



# Lactic Acid Bacteria in Wine: Technological Advances and Evaluation of Their Functional Role

Carla Virdis<sup>1</sup>, Krista Sumbly<sup>1,2</sup>, Eveline Bartowsky<sup>2,3</sup> and Vladimir Jiranek<sup>1,2\*</sup>

<sup>1</sup> Department of Wine Science, University of Adelaide, Urrbrae, SA, Australia, <sup>2</sup> Australian Research Council Training Centre for Innovative Wine Production, Urrbrae, SA, Australia, <sup>3</sup> Lallemand Australia, Edwardstown, SA, Australia

## OPEN ACCESS

### Edited by:

Cristina Reguant,  
University of Rovira i Virgili, Spain

### Reviewed by:

Vittorio Capozzi,  
Italian National Research Council, Italy  
Giuseppe Spano,  
University of Foggia, Italy

### \*Correspondence:

Vladimir Jiranek  
vladimir.jiranek@adelaide.edu.au

### Specialty section:

This article was submitted to  
Food Microbiology,  
a section of the journal  
Frontiers in Microbiology

**Received:** 30 September 2020

**Accepted:** 23 November 2020

**Published:** 15 January 2021

### Citation:

Virdis C, Sumbly K, Bartowsky E  
and Jiranek V (2021) Lactic Acid  
Bacteria in Wine: Technological  
Advances and Evaluation of Their  
Functional Role.  
Front. Microbiol. 11:612118.  
doi: 10.3389/fmicb.2020.612118

Currently, the main role of Lactic Acid Bacteria (LAB) in wine is to conduct the malolactic fermentation (MLF). This process can increase wine aroma and mouthfeel, improve microbial stability and reduce the acidity of wine. A growing number of studies support the appreciation that LAB can also significantly, positively and negatively, contribute to the sensorial profile of wine through many different enzymatic pathways. This is achieved either through the synthesis of compounds such as diacetyl and esters or by liberating bound aroma compounds such as glycoside-bound primary aromas and volatile thiols which are odorless in their bound form. LAB can also liberate hydroxycinnamic acids from their tartaric esters and have the potential to break down anthocyanin glucosides, thus impacting wine color. LAB can also produce enzymes with the potential to help in the winemaking process and contribute to stabilizing the final product. For example, LAB exhibit peptidolytic and proteolytic activity that could break down the proteins causing wine haze, potentially reducing the need for bentonite addition. Other potential contributions include pectinolytic activity, which could aid juice clarification and the ability to break down acetaldehyde, even when bound to SO<sub>2</sub>, reducing the need for SO<sub>2</sub> additions during winemaking. Considering all these findings, this review summarizes the novel enzymatic activities of LAB that positively or negatively affect the quality of wine. Inoculation strategies, LAB improvement strategies, their potential to be used as targeted additions, and technological advances involving their use in wine are highlighted along with suggestions for future research.

**Keywords:** MLF, LAB, wine aroma, quality, enzymes, management

## INTRODUCTION

The fruit of the grapevine, *Vitis vinifera*, was first transformed into wine sometime between 8500BC and 4000BC (Varriano, 2010). However, wine, as we know it nowadays, is a complex beverage in which many key elements shape the final product. These key elements include the quality of the grapes and their varietal and clonal genotype, the yeasts and bacteria conducting the alcoholic fermentation (AF) and the malolactic fermentation (MLF), respectively, the aging vessels and the winemaking techniques (Styger et al., 2011).

The main role of lactic acid bacteria (LAB) in wine has traditionally been to perform the conversion of malic acid to lactic acid. In the last decades, various papers have shown that LAB metabolism also involves a large array of secondary enzymatic activities capable of generating many

volatile secondary compounds (Matthews et al., 2004; Sumbly et al., 2010; Bartowsky and Borneman, 2011; Michlmayr et al., 2012; Cappello et al., 2017; Takase et al., 2018).

Although it is evident that wines originating from a specific grape variety display particular characters that distinguish them from other varieties, in many cases these active-flavor compounds are not detectable at pre-fermentative stages (Swiegers et al., 2005). Often, they are instead the product of the metabolism of yeast and bacteria and are modified and released in wine during the fermentation processes. Enzymes from LAB that can exert their activity in wine include glycosidases, esterases, proteases and others (Liu, 2002). The activity of these enzymes can significantly add to the appearance, flavor, texture and aroma of wine, ultimately, helping to define its structure (Swiegers et al., 2005).

In this review we examine the current literature with regards to the functional role of LAB in winemaking (**Figure 1**) including; inoculation strategies, modification of wine aroma, impact on wine color, potential novel uses to aid winemaking processes, and their overall effect on the wholesomeness of wine. Finally, we discuss current limitations and future prospects.

## THE ROLE OF LACTIC ACID BACTERIA IN WINEMAKING

### The Malolactic Fermentation

Once the primary fermentation has finished, or simultaneously in some cases, most red, some white and sparkling wines undergo a secondary microbial fermentation. During MLF, LAB convert the dicarboxylic malic acid into the monocarboxylic lactic acid and carbon dioxide (**Figure 1**) with no free intermediary products (Ribéreau-Gayon et al., 2006a). MLF typically achieves a more palatable wine by reducing the tart taste of malic acid. Additionally, MLF reduces the amount of residual nutrients able to support microbial growth making the wine more stable prior to being bottled (Lonvaud-Funel, 1999).

Several factors affect the performance of LAB in wine, their growth, their ability to complete MLF and their metabolic properties (Sumbly et al., 2019). Some known limiting factors are high ethanol content, low pH, sub-optimal temperatures and high SO<sub>2</sub>, however, their inhibiting effect can be potentiated by their synergistic action (Lerm et al., 2010).

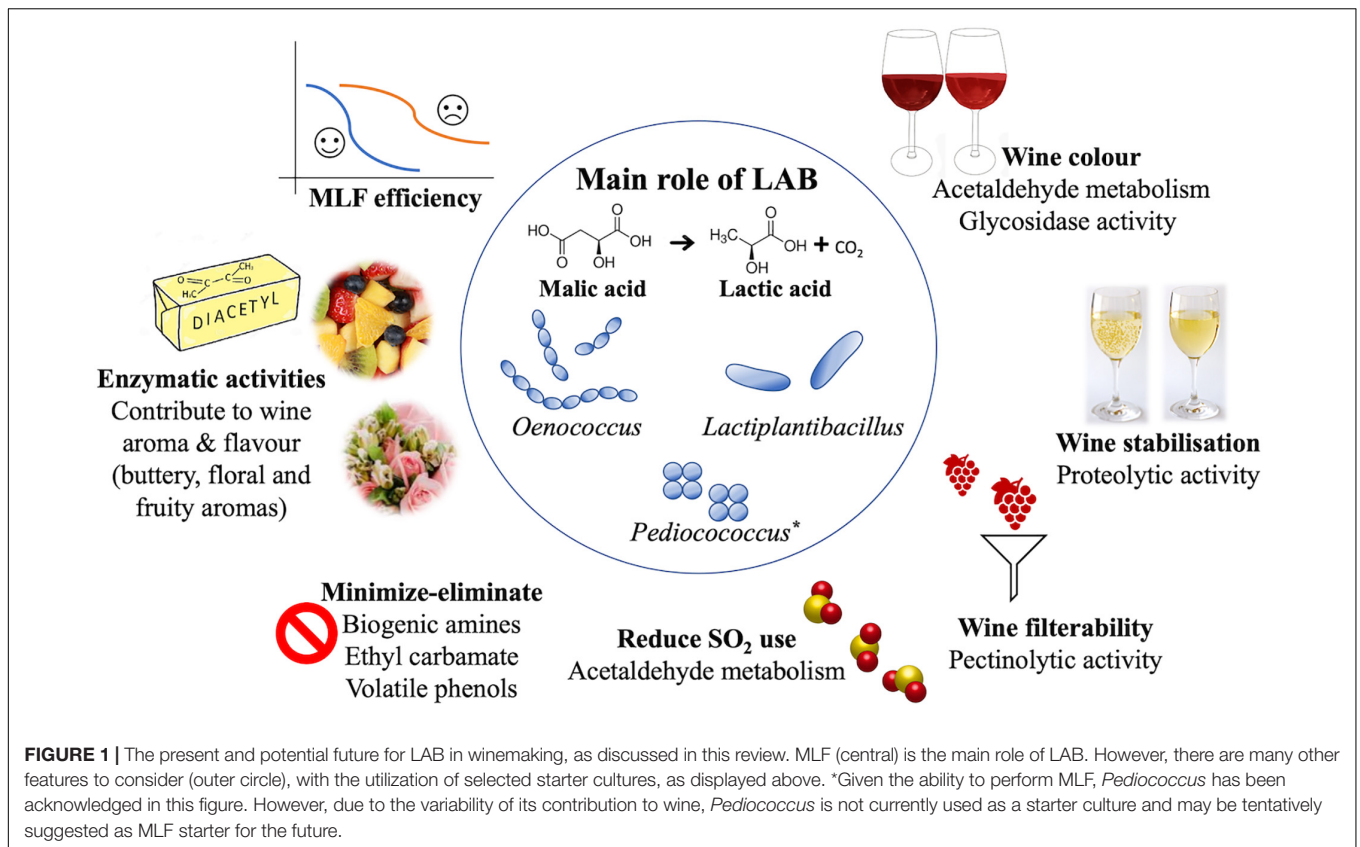
### Inoculation Strategies

Malolactic fermentation can be facilitated by autochthonous LAB or can be induced by inoculating the wine with selected bacterial starters (**Table 1**). Before the use of MLF starters became a common winemaking practice, a secondary fermentation in wine was often enabled by the microbial populations that originated in the vineyard or winery and were brought to the wine via the grape skin or the winery vessels (Bokulich et al., 2016). Although not common, uninoculated MLF performed by autochthonous LAB is still used in winemaking (**Table 1**), mainly with the aim of producing wines with a regional character or a particular style (Bokulich et al., 2016). However, when MLF is uninoculated and carried out by autochthonous bacteria, the conversion

of malic acid into lactic acid can be slow or incomplete, or undesired volatile compounds and potentially hazardous compounds can be produced (Lonvaud-Funel, 1999). The use of bacterial starters can help minimize these risks (Lonvaud-Funel, 1999). Considerable research has been dedicated to identifying robust starters with desirable metabolic activity (Lerm et al., 2011). More recently, efforts have been directed to exploring the biodiversity of wine associated geographic areas, with the aim of finding new LAB starters with a high degree of adaptation to the specific niches (Miranda-Castilleja et al., 2016; López-Seijas et al., 2020). For example, two potential new autochthonous MLF starters with interesting  $\beta$ -glucosidase activity, *Lactocaseibacillus paracasei* (formerly known as *Lactobacillus paracasei*) UVI-2 and *Lentilactobacillus hilgardii* (formerly known as *L. hilgardii*) UVI-23, have been identified from Albariño grapes in Val do Salnés, Spain (López-Seijas et al., 2020). In another study, the isolation of numerous *Lactiplantibacillus plantarum* (formerly known as *Lactobacillus plantarum*) highlighted a great intra-species genetic variability in North Patagonia, Argentina, and suggested also their potential use as MLF starter cultures for regional wines (Bravo-Ferrada et al., 2013). This is especially interesting considering that the regional identity of wines, or terroir, can be an important factor in increasing the value of the final product (Bartowsky et al., 2015).

When selected bacterial starters are used (**Table 1**), the winemaker can opt for a sequential inoculation with yeast, where *Oenococcus oeni* is inoculated at the end of AF, or for a coinoculation strategy, where LAB starters are inoculated simultaneously or shortly (24–48 h) after the beginning of AF (Lucio et al., 2017; Bartle et al., 2019; Sumbly et al., 2019). The main advantages of the coinoculation technique, compared to the more traditional sequential inoculation, are the potentially positive contribution to wine composition and the reduction of microbial spoilage risk (Sumbly et al., 2019). Reducing the overall AF and MLF time allows the winemaker to protect the wine with SO<sub>2</sub> additions earlier in the winemaking process, thus avoiding the production of volatile phenols from spoilage microorganisms such as *Brettanomyces bruxellensis* (Sumbly et al., 2019). During coinoculation, the interactions between yeasts and bacteria may affect the efficiency of MLF and the sensory properties of the final wine (Bartle et al., 2019; Du Plessis et al., 2019). For more information regarding yeast and bacteria interactions, refer to the recent review from Bartle et al. (2019). Lastly, bacteria can be inoculated before yeast. Although not common this technique has proven to be very efficient in promoting MLF, especially when *Liquorilactobacillus mali* (formerly known as *Lactobacillus mali*), *L. paracasei*, *Liquorilactobacillus satsumensis* (formerly known as *Lactobacillus satsumensis*) and *L. plantarum* strains were used as starters (Lucio et al., 2017). Amongst the above mentioned genera, the best results in terms of growth in juice and malic consumption efficiency were obtained with *L. plantarum* (Lucio et al., 2017).

In recent years, mixed inoculation strategies have also been trialed (**Table 1**). The use of blended cultures of *L. plantarum* and *O. oeni* as MLF starters, can facilitate a rapid consumption of malic acid, whilst contributing significantly to the volatile profile of wine (Brizuela et al., 2018). Preparations comprising mixtures



**TABLE 1** | Overview of the main LAB of enological interest, and their main roles in wine; past, present, and future.

	Strains used	When	Major roles	Other roles	References
Past	• Autochthonous LAB	Not controllable	Softer and more aromatic wine		Bokulich et al., 2016
Present	• Single bacterial starter cultures: <i>O. oeni</i> <i>L. plantarum</i>	During and after AF	Softer mouthfeel, Higher microbiological stability Lower acidity	Production of diacetyl, esters, aromatic thiols and other aromatic compounds, Liberation of glycoside-bound aromatic compounds	Bartowsky and Borneman, 2011; Krieger-Weber et al., 2020
	• Mixed bacterial starter cultures				
	• Autochthonous LAB	Not controllable	Softer and more aromatic wine	Development of regional character	Bokulich et al., 2016
Future	• Single bacterial starter cultures: <i>O. oeni</i> <i>L. plantarum</i> <i>Pediococcus spp.</i> Other LAB?	Before, during and after AF	Softer mouthfeel, Higher microbiological stability, Lower acidity, Production of diacetyl, esters, aromatic thiols and other aromatic compounds, Liberation of glycoside-bound aromatic compounds	Reduction of Acetaldehyde content in wine, Reduction of biogenic amine content in wine, Protein stability, Juice clarification, Reduction of SO <sub>2</sub> required in winemaking, More properties?	Wade et al., 2018; Iorizzo et al., 2016; Wells and Osborne, 2011; López-Seijas et al., 2020
	• Mixed bacterial starter cultures • Autochthonous, LAB starters				

of *L. plantarum* and *O. oeni*, recommended for coinoculation use, are commercially available (for example Anchor Oenology).

