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Surveillance of Erythrovirus B19 (B19V) in patients with acute febrile illness suspected of arboviruses in Mato Grosso do Sul state, Brazil

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Introduction: Human Erythrovirus (parvovirus) B19 infection can produce symptoms similar to those produced by Dengue, Chikungunya, and Zika viruses, making clinical diagnosis difficult. The importance of erythrovirus B19 in human pathology has been increased and reported in numerous studies published globally.

Methods: The B19V infection was investigated by real-time PCR in sera samples from patients with signs and symptoms related to classic arboviral symptoms. This study was conducted to provide information on the genetic diversity of Human Erythrovirus B19 (B19V) circulating in the state of Mato Grosso do Sul, Midwest region of Brazil, from 2017 to 2022. A total of 773 sera samples of patients with negative diagnostic results for Dengue, Chikungunya, and Zika, during the study period were analyzed.

Results: Erythrovirus DNA was found in 10.6% (82/773) of patients, among them 10 were pregnant women. Four samples were completely sequenced, and the other five partially, to genotype by phylogenetic reconstruction. All samples belong to worldwide dispersed genotype 1, subgenotype 1a.

Discussion: The findings of the study demonstrate the importance of including B19V in differential laboratory diagnosis for epidemiological purposes and appropriate patient management. The diagnosis for B19V should be performed,

particularly among pregnant women, immunocompromised patients, and individuals with hemolytic diseases, given that the infection is more severe in these cases.

KEYWORDS

Parvovirus B19, Dengue, Chikungunya, Zika, differential diagnosis, human, surveillance

1 Introduction

Human Erythrovirus B19 (parvovirus B19), of the Parvoviridae family, a member of the Erythroparvovirus genus, was discovered in England by Cossart et al. (1975) in serum samples from blood donors who performed serological testing for hepatitis B virus. This sample was coded as "number 19 in panel B", and later the virus was called B19 (Cossart et al., 1975). After its discovery, the B19V infection was associated with an asymptomatic or benign acute pediatric infection known as erythema infectiosum and other clinical manifestations such as transient aplastic crisis, and arthropathies (Moore, 2000).

The transmission of B19V is through the respiratory droplets. However, the virus can also be transmitted parenterally, especially by haemoderivative applications (Cotmore et al., 2013). The Erythrovirus B19 has a tropism for human erythroid progenitor cells and it is involved in suppressing blood cell formation during infection (Servant et al., 2002). In 2002, the virus was classified into three genotypes, defined as genotype 1 B19 classic (represented by the prototype strain Au), genotype 2 (prototype K71 and strain A6-similar), and genotype 3 (prototype V9) (Cruz et al., 1989).

The infection by Human Erythrovirus B19 during pregnancy has been widely studied, and it is known to cause a range of complications, including spontaneous abortion (Moore, 2000). The virus acts on the inhibition of red blood cell formation, generating cytotoxic effects that lead to variable clinical conditions, such as intrauterine growth retardation, myocarditis, and pericardial effusions, which will depend on the patient's hematological and immunological status. The age group most affected is children/adolescents under 14 years old (Garcia and Leon, 2021). However, many infections are asymptomatic and are not correctly diagnosed.

In Brazil, the B19V and some arboviruses (Dengue, Chikungunya, and Zika) co-circulate in several regions of the country, making the differential diagnosis difficult, mainly because these viruses can cause similar symptoms associated with exanthema and acute febrile diseases (Alves, 2022).

The disease caused by B19V was not classified as national or state compulsory notification, in Brazil. The infection was detected in surveillance of exanthematous diseases, as a differential diagnosis in suspected cases of measles or rubella, according to the characteristics of symptoms (Cubel et al., 1997). In 2020, the State Secretary of Health of Mato Grosso do Sul (SES) and the Central Laboratory of Public Health (LACEN) recommended that suspected cases of Erythema Infectiosum (B19V) should be notified, but it was only from 2021 that some cases were included in the Notification Diseases Information System (SINAN). The detection of IgM antibodies against parvovirus B19 is carried out at the LACEN following national guidelines and protocols provided by the Ministry of Health. In addition, the use of real-time PCR for the molecular diagnosis of B19V enables its prompt detection, ruling out concurrent arboviral infections or other viruses with similar symptoms.

This is particularly relevant in Brazilian regions with a high potential for annual dengue cases. The implementation in the laboratory routine can clarify whether erythrovirus B19 may be one of the etiological agents of exanthematous diseases, as well as determine the frequency of their infection in patients, particularly among pregnant women, immunocompromised patients, and individuals with hemolytic diseases since the infection is more serious in these cases.

Our study describes the presence of Human Erythrovirus B19 virus in samples from patients with acute febrile syndromes and negative diagnostic results for Dengue, Chikungunya, and Zika collected in several municipalities in the state of Mato Grosso do Sul, Brazil, during the period from 2017 to 2022. We demonstrate that molecular approaches contribute to differential diagnosis of Human Erythrovirus B19 and epidemiological purposes, monitoring the characterization and genetic variability of circulating strains in the region stu.

