Check for updates

#### OPEN ACCESS

EDITED BY Alina Arulsamy, Monash University, Malaysia

REVIEWED BY Ching Soong Khoo, National University of Malaysia, Malaysia Si Lei Fong, University of Malaya, Malaysia

\*CORRESPONDENCE Yuxin Zhang [baby20050218@163.com](mailto:baby20050218@163.com)

RECEIVED 13 August 2024 ACCEPTED 23 October 2024 PUBLISHED 14 November 2024

#### CITATION

He X and Zhang Y (2024) Changes in gut flora in patients with epilepsy: a systematic review and meta-analysis. *Front. Microbiol.* 15:1480022. [doi: 10.3389/fmicb.2024.1480022](https://doi.org/10.3389/fmicb.2024.1480022)

#### COPYPICHT

© 2024 He and Zhang. This is an open-access article distributed under the terms of the [Creative Commons Attribution](http://creativecommons.org/licenses/by/4.0/)  [License \(CC BY\)](http://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# [Changes in gut flora in patients](https://www.frontiersin.org/articles/10.3389/fmicb.2024.1480022/full)  [with epilepsy: a systematic review](https://www.frontiersin.org/articles/10.3389/fmicb.2024.1480022/full)  [and meta-analysis](https://www.frontiersin.org/articles/10.3389/fmicb.2024.1480022/full)

## Xingyan He<sup>1,2</sup> and Yuxin Zhang<sup>1\*</sup>

1 Department of Pediatrics, Guangdong Provincial People's Hospital (Guangdong Academy of Medical Sciences), Southern Medical University, Guangzhou, Guangdong, China, 2Shantou University Medical College, Shantou, Guangdong, China

**Background:** Epilepsy is a prevalent chronic neurological disorder that is strongly associated with a wide range of psychological, cognitive and social problems. It affects a significant proportion of the global population and has a number of complex etiologies. A growing body of research indicates that there is a strong association between epilepsy and the gut microbiota. Indeed, a substantial body of research has investigated the potential role of epilepsy in relation to the gut microbiota, examining alterations in the abundance, diversity, and relative abundance of the gut microbiota in patients with epilepsy.

Methods: This study was conducted in accordance with the PRISMA guidelines and included multiple studies that met specific criteria. A keyword search was conducted in the following databases: PubMed, Embase, and Web of Science. The data extraction and quality assessment were conducted by two independent researchers. A systematic review and meta-analysis of the relationship between patients with epilepsy and gut flora was conducted using the R 4.3.4 software.

Results: The results of the analyses indicated that the intestinal flora of patients with epilepsy did not differ significantly in alpha diversity compared to healthy controls. However, the relative abundance of specific flora, such as *Verrucomicrobia* and *Ackermannia* was significantly increased in patients, whereas Lactobacillus was significantly decreased.

Conclusion: The relationship between epilepsy and gut flora is reciprocal. The present meta-analysis demonstrated that there were no statistically significant alterations in the overall characteristics of the intestinal flora of the patients. However, significant changes were observed in the relative abundance of certain phyla and genera. Consequently, it is hypothesized that epilepsy can cause changes in the relative abundance of specific flora in patients. Furthermore, in conjunction with previous studies, it is believed that changes in intestinal flora can also have an effect on seizures. For example, *Lactobacillus* may be a beneficial genus that potentially reduces seizures. Conversely, the effect of Akkermansia is two-sided.

#### KEYWORDS

epilepsy, intestinal flora, meta-analysis, gut-brain axis, gut microbiome

## 1 Background

Epilepsy is a chronic and devastating neurological disorder characterized by seizures (Thijs et al., 2019). Epilepsy is often associated with psychological, cognitive, and social consequences ([Fisher et al., 2014](#page-16-1)). Approximately 50 million people and 0.5–1% of children worldwide suffer from epilepsy [\(Aaberg et al., 2017](#page-15-0)). The incidence of epilepsy is highest in infancy, with approximately 0.67% of children diagnosed with epilepsy in the first 10years of life [\(Elliott](#page-16-2)  [et al., 2019](#page-16-2)). The causes of epilepsy can be broadly categorized into six groups: genetic, structural, metabolic, immunologic, immune, infectious, and unknown factors [\(Scheffer et al., 2017](#page-16-3); [Fisher et al.,](#page-16-4)  [2017;](#page-16-4) [Fisher et al., 2017\)](#page-16-5).

Most people with epilepsy live in areas with poor sanitation ([World](#page-16-6)  [Health Organization, 2024\)](#page-16-6), and environmental factors have been shown to have a significant impact on the composition of the gut microbiota [\(Gacesa et al., 2022\)](#page-16-7). Prior research has demonstrated that the utilization of antibiotics can potentially exacerbate seizures [\(Sutter et al., 2015\)](#page-16-8). Furthermore, a high-fat ketogenic diet has been demonstrated to be an efficacious treatment for medically refractory epilepsy [\(Kwan and Brodie,](#page-16-9)  [2000](#page-16-9); [Newman, 2000\)](#page-16-10). Additionally, studies have indicated that the gut flora is a mediator of the antiepileptic effects of the ketogenic diet. In conclusion, the role of gut flora in the development or remission of epilepsy is significant. The gut-brain axis has been demonstrated to perceive and respond to alterations in the dynamic ecosystem by translating chemical signals from the environment and gut microbes into neural signals, thereby indicating a pivotal function for the gut microbiota in epileptogenesis ([Tremlett et al., 2017](#page-16-11)).

To date, a number of systematic evaluations have demonstrated an association between epilepsy and the gut microbiota. This systematic review and meta-analysis aimed to examine the relationship between epilepsy and gut flora, with the objective of synthesizing and critically evaluating the available evidence supporting the interaction between gut microbiota and epilepsy, and elucidating potential interventions.

## 2 Methods

## 2.1 Search strategy

This systematic review and meta-analysis is conducted in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines ([Page et al., 2021](#page-16-12)).

The following keywords were used for the analysis: epilepsy and intestinal flora. The keywords are presented in the following format: Medical Subject Headings + free word. A comprehensive search was conducted in PubMed, Embase, and Web of Science using the keywords "intestinal flora" and "epilepsy" to identify studies involving valuable epilepsy and intestinal flora analyses from database creation to 9 June 2024 (the search strategy refers to [Table 1\)](#page-2-0). The search language was English [\(Figure 1](#page-7-0)).

## 2.2 Eligibility criteria

The inclusion criteria were as follows: (1) Control for the type of study design. (2) The study was conducted on patients with primary epilepsy. (3) The outcome metrics were gut flora abundance or gut flora richness or diversity metrics. (4) The source was published primary literature.

