great importance to understand the mechanisms orchestrating CSC-mediated immune evasion, and to elucidate the regulatory roles of immune cells in shaping CSC phenotypes.

5.1.1 Tumor-infiltrating lymphocytes

Tumor-infiltrating lymphocytes, one of the major constituents of TME, refers to the lymphocytes that migrate from the blood to the site of the tumor, including CD8⁺ T cells, CD4⁺ T cells, B lymphocytes and NK cells (Yang et al., 2021). Among the trending cancer therapies, immune check point inhibitors is one of the most promising avenues. PD-L1, one of the most studied targets, can attenuate T-cell activity signals both by directly binding PD-1 and by interacting with inhibitory CD80 (Keir et al., 2008; Sun et al., 2018). Another immune checkpoint mediator, CTLA-4, competes with the CD28 receptor on T cells for binding to CD80, thereby curbing CD8⁺ T cell activity (Farhood et al., 2019).

CD8⁺ T cells are the main immune effectors exhibiting cytotoxicity. However, CSCs can escape CD8⁺ recognition by decreasing the expression of MHC-1 molecule (Morrison et al., 2018), and by elevating CD80 level in a TGF β -dependent, CTLA-4 mediated manner (Miao et al., 2019), thereby acquiring immune resistance. Meanwhile, T cell activity is suppressed via upregulated PD-L1 expression on CSC surfaces (Wu et al., 2017b), as observed in various cancer types including CRC (Wei et al., 2019; Zheng et al., 2022) and GC (Sun et al., 2020).

In contrast, CD4⁺ T cells act not as effectors but regulators of immune responses, mediating the proliferations and activities of cytotoxic cells. Regulatory T (Treg) cells, among the CD4⁺ T cell subtypes, primarily suppress immune responses, inhibiting the cytotoxic functions of CD8⁺ T cells and NK cells (Liu et al., 2019). Particularly, the CD4⁺CD25⁺ T cells, marked by upregulated FOXP3 mRNA expression and intracellular CTLA-4 molecules, are related to poorer prognosis and survival rates in patients with GC and EC (Kono et al., 2006). The crosstalk between CSC and Treg significantly contributes to the immunosuppressive milieu of the TME. Liver CSCs, for instance, elevate Treg infiltration by upregulating PD-L1 and TGF β 1 (Wu et al., 2023). Reciprocally, Th17, another CD4+T cell subtype with occasional antiinflammatory phenotype (Sharma et al., 2009), is found in GC, CRC and PDAC as modulator of CSC self-renewal capacity (Su et al., 2014). By releasing IL17, Th17 can induce EMT-associated mitogenactivated protein kinase (MAPK) and AKT kinase activation, further augmenting CSC properties (Becerril-Rico et al., 2021).

5.1.2 Tumor-associated macrophages

Macrophages display remarkable plasticity and abundance in the CSC niche. TAMs exist with a wide range of spatial and temporal variance (Huang et al., 2019b), with TME-resided substances mediating their differentiation, dedifferentiation, and even reeducation (Bowman et al., 2016). TAMs are generally defined into two polarized types, the classically activated M1 with antitumor properties, and the alternatively activated M2 with immunosuppressive effects (Najafi et al., 2019), the later mainly manifested in established tumors.

CSCs play a critical role in recruiting M2 macrophages. In a comprehensive comparison of 26 CSC subsets within a CRC patient cohort, M2 macrophages stood out as one of the most substantial cell types in groups exhibiting a high stemness signature (Zheng et al., 2022), indicating its close association to CSC stemness. Many pro-tumorigenic macrophage factors are elevated in a CSC-enriched environment, including CCL2 and macrophage colony-stimulating factor 1 (CSF1), the main governors of TAM survival and metastasis. In EC colony, the CCL2-CCR2 axis had been demonstrated as a critical M2 polarization factor, the blockade of which leads to decreased PD-L2 expression in TAMs and suppressed tumor invasiveness (Yang et al., 2020). Research on CRC assays underscore the similar M2-polarizing effects of CSF1, where an upregulated level of IL-10 and CCL2 was also observed, further linking CSC-derived substances with M2 population dynamics (Huang et al., 2021).

Reciprocally, TAM activity increases CSC survival rate and reinforces CSC phenotypes, as a study using HCC mouse model documented an increase in CSC-like traits in TAM coculture group (Fan et al., 2014). M2-dominated TAMs regulate immune responses within the tumor niche, actively contributing to cancer cell immune evasion. Interactions between TAM and other immunocytes display an overall immunosuppressive effect, suggesting TAM's role in neutralizing anti-tumor properties. In GC, TAMs impair NK-cell function and compromises its cytotoxicity by downregulating the expression level of IFNy, TNFa, and Ki-67 (Peng et al., 2017). Literature investigating TAM also suggested its close relationship with EMT. In GC, TAMs expressing CD68 correlate with reduced epithelial marker E-cadherin (Lin et al., 2015a) and upregulated forkhead box Q1 (FOXQ1) expression (Guo et al., 2017). TAM can also induce the secretion of multiple soluble factors including IL-6 and TGFB, respectively promoting CSC self-renewal and inducing EMT (Wan et al., 2014). Interestingly, although M2 macrophages exhibit comparably more phagocytotic phenotype (Najafi et al., 2019), it does not affect CSC survival. This is mediated by the upregulation of CD47 in CSCs, with a "do not eat me" signal observed in various cancer types including HCC and PDAC (Lee et al., 2014; Cioffi et al., 2015).

5.1.3 Dendritic cells

DC is the most powerful antigen presenting cell in human immune system. Mature DCs can effectively stimulate T cell toxicity and cross-present tumor antigens (Bayik and Lathia, 2021). However, DCs residing in TME often exhibit compromised maturation and display tolerogenic phenotypes. Meanwhile, DCs have been shown to promote early metastasis by inducing the proliferation of Treg and suppressing the activities of CD8⁺ T cells. In GC, DC is able to convert effector T cells into CD4⁺CD25+FOXP3+Treg (Liu et al., 2019), thus amplifying the immunosuppressive milieu. In PDAC models, DC-expressed PD-L2 and MGL2 are also found to govern tumorigenesis, the blockade of which in turn suppressed metastasis formation (Kenkel et al., 2017).

Latest evidence suggests reciprocal interactions between DCs and CSCs in GI tumors. DCs obtained from CRC patients were recorded to exhibit elevated expression level of CXCL-1, a substance responsible for activating proteins within the CSC signaling network including SOX2 (Hsu et al., 2018). On the other hand, while the precise mechanism through which CSCs enhance DC tumorigenicity remains unclear, several studies managed to establish positive connections indicating CSC's promotive effects in tumor-associated DCs(Grange et al., 2015).

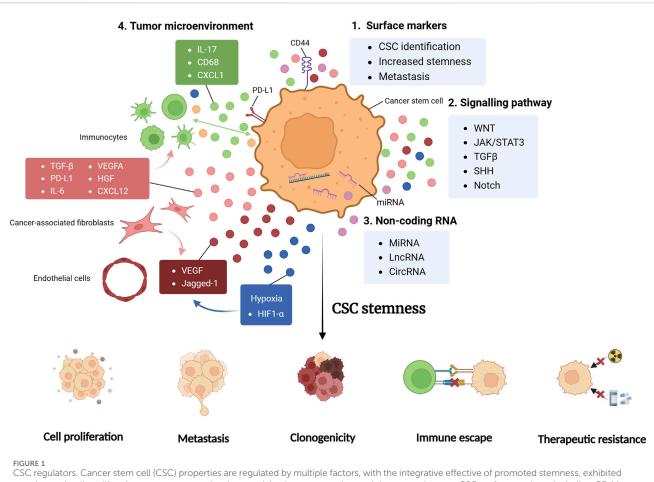
5.2 Cancer-associated fibroblast

CAFs are defined as a group of elongated cells located in cancer tissues that do not fall under epithelial, endothelial, or immune cell categories (Sahai et al., 2020). As the major constituent of ECM in GI organs (Kobayashi et al., 2019), they are characterized by high resilience (Kalluri, 2016) and heterogeneity, and are active participants in cell signaling mechanisms (Ohlund et al., 2014). CAFs have been shown to reshape ECM and establish communication networks with both tumorigenic and immune cells through the secretion of diverse chemokines and cytokines (Nurmik et al., 2020). This multifaceted interaction facilitates cell proliferation, migration, and angiogenesis, exerting a significant influence on the modulation of tumor immune responses while enhancing tumorigenic traits (Figure 1 and Figure 2).

5.2.1 Promoting CSC phenotype

CAFs actively participate in CSC phenotype promotion and stemness maintenance by secreting an array of cytokines, chemokines and growth factors, including IL-6, CXCL12, TGF β and hepatocyte growth factor (HGF).

As a key tumorigenesis factor, TGF\beta stands out as a major mediator of the crosstalk between CAF and other tumor cells (Wu et al., 2013). Once activated, TGF^β pathway not only triggers fibrosis within the TME but also serves as an initiator for metastasis, reshaping the TME dynamics, fueling CSC proliferation, and propelling tumor advancement (Calon et al., 2012; Shi et al., 2020). Moreover, CAF-derived CXCL12 can bind to cancer cellderived CXCR4, activating the CXCR4/CXCL12 axis and promoting CSC phenotypes, including its proliferation, invasion and metastasis (Orimo and Weinberg, 2006). Research targeting CRC cohorts revealed a poor prognosis related to an elevated CXCR4 and CXCL12 expression level (Saigusa et al., 2010), emphasizing their potential as malignancy markers. Studies in EC and GC have also demonstrated the cancer promoting properties of CAF-secreted IL-6, which promotes EMT and metastasis via JAK2/STAT3 signaling pathway (Wu et al., 2017a). Furthermore, various literature has



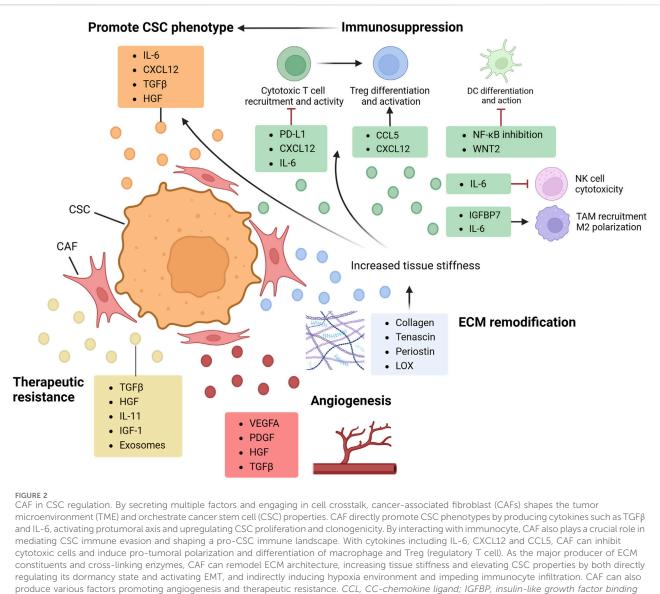
as enhanced cell proliferation, tumor metastasis, clonogenicity, immune evasion and therapy resistance. CSC surface markers, including CD44, are critical for the stemness maintenance and metastasis enhancement of CSC, and can serve as means of CSC identification in researches (1). Signaling pathways are formed by cytokines and chemokines secreted from tumor cells and various cells in tumor microenvironment (TME). These substances are involved in complex crosstalk and cellular interactions, shaping the CSC niche significantly (2). Non-coding RNA (ncRNA) including miRNA, lncRNA and circRNA are found to exert both pro-tumorigenic and antitumoral effects, fine-tuning TME and CSC properties. Some ncRNAs directly modulates genetic alterations, while others function by paracrine signaling (3). TME harbors various cell types, each secreting multiple factors and engages in complex crosstalk regulating CSC stemness (4). JAK/STAT3, Janus kinase/signal transducer and activator of transcription 3; TGF β , transforming growth factor beta; SHH, sonic hedgehog; miRNA, micro RNA; lncRNA, long non-coding RNA; circRNA, circular RNA; CXCL, CXC-chemokine ligand; PD-L1, programmed cell death 1 ligand 1; VEGFA, vascular endothelial growth factor A; HGF, hepatocyte growth factor; HIF1- α , hypoxia-inducible factor-1 α .

reported HGF's critical role in tumorigenesis. In HCC model, CAF can produce HGF, which regulates liver CSCs via activation of downstream effector FRA1, and promotes tumor growth process in a fibrosis-dependent manner (Lau et al., 2016). Similarly in CRC, HGF orchestrates CSC stemness via augmenting WNT pathway, restoring its phenotype even in well-differentiated tissues (Vermeulen et al., 2010).

5.2.2 ECM remodification

As the main producer of collagen, tenascin and periostin (Calon et al., 2012), CAF is a key factor in the remodeling and reorganization of ECM. By secreting matrix-crosslinking enzymes, CAF can induce force-mediated reconstruction and amplify the stiffness of tumor tissues. This altered ECM architecture triggers pro-survival signaling (Rice et al., 2017) and further intensifies the proliferation, metastasis and therapy resistance of cancer cells (Schrader et al., 2011; Mohammadi and Sahai, 2018). In HCC, fibrotic liver tissues display higher proliferative indices and reduced cancer cell apoptosis, suggesting an association between increased tissue stiffness and elevated cancer cell proliferation rates. Interestingly, cell clonogenicity have been found to increase in a soft, rather than stiff, microenvironment. Schrader et al. discovered that, in HCC cells treated with chemotherapeutic agent cisplatin and 5-Fluorouracil (5-FU), an enrichment of cells with surface marker CD133, CXCR4 and CD44 is observed in the soft supports instead of the hard ones. However, this phenomenon was absent in untreated cell groups, indicating the role of CSCs specifically in response to treatment. Meanwhile, escalated tissue stiffness may trigger the collapse of vasculatures (Bordeleau et al., 2017), inducing tissue hypoxia and blocking T cell migration toward tumor sites (Salmon et al., 2012).

CAF-mediated matrix remodification is also a potent trigger of local metastasis. In a study discussing CRC metastasis, Calon et al. proposed a Darwinian model in which cells with tumor-initiating competence need to acquire TGF β -associated functions and interact with CAF. The microenvironment shaped by TGF β , known as the TGF β niche, is filled with colonization-required stroma and prometastasis chemokines, both secreted by TGF β -stimulated CAFs



protein; LOX, lysyl oxidase; PDGF, platelet-derived growth factor.

(Calon et al., 2012). Another study identified PDAC-derived SPOCK-1 as a key stromal protein, and documented its capacity to elevate cancer cell growth through matrix reorganization (Veenstra et al., 2017).

5.2.3 Immunosuppression

Considering the CSC-promoting effects of immunocytes in TME, it is imperative to investigate CAF's engagement in immunocyte crosstalk as an indirect means of CSC regulation, including its role in evading immune surveillance and shaping the tumor immune landscape.

By interacting with collagen and reorganizing ECM structure, CAF can impede immune cell immigration (Sahai et al., 2020). Abnormal vasculature blocks T cell extravasation, while an altered intercellular adhesion state may disrupt functional diapedesis (Turley et al., 2015). CAF also modulates the immune landscape via the production of PD-L1, TGF β , CCL5 and CXCL12 (Ng et al., 2005; Ahamed et al., 2008; Turley et al., 2015). These cytokines collectively contribute to shaping an immunosuppressive TME, either by affecting downstream pathways, recruiting immunosuppressive factors, or by directly regulating immune cell properties.

