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The progress of microenvironment-targeted therapies in brain metastases

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The incidence of brain metastases (BrM) has become a growing concern recently. It is a common and often fatal manifestation in the brain during the end-stage of many extracranial primary tumors. Increasing BrM diagnoses can be attributed to improvements in primary tumor treatments, which have extended patients' lifetime, and allowed for earlier and more efficient detection of brain lesions. Currently, therapies for BrM encompass systemic chemotherapy, targeted therapy, and immunotherapy. Systemic chemotherapy regimens are controversial due to their associated side effects and limited efficacy. Targeted and immunotherapies have garnered significant attention in the medical field: they target specific molecular sites and modulate specific cellular components. However, multiple difficulties such as drug resistance and low permeability of the blood-brain barrier (BBB) remain significant challenges. Thus, there is an urgent need for novel therapies. Brain microenvironments consist of cellular components including immune cells, neurons, endothelial cells as well as molecular components like metal ions, nutrient molecules. Recent research indicates that malignant tumor cells can manipulate the brain microenvironment to change the anti-tumoral to a pro-tumoral microenvironment, both before, during, and after BrM. This review compares the characteristics of the brain microenvironment in BrM with those in other sites or primary tumors. Furthermore, it evaluates the preclinical and clinical studies of microenvironment-targeted therapies for BrM. These therapies, due to their diversity, are expected to overcome drug resistance or low permeability of the BBB with low side effects and high specificity. This will ultimately lead to improved outcomes for patients with secondary brain tumors.

KEYWORDS

brain metastases, brain microenvironment, targeted therapy, immunotherapy, glioma, microenvironment-targeted therapy

1 Introduction

The prevalence of brain metastasis (BrM) is approximately 8.5%–9.6%. However, this likely underestimates the true incidence of BrM because many cancer patients with this condition are not included within the statistics. BrM is becoming more frequent even more frequent than any other primary tumor in the brain, with lung cancer having the highest incidence at (19.9%), followed by melanoma (6.9%), renal cancer (6.5%), breast cancer (5.1%), and colorectal cancer (1.8%) (Valiente et al., 2018). The increased incidence of BrM may be attributed to the prolonged overall survival (OS) of patients with cancer because of novel cancer therapies. Current medical treatments for BrM

primarily include systemic chemotherapy, targeted therapy, and immunotherapy (Achrol et al., 2019). Traditional systemic agents chemotherapeutic such as pemetrexed and temozolomide have a limited role in management of BrM due to the BBB or the blood-tumor barrier (BTB). Targeted therapy has dramatically improved the OS of patients with BrM (Valiente et al., 2018), for instance, osimertinib targeting epidermal growth factor receptor (EGFR) mutated non-small cell lung cancer (NSCLC) BrM, lapatinib targeting human epidermal growth factor receptor 2 (HER-2) mutated breast cancer, and B-Raf kinase gene (BRAF) inhibitor dabrafenib combined with the mitogen-activated protein kinase (MAPK) pathway inhibitor trametinib targeting melanoma (Achrol et al., 2019). Additionally, novel drugs targeting immune checkpoints, including cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) (ipilimumab), programmed cell death protein 1 (PD-1) (pembrolizumab and nivolumab), and programmed deathligand 1 (PD-L1) (atezolizumab), also have incredibly prolonged OS of BrM patients (Achrol et al., 2019).

Despite advancements in BrM therapy, there are significant challenges such as drug resistance and low permeability of the BBB. These challenges have led to growing concerns in the development of novel therapies with high efficacy. Cancer metastasis is a complex process involving multiple interactions and mechanisms, including but not limited to dissemination, intravasation, circulating, extravasation, and colonization (Fares et al., 2020). The unique microenvironment of BrM promotes the expansion of tumor cells and protects them from anti-tumoral insults. This specific microenvironment mainly includes cellular and non-cellular components, the former being the focus of recent research. Cellular components include astrocytes, neurons, tumor-infiltrating lymphocytes, tumor-associated macrophages, natural killer cells (Tomaszewski et al., 2019). Non-cellular components mainly refer to the extracellular matrix (ECM) and BBB (Achrol et al., 2019). The ECM includes nutrients such as oxygen and electrolytes such as iron and calcium ions, exosomes, etc., (Hoshino et al., 2015; Basnet et al., 2019; Chi et al., 2020). Despite the emergence of many cutting-edge therapeutic targets and drugs for various types of BrM, the therapeutic effects have been unsatisfactory. This can be attributed to the complex microenvironment of BrM, which poses significant challenges for effective treatment. Firstly, the BBB separates the brain from the peripheral circulation in the microenvironment, preventing the drugs that target the primary tumor from penetrating the brain tissue. Secondly, the structure of the BBB changes after brain metastasis, forming the bloodtumor barrier (BTB). Then the BTB makes it difficult for drugs that initially passed through the BBB to enter the brain tissue (Achrol et al., 2019). Thirdly, BrM is heterogeneous, with differences in the molecular traits of primary tumors and metastases leading to a loss of original targets of cancer cells (Achrol et al., 2019). Furthermore, the microenvironment of metastases is also heterogeneous with its microenvironment modified by metastatic cells which differ from the primary tumour (Mansfield et al., 2016; Priego et al., 2018; Kim et al., 2019; Kudo et al., 2019; Vareslija et al., 2019; Sirkisoon et al., 2020). This renders drugs that were initially effective against the primary tumor microenvironment ineffective. Additionally, it was previously believed that the brain was 'immune privileged. However, recent research has shown that immune cells exist in the brain microenvironment (Jiang et al., 2020), and their role in promoting metastatic tumor outgrowth is still being studied in detail (Jiang et al., 2020).

The microenvironmental therapy that we are proposing includes a series of therapies which target components of the BrM microenvironment or inhibit the interaction between cancer cells and the microenvironment components to inhibit tumor growth. This is a different strategy to traditional chemotherapy in that it acts precisely on specific cells or molecules as well as not directly targeting and killing cancer cells and through induced cytotoxicity. Although this therapy is still being debated regarding its efficacy and side effects, it has been demonstrated in cell experiments to have advantages in overcoming drug resistance or low permeability of the BBB with low side effects and high specificity (Tomaszewski et al., 2019). This therapy showed promising drug efficacy when administered alone and can also be combined with other traditional drugs for improved effectiveness (Tomaszewski et al., 2019).

In addition, single cell sequencing technology develop fast. In the field of BrM, this technology gives a deep insight into the expression of specific cells in BrM microenvironment (Gonzalez et al., 2022). Single cell sequence not only analyses the genomic and proteomic profile of BrM microenvironment, especially immune cells, but also provides various prognosis and treatment approaches (Parker et al., 2020; Gonzalez et al., 2022; Jin et al., 2022). It is believed that development of single cell sequencing technology will contribute to the exploration in BrM microenvironment (Gonzalez et al., 2022).

It is important to note that microenvironment therapy must consider the heterogeneity of BrM. Previous metastatic cancer antagonists were designed based on the assumption that primary tumor and metastases are the same on a molecular level; however, they differ significantly (Valiente et al., 2018). Microenvironment targeted therapy has shown promising prospects in the preclinical and clinical stages, but there are still substantial unexplored fields with many medicine designs still in therapeutic setting. In this review, we highlight the uniqueness of the BrM microenvironment targeted therapy according to cell types, including astrocytes, macrophages and microglia, T lymphocytes, and endothelial cells, and other components.

2 The uniqueness of BrM microenvironment

The tumor microenvironment is essential for tumor cells to survive. It undergoes dynamic cellular and non-cellular components changes, which could promote the heterogeneity of cancer cells, clonal evolution, and drug resistance. The microenvironment of BrM differs from that of the primary brain tumors. Understanding these differences between BrM may aid in the development of targeted treatments strategies for different types of BrM as well as drugs with broad anti-cancer capabilities for common targets.

BrMs from different primary sites					
	NSCLC BrM	Breast cancer BrM	Melanoma BrM	Colon cancer BrM	
Checkpoint pathways genes	Ť	-	-	-	
Lymphocyte infiltration genes	1	-	-	-	
CD4 ⁺ and CD8+T cells	_	-	1	-	
Neutrophils	—	Î	-	-	

TABLE 1 Comparison between BrMs from different primary sites. Checkpoint pathways genes include genes of TNFRSF9, TNFRSF4, PDCDILG2, IDO1, ICOS, CD274. Lymphocyte infiltration genes include genes of TARP, PTPRC, PTPN7, IL10RA, GZMK, CD52, CD2, CCR5, CCL5. "-." represents that the value is relatively normal compared with the decreased or increased one. "/" means no outcomes available. "^" means overexpression. "\]" means relatively low expression.