The exclusion criteria were as follows: (1) Animal subjects. (2) Non-epileptic subjects. (3) Secondary epilepsy such as post-traumatic epilepsy, post-stroke epilepsy, and postencephalitis epilepsy. (4) Incomplete data that prevented the extraction of raw data for statistical literature. (5) Conference papers, dissertations, and review articles. (6) Non-English papers.

## 2.3 Data extraction

The data were extracted independently by both authors from the original text or graphs. (1) General characteristics: first author, year of publication, population size, age, diagnosis. (2) Outcome metrics studied: alpha diversity (observed species index, Chao1 index, Shannon index, Simpson index, ACE index), bacterial phylum, bacterial genus.

The data were extracted directly from the original article, or the raw data were requested by email from the corresponding author of the study. In the event of a lack of response from the authors, the GetData chart digitizer was employed to procure pertinent and precise raw data that could not be obtained from the corresponding author ([Liu et al., 2017](#page-16-13)).

## 2.4 Quality assessment

The potential for bias in the included studies was evaluated using a scale developed by the Agency for Healthcare Research and Quality (AHRQ) for the assessment of cross-sectional studies. Two independent authors evaluated the risk of bias based on the following criteria: (1) Is the source of data specified? (2) Whether the inclusion and exclusion criteria for the exposed and non-exposed groups were clearly defined. (3) Whether the time period for identifying patients was specified. (4) Whether the study population was consecutive. (5) Whether the assessor of subjectively-indicated patient outcomes was isolated from other objective indicators of the patient. (6) Describing any assessments made for quality assurance purposes. (7) Providing explanations for the exclusion of some patients. (8) Describing how confounders were controlled for. Each criterion could be classified as low, unclear, or high risk of bias. Two investigators conducted an independent assessment of the risk of bias and cross-validation. In the event of a discrepancy, a third party was consulted for a resolution after a discussion. The generation of random numbers using tables or computer software was deemed a low-risk method for ensuring the reliability of the random sequence. Methods deemed unreliable, such as those involving the selection of numbers artificially, were considered to pose a high risk, while those with unknown generation methods were considered to be of unclear risk. The potential for the assessor of a patient's subjectively-derived indicators to be isolated from other objective indicators of the patient is contingent upon the rigor of the employed blinding methodology and its correct implementation. A potentially compromised blinding is defined as a failure to report all pre-specified primary outcome indicators during the study period, which is considered high risk; otherwise, it is considered low risk. The failure to report all pre-specified key outcome indicators is regarded as a high-risk factor, whereas the outcome indicators included in the fully published study plan are considered low risk.

<span id="page-2-0"></span>



[Frontiers in Microbiology](https://www.frontiersin.org/journals/microbiology)

*(Continued)*

[10.3389/fmicb.2024.1480022](https://doi.org/10.3389/fmicb.2024.1480022)

10.3389/fmicb.2024.1480022



*(Continued)*







<span id="page-7-0"></span>

## 2.5 Data analysis

All extracted data were converted to the same unit prior to calculation. Continuous variables were expressed as mean±standard deviation (m=median, a=minimum, b=maximum) in accordance with the formula detailed in [Supplementary Image 3](#page-15-1) [\(Hozo et al.,](#page-16-14)  [2005\)](#page-16-14). The data were analyzed using the R 4.3.3 software. The results were expressed as a Standardized Mean Difference (SMD) with a 95% confidence interval (CI). The presence of heterogeneity was evaluated through the utilization of the chi-square test and the  $I^2$  statistic. A  $p$ -value of less than 0.1 and an  $I^2$  value exceeding 50% were indicative of a considerable probability of heterogeneity within the data. Given the anticipated presence of heterogeneity, a random-effects model was

employed. Furthermore, subgroup analyses were conducted when deemed necessary to facilitate a more comprehensive interpretation of the results. A *p*-value of less than 0.05 was considered statistically significant.

## 3 Results

## 3.1 Search results

A total of 1,254 studies were retrieved from Web of Science, PubMed, and Embase on 9 June 2024, following a comprehensive search. Two researchers employed the EndNote software to screen the titles and abstracts of the references, resulting in the removal of 363 duplicates. A total of 845 references were excluded from further consideration as they did not meet the pre-established inclusion criteria. Additionally, 46 potential references were included. Following a full-text review by two researchers, 14 studies were included in the meta-analysis ([Table 2\)](#page-9-0).

## 3.2 Characteristics of the selected studies

The experimental group was diagnosed with epilepsy in 14 studies, eight of which involved children. In five studies, the mean age of the experimental group was less than 18years, while in seven studies, it was greater than 18years. In two studies, the mean age could not be calculated due to the lack of specification regarding the age of the control group.

Six studies did not specify the type of epilepsy. Two studies focused on refractory epilepsy in children, two studies investigated drug-resistant and drug-sensitive epilepsy, two studies explored West's syndrome, one study examined focal epilepsy, and the cause of epilepsy in one study was unclear.

The six principal outcome indicators are alpha diversity (Chao1 index, Shannon's index, Simpson's index, and Ace's index), phylum, and genus. These are presented in [Table 2](#page-9-0).

## 3.3 Risk of bias assessment

All studies were deemed to have a low risk of bias in terms of the source of information, inclusion and exclusion criteria, assessment of quality valuation, explanation for exclusion of each individual patient, and measures to control for confounders. Thirteen studies were identified as being at high risk of bias with regard to the question of whether the assessor of the patient's subjectivized indicator was isolated from the patient's other objective indicators. In addition, some of the studies were at high risk of bias with respect to whether the subjects were consecutive or not, and whether a time period for identifying the patients was provided. The risk of bias was unclear in terms of whether the subjects were consecutive or not, quality assessment, and control of confounders. In some cases, the risk of bias was unclear.

The overall assessment of risk of bias was low in 11 trials and uncertain in 2 trials ([Figure 2\)](#page-10-0).

## 3.4 Differential assessment of flora outcomes in epileptic and non-epileptic patients

### 3.4.1 Alpha diversity

In this study, a range of alpha diversity indices were employed to evaluate microbial diversity within a given group. These included estimates of richness (such as the observed species index and the Chao1 index) and indices that combine richness and evenness (such as the Shannon index, the Simpson index and the ACE index) ([Figure 3\)](#page-11-0).

A total of 12 controlled trials provided data on the Chao1 index of richness. Of these, five trials revealed a significant difference between the epilepsy group and the control group with regard to the Chao1 index. The meta-analysis of this study revealed no significant difference in the SMD of the species index Chao1 (SMD=0.30,  $p=0.47$ , 95% CI = -0.52 to 1.11, I<sup>2</sup> = 93%). A subgroup analysis according to age revealed no significant difference between minors (aged <18years) and adults. The former exhibited a standardized mean difference (SMD) of −0.18 ( $p = 0.73$ , 95% CI = −1.18 to 0.83,  $I^2 = 85\%$ ), while the latter displayed a SMD of 0.63 ( $p = 0.30$ , 95% CI= $-56$  to 1.86, I<sup>2</sup> = 95) [\(Supplementary Images 1, 2\)](#page-15-1).

