Check for updates

OPEN ACCESS

EDITED AND REVIEWED BY Axel Cloeckaert, Institut National de recherche pour l'agriculture, l'alimentation et l'environnement (INRAE), France

*CORRESPONDENCE Svetlana Khaiboullina Sv.khaiboullina@gmail.com

RECEIVED 25 March 2023 ACCEPTED 10 April 2023 PUBLISHED 04 May 2023

CITATION

Foster T and Khaiboullina S (2023) Editorial: Community series - innovative approaches in diagnosis of emerging/re-emerging infectious diseases, volume II. *Front. Microbiol.* 14:1193841. doi: 10.3389/fmicb.2023.1193841

COPYRIGHT

© 2023 Foster and Khaiboullina. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Community series innovative approaches in diagnosis of emerging/re-emerging infectious diseases, volume II

Toshana Foster¹ and Svetlana Khaiboullina^{2*}

¹Faculty of Medicine and Health Sciences, School of Veterinary Medicine and Science, Wolfson Centre for Global Virus Research, The University of Nottingham, Loughborough, United Kingdom, ²Department of Microbiology, University of Nevada, Reno, Reno, NV, United States

KEYWORDS

emerging infection, diagnostics, innovative approaches, rapid diagnostics, innovative methods, protozoan, virus, fungus

Editorial on the Research Topic

Community series - innovative approaches in diagnosis of emerging/re-emerging infectious diseases, volume II

Emerging and re-emerging infections remain a rising burden on public health and global economies. Several outbreaks of emerging infectious diseases and notably, the COVID-19 pandemic have had significant impacts throughout the 21st century. Across multiple countries, each infection outbreak made a remarkable influence on population health and healthcare facilities, despite recent improved medical and diagnostic advances in countering disease epidemics: a high mortality rate was characteristic of the Ebola virus disease epidemic in 2012–2016 (Li et al., 2016; Forna et al., 2020), the teratogenic effects of Zika virus (ZIKV) were reported during the outbreak in French Polynesia in 2013–2014 and Brazil in 2015–2016 (Gilbert et al., 2023), and the highly contagious nature of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) contributed to its rapid spread and the global pandemic in 2020 (Mohapatra et al., 2020).

The risk of emerging and re-emerging pathogens with epidemic and pandemic potential is expected to remain high (Baker et al., 2022). The rapid spread of these pathogens is driven by the variable dynamic balance/imbalance between the pathogen, host and environment, influenced by demographic change as a catalyst for increased transmission dynamics, changes in reservoir species range and density due to climate change particularly for vector-borne disease spread (Brashares et al., 2004; Viboud et al., 2006; McDermott, 2022), drug resistance and declining immunity due to the aging global population (Schaible and Kaufmann, 2007; Dropulic and Lederman, 2016; Harpaz et al., 2016; Corey et al., 2022). Understanding the drivers and contributors of disease spread and pathogenesis is important when designing successful measures of control. Given that emerging and re-emerging and diseases are characterized by acute onset and rapid development of symptoms, early and reliable diagnosis of infection is essential to reduce mortality and to select the most effective treatment strategy. The fast development of ultra-high throughput systems for detecting SARS-CoV-2 (Biosearchtech, 2022) and the development of the point-of-care test that allows healthcare providers to reduce the time for diagnosis (Song et al., 2021) are a reflection of the recent, rapid diagnostics advances made that can influence outbreak dynamics.

The aim of this Research Topic was to facilitate the sharing of innovative approaches in diagnosing emerging and re-emerging diseases. We focused on highlighting the importance and urgency of developing novel, rapid and more accurate diagnostic tools. The scope of the four original articles published under this topic included research on identifying diagnostic targets for fungal, bacterial, and viral pathogens.

