#### Check for updates

#### **OPEN ACCESS**

EDITED BY Weiqi He, Soochow University, China

REVIEWED BY Yukiko Miyamoto, University of California, San Diego, United States Graham J. Britton, Icahn School of Medicine at Mount Sinai, United States

\*CORRESPONDENCE Ning Liu leirum\_ln@qdu.edu.cn Dongming Xing xdm\_tsinghua@163.com

SPECIALTY SECTION This article was submitted to Microorganisms in Vertebrate Digestive Systems, a section of the journal Frontiers in Microbiology

RECEIVED 10 October 2022 ACCEPTED 30 November 2022 PUBLISHED 16 December 2022

#### CITATION

Liang B, Wu C, Wang C, Sun W, Chen W, Hu X, Liu N and Xing D (2022) New insights into bacterial mechanisms and potential intestinal epithelial cell therapeutic targets of inflammatory bowel disease. *Front. Microbiol.* 13:1065608. doi: 10.3389/fmicb.2022.1065608

#### COPYRIGHT

© 2022 Liang, Wu, Wang, Sun, Chen, Hu, Liu and Xing. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# New insights into bacterial mechanisms and potential intestinal epithelial cell therapeutic targets of inflammatory bowel disease

Bing Liang<sup>1</sup>, Changhao Wu<sup>2</sup>, Chao Wang<sup>1</sup>, Wenshe Sun<sup>1</sup>, Wujun Chen<sup>1</sup>, Xiaokun Hu<sup>3</sup>, Ning Liu<sup>1\*</sup>and Dongming Xing<sup>1,4\*</sup>

<sup>1</sup>Cancer Institute, The Affiliated Hospital of Qingdao University, Qingdao, China, <sup>2</sup>Department of Biochemistry and Physiology, Faculty of Health and Medical Sciences, University of Surrey, Guildford, United Kingdom, <sup>3</sup>Intervention Neurosurgery, The Affiliated Hospital of Qingdao University, Qingdao, China, <sup>4</sup>School of Life Sciences, Tsinghua University, Beijing, China

The global incidence of inflammatory bowel disease (IBD) has increased rapidly in recent years, but its exact etiology remains unclear. In the past decade, IBD has been reported to be associated with dysbiosis of gut microbiota. Although not yet proven to be a cause or consequence of IBD, the common hypothesis is that at least some alterations in the microbiome are protective or pathogenic. Furthermore, intestinal epithelial cells (IECs) serve as a protective physical barrier for gut microbiota, essential for maintaining intestinal homeostasis and actively contributes to the mucosal immune system. Thus, dysregulation within the intestinal epithelium increases intestinal permeability, promotes the entry of bacteria, toxins, and macromolecules, and disrupts intestinal immune homeostasis, all of which are associated with the clinical course of IBD. This article presents a selective overview of recent studies on bacterial mechanisms that may be protective or promotive of IBD in biological models. Moreover, we summarize and discuss the recent discovery of key modulators and signaling pathways in the IECs that could serve as potential IBD therapeutic targets. Understanding the role of the IECs in the pathogenesis of IBD may help improve the understanding of the inflammatory process and the identification of potential therapeutic targets to help ameliorate this increasingly common disease.

#### KEYWORDS

inflammatory bowel disease, gut microbiota, bacterial mechanisms, intestinal epithelial cells (IECs), therapeutic targets

# Introduction

Inflammatory bowel disease (IBD) is a multifactorial chronic inflammatory disease, which primarily includes Crohn's disease (CD) and Ulcerative colitis (UC). IBD causes severe gastrointestinal (GI) tract symptoms, including diarrhea, abdominal pain, bleeding, anemia, and weight loss, and poses a severe but incurable threat to human life and health. The etiology and pathogenesis of IBD remain unknown. However, they may be linked to genetic susceptibility, intestinal microbial homeostasis imbalance, impaired intestinal mucosal barrier function, disorders of innate and adaptive immune regulation in the intestine, and stimulation by external environmental factors. IBD primarily affects genetically predisposed individuals in a specific environment where the intestinal mucosal immune system produces an abnormally amplified immune response to intestinal microbial antigens, resulting in inflammatory damage to the intestinal mucosa.

There are ~400 species of bacteria in the human intestine, including resident and some transient bacteria. The sites of IBD are the colon, rectum, and ileum, where the intestine is most exposed to bacteria. Some clinical features of IBD are similar to those of some infectious diseases of the GI. Several studies have shown that a large quantity of bacteria in the human intestine affects the function of the intestinal mucosal immune system and that a large number of intestinal antigens, such as enteric bacteria and bacterial products, induce abnormal immune responses in the intestine of individuals with genetic susceptibility to IBD, leading to the development of IBD. Although the current research data do not support that infection with a specific pathogenic microorganism causes the development of IBD, clinical observations suggest that intestinal infections caused by these microorganisms can induce the recurrence of IBD in remission. In addition, many experiments have confirmed the involvement of intestinal bacteria in the development of experimental colitis. About 99.9% of human GI microbiota are bacteria, and the other 0.1% are fungi, archaea, and viruses (Qin et al., 2010; Sommer and Backhed, 2013). Bacteria have been extensively reported to have an important role in the pathogenesis of IBD. Intestinal epithelial cells (IECs) line the surface of the intestinal epithelium and perform a variety of functions, including the physical isolation of commensal bacteria and the integration of microbial signals (Peterson and Artis, 2014). IECs also react to immune cellproduced factors that regulate epithelial barrier function, proliferation, and differentiation (Soderholm and Pedicord, 2019). Dysregulation of the IECs can lead to increased intestinal permeability, abnormal IEC interactions with immune cells, and disruption of intestinal immune homeostasis, all of which are associated with the clinical course of IBD (Coskun, 2014).

Many medications, including aminosalicylates, corticosteroids, immunomodulatory drugs, biologics, and antibiotics, can help reduce inflammation and alleviate IBD symptoms. Among them, the introduction of tumor necrosis factor (TNF) inhibitors is an outstanding achievement, allowing for long-term remission and modification of the IBD course in many patients (Nielsen and Ainsworth, 2013). The clinical treatment of IBD has advanced dramatically due to the development and application of these medications and the updating of therapeutic targets. However, these conventional treatments still focus on inducing and maintaining clinical remission, which is not yet wholly curable. A substantial proportion of patients still have long-term chronic active inflammatory reactions and the need for surgical procedures, which significantly impacts patients' quality of life and imposes a major social and medical burden. Therefore, it is critical to investigate the pathogenesis of IBD, identify new targets for IBD treatment and implement precision medicine to develop the best treatment for patients.

## Bacteria species and functional pathways involved in the protection and pathogenicity of inflammatory bowel disease

The gut microbiota is considered to be a nonnegligible factor in the pathogenesis of IBD and an important target for the research of therapeutic drugs for IBD, which can regulate the host's vital activities, modulate the immune response and counteract the dysbiosis of gut microbiota, and play a vital role in protecting the host's health. Many studies have found that IBD patients have a disturbed intestinal microecological balance, characterized by reduced diversity, increased bacterial instability, increased Actinobacteria, and Proteobacteria, and decreased Bacteroidetes and Firmicutes, especially a significant decrease in bacteria producing short-chain fatty acids (SCFAs; Caruso et al., 2020). The pathophysiological mechanisms by which gut microbiota antigens induce abnormal responses in the innate and adaptive immune systems of the intestinal mucosa remain unknown. In addition to SCFAs, tryptophan metabolism, bile acid metabolism, Polysaccharide A (PSA) production, and virulence factor genes are balanced in the normal intestine, whereas tryptophan derivatives are reduced, secondary bile acids are reduced, PSA gene expression is reduced, and microbial virulence gene expression is increased in IBD patients' intestines.

## Bacteria species and functional pathways that are potentially protective of inflammatory bowel disease

The first bacterial metabolite to play a potentially protective role for IBD is SCFA. The major SCFAs, including acetate, propionate, and butyrate, account for over 95% of the total SCFA content in feces (Kim et al., 2019). SCFAs can be produced by bacteria such as *Roseburia* and *Faecalibacterium prausnitzii*. Intestinal microorganisms convert dietary fiber fatty acids into SCFAs, which then enter the intestinal lumen. SCFAs can bind to G protein-coupled receptors (GPCRs), like GPR41 (Brown et al., 2003), GPR43 (Maslowski et al., 2009), and GPR109A (Thangaraju et al., 2009), to activate signaling cascades that control immune functions and improving intestinal barrier integrity (Vieira et al., 2015). SCFAs can also modulate immune cells, reduce pro-inflammatory factors, and mitigate the development of IBD (Luis et al., 2021); (Renga et al., 2022); (Rösch et al., 2017); (Nicolas and Chang, 2019). Back in 2013, butyrate, a by-product

from Clostridia fermentation of indigestive dietary fiber, was discovered to increase histone H3 acetylation at the Foxhead box P3 (Foxp3) promoter and conserved noncoding regions, promoting Treg differentiation and alleviate the development of colitis in mice (Furusawa et al., 2013). Similarly, Li et al. reported that butyrate inhibited neutrophil migration, formation of neutrophil extracellular traps (NETs), and production of pro-inflammatory cytokines, chemokines, and calmodulin in IBD patients by inhibiting histone deacetylase (HDAC). RNA sequencing analysis revealed that butyrate's immunomodulatory effects on neutrophils included leukocyte activation, innate immune response regulation, and oxidative stress (Li et al., 2021). Recent studies demonstrated previously unknown intracellular signaling network mediated by butyrate. For instance, butyrate could exert a protective effect against colitis by downregulating Hexokinase 2 (HK2) in the intestinal epithelium via HDAC8 (Hinrichsen et al., 2021). Moreover, a subsequent study revealed that butyrate increased P-glycoprotein (P-gp) transcription by inhibiting HDACs and activating nuclear receptors, pregnane X receptor (PXR) and vitamin D3 receptor (VDR). P-gp is a component of Intestinal epithelial cells (IECs) that plays an essential role in the excretion of endogenous cannabinoid toxins, the prevention of excessive inflammation, and the maintenance of endocytosis (Foley et al., 2022; Figure 1). The second pathway that protects against IBD is the metabolism of bile acids. Bile acids are synthesized in the liver, then processed and modified before being secreted into the duodenum, where intestinal microorganisms convert them to secondary bile acids (Ahmad and Haeusler, 2019). There are two secondary bile acids: lithocholic acid (LCA) and chenodeoxycholic acid (CDCA). They modulate immune cells, reduce pro-inflammatory factors, decrease systemic inflammation and alleviate the development of IBD (Postler and Ghosh, 2017); (Sinha et al., 2020). Bile acids can affect host metabolism, cancer progression, and innate immunity, but their effects on adaptive immune cells such as Helper T-cell 17 (Th17) cells and Regulatory T (Treg) cells have not been clarified. Previous research has shown that 3-oxoLCA and isoalloLCA, 2 lithocholic acid (LCA) derivatives, can inhibit the differentiation of Th17 cells and promote the differentiation of Treg, respectively. 3-oxoLCA inhibits the differentiation of Th17 cells by directly binding to the key transcription factor RAR-related orphan receptor gamma t (RORyt) and isoalloLCA promotes Treg differentiation by enhancing *Foxp3* expression and inducing the production of mitochondrial reactive oxygen species (mROS; Hang et al., 2019); however, the relationship between 3-oxoLCA and IBD remains unknown. A recent study discovered Enterobacteria and bacterial enzyme 3-hydroxysteroid dehydrogenase (3-HSDH) that convert LCA to 3-oxoLCA and then to isoLCA through in vitro culture screening. IsoLCA, like 3-oxoLCA, inhibits the transcriptional activity of RORyt and thus reduces Th17 cell differentiation (Paik et al., 2022). This study adds to our understanding of Enterobacteria-metabolite-host immune interactions and highlight the critical role of microbederived bile acid metabolites in regulating host immunity,

