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β-Lactam potentiators to re-sensitize resistant pathogens: Discovery, development, clinical use and the way forward

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β-lactam antibiotics are one of the most widely used and diverse classes of antimicrobial agents for treating both Gram-negative and Gram-positive bacterial infections. The β -lactam antibiotics, which include penicillins, cephalosporins, monobactams and carbapenems, exert their antibacterial activity by inhibiting the bacterial cell wall synthesis and have a global positive impact in treating serious bacterial infections. Today, β -lactam antibiotics are the most frequently prescribed antimicrobial across the globe. However, due to the widespread use and misapplication of β -lactam antibiotics in fields such as human medicine and animal agriculture, resistance to this superlative drug class has emerged in the majority of clinically important bacterial pathogens. This heightened antibiotic resistance prompted researchers to explore novel strategies to restore the activity of β -lactam antibiotics, which led to the discovery of β -lactamase inhibitors (BLIs) and other β -lactam potentiators. Although there are several successful β -lactam- β -lactamase inhibitor combinations in use, the emergence of novel resistance mechanisms and variants of β -lactamases have put the quest of new β -lactam potentiators beyond precedence. This review summarizes the success stories of β -lactamase inhibitors in use, prospective β -lactam potentiators in various phases of clinical trials and the different strategies used to identify novel β -lactam potentiators. Furthermore, this review discusses the various challenges in taking these β -lactam potentiators from bench to bedside and expounds other mechanisms that could be investigated to reduce the global antimicrobial resistance (AMR) burden.

KEYWORDS

antibiotic resistance, antibiotic potentiators, β -lactamas, β -lactamases, β -lactamase inhibitors, molecular docking, multidrug resistance

1. Introduction

Since the discovery of benzylpenicillin by Sir Alexander Fleming, antibiotics are considered the most important lifesaving drugs and have been nick-named the "magic bullets." Today, there are different classes of antibiotics used to treat several bacterial infections, and they are classified based on their structure, spectrum, and mode of action (Bush and Jacoby, 2010). Among the different classes of antibiotics, the β -lactam antibiotics are the most widely used and prescribed (Wilke et al., 2005). In the past decade, β -lactams represented more than 60% of all prescriptions (Damlin et al., 2020). This class of antibiotics is characterized by the β -lactam ring in their

structure and includes the penems (penicillins), cephems (cephalosporins and cephamycins), carbacephems, carbapenems and monobactams. However, with the growing use of antibiotics, the threat of antimicrobial resistance (AMR) among the bacterial pathogens has also loomed. Due to the antibiotic selection pressure, microorganisms gain AMR mostly by horizontal gene transfer or by accumulating spontaneous mutations in the target genes (Munita and Arias, 2016; Narendrakumar et al., 2020). Enterococcus spp., Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, Pseudomonas aeruginosa, and Escherichia coli, collectively termed the ESKAPE pathogens, are predominantly responsible for infectious diseases in humans and have been identified to be the major contributors to resistance and resistance dissemination (Zhen et al., 2019). Currently, there are increasing reports of difficult to treat (DTR) pathogens resistant to most first-line therapy agents from around the world, and experts have predicted that this growing AMR can possibly cause an "antibiotic apocalypse" where bacterial infections will become no longer treatable by antibiotics.

The World Health Organization (WHO) has stated that AMR is one of the top 10 global public health threats that humanity is facing today (World Health Organization, 2014; Munita and Arias, 2016). The economic cost of AMR is significant, and it has been identified as a major burden for low-and middle-income communities (World Bank, 2017). Considering the impact of AMR in the health sector and the economy, there have been several initiatives to contain the growing AMR burden. This included enforcement of strict rules on the indiscriminate use of antibiotics, surveillance of AMR and AMR drivers (Infectious Diseases Society of America (IDSA), 2011), "One Health" approach to address and avert AMR (Zhen et al., 2019; Mares et al., 2021), extensive research on identifying novel antibiotics and alternatives to antibiotics like vaccines, probiotics, and anti-virulence agents to curb AMR.

The present review expounds one such major strategy, "antibiotic potentiation" with special emphasis to the most used class of antibiotics, the β -lactams. The term antibiotic potentiation refers to the technique by which the activity of an antibiotic is increased, improved or restored. Compounds, either natural or synthetic, that do not exert direct antibacterial property but increase the efficacy of the antibiotic have been used to revive the activity of the antibiotics. Such compounds work either by blocking the primary resistance mechanism of the bacterial pathogen thus rendering the antibiotic effective, or work on the antibiotic to improve its antibacterial activity (Chawla et al., 2022). Several β -lactam potentiators, primarily the β-lactamase inhibitors (BLI) are already in the clinical use. Clinically approved BLIs include clavulanic acid, sulbactam, tazobactam, avibactam, relebactam and vaborbactam. In the present review, the first section recapitulates the various β -lactam antibiotics, their modes of action, and bacterial resistance to them, while the second section summarizes recent developments in β-lactam potentiator discovery, novel potentiators, various strategies used to identify them and the major roadblocks in bringing them from bench to bed-side.

2. β -Lactam antibiotics and their mechanism of action

From early 1920s to late 1940s, various research groups had observed antibacterial activity of several species of fungi against

different clinically important bacteria. The most common fungi observed to exhibit potential antibacterial activity was the Penicillium. Penicillin was isolated from Penicillium notatum, but the active compound for clinical trials was produced on a large scale from Penicillium chrysogenum (Chain, 1979). Though different Penicillium strains produced penicillin, all compounds were identified to have a common four-membered ring with an amide function called the "β-lactam ring." Penicillin was discovered in 1928 and used clinically by 1940. Later, cephalosporin C was identified and isolated from Cephalosporium acremonium in 1945 and the prototype drug for carbapenem was isolated from Streptomyces spp. in 1980s (Burton and Abraham, 1951). Monobactams, produced by Pseudomonas acidophila and Pseudomonas mesoacidophila were discovered more recently (Gwynn et al., 1988). Unlike the 6-aminopenicillanic acid (6-APA) the cephalosporins nucleus of penicillins, have а 7-aminocephalosporinic acid (7-ACA) nucleus while the carbapenems have a five-membered carbon ring associated with a β -lactam ring and monobactams contain a monocyclic ring structure. The β-lactam antibiotics were identified to have a broad spectrum of action and are effective in treating Gram-negative, Gram-positive and anaerobic bacteria. The β -lactam antibiotics have been used to treat infections such as pneumonia, sinusitis, pharyngitis, urinary tract infections (UTIs), skin and soft tissue infections and bloodstream infections (Bush and Bradford, 2016). According to a study published in 2020 by Vincent et al., β -lactam antibiotics were also the most commonly used antibiotics in intensive care units (ICUs), accounting for ~36% of all antibiotics used in treating critically ill patients (Vincent et al., 2020).

The β-lactam antibiotics are bactericidal in nature and the primary mechanism of action of the β -lactam antibiotics is by binding to a group of inner-membrane bound enzymes known as the "penicillin binding proteins (PBPs)" which are important in bacterial peptidoglycan wall synthesis. In Gram-negative bacteria, this peptidoglycan cell wall is ensconced by the bacterial outer membrane (OM), which to an extent hinders the antibiotic penetration (Bush and Macielag, 2010). However β -lactam antibiotics pass through the Gram-negative bacterial porins, reach the peptidoglycan and exert their action (Kim et al., 2020). The mechanism of action of β -lactam antibiotics against Gram negative and Gram positive bacteria is presented in Supplementary Figure 1. The major classes of β-lactam antibiotics, their spectrum of activity, major molecular targets and been resistance mechanisms have summarized in Supplementary Table 1. Structures of major classes of β-lactam antibiotics is presented in Supplementary Figure 2.

2.1. Penicillins

Penicillin was the first β -lactam antibiotic discovered and the second antibiotic to be used in human medicine after the sulfonamides. The group contains several natural and semisynthetic compounds and their pharmacokinetics and spectra of action differs significantly. Penicillin Penicillin (benzylpenicillin) and G V (phenoxymethylpenicillin) are the two natural penicillins used in human medicine mostly used to treat Gram-positive bacterial infections (Kulkarni et al., 2019). Among the semisynthetic penicillins, aminopenicillins like ampicillin, developed by the addition of an amino group to the benzylpenicillin, are the most clinically used antibiotics. Amoxicillin, developed by adding hydroxyl group to ampicillin was among the first broad spectrum penicillins that displayed activity against both Gram-positive and Gram-negative bacteria due to its better water solubility and absorbed from gastrointestinal tract. However, emergence of penicillinase producing bacteria prompted researchers to look for novel molecules that resisted penicillinase activity and thus methicillin was developed. Sooner, isoxazolyl penicillins which included the oxacillin, cloxacillin and dicloxacillin also known as antistaphylococcal penicillins, which had better oral absorption than methicillin were introduced to treat infections caused by Penicillin G resistant staphylococci (Smith et al., 1962). In short order, several other semisynthetic penicillins such as carboxypenicillins, acylureidopenicillins and amidino penicillins with improved action and bioavailability were developed, collectively known as the extended spectrum penicillins.

2.2. Cephalosporins

The cephalosporin antibiotics have a dihydrothiazine ring fused to the β -lactam nucleus with a 6-membered sulfur containing ring. Cephalosporins are the most frequently prescribed class of β -lactam antibiotics because of their broad spectrum of activity and lower allergenic and toxicity risks (Bush and Bradford, 2016). The use of cephalosporins extensively improved the treatment of infectious diseases and drastically reduced morbidity and mortality rates in pneumonia, meningitis, secondary infections in cancer patients, complicated skin and soft tissue infections, urogenital infections, infective endocarditis, serious bone and joint infections and *Salmonella* infections in children (Kwak et al., 2017; Kapil et al., 2019). Based on the spectra of action, cephalosporin groups of antibiotics have been classified into five generations.

2.3. Monobactams

Aztreonam is the most used monobactam antibiotic (Groman, 2009). Monobactam antibiotics are characterized by a single β -lactam ring not fused to another ring like other β -lactams. These antibiotics were identified to be more effective against the Gram-negative bacteria as compared to the Gram-positive bacteria due to their lower affinity to PBPs (Brogden and Heel, 1986). Surprisingly, monobactam antibiotics are resistant to bacterial metallo- β -lactamases (MBL), but susceptible to serine β -lactamases (SBL). Interestingly, Blais et al. had recently reported the discovery of a novel monobactam antibiotic, LYS228 effective against both MBL and SBL producing *Enterobacteriaceae* (Blais et al., 2018). The antibiotic is now in phase II clinical trials in patients with UTI and intra-abdominal infections.

2.4. Penems

Penems, the broad spectrum β -lactam antibiotics are majorly synthetic in nature and are sub-grouped as alkylpenems, aminopenems, arylpenems, oxypenems and thiopenems which are distinguished by the side chain of the unsaturated five-membered ring. Carbapenems are the most common penem antibiotic used which differs from other groups by possessing double bond between C2 and C3 and a substitution of carbon for sulfur at C1 (Papp-Wallace et al., 2011). One among the most used carbapenem antibiotics is the amidine derivative of thienamycin, Imipenem (N-formimidoylthienamycin) which is a broad spectrum β -lactam antibiotic effective against both Gram-positive and Gram-negative bacteria. However, imipenem is rapidly degraded by kidney dehydropeptidase-I (DHP-I). Hence, another broad spectrum carbapenem antibiotic, meropenem was developed which was relatively stable to DHP-I and that possessed enhanced activity against *Enterobacteriaceae*, *P. aeruginosa, Haemophilus influenzae*, and *Neisseria gonorrhoeae*. Other newly developed carbapenem antibiotics include doripenem, panipenem and biapenem (Perry and Ibbotson, 2002; Goa and Noble, 2003; Greer, 2008).

3. Resistance against β -lactam antibiotics: Major threat to the mighty class

It has been estimated that the annual sales of the β -lactam antibiotics is US \$15 billion in the international market accounting for 65% of the total antibiotics sold (Thakuria, 2013; Graham et al., 2016). While third and fourth generation cephalosporins, carbapenems and monobactams are the most used β -lactam antibiotics in human medicine, penicillin G is the β -lactam antibiotic used in veterinary medicine. With the increase in use of these antibiotics in various sectors, there has been a surge in resistance acquisition by diverse bacterial species as part of their evolution for survival. Bacteria gain resistance to β -lactam antibiotics mainly by three mechanisms. While production of β -lactamase enzymes (EC 3.5.2.6) serves as the most common cause of resistance against this diverse antibiotic group, loss or alteration of porins and upregulation of efflux pumps are other common mechanisms.

High prevalence of β -lactam antibiotic resistance, especially toward last generation cephalosporins and carbapenem antibiotics has been widely reported in clinically important bacterial pathogens worldwide. In a study conducted by Feretzakis and group in South Eastern Europe, more than 55% of P. aeruginosa, one of the major pathogen causing infections in cystic fibrosis (CF) patients was identified to be resistant to carbapenem antibiotics, imipenem, meropenem and doripenem and 15% of the isolates were resistant to cephalosporin antibiotics, cefepime and ceftazidime (Feretzakis et al., 2019). Pseudomonas aeruginosa is also known to cause infection in patients with chronic obstructive pulmonary disease (COPD), diabetes mellitus and also cause nosocomial infections. Another major pathogen imparting high AMR burden is the K. pneumoniae causing sepsis, pneumonia, surgical site infections and UTIs (Fursova et al., 2021). About 60% of K. pneumoniae isolated from India were reported to be resistant to carbapenem antibiotics (OECD and World Health Organization, 2020). Emergence and spread of carbapenem resistant A. baumanii has caused significant alarm in recent times. A. baumanii is associated with many nosocomial infections including ventilator associated pneumonia (VAP), bloodstream infections, UTI and wound infections (Necati Hakyemez et al., 2013). A plethora of β-lactam resistance determinants were identified in these resistant pathogens including genes coding for various β-lactamase enzymes and efflux pump proteins (Necati Hakyemez et al., 2013). Many of these resistance determinants are transferred among the bacteria through horizontal gene transfer and spread to inter and intra bacterial species (Hull et al., 2021).

Additionally, high prevalence of β-lactam heterogenous drug resistance or heteroresistance has been observed in several clinically important pathogens such as K. pneumoniae and Methicillin resistant S. aureus (MRSA). Heteroresistance is defined as the existence of two different subpopulation of the same microorganism within a single population with difference in resistance profiles (susceptible and resistant) against a common antibiotic. Majority of the Methicillin resistant Staphylococcus aureus (MRSA) cultures are identified to have a phenotypically heterogenous population of S. aureus strains with one subpopulation with extremely high β -lactam minimum inhibitory concentration (MIC) values and another subpopulation which exhibit very poor resistance (Kim et al., 2013). Similarly, heterogenous drug resistance was observed in MRSA against oxacillin which on single induction with the antibiotic converted into a homogenously resistant population (Chung et al., 2016). Nodari and his team reported high prevalence of imipenem heteroresistance among carbapenemaseproducing Enterobacteriaceae (CPE; Nodari et al., 2015). Heterogenous drug resistance among bacterial populations are often under-detected and have significant clinical implications as it is one of the major reasons for treatment failure.

3.1. β-Lactamase enzymes

β-Lactamases are a diverse group of enzymes that have the ability to hydrolyze the amide bond of the β -lactam ring rendering the antibiotic ineffective. β-Lactamase producing E. coli was first identified in 1940's even before the first β -lactam antibiotic was introduced for clinical use. Currently, there are numerous β-lactamases identified that have narrow or broad-spectrum activity against different β-lactam antibiotics. The sequence information of 7,537 β -lactamases, 1,526 structural information, and kinetics of 47 β-lactamases evolved over the years have been listed in the Beta-Lactamase DataBase (BLDB) revealing the diverse nature of the enzymes (Naas et al., 2017). These enzymes are usually located in the periplasmic space of the bacteria and they hydrolyse the β -lactam antibiotic before they reach the target molecule. Many Gram-negative bacteria also produce membrane vesicles that bear high levels of these enzymes that act on the antibiotics and reduce the concentration even before it reaches the infection foci (Ciofu et al., 2000).

Various characteristics like molecular structure, substrate profile, biochemical properties and functional characteristics were initially used to classify the β-lactamase enzymes. The Amber classification classifies the enzymes into four different classes (class A, B, C, and D) based on its sequence motifs and hydrolytic properties. Class A, C, and D constitute enzymes that employ their serine residue for the nucleophilic attack and hydrolyze of the β-lactam antibiotic while class B enzymes utilize a metal-activated water nucleophile to drive the hydrolytic reaction. There have been various reviews that summarize the different classes of β-lactamases and their substrates (Page, 2002; Nagshetty et al., 2021; Akhtar et al., 2022). Some β-lactamases have been identified to be specific to certain species of bacteria and some have been identified to be successful in global dissemination and confer selective fitness to bacterial clones (Fernández et al., 2012). Many ESKAPE pathogens have been identified to possess more than one class of β-lactamase resistance gene rendering them multidrug resistant and hard to treat (Oliveira et al., 2020). In 1980s, Enterobacteriaceae isolates capable of hydrolysing extended spectrum β -lactam antibiotics were reported from around the world and were identified to possess extended spectrum β -lactamases (ESBLs) such as TEM, SHV, and CTX-M. Carbapenem antibiotics were the treatment options for such ESBL producing isolates. However, bacterial isolates resistant to carbapenem antibiotics soon evolved capable of producing carbapenemase enzymes. Genes that confer ESBL activity (bla_{TEM} , bla_{SHV} , $bla_{\text{CTX-M}}$) and carbapenamase activity (bla_{KPC}) in the class A, carbapenamases (bla_{NDM} , bla_{VIM} and bla_{IMP}) in the class B, ESBLs (bla_{CMY} , bla_{AMP} and bla_{ADC}) in class C and the carbapenemase encoding bla_{OXA} genes in class D have been explicitly been successful in transmission among the *Enterobacteriaceae* family, *A. baumannii* and *P. aeruginosa* (Raible et al., 2017; Ahmed et al., 2020).