The timing of inoculation leads to different aroma compounds being released in wine, qualitatively and quantitatively modifying wine profiles (Lasik-Kurdyś et al., 2018). However, independently from the inoculation time, the key of the success of MLF seems to be the correct management of the inoculation technique

(Lombardi et al., 2020). For example, pre-adapting the bacterial starters to sub-optimal pH (5.0) can improve the consumption of malic acid (Lombardi et al., 2020).

## Lactic Acid Bacteria Used in Winemaking

Lactic Acid Bacteria are Gram-positive bacteria, grouped in the phylum Firmicutes, class Bacilli, order Lactobacillales (Holzapfel

et al., 2014). Several genera of the family *Lactobacillaceae* are used in the food industry and are involved in the production of numerous fermented foods, such as yogurt, cheese and sauerkraut (Szutowaska, 2020). *Oenococcus*, *Leuconostoc*, and *Pediococcus*, within the formerly known Leuconostocaceae family, and the formerly known genus *Lactobacillus*, within the formerly known Lactobacillaceae family, are the only genera associated to wine (Holzapfel et al., 2014; Zheng et al., 2020).

Recently the description of LAB has been amended, following a modern multifactorial approach used to re-evaluate the taxonomy of these microorganisms (Zheng et al., 2020). Core genome phylogeny, physiological and metabolic criteria, and the ecology of the organisms are some of the parameters used for this evaluation (Zheng et al., 2020). The formerly known genus *Lactobacillus* has been restructured into 25 new genera and the former *Lactobacillaceae* and *Leuconostocaceae* families have been fused into a new larger *Lactobacillaceae* family (Zheng et al., 2020).

Lactic acid bacteria can be both detrimental and beneficial to the quality of wine (Bartowsky, 2009). Their performance in wine is related to the specific species and strain genetics but also to many other factors including environmental conditions and microbial interactions (Cappello et al., 2017; Devi and Ka, 2019). At present, *O. oeni* is one of the three, and the most known, species in the *Oenococcus* genus (Lorentzen and Lucas, 2019). Due to its high tolerance for low pH, high ethanol concentrations and scarcity of nutrients, *O. oeni* is the main LAB of choice in winemaking (Bartowsky, 2005). However, with increasing temperatures during growth and harvest, and a consequent rising pH trend for many wines, other LAB have the potential to become a valid alternative to *Oenococcus*, playing an important role in the modifications of wine aroma (Du Toit et al., 2010; Mira and de Orduña, 2010; Berbegal et al., 2019; Krieger-Weber et al., 2020; López-Seijas et al., 2020; Shao-Yang et al., 2020; Sun et al., 2020). Above all, *Lactiplantibacillus* strains, with their fast consumption of malic acid (up to 3 g/L in 2–4 days) and the suppression of the activity of other spontaneous LAB populations, are an ideal starter choice for the winemaker (Du Toit et al., 2010; Krieger-Weber et al., 2020). Furthermore, as *L. plantarum* is homofermentative for hexoses it does not produce volatile acidity (VA) through sugar metabolism. Currently, only a few freeze-dried starter cultures of *L. plantarum* are commercially available and their use is especially recommended for coinoculation strategies in wines with high pH (>3.4) and a high risk of autochthonous LAB contamination<sup>1,2</sup>.

Lastly, the genus *Pediococcus* is generally considered a spoilage microorganism in wine (Wade et al., 2018). *P. damnosus*, *P. inopinatus*, *P. parvulus*, and *P. pentosaceus* have been reported to produce excessive diacetyl, exopolysaccharides, biogenic amines, acrolein and more generally off-odors, flavors and textures, thus contributing detrimentally to wine quality (Wade et al., 2018). However, recent findings have shown that the presence of *Pediococcus* species in wine does not always lead to spoilage and that some species and strains

within this genus may contribute positively to wine aroma and can inhibit the formation of 4-ethylphenol from the spoilage yeast, *B. bruxellensis* (Strickland et al., 2016; Wade et al., 2018). Given the variability of the contribution to wine from *Pediococcus* strains, further studies on this genus are of crucial importance. Understanding the differences between strains, and the interactions of this genus with other microorganisms, could open up the possibility of using selected *Pediococcus* starter cultures, potentially reconsidering its role in winemaking.

## HOW CAN LAB IMPROVE WINE AROMA?

### Citrate Metabolism

Oenologically, one of the major aroma compounds associated with LAB is diacetyl, which originates from citrate fermentation (Bartowsky and Borneman, 2011). At low concentrations (1–4 mg/L) diacetyl confers the typical buttery character to wine, while at high concentrations (>5–7 g/L) it is associated with undesirable aromas (Bartowsky and Henschke, 2004). A 2002 survey showed that the concentration of diacetyl in wine varies widely, ranging from 0.3 to 0.6 mg/L in Chardonnay wines and from 0.3 to 2.5 mg/L in red wines (Bartowsky et al., 2002). The sensory threshold can vary greatly across different types of wines and is greatly affected by the presence in wine of other compounds such as sulfur dioxide (Bartowsky et al., 2002).

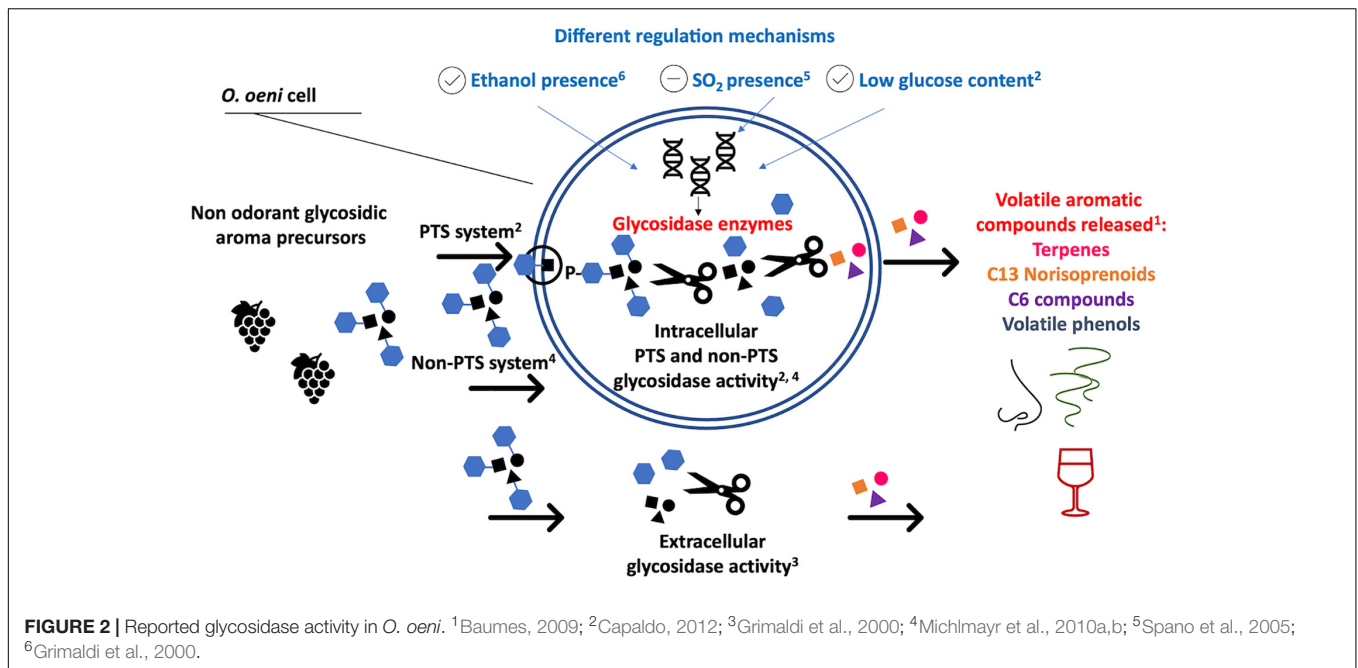
Referring to their ability to degrade citrate, LAB can be classified in cit<sup>+</sup> and cit<sup>-</sup>. Cit<sup>+</sup> strains have all the genes that encode for the necessary permease and the lyase subunits needed to degrade citrate, leading to the production of pyruvate. Conversely, cit<sup>-</sup> strains may lack one or more genes in this metabolic pathway and cannot degrade citrate, however, they can still produce diacetyl from the pyruvate that originates in the glycolysis pathway (Lerm et al., 2011; Mink et al., 2015). Cit<sup>+</sup> LAB strains typically produce more D-lactate, acetate, diacetyl and acetoin from pyruvate (Pretorius et al., 2019). The content of diacetyl found in wine is therefore dependent on the LAB species used as a starter for MLF (Lonvaud-Funel, 1999; Bartowsky and Henschke, 2004).

### Glycosidase Activity

Odorless, sugar-bound aromas represent a reservoir of wine aroma that can be released by cleaving the bonds between the glycosidic molecule and the volatile, aromatic aglycone (**Figure 2**), such as terpenes, C13 norisoprenoids, volatile phenols, C6 compounds and others (Baumes, 2009). A group of enzymes called glycosidases are responsible for the enzymatic hydrolysis of these compounds. The type of aroma precursor determines which specific glycosidase is needed to break the bonds. The aromatic volatiles can be conjugated to glucose ( $\beta$ -D-glucopyranoside) or to a disaccharide. In the latter case, the inner molecule is a glucose unit and the outer one is a second sugar unit (e.g.,  $\alpha$ -L-arabinofuranose,  $\alpha$ -L-rhamnopyranose,  $\beta$ -D-xylopyranose, or  $\beta$ -D-apiofuranose) (Williams et al., 1982; Gunata et al., 1988). When the aromatic compound is linked

<sup>1</sup>www.lallemandwine.com

<sup>2</sup>www.chr-hansen.com



to a glucose, a  $\beta$ -glucosidase is required to hydrolyze the bond. When the aromatic compound is linked to a disaccharide two enzymes are needed to break the bond: an exo-glycosidase e.g.,  $\alpha$ -L-arabinofuranosidase,  $\alpha$ -L-rhamnopyranosidase,  $\beta$ -D-xylopyranosidase, or  $\beta$ -D-apiofuranosidase, cleaves the second sugar unit and subsequently a  $\beta$ -glucosidase removes the remaining glucose (Gunata et al., 1988).

Glycosidases can originate from different sources, including grapes (Aryan et al., 1987), yeasts (Swiegers et al., 2005) and bacteria (Bartowsky and Borneman, 2011; Capaldo, 2012; Cappello et al., 2017). The addition of enzymatic preparations with hydrolytic activity to musts and wines can increase the content of free aromatic compounds such as terpenes and norisoprenoids (Pretorius and Rensburg, 2000; Codresi et al., 2012). However, purified enzymes can be expensive and may have low specificity and efficiency (Tavernini et al., 2020). To overcome these issues, polymeric chitosan beads have been successfully trialed as matrix for the immobilization of  $\beta$ -D-glucosidases and  $\alpha$ -L-arabinofuranosidases sourced from a commercial, *Aspergillus niger*-derived preparation and previously organized in a cross-linked matrix (Tavernini et al., 2020). There is also a high potential for the use of glycosidases from bacteria, especially LAB, because these enzymes could perform better under stressful conditions such as high concentrations of alcohol and acidity (Bartowsky and Borneman, 2011).

Several authors have shown that LAB are able to enzymatically hydrolyze glycosides and that this activity is widespread amongst different *O. oeni* strains (Grimaldi et al., 2000, 2005a; Barbagallo et al., 2004; D'Incecco et al., 2004; Spano et al., 2005; Hernandez-Orte et al., 2009; Fia et al., 2018) and other LAB genera (Grimaldi et al., 2005b; Spano et al., 2005; Hernandez-Orte et al., 2009). *Lactiplantibacillus* strains show a varied array of enzymes that can contribute to wine aroma, and particularly, a great ability to

release volatile aromas from glycosidic precursors (Lerm et al., 2011; Iorizzo et al., 2016).

Although considerable efforts have been directed toward understanding the genetics and the expression mechanisms, the glycosidase activity of LAB is not yet clear. The carbohydrate phosphotransferase system (PTS) seems to play a key role in *O. oeni*, by phosphorylating and transporting the glycosides into the cell (Figure 2) through a membrane-associated enzymatic system (Capaldo, 2012). The activated glycosides can then be hydrolyzed by intracellular phosphoglycosidases (Capaldo, 2012). Some phosphoglycosidases have been characterized and their hydrolytic activity against *p*-nitrophenol- $\beta$ -D-glucopyranoside-6-phosphate confirmed (Capaldo, 2012). Intracellular non-PTS glycosidase activity has been reported by a number of authors (Figure 2) and some of these glycosidases have also been characterized (Michlmayr et al., 2010a,b; Capaldo, 2012). However, other authors reported extra-cellular and parietal glycosidase activity, suggesting that the glycosidase activity in *O. oeni* is the consequence of a complex enzymatic system, involving many different genes, enzymes and regulation mechanisms (Grimaldi et al., 2000; Barbagallo et al., 2004). Little is known regarding the molecular mechanisms involved in the glycosidase activity of other LAB, such as *Lactiplantibacillus* and *Pediococcus*. It is known that abiotic stresses have a key role in upregulating the expression of these enzymes, although data are controversial. Low levels of glucose and the presence of ethanol seem to lead to an enhancement of the glycosidase activity, while low pH values and sulfite content seem to repress the activity (D'Incecco et al., 2004; Grimaldi et al., 2005a,b; Spano et al., 2005; Capaldo, 2012). See Figure 2 for an overview of the glycosidase activity of LAB in wine.

Finally, it is important to note that glycosidase activities in wine can also have a negative effect on the final product. Similar

to yeast, the bacterial starters, with their ability to hydrolyse the glycoconjugate compounds, can also heavily affect the evolution of volatile phenols associated with smoke taint, influencing the intensity of smoke taint-associated aromas (Kennison et al., 2008; Jiranek, 2011). The aroma compounds associated with the smoky and earthy notes that characterize smoke tainted wines, such as guaiacol, 4-methylguaiacol, 4-ethylguaiacol and 4-ethylphenol, typically peak in finished wine (388  $\mu\text{g/L}$  for guaiacol and 93  $\mu\text{g/L}$  for methylguaiacol) (Kennison et al., 2007). This can be explained by the presence of conjugated precursor compounds, particularly disaccharide conjugates, which can be liberated via enzymatic hydrolysis (Kennison et al., 2008; Hayasaka et al., 2010; Dungey et al., 2011). For the impact that LAB glycosidase activity can have on wine color see the dedicated paragraph: “how can LAB impact the wine colour?”.