shedding new light on the treatment of immune diseases such as IBD (Figure 1). Treg cells have an important role in maintaining intestinal homeostasis, and their differentiation is influenced by the products of gut microbiota. A recently published study identified two new secondary bile acids, ω-muricholic acid (ω-MCA) and 3β-hydroxydeoxycholic acid (isoDCA) that effectively induce Treg differentiation in vitro, with isoDCA being more abundant in the intestine (Campbell et al., 2020). Further studies showed that isoDCA enhances its Treg induction by interacting with the farnesol X receptor (FXR) in dendritic cells (DCs), and by constructing engineered bacteria and colony colonization experiments, it was confirmed in mice that isoDCAproducing colonies promote peripheral Treg. The third pathway is tryptophan metabolism. Tryptophan is a necessary amino acid. Dietary tryptophan can undergo the kynurenine ammonia pathway in response to microbial and other inflammatory stimuli, producing a variety of compounds (Ma et al., 2020). Microbial action breaks down tryptophan to an indole derivative, which can modulate immune cells by stimulating IL-22, secreting antimicrobial peptides, and inhibiting the expression of inflammatory factors (Zelante et al., 2013). Clostridium perfringens can participate in the metabolism of tryptophan, which is also protective of IBD. According to a recent study, a tryptophan metabolite, indole-3-carboxaldehyde (3-IAld) reduces immune checkpoint inhibitor (ICI)-induced colitis in mice by influencing the structure and function of the mouse gut microbiota, resulting in an increase in sugar-fermenting bacteria and SCFA-producing bacteria and regulating Aryl hydrocarbon receptor (Ahr)/IL-22, and IL-10+ Treg cells (Renga et al., 2022; Figure 1). PSA production, the fourth potentially protective pathway for IBD, is primarily produced by Bacteroides fragilis. B. fragilis can deliver the immunomodulatory molecule PSA to immune cells by releasing outer membrane vesicles (OMVs), activate TLR 2 on the surface of DCs, resulting in the release of cytokines that promote IL-10 production by T cells (Wang et al., 2006; Shen et al., 2012; Dasgupta et al., 2014). This process is enhanced by direct binding of PSA to TLR2 expressed on Foxp3+ Tregs, which increases IL-10 production even further (Round and Mazmanian, 2010; Figure 1). PSA-induced IL-10 production inhibits the activity of mucosal effector T cells, particularly TH17 cells, and thus protects mice in colitis models (Mazmanian et al., 2008; Dasgupta et al., 2014). Here, B. fragilis refers mainly to non-toxigenic B. fragilis, or NTBF. On the other hand, Enterotoxigenic B. fragilis (ETBF) is toxin-producing, which is likely to promote the development of IBD (Chung et al., 2018).

In addition to the four metabolite pathways associated with gut microbiota mentioned previously, other microbial species and functional molecules/pathways have been studied extensively recently (Figure 2). *Akkermansia muciniphila* (Akk) binds to a receptor on the membrane of IECs, affecting downstream immune regulatory responses, lowering lipopolysaccharide (LPS), reducing inflammation, regulating lipids, and lowering blood glucose in mouse models (Plovier et al., 2017). The literature has reported that Akk and its outer membrane protein, Amuc\_1100, can



alleviate enteritis in mice (Zhai et al., 2019). Toll-like receptor 4 (TLR4) is a critical mediator in the interactions between gut microbiota and host immunity. Yang et al. recently discovered that TLR4 protects mice from colitis by promoting Akk colonization in the intestine, which upregulates RORt+ Treg cell-mediated immune responses (Liu et al., 2022). Furthermore, in a previous study, oral administration of Akk bacteria strain BAA-835 significantly improved the phenotype of Dextran sulfate sodium (DSS)-induced acute colitis in mice. Mechanistically, Akk increased the expression of NLR family pyrin domain containing 3 (NLRP3), caspase-1 p20, and IL-1 p17 improved intestinal barrier function, and inhibited the expression of pro-inflammatory factors such as *TNF-* $\alpha$  and *IL-*6 (Qu et al., 2021). However, an earlier study found that Akk can cause colitis in sterile IL-10 genedeficient mice (Seregin et al., 2017). Therefore, whether Akk can be used to treat IBD is still debatable. Furthermore, according to a recent study by Wang et al., Enterobacter ludwigii isolated from metronidazole-treated mouse feces could enhance DCs and promote Tregs differentiation through its metabolite choline and its receptor a7 nicotinic acetylcholine receptor (a7nAChR)mediated upregulation of retinoic acid (RA) and TGF-β, thereby increasing the CD103+ DC/Treg-dependent tolerance response

and ultimately reducing the susceptibility of mice to DSS-induced colitis. Thus, this study provides a potential therapeutic approach for IBD (Li et al., 2022).

## Bacteria species and functional pathways that are potentially promotive of inflammatory bowel disease

Previous research has found that some pathobionts play a pro-IBD role in genetically susceptible individuals, such as *Mycobacterium paratuberculosis* and *Helicobacter pylori*. In addition, the efficacy of antibiotics and probiotics targeting gut microbiota further suggests a role for microorganisms in IBD. New research has recently emerged identifying many new bacteria species and the virulence factors they contain that potentially promote IBD (Figure 3). Adherent-invasive *E. coli* (AIEC) is distinct from typical *E. coli*, which AIEC is enriched in the intestinal microbiota of CD patients and promotes intestinal inflammation, development of IBD and even colorectal cancer (CRC). However, it is unclear how the nutrient metabolism of AIEC affects intestinal homeostasis. AIEC can adhere to the IECs



layer via the FimH and carcinoembryonic antigen-related cell adhesion molecule 6 (CEACAM6), enter the lamina propria and Peyer's patch via Microfold (M) cells, and interact with immune cells in the intestinal lumen to increase the production of inflammatory factors (Barnich et al., 2007; Palmela et al., 2018; Viladomiu et al., 2021). A recent article reported that in CD, propionate metabolism by AIEC is critical in promoting intestinal T cell inflammation. Meanwhile, CX3CR1+ mononuclear phagocytes produce IL-1ß upon sensing the AIEC metabolite propionate, which induces Th17 cells and drives intestinal inflammation (Viladomiu et al., 2021). Moreover, ETBF is strongly associated with the development of IBD, colitis associated cancer (CAC), and CRC. However, the mechanisms by which ETBF induces intestinal inflammation and tumorigenesis are unclear. B. fragilis contains the virulence gene B. fragilis toxin (BFT), which encodes the virulence protein BFT, interacting with the membrane receptors of IECs and immune cells via the STAT3 and NF-KB pathways, causing the expression of IL-17 downstream, promoting the expression of inflammatory factors and thus influencing the development of CRC (Chung et al., 2018). Recent study has discovered that ETBF-stimulated cells deliver miR-149-3p via exosomes and promote Th17 cell differentiation. ETBF induced CRC by suppressing miR-149-3p and promoting PHD finger protein 5A (PHF5A)-mediated histone acetyltransferase KAT2A mRNA alternative splicing (Cao et al., 2021). Campylobacter concisus is involved in a wide range of inflammatory diseases, including IBD. The zonula occludens

toxin (Zot) gene, discovered in Asiatic cholera and had a pathogenic effect, is found in 30% of C. concisus (Zhang et al., 2014). In a large study of Danish subjects, 962 people were infected with C. concisus and 1725 with C. jejuni. The risk of colitis was significantly higher in C. concisus-infected subjects than in controls, with a risk ratio of 32.4 (Nielsen et al., 2020). This also implies that C. Concisus likely hasten the onset of IBD. Further research has revealed that the enteropathogenic Zot virulence protein of C. concisus disrupts the intestinal epithelial barrier and induces the production of pro-inflammatory cytokines, particularly TNF- $\alpha$ , by intestinal epithelial and macrophage cells (Mahendran et al., 2016). Fusobacterium varium was found in up to 54.6% of 163 inflamed mucosal samples from 152 UC patients in Japan using real-time quantitative PCR (Tahara et al., 2015). A genome-wide analysis of an F. varium Fv113-g1 isolate from a UC patient's intestine revealed numerous virulence factors, including 44 Type 5 secretion systems (T5SS) and 13 FadA adhesion gene homologs in mucosal inflammation (Sekizuka et al., 2017). Some gut microbiota (for example, pks+E. coli) produce colibactin, a chemically unstable small molecule genotoxin whose genotoxicity to host cells can increase the host's risk of CRC. The effects of colibactin on other microorganisms in the gut have received little attention. A recent study discovered that colibactin could target surrounding bacteria containing prophages and activate prophages within the bacteria, causing phage replication and bacterial lysis. The induction of prophages by colibactin-producing bacteria has been observed in



various human and gut-associated phage-bacterial systems and in mouse fecal flora. These findings shed light on the potential mechanisms by which colibactin influences gut microbiota and the relationship between bacterial products and phagosomal behavior. As a result, it is possible that bacteria evolved this genotoxin to inhibit other bacteria rather than poison the host and that the cancer-promoting effects of colibactin are a "misuse" of the host (Silpe et al., 2022). Enterococci are a group of bacteria widely found in the intestines of humans and other animals that cause multi-drug resistant infections. However, a family of potent protein-like toxins specifically targeting human and animal cells has not been previously identified in Enterococcus spp. Min's group have recently identified and resolved a novel family of Poreforming toxins (Epxs) expressed in Enterococcus spp. They discovered that Epx is a subclass of small  $\beta$ -barrel pore-forming toxins, with the  $\beta$ -barrel serving as the top structural domain and forming homo-octameric pores. The whole-genome CRISPR-Cas9 screen revealed that Epx2/3 binds to human leukocyte antigen class I (HLA-I). Through Epx2 virulence, Epx2+ E. faecalis strains harm peripheral blood mononuclear cells (PBMCs) and intestinal-like organs (Xiong et al., 2022). Ruminococcus gnavus is a type of commensal bacteria found in the human intestine. Several studies have found that they are

