



# Biophysical Characterization of Membrane Phase Transition Profiles for the Discrimination of Outer Membrane Vesicles (OMVs) From *Escherichia coli* Grown at Different Temperatures

## OPEN ACCESS

### Edited by:

Araceli Contreras-Rodriguez,  
National Polytechnic Institute, Mexico

### Reviewed by:

Scott Allen Walper,  
United States Naval Research  
Laboratory, United States  
Medicharla Venkata  
Jagannadham,  
Centre for Cellular and Molecular  
Biology (CCMB), India

### \*Correspondence:

Federico Bordi  
federico.bordi@roma1.infn.it  
Andrea Masotti  
andrea.masotti@opbg.net

† These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Microbial Physiology and Metabolism,  
a section of the journal  
Frontiers in Microbiology

**Received:** 25 November 2019

**Accepted:** 10 February 2020

**Published:** 27 February 2020

### Citation:

Sarra A, Celluzzi A, Bruno SP,  
Ricci C, Sennato S, Ortore MG,  
Cascardi S, Del Chierico F,  
Postorino P, Bordi F and Masotti A  
(2020) Biophysical Characterization  
of Membrane Phase Transition  
Profiles for the Discrimination of Outer  
Membrane Vesicles (OMVs) From  
*Escherichia coli* Grown at Different  
Temperatures.  
*Front. Microbiol.* 11:290.  
doi: 10.3389/fmicb.2020.00290

Angelo Sarra<sup>1†</sup>, Antonella Celluzzi<sup>2†</sup>, Stefania Paola Bruno<sup>2</sup>, Caterina Ricci<sup>3</sup>,  
Simona Sennato<sup>4</sup>, Maria Grazia Ortore<sup>3</sup>, Stefano Cascardi<sup>5</sup>, Federica Del Chierico<sup>6</sup>,  
Paolo Postorino<sup>4</sup>, Federico Bordi<sup>4\*\*</sup> and Andrea Masotti<sup>2\*†</sup>

<sup>1</sup> Department of Science, University of Roma Tre, Rome, Italy, <sup>2</sup> Research Laboratories, Bambino Gesù Children's Hospital, IRCCS, Rome, Italy, <sup>3</sup> Department of Life and Environmental Sciences, Marche Polytechnic University, Ancona, Italy,

<sup>4</sup> CNR-ISC UOS Sapienza and Department of Physics, Sapienza University of Rome, Rome, Italy, <sup>5</sup> Department of Occupational & Environmental Medicine, Epidemiology and Hygiene, National Institute for Insurance Against Accidents at Work (INAIL), Monte Porzio Catone, Italy, <sup>6</sup> Unit of Human Microbiome, Bambino Gesù Children's Hospital, IRCCS, Rome, Italy

Dynamic Light Scattering (DLS), Small Angle X-ray Scattering (SAXS) and Transmission Electron Microscopy (TEM) are physical techniques widely employed to characterize the morphology and the structure of vesicles such as liposomes or human extracellular vesicles (exosomes). Bacterial extracellular vesicles are similar in size to human exosomes, although their function and membrane properties have not been elucidated in such detail as in the case of exosomes. Here, we applied the above cited techniques, in synergy with the thermotropic characterization of the vesicles lipid membrane using a turbidimetric technique to the study of vesicles produced by Gram-negative bacteria (Outer Membrane Vesicles, OMVs) grown at different temperatures. This study demonstrated that our combined approach is useful to discriminate vesicles of different origin or coming from bacteria cultured under different experimental conditions. We envisage that in a near future the techniques employed in our work will be further implemented to discriminate complex mixtures of bacterial vesicles, thus showing great promises for biomedical or diagnostic applications.

**Keywords:** outer membrane vesicle, dynamic light scattering, transmission electron microscopy, small-angle X-ray scattering, gram negative bacteria

## INTRODUCTION

In all domains of life Eukarya, Archaea, and Bacteria produce and release membrane vesicles for reasons that are still not completely understood (Deatherage and Cookson, 2012). In humans, many cells such as dendritic cells, lymphocytes, and tumor cells actively release (i.e. by exocytosis) small (~30–100 nm of diameter) membrane vesicles, referred as exosomes, into biofluids

(i.e. plasma/serum, urine, cerebrospinal fluid, and saliva). These vesicles are powerful cell-to-cell messengers as they transfer lipids, proteins, DNA, and ribonucleic acids (i.e. mRNA, microRNA, lncRNA, and other RNA species) between cells (Valadi et al., 2007; Simpson et al., 2009; Chen et al., 2012). In the last few years, exosomes and their inner content have been also exploited as innovative and effective biomarkers for the diagnosis of many different diseases (i.e. tumors, obesity, gastrointestinal disorders, fibromyalgia, etc.) (Taylor and Gercel-Taylor, 2008; Logozzi et al., 2009; Felli et al., 2017; Masotti et al., 2017).

Similarly to human cells, either Gram-negative and Gram-positive bacteria produce extracellular vesicles, referred as Outer-membrane vesicles (OMVs) and membrane vesicles (MVs), respectively. The production and release of vesicles by bacteria is a natural process and it is necessary for inter-species (bacteria-bacteria) and inter-kingdom (bacteria-host) interactions (Leitão and Enguita, 2016).

One of the main structural differences between Gram-positive and Gram-negative bacteria relies on the composition of their cell envelope. In both cases, the bacterial envelope comprises the plasma membrane and a layer of peptidoglycans. However, in the case of Gram-negative bacteria an additional layer (i.e. the outer membrane) is also observed. The space between the cytoplasmic membrane and the outer membrane is the periplasmic space or periplasm. The outer membrane is made by negatively charged phospholipids and lipopolysaccharides that confer to the Gram-negative wall an overall negative charge. Owing to the presence of these lipopolysaccharides on the outer membranes, many gram-negative bacteria are often pathogenic.

Gram-negative bacteria (e.g. *Escherichia coli*, *Pseudomonas aeruginosa*, etc.) spontaneously secrete OMVs, small (20–100 nm of diameter) spherical bi-layered vesicles, delivered in a variety of environments, including planktonic cultures, fresh and salt water, biofilms, inside eukaryotic cells and within mammalian hosts (Beveridge et al., 1997; Beveridge, 1999; Hellman et al., 2000; Biller et al., 2014). Gram positive bacteria as well, such as *Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus anthracis*, *Streptomyces coelicolor*, *Listeria monocytogenes*, *Clostridium perfringens*, *Streptococcus mutants*, and *Streptococcus pneumoniae* spontaneously produce MVs (Lee et al., 2009, 2013; Rivera et al., 2010; Schrempf et al., 2011; Jiang et al., 2014; Liao et al., 2014).

Recently, it has been observed that also non-pathogenic bacteria (i.e. probiotics), such as Lactobacilli, release extracellular vesicles. Given the importance and the positive impact that probiotic effects have on human health, the study of Lactobacilli MVs may be an interesting opportunity for various applications, from vaccines to therapeutic delivery (van der Pol et al., 2015; Dean et al., 2017). In this context, a detailed physicochemical characterization of MVs from *Lactobacillus acidophilus* ATCC 53544, *Lactobacillus casei* ATCC 393, and *Lactobacillus reuteri* ATCC 23272, was recently reported (Grande et al., 2017; Dean et al., 2019). Among the Lactobacilli, the *Lactobacillus rhamnosus* GG (LGG), is emerging as an important probiotic strain due to its validated effects in both treating and preventing some gastro-intestinal diseases (Wolvers et al., 2010). Behzadi et al. (2017) revealed the cytotoxic and inhibitor role of

LGG-derived extracellular vesicles on hepatic cancer cells, but the characterization of these vesicles has not still been reported. Moreover, owing to the recent employ of OMVs and MVs as adjuvants in vaccines (van der Pol et al., 2015; Bottero et al., 2016) or as novel vaccines platform (Wang et al., 2018) to regulate host activity processes (Chen et al., 2017; Grandi et al., 2017) or other pathogenic processes (Ellis and Kuehn, 2010), an in depth biophysical characterization (i.e. identification, discrimination and quantification) of these types of vesicles is highly desired for biomedical applications.