A total of 10 studies provided data on Shannon's index, and in seven of these studies, a significant difference was observed between the epilepsy group and the control group. In all five studies showing a significant reduction in Shannon's index in the epilepsy group, compared to the control group, the opposite was true in two studies. The results of the meta-analysis demonstrated no statistically significant difference in the SMD of the Shannon index (SMD=−0.92,  $p=0.14$ , 95% CI = -2.14 to 0.30, I<sup>2</sup> = 94%), nor in the subgroup analyses (minors: SMD=−0.11, *p*=0.85, 95% CI=−1.24 to 1.03, I<sup>2</sup> = 88%). The SMD was −1.74 ( $p$  = 0.10), with a 95% CI of −3.80 to 0.32 and an  $I^2$  value of 96%.

Seven studies provided data on Simpson's index. Five studies demonstrated a statistically significant difference between patients with epilepsy and a control group. Three of these studies showed a significant increase in Simpson's index in the epilepsy group compared to the control group, while the other two studies yielded opposite results. The meta-analysis of the present study revealed no statistically significant difference in Simpson's index between patients with epilepsy and the healthy population [standardized mean difference (SMD) 0.05, *p*=0.96, 95% confidence interval (CI) −2.09 to 2.00,  $I^2 = 96.5\%$ ]. The ACE index was provided as an indicator of outcome by only three studies. The meta-analysis of these studies showed a non-significant difference (SMD=2.45, *p*=0.06, 95% CI=−0.11 to  $5.00, I^2 = 96.4\%$ ).

#### 3.4.2 Phylum of bacteria

The present study identified five distinct clades ([Figure 4\)](#page-12-0). The identified clades were *Bacteroidetes*, *Actinobacteria*, *Verrucomicrobia*, *Firmicutes*, and *Proteobacteria*. Additionally, the *Firmicutes/ Bacteroidetes* (F/B) ratio was included in the outcome index. The results of the meta-analysis indicated that there was a statistically significant association between *Verrucomicrobia* and the outcome variable (SMD=0.35, 95% CI 0.08 to 0.62,  $p=0.01$ ,  $I^2=0$ %). No statistically significant differences were observed in the levels or F/B ratios for the remaining clades. Only data on *Acidobacteria* levels were available from the study by [Liu et al. \(2023\)](#page-16-15), which demonstrated a significant difference between the upper epilepsy group and the control population (SMD=0.35,  $p=0.003$ , 95%  $CI = -2.68$  to  $-0.54$ ).

#### 3.4.3 Genera of bacteria

A total of 14 genera have been identified. The identified genera are as follows: *Bifidobacterium*, *Faecalibacterium*, *Akkermansia*, *Enterococcus*, *Collinsella*, *Streptococcus*, *Escherichia/Shigella*, *Veillonella*, *Clostridium sensu strict*, *Bacteroides*, *Klebsiella, Lactobacillus*, *Enterobacter*. Significant differences were observed in



<span id="page-9-0"></span>–, Not mentioned in relevant literature or data not available.

<span id="page-10-0"></span>

the abundance of *Akkermansia* (SMD=0.31, *p*=0.02, 95% CI=−0.04 to 0.58, I2=0%) and *Lactobacillus* (SMD=−0.55, *p*=0.02, 95% CI= $-1.10$  to 0.09, I<sup>2</sup>=0%) between epileptic patients and controls ([Figure 5\)](#page-13-0).

Figure 6 illustrates the phylogenetic characteristics of the abundance of taxonomic differences between epileptic and normal populations at the phylum, order and genus levels, with colors indicating different bacterial variants. The colors red, blue and orange indicate increased abundance, decreased abundance and inconsistent variation, respectively.

## 4 Discussion

In order to identify changes in the diversity, abundance, and homogeneity of the gut microbiota after a diagnosis of epilepsy, as well as possible characteristics of bacteria that reduce seizures, three of the most commonly used databases were searched.

## 4.1 About alpha-diversity

The present study encompasses 14 high-quality studies [\(Figure 7\)](#page-14-0), encompassing alpha diversity and relative abundance at the phylum and genus levels. Alpha diversity is associated with the number of species in a local, homogeneous habitat, also referred to as intrahabitat diversity. In the context of the human gut flora, higher alpha diversity is typically indicative of a more complex and diverse microbial community, exhibiting greater metabolic versatility and stability, and a healthier metabolic profile, particularly with regard to drug metabolism [\(Li et al., 2022](#page-16-29); [Knight et al., 2018\)](#page-16-30). It is also a marker of intestinal health. The Chao1 index, the Shannon index, the Simpson index, and the ACE index are frequently employed as measures of alpha diversity. The Chao1 index is of particular significance in that it is capable of deriving a theoretical abundance from the number of species observed, which is closer to the true abundance. Furthermore, it is particularly sensitive to rare species. The Shannon index is of significance in that it quantifies the uncertainty in the diversity of the community, that is to say, the difficulty of predicting the species identity of an individual selected at random. The Simpson index is of significance in that it reflects the degree of dominance and the even

distribution of species diversity in the community ([Supplementary Presentation 1](#page-15-1)). In a previous study of gut flora in epileptic patients, [Zhou et al. \(2022\)](#page-17-1) demonstrated that the alpha diversity of the epileptic group differed significantly from that of the healthy control group, with a notable decrease in the Shannon index and an increase in the Simpson index. Similarly, [Xie et al. \(2017\)](#page-16-31) reported that the alpha diversity of gut flora in infants with refractory epilepsy was reduced in comparison to healthy subjects. Additionally, [Xu et al. \(2021\)](#page-16-32) and [Gong et al. \(2021\)](#page-16-33) demonstrated that there were no statistically significant differences between the refractory epilepsy group and the refractory epilepsy group after ketogenic diet treatment compared to healthy controls. However, the study did indicate a potential trend toward a decrease. Another study by the same author revealed that the alpha diversity index of children in the epilepsy group was significantly lower than that of the family control group ([Gong et al., 2020](#page-16-34)). The results were inconclusive due to the heterogeneity of the samples, the diversity of the experimental approaches, the variability in the etiology of epilepsy, the differing sensitivities to antiepileptic drugs, and the age range of the participants. It is therefore recommended that further studies be conducted using more specialized experiments with larger sample sizes.