significantly more abundant in IBD patients than in healthy subjects. R. gnavus. Can reduce intestinal mucin, which is mediated by intramolecular trans-sialidase (RgNanH; Tailford et al., 2015). Moreover, R. gnavus. Can stimulate DCs via the production of inflammatory polysaccharides, increasing the inflammatory cytokine TNF- $\alpha$ , promoting IBD's development (Henke et al., 2019). Other bacteria that potentially contribute to IBD are Helicobacter hepaticus and H. bilis, both of which are from the genus Helicobacter. Bacteria from the genus Helicobacter that are not H. pylori often positively associated with IBD. In contrast, H. pylori and IBD show a negative correlation, meaning that people infected with H. pylori are less likely to develop IBD, possibly due to the ability of H. pylori to inhibit the growth of other bacteria. According to one study, in a mouse model of IL-10 gene-deficient colitis, H. hepaticus induced inflammatory Th17, increased IL-17A and IFN-y expression, and aided in the development of colitis (Xu et al., 2018). Some gut microbiota can colonize the intestinal mucus layer and feed on mucin, a major component of the mucus layer. Mucin is a heavily glycosylated protein with numerous O-glycan chains. Mucin O-glycan ends are frequently sulfated in the distal colon. Bacterial degradation of colonic mucin requires specific sulfatases to remove sulfate groups on O-glycans. However, little is known about the bacterial

sulfate esterases that act on colonic mucin and how they work. A recently published study discovered a sulfate esterase expressed by Bacillus thetaiotaomicron, a common human intestinal commensal that uses sulfated colonic mucin O-glycans as its sole carbon source. The knockout of sulfate esterases from B. thetaiotaomicron revealed that one of the enzymes (BT1636^3S-Gal) is required to utilize sulfated mucin O-glycan and plays an important role in the intestinal colonization of the bacterium (Luis et al., 2021). These findings shed light on how gut microbiota degrade colonic mucins and the mechanisms associated with bacterial colonization and IBD. Blocking this enzymatic pathway could be a potential intervention strategy to prevent pro-IBD bacteria from disrupting the intestinal mucus layer barrier. Fusobacterium nucleatum infection may contribute to the worsening of UC, but its mechanism is not yet clear. Dong's team recently published findings that EVs of F. nucleatum could promote the expression of pro-inflammatory factors (IL-1β, IL-6, TNF- $\alpha$ ) and down-regulate the expression of anti-inflammatory factors (IL-10) and tight junction proteins (ZO-1 and occludin) in IECs by down-regulating the expression of miR-574-5p to promote the activation of autophagy, leading to barrier dysfunction and worsen DSS-induced colitis in mice (Wei et al., 2022). Furthermore, another recent study discovered that Eggerthella lenta could promote colitis in mice by activating Th17 cells and promoting IL-17a production through its expression of a drug-metabolizing enzyme, Cardiac glycoside reductase (cgr2). E. lenta was found to be enriched in patients with IBD, while the expression of the cgr2 gene was increased in patients with rheumatoid arthritis. Cgr2 metabolizes steroidal saponins, which inhibit Th17 cell activity and are negatively associated with IBD severity. Furthermore, dietary arginine supplementation inhibited cgr2-induced Th17 cell activation and colitis in mice (Alexander et al., 2022).

## Targeting potential modulators and signaling pathways in intestinal epithelial cells to treat inflammatory bowel disease

Recent advances in various treatments have directly changed the clinical management model for IBD patients. The introduction of immunosuppressants and biologics significantly reduces the use of corticosteroids. In addition, the alpha  $4\beta7$  integrin blocker vedolizumab has also been included in the clinical IBD treatment. While these strategies expand the arsenal of IBD physicians, a significant percentage of patients do not respond to these treatments (Singh et al., 2018). Several new cytokine inhibitors, modulators of cytokine signaling pathways, inhibitors of transcription factors, and new anti-adhesion and anti-T cell activation and migration strategies are undergoing clinical trial evaluation. More therapeutic targets and signaling pathways need to be found to provide patients with more choices and personalized services in treatment decisions (Table 1). An essential function of the intestinal epithelium is to act as a barrier limiting the interaction between the luminal contents, such as intestinal microorganisms, and the immune system, absorbing nutrients while limiting microbial translocation (Allaire et al., 2018). Impaired intestinal barrier function leads to increased membrane permeability of IECs, which promotes the entry of intraperitoneal bacteria, toxins, and macromolecules into the body, thus inducing an inflammatory immune response. Intestinal epithelial barrier injury is of great interest in the study of the cellular and molecular pathogenesis of IBD, and reconstruction and regeneration of the intestinal epithelium is essential to restore intestinal homeostasis after injury (Sommer et al., 2021). Patients with IBD exhibit defects in multiple components of the mucosal barrier. Future research should focus on the modulation of the intestinal epithelial barrier as a possible therapeutic target for IBD.

By analyzing the function of the creatine transporter protein SLC6A8 (also known as CRT) in IECs, a recent study found that CRT is localized around tight junction proteins and can maintain intestinal epithelial barrier function by regulating the energy balance of IECs to promote the expression and localization of tight junction proteins. The deficiency of CRT inhibited creatine uptake, barrier formation, and healing function in IECs (Hall et al., 2020). Mechanistically, CRT deletion increased Claudin-2 expression and decreased Claudin-1 expression to promote the "leaky gut" phenotype, altered the localization of tight junction proteins, and inhibited actin polymerization. The study also found that the expression of CRT was significantly reduced in the colon tissue of IBD patients, suggesting that CRT may be a potential target for treating IBD (Hall et al., 2020). Recently, the protective role of IECs Golgi membrane protein 1 (GOLM1) in IBD has been demonstrated. IECs-specific deletion of GOML1 leads to greater susceptibility to DSS-induced colitis and Azoxymethane (AOM)/ DSS-induced colonic carcinogenesis in mice. GOLM1 interacts with the Notch2 intracellular structural domain to maintain intestinal epithelial barrier homeostasis by regulating the balance of the Notch signaling pathway in IECs, thereby inhibiting the development of colitis and colorectal carcinogenesis (Pu et al., 2021). Moreover, myosin light chain kinase (MLCK) is also a key regulator of intestinal barrier function. A unique structural domain within the MLCK splice variant MLCK1 mediates its recruitment to the tightly linked peri-junctional actomyosin ring (PAMR). A small molecule divertin was identified previously through screening that blocked acute TNF-induced MLCK1 recruitment, downstream myosin light chain (MLC) phosphorylation, barrier dysfunction, and diarrhea in vitro and in vivo, and inhibited the progression of experimental IBD (Graham et al., 2019). Another recent study has found that MLCK1 can interact with the tacrolimus-binding protein FKBP8 peptidylprolyl cis/trans isomerase (PPIase) structural domain and that these interactions are critical for MLCK1 recruitment, MLC phosphorylation, and TNF-induced barrier loss. Tissue biopsies from CD patients revealed an increased number of intercellularly linked MLCK1-FKBP8 interactions compared to controls. Blocking MLCK1-FKBP8 binding reversed MLCK1-FKBP8

TABLE 1 Targeting IEC modulators or signaling pathways to treat IBD.

Modulators/pathways	Mode of action	<b>Biological process</b>	References
Creatine transporter protein SLC6A8	Promote expression and localization of tight junction	Regulate energy balance and maintain	Hall et al. (2020)
	proteins Claudin1/2 and actin polymerization	barrier formation and healing function	
Golgi membrane protein 1 (GOLM1)	Regulate the balance of the Notch signaling pathway	Maintain barrier homeostasis	Pu et al. (2021)
Myosin light chain kinase splice variant	TNF-induced MLCK1 recruitment to peri-junctional	Maintain barrier function	Graham et al. (2019)
(MLCK1)	actomyosin ring (PAMR) mediates MLC		
	phosphorylation.		
	Interact with FKBP8 PPIase for MLCK1 recruitment and		
	MLC phosphorylation		
miR-181 family members	Regulate Wnt and immune processes	Promote intestinal recovery after injury	Jimenez et al. (2022)
Histone H3K9 methyltransferase	Silence endogenous retroviruses to inhibit DNA damage	Maintain IECs survival, barrier function,	Juznic et al. (2021)
SETDB1		and homeostasis	
m6A methyltransferase METTL14	Regulate NF-kB mRNA stability, affecting the activity of	Regulate the development and apoptosis of	Zhang et al. (2022)
	the NF-kB pathway and inhibiting TNF-mediated	colonic epithelial cells and colonic stem cells	
	apoptosis		
DNA methyltransferase 3A (DNMT3A)	DNMT3A deficiency leads to hypomethylation	Maintain intestinal epithelial barrier	Fazio et al. (2022)
		function and regeneration	
Special AT-rich sequence-binding protein	Increase the expression of the Cl-/HCO3-transport	Maintain diversity and composition of the	Ni et al. (2021)
2 (SATB2)	protein SLC26A3	gut microbiota	
5-Hydroxytryptamine (5-HT)	Reduce TGF-β secretion and inhibit the TGF-β/SMAD	Tumor-suppressive effects at the initiation	Mao et al. (2022)
	pathway	stage of CAC	
	Enhance AKT activity through 5-HT2B and activate the	Promote tumor progression in the later	
	IL-6/STAT3 pathway	stages of CAC	
Spermidine oxidase (SMOX)	Promote spermidine production and inhibit alpha-	Maintain diversity and composition of the	Gobert et al. (2022)
-	defensin expression	gut microbiota	
Stromal interaction molecule 1 (STIM1)	STIM1 deficiency controls the stimulation of the	- Maintain intestinal barrier	Liang et al. (2022)
	intestinal epithelium by reducing endoplasmic reticulum		
	stress caused by an imbalance in Ca2+ homeostasis		
RhoB	Reduced or absent RhoB inhibits the Wnt pathway and	Promote goblet cell differentiation and IECs	Yang et al. (2022)
	activates the p38 MAPK pathway	proliferation, increase SCFA-producing	Ē
		bacteria abundance and elevate the levels of	
		SCFAs and receptors	
Leukemia inhibitory factor (LIF)	Inhibit Th17 cells differentiation by phosphorylating and	Alleviate colonic inflammation	Zhang et al. (2019)
· · · ·	activating STAT4 and suppressing STAT3-induced IL17		0
	gene expression		
Alpha B-crystallin (CRYAB)	Reduce cellular inflammatory factor (TNF-α, IL-6, IL-1β,	Protect barrier integrity	Xu et al. (2019)
<b>A 7 - - -</b>	IL-8) production by inhibiting IKK $\beta$ activity	0,1	
deSUMOylase SENP7	Ubiquitin ligase SIAH2 downregulates SENP7 through	Control intestinal inflammation	Suhail et al. (2019)
	ubiquitination, which reduces pro-inflammatory		
	mechanisms through γδ T cells.		
Epithelial NF-κB2 signaling	Amplify RelA activity and increase RelA-driven	Induce abnormal intestinal inflammation	Chawla et al. (2021)
	inflammatory gene expression		
Guanylate cyclase C (GC-C)	Regulate intestinal ion and fluid secretion through	Loss of overall internal environmental	Mishra et al. (2021)
	cGMP production and activation of cGMP-dependent	homeostasis, fluid ion imbalance, and	
	protein kinase II	dysregulation of the intestinal microbiota	
Epithelial autonomous NAIP/NLRC4	Drive IEC pyroptosis or apoptosis and promote IEC	Mitigate TNF-induced disruption of the	Fattinger et al. (2021)
inflammasomes	expulsion	intestinal epithelial barrier	0 · · · · · · · ·/
NLRP3 Inflammasomes	miR223 mediates NLRP3 regulation and NLRP3 pathway	Inhibition of inflammasomes suppresses	Kanneganti (2017)
	blockage reduces IL-1β production	excessive inflammatory responses in the	
	5 FI	intestine	