2.1 Differences in brain microenvironment among BrMs coming from different primary sites

BrMs from different primary sites experience differences at both the molecular and cellular level. For example, BrMs from lung cancer, breast cancer, and colon cancer. BrM in lung cancer showed significantly higher expression of commonly mutated genes, especially the genes regulating checkpoint pathways [tumor necrosis factor receptor superfamily 9 (TNFRSF9), tumor necrosis factor receptor superfamily 4 (TNFRSF4), programmed cell death 1 ligand 2 (PDCDILG2), indoleamine 2,3-dioxygenase 1 (IDO1), inducible T Cell costimulator (ICOS), cluster of differentiation 274 (CD274)] and lymphocyte infiltration [T cell receptor gamma alternate reading frame protein (TARP), protein tyrosine phosphatase receptor type c (PTPRC), protein tyrosine phosphatase non-receptor type 7 (PTPN7), interleukin 10 receptor subunit alpha (IL10RA), granzyme l (GZMK), cluster of differentiation 52 (CD52), cluster of differentiation (CD2), C-C motif chemokine receptor 5 (CCR5), C-C motif chemokine ligand 5 (CCL5)], indicating that lung cancer brain metastasis may be more sensitive to immunotherapy (Jiang et al., 2020). In contrast, BrM from melanoma have higher presence of CD4 and CD8 positive T cells, while BrM in breast cancer has a higher presence of neutrophils and macrophages, indicating that breast cancer BrM may show increased drug resistance compared with melanoma BrM (Klemm et al., 2020; Gonzalez et al., 2022). Besides, lung cancer BrM has more T lymphocytes, while melanoma and oval cancer have more B lymphocytes (Gonzalez et al., 2022). Through single cell sequencing technology, Sudmeier, et al. also found that CD8+ T cell phenotype is linked with spatial distribution within the tumor (Gonzalez et al., 2022). Additionally, hypoxia was found to be a characteristic in lung cancer BrM compared with other types of BrMs (Corroyer-Dulmont et al., 2021). Despite differences in the microenvironment of BrMs from different primary sites, several similarities have been found. In vivo experiments have reported that both breast cancer cells and lung cancer cells interact with astrocytes in the central nervous system to improve the expression of survival genes, including B-cell lymphoma-2-like protein 1 (BCL2L1), twist family BHLH transcription factor 1 (TWIST1), and glutathione S-transferase alpha 5 (GSTA5) to obtain drug resistance. (Table 1) (Kim et al., 2011; Vareslija et al., 2019). These differences in the microenvironment of BrMs from primary sites have important implications for treatment options and highlight the need for tailored therapies.

2.2 Differences between BrMs and glioma

Primary and metastatic tumors, such as gliomas and BrM have distinct differences in tumor microenvironment. For example, Klemm, et al. reported that gliomas contain many tumor-associated macrophages (TAMs), but have few T cells, especially in isocitrate dehydrogenase (IDH)-mutated tumors. However, lymphocytes and neutrophils are abundant in BrM, and the microenvironment of BrM contains more immune regulatory factors including CD40L, interleukin 6 receptor (IL6R), inhibin subunit beta a (INHBA), and amphiregulin (AREG) according to experiment data from Klemm, et al. (Klemm et al., 2020). Plasma tumors (especially IDH mutant gliomas) have a lower presence of immune cells (Klemm et al., 2020) (Table 2). However, data from Sun, et al. showed different results. The experiment showed that lung cancer BrM contains more macrophages, T lymphocytes, and mastocytes, while gliomas contains more astrocytes and microglia (Perelroizen et al., 2022; Sun et al., 2022), possibly because microenvironment can change as the glioma progression goes on (increases in macrophages, lymphocytes, monocytes and NK cells over time) (Varn et al., 2022; Yeo et al., 2022). Besides, subclones of stroma cells in glioma were found to be much more than that in BrM (Schaettler et al., 2022). This is possibly because gliomas arise from the slow accumulation of somatic mutations, while BrM likely develop quickly from the rapid growth of an already transformed subclone upon arrival into the central nervous system (CNS). This malignant clone will quickly develop into an apparent tumor, allowing less time for the development of genetically disparate subclones (Schaettler et al., 2022).

2.3 Differences in microenvironments between primary tumor sites and the matched BrMs

Differences in immune microenvironments between primary tumor and BrM, especiallyNSCLC, have been explored. In NSCLC BrM, dendritic cell maturation procession and leukocyte extravasation signalling pathways in innate immunity are inhibited, while tumor-associated macrophages are increased. Regarding specific immunity, PD-1+ lymphocytes, helper T cells 1, and PD-L1+ immune cells are reduced in BrM, which may explain why PD-L1 targeted immunotherapy is a poor therapeutic strategy for patients with BrM. Interestingly, due to the accordance of antigen between

TABLE 2 Comparison between BrMs and glioma. NSCLC represents non-small cell lung cancer.

	BrMs				Intracranial primary cancer	
	NSCLC BrM	Breast cancer BrM	Melanoma cancer BrM	Colon cancer BrM	Liver cancer BrM	Glioma
CD4 ⁺ and CD8+T cells	—	-	Î	-	—	\downarrow
Neutrophils	_	1			_	-
TAMs	Ť	-			—	Î

TABLE 3 Comparison between BrMs and primary tumors. Anti-inflammatory markers include TOLLIP, HLA-G. Cell cycle dysregulation genes include E2F3 and RB. Proto-oncogenes stand for KRAS and ALK. Kinase-driven pathways represent SRC, mTOR and HER2. Metastasis formation-associated genes include CXCR4, PLLP, TNFSF4, VCAM1, SLC8A2, and SLC7A11. BrM/primary tumor represents relative relationship between BrM and primary tumor. TAMs, tumor-associated macrophages. NSCLC, non-small cell lung cancer.

	NSCLC BrM/primary tumor	Breast cancer BrM/primary tumor	Melanoma BrM/primary tumor
PD-1+ and PD-L1+ T cells	\downarrow	_	_
TAMs	<u>↑</u>	_	_
Dendritic cell maturation	\downarrow	_	_
Leukocyte extravasation	Ļ	_	_
TCR clonality	↑ (_	_
Anti-inflammatory markers	1	_	_
Interferon-y-related gene signature	Ļ	_	_
Cell cycle dysregulation genes	-	1	-
Proto-oncogenes	-	1	_
Kinase-driven pathways genes	-	↑	_
Metastasis formation-associated genes	_	Î	Î

BrM and primary tumor, T cell receptor (TCR) repertoire remains relatively unchanged (Mansfield et al., 2016; Kim et al., 2019; Kudo et al., 2019). Additionally, astrocytes which express signal transducer and activator of transcription 3 (STAT3) contribute to BrM by regulating innate and specific immunity as well as establish gap junctions with metastases cells to enhance the growth ability and drug resistance of tumor cells (Chen et al., 2016; Sato et al., 2017; Priego et al., 2018; Kim et al., 2019; Song S. G. et al., 2021; Song Z. et al., 2021). When compared with primary breast cancer, breast cancer BrM tends to have lower infiltration of immune cells (macrophages, microglia, lymphocytes, and monocytes), lower protein and gene expression of immune activation markers (CD27, T cell immunoglobulin and mucin-domain containing-3 (Tim-3), and CD137), and lower expression of immune-related genes (PD-L1 and CTLA-4) (Schlam et al., 2021; Giannoudis et al., 2022). In melanoma BrM, oxidative phosphorylation (OXPHOS) gene set in Kyoto Encyclopedia of Genes and Genomes (KEGG) database is enriched compared with primary cancer (Fischer et al., 2019), while melanoma BrM has lower T-cell content and microvessel density (Weiss et al., 2021). Besides, Sato, et al. found several genes upregulation in breast cancer and melanoma BrM compared with their primary tumors. In breast cancer BrM, cell cycle dysregulation genes [E2F transcription factor 3 (E2F3) and retinoblastoma protein (RB)], proto-oncogenes [Kirsten rat sarcoma virus (KRAS) and anaplastic lymphoma kinase (ALK)], kinase-driven pathways genes [src proto-oncogene (SRC), mechanistic target of rapamycin kinase (mTOR) and HER2], metastases formation-associated genes [C-X-C motif chemokine receptor 4 (CXCR4), plasmolipin (PLLP), tumor necrosis factor superfamily member 4 (TNFSF4), vascular cell adhesion molecule 1 (VCAM-1), solute carrier family 8 member a2 (SLC8A2), and solute carrier family 7 member a11 (SLC7A11)] were overexpressed (Chen et al., 2016; Song S. G. et al., 2021; Song Z. et al., 2021). Only metastasis formation-associated genes were upregulated in melanoma BrM compared with melanoma primary tumor (Sato et al., 2017). Nevertheless, immunohistochemical similarity of immune cells including tumor infiltrating lymphocytes (TIL) and TAMs is reported in gynaecological malignancies/renal cell carcinoma and their matched BrM (Gill et al., 2021; Steindl et al., 2021) (Table 3).

2.4 Differences in microenvironments between intracranial metastases and extracranial metastases

There are many differences in microenvironment between intracranial metastases and extracranial metastases, including differences in physical condition, immune cells, extracellular matrix, and exosomes.

Breast cancer					Melanoma	
	Brain metastasis	Liver metastasis	Lung metastasis	Bone metastasis	Brain metastasis	Extracranial metastasis
Characterized tumor cell-derived proteins	CD109, SERPINB1, HCFC1 and cerebellin-1	COL6A5	collagen COL4A4 and laminin-121	\$100A6 and \$100A11	—	_
Characterized stroma cell-derived proteins	Secreted neuronal glycoprotein Lgi1, Adam22 and brevican	Tnc, Fn1, fibrinogens, thrombin and von Willebrand factor	Laminin chains, type IV collagens and pulmonary surfactantassociated protein A1	Thrombospondin-1, another S100 protein, the protease cathespin-G, the protease inhibitors cystatin C and stefin-2	_	_
Exosome integrins	ITGβ3	ΙΤGανβ5	ITGα6β4 and ITGα6β1	—	—	_
CD4 ⁺ and CD8 ⁺ T cells	Ļ	-	-	-	Ļ	-
Overall extracellular matrix protein quantity	Ļ	-	-	-	_	_
Glycoproteins proportion	-	Î	-	-	_	_
Immunosuppression	_	_	—	—	1	_
Oxidative phosphorylation	_	_	_	_	_	Î

TABLE 4 Comparison between BrMs and extracranial metastasis. ITG, integrin.

2.4.1 Physical condition

Physical conditions also play a role in molecular expression in metastasis with different organotropism. For example, breast cancer lung metastasis has an increased expression of mitochondrial electron transport Complex I, oxidative stress, and counteracting antioxidant programs compared to primary breast cancer and breast cancer brain metastasis, which may be related to more oxidative stress in the lung (Basnet et al., 2019; Hebert et al., 2020; Weiss et al., 2021).