### Release of Volatile Thiols From Precursor Compounds

Volatile thiols, such as 3-sulfanylohexan-1-ol and 3-sulfanylohexyl acetate, first identified in Sauvignon Blanc, are important aromatic compounds that contribute varietal aromas like grapefruit and passionfruit (Takase et al., 2018). Their non-odorant precursors, such as glutathione S-conjugate, cysteinyl-glycine S-conjugate and cysteine S-conjugate, are found in grapes and are enzymatically released by yeast during AF (Thibon et al., 2008). A 2019 study has shown that *L. plantarum* is able to enzymatically convert the cysteine S-conjugates and cysteinyl-glycine S-conjugates, with a noticeable preference for the latter compound under experimental conditions (Takase et al., 2018). The enzymes responsible for this transformation and the underlying regulatory mechanisms have not yet been characterized. However, this represents an opportunity to improve the varietal aroma of wines, and further research is warranted to provide a better understanding of this metabolic pathway.

### Esterase Activity

Esters are secondary or tertiary aroma compounds that contribute significantly to wine aroma. In wine, esters can either be formed in a process called esterification, or broken down via ester hydrolysis (Waterhouse et al., 2016). They can be formed during the primary or secondary fermentation by yeast or bacteria, and their concentration and composition is slowly changed during wine aging (Liu, 2002; Sumbly et al., 2010, 2013; Antalick et al., 2012). The contribution of LAB to the ester composition of wine has been highlighted with several wine volatile profiling studies conducted after MLF (Ugliano and Moio, 2005; Antalick et al., 2012; Costello et al., 2012; Sumbly et al., 2012; Lytra et al., 2020). In wine, LAB can both increase and decrease the content of esters (Sumbly et al., 2013). The degree of LAB contribution to the ester profile of wine is strain-specific (Sumbly et al., 2013; Gammacurta et al., 2018) and MLF inoculation strategy can affect the quantity and quality of esters released by the bacteria (Lasik-Kurdyś et al., 2018). It seems that the use of a coinoculation technique increases the release of ethyl esters, particularly ethyl lactate, diethyl succinate and ethyl

acetate, thus (depending on their concentration) enriching the wine with floral and fruity notes (Lasik-Kurdyś et al., 2018).

### Activities That Can Affect the Content of Volatile Phenols in Wine

Ethyl phenols are crucial aromatic compounds associated with unpleasant odors in wine such as horse sweat, leather and stable. Ethyl phenols are produced by some yeasts within the genus *Brettanomyces/Dekkera* from hydroxycinnamic acids (HCAs) precursors (Chatonnet et al., 1992). *Brettanomyces* yeasts can decarboxylase the free HCAs, converting them into vinylphenols first, via an hydroxycinnamate decarboxylase enzyme (HCDC), and then into the unpleasant ethyl phenol, via a vinylphenol reductase enzyme (VPhR) (Chatonnet et al., 1992). HCAs can be found in the must as free acids but, most commonly, as their tartrate esters, which are hydrolyzed slowly throughout the winemaking process (Waterhouse et al., 2016; Lima et al., 2018). This leads to small amounts of ethyl phenol precursors being continuously released in wine and made available to *Brettanomyces* metabolism.

It has been reported that the use of *O. oeni* strains with cinnamoyl esterase activity can lead to higher amounts of 4-ethylphenol and 4-ethylguaiacol in Pinot Noir after MLF, compared to wine that did not undergo MLF, or underwent MLF with bacterial starters with no cinnamoyl esterase activity (Chescheir et al., 2015). Cinnamoyl esterase activity is strain-specific in *O. oeni* and may be constitutively expressed (Chescheir et al., 2015; Collombel et al., 2019). Thus, to avoid the faults that arise from *Brettanomyces* spoilage, it can be beneficial to use bacterial starters with low cinnamoyl esterase activity (Chescheir et al., 2015).

LAB have also been reported to be directly responsible for the production of 4-vinylphenol (Silva et al., 2011). In *L. plantarum*, *Secundilactobacillus collinoides* (formerly known as *Lactobacillus collinoides*) and *Pediococcus pentosaceus*, this activity is enhanced by the presence of hydroxycinnamic acids, especially caffeic acid, in the growth medium (Silva et al., 2011). Conversely, a tannin content of 1 g/L (the average range for red wines is 1–4 g/L) can inhibit the release of volatile phenols in wine by *L. plantarum* (Silva et al., 2011). However, the literature regarding this matter is still controversial. It has also been suggested that the addition to the must of cinnamoyl esterase enzymes can help reduce the formation of ethyl phenols (Morata et al., 2013). The intermediate vinylphenols are also able to react with anthocyanins, producing very stable vinylphenolic pyranoanthocyanins (Rentzsch et al., 2007). When vinylphenols are present in their bound form, they can help reduce the number of precursors for the formation of ethyl phenols and also stabilize the color of wine (Morata et al., 2013). In support of this, wines fermented with *Saccharomyces* yeast with an increased hydroxycinnamate decarboxylase activity (HCDC+) have an increased content of vinylphenolic pyranoanthocyanins and a reduced content of ethyl phenols (Morata et al., 2013). The addition to the must of cinnamoyl esterase enzymes, to help release more quickly the free HCAs paired with the use of HCDC+ *Saccharomyces cerevisiae* strains, could be a way

to reduce the formation of ethylphenols (Morata et al., 2013). Similarly, the use of LAB with enhanced cinnamoyl esterase activity could be beneficial for the quality of wine, helping to prevent the formation of unwanted off flavors and favoring the development of stable color compounds.

Furthermore, the levels of volatile phenols in the finished wine seem to depend more on differences between *Brettanomyces* strains rather than on the cinnamoyl esterase activity of the LAB employed for MLF (Madsen et al., 2017). Two different *O. oeni* strains, with and without cinnamoyl esterase activity, were tested on Cabernet Sauvignon and, although there were differences in the degradation rate of tartaric esters of HCAs into free HCAs, this did not affect the final content of volatile phenols (Madsen et al., 2017).

Further studies are needed to characterize the cinnamoyl esterase activity in LAB. Understanding the basis for the differential cinnamoyl esterase activity of LAB could enable the winemaker to make appropriate decisions in regards the choice of bacteria to employ.

### Acetaldehyde Metabolism

Acetaldehyde is an important compound in wine, affecting aroma, color, stability and microbiological properties (Liu and Pilone, 2000). Wine typically contains 20–100 mg/L of acetaldehyde, and, at low concentrations, it can enhance the fruity character of wine (Waterhouse et al., 2016). However, at higher concentrations it is associated with an unpleasant rotten apple aroma (Waterhouse et al., 2016). Acetaldehyde is an intermediate compound produced by yeast during AF and also plays a vital role in the stabilization of wine color (see next section “how can LAB impact the wine color?”) (Waterhouse et al., 2016; Forino et al., 2020). During AF acetaldehyde is readily reduced to ethanol, thus at this stage, its content in wine is typically low (25–40 mg/L) depending on yeast strain and the concentration of SO<sub>2</sub> in the must (Waterhouse et al., 2016). However, in later stages, the oxidation of alcohol can lead to an increase in acetaldehyde content (Waterhouse et al., 2016).

Excessive accumulation of acetaldehyde is not desirable because it strongly binds to sulfur dioxide making it less active (see section “reducing the SO<sub>2</sub> required in winemaking”) and requiring the winemaker to add additional SO<sub>2</sub> to protect the wine (Wells and Osborne, 2011). Additionally, as acetaldehyde has potential toxic and carcinogenic effects, high amounts are undesirable in beverages for human consumption (Lachenmeier and Sohnius, 2008).

Acetaldehyde can exert a stimulating or inhibiting effect on LAB growth in wine (Liu and Pilone, 2000). Both heterofermentative and homofermentative LAB strains can degrade free and SO<sub>2</sub>-bound acetaldehyde into small amounts of ethanol and acetic acid (Osborne et al., 2000). However, this could also lead to higher VA in wine (Waterhouse et al., 2016). The efficiency of this activity seems to be strain-specific (Mira and de Orduña, 2010). The ability of probiotics, including LAB, to break down acetaldehyde, is therefore of interest to the wine industry and could be used to reduce acetaldehyde levels in wine. Indeed, a patent describing “a composition of probiotics including LAB, able to degrade alcohol and acetaldehyde,” to

help prevent and treat alcohol-related diseases, has been filed (Chung, 2018).

### HOW CAN LAB IMPACT THE WINE COLOR?

Wine color is an important sensory attribute (**Figure 1**) that is relevant to red wines, largely related to the grape variety and the vintage, and to a minor extent to the winemaking practices (González-Neves et al., 2013). Wine color can also be affected by the activity of yeast and LAB (Morata et al., 2005; Benito et al., 2011; Burns and Osborne, 2013, 2015; Devi and Ka, 2019; Devi et al., 2019). Color loss is common in wines that have undergone MLF (Abrahamse and Bartowsky, 2012; Burns and Osborne, 2013, 2015). Independently from pH, wines post-MLF have lower levels of polymeric pigments, lower Visitin A and B content and a higher content of monomeric anthocyanins than their respective controls that did not undergo MLF (Burns and Osborne, 2013). An explanation for the color loss in wine post-MLF is the LAB metabolism of acetaldehyde (see acetaldehyde metabolism above) (Burns and Osborne, 2015). Acetaldehyde is crucial for the formation and stabilization of wine color because it mediates the formation of stable ethylene-linked pigments, which are more stable than their respective monomeric anthocyanins and show better colorimetric properties (Forino et al., 2020). Furthermore, pyruvic acid and acetaldehyde can react with pigments such as malvidin-3-glucoside, generating the relatively stable pyranoanthocyanins Visitin A and Visitin B (Waterhouse et al., 2016).

Recent studies have shown that *O. oeni* and *L. plantarum* strains can also absorb anthocyanin glucosides such as delphinidin-3-glucoside, malvidin-3-glucoside and peonidin-3-glucoside through the cell wall (Devi et al., 2019). These LAB can produce  $\beta$ -glycosidase enzymes that cleave the anthocyanin glucoside glycosidic bonds and can further degrade the aglycons into phloroglucinol aldehyde and corresponding phenolic acids (Devi et al., 2019). The absorption rate of anthocyanin glucosides, the  $\beta$ -glycosidase activity and the degradation rate of anthocyanins are dependent on the species and strains of LAB (Devi et al., 2019). Different inoculation regimes, such as coinoculation or sequential inoculation of yeast and bacteria, can determine different wine color outcomes, seemingly due to microbial interactions, rather than to absorption mechanisms (Devi and Ka, 2019). A higher color loss occurs when MLF is performed with sequential inoculation regimes, rather than with coinoculation regimes (Abrahamse and Bartowsky, 2012).

Little has been reported regarding the long-term effects of MLF on wine color. A 2016 study reported that, nine months after MLF completion, up to 9% color intensity loss and lower acylated and non-acylated anthocyanins levels were detected in wines (Izquierdo-Cañas et al., 2016). Interestingly an increase of the pyranoanthocyanin concentration was also observed (Izquierdo-Cañas et al., 2016).

Further research is warranted to understand the specific yeast-bacteria interactions during the secondary fermentation, and how they subsequently affect wine color. Considering the multitude

of factors involved, understanding the basis of wine color is not simple and far from fully achieved. However, this knowledge would greatly benefit the industry by enabling the winemaker to plan MLF in order to obtain the desired color outcome even in wines naturally low in pigments.

## POSSIBLE CONTRIBUTIONS OF LAB TO WINEMAKING

### Reducing the Need for Bentonite

Residual grape proteins can represent a problem for the quality of wine. Thaumatin-like proteins, chitinases and, to a lesser extent,  $\beta$ -glucanases, are the main classes of proteins responsible for wine protein instability (Van Sluyter et al., 2015; Cosme et al., 2020). Protein aggregation, particularly during wine storage, can lead to the formation of an unwanted sediment or haze (**Figure 1**) once the wine is bottled (Ferreira et al., 2001). Haze formation in white wines is mainly an esthetic issue, but it can economically depreciate the final product (Van Sluyter et al., 2015).

To alleviate this problem, the main treatment currently applied during winemaking is the addition of bentonite. However, this is a costly treatment and it involves a loss of wine volume, wine aroma and assimilable nitrogen resources (Muhlack et al., 2006). It has been shown that proteolytic and peptidolytic activities are common amongst LAB (Savijoki et al., 2006; Moslehishad et al., 2013; Atanasova et al., 2014; García-Cano et al., 2019). LAB proteolytic activities could be exploited in winemaking and have the potential to replace or reduce the use of fining agents such as bentonite. However, more studies are needed as most of the literature refers to work done in the dairy industry, at pH values of 6.5 and with considerably different substrates (Moslehishad et al., 2013; Atanasova et al., 2014; García-Cano et al., 2019).

### Aiding Clarification and Filtration of Juice

Grape juice is naturally rich in polysaccharides, such as pectin, cellulose, hemicellulose, and other substances, originating mainly from grape cell walls and the middle lamellae. In particular, the high content of pectins leads to the formation of a colloid structure, which can make processing of juice difficult (Sandri et al., 2011). The addition of fungal pectinase enzymes can help with the clarification and filtration of the juice, increasing must yield, and favoring the extraction of polyphenols, pigments and aromas (Merín et al., 2011). Some yeast strains are capable of degrading polysaccharides (Merín et al., 2014; Belda et al., 2016). Similarly, LAB may also possess pectinolytic activity, however, little research has been done in this field (Ruiz Rodríguez et al., 2019).

### Reducing the SO<sub>2</sub> Required in Winemaking

Due to its antimicrobial and antioxidant properties, SO<sub>2</sub> is one of the most common agents added to wine (Lisanti et al., 2019). Despite the benefits that come from SO<sub>2</sub>, the sensitivity

of some consumers to this compound, have raised concerns regarding its safety in the food and beverage industries (Benito, 2019). Thus, the addition of potassium metabisulfite to wine is strictly regulated, with maximum limits varying depending on local regulations. As a general guide, the international code of enological practices recommends a residual limit of 150 mg/L total sulfur dioxide for red wines and 200 mg/L for white and rosé wines with up to 4 g/L of reducing substances<sup>3</sup>.

As mentioned already, acetaldehyde strongly binds free SO<sub>2</sub>, making it less effective in its antiseptic, antioxidant and antioxidasic roles (Ribéreau-Gayon et al., 2006b). Thus, reducing the content of acetaldehyde in wine could help minimize the amount of SO<sub>2</sub> required in winemaking (**Figure 1**) and, at the same time, could improve the aroma of wine (Lisanti et al., 2019). The use of LAB starters that are able to degrade SO<sub>2</sub>-bound acetaldehyde (see acetaldehyde metabolism above) could represent a winemaking strategy to minimize SO<sub>2</sub> additions to wine by converting the bound SO<sub>2</sub> into the more effective free SO<sub>2</sub>. Importantly, acetaldehyde also plays a crucial role in the development of wine color. Thus, its content in wine has to be optimized to determine the balance between optimizing color, flavors and aromas, and health associated risks.