(Continued)

#### TABLE1 (Continued)

Modulators/pathways	Mode of action	Biological process	References
ADP/P2Y1	ADP activates NLRP3 inflammasomes via P2Y1		Zhang et al. (2020)
	receptors and promotes NLRP3 inflammasomes		
	component ASC phosphorylation to increase IL-1 $\beta$		
	production		
NLRP6 inflammasomes	Deubiquitinating enzyme Cyld binds to NLRP6, removes		Mukherjee et al.
	its ubiquitination modifications, inhibits NLRP6-ASC		(2020)
	inflammasome formation, and regulates the maturation		
	process of IL-18.		
Nicotinamide adenine dinucleotide	The absence of intestinal epithelial NOX1 combined with	Promote lymphoplasmacytosis	Hsu et al. (2022)
phosphate (NADPH) oxidase 1 (NOX1)	$\mathrm{TNF}\alpha$ stimulation alters the stem cell microenvironment		
	and stem cell differentiation		
Superoxide dismutase 1 (SOD1)	Restoring SOD activity inhibits p38-MAPK/NF-ĸB	SOD1 deletion enhances oxidative stress	Hwang et al. (2020)
	signal-mediated inflammation and apoptosis.	and disrupts the intestinal epithelial barrier,	
		reduces antioxidant enzyme activity, and	
		increases colonic infiltration of pro-	
		inflammatory immune cells	
Multidrug resistance 1 (MDR1)	Reduce mROS	Protect mitochondria against xenotoxins	Ho et al. (2018)
		through its efflux function	
3-mercaptopyruvate sulfurtransferase	Regulate the AKT/apoptotic axis in IECs, inhibit pro-	Protect the intestine from inflammation and	Zhang et al. (2022)
(MPST)	inflammatory cytokines expression, ROS production	apoptosis incidence	
Reductively modified albumin (r-Alb)	Inhibit cellular ROS and superoxide production through	Reduce the cellular damage caused by	Yang et al. (2021)
	sulfhydryl (-SH)	oxidative stress	

interactions, MLCK1 recruitment, and barrier loss *in vitro* and *in vivo* (Zuo et al., 2022). These studies provide a new potential therapeutic target for diseases related to intestinal barrier dysfunction, such as IBD.

There are fewer studies on post-transcriptional regulation in the maintenance of intestinal homeostasis and the development of colitis. A recently published study showed that the conserved miR-181 family members in the intestinal epithelium are downregulated in IBD and mouse colitis samples, and their expression promotes intestinal recovery after injury by regulating Wnt and immune processes, which could be a potential new target for the treatment of intestinal inflammation (Jimenez et al., 2022). Furthermore, SETDB1 is a histone H3K9 methyltransferase that plays a regulatory role in intestinal epithelial homeostasis and IBD. Recent studies reveal that SETDB1 can maintain intestinal epithelial homeostasis by silencing endogenous retroviruses to inhibit DNA damage. Mice specifically deficient in SETDB1 in the intestinal epithelium exhibited impaired intestinal epithelial differentiation, impaired intestinal barrier, enhanced intestinal inflammation, and reduced survival. Meanwhile, some missense mutations associated with SETDB1 loss of function were significantly enriched in IBD patients (Juznic et al., 2021). Taken together, the results of this study suggest that SETDB1 may be a potential target for the treatment of IBD. In recent years, many studies have found that the methylation modification of RNA N6-methyladenosine (m6A) is involved in the maintenance of many types of stem cells, but whether RNA m6A also plays a vital

role in intestinal stem cells is unclear. Recently, Li's group found that m6A methyltransferase METTL14 plays an essential role in colonic stem cell self-renewal, and it may regulate the development and apoptosis of colonic epithelial cells and colonic stem cells by regulating the stability of NF-kB mRNA, affecting the activity of NF-kB pathway, and inhibiting TNF-mediated apoptosis (Zhang et al., 2022). Therefore, m6A is closely related to the development of colitis and can be used as a potential clinical target for the treatment of colitis. DNA methyltransferase 3A (DNMT3A) is involved in DNA methylation modifications and its genetic variants are associated with IBD. A recent study observed downregulation of DNMT3A expression in IECs in CD patients. Mechanistically, DNMT3A deficiency leads to hypomethylation, which impairs the function and regeneration of the intestinal epithelial barrier, making mice more susceptible to DSS-induced colitis, implying a role for impaired epithelial DNMT3A function in the etiology of IBD (Fazio et al., 2022).

Recently, the role of some regulatory factors known to be associated with colorectal carcinogenesis and development in IBD has also been revealed. Special AT-rich sequence-binding protein 2 (SATB2) is a potential diagnostic and prognostic marker for CRC, but its role in colitis and CAC is unclear. A recent study found that intestinal epithelial cell-specific SATB2 deficiency worsened colitis and CAC in mice by altering the diversity and composition of the gut microbiota and decreasing the expression of the Cl-/HCO3-transport protein SLC26A3 (Ni et al., 2021). Moreover, 5-hydroxytryptamine (5-HT) is commonly associated with CAC, and the conclusions on how 5-HT affects CAC are not vet uniform. 5-HT2B is one of the receptors for 5-HT, which is expressed in IECs. A recent study revealed that 5-HT/5-HT2B/ TGF-β signaling exerts tumor-suppressive effects at the initiation stage of CAC while promoting tumor progression in the later stages. 5-HT2B-deficient IECs, on the one hand, reduce TGF-β secretion and inhibit the TGF- $\beta$ /SMAD pathway; on the other hand, it enhances AKT activity through 5-HT2B and thus activate the IL-6/STAT3 pathway, enhance the intestinal inflammatory response, and exacerbates IEC destruction, thus promoting CAC tumorigenesis. Antibody blockade of IL-6 reverses the promoting effect of 5-HT2B deficiency on CAC development. TCGA data show a positive correlation between 5-HT2B expression and prognosis in CRC patients (Mao et al., 2022). Previous studies have shown that spermidine oxidase (SMOX) promotes spermidine production and regulates colitis. In a recent study, SMOX deficiency in mice worsened DSS-induced colitis and AOM/DSS-induced colonic tumorigenesis, increased alphadefensin expression, and induced dysbiosis of gut microbiota, leading to reduced abundance of Prevotella and increased Proteobacteria and Deferribacteres. In contrast, spermidine supplementation reversed the above phenotype. In addition, down-regulation of SMOX expression was observed in inflamed colonic tissues from patients with UC and patients with colitisassociated heterotypic hyperplasia (Gobert et al., 2022). Furthermore, stromal interaction molecule 1 (STIM1) is an integral component of the store-operated calcium entry (SOCE) process that promotes CRC and T-cell-mediated inflammatory diseases. However, it is unclear whether STIM1 in IECs is involved in the pathological process of IBD. Recent studies have found that STIM1 deficiency in the intestinal epithelium in colitis controls the stimulation of the intestinal epithelium by commensal bacteria by reducing endoplasmic reticulum stress caused by an imbalance in Ca2+ homeostasis, reducing loss of goblet cells, and maintaining the integrity of the mucus layer (Liang et al., 2022). STIM1 is a vital regulator of maintaining the intestinal barrier and a potential target for IBD treatment. RhoB, a member of the small Rho GTPase family, exhibits rapid upregulation when the organism is induced by genotoxic stress, LPS, inflammatory cytokines, growth factors and toxins, and is involved in a variety of cellular processes; however, the role of RhoB in colitis remains unclear. Recently, Wang Quan's team found that RhoB is significantly upregulated in UC, and that reduced or absent RhoB promotes goblet cell differentiation and proliferation of IECs by inhibiting the Wnt pathway and activating the p38 MAPK pathway. In addition, decreased RhoB increased the abundance of SCFA-producing bacteria and elevated the levels of SCFAs and their receptors. These findings suggest that RhoB is a potential molecular marker for the diagnosis of IBD, as well as a potential therapeutic target

The intestine is the fastest self-renewing tissue in the adult mammalian organism, and its primary stem cells are located in the crypt of the intestinal villi. Stem cells in this location have a high capacity for self-renewal and differentiation and can differentiate into intestinal epithelial absorptive cells and intestinal epithelial endocrine cells after 4-5 mitotic divisions. Among them, Paneth cells and Tuff cells are involved in the maintenance of intestinal stem cells. Different subtypes of cells differentiated from intestinal stem cells are essential in maintaining intestinal homeostasis by producing a mucus layer covering the surface of the intestinal epithelial cells, recognizing pathogens, and producing antimicrobial peptides to ensure an effective immune response (Peterson and Artis, 2014). Moreover, a recent study found that leukemia inhibitory factor (LIF), a cytokine in the IL-6 family secreted by IECs, alleviates colonic inflammation by inhibiting the differentiation of Th17 cells in a mouse model of colitis. Mechanistically, LIF inhibits the differentiation of Th17 cells by phosphorylating and activating STAT4 and suppressing STAT3induced IL17 gene expression (Zhang et al., 2019). The results of this study provide new potential therapeutic targets for the treatment of IBD. Alpha B-crystallin (CRYAB) is a small heat shock protein that plays a protective role in intestinal inflammation. A recent study found that CRYAB expression was significantly reduced in both IBD patients and DSS-induced colitis mouse models and negatively correlated with TNF- $\alpha$  and IL-6 levels; CRYAB could reduce cellular inflammatory factors (TNF-α, IL-6, IL-1β, IL-8) production *in vitro* by inhibiting IKKβ activity, and in colitis mice in vivo (Xu et al., 2019). This suggests that CRYAB may be a potential therapeutic target for IBD, as it protects the integrity of the intestinal barrier and alleviates colitis symptoms. Previous studies have found that epithelial deSUMOylation is associated with IBD. The deSUMOylase SENP7 and its interacting group regulate epithelial-immune cross-reactivity. In healthy cells, the ubiquitin ligase SIAH2 negatively regulates SENP7 through ubiquitination. Upregulation of epithelial SENP7 induces pro-inflammatory mechanisms through  $\gamma\delta$  T cells. Knockdown of SENP7 or clearance of  $\gamma\delta$  T cells suppressed DSS-induced intestinal inflammation; upregulation of SENP7 expression was strongly statistically correlated with higher clinical disease indices in IBD patients (Suhail et al., 2019). This study shows that epithelial SENP7 is necessary to control intestinal inflammation and highlights its importance as a potential drug target.

