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### \*CORRESPONDENCE

Diana Bahia ⊠ dianabahia@ufmg.br; ⊠ dianabahia@gmail.com Renan P. Souza ⊠ renanpedra@gmail.com

<sup>†</sup>These authors have contributed equally to this work and share senior authorship

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# The frequency of mutations in the penA, mtrR, gyrA and parC genes of Neisseria gonorrhoeae, the presence of tetM gene and antibiotic resistance/ susceptibility: a systematic review and meta-analyses

### Ana Clara Mendes, Renan Pedra de Souza\*<sup>†</sup> and Diana Bahia\*<sup>†</sup>

Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Minas Gerais, Brazil

Gonorrhoea is currently one of the most important sexually transmitted infections (STIs) due to the increasing spread of multidrug-resistant strains of N. gonorrhoeae. The aim of this study was to analyse the association between resistance or decreased susceptibility to antibiotics in N. gonorrhoeae and the presence of mutations in the penA, mtrR, gyrA and parC genes, and the presence of tetM gene. We conducted a systematic review according to the PRISMA guidelines. We selected 19 studies for the penA gene, 23 for gyrA and parC, 18 for mtrR and 12 for tetM using the Science Direct and PubMed databases. Meta-analyses of isolates resistant to penicillin, cefixime and ceftriaxone showed that more than 50% of isolates had mutations in the penA and mtrR genes. More than 50% of azithromycin-resistant isolates had mutations in the mtrR gene, while more than 50% of ciprofloxacin-resistant and intermediate-resistant isolates had mutations in gyrA. Less than 50% of the isolates with intermediate resistance to ciprofloxacin had mutations in parC. The plasmid containing the tetM gene was found in more than 50% of tetracycline-resistant isolates. Infection surveillance and genetic studies are important for controlling the spread of the disease, which can improve the quality of life of infected people and reduce the financial burden on public health systems.

### KEYWORDS

Neisseria gonorrhoeae, antibiotic resistance, mutation, penA, mtrR, tetM, gyrA, parC gene

### **1** Introduction

Sexually transmitted infections (STIs) are one of the main global public health problems and among the main transmissible diseases, affecting the health of men, women, and infants around the world (World Health Organization, 2019b). It is estimated that, worldwide, over 1 million new STI cases occur daily (376.4 million new infections annually) due to the four currently treatable STIs: chlamydia, gonorrhoea, syphilis, and trichomoniasis (World Health Organization, 2019a). On average, approximately 1 in 25 people has at least one STI. Since the last published data, there has been no significant decline in existing or new infections (World Health Organization, 2019b).

Gonorrhoea is an STI that exclusively affects humans and is caused by the bacterium *Neisseria gonorrhoeae*, a Gram-negative diplococcus, also known as gonococcus (Quillin and

Seifert, 2018). The infection causes urethritis in males and cervicitis in females. STIs, if unidentified or untreated, can ascend in the genital tract and result in complications (Rice et al., 2017). In addition, the infection increases the rates of transmission and acquisition of HIV (Cohen et al., 1996). In 2020, the WHO estimated that there were 82.4 million cases of gonorrhoea worldwide (World Health Organization, 2024). Sexual orientation, sexual behavior, socioeconomic status, geographical location, culture, and access to sex education are directly related to the epidemiological variety of the disease (World Health Organization, 2019a,2019b).

Gonorrhoea is currently one of the most important STIs due to the increase in the spread and emergence of multidrug-resistant strains; it is also the second most common STI globally (Unemo et al., 2019). For this reason, the WHO has included the bacterium *N. gonorrhoeae* in the list of priorities for combatting transmission and researching new drugs (World Health Organization, 2017). Currently, *N. gonorrhoeae* is resistant to penicillin, tetracycline, macrolides, azithromycin, sulphonamides, quinolones and cephalosporins such as cefixime and ceftriaxone. Many cephalosporin-resistant strains are also resistant to other antibiotics, making *N. gonorrhoeae* a multidrugresistant bacterium (Unemo and Shafer, 2014; World Health Organization, 2017).

Neisseria gonorrhoeae can change its genetic material through several types of mutations, which it uses to adapt and survive in the human host. This species has evolved and acquired or developed mechanisms of resistance to almost all types of antibiotics recommended for use in treatment (Unemo and Shafer, 2014). In addition, N. gonorrhoeae is naturally competent for transformation, referring to the ability to take up and incorporate DNA, which occurs via DNA donation, DNA binding and uptake, processing and homologous recombination. Transformation occurs particularly often between N. gonorrhoeae and other Neisseria species and is an important mechanism to generate genetic diversity. It has also been shown that commensal Neisseria is a reservoir of genetic material for pathogenic Neisseria (Rotman and Seifert, 2014; Quillin and Seifert, 2018; Unemo et al., 2019). The main genes related to antibiotic resistance of N. gonorrhoeae are penA, blaTEM, mtrR, tetM, gyrA and parC (Unemo et al., 2016).

Beta-lactam antibiotics, such as penicillin and cephalosporins, inhibit peptidoglycan synthesis in the bacterial cell wall by binding the beta-lactam ring to transpeptidase enzymes called penicillin-binding proteins (PBPs) located in the periplasm. Resistance results from cumulative chromosomal mutations in genes involved in cell wall synthesis, such as *penA*. Mutations in the *penA* gene alter the PBP protein, preventing antibiotic binding. *penA* allelic mosaics are associated with reduced susceptibility to cephalosporins. These alleles in *penA* are called "mosaics" because their DNA sequence appears to have been formed by homologous recombination with DNA from other species of *Neisseria* that are naturally resistant to thirdgeneration cephalosporins (Gose et al., 2013).

Tetracyclines inhibit the binding of aminoacyl-tRNA to the mRNA-ribosome complex by binding to the 30S ribosomal subunit, thereby inhibiting protein synthesis. The *tet*M gene, carried by plasmids in *N. gonorrhoeae*, confers resistance to tetracyclines because it encodes the *tet*M protein, which binds to the ribosome and protects it from antibiotic binding, allowing the process of protein synthesis (Morse et al., 1986; Wi et al., 2017).

Quinolones/fluoroquinolones, such as ciprofloxacin, inhibit the activity of DNA gyrase and topoisomerase IV, enzymes essential for DNA replication, transcription, recombination and repair in bacteria. This class of antibiotics acts by forming a drug-enzyme-DNA complex, causing double-strand breaks in the DNA (Costa-Lourenço et al., 2017). Gonococcal resistance to ciprofloxacin is mediated by mutations in the quinolone resistance-determining region (QRDR). Mutations in the DNA gyrase enzyme (encoded by the *gyr*A gene) and topoisomerase IV (encoded by the *par*C gene) reduce the binding of the antibiotic to these enzymes and prevent its action (Belland et al., 1994; Sánchez-Busó et al., 2019; Wi et al., 2017).

Mtr (multiple transferable resistance) efflux pumps are triple efflux pumps (MtrCDE) that export a variety of antimicrobial agents such as antimicrobial peptides, antibiotics, bile salts and fatty acids. MtrCDE, whose expression is controlled by the *mtr*R repressor, is composed of inner membrane (MtrD) and outer membrane (MtrE) channels connected by a periplasmic membrane fusion lipoprotein (MtrC) (Handing et al., 2018). These efflux pumps control the concentration of drugs in the periplasm and regulate the efflux of antibiotics such as tetracyclines, penicillins, quinolones, cephalosporins and macrolides (Cousin et al., 2003; Unemo and Shafer, 2014). Mutations in *mtr*R, in the promoter or coding sequence, promote overexpression and increased efflux of the MtrCDE efflux pump, leading to resistance to these antibiotics (Costa-Lourenço et al., 2017).