## 4.2 About levels of genus and phylum

This study examines characteristic gut microbiota taxa by selecting flora from multiple studies with sufficient data for metaanalysis. The findings of different studies have yielded disparate conclusions, with some studies even reaching completely opposite conclusions. The two most abundant phyla are *Anabaena* and *Firmicutes* ([Zhang et al., 2023](#page-17-2)). A significant decrease in Firmicutes was observed in a study by [Cui et al. \(2022\)](#page-16-35). This group is complex, comprising intricate sub-phylum, orders, and genera under the genus *Firmicutes*. Additionally, relatively few studies have been conducted on this group. The *Firmicutes/Bacteroidetes* (F/B) ratio is regarded as a key indicator for depicting intestinal disorders [\(Mohamed Qadir and](#page-16-36)  [Assafi, 2021\)](#page-16-36). Prior research has revealed an elevated F/B ratio in the intestinal microbiota of individuals diagnosed with epilepsy ([Arulsamy et al., 2020](#page-15-2)), indicating a potential correlation between this alteration in microbiota structure and the production of short-chain fatty acids, which may contribute to the development of severe

<span id="page-11-0"></span>

inflammation and complex infections [\(Bazzocchi et al., 2021](#page-15-3); [Rodenhouse et al., 2022](#page-16-37)). However, in the meta-analysis of the present study, the F/B ratio was not significantly different in the epileptic and control groups, which may be related to the small sample size. In this study, there was no significant difference between epileptic patients and healthy controls in the phylum Bacteroidetes, but *Lactobacillus* under *Bacteroidetes* was significantly reduced in the group of epileptic patients, and *Lactobacillus* is able to regulate the balance of the intestinal microbial community, promote the digestion of food and absorption of nutrients, reduce the cholesterol level of the body, and alleviate the intestinal inflammation and infection,. It is also implied that there exists the possibility of decreased intestinal anti-inflammatory capacity in epilepsy patients ([Schloissnig et al.,](#page-16-38)  [2013\)](#page-16-38). The results of the present study showed a significant increase in *Verrucomicrobia* and subordinate *Akkermansia* in the intestinal microbiota of epileptic patients compared to healthy controls. The abundance of *Verrucomicrobia* is closely related to intestinal health, contributing to glucose homeostasis in the human gut; and also possesses anti-inflammatory properties, which may further aid in intestinal health, as demonstrated by studies showing that a positive correlation between *Verrucomicrobia* and the foxp3 gene, a gene that expresses anti-inflammatory and immunity properties in humans ([Yan et al., 2023\)](#page-17-3). *Akkermansia*, which is under the phylum of *Verrucomicrobia*, is present in large quantities under normal

<span id="page-12-0"></span>

conditions and is considered to be used as a probiotic [\(Zhai et al.,](#page-17-4)  [2019\)](#page-17-4). The relative abundance of *Akkermansia* is reduced in glucose metabolism, obesity and inflammatory bowel disease [\(Cani and de](#page-16-39)  [Vos, 2017](#page-16-39); [Rajilić-Stojanović et al., 2013\)](#page-16-40), and studies have shown ([Zhai et al., 2019](#page-17-4); [Derrien et al., 2017](#page-16-41)) that it promotes intestinal barrier integrity, increases mucus thickness, and is conducive to metabolic and immune responses. The precise mechanism of interaction between *Akkermansia* and epilepsy remains unclear. [Olson](#page-16-42)  [et al. \(2018\)](#page-16-42) showed that in a mouse model of refractory epilepsy, *Akkermansia* mediated a ketogenic diet-induced protective effect

<span id="page-13-0"></span>

<span id="page-13-1"></span>

<span id="page-14-0"></span>

against 6-hz-induced seizures. However, the results of a Mendelian randomized study on epilepsy risk and intestinal flora [\(Zeng et al.,](#page-17-5)  [2023\)](#page-17-5) showed that *Akkermansia* was able to increase the risk of epilepsy. In conclusion, *Akkermansia* has both a protective effect and a pathogenic risk, and therefore its study needs to be further deepened to clarify its specific mechanism of action.

## 4.3 Interaction of the gut-brain axis

The experimental results included in this study pertain to the changes in gut microbiology that occur subsequent to the diagnosis of epilepsy. However, the specific causal relationship between seizure and gut flora remains unclear. It has been proposed that the gut microbiome may play a role in the etiology of epilepsy [\(Gong](#page-16-34)  [et al., 2020](#page-16-34)). Medel-Matus et al. demonstrated that altered gut microbiome can trigger seizures by transplanting feces from epileptic mice into non-epileptic mice ([Medel-Matus et al., 2018](#page-16-43)). Lum et al. explored the potential mechanisms of the gut microbiome in the treatment of epilepsy by investigating the effects of a ketogenic diet on the gut microbiome in epileptic patients [\(Lum](#page-16-44)  [et al., 2020](#page-16-44)). Animal and human studies have demonstrated that *γ*-aminobutyric acid (GABA) plays a pivotal role in the alteration of intestinal flora in relation to seizures in temporal lobe epilepsy ([Goldberg and Coulter, 2013;](#page-16-45) [Avoli et al., 2016](#page-15-4)). Additionally, these studies have indicated that the intestinal flora may alter neurotransmitter content to facilitate brain-gut interactions via the vagus nerve. Lactobacillus has been demonstrated to modulate mood and central GABA receptor levels via sympathetic nerves ([Bravo et al., 2011\)](#page-16-46). It is possible that intestinal flora and their products modulate sympathetic activity by influencing the secretion of enteroendocrine cells [\(Bonaz et al., 2018\)](#page-15-5). Furthermore, epigenetic modifications of genes have been identified as a contributing factor in the pathogenesis and anti-epileptic mechanisms of epilepsy [\(Younus and Reddy, 2017\)](#page-17-6). This process is

regulated by metabolites associated with gut flora [\(Reddy et al.,](#page-16-47)  [2018](#page-16-47)). Abnormalities in immune regulation represent a significant contributing factor to seizure. Additionally, immune disorders have the potential to induce alterations in the gut commensal flora (Weis [and Round, 2021](#page-16-48)), suggesting a potential convergence between seizure and gut disorders as a consequence of abnormalities in immune regulation.