## THE IMPACT OF LAB ON THE WHOLESOMENESS OF THE WINE

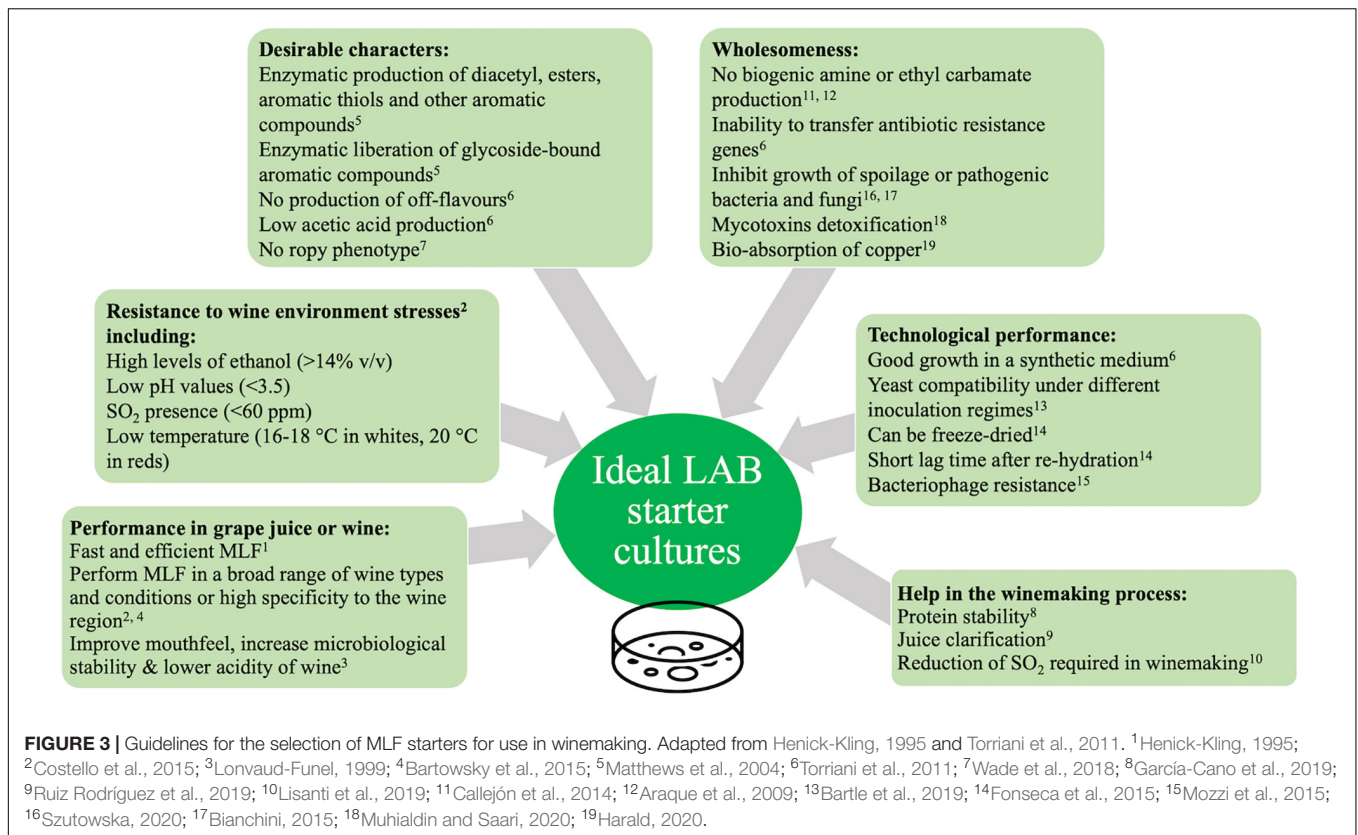
LAB can positively and negatively affect the impact that wine may have on human health (**Figure 3**). LAB can produce numerous metabolites, including organic acids (mainly lactic and acetic), phenyllactic acid, diacetyl, cyclic dipeptides, and bacteriocins that can inhibit the growth of spoilage or pathogenic bacteria (Corsetti et al., 2015; Szutowaska, 2020). They have also been reported to have antifungal activity, although the mechanisms of action are not clear (Bianchini, 2015). LAB can exert a high detoxifying action toward mycotoxins in various foods including wine (Muhialdin and Saari, 2020). This is likely to be due to the proteolytic activity of LAB, however, other detoxification mechanisms have been suggested, including the binding of mycotoxins to LAB metabolites such as acids, phenolic compounds and small bioactive peptides, and the absorption of mycotoxins by the bacterial cell walls (Muhialdin and Saari, 2020). Recent studies show that wines that underwent MLF have higher contents of melatonin and other tryptophan related compounds, which are associated with several human health benefits (Fracassetti et al., 2020). LAB could also have a role in the bio-absorption of copper, which, at high concentrations, is associated with health risks and negative effects in wine too (Harald, 2020).

### Biogenic Amines

Biogenic amines (BA) are potentially hazardous compounds that are commonly found in wine (**Figure 1**) and other fermented foods (Benito, 2019). Although BA can be produced in the grapes (Del Prete et al., 2009) and by yeast (Tristezza et al., 2013), their

<sup>3</sup><http://www.oiv.int>





presence in wine is primarily due to LAB metabolism (Costantini et al., 2019). The main BAs produced during MLF are histamine, putrescine and tyramine (Landete et al., 2007).

These BAs are formed through the activity of the LAB enzymes, histidine decarboxylase (*hdc*), ornithine decarboxylase (*odc*), and tyrosine decarboxylase (*tdc*), which decarboxylate the precursor compounds histidine, ornithine and tyramine, with both the enzymatic activity and presence of the genes encoding these proteins being strain dependent (Coton et al., 2010; Costantini et al., 2019). In addition to this, several LAB genera can produce putrescine via the agmatine deiminase, from its precursor agmatine (*agdi*) (Lucas et al., 2007).

Some LAB can degrade BA in culture media, and to different degrees in wine, through the action of multicopper oxidases or amine oxidases (García-Ruiz et al., 2011; Callejón et al., 2014). Many species, including *Levilactobacillus brevis* (formerly *Lactobacillus brevis*), *L. delbrueckii*, *L. hilgardii*, *L. paracasei*, *Lactiplantibacillus pentosus* (formerly *Lactobacillus pentosus*), *L. plantarum*, *P. parvulus*, and *P. pentosaceus* have been biochemically tested and showed degrading activity against BA (García-Ruiz et al., 2011; Callejón et al., 2014). Seemingly this property of LAB is strain dependent for most species tested, except in the case of *L. plantarum*, in which a high level of activity was widespread amongst all strains (Callejón et al., 2014). Similar results were obtained in a previous study which aimed to investigate the BA degrading activity of two *L. plantarum* strains (NDT 09 and NDT16) (Capozzi et al., 2012). Interestingly, the degrading activity of *L. plantarum* NDT09 and *L. plantarum*

NDT16 toward putrescine and tyramine, in synthetic media, was higher when the two strains were used in conjunction (Capozzi et al., 2012). Recently, a *L. plantarum* with good MLF and stress tolerance properties, that is also able to decrease the histamine, tyramine and cadaverine content by over 57%, has been obtained, through successive screenings (Sun et al., 2020).

Currently, the options available to control the BA content in wine are the reduction of precursor compounds, the inhibition of spoilage bacteria growth and the use of selected non biogenic amine-producer starters (Callejón et al., 2014). Given the recent findings on the ability of LAB, particularly *L. plantarum* strains, to degrade BA, the use of MLF starters that are able to reduce BA in wine would represent an innovative and alternative option for the winemaker.

## Ethyl Carbamate

Ethyl carbamate (Figure 1) is a carcinogenic compound found in many fermented foods and beverages (Benito, 2019). The reaction of ethanol with N-carbamyl compounds, such as urea (produced by yeast) and citrulline (produced by some LAB), is responsible for the main formation of ethyl carbamate in wine (Liu et al., 1994).

Some species of LAB can degrade L-arginine, through the arginine deaminase pathway, producing ATP, CO<sub>2</sub> and ammonium as final products (Araque et al., 2009). The intermediate steps in this pathway yield potential ethyl carbamate precursors like citrulline and carbamyl phosphate (Liu et al., 1994; Araque et al., 2009). The genes *arcA*, *arcB*, and *arcC*,

which encode for the enzymes arginine deiminase, ornithine transcarbamylase and carbamate kinase, respectively, have been characterized in many genera (Araque et al., 2009). The activity of these genes, and the production of ethyl carbamate precursors, has been reported for all wine-related genera (Liu et al., 1994, 1995; Araque et al., 2009).

The presence of the *arc* genes is generally a good predictor of the capability of LAB to degrading arginine (Liu et al., 1995; Araque et al., 2009); however, it is not indicative of the expression of the genes under particular circumstances. Both genotypic and phenotypic screenings are recommended to correctly characterize the capability of LAB to produce ethyl carbamate precursors in wine. On the other hand, LAB can produce urea-degrading enzymes that can selectively hydrolyze urea, contributing to reduction of ethyl carbamate in wine (Fang et al., 2016).

## IMPROVEMENT OF LAB AND TECHNOLOGICAL ADVANCES

### Strain Selection

The selection of new LAB starters requires the assessment of many properties related to the new strain including resistance to biotic and abiotic stresses, technological performances and safety aspects (Henick-Kling, 1995; Torriani et al., 2011; Costello et al., 2015). See **Figure 3** for a summary of the properties of interest. Traditionally a phenotypic approach has been used for the purpose of selecting new starters (Torriani et al., 2011). In recent years, phenotypic and genotypic approaches are used in tandem to rapidly characterise new potential starter candidates (Torriani et al., 2011).

### Microbial Integrity: Phage Resistance

One of the major issues that can arise during fermentation processes is the contamination of the LAB starter cultures with bacteriophages (Mozzi et al., 2015). Most of the knowledge regarding phage infections in LAB comes from the dairy industry, with *Lactococcus lactis* and *Streptococcus thermophilus* being the most susceptible and studied species (Mozzi et al., 2015). However, since *O. oeni*, *Lacticaseibacillus casei* (formerly known as *Lactobacillus casei*), *L. delbrueckii* and other species have become more prominent in the food industry, more research has been directed toward investigating their phages, disclosing multiple phage infections within these genera (Mozzi et al., 2015). In a 2017 study, 15 *O. oeni* phages have been studied, containing more than one type of integrase genes each and varying in genome size (Costantini et al., 2017). In the same study, it was shown that the presence of ethanol and low pH values inhibited the phages from attacking the cells, suggesting that inoculation time of the bacterial starters could play a significant role in preventing bacteriophage infections (Costantini et al., 2017). The first sequence of an *O. oeni* phage, E33PA, has been published in 2018 by Jaomanjaka and co-workers (Jaomanjaka et al., 2018). Interestingly, although most of phage isolated from the *Oenococcus* genus are temperate, E33PA was found to have a predominately lytic lifestyle (Jaomanjaka et al., 2018).

The adaptive immune system of bacteria is a potential mechanism for defense against phage, which has been highlighted by recent studies (Barrangou and Marraffini, 2014). For example, it is now known that Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) and CRISPR-associated genes, are essential elements in phage defense (Barrangou and Marraffini, 2014). CRISPR-Cas systems can confer phage resistance to the bacterial cell through three steps: during the adaptation, foreign genomic sequences can be integrated in the CRISPR arrays as new spacers, bordered by the palindromic repeats. During the expression the CRISPR locus is transcribed and processed, generating multiple copies of small CRISPR RNAs, able to direct the Cas proteins to foreign, complimentary DNA. Lastly, during the interference, foreign nucleic acid is targeted and cleaved (Crawley et al., 2018).

The CRISPR-mediated immunity mechanism causes phages to mutate at high rates, to maintain their ability to infect the hosts; furthermore, it's likely that phages and CRISPR systems are continuously co-evolving (Sorek et al., 2008). Interestingly, there is a rich variety of type II CRISPR-Cas systems within LAB (Sun et al., 2015); in a 2015 study comprising 213 different LAB genera, type II systems were found in 36% of them, representing a wealth of tools for LAB genetic engineering (Sun et al., 2015).

Conversely, CRISPR-Cas systems are either uncommon or absent in *O. oeni*. We searched six different *O. oeni* whole genome sequences from NCBI on the CRISPR finder database (version 4.2.20) (Couvin et al., 2018) showing that there are no CRISPR arrays or Cas clusters.

However, two closely related *Oenococcus* species contained both Cas clusters and CRISPR arrays at a high-quality score (quality score of 4). These were *O. kitaharae* DSM 17330 (CAS clusters = 1, CRISPR arrays = 1) and *O. sicerae* UCMA 15228 (CAS clusters = 2, CRISPR arrays = 2).

This inconsistent distribution of CRISPR-Cas immune systems within and between bacterial species is not uncommon and it may be explained by the fact that bacterial cells that have lost the CRISPR-Cas system from their genome avoid damage caused by autoimmune targeting (Rollie et al., 2020). Additionally, the efficacy of CRISPR-Cas is dependent on viral mutation rate and the frequency of spacer incorporation. *O. oeni* has a slow growth rate (~7 days to reach OD<sub>600</sub> of 1.0) and may not be able to mount a response fast enough or acquire the spacers quickly enough for the system to be effective (Sorek et al., 2008).

### Strategies to Harness LAB Enzymatic Activity

Directed evolution of microorganisms is a widely used technique for improving desired properties of bacterial strains (Bachmann et al., 2017). It is based on the concept that, during evolution, the environment selects the fittest variants. Growth limiting conditions, such as scarcity of nutrients, and environmental stresses induce a stress response and can lead to the production of genetic mutations (Rosenberg and Hastings, 2003). To generate an array of genetic and phenotypic variants treatments of UV and ethylmethanesulfonate (EMS) can also be applied to the culture (Dragosits and Mattanovich, 2013; Li et al., 2015).

As part of a broader study involving metabolic networks, a strain of *L. plantarum* has been successfully adapted, through serial dilutions, to grow well in a medium containing glycerol as main carbon source (Teusink et al., 2009). Directed evolution has been used to successfully generate acid-resistant mutants of *L. casei* and *Leuconostoc mesenteroides* (Zhang et al., 2012; Ju et al., 2016). An ethanol tolerant *O. oeni* strain, A90, has been generated after exposure to increasing ethanol concentrations, in approximately 330 generations (Betteridge et al., 2018). This strain was further evolved to withstand the multiple stresses typical of wine environment, obtaining after approximately 350 generations, an alcohol tolerant, acid tolerant (3.35 pH) and SO<sub>2</sub> tolerant (26 mg/L) strain (Jiang et al., 2018).