These classes of resistance genes are either chromosomally encoded or plasmid encoded. Plasmid encoded resistance genes have demonstrated significant success in the global spread of AMR among bacteria in clinical and non-clinical settings. KPC encoded by *bla*_{KPC} gene has been identified to have the potential for inter-species and geographical dissemination due to its association with Tn-3 type transposon Tn4401 capable of inserting into different types of plasmids of Gram-negative bacteria (Arnold et al., 2011). Additionally, the global success of KPC dissemination is also attributed to the clonal expansion of K. pneumoniae ST258, ST512, ST304, and ST11 (Munoz-Price et al., 2013). Mostly, *bla*_{NDM} variants are plasmid encoded and are rapidly transferred between enterobacterial isolates K. pneumoniae and E. coli. NDM encoding isolates are resistant to almost all β-lactam antibiotics with tigecycline and colistin being the only treatment options (Wu et al., 2019). Recently, there have been various reports of chromosomal integration of usually plasmid encoded β-lactamase resistance genes like bla_{CTX-M} and bla_{OXA-48} in E. coli isolates (Rodríguez et al., 2014; Turton et al., 2016). In many other cases, co-existence of more than one class of β-lactamase resistance gene in both chromosome and plasmid was observed in Gram-negative bacterial pathogens (Poirel et al., 2010). The β -lactamase resistance genes have been found associated with various mobile genetic elements (MGEs) like plasmids, transposons, insertion sequences (IS) and integrons. bla_{IMP-1} has been identified to be associated with Class I integrons which may have chromosomal or plasmid location in different species (Watanabe et al., 1991; Arakawa et al., 1995). Verona integronencoded metallo-β-lactamase (VIM) is another prevalent integron associated MBL followed by German imipenemase (GIM-1) and Seoul imipenemase (SIM-1) located in class 1 integrons of Pseudomonas spp. Insertion sequences like ISAba1, ISAba4 and transposons such as Tn2006 and Tn2008 are found associated with the bla_{OXA-23} gene in A. baumannii chromosome while ISAba3 and IS18 are found associated with bla_{OXA-58} (Poirel and Nordmann, 2006; Corvec et al., 2007). Recently Chen et al., reported the identification of a novel ESBL encoded by the gene $bla_{OXA-830}$ in the chromosome of environmental bacteria Aeromonas simiae (Chen et al., 2019). The OXA-48 like carbapenemases in Enterobacterales has been found to be associated with various transposable elements and insertion sequences. Klebsiella pneumoniae and E. coli circulating in the North Africa and Middle East have been reported to possess OXA-48 associated with Tn1999 variants on IncL plasmids (Giani et al., 2012). OXA-181 and OXA-232 endemic to the Indian subcontinent and few sub-Saharan African countries are associated with ISEcp1, Tn2013 on ColE2, and IncX3 types of plasmids. Additionally, clonal dissemination of certain high risk clones of K. pneumoniae (ST147, ST307, ST15, and

ST14) and *E. coli* (ST38 and ST410) are also a minor cause of the spread of OXA-48 like carbapenemases (Pitout et al., 2019). Many ESBL genes such as bla_{TEM} bla_{SHV} , $bla_{\text{CTX-M}}$, carbapenemase genes such as bla_{SPM} , bla_{IMP} , bla_{NDM} , bla_{KPC} and $bla_{\text{OXA-48}}$ are conjugative plasmid encoded in ESKAPE pathogens which allows in the easy transmission of these genes among the bacterial isolates (Kim et al., 2011; Veeraraghavan et al., 2019). Thus, β -lactamase enzymes form the most common method of β -lactam resistance in clinical pathogens.

3.2. PBP modification

The PBPs are the target of the β -lactam antibiotics. The PBPs are transpeptidases or carboxypeptidases involved in peptidoglycan metabolism. Hence PBP modification serves as another major mechanism by which bacteria resist the action of the antibiotic. The mutational alterations in PBPs can confer resistance to β-lactam antibiotics (Chesnel et al., 2002). Unemo et al., in 2012 had reported the isolation of a high level cefixime and ceftriaxone resistant Neisseria gonorrhoeae having a penA mosaic allele with an additional A501P mutation in the PBP2 (Unemo et al., 2012). Moya et al. had previously reported an unusual mechanism of β-lactam resistance by mutational inactivation of non-essential PBPs (PBP4) coded by dacB which behaves as a trap target for β -lactams (Moya et al., 2009). Additionally, the mutation on *dacB* also triggers over-expression of AmpC coding β -lactamase enzymes and activating the CreBC (BlrAB) two-component regulator which together results in high level β-lactam resistance. Mutation in the PBP coding gene down regulates its expression which in turn reduces the peptidoglycan turnover. The reduced peptidoglycan synthesis is considered as a cue for over production of β-lactamase enzymes. Recently, Lazzaro et al. reported high-level resistance of Enterococcus faecalis to Ceftobiprole (BPR), a new-generation cephalosporin as a result of alterations in the pbp4 gene (Lazzaro et al., 2022). The mutations revealed to over-express PBP4 leading to increased transpeptidation causing higher peptidoglycan synthesis that prevented Ceftobiprole penetration into the bacterial cell. Thus, mutations leading to gain of function or loss of function of PBP have been determined to play a major role in β-lactam resistance in bacteria.

3.3. Selective permeability by porin regulation

Gram-negative bacteria possess the OM that acts as an additional barrier for the transport of solutes into the cell. The selective transport of molecules is achieved by specialized proteins called porins present on the OM. Modulation of porin expression is another mechanism of β -lactam resistance particularly in Gram-negative bacteria. Various insertional elements and point mutations in the promoter and protein coding sequences have been identified to alter porin protein expression and structure. *Escherichia coli* mutants lacking OmpC and OmpF proteins were documented to demonstrate high level resistance to cefoxitin and other cephalosporin antibiotics (Jaffe et al., 1982). Similarly, characterization of meropenem resistant *Serratia marcescens* isolates revealed insertional inactivation of *ompF* gene by an IS element, IS1 (Suh et al., 2010). Mutations in *ompK*35 and *ompK*36 in *K. pneumoniae* and *ompC* and *ompF* in *Enterobacter* spp. have been associated with increased resistance to the majority of carbapenem antibiotics (Doumith et al., 2009). Pagel et al., noticed that mutations at the L3 loop of OmpU in the cholera causing pathogen, Vibrio cholerae resulted in decreased susceptibility of the bacteria to cephalosporin antibiotics (Pagel et al., 2007). Likewise, several studies utilizing either site directed mutagenesis, knockdown or knockout of porin encoding genes in several clinically important pathogens revealed that these pathogens could gain reduced susceptibility to even last generation cephalosporins and carbapenems (Smani et al., 2014; Choi and Lee, 2019). A recent study by Khalifa et al. revealed that 93.3% of E. coli and 95.7% of K. pneumoniae isolated from hospitals in Egypt were identified to have lost their porins or showed modified porins resulting in heightened resistance to all carbapenem antibiotics (Khalifa et al., 2021). Interestingly, there have also been reports of association of porin regulation and expression of efflux pumps. Carbapenem resistance in P. aeruginosa has been primarily due to a combination of decreased transcriptional expression of OprD and upregulation of MexAB-OprM active efflux system (Muderris et al., 2018). Thus, in clinical isolates, interrelation of different mechanisms of resistance has been identified and mutations in some genes have been identified to activate multiple other resistance pathways.

3.4. Efflux pumps

Decreasing passive influx and increasing active efflux of antibiotics is a common mechanism of antibiotic resistance in bacteria. In clinically important pathogens, especially Gram-negative bacteria, a large number of promiscuous/polyselective efflux pumps such as the Resistance-Nodulation-Division (RND) efflux pumps that can recognize and expel a large variety of antibiotics have been identified. Unlike the β -lactamase coding genes that are mostly acquired, genes coding for efflux pumps are mostly intrinsic and found in both susceptible as well as resistant clones. However, acquisition of mutations at the regulatory region, promoter or open reading frame (ORF) of the gene leads to over-expression of these genes that cause resistance. Though the role of efflux pumps in β-lactam drug resistance was widely described in P. aeruginosa, it was not until recently that its role was identified in the members of the Enterobacteriaceae family (Glen and Lamont, 2021). About 12 different RND efflux pumps have been identified in P. aeruginosa of which MexAB-OprM, MexXY-OprM, and MexCD-OprJ play important role in β-lactam resistance and at least one of these efflux pumps have revealed increased expression during antibiotic treatment (Masuda et al., 2000). RND efflux pumps have also been reported in K. pneumoniae conferring β -lactam drug resistance. KexD expression was identified to be high in imipenem resistant K. pneumoniae and KpnEF (SMR type efflux pump) over-expression was noticed in cephalosporin resistant isolates (Srinivasan and Rajamohan, 2013). Uddin and Ahn recently reported multiple efflux pumps (AcrAB, AcrEF, EmrAB, MdfA, and MdtK) involved in conferring reduced susceptibility of β-lactam antibiotics in Salmonella Typhimurium (Uddin and Ahn, 2018). The expression of efflux pumps has been identified to be highly increased in bacterial pathogens during antibiotic treatment thereby significantly increasing the MIC of the drug required for the treatment. The treatment of Klebsiella aerogenes with imipenem was identified to drastically increase the expression of the marA gene that codes for an efflux

pump protein (Bornet et al., 2003). Many other efflux pumps that cause co-resistance to other antibiotics have been identified to co-spread with β -lactam resistance genes (Li et al., 2019). The OqxAB, a multi-drug efflux pump widely attributed to resistance against quinolones, nitrofurantoin and chloramphenicol is co-transferable with *bla*_{CTX-M}, *rmtB* and *aac*(6')-1b etc. The *oqxAB* genes are located either on the chromosome and/or on plasmids and are flanked by IS26-like element in clinical isolates of *Enterobacteriaceae* which makes them transferable (Li et al., 2019). Efflux pumps that expel out multiple antibiotics thus posing threat of multidrug resistance (MDR) in bacteria have been a major concern to both researchers and clinicians alike.

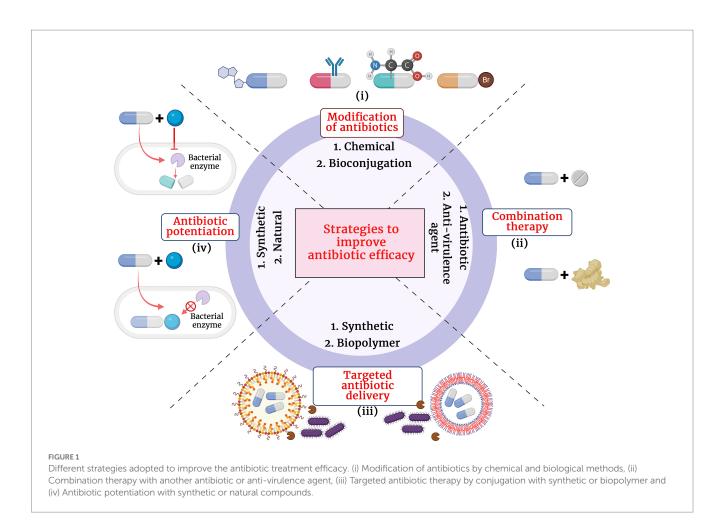
4. Strategies to improve efficacy of existing antibiotics

The growing antibiotic resistance and the discovery void of novel classes of antibiotics have caused immense pressure and challenge to clinicians and researchers across the globe. Since 1987, for over 30 years, very few new antibiotics with potential antibacterial activity have reached the final phases of clinical trials and have made their way to the market. Though retapamulin, the topically used pleuromutilin antibacterial, was introduced into the market in 2007, the chemical class was first described in 1952 (Novak and Shlaes, 2010). Though there exists a discovery void of novel classes of antibacterial compounds, the antibacterial pipeline has not been empty during the past decades and there have been many antibacterial compounds with improved activity being introduced for trials. The different strategies adopted to improve antibiotic resistance pathogens are presented in Figure 1.

- i. Structural modification of antibiotics: Remarkable improvement in the antibacterial activity and stability of the β-lactam antibiotics have been achieved by structural modifications. A modification in the dihydrothiazine ring fused to the β -lactam ring in the cephalosporin nucleus was identified to improve cephalosporin activity against S. aureus and P. aeruginosa isolates (Neu, 1985). Similarly, introduction of acyl side chains to β-lactam antibiotics ameliorated its antibacterial activity by increasing its affinity toward PBPs (Neu, 1985). Additionally, chemical modifications of β-lactam antibiotics have also been identified to impart multifaceted properties to the class such as antifungal, anti-parasitic and anticancer (Neu, 1985; Hamilton-Miller, 1999). Novel modifications of β-lactam antibiotics with different pharmacophophoric groups and their biological activities were recently reviewed by De Rosa et al. (2021).
- ii. Combination therapy: Combinations of drugs that have different modes of action or targets in the bacterial cell are used to combat antibiotic resistance. Antibiotic combinations that impart synergistic activities have been identified to possess broad spectrum antibacterial activity and less resistance evolution of bacterial pathogens. Combination therapy of β -lactam antibiotics with aminoglycoside antibiotics have revealed improved activity against MDR *P. aeruginosa, S. aureus, Enterococcus* sp. and *Streptococcus* sp.

(Paul et al., 2006). β-lactam antibiotics exert its antibacterial action by inhibiting the bacterial cell wall synthesis while the aminoglycoside antibiotics inhibit the bacterial growth by interfering with the protein synthesis by binding to the aminoacyl site of 16S ribosomal RNA of the 30S ribosomal subunit. Conventionally, combination therapy with a β-lactam antibiotic and aminoglycoside works due to the cell damage caused by β-lactam antibiotic that allow aminoglycoside to penetrate more efficiently into the bacterial cell and exert its action. However, the exact detailed mechanism of the synergism is not clear. Also, recent reports revealed enhanced synergistic activity of carbapenem antibiotics with peptide antibiotic daptomycin (Sakoulas et al., 2017). Daptomycin is a cyclin anionic lipopeptide which is usually ineffective against Gram negative pathogens due to the presence of its OM. However, daptomycin along with β -lactam antibiotics show synergism as the peptidoglycan damage caused by β-lactam antibiotic could help daptomycin to enter the bacterial cell and cause membrane depolarization and also inhibit DNA, RNA and protein synthesis. However, the exact molecular bases and mechanism of action involved in carbapenem antibiotic and daptomycin interaction remain unknown (Gagetti et al., 2016). Siriyong et al., had recently reported the effectiveness of dual β-lactam antibiotic therapy to be effective against MDR P.aeruginosa (Siriyong et al., 2019). This activity is due to the double dose effect. Combining two β -lactam antibiotics enables inactivation of multiple PBPs to achieve synergistic bacterial killing. Nevertheless, clinical data to support combination therapy of β-lactam antibiotics with other classes of antibiotics is sparse and recent reports suggested that dual β-lactam or carbapenem therapy was not superior to single carbapenem regimens (Rigatto et al., 2022). Apart from antibiotic-antibiotic combination therapies, combining an antibiotic with an anti-virulence compound has also been identified to be an efficient way to combat MDR pathogens with minimal resistance progression (Rezzoagli et al., 2020). Anti-virulence compounds are those that target the pathogen's virulence pathways rather than pathways for its survival. Cefiderocol that combines a catechol type siderophore and cephalosporin core with side chains similar to cefepime and ceftazidime has been identified to be effective against Gram-negative pathogens, including carbapenemresistant Enterobacterales (Wu et al., 2020). The combined structure of siderophore-cephalosporin confer enhanced stability against hydrolysis by ESBL, CTX-M, and carbapenemases, such as KPC, NDM, VIM, IMP, OXA-23, OXA-51-like and OXA-58. Moreover, the siderophore that chelate extracellular iron enhances the penetration of the molecule into the bacterial cells through the iron transporter channels in addition to the passive diffusion through membrane porins (Wu et al., 2020).

iii. Targeted antibiotic delivery: Apart from the above strategies, another method to increase the efficacy of antibiotics is to achieve targeted and durable drug delivery. Targeted drug delivery increases the amount of antibiotic at the infection foci and increased durability allows more time for the drug to act upon the bacteria. Complete killing by the antibiotics prevents



bacterial evolution to attain resistance. Nanoscale delivery systems which are either biological lipid based, or synthetic polymer based are popularly used to deliver the antibiotic to the site of infection (Stebbins et al., 2014). These delivery systems protect the antibiotic from enzymatic degradation by antibiotic modifying enzymes and additionally, antibody conjugated delivery systems specifically target the pathogenic bacteria and also activate host immunity at the site of infection thereby enhancing pathogen killing (Abed et al., 2015).

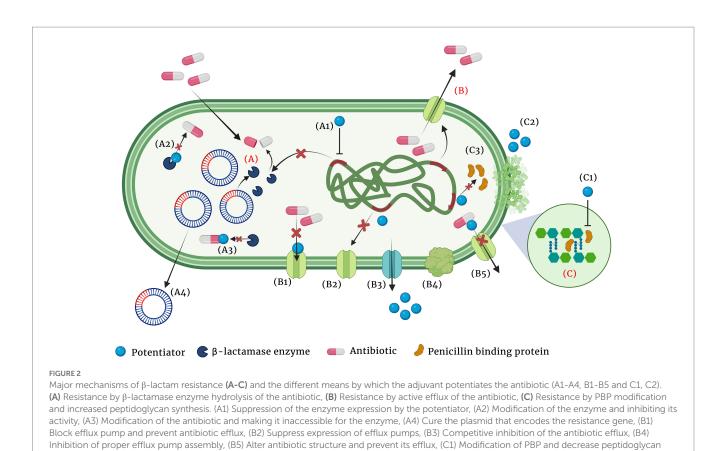
iv. Antibiotic potentiators: Another strategy used to battle the growing antibiotic resistance is to preserve the activity of existing antibiotics with non-antibiotic compounds that could restore their activity. Such compounds that do not exert direct antibacterial property, but increase the efficacy of the antibiotic or restore the activity of the drug are collectively known as antibiotic potentiators, adjuvants or resistance breakers (Laws et al., 2019). Figure 2 depicts the various mechanisms by which bacteria resist antibiotics and how potentiators function. In addition to not having antibacterial activity, it is important that the antibiotic potentiators should have other characteristics such as ideal pharmacological features when administered along with the antibiotic, high serum stability, high bioavailability, and low toxicity. The potentiators should also ideally be highly selective and have no eukaryotic targets. Additionally, it is ideal that the potentiator compounds be easily available and economical.

5. β-Lactam potentiators

 β -lactam potentiators are mainly of three types, *viz*. (i) β -lactamase inhibitors (BLIs), (ii) efflux pump inhibitors (EPI), and (iii) membrane permeabilizers. Among the three, the BLIs are the most successful and widely used in the clinical setting. There are several natural and synthetic β -lactam potentiators developed. In the context of rising AMR, many non-conventional potentiators are also being studied to evaluate their potency in clinical settings.