CD8⁺ cytotoxic T cell activity is inhibited by CAF-derived TGF β , CXCL12 and IL-6. While CAF can directly produce PD-L1 (Turley et al., 2015), substances like TGF β and CXCL12 can also attenuate the anti-PD-L1 response (Cheng et al., 2018; Mariathasan et al., 2018). Moreover, CAFs recruit and activate FOXP3+ Tregs to suppress antitumor immune responses (Wang et al., 2023). CAFderived CCL5 is found to promote the survival, differentiation and activation of Tregs (Wang et al., 2017; Kieffer et al., 2020), further restraining immune surveillance. In HCC and PDAC, CAFs induce M2 polarization of macrophages, fostering an immunosuppressive environment (Mathew et al., 2016; Chen et al., 2021). By upregulating the level of IL-10, CAF in PDAC is found to

Secreted factor	Targeted pathway	Mechanism	Tumor type	Citation
IGF-1	IGF1/IGF1R	Induces metabolic changes; reduces cancer cell sensitivity to irradiation	CRC	Tommelein et al. (2018)
CXCL1	Mek/Erk	Regulates an ROS-dependent DNA damage repair	EC	Zhang et al. (2017b)
TGFβ	FOXO1/TGFβ1	Elevates epithelial cell proliferation and angiogenesis	EC	Zhang et al. (2017a)
HGF	ERK-MAPK	Provides BRAF-independent mechanism regarding BRAF-targeted therapies; resist tyrosine kinase inhibitors	CRC	Straussman et al. (2012), Luraghi et al. (2014)
IL-11	JAK/STAT3/Bcl2	Activates anti-apoptosis signaling pathway	GC	Ma et al. (2019a)
IL-17A	-	Induces viability of CIC	CRC	Lotti et al. (2013)
lncRNA H19	β-catenin	Competes with stemness-inhibiting factor miR-141	CRC	Ren et al. (2018)
miR-522	ALOX15	Suppresses ferroptosis	GC	Zhang et al. (2020)
Proteins (unidentified)	mTOR/4E-BP1	Secretes proteins that activate tumor cell survival	PDAC	Duluc et al. (2015)
Exosomes (unidentified)	WNT	Increases the percentage, clonogenicity and growth of CSCs	CRC	Hu et al. (2015)

TABLE 1 CAF-secreted soluble factors in therapeutic resistance of GI tumors.

activate Arg1 expression in macrophages, hence inducing alternative polarization. Furthermore, CAF in GC has been shown to secrete insulin-like growth factor binding protein 7 (IGFBP7), elevating TAM infiltration level and promoting tumor growth (Li et al., 2023a). Notably, TAMs are engaged in a crosstalk with CAFs, producing factors that further activate CAFs (Comito et al., 2014) and creating an intercellular network that fuels cancer growth and metastasis (Sathe et al., 2023).

5.2.4 Angiogenesis

Neovascularization, one of the major conditions for CSC maintenance, is regulated by various CAF-derived factors. As a critical angiogenetic factor, vascular endothelial growth factor A (VEGFA) is directly produced by CAFs (Kobayashi et al., 2019) or indirectly activated by CAF-secreted molecules (Wang et al., 2019a). A study in CRC discovered that, combined blockade of VEGFR and COX2 augmented antiangiogenic, antitumoral and anti-CSC effects, indicating VEGF's role in maintaining CSC properties (Zhao et al., 2015a). Other pro-angiogenic substances secreted by CAFs include platelet-derived growth factor (PDGF), CXCL12, fibroblast growth factor 2 (FGF2), HGF and TGF β (Anderberg et al., 2009; Wang et al., 2019a; Kobayashi et al., 2019; Hjazi et al., 2023).

5.2.5 Therapeutic resistance

An extensive body of research underscores the pivotal contribution of CAFs in fostering therapeutic resistance in cancer cells. Remodeled by CAF, the dense ECM acts as a physical barrier to impede drug deliveries. A study using PDAC mice model revealed notably elevated interstitial fluid pressures (IFP) and occurrences of vascular collapse in tumor tissues, (Provenzano et al., 2012), which are presented as delivery barriers induced by CAF's desmoplastic reactions.

Various CAF-secreted soluble factors also play significant roles in eliciting therapeutic resistance of GI tumors (listed in Table.1). These factors exert their effects by either obstructing therapy pathways, inducing rapid adaptations to altered milieu, or accelerating damage repair processes. Meanwhile, CAF's immunosuppressive properties, highlighted earlier, dampen the responses to immune checkpoint inhibitors, consequently diminishing the efficacy of conventional immunotherapies.

5.3 Angiogenesis and endothelial transformation

In TME, stress-induced angiogenesis provides a conducive niche for CSCs, while endothelial cells themselves exhibit properties that foster stemness (Yang et al., 2021). Studies on early tumor stages highlighted the spatial proximity between CSC niche and endothelial cells (Mercurio, 2019), indicating a reciprocal relationship established by paracrine signaling.

VEGF, a potent endothelial cell-specific mitogen, orchestrates endothelial development and pathological angiogenesis (Leung et al., 1989). It also plays the role of CSC promoter, as VEGF blockade results in declined CSC population and activities (Mercurio, 2019). Previous research primarily focused on squamous carcinoma and skin cancer, while recent findings in GC mouse models uncovered a KRAS-activated VEGF production in CSC (Yoon et al., 2023), suggesting CSC's role in regulating endothelial growth and neovascularization. Thus, antivascular therapies emerge as a promising approach to combat cancer metastasis (Lu et al., 2013).

Meanwhile, endothelial cells can perform CSC-promoting properties. Previous study has identified that Jagged-1, a soluble factor secreted by CRC endothelial cells, activates the Notch signaling pathway and promote CD133+ CSC self-renewal and proliferation *in vitro* and *in vivo* (Lu et al., 2013). Moreover, recent literature correlates a reduction in CSCs to the inhibition of a CD44 ligand, DLL4, derived from endothelial cells. Meconca et al. demonstrated that suppressing DLL4 in endothelial cells, even during induced tumoral neovascularization, resulted in insubstantially perfused vessels unable to support normal vascular functions, thereby inhibiting EMT and CSC activity (Mendonça et al., 2019).

5.4 Hypoxia

Aberrant ECM structure and vasculature in tumor tissues frequently leads to hypoxia, a major stressor that drives the adaptive properties and tumorigenicity of CSCs. Consequently, hypoxic environment favors cells with CSC-like features due to clonal selection, fostering malignant properties (Kim et al., 2018). A study using differential gene expression analysis revealed a strong correlation between tumor diversity and hypoxia-related genes (Ma et al., 2019b). Upregulated levels of hypoxia-inducible factor-1a (HIF1-a), when stabilized by CAF-derived TGFβ, can activate the expression of SHH transcription factor GLI2 in colorectal CSCs, consequently maintaining CSC stemness, promoting dedifferentiation and metastasis (Tang et al., 2018). In various cancers, other proposed mechanisms including the ROS-activated stress response pathway (Liu et al., 2008) and the Notch signaling pathway (Quail et al., 2012) further associate hypoxia with the increased stemness, survival rate and functions of CSC.

The hypoxic environment directly increases HIF1- α levels, triggering downstream hypoxia signaling and stimulating the production of VEGF (Goel and Mercurio, 2013), which plays a major role in angiogenesis and TAM recruitment (Kitamura et al., 2015). Infiltrating TAMs, once migrated to the site, engage in crosstalk with VEGF, further enhancing VEGF-A production and promoting tumor angiogenesis. Additionally, hypoxic CSCs impede T cell activities by producing STAT3, which acts as a transcriptional source for HIF1- α and VEGF, thus exhibiting a hypoxia-induced immunosuppressive effect (Wei et al., 2011).

6 Therapies targeting stemness regulation

Conventional cancer therapies, including chemotherapies and radio therapy, are all currently challenged with severe limitations due to the failure to eliminate CSCs. Without proper eradication, dormant CSCs can be reactivated to trigger cancer relapse. Therefore, novel therapeutic researches are focusing on inhibiting CSCs by targeting the substances engaged in CSC stemness regulation.

6.1 Immunotherapy

6.1.1 Antibodies targeting biomarkers

Bispecific antibodies (BiAbs), representing an innovative strategy in immunotherapy, can simultaneously bind both T cell and tumor cells expressing specific antigens. The binding activates an MHC-independent immune response leading to cell lysis, thereby exhibiting antitumor effects regardless of tumor cell's immune evasiveness (Bargou et al., 2008). Similar approach can be employed to target CSCs utilizing CSC-specific surface markers such as CD133 and EpCAM.

Targeting CD133, Zhao et al. devised a CD133 monoclonal antibody (MS133) with dual-antigen-binding specificity to CD133 and CD3. MS133 exhibited significant cytotoxicity in CD133^{high} CRC cells, demonstrating enhanced stability and prolonged cytotoxic activity (Zhao et al., 2015b). Furthermore, solitomab, a single chain antibody, can bispecifically target EpCAM and CD3. Preclinical evidence revealed its potential in reducing EpCAM + cell group in PDAC and EC both in vitro and in vivo (Cioffi et al., 2012; Yu et al., 2021). Clinically, a Phase I study in patients with refractory solid tumors (including CRC and GC) reported a modest inhibitory effect of solitomab on the metastasis and progression of EpCAM + cancer (Kebenko et al., 2018). Similarly, catumaxomab, a drug originally approved for treating malignant ascites, targets on EpCAM and CD3 in latestage GI tumors. Though withdrawn from the market in 2017, associated studies on catumaxomab have not stopped, with ongoing clinical trials focusing on its efficacy in GC patients (NCT04222114, NCT01504256).

6.1.2 CSC-based vaccines

Compromising DCs' antigen presentation function is one of the most important strategies in CSC immune evasion. However, CSCbased vaccines can directly expose the CSC-unique antigens to the immune system, enhancing CSC recognition and activating robust immune effects (Hashemi et al., 2023), regardless of the decreased MHC-1 expression. The induction of strong immune responses can also elicit an immune memory, which reduces the likelihood of cancer recurrence (Kishi et al., 2021; Hashemi et al., 2023). The therapy features the injection of processed CSCs from autologous or allogeneic origins. The injected substance could be whole CSC-based vaccines made from previously inactivated CSCs, or CSC-lysates loaded in antigen-presenting cells (APCs), mostly DCs (Izadpanah et al., 2023).

The efficacy of this novel approach is supported by abundant preclinical evidence (Perez-Banos et al., 2023). In CRC mouse models, a whole-CSC vaccine made from CD133+ CSC lysates triggered the increase of NK cell cytotoxicity and secretion of inflammatory chemokines such as IFN- γ , which subsequently inhibits the proliferation of CD133+ and ALDH + cell groups, indicating a reduction in CSC proliferation (Guo et al., 2020). In HCC mouse models, Li et al. demonstrated that DC vaccine induced T cell activity *in vivo*, resulting in decreased tumor mass growth (Li et al., 2016). Enhanced T cell cytotoxic activity was observed in GC cell lines when utilizing a DC-CSC mRNA vaccine in cell co-culture (Bagheri et al., 2019).

Progress in clinical studies also supported the potential of CSCbased vaccines. In the first clinical trial of pancreatic CSC vaccine launched by Lin et al., a pancreatic CSC-derived vaccine was injected at low, medium and high dose to three groups of PDAC patients. With a significantly obvious response in the high-dose group, this trial preliminarily validates the efficacy of CSC vaccine as a PDAC therapy (Lin et al., 2015b). Moreover, Lau et al. carried a Phase I study featuring patients with PDAC resection. The injected vaccine, combining autologous DC and allogeneic CSC lysates, has shown a promising outcome in containing cancer metastasis rate (Lau et al., 2022). Ongoing clinical trials are also investigating the efficacy of CSC vaccines in other GI malignancies, including CRC, GC and HCC (NCT01885702, NCT04912765, NCT04147078). Further researches are required regarding DC subtypes and their interactions with substances in the TME, and a refined categorization of CSC biomarkers is needed (Izadpanah et al., 2023) to ensure the targeting efficacy and precision of the vaccines.

6.1.3 CAR therapy

The chimeric antigen receptor (CAR) therapy features the genetic modification of cytotoxic cells, mostly T cell or NK cell. Instead of targeting MHC in the conventional way, the modifiable extracellular region serves as a chimeric receptor, which enables the cell to bind any surface antigen of interest, making it a customizable tool to directly target tumor associated markers (Garza Trevino et al., 2023). This flexibility makes CAR therapy a versatile tool in detecting and eliminating CSCs, with CD133 and EpCAM as the most recruited CSC markers to date.

Preclinical studies highlight the efficacy of CAR therapy in antitumor activities, especially in combination with chemotherapy. Han et al. tested the efficacy of anti-CD133 CAR-T and cisplatin combination therapy in GC xenograft models (Han et al., 2021). Cisplatin is a first line chemotherapeutic drug against GC, but has shown limited effect due to CSC stemness and drug resistance, even upregulating CD133 expression when applied alone. However, with anti-CD133 CAR-T, cisplatin effectively induces the release of cytokine, and with the pronounced cytotoxicity of CAR-T cells, they jointly diminish the growth and infiltration of CD133⁺ cell. Moreover, Zhang et al. investigated the combination therapy of regorafenib and CAR-NK-92 cells in established CRC models (Zhang et al., 2018). Targeting EpCAM⁺ cells significantly increased cytokine levels and suppressed tumor growth, compared to monotherapy with regorafenib or CAR-NK-92.

In a Phase I study of CAR-T cell directed against CD133 (CART-133), patients with advanced solid tumor (PDAC, CRC, HCC) were enrolled and infused with CART-133. Tissue biopsy revealed effective elimination of CD133 cell population, and the trial concluded promising disease stable rate and progression-free survival of CART-133 (Wang et al., 2018b). Another CART-133 Phase II study conducted by Dai et al. has reached similar result. The trial recruited advanced HCC patients, and also recorded promising clinical response and manageable CART-133 safety profile. The level of prognostic indicators, such as proangiogenic and inflammatory factors, were also tracked and reported (Dai et al., 2020).

Further investigation and application of CAR therapy requires comprehensive research on CSC biomarkers, as a large percentage of CSC markers are also expressed on normal tissue cells (Kim and Ryu, 2017). Therefore, recognition of CSC-specific antigens needs to be improved regarding tissue-specificity to enhance therapy efficacy and to avoid on-target/off-tumor toxicity (Izadpanah et al., 2023).

6.1.4 Immune check point inhibitor

As previously highlighted, the overexpression of immune check points, notably CTLA-4 and PD-L1, is crucial in CSC's immune evasion. Thus, inhibiting the immune check points could be an effective approach to reinvigorate anti-tumor immune responses.

Multiple ongoing clinical trials are investigating the safety and feasibility of existing drugs, as well as potential combined therapies. Nivolumab is a PD-1 inhibitor that shows high clinical overall survival rate, and has been applied as first-line solid tumor chemotherapy. It is frequently used in combination therapy with ipilimumab, a human mono-antibody that blocks CTLA-4 activation. A randomized Phase III study showed that, in untreated patients with EC and GC, the nivolumab and therapy achieved ipilimumab combined а significant improvement in overall survival rate (Janjigian et al., 2021). Similarly, a Phase II study in CRC patients exhibited promising progression-free outcomes with the nivolumab plus ipilimumab combination (Lenz et al., 2022). A randomized trial investigating HCC resection's postoperative use of this therapy also concluded a lower recurrence rate, further expanding its application scope (Kaseb et al., 2022). The effect of other immune check point inhibitors in GI cancer, including CXCR4 inhibitor plerixafor, PD-1 inhibitor camrelizumab and pembrolizumab, are all investigated by ongoing clinical trials (Muro et al., 2016; Doi et al., 2018; Xu et al., 2019; Parikh et al., 2021; Li et al., 2023b) (NCT02179970, NCT02658019).

6.2 Targeting signaling pathways

CSCs rely on multiple cellular signaling pathways to maintain its stemness and tumorigenicity. Therefore, therapies targeting aberrantly triggered pathways can downregulate these tumoral effects and eliminate CSCs for cancer eradication.

6.2.1 Salinomycin

Salinomycin, a natural polyether product isolated from *Streptomyces albus*, has been suggested by many preclinical researches as a potent anti-tumor drug. Salinomycin effectively blocks WNT pathway by suppressing the formation of the β -catenin/TCF4E complex, subsequently downregulating the expression of WNT-associated genes LEF1 and LGR5 (Wang et al., 2019c) and inhibiting the growth of cancer cells. Moreover, salinomycin is known as an inhibitor of ATP-binding cassette (ABC) transporters, thereby compromising the drug resistance displayed by CSCs. It also exerts direct effects on CSCs by inducing cell apoptosis and DNA damage, or by promoting CSC differentiation (Naujokat and Steinhart, 2012).