The current research on gut flora is primarily concerned with the pathogenesis of various conditions, with treatments for several diagnoses of epilepsy including ketogenic diets, fecal transplants, antibiotic therapy, probiotic supplementation, prebiotics, and interventions targeting antiepileptic drug-flora interactions [\(Liu et al.,](#page-16-15)  [2023;](#page-16-15) [Dahlin and Prast-Nielsen, 2019](#page-16-49); [Ułamek-Kozioł et al., 2019;](#page-16-50) [Mejía-Granados et al., 2021;](#page-16-51) [Amlerova et al., 2021](#page-15-6)). Over the past decade, research in the field of gut flora has flourished, resulting in significant advancements. However, it is crucial to acknowledge the intricate composition of the intestinal flora and the intestinal immune environment, as well as their distinctive characteristics across different individuals. This indicates the necessity for caution when extrapolating findings from experimental animals to humans. Firstly, although animal models (e.g., mice) are a valuable resource in research, they differ from humans in terms of immune response, gut structure, etc., which may affect the translation from animal models to humans ([Schloissnig et al., 2013](#page-16-38); [Kaiko et al., 2016](#page-16-52); [Falony et al., 2016](#page-16-53)). Secondly, the human gut microbiome comprises over 1,000 microorganisms, with intricate interactions between these microorganisms and their host ([Mabwi et al., 2020\)](#page-16-54). This complexity gives rise to challenges in the precise regulation and application of the microbiome. Moreover, it is essential to consider the inter-individual variability observed across different ethnic, gender, and age groups. It is therefore evident that a significant future development in the exploration and improvement of epilepsy treatments will be the investigation of the changes in gut flora that can have a considerable impact on complex populations with significant heterogeneity. This will facilitate a more precise understanding of the interactions between

gut flora and health and disease, thereby providing a scientific basis for the development of personalized therapeutic regimens. The last, when translating findings from animal models to human therapeutic applications, it is crucial to ensure the safety and efficacy of the treatment. This necessitates the undertaking of extensive clinical studies and validation.

## 5 Limitation

It should be noted that this meta-analysis is not without limitations. Firstly, most studies include experimental analyses regarding beta diversity. Since principal component analysis (PCA) is currently the main method of beta analysis, which selects the eigenvectors corresponding to the top few largest eigenvalues based on the magnitude of the eigenvalues, and these eigenvectors are known as the principal components, however, the eigenvalues of the top few eigenvalues of each study were mixed, so meta-analyses were not performed in this study at the level of beta analysis. Secondly, the study included different populations, including geographic location, gender, weight and age. As a result, it was not possible to accurately assess the effect of these confounding factors on the outcome indicators. Thirdly, the length of time the sample was collected was not known. As a consequence, it was not possible to assess the effect of this factor on the outcome indicators. The initial intention of this study was to subgroup all indicators according to age in order to more effectively illustrate the results. However, due to the heterogeneity of the available data, only subgroup analyses were performed for some of the outcomes.

## 6 Conclusion

The meta-analysis of this study demonstrated that the alteration in alpha-diversity within the intestinal flora of patients with epilepsy was not statistically significant when compared to controls. However, there was a notable increase in *Verrucomicrobia* and its subordinate *Akkermansia*. Furthermore, a notable reduction in Lactobacillus was observed. It is postulated that epilepsy exerts a relatively minor impact on the abundance, homogeneity, and diversity of the intestinal microbiota in patients, whereas a more pronounced influence is exerted on the alteration of the relative abundance of the flora. Furthermore, the collective findings of previous studies indicate that *Lactobacillus* may confer beneficial effects. In contrast, *Akkermansia* are understood to play a complex and multifaceted role, with characteristics that may be two-faced. Given the considerable heterogeneity and restricted sample size, it is imperative to conduct more rigorous methodological studies.

## References

<span id="page-15-0"></span>Aaberg, K. M., Gunnes, N., Bakken, I. J., Lund Søraas, C., Berntsen, A., Magnus, P., et al. (2017). Incidence and prevalence of childhood epilepsy: a Nationwide cohort study. *Pediatrics* 139:e20163908. doi: [10.1542/peds.2016-3908](https://doi.org/10.1542/peds.2016-3908)

<span id="page-15-6"></span>Amlerova, J., Šroubek, J., Angelucci, F., and Hort, J. (2021). Evidences for a role of gut microbiota in pathogenesis and management of epilepsy. *Int. J. Mol. Sci.* 22:5576. doi: [10.3390/ijms22115576](https://doi.org/10.3390/ijms22115576)

<span id="page-15-2"></span>Arulsamy, A., Tan, Q. Y., Balasubramaniam, V., O'Brien, T. J., and Shaikh, M. F. (2020). Gut microbiota and epilepsy: a systematic review on their relationship and possible therapeutics. *ACS Chem. Neurosci.* 11, 3488–3498. doi: [10.1021/acschemneuro.0c00431](https://doi.org/10.1021/acschemneuro.0c00431)

## Data availability statement

The original contributions presented in the study are included in the article/[Supplementary material](#page-15-1), further inquiries can be directed to the corresponding author.

## Author contributions

XH: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. YZ: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing.

## Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## <span id="page-15-1"></span>Supplementary material

The Supplementary material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/fmicb.2024.1480022/](https://www.frontiersin.org/articles/10.3389/fmicb.2024.1480022/full#supplementary-material) [full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fmicb.2024.1480022/full#supplementary-material)

<span id="page-15-4"></span>Avoli, M., de Curtis, M., Gnatkovsky, V., Gotman, J., Köhling, R., Lévesque, M., et al. (2016). Specific imbalance of excitatory/inhibitory signaling establishes seizure onset pattern in temporal lobe epilepsy. *J. Neurophysiol.* 115, 3229–3237. doi: [10.1152/jn.01128.2015](https://doi.org/10.1152/jn.01128.2015)

<span id="page-15-3"></span>Bazzocchi, G., Turroni, S., Bulzamini, M. C., D'Amico, F., Bava, A., Castiglioni, M., et al. (2021). Changes in gut microbiota in the acute phase after spinal cord injury correlate with severity of the lesion. *Sci. Rep.* 11:12743. Published 2021 Jun 17. doi: [10.1038/s41598-021-92027-z](https://doi.org/10.1038/s41598-021-92027-z)

<span id="page-15-5"></span>Bonaz, B., Bazin, T., and Pellissier, S. (2018). The Vagus nerve at the Interface of the microbiota-gut-brain Axis. *Front. Neurosci.* 12:49. doi: [10.3389/fnins.2018.00049](https://doi.org/10.3389/fnins.2018.00049)

<span id="page-16-46"></span><span id="page-16-26"></span><span id="page-16-17"></span>Bravo, J. A., Forsythe, P., Chew, M. V., Escaravage, E., Savignac, H. M., Dinan, T. G., et al. (2011). Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc. Natl. Acad. Sci. USA* 108, 16050–16055. doi: [10.1073/pnas.1102999108](https://doi.org/10.1073/pnas.1102999108)

<span id="page-16-39"></span>Cani, P. D., and de Vos, W. M. (2017). Next-generation beneficial microbes: the case of *Akkermansia muciniphila*. *Front. Microbiol.* 8:1765. doi: [10.3389/](https://doi.org/10.3389/fmicb.2017.01765) [fmicb.2017.01765](https://doi.org/10.3389/fmicb.2017.01765)