Among the most employed biophysical techniques available to characterize such small particles in aqueous suspension, Dynamic Light Scattering (DLS) represents one of the most versatile ones [see for example (Berne and Pecora, 1976)]. DLS is generally used to determine vesicles size but at the same time it provides information about the reorganization of lipid molecules of the membrane bilayer with a turbidimetric method (Michel et al., 2006). Actually, the intensity of light scattered at a fixed angle by particles in a suspension depends on their size and on their optical properties (hence on the structure and on the refractive index of the components). By measuring the time-averaged intensity of scattered light as a function of the temperature, this experiment allows the determination of thermotropic lipid phase transitions (with characteristic transition temperatures  $T_c$ ), related to the lipid organization and their remodeling within the bilayer, which affects the membrane optical properties (refractive index) (Aleandri et al., 2012). Consequently, if different thermotropic behaviors feature different kind of vesicles, DLS experimental set-up can be used as a tool to discriminate them, too. In fact, it was previously known that *E. coli* outer membranes show a phase transition due to conformational changes of their lipid components (Trauble and Overath, 1973). Lipid composition modifications have been also observed in their outer membranes when the environmental conditions are changed (i.e. at growing temperatures) (Marr and Ingraham, 1962; Morein et al., 1996; Mika et al., 2016). A similar behavior can be expected also for OMVs produced by *E. coli* as well.

Therefore, aimed at providing a useful tool to characterize better OMVs and MVs, we propose a solid set of experimental physical techniques, including this turbidimetric approach that is able to discriminate extracellular vesicles originating from bacteria grown at different environmental conditions.

## MATERIALS AND METHODS

### Bacterial Strains and Culturing Conditions

*Escherichia coli* (ATCC 8739) was cultured at three different temperatures: 20°C, 27°C and 37°C in essential M9 microbial growth medium consisting in  $\text{Na}_2\text{HPO}_4$  (6.8 g/L),  $\text{KH}_2\text{PO}_4$  (3 g/L),  $\text{NH}_4\text{Cl}$  (1 g/L) and  $\text{NaCl}$  (0.5 g/L). To M9 medium, a solution containing D-glucose (4 g/L),  $\text{MgSO}_4$  (241 mg/L) and  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$  (15 mg/L) was added and the resulting solution was adjusted to pH = 7. Bacteria were cultured overnight until the culture reached an  $\text{OD}_{600}$  of approximately 1. *Lactobacillus rhamnosus* LGG (ATCC 53103) (Dicoflor 60®, DICOFARM)

was cultured overnight in a De Man, Rogosa and Sharpe medium (SigmaAldrich, MRS broth, 51 g/L) at 37°C in anaerobic conditions up to an OD<sub>600</sub> of approximately 2. Aliquots (5 ml) of the bacterial cultures (both *E. coli* and *L. rhamnosus*) were retained for further analysis and to be used as controls.

To isolate either OMVs or MVs, bacteria were removed from culture media by centrifugation (Beckman Avanti J-25 centrifuge, JA-10 rotor, 6000 × g, 15 min), and the supernatant was filtered through a 0.45 μm filter unit (Sartorius). The supernatant, which contains vesicles, was concentrated by ultrafiltration (Vivaflow 200, Sartorius) up to small volume (50 mL), then filtered through a 0.45 μm filter.

## Isolation of Bacterial Extracellular Vesicles

Bacterial vesicles (OMVs or MVs) were separated by ultracentrifugation (107,000 × g for 3 h at 4°C) (Optima™ XPN-100 Ultracentrifuge, Beckman Coulter, SW 40 Ti-rotor) of the ultrafiltered solution obtained previously. The pellet was washed once in sterile phosphate buffered saline (Dulbecco's Phosphate-Buffered Saline – PBS) and suspended in the same buffer.

At the end of the isolation protocol, a drop of the purified solution was cultured on agar plates at 37°C for one day to exclude the presence of residual bacteria.

The suspension obtained with this method contains a concentrated amount of vesicles (OMVs or MVs) that have been characterized by dynamic light scattering and turbidimetric analyses.

## Dynamic Light Scattering and Turbidimetric Analyses

Dynamic Light Scattering experimental set-up has been employed to provide vesicles size and to characterize the bilayer organization because of its sensitivity to the sample refractive index (Berne and Pecora, 1976). In this case, we use the term “turbidimetric” measurements for this unconventional way of using a light scattering apparatus, although this term is not strictly appropriate, because we observe changes in the refractive index of the lipid membranes, concerning the scattered light and not the transmitted light, as in conventional turbidimetric measurements.

Dynamic Light Scattering and turbidimetric measurements were performed employing a MALVERN Nano Zetasizer apparatus equipped with a 5 mW HeNe laser (Malvern Instruments LTD, United Kingdom). This system uses backscatter detection, i.e. the scattered light is collected at 173°. To obtain the size distribution, the measured autocorrelation functions were analyzed using the CONTIN algorithm (Provencher, 1982). Decay times are used to determine the distribution of the diffusion coefficients  $D_0$  of the particles, which in turn can be converted in a distribution of apparent hydrodynamic diameter,  $D_h$ , using the Stokes-Einstein relationship  $D_h = k_B T / 3\pi\eta D_0$ , where  $k_B$  is the Boltzmann constant,  $T$  the absolute temperature and  $\eta$  the solvent viscosity (Berne and Pecora, 1976). The values of the

radii shown in this work correspond to the average values on several measurements and are obtained from intensity weighted distributions (Provencher, 1982; De Vos et al., 1996). The thermal protocol used for both OMVs, MVs and bacteria in DLS measurements consists of an ascending ramp from 10 to 45°C with temperature step of 1°C. At each step, samples were left to thermalize at the target temperature for few minutes. In particular, for samples grown at 37, 27, and 20°C the thermalization times were 360, 720, and 1080 s, respectively.

For the turbidimetric determination of vesicles' membranes thermotropic behavior, we measured the mean count rate of scattered photons (i.e. the time-averaged intensity of the scattered light),  $I$ , versus temperature,  $T$ , and fitted the data to a Boltzmann sigmoidal curve:

$$I = I_0 + \frac{I_1 - I_0}{1 + \exp\left(\frac{T - T_0}{\Delta T}\right)} \quad (1)$$

where the fitting parameters are  $I_0$  and  $I_1$ , the minimum and maximum intensity, respectively, the transition temperature  $T_0$  and the so called ‘slope’  $\Delta T$  that describes the steepness of the curve.  $\Delta T$  roughly measures the transition width.