2.4.2 Immune cells

Immune cells in BrM, such as CD4 and CD8 positive T cells, are fewer compared to extracranial metastases. However, other immune cells such as B cells and macrophages do not show significant differences. Fewer T lymphocytes lead to less immunoediting of metastatic cells, which may contribute to the lower expression of tyrosinase in BrM (Bartlett et al., 2014). Although there are fewer T lymphocytes in BrM, intracranial and extracranial metastases have similar responses to immune checkpoint inhibitors, indicating that T lymphocytes may not play a crucial role in metastasis. Additionally, BrM cells have been found to have an unstable gene expression and a neural-like cell state, which suggests that BrM cells adapt to the BrM microenvironment through interaction with intracranial stromal cells (Bartlett et al., 2014).

2.4.3 Extracellular matrix

The protein in ECM is composed of both tumor and stromal cell-derived proteins. This is due to the influence of tumor cells on stromal cell protein synthesis. This is due to the influence of tumor cells on stromal cell protein synthesis. Common protein expressions in the ECM of BrM include s100 calcium binding protein a4 (S100A4), annexin a2 (ANXA2), transforming growth factor beta 1 (TGFB1) and CCR2, while different protein expressions include

more serpin family B member 1 (SERPINB1) in the brain (Hebert et al., 2020; Weiss et al., 2021).

2.4.4 Exosome

Articles reported that exosomes expressed by tumor cells could guide tumor metastasis to organotropism (Hoshino et al., 2015). Different integrins (ITGs) on exosomes determine the organotropism of tumor metastases. The type of integrins is different in exosomes with different organotropism and specific cells living in the targeted organs will absorb these exosomes for more precise organ targeting. For example, ITGa6β4 and ITGa6β1 are associated with lung metastasis, while ITGavβ5 is associated with liver metastasis. ITGβ3 was separated from BrM cells *in vitro*, and exosomes from 831-BrT cells were absorbed by cerebral epithelial cells specifically (Hoshino et al., 2015) (Table 4).

3 Therapies targeting BrM microenvironment

Previous research focuses primarily on genomic, epigenomic, and transcriptomic landscapes of cancer cells (Achrol et al., 2019). Understanding the microenvironments and protein expression of different types of cancer can lead to significant improvements in cancer therapies, such as targeted therapies. This has the potential to significantly improve survival rates of cancer such as driver gene mutated lung adenocarcinoma, breast cancer, and melanoma. However, most cancers still develop resistance to this treatment strategy, highlighting the importance of further research in molecular communication between cancer cells and the surrounding microenvironment. However, relatively limited studies focus on the treatment by targeting the microenvironment of metastatic brain tumors. Targeting the

TABLE 5 Recent clinical trials on BrM microenvironment targeted therapies have been listed above. While medicines with new design conception are included, those commonly-seen medicines like VEGF inhibitor apatinib, CTLA-4 inhibitor ipilimumab, PD-1 inhibitor nivolumab, etc. Are not listed. Fulvestrant and tamoxifen citrate are estrogen receptor inhibitor. WP1066 is JAK2/STAT3 inhibitor. BKM120 is PI3K inhibitor. Endostar is recombinant VEGF inhibitor, QBS10072S LAT1-targeted medicine, GRN1005 a novel conjugate of angiopep-2 and paclitaxel, all of which are aim to transport through the BBB. The dendritic cell vaccine has been applied to other fields of cancer in the past, while until recently has it been tested in BrM. Microenvironment targeted therapies used here are marked red.

ANG1005 in breast cancer patients with Recurrent Brain metastases	Drug: GRN 1005	Completed	NCT02048059	Phase 2
ANG1005 in Leptomeningeal Disease From Breast Cancer	Drug: GRN 1005 Drug: Physician's Best Choice	Not yet recruiting	NCT03613181	Phase 3
Expanded Access to ANG1005 for Individual Patients	Drug: GRN 1005	No longer available	NCT02755987	
Dose Escalation Study to Assess the Safety, Tolerability, Pharmacokinetics, and Pharmacodynamics of QBS10072S	Drug: QBS10072S	Recruiting	NCT04430842	Phase 1
Dendritic Cell Vaccines Against Her2/Her3, Cytokine Modulation Regimen, and Pembrolizumab for the Treatment of Brain Metastasis From Triple Negative Breast Cancer or HER2+ Breast Cancer	Biological: Anti-HER2/HER3 Dendritic Cell Vaccine Drug: Celecoxib Biological: Pembrolizumab Biological: Recombinant Interferon Alfa-2b Drug: Rintatolimod	Not yet recruiting	NCT04348747	Phase 2
A Phase II, Open-Label, Multicenter Study of Capmatinib in Subjects With MET Exon 14 Skipping Mutation Positive, Advanced, NSCLC With Brain Metastases	Drug: Capmatinib	Withdrawn	NCT04460729	Phase 2

cellular components of the microenvironment of BrM can be used as a novel strategy to treat cancer.

3.1 Targeting astrocytes

The interaction between astrocytes and cancer cells can occur directly through the gap junction, paracrine, exosome, and certain gene regulations like STAT3. These signalling pathways can be useful drug targets.

3.1.1 Targeting gap junction

One example is targeting gap junctions: astrocytes can establish gap junctions with metastatic cancer cells and activate downstream signalling pathways to promote metastasis formation. Protocadherin 7 (PCDH7) on the cell membranes of these 2 cells interact with each other first to act as an "anchor," and then the connexin 43 (Cx43) molecules on the two membranes are connected to form a gap junction for molecular exchange (Chen et al., 2016). This can then promote cancer metastasis through three pathways: the cyclic guanosine monophosphate–adenosine monophosphate (cGAMP) pathway, calcium ion pathway, and endothelin pathway.

The cGAMP pathway refers to the transfer of cGAMP from cancer cells to astrocytes through gap junctions, stimulating astrocytes to express interferon (IFN) and tumor necrosis factor alpha (TNF- α). These act directly on the surface receptors of cancer cells through the paracrine system, and the downstream pathways such as STAT1 and nuclear factor kappa b (NF- κ B) pathways enhance the proliferation and drug resistance of cancer cells. Meclofenamate and tonabersat are Cx43 gap junction blockers, effectively inhibiting *in vitro* cancer metastasis (Chen et al., 2016).

Calcium ions play a complex role in modulating tumor cell apoptosis. Research has found that persistent increases in calcium benefit tumor cell survival, while transient increases lead to apoptosis. The calcium ion pathway refers to the persistent transmigration of calcium ions from astrocytes into cancer cells through gap junctions which leads to an increase in the activity of phosphodiesterase beta (PLC β) in the cancer cell cytoplasm. PLC β hydrolyzes phosphatidylinositol 4,5-bisphosphate (PIP2) into inositol 1,4,5-trisphosphate (IP3) and 1,2-diacylglycerol (DAG). These molecules store the transiently increased calcium ions in the cell to form a continuously high calcium intracellular environment. These calcium ions promote the activation of calcium-dependent molecules, such as calmodulin-dependent protein kinase II (CaMK2) and S100A4. These promote cancer metastasis. Studies involving the inhibition of PLC with drugs like edelfosine inhibit cancer metastasis in mouse models (Biermann et al., 2022).

Nevertheless, the report implicated that calcium chelator effectively limited BrM by restraining ionic calcium concentrations in the cytoplasm of in vitro tumor cells. One theory is that this resulted in a transient concentration increase of ionic calcium and subsequent tumor cell apoptosis (Fischer et al., 2019). The various results of increased ionic calcium concentration on cancer cells are not fully understood. It is likely that different signalling pathways or astrocyte and cancer cell interactions may be involved. Further research is needed to explore this mechanism and understand the underlying causes. In the endothelin pathway, the expression of interleukin-6 (IL-6) and interleukin-8 (IL-8) molecules in cancer cells increases when gap junctions are established. These molecules promote the expression of endothelin and its receptors [endothelin receptor a (ETAR) and endothelin receptor b (ETBR)] on astrocytes through paracrine signalling. Cancer cells stimulated by endothelin activate the AKT/MAPK pathway, which enhances their drug resistance (Kim et al., 2014). A dual endothelin receptor antagonist called macitentan has been found to increase the effectiveness of the drug paclitaxel in treating BrMs from breast and lung cancer in mice model (Lee et al., 2016). However, clinical trials have not been seen. Further studies are needed.

Astrocytes can also reciprocally transmit signals to cancer cells through direct small molecule signalling. Cancer cells secrete molecules such as macrophage migration inhibitory factor (MIF), interleukin-8 (IL-8), and plasminogen activator inhibitor-1 (PAI-1) subsequently activating astrocytes. Astrocytes then secrete IL-6, TNF-a, IL-1β, IL-23, and other signalling molecules that promote cancer cell proliferation (Jaffe, 2005; Lee et al., 2016; Ren et al., 2018). IL-3 plays a vital role in the extravasation of cancer cells by promoting the production of matrix metallopeptidase 2 (MMP-2) by cancer cells, enhancing the invasiveness of cancer cells (Fischer et al., 2019). Paracrine signalling between these 2 cells is not necessarily reciprocal, it can be unilateral instead. Another article found that estradiol activates astrocytes during triple-negative breast cancer (TNBC) metastasis, activating astrocytes, promoting metastasis. This pathway promotes the expression of brain derived neurotrophic factor (BDNF) and epidermal growth factor (EGF) after estradiol binds to the intracellular receptors of astrocytes. BDNF and EGF act on downstream receptors [tropomyosin receptor kinase b (TrkB) and EGFR] on the surface of cancer cells and activate pathways ensuring cancer cell invasiveness (Embi et al., 2012; Kim et al., 2014). Anti-IL-6 receptor antibody tocilizumab (Lee et al., 2016), IL-23 neutralizing antibody, and anti-TrkB antibody ANA-12 have been identified as possible drugs in the treatment of metastatic cancer in mice model (Lee et al., 2016). Further toxicity tests and clinical trials may come into sight in the future.