In reviewing the antimicrobial resistance mechanisms of *N. gonorrhoeae*, Unemo and Shafer (2014) suggested that the world may be entering an era of intractable *N. gonorrhoeae* due to multidrug resistance. Antibiotic-resistant *N. gonorrhoeae* can appear as a silent epidemic, and the disease and its complications can cause morbidity and economic consequences, as treatment can become more expensive when complications occur.

Several studies, such as those by Lee et al., (2015) and Calado et al., (2019), evaluated the minimum inhibitory concentrations (MICs) of *N. gonorrhoeae* isolates and searched for *N. gonorrhoeae* resistance genes and resistance gene mutations of resistant and/or reduced susceptibility isolates. They also correlated the mutations or presence of resistance genes with resistance or reduced susceptibility to antibiotics. Building on these previous studies, this work involved a systematic review aimed at identifying critical articles focusing on these characteristics and selecting appropriate ones for analysis using a set of criteria. The meta-analysis performed here should deepen our understanding of gonorrhoea and help develop solutions to this urgent public health problem.

### 2 Methods

This systematic review was conducted considering the protocol proposed by PRISMA (Moher et al., 2009). The phases for the selection of studies were identification, selection, eligibility and inclusion (Table 1, see also Figure 7). The search focused on the databases PubMed and Science Direct, including all articles within them published to date. The first screening was conducted based on reading of the title and abstract, after which the eligibility was determined through complete reading of the article, with this work being conducted by two researchers independently. This review includes

### TABLE 1 Phases of the studies selection, see also Figure 7.

	Identifica	ation	Screening							
Gene	Identification from databases	Screened	Excluded	Assessed for eligibility	Excluded	Studies included in qualitative and quantitative analysis				
penA	226	62	164	19	43	19				
gyrA and parC	185	53	132	23	30	23				
mtrR	180	66	114	18	48	18				
tetM	102	26	76	12	14	12				

The gene penA (PenA family class A beta-lactamase) is related to resistance to penicillins and cephalosporins. The genes gyrA (DNA gyrase subunit A) and parC (DNA topoisomerase IV subunit A) are related to resistance to quinolones, the mtrR (multidrug efflux system transcriptional repressor) gene is related to penicillin, tetracycline, cephalosporins and macrolides, and the gene tetM (tetracycline resistance ribosomal protection protein) is related to resistance to tetracyclines.

studies on the *N. gonorrhoeae* genes *penA*, *mtrR*, *tetM*, *parC* and *gyrA*, which are related to resistance, intermediate resistance or reduced susceptibility to penicillin, cefixime, ceftriaxone, azithromycin, ciprofloxacin and tetracycline.

### 2.1 Search argument

For the PubMed search, the following search argument was used: ["Gonorrhea"(Mesh) OR "Gonorrhea"(Title/Abstract) OR "Neisseria gonorrhoeae"(Mesh) OR "Neisseria gonorrhoeae"(Title/ Abstract) OR "N. gonorrhoeae"(Title/Abstract)] AND ["Antimicrobial resistance"(Title/Abstract) OR "Antimicrobial susceptibility"(Title/Abstract) OR "Resistance Profile"(Title/ Abstract) OR "Drug Resistance, Microbial"(Mesh)] AND ["penA"(Title/Abstract)]. The argument was adapted for each gene.

For the search in the Science Direct database, the keywords "*Neisseria gonorrhoeae*, Antimicrobial resistance, *penA*, *mtrR*, *tetM*, *gyrA* and *parC*" were used. In terms of the manuscript type, the search was limited to research articles.

For the articles, no restrictions on the year or site of publication were applied, while the inclusion criteria were articles in either Portuguese or English that address susceptibility and resistance to antibiotics and mutations in the selected genes. The exclusion criteria were articles in languages other than English and Portuguese, articles about other *Neisseria* species, studies that did not analyze antibiotic susceptibility profiles, case reports, review articles, studies that analyzed the application of a technique, articles that detected susceptibility and mutations in non-clinical isolates and articles with incomplete genotypic data.

### 2.2 Meta-analysis

Meta-analysis was conducted when two or more studies were found for the same gene and the same antibiotic. For this, the metaprop function of the meta package was used in the R program (version 4.0.2). The original proportions were combined using the inverse variance method with both fixed- and random-effects models. The fixed-effects model assumes that all studies estimate the same underlying effect size, meaning that any observed differences between study results are due solely to chance. In contrast, the random-effects model assumes that the true effect size may vary between studies due to differences in study populations, methodologies or other factors, reflecting real variations in effect size. This model incorporates both within-study variance and an additional between-study variance component to account for this heterogeneity (Dettori et al., 2022). Heterogeneity across studies was assessed using the  $l^2$  statistic, which represents the percentage of total variation attributable to heterogeneity, and tau<sup>2</sup>, which quantifies the absolute between-study variance. The significance of heterogeneity was evaluated using Cochran's Q test. A significance level of 5% was set for all analyses.

### **3** Results

### 3.1 Selection of studies

The screening and selection processes are summarized in the are summarized in Table 1 and Figure 7.

### 3.2 Studies' characteristics

The studies' characteristics are summarized in Supplementary Tables S1-S6 according to the phenotypes. In general, we observed many common characteristics across studies. The earliest included studies investigating antibiotic resistance, decreased susceptibility and mutations in resistance genes were performed in 1998. Research was conducted in various countries, but particularly in East Asia, mainly China and Japan (Supplementary Tables S1-S6, column "Local"). In most of the studies, samples were collected from urethra, rectum, cervix, endocervix and pharynx; in only one study were samples collected from other sites such as eye, blood, surgical wound, gastric juice, synovial fluid and Bartholin abscess. Moreover, the results show a lack of studies in Latin America and Africa. Many papers do not provide details on clinical isolates beyond the study period (second column). Although fewer in number, some papers detail sex, sexuality and sexual behavior. In studies providing information on sex, most of the isolates were obtained from samples collected from men.

The papers also reveal which mutations were found most frequently (Supplementary Tables S7–S10). The most common mutations in the *penA* gene were A501V substitution and mosaic-type mutations. In the *mtr*R gene, deletion of A (adenine), G45D, H105Y and A39T substitutions were the most common mutations. Meanwhile, the *gyrA* gene presented the S91F and D95G substitutions and the *par*C gene the D86N substitution.

### 3.3 Meta-analysis of studies evaluating penicillin resistance

The results of the meta-analysis for the antibiotic penicillin with the proportions of mutations in the *penA* and *mtrR* genes are shown in Supplementary Figure S1. The results of the random-effects model indicate that 87% (CI 42–98%) of the analyzed isolates had mutations in the *penA* gene (Supplementary Figure 1A). Fixed-effect model results indicate that 95% (CI 86–98%) had mutations in the *mtrR* gene (Supplementary Figure 1B).

The meta-analysis results of studies on the antibiotic penicillin with the proportions of mutations in the *mtr*R gene are shown in Supplementary Figure S2. The results of the fixed model indicate that 85% (CI 71–93%) of the analyzed isolates had mutations in the *mtr*R gene.