## Small Angle X-ray Scattering

Small Angle X-ray Scattering (SAXS) experiments were performed at the Austrian SAXS beamline of the Elettra Synchrotron in Trieste, Italy (Amenitsch et al., 1995). SAXS images were recorded with a 2D pixel detector Pilatus3 1M spanning the Q-range between 0.1 and 6 nm<sup>-1</sup>, with  $Q = 4\pi\sin(2\Theta)/\lambda$ , where  $2\Theta$  is the scattering angle and  $\lambda = 0.0995$  nm the X-ray wavelength. The image conversion to one-dimensional SAXS pattern was performed with FIT2D (Hammersley, 2004). Detector calibration was performed with silver behenate powder (d-spacing = 0.05838 nm). Samples were held in a 1 mm glass capillary (Hilgenberg, Malsfeld, Germany). Experiments were carried out between 10 and 45°C, by using the same heating rate already adopted in DLS experiments. Each measurement was performed for 1 s and followed by a dead time of at least 10 s in order to avoid radiation damage. We obtained a set of SAXS curves resulting from the average of several overlapping SAXS spectra obtained at about the same temperature ( $\pm 1^\circ\text{C}$ ), in order to analyze data with an improved signal-to-noise ratio. SAXS data were fitted according to a core-shell model (Berndt et al., 2005) by GENFIT software (Spinozzi et al., 2014), and taking into account the vesicles polydispersion, in agreement with the DLS results. The fitting parameters are OMVs radius  $R$  and thickness  $d$ , as well as the electron densities of the core and of the shell,  $\rho_i$  and  $\rho_e$ , respectively.

## Transmission Electron Microscopy

Transmission electron microscopy (TEM) was used for the morphological characterization of OMVs and MVs. TEM images were performed at Department of Occupational and Environmental Medicine, Epidemiology and Hygiene of INAIL-Research in Rome. Samples for TEM measurements were prepared at room temperature by depositing 20 μl of vesicles suspensions on a 300-mesh copper grid for electron microscopy

covered by a thin amorphous carbon film. A negative staining was realized by addition of 10  $\mu\text{l}$  of 2% aqueous phosphotungstic acid (PTA) solution (pH-adjusted to 7.3 using 1 N NaOH). Measurements were carried out by using a FEI TECNAI 12 G2 Twin (FEI Company, Hillsboro, OR, United States), operating at 120 kV and equipped with an electron energy loss filter (Biofilter, Gatan Inc., Pleasanton, CA, United States) and a slow-scan charge-coupled device camera (794 IF, Gatan Inc., Pleasanton, CA, United States).

## Scattering Cross Section Simulations

To better understand the behavior of phase transitions in OMVs and bacteria, Mie scattering simulations were performed using the MiePlot 4.6 simulator by Laven (2003). This simulator allows to calculate the scattered intensity as a function of scattering angle for inhomogeneous spheres where the refractive index is a function of the radius. For simplicity sake, both OMVs and bacteria were described as multiple-shelled spheres. In all calculations, we assumed a wavelength of 633 nm for the incident light and a scattering angle of  $173^\circ$ . For the refractive index of the aqueous core (of both vesicles and bacteria) we used the value  $n_{\text{cytoplasm}} = 1.367$  according to the literature (Marquis, 1973; Choi et al., 2007), whereas for the bacterial cell wall we used  $n_{\text{cellwall}} = 1.455$  (Marquis, 1973). Finally, we assumed that the dependence of the refractive index as a function of the temperature of the lipid membrane (for vesicles and bacteria) was described by a Boltzmann sigmoidal curve (Eq. 1).

## RESULTS AND DISCUSSION

### Isolation of OMVs and MVs

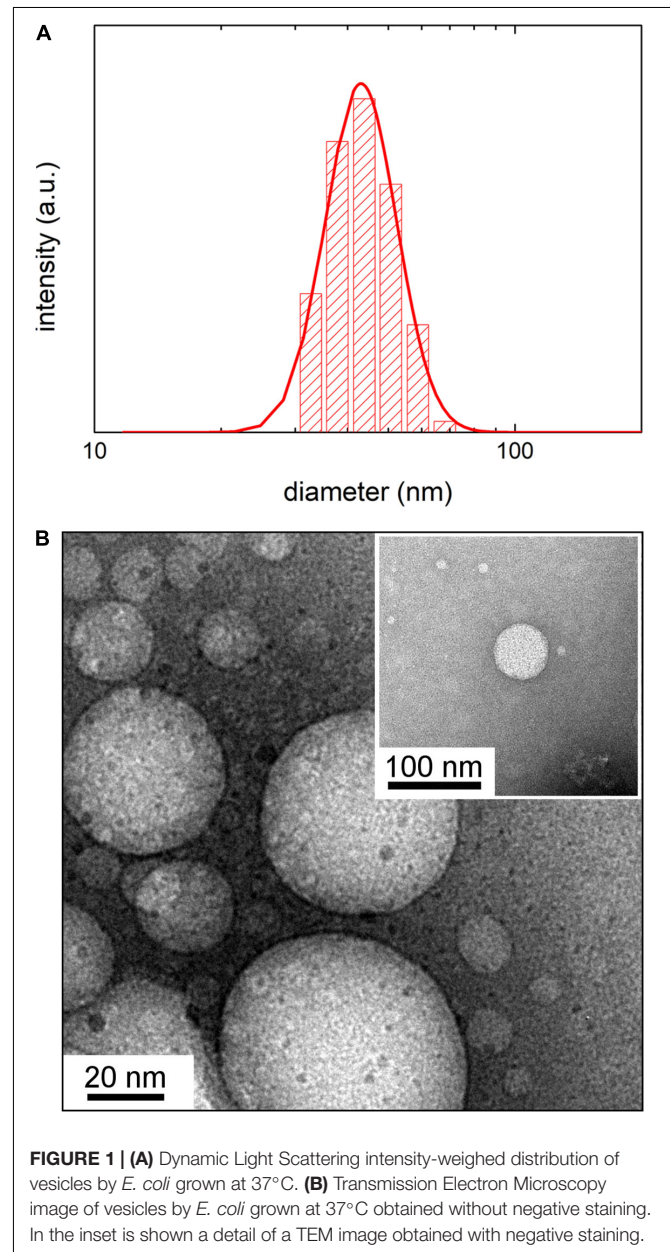
The overall workflow followed for the isolation of bacterial extracellular vesicles comprehensive of the quality control steps has been depicted in **Supplementary Figure S1**. Briefly, to have an enriched solution of bacterial vesicles, we started from a large amount of culture medium that has been first centrifuged and filtered to remove bacteria. Then the tangential flow ultrafiltration procedure allowed us to concentrate the sample up to 100 times and reduce the operating time compared to traditional ultrafiltration techniques. The ultrafiltered solution undergone ultracentrifugation to obtain a transparent pellet of bacterial vesicles that after a washing step afforded a very clean sample without residual proteins contaminants. Our extraction and purification protocol allowed us to obtain pure and concentrated samples of OMVs from cultures of *E. coli* grown at three different temperatures (i.e. 20, 27, and  $37^\circ\text{C}$ ). In fact, following the procedure reported in this work, we were able to reach a concentration of  $\sim 0.5$  mg/ml of proteins (measured by BCA assay) for OMV samples obtained after culturing bacteria at  $37^\circ\text{C}$ ,  $\sim 0.2$  mg/ml for OMVs obtained at  $27^\circ\text{C}$ , and  $\sim 0.1$  mg/ml for OMVs obtained at  $20^\circ\text{C}$ . Similar procedures were adopted for the isolation of MVs from LGG.