2 Materials and methods

2.1 Study design and sample collection

Biological samples were selected from the Biobank of the Central Laboratory of Public Health of Mato Grosso do Sul state (LACEN/MS), based on the following criteria: (i) serum, urine, and cerebrospinal fluid samples collected between January 2017 and October 2022; (ii) with clinical suspicion of chikungunya, dengue or zika disease but with negative test results ("not detectable" by the RT-qPCR and "non-reactive" when using the NS1 and enzyme-linked immunosorbent assay (ELISA) for these arboviruses; (iii) reported onset of symptoms within 5 days for the NS1 test; and (iv) availability at the time of sample selection, since a substantial number of samples were discarded during the laboratory routine due to a lack of storage capacity, a condition exacerbated by the SARS-CoV-2 pandemic; (v) samples with a volume equal or more than 200 microliters.

Socio-demographic and clinical data of each sample were obtained from the medical request forms to assess associations with Erythrovirus B19 infection, including, gender, age, city of residence, reported symptoms, pregnancy, and confirmation of death. In addition, the records of positive cases of CHIKV, DENV, and ZIKV during the routine diagnosis of LACEN/MS were compared with the qPCR results obtained for B19V in the present study.

2.2 Ethical approval

This study was conducted with the authorization of the Research Ethics Committee involving Human Subjects at the Dom Bosco Catholic University of Mato Grosso do Sul, Brazil (CAAE: 54019821.2.0000.5162) and with the approvals of the Central Laboratory of Public Health of Mato Grosso do Sul (LACEN/MS) and the State Health Department of Mato Grosso do Sul. The study was conducted in accordance with the National legislation and state authorities.

2.3 Erythrovirus real-time PCR (qPCR)

The nucleic acids of each sample were extracted using the QIAsymphony DSPVirus/ Pathogen Mini Kit (QIAGEN, Germany), according to the manufacturer's protocol. The extracted DNA was tested for B19V using the protocol developed by Naveca et al. (data not yet published) from the Leônidas and Maria Deane Institute (ILMD), Fiocruz Amazônia. Primers and probes are presented in Figure 1 and Supplementary Table 1. Real-time PCR (qPCR) was performed using the KAPA PROBE FAST qPCR Master Mix (2X) (Roche), following the calculations shown in Supplementary Table 2. In all the reactions, the amplification of the human internal control gene (RNase P) was used to rule out false negatives, thereby confirming the accuracy of the results. A no-template control and positive control for B19V were used to discard possible contamination and validate the reaction. The qPCR was performed using the ABI 7500 and QuantStudioTM 5 (Applied Biosystems). Following the manufacturer's protocol, it applied an initial denaturation at 95°C for 5 min, with 40 cycles of denaturation at 95°C for 3 seg and combined annealing/extension/data acquisition at 60°C for 30 seg. The threshold for the quantification cycle (Cq) was calculated automatically with default settings using equipment software. Any sample was considered positive for B19V when the Cq value was <37.0.

2.4 Sequencing

Samples with positive results for B19V were amplified by conventional PCR using three pairs of primers (Supplementary Table 3), covering almost the entire viral genome. Nine samples were separated for sequencing (Table 1). They were indicative of the first 3 years of the study and had a high viral load. PCR reactions were performed using SuperFi II green enzyme (Thermo Fisher Scientific), primers at 0.5 μ M, and 1 μ L of viral DNA in 10 μ L of the final volume. The recommended cycling parameters were as follows, 98°C of initial denaturation for 30seg, 35 cycles of 10seg denaturation at 98°C, 10seg annealing at 55°C, 2 min extension at 72°C, ending with 5 min final extension at 72°C. At the end of the reactions, each amplicon was subjected to electrophoresis at 80V for 1 h, visualized on a 1% agarose gel stained with GelRed (Biotium), and the GeneRuler 1 kb DNA Ladder (Thermo Fisher Scientific) was used to confirm the expected size of the product. Subsequently, the PCR products were precipitated using polyethylene glycol 8000 following an adapted protocol. Initially, 20% PEG8000 (Promega) was added at a 1:1 ratio to the tube containing the PCR product, followed by incubation at 37°C for 15 min. After incubation, centrifugation was performed at 13,000 \times g for 15 min. The supernatant was then discarded, 125 µL of 80% ethanol was added, followed by a new centrifugation step at 12,000 \times g for 2 min, and the supernatant was discarded. The microtube was then placed in a Mivac DNA concentrator (Genevc SP Scientific) for 15 min at 37°C. DNA was then resuspended in nuclease-free water and quantified using a microvolume spectrophotometer (Biodrop Duo Biochrom). All amplicons from the same sample were pooled together and quantified using the Qubit 2.0 and dsDNA HS Assay kit (Thermo Fisher Scientific).

Additionally, 4 samples were suitable for whole-genome sequencing. Library preparation was performed using a Nextera XT DNA Library Prep Kit (Illumina). Enzymatic tagmentation, adapter, and index addition, amplification, normalization, and pool libraries were performed as described in the manufacturer's manual using 1ng of DNA. All libraries were quantified using a Qubit dsDNA HS Assay Kit. Sequencing was performed on MiSeq equipment (Illumina) using the Reagent v2 500-cycle sequencing kit. The data generated after sequencing were analyzed in Geneious Prime 2022.0.1 software for the assembly of contigs of each sample, using the genome NC_000883 as the reference and the BBMap v 38.84 tool under "default" conditions. The remaining five samples were partially sequenced (ranging from 585 to 1281 base pairs of the VP1/VP2 gene) using standard procedures for capillary sequencing, as recommended for ABI3130 (ThermoFisher). Trace files were edited for quality and primer removal and assembled using the B19V. The nucleotide sequences obtained during this study were deposited in the GenBank database (Table 1).