Preclinical studies have highlighted salinomycin as a promising agent for eliminating CSC in CRC, GC and HCC (Dong et al., 2011; Zhi et al., 2011; Wang et al., 2020c). Additionally, combination therapies have gained interests in experimental studies. In PDAC models, Zhang et al. demonstrated that simultaneous administration of salinomycin and gemcitabine, a first-line drug commonly targeting non-CSC tumor cells, resulted in better containment of tumor growth and cancer cell viability compared to using either drug alone (Zhang et al., 2011). Current studies are also exploring novel drug delivery methods including nanocapsules or nanocrystals, with evidence validating for their capacity to enhance efficacy and bioavailability in CRC patients (Wang et al., 2020b; Tsakiris et al., 2020).

6.2.2 Sulforaphane

Sulforaphane is an isothiocyanate found in cruciferous vegetables like broccoli and cauliflower. Extracted and purified sulforaphane is reported to exert cancer-preventive functions including apoptosis induction, angiogenesis inhibition and metastasis suppression (Elkashty and Tran, 2021). Recent

evidence suggests sulforaphane as a potent CSC inhibitor across multiple cancer types, as it targets crucial signaling pathways including WNT, Notch, SHH and NF- κ B (Naujokat and McKee, 2021). Moreover, in PDAC, sulforaphane is found to deter CSC selfrenewal by inhibiting EMT-associated proteins such as vimentin, TWIST-1 and ZEB1 (Srivastava et al., 2011).

Sulforaphane administration in PDAC xenograft models was associated to NF-kB binding and subsequent apoptosis activation. In the preclinical study conducted by Kallifatidis et al., treated groups are found with suppressed angiogenesis and tumor growth, with no apparent cytotoxicity observed in normal cells (Kallifatidis et al., 2009). Tracking SHH pathway components in vivo also reveals sulforaphane's function in SHH blockade, accompanied with reduced PDAC tumor size and inhibited EMT activities (Li et al., 2013). Sulforaphane's anti-tumor effects has also been preclinically demonstrated in CRC and HCC (Liu et al., 2017; Ren et al., 2017; Chen et al., 2018; Chen et al., 2020). Large-scale clinical transformation, however, is challenged by the limited efficiency of drug delivery (Elkashty and Tran, 2021). The most recent clinical investigation is conducted by Lozanovski et al., with capsules of pulverized broccoli sprouts (containing 90 mg/ 508 µmol sulforaphane) administered to PDAC patients (Lozanovski et al., 2020). Though the results were not statistically significant, treatment group showed slightly prolonged survival, which provides preliminary evidence for the feasibility of sulforaphane application against GI cancer.

6.2.3 Napabucasin

Napabucasin, also known as BBI608, is an orally available STAT3 inhibitor. By reducing STAT3-induced gene transcription, it can exert anti-CSC effects and effectively prevent cancer metastasis and recurrence (Sonbol et al., 2019). Li et al. demonstrated its capability to impede spherogenesis and eliminate CSC-like cells, as well as inhibit cancer metastasis and relapse (Li et al., 2015b). These effects are observed across multiple cancer cell lines, including those originated from liver, pancreas and colon, suggesting napabucasin as a potent CSC-inhibitor in GI malignancies. Further evidence indicates that the administration of napabucasin facilitates the overall responsiveness of cancer cells to conventional therapies (Flebbe et al., 2022), providing new strategies for designing efficient combination therapies.

In a Phase lb/ll study, the combination therapy of napabucasin and bevacizumab was administered to patients with metastatic CRC, which demonstrated prolonged survival rate and contained tumor growth in treated group compared to placebo group (Bendell et al., 2017). In an international multi-center randomized Phase III study (NCT02753127) with 1253 CRC patients, napabucasin efficacy in combination with 5-FU, Leucovorin, and Irinotecan (FOLFIRI) was evaluated among adult patients (Grothey et al., 2016). Another open label, multi-center, Phase I/II dose escalation study (NCT02024607) included 495 adult patients with advanced GI tumor. Results highlighted that napabucasin administration effectively controlled cancer metastasis and significantly promoted progression-free survival rate. However, several completed clinical trials fail to acquire anticipated outcomes, including a Phase III study on PDAC (NCT02993731) and the Phase III BRIGHTER trial on GC (NCT02178956). These studies either detected no significant differences in treatment groups or recorded reverse effects.

6.3 Targeting ncRNA

NcRNAs can regulate multiple CSC properties, including cell stemness, therapy resistance and metastatic abilities. Therefore, targeting ncRNAs, either by silencing or upregulating specific ncRNAs, stands as a promising avenue for novel therapeutic approaches (Hwang et al., 2021). In a Phase I study in patients with advanced solid tumors, MRX34, a liposomal miR-34a mimic, was intravenously administered in a dosage escalating regimen (Hong et al., 2020). Despite participants encountering manageable adverse effects including fever, chills, and nausea, the treatment showcased a dose-dependent capability to downregulate targeted oncogenes. Though the trial was terminated due to fatal immune-mediated adverse effects, it provided preliminary proof validating the feasibility of miRNA-based therapies. Moreover, a multitude of ongoing clinical studies use ncRNAs as a prognostic tool in the early detection of GI cancers, including Project CADENCE in Singapore (NCT05633342), the Sibling AN Study in Hong Kong, China (NCT01593098), and a single blind, case control, multi-center study in Shanghai, China (NCT05431621).

7 Conclusion

Extensive research efforts have shed light on the profound impact CSCs exert in the initiation, progression, metastasis and recurrence of malignancies. This theory, accommodating the factors behind tumor heterogeneity, has no doubt enriched our understanding of the molecular processes underlying cancer progressions. Emerging researches targeting CSC regulation has provided new insights in recruiting prognostic tools and developing novel therapeutic targets.

However, investigations into CSCs confront multifaceted challenges. Identifying CSCs primarily relies on cell surface markers, a methodology prone to inaccuracy due to their nonuniversal distribution. Even widely-located markers, such as CD44 and CD133, have been found to display diverse effects influenced by spatial and temporal heterogeneity (Huang et al., 2013; Miwa et al., 2017). Thus, questions addressing a more reliable strategy for CSC identification urge answers, including the number of exhibited markers needed to signify a CSC population; whether each cancer type possesses an exclusive set of markers; and whether CSC biomarker distribution alter with cancer progression.

Moreover, current findings on CSC regulation often overlook the heterogeneity among GI cancers. Due to the plasticity of CSC and the dynamic TME, regulators could exhibit diverse effects regarding spatial or temporal differences. Certain signaling pathways exhibit both tumorigenicity and anti-tumor effects (Katsuno et al., 2012; Esebanmen and Langridge, 2017; Peng et al., 2022), so the complete blockade of which may diminish its effect in tumor suppression. Furthermore, the intricate interplay within the TME suggests that inhibiting one factor might alter the effectiveness of pathways, making therapeutic outcomes unpredictable. On the other hand, this interactive complexity could lead to ineffectiveness in targeting sole factor, as alternative pathways could compensate tumorigenesis.

Therefore, dynamic updates are needed regarding CSC heterogeneity and the comprehensive interactions among influencing factors. Meanwhile, novel cancer treatments could

recruit a comprehensive strategy, combining CSC eradication, blockade of regulatory pathways and chemo-sensitization, while customizing therapies based on specific cancer types (Chen et al., 2013a; Du et al., 2022).

Author contributions

KY: Writing-original draft. TY: Writing-review and editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

References

Abdullah, L. N., and Chow, E. K. (2013). Mechanisms of chemoresistance in cancer stem cells. *Clin. Transl. Med.* 2, 3. doi:10.1186/2001-1326-2-3

Aguirre-Ghiso, J. A. (2007). Models, mechanisms and clinical evidence for cancer dormancy. *Nat. Rev. Cancer* 7, 834–846. doi:10.1038/nrc2256

Ahamed, J., Burg, N., Yoshinaga, K., Janczak, C. A., Rifkin, D. B., and Coller, B. S. (2008). *In vitro* and *in vivo* evidence for shear-induced activation of latent transforming growth factor-beta1. *Blood* 112, 3650–3660. doi:10.1182/blood-2008-04-151753

Akil, A., GutiéRREZ-GarcíA, A. K., Guenter, R., Rose, J. B., Beck, A. W., Chen, H., et al. (2021). Notch signaling in vascular endothelial cells, angiogenesis, and tumor progression: an update and prospective. *Front. Cell Dev. Biol.* 9, 642352. doi:10.3389/ fcell.2021.642352

Anderberg, C., Li, H., Fredriksson, L., Andrae, J., Betsholtz, C., Li, X., et al. (2009). Paracrine signaling by platelet-derived growth factor-CC promotes tumor growth by recruitment of cancer-associated fibroblasts. *Cancer Res.* 69, 369–378. doi:10.1158/ 0008-5472.CAN-08-2724

Andersson, E. R., Sandberg, R., and Lendahl, U. (2011). Notch signaling: simplicity in design, versatility in function. *Development* 138, 3593–3612. doi:10.1242/dev.063610

Bagheri, V., Abbaszadegan, M. R., Memar, B., Motie, M. R., Asadi, M., Mahmoudian, R. A., et al. (2019). Induction of T cell-mediated immune response by dendritic cells pulsed with mRNA of sphere-forming cells isolated from patients with gastric cancer. *Life Sci.* 219, 136–143. doi:10.1016/j.lfs.2019.01.016

Bailey, J. M., Swanson, B. J., Hamada, T., Eggers, J. P., Singh, P. K., Caffery, T., et al. (2008). Sonic hedgehog promotes desmoplasia in pancreatic cancer. *Clin. Cancer Res.* 14, 5995–6004. doi:10.1158/1078-0432.CCR-08-0291

Bargou, R., Leo, E., Zugmaier, G., Klinger, M., Goebeler, M., Knop, S., et al. (2008). Tumor regression in cancer patients by very low doses of a T cell-engaging antibody. *Science* 321, 974–977. doi:10.1126/science.1158545

Bartel, D. P. (2004). MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 116, 281–297. doi:10.1016/s0092-8674(04)00045-5

Bayik, D., and Lathia, J. D. (2021). Cancer stem cell-immune cell crosstalk in tumour progression. *Nat. Rev. Cancer* 21, 526–536. doi:10.1038/s41568-021-00366-w

Becerril-Rico, J., Alvarado-Ortiz, E., Toledo-Guzman, M. E., Pelayo, R., and Ortiz-Sanchez, E. (2021). The cross talk between gastric cancer stem cells and the immune microenvironment: a tumor-promoting factor. *Stem Cell Res. Ther.* 12, 498. doi:10.1186/s13287-021-02562-9

Bendell, J., Hubbard, J., O'Neil, B. H., Jonker, D., Starodub, A., Peyton, J., et al. (2017). Phase 1b/II study of cancer stemness inhibitor napabucasin in combination with FOLFIRI +/– bevacizumab (bev) in metastatic colorectal cancer (mCRC) patients (pts). *Ann. Oncol.* 28, 150–151. doi:10.1093/annonc/mdx302.002

Bordeleau, F., Mason, B. N., Lollis, E. M., Mazzola, M., Zanotelli, M. R., Somasegar, S., et al. (2017). Matrix stiffening promotes a tumor vasculature phenotype. *Proc. Natl. Acad. Sci. U. S. A.* 114, 492–497. doi:10.1073/pnas.1613855114

Bowman, R. L., Klemm, F., Akkari, L., Pyonteck, S. M., Sevenich, L., Quail, D. F., et al. (2016). Macrophage ontogeny underlies differences in tumor-specific education in brain malignancies. *Cell Rep.* 17, 2445–2459. doi:10.1016/j.celrep. 2016.10.052

Busenhart, P., Montalban-Arques, A., Katkeviciute, E., Morsy, Y., Van Passen, C., Hering, L., et al. (2022). Inhibition of integrin $\alpha\nu\beta 6$ sparks T-cell antitumor response and enhances immune checkpoint blockade therapy in colorectal cancer. *J. Immunother. Cancer* 10, e003465. doi:10.1136/jitc-2021-003465

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Calon, A., Espinet, E., Palomo-Ponce, S., Tauriello, D. V., Iglesias, M., Cespedes, M. V., et al. (2012). Dependency of colorectal cancer on a TGF- β -driven program in stromal cells for metastasis initiation. *Cancer Cell* 22, 571–584. doi:10.1016/j.ccr.2012. 08.013

Calon, A., Lonardo, E., Berenguer-Llergo, A., Espinet, E., Hernando-Momblona, X., Iglesias, M., et al. (2015). Stromal gene expression defines poor-prognosis subtypes in colorectal cancer. *Nat. Genet.* 47, 320–329. doi:10.1038/ng.3225

Castro, F., Cardoso, A. P., GonçALVES, R. M., Serre, K., and Oliveira, M. J. (2018). Interferon-gamma at the crossroads of tumor immune surveillance or evasion. *Front. Immunol.* 9, 847. doi:10.3389/fimmu.2018.00847

Castro-Oropeza, R., Melendez-Zajgla, J., Maldonado, V., and Vazquez-Santillan, K. (2018). The emerging role of lncRNAs in the regulation of cancer stem cells. *Cell Oncol.* (*Dordr*) 41, 585–603. doi:10.1007/s13402-018-0406-4

Chen, B., Mu, C., Zhang, Z., He, X., and Liu, X. (2022). The love-hate relationship between TGF- β signaling and the immune system during development and tumorigenesis. *Front. Immunol.* 13, 891268. doi:10.3389/fimmu.2022.891268

Chen, D., Liu, L., Wang, K., Yu, H., Wang, Y., Liu, J., et al. (2017). The role of MALAT-1 in the invasion and metastasis of gastric cancer. *Scand. J. Gastroenterol.* 52, 790–796. doi:10.1080/00365521.2017.1280531

Cheng, Y., Li, H., Deng, Y., Tai, Y., Zeng, K., Zhang, Y., et al. (2018). Cancerassociated fibroblasts induce PDL1+ neutrophils through the IL6-STAT3 pathway that foster immune suppression in hepatocellular carcinoma. *Cell Death Dis.* 9, 422. doi:10. 1038/s41419-018-0458-4

Chen, K., Huang, Y. H., and Chen, J. L. (2013a). Understanding and targeting cancer stem cells: therapeutic implications and challenges. *Acta Pharmacol. Sin.* 34, 732–740. doi:10.1038/aps.2013.27

Chen, S., Morine, Y., Tokuda, K., Yamada, S., Saito, Y., Nishi, M., et al. (2021). Cancer-associated fibroblast-induced M2-polarized macrophages promote hepatocellular carcinoma progression via the plasminogen activator inhibitor-1 pathway. *Int. J. Oncol.* 59, 59. doi:10.3892/ijo.2021.5239

Chen, S., Song, X., Chen, Z., Li, X., Li, M., Liu, H., et al. (2013b). CD133 expression and the prognosis of colorectal cancer: a systematic review and meta-analysis. *PLoS One* 8, e56380. doi:10.1371/journal.pone.0056380

Chen, Y., Li, Y., Wang, X. Q., Meng, Y., Zhang, Q., Zhu, J. Y., et al. (2018). Phenethyl isothiocyanate inhibits colorectal cancer stem cells by suppressing Wnt/ β -catenin pathway. *Phytother. Res.* 32, 2447–2455. doi:10.1002/ptr.6183

Chen, Y., Wang, M. H., Zhu, J. Y., Xie, C. F., Li, X. T., Wu, J. S., et al. (2020). TAp63 α targeting of Lgr5 mediates colorectal cancer stem cell properties and sulforaphane inhibition. *Oncogenesis* 9, 89. doi:10.1038/s41389-020-00273-z

Chu, C. Y., Wang, R., and Liu, X. L. (2022). Roles of Wnt/β-catenin signaling pathway related microRNAs in esophageal cancer. *World J. Clin. Cases* 10, 2678–2686. doi:10. 12998/wjcc.v10.i9.2678

Chung, W. C., and Xu, K. (2023). Notch signaling pathway in pancreatic tumorigenesis. Adv. Cancer Res. 159, 1-36. doi:10.1016/bs.acr.2023.02.001

Chu, P., Clanton, D. J., Snipas, T. S., Lee, J., Mitchell, E., Nguyen, M. L., et al. (2009). Characterization of a subpopulation of colon cancer cells with stem cell-like properties. *Int. J. Cancer* 124, 1312–1321. doi:10.1002/ijc.24061