### Morphological Characterization

Dynamic Light Scattering intensity weighed distribution (**Figure 1**) and TEM analysis of OMVs from *E. coli* grown at

$37^\circ\text{C}$  (inset of **Figure 1**) revealed that vesicles have a spherical shape with a mean hydrodynamic diameter of  $48 \pm 3$  nm (FWHM =  $24 \pm 2$  nm). Notably, the DLS analysis revealed that the vesicle size decreases as the growth temperature is lowered (**Table 1**). OMVs grown at 27 and  $20^\circ\text{C}$  have a mean diameter of  $37 \pm 4$  nm (FWHM =  $32 \pm 2$  nm) and  $24 \pm 2$  nm (FWHM =  $20 \pm 3$  nm), respectively. Typical TEM images and DLS intensity weighed distributions for vesicles growth at 27 and  $20^\circ\text{C}$  are reported in **Supplementary Figure S2**.

It is known that *E. coli* can undergo remarkable modifications (i.e. conformational and/or compositional changes of the lipid components) of the outer membrane as a function of the different environmental conditions such as the growth temperature. Therefore, to investigate if the size reduction of OMVs as a



**FIGURE 1 | (A)** Dynamic Light Scattering intensity-weighted distribution of vesicles by *E. coli* grown at  $37^\circ\text{C}$ . **(B)** Transmission Electron Microscopy image of vesicles by *E. coli* grown at  $37^\circ\text{C}$  obtained without negative staining. In the inset is shown a detail of a TEM image obtained with negative staining.

**TABLE 1** | Mean hydrodynamic diameter and Full Width Half Maximum (FWHM) of DLS intensity weighed size distribution obtained by CONTIN analysis of correlation function measured at 23°C of OMVs by *Escherichia coli* growth at different temperature; reported errors are the standard deviation of three measurements.

Growth temperature (°C)	Mean diameter (nm)	FWHM (nm)
37	48 ± 3	24 ± 2
27	37 ± 4	32 ± 2
20	24 ± 2	20 ± 3

function of the growing temperature could be related to a different conformational state of the lipids that form the vesicle itself or their remodeling within the bilayer, we performed turbidimetric measurements on OMV samples extracted from bacteria grown at the same temperatures.

### Turbidimetric Experiments

The intensity of light scattered at a fixed angle by the particles in a suspension depends on their size, geometry and optical properties (structure and refractive index of the components). In particular, the refractive index of a lipid bilayer depends on the conformational state of the lipids and on their ordering in the bilayer. Hence, by measuring the time-averaged intensity of scattered light as a function of the temperature, the instrument allowed us to determine the thermotropic lipid phase transitions (with characteristic transition temperatures  $T_c$ ), related to the lipid organization and their remodeling within the bilayer (Michel et al., 2006; Aleandri et al., 2012). We used the term “turbidimetric measurements” for this unconventional way of using a light scattering apparatus although this term is not strictly appropriate, since in the present case to reveal possible changes in the refractive index of the lipid membranes as a function of temperature, the scattered light and not the transmitted light, as in conventional turbidimetric measurements, is monitored. Turbidimetric measurements were performed in the temperature range 10–45°C on OMVs samples and on bacteria grown at 37, 27, and 20°C.

### Characterization of OMVs Extracted From *E. coli* Cultured at 37°C

In **Figure 2** the results obtained from turbidimetric measurements for vesicles (**Figure 2A**) and *E. coli* (**Figure 2B**) (as comparison) grown at 37°C have been reported. As far as the vesicles are concerned, the mean scattering intensity (**Figure 2A**, upper panel) decreased significantly (about 25%) when the temperature was increased. We determined a transition temperature by fitting a Boltzmann sigmoidal curve [Eqn. 1] to the experimental data (red line in **Figure 2A**, upper panel). The fitting procedure provided a transition temperature of  $32 \pm 2^\circ\text{C}$ , where the uncertainty is the fitted slope  $\Delta T$ .

The observed change of the scattered intensity could be ascribed to different factors (Berne and Pecora, 1976) (i.e. to a decrease of the concentration and/or the size of the suspended particles) or to a variation of vesicles refractive index (Michel et al., 2006). However, we did not observe any phase separation (flocculation or precipitation of the particles) and the hydrodynamic diameter of the particles determined by

DLS (lower panel of **Figure 2A**) did not show any appreciable dependence on temperature. Hence, we ascribed the observed decrease of the scattered intensity to a change of the refractive index of the particles, likely due to a phase transition of the membrane lipid bilayer or, more in general, to a structural reorganization of the membrane (Berne and Pecora, 1976). **Figure 2B** shows the results obtained from a suspension of *E. coli*. Again, also with bacteria a rather steep variation of the scattered intensity was observed within the same temperature range observed in the case of OMVs suggesting the occurrence of a similar membrane transition. However, in the case of *E. coli* the scattered intensity increased, whereas in the case of OMVs decreased. Although the scattered intensity measurements were unreliable above 35°C (owing to the proliferation of bacteria that altered the optical properties of the suspension), a transition temperature of  $30 \pm 3^\circ\text{C}$  was obtained (**Figure 2B**). We attributed the opposite behavior of the scattered intensity to the different structures of the bacteria cell wall and, as a consequence, of the OMVs membrane.

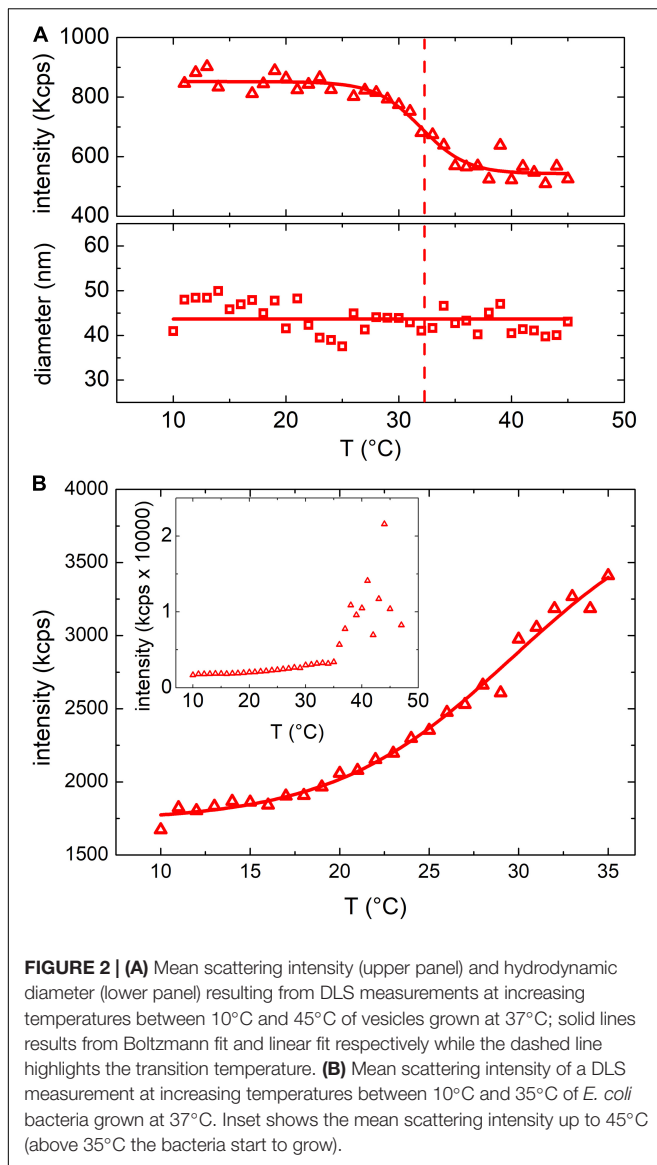
### Characterization of OMVs Extracted From *E. coli* Cultured at 27°C and 20°C