# 3.4 Meta-analysis of studies evaluating resistance and reduced susceptibility to cefixime

Supplementary Figure S3 presents the meta-analysis results of studies on cefixime resistance and the proportions of mutations in the *mtr*R and *penA* genes. The results of the fixed-effects model indicate that 94% (CI 65–99%) of the analyzed isolates had mutations in the *mtr*R gene (Supplementary Figure 3A) and 93% (CI 77–98%) of them had mutations in the *penA* gene (Supplementary Figure 3B).

Supplementary Figure S4 presents the meta-analysis results of studies on reduced susceptibility to cefixime. The results of the randomeffects model indicate that 93% (CI 73–99%) of the analyzed isolates had mutations in the gene *penA* (Supplementary Figure 4A). Meanwhile, the results of the fixed-effects model indicate that 96% (CI 88–99%) of the isolates had mutations in the *mtr*R gene (Supplementary Figure 4B).

Supplementary Figures S5, S6 present the results of the metaanalysis of the systematic review of studies on the *mtr*R gene. Supplementary Figure S5 presents the results of the meta-analysis of studies on reduced susceptibility to the antibiotic cefixime with the proportions of mutations in the *mtr*R gene. The results of the fixedeffects model show that 96% (CI 88–99%) of the analyzed isolates had mutations in the *mtr*R gene. Supplementary Figure S6 presents the result of the meta-analysis of cefixime-resistant isolates with the proportions of mutations in the *mtr*R gene. The results of the fixedeffects model indicate that 94% (CI 65–99%) of the analyzed isolates had mutations in the *mtr*R gene.

# 3.5 Meta-analysis of studies evaluating resistance and reduced susceptibility to ceftriaxone

Supplementary Figure S7 presents the meta-analysis results on ceftriaxone resistance with the proportions of mutations in the *penA* (Supplementary Figure 7A) and *mtr*R (Supplementary Figure 7B) genes. The results of the fixed-effects model show that 93% (CI 82–97%) of the analyzed isolates had mutations in the *penA* gene (Supplementary Figure 7A), and 90% (CI 72–97%) of the isolates had mutations in the *mtr*R gene (Supplementary Figure 7B).

Supplementary Figure S8 shows the meta-analysis results of studies on reduced susceptibility to ceftriaxone with the proportions of

mutations in the *penA* (Supplementary Figure 8A) and *mtrR* (Supplementary Figure 8B) genes. The results of the random-effects models indicate that 95% (CI 85–99%) of the isolates had mutations in the *penA* gene (Supplementary Figure 8A) and 94% (CI 80–99%) of the isolates had mutations in the *mtrR* gene (Supplementary Figure 8B).

Supplementary Figures S9, S10 present the meta-analysis results of the systematic review of the *mtr*R gene. Supplementary Figure S9 presents the meta-analysis results of studies on reduced susceptibility to ceftriaxone with the proportions of mutations in the *mtr*R gene. Results from the random-effects model indicate that 71% (CI 58–82%) of the isolates had mutations in the *mtr*R gene.

Supplementary Figure S10 presents the meta-analysis results of studies on ceftriaxone resistance with the proportions of *mtr*R mutations. Fixed-effects model results show that 93% (CI 72–99%) of the isolates had mutations in the *mtr*R gene.

### 3.6 Meta-analysis of studies evaluating resistance to azithromycin

The meta-analysis results of studies of azithromycin-resistant isolates with the proportions of mutations in the *mtr*R gene are shown in Figure 1. The results of the fixed-effects model show that 95% (CI 88–98%) of the analyzed isolates had mutations in the *mtr*R gene.

# 3.7 Meta-analysis of studies evaluating resistance and intermediate resistance to ciprofloxacin

The meta-analysis results of studies on ciprofloxacin resistance with the proportions of mutations in the *gyr*A gene are shown in Figure 2. The results of the random-effects model show that 97% (95–99% CI) of the isolates had mutations in the *gyr*A gene.

Figure 3 presents the meta-analysis results of studies on ciprofloxacin resistance with the proportions of mutations in the *parC* gene. The results of the random-effects model indicate that 88% (CI 83–92%) of the analyzed isolates have mutations in the *parC* gene.

Figure 4 presents the results of the meta-analysis of studies on intermediate resistance to ciprofloxacin with the proportions of mutations in the *gyr*A gene. The results of the random-effects model indicate that 91% (CI 78–97%) of the isolates had mutations in the *gyr*A gene.

The meta-analysis results of studies on intermediate resistance to ciprofloxacin with the proportions of mutations in the *par*C gene are shown in Figure 5. Random-effects model results indicate that 29% (CI 11–59%) of the isolates had mutations in the *par*C gene.

### 3.8 Meta-analysis of studies evaluating tetracycline resistance

The presence of the plasmid containing the *tet*M gene and tetracycline resistance was also evaluated. Figure 6 presents the results of the meta-analysis of studies on the antibiotic tetracycline with the proportions of plasmids containing the *tet*M gene. Little variability and random error are observed. The random-effects

Study	Events	Total					Pro	portion	95%-CI	Weight (fixed)	Weight (random)
Belkacem, 2016	9	9			-			1.00	[0.66; 1.00]	14.3%	14.3%
Liu, 2019	50	52				-		0.96	[0.87; 1.00]	57.7%	57.7%
Zheng, 2019	13	13					-	1.00	[0.75; 1.00]	14.5%	14.5%
Maduna, 2020	4	4						1.00	[0.40; 1.00]	13.5%	13.5%
Fixed effect model Random effects mode	I	78				~		0.95 0.95	[0.88; 0.98] [0.88; 0.98]	100.0% 	 100.0%
Heterogeneity: $I^2 = 0\%$ , $\tau^2$	= 0, <i>p</i> = 0	.94	0.2	0.4	0.6		1				

### FIGURE 1

Meta-analysis of the proportion of gene mutations in azithromycin-resistant isolates (*mtr*R gene). The results are presented in forest plots. In its first column ("study") are the citations of the selected studies. The second column ("events") contains the number of isolates with mutations in the evaluated gene, while the third column ("total") shows the total number of samples analyzed in that study. The fourth column presents the proportion estimate followed by their confidence intervals. The sixth and seventh columns present, respectively, the weights for the fixed and random models of the meta-analysis.

Study	Events	Total			Proportion	95%-CI	Weight (fixed)	Weight (random)
Trees, 1999	74	74			1.00	[0.95; 1.00]	1.6%	3.5%
De Neeling, 2000	11	11			1.00	[0.72; 1.00]	1.5%	3.4%
Tanaka, 2000	14	14			1.00	[0.77; 1.00]	1.6%	3.4%
Tanaka, 2000	29	29			1.00	[0.88; 1.00]	1.6%	3.5%
Chaudhry, 2002	35	35			1.00	[0.90; 1.00]	1.6%	3.5%
Alcala, 2003	11	11			1.00	[0.72; 1.00]	1.5%	3.4%
Saika, 2004	57	57			1.00	[0.94; 1.00]	1.6%	3.5%
Yoo, 2004	123	123		-	1.00	[0.97; 1.00]	1.6%	3.5%
Vereshchagin, 2004	16	21			0.76	[0.53; 0.92]	12.2%	10.0%
Dewi, 2004	32	32			1.00	[0.89; 1.00]	1.6%	3.5%
Uthman, 2004	58	58		i i i i i i i i i i i i i i i i i i i	1.00	[0.94; 1.00]	1.6%	3.5%
Giles, 2004	42	42		÷++	1.00	[0.92; 1.00]	1.6%	3.5%
Yang, 2006	101	101			1.00	[0.96; 1.00]	1.6%	3.5%
Wang, 2006	54	54		H•	1.00	[0.93; 1.00]	1.6%	3.5%
llina, 2008	194	211			0.92	[0.87; 0.95]	50.2%	12.6%
Chen, 2010	35	35			1.00	[0.90; 1.00]	1.6%	3.5%
Allen, 2011	40	42			0.95	[0.84; 0.99]	6.1%	7.8%
Uehara, 2011	19	19			1.00	[0.82; 1.00]	1.6%	3.5%
Kulkarni, 2012	62	62		<u> </u>	1.00	[0.94; 1.00]	1.6%	3.5%
Sethi, 2013	61	61		<u> </u>	1.00	[0.94; 1.00]	1.6%	3.5%
Calado, 2019	11	11			1.00	[0.72; 1.00]	1.5%	3.4%
Kivata, 2019	20	20			1.00	[0.83; 1.00]	1.6%	3.5%
Boiko, 2019	17	17			1.00	[0.80; 1.00]	1.6%	3.5%
Fixed effect model		1140		\$	0.95	[0.93; 0.96]	100.0%	
Random effects mode	əl			\$	0.97	[0.95; 0.99]		100.0%
Heterogeneity: $I^2 = 41\%$ ,	$\tau^2 = 0.6869$	, p = 0.'02 0	0.2 0.4	06 08 1				