3.1.3 Targeting exosome

The exosome pathway is a recently discovered mechanism by which astrocytes can promote cancer growth. Astrocytes deliver PTEN-targeted miRNAs to metastatic cancer cells through exosomes, reducing PTEN expression. This leads to increased expression of chemokine CCL2 in cancer cells and recruits many myeloid cells (microglia and macrophages) that are ionized calcium binding adaptor molecule 1 (IBA1) positive. The latter cells can promote cancer cell proliferation and reduce apoptosis. Targeting PTEN, exosomes, and CCL2 through drug development may be a potential therapeutic strategy for treating BrM(Seike et al., 2011; Klein et al., 2015).

3.1.4 Targeting STAT3

Astrocytes can also promote cancer growth by affecting the immunity in the brain microenvironment. Neibla Priego et al. found that STAT3-positive astrocytes can inhibit the anti-tumor effect of CD8-positive T cells and promote aggregation of CD74-positive cells (macrophages and microglia) to interfere with innate immunity. Silibinin (trade name Legasil) is a drug that specifically binds to the src homology 2 (SH2) domain on the pSTAT3 molecule of astrocytes, inhibiting the action of STAT3, and enhancing the efficacy of chemotherapy drugs in brain tumors when used in combination (Priego et al., 2018; Contreras-Zarate et al., 2019). Case report of Silibinin used in BrM was reported by Neibla, et al. (Priego et al., 2018). As silibinin has already been applied in clinical trials in other diseases like hepatitis and breast cancer, the prospect of silibinin can be promising (Ferenci et al., 2008; Lazzeroni et al., 2016).

3.2 Targeting macrophages and microglia

Unlike astrocytes, macrophages promote tumor BrM by affecting the ability or number of other immune cells. Therefore, drug treatments should aim to reduce the number of macrophages in the brain or to change the expression of macrophages in the brain.

3.2.1 PI3K

Phosphoinositide 3-kinase (PI3K) promotes BrM by driving the differentiation of macrophages into a tumor-promoting phenotype. PI3K is a master regulator of brain-metastases-promoting macrophages and microglia. The infiltration of marrow-derived-macrophages (MDM) is inhibited by PI3K inhibitor buparlisib, and the MDM-induced tumor invasion can be reduced significantly (Caetano et al., 2016). PI3K inhibitor buparlisib can reduce both macrophage/microglia induced metastases, implicating those microglia involved in metastases promotion (Benbenishty et al., 2019).

3.2.2 CSF-1

Colony stimulator factor-1 (CSF-1) plays a crucial role as a growth factor of macrophages and microglia in a paracrine loop between metastases cells and monocyte-derived cells. Although studies have shown that anti-CSF-1 (5A1, a CSF-1 antagonist) treatment effectively limits macrophage proliferation in BrM *in vitro*, an alternative CSF-1 ligand (IL-34) released by both metastatic cells and normal brain microenvironment can have a CSF-1-like effect rendering anti-CSF-1 treatment ineffective. Therefore, when considering treatment options, targeting IL-34 in addition to anti-CSF-1 treatment should be taken into account in order to achieve optimal results (Noda et al., 2012).

3.2.3 LncRNA

Long non-coding RNAs (lncRNAs) have emerged as a new target in BrM research. LncRNAs are expressed by metastatic cells and activate a signalling pathway involving Janus kinase 2 (JAK2), oncostatin M, IL-6, and STAT3 to increase the expression of ICAM-1 and CCL2. This enhances the ability of the cancer cells to attach to capillary endothelial cells and recruit macrophages. These recruited macrophages can further promote lncRNA expression by releasing oncostatin M and IL-6. Depletion of Lnc-BM with nanoparticle-encapsulated siRNAs in mice model effectively treated BrM^{51.} Therefore, developing therapies that target oncostatin M and IL-6 or directly target lncRNA may be a promising strategy for treating BrM (Zhang et al., 2015).

3.2.4 Toll-like receptor 9

Microglia are a type of immune cell found in the brain microenvironment which exert anti-metastases effects. They are activated by CpG oligodeoxynucleotides (CpG-ODN), a toll-like receptor (TLR) 9 agonist, causing the microglia to display an increased expression of anti-tumor genes, inducing apoptosis of metastases cells by direct contact with tumor or early metastases focus (Rietkotter et al., 2015). Although inhibition of primary tumor was proved to be valid in CpG ODN administration, Xiong, et al. found that in mice model, the therapeutic effect was limited in treating BrM, due to the increase in regulatory T lymphocytes (T_{reg} S) in BrM (Marabelle et al., 2013). T_{reg}S play an important role in

immune suppression in BrM. The suppression of other immune cells like microglia may be the reason why CpG ODN cannot limit the BrM progression (Xiong et al., 2008; Marabelle et al., 2013).

3.2.5 Nanocarrier

Drug delivery *via* nanocarrier is a new direction in current research for BrM treatment. Nanocarrier-mediated drug delivery is a promising approach that could lead to the development of more effective and targeted therapies for BrM in the future. This new technology has several advantages over traditional drugs: multiple signal molecules can be set on the membrane of nanoparticles allowing for multiple cells to be targeted, the nanoparticle can carry a great amount of traditional anti-tumor drugs once completed decreasing time spent on finding drug targets and nanoparticles (unlike brain tumor antagonists) have a more complicated design allowing them to cross the BBB more effectively (Zhang et al., 2019).

Tian Zhang et al. designed a nanoparticle, iRGD-terpolymerlipid hybrid nanoparticle with coloaded doxorubicin and mitomycin c (iRGD-DMTPLN), with an iRGD signal molecule iRGD on it, which targets endothelial and BrM cells as well as TAMs. After interacting with endothelial cells, the particle can be endocytosed by the cells, transferred from peripheral circulation to the brain microenvironment, and then be endocytosed by TAMs and tumor cells, releasing the contained drug. This nanoparticle effectively reduces metastases burden and TAMs, significantly restoring the brain microenvironment (Zhang et al., 2019).

Pengfei Zhao et al. designed a T12 peptide-modified albumin nanoparticle coloaded with regorafenib and disulfiram/copper ion chelant. This particle also has multiple target cells, including brain tumor capillary endothelial cells, tumor vessel endothelial cells, cancer cells, and TAMs. The drug acting on TAMs repolarized $M2\Phi$ into $M1\Phi$ instead of killing macrophages (Zhao et al., 2021).

3.2.6 Ionic iron

Macrophages have both an indirect and direct pathway to affect tumor outgrowth. In Leptomeningeal metastases (LM), inflammatory cytokines secreted from macrophages induce ironbinding protein lipocalin-2 (LCN2) and its receptor solute carrier family 22 member 17 (SCL22A17) expression in metastatic cells. Tumor cells can then obtain ionic iron from the extracellular matrix and contribute to chemical synthesis. An experiment in a mouse model showed that iron chelator deferoxamine inhibited LM (Chi et al., 2020). Microglia also have a role in metastasis formation, so their inhibition is crucial in the treatment of BrM.

3.3 Targeting T lymphocytes

T lymphocytes are critical in the anti-tumor effect but are constantly inhibited by tumor cells and tumor-associated macrophages. T lymphocytes activation pathways are essential to immunotherapy including: target drug treatment, virus transfection, and adoptive cell transfer therapy.

3.3.1 PD-1

Anti-PD-1 treatment has been a popular approach in clinical drugs, as it rescues T lymphocytes, particularly CD8 $^+$ T

lymphocytes, from an immunosuppressive state and is specific in its action. Additionally, this treatment not only improves the antitumor ability but also increases the number of T lymphocytes, aggregation rather through peripheral primarily than proliferation within the brain microenvironment. The anti-PD-1 treatment has also been combined with other therapies to enhance the drug's effect and elevate the number of CD44+CD62L-effector cells. It has been proposed that after stereotactic radiosurgery (SRS) or whole brain radiation therapy, immunotherapy including anti-PD-1 treatment can accelerate radiation-induced brain tissue changes by enhancing the immune microenvironment, as reflected by increased infiltration of T lymphocytes, mainly CD8⁺ T cells and reactive astrocytosis. However, it is been also reported that whole brain radiotherapy can sensitize breast cancer BrM to anti-PD-1 treatment in mouse models (Qian et al., 2011).

3.3.2 Oncolytic viruses'

Oncolytic viruses (OVs) have recently emerged as a promising option in the anti-tumor field. OVs specifically target tumor cells as they are artificially modified. Upon infection of tumor cells, cytotoxic T cells are recruited to the infected lesion and kill the infected metastatic cells (Benencia et al., 2005; Gao et al., 2015; Du et al., 2017). Furthermore, OVs stimulate tumor cells to secrete IFN, which plays a key role in the PD-1/PD-L1 axis and promotes the expression of PD-L1 on metastatic cells. Research has shown that the effectiveness of anti-PD-1 therapy is determined by the expression of PD-L1 on the tumor, highlighting the importance of the PD-1/PD-L1 axis in treatment response (Steele et al., 2011).

Anti-PD-L1 treatment can have a more effective therapeutic action on metastases after OVs administration (Zhao et al., 2021). However, OVs immunotherapy is associated with virus-related toxicity (Benbenishty et al., 2019). Wanlu Du et al. invented oncolytic herpes simplex virus (oHSV)-armed mesenchymal stem cells (MSCs), which utilize MSCs as a virus carrier (Alomari et al., 2016; Taggart et al., 2018). This method avoids virus-related toxicity while keeping BBB penetration and high specificity. Clinically, OVs therapy has recently proven effective through intracarotid injection and local injection (Taggart et al., 2018).