2.5 Phylogenetic analysis

The consensus B19V sequences obtained in this study and the reference sequences available on GenBank [G1a: M13178 (isolated Au); G1b: DQ357064 (isolated Vn147); G2: AY064475 (isolated A6); G2 AY044266 (isolated LaLi); G3a: AX003421 (isolated V9) and G3b: AY083234 (isolated D91.1)] were aligned using the MAFFT v7.490 tool with automatic algorithm selection. Subsequently, the alignment file was subjected to maximumlikelihood phylogenetic reconstruction with the FastTree 2.1.11 program using the GTR evolutionary model. An approximate likelihood ratio test (aLRT) (Anisimova and Gascuel, 2006) was used to assess the branch supports. The phylogenetic tree had the root placed at the central point, with increasing node order, and was edited in the FigTree 1.4.4 program. Tip labels were aligned for clarity.

2.6 Statistical analyses

Mixed Generalized Linear Models (GLMMs) with random effects for municipalities and binomial distribution were used to

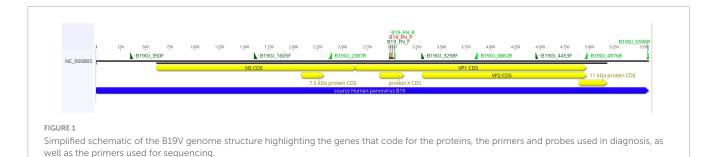


TABLE 1 Characteristics of samples sequenced to identify the circulating B19V genotype.

Code ID	Age	Gender	Municipality	Collection date	Pregnancy	Accession number
MS-B19-17-201-532	21	Woman	Dourados	03/26/2017	Yes	OR542579
MS-B19-17-32-174	39	Woman	Chapadão do Sul	05/05/2017	No	OR542583
MS-B19-17-25-839	20	Woman	Corumbá	05/12/2017	Yes	OR542582
MS-B19-17-201-1400	19	Woman	Dourados	07/05/2017	Yes	OR578525
MS-B19-17-18-408	38	Woman	Caarapó	08/03/2017	No	OR542580
MS-B19-17-31-100	20	Woman	Eldorado	10/22/2017	No	OR578526
MS-B19-17-31-148	26	Man	Eldorado	11/15/2017	-	OR578523
MS-B19-18-102-7397	32	Man	Campo Grande	07/25/2018	-	OR578524
MS-B19-19-04-3	1	Woman	Alcinópolis	01/18/2019	No	OR542581

-, not applicable. All municipalities are in the state of Mato Grosso do Sul, Brazil. The GenBank access numbers are shown for each sample.

understand how PCR test results could be related to (1) the age and gender of patients, as well as the year of sampling; and (2) the symptoms recorded in patient records. Symptoms were evaluated in a separate model, owing to fewer records. Multicollinearity was checked by examining the Pearson correlation coefficient (Pearson's r) between each pair of explanatory variables, using the R package 'correlation' (Makowski et al., 2019; Makowski et al., 2022), and computing the variance inflation factor (VIF), with the R package 'performance' (Lüdecke et al., 2021). The analyses were performed using the 'lme4' package (Bates et al., 2015) in the R software (R Core Team, 2023). Tables and estimation plots were generated using the 'jtools' package (Long, 2022). The models were checked for normality of residuals, normality of random effects, homogeneity of variance, and residual dispersion using the packages 'DHARMa' (Hartig, 2022) and 'performance'.

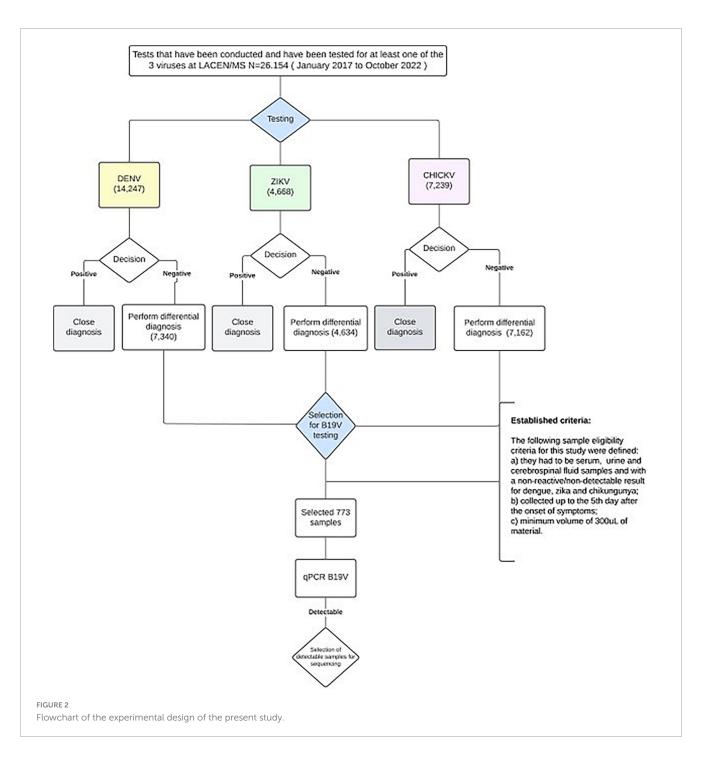
In years when sampling was conducted for all months, circular statistics were employed to test seasonal patterns in the number of cases in Mato Grosso do Sul (Almeida et al., 2018; Landler et al., 2019, 2020). Two approaches were used: (1) a hypothesis test and (2) a model-based approach, both implemented in the R package 'CircMLE' (Fitak and Jonhsen, 2017). In the hypothesis test, the null hypothesis is that the data are uniformly distributed throughout the year (no clear pattern of occurrence), versus some form of concentration, be it unimodal (occurrence concentrated in a single specific period), bimodal (occurrence concentrated in two different periods), or multimodal (occurrence concentrated in more than two different periods) (Landler et al., 2020). The Hermans-Rasson test was used to verify the uniformity of the distributions, as it has high statistical power for grouped multimodal distributions (Landler et al., 2019). In the model-based approach, the package calculates the maximum likelihood of 10 models described by