#### FIGURE 2

Meta-analysis of the proportion of gene mutations in isolates resistant to ciprofloxacin (*gyrA* gene). The results are presented in forest plots. In its first column ("study") are the citations of the selected studies. The second column ("events") contains the number of isolates with mutations in the evaluated gene, while the third column ("total") shows the total number of samples analyzed in that study. The fourth column presents the proportion estimate followed by their confidence intervals. The sixth and seventh columns present, respectively, the weights for the fixed and random models of the meta-analysis.

Ctudu	Evente	Total			Dronortion	0.59/ 01	Weight	Weight
Study	Events	TOLAI			Froportion	95%-01	(lixeu)	(ranuom)
Trees, 1999	66	74		- <u></u>	0.89	[0.80; 0.95]	6.8%	6.7%
De Neeling, 2000	11	11			1.00	[0.72; 1.00]	0.5%	1.7%
Tanaka, 2000	14	14			1.00	[0.77; 1.00]	0.5%	1.8%
Tanaka, 2000	29	29			1.00	[0.88; 1.00]	0.5%	1.8%
Chaudhry , 2002	35	35			1.00	[0.90; 1.00]	0.5%	1.8%
Alcala, 2003	10	11			0.91	[0.59; 1.00]	0.9%	2.8%
Saika, 2004	53	57		<u>.</u>	0.93	[0.83; 0.98]	3.5%	5.7%
Yoo, 2004	100	123			0.81	[0.73; 0.88]	17.8%	7.7%
Vereshchagin, 2004	10	21		— II	0.48	[0.26; 0.70]	5.0%	6.3%
Dewi, 2004	27	32		<u>#</u>	0.84	[0.67; 0.95]	4.0%	5.9%
Uthman, 2004	58	58		ii →	1.00	[0.94; 1.00]	0.5%	1.8%
Giles, 2004	42	42			1.00	[0.92; 1.00]	0.5%	1.8%
Yang, 2006	77	101			0.76	[0.67; 0.84]	17.4%	7.7%
Wang, 2006	41	54			0.76	[0.62; 0.87]	9.4%	7.1%
llina, 2008	194	211		·	0.92	[0.87; 0.95]	14.9%	7.6%
Chen, 2010	29	35			0.83	[0.66; 0.93]	4.7%	6.2%
Allen, 2011	39	42		<u> </u>	0.93	[0.81; 0.99]	2.7%	5.1%
Uehara, 2011	17	19			0.89	[0.67; 0.99]	1.7%	4.2%
Kulkarni, 2012	62	62			1.00	[0.94; 1.00]	0.5%	1.8%
Sethi, 2013	61	61			1.00	[0.94; 1.00]	0.5%	1.8%
Calado, 2019	11	11			1.00	[0.72; 1.00]	0.5%	1.7%
Kivata, 2019	14	20		-	0.70	[0.46; 0.88]	4.0%	5.9%
Boiko, 2019	13	17			0.76	[0.50; 0.93]	2.9%	5.3%
Fixed effect model		1140		$\diamond$	0.84	[0.81; 0.87]	100.0%	
Random effects mode	el			<u> </u>	0.88	[0.83; 0.92]		100.0%
Heterogeneity: $I^2 = 70\%$ ,	$\tau^2 = 0.5419$	, p < 0.01						
		0	0.2 0.4 0	0.6 0.8 1				

### FIGURE 3

Meta-analysis of the proportion of gene mutations in isolates resistant to ciprofloxacin (*parC* gene). The results are presented in forest plots. In its first column ("study") are the citations of the selected studies. The second column ("events") contains the number of isolates with mutations in the evaluated gene, while the third column ("total") shows the total number of samples analyzed in that study. The fourth column presents the proportion estimate followed by their confidence intervals. The sixth and seventh columns present, respectively, the weights for the fixed and random models of the meta-analysis.

Study	Events	Total					Proportion	95%-CI	Weight (fixed)	Weight (random)
Trees, 1999	160	160					1.00	[0.98; 1.00]	3.7%	8.3%
Tanaka, 2000	36	36					1.00	[0.90: 1.00]	3.7%	8.2%
Chaudhry, 2002	23	23				4	1.00	[0.85; 1.00]	3.7%	8.2%
Dewi, 2004	53	62					0.85	[0.74; 0.93]	57.7%	16.2%
Saika, 2004	103	103					1.00	[0.96; 1.00]	3.7%	8.3%
Yang, 2006	1	1 —					1.00	[0.03; 1.00]	2.8%	7.1%
Zhang, 2009	2	2					1.00	[0.16; 1.00]	3.1%	7.5%
Chen, 2010	2	2					1.00	[0.16; 1.00]	3.1%	7.5%
Allen, 2011	1	2 —					0.50	[0.01; 0.99]	3.8%	8.3%
Uehara, 2011	3	6			6		0.50	[0.12; 0.88]	11.3%	12.7%
Sethi, 2013	4	4					1.00	[0.40; 1.00]	3.4%	7.8%
Fixed effect model		401					0.87	[0.80; 0.92]	100.0%	
Random effects mode	əl					$\sim$	0.91	[0.78; 0.97]		100.0%
Heterogeneity: $I^2 = 61\%$ ,	$\tau^2 = 1.8284$	, p < 0.01								
		0	0.2	0.4	0.6	0.8 1				

### FIGURE 4

Meta-analysis of the proportion of gene mutations in isolates with intermediate resistance to ciprofloxacin (*gyrA* gene). The results are presented in forest plots. In its first column ("study") are the citations of the selected studies. The second column ("events") contains the number of isolates with mutations in the evaluated gene, while the third column ("total") shows the total number of samples analyzed in that study. The fourth column presents the proportion estimate followed by their confidence intervals. The sixth and seventh columns present, respectively, the weights for the fixed and random models of the meta-analysis.