Directed evolution, as a strategy to harness LAB enzymatic activity, offers several advantages (Bennett, 2002). It does not involve the use of recombinant technology, posing less problems in the public acceptance of the products. It does not require a specific knowledge of the underlying genetics behind the specific phenotypic traits that are object of interest. Finally, whereas bioengineering approaches often target one gene at the time, directed evolution can result in multiple beneficial mutations, broadening the possible outcomes (Bloom and Arnold, 2009). However, the specific application of directed evolution to the LAB species associated with wine, is still a relatively new but promising technique. By using this approach, it could be possible to produce LAB strains that exhibit a higher desired enzymatic activity and/or are more tolerant to the harsh wine environment.

Another way to enhance the enzymatic activity of bacteria is the immobilization of the organisms onto supports such as alginate beads or apple pieces, corn cobs, delignified cellulosic material, grape skins and grape stems (Genisheva et al., 2013; Bleve et al., 2016; Simó et al., 2017; Nikolaou et al., 2020). Immobilized bacteria can perform MLF twice as fast, compared to free cells, and are also more resistant to ethanol content, SO<sub>2</sub> content and elevated temperatures (Genisheva et al., 2013).

Currently, it may not be practical to apply this methodology in a traditional winemaking system, considering the large amounts of required supporting material (up to 30 g/L). Nonetheless this is a promising technique, with a great potential to improve MLF in wine. Furthermore, it would be interesting to know if secondary enzymatic activities of LAB would also be affected by the use of immobilized cells.

## Genome Editing of LAB

In addition to strain selection and non-GMO strategies to harness enzymatic activity of LAB, genome editing is a promising strategy to develop high performing bacterial starters. Transformation with plasmids, transduction and conjugative transposons systems are the main recombinant methods used to genetically manipulate LAB (Sumbly et al., 2014). The transformation of *Lactiplantibacillus* and *Lacticaseibacillus* strains has been successfully performed by many authors (Spath et al., 2012; Xin et al., 2018). However, the rate of success of this method depends on the plasmid vector, on the bacterial strain and the possible presence of a enzymatic restriction system in the host cell (Teresa Alegre et al., 2004). Currently, not many expression vectors are available for *O. oeni* (Sumbly et al., 2014).

A plasmid, pGID052, was developed in 2004 and successfully mobilized from *L. lactis* to *O. oeni* (Beltramo et al., 2004). The same plasmid, pGID052, encoding a truncated form of the ClpL2 protein was later introduced in *O. oeni* ATCC BAA-1163, trialing an optimized electroporation method (Assad-García et al., 2008). Although promising, there has been no follow up research to these findings, perhaps due to the low copy level of this plasmid (Sumbly et al., 2014).

The most promising technology is currently based on CRISPR-Cas systems. This strategy exploits the functions of CRISPR and CRISPR-associated genes (Doudna and Charpentier, 2014; van Pijkeren and Barrangou, 2017). Desired edits can be obtained with different methods, e.g., utilizing a plasmid-encoded recombineering sequence or an oligonucleotide sequence with an inducible DNA recombinase (Leenay et al., 2019). The efficiency of the selected method varies dramatically across different strains of *Lactoplantibacillus plantarum* (Leenay et al., 2019).

## Bacterial Starter Implementation

Freeze drying is the method of choice for long term storage of bacteria and continuous research has been done toward the optimization of the process (Fonseca et al., 2015, 2019; Polo et al., 2017). Freeze dried bacterial starters have a good rate of cell viability over long periods and are easy to use, store and transport (Fonseca et al., 2015). To protect the integrity of the cells membranes and the structural conformation of proteins and DNA during the dehydration process, lyoprotectants such as trehalose, sucrose and monosodium glutamate, are typically added (Wang et al., 2019). More recently, soluble extracellular polymeric substances (sEPS) from *O. oeni* have been successfully tested as lyoprotectants. Notably, a mixture of 5% sEPS, 15% sucrose, 15% trehalose, and 0.5% MSG, increased the cell viability to 92.83% (Wang et al., 2019). The positive effects of ethanol acclimatization on cell survival rate of lyophilized *O. oeni* have also been reported (Yang et al., 2020). Due to the length and costs associated with this process, other techniques have been investigated, including vacuum drying, spray drying, drum drying, fluidized bed drying and air drying (Santivarangkna et al., 2007). Currently these alternative drying processes do not perform as well as freeze drying, especially regarding the cell viability and the lag time required to restore the metabolic activity (Santivarangkna et al., 2007). However, they could represent a cheaper, fast and efficient way to preserve commercial bacterial starters. Promising results have also been obtained by freeze drying bacterial cultures previously immobilized onto natural supports (Nikolaou et al., 2020).

## LIMITATIONS AND FUTURE PROSPECTS

Currently the main enological role of LAB is performing MLF. Thus, the use of LAB, with the intention of increasing wine aroma and facilitating the winemaking process, is an unexplored and promising field. In this new broader

role, LAB could replace, or partially substitute, the use of purified enzymatic preparations. Furthermore, using whole cells, rather than purified enzymes, provides a series of advantages (De Carvalho and da Fonseca, 2007).

Firstly, whole cells are a natural environment for enzymes, preventing loss of activity in non-conventional media. Secondly, they are able to efficiently regenerate co-factors. However, more studies are needed in order to improve the knowledge of LAB's secondary enzymatic activities and their potential usability in the production of wine.

To benefit from the LAB's secondary metabolism a system to control and enhance the expression of these activities is needed.

Currently, disregarding studies on recombinant technology, little research has been done on this point (Renault, 2002). Directed evolution has been successfully used to produce highly performing LAB strains in the dairy industry, but seldomly in the wine industry (Betteridge et al., 2015). However, without resorting to genetic engineering techniques, no other methods are currently available to enhance a chosen secondary enzymatic activity in a LAB strain.

Overall, the benefits of harnessing the enzymatic activities of LAB will increase the options open to winemakers to sculpt the

wine of their choice. The industry could benefit greatly from the advent of these extra tools.

## AUTHOR CONTRIBUTIONS

CV and KS prepared the first draft. All authors reviewed and finalized the manuscript. All authors contributed to the development of the outline and scope of the review.

## FUNDING

Financial support for this work was provided by the Lallemand Inc. and the Australian Research Council Training Center for Innovative Wine Production ([www.arcwinecentre.org.au](http://www.arcwinecentre.org.au); IC170100008), which is funded by the Australian Government with additional support from Wine Australia and industry partners. CV is supported by a scholarship provided by Lallemand Inc. The University of Adelaide is a member of the Wine Innovation Cluster (<http://www.thewaite.org.waite-partners/wine-innovation-cluster/>).

## REFERENCES

- Abrahamse, C., and Bartowsky, E. (2012). Timing of malolactic fermentation inoculation in Shiraz grape must and wine: influence on chemical composition. *World J. Microbiol. Biotechnol.* 28, 255–265. doi: 10.1007/s11274-011-0814-813
- Antalick, G., Perello, M.-C., and de Revel, G. (2012). Characterization of fruity aroma modifications in red wines during malolactic fermentation. *J. Agric. Food Chem.* 60, 12371–12383. doi: 10.1021/jf303238n
- Araque, I., Gil, J., Carreté, R., Bordons, A., and Reguant, C. (2009). Detection of *Arc* genes related with the ethyl carbamate precursors in wine lactic acid bacteria. *J. Agric. Food Chem.* 57, 1841–1847. doi: 10.1021/jf803421w
- Aryan, A. P., Wilson, B., Strauss, C. R., and Williams, P. J. (1987). The properties of glycosidases of *Vitis vinifera* and a comparison of their  $\beta$ -glucosidase activity with that of exogenous enzymes. An assessment of possible applications in enology. *Am. J. Enol. Vitic.* 38, 182–188.
- Assad-García, J. S., Bonnín-Jusserand, M., Garmyn, D., Guzzo, J., Alexandre, H., and Grandvalet, C. (2008). An improved protocol for electroporation of *Oenococcus oeni* ATCC BAA-1163 using ethanol as immediate membrane fluidizing agent. *Letts. Appl. Microbiol.* 47, 333–338. doi: 10.1111/j.1472-765X.2008.02435.x
- Atanasova, J., Moncheva, P., and Ivanova, I. (2014). Proteolytic and antimicrobial activity of lactic acid bacteria grown in goat milk. *Biotechnol. Biotechnol. Equip.* 28, 1073–1078. doi: 10.1080/13102818.2014.971487
- Bachmann, H., Molenaar, D., Branco, dos Santos, F., and Teusink, B. (2017). Experimental evolution and the adjustment of metabolic strategies in lactic acid bacteria. *FEMS Microbiol. Rev.* 41, S201–S219. doi: 10.1093/femsre/fux024
- Barbagallo, R. N., Spagna, G., Palmeri, R., and Torriani, S. (2004). Assessment of  $\beta$ -glucosidase activity in selected wild strains of *Oenococcus oeni* for malolactic fermentation. *Enzyme Microb. Technol.* 34, 292–296. doi: 10.1016/j.enzmictec.2003.11.013
- Barrangou, R., and Marraffini, L. A. (2014). CRISPR-Cas systems: prokaryotes upgrade to adaptive immunity. *Mol. Cell* 54, 234–244. doi: 10.1016/j.molcel.2014.03.011
- Bartle, L., Sumby, K., Sundstrom, J., and Jiranek, V. (2019). The microbial challenge of winemaking: yeast-bacteria compatibility. *FEMS Yeast Res.* 19:foz040. doi: 10.1093/femsyr/foz040
- Bartowsky, E. J. (2005). *Oenococcus oeni* and malolactic fermentation - moving into the molecular arena. *Aust. J. Grape Wine Res.* 11, 174–187. doi: 10.1111/j.1755-0238.2005.tb00286.x
- Bartowsky, E. J. (2009). Bacterial spoilage of wine and approaches to minimize it. *Letts. Appl. Microbiol.* 48, 149–156. doi: 10.1111/j.1472-765X.2008.02505.x
- Bartowsky, E. J., and Borneman, A. R. (2011). Genomic variations of *Oenococcus oeni* strains and the potential to impact on malolactic fermentation and aroma compounds in wine. *Appl. Microbiol. Biotechnol.* 92, 441–447. doi: 10.1007/s00253-011-3546-3542
- Bartowsky, E. J., and Henschke, P. A. (2004). The 'buttery' attribute of wine - diacetyl - desirability, spoilage and beyond. *Int. J. Food Microbiol.* 96, 235–252. doi: 10.1016/j.ijfoodmicro.2004.05.013
- Bartowsky, E. J., Costello, P. J., and Chambers, P. J. (2015). Emerging trends in the application of malolactic fermentation. *Aust. J. Grape Wine Res.* 21, 663–669. doi: 10.1111/ajgw.12185
- Bartowsky, E. J., Francis, I. L., Bellon, J. R., and Henschke, P. A. (2002). Is buttery aroma perception in wines predictable from the diacetyl concentration? *Aust. J. Grape Wine Res.* 8, 180–185. doi: 10.1111/j.1755-0238.2002.tb00254.x
- Baumes, R. (2009). "Wine aroma precursors," in *Wine Chemistry and Biochemistry*, eds M. V. Moreno-Arribas and M. C. Polo (New York, NY: Springer), 251–274.
- Belda, I., Conchillo, L. B., Ruiz, J., Navascués, E., Marquina, D., and Santos, A. (2016). Selection and use of pectinolytic yeasts for improving clarification and phenolic extraction in winemaking. *Int. J. Food Microbiol.* 223, 1–8. doi: 10.1016/j.ijfoodmicro.2016.02.003
- Beltramo, C., Oraby, M., Bourel, G., Garmyn, D., and Guzzo, J. (2004). A new vector, pGID052, for genetic transfer in *Oenococcus oeni*. *FEMS Microbiol. Lett.* 236, 53–60. doi: 10.1016/j.femsle.2004.05.029
- Benito, S. (2019). The management of compounds that influence human health in modern winemaking from an HACCP point of view. *Fermentation* 5:33. doi: 10.3390/fermentation5020033
- Benito, S., Morata, A., Palomero, F., González, M. C., and Suárez-Lepe, J. A. (2011). Formation of vinylphenolic pyranoanthocyanins by *Saccharomyces cerevisiae* and *Pichia guilliermondii* in red wines produced following different fermentation strategies. *Food Chem.* 124, 15–23. doi: 10.1016/j.foodchem.2010.05.096
- Bennett, A. (2002). Experimental evolution and the Krogh principle: generating biological novelty for functional and genetic analyses. *Physiol. Biochem. Zool.* 76, 1–11. doi: 10.1086/374275
- Berbegal, C., Fragasso, M., Russo, P., Bimbo, F., Grieco, F., Spano, G., et al. (2019). Climate changes and food quality: the potential of microbial activities as mitigating strategies in the wine sector. *Fermentation* 5:85. doi: 10.3390/fermentation5040085
- Betteridge, A. L., Sumby, K. M., Sundstrom, J. F., Grbin, P. R., and Jiranek, V. (2018). Application of directed evolution to develop ethanol tolerant *Oenococcus oeni* for more efficient malolactic fermentation. *Appl. Microbiol. Biotechnol.* 102, 921–932. doi: 10.1007/s00253-017-8593-x