Schnute and Groot (1992) and compares them using model selection criteria. Thus, this approach allows the identification of patterns of occurrence. Briefly, the models fall into three main categories: (i) a uniform model (M1) of random orientation; (ii) unimodal models (M2A, M2B, M2C) with a single preferred direction; and (iii) bimodal models (M3, M4, and M5) with two preferred directions. Bimodal models can also be divided into axial (M3A, M3B, M4A, M4B) and non-axial (M5A, M5B) (Schnute and Groot, 1992; Fitak and Jonhsen, 2017; Landler et al., 2020; Ojeda et al., 2021). Herein, the models were compared based on Akaike's Information Criterion with small-sample correction (AICc). Additionally, the Akaike weight (w_i) was used, which describes the probability that a particular model is the best model (approximate), given the experimental data and the collection of models considered (Burnham et al., 2011; Portet, 2020; Ojeda et al., 2021). For each year, it was also calculated the circular standard deviation (sd), circular mean (μ), and the length of the mean vector (r) using the R package 'circular' (Agostinelli and Lund, 2023). The μ represents the mean date of cases, and r represents how the data is clustered around the mean (0, perfectly uniformly distributed; and 1, perfectly clustered). All statistical analyses were performed using the R software v4.2.1.

3 Results

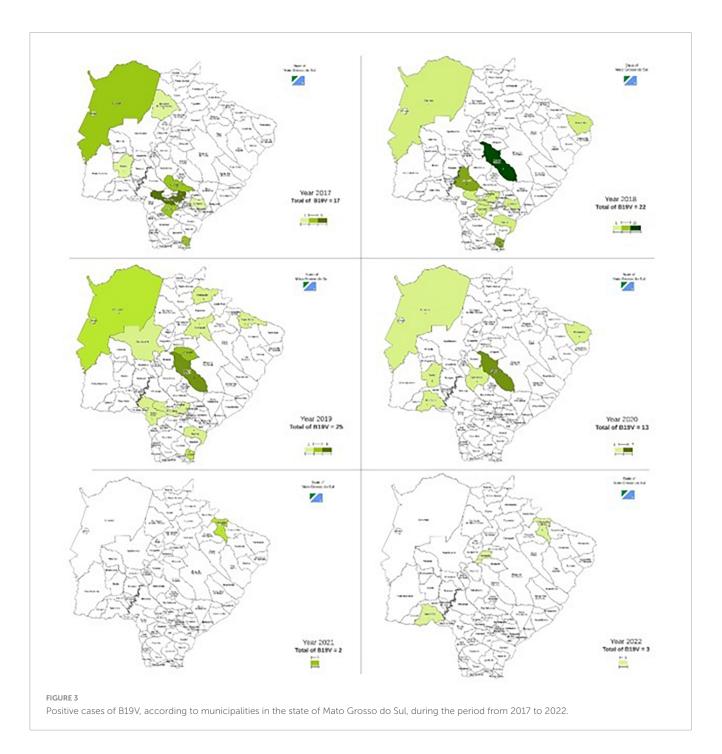
3.1 Erythrovirus DNA detection

Between January 2017 and October 2022, LACEN/MS performed 26,154 tests for at least one of the three arboviruses.



Some of the samples received for diagnosis of dengue were also tested for CHIKV and/or ZIKV, and others were just evaluated for CHIKV and/or ZIKV. Of the total samples analyzed, 7,239 were tested for CHIKV, 14,247 for DENV, and 4,668 for ZIKV. Of those, 7,162 were negative for CHIKV (98.9%), 7,340 for DENV (51.5%), and 4,634 for ZIKV (99.3%) (Figure 2). Despite the high number of tests performed, few samples were available at the time of sample selection. Therefore, only 773 samples from 62 municipalities in the state of Mato Grosso do Sul met the previously defined criteria, had negative results for dengue, zika, and chikungunya, and were chosen for B19V research (Figure 2). Of the total samples analyzed, 82 were positive for B19V (10.6%), from 24 municipalities (Figure 3 and Table 2), surpassing the number of cases registered for Chikungunya and Zika during the same period in the state of Mato Grosso do Sul (Figure 4).