Chu, Y., Fan, W., Guo, W., Zhang, Y., Wang, L., Guo, L., et al. (2017). miR-1247-5p functions as a tumor suppressor in human hepatocellular carcinoma by targeting Wnt3. *Oncol. Rep.* 38, 343–351. doi:10.3892/or.2017.5702

Cioffi, M., Dorado, J., Baeuerle, P. A., and Heeschen, C. (2012). EpCAM/CD3-Bispecific T-cell engaging antibody MT110 eliminates primary human pancreatic cancer stem cells. *Clin. Cancer Res.* 18, 465–474. doi:10.1158/1078-0432.CCR-11-1270

Cioffi, M., Trabulo, S., Hidalgo, M., Costello, E., Greenhalf, W., Erkan, M., et al. (2015). Inhibition of CD47 effectively targets pancreatic cancer stem cells via dual mechanisms. *Clin. Cancer Res.* 21, 2325–2337. doi:10.1158/1078-0432.CCR-14-1399

Clara, J. A., Monge, C., Yang, Y., and Takebe, N. (2020). Targeting signalling pathways and the immune microenvironment of cancer stem cells - a clinical update. *Nat. Rev. Clin. Oncol.* 17, 204–232. doi:10.1038/s41571-019-0293-2

Comito, G., Giannoni, E., Segura, C. P., Barcellos-De-Souza, P., Raspollini, M. R., Baroni, G., et al. (2014). Cancer-associated fibroblasts and M2-polarized macrophages synergize during prostate carcinoma progression. *Oncogene* 33, 2423–2431. doi:10. 1038/onc.2013.191

Dagogo-Jack, I., and Shaw, A. T. (2018). Tumour heterogeneity and resistance to cancer therapies. *Nat. Rev. Clin. Oncol.* 15, 81–94. doi:10.1038/nrclinonc.2017.166

Dai, H., Tong, C., Shi, D., Chen, M., Guo, Y., Chen, D., et al. (2020). Efficacy and biomarker analysis of CD133-directed CAR T cells in advanced hepatocellular carcinoma: a single-arm, open-label, phase II trial. *Oncoimmunology* 9, 1846926. doi:10.1080/2162402X.2020.1846926

Dai, M., Yuan, F., Fu, C., Shen, G., Hu, S., and Shen, G. (2017). Relationship between epithelial cell adhesion molecule (EpCAM) overexpression and gastric cancer patients: a systematic review and meta-analysis. *PLoS One* 12, e0175357. doi:10.1371/journal.pone. 0175357

Dalerba, P., Dylla, S. J., Park, I. K., Liu, R., Wang, X., Cho, R. W., et al. (2007). Phenotypic characterization of human colorectal cancer stem cells. *Proc. Natl. Acad. Sci.* U. S. A. 104, 10158–10163. doi:10.1073/pnas.0703478104

Dean, M., Fojo, T., and Bates, S. (2005). Tumour stem cells and drug resistance. Nat. Rev. Cancer 5, 275-284. doi:10.1038/nrc1590

De Craene, B., and Berx, G. (2013). Regulatory networks defining EMT during cancer initiation and progression. *Nat. Rev. Cancer* 13, 97–110. doi:10.1038/nrc3447

Doi, T., Piha-Paul, S. A., Jalal, S. I., Saraf, S., Lunceford, J., Koshiji, M., et al. (2018). Safety and antitumor activity of the anti-programmed death-1 antibody pembrolizumab in patients with advanced esophageal carcinoma. *J. Clin. Oncol.* 36, 61–67. doi:10.1200/ JCO.2017.74.9846

Dong, T. T., Zhou, H. M., Wang, L. L., Feng, B., Lv, B., and Zheng, M. H. (2011). Salinomycin selectively targets 'CD133+' cell subpopulations and decreases malignant traits in colorectal cancer lines. *Ann. Surg. Oncol.* 18, 1797–1804. doi:10.1245/s10434-011-1561-2

Du, L., Cheng, Q., Zheng, H., Liu, J., Liu, L., and Chen, Q. (2022). Targeting stemness of cancer stem cells to fight colorectal cancers. *Semin. Cancer Biol.* 82, 150–161. doi:10. 1016/j.semcancer.2021.02.012

Duluc, C., Moatassim-Billah, S., Chalabi-Dchar, M., Perraud, A., Samain, R., Breibach, F., et al. (2015). Pharmacological targeting of the protein synthesis mTOR/4E-BP1 pathway in cancer-associated fibroblasts abrogates pancreatic tumour chemoresistance. *EMBO Mol. Med.* 7, 735–753. doi:10.15252/emmm. 201404346

Elkashty, O. A., and Tran, S. D. (2021). Sulforaphane as a promising natural molecule for cancer prevention and treatment. *Curr. Med. Sci.* 41, 250–269. doi:10.1007/s11596-021-2341-2

Esebanmen, G. E., and Langridge, W. H. R. (2017). The role of TGF-beta signaling in dendritic cell tolerance. *Immunol. Res.* 65, 987–994. doi:10.1007/s12026-017-8944-9

Fan, Q. M., Jing, Y. Y., Yu, G. F., Kou, X. R., Ye, F., Gao, L., et al. (2014). Tumorassociated macrophages promote cancer stem cell-like properties via transforming growth factor-beta1-induced epithelial-mesenchymal transition in hepatocellular carcinoma. *Cancer Lett.* 352, 160–168. doi:10.1016/j.canlet.2014.05.008

Farhood, B., Najafi, M., and Mortezaee, K. (2019). CD8(+) cytotoxic T lymphocytes in cancer immunotherapy: a review. J. Cell Physiol. 234, 8509–8521. doi:10.1002/jcp.27782

Felices, M., Ankarlo, D. E., Lenvik, T. R., Nelson, H. H., Blazar, B. R., Verneris, M. R., et al. (2014). Notch signaling at later stages of NK cell development enhances KIR expression and functional maturation. *J. Immunol.* 193, 3344–3354. doi:10.4049/jimmunol.1400534

Flebbe, H., Spitzner, M., Marquet, P. E., Gaedcke, J., Ghadimi, B. M., Rieken, S., et al. (2022). Targeting STAT3 signaling facilitates responsiveness of pancreatic cancer cells to chemoradiotherapy. *Cancers (Basel)* 14, 1301. doi:10.3390/cancers14051301

Fridlender, Z. G., Sun, J., Kim, S., Kapoor, V., Cheng, G., Ling, L., et al. (2009). Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell* 16, 183–194. doi:10.1016/j.ccr.2009.06.017

Fuyuhiro, Y., Yashiro, M., Noda, S., Kashiwagi, S., Matsuoka, J., Doi, Y., et al. (2011). Upregulation of cancer-associated myofibroblasts by TGF- β from scirrhous gastric carcinoma cells. *Br. J. Cancer* 105, 996–1001. doi:10.1038/bjc.2011.330

Garza Trevino, E. N., Quiroz Reyes, A. G., Rojas Murillo, J. A., De La Garza Kalife, D. A., Delgado Gonzalez, P., Islas, J. F., et al. (2023). Cell therapy as target therapy against colon cancer stem cells. *Int. J. Mol. Sci.* 24, 8163. doi:10.3390/ijms24098163

Gattinoni, L., Ji, Y., and Restifo, N. P. (2010). Wnt/beta-catenin signaling in T-cell immunity and cancer immunotherapy. *Clin. Cancer Res.* 16, 4695–4701. doi:10.1158/1078-0432.CCR-10-0356

Goel, H. L., and Mercurio, A. M. (2013). VEGF targets the tumour cell. Nat. Rev. Cancer 13, 871-882. doi:10.1038/nrc3627

Grange, C., Tapparo, M., Tritta, S., Deregibus, M. C., Battaglia, A., Gontero, P., et al. (2015). Role of HLA-G and extracellular vesicles in renal cancer stem cell-induced inhibition of dendritic cell differentiation. *BMC Cancer* 15, 1009. doi:10.1186/s12885-015-2025-z

Grothey, A., Tebbutt, N., Van Cutsem, E., Taieb, J., Falcone, A., Yoshino, T., et al. (2016). CanStem303C trial: a phase III study of BBI-608 (napabucasin) in combination with 5-fluorouracil (5-FU), leucovorin, irinotecan (FOLFIRI) in adult patients with previously treated metastatic colorectal cancer (mCRC). *Ann. Oncol.* 27, vi206. doi:10. 1093/annonc/mdw370.156

Guo, M., Luo, B., Pan, M., Li, M., Xu, H., Zhao, F., et al. (2020). Colorectal cancer stem cell vaccine with high expression of MUC1 serves as a novel prophylactic vaccine for colorectal cancer. *Int. Immunopharmacol.* 88, 106850. doi:10.1016/j.intimp.2020. 106850

Guo, X., Zhao, Y., Yan, H., Yang, Y., Shen, S., Dai, X., et al. (2017). Single tumorinitiating cells evade immune clearance by recruiting type II macrophages. *Genes Dev.* 31, 247–259. doi:10.1101/gad.294348.116

Han, S., Shi, Y., Sun, L., Liu, Z., Song, T., and Liu, Q. (2019). MiR-4319 induced an inhibition of epithelial-mesenchymal transition and prevented cancer stemness of HCC through targeting FOXQ1. *Int. J. Biol. Sci.* 15, 2936–2947. doi:10.7150/ijbs.38000

Han, Y., Sun, B., Cai, H., and Xuan, Y. (2021). Simultaneously target of normal and stem cells-like gastric cancer cells via cisplatin and anti-CD133 CAR-T combination therapy. *Cancer Immunol. Immunother.* 70, 2795–2803. doi:10.1007/s00262-021-02891-x

Hashemi, F., Razmi, M., Tajik, F., Zoller, M., Dehghan Manshadi, M., Mahdavinezhad, F., et al. (2023). Efficacy of whole cancer stem cell-based vaccines: a systematic review of preclinical and clinical studies. *Stem Cells* 41, 207–232. doi:10. 1093/stmcls/sxac089

Hassn Mesrati, M., Syafruddin, S. E., Mohtar, M. A., and Syahir, A. (2021). CD44: a multifunctional mediator of cancer progression. *Biomolecules* 11, 1850. doi:10.3390/biom11121850

Hayashi, H., Inoue, Y., Tsutsui, H., Okamura, H., Nakanishi, K., and Onozaki, K. (2003). TGFbeta down-regulates IFN-gamma production in IL-18 treated NK cell line LNK5E6. *Biochem. Biophys. Res. Commun.* 300, 980–985. doi:10.1016/s0006-291x(02) 02939-x

Hibdon, E. S., Razumilava, N., Keeley, T. M., Wong, G., Solanki, S., Shah, Y. M., et al. (2019). Notch and mTOR signaling pathways promote human gastric cancer cell proliferation. *Neoplasia* 21, 702–712. doi:10.1016/j.neo.2019.05.002

Higashino, N., Koma, Y. I., Hosono, M., Takase, N., Okamoto, M., Kodaira, H., et al. (2019). Fibroblast activation protein-positive fibroblasts promote tumor progression through secretion of CCL2 and interleukin-6 in esophageal squamous cell carcinoma. *Lab. Invest.* 99, 777–792. doi:10.1038/s41374-018-0185-6

Hjazi, A., Nasir, F., Noor, R., Alsalamy, A., Zabibah, R. S., Romero-Parra, R. M., et al. (2023). The pathological role of C-X-C chemokine receptor type 4 (CXCR4) in colorectal cancer (CRC) progression; special focus on molecular mechanisms and possible therapeutics. *Pathol. Res. Pract.* 248, 154616. doi:10.1016/j.prp.2023.154616

Hong, D. S., Kang, Y. K., Borad, M., Sachdev, J., Ejadi, S., Lim, H. Y., et al. (2020). Phase 1 study of MRX34, a liposomal miR-34a mimic, in patients with advanced solid tumours. *Br. J. Cancer* 122, 1630–1637. doi:10.1038/s41416-020-0802-1

Hong, Y., Qin, H., Li, Y., Zhang, Y., Zhuang, X., Liu, L., et al. (2019). FNDC3B circular RNA promotes the migration and invasion of gastric cancer cells via the regulation of E-cadherin and CD44 expression. *J. Cell. Physiology* 234, 19895–19910. doi:10.1002/jcp. 28588

Hsu, Y. L., Chen, Y. J., Chang, W. A., Jian, S. F., Fan, H. L., Wang, J. Y., et al. (2018). Interaction between tumor-associated dendritic cells and colon cancer cells contributes to tumor progression via CXCL1. *Int. J. Mol. Sci.* 19, 2427. doi:10.3390/ijms19082427

Hu, Z. Y., Wang, X. Y., Guo, W. B., Xie, L. Y., Huang, Y. Q., Liu, Y. P., et al. (2016). Long non-coding RNA MALAT1 increases AKAP-9 expression by promoting SRPK1catalyzed SRSF1 phosphorylation in colorectal cancer cells. *Oncotarget* 7, 11733–11743. doi:10.18632/oncotarget.7367

Huang, S. D., Yuan, Y., Tang, H., Liu, X. H., Fu, C. G., Cheng, H. Z., et al. (2013). Tumor cells positive and negative for the common cancer stem cell markers are capable of initiating tumor growth and generating both progenies. *PLoS One* 8, e54579. doi:10. 1371/journal.pone.0054579

Huang, Y. K., Wang, M., Sun, Y., Di Costanzo, N., Mitchell, C., Achuthan, A., et al. (2019b). Macrophage spatial heterogeneity in gastric cancer defined by multiplex immunohistochemistry. *Nat. Commun.* 10, 3928. doi:10.1038/s41467-019-11788-4

Huang, C., Ou, R., Chen, X., Zhang, Y., Li, J., Liang, Y., et al. (2021). Tumor cellderived SPON2 promotes M2-polarized tumor-associated macrophage infiltration and cancer progression by activating PYK2 in CRC. J. Exp. Clin. Cancer Res. 40, 304. doi:10. 1186/s13046-021-02108-0 Huang, L., Zhang, Y., Li, Z., Zhao, X., XI, Z., Chen, H., et al. (2019a). MiR-4319 suppresses colorectal cancer progression by targeting ABTB1. United Eur. Gastroenterol. J. 7, 517–528. doi:10.1177/2050640619837440

Huang, T., Song, X., Xu, D., Tiek, D., Goenka, A., Wu, B., et al. (2020). Stem cell programs in cancer initiation, progression, and therapy resistance. *Theranostics* 10, 8721–8743. doi:10.7150/thno.41648

Hu, X., Wang, M., Cao, L., Cong, L., Gao, Y., Lu, J., et al. (2020). miR-4319 suppresses the growth of esophageal squamous cell carcinoma via targeting NLRC5. *Curr. Mol. Pharmacol.* 13, 144–149. doi:10.2174/1874467212666191119094636

Hu, Y., Yan, C., Mu, L., Huang, K., Li, X., Tao, D., et al. (2015). Fibroblast-derived exosomes contribute to chemoresistance through priming cancer stem cells in colorectal cancer. *PLoS One* 10, e0125625. doi:10.1371/journal.pone.0125625

Hwang, G. R., Yuen, J. G., and Ju, J. (2021). Roles of microRNAs in gastrointestinal cancer stem cell resistance and therapeutic development. *Int. J. Mol. Sci.* 22, 1624. doi:10.3390/ijms22041624

Ishimoto, T., Oshima, H., Oshima, M., Kai, K., Torii, R., Masuko, T., et al. (2010). CD44+ slow-cycling tumor cell expansion is triggered by cooperative actions of Wnt and prostaglandin E2 in gastric tumorigenesis. *Cancer Sci.* 101, 673–678. doi:10.1111/j. 1349-7006.2009.01430.x

Ishimoto, T., Sugihara, H., Watanabe, M., Sawayama, H., Iwatsuki, M., Baba, Y., et al. (2014). Macrophage-derived reactive oxygen species suppress miR-328 targeting CD44 in cancer cells and promote redox adaptation. *Carcinogenesis* 35, 1003–1011. doi:10.1093/carcin/bgt402