- Betteridge, A., Grbin, P., and Jiranek, V. (2015). Improving *Oenococcus oeni* to overcome challenges of wine malolactic fermentation. *Trends Biotechnol.* 33, 547–553. doi: 10.1016/j.tibtech.2015.06.008
- Bianchini, A. (2015). “Lactic acid bacteria as antifungal agents,” in *Advances in Fermented Foods and Beverages*, ed. W. Holzapfel (Cambridge: Elsevier Ltd.), 333–353. doi: 10.1016/B978-1-78242-015-6.00014-1
- Bleve, G., Tufariello, M., Vetrano, C., Mita, G., and Grieco, F. (2016). Simultaneous alcoholic and malolactic fermentations by *Saccharomyces cerevisiae* and *Oenococcus oeni* cells co-immobilized in alginate beads. *Front. Microbiol.* 7:943. doi: 10.3389/fmicb.2016.00943
- Bloom, J. D., and Arnold, F. H. (2009). In the light of directed evolution: pathways of adaptive protein evolution. *Proc. Natl. Acad. Sci. U.S.A.* 106, 9995–10000. doi: 10.1073/pnas.0901522
- Bokulich, N. A., Collins, T. S., Masarweh, C., Allen, G., Heymann, H., Ebeler, S. E., et al. (2016). Associations among wine grape microbiome, metabolome, and fermentation behavior suggest microbial contribution to regional wine characteristics. *mBio* 7:e00631-16. doi: 10.1128/mBio.00631-16
- Bravo-Ferrada, B., Hollmann, A., Delfederico, L., Valdés, La Hens, D., Caballero, A., et al. (2013). Patagonian red wines: selection of *Lactobacillus plantarum* isolates as potential starter cultures for malolactic fermentation. *World J. Microbiol. Biotechnol.* 29, 1537–1549. doi: 10.1007/s11274-013-1337-x
- Brizuela, N. S., Bravo-Ferrada, B. M., Curilén, Y., Delfederico, L., Caballero, A., and Semorile, L. (2018). Advantages of using blend cultures of native *L. plantarum* and *O. oeni* strains to induce malolactic fermentation of Patagonian Malbec wine. *Front. Microbiol.* 9:2109. doi: 10.3389/fmicb.2018.02109
- Burns, T. R., and Osborne, J. P. (2013). Impact of malolactic fermentation on the color and color stability of Pinot noir and Merlot wine. *Am. J. Enol. Vitic.* 64, 370–377. doi: 10.5344/ajev.2013.13001
- Burns, T. R., and Osborne, J. P. (2015). Loss of Pinot noir wine color and polymeric pigment after malolactic fermentation and potential causes. *Am. J. Enol. Vitic.* 66, 130–137. doi: 10.5344/ajev.2014.14061
- Callejón, S., Sendra, R., Ferrer, S., and Pardo, I. (2014). Identification of a novel enzymatic activity from lactic acid bacteria able to degrade biogenic amines in wine. *Appl. Microbiol. Biotechnol.* 98, 185–198. doi: 10.1007/s00253-013-4829-4826
- Capaldo, A. A. (2012). *Genes and Mechanisms Responsible for  $\beta$ -Glucoside Metabolism in the Oenologically Important Lactic Acid Bacterium *Oenococcus oeni**. Ph.D. thesis, University of Adelaide, Australia.
- Capozzi, V., Russo, P., Ladero, V., Fernández, M., Fiocco, D., Alvarez, M. A., et al. (2012). Biogenic amines degradation by *Lactobacillus plantarum*: toward a potential application in wine. *Front. Microbiol.* 3:122. doi: 10.3389/fmicb.2012.00122
- Cappello, M. S., Zapparoli, G., Logrieco, A., and Bartowsky, E. J. (2017). Linking wine lactic acid bacteria diversity with wine aroma and flavour. *Int. J. Food Microbiol.* 243, 16–27. doi: 10.1016/j.ijfoodmicro.2016.11.025
- Chatonnet, P., Dubourdieu, D., Boidron, J. N., and Pons, M. (1992). The origin of ethylphenols in wines. *J. Sci. Food Agric.* 60, 165–178. doi: 10.1002/jsfa.2740600205
- Chescheir, S., Philbin, D., and Osborne, J. P. (2015). Impact of *Oenococcus oeni* on wine hydroxycinnamic acids and volatile phenol production by *Brettanomyces bruxellensis*. *Am. J. Enol. Vitic.* 66, 357–362. doi: 10.5344/ajev.2015.14108
- Chung, M. J. (2018). Composition for Alcohol or Acetaldehyde Degradation Comprising Probiotics. European patent no. 3 415 155 A1, filed May 17, 2018, and issued December 19, 2018.
- Codres, C., Răpeanu, G., and Alexe, P. (2012). Effect of  $\beta$ -glucosidases in the making of Chardonnay wines. *Ann. Univ.* 36, 9–17.
- Collombel, I., Melkonian, C., Molenaar, D., Campos, F. M., and Hogg, T. (2019). New insights into cinnamoyl esterase activity of *Oenococcus oeni*. *Front. Microbiol.* 10:2597. doi: 10.3389/fmicb.2019.02597
- Corsetti, A., Perpetuini, G., and Tofalo, R. (2015). “Biopreservation effects in fermented foods,” in *Advances in Fermented Foods and Beverages*, ed. W. Holzapfel (Cambridge: Elsevier Ltd.), 311–332. doi: 10.1016/B978-1-78242-015-6.00013-X
- Cosme, F., Fernandes, C., Ribeiro, T., Filipe-Ribeiro, L., and Nunes, F. (2020). White wine protein instability: mechanism, quality control and technological alternatives for wine stabilisation - an overview. *Beverages* 6:19. doi: 10.3390/beverages6010019
- Costantini, A., Doria, F., Saiz, J.-C., and Garcia-Moruno, E. (2017). Phage-host interactions analysis of newly characterized *Oenococcus oeni* bacteriophages: implications for malolactic fermentation in wine. *Int. J. Food Microbiol.* 246, 12–19. doi: 10.1016/j.ijfoodmicro.2017.01.020
- Costantini, A., Vaudano, E., Pulcini, L., Carafa, T., and Garcia-Moruno, E. (2019). An overview on biogenic amines in wine. *Beverages* 5:19. doi: 10.3390/beverages5010019
- Costello, P. J., Siebert, T. E., Solomon, M. R., and Bartowsky, E. J. (2012). Synthesis of fruity ethyl esters by acyl coenzyme a : alcohol acyltransferase and reverse esterase activities in *Oenococcus oeni* and *Lactobacillus plantarum*. *J. Appl. Microbiol.* 114, 797–806. doi: 10.1111/jam.12098
- Costello, P., Deléris-Bou, M., Descenzo, R., Hall, N., Krieger, S., and Lonvaud Funel, A. (2015). “Strain selection techniques,” in *Malolactic Fermentation – Importance of Wine Lactic Acid Bacteria in Winemaking*, eds R. Morenzone and K. S. Specht (Montréal: Lallemand Inc).
- Coton, M., Romano, A., Spano, G., Ziegler, K., Vetrano, C., and Desmarais, C. (2010). Occurrence of biogenic amine-forming lactic acid bacteria in wine and cider. *Food Microbiol.* 27, 1078–1085. doi: 10.1016/j.fm.2010.07.012
- Couvin, D., Bernheim, A., Toffano-Nioche, C., Touchon, M., Michalik, J., and Néron, B. (2018). CRISPR-CasFinder, an update of CRISPRFinder, includes a portable version, enhanced performance and integrates search for cas proteins. *Nucleic Acids Res.* 46, W246–W251. doi: 10.1093/nar/gky425
- Crawley, A. B., Henriksen, E. D., Stout, E., Brandt, K., and Barrangou, R. (2018). Characterizing the activity of abundant, diverse and active CRISPR-Cas systems in lactobacilli. *Sci. Rep.* 8, 1–12. doi: 10.1038/s41598-018-29746-29743
- D’Incecco, N., Bartowsky, E., Kassara, S., Lante, A., Spetoli, P., and Henschke, P. (2004). Release of glycosidically bound flavour compounds of Chardonnay by *Oenococcus oeni* during malolactic fermentation. *Food Microbiol.* 21, 257–265. doi: 10.1016/j.fm.2003.09.003
- De Carvalho, C., and da Fonseca, M. M. R. (2007). Bacterial whole cell biotransformations: in vivo reactions under in vitro conditions. *Dyn. Biochem. Process. Biotechnol. Mol. Biol.* 1, 32–39.
- Del Prete, V., Costantini, A., Cecchini, F., Morassut, M., and Garcia-Moruno, E. (2009). Occurrence of biogenic amines in wine: the role of grapes. *Food Chem.* 112, 474–481. doi: 10.1016/j.foodchem.2008.05.102
- Devi, A., Aiyappaa, A. A. K., and Waterhouse, A. L. (2019). Adsorption and biotransformation of anthocyanin glucosides and quercetin glucosides by *Oenococcus oeni* and *Lactobacillus plantarum* in model wine solution. *J. Sci. Food Agric.* 100, 2110–2120. doi: 10.1002/jsfa.10234
- Devi, A., and Ka, A.-A. (2019). Yeast-Bacterial Interactions during malolactic inoculations affecting anthocyanin adsorption and content in Shiraz wine. *Am. J. Enol. Vitic.* 71, 105–112. doi: 10.5344/ajev.2019.19033
- Doudna, J. A., and Charpentier, E. (2014). The new frontier of genome engineering with CRISPR-Cas9. *Science* 346:1258096. doi: 10.1126/science.1258096
- Dragosits, M., and Mattanovich, D. (2013). Adaptive laboratory evolution – principles and applications for biotechnology. *Microb. Cell Fact.* 12:64. doi: 10.1186/1475-2859-12-64
- Du Plessis, H., Du Toit, M., Nieuwoudt, H., Van der Rijst, M., Hoff, J., and Jolly, N. (2019). Modulation of wine flavor using *Hanseniaspora uvarum* in combination with different *Saccharomyces cerevisiae*, lactic acid bacteria strains and malolactic fermentation strategies. *Fermentation* 5:64. doi: 10.3390/fermentation5030064
- Du Toit, M., Engelbrecht, L., Lerm, E., and Krieger-Weber, S. (2010). *Lactobacillus*: the next generation of malolactic fermentation starter cultures - an overview. *Food Bioproc. Tech.* 4, 876–906. doi: 10.1007/s11947-010-0448-448
- Dungey, K. A., Hayasaka, Y., and Wilkinson, K. L. (2011). Quantitative analysis of glycoconjugate precursors of gualaicol in smoke-affected grapes using liquid chromatography–tandem mass spectrometry based stable isotope dilution analysis. *Food Chem.* 126, 801–806. doi: 10.1016/j.foodchem.2010.11.094
- Fang, F., Feng, T., Du, G., and Chen, J. (2016). Evaluation of the impact on food safety of a *Lactobacillus coryniformis* strain from pickled vegetables with degradation activity against nitrite and other undesirable compounds. *Food Addit. Contam. Part A* 33, 623–630. doi: 10.1080/19440049.2016.1156774
- Ferreira, R. B., Piçarra-Pereira, M. A., Monteiro, S., Loureiro, V. B., and Teixeira, A. R. (2001). The wine proteins. *Trends Food Sci. Technol.* 12, 230–239. doi: 10.1016/S0924-2244(01)00080-82
- Fia, G., Millarini, V., Granchi, L., Bucalossi, G., Guerrini, S., Zanon, B., et al. (2018). Beta-glucosidase and esterase activity from *Oenococcus oeni*: screening and evaluation during malolactic fermentation in harsh conditions. *Food Sci. Technol.* 89, 262–268. doi: 10.1016/j.lwt.2017.10.060