Of the total positive samples, 28 were collected in men and 54 in women (Supplementary Figure 1), with 10 pregnant women. The ages of patients ranged from 1 to 70 years, however, B19V detection was not significantly associated with gender and age group, although it was significantly different between the years of the period evaluated (Figure 5A). The positive results in the years 2019, 2021, and 2022 were significantly lower than in 2017 (Figure 5A and Table 2). The notification forms of 598 patients contained clinical



information, identifying a total of 20 symptoms. Among B19Vpositive patients, fever was the most frequent symptom, followed by myalgia, headache, arthralgia, retro-orbital pain, nausea, and rash (Figure 6). Despite this, B19V detection was significantly associated only with retro-orbital pain, leukopenia, petechiae, and malaise (Figure 5B and Table 2). The symptom "abdominal discomfort" was not included in the model because it was correlated with "diarrhea". The symptoms "gingival bleeding", "renal failure", "sore throat" and "thrombocytopenia" were not included because they only had 2 cases in the notification forms, causing problems in the estimates.

The phylogenetic reconstruction showed that all samples sequenced in this study were recovered with Genotype 1, subgenotype 1a (RefSeq M13178), with high support (aLRT = 0.94)

(Figure 7). The B19V samples that underwent sequencing originate from different municipalities (Corumbá, Campo Grande, Dourados, Deodápolis, Caarapó, Chapadão do Sul, and Alcinópolis) in various geographic areas, demonstrating the wide spread of the virus by the whole state (Table 1 and Supplementary Figure 2).

The results of the Hermans-Rasson test demonstrated that the number of cases was seasonal for all the years evaluated, 2017 (7.29; p = 0.033), 2018 (13.02; p < 0.001), and 2019 (13.90; p < 0.001). For 2017, despite the best model having presented an AICc w_i with more than twice the second, the following models can still be considered plausible choices. When considering the first two models (M3A and M4A, AICc $w_i = 0.38$ and 0.17; Figure 8

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TABLE 2 Population and municipalities studied by year and positive cases for B19V.

Year	Samples	Positives (%)	Municipali- ties sampled	Municipali- ties with positive cases (%)
2017	82	17 (20.7)	28	10 (35.7)
2018	160	22 (13.8)	27	10 (37.0)
2019	285	25 (8.8)	39	11 (28.2)
2020	71	13 (18.3)	22	6 (27.3)
2021	86	2 (2.3)	15	1 (6.7)
2022	89	3 (3.4)	22	3 (13.6)
Total	773	82 (10.6)	62	24 (38.7)

and Supplementary Table 4), they together presented an AICc $w_i = 0.55$, which means that there is at least a 55% chance that it is the best approximation that describes the data, given the candidate set of orientations considered. Both models are axial bimodal, suggesting a distribution in two equally sized points in 2017, one in each semester, with the mean date of cases in September (mean date = September 01, 2017; sd = 1.67; r = 0.25). In 2018, no model stood out, and the first two were equally supported, with close AICc w_i values, which, together, presented a value of AICc $w_i = 0.71$ (M5A and M3A, AICc $w_i = 0.38$ and 0.33; Figure 8 and Supplementary Table 5). Although both have a bimodal distribution, one is non-axial (M5A), while the other is (M3A), making it more complicated to determine a pattern for the year 2018. Even so, both models showed a division of distribution between the two semesters, with a greater presence of cases closer to the end of each semester, with the mean date of cases in July (mean date = July 23, 2018; sd = 1.68; r = 0.25). In contrast, the pattern shown in 2019 was unequivocal, with an unimodal distribution (M2A, AICc $w_i = 0.72$; Figure 8 and Supplementary Table 6) concentrated in the first half, with the average date in March (mean date = March 17, 2019; sd = 1.03; r = 0.59). There was a greater number of cases in 2017 and 2018, having decreased in 2019, rising again in 2020, and decreasing again in 2021 and 2022, with a positivity percentage of 20.7, 13.75, 8.8, 18.3, 2.3 and 3.4%, respectively (Figures 4, 5A and Table 2).