Study	Events	Fotal						Proportion	95%-CI	Weight (fixed)	Weight (random)
Trees, 1999	5	160 🛨		1				0.03	[0.01; 0.07]	12.9%	12.3%
Tanaka, 2000	11	36		*				0.31	[0.16; 0.48]	20.3%	12.6%
Chaudhry, 2002	22	23		1			+	0.96	[0.78; 1.00]	2.5%	9.8%
Dewi, 2004	38	62		1 1 -		_		0.61	[0.48; 0.73]	39.1%	12.9%
Saika, 2004	7	103 -+	-	1				0.07	[0.03; 0.14]	17.3%	12.5%
Yang, 2006	1	1 —						1.00	[0.03; 1.00]	1.0%	7.0%
Chen, 2010	0	2 ⊷		1				0.00	[0.00; 0.84]	1.1%	7.3%
Allen, 2011	0	2 ⊷		1				0.00	[0.00; 0.84]	1.1%	7.3%
Uehara, 2011	2	6 —						0.33	[0.04; 0.78]	3.5%	10.6%
Sethi, 2013	0	4 ⊷		2				0.00	[0.00; 0.60]	1.2%	7.6%
Fixed effect model Random effects mode Heterogeneity: $I^2 = 91\%$ ,	<b>Ι</b> τ <sup>2</sup> = 3.0341,	<b>399</b> p < 0.01	-					0.29 0.29	[0.23; 0.36] [0.11; 0.59]	100.0% 	 100.0%
		0	0.2	0.4	0.6	0.8	1				

### FIGURE 5

Meta-analysis of the proportion of gene mutations in isolates with intermediate resistance to ciprofloxacin (*parC* gene). The results are presented in forest plots. In its first column ("study") are the citations of the selected studies. The second column ("events") contains the number of isolates with mutations in the evaluated gene, while the third column ("total") shows the total number of samples analyzed in that study. The fourth column presents the proportion estimate followed by their confidence intervals. The sixth and seventh columns present, respectively, the weights for the fixed and random models of the meta-analysis.



### FIGURE 6

Meta-analysis of the proportion of gene mutations in tetracycline-resistant isolates and the presence of the *tetM* gene. The results are presented in forest plots. In its first column ("study") are the citations of the selected studies. The second column ("events") contains the number of isolates with mutations in the evaluated gene, while the third column ("total") shows the total number of samples analyzed in that study. The fourth column presents the proportion estimate followed by their confidence intervals. The sixth and seventh columns present, respectively, the weights for the fixed and random models of the meta-analysis.

model results indicate that 98% of the analyzed isolates had the *tet*M gene.

### **4** Discussion

The present study estimated the frequency of mutations in the genes of *N. gonorrhoeae* isolates conferring resistance to the antibiotics penicillin, cefixime, ceftriaxone, azithromycin and ciprofloxacin, and

the presence of the *tetM* gene conferring resistance to tetracycline. As shown by the results obtained in this study, researchers in several countries have studied mechanisms of resistance of *N. gonorrhoeae* to antibiotics. It is evident that *N. gonorrhoeae* has developed or acquired resistance to several classes of antibiotics. Here, it was possible to determine the existence of resistance and/or reduced susceptibility to penicillin, ceftriaxone, cefixime, tetracycline, azithromycin and ciprofloxacin and show that mutations in genes encoding target proteins of these antibiotics are present in more than 50% of the



isolates. Cases with elevated MICs for third-generation cephalosporins have been reported worldwide. These isolates are of concern, as these antibiotics are the last option for treating gonorrhoea.

The results of meta-analyses of the *penA* and *mtrR* genes revealed that there is a relationship between reduced susceptibility and/or resistance to penicillin, ceftriaxone and cefixime and mutations in these genes in different countries over time. In addition, the meta-analyses of studies on the *gyrA* and *parC* genes showed that mutations in these genes are involved in ciprofloxacin resistance. Moreover, the presence of a plasmid containing the *tetM* gene is related to tetracycline resistance, as shown by the meta-analysis of studies on this gene. The results also showed that mutations in the *mtrR* gene are involved in the development of resistance to cefixime, ceftriaxone and azithromycin. The relationship between the presence of mutations and antibiotic resistance was supported by the presence of mutations in more than 50% of isolates with resistance, intermediate resistance or reduced susceptibility.

The studies also evaluated the main mutations found (Supplementary materials). The most common mutations in *penA* contribute to resistance to  $\beta$ -lactams. The A501V, A501T and G545S substitutions may increase the MICs across the spectrum of cephalosporins (Unemo and Golparian, 2019). Mosaic-like structures in PBP2 were found in several isolates (Table 1 and 2, column 3). Meanwhile, Takahata et al. (2006) found that the presence of mosaicism in PBP2 was strongly associated with reduced susceptibility to cefixime and other cephalosporins. Commensal *Neisseria* are reservoirs of resistance genes and are linked to the increase in *N. gonorrhoeae* resistant isolates. Takahata et al. (2006) proposed that horizontal transfer of *penA* genes among *Neisseria* resulted in allelic mosaicism in *N. gonorrhoeae* and *N. meningitidis*. However, some substitutions such as G545S are the result of selective pressure by antibiotics.

Mutations in *mtr*R, such as the deletion of alanine (A) in the 13 bp inverted repeat sequence in the promoter region and more common

substitutions such as G45D in the coding region, were shown to cause the overexpression of MtrCDE efflux pumps and the increase in efflux. These were in turn revealed to be related to resistance to penicillin and azithromycin and reduced susceptibility to cefixime and ceftriaxone (Costa-Lourenço et al., 2017). The efflux pump system is one of the essential factors behind multi-drug resistance (Tanaka et al., 2004). The presence of other mutations such as in *mtr*R may be necessary for the high MIC level of ceftriaxone (Jamaludin et al., 2019).

Meta-analysis results of the *gyr*A and *par*C genes showed that there is a higher proportion of mutations in *gyr*A than in *par*C, mainly in isolates with intermediate resistance. Costa-Lourenço et al. (2017) suggested that mutations in *gyr*A are more important for resistance to the quinolone ciprofloxacin as, in most cases, there is no mutation only in *par*C. In addition, Yang et al. (2006) suggested that mutations in *gyr*A determine the resistance in *N. gonorrhoeae*, while mutations in *par*C are related to a high level of resistance. The most common mutations in *gyr*A, such as S91F and D95N substitutions, decreased the binding of fluoroquinolones to the DNA gyrase subcomponent, while changes in *par*C such as D86N and S88P substitutions decreased the binding of fluoroquinolones to the topoisomerase IV subcomponents (Unemo and Shafer, 2014).

One of the characteristics of the studies that drew attention is the greater number of samples collected from male individuals in the studies that provide information on the sex of the subjects. Given that men are generally more symptomatic than women (Quillin and Seifert, 2018), the demand for care by men is greater, which explains the greater amount of information related to this group. Data on sex, sexual behavior and age may be relevant given that there is a higher incidence of gonorrhoea cases in particular populations, such as men who have sex with men, sex workers and young adults. Highlighting these data may be important to achieve a more complete analysis of the epidemiology of infection. However, few studies have reported this information. The incompleteness of the data makes it difficult to assess the quality of studies in the literature. The STROBE (Strengthening the reporting of observational studies in epidemiology) statement (STROBE, 2022) has recommendations that can improve the quality of epidemiological research reports.

Notably, there is an abundance of studies from Asia, such as China and Japan, but limited research from Latin America and Africa. The lack of Latin American and African epidemiological studies makes it difficult to monitor the infection and the advance of antibiotic resistance in these regions, while also affecting the clinical management of infections there.

The results also demonstrate that the relevant mutations are consistent over time and between different countries and regions. These epidemiological analyses are essential for the control of gonorrhoea, as measures to prevent the spread can be focused on the populations of interest. Studies on mutations and their connection with antibiotic resistance are essential for monitoring the infection.