3.3.3 Adoptive cell transfer therapy

Adoptive cell transfer therapy (ACT therapy) includes various techniques such as chimeric antigen receptor-engineered T cell therapy (CAR T cell therapy), TIL therapy, and T cell receptor (TCR) therapy. These therapies have been applied clinically, but until recently, have not been widely tested in the context of BrM (Priceman et al., 2018). However, recent studies have shown that HER2+ breast cancer, NSCLC, and melanoma BrM are responsive to ACT therapy. For example, a study by Priceman et al. compared CD137 CAR T cells and CD28 CAR T cells in treating HER2+ breast cancer and found that CD137 CAR T cells had improved tumor killing effects, with reduced T-cell exhaustion and greater proliferative capacity (Priceman et al., 2018). Additionally, intraventricular injection was found to be as effective as intratumoral injection, offering a new administration option. Another study by Wang et al. used recombinant adenoassociated virus-denditic cells (DCs) to stimulate T lymphocytes and successfully limited tumor outgrowth in 3 patients (Wang



immunity by activating pSTAT3. The green boxes contain drugs used in experiments targeting microenvironment *in vitro* or *in vivo*. $\bigcirc - @$ belong to gunction-dependent pro-tumoral interaction. \bigcirc belong to exosome-dependent pro-tumoral interaction. \bigcirc belong to exosome-dependent pro-tumoral interaction.

et al., 2020). These DCs contained several specific tumorantigenic determinant associated genes, including carcinoembryonic antigen (CEA), cytokeratin 19 (CK19), and prostate specific membrane antigen (PSMA), highlighting the potential of ACT therapies in treating BrM (Priceman et al., 2018; Wang et al., 2020). Recent study showed that CD19targeted CAR T cells treat BrM not only through directly causing tumor cell apoptosis. Through single cell sequence, Parker, et al. found that brain mural cells which sustain integrity of BBB express CD19. When CD19-targeted CAR T cells attack mural cells and break BBB, cytokines can easily leak through the BBB and limit the BrM progression (Parker et al., 2020).

3.4 Targeting endothelial cells

Endothelial cells are a component of the BBB and play a crucial role in the extravasation of metastatic cells as well as giving metastatic cells several protections to promote tumor growth. The interaction between endothelial cells and metastatic cells can be divided into three parts, based on which drugs are designed (Haymaker et al., 2017).

First, metastatic cells can disrupt the inter-endothelial junction. MicroRNA (miR-101-3p), inhibits the cyclooxygenase-2 (COX-2)/ MMP-1 pathway, where MMP-1 degrades the gap junction between endothelial cells. Loss of miR-101-3p therefore significantly increases the metastatic ability of metastatic cells. Experiments *in vitro* reported that restoring these microRNA in tumor cells decreased the metastatic burden in the brain (Niesel et al., 2021).

Secondly, metastatic cells cross the BBB through adhesion and metamorphosis. Adhesion is a vital step during this progress. In terms of tumor cells, $\alpha 3\beta 1$ integrins can be elevated by α B-crystallin expressed in metastatic cells (Xiong et al., 2008). Silencing α B-crystallin through lentiviral and retroviral transduction *in vitro* blocked BrM progression (Malin et al., 2014). VCAM-1 is constantly expressed in endothelial cells, but significantly increased during inflammation (Marabelle et al., 2013). Hence, silencing these proteins on these 2 cells is proved to be effective in inhibiting BrM. microparticles of iron oxide (MPIOs)_{-VCAM-1}, a VCAM-1 antagonist, succeed in limiting BrM progression in mice model (Sikpa et al., 2020).



endothelial cells and potential drug targets in brain microenvironment. The green boxes contain drugs used in experiments targeting microenvironment in vitro or in vivo.

Thirdly, after tumor cells extravasate into the brain stroma, tumor cells tend to stay close to endothelial cells for protection from immune cells and drugs. The gap junction between these 2 cells is essential for tumor growth. After the gap junction is established, endothelial cells receive IL-6 and IL-8 from tumor cells and secrete endothelin to tumor cells both through the paracrine pathway, resulting in an increased expression of survival protein in tumor cells (Benencia et al., 2005; Du et al., 2017). In this way, HER-2+ breast cancer BrM gains drug resistance against the targeted drug trastuzumab emtansine (T-DM1). Therefore, macitentan, a dual endothelin receptor (ETAR and ETBR) antagonist, is also applied, in order to inhibit this protective effect and can also be combined with a cytotoxic drug to increase drug effect (Gao et al., 2015). Furthermore, endothelial cells can secrete exosomes to tumor cells, resulting in an elevated expression of s100 calcium binding protein a16 (S100A16), which promotes prohibitin-1 (PHB-1) expression and inhibits apoptosis. siRNA inhibiting prohibitin-1

(PHB-1) to promote apoptosis of tumor cells is proved valid (Steele et al., 2011).

Additionally, some medicines are designed to transport more easily through the endothelial cells, thus acting more accurately on cancer cells. For instance, GRN1005, a novel conjugate of angiopep-2, a peptide facilitating brain penetration, and paclitaxel, is proved to be effective in endothelial transcytosis due to amplification of angiopep-2 on the surface of endothelial cells with BrM(Xu et al., 2019).

3.5 Targeting other cellular components of microenvironments

There are a variety of other cellular components that have been identified as potential targets in the microenvironment of BrM, but few studies have investigated these. For example, choroid plexus

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epithelial cells can be affected by complement component 3 (C3) produced by leptomeningeal metastases (LM) cells. C3 molecules act on C3a receptors on choroid plexus epithelial cells, resulting in the degradation of the intercellular tight junction. This permeabilization of the blood-brain barrier alters, allowing growth factors such as amphiregulin and other mitogens to enter and promote the growth of LM cells. C3aR inhibitor (SB290157) has been found to limit the growth of LM in mouse models (Boire et al., 2017). Additionally, neutrophils, which are PD-L1 positive, have been identified as immunosuppressive cells in BrM. Studies have shown that metastatic cells recruit immunosuppressive neutrophils through the Src/enhancer of zeste homolog 2 (EZH2)/c-JUN/granulocyte colony-stimulating factor (G-CSF) pathway, inhibiting the cytotoxic effect of CD8⁺ T lymphocytes. Inhibition of metastatic growth in mouse models has been achieved through blockade of G-CSF or Src (Zhang et al., 2020). Furthermore, research has also suggested that pericytes in the blood-brain barrier may have an anti-tumor role, but the underlying mechanisms are yet to be fully explored (Samson et al., 2018). What's more, Jin, et al. found that microglia were the dominant cell population in NSCLC BrM microenvironment through single cell sequencing. They identified IL-6 as the key regulator in BrM cells to induce anti-inflammatory microglia via JAK2/STAT3 signaling, which in turn promoted the colonization process in BrM cells. IL6 inhibitors (tocilizumab or fedratinib) can block JAK2/STAT3 activation and impede BrM of NSCLC cells in mice model (Jin et al., 2022).

3.6 Summary

In conclusion, current microenvironment-targeted therapies mainly focus on cellular component, including astrocytes, microglia, macrophages, T lymphocytes, and endothelial cells. Besides, we provide two figures to conclude all the mentioned pro-tumoral mechanisms and the corresponding targeted therapies (Figures 1, 2).

Tumor cells interact with astrocytes through gap junction, paracrine, exosomes, and STAT3 expression. Meclofenamate, tonabersat, edelfosine, and macitentan have the potential to disrupt the gap junction. Tocilizumab and ANA-12 inihibit IL-6 and TrkB paracrine pathway, respectively. Silibinin is an anti-STAT3 medicine. Besides, medicine targeting exosomes may be a potential therapeutic strategy.

Macrophages and microglia play an important role in BrM progression. PI3K, CSF-1, LncRNA, TLR9, and ionic iron are target molecule. Buparlisib and deferoxamine are PI3K and ionic iron inhibitors. In addition, nanocarriers targeting macrophages are proved to be effective *in vitro* and in mice model.

Several therapies targeting T lymphocytes include anti-PD-1 treatment (pembrolizumab and nivolumab), OVs therapy, and ACT therapy (CAR T cell therapy, TIL therapy, and TCR therapy).

Endothelial cells promote extravasation of metastatic cells in three steps. MPIOs-VCAM-1, macitentan, S100A16, and GRN1005 can inhibit one of these steps.

Also, SB290157 act on choroid plexus epithelial cells to inhibit metastatic progress. Therapies targeting other components are to be explored. Although some of the medicines are proved to be effective only *in vitro* or in mouse models, some are applied in clinical trials. Here, we provide a table concluding concurrent clinical trials which use microenvironment-targeted medicines (Table 5).

4 Perspective

With a survival expectancy of patients is 17.6 months with single BrM and 17.9 months with multiple BrMs (Ferguson et al., 2012), BrM threaten the survival of cancer patients. Nowadays, current therapies cannot meet patients' need to cure or prevent BrM. Microenvironment therapies provide a novel target with cellular microenvironment components leading to a complemented efficacy when combined with systemic treatments or other targeted therapies. Due to the lack of cytotoxic effects associated with microenvironment therapies, they typically do not have severe side effects unlike systemic chemotherapies or some targeted therapies. Furthermore, some microenvironment therapies enhance the efficacy of conventional drugs by reducing drug resistance and/or alternating the permeability of the BBB. Due to their indirect interaction pathway to inhibit tumor growth, whether these drugs can have a good response when clinically used is still not confirmed. Progress has been made in exploring mechanisms of interaction cells between and metastatic the brain microenvironment, drugs mentioned in these articles aim mainly at tumor cells instead of the microenvironment. Besides, the contribution of single cell sequence technology in the analysis, diagnosis, and treatment of BrM is promising and more researches about this approach are needed. Brain microenvironment deserves more concern due to its potential to cure or limit the outgrowth of BrM. Medicines targeting the microenvironment directly should be considered as novel therapeutics.