<span id="page-16-35"></span>Cui, G., Liu, S., Liu, Z., Chen, Y., Wu, T., Lou, J., et al. (2022). Gut microbiome distinguishes patients with epilepsy from healthy individuals. *Front. Microbiol.* 12:696632. doi: [10.3389/fmicb.2021.696632](https://doi.org/10.3389/fmicb.2021.696632)

<span id="page-16-49"></span>Dahlin, M., and Prast-Nielsen, S. (2019). The gut microbiome and epilepsy. *EBioMedicine* 44, 741–746. doi: [10.1016/j.ebiom.2019.05.024](https://doi.org/10.1016/j.ebiom.2019.05.024)

Dai, Y., Wang, M., Zhong, D., and Xu, X. (2022). *Bacillus subtilis* plays a role in the inhibition of transporter ABCB1 in Caco-2 cells. *Epilepsy Res.* 183:106925. doi: [10.1016/j.eplepsyres.2022.106925](https://doi.org/10.1016/j.eplepsyres.2022.106925)

<span id="page-16-41"></span>Derrien, M., Belzer, C., and de Vos, W. M. (2017). Akkermansia muciniphila and its role in regulating host functions. *Microb. Pathog.* 106, 171–181. doi: [10.1016/j.](https://doi.org/10.1016/j.micpath.2016.02.005) [micpath.2016.02.005](https://doi.org/10.1016/j.micpath.2016.02.005)

Dong, L., Zheng, Q., Cheng, Y., Zhou, M., Wang, M., Xu, J., et al. (2022). Gut microbial characteristics of adult patients with epilepsy. *Front. Neurosci.* 16:803538. doi: [10.3389/](https://doi.org/10.3389/fnins.2022.803538) [fnins.2022.803538](https://doi.org/10.3389/fnins.2022.803538)

<span id="page-16-2"></span>Elliott, J., DeJean, D., Clifford, T., Coyle, D., Potter, B. K., Skidmore, B., et al. (2019). Cannabis-based products for pediatric epilepsy: a systematic review. *Epilepsia* 60, 6–19. doi: [10.1111/epi.14608](https://doi.org/10.1111/epi.14608)

<span id="page-16-53"></span>Falony, G., Joossens, M., Vieira-Silva, S., Wang, J., Darzi, Y., Faust, K., et al. (2016). Population-level analysis of gut microbiome variation. *Science* 352, 560–564. doi: [10.1126/science.aad3503](https://doi.org/10.1126/science.aad3503)

<span id="page-16-28"></span><span id="page-16-21"></span><span id="page-16-1"></span>Fisher, R. S., Acevedo, C., Arzimanoglou, A., Bogacz, A., Cross, J. H., Elger, C. E., et al. (2014). ILAE official report: a practical clinical definition of epilepsy. *Epilepsia* 55, 475–482. doi: [10.1111/epi.12550](https://doi.org/10.1111/epi.12550)

<span id="page-16-5"></span>Fisher, R. S., Cross, J. H., D'Souza, C., French, J. A., Haut, S. R., Higurashi, N., et al. (2017). Instruction manual for the ILAE 2017 operational classification of seizure types. *Epilepsia* 58, 531–542. doi: [10.1111/epi.13671](https://doi.org/10.1111/epi.13671)

<span id="page-16-4"></span>Fisher, R. S., Cross, J. H., French, J. A., Higurashi, N., Hirsch, E., Jansen, F. E., et al. (2017). Operational classification of seizure types by the international league against epilepsy: position paper of the ILAE Commission for Classification and Terminology. *Epilepsia* 58, 522–530. doi: [10.1111/epi.13670](https://doi.org/10.1111/epi.13670)

<span id="page-16-7"></span>Gacesa, R., Kurilshikov, A., Vich Vila, A., Sinha, T., Klaassen, M. A. Y., Bolte, L. A., et al. (2022). Environmental factors shaping the gut microbiome in a Dutch population. *Nature* 604, 732–739. doi: [10.1038/s41586-022-04567-7](https://doi.org/10.1038/s41586-022-04567-7)

<span id="page-16-45"></span>Goldberg, E. M., and Coulter, D. A. (2013). Mechanisms of epileptogenesis: a convergence on neural circuit dysfunction. *Nat. Rev. Neurosci.* 14, 337–349. doi: [10.1038/nrn3482](https://doi.org/10.1038/nrn3482)

<span id="page-16-33"></span>Gong, X., Cai, Q., Liu, X., An, D., Zhou, D., Luo, R., et al. (2021). Gut flora and metabolism are altered in epilepsy and partially restored after ketogenic diets. *Microb.*  Pathog. 155:105832. doi: [10.1016/j.micpath.2022.105832](https://doi.org/10.1016/j.micpath.2022.105832)

<span id="page-16-34"></span>Gong, X., Liu, X., Chen, C., Lin, J., Li, A., Guo, K., et al. (2020). Alteration of gut microbiota in patients with epilepsy and the potential index as a biomarker. *Front. Microbiol.* 11:517797. doi: [10.3389/fmicb.2020.517797](https://doi.org/10.3389/fmicb.2020.517797)

<span id="page-16-14"></span>Hozo, S. P., Djulbegovic, B., and Hozo, I. (2005). Estimating the mean and variance from the median, range, and the size of a sample. *BMC Med. Res. Methodol.* 5:13. Published 2005. doi: [10.1186/1471-2288-5-13](https://doi.org/10.1186/1471-2288-5-13)

<span id="page-16-52"></span>Kaiko, G. E., Ryu, S. H., Koues, O. I., Pearce, E. L., Oltz, E. M., Stappenbeck, T. S., et al. (2016). The colonic crypt protects stem cells from microbiota-derived metabolites [published correction appears in cell. 2016; 167(4): 1137. Doi:10.1016/j.cell.2016.10.034]. *Cell* 165, 1708–1720. doi: [10.1016/j.cell.2016.05.018](https://doi.org/10.1016/j.cell.2016.05.018)

<span id="page-16-30"></span>Knight, R., Vrbanac, A., Taylor, B. C., Aksenov, A., Callewaert, C., Debelius, J., et al. (2018). Best practices for analysing microbiomes. *Nat. Rev. Microbiol.* 16, 410–422. doi: [10.1038/s41579-018-0029-9](https://doi.org/10.1038/s41579-018-0029-9)

<span id="page-16-9"></span>Kwan, P., and Brodie, M. J. (2000). Early identification of refractory epilepsy. *N. Engl. J. Med.* 342, 314–319. doi: [10.1056/NEJM200002033420503](https://doi.org/10.1056/NEJM200002033420503)