5.1. β -Lactamase inhibitors – success stories and clinical status

The BLIs are compounds that render the β -lactamase enzymes produced by the bacterial isolates inactive to hydrolyze the antibiotic. The emergence of β -lactamase producing isolates were reported from clinical settings soon after the introduction of β -lactam antibiotics. Though several macromolecular fractions (MM4550, MM13902, MM17880, BRL1437) from different *Streptomyces* sp. were identified to pose β -lactamase inhibitory activity, they were not clinically translated due to their unstable nature and undesirable pharmacological profiles. Reading and Cole in the late 1970s reported the detection of a compound with novel clavum structure isolated from *Streptomyces clavuligerus* ATCC 27064 (Reading and Cole, 1977). The compound when used in combination with β -lactam



antibiotics significantly enhanced its antibacterial activity against penicillinase and cephalosporinase producing *E. coli*, *K. aerogenes*, *Proteus mirabilis*, and *S. aureus* isolates. This compound identified to be clavulanic acid was the first clinically potent BLI. Soon, a combination of amoxicillin and clavulanic acid (Amoxiclauv/ Augmentin) was used clinically and found highly successful against class A β -lactamase producing bacterial isolates. The similarity of stereochemistry between clavulanic acid and β -lactamase enzymes was identified as the major contributing factor for its potentiation activity (Reading and Cole, 1977). Some BLIs are by themselves β -lactam antibiotics that at sub-inhibitory concentration augment the activity of the partner β -lactam antibiotic.

synthesis, (C2) Chelation of divalent cations and (C3) Suppression of PBP synthesis.

Following the identification of clavulanic acid, the β-lactamase inhibitory activities of various synthetic penicillin-based sulfones were screened which led to the discovery of sulbactam and tazobactam (YTR 830; Akova, 2008). Sulbactam, the halogenated derivatives of penicillanic acid (6-β-bromo-and 6-β-iodopenicillanic acid), was identified to be effective against class A β-lactamase producing Gramnegative isolates (Rolinson, 1991). However, it was identified to be less potent as compared to clavulanic acid in its activity against S. aureus β-lactamases and bacteria that possess TEM variants. While sulbactam was combined with ampicillin, amoxicillin, piperacillin, cefoperazone, cefotaxime and cefepime, tazobactam is used in combination with piperacillin and ceftolozane. The piperacillin-tazobactam combination was identified to be effective against broad-spectrum β-lactamasesproducing and some extended-spectrum β-lactamases-producing Enterobacteriaceae. However, the combination was not effective against AmpC β-lactamases producing Gram-negative bacteria (Rolinson, 1991; Gin et al., 2007). Ceftolozane-tazobactam combination was identified to be even effective against Pseudomonas. As a therapeutic value addition, recent preliminary *in vitro* studies by Sakoulas et al., revealed that tazobactam potentiate peptide antibiotic, daptomycin against MRSA and colistin resistant *A. baumanii* (Sakoulas et al., 2017). An interesting finding by Urban et al. revealed that different concentrations of the BLIs have different targets. Clavulanic acid, sulbactam and tazobactam at lower concentration were identified to manifest the potentiation activity by inhibiting the β -lactamase activity, while at higher concentration, it exerted its potentiation activity through PBP modulation (Urban et al., 1995). Important BLIs, their class, inhibition spectrum and clinical status are listed in Table 1.

Recently, two new BLIs of the diazabicyclooctanones (DBOs) group namely avibactam and relebactam were identified with enhanced carbapenamase inhibitory activity. They were the first non β -lactam BLI to be used clinically and they exerted their β -lactamase inhibitory activity by acylating the β -lactamases reveribily unlike clavulanic acid, sulbactam or tazobactam that exerted irreversible inactivation of the resistance enzyme (Ehmann et al., 2012). Avibactam potentiated ceftaroline, ceftazidime, aztreonam while relebactam was used in combination with imipenem (Yahav et al., 2020). Interestingly, relebactam at the same concentration potentiates imipenem by its β -lactamase inhibitory activity and also PBP inhibitory activity (Blizzard et al., 2014). Successive to DBO BLIs, several boronic acid based BLIs were developed which were found to be effective against β -lactamase producing bacteria and they are at different stages of clinical trials. Most of the boronic acid based BLIs were identified to have dual mode of

TABLE 1 List of important BLIs, their class, inhibition spectrum, and clinical status.

Class of β-lactamase inhibitor	Inhibitor	Partner β-lactams	Inhibition spectrum	Current status	Isolated from	References
First generation						
Irreversible acylation of catalytic serine residue	Clavulanic acid	Amoxicillin, Ticarcillin	Plasmid encoded Class A ESBLs	Completed, marketed as Augmentin [®] , Timentin [®]	Streptomyces clavuligerus	Bush (1988)
Penicillin-based sulfones	Sulbactam	Ampicillin, Piperacillin, Amoxicillin, Cefoperazone	Primarily to Class A (TEM-type β-lactamases), less effective against SHV and OXA-type β-lactamases	Completed, marketed as Unasyn [®]	Synthetic	Benson and Nahata (1988)
	Tazobactam (YTR 830)	Piperacillin, Ceftolozane	$\begin{array}{c} Class \ A \ ESBLs \ (TEM, \\ SHV), \ Class \ C \ enzymes \\ and \ OXA-type \ \beta \\ lactamases \end{array}$	Completed, marketed as Zosyn [®]	Synthetic derivative of penicillin	Wise et al. (1991) and Busl (2015)
Second generation	·					
Diazabicyclooctane (DABCO)	Avibactam (AVE1330A, NZL 104)	Ceftaroline, Ceftazidime, Aztreonam	Class A ESBLs (TEM, KPC and SHV) Class A carbapenemases, Class C and D enzymes,	Completed, marketed as Avycaz [®] , Zavicefta [®]	Synthetic	Bonnefoy (2004) and Ehmann et al. (2012)
	Relebactam (MK-7655)	Imipenem	Class A ESBLs, Class A carbapenemases Class C enzymes	Completed, marketed as Recarbrio [®]	Synthetic	Blizzard et al. (2014) and FDA (2019)
Third generation						
Boronic acid inhibitor	Vaborbactam	Biapenem, Meropenem, Doripenem, Ertapenem	Broad spectrum of class A (KPC, CTX-M, SHV), C (CMY) and D enzymes	Completed phase III trials, marketed as Vabomere TM , Carbavance [®]	Synthetic	Hecker et al. (2015)
	(VNRX-5133)	Cefepime	Class A, C and D serine β lactamases and class B MBLs (VIM and NDM) in both carbapenem- resistant Enterobacteriaceae and <i>P. aeruginosa</i>	Undergoing phase III drug– drug interaction study	Synthetic	Liu et al. (2020)
Bicyclo-acyl hydrazide	Zidebactam (WCK 5107)	Cefepime	AmpC, ESBL, KPC, and OXA-48-like β-lactamases	Under phase III clinical trials	Synthetic	Yahav et al. (2020) and Karlowsky et al. (2020)
Diazabicyclooctane	Nacubactam (OP0595, RO7079901, RG6080)	Meropenem	Class A, Class C and some Class D β-lactamases and against carbapenem-resistant <i>K</i> . <i>pneumoniae</i>	Phase I clinical trial completed	Synthetic	Mallalieu et al. (2020)
Diazabicyclooctane	Durlobactam (ETX2514)	Sulbactam	Class A, C and D serine β-lactamases.	Phase III clinical trial completed	Synthetic	Shapiro et al. (2021)
Diazabicyclooctane	ETX1317/ Prodrug ETX0282	Cefpodoxime proxetil	Class A, C, and D serine β-lactamases	Phase I clinical trial completed	Synthetic	Durand-Réville et al. (2020)

(Continued)

Class of β -lactamase inhibitor	Inhibitor	Partner β-lactams	Inhibition spectrum	Current status	Isolated from	References
Penicillanic acid sulfone	Enmetazobactam (AAI101)	Cefepime	CTX-M, TEM, SHV, and other class A β-lactamases	Phase III clinical trial completed	Synthetic	Papp-Wallace et al. (2019)
Metallo-beta- lactamases (MBL) inhibitors	Elores (CSE:1034)	Ceftriaxone, sulbactam, disodium edetate (EDTA)	MBLs	Completed, marketed in India as Elores	EDTA	Chaudhary et al. (2017)
	ANT 431	Meropenem	MBLs and MBL- producing carbapenem resistant <i>Enterobacteriaceae</i> (CRE)	Undergoing clinical trials	Synthetic	Everett et al. (2018)
	QPX7728	Meropenem	VIM-1, NDM-1 and IMP-1	Under phase I clinical trials	Synthetic	Lomovskaya et al. (2021)
	Magnolol	Meropenem	NDM	Undergoing clinical trials	Magnolia officinalis	Liu et al. (2018)
	Aspergillomarasmine A	Meropenem	NDM-1 enzyme and other MBLs, VIM-2	Undergoing clinical trials	Aspergillus versicolor	King et al. (2014)
	Bisthiazolidines	Meropenem	Class B1,B2 and B3 MBLs	Undergoing laboratory studies	Synthetic	Hinchliffe et al. (2016)
	Nitrilotriacetic acid and <i>N</i> - (phosphonomethyl) iminodiacetic acid	Meropenem	VIM, NDM and IMP classes		6-phosphonomethylpyridine- 2-carboxylates	Tehrani et al. (2020)

TABLE 1 (Continued)

action like the relebactam. They exert their potentiation activity by β -lactamase inhibition and also PBP modification thereby enhancing their spectrum. Boronic acid based BLIs were active against most of the Class A, C and D (Oxa-48) β -lactamases (Lepak et al., 2019; Liu et al., 2020). Some boronic acid based BLIs such as zidebactam (WCK 5222) and taniborbactam (VNRX-5133) were active against Gram-negative organisms producing various carbapenemases, including MBLs producers (Lepak et al., 2019; Liu et al., 2020).

Though there are various BLIs developed against SBL (Amber classes A, C and D), effective MBL inhibitors are sparse. Despite a large amount of research being carried out to identify a potent MBL inhibitor, very few have reached the clinical trials and none have been commercialized. Elores (CSE-1034: ceftriaxone+sulbactam+disodium edetate) is the only MBL inhibitor used in patients with ESBL/MBL producing bacterial infections. The drug is approved in India by the Indian FDA after phase III clinical trials (Chaudhary et al., 2017). The major mechanism by which MBL inhibitors exert their activity is metal chelation. Elores showed high treatment success (83%) against both ESBL as well as MBL producing pathogens and lower rate of adverse effects (8.4%) caused by other drugs of choice to treat the infections (Chaudhary et al., 2017). However, long term usage of sodium edetate is not advisable as the chelating agent could interfere with eukaryotic enzymes that utilizes divalent cations for their activity. Aspergillomarasmine A, a natural compound discovered to have inhibitory activity against NDM-1 and VIM-2 exert its action by chelating the zinc ion present at the enzyme's active site (King et al., 2014). Bisthiazolidines and small bicyclic compounds also utilize the same mechanism and were found to be effective against B1, B2, and B3 MBLs (Hinchliffe et al., 2016). Currently, development of covalent inhibitors that can bind to specific conserved amino acids at the active site of MBLs are gaining momentum. Irreversible inhibition of MBL (IMP-1) by 3-(3-mercaptopropionylsulfanyl) propionic acid pentafluorophenyl ester by binding to Lys224 and activity of ebselen against NDM-1 by binding to Cys221 are examples for such strategies (Kurosaki et al., 2005; Chiou et al., 2015). The major MBL inhibitors have been recently reviewed by Li et al. (2022). Ceftazidime-avibactam, ceftolozane-tazobactam, ceftazidime-tazobactam, meropenemvaborbactam and imipenem-relebactam are the recently developed β -lactam-BLI combinations used to treat ESBL and carbapenemase producing bacterial pathogens (Yahav et al., 2020). However, very limited BLIs have been identified that pose both SBL and MBL inhibitory activity and the quest for such compounds continues.

5.2. β-Lactam efflux pump inhibitors

Efflux pumps are a mechanism by which bacteria counteract the action of antibiotics by pumping out the drug thereby significantly decreasing its concentration and effectiveness. EPI or blockers have grown as a potential and attractive strategy to overcome bacterial resistance on account of the dwindling novel antibiotics. Efflux pump blocker, Phenylalanyl arginyl β -naphthylamide (PA β N) has been widely explored for its potentiation activity in clinical pathogens possessing polyspecific efflux pumps (Lamers et al., 2013).

1-(1-napthylmethyl)-piperazine (NMP), another putative EPI was identified to potentiate oxacillin antibiotic by inhibiting direct binding of the substrate to the efflux pump (AcrAB and AcrEF) of pathogens in the Enterobacteriaceae family. However, the compound did not have action on E. coli isolates and MexAB-OprM or MexCD-OprJ efflux pumps of P. aeruginosa (Bohnert and Kern, 2005). Further, artesunate, an anti-malarial drug was identified to possess EPI activity by binding to AcrAB-TolC of E. coli and thereby potentiating several β -lactam antibiotics such as penicillin G, ampicillin, cefazolin, cefuroxime and cefoperazone (Li et al., 2011). A plant based 5,6,7-trihydroflavone EPI, Baicalein, was identified to potentiate β-lactam antibiotics including oxacillin, cefmetazole and ampicillin while Conessine, another plantbased compound interfered with MexAB-OprM efflux pump in MDR *P. aeruginosa* potentiating cefotaxime antibiotic (Siriyong et al., 2017). Karumathil et al., had identified the potentiation efficacy of transcinnamaldehyde, isolated from cinnamon, eugenol isolated from clove, carvacrol isolated from oregano oil and thymol obtained from thyme against MDR A. baumanii. The natural compounds exerted their action by acting on the AdeABC efflux pump of A. baumanii making it sensitive to seven β -lactam antibiotics toward which it was previously resistant (Karumathil et al., 2018). Catechin gallate, a component from tea extract, was identified to potentiate oxacillin against MRSA isolates by reducing their MIC from 64–512 μ g mL⁻¹ to \leq 0.5–1 μ g mL⁻¹ (Stapleton et al., 2004). However, no EPIs which specifically potentiate β -lactam antibiotics have been commercialized and clinically used. Today, dual potentiation therapy by combining EPI-antibiotic combinations with antimicrobial photodynamic inactivation (APDI) that utilizes a photosensitizer to activate reactive oxygen species within the bacteria in presence of visible light and kill the pathogen are gaining popularity to target MDR and XDR strains (Tegos et al., 2008; Hamblin, 2016).

5.3. Outer membrane permeabilizers that potentiate β -lactam therapy

Permeabilizers are compounds that debilitate the OM thereby allowing the antibiotic to enter the bacterial cell and exert its action. Unlike the BLIs, research on β -lactam potentiators that act by permeabilizing OM and cell membranes of bacterial pathogens are at its infancy. There have been sparse studies that have identified OM permeabilizers that work in conjunction with β-lactam antibiotics though it is a promising strategy. Several natural compounds have been screened for its OM permeabilizing ability against β-lactamase producing and carbapenamase producing bacterial pathogens. Recently, Farrag et al. demonstrated the permeabilizing ability of gallic acid against β -lactamase producing *P. aeruginosa* (Farrag et al., 2019). The irreversible OM destabilization ability of gallic acid was determined to be the result of chelation of divalent cations from the OM and lipopolysaccharide components of the pathogen. A combinatorial potentiating strategy by combining an OM permeabilizing agent, a BLI and a suitable β -lactam antibiotic has been hypothesized to be an effective way to overcome infections caused by β -lactam resistant pathogens (Farrag et al., 2015). Interestingly, many of the BLIs that have been identified and clinically used also have PBP binding and modifying activity (Moya et al., 2017). Zidebactam and WCK 5153 have been identified to specifically bind to PBP2 apart from inhibiting class A, C and some class D β-lactamases which together enhance the activity of β -lactam antibiotic (Moya et al.,

2017). Anti-parasite compound pentamidine was identified to potentiate Gram-positive drugs against the Gram-negative pathogen *A. baumanii* in animal models through a mechanism of membrane destabilization (Ando et al., 2012). Polymyxin antibiotics and nanopeptide derivatives of polymyxin B were identified to portray significant OM permeabilizing activity thereby potentiating β -lactam antibiotics (Vaara, 2019). Apart from the BLIs that exert OM permeabilizing activity, nanopeptide named NAB74 developed from polymyxin B has entered the initial phases of clinical trials making the strategy encouraging and optimistic.

5.4. Novel β -lactam potentiators

Apart from the classical β-lactam potentiators, many natural, synthetic, and semisynthetic compounds have been identified to potentiate β-lactam antibiotics against β-lactamase producing bacteria. Few promising β -lactam potentiators discovered within the last 5 years include farnesol, bulgecins, thiosemicarbazones and octyl gallate. Synthetic structural variants of farnesol, a sesquiterpene alcohol was identified to potentiate ampicillin and oxacillin against MRSA strains both in-vitro and in-vivo (Kim et al., 2018). Farnesol have been identified to upregulate and downregulate genes involved in cell membrane biogenesis and efflux pump proteins of Gramnegative pathogens A. baumannii and E. coli (Kostoulias et al., 2016). However, the exact molecular targets and mechanism by which these compounds potentiate β -lactams against MRSA are not elucidated. Bulgecins which are iminosaccharide secondary metabolites of Paraburkholderia acidophila have been reported to be a potential β -lactam enhancer against β -lactam resistant *P. aeruginosa* strains. The putative mechanism of ceftazidime and meropenem potentiation by bulgecin is understood to be the inhibition of periplasmic enzymes, Slt, MltD and MltG lytic transglycosylases that are very important for bacterial cell wall synthesis (Tomoshige et al., 2018). Additionally, diaryl and dipyridyl-substituted thiosemicarbazones were identified to possess synergistic effects with carbapenem antibiotic, meropenem against broad spectrum of MBL possessing bacterial pathogens (Zhao et al., 2021). Dipyridyl-substituted thiosemicarbazones competitively and reversibly inhibited NDM-1. The inhibition of MBLs by thiosemicarbazones was identified to be by the Zn(II) binding ability of a sulfur group in the compound (Li et al., 2021). Octyl gallate, a FDA approved antioxidant was identified to potentiate β-lactam antibiotics, penicillin, ampicillin and cephalothin against MRSA by increasing the bacterial cell wall permeability (Tamang et al., 2022). The galloyl moiety of octyl gallate is identified to interact with multiple targets in the bacterial cell wall which inhibit effective cell wall synthesis. Though the in-depth mechanistic evaluation of these molecules is still under evaluation, the differential targets of these compounds other than the conventional β -lactam drug targets make them potential futuristic β -lactam potentiators.