Author contributions

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

Achrol, A. S., Rennert, R. C., Anders, C., Soffietti, R., Ahluwalia, M. S., Nayak, L., et al. (2019). Brain metastases. *Nat. Rev. Dis. Prim.* 5 (1), 5. doi:10.1038/s41572-018-0055-y

Alomari, A. K., Cohen, J., Vortmeyer, A. O., Chiang, A., Gettinger, S., Goldberg, S., et al. (2016). Possible interaction of anti-PD-1 therapy with the effects of radiosurgery on brain metastases. *Cancer Immunol. Res.* 4 (6), 481–487. doi:10.1158/2326-6066.CIR-15-0238

Bartlett, E. K., Fetsch, P. A., Filie, A. C., Abati, A., Steinberg, S. M., Wunderlich, J. R., et al. (2014). Human melanoma metastases demonstrate nonstochastic site-specific antigen heterogeneity that correlates with T-cell infiltration. *Clin. Cancer Res.* 20 (10), 2607–2616. doi:10.1158/1078-0432.CCR-13-2690

Basnet, H., Tian, L., Ganesh, K., Huang, Y. H., Macalinao, D. G., Brogi, E., et al. (2019). Flura-seq identifies organ-specific metabolic adaptations during early metastatic colonization. *Elife* 8, e43627. doi:10.7554/eLife.43627

Benbenishty, A., Gadrich, M., Cottarelli, A., Lubart, A., Kain, D., Amer, M., et al. (2019). Prophylactic TLR9 stimulation reduces brain metastasis through microglia activation. *PLoS Biol.* 17 (3), e2006859. doi:10.1371/journal.pbio.2006859

Benencia, F., Courreges, M. C., Conejo-Garcia, J. R., Mohamed-Hadley, A., Zhang, L., Buckanovich, R. J., et al. (2005). HSV oncolytic therapy upregulates interferon-inducible chemokines and recruits immune effector cells in ovarian cancer. *Mol. Ther.* 12 (5), 789–802. doi:10.1016/j.ymthe.2005.03.026

Biermann, J., Melms, J. C., Amin, A. D., Wang, Y., Caprio, L. A., Karz, A., et al. (2022). Dissecting the treatment-naive ecosystem of human melanoma brain metastasis. *Cell* 185 (14), 2591–2608.e30. doi:10.1016/j.cell.2022.06.007

Boire, A., Zou, Y., Shieh, J., Macalinao, D. G., Pentsova, E., and Massague, J. (2017). Complement component 3 adapts the cerebrospinal fluid for leptomeningeal metastasis. *Cell* 168 (6), 1101–1113.e13. doi:10.1016/j.cell.2017.02.025

Caetano, M. S., Zhang, H., Cumpian, A. M., Gong, L., Unver, N., Ostrin, E. J., et al. (2016). IL6 blockade reprograms the lung tumor microenvironment to limit the development and progression of K-ras-Mutant lung cancer. *Cancer Res.* 76 (11), 3189–3199. doi:10.1158/0008-5472.CAN-15-2840

Chen, Q., Boire, A., Jin, X., Valiente, M., Er, E. E., Lopez-Soto, A., et al. (2016). Carcinoma-astrocyte gap junctions promote brain metastasis by cGAMP transfer. *Nature* 533 (7604), 493–498. doi:10.1038/nature18268

Chi, Y., Remsik, J., Kiseliovas, V., Derderian, C., Sener, U., Alghader, M., et al. (2020). Cancer cells deploy lipocalin-2 to collect limiting iron in leptomeningeal metastasis. *Science* 369 (6501), 276–282. doi:10.1126/science.aaz2193

Contreras-Zarate, M. J., Day, N. L., Ormond, D. R., Borges, V. F., Tobet, S., Gril, B., et al. (2019). Estradiol induces BDNF/TrkB signaling in triple-negative breast cancer to promote brain metastases. *Oncogene* 38 (24), 4685–4699. doi:10.1038/s41388-019-0756-z

Corroyer-Dulmont, A., Valable, S., Fantin, J., Chatre, L., Toutain, J., Teulier, S., et al. (2021). Multimodal evaluation of hypoxia in brain metastases of lung cancer and interest of hypoxia image-guided radiotherapy. *Sci. Rep.* 11 (1), 11239. doi:10.1038/s41598-021-90662-0

Du, W., Seah, I., Bougazzoul, O., Choi, G., Meeth, K., Bosenberg, M. W., et al. (2017). Stem cell-released oncolytic herpes simplex virus has therapeutic efficacy in brain metastatic melanomas. *Proc. Natl. Acad. Sci. U. S. A.* 114 (30), E6157–E6165. doi:10. 1073/pnas.1700363114

Embi, A., Scherlag, B. J., Embi, P. J., Menes, M., and Po, S. S. (2012). Targeted cellular ionic calcium chelation by oxalates: Implications for the treatment of tumor cells. *Cancer Cell Int.* 12 (1), 51. doi:10.1186/1475-2867-12-51

Fares, J., Fares, M. Y., Khachfe, H. H., Salhab, H. A., and Fares, Y. (2020). Molecular principles of metastasis: A hallmark of cancer revisited. *Signal Transduct. Target Ther.* 5 (1), 28. doi:10.1038/s41392-020-0134-x

Ferenci, P., Scherzer, T. M., Kerschner, H., Rutter, K., Beinhardt, S., Hofer, H., et al. (2008). Silibinin is a potent antiviral agent in patients with chronic hepatitis C not responding to pegylated interferon/ribavirin therapy. *Gastroenterology* 135 (5), 1561–1567. doi:10.1053/j.gastro.2008.07.072

Ferguson, M. S., Lemoine, N. R., and Wang, Y. (2012). Systemic delivery of oncolytic viruses: Hopes and hurdles. *Adv. Virol.* 2012, 805629. doi:10.1155/2012/805629

Fischer, G. M., Jalali, A., Kircher, D. A., Lee, W. C., McQuade, J. L., Haydu, L. E., et al. (2019). Molecular profiling reveals unique immune and metabolic features of melanoma brain metastases. *Cancer Discov.* 9 (5), 628–645. doi:10.1158/2159-8290.CD-18-1489

Gao, Y., Whitaker-Dowling, P., Barmada, M. A., Basse, P. H., and Bergman, I. (2015). Viral infection of implanted meningeal tumors induces antitumor memory T-cells to organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

travel to the brain and eliminate established tumors. Neuro Oncol. 17 (4), 536-544. doi:10.1093/neuonc/nou231

Giannoudis, A., Vareslija, D., Sharma, V., Zakaria, R., Platt-Higgins, A., Rudland, P. S., et al. (2022). Characterisation of the immune microenvironment of primary breast cancer and brain metastasis reveals depleted T-cell response associated to ARG2 expression. *ESMO Open* 7 (6), 100636. doi:10.1016/j.esmoop.2022.100636

Gill, C. M., D'Andrea, M. R., Tomita, S., Suhner, J., Umphlett, M., Zakashansky, K., et al. (2021). Tumor immune microenvironment in brain metastases from gynecologic malignancies. *Cancer Immunol. Immunother.* 70 (10), 2951–2960. doi:10.1007/s00262-021-02909-4

Gonzalez, H., Mei, W., Robles, I., Hagerling, C., Allen, B. M., Hauge Okholm, T. L., et al. (2022). Cellular architecture of human brain metastases. *Cell* 185 (4), 729–745.e20. doi:10.1016/j.cell.2021.12.043

Haymaker, C. L., Kim, D., Uemura, M., Vence, L. M., Phillip, A., McQuail, N., et al. (2017). Metastatic melanoma patient had a complete response with clonal expansion after whole brain radiation and PD-1 blockade. *Cancer Immunol. Res.* 5 (2), 100–105. doi:10.1158/2326-6066.CIR-16-0223

Hebert, J. D., Myers, S. A., Naba, A., Abbruzzese, G., Lamar, J. M., Carr, S. A., et al. (2020). Proteomic profiling of the ECM of xenograft breast cancer metastases in different organs reveals distinct metastatic niches. *Cancer Res.* 80 (7), 1475–1485. doi:10.1158/0008-5472.CAN-19-2961

Hoshino, A., Costa-Silva, B., Shen, T. L., Rodrigues, G., Hashimoto, A., Tesic Mark, M., et al. (2015). Tumour exosome integrins determine organotropic metastasis. *Nature* 527 (7578), 329–335. doi:10.1038/nature15756

Jaffe, L. F. (2005). A calcium-based theory of carcinogenesis. Adv. Cancer Res. 94, 231–263. doi:10.1016/S0065-230X(05)94006-2

Jiang, J., Wu, L., Yuan, F., Ji, J., Lin, X., Yang, W., et al. (2020). Characterization of the immune microenvironment in brain metastases from different solid tumors. *Cancer Med.* 9 (7), 2299–2308. doi:10.1002/cam4.2905

Jin, Y., Kang, Y., Wang, M., Wu, B., Su, B., Yin, H., et al. (2022). Targeting polarized phenotype of microglia via IL6/JAK2/STAT3 signaling to reduce NSCLC brain metastasis. *Signal Transduct. Target Ther.* 7 (1), 52. doi:10.1038/s41392-022-00872-9

Kim, R., Keam, B., Kim, S., Kim, M., Kim, S. H., Kim, J. W., et al. (2019). Differences in tumor microenvironments between primary lung tumors and brain metastases in lung cancer patients: Therapeutic implications for immune checkpoint inhibitors. *BMC Cancer* 19 (1), 19. doi:10.1186/s12885-018-5214-8

Kim, S. J., Kim, J. S., Park, E. S., Lee, J. S., Lin, Q., Langley, R. R., et al. (2011). Astrocytes upregulate survival genes in tumor cells and induce protection from chemotherapy. *Neoplasia* 13 (3), 286–298. doi:10.1593/neo.11112