Ishiwata, T., Matsuda, Y., Yoshimura, H., Sasaki, N., Ishiwata, S., Ishikawa, N., et al. (2018). Pancreatic cancer stem cells: features and detection methods. *Pathol. Oncol. Res.* 24, 797–805. doi:10.1007/s12253-018-0420-x

Islam, F., Gopalan, V., Wahab, R., Smith, R. A., and Lam, A. K. (2015). Cancer stem cells in oesophageal squamous cell carcinoma: identification, prognostic and treatment perspectives. *Crit. Rev. Oncol. Hematol.* 96, 9–19. doi:10.1016/j.critrevonc.2015.04.007

Izadpanah, A., Mohammadkhani, N., Masoudnia, M., Ghasemzad, M., Saeedian, A., Mehdizadeh, H., et al. (2023). Update on immune-based therapy strategies targeting cancer stem cells. *Cancer Med.* 12, 18960–18980. doi:10.1002/cam4.6520

Janjigian, Y. Y., Shitara, K., Moehler, M., Garrido, M., Salman, P., Shen, L., et al. (2021). First-line nivolumab plus chemotherapy versus chemotherapy alone for advanced gastric, gastro-oesophageal junction, and oesophageal adenocarcinoma (CheckMate 649): a randomised, open-label, phase 3 trial. *Lancet* 398, 27–40. doi:10.1016/S0140-6736(21)00797-2

Jeng, K. S., Chang, C. F., and Lin, S. S. (2020). Sonic hedgehog signaling in organogenesis, tumors, and tumor microenvironments. *Int. J. Mol. Sci.* 21, 758. doi:10.3390/ijms21030758

Jeng, K. S., Chang, C. F., Sheen, I. S., Jeng, C. J., and Wang, C. H. (2023). Cellular and molecular biology of cancer stem cells of hepatocellular carcinoma. *Int. J. Mol. Sci.* 24, 1417. doi:10.3390/ijms24021417

Jeng, K. S., Jeng, C. J., Jeng, W. J., Sheen, I. S., Chang, C. F., Hsiau, H. I., et al. (2015). Sonic hedgehog pathway inhibitor mitigates mouse hepatocellular carcinoma. *Am. J. Surg.* 210, 554–560. doi:10.1016/j.amjsurg.2015.03.001

Jiao, F., Hu, H., Han, T., Yuan, C., Wang, L., Jin, Z., et al. (2015). Long noncoding RNA MALAT-1 enhances stem cell-like phenotypes in pancreatic cancer cells. *Int. J. Mol. Sci.* 16, 6677–6693. doi:10.3390/ijms16046677

Ji, L., Qian, W., Gui, L., Ji, Z., Yin, P., Lin, G. N., et al. (2020). Blockade of β -catenininduced CCL28 suppresses gastric cancer progression via inhibition of Treg cell infiltration. *Cancer Res.* 80, 2004–2016. doi:10.1158/0008-5472.CAN-19-3074

Jin, W. (2020). Role of JAK/STAT3 signaling in the regulation of metastasis, the transition of cancer stem cells, and chemoresistance of cancer by epithelial-mesenchymal transition. *Cells* 9, 217. doi:10.3390/cells9010217

Jones, S. A., and Jenkins, B. J. (2018). Recent insights into targeting the IL-6 cytokine family in inflammatory diseases and cancer. *Nat. Rev. Immunol.* 18, 773–789. doi:10. 1038/s41577-018-0066-7

Ju, X., Zhang, H., Zhou, Z., Chen, M., and Wang, Q. (2020). Tumor-associated macrophages induce PD-L1 expression in gastric cancer cells through IL-6 and TNF-a signaling. *Exp. Cell Res.* 396, 112315. doi:10.1016/j.yexcr.2020.112315

Kallifatidis, G., Rausch, V., Baumann, B., Apel, A., Beckermann, B. M., Groth, A., et al. (2009). Sulforaphane targets pancreatic tumour-initiating cells by NF-kappaB-induced antiapoptotic signalling. *Gut* 58, 949–963. doi:10.1136/gut.2008.149039

Kalluri, R. (2016). The biology and function of fibroblasts in cancer. *Nat. Rev. Cancer* 16, 582–598. doi:10.1038/nrc.2016.73

Kaseb, A. O., Hasanov, E., Cao, H. S. T., Xiao, L., Vauthey, J. N., Lee, S. S., et al. (2022). Perioperative nivolumab monotherapy versus nivolumab plus ipilimumab in resectable hepatocellular carcinoma: a randomised, open-label, phase 2 trial. *Lancet Gastroenterol. Hepatol.* 7, 208–218. doi:10.1016/S2468-1253(21)00427-1

Katoh, M. (2017). Canonical and non-canonical WNT signaling in cancer stem cells and their niches: cellular heterogeneity, omics reprogramming, targeted therapy and tumor plasticity (Review). *Int. J. Oncol.* 51, 1357–1369. doi:10.3892/ijo.2017.4129

Katoh, M., and Katoh, M. (2007). WNT signaling pathway and stem cell signaling network. Clin. Cancer Res. 13, 4042-4045. doi:10.1158/1078-0432.CCR-06-2316

Katoh, M., and Katoh, M. (2017). Molecular genetics and targeted therapy of WNTrelated human diseases (Review). *Int. J. Mol. Med.* 40, 587–606. doi:10.3892/ijmm.2017. 3071

Katoh, M., and Katoh, M. (2022). WNT signaling and cancer stemness. *Essays Biochem.* 66, 319–331. doi:10.1042/EBC20220016

Katsuno, Y., Ehata, S., Yashiro, M., Yanagihara, K., Hirakawa, K., and Miyazono, K. (2012). Coordinated expression of REG4 and aldehyde dehydrogenase 1 regulating tumourigenic capacity of diffuse-type gastric carcinoma-initiating cells is inhibited by TGF-β. J. Pathol. 228, 391–404. doi:10.1002/path.4020

Kebenko, M., Goebeler, M. E., Wolf, M., Hasenburg, A., Seggewiss-Bernhardt, R., Ritter, B., et al. (2018). A multicenter phase 1 study of solitomab (MT110, AMG 110), a bispecific EpCAM/CD3 T-cell engager (BiTE[®]) antibody construct, in patients with refractory solid tumors. *Oncoimmunology* 7, e1450710. doi:10.1080/2162402X.2018. 1450710

Keir, M. E., Butte, M. J., Freeman, G. J., and Sharpe, A. H. (2008). PD-1 and its ligands in tolerance and immunity. *Annu. Rev. Immunol.* 26, 677–704. doi:10.1146/annurev. immunol.26.021607.090331

Kenkel, J. A., Tseng, W. W., Davidson, M. G., Tolentino, L. L., Choi, O., Bhattacharya, N., et al. (2017). An immunosuppressive dendritic cell subset accumulates at secondary sites and promotes metastasis in pancreatic cancer. *Cancer Res.* 77, 4158–4170. doi:10. 1158/0008-5472.CAN-16-2212

Khosla, R., Rastogi, A., Ramakrishna, G., Pamecha, V., Mukhopadhyay, A., Vasudevan, M., et al. (2017). EpCAM+ liver cancer stem-like cells exhibiting autocrine Wnt signaling potentially originate in cirrhotic patients. *Stem Cells Transl. Med.* 6, 807–818. doi:10.1002/sctm.16-0248

Kieffer, Y., Hocine, H. R., Gentric, G., Pelon, F., Bernard, C., Bourachot, B., et al. (2020). Single-cell analysis reveals fibroblast clusters linked to immunotherapy resistance in cancer. *Cancer Discov.* 10, 1330–1351. doi:10.1158/2159-8290.CD-19-1384

Kim, W. T., and Ryu, C. J. (2017). Cancer stem cell surface markers on normal stem cells. *BMB Rep.* 50, 285–298. doi:10.5483/bmbrep.2017.50.6.039

Kim, H., Lin, Q., Glazer, P. M., and Yun, Z. (2018). The hypoxic tumor microenvironment *in vivo* selects the cancer stem cell fate of breast cancer cells. *Breast Cancer Res.* 20, 16. doi:10.1186/s13058-018-0944-8

Kishi, M., Asgarova, A., Desterke, C., Chaker, D., Artus, J., Turhan, A. G., et al. (2021). Evidence of antitumor and antimetastatic potential of induced pluripotent stem cellbased vaccines in cancer immunotherapy. *Front. Med. (Lausanne)* 8, 729018. doi:10. 3389/fmed.2021.729018

Kitamura, T., Qian, B. Z., and Pollard, J. W. (2015). Immune cell promotion of metastasis. *Nat. Rev. Immunol.* 15, 73-86. doi:10.1038/nri3789

Kobayashi, H., Enomoto, A., Woods, S. L., Burt, A. D., Takahashi, M., and Worthley, D. L. (2019). Cancer-associated fibroblasts in gastrointestinal cancer. *Nat. Rev. Gastroenterology Hepatology* 16, 282–295. doi:10.1038/s41575-019-0115-0

Kono, K., Kawaida, H., Takahashi, A., Sugai, H., Mimura, K., Miyagawa, N., et al. (2006). CD4(+)CD25high regulatory T cells increase with tumor stage in patients with gastric and esophageal cancers. *Cancer Immunol. Immunother*. 55, 1064–1071. doi:10. 1007/s00262-005-0092-8

Krzysiek-Maczka, G., Wrobel, T., Targosz, A., Szczyrk, U., Strzalka, M., Ptak-Belowska, A., et al. (2019). Helicobacter pylori-activated gastric fibroblasts induce epithelial-mesenchymal transition of gastric epithelial cells *in vitro* in a TGF- β -dependent manner. *Helicobacter* 24, e12653. doi:10.1111/hel.12653

Kubiczkova, L., Sedlarikova, L., Hajek, R., and Sevcikova, S. (2012). TGF- β - an excellent servant but a bad master. *J. Transl. Med.* 10, 183. doi:10.1186/1479-5876-10-183

Lapidot, T., Sirard, C., Vormoor, J., Murdoch, B., Hoang, T., Caceres-Cortes, J., et al. (1994). A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* 367, 645–648. doi:10.1038/367645a0

Lau, E. Y., Lo, J., Cheng, B. Y., Ma, M. K., Lee, J. M., Ng, J. K., et al. (2016). Cancerassociated fibroblasts regulate tumor-initiating cell plasticity in hepatocellular carcinoma through c-met/FRA1/HEY1 signaling. *Cell Rep.* 15, 1175–1189. doi:10. 1016/j.celrep.2016.04.019

Lau, S. P., Klaase, L., Vink, M., Dumas, J., Bezemer, K., Van Krimpen, A., et al. (2022). Autologous dendritic cells pulsed with allogeneic tumour cell lysate induce tumourreactive T-cell responses in patients with pancreatic cancer: A phase I study. *Eur. J. Cancer* 169, 20–31. doi:10.1016/j.ejca.2022.03.015

Lee, J. H., and Massague, J. (2022). TGF- β in developmental and fibrogenic EMTs. Semin. Cancer Biol. 86, 136–145. doi:10.1016/j.semcancer.2022.09.004

Lee, J. L., Wang, M. J., Sudhir, P. R., and Chen, J. Y. (2008). CD44 engagement promotes matrix-derived survival through the CD44-SRC-integrin axis in lipid rafts. *Mol. Cell Biol.* 28, 5710–5723. doi:10.1128/MCB.00186-08

Lee, T. K., Cheung, V. C., Lu, P., Lau, E. Y., Ma, S., Tang, K. H., et al. (2014). Blockade of CD47-mediated cathepsin S/protease-activated receptor 2 signaling provides a therapeutic target for hepatocellular carcinoma. *Hepatology* 60, 179–191. doi:10. 1002/hep.27070

Lenz, H. J., Van Cutsem, E., Luisa Limon, M., Wong, K. Y. M., Hendlisz, A., Aglietta, M., et al. (2022). First-line nivolumab plus low-dose ipilimumab for microsatellite

instability-high/mismatch repair-deficient metastatic colorectal cancer: the phase II CheckMate 142 study. J. Clin. Oncol. 40, 161–170. doi:10.1200/JCO.21.01015

Leung, D. W., Cachianes, G., Kuang, W. J., Goeddel, D. V., and Ferrara, N. (1989). Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science* 246, 1306–1309. doi:10.1126/science.2479986

Li, S. H., Fu, J., Watkins, D. N., Srivastava, R. K., and Shankar, S. (2013). Sulforaphane regulates self-renewal of pancreatic cancer stem cells through the modulation of Sonic hedgehog-GLI pathway. *Mol. Cell Biochem.* 373, 217–227. doi:10.1007/s11010-012-1493-6

Li, D., Xia, L., Huang, P., Wang, Z., Guo, Q., Huang, C., et al. (2023a). Cancerassociated fibroblast-secreted IGFBP7 promotes gastric cancer by enhancing tumor associated macrophage infiltration via FGF2/FGFR1/PI3K/AKT axis. *Cell Death Discov.* 9, 17. doi:10.1038/s41420-023-01336-x

Lin, C. N., Wang, C. J., Chao, Y. J., Lai, M. D., and Shan, Y. S. (2015a). The significance of the co-existence of osteopontin and tumor-associated macrophages in gastric cancer progression. *BMC Cancer* 15, 128. doi:10.1186/s12885-015-1114-3

Lin, M., Yuan, Y. Y., Liu, S. P., Shi, J. J., Long, X. A., Niu, L. Z., et al. (2015b). Prospective study of the safety and efficacy of a pancreatic cancer stem cell vaccine. *J. Cancer Res. Clin. Oncol.* 141, 1827–1833. doi:10.1007/s00432-015-1968-4

Lin, X., Zuo, S., Luo, R., Li, Y., Yu, G., Zou, Y., et al. (2019). HBX-induced miR-5188 impairs FOXO1 to stimulate β -catenin nuclear translocation and promotes tumor stemness in hepatocellular carcinoma. *Theranostics* 9, 7583–7598. doi:10.7150/thno. 37717

Li, S., Yu, W., Xie, F., Luo, H., Liu, Z., Lv, W., et al. (2023b). Neoadjuvant therapy with immune checkpoint blockade, antiangiogenesis, and chemotherapy for locally advanced gastric cancer. *Nat. Commun.* 14, 8. doi:10.1038/s41467-022-35431-x

Liu, C. X., and Chen, L. L. (2022). Circular RNAs: characterization, cellular roles, and applications. *Cell* 185, 2390–2034. doi:10.1016/j.cell.2022.06.001

Liu, L., Wise, D. R., Diehl, J. A., and Simon, M. C. (2008). Hypoxic reactive oxygen species regulate the integrated stress response and cell survival. *J. Biol. Chem.* 283, 31153–31162. doi:10.1074/jbc.M805056200

Liu, P., Atkinson, S. J., Akbareian, S. E., Zhou, Z., Munsterberg, A., Robinson, S. D., et al. (2017). Sulforaphane exerts anti-angiogenesis effects against hepatocellular carcinoma through inhibition of STAT3/HIF-1a/VEGF signalling. *Sci. Rep.* 7, 12651. doi:10.1038/s41598-017-12855-w

Liu, X., Zhang, Z., and Zhao, G. (2019). Recent advances in the study of regulatory T cells in gastric cancer. *Int. Immunopharmacol.* 73, 560–567. doi:10.1016/j.intimp. 2019.05.009

Li, X., Yan, X., Wang, Y., Kaur, B., Han, H., and Yu, J. (2023c). The Notch signaling pathway: a potential target for cancer immunotherapy. *J. Hematol. Oncol.* 16, 45. doi:10. 1186/s13045-023-01439-z

Li, X., Zhang, Z., Lin, G., Gao, Y., Yan, Z., Yin, H., et al. (2016). Antigen-specific T cell response from dendritic cell vaccination using side population cell-associated antigens targets hepatocellular carcinoma. *Tumour Biol.* 37, 11267–11278. doi:10.1007/s13277-016-4935-z

Li, Y., Lv, Z., He, G., Wang, J., Zhang, X., Lu, G., et al. (2015a). The SOX17/miR-371-5p/SOX2 axis inhibits EMT, stem cell properties and metastasis in colorectal cancer. *Oncotarget* 6, 9099–9112. doi:10.18632/oncotarget.3603