5.5. Non-conventional β -lactam antibiotic potentiators

5.5.1. Antimicrobial peptides

Antimicrobial peptides (AMPs), also known as host defence peptides are positively charged short peptides found in a wide range of living organisms including bacteria. Against the backdrop of rapidly developing antibiotic resistance, attempts to put AMPs into clinical use as potentiators is accelerating. Thanatin, a single - disulfie-bond containing β -hairpin antimicrobial peptide was found to damage the OM of NDM-1-producing bacteria by displacing divalent cations competitively and promoting lipopolysaccharide leakage (Moura et al., 2020). Additionally, thanatin was identified to reduces NDM-1 enzymatic activity by displacing zinc ions from the active site of the enzyme and thereby restore carbapenem antibiotics against carbapenem resistant NDM-1-coding bacteria (Ma et al., 2019). Temporin A isolated from the skin excretions of the frog Rana temporaria was identified to potentiate imipenem and amoxicillinclavulanic acid antibiotics and potentiate these antibiotics to re-sensitize MDR E. faecalis (Giacometti et al., 2005), Previously, Mandal et al., had created two novel short AMP BLIs, dBLIP-1 and dBLIP-2 by peptide rational designing technique which could potentiated β-lactam antibiotics (amoxicillin, ampicillin and cefotaxime) against plasmid mediated CTX-M-14 coding Gramnegative pathogens (Mandal et al., 2014). In another study employing natural AMPs having different structure and mode of action, PG-1, β-defensins, LL-37 were identified to have good membrane permeabilizing potential (Zharkova et al., 2019). Additionally, sub-inhibitory concentrations of a bivalent AMP, ASU014, used in combination with sub-inhibitory concentrations of oxacillin was identified to be effective against MRSA in both in-vitro and in-vivo studies (Lainson et al., 2017). Though AMPs are largely screened for discovering novel BLIs and OM permeabilizers, not many have been successfully translated clinically due to their host cell toxicity. To minimize toxicity and poor pharmacokinetics, AMPs are structurally modified that at many a times reduce its potency (Wang et al., 2021).

Peptidomimetics, which are synthetic analogues of peptides, capable of functionally being active and engaging in the same biological functions as that of AMPs, with reduced toxicity, is yet another popular method of antibiotic potentiation. Mood et al., described the potentiation activity of the peptidomimetic compound CEP-136 in combination with carbapenam antibiotics against carbapenamase producing clinical isolates *P. aeruginosa, K. pneumoniae, E. coli*, and *A. baumannii* (Mood et al., 2021). Yet in another study, a class of peptidomimetics were developed from human α -defensin 5 (HD5) which displayed promising potentiating activity of β -lactam antibiotics against MDR Gram-negative pathogens (Luo et al., 2022). The different mechanisms by which AMPs exert their potentiating activities is presented in Figure 3i.

5.5.2. Nanoparticles

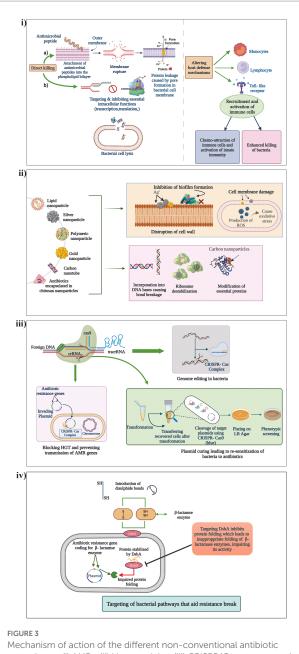
Nanoparticles (Nps) have long been investigated for their various therapeutic characteristics including antibacterial activity against drug resistant pathogens. Many Nps are known for their ability to penetrate the cell membranes of bacteria and interfere with their molecular functions thus devising unique antimicrobial activities (Mba and Nweze, 2021). Recent studies have also identified antibiotic potentiation ability of few Nps against β -lactam resistant pathogens. Graphene oxide and carbon nanotubes have displayed potent inhibitory effects on the MBL, VIM-2 by non-competitive adsorption of the enzyme to the surface of the Nps (Huang et al., 2015). In another study, silver nanoparticles (AgNPs) stabilized using bio-surfactin revealed good potentiation activity of amoxicillin, cefoperazone and doripenem against NDM-1, NDM-4, NDM-5, and NDM-7 producing E. coli (Kumar et al., 2019). AgNPs has also been identified to potentiate cefotaxime against ESBL producing E. coli (Mohammed et al., 2021). Furthermore, metal nanoparticles can be used as nanocarriers of antibiotics to minimize toxicity, and also lower the instances of resistance development through approaches of dual drug action which include antibiotic action and the nanoparticle activity targeting another pathway (Kotrange et al., 2021). Fan et al. encapsulated cephalosporin antibiotics (cefotaxime and ceftiofur) and BLIs (tazobactam and clavulanate) with cross-linked chitosan nanoparticles (CNAIs) which revealed excellent drug stability and antimicrobial activity against MDR ESBL-producing Enterobacteriaceae as compared to conventional antibiotic plus BLI therapy (Fan et al., 2022). Thus, nanoparticles carry a wide range of applications as potentiators and carriers that should be explored meticulously for therapeutic application. Mechanisms by which nanoparticles exert their potentiating activities is presented in Figure 3ii.

5.5.3. CRISPR/Cas systems

Using CRISPR/Cas (Clustered regularly interspaced short palindromic repeats and CRISPR-associated protein) for countering AMR is one of the most exciting and latest strategies. The CRISPR/Cas system has the ability to degrade foreign incoming nucleic acids such as AMR gene encoding MGEs thereby acting as a bacterial immune system. CRISPR/Cas systems have shown potency to limit entry of plasmids into the bacterial cells and also cure them, which is explored to potentiate antibiotics (Butiuc-Keul et al., 2022). A study conducted by Tagliaferr et al., found a decreased expression of *bla*_{TEM-1} gene and decreased *bla*_{TEM-1} coding plasmid copy number in *E. coli* treated with CRISPR/Cas which thereby potentiated five β-lactam antibiotics (Tagliaferri et al., 2020). It was identified that the CRISPR/Cas system could cure even high copy number plasmids leading to complete sensitization of the antibiotics. In another study, Hao et al., recently developed a CRISPR/Cas system (pCasCure) that could re-sensitize carbapenem antibiotics against carbapenemase producing clinical isolates. The authors were able to obtain the deletion of plasmid mediated carbapenem resistance genes such as $bla_{\rm KPC}$, $bla_{\rm NDM}$ and bla_{OXA-48} by a rate of effectiveness of over 94 percent (Hao et al., 2020). Mechanisms by which CRISPR/Cas is utilized for potentiating antibiotics is presented in Figure 3iii. Though the strategy is effective in-vitro, the major challenge for its applicability in the clinical setting is the lack of an optimal deliver system of CRISPR-Cas9 into target bacterial populations.

5.5.4. Identification of bacterial pathways that aid resistance break

New antibacterial agents, antibiotic potentiators and novel techniques to potentiate antibiotics are the need of the hour. Apart from the discovery of antibiotic adjuvants that either neutralize resistance enzymes or undermine cellular integrity, exploration of other bacterial pathways that aid in resistance break is becoming increasingly popular. DsbA is a protein present in bacteria which facilitates correct protein folding by presenting di-sulfide bonds leading to their stability. Furniss et al., investigated the role of DsbA modulation on incapacitating diverse β -lactamases in *E. coli* mutants. The study revealed that chemical inhibition of DsbA leads to resensitization of MDR Gram-negative pathogens to β -lactam antibiotics and increased the survival of animals challenged with



Mechanism of action of the different non-conventional antibiotic potentiators. (i) AMPs, (ii) Nanoparticles, (iii) CRISPR/Cas systems and (iv) Targeting of different bacterial pathways that aid in resistance break.

MDR *P.aeruginosa* (Furniss et al., 2022). Similarly, experiments of Skredenske et al., had earlier demonstrated that targeting the global transcriptional activator, RhaS of XylS/AraC family proteins, substantially inhibited the transcription of various antibiotic resistance genes (Skredenske et al., 2013; Furniss et al., 2022). Screening of small molecule libraries against RhaS identified a highly selective compound able to potentiate various antibiotics. Thus, identification of novel unconventional potentiators of β -lactam antibiotics that target previously unrecognized targets have gained importance because of the evolution and diversification of β -lactam resistance genes. A representation of how targeting of a different bacterial pathway could aid in potentiating an antibiotic is presented in Figure 3iv.

6. β-Lactam potentiators-screening/ identification strategies

There are various techniques by which a potential β -lactam potentiator could be screened and identified. In early times, for the identification of BLIs, screening was done by conventional agar well diffusion assays (Rolinson, 1991). However, recently, various less laborious techniques are developed for the fast, effective and accurate screening of large numbers of compounds. In the following section, we discuss the classical as well as emerging techniques used for the screening of BLIs and other potentiators that can reinstate the activity of β -lactam antibiotics.

6.1. Colorimetric assays for screening novel $\beta\text{-lactam}$ potentiators

Colorimetric assays that analyze the growth inhibition of the pathogens in presence of antibiotic, compound and combinations of antibiotic and compounds is a conventional method of potentiator screening. Microwell plates are used to screen multiple compounds at a time. Reporter strains that over-express the enzyme or protein against which the compounds are being screened and chromogenic substrates that allow easy detection of the desired enzymatic activity are used to develop the assays. Compounds are initially screened for their potentiation activity and further assays are performed to characterize their mode of action. BLIs, EPIs and OM permeabilizers can be identified by this technique. Colorimetric assay with CENTA and Nitrocefin as chromogenic substrate revealed the antibacterial activity of Syzygium aromaticum oil and fresh juice of Ocimum sanctum leaves against ESBL producing E. coli (Shrivastav et al., 2019). A high throughput whole cell turbidity assay was used for screening a 645,000 compounds library against AmpC overproducing reporter strain to identify small molecule inhibitors of AmpG which is necessary for the induction of AmpC (Li et al., 2016). The study discovered 8 potential AmpG inhibitors that potentiate the activity of cefoxitin antibiotic inhibiting the growth of ESBL producing P. aeruginosa (Collia et al., 2018). Modified whole cell colorimetric assays such as the UV-Vis spectroscopy that can evaluate real time activity of β -lactamases and inhibitory activity of β -lactam potentiators are used widely for screening potential antibiotic adjuvants (Yang et al., 2017).

6.2. In-silico predictions

Molecular docking and molecular dynamic simulations (MDS) are very popular and well-established techniques for virtual screening of compounds. *In-silico* predictions not only predict the mechanism of resistance and identify novel potentiators, but also help in understanding structural restructuring capabilities for enhancing compound activity. During the screening of lead molecules, parameters of GRID Molecular Interaction Fields (MIFs) such as H-bond donor, H-bond acceptor and hydrophobicity are inspected to identify favorable active sites in the protein, and the molecule which gives the highest interactions is selected. Several studies have used *in-silico* screening to identify BLIs and EPIs that potentiate β -lactam antibiotics. FLAPdock

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(Fingerprint for Ligands and Proteins) is one such molecular docking tool that has identified a number of potential molecules active against class A β -lactamases (CTX-M, KPC-2, AmpC), class C β -lactamases, and MBLs (NDM-1 and VIM-2; Spyrakis et al., 2020). Ten natural compounds exhibiting potent MBL inhibiting activity targeting NDM-1 were identified by *in-silico* screening using Autodock Vina and Schrödinger suite software Extra Precision (XP; Spyrakis et al., 2020; Salari-Jazi et al., 2021). In another recent study, *in-silico* screening using Auto Dock4 software revealed Carsonic acid to exhibit promising inhibitory effects against NDM-1 (Yang et al., 2020). *In-silico* predictions thus are a promising strategy to identify broad-spectrum BLIs and EPIs though their *in-vitro* success has to be analyzed through various other assays.

6.3. Artificial intelligence – based screening

By being able to analyze massive amounts of data instantly, machine learning (ML), artificial intelligence (AI), and neural networks (NN) have ushered in a golden era of drug discovery and synthesis. Bhadra et al., had used ML to analyze distribution of amino acid sequences in AMPs to predict their novel antibacterial and potentiating activities (Bhadra et al., 2018; Cunha et al., 2021). Mansbach et al. used ML coupled with the Hunting Fox Algorithm to study the membrane penetration activity of compounds in MDR P. aeruginosa (Mansbach et al., 2020). ML integrated with highthroughput Fourier-transform infrared spectroscopy has been applied to detect novel β -lactam resistance genes and β -lactam potentiators (Cunha et al., 2021). Parvaiz et al. specifically screened compounds potentiating β -lactam antibiotics by targeting bla_{CMY} genes from a 700,000 compounds library by using Site-Identification by Ligand Competitive Saturation (SILCS) technology, Ligand-grid Free Energy (LGFE) analysis and ML based random-forest (RF) scoring (Parvaiz et al., 2021). Thus, the intersection of ML and AMR has enabled the development and enhancement of several models that have aided new antibiotic and antibiotic potentiator discovery.

6.4. Omics based approaches

In many cases, antibiotic potentiating molecules which are not synthetic are either derived from microorganisms itself such as secondary metabolites, plant based such as polyphenols or alkaloids or animal based such as AMPs. The presence of such antibiotic potentiating molecules could be detected by various omics techniques. The major omics techniques utilized for novel drug and potentiating molecule discovery include genomics, metagenomics, transcriptomics, proteomics and metabolomics.

i. Genomics and metagenomics: Genomics techniques such as DNA microarray and next generation sequencing along with genome mining is useful in discovering biosynthetic gene clusters (BGCs) coding for novel bioactive molecules (Chandra Mohana et al., 2018). Many polyketides and non-ribosomal peptides with antibacterial and antibiotic potentiating activity has been identified through genome mining. As many as 17

novel BGCs coding for polyenes including salinilactam, a novel macrolactam molecule with potential antibacterial activity was identified through genome mining of marine actinomycete Salinispora tropica (Udwary et al., 2007). Likewise, several antiinfective molecules such as clarepoxcins, cypemycin, haloduracin, lactocillin, microcyclamide, teixobactin, reistomycin have been identified from different bacteria and actinomycetes through genomic approaches (Claesen and Bibb, 2010; Velásquez and van der Donk, 2011; Donia et al., 2014). The antibiotic potentiating activity of these molecules is worth exploring. Lactocillin was identified through metagenomic mining of human microbiome while clarepoxin was identified through multiplexed metagenomic mining of soil microbiome (Donia et al., 2014; Owen et al., 2015). Similarly, metagenomics coupled with synthetic biology approaches could be used to identify various non-ribosomal peptides with different bioactivities (Stevenson et al., 2019). Additionally, Santana-Pereira et al., had recently used shotgun sequencing of forest soil metagenomic libraries to identify 1015 BGCs (Santana-Pereira et al., 2020). Molecules from thus identified novel gene clusters could be screened for antibiotic potentiating activity. Hence, it is important to validate the expression of these gene clusters identified by genomics to understand the potency of biomolecules.

- ii. Transcriptomics: Understanding the mode of action of prospective potentiating molecules is one of the major steps in developing the candidate molecule into a drug combination. Through transcriptomics, it is possible to identify and evaluate the molecule's putative targets. Also, adverse drug target effects could be identified through transcriptomics (O'Rourke et al., 2020). Recently, Aunins and his team designed novel anti-sense inhibitors through transcriptomics-based approach which could potentiate carbapenem efficiency in carbapenem resistant *E. coli* (Aunins et al., 2020).
- iii. Proteomics: Like the transcriptomics approach proteomics data is largely used to understand the dynamics of drug targets at the protein level upon candidate molecule treatment. A novel β-lactamase (Axc) with carbapenemase activity was identified through proteomics approach in a meropenem resistant clinical isolate of Achromobacter xylosoxidans (Fleurbaaij et al., 2018). Recently, Guan et al., had utilized proteomics to reveal synergistic action of theaflavin, a polyphenolic compound present in fermented tea with β-lactam antibiotics against MRSA (Guan et al., 2022). Cassiano et al., had recently utilized chemical proteomics guided approach to identify novel targets of the MBL inhibitor, magnolol (Cassiano et al., 2019). Proteomics could be thus used for understanding protein targets of synergistic and potentiating molecules. Most of the proteomics-based research studies in drug discovery have been performed to characterize protein expression profiling, functional proteomics and phosphoproteomics.
- iv. Metabolomics: Identification of the metabolic pathways of the lead molecule through metabolomics has reduced the cost of toxicological screening. This has enabled improved clinical trials which has shortened the time needed for drugs to move through the development pipeline. Though metabolomics has been widely used to identify novel antibiotics, it is under explored for identifying β -lactam adjuvants (Tuyiringire et al.,

2018; Aminov, 2022). Recently, Campos and Zampieri, had utilized high throughput metabolomic approach to test the activity of 1,279 pharmacologically diverse drugs with no known antibacterial activity on *E. coli*. The metabolome profiles of drug treated *E. coli* was compared with metabolome profiles of untreated *E. coli* to understand the activity of the small molecules. The study revealed nine drugs with non-promiscuous enzyme targets in the bacteria (Campos and Zampieri, 2019). Also Zampieri and his team had developed a rapid systemic metabolome profiling strategy to identify the different modes of action of new bioactive compounds (Zampieri et al., 2018).

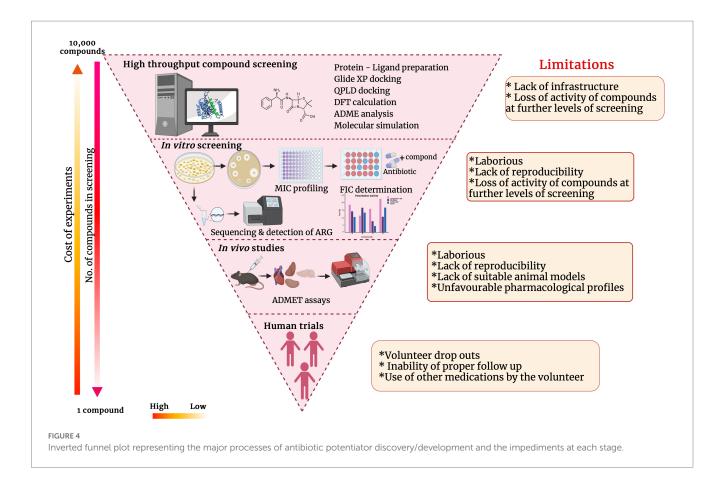
7. Challenges of taking β -lactam potentiators from bench to bedside

Potentiators for sure obviate the urgent need to discover novel antibiotics. However, after the identification of a potential antibiotic adjuvant, development, and commercialization of it have a lot of roadblocks. Though the time taken and capital associated with novel antibiotic discovery and commercialization is considered higher than potentiator discovery, the process is invariable and equally strenuous. The major processes of antibiotic potentiator discovery/development and the impediments at each stage are represented in the inverted funnel plot (Figure 4). Though there have been many lead molecules identified as β-lactam potentiators through in-silico predictions, very few of them have been analyzed in-vitro and in-vivo on clinical pathogens to determine their efficacy. Automation of in-vitro screening and cutting down the labor would encourage more labs to translate the hit molecules they have identified through in-silico techniques to further levels. Interaction and collaboration between computational biologists and wet lab researchers can accelerate the hit molecule screening processes.