Similar results were obtained also for samples grown at 27 and 20°C. At these growth temperatures, OMVs were smaller (diameters were  $37 \pm 4$  and  $24 \pm 2$  nm, respectively, compared to  $48 \pm 3$  nm at 37°C) and their concentration was found to be systematically lower than that obtained for the samples grown at 37°C. As a consequence, also the mean scattering intensity was significantly lower. Nevertheless, also in these conditions the trend of the registered data points suggested the presence of a phase transition. In **Figure 3** we compared the scattering intensity of vesicles (**Figure 3A**) and bacteria (**Figure 3B**) grown at different temperatures. For each sample we reported the normalized scattering intensity, the fitting curve (solid line) and the value of the fitted transition temperature. All the samples analyzed shown a similar behavior in terms of phase transitions. Interestingly, for both OMVs and bacteria a strong correlation between the growth temperature at which *E. coli* was cultured and the transition temperature of the membrane was found (**Figure 4**).

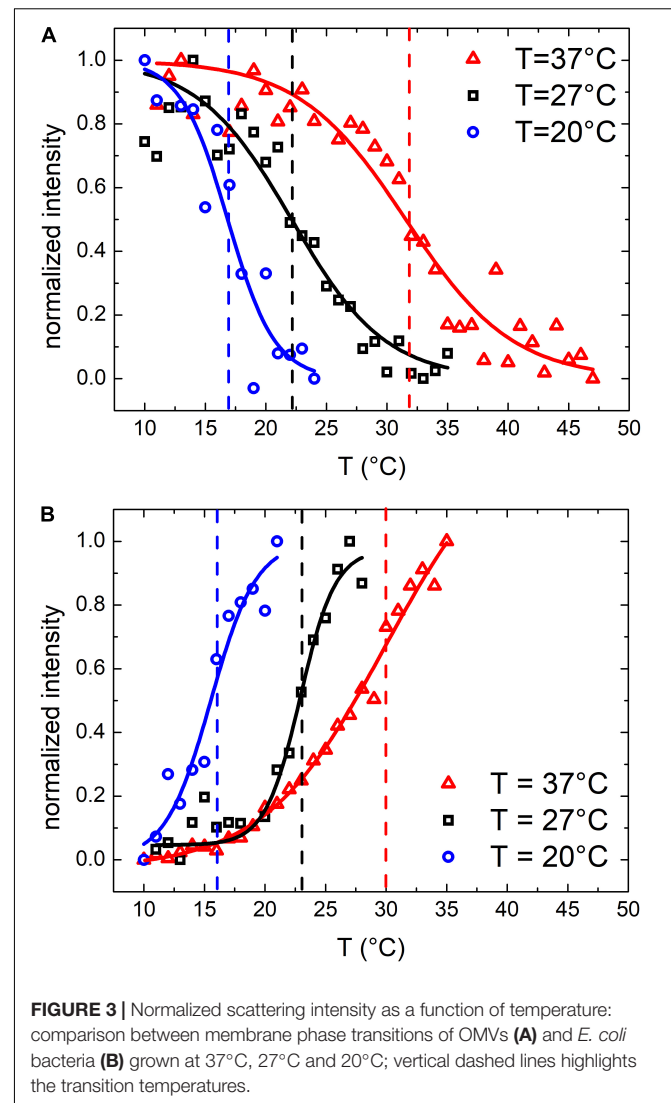
From previous *E. coli* studies, we know that by reducing the temperature of the growth medium the content of unsaturated lipids, which confer a higher flexibility to the membrane, increases consequently. Moreover, bacteria membranes show thermotropic phase transitions whose characteristic transition temperature depends on the lipid composition (Trauble and Overath, 1973) which in turn is related to the growth temperature (Morein et al., 1996). It is reasonable to think that OMVs, that originate from the bacterial outer membrane, may have a similar composition and similar thermotropic behaviors.

### Simulations of Bacteria and OMVs Scattering Cross Section

To better understand the different optical response observed for vesicles and bacteria, we calculated the scattering cross section

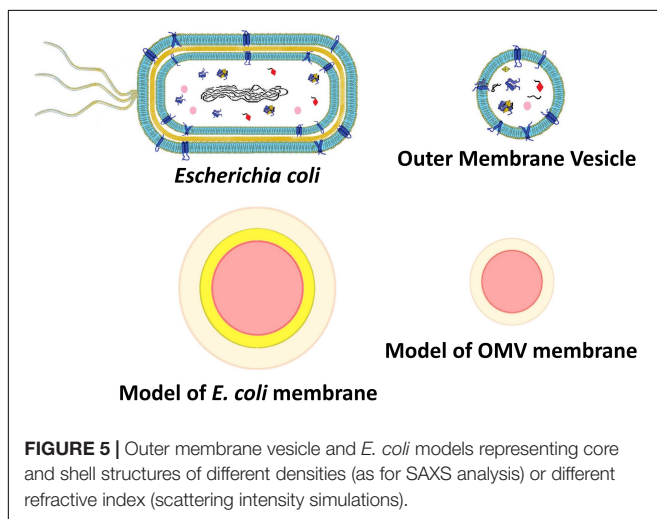
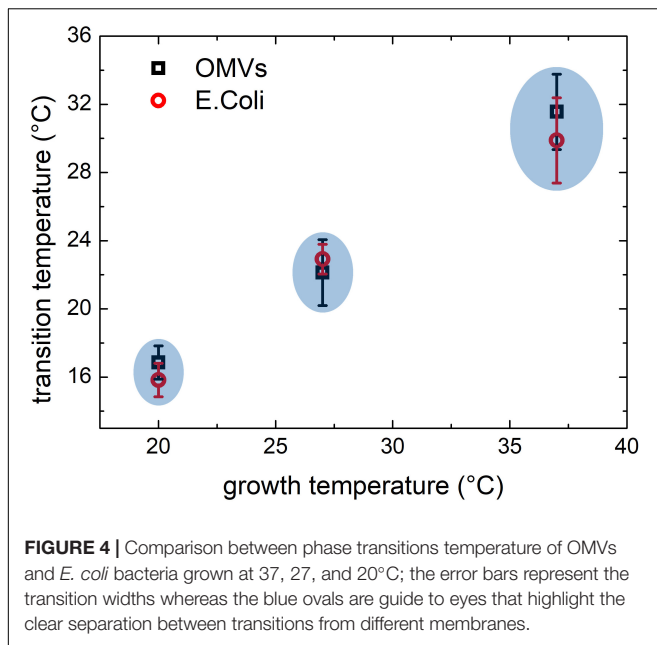


of different structures simulating vesicles and bacteria by using the Mie theory (Laven, 2003). Vesicles and bacteria have been represented and modeled as shown in **Figure 5**. In particular, OMVs have been modeled as a single-shelled sphere, where the core is the cytoplasm (assumed homogeneous) and the shell represents the lipid membrane, whereas in the case of *E. coli* also a second (internal) shell is added, which represents the cell wall, mainly composed of peptidoglycans. To take into account the thermotropic behavior and evaluate the refractive index  $n(T)$  of the lipid bilayer, we assumed the Boltzmann function dependence on the temperature (see Eqn. 1) with the  $T_0$  and  $\Delta T$  values determined from experimental turbidimetric measurements. We also imposed the ratio of the asymptotic values of the refractive index ( $n_0/n_1$ ) equal to the ratio of the measured intensities ( $I_0/I_1$ ). Finally, we constrained the Boltzmann curve to assume the value of 1.44 at  $T = 23^\circ\text{C}$  according to the literature (White et al., 2000).