Kim, S. W., Choi, H. J., Lee, H. J., He, J., Wu, Q., Langley, R. R., et al. (2014). Role of the endothelin axis in astrocyte- and endothelial cell-mediated chemoprotection of cancer cells. *Neuro Oncol.* 16 (12), 1585–1598. doi:10.1093/neuonc/nou128

Klein, A., Schwartz, H., Sagi-Assif, O., Meshel, T., Izraely, S., Ben Menachem, S., et al. (2015). Astrocytes facilitate melanoma brain metastasis via secretion of IL-23. *J. Pathol.* 236 (1), 116–127. doi:10.1002/path.4509

Klemm, F., Maas, R. R., Bowman, R. L., Kornete, M., Soukup, K., Nassiri, S., et al. (2020). Interrogation of the microenvironmental landscape in brain tumors reveals disease-specific alterations of immune cells. *Cell* 181 (7), 1643–1660.e17. doi:10.1016/j. cell.2020.05.007

Kudo, Y., Haymaker, C., Zhang, J., Reuben, A., Duose, D. Y., Fujimoto, J., et al. (2019). Suppressed immune microenvironment and repertoire in brain metastases from patients with resected non-small-cell lung cancer. *Ann. Oncol.* 30 (9), 1521–1530. doi:10.1093/annonc/mdz207

Lazzeroni, M., Guerrieri-Gonzaga, A., Gandini, S., Johansson, H., Serrano, D., Cazzaniga, M., et al. (2016). A presurgical study of oral silybin-phosphatidylcholine in patients with early breast cancer. *Cancer Prev. Res. (Phila)* 9 (1), 89–95. doi:10.1158/1940-6207.CAPR-15-0123

Lee, H. J., Hanibuchi, M., Kim, S. J., Yu, H., Kim, M. S., He, J., et al. (2016). Treatment of experimental human breast cancer and lung cancer brain metastases in mice by macitentan, a dual antagonist of endothelin receptors, combined with paclitaxel. *Neuro Oncol.* 18 (4), 486–496. doi:10.1093/neuonc/now037

Malin, D., Strekalova, E., Petrovic, V., Deal, A. M., Al Ahmad, A., Adamo, B., et al. (2014). αB-crystallin: A novel regulator of breast cancer metastasis to the brain. *Clin. Cancer Res.* 20 (1), 56–67. doi:10.1158/1078-0432.CCR-13-1255

Mansfield, A. S., Aubry, M. C., Moser, J. C., Harrington, S. M., Dronca, R. S., Park, S. S., et al. (2016). Temporal and spatial discordance of programmed cell death-ligand 1 expression and lymphocyte tumor infiltration between paired primary lesions and brain metastases in lung cancer. *Ann. Oncol.* 27 (10), 1953–1958. doi:10.1093/annonc/mdw289

Marabelle, A., Kohrt, H., Sagiv-Barfi, I., Ajami, B., Axtell, R. C., Zhou, G., et al. (2013). Depleting tumor-specific Tregs at a single site eradicates disseminated tumors. *J. Clin. Invest.* 123 (6), 2447–2463. doi:10.1172/JCI64859

Niesel, K., Schulz, M., Anthes, J., Alekseeva, T., Macas, J., Salamero-Boix, A., et al. (2021). The immune suppressive microenvironment affects efficacy of radioimmunotherapy in brain metastasis. *EMBO Mol. Med.* 13 (5), e13412. doi:10.15252/ emmm.202013412

Noda, M., Yamakawa, Y., Matsunaga, N., Naoe, S., Jodoi, T., Yamafuji, M., et al. (2012). IL-6 receptor is a possible target against growth of metastasized lung tumor cells in the brain. *Int. J. Mol. Sci.* 14 (1), 515–526. doi:10.3390/ijms14010515

Parker, K. R., Migliorini, D., Perkey, E., Yost, K. E., Bhaduri, A., Bagga, P., et al. (2020). Single-cell analyses identify brain mural cells expressing CD19 as potential off-tumor targets for CAR-T immunotherapies. *Cell* 183 (1), 126–142.e17. doi:10.1016/j.cell.2020. 08.022

Perelroizen, R., Philosof, B., Budick-Harmelin, N., Chernobylsky, T., Ron, A., Katzir, R., et al. (2022). Astrocyte immunometabolic regulation of the tumour microenvironment drives glioblastoma pathogenicity. *Brain* 145 (9), 3288–3307. doi:10.1093/brain/awac222

Priceman, S. J., Tilakawardane, D., Jeang, B., Aguilar, B., Murad, J. P., Park, A. K., et al. (2018). Regional delivery of chimeric antigen receptor-engineered T cells effectively targets HER2(+) breast cancer metastasis to the brain. *Clin. Cancer Res.* 24 (1), 95–105. doi:10.1158/1078-0432.CCR-17-2041

Priego, N., Zhu, L., Monteiro, C., Mulders, M., Wasilewski, D., Bindeman, W., et al. (2018). STAT3 labels a subpopulation of reactive astrocytes required for brain metastasis. *Nat. Med.* 24 (7), 1024–1035. doi:10.1038/s41591-018-0044-4

Qian, B. Z., Li, J., Zhang, H., Kitamura, T., Zhang, J., Campion, L. R., et al. (2011). CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. *Nature* 475 (7355), 222–225. doi:10.1038/nature10138

Ren, D., Zhu, X., Kong, R., Zhao, Z., Sheng, J., Wang, J., et al. (2018). Targeting brainadaptive cancer stem cells prohibits brain metastatic colonization of triple-negative breast cancer. *Cancer Res.* 78 (8), 2052–2064. doi:10.1158/0008-5472.CAN-17-2994

Rietkotter, E., Bleckmann, A., Bayerlova, M., Menck, K., Chuang, H. N., Wenske, B., et al. (2015). Anti-CSF-1 treatment is effective to prevent carcinoma invasion induced by monocyte-derived cells but scarcely by microglia. *Oncotarget* 6 (17), 15482–15493. doi:10.18632/oncotarget.3855

Samson, A., Scott, K. J., Taggart, D., West, E. J., Wilson, E., Nuovo, G. J., et al. (2018). Intravenous delivery of oncolytic reovirus to brain tumor patients immunologically primes for subsequent checkpoint blockade. *Sci. Transl. Med.* 10 (422), eaam7577. doi:10.1126/scitranslmed.aam7577

Sato, R., Nakano, T., Hosonaga, M., Sampetrean, O., Harigai, R., Sasaki, T., et al. (2017). RNA sequencing analysis reveals interactions between breast cancer or melanoma cells and the tissue microenvironment during brain metastasis. *Biomed. Res. Int.* 2017, 8032910. doi:10.1155/2017/8032910

Schaettler, M. O., Richters, M. M., Wang, A. Z., Skidmore, Z. L., Fisk, B., Miller, K. E., et al. (2022). Characterization of the genomic and immunologic diversity of malignant brain tumors through multisector analysis. *Cancer Discov.* 12 (1), 154–171. doi:10.1158/2159-8290.CD-21-0291

Schlam, I., Church, S. E., Hether, T. D., Chaldekas, K., Hudson, B. M., White, A. M., et al. (2021). The tumor immune microenvironment of primary and metastatic HER2positive breast cancers utilizing gene expression and spatial proteomic profiling. *J. Transl. Med.* 19 (1), 480. doi:10.1186/s12967-021-03113-9

Seike, T., Fujita, K., Yamakawa, Y., Kido, M. A., Takiguchi, S., Teramoto, N., et al. (2011). Interaction between lung cancer cells and astrocytes via specific inflammatory cytokines in the microenvironment of brain metastasis. *Clin. Exp. Metastasis* 28 (1), 13–25. doi:10.1007/s10585-010-9354-8

Sikpa, D., Whittingstall, L., Fouquet, J. P., Radulska, A., Tremblay, L., Lebel, R., et al. (2020). Cerebrovascular inflammation promotes the formation of brain metastases. *Int. J. Cancer* 147 (1), 244–255. doi:10.1002/ijc.32902

Sirkisoon, S. R., Carpenter, R. L., Rimkus, T., Doheny, D., Zhu, D., Aguayo, N. R., et al. (2020). TGL11 transcription factor mediates breast cancer brain metastasis via activating metastasis-initiating cancer stem cells and astrocytes in the tumor microenvironment. *Oncogene* 39 (1), 64–78. doi:10.1038/s41388-019-0959-3

Song, S. G., Kim, S., Koh, J., Yim, J., Han, B., Kim, Y. A., et al. (2021a). Comparative analysis of the tumor immune-microenvironment of primary and brain metastases of

non-small-cell lung cancer reveals organ-specific and EGFR mutation-dependent unique immune landscape. *Cancer Immunol. Immunother.* 70 (7), 2035–2048. doi:10.1007/s00262-020-02840-0

Song, Z., Yang, L., Zhou, Z., Li, P., Wang, W., Cheng, G., et al. (2021b). Genomic profiles and tumor immune microenvironment of primary lung carcinoma and brain oligo-metastasis. *Cell Death Dis.* 12 (1), 106. doi:10.1038/s41419-021-03410-7

Steele, L., Errington, F., Prestwich, R., Ilett, E., Harrington, K., Pandha, H., et al. (2011). Pro-inflammatory cytokine/chemokine production by reovirus treated melanoma cells is PKR/NF-κB mediated and supports innate and adaptive anti-tumour immune priming. *Mol. Cancer* 10, 20. doi:10.1186/1476-4598-10-20

Steindl, A., Alpar, D., Heller, G., Mair, M. J., Gatterbauer, B., Dieckmann, K., et al. (2021). Tumor mutational burden and immune infiltrates in renal cell carcinoma and matched brain metastases. *ESMO Open* 6 (2), 100057. doi:10.1016/j.esmoop.2021. 100057

Sun, H. F., Li, L. D., Lao, I. W., Li, X., Xu, B. J., Cao, Y. Q., et al. (2022). Single-cell RNA sequencing reveals cellular and molecular reprograming landscape of gliomas and lung cancer brain metastases. *Clin. Transl. Med.* 12 (11), e1101. doi:10.1002/ ctm2.1101