In addition, excessive activation of RelA/NF- $\kappa$ B causes abnormal inflammation associated with IBD. The canonical NF- $\kappa$ B module regulates intranuclear activation of RelA dimers and induces e pro-inflammatory gene expression, and uncontrolled RelA activity induces abnormal intestinal inflammation. A recently published study found that atypical NF- $\kappa$ B2 signaling amplifies RelA activity in colonic epithelial cells, increases RelA-driven inflammatory gene expression, and exacerbates intestinal pathogenesis in IBD patients and mice with colitis, suggesting that the atypical NF- $\kappa$ B2 pathway may be a potential therapeutic target for inflammatory diseases (Chawla et al., 2021). Activating mutations in guanylate cyclase C (GC-C), a target receptor for the gastrointestinal peptide hormone guanylate and urinary guanylate, and bacterial heat-stable enterotoxins can both cause early-onset diarrhea and chronic

(Yang et al., 2022).

IBD. GC-C regulates intestinal ion and fluid secretion through cGMP production and activation of cGMP-dependent protein kinase II. A recent study constructing activating mutant mice with the *guanylate cyclase C (Gucy2c)* gene found that mutant mice have increased intestinal cGMP, leading to loss of overall internal environmental homeostasis, fluid ion imbalance, dysregulation of the intestinal microbiota, and susceptibility to colitis, suggesting that gut-associated cGMP signaling pathway may mediate colitis and flora dysbiosis (Mishra et al., 2021).

Inflammasomes are an important natural immune component expressed in both immune cells and non-immune cells (Lamkanfi and Dixit, 2014). By regulating the degree of inflammation and cell death in response to pathogen-associated molecular patterns (PAMPs) or danger-associated molecular patterns (DAMPs), they can either protect or harm the host (Man, 2018). Inflammasomes play an important role in inflammation and pathogen clearance. However, the mechanisms by which endogenous danger signals activate inflammasomes and the association of inflammasomes with inflammatory diseases have not been clarified. Earlier studies have shown that Salmonella typhimurium expresses a specific PAMP, a ligand for NLR family of apoptosis inhibitory proteins (NAIPs), during infection of IECs and high expression of NAIP/ NLRC4 inflammasomes in IECs specifically inhibits the dissemination of S. typhimurium from the intestine to the whole body (Rauch et al., 2017; Hausmann et al., 2020). A recent study demonstrated that epithelial autonomous NAIP/NLRC4 inflammasomes mitigate TNF-induced disruption of the intestinal epithelial barrier by driving IEC pyroptosis or apoptosis and promoting IEC expulsion (Fattinger et al., 2021). miR-223 expression is increased in inflammatory tissues in IBD (Zhang et al., 2021). NLRP3 is a component of inflammasomes, and NLRP3 expression is elevated in colon and myeloid cells in miR-223-deficient environments. Drugs block the NLRP3 pathway, reduce IL-1ß production, and attenuate colitis (Kanneganti, 2017). Recently, Zhang et al. revealed the mechanism by which endogenous danger signals activate inflammasomes to promote IBD. ADP, a danger signal released during colonic injury from IECs, activates NLRP3 inflammasomes via P2Y1 receptors and promotes phosphorylation of NLRP3 inflammasomes component ASC to increase IL-1ß production, thereby worsening DSS-induced colitis in mice. Extracellular ADP activates NLRP3 inflammasomes via P2Y1 receptor-mediated calcium signaling, and deletion of P2Y1 receptors or treatment with P2Y1 receptor inhibitors inhibits NLRP3 inflammasomes activation to alleviate colitis (Zhang et al., 2020). The results of this study suggest that ADP/P2Y1 may serve as a potential therapeutic target for IBD. NLRP6 inflammasomes regulate the intestinal inflammatory response and the organism's resistance to microbial responses, but the molecular mechanism of how to inhibit NLRP6 function to avoid tissue damage caused by excessive inflammatory response is not clear. A recent study showed that the deubiquitinating enzyme Cyld binds to NLRP6, removes its ubiquitination modifications, inhibits NLRP6-ASC inflammasomes formation, regulates the maturation process of IL-18, and suppresses excessive inflammatory responses in the intestine (Mukherjee et al., 2020). This finding may provide new ideas for the treatment of intestinal inflammation.

The intestinal epithelium is constantly exposed to inducers of ROS, such as commensal microorganisms (Kajino-Sakamoto et al., 2010). In addition to an excessive inflammatory response, oxidative stress is considered a major feature of IBD (Tian et al., 2017). In the gastrointestinal tract, ROS production by nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (NOX/DUOX) is a key biological mechanism regulating pathogen killing, host-microbe interactions and tissue repair after injury (Aviello et al., 2019). ROS production via enterocyte NOX1 has been shown to transduce microbial signals that promote epithelial wound healing (Alam et al., 2014) and mutations in genes that induce ROS production, such as Nox1, are highly associated with IBD (Aviello et al., 2019; Hsu et al., 2022). Mechanistically, the absence of intestinal epithelial NOX1 combined with  $TNF\alpha$ stimulation can alter the stem cell microenvironment and stem cell differentiation, promote the number of lymphoplasmacytes and thus accelerate the progression of colitis, which may provide new insights into the prevention and treatment of IBD (Hsu et al., 2022). Superoxide dismutase 1 (SOD1) is one of the three superoxide dismutases responsible for the destruction of free superoxide radicals in the body. However, the role of SOD1 in oxidative stress in colitis is unclear. Hwang et al. found that SOD1 deletion enhances oxidative stress in mice and disrupts the intestinal epithelial barrier, reduces antioxidant enzyme activity, and increases colonic infiltration of pro-inflammatory immune cells to worsen DSS-induced colitis in mice. Restoring SOD activity can inhibit p38-MAPK/NF-kB signal-mediated inflammation and apoptosis, thereby alleviating colitis (Hwang et al., 2020). In mice, loss of multidrug resistance 1 (MDR1) function leads to colitis similar to human IBD. Ho et al. showed that MDR1 has a protective effect on mitochondria, where MDR1 deficiency leads to mitochondrial dysfunction, while increased mROS drive the development of colitis. Since MDR1 protects against xenotoxins primarily through its efflux function, the results of this study suggest a unique mitochondrial toxin genetic susceptibility interaction leading to mitochondrial dysfunction, a novel pathogenic mechanism that could provide many new therapeutic opportunities for IBD (Ho et al., 2018). Moreover, endogenous hydrogen sulfide (H2S) has anti-inflammatory activity in IBD, and the role of 3-mercaptopyruvate sulfurtransferase (MPST), a key enzyme regulating endogenous H2S biosynthesis, in IBD is not yet clear. A recent study found that MPST likely protects the intestine from inflammation by regulating the AKT/apoptotic axis in IECs, inhibiting the expression of pro-inflammatory cytokines, ROS production, and the incidence of apoptosis, providing a new therapeutic strategy for colitis (Zhang et al., 2022). Albumin is the most abundant matrix protein in the body and performs a variety of biological functions, including the ability to resist oxidative stress through free sulfhydryl groups (-SH). Oxidative stress is an important feature of colitis, and studies have shown that colitis can

10.3389/fmicb.2022.1065608

significantly reduce albumin and increase oxidized albumin (Krzystek-Korpacka et al., 2008; Khan et al., 2017). However, no studies have yet explored the potential efficacy of albumin in the treatment of colitis. A recent work showed that DSS-induced administration of reductively modified albumin (r-Alb) to mice with colitis effectively improved the colitis phenotype by the mechanism that r-Alb inhibits cellular ROS and superoxide production through sulfhydryl (-SH) and reduces the cellular damage caused by oxidative stress. The results of this study provide a new intervention technique for the clinical treatment of colitis, while r-Alb has a promising application as an effective antioxidant (Yang et al., 2021).

## Conclusion

Multiple factors are involved in the pathogenesis of IBD, such as genetic, environmental, infectious, and immunological factors, among which intestinal inflammation and immune dysfunction play an important role. In IBD patients, molecules such as SCFAs, bile acids and microbial tryptophan metabolites are altered (Sun et al., 2017; Heinken et al., 2019; Scott et al., 2020). These microbe-derived compounds function as signaling molecules, mediating host-microbiota communication and regulating immune homeostasis. Recent research suggests that SCFAs, particularly butyrate, have immunomodulatory properties. Furthermore, SCFAs can activate the signaling cascade that controls immune function via GPCRs. With the advancement of molecular biology techniques, the value of epigenetics as one of the pathways regulating gene expression in IBD has been gradually explored and established to be particularly relevant to IBD. Butyrate has recently been discovered to regulate epithelial gene expression or immune cells via epigenetic pathways. Besides, the latest evidence suggests that specific bile acid metabolites, tryptophan metabolites, and PSA interact with adaptive immune cells to improve the inflammatory environment of IBD. In addition to traditional bacterial metabolites, certain new bacteria species and their effector molecules have been identified to regulate immune factors release and improve IBD by binding to IECs or immune cell surface receptors. On the contrary, many new bacteria species and the virulence factors they contain potentially promote IBD by disrupting epithelial barrier, promoting pro-inflammatory factors secretion, and interfering normal gut microbiota. Recent research has confirmed the importance of miRNAs in targeting specific molecules in signaling pathways that regulate intestinal barrier homeostasis, inflammation, and autophagy. Several studies have found specific miRNAs linked to IBD and attempted to use them as diagnostic biomarkers (Soroosh et al., 2018).

The long-term persistence of chronic inflammation in IBD is a major contributor to tumor transformation and the development of CAC. In the past decades, significant progress has been made in the immunological mechanisms of IBD, providing new strategies and ideas for treating IBD. The subsequent introduction of biologics, such as TNF- $\alpha$  blocker, replaced nonselective anti-inflammatory corticosteroids in IBD management. However, these therapies still have the potential for primary unresponsiveness, secondary loss of response, opportunistic infections, and cancer development. Therefore, there is an urgent need to develop novel and effective therapies that target specific signaling pathways in the pathogenesis of IBD. Disruption of the intestinal barrier, abnormal death of epithelial cells, and subsequent inflammation are at the heart of chronic inflammatory and infectious gastrointestinal diseases (Patankar and Becker, 2020). Recent studies have shown that the Notch, Wnt, and NfkB signaling pathways are involved in the pathogenesis of IBD and different upstream signaling molecules can activate these pathways, thus serving as potential targets for the treatment of IBD. Moreover, RNA methylation is a new class of RNA epigenetic modifications discovered in recent years and can regulate mRNA expression. One of the most common modifications is m6A. There is a growing body of research involving RNA methylation and disease, covering tumors (Su et al., 2019), neurological disorders (Jiang et al., 2022), obesity (Li et al., 2020), and immune response (Wang et al., 2020), but the relationship with IBD needs further investigation. Elevated levels of NLRP3 inflammasome and pro-inflammatory cytokines are the main pathological mechanisms of IBD (Zhen and Zhang, 2019). Therefore, targeting NLRP3 inflammasome offers a promising strategy for the treatment of IBD. Chronic intestinal inflammation is related with the overproduction of ROS, which leads to oxidative stress. Recent studies have identified several key molecules that play varying degrees of regulatory functions in the production of ROS and could be explored as possible targets for IBD therapy. In the future, altering specific genetic loci may be a promising therapeutic approach for IBD. In addition, novel antibodies or inhibitors, combination therapy regimens, and multifactor blockers are also expected to break the bottleneck of IBD treatment and improve disease for patients with IBD. Furthermore, in the past three decades, there has been a rapid development in the clinical diagnosis and treatment of IBD. However, numerous issues still need to be explored in depth, such as genetic testing, serological markers in the disease population, imaging of the disease, endoscopic assessment, monitoring, drug specificity, and FMT. It is the future trend to further develop relatively specific novel drugs, develop IBD assessment models through extensive sample population studies, and provide personalized treatment to patients.