Lee, K., Kim, N., Shim, J. O., and Kim, G. H. (2020). Gut bacterial dysbiosis in children with intractable epilepsy. *J. Clin. Med.* 10:5. doi: [10.3390/jcm10010005](https://doi.org/10.3390/jcm10010005)

<span id="page-16-29"></span>Li, Z., Zhou, J., Liang, H., Ye, L., Lan, L., Lu, F., et al. (2022). Differences in alpha diversity of gut microbiota in neurological diseases. *Front. Neurosci.* 16:879318. doi: [10.3389/fnins.2022.879318](https://doi.org/10.3389/fnins.2022.879318)

Lindefeldt, M., Eng, A., Darban, H., Bjerkner, A., Zetterström, C. K., Allander, T., et al. (2019). The ketogenic diet influences taxonomic and functional composition of the gut microbiota in children with severe epilepsy. *NPJ Biof. Microb.* 5:5. doi: [10.1038/](https://doi.org/10.1038/s41522-018-0073-2) [s41522-018-0073-2](https://doi.org/10.1038/s41522-018-0073-2)

<span id="page-16-15"></span>Liu, T., Jia, F., Guo, Y., Wang, Q., Zhang, X., Chang, F., et al. (2023). Altered intestinal microbiota composition with epilepsy and concomitant diarrhea and potential indicator biomarkers in infants. *Front. Microbiol.* 13:1081591. doi: [10.3389/fmicb.2022.](https://doi.org/10.3389/fmicb.2022.1081591) [1081591](https://doi.org/10.3389/fmicb.2022.1081591)

<span id="page-16-25"></span><span id="page-16-24"></span><span id="page-16-22"></span><span id="page-16-20"></span><span id="page-16-19"></span><span id="page-16-18"></span><span id="page-16-13"></span>Liu, H., Wu, H., Yao, C., et al. (2017). Advanced methods of data extraction for continuous outcomes in meta-analysis. *Chin. J. Evid. Based Med.* 17, 117–121. doi: [10.7507/1672-2531.201612004](https://doi.org/10.7507/1672-2531.201612004)

<span id="page-16-44"></span>Lum, G. R., Olson, C. A., and Hsiao, E. Y. (2020). Emerging roles for the intestinal microbiome in epilepsy. *Neurobiol. Dis.* 135:104576. doi: [10.1016/j.nbd.2019.104576](https://doi.org/10.1016/j.nbd.2019.104576)

<span id="page-16-54"></span>Mabwi, H. A., Kim, E., Song, D. G., Yoon, H. S., Pan, C. H., Komba, E. V. G., et al. (2020). Synthetic gut microbiome: advances and challenges. *Comput. Struct. Biotechnol. J.* 19, 363–371. doi: [10.1016/j.csbj.2020.12.029](https://doi.org/10.1016/j.csbj.2020.12.029)

<span id="page-16-43"></span>Medel-Matus, J. S., Shin, D., Dorfman, E., Sankar, R., and Mazarati, A. (2018). Facilitation of kindling epileptogenesis by chronic stress may be mediated by intestinal microbiome. *Epilepsia Open.* 3, 290–294. doi: [10.1002/epi4.12114](https://doi.org/10.1002/epi4.12114)

<span id="page-16-51"></span>Mejía-Granados, D. M., Villasana-Salazar, B., Lozano-García, L., Cavalheiro, E. A., and Striano, P. (2021). Gut-microbiota-directed strategies to treat epilepsy: clinical and experimental evidence. *Seizure* 90, 80–92. doi: [10.1016/j.seizure.2021.03.009](https://doi.org/10.1016/j.seizure.2021.03.009)

<span id="page-16-36"></span>Mohamed Qadir, R., and Assafi, M. S. (2021). The association between body mass index and the oral Firmicutes and Bacteroidetes profiles of healthy individuals. *Malays. Fam. Physician.* 16, 36–43. doi: [10.51866/oa1129](https://doi.org/10.51866/oa1129)

<span id="page-16-10"></span>Newman, L. (2000). AHRQ's evidence-based practice centres prove viable. Agency<br>for Healthcare Research and Quality *Lancet* 356:1990 doi: 10.1016/ for Healthcare Research and Quality. *Lancet* 356:1990. doi: [10.1016/](https://doi.org/10.1016/s0140-6736(05)72963-9) [s0140-6736\(05\)72963-9](https://doi.org/10.1016/s0140-6736(05)72963-9)

<span id="page-16-42"></span>Olson, C. A., Vuong, H. E., Yano, J. M., Liang, Q. Y., Nusbaum, D. J., and Hsiao, E. Y. (2018). The gut microbiota mediates the anti-seizure effects of the ketogenic diet. *Cell* 174:497. doi: [10.1016/j.cell.2018.06.051](https://doi.org/10.1016/j.cell.2018.06.051)

<span id="page-16-12"></span>Page, M. J., McKenzie, J. E., Bossuyt, P. M., et al. (2021). The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ* 372:n71. doi: [10.1136/bmj.n71](https://doi.org/10.1136/bmj.n71)

<span id="page-16-27"></span><span id="page-16-23"></span><span id="page-16-16"></span>Peng, A., Qiu, X., Lai, W., Li, W., Zhang, L., Zhu, X., et al. (2018). Altered composition of the gut microbiome in patients with drug-resistant epilepsy. *Epilepsy Res.* 147, 102–107. doi: [10.1016/j.eplepsyres.2018.09.013](https://doi.org/10.1016/j.eplepsyres.2018.09.013)

<span id="page-16-40"></span>Rajilić-Stojanović, M., Shanahan, F., Guarner, F., and de Vos, W. M. (2013). Phylogenetic analysis of dysbiosis in ulcerative colitis during remission. *Inflamm. Bowel Dis.* 19, 481–488. doi: [10.1097/MIB.0b013e31827fec6d](https://doi.org/10.1097/MIB.0b013e31827fec6d)

<span id="page-16-47"></span>Reddy, S. D., Clossen, B. L., and Reddy, D. S. (2018). Epigenetic histone deacetylation inhibition prevents the development and persistence of temporal lobe epilepsy. *J. Pharmacol. Exp. Ther.* 364, 97–109. doi: [10.1124/jpet.117.244939](https://doi.org/10.1124/jpet.117.244939)

<span id="page-16-37"></span>Rodenhouse, A., Talukder, M. A. H., Lee, J. I., Govindappa, P. K., O'Brien, M., Manto, K. M., et al. (2022). Altered gut microbiota composition with antibiotic treatment impairs functional recovery after traumatic peripheral nerve crush injury in mice: effects of probiotics with butyrate producing bacteria. *BMC. Res. Notes* 15:80. doi: [10.1186/](https://doi.org/10.1186/s13104-022-05967-8) [s13104-022-05967-8](https://doi.org/10.1186/s13104-022-05967-8)