Various factors have to be considered before commercializing and using the antibiotic-antibiotic potentiator combination for therapeutic purposes. One major factor that has to be considered while combining the antibiotic and antibiotic potentiator is its compatibility. It has to be made sure that there are no unfavorable pharmacological profiles when the combination is used. For this, the concentrations of the antibiotic and antibiotic adjuvant have to be standardized. In many instances, it has been noticed that β-lactam-BLI combinations require a high concentration of either of the components for the desired activity in *in-vivo* studies, which can lead to toxicity or undesirable pharmacological outcomes (Papp-Wallace, 2019). Additionally, many BLIs exhibit low oral absorption, especially in their native form, and demand chemical modifications to stabilize them and exhibit better oral absorption (Luci et al., 2021). For instance, sulbactam in its native form was identified to have poor oral absorption as compared to pivaloyloxymethyl ester of sulbactam (Campoli-Richards and Brogden, 1987). Thus, structural modifications of the potentiator molecule could improve its absorption. Potentiator molecules both existing and newly identified could be used as the core to construct a structurally modified library which could be checked for improvement of activity at lower concentrations, improvement in absorption and other physiological parameters.

Other factors that have to be contemplated are target specificity and substrate specificity. Target specificity is very important, especially while developing EPI and it has to be ratified that it does not inhibit any of the eukaryotic efflux pumps. There are reports of many antibacterial and potentiating molecules that target both bacterial and human pathways (Dalhoff, 2021). Reserpine, known to be a potent Norfloxacin potentiator has been identified to have direct action as calcium antagonist on mammalian smooth muscle cells (Casteels and Login, 1983). Similarly, Verapamil, another mammalian calcium channel blocker has been identified to potentiate carbenicillin against P. aeruginosa (Hulen et al., 2020). Verampil is used to treat high blood pressure, angina and supraventricular tachycardia in humans. Further, it has been identified that many prokaryotic and eukaryotic enzymes share key structural features. Sevier et al., had previously revealed that the catalytic domain of eukaryotic oxidoreductases Ero1 and Erv2 share sequence homology and structural features similar to that of the catalytic core of DsbB (Sevier et al., 2005). Hence, molecules targeting DsbB catalytic core could also affect eukaryotic oxidoreductases which could hamper disulfide bond formation of eukaryotic peptides also. Thus, it is very important to develop molecules that have lower affinity to mammalian proteins and those which have higher specificity to prokaryotic targets. This could be possible by generating precise protein interaction networks and binding affinity studies of prokaryotic and eukaryotic targets with the lead molecule. Similarly, identification of a universal BLI that can target all β-lactamases is a herculean task and hence different combinations have to be used to treat different bacterial infections. Understanding the type of β-lactamases produced by the bacterial pathogens is warranted to determine the appropriate antibiotic-BLI combination. For the same, rigorous surveillance and documentation of different bacterial pathogens and their resistance profile both phenotypic and genotypic is necessary.

Another major problem of β-lactam potentiator commercialization is the rapid emergence of resistant microorganisms to them. Previously, more than 90% of isolates were susceptible to ampicillinsulbactam combinations. But, now a days, extensive resistance of clinical pathogens against ampicillin-sulbactam has been reported (Ferreira et al., 2019). It has been only more than a decade that avibactam and relebactam potentiators were introduced for clinical use. However, bacterial pathogens especially the Gram-negatives have evolved mechanisms of resistance against them. Isolates harboring *bla*_{KPC} genes with specific mutations that confer DBO BLIs ineffective have been isolated from various parts of the world (Both et al., 2017; Giddins et al., 2018). There are various methods by which bacteria have developed methods to resist even β -lactam-BLI combinations. Overproduction of β -lactamase enzymes has been described as a major mechanism by which microorganisms resist β -lactam-BLI combinations. Additionally, mutations in the efflux pumps can reduce the potentiating action of EPIs. Schuster et al., had reported that random mutagenesis of AcrB efflux pump protein of E. coli resulted in reduced effectiveness of the most commonly used EPI, 1-(1-napthylmethyl)-piperazine (NMP; Schuster et al., 2014). To counteract this, molecules can be screened to target conserved protein domains in enzymes and efflux pumps that have lesser chance of resistance accumulations. Further, adaptive laboratory evolution studies of bacterial pathogens in the presence of the antibiotic and the lead molecule can be performed in-vivo and in-vitro to determine the possibilities of resistance developments.



8. Future directions and the way forward

Today, antibiotics are considered a double-edged sword. Due to the indiscriminate and inappropriate use of the antibiotics, bacterial pathogens have evolved multiple resistance mechanisms. To counteract the bacterial resistance and restore the efficacy of antibiotics, natural and synthetic compounds that do not possess antibacterial activity by themselves are being screened for their antibiotic potentiating activity. However, the vast structural and mechanical diversification of β-lactam resistance genes have caused bacterial pathogens to develop resistance to even the antibiotic-βlactam potentiator combinations. In this context, screening of novel potential β-lactam potentiators to maintain the efficacy of this powerful class of antibiotics is highly warranted. Soon after the introduction of penicillin for clinical use in 1941, bacterial persistence was reported by Joseph Bigger in 1944 where he observed that a sub-population of S. aureus was not killed by penicillin. Today, persister population of bacteria cause a major cause of concern for clinicians as they cause treatment failures by alleviating bacterial resistance. Hence it is very important to identify an effective way to eradicate the persister population. A probable way would be identification of a β-lactam potentiators effective against persister bacterial population. Though there are no β-lactam potentiators found effective against persister bacterial populations, bithionol, an anthelmintic drug was identified to potentiate gentamicin against persister population of MRSA by increasing bacterial membrane permeability (Kim et al., 2019).

Natural and marine sources have been identified to serve as a vast resource for discovering novel bioactive compounds. Marine pharmacology is an expanding but underexplored field in which novel β -lactam potentiators could be discovered. The active interaction between the biotic and abiotic systems within the marine environment allows the marine flora and fauna to produce structurally and functionally distinct bioactive compounds. Multi-omics approaches like genomics, transcriptomics, proteomics and metabolomics can be integrated and used to discover potent β-lactam adjuvants. Machine learning and AI can be utilized to screen large datasets of known marine and natural bioactive molecules in-silico, apart from conventional in-vitro laboratory screening. Structural similarity to the existing β -lactam antibiotics can be one of the positive filtering options to identify a potent β -lactam potentiator. Furthermore, chemical or biological modifications of existing β-lactam potentiators to increase their efficacy when combined with antibiotics could be used to avoid the time-consuming process of screening and identifying potentiators from natural sources. Repurposing previously approved drugs for potentiation activity could also be used to develop effective β-lactam potentiators.

In addition to the screening of novel potentiators, ways to decrease the frantic use of antibiotics by means of non-antibiotic therapies have to be promoted to put a brake on the growing AMR burden. Awareness among the general public regarding the harmful effects of AMR is of prime importance to reduce antibiotic use. Traditional non-antibiotic therapies such as phage therapy and probiotic therapy could be utilized to treat infections. The specificity of phages to target specific bacteria could be utilized for effective pathogen elimination without disturbing the normal flora. Probiotic bacteria can be also used as a prophylactic agent as they either competitively exclude the pathogen from successful colonization, or create an unfavorable condition for the pathogen's growth thereby preventing the infection. Furthermore, vaccines continue to be the most important non-antibiotic intervention that can prevent infections. Multiple vaccines have been developed against various pathogenic bacteria. Cocktails of β -lactamases that can create preventive immunity could be used to develop potential vaccine candidates to prevent infection by MDR pathogens.

9. Conclusion

Antibiotic therapy and resistance acquisition by microorganisms are two sides of the same coin. Since long, there have been limited potential antibiotics that have reached the market. The only option we are left with is to opt for novel antibiotic based and non-antibiotic based strategies that can improve the efficacy of existing antibiotics. Screening for antibiotic potentiators that can restore the activity of existing antibiotics is a promising strategy for the same. Though the screening of antibiotic potentiators are equally strenuous, it is considered to be a wiser and coherent option to limit bacterial resistance evolution to even last resort antibiotics.

Author contributions

BD conceived the idea. LN collected the literature and drafted the review. MC contributed to the " β -lactam potentiators screening strategies" section and prepared Figure 3. SK and DP prepared the tables. All authors contributed to the article and approved the submitted version.

References

Abed, N., Saïd-Hassane, F., Zouhiri, F., Mougin, J., Nicolas, V., Desmaële, D., et al. (2015). An efficient system for intracellular delivery of beta-lactam antibiotics to overcome bacterial resistance. *Sci. Rep.* 5:13500. doi: 10.1038/srep13500

Ahmed, M. A. S., Sid Ahmed, M. A., Khan, F. A., Sultan, A. A., Söderquist, B., Ibrahim, E. B., et al. (2020). β -Lactamase-mediated resistance in MDR-Pseudomonas aeruginosa from Qatar. *Antimicrob. Resist. Infect. Control* 9:170. doi: 10.1186/s13756-020-00838-y

Akhtar, A., Fatima, N., and Khan, H. M. (2022). "Beta-lactamases and their classification: an overview" in *Beta-lactam resistance in gram-negative bacteria*. eds. M. Shahid, A. Singh and H. Sami (Singapore: Springer)

Akova, M. (2008). Sulbactam-containing beta-lactamase inhibitor combinations. *Clin. Microbiol. Infect.* 14, 185–188. doi: 10.1111/j.1469-0691.2007.01847.x

Aminov, R. (2022). Metabolomics in antimicrobial drug discovery. *Expert Opin. Drug Discov.* 17, 1047–1059. doi: 10.1080/17460441.2022.2113774

Ando, M., Kamei, R., Komagoe, K., Inoue, T., Yamada, K., and Katsu, T. (2012). In situ potentiometric method to evaluate bacterial outer membrane-permeabilizing ability of drugs: example using antiprotozoal diamidines. *J. Microbiol. Methods* 91, 497–500. doi: 10.1016/j.mimet.2012.09.033

Arakawa, Y., Murakami, M., Suzuki, K., Ito, H., Wacharotayankun, R., Ohsuka, S., et al. (1995). A novel integron-like element carrying the metallo-β-lactamase gene bla_{IMP}. *Antimicrob. Agents Chemother.* 39, 1612–1615. doi: 10.1128/AAC.39.7.1612

Arnold, R. S., Thom, K. A., Sharma, S., Phillips, M., Kristie Johnson, J., and Morgan, D. J. (2011). Emergence of *Klebsiella pneumoniae* carbapenemase-producing bacteria. *South. Med. J.* 104, 40–45. doi: 10.1097/SMJ.0b013e3181fd7d5a

Aunins, T. R., Erickson, K. E., and Chatterjee, A. (2020). Transcriptome-based design of antisense inhibitors potentiates carbapenem efficacy in CRE

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmicb.2022.1092556/ full#supplementary-material

Escherichia coli. Proc. Natl. Acad. Sci. U. S. A. 117, 30699-30709. doi: 10.1073/ pnas.1922187117

Benson, J. M., and Nahata, M. C. (1988). Sulbactam/ampicillin, a new Beta-lactamase inhibitor/Beta-lactam antibiotic combination. *Drug Intell. Clin. Pharm.* 22, 534–541. doi: 10.1177/106002808802200702

Bhadra, P., Yan, J., Li, J., Fong, S., and Siu, S. W. I. (2018). AmPEP: sequence-based prediction of antimicrobial peptides using distribution patterns of amino acid properties and random forest. *Sci. Rep.* 8:1697. doi: 10.1038/s41598-018-19752-w

Blais, J., Lopez, S., Li, C., Ruzin, A., Ranjitkar, S., Dean, C. R., et al. (2018). In vitro activity of LYS228, a novel Monobactam antibiotic, against multidrug-resistant Enterobacteriaceae. *Antimicrob. Agents Chemother.* 62:e00552-18. doi: 10.1128/aac.00552-18

Blizzard, T. A., Chen, H., Kim, S., Wu, J., Bodner, R., Gude, C., et al. (2014). Discovery of MK-7655, a β -lactamase inhibitor for combination with Primaxin[®]. *Bioorg. Med. Chem. Lett.* 24, 780–785. doi: 10.1016/j.bmcl.2013.12.101

Bohnert, J. A., and Kern, W. V. (2005). Selected Arylpiperazines are capable of reversing multidrug resistance in *Escherichia coli* overexpressing RND efflux pumps. *Antimicrob. Agents Chemother.* 49, 849–852. doi: 10.1128/aac.49.2.849-852.2005

Bonnefoy, A. (2004). In vitro activity of AVE1330A, an innovative broad-Spectrum non-lactam β -lactamase inhibitor. J. Antimicrob. Chemother. 54, 410–417. doi: 10.1093/jac/dkh358

Bornet, C., Chollet, R., Malléa, M., Chevalier, J., Davin-Regli, A., Pagès, J.-M., et al. (2003). Imipenem and expression of multidrug efflux pump in *Enterobacter* aerogenes. Biochem. Biophys. Res. Commun. 301, 985–990. doi: 10.1016/S0006-291X(03)00074-3

Both, A., Büttner, H., Huang, J., Perbandt, M., Campos, C. B., Christner, M., et al. (2017). Emergence of ceftazidime/avibactam non-susceptibility in an MDR *Klebsiella* pneumoniae isolate. J. Antimicrob. Chemother. 72, 2483–2488. doi: 10.1093/jac/dkx179

Brogden, R. N., and Heel, R. C. (1986). Aztreonam. A review of its antibacterial activity, pharmacokinetic properties and therapeutic use. *Drugs* 31, 96–130. doi: 10.2165/00003495-198631020-00002

Burton, H. S., and Abraham, E. P. (1951). Isolation of antibiotics from a species of Cephalosporium. Cephalosporins P1, P2, P3, P4 and P5. *Biochem. J.* 50, 168–174. doi: 10.1042/bj0500168

Bush, K. (1988). Beta-lactamase inhibitors from laboratory to clinic. *Clin. Microbiol. Rev.* 1, 109–123. doi: 10.1128/CMR.1.1.109

Bush, K. (2015). A resurgence of β -lactamase inhibitor combinations effective against multidrug-resistant Gram-negative pathogens. *Int. J. Antimicrob. Agents* 46, 483–493. doi: 10.1016/j.ijantimicag.2015.08.011

Bush, K., and Bradford, P. A. (2016). β-Lactams and β-lactamase inhibitors: an overview. *Cold Spring Harb. Perspect. Med.* 6:a025247. doi: 10.1101/cshperspect.a025247

Bush, K., and Jacoby, G. A. (2010). Updated functional classification of betalactamases. Antimicrob. Agents Chemother. 54, 969–976. doi: 10.1128/AAC.01009-09

Bush, K., and Macielag, M. J. (2010). New β -lactam antibiotics and β -lactamase inhibitors. *Expert Opin. Ther. Pat.* 20, 1277–1293. doi: 10.1517/13543776.2010.515588

Butiuc-Keul, A., Farkas, A., Carpa, R., and Iordache, D. (2022). CRISPR-Cas system: the powerful modulator of accessory genomes in prokaryotes. *Microb. Physiol.* 32, 2–17. doi: 10.1159/000516643

Campoli-Richards, D. M., and Brogden, R. N. (1987). Sulbactam/ampicillin. A review of its antibacterial activity, pharmacokinetic properties, and therapeutic use. *Drugs* 33, 577–609. doi: 10.2165/00003495-198733060-00003, Erratum in: Drugs.(1987). 34(5), preceding 515

Campos, A. I., and Zampieri, M. (2019). Metabolomics-driven exploration of the chemical drug space to predict combination antimicrobial therapies. *Mol. Cell* 74, 1291–1303.e6. doi: 10.1016/j.molcel.2019.04.001

Cassiano, C., Esposito, R., Tosco, A., Casapullo, A., Mozzicafreddo, M., Tringali, C., et al. (2019). Chemical proteomics-guided identification of a novel biological target of the bioactive Neolignan Magnolol. *Front. Chem.* 7:53. doi: 10.3389/fchem.2019.00053

Casteels, R., and Login, I. S. (1983). Reserpine has a direct action as a calcium antagonist on mammalian smooth muscle cells. *J. Physiol.* 340, 403–414. doi: 10.1113/jphysiol.1983.sp014769

Chain, E. (1979). The early years of the penicillin discovery. *Trends Pharmacol. Sci.* 1, 6–11. doi: 10.1016/0165-6147(79)90004-x

Chandra Mohana, N., Yashavantha, R. H. C., Rakshith, D., Mithun, P. R., Nuthan, B. R., and Satish, S. (2018). Omics based approach for biodiscovery of microbial natural products in antibiotic resistance era. *J. Genet. Eng. Biotechnol.* 16, 1–8. doi: 10.1016/j. jgeb.2018.01.006

Chaudhary, M., Mir, M. A., and Ayub, S. G.Protocol 06 Group (2017). Safety and efficacy of a novel drug elores (ceftriaxone+sulbactam+disodium edetate) in the management of multi-drug resistant bacterial infections in tertiary care centers: a post-marketing surveillance study. *Braz. J. Infect. Dis.* 21, 408–417. doi: 10.1016/j. bid.2017.02.007

Chawla, M., Verma, J., Gupta, R., and Das, B. (2022). Antibiotic Potentiators against multidrug-resistant bacteria: discovery, development, and clinical relevance. *Front. Microbiol.* 13:887251. doi: 10.3389/fmicb.2022.887251

Chen, Q., Zhou, W., Qian, C., Shen, K., Zhu, X., Zhou, D., et al. (2019). OXA-830, a Novel Chromosomally Encoded Extended-Spectrum Class D β -Lactamase in Aeromonas simiae. *Front Microbiol.* 10:2732. doi: 10.3389/fmicb.2019.02732

Chesnel, L., Zapun, A., Mouz, N., Dideberg, O., and Vernet, T. (2002). Increase of the deacylation rate of PBP2x from *Streptococcus pneumoniae* by single point mutations mimicking the class a beta-lactamases. *Eur. J. Biochem.* 269, 1678–1683. doi: 10.1046/j. 1432-1327.2002.02815.x