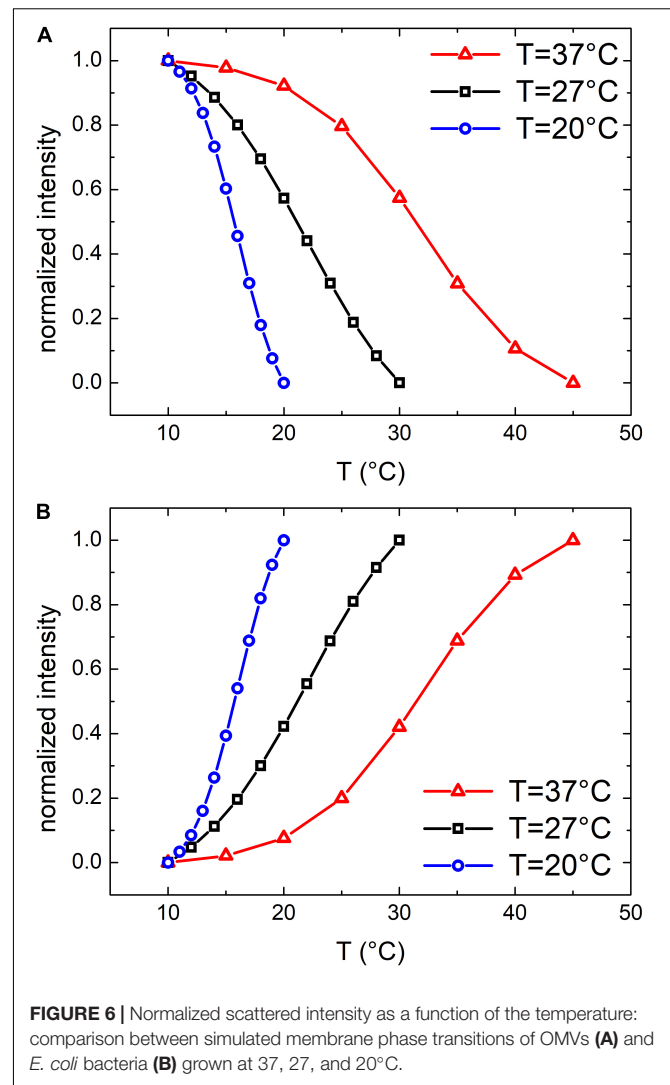
The results of the calculations obtained using MiePlot 4.6 simulator were reported in **Figure 6**. In particular, **Figure 6A** showed the transitions of OMV models, whereas **Figure 6B** the corresponding curves calculated for the *E. coli* models. Notably, the simulations clearly confirmed not only the temperature-dependent transition trend but also the opposite scattering intensity behavior of OMVs and bacteria, as we previously observed experimentally. Simulations gave us the possibility to identify the reason behind the transition differences observed in the case of OMVs and bacteria and ascribe this behavior to the presence of a second ‘layer’ beneath the outer membrane (the cell wall comprising the peptidoglycan layer) with a fixed refractive index. In fact, this is the only difference between these two models (at least from the optical point of view).

This strongly support the concept the trend of the transition temperature may represent a characteristic “fingerprint” of OMVs and this behavior could be employed as a characteristic “marker” to distinguish vesicles from the bacteria that originated them.

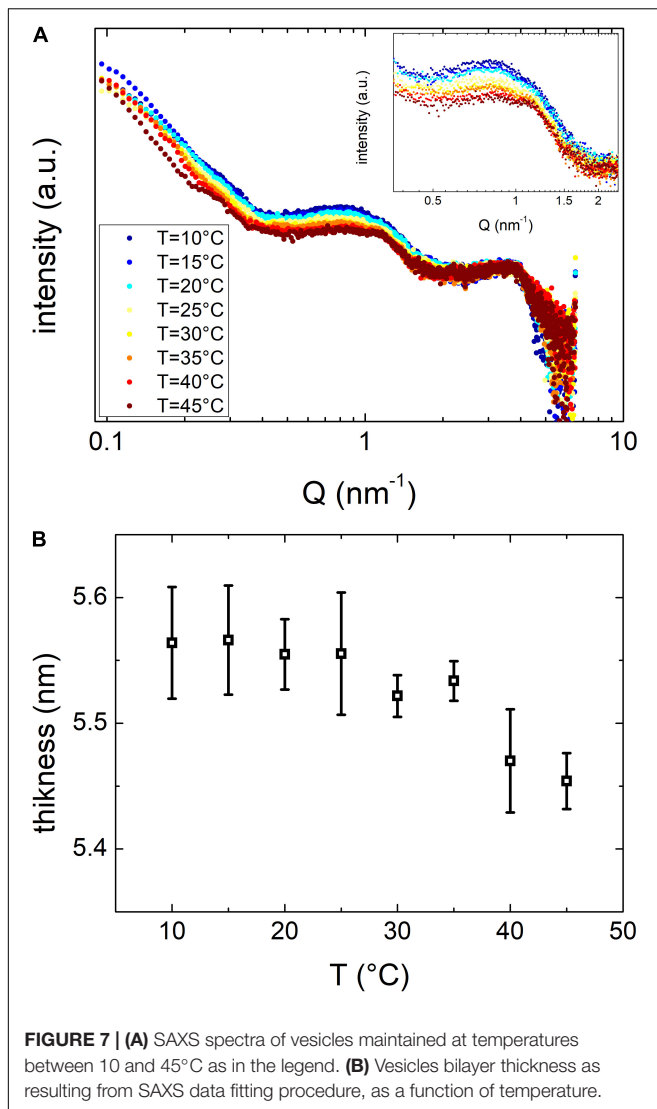


## Small Angle X-ray Scattering Experiments

To investigate in more details the effects of the temperature on OMVs structure we performed SAXS experiments. Due to the low concentration of vesicles obtained at 27 and 20°C, we performed these measurements only on vesicles grown at 37°C. We reported SAXS patterns in **Figure 7A**. The overall SAXS pattern, which is typical of shelled spheres, did not change significantly when the temperature increased. These data confirmed that OMVs size did not vary in the temperature range investigated and no membrane damages were reported. However, the smooth peak centered at  $Q \approx 1 \text{ nm}^{-1}$  (inset of **Figure 7A**) is related to the thickness of the membrane and has been already detected by SAXS on Alix-positive exosome-like small extracellular vesicles (Romancino et al., 2018). This peak



significantly changed the shape and moved toward higher  $Q$  values at increasing temperatures, although this variation was modest. In order to better understand the significance of this peak position shift, SAXS curves were analyzed considering the simplified model of OMVs reported in **Figure 5**. Although OMVs membrane contain a mixture of different lipids (i.e. sterols, polyisoprenoids, etc.) (Bramkamp and Lopez, 2015) and proteins, this simple model appeared to be able to reproduce reasonably our SAXS data. The thickness as a function of temperature obtained from the fitting procedure was reported in **Figure 7B**. The radius and the electron densities as a function of temperature were reported in the **Supplementary Figure S3**. The thickness of the OMVs membrane changed significantly as the temperature increased, whereas vesicles size was not affected by temperature, remained intact in this temperature range as indicated by the core electron density as well as the shell's one that remained constant. These findings suggested that temperature was able to induce a general rearrangement of the membrane structural motifs