The original article by Hsiao et al. elucidate the universal optimal cut-off values used in Aspergillus-specific antibody tests as diagnostic tools for the three classified types of aspergillosis, allergic bronchopulmonary aspergillosis (ABPA), chronic pulmonary aspergillosis (CPA), and invasive aspergillosis (IA). Aspergillus infection is frequently diagnosed in immunocompromised individuals (Gletsou et al., 2018). In immunocompromised patients, the disease could be in an invasive form with fatality rates as high as 50-85% and that can remain high even after specific treatment (Lin et al., 2001; Lowes et al., 2017). Therefore, early diagnosis of the disease could improve patient survival, where currently a single test does not exist to definitively confirm Aspergillus infection. The diagnosis of CPA is based on detecting fungus-specific IgG (Shin et al., 2014). However, substantial differences in anti-Aspergillus antibodies based on ethnicity, geographic location, and frequency of exposure have been reported (Rayens et al., 2022). This makes it challenging to establish a biological reference range for aspergillosis and thus the universal cut-off values for Aspergillus antibody tests. In the current study, Hsiao et al. aimed to develop the cut-off values for Aspergillusspecific antibodies in the Taiwanese population and compare and report on geographic variations. The authors collected serum samples from 118 controls and 128 patients with IA, CPA, and ABPA between June 2018 to September 2021. All serum samples were probed for Aspergillus (A.) fumigatus and A. niger-specific IgG and IgE using ImmunoCAP technology. Optimal cut-offs were determined using receiver operating characteristic curve (ROC) analysis.

The authors found that CPA patients had the highest *A. fumigatus*-specific IgG levels compared to other forms of aspergillosis. In contrast, patients with ABPA had the highest *A. fumigatus*-specific IgE and *A. niger*-specific IgG and IgE serum levels compared to IA, with *A. niger*-specific IgE demonstrated in this study to be a potential a diagnostic tool for ABPA. Based on collected data, the authors determined the optimal cut-off of *A. fumigatus* and *A. niger*-specific IgG for patients with CPA and ABPA in the Taiwanese population. The authors also described that the cut-off differences for IgG antibodies to *A. fumigatus* were higher in patients living in different eco-climatic zones. These findings infer that geographic variations may affect antibody levels in patients and suggest that further work is needed to establish reference ranges across affected countries.

The genotyping of *Chlamydia (Ch.) trachomatis* (CT) and the spatiotemporal spread of lymphogranuloma venereum (LGV) in Madrid were the focus of the original manuscript by Martinez-Garcia et al. LGV, which clinically commonly presents with inguinal femoral lymphadenopathy, proctitis, proctocolitis and ulcers is increasingly endemic in vulnerable populations in Europe (Stoner and Cohen, 2015; Control, 2019). However, our understanding of the disease epidemic remains incomplete due to the limited molecular epidemiology data on Ch. trachomatis, the causative agent of a broad spectrum of diseases (Dean et al., 1991; Rodríguez-Domínguez et al., 2014). LGV is caused by the L-genotypes (L1, L2, and L3) of CT (Ceovic and Gulin, 2015) and current the molecular epidemiology of the LGV epeidemic is based on sequence analysis of the ompA and ompA-pmpH genes. However, this test has limited discriminatory power across the different genotypes of LGV. The authors aimed to use multilocus sequence typing (MLST) methods and a novel combination of molecular markers with high diversity derived from variable genes of L-genotypes genomes to improve the specificity of Ch. trachomatis genotyping in Madrid. Martinez-Garcia et al. selected four genes, CTLon_0054, CTLon_0087, CTLon_0243, and CTLon_0301, in addition to ompA, to complete the genotype analysis of Ch. trachomatis. In silico and experimental studies were performed to compare the previously described MLST schemes and genes selected by authors. The diversity analysis approach tested by authors identified higher diversity of Ch. Trachomatis than previously reported and identified 3 main transmission chains. Subsequent spacio-temporal analysis revealed the major cluster was characterized by high diversification of Ch. Trachomatis genome and an active transmission chain. The authors also detected the L2b genome identical to the epidemic's origin, suggesting re-introductions or low screening rates in the studied population. Overall, this study proposes new methods to monitor sexual networks with higher precision, offers the possibility of differentiating between reinfection and persistence and identifying hotspots that could be crucial for contact tracing.