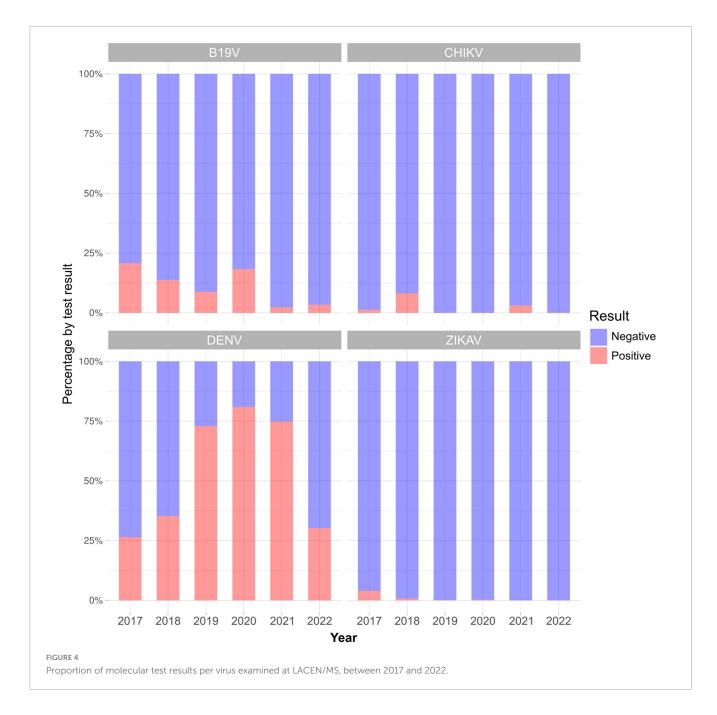
4 Discussion

A wide variety of viruses can cause rashes and joint pain in children, adolescents, and adults, leading to etiological identification exclusively through clinical examination, which makes it a challenging task. The most common agents of exanthematous disease (ED) include Measles, Rubella, Dengue, Chickenpox, Cytomegalovirus, Epstein Barr, Human HerpesVirus 6, Enterovirus, Erythrovirus (Human Parvovirus B19), Chikungunya and Zika (Holterhus et al., 2023). Although these injuries are common in Brazil, there are several difficulties in determining an accurate etiological diagnosis, especially in those regions where there is co-circulation of exanthematous diseases and arboviruses. Therefore, the differential diagnosis of these diseases is extremely important, not only for epidemiological purposes but also for the control and treatment of infections. Even if most of them have a benign course, for certain age groups, pregnant women, and immunocompromised people, some infections represent an important risk, as they can evolve into serious cases, which may require emergency hospitalization, burdening the public health service (Garcia and Leon, 2021).

Molecular diagnosis of B19V infection is not commonly used as a differential diagnosis, and there are no studies dealing with the detection of B19V using the qPCR methodology in the state of Mato Grosso do Sul, Brazil. The continuous lack of specific studies for B19V in the state of Mato Grosso do Sul may be contributing to the high number of cases that are not laboratory-confirmed (Figure 4). This highlights the need to expand laboratory diagnosis so that B19V can be monitored. However, it is important to point out that the use of a sensitive and specific molecular method is of paramount importance for clarifying the laboratory diagnosis of different viral infections. Thus, our study is the first one showing the circulation of B19V in the state of Mato Grosso do Sul, in the Midwest region of Brazil, using the new protocol developed by F.G. Naveca and his team. Also, our study demonstrates the need and feasibility of implementing B19V detection as a differential diagnosis in patients with acute febrile illness or suspicion of arbovirus infection.

Erythrovirus B19 can occur in individuals of all age groups and from different population groups (Qiu et al., 2017), which is supported by our results (Figure 5A). The B19V infection is usually characterized as acute and self-limited, but clinical manifestations might vary according to the immunological and hematological profile of each person (Qiu et al., 2017). Some studies describe arthralgia as the most characteristic symptom of B19V infection, especially in adults (Young and Brown, 2004; Suzuki and Watari, 2023). Herein, the most common symptoms recorded were, in decreasing order of frequency, fever, myalgia, headache, and, only then, arthralgia (Figure 6). Also, the detection of B19V was significantly associated with the symptoms of "retroorbital pain", "leukopenia", "petechiae" and "malaise" (Figure 5B), thus, diverging from previous studies. However, regardless of the existence of symptoms that might be more associated with B19V infection, there is immense difficulty in distinguishing it from other exanthematous diseases due to the similarity of symptoms. The similarity of the symptoms of erythema infectiosum to other cutaneous exanthematous diseases, together with the wide circulation of arboviruses in Brazil, make the clinical diagnosis a difficult challenge to overcome (Garcia and Leon, 2021). In this sense, a study published by Alonso et al. (2003) showed that about 19% of cases of exanthematous diseases remain without a confirmatory diagnosis; among them, B19V infection is the major cause (33%). Hence, it is essential to carry out a specific laboratory diagnosis for the correct identification of the exanthematous disease agent.

In our study, ten pregnant women tested positive for B19V. Unfortunately, these women were being monitored under the initial clinical hypothesis of arboviruses and were therefore not diagnosed promptly for adequate monitoring during pregnancy. There was an attempt to track these positive pregnant women to verify the outcome of each case, but the data reported in the notification forms were out of date.



Infection by B19V during pregnancy is a serious public health problem, as abortion can occur in the first trimester of pregnancy and in the second trimester, the fetus can develop non-immune hydrops fetalis (Bouraddane et al., 2021). The infection can result in vertical transmission to the fetus, causing infection of erythroid precursors and intense hemolysis, leading to severe anemia, fetal hydrops, and death. Rapid correction of anemia by transfusion of packed red blood cells *in utero* largely prevents fetal death (Slavov et al., 2012). Women of reproductive age have an annual seroconversion rate of 1.5%, and as in the population group of pregnant women the disease tends to be more severe, so the pregnancy must be monitored medically with the detection of B19V (Koch and Sp Adler, 2023).