Chiou, J., Wan, S., Chan, K.-F., So, P.-K., He, D., Chan, E. W.-C., et al. (2015). Ebselen as a potent covalent inhibitor of New Delhi metallo- β -lactamase (NDM-1). *Chem Comm.* 51, 9543–9546. doi: 10.1039/c5cc02594j

Choi, U., and Lee, C.-R. (2019). Distinct roles of outer membrane Porins in antibiotic resistance and membrane integrity in *Escherichia coli. Front. Microbiol.* 10:953. doi: 10.3389/fmicb.2019.00953

Chung, M., Kim, C. K., Conceição, T., Aires-De-Sousa, M., De Lencastre, H., and Tomasz, A. (2016). Heterogeneous oxacillin-resistant phenotypes and production of PBP2A by oxacillin-susceptible/mec a-positive MRSA strains from Africa. *J. Antimicrob. Chemother.* 71, 2804–2809. doi: 10.1093/jac/dkw209

Ciofu, O., Beveridge, T. J., Kadurugamuwa, J., Walther-Rasmussen, J., and Høiby, N. (2000). Chromosomal beta-lactamase is packaged into membrane vesicles and secreted from *Pseudomonas aeruginosa*. J. Antimicrob. Chemother. 45, 9–13. doi: 10.1093/jac/45.1.9

Claesen, J., and Bibb, M. (2010). Genome mining and genetic analysis of cypemycin biosynthesis reveal an unusual class of post-translationally modified peptides. *Proc. Natl. Acad. Sci. U. S. A.* 107, 16297–16302. doi: 10.1073/pnas.1008608107

Collia, D., Bannister, T. D., Tan, H., Jin, S., Langaee, T., Shumate, J., et al. (2018). A rapid phenotypic whole-cell screening approach for the identification of small-molecule inhibitors that counter β -lactamase resistance in *Pseudomonas aeruginosa. SLAS Discov.* 23, 55–64. doi: 10.1177/2472555217728489

Corvec, S., Poirel, L., Naas, T., Drugeon, H., and Nordmann, P. (2007). Genetics and expression of the Carbapenem-hydrolyzing Oxacillinase gene Bla _{0XA-23} in *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* 51, 1530–1533. doi: 10.1128/aac.01132-06

Cunha, B. R. D., da Cunha, B. R., Fonseca, L. P., and Calado, C. R. C. (2021). Simultaneous elucidation of antibiotic mechanism of action and potency with high-throughput Fourier-transform infrared (FTIR) spectroscopy and machine learning. *Appl. Microbiol. Biotechnol.* 105, 1269–1286. doi: 10.1007/s00253-021-11102-7

Dalhoff, A. (2021). Are antibacterial effects of non-antibiotic drugs random or purposeful because of a common evolutionary origin of bacterial and mammalian targets? *Infection* 49, 569–589. doi: 10.1007/s15010-020-01547-9. Epub 2020 Dec 15

Damlin, A., Sharma, M., Marrone, G., and Stålsby Lundborg, C. (2020). Antibiotic prescribing among patients with severe infectious diseases in two private sector hospitals in Central India – a time series analysis over 10 years. *BMC Infect. Dis.* 20:340. doi: 10.1186/s12879-020-05059-7

De Rosa, M., Verdino, A., Soriente, A., and Marabotti, A. (2021). The odd couple(s): an overview of Beta-lactam antibiotics bearing more than one Pharmacophoric group. *Int. J. Mol. Sci.* 22:617. doi: 10.3390/ijms22020617

Donia, M. S., Cimermancic, P., Schulze, C. J., Wieland Brown, L. C., Martin, J., Mitreva, M., et al. (2014). A systematic analysis of biosynthetic gene clusters in the human microbiome reveals a common family of antibiotics. *Cells* 158, 1402–1414. doi: 10.1016/j.cell.2014.08.032

Doumith, M., Ellington, M. J., Livermore, D. M., and Woodford, N. (2009). Molecular mechanisms disrupting porin expression in ertapenem-resistant Klebsiella and *Enterobacter* spp. clinical isolates from the UK. *J. Antimicrob. Chemother.* 63, 659–667. doi: 10.1093/jac/dkp029

Durand-Réville, T. F., Comita-Prevoir, J., Zhang, J., Xiaoyun, W., May-Dracka, T. L., Jan, A. C., et al. (2020). Discovery of an orally available Diazabicyclooctane inhibitor (ETX 0282) of class A, C, and D serine β -lactamases. *J. Med. Chem.* 63, 12511–12525. doi: 10.1021/acs.jmedchem.0c00579

Ehmann, D. E., Jahić, H., Ross, P. L., Gu, R.-F., Hu, J., Kern, G., et al. (2012). Avibactam is a covalent, reversible, non- β -lactam β -lactamase inhibitor. *Proc. Natl. Acad. Sci. U. S. A.* 109, 11663–11668. doi: 10.1073/pnas.1205073109

Everett, M., Sprynski, N., Coelho, A., Castandet, J., Bayet, M., Bougnon, J., et al. (2018). Discovery of a novel Metallo- β -lactamase inhibitor that potentiates Meropenem activity against Carbapenem-resistant Enterobacteriaceae. *Antimicrob. Agents Chemother.* 62, e00074–e00018. doi: 10.1128/AAC.00074-18

Fan, P., Ma, Z., Partow, A. J., Kim, M., Shoemaker, G. M., Tan, R., et al. (2022). A novel combination therapy for multidrug resistant pathogens using chitosan nanoparticles loaded with β -lactam antibiotics and β -lactamase inhibitors. *Int. J. Biol. Macromol.* 195, 506–514. doi: 10.1016/i.iibiomac.2021.12.035

Farrag, H. A., Abdallah, N., Shehata, M. M. K., and Awad, E. M. (2019). Natural outer membrane permeabilizers boost antibiotic action against irradiated resistant bacteria. *J. Biomedl. Sci.* 26:69. doi: 10.1186/s12929-019-0561-6

Farrag, H., Shehata, M., Abdallah, N., and Awad, E. (2015). Synergistic effect of certain natural permeabilizers with antimicrobial agents on outer membrane of some multidrug gram-negative pathogenic bacteria. *J. Sci. Res. Sci.* 32, 107–121. doi: 10.21608/jsrs.2015.18358

FDA (2019). Drug trial snapshot: Recarbrio. Available at: https://www.fda.gov/drugs/resources-information-approved-drugs/drug-trial-snapshot-recarbrio

Feretzakis, G., Loupelis, E., Sakagianni, A., Skarmoutsou, N., Michelidou, S., Velentza, A., et al. (2019). A 2-year single-Centre audit on antibiotic resistance of *Pseudomonas aeruginosa, Acinetobacter baumannii* and *Klebsiella pneumoniae* strains from an intensive care unit and other wards in a general public hospital in Greece. *Antibiotics* 8:62. doi: 10.3390/antibiotics8020062

Fernández, A., Pérez, A., Ayala, J. A., Mallo, S., Rumbo-Feal, S., Tomás, M., et al. (2012). Expression of OXA-type and SFO-1 β -lactamases induces changes in peptidoglycan composition and affects bacterial fitness. *Antimicrob. Agents Chemother.* 56, 1877–1884. doi: 10.1128/AAC.05402-11

Ferreira, R. L., da Silva, B. C. M., Rezende, G. S., Nakamura-Silva, R., Pitondo-Silva, A., Campanini, E. B., et al. (2019). High prevalence of multidrug-resistant *Klebsiella pneumoniae* harboring several virulence and β -lactamase encoding genes in a Brazilian intensive care unit. *Front. Microbiol.* 9:3198. doi: 10.3389/fmicb.2018.03198

Fleurbaaij, F., Henneman, A. A., Corver, J., Knetsch, C. W., Smits, W. K., Nauta, S. T., et al. (2018). Proteomic identification of Axc, a novel beta-lactamase with carbapenemase activity in a meropenem-resistant clinical isolate of *Achromobacter xylosoxidans. Sci. Rep.* 8:8181. doi: 10.1038/s41598-018-26079-z

Furniss, R. C. D., Kaderabkova, N., Barker, D., Bernal, P., Maslova, E., Antwi, A. A. A., et al. (2022). Breaking antimicrobial resistance by disrupting extra-cytoplasmic protein folding. *eLife* 11:57974. doi: 10.7554/elife.57974

Fursova, N. K., Astashkin, E. I., Ershova, O. N., Aleksandrova, I. A., Savin, I. A., Novikova, T. S., et al. (2021). Multidrug-resistant *Klebsiella pneumoniae* causing severe infections in the neuro-ICU. *Antibiotics (Basel)*. 10:979. doi: 10.3390/antibiotics10080979 Gagetti, P., Pasteran, F., Martinez, M. P., Fatouraei, M., Gu, J., Fernandez, R., et al. (2016). Modeling Meropenem treatment, alone and in combination with Daptomycin, for KPC-producing *Klebsiella pneumoniae* strains with unusually low Carbapenem MICs. *Antimicrob. Agents Chemother.* 60, 5047–5050. doi: 10.1128/AAC.00168-16

Giacometti, A., Cirioni, O., Kamysz, W., D'Amato, G., Silvestri, C., Del Prete, M. S., et al. (2005). In vitro activity and killing effect of temporin a on nosocomial isolates of enterococcus faecalis and interactions with clinically used antibiotics. *J. Antimicrob. Chemother.* 55, 272–274. doi: 10.1093/jac/dkh545

Giani, T., Conte, V., Di Pilato, V., Aschbacher, R., Weber, C., Larcher, C., et al. (2012). *Escherichia coli* from Italy producing OXA-48 carbapenemase encoded by a novel Tn1999 transposon derivative. *Antimicrob. Agents Chemother.* 56, 2211–2213. doi: 10.1128/AAC.00035-12

Giddins, M. J., Macesic, N., Annavajhala, M. K., Stump, S., Khan, S., McConville, T. H., et al. (2018). Successive emergence of ceftazidime-avibactam resistance through distinct genomic adaptations in-harboring *Klebsiella pneumoniae* sequence type 307 isolates. *Antimicrob. Agents Chemother.* 62, e02101–e02117. doi: 10.1128/AAC.02101-17

Gin, A., Dilay, L., Karlowsky, J. A., Walkty, A., Rubinstein, E., and Zhanel, G. G. (2007). Piperacillin-tazobactam: a beta-lactam/beta-lactamase inhibitor combination. *Expert Rev. Anti-Infect. Ther.* 5, 365–383. doi: 10.1586/14787210.5.3.365

Glen, K. A., and Lamont, I. L. (2021). β-Lactam resistance in *Pseudomonas aeruginosa*: current status, Future Prospects. *Pathogens* 10:1638. doi: 10.3390/pathogens10121638

Goa, K. L., and Noble, S. (2003). Panipenem/betamipron. Drugs 63, 913-925. doi: 10.2165/00003495-200363090-00005

Graham, D. W., Knapp, C. W., Christensen, B. T., McCluskey, S., and Dolfing, J. (2016). Appearance of β -lactam Resistance Genes in Agricultural Soils and Clinical Isolates over the 20th Century. *Sci Rep.* 6:21550. doi: 10.1038/srep21550

Greer, N. D. (2008). Doripenem (Doribax): the newest addition to the carbapenems. Proc. (Bayl. Univ. Med. Cent). 21, 337-341. doi: 10.1080/08998280.2008.11928422

Groman, R. P. (2009). "Chapter 200- miscellaneous antibiotics" in Small animal critical care medicine. eds. D. C. Silverstein and K. Hopper (St. Louis, MO: W.B. Saunders), 845–849.

Guan, S., Zhong, L., Yu, H., Wang, L., Jin, Y., Liu, J., et al. (2022). Molecular docking and proteomics reveals the synergistic antibacterial mechanism of theaflavin with β -lactam antibiotics against MRSA. *Front. Microbiol.* 13:993430. doi: 10.3389/ fmicb.2022.993430

Gwynn, M. N., Box, S. J., Brown, A. G., and Gilpin, M. L. (1988). MM 42842, a new member of the monobactam family produced by pseudomonas cocovenenans. I. Identification of the producing organism. *J. Antibiot. (Tokyo)* 41, 1–6. doi: 10.7164/ antibiotics.41.1. PMID: 3346181

Hamblin, M. R. (2016). Antimicrobial photodynamic inactivation: a bright new technique to kill resistant microbes. *Curr. Opin. Microbiol.* 33, 67–73. doi: 10.1016/j. mib.2016.06.008

Hamilton-Miller, J. M. (1999). Beta-lactams: variations on a chemical theme, with some surprising biological results. *J. Antimicrob. Chemother.* 44, 729–734. doi: 10.1093/jac/44.6.729

Hao, M., He, Y., Zhang, H., Liao, X. P., Liu, Y. H., Sun, J., et al. (2020). CRISPR-Cas9mediated Carbapenemase gene and Plasmid curing in Carbapenem-resistant Enterobacteriaceae. *Antimicrob. Agents Chemother.* 64, e00843–e00820. doi: 10.1128/ AAC.00843-20

Hecker, S. J., Raja Reddy, K., Totrov, M., Hirst, G. C., Lomovskaya, O., Griffith, D. C., et al. (2015). Discovery of a cyclic Boronic acid β -lactamase inhibitor (RPX7009) with utility vs class A serine Carbapenemases. *J. Med. Chem.* 58, 3682–3692. doi: 10.1021/acs. jmedchem.5b00127

Hinchliffe, P., González, M. M., Mojica, M. F., González, J. M., Castillo, V., Saiz, C., et al. (2016). Cross-class metallo-β-lactamase inhibition by bisthiazolidines reveals multiple binding modes. *Proc. Natl. Acad. Sci. U. S. A.* 113, E3745–E3754. doi: 10.1073/ pnas.1601368113

Huang, P.-J. J., Pautler, R., Shanmugaraj, J., Labbé, G., and Liu, J. (2015). Inhibiting the VIM-2 Metallo-β-lactamase by graphene oxide and carbon nanotubes. *ACS Appl. Mater. Interfaces* 7, 9898–9903. doi: 10.1021/acsami.5b01954

Hulen, C., Racine, P. J., Chevalier, S., Feuilloley, M., and Lomri, N. E. (2020). Identification of the PA1113 gene product as an ABC transporter involved in the uptake of Carbenicillin in *Pseudomonas aeruginosa* PAO1. *Antibiotics (Basel)*. 9:596. doi: 10.3390/antibiotics9090596

Hull, D. M., Harrell, E., van Vliet, A. H. M., Correa, M., and Thakur, S. (2021). Antimicrobial resistance and interspecies gene transfer in campylobacter coli and campylobacter jejuni isolated from food animals, poultry processing, and retail meat in North Carolina, 2018–2019. *PLoS One* 16:e0246571. doi: 10.1371/journal.pone.0246571

Infectious Diseases Society of America (IDSA) (2011). Combating antimicrobial resistance: policy recommendations to save lives. *Clin. Infect. Dis.* 52, S397–S428. doi: 10.1093/cid/cir153

Jaffe, A., Chabbert, Y. A., and Semonin, O. (1982). Role of porin proteins OmpF and OmpC in the permeation of beta-lactams. *Antimicrob. Agents Chemother.* 22, 942–948. doi: 10.1128/AAC.22.6.942

Kapil, A., Dahiya, S., Malik, R., Sharma, P., Sashi, A., Lodha, R., et al. (2019). Current antibiotic use in the treatment of enteric fever in children. *Indian J. Med. Res.* 149, 263–269. doi: 10.4103/ijmr.ijmr_199_18

Karlowsky, J. A., Hackel, M. A., Bouchillon, S. K., and Sahm, D. F. (2020). Activity of WCK 5222 (Cefepime-Zidebactam) against worldwide collected gram-negative bacilli not susceptible to Carbapenems. *Antimicrob. Agents Chemother.* 64, e01432–e01420. doi: 10.1128/AAC.01432-20

Karumathil, D. P., Nair, M. S., Gaffney, J., Kollanoor-Johny, A., and Venkitanarayanan, K. (2018). Trans-Cinnamaldehyde and eugenol increase *Acinetobacter baumannii* sensitivity to Beta-lactam antibiotics. *Front. Microbiol.* 9:1011. doi: 10.3389/fmicb.2018.01011

Khalifa, S. M., Abd El-Aziz, A. M., Hassan, R., and Abdelmegeed, E. S. (2021). β -Lactam resistance associated with β -lactamase production and porin alteration in clinical isolates of *E. coli* and *K. pneumoniae. PLoS One* 16:e0251594. doi: 10.1371/journal.pone.0251594

Kim, J., Bae, I. K., Jeong, S. H., Chang, C. L., Lee, C. H., and Lee, K. (2011). Characterization of IncF plasmids carrying the blaCTX-M-14 gene in clinical isolates of *Escherichia coli* from Korea. *J. Antimicrob. Chemother.* 66, 1263–1268. doi: 10.1093/ jac/dkr106

Kim, C., Hesek, D., Lee, M., and Mobashery, S. (2018). Potentiation of the activity of β -lactam antibiotics by farnesol and its derivatives. *Bioorg. Med. Chem. Lett.* 28, 642–645. doi: 10.1016/j.bmcl.2018.01.028

Kim, S. W., Lee, J. S., Park, S. B., Lee, A. R., Jung, J. W., Chun, J. H., et al. (2020). The importance of Porins and β -lactamase in outer membrane vesicles on the hydrolysis of β -lactam antibiotics. *Int. J. Mol. Sci.* 21:2822. doi: 10.3390/ijms21082822

Kim, C., Mwangi, M., Chung, M., Milheiriço, C., de Lencastre, H., and Tomasz, A. (2013). The mechanism of heterogeneous beta-lactam resistance in MRSA: key role of the stringent stress response. *PLoS One* 8:e82814. doi: 10.1371/journal.pone.0082814. Erratum in: PLoS One. 2014; 9(1). doi:10.1371/annotation/7775dee3-7a91-4c44-9125c7634bf909e4. Milheirço, Catarina [corrected to Milheiriço, Catarina]

Kim, W., Zou, G., Hari, T. P. A., Wilt, I. K., Zhu, W., Galle, N., et al. (2019). A selective membrane-targeting repurposed antibiotic with activity against persistent methicillinresistant *Staphylococcus aureus*. *Proc. Natl. Acad. Sci. U. S. A.* 116, 16529–16534. doi: 10.1073/pnas.1904700116

King, A. M., Reid-Yu, S. A., Wang, W., King, D. T., De Pascale, G., Strynadka, N. C., et al. (2014). Aspergillomarasmine A overcomes metallo- β -lactamase antibiotic resistance. *Nature* 510, 503–506. doi: 10.1038/nature13445