- Fonseca, F., Cenard, S., and Passot, S. (2015). *Cryopreservation and Freeze-Drying Protocols*, 3rd edn., eds W. F. Wolkers and H. Oldenhof (New York, NY: Springer). doi: 10.1007/978-1-4939-2193-5
- Fonseca, F., Pénicaud, C., Tymczyszyn, E. E., Gómez-Zavaglia, A., and Passot, S. (2019). Factors influencing the membrane fluidity and the impact on production of lactic acid bacteria starters. *Appl. Microbiol. Biotechnol.* 103, 6867–6883. doi: 10.1007/s00253-019-10002-10001
- Forino, M., Picariello, L., Lopatriello, A., Moio, L., and Gambuti, A. (2020). New insights into the chemical bases of wine color evolution and stability: the key role of acetaldehyde. *Eur. Food Res. Technol.* 246, 733–743. doi: 10.1007/s00217-020-03442-x
- Fracassetti, D., Francesco, Lo Faro, A. F., Moiola, S., Orioli, M., Tirelli, A., et al. (2020). Production of melatonin and other tryptophan derivatives by *Oenococcus oeni* under winery and laboratory scale. *Food Microbiol.* 86:103265. doi: 10.1016/j.fm.2019.103265
- Gammacurta, M., Lytra, G., Marchal, A., Marchand, S., Christophe Barbe, J., and Moine, V. (2018). Influence of lactic acid bacteria strains on ester concentrations in red wines: specific impact on branched hydroxylated compounds. *Food Chem.* 239, 252–259. doi: 10.1016/j.foodchem.2017.06.123
- García-Cano, I., Rocha-Mendoza, D., Ortega-Anaya, J., Wang, K., Kosmerl, E., and Jiménez-Flores, R. (2019). Lactic acid bacteria isolated from dairy products as potential producers of lipolytic, proteolytic and antibacterial proteins. *Appl. Microbiol. Biotechnol.* 103, 5243–5257. doi: 10.1007/s00253-019-09844-9846
- García-Ruiz, A., González-Rompinelli, E. M., Bartolomé, B., and Moreno-Arribas, M. V. (2011). Potential of wine-associated lactic acid bacteria to degrade biogenic amines. *Int. J. Food Microbiol.* 148, 115–120. doi: 10.1016/j.ijfoodmicro.2011.05.009
- Genisheva, Z., Mussatto, S. I., Oliveira, J. M., and Teixeira, J. A. (2013). Malolactic fermentation of wines with immobilised lactic acid bacteria – Influence of concentration, type of support material and storage conditions. *Food Chem.* 138, 1510–1514. doi: 10.1016/j.foodchem.2012.11.058
- González-Neves, G., Gil, G., Favre, G., Baldi, C., Hernández, N., and Traverso, S. (2013). Influence of winemaking procedure and grape variety on the colour and composition of young red wines. *S. Afr. J. Enol. Vitic.* 34, 138–146.
- Grimaldi, A., Bartowsky, E., and Jiranek, V. (2005a). A survey of glycosidase activities of commercial wine strains of *Oenococcus oeni*. *Int. J. Food Microbiol.* 105, 233–244. doi: 10.1016/j.ijfoodmicro.2005.04.011
- Grimaldi, A., Bartowsky, E., and Jiranek, V. (2005b). Screening of *Lactobacillus* spp. and *Pediococcus* spp. for glycosidase activities that are important in oenology. *J. Appl. Microbiol.* 99, 1061–1069. doi: 10.1111/j.1365-2672.2005.02707.x
- Grimaldi, A., McLean, H., and Jiranek, V. (2000). Identification and partial characterization of glycosidase activities of commercial strains of the lactic acid bacterium, *Oenococcus oeni*. *Am. J. Enol. Vitic.* 51, 362–369.
- Gunata, Z., Bitteur, S., Brillouet, J.-M., Bayonove, C., and Cordonnier, R. (1988). Sequential enzymic hydrolysis of potentially aromatic glycosides from grape. *Carbohydr. Res.* 184, 139–149.
- Harald, C. (2020). How to deal with uninvited guests in wine: copper and copper-containing oxidases. *Fermentation* 6:38. doi: 10.3390/fermentation6010038
- Hayasaka, Y., Baldock, G. A., Pardon, K. H., Jeffery, D. W., and Herderich, M. J. (2010). Investigation into the formation of guaiaicol conjugates in berries and leaves of grapevine *Vitis vinifera* L. Cv. Cabernet Sauvignon using stable isotope tracers combined with HPLC-MS and MS/MS analysis. *J. Agric. Food Chem.* 58, 2076–2081. doi: 10.1021/jf903732p
- Henick-Kling, T. (1995). Control of malo-lactic fermentation in wine: energetics, flavour modification and methods of starter culture preparation. *J. Appl. Bacteriol. Symp. Suppl.* 79, 29–37.
- Hernandez-Orte, P., Cersosimo, M., Loscos, N., Cacho, J., Garcia-Moruno, E., and Ferreira, V. (2009). Aroma development from non-floral grape precursors by wine lactic acid bacteria. *Food Res. Int.* 42, 773–781.
- Holzappel, W., Abriouel, H., and Wood, B. J. B. (2014). *Lactic Acid Bacteria: Biodiversity and Taxonomy*, eds W. Holzappel and B. J. B. Wood (Chichester, UK: Wiley Blackwell).
- Iorizzo, M., Testa, B., Lombardi, S. J., García-Ruiz, A., Muñoz-González, C., and Bartolomé, B. (2016). Selection and technological potential of *Lactobacillus plantarum* bacteria suitable for wine malolactic fermentation and grape aroma release. *Food Sci. Technol.* 73, 557–566. doi: 10.1016/j.lwt.2016.06.062
- Izquierdo-Cañas, P. M., García-Romero, E., Mena-Morales, A., and Gómez-Alonso, S. (2016). Effects of malolactic fermentation on colour stability and phenolic composition of Petit Verdot red wines. *Wine Stud.* 5:5795. doi: 10.4081/ws.2016.5795
- Jaomanjaka, F., Claisse, O., Philippe, C., and Le Marrec, C. (2018). Complete genome sequence of lytic *Oenococcus oeni* bacteriophage OE33PA. *Microbiol. Resour. Announc.* 7:e818. doi: 10.1128/MRA.00818-18
- Jiang, J., Sumby, K. M., Sundstrom, J. F., Grbin, P. R., and Jiranek, V. (2018). Directed evolution of *Oenococcus oeni* strains for more efficient malolactic fermentation in a multi-stressor wine environment. *Food Microbiol.* 73, 150–159. doi: 10.1016/j.fm.2018.01.005
- Jiranek, V. (2011). Smoke taint compounds in wine: nature, origin, measurement and amelioration of affected wines: smoke taint compounds in wine. *Aust. J. Grape Wine Res.* 17, S2–S4. doi: 10.1111/j.1755-0238.2011.00154.x
- Ju, S. Y., Kim, J. H., and Lee, P. C. (2016). Long-term adaptive evolution of *Leuconostoc mesenteroides* for enhancement of lactic acid tolerance and production. *Biotechnol. Biofuels* 9:240. doi: 10.1186/s13068-016-0662-663
- Kennison, K. R., Gibberd, M. R., Pollnitz, A. P., and Wilkinson, K. L. (2008). Smoke-derived taint in wine: the release of smoke-derived volatile phenols during fermentation of Merlot juice following grapevine exposure to smoke. *J. Agric. Food Chem.* 56, 7379–7383. doi: 10.1021/jf800927e
- Kennison, K. R., Wilkinson, K. L., Williams, H. G., Smith, J. H., and Gibberd, M. R. (2007). Smoke-derived taint in wine: effect of postharvest smoke exposure of grapes on the chemical composition and sensory characteristics of wine. *J. Agric. Food Chem.* 55, 10897–10901. doi: 10.1021/jf800927e
- Krieger-Weber, S., Heras, J., and Suarez, C. (2020). *Lactobacillus plantarum*, a new biological tool to control malolactic fermentation: a review and an outlook. *Beverages* 6:23. doi: 10.3390/beverages6020023
- Lachenmeier, D. W., and Sohnius, E.-M. (2008). The role of acetaldehyde outside ethanol metabolism in the carcinogenicity of alcoholic beverages: evidence from a large chemical survey. *Food Chem. Toxicol.* 46, 2903–2911. doi: 10.1016/j.fct.2008.05.034
- Landete, J. M., Ferrer, S., and Pardo, I. (2007). Biogenic amine production by lactic acid bacteria, acetic bacteria and yeast isolated from wine. *Food Control* 18, 1569–1574. doi: 10.1016/j.foodcont.2006.12.008
- Lasik-Kurdyś, M., Majcher, M., and Nowak, J. (2018). Effects of different techniques of malolactic fermentation induction on diacetyl metabolism and biosynthesis of selected aromatic esters in cool-climate grape wines. *Molecules* 23:2549. doi: 10.3390/molecules23102549
- Leenay, R. T., Vento, J. M., Shah, M., Martino, M. E., Leulier, F., and Beisel, C. L. (2019). Genome editing with CRISPR-Cas9 in *Lactobacillus plantarum* revealed that editing outcomes can vary across strains and between methods. *Biotechnol. J.* 4:e1700583.
- Lerm, E., Engelbrecht, L., and Du Toit, M. (2010). Malolactic fermentation: the Abc's of MLF. *J. Enol. Vitic.* 31, 186–212. doi: 10.21548/31-2-1417
- Lerm, E., Engelbrecht, L., and du Toit, M. (2011). Selection and characterisation of *Oenococcus oeni* and *Lactobacillus plantarum* South African wine isolates for use as malolactic fermentation starter cultures. *S. Afr. J. Enol. Vitic.* 32, 280–295. doi: 10.21548/32-2-1388
- Li, N., Duan, J., Gao, D., Luo, J., Zheng, R., Bian, Y., et al. (2015). Mutation and selection of *Oenococcus oeni* for controlling wine malolactic fermentation. *Eur. Food Res. Technol.* 240, 93–100. doi: 10.1007/s00217-014-2310-2310
- Lima, A., Oliveira, C., Santos, C., Campos, F., and Couto, J. (2018). Phenolic composition of monovarietal red wines regarding volatile phenols and its precursors. *Eur. Food Res. Technol.* 244, 1985–1994. doi: 10.1007/s00217-018-3110-3118
- Lisanti, M. T., Blaiotta, G., Nioi, C., and Moio, L. (2019). Alternative methods to SO<sub>2</sub> for microbiological stabilization of wine. *Compr. Rev. Food. Sci. Food. Saf.* 18, 455–479. doi: 10.1111/1541-4337.12422
- Liu, S. Q. (2002). Malolactic fermentation in wine – beyond deacidification. *J. Appl. Microbiol.* 92, 589–601. doi: 10.1046/j.1365-2672.2002.01589.x
- Liu, S. Q., and Pilone, G. J. (2000). An overview of formation and roles of acetaldehyde in winemaking with emphasis on microbiological implications. *Intern. J. Food Sci. Technol.* 35, 49–61. doi: 10.1046/j.1365-2621.2000.00341.x
- Liu, S. Q., Pritchard, G. G., Hardman, M. J., and Pilone, G. J. (1994). Citrulline production and ethyl carbamate (urethane) precursor formation from arginine degradation by wine lactic acid bacteria *Leuconostoc oenos* and *Lactobacillus buchneri*. *Am. J. Enol. Vitic.* 45, 235–242.
- Liu, S., Pritchard, G. G., Hardman, M. J., and Pilone, G. J. (1995). Occurrence of arginine deiminase pathway enzymes in arginine catabolism by wine lactic

- acid bacteria. *Appl. Environ. Microbiol.* 61:310. doi: 10.1128/AEM.61.1.310-316.1995
- Lombardi, S. J., Pannella, G., Iorizzo, M., Testa, B., Succi, M., and Tremonte, P. (2020). Inoculum strategies and performances of malolactic starter *Lactobacillus plantarum* M10: impact on chemical and sensorial characteristics of Fiano wine. *Microorganisms* 8:516. doi: 10.3390/microorganisms8040516
- Lonvaud-Funel, A. (1999). Lactic acid bacteria in the quality improvement and depreciation of wine. *Antonie Van Leeuwenhoek* 76, 317–331. doi: 10.1023/A:1002088931106
- López-Seijas, J., García-Fraga, B., da Silva, A. F., Zas-García, X., Lois, L. C., and Gago-Martínez, A. (2020). Evaluation of malolactic bacteria associated with wines from Albariño variety as potential starters: screening for quality and safety. *Foods* 9:99. doi: 10.3390/foods9010099
- Lorentzen, M. P. G., and Lucas, P. M. (2019). Distribution of *Oenococcus oeni* populations in natural habitats. *Appl. Microbiol. Biotechnol.* 103, 2937–2945. doi: 10.1007/s00253-019-09689-z
- Lucas, P., Blancato, V., Claisse, O., Magni, C., Lolkema, J., and Lonvaud-Funel, A. (2007). Agmatine deiminase pathway genes in *Lactobacillus brevis* are linked to the tyrosine decarboxylation operon in a putative acid resistance locus. *Microbiology* 153, 2221–2230. doi: 10.1099/mic.0.2007/006320-6320
- Lucio, O., Pardo, I., Heras, J. M., Krieger-Weber, S., and Ferrer, S. (2017). Use of starter cultures of *Lactobacillus* to induce malolactic fermentation in wine. *Aust. J. Grape Wine Res.* 23, 15–21. doi: 10.1111/ajgw.12261
- Lytra, G., Miot-Sertier, C., Moine, V., Coulon, J., and Barbe, J.-C. (2020). Influence of must yeast-assimilable nitrogen content on fruity aroma variation during malolactic fermentation in red wine. *Food Res. Int.* 135:109294. doi: 10.1016/j.foodres.2020.109294
- Madsen, M. G., Edwards, N., Petersen, M. A., Mokwena, L., Swiegers, J. H., and Arneborg, N. (2017). Influence of *Oenococcus oeni* and *Brettanomyces bruxellensis* on hydroxycinnamic acids and volatile phenols of aged wine. *Am. J. Enol. Vitic.* 68, 23–29. doi: 10.5344/ajev.2016.16015
- Matthews, A., Grimaldi, A., Walker, M., Bartowsky, E., Grbin, P., and Jiranek, V. (2004). Lactic acid bacteria as a potential source of enzymes for use in vinification. *Appl. Environ. Biotechnol.* 70:5715. doi: 10.1128/AEM.70.10.5715-5731.2004
- Merin, M. G., Mendoza, L. M., and Ambrosini, V. I. M. D. (2014). Pectinolytic yeasts from viticultural and enological environments: novel finding of *Filobasidium capsuligenum* producing pectinases. *J. Basic Microbiol.* 54, 835–842. doi: 10.1002/jobm.201200534
- Merin, M. G., Mendoza, L. M., Fariás, M. E., Morata, and de Ambrosini, V. I. (2011). Isolation and selection of yeasts from wine grape ecosystem secreting cold-active pectinolytic activity. *Int. J. Food Microbiol.* 147, 144–148. doi: 10.1016/j.ijfoodmicro.2011.04.004
- Michlmayr, H., Eder, R., Kulbe, K. D., and Del Hierro, A. (2012).  $\beta$ -Glycosidase activities of *Oenococcus oeni*: current state of research and future challenges. *Mitt. Klosterneuburg* 62, 87–96.
- Michlmayr, H., Schümann, C., Barreira Braz Da Silva, N. M., Kulbe, K. D., and Del Hierro, A. M. (2010a). Isolation and basic characterization of a  $\beta$ -glucosidase from a strain of *Lactobacillus brevis* isolated from a malolactic starter culture. *J. Appl. Microbiol.* 108, 550–559. doi: 10.1111/j.1365-2672.2009.04461.x
- Michlmayr, H., Schümann, C., Wurbs, P., Barreira Braz, da Silva, N. M., Rogl, V., et al. (2010b). A  $\beta$ -glucosidase from *Oenococcus oeni* ATCC BAA-1163 with potential for aroma release in wine: cloning and expression in *E. coli*. *World J. Microbiol. Biotechnol.* 26, 1281–1289. doi: 10.1007/s11274-009-0299-295
- Mink, R., Kölling, R., Sommer, S., Schmarr, H.-G., and Scharfenberger Schmeier, M. (2015). Diacetyl formation by *Oenococcus oeni* during winemaking induced by exogenous pyruvate. *Am. J. Enol. Vitic.* 66, 85–90.
- Mira and de Orduña, R. (2010). Climate change associated effects on grape and wine quality and production. *Food Res. Int.* 43, 1844–1855.
- Miranda-Castilleja, D. E., Martínez-Peniche, R. Á., Aldrete-Tapia, J. A., Soto-Muñoz, L., Iturriaga, M. H., and Pacheco-Aguilar, J. R. (2016). Distribution of native lactic acid bacteria in wineries of Queretaro, Mexico and their resistance to wine-like conditions. *Front. Microbiol.* 7:1769. doi: 10.3389/fmicb.2016.01769
- Morata, A., Gómez-Cordovés, M., Colomo, B., and Suárez, J. (2005). Cell wall anthocyanin adsorption by different *Saccharomyces* strains during the fermentation of *Vitis vinifera* L. cv Graciano grapes. *Eur. Food Res. Technol.* 220, 341–346. doi: 10.1007/s00217-004-1053-1058
- Morata, A., Vejarano, R., Ridolfi, G., Benito, S., Palomero, F., and Uthurry, C. (2013). Reduction of 4-ethylphenol production in red wines using HCDC+ yeasts and cinnamyl esterases. *Enzyme Microb. Technol.* 52, 99–104. doi: 10.1016/j.enzymitec.2012.11.001
- Moslehisshad, M., Mirdamadi, S., Ehsani, M. R., Ezzatpanah, H., and Moosavi-Movahedi, A. A. (2013). The proteolytic activity of selected lactic acid bacteria in fermenting cow's and camel's milk and the resultant sensory characteristics of the products. *Int. J. Dairy Technol.* 66, 279–285. doi: 10.1111/1471-0307.12017
- Mozzi, F., Raya, R. R., and Vignolo, G. M. (2015). *Biotechnology of Lactic Acid Bacteria: Novel Applications*, 2nd edn., eds F. Mozzi, R. R. Raya, and G. M. Vignolo (Iowa: Wiley-Blackwell). doi: 10.1002/9780813820866.ch6
- Muhalidin, B., and Saari, N. (2020). Review on the biological detoxification of mycotoxins using lactic acid bacteria to enhance the sustainability of foods supply. *Molecules* 25:2655. doi: 10.3390/molecules25112655
- Muhlack, R., Nordestgaard, S., Waters, E. J., O' Neill, B. K., Lim, A., et al. (2006). In-line dosing for bentonite fining of wine or juice: contact time, clarification, product recovery and sensory effects. *Aust. J. Grape Wine Res.* 12, 221–234. doi: 10.1111/j.1755-0238.2006.tb00062.x
- Nikolaou, A., Sgouros, G., Mitropoulou, G., Santarmaki, V., and Kourkoutas, Y. (2020). Freeze-dried immobilized kefir culture in low alcohol winemaking. *Foods* 9:115. doi: 10.3390/foods9020115
- Osborne, J. P., Mira, de Orduña, R., Pilone, G. J., and Liu, S.-Q. (2000). Acetaldehyde metabolism by wine lactic acid bacteria. *FEMS Microbiol. Lett.* 191, 51–55. doi: 10.1016/S0378-1097(00)00369-364
- Polo, L., Mañes-Lázaro, R., Olmeda, I., Cruz-Pio, L. E., Medina, Á, Ferrer, S., et al. (2017). Influence of freezing temperatures prior to freeze-drying on viability of yeasts and lactic acid bacteria isolated from wine. *J. Appl. Microbiol.* 122, 1603–1614. doi: 10.1111/jam.13465
- Pretorius, I. S., and Rensburg, P. V. (2000). Enzymes in winemaking: harnessing natural catalysts for efficient biotransformations a review. *S. Afr. J. Enol. Vitic.* 21, 52–73.
- Pretorius, N., Engelbrecht, L., and Du Toit, M. (2019). Influence of sugars and pH on the citrate metabolism of different lactic acid bacteria strains in a synthetic wine matrix. *J. Appl. Microbiol.* 127, 1490–1500. doi: 10.1111/jam.14401
- Renault, P. (2002). Genetically modified lactic acid bacteria: applications to food or health and risk assessment. *Biochimie* 84, 1073–1087. doi: 10.1016/S0300-9084(02)00029-29
- Rentzsch, M., Schwarz, M., and Winterhalter, P. (2007). Pyranoanthocyanins – an overview on structures, occurrence, and pathways of formation. *Trends Food Sci. Technol.* 18, 526–534. doi: 10.1016/j.tifs.2007.04.014
- Ribéreau-Gayon, P., Dubourdieu, D., Donèche, B., and Lonvaud, A. (2006a). *Handbook of Enology, Volume 1: The Microbiology of Wine and Vinifications*, 2nd edn., eds P. Ribéreau-Gayon, D. Dubourdieu, B. Donèche, and A. Lonvaud (Chichester, UK: John Wiley & Sons).
- Ribéreau-Gayon, P., Glories, Y., Maujean, A., and Dubourdieu, D. (2006b). *Handbook of Enology, Volume 2: The Chemistry of Wine – Stabilization and Treatments*, 2nd edn., eds P. Ribéreau-Gayon, D. Dubourdieu, B. Donèche, and A. Lonvaud (Chichester, UK: John Wiley & Sons).
- Rollie, C., Chevallereau, A., Watson, B. N. J., Chyou, T.-Y., Fradet, O., and McLeod, I. (2020). Targeting of temperate phages drives loss of type I CRISPR–Cas systems. *Nature* 578, 149–153. doi: 10.1038/s41586-020-1936-1932
- Rosenberg, S. M., and Hastings, P. J. (2003). Modulating mutation rates in the wild. *Science* 300, 1382–1383. doi: 10.1126/science.1085691
- Ruiz Rodríguez, L. G., Mohamed, F., Bleckwedel, J., Medina, R., De Vuyst, L., Hebert, E. M., et al. (2019). Diversity and functional properties of lactic acid bacteria isolated from wild fruits and flowers present in northern Argentina. *Front. Microbiol.* 10:1091. doi: 10.3389/fmicb.2019.01091
- Sandri, I. G., Fontana, R. C., Barfknecht, D. M., and da Silveira, M. M. (2011). Clarification of fruit juices by fungal pectinases. *Food Sci. Technol.* 44, 2217–2222. doi: 10.1016/j.lwt.2011.02.008
- Santivarangkna, C., Kulozik, U., and Foerst, P. (2007). Alternative drying processes for the industrial preservation of lactic acid starter cultures. *Biotechnol. Prog.* 23, 302–315. doi: 10.1021/bp060268f
- Savijoki, K., Ingmer, H., and Varmanen, P. (2006). Proteolytic systems of lactic acid bacteria. *Appl. Microbiol. Biotechnol.* 71, 394–406. doi: 10.1007/s00253-006-0427-421
- Shao-Yang, W., Hai-Zhen, Z., Yi-Bin, L., Ruo-Jin, L., Ya-Ran, L., and Bo-Lin, Z. (2020). Modifications of phenolic compounds, biogenic amines, and volatile compounds in Cabernet Gernischt wine through malolactic fermentation by *Lactobacillus plantarum* and *Oenococcus oeni*. *Fermentation* 6:15. doi: 10.3390/fermentation6010015