The identification of B19V "Genotype 1a" (Figure 7) demonstrates the importance of differential laboratory diagnosis

using molecular techniques in patients with fever and characteristic symptoms, to assist the health surveillance system through continuous monitoring to detect all circulating genotypes and the spread of the virus in the population. There are three recognized genotypes of B19V (1, 2, and 3), segregated into subtypes 1a, 1b, 2, 3a, and 3b (Servant et al., 2002; Toan et al., 2006; Parsyan et al., 2007; Conteville et al., 2023). All three genotypes have been reported in both symptomatic and asymptomatic persons, and no association has been established between genotype and clinical manifestations (Sanabani et al., 2006; Molenaar-de Backer et al., 2012; Rahiala et al., 2013; Katoh, 2020; Seetha et al., 2021; Oliveira et al., 2023). Genotype 1 is the most prevalent in the world. In Brazil, the three genotypes have already been detected, but there is also a predominance of Genotype 1 (Freitas et al., 2008; Garcia et al., 2023). The first complete genome of B19V

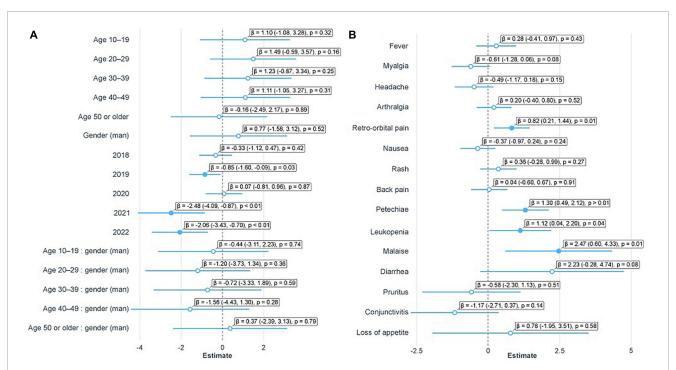
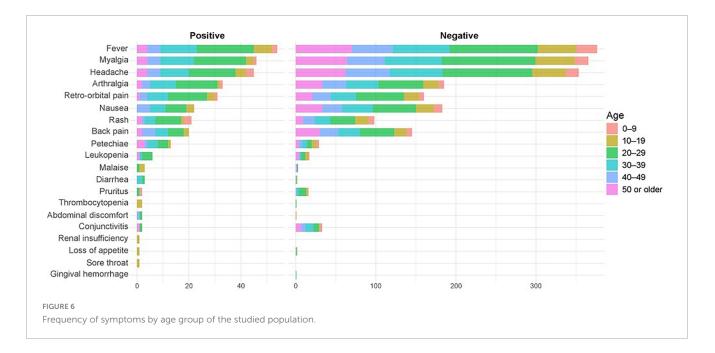


FIGURE 5

Estimates of mixed generalized linear models. (A) Estimates of patient characteristics and period. (B) Estimates of symptoms. Filled dots indicate significant variables (p < 0.05); bars represent the 95% confidence interval; β , estimate; numbers in parentheses indicate the estimated 95% confidence intervals; p, p-value.



genotype 1 a was from a serum sample suspected of dengue infection, from a fatal case of a 12-year-old boy in Rio de Janeiro, Brazil (Conteville et al., 2023). The viral genome of erythrovirus B19 is highly conserved, with 98–99% similarity between isolates (Cubel et al., 1997; Hicks et al., 2023), which means that a few sequenced samples are enough to know the circulating genotype.

An important characteristic of B19V infection in Brazil is its cyclic pattern of occurrence every 3-5 years, with years with high infection rates followed by periods with low circulation, as occurred in 1988-1989, 1995-1996, 1999-2000, 2004-2005, 2009-2010, 2013-2014 (Almada et al., 2022; Reno et al., 2022). Our results corroborate those of previous studies suggesting that B19V has a cyclical pattern of occurrence in Mato Grosso do Sul, with changes between years. Interestingly, in both 2017 and 2018, the highest average concentration occurred in the second semester, between July and September (red and green vectors). This period marks the end of winter and the beginning of spring, but a

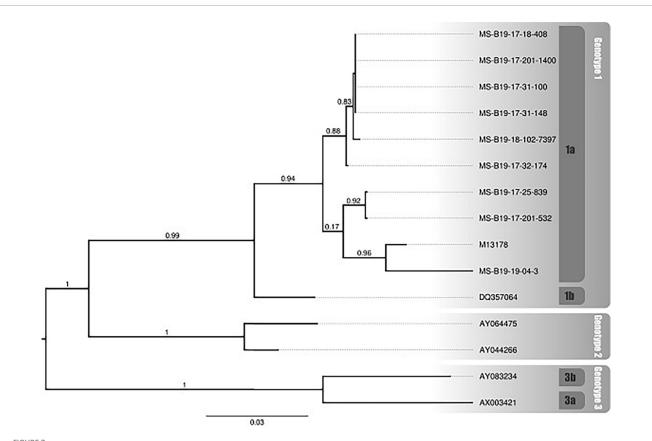


FIGURE 7

Phylogenetic relationship of sequenced samples and B19V genotypes (1a, 1b, 2, 3a and 3b). The samples sequenced in this study start with "MS-B19". The reference sequences obtained from GenBank are identified by their accession number. The numbers above branches indicate the approximate likelihood ratio test (aLRT).