Li, Y., Rogoff, H. A., Keates, S., Gao, Y., Murikipudi, S., Mikule, K., et al. (2015b). Suppression of cancer relapse and metastasis by inhibiting cancer stemness. *Proc. Natl. Acad. Sci. U. S. A.* 112, 1839–1844. doi:10.1073/pnas.1424171112

Li, Z., Chen, K., Jiang, P., Zhang, X., Li, X., and Li, Z. (2014). CD44v/CD44s expression patterns are associated with the survival of pancreatic carcinoma patients. *Diagn Pathol.* 9, 79. doi:10.1186/1746-1596-9-79

Lotti, F., Jarrar, A. M., Pai, R. K., Hitomi, M., Lathia, J., Mace, A., et al. (2013). Chemotherapy activates cancer-associated fibroblasts to maintain colorectal cancerinitiating cells by IL-17A. *J. Exp. Med.* 210, 2851–2872. doi:10.1084/jem.20131195

Lozanovski, V. J., Polychronidis, G., Gross, W., Gharabaghi, N., Mehrabi, A., Hackert, T., et al. (2020). Broccoli sprout supplementation in patients with advanced pancreatic cancer is difficult despite positive effects-results from the POUDER pilot study. *Invest. New Drugs* 38, 776–784. doi:10.1007/s10637-019-00826-z

Lugli, A., Iezzi, G., Hostettler, I., Muraro, M. G., Mele, V., Tornillo, L., et al. (2010). Prognostic impact of the expression of putative cancer stem cell markers CD133, CD166, CD44s, EpCAM, and ALDH1 in colorectal cancer. *Br. J. Cancer* 103, 382–390. doi:10.1038/sj.bjc.6605762

Lu, J., Ye, X., Fan, F., Xia, L., Bhattacharya, R., Bellister, S., et al. (2013). Endothelial cells promote the colorectal cancer stem cell phenotype through a soluble form of Jagged-1. *Cancer Cell* 23, 171–185. doi:10.1016/j.ccr.2012.12.021

Luo, Y., Wu, J., Wu, Q., Li, X., Wu, J., Zhang, J., et al. (2019). miR-577 regulates TGF- β induced cancer progression through a SDPR-modulated positive-feedback loop with ERK-NF- κ b in gastric cancer. *Mol. Ther.* 27, 1166–1182. doi:10.1016/j.ymthe.2019.02.002

Luraghi, P., Reato, G., Cipriano, E., Sassi, F., Orzan, F., Bigatto, V., et al. (2014). MET signaling in colon cancer stem-like cells blunts the therapeutic response to EGFR inhibitors. *Cancer Res.* 74, 1857–1869. doi:10.1158/0008-5472.CAN-13-2340-T

Ma, J., Song, X., Xu, X., and Mou, Y. (2019a). Cancer-associated fibroblasts promote the chemo-resistance in gastric cancer through secreting IL-11 targeting JAK/STAT3/ Bcl2 pathway. *Cancer Res. Treat.* 51, 194–210. doi:10.4143/crt.2018.031

Ma, L., Hernandez, M. O., Zhao, Y., Mehta, M., Tran, B., Kelly, M., et al. (2019b). Tumor cell biodiversity drives microenvironmental reprogramming in liver cancer. *Cancer Cell* 36, 418–430. doi:10.1016/j.ccell.2019.08.007

Malfettone, A., Soukupova, J., Bertran, E., Crosas-Molist, E., Lastra, R., Fernando, J., et al. (2017). Transforming growth factor-β-induced plasticity causes a migratory stemness phenotype in hepatocellular carcinoma. *Cancer Lett.* 392, 39–50. doi:10. 1016/j.canlet.2017.01.037

Mani, S. A., Guo, W., Liao, M. J., Eaton, E. N., Ayyanan, A., Zhou, A. Y., et al. (2008). The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell* 133, 704–715. doi:10.1016/j.cell.2008.03.027

Mariathasan, S., Turley, S. J., Nickles, D., Castiglioni, A., Yuen, K., Wang, Y., et al. (2018). TGF β attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. *Nature* 554, 544–548. doi:10.1038/nature25501

Mathew, E., Brannon, A. L., Del Vecchio, A., Garcia, P. E., Penny, M. K., Kane, K. T., et al. (2016). Mesenchymal stem cells promote pancreatic tumor growth by inducing alternative polarization of macrophages. *Neoplasia* 18, 142–151. doi:10.1016/j.neo.2016. 01.005

Mccabe, E. M., and Rasmussen, T. P. (2021). lncRNA involvement in cancer stem cell function and epithelial-mesenchymal transitions. *Semin. Cancer Biol.* 75, 38–48. doi:10. 1016/j.semcancer.2020.12.012

Mele, L., Liccardo, D., and Tirino, V. (2018). Evaluation and isolation of cancer stem cells using ALDH activity assay. *Methods Mol. Biol.* 1692, 43–48. doi:10.1007/978-1-4939-7401-6_4

MendonçA, L., Trindade, A., Carvalho, C., Correia, J., Badenes, M., Gigante, J., et al. (2019). Metastasis is impaired by endothelial-specific Dll4 loss-of-function through inhibition of epithelial-to-mesenchymal transition and reduction of cancer stem cells and circulating tumor cells. *Clin. Exp. Metastasis* 36, 365–380. doi:10.1007/s10585-019-09973-2

Meng, J., Jiang, Y. Z., Zhao, S., Tao, Y., Zhang, T., Wang, X., et al. (2022). Tumorderived Jagged1 promotes cancer progression through immune evasion. *Cell Rep.* 38, 110492. doi:10.1016/j.celrep.2022.110492

Meng, L., Bai, Z., He, S., Mochizuki, K., Liu, Y., Purushe, J., et al. (2016). The Notch ligand DLL4 defines a capability of human dendritic cells in regulating Th1 and Th17 differentiation. *J. Immunol.* 196, 1070–1080. doi:10.4049/jimmunol.1501310

Merchant, J. L. (2012). Hedgehog signalling in gut development, physiology and cancer. J. Physiol. 590, 421-432. doi:10.1113/jphysiol.2011.220681

Mercurio, A. M. (2019). VEGF/Neuropilin signaling in cancer stem cells. Int. J. Mol. Sci. 20, 490. doi:10.3390/ijms20030490

Miao, Z. F., Xu, H., Xu, H. M., Wang, Z. N., Zhao, T. T., Song, Y. X., et al. (2017). DLL4 overexpression increases gastric cancer stem/progenitor cell self-renewal ability and correlates with poor clinical outcome via Notch-1 signaling pathway activation. *Cancer Med.* 6, 245–257. doi:10.1002/cam4.962

Miao, Y., Yang, H., Levorse, J., Yuan, S., Polak, L., Sribour, M., et al. (2019). Adaptive immune resistance emerges from tumor-initiating stem cells. *Cell* 177, 1172–1186. doi:10.1016/j.cell.2019.03.025

Mima, K., Okabe, H., Ishimoto, T., Hayashi, H., Nakagawa, S., Kuroki, H., et al. (2012). CD44s regulates the TGF-β-mediated mesenchymal phenotype and is associated with poor prognosis in patients with hepatocellular carcinoma. *Cancer Res.* 72, 3414–3423. doi:10.1158/0008-5472.CAN-12-0299

Miranda, A., Hamilton, P. T., Zhang, A. W., Pattnaik, S., Becht, E., Mezheyeuski, A., et al. (2019). Cancer stemness, intratumoral heterogeneity, and immune response across cancers. *Proc. Natl. Acad. Sci. U. S. A.* 116, 9020–9029. doi:10.1073/pnas.1818210116

Miwa, T., Nagata, T., Kojima, H., Sekine, S., and Okumura, T. (2017). Isoform switch of CD44 induces different chemotactic and tumorigenic ability in gallbladder cancer. *Int. J. Oncol.* 51, 771–780. doi:10.3892/ijo.2017.4063

Mohammadi, H., and Sahai, E. (2018). Mechanisms and impact of altered tumour mechanics. Nat. Cell Biol. 20, 766-774. doi:10.1038/s41556-018-0131-2

Morrison, B. J., Steel, J. C., and Morris, J. C. (2018). Reduction of MHC-I expression limits T-lymphocyte-mediated killing of Cancer-initiating cells. *BMC Cancer* 18, 469. doi:10.1186/s12885-018-4389-3

Mortezaee, K. (2021). Enriched cancer stem cells, dense stroma, and cold immunity: interrelated events in pancreatic cancer. J. Biochem. Mol. Toxicol. 35, e22708. doi:10. 1002/jbt.22708

Mozammel, N., Amini, M., Baradaran, B., Mahdavi, S. Z. B., Hosseini, S. S., and Mokhtarzadeh, A. (2023). The function of miR-145 in colorectal cancer progression; an updated review on related signaling pathways. *Pathol. Res. Pract.* 242, 154290. doi:10. 1016/j.prp.2022.154290

Muro, K., Chung, H. C., Shankaran, V., Geva, R., Catenacci, D., Gupta, S., et al. (2016). Pembrolizumab for patients with PD-L1-positive advanced gastric cancer (KEYNOTE-012): a multicentre, open-label, phase 1b trial. *Lancet Oncol.* 17, 717–726. doi:10.1016/ S1470-2045(16)00175-3 Najafi, M., Hashemi Goradel, N., Farhood, B., Salehi, E., Nashtaei, M. S., Khanlarkhani, N., et al. (2019). Macrophage polarity in cancer: a review. J. Cell Biochem. 120, 2756–2765. doi:10.1002/jcb.27646

Naujokat, C., and Mckee, D. L. (2021). The "big five" phytochemicals targeting cancer stem cells: curcumin, EGCG, sulforaphane, resveratrol and genistein. *Curr. Med. Chem.* 28, 4321–4342. doi:10.2174/0929867327666200228110738

Naujokat, C., and Steinhart, R. (2012). Salinomycin as a drug for targeting human cancer stem cells. J. Biomed. Biotechnol. 2012, 950658. doi:10.1155/2012/950658

Ng, C. P., Hinz, B., and Swartz, M. A. (2005). Interstitial fluid flow induces myofibroblast differentiation and collagen alignment *in vitro*. J. Cell Sci. 118, 4731–4739. doi:10.1242/jcs.02605

Nishikawa, S., Konno, M., Hamabe, A., Hasegawa, S., Kano, Y., Ohta, K., et al. (2013). Aldehyde dehydrogenase high gastric cancer stem cells are resistant to chemotherapy. *Int. J. Oncol.* 42, 1437–1442. doi:10.3892/ijo.2013.1837

Nurmik, M., Ullmann, P., Rodriguez, F., Haan, S., and Letellier, E. (2020). In search of definitions: cancer-associated fibroblasts and their markers. *Int. J. Cancer* 146, 895–905. doi:10.1002/ijc.32193

Ohlund, D., Elyada, E., and Tuveson, D. (2014). Fibroblast heterogeneity in the cancer wound. J. Exp. Med. 211, 1503–1523. doi:10.1084/jem.20140692

Olver, S., Groves, P., Buttigieg, K., Morris, E. S., Janas, M. L., Kelso, A., et al. (2006). Tumor-derived interleukin-4 reduces tumor clearance and deviates the cytokine and granzyme profile of tumor-induced CD8+ T cells. *Cancer Res.* 66, 571–580. doi:10.1158/ 0008-5472.CAN-05-1362

Orimo, A., and Weinberg, R. A. (2006). Stromal fibroblasts in cancer: a novel tumorpromoting cell type. *Cell Cycle* 5, 1597–1601. doi:10.4161/cc.5.15.3112

Osipov, A., Saung, M. T., Zheng, L., and Murphy, A. G. (2019). Small molecule immunomodulation: the tumor microenvironment and overcoming immune escape. *J. Immunother. Cancer* 7, 224. doi:10.1186/s40425-019-0667-0

PáDUA Alves, C., Fonseca, A. S., Muys, B. R., De Barros, E. L. B. R., BüRGER, M. C., De Souza, J. E., et al. (2013). Brief report: the lincRNA Hotair is required for epithelial-to-mesenchymal transition and stemness maintenance of cancer cell lines. *Stem Cells* 31, 2827–2832. doi:10.1002/stem.1547

Palaga, T., Wongchana, W., and Kueanjinda, P. (2018). Notch signaling in macrophages in the context of cancer immunity. *Front. Immunol.* 9, 652. doi:10. 3389/fmmu.2018.00652

Pamudurti, N. R., Bartok, O., Jens, M., Ashwal-Fluss, R., Stottmeister, C., Ruhe, L., et al. (2017). Translation of CircRNAs. *Mol. Cell* 66, 9–21. doi:10.1016/j.molcel.2017. 02.021

Pan, G., Liu, Y., Shang, L., Zhou, F., and Yang, S. (2021). EMT-associated microRNAs and their roles in cancer stemness and drug resistance. *Cancer Commun. (Lond)* 41, 199–217. doi:10.1002/cac2.12138

Pan, T., Xu, J., and Zhu, Y. (2017). Self-renewal molecular mechanisms of colorectal cancer stem cells. *Int. J. Mol. Med.* 39, 9–20. doi:10.3892/ijmm.2016.2815

Parikh, A. R., Szabolcs, A., Allen, J. N., Clark, J. W., Wo, J. Y., Raabe, M., et al. (2021). Radiation therapy enhances immunotherapy response in microsatellite stable colorectal and pancreatic adenocarcinoma in a phase II trial. *Nat. Cancer* 2, 1124–1135. doi:10. 1038/s43018-021-00269-7

Park, Y. R., Kim, S. L., Lee, M. R., Seo, S. Y., Lee, J. H., Kim, S. H., et al. (2017). MicroRNA-30a-5p (miR-30a) regulates cell motility and EMT by directly targeting oncogenic TM4SF1 in colorectal cancer. J. Cancer Res. Clin. Oncol. 143, 1915–1927. doi:10.1007/s00432-017-2440-4

Park, Y. R., Lee, S. T., Kim, S. L., Liu, Y. C., Lee, M. R., Shin, J. H., et al. (2016). MicroRNA-9 suppresses cell migration and invasion through downregulation of TM4SF1 in colorectal cancer. *Int. J. Oncol.* 48, 2135–2143. doi:10.3892/ijo.2016.3430

Peng, L. S., Zhang, J. Y., Teng, Y. S., Zhao, Y. L., Wang, T. T., Mao, F. Y., et al. (2017). Tumor-associated monocytes/macrophages impair NK-cell function via TGFβ1 in human gastric cancer. *Cancer Immunol. Res.* 5, 248–256. doi:10.1158/2326-6066. CIR-16-0152

Peng, D., Fu, M., Wang, M., Wei, Y., and Wei, X. (2022). Targeting TGF-β signal transduction for fibrosis and cancer therapy. *Mol. Cancer* 21, 104. doi:10.1186/s12943-022-01569-x

Perez-Banos, A., Gleisner, M. A., Flores, I., Pereda, C., Navarrete, M., Araya, J. P., et al. (2023). Whole tumour cell-based vaccines: tuning the instruments to orchestrate an optimal antitumour immune response. *Br. J. Cancer* 129, 572–585. doi:10.1038/s41416-023-02327-6

Phan, T. G., and Croucher, P. I. (2020). The dormant cancer cell life cycle. *Nat. Rev. Cancer* 20, 398-411. doi:10.1038/s41568-020-0263-0

Plaks, V., Kong, N., and Werb, Z. (2015). The cancer stem cell niche: how essential is the niche in regulating stemness of tumor cells? *Cell Stem Cell* 16, 225–238. doi:10.1016/j.stem.2015.02.015

Prasetyanti, P. R., and Medema, J. P. (2017). Intra-tumor heterogeneity from a cancer stem cell perspective. *Mol. Cancer* 16, 41. doi:10.1186/s12943-017-0600-4

Provenzano, P. P., Cuevas, C., Chang, A. E., Goel, V. K., Von Hoff, D. D., and Hingorani, S. R. (2012). Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma. Cancer Cell 21, 418–429. doi:10.1016/j. ccr.2012.01.007

Quail, D. F., Taylor, M. J., and Postovit, L. M. (2012). Microenvironmental regulation of cancer stem cell phenotypes. *Curr. Stem Cell Res. Ther.* 7, 197–216. doi:10.2174/157488812799859838