In a study by Liu et al., the clinical validity of the combination of gut microbiome data and common clinical indicators was analyzed as a predictor of myocardial infarction in young males. ST-segment elevation myocardial infarction (STEMI) in young male patients could indicate high heart attack risk (Zhang et al., 2016). Studies have demonstrated that changes in the gut microbiome could contribute to the incidence of coronary artery disease (CAD) (Koeth et al., 2013; Yoshida et al., 2018; Liu et al., 2019). This hypothesis was further applied to the pathogenesis of STEMI, a severe form of CAD in young males. The authors, Liu et al., therefore aimed to develop an early risk prediction model based on a comprehensive analysis of the gut microbiome in young male patients with STEMI combined with clinical data to address this hypothesis. They conducted the Illumina sequencing of 16S rRNA of the gut microbiome of 41 age and gender-matched STEMI patients and 40 non-CAD controls. The collected data was used to evaluate the efficacy and practicability by generating a nomogram and corresponding web page with K-fold crossvalidation, calibration curves and decision curve analysis (DCA). The authors demonstrated a substantially decreased α and β diversity and significantly altered gut microbiome in the young male patients with STEMI compared to controls. Also, the STMEI patients had higher body mass index (BMI), systolic blood pressure (SBP), triglyceride (TG), alanine aminotransferase (ALT), and aspartate aminotransferase (AST) parameters, while blood urea nitrogen (BUN) was lower compared to the control. Additionally, authors found that BMI and SBP had a positive correlation with Streptococcus and [Ruminococcus] and a negative correlation with Prevotella and Megasphaera, implying that alterations in

microbiome composition and abundances could play a role in the biochemical and metabolic changes in young male STEMI patients. Analysis of the calibration curves of the microbiome prediction model demonstrated consistency between the actual and predicted probabilities, with an indication that the microbiome model was superior to the clinical model. The authors concluded that combining data on the gut microbiome and clinical signs could be a valid and reliable non-invasive tool to predict STEMI in young males.

The original manuscript by Wang et al. addressed the urgent need for a sensitive and cost-effective novel approach to identify subtypes of influenza A viruses (IAVs). There are several subtypes of IAVs based on the expression of surface proteins hemagglutinin (HA) and neuraminidase (NA) (Schulze et al., 2010). The IAV variants emerge as a result of reassortment between genes coding for various HA and NA genes (Bhoumik and Hughes, 2010). The emergence of these new IAVs subtypes could be linked to different clinical manifestations of the disease. H1N1 (pH1N1) genotype usually causes a mild flu infection (Huo et al., 2019). In contrast, H5N1 viruses are highly pathogenic (HP), with a mortality rate of 60% (Yin et al., 2013). A rapid, sensitive, and cost-effective test which can differentiate across H5N1 and H1N1 subtypes, which present with similar symptoms upon initial infection, could aid in the management of influenza pandemic and control of HP infection spread. To develop a test that improved upon current virus culture, serology and genetic testing methods, the authors developed an approach using the spectral tool, Fluorescence resonance energy transfer (FRET), that is widely used for intermolecular interaction studies. They designed a homogeneous fluorescence method for dual detection of HA proteins of H1N1 and H5N1 influenza viruses-an aptasensor for HA using FRET strategy combined with DNase I-assisted cyclic enzymatic signal amplification. Fluorescent-labeled HA aptamers of H1N1 and H5N1 IAVs were designed and used as probes. Graphene oxide (GO) was used as a FRET acceptor and protected aptamers from DNase I cleavage. Increased sensitivity of the test was based on the amplification of fluorescence signal by liberated aptamers and HA proteins after DNase I digest. This test system was approximately 20-fold greater in sensitivity than traditional fluorometric methods without amplification. The authors also state no cross-reaction between H1N1 and H5N1 IAV subtypes with good reproducibility and stability of the aptasensor probes. This tool could be applied to the clinical setting to aid in the typing and early detection of influenza virus and surveillance for early pandemic preparedness efforts.

This Research Topic highlights the novel approaches in the detection of emerging infections. The development of novel diagnosis methods addresses the most urgent needs, which are sensitivity, specificity, speed of detection and affordability. The knowledge attained from the articles on this Research Topic could facilitate the development of diagnostic tools for the detection of emerging infections. This research is essential and urgently needed to improve our preparedness for future pandemics.

We are grateful to the authors and reviewers for their valuable contribution to the research and discussion that will leverage new frameworks for infectious disease diagnostics and control that responds to the increasing risk from emerging and reemerging pathogens.