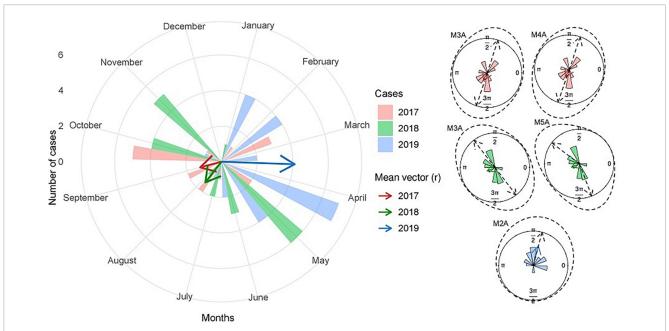


FIGURE 8

Circular histograms of several B19 cases per month for the years 2017, 2018, and 2019. In the larger histogram, the numbers indicate the number of cases in the respective month, and the mean vector (arrow) is the length and direction of the mean date of cases. The smaller histograms represent the best models found, with the average vector (arrow), the density (dashed line), and the average direction (dashed arrows).

climate-related interpretation may be misleading because of the few years sampled in this study. Therefore, further studies over a longer period are needed, as it is suggested that the cycle for B19V may last 4–5 years (Almada et al., 2022). Additionally, as 3 years of the study overlapped with the COVID-19 pandemic, this may have influenced the dynamics of the circulation and transmission of B19V. The knowledge of the virus's seasonality is important to demonstrate to clinicians that during this period, the diagnostic hypothesis for B19V should be more valued, even a period marked by widespread outbreaks of DENV, CHIKV and ZIKV.

Timely information represents an essential tool for epidemiological surveillance, as it triggers the "informationdecision-action" process, a triad that summarizes the dynamics of its activities, which, as is known, must be initiated from the information of an indication or suspected case of any disease or injury. This study highlights the fundamental importance of molecular detection to monitor and manage diseases, strengthening their surveillance and control. Although not a new concern, our results emphasize the relevance of monitoring the simultaneous presence of pathogens, such as B19V, which can significantly impact clinical management, especially in pregnant women. This points to its usefulness in diagnosing individuals with undefined disease causes, especially during periods of seasonal outbreaks of arboviral diseases that manifest with acute fever and other symptoms.

As an example, clinical and epidemiological studies carried out in the North region, in 1993, already showed that B19V was becoming a public health problem on the rise at that time (Freitas et al., 1993). The B19V is often underestimated from a clinical standpoint. Its widespread dissemination and generally benign, self-limiting clinical course itself lead to a diminished appreciation of its pathogenic potential. Our study population did not have a laboratory-confirmed diagnosis and, therefore, surveillance did not carry out control actions. In laboratory routine during suspected outbreaks of arbovirus infections, including testing for B19V as part of the differential investigation in pregnant women with fever, joint pain, and rash could accelerate the development of effective prophylactic interventions and new therapeutic options, ensuring a significant increase in in fetal survival. Further, continuous monitoring of B19V is needed to detect all known genotypes and the emergence of new genotypes for the control of cases.

Data availability statement

The authors confirm that the data supporting the findings of this study are available within the article and its Supplementary materials. The sequences of the sequenced samples were deposited in the GenBank database (www.ncbi.nlm.nih.gov/genbank/), under the accession numbers: "OR542579", "OR542583", "OR542582", "OR578525", "OR542580", "OR578526", "OR578523", "OR578524", "OR542581". The reference sequences used for phylogenetic analysis were obtained from the GenBank database, and are identified by the following accession numbers: "M13178", "DQ357064", "AY064475", "AY044266", "AY083234", "AX003421".

Ethics statement

The studies involving humans were approved by the Research Ethics Committee involving Human Subjects at the Dom Bosco Catholic University of Mato Grosso do Sul, Brazil (CAAE: 54019821.2.0000.5162). The studies were conducted in accordance with the local legislation and institutional requirements. The ethics committee/institutional review board waived the requirement of written informed consent for participation from the participants or the participants' legal guardians/next of kin because the Human Biological Material used in the research is stored and coded, in the biobank under the responsibility of the Central Laboratory of Mato Grosso do Sul (LACEN) of the State Health Department of Mato Grosso do Sul, which authorized access to the samples allowed for the proposed research and only for the purpose described in the project, respecting all guidelines and regulatory standards for research relating to human beings as set out in Resolution 466/12 and its supplements. The LACEN authorization document was submitted and approved by the CEP.

Authors contributions

GL: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Writing - original draft, Writing review and editing. ZF: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Writing - original draft, Writing review and editing, Visualization. VN: Data curation, Formal analysis, Investigation, Methodology, Writing - original draft, Writing - review and editing. DA: Data curation, Formal analysis, Investigation, Methodology, Writing - original draft, Writing - review and editing. EL: Data curation, Formal analysis, Methodology, Writing - review and editing. CC: Resources, Writing - review and editing. LD: Funding acquisition, Resources, Writing - review and editing. CG: Funding acquisition, Resources, Writing - review and editing. FN: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Resources, Writing - original draft, Writing - review and editing, Visualization. AF: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Validation, Visualization, Writing - original draft, Writing - review and editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmicb.2024. 1417434/full#supplementary-material

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