## Author contributions

BL and CnW wrote the paper. CoW, WS, and XH made figures. WC made the table. NL and DX conceived of the presented idea. All authors contributed to the article and approved the submitted version.

# Funding

This work is financially supported by the Qingdao Postdoctoral Applied Research Project (grant no. RZ2200001423).

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships

## References

Ahmad, T. R., and Haeusler, R. A. (2019). Bile acids in glucose metabolism and insulin signalling – mechanisms and research needs. *Nat. Rev. Endocrinol.* 15, 701–712. doi: 10.1038/s41574-019-0266-7

Alam, A., Leoni, G., Wentworth, C. C., Kwal, J. M., Wu, H., Ardita, C. S., et al. (2014). Redox signaling regulates commensal-mediated mucosal homeostasis and restitution and requires formyl peptide receptor 1. *Mucosal Immunol.* 7, 645–655. doi: 10.1038/mi.2013.84

Alexander, M., Ang, Q. Y., Nayak, R. R., Bustion, A. E., Sandy, M., Zhang, B., et al. (2022). Human gut bacterial metabolism drives Th17 activation and colitis. *Cell Host Microbe* 30, 17–30.e9. doi: 10.1016/j.chom.2021.11.001

Allaire, J. M., Crowley, S. M., Law, H. T., Chang, S. Y., Ko, H. J., and Vallance, B. A. (2018). The intestinal epithelium: central coordinator of mucosal immunity. *Trends Immunol.* 39, 677–696. doi: 10.1016/j.it.2018.04.002

Aviello, G., Singh, A. K., O'Neill, S., Conroy, E., Gallagher, W., D'Agostino, G., et al. (2019). Colitis susceptibility in mice with reactive oxygen species deficiency is mediated by mucus barrier and immune defense defects. *Mucosal Immunol.* 12, 1316–1326. doi: 10.1038/s41385-019-0205-x

Barnich, N., Carvalho, F. A., Glasser, A. L., Darcha, C., Jantscheff, P., Allez, M., et al. (2007). CEACAM6 acts as a receptor for adherent-invasive *E. coli*, supporting ileal mucosa colonization in Crohn disease. *J. Clin. Invest.* 117, 1566–1574. doi: 10.1172/JCI30504

Brown, A. J., Goldsworthy, S. M., Barnes, A. A., Eilert, M. M., Tcheang, L., Daniels, D., et al. (2003). The orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *J. Biol. Chem.* 278, 11312–11319. doi: 10.1074/jbc.M211609200

Campbell, C., McKenney, P. T., Konstantinovsky, D., Isaeva, O. I., Schizas, M., Verter, J., et al. (2020). Bacterial metabolism of bile acids promotes generation of peripheral regulatory T cells. *Nature* 581, 475–479. doi: 10.1038/s41586-020-2193-0

Cao, Y., Wang, Z., Yan, Y., Ji, L., He, J., Xuan, B., et al. (2021). Enterotoxigenic Bacteroidesfragilis promotes intestinal inflammation and malignancy by inhibiting exosome-packaged miR-149-3p. *Gastroenterology* 161, 1552–1566.e12. doi: 10.1053/j.gastro.2021.08.003

Caruso, R., Lo, B. C., and Nunez, G. (2020). Host-microbiota interactions in inflammatory bowel disease. *Nat. Rev. Immunol.* 20, 411–426. doi: 10.1038/s41577-019-0268-7

Chawla, M., Mukherjee, T., Deka, A., Chatterjee, B., Sarkar, U. A., Singh, A. K., et al. (2021). An epithelial Nfkb2 pathway exacerbates intestinal inflammation by supplementing latent RelA dimers to the canonical NF-kappaB module. *Proc. Natl. Acad. Sci. U. S. A.* 118:e2024828118. doi: 10.1073/pnas.2024828118

Chung, L., Thiele Orberg, E., Geis, A. L., Chan, J. L., Fu, K., DeStefano Shields, C. E., et al. (2018). Bacteroides fragilis toxin coordinates a pro-carcinogenic inflammatory Cascade via targeting of colonic epithelial cells. *Cell Host Microbe* 23, 203–214.e5. doi: 10.1016/j.chom.2018.01.007

Coskun, M. (2014). Intestinal epithelium in inflammatory bowel disease. *Front. Med.* (*Lausanne*). 1:24. doi: 10.3389/fmed.2014.00024

Dasgupta, S., Erturk-Hasdemir, D., Ochoa-Reparaz, J., Reinecker, H. C., and Kasper, D. L. (2014). Plasmacytoid dendritic cells mediate anti-inflammatory responses to a gut commensal molecule via both innate and adaptive mechanisms. *Cell Host Microbe* 15, 413–423. doi: 10.1016/j.chom.2014.03.006

Fattinger, S. A., Geiser, P., Samperio Ventayol, P., Di Martino, M. L., Furter, M., Felmy, B., et al. (2021). Epithelium-autonomous NAIP/NLRC4 prevents TNF-driven

that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

inflammatory destruction of the gut epithelial barrier in salmonella-infected mice. *Mucosal Immunol.* 14, 615–629. doi: 10.1038/s41385-021-00381-y

Fazio, A., Bordoni, D., Kuiper, J. W. P., Weber-Stiehl, S., Stengel, S. T., Arnold, P., et al. (2022). DNA methyltransferase 3A controls intestinal epithelial barrier function and regeneration in the colon. *Nat. Commun.* 13:6266. doi: 10.1038/ s41467-022-33844-2

Foley, S. E., Dente, M. J., Lei, X., Sallis, B. F., Loew, E. B., Meza-Segura, M., et al. (2022). Microbial metabolites orchestrate a distinct multi-tiered regulatory network in the intestinal epithelium that directs P-glycoprotein expression. *mBio* 13:e0199322. doi: 10.1128/mbio.01993-22

Furusawa, Y., Obata, Y., Fukuda, S., Endo, T. A., Nakato, G., Takahashi, D., et al. (2013). Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 504, 446–450. doi: 10.1038/nature12721

Gobert, A. P., Latour, Y. L., Asim, M., Barry, D. P., Allaman, M. M., Finley, J. L., et al. (2022). Protective role of Spermidine in colitis and colon carcinogenesis. *Gastroenterology* 162, 813–827.e8. doi: 10.1053/j.gastro.2021.11.005

Graham, W. V., He, W., Marchiando, A. M., Zha, J., Singh, G., Li, H. S., et al. (2019). Intracellular MLCK1 diversion reverses barrier loss to restore mucosal homeostasis. *Nat. Med.* 25, 690–700. doi: 10.1038/s41591-019-0393-7

Hall, C. H. T., Lee, J. S., Murphy, E. M., Gerich, M. E., Dran, R., Glover, L. E., et al. (2020). Creatine transporter, reduced in colon tissues from patients with inflammatory bowel diseases, regulates energy balance in intestinal epithelial cells, epithelial integrity, and barrier function. *Gastroenterology* 159, 984–998e1. doi: 10.1053/j.gastro.2020.05.033

Hang, S., Paik, D., Yao, L., Kim, E., Trinath, J., Lu, J., et al. (2019). Bile acid metabolites control T(H)17 and T(reg) cell differentiation. *Nature* 576, 143–148. doi: 10.1038/s41586-019-1785-z

Hausmann, A., Böck, D., Geiser, P., Berthold, D. L., Fattinger, S. A., Furter, M., et al. (2020). Intestinal epithelial NAIP/NLRC4 restricts systemic dissemination of the adapted pathogen salmonella Typhimurium due to site-specific bacterial PAMP expression. *Mucosal Immunol.* 13, 530–544. doi: 10.1038/s41385-019-0247-0

Heinken, A., Ravcheev, D. A., Baldini, F., Heirendt, L., Fleming, R. M., and Thiele, I. (2019). Systematic assessment of secondary bile acid metabolism in gut microbes reveals distinct metabolic capabilities in inflammatory bowel disease. *Microbiome.* 7, 1–18.

Henke, M. T., Kenny, D. J., Cassilly, C. D., Vlamakis, H., Xavier, R. J., and Clardy, J. (2019). Ruminococcus gnavus, a member of the human gut microbiome associated with Crohn's disease, produces an inflammatory polysaccharide. *Proc. Natl. Acad. Sci. U. S. A.* 116, 12672–12677. doi: 10.1073/pnas.1904099116

Hinrichsen, F., Hamm, J., Westermann, M., Schroder, L., Shima, K., Mishra, N., et al. (2021). Microbial regulation of hexokinase 2 links mitochondrial metabolism and cell death in colitis. *Cell Metab.* 33, 2355–2366.e8. doi: 10.1016/j. cmet.2021.11.004

Ho, G. T., Aird, R. E., Liu, B., Boyapati, R. K., Kennedy, N. A., Dorward, D. A., et al. (2018). MDR1 deficiency impairs mitochondrial homeostasis and promotes intestinal inflammation. *Mucosal Immunol.* 11, 120–130. doi: 10.1038/mi.2017.31

Hsu, N.-Y., Nayar, S., Gettler, K., Talware, S., Giri, M., Alter, I., et al. (2022). NOX1 is essential for TNF $\alpha$ -induced intestinal epithelial ROS secretion and inhibits M cell signatures. *Gut*:gutjnl-2021-326305. doi: 10.1136/gutjnl-2021-326305

Hwang, J., Jin, J., Jeon, S., Moon, S. H., Park, M. Y., Yum, D.-Y., et al. (2020). SOD1 suppresses pro-inflammatory immune responses by protecting against oxidative stress in colitis. *Redox Biol.* 37:101760. doi: 10.1016/j.redox.2020.101760

Jiang, L., Li, X., Wang, S., Yuan, Z., and Cheng, J. (2022). The role and regulatory mechanism of m(6)a methylation in the nervous system. *Front. Genet.* 13:962774. doi: 10.3389/fgene.2022.962774

Jimenez, M. T., Clark, M. L., Wright, J. M., Michieletto, M. F., Liu, S., Erickson, I., et al. (2022). The miR-181 family regulates colonic inflammation through its activity in the intestinal epithelium. *J. Exp. Med.* 219:e20212278. doi: 10.1084/jem.20212278

Juznic, L., Peuker, K., Strigli, A., Brosch, M., Herrmann, A., Hasler, R., et al. (2021). SETDB1 is required for intestinal epithelial differentiation and the prevention of intestinal inflammation. *Gut* 70, 485–498. doi: 10.1136/gutjnl-2020-321339