Several countries reported lower effectiveness of azithromycin and ceftriaxone. Given that *N. gonorrhoeae* is naturally competent for transformation, as with other *Neisseria*, monitoring resistant isolates and adequate clinical management for gonorrhoea are essential. In addition, inadequate clinical management for other bacterial infections also accelerates the increase in resistance, favoring the transfer of resistance genes among bacteria. The emergence and spread of antibiotic-resistant *N. gonorrhoeae* may result in irreversible intractable gonorrhoea.

### **5** Conclusion

The development of resistance of gonorrhoea to various antibiotics has led to changes in treatment over the years. Genetic studies, which identify the mechanisms of resistance, and studies on pathology, evolution and epidemiology are necessary to establish adequate treatment for this disease. Moreover, such studies in combination with the monitoring of transmission and increased resistance may even contribute to the development of new antibiotics. Furthermore, until an effective treatment or vaccine for gonorrhoea is developed, activities to contain its spread and slow its rise of resistance are essential. These activities include prevention, early diagnosis, epidemiological monitoring, adequate treatment and awareness programs. The results of this study highlight the urgency of continuously monitoring these resistance genes. Putting these measures into practice will positively affect the quality of life of infected individuals and reduce the financial burden on healthcare systems.

### Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary material.

### Author contributions

AM: Formal analysis, Writing – original draft, Writing – review & editing, Data curation, Validation. RS: Data curation, Formal analysis, Validation, Writing – review & editing, Conceptualization, Methodology, Resources, Software, Supervision, Visualization. DB: Conceptualization, Formal analysis, Supervision, Visualization, Funding acquisition, Investigation, Project administration, Writing – original draft, Writing – review & editing.

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### **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

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### References

Alcalá, L., Arreaza, C., Salcedo, I., Antolín, N., Borrell, J., Cacho, C., et al. (2003). Molecular characterization of ciprofloxacin resistance of gonococcal strains in Spain. *Sex. Transm. Dis.* 30, 395–398. doi: 10.1097/00007435-200305000-00004

Allen, V. G., Farrell, D. J., Rebbapragada, A., Tan, J., Tijet, N., Perusini, S. J., et al. (2011). Molecular analysis of antimicrobial resistance mechanisms in *Neisseria* gonorrhoeae isolates from Ontario, Canada. *Antimicrob. Agents Chemother.* 55, 703–712. doi: 10.1128/AAC.00788-10

Belkacem, A., Jacquier, H., Goubard, A., Mougari, F., La Ruche, G., Patey, O., et al. (2016). Molecular epidemiology and mechanisms of resistance of azithromycin-resistant *Neisseria gonorrhoeae* isolated in France during 2013-14. *J. Antimicrob. Chemother.* 71, 2471–2478. doi: 10.1093/jac/dkw182

Belland, R. J., Morrison, S. G., Ison, C., and Huang, W. M. (1994). Neisseria gonorrhoeae acquires mutations in analogous regions of gyrA and parC in fluoroquinolone-resistant isolates. *Mol. Microbiol.* 14, 371–380. doi: 10.1111/j.1365-2958.1994.tb01297.x

Boiko, I., Golparian, D., Jacobsson, S., Krynytska, I., Frankenberg, A., Shevchenko, T., et al. (2020). Genomic epidemiology and antimicrobial resistance determinants of *Neisseria gonorrhoeae* isolates from Ukraine, 2013–2018. *Apmis* 128, 465–475. doi: 10.1111/apm.13060

Calado, J., Castro, R., Lopes, Â., Campos, M. J., Rocha, M., and Pereira, F. (2019). Antimicrobial resistance and molecular characteristics of *Neisseria gonorrhoeae* isolates from men who have sex with men. *Int. J. Infect. Dis.* 79, 116–122. doi: 10.1016/j. ijid.2018.10.030

Chalkley, L. J., Janse Van Rensburg, M. N., Matthee, P. C., Ison, C. A., and Botha, P. L. (1997). Plasmid analysis of *Neisseria gonorrhoeae* isolates and dissemination of tetM genes in southern Africa 1993-1995. *J. Antimicrob. Chemother.* 40, 817–822. doi: 10.1093/jac/40.6.817

Chaudhry, U., Ray, K., Bala, M., and Saluja, D. (2002). Mutation patterns in gyrA and parC genes of ciprofloxacin resistant isolates of *Neisseria gonorrhoeae* from India. *Sex. Transm. Infect.* 78, 440–444. doi: 10.1136/sti.78.6.440

Chen, P. L., Lee, H. C., Yan, J. J., Hsieh, Y. H., Lee, N. Y., Ko, N. Y., et al. (2010). High Prevalence of Mutations in Quinolone-resistance-determining Regions and mtrR Loci in Polyclonal *Neisseria gonorrhoeae* Isolates at a Tertiary Hospital in Southern Taiwan. *J. Formos. Med. Assoc.* 109, 120–127. doi: 10.1016/S0929-6646(10)60032-0

Cohen, M. S., Hoffman, I. F., Royce, R. A., Kazembe, P., Dyer, J. R., Daly, C. C., et al. (1996). Reduction of concentration of HIV-1 in semen after treatment of urethritis: Implications for prevention of sexual transmission of HIV-1. *The Lancet* 349, 1868–1873. doi: 10.1016/S0140-6736(97)02190-9

Costa-Lourenço, A. P. R., Barros dos Santos, K. T., Moreira, B. M., Fracalanzza, S. E. L., and Bonelli, R. R. (2017). Antimicrobial resistance in Neisseria gonorrhoeae: History, molecular mechanisms, and epidemiological aspects of an emerging global threat. *Braz. J. Microbiol.* 48, 617–628. doi: 10.1016/j.bjm.2017.06.001

Cousin, S. L., Whittington, W. L. H., and Roberts, M. C. (2003). Acquired macrolide resistance genes and the 1 bp deletion in the mtrR promoter in Neisseria gonorrhoeae. *J. Antimicrob. Chemother*. 51, 131–133. doi: 10.1093/jac/dkg040

De Neeling, A. J., Van Santen-Verheuvel, M., Spaargaren, J., and Willems, R. J. L. (2000). Antimicrobial resistance of *Neisseria gonorrhoeae* and emerging ciprofloxacin resistance in The Netherlands, 1991 to 1998. *Antimicrob. Agents Chemother.* 44, 3184–3185. doi: 10.1128/AAC.44.11.3184-3185.2000

Dettori, J. R., Norvell, D. C., and Chapman, J. R. (2022). Fixed-effect vs random-effects models for meta-analysis: 3 points to consider. *Global Spine J.* 12, 1624–1626. doi: 10.1177/21925682221110527

Dewi, B. E., Akira, S., Hayashi, H., and Ba-thein, W. (2004). High occurrence of simultaneous mutations in target enzymes and MtrRCDE efflux system in quinolone-resistant *Neisseria gonorrhoeae. Sex. Transm. Dis.* 31, 353–359. doi: 10.1097/00007435-200406000-00007

Dillon, J. A. R., Li, H., Sealy, J., Ruben, M., Prabhakar, P., Swantson, W. H., et al. (2001). Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from three Caribbean reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

### Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmicb.2024.1414330/ full#supplementary-material

countries: Trinidad, Guyana, and St. Vincent. Sex. Transm. Dis. 28, 508-514. doi: 10.1097/00007435-200109000-00006

Dyck, E., Alary, M., Guédou, A., Abdellati, S., Lafia, E., and Anagonou, S. (2001). Antimicrobial susceptibilities and plasmid patterns of *Neisseria gonorrhoeae* in Bénin. *Int. J. STD. AIDS.* 12, 89–93. doi: 10.1258/0956462011916848