Rasheed, Z. A., Yang, J., Wang, Q., Kowalski, J., Freed, I., Murter, C., et al. (2010). Prognostic significance of tumorigenic cells with mesenchymal features in pancreatic adenocarcinoma. J. Natl. Cancer Inst. 102, 340–351. doi:10.1093/jnci/djp535

Regan, J. L., Schumacher, D., Staudte, S., Steffen, A., Haybaeck, J., Keilholz, U., et al. (2017). Non-canonical hedgehog signaling is a positive regulator of the WNT pathway and is required for the survival of colon cancer stem cells. *Cell Rep.* 21, 2813–2828. doi:10.1016/j.celrep.2017.11.025

Rengganaten, V., Huang, C.-J., Tsai, P.-H., Wang, M.-L., Yang, Y.-P., Lan, Y.-T., et al. (2020). Mapping a circular RNA-microRNA-mRNA-signaling regulatory Axis that modulates stemness properties of cancer stem cell populations in colorectal cancer spheroid cells. *Int. J. Mol. Sci.* 21, 7864. doi:10.3390/ijms21217864

Ren, J., Ding, L., Zhang, D., Shi, G., Xu, Q., Shen, S., et al. (2018). Carcinomaassociated fibroblasts promote the stemness and chemoresistance of colorectal cancer by transferring exosomal lncRNA H19. *Theranostics* 8, 3932–3948. doi:10.7150/thno. 25541

Ren, K., Li, Z., Li, Y., Zhang, W., and Han, X. (2017). Sulforaphene enhances radiosensitivity of hepatocellular carcinoma through suppression of the NF-κB pathway. J. Biochem. Mol. Toxicol. 31. doi:10.1002/jbt.21917

Reya, T., Morrison, S. J., Clarke, M. F., and Weissman, I. L. (2001). Stem cells, cancer, and cancer stem cells. *Nature* 414, 105–111. doi:10.1038/35102167

Rice, A. J., Cortes, E., Lachowski, D., Cheung, B. C. H., Karim, S. A., Morton, J. P., et al. (2017). Matrix stiffness induces epithelial-mesenchymal transition and promotes chemoresistance in pancreatic cancer cells. *Oncogenesis* 6, e352. doi:10.1038/oncsis. 2017.54

Sahai, E., Astsaturov, I., Cukierman, E., Denardo, D. G., Egeblad, M., Evans, R. M., et al. (2020). A framework for advancing our understanding of cancer-associated fibroblasts. *Nat. Rev. Cancer* 20, 174–186. doi:10.1038/s41568-019-0238-1

Saigusa, S., Toiyama, Y., Tanaka, K., Yokoe, T., Okugawa, Y., Kawamoto, A., et al. (2010). Stromal CXCR4 and CXCL12 expression is associated with distant recurrence and poor prognosis in rectal cancer after chemoradiotherapy. *Ann. Surg. Oncol.* 17, 2051–2058. doi:10.1245/s10434-010-0970-y

Salmon, H., Franciszkiewicz, K., Damotte, D., Dieu-Nosjean, M. C., Validire, P., Trautmann, A., et al. (2012). Matrix architecture defines the preferential localization and migration of T cells into the stroma of human lung tumors. *J. Clin. Invest.* 122, 899–910. doi:10.1172/JCI45817

Samavarchi Tehrani, S., Esmaeili, F., Shirzad, M., Goodarzi, G., Yousefi, T., Maniati, M., et al. (2023). The critical role of circular RNAs in drug resistance in gastrointestinal cancers. *Med. Oncol.* 40, 116. doi:10.1007/s12032-023-01980-4

Sathe, A., Mason, K., Grimes, S. M., Zhou, Z., Lau, B. T., Bai, X., et al. (2023). Colorectal cancer metastases in the liver establish immunosuppressive spatial networking between tumor-associated SPP1+ macrophages and fibroblasts. *Clin. Cancer Res.* 29, 244–260. doi:10.1158/1078-0432.CCR-22-2041

Schrader, J., Gordon-Walker, T. T., Aucott, R. L., Van Deemter, M., Quaas, A., Walsh, S., et al. (2011). Matrix stiffness modulates proliferation, chemotherapeutic response, and dormancy in hepatocellular carcinoma cells. *Hepatology* 53, 1192–1205. doi:10. 1002/hep.24108

Sharma, M. D., Hou, D. Y., Liu, Y., Koni, P. A., Metz, R., Chandler, P., et al. (2009). Indoleamine 2,3-dioxygenase controls conversion of Foxp3+ Tregs to TH17-like cells in tumor-draining lymph nodes. *Blood* 113, 6102–6111. doi:10.1182/blood-2008-12-195354

Sharma, N. S., Gupta, V. K., Dauer, P., Kesh, K., Hadad, R., Giri, B., et al. (2019). O-GlcNAc modification of Sox2 regulates self-renewal in pancreatic cancer by promoting its stability. *Theranostics* 9, 3410–3424. doi:10.7150/thno.32615

Shibue, T., and Weinberg, R. A. (2017). EMT, CSCs, and drug resistance: the mechanistic link and clinical implications. *Nat. Rev. Clin. Oncol.* 14, 611–629. doi:10.1038/nrclinonc.2017.44

Shi, X., Young, C. D., Zhou, H., and Wang, X. (2020). Transforming growth factor- β signaling in fibrotic diseases and cancer-associated fibroblasts. *Biomolecules* 10, 1666. doi:10.3390/biom10121666

Siegel, R. L., Miller, K. D., Wagle, N. S., and Jemal, A. (2023). Cancer statistics, 2023. *CA Cancer J. Clin.* 73, 17–48. doi:10.3322/caac.21763

Soleimani, A., Dadjoo, P., Avan, A., Soleimanpour, S., Rajabian, M., Ferns, G., et al. (2022). Emerging roles of CD133 in the treatment of gastric cancer, a novel stem cell biomarker and beyond. *Life Sci.* 293, 120050. doi:10.1016/j.lfs.2021.120050

Sonbol, M. B., Ahn, D. H., and Bekaii-Saab, T. (2019). Therapeutic targeting strategies of cancer stem cells in gastrointestinal malignancies. *Biomedicines* 7, 17. doi:10.3390/biomedicines7010017

Song, M., He, J., Pan, Q. Z., Yang, J., Zhao, J., Zhang, Y. J., et al. (2021). Cancerassociated fibroblast-mediated cellular crosstalk supports hepatocellular carcinoma progression. *Hepatology* 73, 1717–1735. doi:10.1002/hep.31792 Spranger, S., Dai, D., Horton, B., and Gajewski, T. F. (2017). Tumor-residing Batf3 dendritic cells are required for effector T cell trafficking and adoptive T cell therapy. *Cancer Cell* 31, 711–723. doi:10.1016/j.ccell.2017.04.003

Srivastava, R. K., Tang, S. N., Zhu, W., Meeker, D., and Shankar, S. (2011). Sulforaphane synergizes with quercetin to inhibit self-renewal capacity of pancreatic cancer stem cells. *Front. Biosci. Elite Ed.* 3, 515–528. doi:10.2741/e266

Straussman, R., Morikawa, T., Shee, K., Barzily-Rokni, M., Qian, Z. R., Du, J., et al. (2012). Tumour micro-environment elicits innate resistance to RAF inhibitors through HGF secretion. *Nature* 487, 500–504. doi:10.1038/nature11183

Su, Y. J., Lai, H. M., Chang, Y. W., Chen, G. Y., and Lee, J. L. (2011). Direct reprogramming of stem cell properties in colon cancer cells by CD44. *Embo J.* 30, 3186–3199. doi:10.1038/emboj.2011.211

Sun, C., Mezzadra, R., and Schumacher, T. N. (2018). Regulation and function of the PD-L1 checkpoint. *Immunity* 48, 434–452. doi:10.1016/j.immuni.2018.03.014

Sun, L., Huang, C., Zhu, M., Guo, S., Gao, Q., Wang, Q., et al. (2020). Gastric cancer mesenchymal stem cells regulate PD-L1-CTCF enhancing cancer stem cell-like properties and tumorigenesis. *Theranostics* 10, 11950–11962. doi:10.7150/thno.49717

Sun, M., Zhou, W., Zhang, Y. Y., Wang, D. L., and Wu, X. L. (2013). CD44(+) gastric cancer cells with stemness properties are chemoradioresistant and highly invasive. *Oncol. Lett.* 5, 1793–1798. doi:10.3892/ol.2013.1272

Su, Z., Sun, Y., Zhu, H., Liu, Y., Lin, X., Shen, H., et al. (2014). Th17 cell expansion in gastric cancer may contribute to cancer development and metastasis. *Immunol. Res.* 58, 118–124. doi:10.1007/s12026-013-8483-y

Swiderska-Syn, M., Xie, G., Michelotti, G. A., Jewell, M. L., Premont, R. T., Syn, W. K., et al. (2016). Hedgehog regulates yes-associated protein 1 in regenerating mouse liver. *Hepatology* 64, 232–244. doi:10.1002/hep.28542

Takaishi, S., Okumura, T., Tu, S., Wang, S. S., Shibata, W., Vigneshwaran, R., et al. (2009). Identification of gastric cancer stem cells using the cell surface marker CD44. *Stem Cells* 27, 1006–1020. doi:10.1002/stem.30

Takeda, K., Mizushima, T., Yokoyama, Y., Hirose, H., Wu, X., Qian, Y., et al. (2018). Sox2 is associated with cancer stem-like properties in colorectal cancer. *Sci. Rep.* 8, 17639. doi:10.1038/s41598-018-36251-0

Talukdar, S., Bhoopathi, P., Emdad, L., Das, S., Sarkar, D., and Fisher, P. B. (2019). Dormancy and cancer stem cells: an enigma for cancer therapeutic targeting. *Adv. Cancer Res.* 141, 43–84. doi:10.1016/bs.acr.2018.12.002

Tamada, M., Nagano, O., Tateyama, S., Ohmura, M., Yae, T., Ishimoto, T., et al. (2012). Modulation of glucose metabolism by CD44 contributes to antioxidant status and drug resistance in cancer cells. *Cancer Res.* 72, 1438–1448. doi:10.1158/0008-5472. CAN-11-3024

Tanaka, T., Narazaki, M., Masuda, K., and Kishimoto, T. (2016). Regulation of IL-6 in immunity and diseases. *Adv. Exp. Med. Biol.* 941, 79–88. doi:10.1007/978-94-024-0921-5_4

Tang, Y. A., Chen, Y. F., Bao, Y., Mahara, S., Yatim, S., Oguz, G., et al. (2018). Hypoxic tumor microenvironment activates GL12 via HIF-1 α and TGF- β 2 to promote chemoresistance in colorectal cancer. *Proc. Natl. Acad. Sci. U. S. A.* 115, E5990–e5999. doi:10.1073/pnas.1801348115

Tang, Q., Chen, J., Di, Z., Yuan, W., Zhou, Z., Liu, Z., et al. (2020). TM4SF1 promotes EMT and cancer stemness via the Wnt/ β -catenin/SOX2 pathway in colorectal cancer. *J. Exp. Clin. Cancer Res.* 39, 232. doi:10.1186/s13046-020-01690-z

Toledo-GuzmáN, M. E., HernáNDEZ, M. I., GóMEZ-Gallegos Á, A., and Ortiz-SáNCHEZ, E. (2019). ALDH as a stem cell marker in solid tumors. *Curr. Stem Cell Res. Ther.* 14, 375–388. doi:10.2174/1574888X13666180810120012

Tommelein, J., De Vlieghere, E., Verset, L., Melsens, E., Leenders, J., Descamps, B., et al. (2018). Radiotherapy-activated cancer-associated fibroblasts promote tumor progression through paracrine IGF1R activation. *Cancer Res.* 78, 659–670. doi:10. 1158/0008-5472.CAN-17-0524

Trelford, C. B., Dagnino, L., and Di Guglielmo, G. M. (2022). Transforming growth factor- β in tumour development. *Front. Mol. Biosci.* 9, 991612. doi:10.3389/fmolb.2022. 991612

Tsakiris, N., Fauvet, F., Ruby, S., Puisieux, A., Paquot, A., Muccioli, G. G., et al. (2020). Combined nanomedicines targeting colorectal cancer stem cells and cancer cells. *J. Control Release* 326, 387–395. doi:10.1016/j.jconrel.2020.07.025

Turley, S. J., Cremasco, V., and Astarita, J. L. (2015). Immunological hallmarks of stromal cells in the tumour microenvironment. *Nat. Rev. Immunol.* 15, 669–682. doi:10. 1038/nri3902

Tyagi, A., Sharma, A. K., and Damodaran, C. (2020). A review on Notch signaling and colorectal cancer. *Cells* 9, 1549. doi:10.3390/cells9061549

Ullmann, P., Nurmik, M., Schmitz, M., Rodriguez, F., Weiler, J., Qureshi-Baig, K., et al. (2019). Tumor suppressor miR-215 counteracts hypoxia-induced colon cancer stem cell activity. *Cancer Lett.* 450, 32–41. doi:10.1016/j.canlet.2019.02.030

Uthaya Kumar, D. B., Chen, C. L., Liu, J. C., Feldman, D. E., Sher, L. S., French, S., et al. (2016). TLR4 signaling via NANOG cooperates with STAT3 to activate Twist1 and promote formation of tumor-initiating stem-like cells in livers of mice. *Gastroenterology* 150, 707–719. doi:10.1053/j.gastro.2015.11.002

Van Den Brink, G. R., Hardwick, J. C., Nielsen, C., Xu, C., Ten Kate, F. J., Glickman, J., et al. (2002). Sonic hedgehog expression correlates with fundic gland differentiation in the adult gastrointestinal tract. *Gut* 51, 628–633. doi:10.1136/gut.51.5.628

Van De Wetering, M., Sancho, E., Verweij, C., De Lau, W., Oving, I., Hurlstone, A., et al. (2002). The beta-catenin/TCF-4 complex imposes a crypt progenitor phenotype on colorectal cancer cells. *Cell* 111, 241–250. doi:10.1016/s0092-8674(02)01014-0

Veenstra, V. L., Damhofer, H., Waasdorp, C., Steins, A., Kocher, H. M., Medema, J. P., et al. (2017). Stromal SPOCK1 supports invasive pancreatic cancer growth. *Mol. Oncol.* 11, 1050–1064. doi:10.1002/1878-0261.12073

Vermeulen, L., De Sousa, E. M. F., Van Der Heijden, M., Cameron, K., De Jong, J. H., Borovski, T., et al. (2010). Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *Nat. Cell Biol.* 12, 468–476. doi:10.1038/ncb2048

Vinson, K. E., George, D. C., Fender, A. W., Bertrand, F. E., and Sigounas, G. (2016). The Notch pathway in colorectal cancer. *Int. J. Cancer* 138, 1835–1842. doi:10.1002/ijc. 29800

Walcher, L., Kistenmacher, A. K., Suo, H., Kitte, R., Dluczek, S., Strauss, A., et al. (2020). Cancer stem cells-origins and biomarkers: perspectives for targeted personalized therapies. *Front. Immunol.* 11, 1280. doi:10.3389/fimmu.2020.01280

Wang, F. T., Sun, W., Zhang, J. T., and Fan, Y. Z. (2019a). Cancer-associated fibroblast regulation of tumor neo-angiogenesis as a therapeutic target in cancer. *Oncol. Lett.* 17, 3055–3065. doi:10.3892/ol.2019.9973

Wang, J. H., Huang, S. T., Zhang, L., Liu, Z. G., Liang, R. X., Jiang, S. W., et al. (2019b). Combined prognostic value of the cancer stem cell markers CD47 and CD133 in esophageal squamous cell carcinoma. *Cancer Med.* 8, 1315–1325. doi:10.1002/cam4. 1894

Wang, T. W., Chern, E., Hsu, C. W., Tseng, K. C., and Chao, H. M. (2020a). SIRT1-Mediated expression of CD24 and epigenetic suppression of novel tumor suppressor miR-1185-1 increases colorectal cancer stemness. *Cancer Res.* 80, 5257–5269. doi:10. 1158/0008-5472.CAN-19-3188