Kajino-Sakamoto, R., Omori, E., Nighot, P. K., Blikslager, A. T., Matsumoto, K., and Ninomiya-Tsuji, J. (2010). TGF- $\beta$ -activated kinase 1 signaling maintains intestinal integrity by preventing accumulation of reactive oxygen species in the intestinal epithelium. J. Immunol. 185, 4729–4737. doi: 10.4049/jimmunol.0903587

Kanneganti, T.-D. (2017). Inflammatory bowel disease and the NLRP3 Inflammasome. N. Engl. J. Med. 377, 694–696. doi: 10.1056/NEJMcibr1706536

Khan, N., Patel, D., Shah, Y., Trivedi, C., and Yang, Y. X. (2017). Albumin as a prognostic marker for ulcerative colitis. *World J. Gastroenterol.* 23, 8008–8016. doi: 10.3748/wjg.v23.i45.8008

Kim, K. N., Yao, Y., and Ju, S. Y. (2019). Short chain fatty acids and fecal microbiota abundance in humans with obesity: a systematic review and metaanalysis. *Nutrients* 11:2512. doi: 10.3390/nu11102512

Krzystek-Korpacka, M., Neubauer, K., Berdowska, I., Boehm, D., Zielinski, B., Petryszyn, P., et al. (2008). Enhanced formation of advanced oxidation protein products in IBD. *Inflamm. Bowel Dis.* 14, 794–802. doi: 10.1002/ibd.20383

Lamkanfi, M., and Dixit, V. M. (2014). Mechanisms and functions of inflammasomes. *Cells* 157, 1013–1022. doi: 10.1016/j.cell.2014.04.007

Li, G., Lin, J., Zhang, C., Gao, H., Lu, H., Gao, X., et al. (2021). Microbiota metabolite butyrate constrains neutrophil functions and ameliorates mucosal inflammation in inflammatory bowel disease. *Gut Microbes* 13:1968257. doi: 10.1080/19490976.2021.1968257

Li, Q., Sun, X., Yu, K., Lv, J., Miao, C., Yang, J., et al. (2022). Enterobacter ludwigii protects DSS-induced colitis through choline-mediated immune tolerance. *Cell Rep.* 40:111308. doi: 10.1016/j.celrep.2022.111308

Li, Y., Wang, J., Huang, C., Shen, M., Zhan, H., and Xu, K. (2020). RNA N6methyladenosine: a promising molecular target in metabolic diseases. *Cell Biosci.* 10:19. doi: 10.1186/s13578-020-00385-4

Liang, X., Xie, J., Liu, H., Zhao, R., Zhang, W., Wang, H., et al. (2022). STIM1 deficiency in intestinal epithelium attenuates colonic inflammation and tumorigenesis by reducing ER stress of goblet cells. *Cell. Mol. Gastroenterol. Hepatol.* 14, 193–217. doi: 10.1016/j.jcmgh.2022.03.007

Liu, Y., Yang, M., Tang, L., Wang, F., Huang, S., Liu, S., et al. (2022). TLR4 regulates RORgammat(+) regulatory T-cell responses and susceptibility to colon inflammation through interaction with *Akkermansia muciniphila*. *Microbiome*. 10:98, doi: 10.1186/s40168-022-01296-x

Luis, A. S., Jin, C., Pereira, G. V., Glowacki, R. W. P., Gugel, S. R., Singh, S., et al. (2021). A single sulfatase is required to access colonic mucin by a gut bacterium. *Nature* 598, 332–337. doi: 10.1038/s41586-021-03967-5

Ma, N., He, T., Johnston, L. J., and Ma, X. (2020). Host-microbiome interactions: the aryl hydrocarbon receptor as a critical node in tryptophan metabolites to brain signaling. *Gut Microbes* 11, 1203–1219. doi: 10.1080/19490976.2020.1758008

Mahendran, V., Liu, F., Riordan, S. M., Grimm, M. C., Tanaka, M. M., and Zhang, L. (2016). Examination of the effects of campylobacter concisus zonula occludens toxin on intestinal epithelial cells and macrophages. *Gut Pathog.* 8:18. doi: 10.1186/s13099-016-0101-9

Man, S. M. (2018). Inflammasomes in the gastrointestinal tract: infection, cancer and gut microbiota homeostasis. *Nat. Rev. Gastro Hepat.* 15, 721–737. doi: 10.1038/ s41575-018-0054-1

Mao, L., Xin, F., Ren, J., Xu, S., Huang, H., Zha, X., et al. (2022). 5-HT2B-mediated serotonin activation in enterocytes suppresses colitis-associated cancer initiation and promotes cancer progression. *Theranostics*. 12, 3928–3945. doi: 10.7150/thno.70762

Maslowski, K. M., Vieira, A. T., Ng, A., Kranich, J., Sierro, F., Yu, D., et al. (2009). Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 461, 1282–1286. doi: 10.1038/nature08530

Mazmanian, S. K., Round, J. L., and Kasper, D. L. (2008). A microbial symbiosis factor prevents intestinal inflammatory disease. *Nature* 453, 620–625. doi: 10.1038/ nature07008

Mishra, V., Bose, A., Kiran, S., Banerjee, S., Shah, I. A., Chaukimath, P., et al. (2021). Gut-associated cGMP mediates colitis and dysbiosis in a mouse model of an activating mutation in GUCY2C. *J. Exp. Med.* 218:e20210479. doi: 10.1084/jem.20210479

Mukherjee, S., Kumar, R., Tsakem Lenou, E., Basrur, V., Kontoyiannis, D. L., Ioakeimidis, F., et al. (2020). Deubiquitination of NLRP6 inflammasome by Cyld

critically regulates intestinal inflammation. Nat. Immunol. 21, 626-635. doi: 10.1038/s41590-020-0681-x

Ni, H., Chen, Y., Xia, W., Wang, C., Hu, C., Sun, L., et al. (2021). SATB2 defect promotes colitis and colitis-associated colorectal cancer by impairing cl-/HCO3-exchange and homeostasis of gut microbiota. *J. Crohns Colitis* 15, 2088–2102. doi: 10.1093/ecco-jcc/jjab094

Nicolas, G. R., and Chang, P. V. (2019). Deciphering the chemical lexicon of hostgut microbiota interactions. *Trends Pharmacol. Sci.* 40, 430–445. doi: 10.1016/j. tips.2019.04.006

Nielsen, O. H., and Ainsworth, M. A. (2013). Tumor necrosis factor inhibitors for inflammatory bowel disease. *N. Engl. J. Med.* 369, 754–762. doi: 10.1056/ NEJMct1209614

Nielsen, H. L., Dalager-Pedersen, M., and Nielsen, H. (2020). High risk of microscopic colitis after campylobacter concisus infection: population-based cohort study. *Gut* 69, 1952–1958. doi: 10.1136/gutjnl-2019-319771

Paik, D., Yao, L., Zhang, Y., Bae, S., D'Agostino, G. D., Zhang, M., et al. (2022). Human gut bacteria produce TauEta17-modulating bile acid metabolites. *Nature* 603, 907–912. doi: 10.1038/s41586-022-04480-z

Palmela, C., Chevarin, C., Xu, Z., Torres, J., Sevrin, G., Hirten, R., et al. (2018). Adherent-invasive Escherichia coli in inflammatory bowel disease. *Gut* 67, 574–587. doi: 10.1136/gutjnl-2017-314903

Patankar, J. V., and Becker, C. (2020). Cell death in the gut epithelium and implications for chronic inflammation. *Nat Rev Gastro Hepat.* 17, 543–556. doi: 10.1038/s41575-020-0326-4

Peterson, L. W., and Artis, D. (2014). Intestinal epithelial cells: regulators of barrier function and immune homeostasis. *Nat. Rev. Immunol.* 14, 141–153. doi: 10.1038/nri3608

Plovier, H., Everard, A., Druart, C., Depommier, C., Van Hul, M., Geurts, L., et al. (2017). A purified membrane protein from Akkermansia muciniphila or the pasteurized bacterium improves metabolism in obese and diabetic mice. *Nat. Med.* 23, 107–113. doi: 10.1038/nm.4236

Postler, T. S., and Ghosh, S. (2017). Understanding the Holobiont: how microbial metabolites affect human health and shape the immune system. *Cell Metab.* 26, 110–130. doi: 10.1016/j.cmet.2017.05.008

Pu, Y., Song, Y., Zhang, M., Long, C., Li, J., Wang, Y., et al. (2021). GOLM1 restricts colitis and colon tumorigenesis by ensuring notch signaling equilibrium in intestinal homeostasis. *Signal Transduct. Target. Ther.* 6:148. doi: 10.1038/ s41392-021-00535-1

Qin, J., Li, R., Raes, J., Arumugam, M., Burgdorf, K. S., Manichanh, C., et al. (2010). A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 464, 59–65. doi: 10.1038/nature08821

Qu, S., Fan, L., Qi, Y., Xu, C., Hu, Y., Chen, S., et al. (2021). Akkermansia muciniphila alleviates dextran sulfate sodium (DSS)-induced acute colitis by NLRP3 activation. *Microbiol Spectr.* 9:e0073021. doi: 10.1128/Spectrum.00730-21

Rauch, I., Deets, K. A., Ji, D. X., von Moltke, J., Tenthorey, J. L., Lee, A. Y., et al. (2017). NAIP-NLRC4 Inflammasomes coordinate intestinal epithelial cell expulsion with eicosanoid and IL-18 release via activation of Caspase-1 and -8. *Immunity* 46, 649–659. doi: 10.1016/j.immuni.2017.03.016

Renga, G., Nunzi, E., Pariano, M., Puccetti, M., Bellet, M. M., Pieraccini, G., et al. (2022). Optimizing therapeutic outcomes of immune checkpoint blockade by a microbial tryptophan metabolite. *J. Immunother. Cancer* 10:e003725. doi: 10.1136/jitc-2021-003725

Rösch, C., Taverne, N., Venema, K., Gruppen, H., Wells, J. M., and Schols, H. A. (2017). Effects of in vitro fermentation of barley  $\beta$ -glucan and sugar beet pectin using human fecal inocula on cytokine expression by dendritic cells. *Mol. Nutr. Food Res.* 61:1600243. doi: 10.1002/mnfr.201600243

Round, J. L., and Mazmanian, S. K. (2010). Inducible Foxp3<sup>+</sup> regulatory T-cell development by a commensal bacterium of the intestinal microbiota. *Proc. Natl. Acad. Sci. U. S. A.* 107, 12204–12209. doi: 10.1073/pnas.0909122107

Scott, S. A., Fu, J., and Chang, P. V. (2020). Microbial tryptophan metabolites regulate gut barrier function via the aryl hydrocarbon receptor. *Proc. Natl. Acad. Sci. U. S. A.* 117, 19376–19387. doi: 10.1073/pnas.2000047117

Sekizuka, T., Ogasawara, Y., Ohkusa, T., and Kuroda, M. (2017). Characterization of Fusobacterium varium Fv113-gl isolated from a patient with ulcerative colitis based on complete genome sequence and transcriptome analysis. *PLoS One* 12:e0189319. doi: 10.1371/journal.pone.0189319