Taggart, D., Andreou, T., Scott, K. J., Williams, J., Rippaus, N., Brownlie, R. J., et al. (2018). Anti-PD-1/anti-CTLA-4 efficacy in melanoma brain metastases depends on extracranial disease and augmentation of CD8(+) T cell trafficking. *Proc. Natl. Acad. Sci. U. S. A.* 115 (7), E1540–E1549. doi:10.1073/pnas.1714089115

Tomaszewski, W., Sanchez-Perez, L., Gajewski, T. F., and Sampson, J. H. (2019). Brain tumor microenvironment and host state: Implications for immunotherapy. *Clin. Cancer Res.* 25 (14), 4202–4210. doi:10.1158/1078-0432.CCR-18-1627

Valiente, M., Ahluwalia, M. S., Boire, A., Brastianos, P. K., Goldberg, S. B., Lee, E. Q., et al. (2018). The evolving landscape of brain metastasis. *Trends Cancer* 4 (3), 176–196. doi:10.1016/j.trecan.2018.01.003

Vareslija, D., Priedigkeit, N., Fagan, A., Purcell, S., Cosgrove, N., O'Halloran, P. J., et al. (2019). Transcriptome characterization of matched primary breast and brain metastatic tumors to detect novel actionable targets. *J. Natl. Cancer Inst.* 111 (4), 388–398. doi:10.1093/jnci/djy110

Varn, F. S., Johnson, K. C., Martinek, J., Huse, J. T., Nasrallah, M. P., Wesseling, P., et al. (2022). Glioma progression is shaped by genetic evolution and microenvironment interactions. *Cell* 185 (12), 2184–2199.e16. doi:10.1016/j.cell.2022.04.038

Wang, H., Liu, P., Mi, J., Chen, J., and Liu, Y. (2020). A preliminary study of adoptive T-cell transfer therapy for patients with non-small-cell lung adenocarcinoma with brain metastasis: A case report of 3 patients. *Clin. Lung Cancer* 21 (4), e270–e273. doi:10.1016/ j.cllc.2020.02.015

Weiss, S. A., Zito, C., Tran, T., Heishima, K., Neumeister, V., McGuire, J., et al. (2021). Melanoma brain metastases have lower T-cell content and microvessel density compared to matched extracranial metastases. *J. Neurooncol* 152 (1), 15–25. doi:10. 1007/s11060-020-03619-0

Xiong, Z., Gharagozlou, S., Vengco, I., Chen, W., and Ohlfest, J. R. (2008). Effective CpG immunotherapy of breast carcinoma prevents but fails to eradicate established brain metastasis. *Clin. Cancer Res.* 14 (17), 5484–5493. doi:10.1158/1078-0432.CCR-07-4139

Xu, Z. H., Miao, Z. W., Jiang, Q. Z., Gan, D. X., Wei, X. G., Xue, X. Z., et al. (2019). Brain microvascular endothelial cell exosome-mediated \$100A16 up-regulation confers small-cell lung cancer cell survival in brain. *FASEB J.* 33 (2), 1742–1757. doi:10.1096/fj. 201800428R

Yeo, A. T., Rawal, S., Delcuze, B., Christofides, A., Atayde, A., Strauss, L., et al. (2022). Single-cell RNA sequencing reveals evolution of immune landscape during glioblastoma progression. *Nat. Immunol.* 23 (6), 971–984. doi:10.1038/s41590-022-01215-0

Zhang, L., Yao, J., Wei, Y., Zhou, Z., Li, P., Qu, J., et al. (2020). Blocking immunosuppressive neutrophils deters pY696-EZH2-driven brain metastases. *Sci. Transl. Med.* 12 (545), eaaz5387. doi:10.1126/scitranslmed.aaz5387

Zhang, L., Zhang, S., Yao, J., Lowery, F. J., Zhang, Q., Huang, W. C., et al. (2015). Microenvironment-induced PTEN loss by exosomal microRNA primes brain metastasis outgrowth. *Nature* 527 (7576), 100–104. doi:10.1038/nature15376

Zhang, T., Lip, H., He, C., Cai, P., Wang, Z., Henderson, J. T., et al. (2019). Multitargeted nanoparticles deliver synergistic drugs across the blood-brain barrier to brain metastases of triple negative breast cancer cells and tumorassociated macrophages. *Adv. Healthc. Mater* 8 (18), e1900543. doi:10.1002/ adhm.201900543

Zhao, P., Zhang, J., Wu, A., Zhang, M., Zhao, Y., Tang, Y., et al. (2021). Biomimetic codelivery overcomes osimertinib-resistant NSCLC and brain metastasis via macrophage-mediated innate immunity. *J. Control Release* 329, 1249–1261. doi:10. 1016/j.jconrel.2020.10.052

Glossary

BrM brain metastases; **BBB** blood-brain barrier; BTB blood-tumor barrier; OS overall survival; HER-2 human epidermal growth factor receptor 2; BRAF B-Raf kinase gene; MAPK mitogen-activated protein kinase; CTLA-4 cytotoxic T-lymphocyte-associated protein 4; PD-1 programmed cell death protein 1; PD-L1 programmed death-ligand 1; ECM extracellular matrix; TNFRSF tumor necrosis factor receptor superfamily; PDCDILG2 programmed cell death 1 ligand 2; IDO indoleamine 2,3-dioxygenase; ICOS inducible T Cell costimulator; CD cluster of differentiation; TARP T cell receptor gamma alternate reading frame protein; PTPRC protein tyrosine phosphatase receptor type c; PTPN7 protein tyrosine phosphatase non-receptor type 7; IL10RA interleukin 10 receptor subunit alpha; GZMK granzyme k; CCR C-C motif chemokine receptor; CCL C-C motif chemokine ligand; BCL2L1 B-cell lymphoma-2-like protein 1; TWIST1 twist family BHLH transcription factor 1; GSTA5 glutathione S-transferase alpha 5; TAMs tumor-associated macrophages; IL6R interleukin 6 receptor; INHBA inhibin subunit beta a; AREG amphiregulin; IDH isocitrate dehydrogenase; CNS central nervous system; NSCLC non-small cell lung cancer; TCR T cell receptor; STAT signal transducer and activator of transcription; Tim-3 T cell immunoglobulin and mucin-domain containing-3; **OXPHOS** oxidative phosphorylation; KEGG Kyoto Encyclopedia of Genes and Genomes; E2F3 E2F transcription factor 3; **RB** retinoblastoma protein; KRAS Kirsten rat sarcoma virus; ALK anaplastic lymphoma kinase; SRC src proto-oncogene; mTOR mechanistic target of rapamycin kinase;

CXCR4 C-X-C motif chemokine receptor 4; **PLLP** plasmolipin; TNFSF4 tumor necrosis factor superfamily member 4; VCAM-1 vascular cell adhesion molecule 1; SLC8A2 solute carrier family 8 member a2; SLC7A11 solute carrier family 8 member a11; TIL tumor infiltrating lymphocytes; S100A4 s100 calcium binding protein a4; ANXA2 annexin a2; TGFB1 transforming growth factor beta 1; SERPINB1 serpin family B member 1; **ITGs** integrins; PCDH7 protocadherin 7; Cx43 connexin 43; cGAMP cyclic guanosine monophosphate-adenosine monophosphate; IFN interferon; TNF-a tumor necrosis factor alpha; **NF-κB** nuclear factor kappa b; **PLC**β phosphodiesterase beta; PIP2 phosphatidylinositol 4,5-bisphosphate; IP3 inositol 1,4,5-trisphosphate; DAG 1,2-diacylglycerol; CaMK2 calmodulin-dependent protein kinase II; ETAR endothelin receptor a; ETBR endothelin receptor b; MIF migration inhibitory factor; IL-8 interleukin-8; PAI-1 plasminogen activator inhibitor-1; MMP matrix metallopeptidase; TNBC triple-negative breast cancer; BDNF brain derived neurotrophic factor; EGF epidermal growth factor; TrkB tropomyosin receptor kinase b; PTEN phosphatase and tensin homolog; SH2 src homology 2; **IBA1** ionized calcium binding adaptor molecule 1; PI3K phosphoinositide 3-kinase; MDM marrow-derived-macrophages; CSF-1 colony stimulator factor-1; IncRNAs long non-coding RNAs; JAK2 Janus kinase 2; CpG-ODN CpG oligodeoxynucleotides; TLR toll-like receptor; iRGD-DMTPLN iRGD-terpolymer-lipid hybrid nanoparticle with coloaded doxorubicin and mitomycin c;

IM leptomeningeal metastases:	CEA carcino
EM repromeningen metastases,	CLA carcino
LCN2 lipocalin-2;	CK19 cytoke
SCL22A17 solute carrier family 22 member 17;	PSMA prosta
SRS stereotactic radiosurgery;	COX-2 cyclo
OVs oncolytic viruses;	MPIO micro
oHSV oncolytic herpes simplex virus;	T-DM1 trast
MSCs mesenchymal stem cells;	S100A16 s10
ACT therapy adoptive cell transfer therapy;	PHB-1 prohi
CAR T cell therapy chimeric antigen receptor-engineered T cell	C3 complem
therapy;	EZH2 enhan
DCs denditic cells;	G-CSF granu

EA carcinoembryonic antigen; **EA** carcinoembryonic antigen; **EX19** cytokeratin 19; **SMA** prostate specific membrane antigen; **COX-2** cyclooxygenase-2; **IPIO** microparticles of iron oxide; **CDM1** trastuzumab emtansine; **100A16** s100 calcium binding protein a16; **HB-1** prohibitin-1; **C3** complement component 3; **ZH2** enhancer of zeste homolog 2; **CSF** granulocyte colony-stimulating factor