Kostoulias, X., Murray, G. L., Cerqueira, G. M., Kong, J. B., Bantun, F., Mylonakis, E., et al. (2016). Impact of a Cross-kingdom signaling molecule of Candida albicans on Acinetobacter baumannii physiology. Antimicrob. Agents Chemother. 60, 161–167. doi: 10.1128/AAC.01540-15

Kotrange, H., Najda, A., Bains, A., Gruszecki, R., Chawla, P., and Tosif, M. M. (2021). Metal and metal oxide nanoparticle as a novel antibiotic carrier for the direct delivery of antibiotics. *Int. J. Mol. Sci.* 22:9596. doi: 10.3390/ijms22179596

Kulkarni, A. P., Nagvekar, V. C., Veeraraghavan, B., Warrier, A. R., Ts, D., Ahdal, J., et al. (2019). Current perspectives on treatment of gram-positive infections in India: what is the way forward? *Interdiscip. Perspect. Infect. Dis.* 2019, 1–8. doi: 10.1155/2019/7601847

Kumar, N. G., Kumar, G., Mallick, S., Ghosh, S. K., Pramanick, P., and Ghosh, A. S. (2019). Bio-surfactin stabilised silver nanoparticles exert inhibitory effect over new-Delhi metallo-beta-lactamases (NDMs) and the cells harbouring NDMs. *FEMS Microbiol. Lett.* 366:fnz 118. doi: 10.1093/femsle/fnz118

Kurosaki, H., Yamaguchi, Y., Higashi, T., Soga, K., Matsueda, S., Yumoto, H., et al. (2005). Irreversible inhibition of Metallo-β-lactamase (IMP-1) by 3-(3-Mercaptopropionylsulfanyl) propionic acid Pentafluorophenyl Ester. *Angew. Chem. Int. Ed.* 44, 3861–3864. doi: 10.1002/anie.200500835

Kwak, Y. G., Choi, S. H., Kim, T., Park, S. Y., Seo, S. H., Kim, M. B., et al. (2017). Clinical guidelines for the antibiotic treatment for community-acquired skin and soft tissue infection. *Infect. Chemother* 49, 301–325. doi: 10.3947/ic.2017.49.4.301

Lainson, J. C., Daly, S. M., Triplett, K., Johnston, S. A., Hall, P. R., and Diehnelt, C. W. (2017). Synthetic antibacterial peptide exhibits synergy with oxacillin against MRSA. *ACS Med. Chem. Lett.* 8, 853–857. doi: 10.1021/acsmedchemlett.7b00200

Lamers, R. P., Cavallari, J. F., and Burrows, L. L. (2013). The efflux inhibitor phenylalanine-arginine beta-naphthylamide (PAβN) permeabilizes the outer membrane of gram-negative bacteria. *PLoS One* 8:e60666. doi: 10.1371/journal.pone.0060666

Laws, M., Shaaban, A., and Rahman, K. M. (2019). Antibiotic resistance breakers: current approaches and future directions. *FEMS Microbiol. Rev.* 43, 490–516. doi: 10.1093/femsre/fuz014

Lazzaro, L. M., Cassisi, M., Stefani, S., and Campanile, F. (2022). Impact of PBP4 alterations on β -lactam resistance and Ceftobiprole non-susceptibility among enterococcus faecalis clinical isolates. *Front. Cell. Infect. Microbiol.* 11:816657. doi: 10.3389/fcimb.2021.816657

Lepak, A. J., Zhao, M., and Andes, D. R. (2019). WCK 5222 (Cefepime/Zidebactam) Pharmacodynamic target analysis against Metallo-β-lactamase producing in the neutropenic mouse pneumonia model. *Antimicrob. Agents Chemother.* 63, e01648– e01619. doi: 10.1128/AAC.01648-19

Li, J. Q., Gao, H., Zhai, L., Sun, L. Y., Chen, C., Chigan, J. Z., et al. (2021). Dipyridylsubstituted thiosemicarbazone as a potent broad-spectrum inhibitor of metallo-βlactamases. *Bioorg. Med. Chem.* 38:116128. doi: 10.1016/j.bmc.2021.116128 Li, B., Yao, Q., Pan, X.-C., Wang, N., Zhang, R., Li, J., et al. (2011). Artesunate enhances the antibacterial effect of {beta}-lactam antibiotics against *Escherichia coli* by increasing antibiotic accumulation via inhibition of the multidrug efflux pump system AcrAB-TolC. *J. Antimicrob. Chemother.* 66, 769–777. doi: 10.1093/jac/dkr017

Li, P., Ying, J., Yang, G., Li, A., Wang, J., Lu, J., et al. (2016). Structure-function analysis of the transmembrane protein AmpG from *Pseudomonas aeruginosa*. *PLoS One* 11:e0168060. doi: 10.1371/journal.pone.0168060

Li, J., Zhang, H., Ning, J., Sajid, A., Cheng, G., Yuan, Z., et al. (2019). The nature and epidemiology of OqxAB, a multidrug efflux pump. *Antimicrob. Resist. Infect. Control* 8:44. doi: 10.1186/s13756-019-0489-3

Li, X., Zhao, D., Li, W., Sun, J., and Zhang, X. (2022). Enzyme inhibitors: the best strategy to tackle superbug NDM-1 and its variants. *Int. J. Mol. Sci.* 23:197. doi: 10.3390/ ijms23010197

Liu, B., Trout, R. E. L., Chu, G.-H., McGarry, D., Jackson, R. W., Hamrick, J. C., et al. (2020). Discovery of Taniborbactam (VNRX-5133): a broad-Spectrum serine-and Metallo-β-lactamase inhibitor for Carbapenem-resistant bacterial infections. *J. Med. Chem.* 63, 2789–2801. doi: 10.1021/acs.jmedchem.9b01518

Liu, S., Zhou, Y., Niu, X., Wang, T., Li, J., Liu, Z., et al. (2018). Magnolol restores the activity of Meropenem against NDM-1-producing *Escherichia coli* by inhibiting the activity of Metallo-Beta-lactamase. *Cell Death Discov.* 4:28. doi: 10.1038/s41420-018-0029-6

Lomovskaya, O., Tsivkovski, R., Sun, D., Reddy, R., Totrov, M., Hecker, S., et al. (2021). QPX 7728, an ultra-broad-Spectrum B-lactamase inhibitor for intravenous and Oral therapy: overview of biochemical and microbiological characteristics. *Front. Microbiol.* 12:697180. doi: 10.3389/fmicb.2021.697180

Luci, G., Mattioli, F., Falcone, M., and Di Paolo, A. (2021). Pharmacokinetics of non-βlactam β-lactamase inhibitors. *Antibiotics (Basel)*. 10:769. doi: 10.3390/antibiotics10070769

Luo, G., Zhang, J., Wang, H., Sun, Y., Cheng, B., Xu, Z., et al. (2022). Human defensininspired discovery of peptidomimetic antibiotics. *Proc. Natl. Acad. Sci. U. S. A.* 119:e2117283119. doi: 10.1073/pnas.2117283119

Ma, B., Fang, C., Lu, L., Wang, M., Xue, X., Zhou, Y., et al. (2019). The antimicrobial peptide thanatin disrupts the bacterial outer membrane and inactivates the NDM-1 metallo-β-lactamase. *Nat. Commun.* 10:3517. doi: 10.1038/s41467-019-11503-3

Mallalieu, N. L., Winter, E., Fettner, S., Patel, K., Zwanziger, E., Attley, G., et al. (2020). Safety and pharmacokinetic characterization of Nacubactam, a novel β -lactamase inhibitor, alone and in combination with Meropenem, in healthy volunteers. *Antimicrob. Agents Chemother.* 64, e02229–e02219. doi: 10.1128/AAC.02229-19

Mandal, S. M., Migliolo, L., Silva, O. N., Fensterseifer, I. C. M., Faria-Junior, C., Dias, S. C., et al. (2014). Controlling resistant bacteria with a novel class of β -lactamase inhibitor peptides: from rational design to in vivo analyses. *Sci. Rep.* 4:6015. doi: 10.1038/srep06015

Mansbach, R. A., Leus, I. V., Mehla, J., Lopez, C. A., Walker, J. K., Rybenkov, V. V., et al. (2020). Machine learning algorithm identifies an antibiotic vocabulary for permeating gram-negative bacteria. *J. Chem. Inf. Model.* 60, 2838–2847. doi: 10.1021/acs. jcim.0c00352

Mares, M., Lim, S. H. E., Lai, K.-S., and Cristina, R.-T. (Eds.). (2021). Antimicrobial resistance: a one health perspective. IntechOpen.

Masuda, N., Sakagawa, E., Ohya, S., Gotoh, N., Tsujimoto, H., and Nishino, T. (2000). Substrate specificities of Mex AB-OprM, MexCD-OprJ, and MexXY-oprM efflux pumps in *Pseudomonas aeruginosa. Antimicrob. Agents Chemother.* 44, 3322–3327. doi: 10.1128/ AAC.44.12.3322-3327.2000

Mba, I. E., and Nweze, E. I. (2021). Nanoparticles as therapeutic options for treating multidrug-resistant bacteria: research progress, challenges, and prospects. *World J. Microbiol. Biotechnol.* 37:108. doi: 10.1007/s11274-021-03070-x

Mohammed, A. S. A., Mourad, M. I., Alsewy, F. Z., and El Moez Azzam, N. F. A. (2021). Combination of silver nanoparticles with ineffective antibiotics against extended spectrum beta-lactamases producing isolates at Alexandria Main university hospital, Egypt. *Beni-Suef Univ. J. Basic Appl. Sci.* 10:58. doi: 10.1186/s43088-021-00147-2

Mood, E. H., Goltermann, L., Brolin, C., Cavaco, L. M., Nejad, A. J., Yavari, N., et al. (2021). Antibiotic potentiation in multidrug-resistant gram-negative pathogenic bacteria by a synthetic Peptidomimetic. *ACS Infect. Dis.* 7, 2152–2163. doi: 10.1021/acsinfecdis.1c00147

Moura, E. C. C. M., Moura, E. C. C., Baeta, T., Romanelli, A., Laguri, C., Martorana, A. M., et al. (2020). Thanatin impairs lipopolysaccharide transport complex assembly by targeting LptC-LptA interaction and decreasing LptA stability. *Front. Microbiol.* 11:909. doi: 10.3389/fmicb.2020.00909

Moya, B., Barcelo, I. M., Bhagwat, S., Patel, M., Bou, G., Papp-Wallace, K. M., et al. (2017). WCK 5107 (Zidebactam) and WCK 5153 are novel inhibitors of PBP2 showing potent "β-lactam enhancer" activity against *Pseudomonas aeruginosa*, including multidrug-resistant Metallo-β-lactamase-producing high-risk clones. *Antimicrob. Agents Chemother.* 61, e02529–e02516. doi: 10.1128/AAC.02529-16

Moya, B., Dötsch, A., Juan, C., Blázquez, J., Zamorano, L., Haussler, S., et al. (2009). β -Lactam resistance response triggered by inactivation of a nonessential penicillinbinding protein. *PLoS Pathog.* 5:e1000353. doi: 10.1371/journal.ppat.1000353

Muderris, T., Durmaz, R., Ozdem, B., Dal, T., Unaldı, O., Aydogan, S., et al. (2018). Role of efflux pump and OprD porin expression in carbapenem resistance of Pseudomonas aeruginosa clinical isolates. J. Infect. Dev. Ctries. 12, 001–008. doi: 10.3855/jidc.9486

Munita, J. M., and Arias, C. A. (2016). Mechanisms of antibiotic resistance. *Microbiol. Spectr.* 4. doi: 10.1128/microbiolspec.VMBF-0016-2015

Munoz-Price, L. S., Poirel, L., Bonomo, R. A., Schwaber, M. J., Daikos, G. L., Cormican, M., et al. (2013). Clinical epidemiology of the global expansion of *Klebsiella pneumoniae* carbapenemases. *Lancet Infect. Dis.* 13, 785–796. doi: 10.1016/ S1473-3099(13)70190-7

Naas, T., Oueslati, S., Bonnin, R. A., Dabos, M. L., Zavala, A., Dortet, L., et al. (2017). Beta-lactamase database (BLDB) – structure and function. *J. Enzyme Inhib. Med. Chem.* 32, 917–919. doi: 10.1080/14756366.2017.1344235

Nagshetty, K., Shilpa, B. M., Patil, S. A., Shivannavar, C. T., and Manjula, N. G. (2021). An overview of extended Spectrum Beta lactamases and Metallo Beta lactamases. *Adv. Microbiol.* 11, 37–62. doi: 10.4236/aim.2021.111004

Narendrakumar, L., Chandrika, S. K., and Thomas, S. (2020). Adaptive laboratory evolution of vibrio cholerae to doxycycline associated with spontaneous mutation. *Int. J. Antimicrob. Agents* 56:106097. doi: 10.1016/j.ijantimicag.2020.106097

Necati Hakyemez, I., Kucukbayrak, A., Tas, T., Burcu Yikilgan, A., Akkaya, A., Yasayacak, A., et al. (2013). Nosocomial Acinetobacter baumannii infections and changing antibiotic resistance. *Pak. J. Med. Sci.* 29, 1245–1248. doi: 10.12669/ pjms.295.3885

Neu, H. C. (1985). Relation of structural properties of beta-lactam antibiotics to antibacterial activity. *Am. J. Med.* 79, 2–13. doi: 10.1016/0002-9343(85)90254-2

Nodari, C. S., Ribeiro, V. B., and Barth, A. L. (2015). Imipenem heteroresistance: high prevalence among Enterobacteriaceae *Klebsiella pneumoniae* carbapenemase producers. *J. Med. Microbiol.* 64, 124–126. doi: 10.1099/jmm.0.081869-0

Novak, R., and Shlaes, D. M. (2010). The pleuromutilin antibiotics: a new class for human use. *Curr. Opin. Investig. Drugs* 11, 182–191.

OECD and World Health Organization (2020). *Challenges to tackling antimicrobial resistance economic and policy responses: economic and policy responses.* Paris, France: OECD Publishing.

Oliveira, D. M. P. D., De Oliveira, D. M. P., Forde, B. M., Kidd, T. J., Harris, P. N. A., Schembri, M. A., et al. (2020). Antimicrobial resistance in ESKAPE pathogens. *Clin. Microbiol. Rev.* 33, e00181–e00119. doi: 10.1128/cmr.00181-19

O'Rourke, A., Beyhan, S., Choi, Y., Morales, P., Chan, A. P., Espinoza, J. L., et al. (2020). Mechanism-of-action classification of antibiotics by global transcriptome profiling. *Antimicrob. Agents Chemother.* 64, e01207–e01219. doi: 10.1128/ AAC.01207-19

Owen, J. G., Charlop-Powers, Z., Smith, A. G., Ternei, M. A., Calle, P. Y., Reddy, B. V., et al. (2015). Multiplexed metagenome mining using short DNA sequence tags facilitates targeted discovery of epoxyketone proteasome inhibitors. *Proc. Natl. Acad. Sci. U. S. A.* 112, 4221–4226. doi: 10.1073/pnas.1501124112

Page, M. G. P. (2002). Beta-Lactamases. Encycl. Mol. Biol. doi: 10.1002/047120918x. emb0150

Pagel, M., Simonet, V., Li, J., Lallemand, M., Lauman, B., and Delcour, A. H. (2007). Phenotypic characterization of pore mutants of the vibrio cholerae Porin OmpU. *J. Bacteriol.* 189, 8593–8600. doi: 10.1128/jb.01163-07

Papp-Wallace, K. M. (2019). The latest advances in β -lactam/ β -lactamase inhibitor combinations for the treatment of gram-negative bacterial infections. *Expert. Opin. Pharmacother.* 20, 2169–2184. doi: 10.1080/14656566.2019.1660772

Papp-Wallace, K. M., Bethel, C. R., Caillon, J., Barnes, M. D., Potel, G., Bajaksouzian, S., et al. (2019). Beyond piperacillin-Tazobactam: Cefepime and AA1101 as a potent β -lactam- β -lactamase inhibitor combination. *Antimicrob. Agents Chemother.* 63, e00105–e00119. doi: 10.1128/AAC.00105-19

Papp-Wallace, K. M., Endimiani, A., Taracila, M. A., and Bonomo, R. A. (2011). Carbapenems: past, present, and future. *Antimicrob. Agents Chemother.* 55, 4943–4960. doi: 10.1128/AAC.00296-11

Parvaiz, N., Ahmad, F., Yu, W., MacKerell, A. D., and Azam, S. S. (2021). Discovery of beta-lactamase CMY-10 inhibitors for combination therapy against multi-drug resistant Enterobacteriaceae. *PLoS One* 16:e0244967. doi: 10.1371/journal.pone.0244967

Paul, M., Grozinsky-Glasberg, S., Soares-Weiser, K., and Leibovici, L. (2006). Beta lactam antibiotic monotherapy versus beta lactam-aminoglycoside antibiotic combination therapy for sepsis. *Cochrane Database Syst. Rev.* 2014:CD003344. doi: 10.1002/14651858.cd003344.pub2

Perry, C. M., and Ibbotson, T. (2002). Biapenem. Drugs 62, 2221-2234. doi: 10.2165/00003495-200262150-00005

Pitout, J. D. D., Peirano, G., Kock, M. M., Strydom, K. A., and Matsumura, Y. (2019). The global ascendency of OXA-48-type Carbapenemases. *Clin. Microbiol. Rev.* 33, e00102–e00119. doi: 10.1128/CMR.00102-19

Poirel, L., Naas, T., and Nordmann, P. (2010). Diversity, epidemiology, and genetics of class D β -lactamases. Antimicrob. Agents Chemother. 54, 24–38. doi: 10.1128/ aac.01512-08

Poirel, L., and Nordmann, P. (2006). Genetic structures at the origin of acquisition and expression of the Carbapenem-hydrolyzing Oxacillinase gene Bla _{OXA-58} in *Acinetobacter*

baumannii. Antimicrob. Agents Chemother. 50, 1442–1448. doi: 10.1128/ aac.50.4.1442-1448.2006

Raible, K. M., Sen, B., Law, N., Bias, T. E., Emery, C. L., Ehrlich, G. D., et al. (2017). Molecular characterization of β -lactamase genes in clinical isolates of carbapenem-resistant *Acinetobacter baumannii. Ann. Clin. Microbiol. Antimicrob.* 16:75. doi: 10.1186/s12941-017-0248-3

Reading, C., and Cole, M. (1977). Clavulanic acid: a beta-lactamase-inhibiting betalactam from *Streptomyces clavuligerus*. *Antimicrob*. *Agents Chemother*. 11, 852–857. doi: 10.1128/AAC.11.5.852