Seregin, S. S., Golovchenko, N., Schaf, B., Chen, J., Pudlo, N. A., Mitchell, J., et al. (2017). NLRP6 protects Il10(–/–) mice from colitis by limiting colonization of *Akkermansia muciniphila*. *Cell Rep.* 19, 733–745. doi: 10.1016/j.celrep.2017. 03.080

Shen, Y., Giardino Torchia, M. L., Lawson, G. W., Karp, C. L., Ashwell, J. D., and Mazmanian, S. K. (2012). Outer membrane vesicles of a human commensal mediate immune regulation and disease protection. *Cell Host Microbe* 12, 509–520. doi: 10.1016/j.chom.2012.08.004

Silpe, J. E., Wong, J. W. H., Owen, S. V., Baym, M., and Balskus, E. P. (2022). The bacterial toxin colibactin triggers prophage induction. *Nature* 603, 315–320. doi: 10.1038/s41586-022-04444-3

Singh, S., George, J., Boland, B. S., Vande Casteele, N., and Sandborn, W. J. (2018). Primary non-response to tumor necrosis factor antagonists is associated with inferior response to second-line biologics in patients with inflammatory bowel diseases: a systematic review and meta-analysis. *J. Crohns Colitis* 12, 635–643. doi: 10.1093/ecco-icc/jiy004

Sinha, S. R., Haileselassie, Y., Nguyen, L. P., Tropini, C., Wang, M., Becker, L. S., et al. (2020). Dysbiosis-induced secondary bile acid deficiency promotes intestinal inflammation. *Cell Host Microbe* 27, 659–670.e5. doi: 10.1016/j.chom.2020.01.021

Soderholm, A. T., and Pedicord, V. A. (2019). Intestinal epithelial cells: at the interface of the microbiota and mucosal immunity. *Immunology* 158, 267–280. doi: 10.1111/imm.13117

Sommer, F., and Backhed, F. (2013). The gut microbiota-masters of host development and physiology. *Nat. Rev. Microbiol.* 11, 227-238. doi: 10.1038/nrmicro2974

Sommer, K., Wiendl, M., Müller, T. M., Heidbreder, K., Voskens, C., Neurath, M. F., et al. (2021). Intestinal mucosal wound healing and barrier integrity in IBDcrosstalk and trafficking of cellular players. *Front. Med. (Lausanne).* 8:643973. doi: 10.3389/fmed.2021.643973

Soroosh, A., Koutsioumpa, M., Pothoulakis, C., and Iliopoulos, D. (2018). Functional role and therapeutic targeting of microRNAs in inflammatory bowel disease. *Am. J. Physiol. Gastrointest. Liver Physiol.* 314, G256–G262. doi: 10.1152/ajpgi.00268.2017

Su, Y., Huang, J., and Hu, J. (2019). m6A RNA methylation regulators contribute to malignant progression and have clinical prognostic impact in gastric cancer. *Front. Oncol.* 9:1038. doi: 10.3389/fonc.2019.01038

Suhail, A., Rizvi, Z. A., Mujagond, P., Ali, S. A., Gaur, P., Singh, M., et al. (2019). DeSUMOylase SENP7-mediated epithelial signaling triggers intestinal inflammation via expansion of Gamma-Delta T cells. *Cell Rep.* 29, 3522–3538.e7. doi: 10.1016/j. celrep.2019.11.028

Sun, M., Wu, W., Liu, Z., and Cong, Y. (2017). Microbiota metabolite short chain fatty acids, GPCR, and inflammatory bowel diseases. *J. Gastroenterol.* 52, 1–8. doi: 10.1007/s00535-016-1242-9

Tahara, T., Shibata, T., Kawamura, T., Okubo, M., Ichikawa, Y., Sumi, K., et al. (2015). Fusobacterium detected in colonic biopsy and clinicopathological features of ulcerative colitis in Japan. *Dig. Dis. Sci.* 60, 205–210. doi: 10.1007/s10620-014-3316-y

Tailford, L. E., Owen, C. D., Walshaw, J., Crost, E. H., Hardy-Goddard, J., Le Gall, G., et al. (2015). Discovery of intramolecular trans-sialidases in human gut microbiota suggests novel mechanisms of mucosal adaptation. *Nat. Commun.* 6:7624. doi: 10.1038/ncomms8624

Thangaraju, M., Cresci, G. A., Liu, K., Ananth, S., Gnanaprakasam, J. P., Browning, D. D., et al. (2009). GPR109A is a G-protein-coupled receptor for the bacterial fermentation product butyrate and functions as a tumor suppressor in colon. *Cancer Res.* 69, 2826–2832. doi: 10.1158/0008-5472. CAN-08-4466

Tian, T., Wang, Z., and Zhang, J. (2017). Pathomechanisms of oxidative stress in inflammatory bowel disease and potential antioxidant therapies. *Oxidative Med. Cell. Longev.* 2017:4535194. doi: 10.1155/2017/4535194

Vieira, A. T., Macia, L., Galvão, I., Martins, F. S., Canesso, M. C., Amaral, F. A., et al. (2015). A role for gut microbiota and the metabolite-sensing receptor GPR43 in a murine model of gout. *Arthritis Rheumatol.* 67, 1646–1656. doi: 10.1002/art.39107

Viladomiu, M., Metz, M. L., Lima, S. F., Jin, W. B., Chou, L., Guo, C. J., et al. (2021). Adherent-invasive *E. coli* metabolism of propanediol in Crohn's disease regulates phagocytes to drive intestinal inflammation. *Cell Host Microbe* 29, 607–19.e8. doi: 10.1016/j.chom.2021.01.002

Wang, Q., McLoughlin, R. M., Cobb, B. A., Charrel-Dennis, M., Zaleski, K. J., Golenbock, D., et al. (2006). A bacterial carbohydrate links innate and adaptive responses through toll-like receptor 2. *J. Exp. Med.* 203, 2853–2863. doi: 10.1084/jem.20062008

Wang, Y. N., Yu, C. Y., and Jin, H. Z. (2020). RNA N(6)-Methyladenosine modifications and the immune response. J Immunol Res 2020:6327614. doi: 10.1155/2020/6327614

Wei, S., Zhang, J., Wu, X., Chen, M., Huang, H., Zeng, S., et al. (2022). Fusobacterium nucleatum extracellular vesicles promote experimental colitis by modulating autophagy via the miR-574-5p/CARD3 Axis. *Inflamm. Bowel Dis.* 23:izac177. doi: 10.1093/ibd/izac177

Xiong, X., Tian, S., Yang, P., Lebreton, F., Bao, H., Sheng, K., et al. (2022). Emerging enterococcus pore-forming toxins with MHC/HLA-I as receptors. *Cells* 185, 1157–1171.e22. doi: 10.1016/j.cell.2022.02.002

Xu, W., Guo, Y., Huang, Z., Zhao, H., Zhou, M., Huang, Y., et al. (2019). Small heat shock protein CRYAB inhibits intestinal mucosal inflammatory responses and protects barrier integrity through suppressing IKKbeta activity. *Mucosal. Immunol.* 12, 1291–1303. doi: 10.1038/s41385-019-0198-5

Xu, M., Pokrovskii, M., Ding, Y., Yi, R., Au, C., Harrison, O. J., et al. (2018). C-MAF-dependent regulatory T cells mediate immunological tolerance to a gut pathobiont. *Nature* 554, 373–377. doi: 10.1038/nature25500

Yang, X., Mao, Z., Huang, Y., Yan, H., Yan, Q., Hong, J., et al. (2021). Reductively modified albumin attenuates DSS-induced mouse colitis through rebalancing systemic redox state. *Redox Biol.* 41:101881. doi: 10.1016/j.redox.2021.101881

Yang, J., Pei, G., Sun, X., Xiao, Y., Miao, C., Zhou, L., et al. (2022). RhoB affects colitis through modulating cell signaling and intestinal microbiome. *Microbiome*. 10:149. doi: 10.1186/s40168-022-01347-3

Zelante, T., Iannitti, R. G., Cunha, C., De Luca, A., Giovannini, G., Pieraccini, G., et al. (2013). Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22. *Immunity* 39, 372–385. doi: 10.1016/j.immuni.2013.08.003

Zhai, R., Xue, X., Zhang, L., Yang, X., Zhao, L., and Zhang, C. (2019). Strain-specific anti-inflammatory properties of two Akkermansia muciniphila strains on chronic colitis in mice. *Front. Cell. Infect. Microbiol.* 9:239. doi: 10.3389/fcimb.2019.00239

Zhang, J., Cen, L., Zhang, X., Tang, C., Chen, Y., Zhang, Y., et al. (2022). MPST deficiency promotes intestinal epithelial cell apoptosis and aggravates inflammatory bowel disease via AKT. *Redox Biol.* 56:102469. doi: 10.1016/j.redox.2022.102469

Zhang, T., Ding, C., Chen, H., Zhao, J., Chen, Z., Chen, B., et al. (2022). M(6)a mRNA modification maintains colonic epithelial cell homeostasis via NF-kappaB-mediated antiapoptotic pathway. *Sci. Adv.* 8:eabl5723. doi: 10.1126/sciadv.abl5723

Zhang, L., Lee, H., Grimm, M. C., Riordan, S. M., Day, A. S., and Lemberg, D. A. (2014). Campylobacter concisus and inflammatory bowel disease. *World J. Gastroenterol.* 20, 1259–1267. doi: 10.3748/wjg.v20.i5.1259

Zhang, C., Qin, J., Zhang, S., Zhang, N., Tan, B., Siwko, S., et al. (2020). ADP/P2Y1 aggravates inflammatory bowel disease through ERK5-mediated NLRP3 inflammasome activation. *Mucosal Immunol.* 13, 931–945. doi: 10.1038/s41385-020-0307-5

Zhang, J., Wang, C., Guo, Z., Da, B., Zhu, W., and Li, Q. (2021). miR-223 improves intestinal inflammation through inhibiting the IL-6/STAT3 signaling pathway in dextran sodium sulfate-induced experimental colitis. *Immun. Inflamm Dis.* 9, 319–327. doi: 10.1002/iid3.395

Zhang, Y. S., Xin, D. E., Wang, Z., Song, X., Sun, Y., Zou, Q. C., et al. (2019). STAT4 activation by leukemia inhibitory factor confers a therapeutic effect on intestinal inflammation. *EMBO J.* 38, 1–20. doi: 10.15252/embj.201899595

Zhen, Y., and Zhang, H. (2019). NLRP3 Inflammasome and inflammatory bowel disease. Front. Immunol. 10:276. doi: 10.3389/fimmu.2019.00276

Zuo, L., Kuo, W. T., Cao, F., Chanez-Paredes, S. D., Zeve, D., Mannam, P., et al. (2022). Tacrolimus-binding protein FKBP8 directs myosin light chain kinasedependent barrier regulation and is a potential therapeutic target in Crohn's disease. *Gut*:gutjnl-2021-326534. doi: 10.1136/gutjnl-2021-326534