- Silva, I., Campos, F. M., Hogg, T., and Couto, J. A. (2011). Wine phenolic compounds influence the production of volatile phenols by wine-related lactic acid bacteria. *J. Appl. Microbiol.* 111, 360–370. doi: 10.1111/j.1365-2672.2011.05052.x
- Simó, G., Vila-Crespo, J., Fernández-Fernández, E., Ruipérez, V., and Rodríguez-Nogales, J. M. (2017). Highly efficient malolactic fermentation of red wine using encapsulated bacteria in a robust biocomposite of silica-alginate. *J. Agric. Food Chem.* 65, 5188–5197. doi: 10.1021/acs.jafc.7b01210
- Sorek, R., Kunin, V., and Hugenholtz, P. (2008). CRISPR a widespread system that provides acquired resistance against phages in bacteria and archaea. *Nat. Rev. Microbiol.* 6, 181–186. doi: 10.1038/nrmicro1793
- Spano, G., Rinaldi, A., Ugliano, M., Moio, L., Beneduce, L., and Massa, S. (2005). A  $\beta$ -glucosidase gene isolated from wine *Lactobacillus plantarum* is regulated by abiotic stresses. *J. Appl. Microbiol.* 98, 855–861. doi: 10.1111/j.1365-2672.2004.02521.x
- Spath, K., Heintl, S., and Grabherr, R. (2012). Direct cloning in *Lactobacillus plantarum*: electroporation with non-methylated plasmid DNA enhances transformation efficiency and makes shuttle vectors obsolete. *Microb. Cell Fact.* 11:141. doi: 10.1186/1475-2859-11-141
- Strickland, M. T., Schopp, L. M., Edwards, C. G., and Osborne, J. P. (2016). Impact of *Pediococcus* spp. on Pinot noir wine quality and growth of *Brettanomyces*. *Am. J. Enol. Vitic.* 67, 188–198. doi: 10.5344/ajev.2015.15011
- Styger, G., Prior, B., and Bauer, F. F. (2011). Wine flavor and aroma. *J. Ind. Microbiol. Biotechnol.* 38, 1145–1159. doi: 10.1007/s10295-011-1018-1014
- Sumby, K. M., Grbin, P. R., and Jiranek, V. (2010). Microbial modulation of aromatic esters in wine: current knowledge and future prospects. *Food Chem.* 121, 1–16. doi: 10.1016/j.foodchem.2009.12.004
- Sumby, K. M., Grbin, P. R., and Jiranek, V. (2012). Characterization of EstCO8 and EstC34, intracellular esterases, from the wine-associated lactic acid bacteria *Oenococcus oeni* and *Lactobacillus hilgardii*. *J. Appl. Microbiol.* 114, 413–422. doi: 10.1111/jam.12060
- Sumby, K. M., Jiranek, V., and Grbin, P. R. (2013). Ester synthesis and hydrolysis in an aqueous environment, and strain specific changes during malolactic fermentation in wine with *Oenococcus oeni*. *Food Chem.* 141, 1673–1680. doi: 10.1016/j.foodchem.2013.03.087
- Sumby, K., Bartle, L., Grbin, P., and Jiranek, V. (2019). Measures to improve wine malolactic fermentation. *Appl. Microbiol. Biotechnol.* 103, 2033–2051. doi: 10.1007/s00253-018-09608-9608
- Sumby, K., Grbin, P., and Jiranek, V. (2014). Implications of new research and technologies for malolactic fermentation in wine. *Appl. Microbiol. Biotechnol.* 98, 8111–8132. doi: 10.1007/s00253-014-5976-5970
- Sun, S., Jiang, D., Fan, M., Li, H., Jin, C., and Liu, W. (2020). Selection of a versatile *Lactobacillus plantarum* for wine production and identification and preliminary characterisation of a novel histamine-degrading enzyme. *Int. J. Food Sci. Technol.* 55, 2608–2618. doi: 10.1111/ijfs.14514
- Sun, Z., Harris, H. M. B., Mccann, A., Guo, C., Argimón, S., and Zhang, W. (2015). Expanding the biotechnology potential of lactobacilli through comparative genomics of 213 strains and associated genera. *Nat. Commun.* 6:8322. doi: 10.1038/ncomms9322
- Swiegers, J. H., Bartowsky, E. J., Henschke, P. A., and Pretorius, I. S. (2005). Yeast and bacterial modulation of wine aroma and flavour. *Aust. J. Grape Wine Res.* 11, 139–173. doi: 10.1111/j.1755-0238.2005.tb00285.x
- Szutowska, J. (2020). Functional properties of lactic acid bacteria in fermented fruit and vegetable juices: a systematic literature review. *Eur. Food Res. Technol.* 246, 357–372. doi: 10.1007/s00217-019-03425-3427
- Takase, H., Sasaki, K., Kiyomichi, D., Kobayashi, H., Matsuo, H., and Takata, R. (2018). Impact of *Lactobacillus plantarum* on thiol precursor biotransformation leading to production of 3-sulfanylhexan-1-ol. *Food Chem.* 259, 99–104. doi: 10.1016/j.foodchem.2018.03.116
- Tavernini, L., Ottone, C., Illanes, A., and Wilson, L. (2020). Entrapment of enzyme aggregates in chitosan beads for aroma release in white wines. *Int. J. Biol. Macromol.* 154, 1082–1090. doi: 10.1016/j.ijbiomac.2020.03.031
- Teresa Alegre, M., Carmen Rodriguez, M., and Mesas, J. M. (2004). Transformation of *Lactobacillus plantarum* by electroporation with in vitro modified plasmid DNA. *FEMS Microbiol. Lett.* 241, 73–77. doi: 10.1016/j.femsle.2004.10.006
- Teusink, B., Wiersma, A., Jacobs, L., Notebaart, R. A., and Smid, E. J. (2009). Understanding the adaptive growth strategy of *Lactobacillus plantarum* by in silico optimisation (Predicting Adaptive Evolution in *L. plantarum*). *PLoS Computat. Biol.* 5:e1000410. doi: 10.1371/journal.pcbi.1000410
- Thibon, C., Marullo, P., Claisse, O., Cullin, C., Dubourdieu, D., and Tominaga, T. (2008). Nitrogen catabolic repression controls the release of volatile thiols by *Saccharomyces cerevisiae* during wine fermentation. *FEMS Yeast Res.* 8, 1076–1086. doi: 10.1111/j.1567-1364.2008.00381.x
- Torriani, S., Felis, G. E., and Fracchetti, F. (2011). Selection criteria and tools for malolactic starters development: an update. *Ann. Microbiol.* 61, 33–39. doi: 10.1007/s13213-010-0072-x
- Tristezza, M., Vetrano, C., Bleve, G., Spano, G., Capozzi, V., and Logrieco, A. (2013). Biodiversity and safety aspects of yeast strains characterized from vineyards and spontaneous fermentations in the Apulia regione. *Italy Food Microbiol.* 36, 335–342. doi: 10.1016/j.fm.2013.07.001
- Ugliano, M., and Moio, L. (2005). Changes in the concentration of yeast-derived volatile compounds of red wine during malolactic fermentation with four commercial starter cultures of *Oenococcus oeni*. *J. Agric. Food Chem.* 53, 10134–10139.
- van Pijkeren, J.-P., and Barrangou, R. (2017). Genome editing of food-grade lactobacilli to develop therapeutic probiotics. *Microbiol. Spectr.* 5, 389–408. doi: 10.1128/microbiolspec.BAD-0013-2016
- Van Sluyter, S. C., McRae, J. M., Falconer, R. J., Smith, P. A., Bacic, A., Waters, E. J., et al. (2015). Wine protein haze: mechanisms of formation and advances in prevention. *J. Agric. Food Chem.* 63, 4020–4030. doi: 10.1021/acs.jafc.5b00047
- Varriano, J., (2010). Wine: a cultural history. *SHAD* 25, 184–186. doi: 10.1086/SHAD25010184
- Wade, M. E., Strickland, M. T., Osborne, J. P., and Edwards, C. G. (2018). Role of *Pediococcus* in winemaking. *Aust. J. Grape Wine Res.* 25, 7–24. doi: 10.1111/ajgw.12366
- Wang, J., He, L., An, W., Yu, D., Liu, S., and Shi, K. (2019). Lyoprotective effect of soluble extracellular polymeric substances from *Oenococcus oeni* during its freeze-drying process. *Process. Biochem.* 84, 205–212. doi: 10.1016/j.procbio.2019.05.026
- Waterhouse, A. L., Sacks, G. L., and Jeffery, D. W. (2016). *Understanding Wine Chemistry* (New York: John Wiley & Sons, Incorporated).
- Wells, A., and Osborne, J. P. (2011). Impact of acetaldehyde- and pyruvic acid-bound sulphur dioxide on wine lactic acid bacteria. *Lett. Appl. Microbiol.* 54, 187–194. doi: 10.1111/j.1472-765X.2011.03193.x
- Williams, P. J., Strauss, C. R., Wilson, B., and Massy-Westropp, R. A. (1982). Novel monoterpene disaccharide glycosides of *Vitis vinifera* grapes and wines. *Phytochemistry* 21, 2013–2020. doi: 10.1016/0031-9422(82)83034-83033
- Xin, Y., Guo, T., Mu, Y., and Kong, J. (2018). A single-plasmid genome editing system for metabolic engineering of *Lactobacillus casei*. *Front. Microbiol.* 9:3024. doi: 10.3389/fmicb.2018.03024
- Yang, K., Liu, M., Yang, J., Wei, X., Fan, M., and Zhang, G. (2020). Physiological and proteomic responses of freeze-dried *Oenococcus oeni* SD-2a with ethanol-acclimations. *Food Sci. Technol.* 129:109425. doi: 10.1016/j.lwt.2020.109425
- Zhang, J., Wu, C., Du, G., and Chen, J. (2012). Enhanced acid tolerance in *Lactobacillus casei* by adaptive evolution and compared stress response during acid stress. *Biotechnol. Bioprocess Eng.* 17, 283–289. doi: 10.1007/s12257-011-0346-346
- Zheng, J., Wittouck, S., Salvetti, E., Franz, C. M. A. P., Harris, H. M. B., and Mattarelli, P. (2020). A taxonomic note on the genus *Lactobacillus*: description of 23 novel genera, emended description of the genus *Lactobacillus* Beijerinck 1901, and union of Lactobacillaceae and Leuconostocaceae. *Int. J. Syst. Evol. Microbiol.* 70, 2782–2858. doi: 10.1099/ijsem.0.004107

**Conflict of Interest:** EB was employed by Lallemand Australia.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Viridis, Sumby, Bartowsky and Jiranek. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.