Rezzoagli, C., Archetti, M., Mignot, I., Baumgartner, M., and Kümmerli, R. (2020). Combining antibiotics with antivirulence compounds can have synergistic effects and reverse selection for antibiotic resistance in *Pseudomonas aeruginosa*. *PLoS Biol*. 18:e3000805. doi: 10.1371/journal.pbio.3000805

Rigatto, M. H., Ramos, F., Barros, A., Pedroso, S., Guasso, I., Gonçalves, L., et al. (2022). Double-, single-and none-carbapenem-containing regimens for the treatment of carbapenem-resistant Enterobacterales (CRE) bloodstream infections: a retrospective cohort. J. Antimicrob. Chemother. 77, 3118–3125. doi: 10.1093/jac/dkac292

Rodríguez, I., Thomas, K., Van Essen, A., Schink, A.-K., Day, M., Chattaway, M., et al. (2014). Chromosomal location of bla_{CTX-M} genes in clinical isolates of *Escherichia coli* from Germany, the Netherlands and the UK. *Int. J. Antimicrob. Agents* 43, 553–557. doi: 10.1016/j.ijantimicag.2014.02.019

Rolinson, G. N. (1991). Evolution of β-lactamase inhibitors. *Clin. Infect. Dis.* 13, S727–S732. doi: 10.1093/clinids/13.supplement_9.s727

Sakoulas, G., Rose, W., Berti, A., Olson, J., Munguia, J., Nonejuie, P., et al. (2017). Classical β -lactamase inhibitors potentiate the activity of Daptomycin against methicillin-resistant *Staphylococcus aureus* and Colistin against *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* 61, e01745–e01716. doi: 10.1128/AAC.01745-16

Salari-Jazi, A., Mahnam, K., Sadeghi, P., Damavandi, M. S., and Faghri, J. (2021). Discovery of potential inhibitors against New Delhi metallo- β -lactamase-1 from natural compounds: in silico-based methods. *Sci. Rep.* 11:2390. doi: 10.1038/s41598-021-82009-6

Santana-Pereira, A. L. R., Sandoval-Powers, M., Monsma, S., Zhou, J., Santos, S. R., Mead, D. A., et al. (2020). Discovery of novel biosynthetic gene cluster diversity from a soil metagenomic library. *Front. Microbiol.* 11:585398. doi: 10.3389/fmicb.2020.585398

Schuster, S., Kohler, S., Buck, A., Dambacher, C., König, A., Bohnert, J. A., et al. (2014). Random mutagenesis of the multidrug transporter AcrB from *Escherichia coli* for identification of putative target residues of efflux pump inhibitors. *Antimicrob. Agents Chemother.* 58, 6870–6878. doi: 10.1128/aac.03775-14

Sevier, C. S., Kadokura, H., Tam, V. C., Beckwith, J., Fass, D., and Kaiser, C. A. (2005). The prokaryotic enzyme DsbB may share key structural features with eukaryotic disulfide bond forming oxidoreductases. *Protein Sci.* 14, 1630–1642. doi: 10.1110/ps.051355705

Shapiro, A. B., Moussa, S. H., McLeod, S. M., Durand-Réville, T., and Miller, A. A. (2021). Durlobactam, a new Diazabicyclooctane β -lactamase inhibitor for the treatment of Acinetobacter infections in combination with Sulbactam. *Front. Microbiol.* 12:709974. doi: 10.3389/fmicb.2021.709974

Shrivastav, A., Sharma, R. K., Shrivastava, N., Gautam, V., and Jain, S. (2019). Study of inhibitory potential and percent inhibition of oil of *Syzygium aromaticum* and leaves of *Ocimum sanctum* on ESBL enzyme from *Escherichia coli* in broilers of Jabalpur. *Indian J. Pharmacol.* 51, 337–342. doi: 10.4103/ijp.ijp_87_17

Siriyong, T., Murray, R. M., Bidgood, L. E., Young, S. A., Wright, F., Parcell, B. J., et al. (2019). Dual β -lactam combination therapy for multi-drug resistant *Pseudomonas aeruginosa* infection: enhanced efficacy in vivo and comparison with monotherapies of penicillin-binding protein inhibition. *Sci. Rep.* 9:9098. doi: 10.1038/s41598-019-45550-z

Siriyong, T., Srimanote, P., Chusri, S., Yingyongnarongkul, B.-E., Suaisom, C., Tipmanee, V., et al. (2017). Conessine as a novel inhibitor of multidrug efflux pump systems in *Pseudomonas aeruginosa*. *BMC Complement. Altern. Med.* 17:405. doi: 10.1186/s12906-017-1913-y

Skredenske, J. M., Koppolu, V., Kolin, A., Deng, J., Kettle, B., Taylor, B., et al. (2013). Identification of a small-molecule inhibitor of bacterial AraC family activators. *J. Biomol. Screen.* 18, 588–598. doi: 10.1177/1087057112474690

Smani, Y., Fàbrega, A., Roca, I., Sánchez-Encinales, V., Vila, J., and Pachón, J. (2014). Role of OmpA in the multidrug resistance phenotype of *Acinetobacter baumannii*. *Antimicrob. Agents Chemother.* 58, 1806–1808. doi: 10.1128/AAC.02101-13

Smith, J. T., Hamilton-Miller, J. M. T., and Knox, R. (1962). Isoxazolyl Penicillins and penicillinase. *Nature* 195, 1300–1301. doi: 10.1038/1951300a0

Spyrakis, F., Santucci, M., Maso, L., Cross, S., Gianquinto, E., Sannio, F., et al. (2020). Virtual screening identifies broad-spectrum β -lactamase inhibitors with activity on clinically relevant serine-and metallo-carbapenemases. *Sci. Rep.* 10:12763. doi: 10.1038/ s41598-020-69431-y

Srinivasan, V. B., and Rajamohan, G. (2013). KpnEF, a new member of the *Klebsiella pneumoniae* cell envelope stress response regulon, is an SMR-type efflux pump involved in broad-spectrum antimicrobial resistance. *Antimicrob. Agents Chemother.* 57, 4449–4462. doi: 10.1128/AAC.02284-12

Stapleton, P. D., Shah, S., Anderson, J. C., Hara, Y., Hamilton-Miller, J. M. T., and Taylor, P. W. (2004). Modulation of β -lactam resistance in *Staphylococcus aureus* by catechins and gallates. *Int. J. Antimicrob. Agents* 23, 462–467. doi: 10.1016/j. ijantimicag.2003.09.027

Stebbins, N. D., Ouimet, M. A., and Uhrich, K. E. (2014). Antibiotic-containing polymers for localized, sustained drug delivery. *Adv. Drug Deliv. Rev.* 78, 77–87. doi: 10.1016/j.addr.2014.04.006

Stevenson, L. J., Owen, J. G., and Ackerley, D. F. (2019). Metagenome driven discovery of nonribosomal peptides. ACS Chem. Biol. 14, 2115–2126. doi: 10.1021/acschembio.9b00618

Suh, B., Bae, I. K., Kim, J., Jeong, S. H., Yong, D., and Lee, K. (2010). Outbreak of Meropenem-resistant Serratia marcescens Comediated by chromosomal AmpC β -lactamase overproduction and outer membrane protein loss. *Antimicrob. Agents Chemother.* 54, 5057–5061. doi: 10.1128/aac.00768-10

Tagliaferri, T. L., Guimarães, N. R., de Paula Martins Pereira, M., Vilela, L. F. F., Horz, H.-P., dos Santos, S. G., et al. (2020). Exploring the potential of CRISPR-Cas9 under challenging conditions: facing high-copy plasmids and counteracting Beta-lactam resistance in clinical strains of Enterobacteriaceae. *Front. Microbiol.* 11:578. doi: 10.3389/ fmicb.2020.00578

Tamang, M. D., Bae, J., Park, M., and Jeon, B. (2022). Potentiation of β -lactams against methicillin-resistant *Staphylococcus aureus* (MRSA) using Octyl Gallate, a food-grade antioxidant. *Antibiotics (Basel)*. 11:266. doi: 10.3390/antibiotics11020266

Tegos, G. P., Masago, K., Aziz, F., Higginbotham, A., Stermitz, F. R., and Hamblin, M. R. (2008). Inhibitors of bacterial multidrug efflux pumps potentiate antimicrobial photoinactivation. *Antimicrob. Agents Chemother.* 52, 3202–3209. doi: 10.1128/AAC.00006-08

Tehrani, K. H. M. E., Brüchle, N. C., Wade, N., Mashayekhi, V., Pesce, D., van Haren, M. J., et al. (2020). Small molecule carboxylates inhibit Metallo- β -lactamases and Resensitize Carbapenem-resistant bacteria to Meropenem. *ACS Infect. Dis.* 6, 1366–1371. doi: 10.1021/acsinfecdis.9b00459

Thakuria, B. (2013). The Beta lactam antibiotics as an empirical therapy in a developing country: an update on their current status and recommendations to counter the resistance against them. *J. Clin. Diagn. Res.* 7, 1207–1214. doi: 10.7860/jcdr/2013/5239.3052

Tomoshige, S., Dik, D. A., Akabane-Nakata, M., Madukoma, C. S., Fisher, J. F., Shrout, J. D., et al. (2018). Total syntheses of Bulgecins a, B, and C and their bactericidal potentiation of the β -lactam antibiotics. ACS Infect Dis. 4, 860–867. doi: 10.1021/acsinfectis.8b00105

Turton, J. F., Doumith, M., Hopkins, K. L., Perry, C., Meunier, D., and Woodford, N. (2016). Clonal expansion of *Escherichia coli* ST38 carrying a chromosomally integrated OXA-48 carbapenemase gene. *J. Med. Microbiol.* 65, 538–546. doi: 10.1099/ jmm.0.000248

Tuyiringire, N., Tusubira, D., Munyampundu, J. P., Tolo, C. U., Muvunyi, C. M., and Ogwang, P. E. (2018). Application of metabolomics to drug discovery and understanding the mechanisms of action of medicinal plants with anti-tuberculosis activity. *Clin. Transl. Med.* 7:29. doi: 10.1186/s40169-018-0208-3

Uddin, M. J., and Ahn, J. (2018). Characterization of β -lactamase- and efflux pumpmediated multiple antibiotic resistance in salmonella typhimurium. *Food Sci. Biotechnol.* 27, 921–928. doi: 10.1007/s10068-018-0317-1

Udwary, D., Zeigler, L., Asolkar, R. N., Singan, V., Lapidus, A., Fenical, W., et al. (2007). Genome sequencing reveals complex secondary metabolome in the marine actinomycete *Salinispora tropica*. *Proc. Natl. Acad. Sci.* 104, 10376–10381. doi: 10.1073/pnas.0700962104

Unemo, M., Golparian, D., Nicholas, R., Ohnishi, M., Gallay, A., and Sednaoui, P. (2012). High-level cefixime- and ceftriaxone-resistant Neisseria gonorrhoeae in France: novel penA mosaic allele in a successful international clone causes treatment failure. *Antimicrob. Agents Chemother.* 56, 1273–1280. doi: 10.1128/AAC.05760-11

Urban, C., Go, E., Mariano, N., and Rahal, J. J. (1995). Interaction of sulbactam, clavulanic acid and tazobactam with penicillin-binding proteins of imipenem-resistant and-susceptible *Acinetobacter baumannii. FEMS Microbiol. Lett.* 125, 193–197. doi: 10.1111/j.1574-6968.1995.tb07357.x

Vaara, M. (2019). Polymyxin derivatives that sensitize gram-negative bacteria to other antibiotics. *Molecules* 24:249. doi: 10.3390/molecules24020249

Veeraraghavan, B., Ragupathi, N. D., Bakthavatchalam, Y., Mathur, P., Pragasam, A., Walia, K., et al. (2019). Plasmid profiles among some ESKAPE pathogens in a tertiary care Centre in South India. *Indian J. Med. Res.* 149, 222–231. doi: 10.4103/ijmr.jjmr_2098_17

Velásquez, J. E., and van der Donk, W. A. (2011). Genome mining for ribosomally synthesized natural products. *Curr. Opin. Chem. Biol.* 15, 11–21. doi: 10.1016/j. cbpa.2010.10.027

Vincent, J.-L., Sakr, Y., Singer, M., Martin-Loeches, I., Machado, F. R., Marshall, J. C., et al. (2020). Prevalence and outcomes of infection among patients in intensive care units in 2017. *JAMA* 323, 1478–1487. doi: 10.1001/jama.2020.2717

Wang, T., Zou, C., Wen, N., Liu, X., Meng, Z., Feng, S., et al. (2021). The effect of structural modification of antimicrobial peptides on their antimicrobial activity, hemolytic activity, and plasma stability. *J. Pept. Sci.* 27:e3306. doi: 10.1002/psc.3306

Watanabe, M., Iyobe, S., Inoue, M., and Mitsuhashi, S. (1991). Transferable imipenem resistance in *Pseudomonas aeruginosa. Antimicrob. Agents Chemother.* 35, 147–151. doi: 10.1128/AAC.35.1.147

Wilke, M. S., Lovering, A. L., and Strynadka, N. C. J. (2005). β -Lactam antibiotic resistance: a current structural perspective. *Curr. Opin. Microbiol.* 8, 525–533. doi: 10.1016/j.mib.2005.08.016

Wise, R., Logan, M., Cooper, M., and Andrews, J. M. (1991). Pharmacokinetics and tissue penetration of Tazobactam administered alone and with piperacillin. *Antimicrob. Agents Chemother*, 35, 1081–1084. doi: 10.1128/aac.35.6.1081

World Bank (2017). Drug-resistant infections: A threat to our economic future. World Health Organization.

World Health Organization (2014). Antimicrobial resistance: Global report on surveillance. World Health Organization.

Wu, W., Feng, Y., Tang, G., Qiao, F., McNally, A., and Zong, Z. (2019). NDM Metallo- β -lactamases and their bacterial producers in health care settings. *Clin. Microbiol. Rev.* 32, e00115–e00118. doi: 10.1128/CMR.00115-18

Wu, J. Y., Srinivas, P., and Pogue, J. M. (2020). Cefiderocol: a novel agent for the management of multidrug-resistant gram-negative organisms. *Infect. Dis. Ther.* 9, 17–40. doi: 10.1007/s40121-020-00286-6

Yahav, D., Giske, C. G., Grāmatniece, A., Abodakpi, H., Tam, V. H., and Leibovici, L. (2020). New β -lactam- β -lactamase inhibitor combinations. *Clin. Microbiol. Rev.* 34:e00115-20. doi: 10.1128/CMR.00115-20

Yang, Y., Guo, Y., Zhou, Y., Gao, Y., Wang, X., Wang, J., et al. (2020). Discovery of a novel natural allosteric inhibitor that targets NDM-1 against *Escherichia coli. Front. Pharmacol.* 11:581001. doi: 10.3389/fphar.2020.581001

Yang, K.-W., Zhou, Y., Ge, Y., and Zhang, Y. (2017). Real-time activity monitoring of New Delhi metallo- β -lactamase-1 in living bacterial cells by UV-vis spectroscopy. *Chem Comm.* 53, 8014–8017. doi: 10.1039/c7cc02774e

Zampieri, M., Szappanos, B., Buchieri, M. V., Trauner, A., Piazza, I., Picotti, P., et al. (2018). High-throughput metabolomic analysis predicts mode of action of uncharacterized antimicrobial compounds. *Sci. Transl. Med.* 10:eaal3973. doi: 10.1126/scitranslmed.aal3973

Zhao, B., Zhang, X., Yu, T., Liu, Y., Zhang, X., Yao, Y., et al. (2021). Discovery of thiosemicarbazone derivatives as effective New Delhi metallo-β-lactamase-1 (NDM-1) inhibitors against NDM-1 producing clinical isolates. *Acta Pharm. Sin. B* 11, 203–221. doi: 10.1016/j.apsb.2020.07.005

Zharkova, M. S., Orlov, D. S., Golubeva, O. Y., Chakchir, O. B., Eliseev, I. E., Grinchuk, T. M., et al. (2019). Application of antimicrobial peptides of the innate immune system in combination with conventional antibiotics-a novel way to combat antibiotic resistance? *Front. Cell. Infect. Microbiol.* 9:128. doi: 10.3389/fcimb.2019.00128

Zhen, X., Lundborg, C. S., Sun, X., Hu, X., and Dong, H. (2019). Economic burden of antibiotic resistance in ESKAPE organisms: a systematic review. *Antimicrob. Resist. Infect. Control* 8:137. doi: 10.1186/s13756-019-0590-7

Glossary

ACA	Aminocephalosporinic acid			
AI	Artificial intelligence			
AMP	Antimicrobial peptides			
AMR	Antimicrobial resistance			
АРА	Aminopenicillanic acid			
APDI	Antimicrobial photodynamic inactivation			
ATCC	American type culture collection			
BGC	Biosynthetic Gene Clusters			
BLDB	Beta-Lactamase DataBase			
BLI	β-Lactamase inhibitors			
CF	cystic fibrosis			
CNAI	Cross-linked chitosan nanoparticles			
СРЕ	Carbapenem-producing Enterobacteriaceae			
CRISPR/Cas	Clustered regularly interspaced short palindromic repeats and CRISPR-associated protein			
DBO	Diazabicyclooctanones			
DHP-I	Dehydropeptidase-I			
DTR	Difficult to treat			
EPI	Efflux pump inhibitor			
ESBL	Extended spectrum β-lactamase			
FLAP	Fingerprint for Ligands and Proteins			
ICU	Intensive care units			
IS	Insertion sequences			
LGFE	Ligand-grid Free Energy			
MBL	Metallo-β-lactamases			
MDR	Multidrug resistance			
MDS	Molecular dynamic simulations			
MGE	Mole genetic elements			
MIC	Minimum inhibitory concentration			
MIF	Molecular Interaction Fields			
ML	Machine learning			
MRSA	Methicillin resistant <i>Staphylococcus aureus</i>			
NMP	1-(1-napthylmethyl)-piperazine			
NN	Neural networks			
Nps	Nanoparticles			
OM	Outer membrane			
ORF	Open reading frame			
ΡΑβΝ	Phenylalanyl arginyl β-naphthylamide			
PBP	Penicillin binding proteins			
RF	Random-forest scoring			
RND	Resistance-Nodulation-Division			
SBL	serine β-lactamases			
SILCS	Site-Identification by Ligand Competitive Saturation			
UTI				
VAP	Urinary tract infections Ventilator associated pneumonia			
WHO	World Health Organization			
XP	Extra Precision			
AI				