Author contributions

TF: conceptualization, investigation, and writing—original draft. SK: writing–original draft and writing–review and editing. All authors contributed to the article and approved the submitted version.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Baker, R. E., Mahmud, A. S., Miller, I. F., Rajeev, M., Rasambainarivo, F., Rice, B. L., et al. (2022). Infectious disease in an era of global change. *Nat. Rev. Microbiol.* 20, 193–205. doi: 10.1038/s41579-021-00639-z

Bhoumik, P., and Hughes, A. L. (2010). Reassortment of ancient neuraminidase and recent hemagglutinin in pandemic (H1N1) 2009 virus. *Emer. Infect. Dis.* 16, 1748. doi: 10.3201/eid1611.100361

Biosearchtech. (2022). Ultra-high-throughput PCR testing system for SARS-CoV-2 detection. Available online at: https://www.biosearchtech.com/ultra-high-throughput-pcr-testing-system-for-sars-cov-2-detection (accessed April 10, 2023).

Brashares, J. S., Arcese, P., Sam, M. K., Coppolillo, P. B., Sinclair, A. R. E., and Balmford, A. (2004). Bushmeat hunting, wildlife declines, and fish supply in West Africa. *Science* 306, 1180–1183. doi: 10.1126/science.1102425

Ceovic, R., and Gulin, S. J. (2015). Lymphogranuloma venereum: diagnostic and treatment challenges. *Infect. Drug Resist.* 8, 39-47. doi: 10.2147/IDR.S57540

Control, E. C. f. D. P. a. (2019). Lymphogranuloma venereum. Available online at: https://www.ecdc.europa.eu/en/lymphogranuloma-venereum (accessed April 10, 2023).

Corey, L., Corbett-Detig, R., and Beyrer, C. (2022). Expanding efforts and support to respond to the HIV and COVID-19 intersecting pandemics. *JAMA* 327, 1227–1228. doi: 10.1001/jama.2022.3517

Dean, D., Patton, M., and Stephens, R. S. (1991). Direct sequence evaluation of the major outer membrane protein gene variant regions of Chlamydia trachomatis subtypes D', I', and L2'. *Infect. Immun.* 59, 1579–1582. doi: 10.1128/iai.59.4.1579-1582.1991

Dropulic, L. K., and Lederman, H. M. (2016). "Overview of infections in the immunocompromised host," in *Diagnostic microbiology of the immunocompromised host*. 1–50. doi: 10.1128/9781555819040.ch1

Forna, A., Nouvellet, P., Dorigatti, I., and Donnelly, C. A. (2020). Case fatality ratio estimates for the 2013–2016 West African Ebola epidemic: application

of Boosted Regression Trees for imputation. Clin. Infect. Dis. 70, 2476-2483. doi: 10.1093/cid/ciz678

Gilbert, R. K., Petersen, L. R., Honein, M. A., Moore, C. A., and Rasmussen, S. A. (2023). Zika virus as a cause of birth defects: Were the teratogenic effects of Zika virus missed for decades? *Birth Defects Res.* 115, 265–274. doi: 10.1002/bdr2.2134

Gletsou, E., Ioannou, M., Liakopoulos, V., Tsiambas, E., Ragos, V., and Stefanidis, I. (2018). Aspergillosis in immunocompromised patients with haematological malignancies. *J. BUON* 23, 7–10.

Harpaz, R., Dahl, R. M., and Dooling, K. L. (2016). Prevalence of immunosuppression among US adults, 2013. JAMA 316, 2547–2548. doi: 10.1001/jama.2016.16477

Huo, C., Wu, H., Xiao, J., Meng, D., Zou, S., Wang, M., et al. (2019). Genomic and bioinformatic characterization of mouse mast cells (P815) upon different influenza a virus (H1N1, H5N1, and H7N2) infections. *Front. Genet* 10, 595. doi: 10.3389/fgene.2019.00595

Koeth, R. A., Wang, Z., Levison, B. S., Buffa, J. A., Org, E., Sheehy, B. T., et al. (2013). Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat. Med.* 19, 576–585. doi: 10.1038/nm.3145

Li, J., Duan, H.-J., Chen, H.-Y., Ji, Y.-J., Zhang, X., Rong, Y.-H., et al. (2016). Age and Ebola viral load correlate with mortality and survival time in 288 Ebola virus disease patients. *Int. J. Infect. Dis.* 42, 34–39. doi: 10.1016/j.ijid.2015.10.021

Lin, S. J., Schranz, J., and Teutsch, S. M. (2001). Aspergillosis case-fatality rate: systematic review of the literature. *Clin. Infect. Dis.* 32, 358–366. doi: 10.1086/318483