<span id="page-16-3"></span>Scheffer, I. E., Berkovic, S., Capovilla, G., Connolly, M. B., French, J., Guilhoto, L., et al. (2017). ILAE classification of the epilepsies: position paper of the ILAE Commission for Classification and Terminology. *Epilepsia* 58, 512–521. doi: [10.1111/](https://doi.org/10.1111/epi.13709) [epi.13709](https://doi.org/10.1111/epi.13709)

<span id="page-16-38"></span>Schloissnig, S., Arumugam, M., Sunagawa, S., Mitreva, M., Tap, J., Zhu, A., et al. (2013). Genomic variation landscape of the human gut microbiome. *Nature* 493, 45–50. doi: [10.1038/nature11711](https://doi.org/10.1038/nature11711)

<span id="page-16-8"></span>Sutter, R., Rüegg, S., and Tschudin-Sutter, S. (2015). Seizures as adverse events of antibiotic drugs: a systematic review. *Neurology* 85, 1332–1341. doi: [10.1212/](https://doi.org/10.1212/WNL.0000000000002023) [WNL.0000000000002023](https://doi.org/10.1212/WNL.0000000000002023)

<span id="page-16-0"></span>Thijs, R. D., Surges, R., O'Brien, T. J., and Sander, J. W. (2019). Epilepsy in adults. *Lancet* 393, 689–701. doi: [10.1016/S0140-6736\(18\)32596-0](https://doi.org/10.1016/S0140-6736(18)32596-0)

<span id="page-16-11"></span>Tremlett, H., Bauer, K. C., Appel-Cresswell, S., Finlay, B. B., and Waubant, E. (2017). The gut microbiome in human neurological disease: a review. *Ann. Neurol.* 81, 369–382. doi: [10.1002/ana.24901](https://doi.org/10.1002/ana.24901)

Türay, S., Cangür, Ş., Kahraman, G., Kayabaşı, E., Çetiner, Ö. F., Aydın, B., et al. (2023). Can the gut microbiota serve as a guide to the diagnosis and treatment of childhood epilepsy? *Pediatr. Neurol.* 145, 11–21. doi: [10.1016/j.pediatrneurol.2023.04.006](https://doi.org/10.1016/j.pediatrneurol.2023.04.006)

<span id="page-16-50"></span>Ułamek-Kozioł, M., Czuczwar, S. J., Januszewski, S., and Pluta, R. (2019). Ketogenic diet and epilepsy. *Nutrients* 11:2510. doi: [10.3390/nu11102510](https://doi.org/10.3390/nu11102510)

Wan, L., Yang, G., Zhang, S., Sun, Y., Li, Z., Wang, J., et al. (2021). Investigation of the association between imbalance of the intestinal flora and infantile spasms: a pilot case-control study. *Transl. Pediatr.* 10, 819–833. doi: [10.21037/tp-20-384](https://doi.org/10.21037/tp-20-384)

<span id="page-16-48"></span>Weis, A. M., and Round, J. L. (2021). Microbiota-antibody interactions that regulate gut homeostasis. *Cell Host Microbe* 29, 334–346. doi: [10.1016/j.chom.2021.02.009](https://doi.org/10.1016/j.chom.2021.02.009)

<span id="page-16-6"></span>World Health Organization. Epilepsy [fact sheet]. (2024-02-07) (2024). Available at: <https://www.who.int/news-room/fact-sheets/detail/epilepsy> (Accessed October 18, 2024).

<span id="page-16-31"></span>Xie, G., Zhou, Q., Qiu, C. Z., Dai, W. K., Wang, H. P., Li, Y. H., et al. (2017). Ketogenic diet poses a significant effect on imbalanced gut microbiota in infants with refractory epilepsy. *World J. Gastroenterol.* 23, 6164–6171. doi: [10.3748/wjg.v23.i33.6164](https://doi.org/10.3748/wjg.v23.i33.6164)

<span id="page-16-32"></span>Xu, L., Chen, D., Zhao, C., Jiang, L., Mao, S., Song, C., et al. (2021). Decreased abundance of Akkermansia after adrenocorticotropic hormone therapy in patients with West syndrome. *BMC Microbiol.* 21:126. doi: [10.1186/s12866-021-02189-z](https://doi.org/10.1186/s12866-021-02189-z)

<span id="page-17-3"></span>Yan, Y., Li, K., Jiang, J., Jiang, L., Ma, X., Ai, F., et al. (2023). Perinatal tissue-derived exosomes ameliorate colitis in mice by regulating the Foxp 3+Treg cells and gut microbiota. *Stem Cell Res Ther* 14:43. Published 2023 Mar 20. doi: [10.1186/s13287-023-03263-1](https://doi.org/10.1186/s13287-023-03263-1)

<span id="page-17-6"></span>Younus, I., and Reddy, D. S. (2017). Epigenetic interventions for epileptogenesis: a new frontier for curing epilepsy. *Pharmacol. Ther.* 177, 108–122. doi: [10.1016/j.](https://doi.org/10.1016/j.pharmthera.2017.03.002) [pharmthera.2017.03.002](https://doi.org/10.1016/j.pharmthera.2017.03.002)

<span id="page-17-5"></span>Zeng, Y., Cao, S., and Yang, H. (2023). Roles of gut microbiome in epilepsy risk: a Mendelian randomization study. *Front. Microbiol.* 14:1115014. doi: [10.3389/](https://doi.org/10.3389/fmicb.2023.1115014) [fmicb.2023.1115014](https://doi.org/10.3389/fmicb.2023.1115014)

<span id="page-17-4"></span>Zhai, Q., Feng, S., Arjan, N., and Chen, W. (2019). A next generation probiotic, *Akkermansia muciniphila*. *Crit. Rev. Food Sci. Nutr.* 59, 3227–3236. doi: [10.1080/10408398.2018.1517725](https://doi.org/10.1080/10408398.2018.1517725)

<span id="page-17-2"></span>Zhang, Z., Cheng, N., Liang, J., Deng, Y., Xiang, P., Hei, Z., et al. (2023). Gut microbiota changes in animal models of spinal cord injury: a preclinical systematic review and meta-analysis. *Ann. Med.* 55:2269379. doi: [10.1080/07853890.2023.2269379](https://doi.org/10.1080/07853890.2023.2269379)

<span id="page-17-1"></span><span id="page-17-0"></span>Zhou, C., Gong, S., Xiang, S., Liang, L., Hu, X., Huang, R., et al. (2022). Changes and significance of gut microbiota in children with focal epilepsy before and after treatment. *Front. Cell. Infect. Microbiol.* 12:965471. doi: [10.3389/fcimb.2022.965471](https://doi.org/10.3389/fcimb.2022.965471)