Giles, J. A., Falconio, J., Yuenger, J. D., Zenilman, J. M., Dan, M., and Bash, M. C. (2004). Quinolone resistance-determining region mutations and por type of *Neisseria* gonorrhoeae isolates: Resistance surveillance and typing by molecular methodologies. *J. Infect. Dis.* 189, 2085–2093. doi: 10.1086/386312

Gose, S., Nguyen, D., Lowenberg, D., Samuel, M., Bauer, H., and Pandori, M. (2013). Neisseria gonorrhoeae and extended-spectrum cephalosporins in California: Surveillance and molecular detection of mosaic penA. *BMC Infect. Dis.* 13, 1–9. doi: 10.1186/1471-2334-13-570

Handing, J. W., Ragland, S. A., Bharathan, U. V., and Criss, A. K. (2018). The MtrCDE efflux pump contributes to survival of Neisseria gonorrhoeae from human neutrophils and their antimicrobial components. *Front. Microbiol.* 9:2688. doi: 10.3389/fmicb.2018.02688

Ilina, E. N., Vereshchagin, V. A., Borovskaya, A. D., Malakhova, M. V., Sidorenko, S. V., Al-Khafaji, N. C., et al. (2008). Relation between genetic markers of drug resistance and susceptibility profile of clinical *Neisseria gonorrhoeae* strains. *Antimicrob. Agents Chemother.* 52, 2175–2182. doi: 10.1128/AAC.01420-07

Jamaludin, N., Gedye, K., Collins-Emerson, J., Benschop, J., and Nulsen, M. (2019). Phenotypic and genotypic characterization of Neisseria gonorrhoeae isolates from New Zealand with reduced susceptibility to ceftriaxone. *Microbial Drug Resist.* 25, 1003–1011. doi: 10.1089/mdr.2018.0111

Karim, S., Bouchikhi, C., Banani, A., El Fatemi, H., Souho, T., Erraghay, S., et al. (2018). Molecular antimicrobial resistance of *Neisseria gonorrhoeae* in a moroccan area. *Infect. Dis. Obstet. Gynecol.* 2018:7263849. doi: 10.1155/2018/7263849

Kivata, M. W., Mbuchi, M., Eyase, F. L., Bulimo, W. D., Kyanya, C. K., Oundo, V., et al. (2019). GyrA and parC mutations in fluoroquinolone-resistant *Neisseria* gonorrhoeae isolates from Kenya. *BMC Microbiol.* 19, 1–9. doi: 10.1186/ s12866-019-1439-1

Kivata, M. W., Mbuchi, M., Eyase, F. L., Bulimo, W. D., Kyanya, C. K., Oundo, V., et al. (2020). Plasmid mediated penicillin and tetracycline resistance among *Neisseria* gonorrhoeae isolates from Kenya. *BMC Infect. Dis.* 20, 1–11. doi: 10.1186/s12879-020-05398-5

Kulkarni, S., Bala, M., Sane, S., Pandey, S., Bhattacharya, J., and Risbud, A. (2012). Mutations in the gyrA and parC genes of quinolone-resistant *Neisseria gonorrhoeae* isolates in India. *Int. J. Antimicrob. Agents* 40, 549–553. doi: 10.1016/j. ijantimicag.2012.08.007

Lee, H., Unemo, M., Kim, H. J., Seo, Y., Lee, K., and Chong, Y. (2015). Emergence of decreased susceptibility and resistance to extended-spectrum cephalosporins in Neisseria gonorrhoeae in Korea. *J. Antimicrob. Chemother.* 70, 2536–2542. doi: 10.1093/jac/dk146

Liu, Y., Wang, Y., Liao, C., and Hsueh, P. (2019). Emergence and Spread of *Neisseria* gonorrhoeae Strains with High-Level Resistance to Azithromycin in Taiwan from 2001 to 2018. *Antimicrob. Agents Chemother.* 63, 1–8. doi: 10.1128/AAC.00773-19

Maduna, L. D., Kock, M. M., van der Veer, B. M. J. W., Radebe, O., McIntyre, J., van Alphen, L. B., et al. (2020). Antimicrobial resistance of *Neisseria gonorrhoeae* isolates from high-risk men in Johannesburg, South Africa. *Antimicrob. Agents Chemother.* 64, 1–34. doi: 10.1128/AAC.00906-20

Márquez, C. M., Dillon, J. A. R., Rodriguez, V., and Borthagaray, G. (2002). Detection of a novel tet M determinant in tetracycline-resistant *Neisseria gonorrhoeae* from Uruguay, 1996-1999. *Sex. Transm. Dis.* 29, 792–797. doi: 10.1097/00007435-200212000-00010

Moher, D., Liberati, A., Tetzlaff, J., Altman, D. G., Altman, D., Antes, G., et al. (2009). Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. *PLoS Med.* 6. doi: 10.1371/journal.pmed.1000097 Morse, S. A., Johnson, S. R., Biddle, J. W., and Roberts, M. C. (1986). High-level tetracycline resistance in Neisseria gonorrhoeae is the result of acquisition of streptococcal tetM determinant. *Antimicrob. Agents Chemother.* 30, 664–670. doi: 10.1128/aac.30.5.664

Quillin, S. J., and Seifert, H. S. (2018). Neisseria gonorrhoeae host adaptation and pathogenesis. Nat Rev Microbiol. 16, 226-240. doi: 10.1038/nrmicro.2017.169

Rambaran, S., Naidoo, K., Dookie, N., Moodley, P., and Sturm, A. W. (2019). Resistance profile of *Neisseria gonorrhoeae* in KwaZulu-Natal, South Africa questioning the effect of the currently advocated dual therapy. *Sex. Transm. Dis.* 46, 266–270.

Rice, P. A., Shafer, W. M., Ram, S., and Jerse, A. (2017). Neisseria gonorrhoeae: Drug resistance, mouse models, and vaccine development. *Annu. Rev. Microbiol.* 71, 665–686. doi: 10.1146/annurev-micro-090816-093530

Rotman, E., and Seifert, H. S. (2014). The genetics of Neisseria species. *Annu. Rev. Genet.* 405–431. doi: 10.1146/annurev-genet-120213-092007

Saika, T., Kobayashi, I., and Inoue, M. (2004). A comparison of the microbiological characteristics of *Neisseria gonorrhoeae* isolated from male and female patients with gonorrhea. *Chemotherapy*. 50, 92–97. doi: 10.1159/000077809

Sánchez-Busó, L., Golparian, D., Corander, J., Grad, Y. H., Ohnishi, M., Flemming, R., et al. (2019). The impact of antimicrobials on gonococcal evolution. *Nat. Microbiol.* 4, 1941–1950. doi: 10.1038/s41564-019-0501-y

Sethi, S., Golparian, D., Bala, M., Dorji, D., Ibrahim, M., Jabeen, K., et al. (2013). Antimicrobial susceptibility and genetic characteristics of *Neisseria gonorrhoeae* isolates from India, Pakistan and Bhutan in 2007-2011. *BMC Infect. Dis.* 13:1.

Shaskolskiy, B., Dementieva, E., Kandinov, I., Filippova, M., Petrova, N., Plakhova, X., et al. (2019). Resistance of *Neisseria gonorrhoeae* isolates to beta-lactam antibiotics (benzylpenicillin and ceftriaxone) in Russia, 2015-2017. *PLoS One.* 14:e0220339. doi: 10.1371/journal.pone.0220339

STROBE (2022). What is STROBE? STROBE - Strengthening the Reporting of Observational Studies in Epidemiology, pp. 1–3. Available at: https://www.strobe-statement.org/.