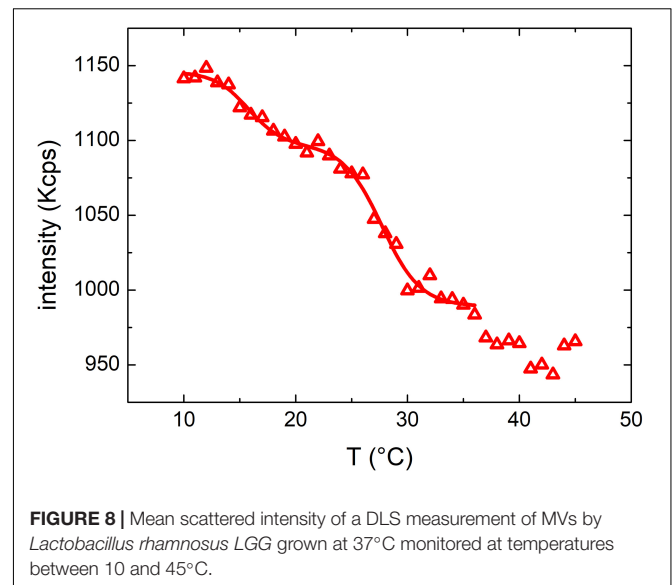


and that these motifs are linked to the reorganization of the lipid membrane and/or to the conformational changes of its components.

## OMVs Have a Distinct Phase Transition Behavior Compared to Other MVs

In order to assess the feasibility of using the phase transition profile to discriminate extracellular vesicles coming from different bacteria (i.e. Gram-negative and Gram-positive bacteria) and potentially use this information for diagnostic purposes in the microbiology field, we decided to measure the phase transition properties of MVs from the Gram-positive LGG comparing the results with those obtained with OMVs from the Gram-negative *E. coli*.

In this specific case, we studied only the phase transition of MVs from LGG by turbidimetric measurements as this technique is easy to reproduce, does not require expensive



equipment, is cheap and also relatively fast. All of these characteristics are highly desirable when thinking to techniques for diagnostics applications.

In **Figure 8** we reported the results of the scattering intensity measurements in the temperature range between 10 and 45°C. By examining the profile of MVs from LGG, we observed the presence of multiple transitions, two of them at a  $T < 35^\circ\text{C}$  and another one likely around 37°C. DLS analysis confirmed that the diameter was  $88 \pm 4$  nm (FWHM =  $110 \pm 10$  nm) and remained constant (as shown in **Supplementary Figure S4**) throughout all the temperature range. Similarly to what observed for OMVs, flocculation or precipitation of MVs did not occur, confirming that the transitions are related only to membrane modifications.

The intensity data were fitted with a double Boltzmann function in the region at  $T < 35^\circ\text{C}$ , providing two transition temperatures of  $16 \pm 2$  and  $28 \pm 3^\circ\text{C}$ . Interestingly, the scattering profile of MVs was completely different than that of OMVs, suggesting that MVs and OMVs have a different lipid composition of the membrane, although these results have been obtained studying vesicles originating only from one Gram positive and one Gram negative bacteria.

## CONCLUSION

In this work, the thermotropic behavior of OMVs and bacteria grown at different temperatures have been studied. The analysis of mean scattering intensity as a function of the temperature has shown the presence of peculiar membrane phase transitions characteristic of both OMVs and bacteria. In particular, the transition temperature that we measured allowed to discriminate not only vesicles of different composition (i.e. OMVs and MVs) but also to correlate them to bacteria that originated them and to their culture conditions (i.e. temperature).



The results we obtained have shown that it is possible to distinguish vesicles coming from a gram-positive (i.e. *E. coli*) and a gram-negative bacteria (i.e. *L. rhamnosus*) by simply studying the phase transitions of their membranes. Although the protein and lipid composition of bacteria tend to change during the culture (i.e. “ageing”), these differences seemed not to interfere in the measurements and did not prevent us to obtain statistically significant differences between OMVs and MVs.

In a close future, by further improving this technique, it could be even possible to perform microbiological analysis by discriminating different bacteria in complex mixtures or matrices (i.e. biological fluids, stool, blood, tissues, etc.). Once implemented and validated on a higher number of samples (i.e. vesicles originating from different Gram negative and Gram positive bacteria), this technique could be also employed in fields such as oncology, for example to discriminate human exosomes originating from tumors from that produced by normal cells and help clinicians to diagnose different form of tumors.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation, to any qualified researcher.

## REFERENCES

- Aleandri, S., Bonicelli, M. G., Bordini, F., Casciardi, S., Diociaiuti, M., Giansanti, L., et al. (2012). How stereochemistry affects the physicochemical features of gemini surfactant based cationic liposomes. *Soft Matter* 8, 5904–5915.
- Amenitsch, H., Bernstorff, S., and Laggner, P. (1995). High-flux beamline for small-angle x-ray scattering at ELETTRA. *Rev. Sci. Instrum.* 66, 1624–1626. doi: 10.1063/1.1145864
- Behzadi, E., Mahmoodzadeh Hosseini, H., and Imani Fooladi, A. A. (2017). The inhibitory impacts of *Lactobacillus rhamnosus* GG-derived extracellular vesicles on the growth of hepatic cancer cells. *Microb. Pathog.* 110, 1–6. doi: 10.1016/j.micpath.2017.06.016
- Berndt, I., Pedersen, J. S., and Richtering, W. (2005). Structure of multiresponsive “intelligent” core-shell microgels. *J. Am. Chem. Soc.* 127, 9372–9373. doi: 10.1021/ja051825h
- Berne, B., and Pecora, R. (1976). *Dynamic Light Scattering*. New York, NY: John Wiley.
- Beveridge, T. J. (1999). Structures of gram-negative cell walls and their derived membrane vesicles. *J. Bacteriol.* 181, 4725–4733. doi: 10.1128/jb.181.16.4725-4733.1999
- Beveridge, T. J., Makin, S. A., Kadurugamuwa, J. L., and Li, Z. (1997). Interactions between biofilms and the environment. *FEMS Microbiol. Rev.* 20, 291–303. doi: 10.1111/j.1574-6976.1997.tb00315.x
- Biller, S. J., Schubotz, F., Roggensack, S. E., Thompson, A. W., Summons, R. E., and Chisholm, S. W. (2014). Bacterial vesicles in marine ecosystems. *Science* 343, 183–186. doi: 10.1126/science.1243457
- Bottero, D., Gaillard, M. E., Zurita, E., Moreno, G., Martinez, D. S., Bartel, E., et al. (2016). Characterization of the immune response induced by pertussis OMVs-based vaccine. *Vaccine* 34, 3303–3309. doi: 10.1016/j.vaccine.2016.04.079
- Bramkamp, M., and Lopez, D. (2015). Exploring the existence of lipid rafts in bacteria. *Microbiol. Mol. Biol. Rev.* 79, 81–100. doi: 10.1128/MMBR.00036-14
- Chen, F., Cui, G., Wang, S., Nair, M. K. M., He, L., Qi, X., et al. (2017). Outer membrane vesicle-associated lipase FtlA enhances cellular invasion and virulence in *Francisella tularensis* LVS. *Emerg. Microbes Infect.* 6:e66. doi: 10.1038/emi.2017.53

## AUTHOR CONTRIBUTIONS

AS performed all the experiments and drafted the manuscript. AC cultured bacteria, helped SB to extract and purify OMVs and together they drafted the manuscript. AS, CR, and MO performed and analyzed SAXS experiments. SS helped AS in DLS experiments. SC helped AS and SS to acquire TEM images. FD suggested the best culturing conditions to grow *E. coli* at different temperatures. FD, SS, and MO edited and critically revised and the manuscript draft. PP, FB, and AM contributed to the experimental plan, coordinated the work, drafted and edited the manuscript, and funded part of the work.