Liu, H., Chen, X., Hu, X., Niu, H., Tian, R., Wang, H., et al. (2019). Alterations in the gut microbiome and metabolism with coronary artery disease severity. *Microbiome* 7, 1–14. doi: 10.1186/s40168-019-0683-9

Lowes, D., Al-Shair, K., Newton, P. J., Morris, J., Harris, C., Rautemaa-Richardson, R., et al. (2017). Predictors of mortality in chronic pulmonary aspergillosis. *Eur. Respir. J.* 49, 1601062. doi: 10.1183/13993003.01062-2016

McDermott, A. (2022). Climate change hastens disease spread across the globe. Proc. Nat. Acad. Sci. 119, e2200481119. doi: 10.1073/pnas.2200481119

Mohapatra, R. K., Pintilie, L., Kandi, V., Sarangi, A. K., Das, D., Sahu, R., et al. (2020). The recent challenges of highly contagious COVID-19, causing respiratory infections: Symptoms, diagnosis, transmission, possible vaccines, animal models, and immunotherapy. *Chem. Biol. Drug Design* 96, 1187–1208. doi: 10.1111/cbdd.13761

Rayens, E., Rayens, M. K., and Norris, K. A. (2022). Demographic and socioeconomic factors associated with fungal infection risk, United States, 2019. *Emer. Infect. Dis.* 28, 1955–1969. doi: 10.3201/eid2810.220391

Rodríguez-Domínguez, M., Puerta, T., Menéndez, B., González-Alba, J. M., Rodríguez, C., Hellín, T., et al. (2014). Clinical and epidemiological characterization of a lymphogranuloma venereum outbreak in Madrid, Spain: co-circulation of two variants. *Clin. Microbiol. Infect.* 20, 219–225. doi: 10.1111/1469-0691.12256

Schaible, U. E., and Kaufmann, S. H. E. (2007). Malnutrition and infection: complex mechanisms and global impacts. *PLoS Med.* 4, e115. doi: 10.1371/journal.pmed.0040115

Schulze, M., Nitsche, A., Schweiger, B., and Biere, B. (2010). Diagnostic approach for the differentiation of the pandemic influenza A (H1N1) v virus from recent human influenza viruses by real-time PCR. *PLoS ONE* 5, e9966. doi: 10.1371/journal.pone.0009966

Shin, B., Koh, W.-J., Jeong, B.-H., Yoo, H., Park, H. Y., Suh, G. Y., et al. (2014). Serum galactomannan antigen test for the diagnosis of chronic pulmonary aspergillosis. *J. Infect.* 68, 494–499. doi: 10.1016/j.jinf.2014.01.005

Song, Q., Sun, X., Dai, Z., Gao, Y., Gong, X., Zhou, B., et al. (2021). Point-of-care testing detection methods for COVID-19. *Lab on a Chip* 21, 1634–1660. doi: 10.1039/D0LC01156H

Stoner, B. P., and Cohen, S. E. (2015). Lymphogranuloma venereum 2015: clinical presentation, diagnosis, and treatment. *Clin. Infect. Dis.* 61, S865–S873. doi: 10.1093/cid/civ756

Viboud, C., Alonso, W. J., and Simonsen, L. (2006). Influenza in tropical regions. *PLoS Med.* 3, e89. doi: 10.1371/journal.pmed.0030089

Yin, J., Liu, S., and Zhu, Y. (2013). An overview of the highly pathogenic H5N1 influenza virus. *Virol. Sinica* 28, 3–15. doi: 10.1007/s12250-013-3294-9

Yoshida, N., Emoto, T., Yamashita, T., Watanabe, H., Hayashi, T., Tabata, T., et al. (2018). Bacteroides vulgatus and Bacteroides dorei reduce gut microbial lipopolysaccharide production and inhibit atherosclerosis. *Circulation* 138, 2486–2498. doi: 10.1161/CIRCULATIONAHA.118.033714

Zhang, Q., Zhao, D., Xie, W., Xie, X., Guo, M., Wang, M., et al. (2016). Recent trends in hospitalization for acute myocardial infarction in Beijing: increasing overall burden and a transition from ST-segment elevation to non-ST-segment elevation myocardial infarction in a population-based study. *Medicine* 95, e2677. doi: 10.1097/MD.000000000002677