Wang, B., Liu, T., Wu, J. C., Luo, S. Z., Chen, R., Lu, L. G., et al. (2018a). STAT3 aggravates TGF-β1-induced hepatic epithelial-to-mesenchymal transition and migration. *Biomed. Pharmacother.* 98, 214–221. doi:10.1016/j.biopha.2017.12.035

Wang, J., Gong, R., Zhao, C., Lei, K., Sun, X., and Ren, H. (2023). Human FOXP3 and tumour microenvironment. *Immunology* 168, 248–255. doi:10.1111/imm.13520

Wang, L., Yang, H., Abel, E. V., Ney, G. M., Palmbos, P. L., Bednar, F., et al. (2015). ATDC induces an invasive switch in KRAS-induced pancreatic tumorigenesis. *Genes Dev.* 29, 171–183. doi:10.1101/gad.253591.114

Wang, X., Lang, M., Zhao, T., Feng, X., Zheng, C., Huang, C., et al. (2017). Cancer-FOXP3 directly activated CCL5 to recruit FOXP3(+)Treg cells in pancreatic ductal adenocarcinoma. *Oncogene* 36, 3048–3058. doi:10.1038/onc.2016.458

Wang, Y., Chen, M., Wu, Z., Tong, C., Dai, H., Guo, Y., et al. (2018b). CD133-directed CAR T cells for advanced metastasis malignancies: A phase I trial. *Oncoimmunology* 7, e1440169. doi:10.1080/2162402X.2018.1440169

Wang, Y., Wang, K., Han, G. C., Wang, R. X., Xiao, H., Hou, C. M., et al. (2014). Neutrophil infiltration favors colitis-associated tumorigenesis by activating the interleukin-1 (IIL-1)/IL-6 axis. *Mucosal Immunol.* 7, 1106–1115. doi:10.1038/mi. 2013.126

Wang, Z., Feng, T., Zhou, L., Jiang, D., Zhang, Y., He, G., et al. (2020b). Salinomycin nanocrystals for colorectal cancer treatment through inhibition of Wnt/ β -catenin signaling. *Nanoscale* 12, 19931–19938. doi:10.1039/d0nr04552g

Wang, Z., Sun, M., Li, W., Fan, L., Zhou, Y., and Hu, Z. (2020c). A novel CD133-and EpCAM-targeted liposome with redox-responsive properties capable of synergistically eliminating liver cancer stem cells. *Front. Chem.* 8, 649. doi:10.3389/fchem.2020.00649

Wang, Z., Zhou, L., Xiong, Y., Yu, S., Li, H., Fan, J., et al. (2019c). Salinomycin exerts anti-colorectal cancer activity by targeting the β -catenin/T-cell factor complex. *Br. J. Pharmacol.* 176, 3390–3406. doi:10.1111/bph.14770

Wan, S., Zhao, E., Kryczek, I., Vatan, L., Sadovskaya, A., Ludema, G., et al. (2014). Tumor-associated macrophages produce interleukin 6 and signal via STAT3 to promote expansion of human hepatocellular carcinoma stem cells. *Gastroenterology* 147, 1393–1404. doi:10.1053/j.gastro.2014.08.039

Wei, F., Zhang, T., Deng, S. C., Wei, J. C., Yang, P., Wang, Q., et al. (2019). PD-L1 promotes colorectal cancer stem cell expansion by activating HMGA1-dependent signaling pathways. *Cancer Lett.* 450, 1–13. doi:10.1016/j.canlet.2019.02.022

Wei, J., Wu, A., Kong, L. Y., Wang, Y., Fuller, G., Fokt, I., et al. (2011). Hypoxia potentiates glioma-mediated immunosuppression. *PLoS One* 6, e16195. doi:10.1371/journal.pone.0016195

Wen, L., Chen, X. Z., Yang, K., Chen, Z. X., Zhang, B., Chen, J. P., et al. (2013). Prognostic value of cancer stem cell marker CD133 expression in gastric cancer: a systematic review. *PLoS One* 8, e59154. doi:10.1371/journal.pone.0059154

Wilson, M. M., Weinberg, R. A., Lees, J. A., and Guen, V. J. (2020). Emerging mechanisms by which EMT programs control stemness. *Trends Cancer* 6, 775–780. doi:10.1016/j.trecan.2020.03.011

Wilusz, J. E., Sunwoo, H., and Spector, D. L. (2009). Long noncoding RNAs: functional surprises from the RNA world. *Genes Dev.* 23, 1494–1504. doi:10.1101/gad.1800909

Wu, B., Shi, X., Jiang, M., and Liu, H. (2023). Cross-talk between cancer stem cells and immune cells: potential therapeutic targets in the tumor immune microenvironment. *Mol. Cancer* 22, 38. doi:10.1186/s12943-023-01748-4

Wu, C., Zhu, X., Tao, K., Liu, W., Ruan, T., Wan, W., et al. (2018). MALAT1 promotes the colorectal cancer malignancy by increasing DCP1A expression and miR203 downregulation. *Mol. Carcinog.* 57, 1421–1431. doi:10.1002/mc.22868

Wuebben, E. L., Wilder, P. J., Cox, J. L., Grunkemeyer, J. A., Caffrey, T., Hollingsworth, M. A., et al. (2016). SOX2 functions as a molecular rheostat to control the growth, tumorigenicity and drug responses of pancreatic ductal adenocarcinoma cells. *Oncotarget* 7, 34890–34906. doi:10.18632/oncotarget.8994

Wu, X., Tao, P., Zhou, Q., Li, J., Yu, Z., Wang, X., et al. (2017a). IL-6 secreted by cancer-associated fibroblasts promotes epithelial-mesenchymal transition and metastasis of gastric cancer via JAK2/STAT3 signaling pathway. *Oncotarget* 8, 20741–20750. doi:10.18632/oncotarget.15119

Wu, Y., Chen, M., Wu, P., Chen, C., Xu, Z. P., and Gu, W. (2017b). Increased PD-L1 expression in breast and colon cancer stem cells. *Clin. Exp. Pharmacol. Physiol.* 44, 602–604. doi:10.1111/1440-1681.12732

Wu, Y., Grabsch, H., Ivanova, T., Tan, I. B., Murray, J., Ooi, C. H., et al. (2013). Comprehensive genomic meta-analysis identifies intra-tumoural stroma as a predictor of survival in patients with gastric cancer. *Gut* 62, 1100–1111. doi:10.1136/gutjnl-2011-301373

Xue, H. G., Yang, A. H., Sun, X. G., Lu, Y. Y., and Tian, Z. B. (2016). Expression of microRNA-328 functions as a biomarker for recurrence of early gastric cancer (EGC) after endoscopic submucosal dissection (ESD) by modulating CD44. *Med. Sci. Monit.* 22, 4779–4785. doi:10.12659/msm.896225

Xu, J., Zhang, Y., Jia, R., Yue, C., Chang, L., Liu, R., et al. (2019). Anti-PD-1 antibody SHR-1210 combined with apatinib for advanced hepatocellular carcinoma, gastric, or esophagogastric junction cancer: an open-label, dose escalation and expansion study. *Clin. Cancer Res.* 25, 515–523. doi:10.1158/1078-0432.CCR-18-2484

Yamashita, T., Ji, J., Budhu, A., Forgues, M., Yang, W., Wang, H. Y., et al. (2009). EpCAM-positive hepatocellular carcinoma cells are tumor-initiating cells with stem/ progenitor cell features. *Gastroenterology* 136, 1012–1024. doi:10.1053/j.gastro.2008. 12.004

Yang, H., Zhang, Q., Xu, M., Wang, L., Chen, X., Feng, Y., et al. (2020). CCL2-CCR2 axis recruits tumor associated macrophages to induce immune evasion through PD-1 signaling in esophageal carcinogenesis. *Mol. Cancer* 19, 41. doi:10.1186/s12943-020-01165-x

Yang, Y., Meng, W. J., and Wang, Z. Q. (2021). Cancer stem cells and the tumor microenvironment in gastric cancer. Front. Oncol. 11, 803974. doi:10.3389/fonc.2021.803974

Ye, M., Zhao, L., Zhang, L., Wu, S., Li, Z., Qin, Y., et al. (2022). LncRNA NALT1 promotes colorectal cancer progression via targeting PEG10 by sponging microRNA-574-5p. *Cell Death Dis.* 13, 960. doi:10.1038/s41419-022-05404-5

Yilmaz, M., and Christofori, G. (2009). EMT, the cytoskeleton, and cancer cell invasion. *Cancer Metastasis Rev.* 28, 15–33. doi:10.1007/s10555-008-9169-0

Yiming, L., Yunshan, G., Bo, M., Yu, Z., Tao, W., Gengfang, L., et al. (2015). CD133 overexpression correlates with clinicopathological features of gastric cancer patients and its impact on survival: a systematic review and meta-analysis. *Oncotarget* 6, 42019–42027. doi:10.18632/oncotarget.5714

Yoon, C., Lu, J., Jun, Y., Suh, Y. S., Kim, B. J., Till, J. E., et al. (2023). KRAS activation in gastric cancer stem-like cells promotes tumor angiogenesis and metastasis. *BMC Cancer* 23, 690. doi:10.1186/s12885-023-11170-0

Yoshida, G. J. (2020). Regulation of heterogeneous cancer-associated fibroblasts: the molecular pathology of activated signaling pathways. *J. Exp. Clin. Cancer Res.* 39, 112. doi:10.1186/s13046-020-01611-0

You, H., Ding, W., and Rountree, C. B. (2010). Epigenetic regulation of cancer stem cell marker CD133 by transforming growth factor-beta. *Hepatology* 51, 1635–1644. doi:10.1002/hep.23544

Yu, P. C., Long, D., Liao, C. C., and Zhang, S. (2018). Association between density of tumor-infiltrating lymphocytes and prognoses of patients with gastric cancer. *Med. Baltim.* 97, e11387. doi:10.1097/MD.000000000011387

Yu, L., Guo, Q. M., Wang, Y., Xu, Y., Liu, L., and Zhang, X. T. (2021). EpCAM expression in esophageal cancer and its correlation with immunotherapy of solitomab. *J. Thorac. Dis.* 13, 2404–2413. doi:10.21037/jtd-21-442

Yu, Q., and Stamenkovic, I. (1999). Localization of matrix metalloproteinase 9 to the cell surface provides a mechanism for CD44-mediated tumor invasion. *Genes Dev.* 13, 35–48. doi:10.1101/gad.13.1.35

Zeng, J. F., Ma, X. Q., Wang, L. P., and Wang, W. (2017). MicroRNA-145 exerts tumor-suppressive and chemo-resistance lowering effects by targeting CD44 in gastric cancer. *World J. Gastroenterol.* 23, 2337–2345. doi:10.3748/wjg.v23.i13.2337

Zhang, G. N., Liang, Y., Zhou, L. J., Chen, S. P., Chen, G., Zhang, T. P., et al. (2011). Combination of salinomycin and gemcitabine eliminates pancreatic cancer cells. *Cancer Lett.* 313, 137–144. doi:10.1016/j.canlet.2011.05.030

Zhang, H., Deng, T., Liu, R., Ning, T., Yang, H., Liu, D., et al. (2020). CAF secreted miR-522 suppresses ferroptosis and promotes acquired chemo-resistance in gastric cancer. *Mol. Cancer* 19, 43. doi:10.1186/s12943-020-01168-8

Zhang, H., Xie, C., Yue, J., Jiang, Z., Zhou, R., Xie, R., et al. (2017a). Cancer-associated fibroblasts mediated chemoresistance by a FOXO1/TGF β 1 signaling loop in esophageal squamous cell carcinoma. *Mol. Carcinog.* 56, 1150–1163. doi:10.1002/mc.22581

Zhang, H., Yue, J., Jiang, Z., Zhou, R., Xie, R., Xu, Y., et al. (2017b). CAF-secreted CXCL1 conferred radioresistance by regulating DNA damage response in a ROS-dependent manner in esophageal squamous cell carcinoma. *Cell Death Dis.* 8, e2790. doi:10.1038/cddis.2017.180

Zhang, N., Ng, A. S., Cai, S., Li, Q., Yang, L., and Kerr, D. (2021). Novel therapeutic strategies: targeting epithelial-mesenchymal transition in colorectal cancer. *Lancet Oncol.* 22, e358–e368. doi:10.1016/S1470-2045(21)00343-0

Zhang, Q., Zhang, H., Ding, J., Liu, H., Li, H., Li, H., et al. (2018). Combination therapy with EpCAM-CAR-NK-92 cells and regorafenib against human colorectal cancer models. *J. Immunol. Res.* 2018, 4263520. doi:10.1155/2018/4263520

Zhang, X., Hua, R., Wang, X., Huang, M., Gan, L., Wu, Z., et al. (2016). Identification of stem-like cells and clinical significance of candidate stem cell markers in gastric cancer. *Oncotarget* 7, 9815–9831. doi:10.18632/oncotarget.6890

Zhan, W., Liao, X., Wang, Y., Li, L., Li, J., Chen, Z., et al. (2019). circCTIC1 promotes the self-renewal of colon TICs through BPTF-dependent c-Myc expression. *Carcinogenesis* 40, 560–568. doi:10.1093/carcin/bgy144

Zhao, D., Pan, C., Sun, J., Gilbert, C., Drews-Elger, K., Azzam, D. J., et al. (2015a). VEGF drives cancer-initiating stem cells through VEGFR-2/Stat3 signaling to upregulate Myc and Sox2. *Oncogene* 34, 3107–3119. doi:10.1038/onc.2014.257

Zhao, H., Chen, S., and Fu, Q. (2020). Exosomes from CD133(+) cells carrying circ-ABCC1 mediate cell stemness and metastasis in colorectal cancer. *J. Cell Biochem.* 121, 3286–3297. doi:10.1002/jcb.29600

Zhao, L., Yang, Y., Zhou, P., Ma, H., Zhao, X., He, X., et al. (2015b). Targeting CD133high colorectal cancer cells *in vitro* and *in vivo* with an asymmetric bispecific antibody. *J. Immunother.* 38, 217–228. doi:10.1097/CJI.0000000000086

Zhao, S., Chen, C., Chang, K., Karnad, A., Jagirdar, J., Kumar, A. P., et al. (2016). CD44 expression level and isoform contributes to pancreatic cancer cell plasticity, invasiveness, and response to therapy. *Clin. Cancer Res.* 22, 5592–5604. doi:10.1158/ 1078-0432.CCR-15-3115

Zheng, H., Liu, H., Li, H., Dou, W., Wang, J., Zhang, J., et al. (2022). Characterization of stem cell landscape and identification of stemness-relevant prognostic gene signature to aid immunotherapy in colorectal cancer. *Stem Cell Res. Ther.* 13, 244. doi:10.1186/s13287-022-02913-0

Zheng, X., Carstens, J. L., Kim, J., Scheible, M., Kaye, J., Sugimoto, H., et al. (2015). Epithelial-to-mesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. *Nature* 527, 525–530. doi:10.1038/nature16064

Zhi, Q. M., Chen, X. H., Ji, J., Zhang, J. N., Li, J. F., Cai, Q., et al. (2011). Salinomycin can effectively kill ALDH(high) stem-like cells on gastric cancer. *Biomed. Pharmacother.* 65, 509–515. doi:10.1016/j.biopha.2011.06.006

Zhou, B., Lin, W., Long, Y., Yang, Y., Zhang, H., Wu, K., et al. (2022a). Notch signaling pathway: architecture, disease, and therapeutics. *Signal Transduct. Target Ther.* 7, 95. doi:10.1038/s41392-022-00934-y

Zhou, Y., Xu, J., Luo, H., Meng, X., Chen, M., and Zhu, D. (2022b). Wnt signaling pathway in cancer immunotherapy. *Cancer Lett.* 525, 84–96. doi:10.1016/j.canlet.2021.10.034

Zhu, Y., Wang, C., Becker, S. A., Hurst, K., Nogueira, L. M., Findlay, V. J., et al. (2018). miR-145 antagonizes SNAI1-mediated stemness and radiation resistance in colorectal cancer. *Mol. Ther.* 26, 744–754. doi:10.1016/j.ymthe.2017.12.023