Su, X., Jiang, F., Qimuge, X., Dai, X., Sun, H., and Ye, S. (2007). Surveillance of antimicrobial susceptibilities in *Neisseria gonorrhoeae* in Nanjing, China, 1999-2006. *Sex. Transm. Dis.* 34, 995–999. doi: 10.1097/OLQ.0b013e3180ca8f24

Takahata, S., Senju, N., Osaki, Y., Yoshida, T., and Ida, T. (2006). Amino acid substitutions in mosaic penicillin-binding protein 2 associated with reduced susceptibility to cefixime in clinical isolates of Neisseria gonorrhoeae. *Antimicrob. Agents Chemother.* 50, 3638–3645. doi: 10.1128/AAC.00626-06

Tanaka, M., Nakayama, H., Haraoka, M., Saika, T., Kobayashi, I., and Naito, S. (2000). Susceptibilities of *Neisseria gonorrhoeae* isolates containing amino acid substitutions in GyrA, with or without substitutions in ParC, to newer fluoroquinolones and other antibiotics. *Antimicrob. Agents Chemother.* 44, 192–195. doi: 10.1128/ AAC.44.1.192-195.2000

Tanaka, M., Nakayama, H., Notomi, T., Irie, S. I., Tsunoda, Y., Okadome, A., et al. (2004). Antimicrobial resistance of Neisseria gonorrhoeae in Japan, 1993–2002: Continuous increasing of ciprofloxacin-resistant isolates. *Int. J. Antimicrob. Agents.* 24, 15–22. doi: 10.1016/j.ijantimicag.2004.02.005

Trees, D. L., Sandul, A. L., Peto-Mesola, V., Aplasca, M. R., Bun Leng, H., Whittington, W. L., et al. (1999). Alterations within the quinolone resistance-determining regions of GyrA and ParC of *Neisseria gonorrhoeae* isolated in the Far East and the United States. *Int. J. Antimicrob. Agents.* 12, 325–332. doi: 10.1016/S0924-8579(99)00081-3

Uehara, A. A., Amorin, E. L. T., Ferreira, M. D. F., Andrade, C. F., Clementino, M. B. M., De Filippis, I., et al. (2011). Molecular characterization of quinolone-resistant *Neisseria gonorrhoeae* isolates from Brazil. *J. Clin. Microbiol.* 49, 4208–4212. doi: 10.1128/JCM.01175-11

Unemo, M., and Golparian, E. (2019). Antimicrobial resistance in Neisseria gonorrhoeae and treatment of gonorrhea. In *Antimicrobial Resistance in Bacteria from Livestock and Companion Animals* (pp. 37–58). doi: 10.1007/978-1-4939-9496-0

Unemo, M., and Shafer, W. M. (2014). Antimicrobial resistance in Neisseria gonorrhoeae in the 21st century: Past, evolution, and future. *Clin. Microbiol. Rev.* 27, 587–613. doi: 10.1128/CMR.00010-14

Unemo, M., del Rio, C., and Shafer, W. M. (2016). Antimicrobial resistance expressed by Neisseria gonorrhoeae: A major global public health problem in the 21st century. *Emerg. Infect. Dis.* 10, 213–237. doi: 10.1128/microbiolspec.ei10-0009-2015

Unemo, M., Seifert, H. S., Hook, E. W., Hawkes, S., Ndowa, F., and Dillon, J. A. R. (2019). Gonorrhoea. *Nat. Rev. Dis. Primers*. 5. doi: 10.1038/s41572-019-0128-6

Uthman, A., Heller-Vitouch, C., Stary, A., Bilina, A., Kuchinka-Koch, A., Söltz-Szöts, J., et al. (2004). High-frequency of quinolone-resistant *Neisseria gonorrhoeae* in Austria with a common pattern of triple mutations in GyrA and ParC genes. *Sex. Transm. Dis.* 31, 616–618. doi: 10.1097/01.olq.0000140019.18390.28

Vereshchagin, V. A., Ilina, E. N., Malakhova, M. V., Zubkov, M. M., Sidorenko, S. V., Kubanova, A. A., et al. (2004). Fluoroquinolone-resistant *Neisseria gonorrhoeae* isolates from Russia: Molecular mechanisms implicated. *J. Antimicrob. Chemother.* 53, 653–656. doi: 10.1093/jac/dkh145

Wang, B., Xu, J. S., Wang, C. X., Mi, Z. H., Pu, Y. P., Hui, M., et al. (2006). Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolated in Jiangsu Province, China, with a focus on fluoroquinolone resistance. *J. Med. Microbiol.* 55, 1251–1255. doi: 10.1099/jmm.0.46401-0

Wi, T., Lahra, M. M., Ndowa, F., Bala, M., Dillon, J.-A. R., Ramon-Pardo, P., et al. (2017). Antimicrobial resistance in Neisseria gonorrhoeae: Global surveillance and a call for international collaborative action. *PLoS Med.* 14, 1–16. doi: 10.1371/journal. pmed.1002344

World Health Organization (2017). WHO publishes list of bacteria for which new antibiotics are urgently needed. *Saudi Med. J.* 38, 444–445. Available at: https://www.who.int/news/item/27-02-2017-who-publishes-list-of-bacteria-for-which-new-antibiotics-are-urgently-needed.

World Health Organization (2024). Multi-drug resistant gonorrhoea. Available at: https://www.who.int/news-room/fact-sheets/detail/multi-drug-resistant-gonorrhoea.

World Health Organization (2019a). More than 1 million new curable sexually transmitted infections every day. *News release*. https://www.who.int/news-room/detail/06-06-2019-more-than-1-million-new-curable-sexually-transmitted-infections-every-day [accessed 10 March 2019].

World Health Organization. (2019b). Sexually transmitted infections (STIs) Prevention of STIs. https://www.who.int/en/news-room/fact-sheets/detail/sexually-transmitted-infections [accessed 10 March 2019].

Yang, Y., Liao, M., Gu, W. M., Bell, K., Wu, L., Eng, N. F., et al. (2006). Antimicrobial susceptibility and molecular determinants of quinolone resistance in *Neisseria gonorrhoeae* isolates from Shanghai. *J. Antimicrob. Chemother.* 58, 868–872. doi: 10.1093/jac/dkl301

Yoo, J., Yoo, C., Cho, Y., Park, H., Oh, H. B., and Seong, W. K. (2004). Antimicrobial Resistance Patterns (1999-2002) and Characterization of Ciprofloxacin-Resistant *Neisseria gonorrhoeae* in Korea. *Sex. Transm. Dis.* 31, 305–310. doi: 10.1097/01. OLQ.0000123650.98303.EB

Zhang, T., Zhou, X., Chen, Y., Gu, W., Zhang, T., and Jiang, Q. (2009). Fluoroquinolone resistance and mutation patterns in gyrA and parC genes in *Neisseria gonorrhoeae* isolates from Shanghai, China. *J. Huazhong Univ. Sci. Technol. - Med. Sci.* 29, 29–34. doi: 10.1007/s11596-009-0106-4

Zheng, Z., Liu, L., Shen, X., Yu, J., Chen, L., Zhan, L., et al. (2019). Antimicrobial resistance and molecular characteristics among *Neisseria gonorrhoeae* clinical isolates in a Chinese tertiary hospital. *Infect. Drug Resist.* 12, 3301–3309. doi: 10.2147/IDR.S221109