## ACKNOWLEDGMENTS

Prof. Heinz Amenitsch is acknowledged for his precious assistance in SAXS experiments.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2020.00290/full#supplementary-material>

- Chen, X., Liang, H., Zhang, J., Zen, K., and Zhang, C. Y. (2012). Horizontal transfer of microRNAs: molecular mechanisms and clinical applications. *Protein Cell* 3, 28–37. doi: 10.1007/s13238-012-2003-z
- Choi, W., Fang-Yen, C., Badizadegan, K., Oh, S., Lue, N., Dasari, R. R., et al. (2007). Tomographic phase microscopy. *Nat. Methods* 4, 717–719.
- De Vos, C., Deriemaeker, L., and Finsy, R. (1996). Quantitative assessment of the conditioning of the inversion of quasi-elastic and static light scattering data for particle size distributions. *Langmuir* 12, 2630–2636. doi: 10.1021/la950852q
- Dean, S. N., Leary, D. H., Sullivan, C. J., Oh, E., and Walper, S. A. (2019). Isolation and characterization of *Lactobacillus*-derived membrane vesicles. *Sci. Rep.* 9:877. doi: 10.1038/s41598-018-37120-6
- Dean, S. N., Turner, K. B., Medintz, I. L., and Walper, S. A. (2017). Targeting and delivery of therapeutic enzymes. *Ther. Deliv.* 8, 577–595. doi: 10.4155/tde-2017-0020
- Deatherage, B. L., and Cookson, B. T. (2012). Membrane vesicle release in bacteria, eukaryotes, and archaea: a conserved yet underappreciated aspect of microbial life. *Infect. Immun.* 80, 1948–1957. doi: 10.1128/IAI.06014-11
- Ellis, T. N., and Kuehn, M. J. (2010). Virulence and immunomodulatory roles of bacterial outer membrane vesicles. *Microbiol. Mol. Biol. Rev.* 74, 81–94. doi: 10.1128/MMBR.00031-09
- Felli, C., Baldassarre, A., and Masotti, A. (2017). Intestinal and circulating microRNAs in coeliac disease. *Int. J. Mol. Sci.* 18:1907. doi: 10.3390/ijms18091907
- Grande, R., Celia, C., Mincione, G., Stringaro, A., Di Marzio, L., Colone, M., et al. (2017). Detection and physicochemical characterization of membrane vesicles (MVs) of *Lactobacillus reuteri* DSM 17938. *Front. Microbiol.* 8:1040. doi: 10.3389/fmicb.2017.01040
- Grandi, A., Tomasi, M., Zanella, I., Ganfani, L., Caproni, E., Fantappie, L., et al. (2017). Synergistic protective activity of tumor-specific epitopes engineered in bacterial outer membrane vesicles. *Front. Oncol.* 7:253. doi: 10.3389/fonc.2017.00253
- Hammersley, A. P. (2004). *Fit2d*. Available online at: <http://www.esrf.eu/computing/scientific/FIT2D/> (accessed November 1, 2019).
- Hellman, J., Loisel, P. M., Zanzot, E. M., Allaire, J. E., Tehan, M. M., Boyle, L. A., et al. (2000). Release of gram-negative outer-membrane proteins into human serum and septic rat blood and their interactions with immunoglobulin in

- antisera to *Escherichia coli* J5. *J. Infect. Dis.* 181, 1034–1043. doi: 10.1086/315302
- Jiang, Y., Kong, Q., Roland, K. L., and Curtiss, R. III (2014). Membrane vesicles of *Clostridium perfringens* type A strains induce innate and adaptive immunity. *Int. J. Med. Microbiol.* 304, 431–443. doi: 10.1016/j.ijmm.2014.02.006
- Laven, P. (2003). Simulation of rainbows, coronas, and glories by use of Mie theory. *Appl. Opt.* 42, 436–444.
- Lee, E. Y., Choi, D. Y., Kim, D. K., Kim, J. W., Park, J. O., Kim, S., et al. (2009). Gram-positive bacteria produce membrane vesicles: proteomics-based characterization of *Staphylococcus aureus*-derived membrane vesicles. *Proteomics* 9, 5425–5436. doi: 10.1002/pmic.200900338
- Lee, J. H., Choi, C. W., Lee, T., Kim, S. I., Lee, J. C., and Shin, J. H. (2013). Transcription factor sigmaB plays an important role in the production of extracellular membrane-derived vesicles in *Listeria monocytogenes*. *PLoS One* 8:e73196. doi: 10.1371/journal.pone.0073196
- Leitão, A. L., and Enguita, F. J. (2016). *Non-Coding RNAs and Inter-Kingdom Communication*. Cham: Springer International Publishing.
- Liao, S., Klein, M. I., Heim, K. P., Fan, Y., Bitoun, J. P., Ahn, S. J., et al. (2014). *Streptococcus mutans* extracellular DNA is upregulated during growth in biofilms, actively released via membrane vesicles, and influenced by components of the protein secretion machinery. *J. Bacteriol.* 196, 2355–2366. doi: 10.1128/JB.01493-14
- Logozzi, M., De Milito, A., Lugini, L., Borghi, M., Calabro, L., Spada, M., et al. (2009). High levels of exosomes expressing CD63 and caveolin-1 in plasma of melanoma patients. *PLoS One* 4:e5219. doi: 10.1371/journal.pone.0005219
- Marquis, R. E. (1973). Immersion refractometry of isolated bacterial cell walls. *J. Bacteriol.* 116, 1273–1279. doi: 10.1128/jb.116.3.1273-1279.1973
- Marr, A. G., and Ingraham, J. L. (1962). Effect of temperature on the composition of fatty acids in *Escherichia Coli*. *J. Bacteriol.* 84, 1260–1267. doi: 10.1128/jb.84.6.1260-1267.1962
- Masotti, A., Baldassarre, A., Fabrizi, M., Olivero, G., Loreti, M. C., Giammaria, P., et al. (2017). Oral glucose tolerance test unravels circulating miRNAs associated with insulin resistance in obese preschoolers. *Pediatr. Obes.* 12, 229–238. doi: 10.1111/ijpo.12133
- Michel, N., Fabiano, A. S., Polidori, A., Jack, R., and Pucci, B. (2006). Determination of phase transition temperatures of lipids by light scattering. *Chem. Phys. Lipids* 139, 11–19. doi: 10.1016/j.chemphyslip.2005.09.003
- Mika, J. T., Thompson, A. J., Dent, M. R., Brooks, N. J., Michiels, J., Hofkens, J., et al. (2016). Measuring the viscosity of the *Escherichia coli* plasma membrane using molecular rotors. *Biophys. J.* 111, 1528–1540. doi: 10.1016/j.bpj.2016.08.020
- Morein, S., Andersson, A., Rilfors, L., and Lindblom, G. (1996). Wild-type *Escherichia coli* cells regulate the membrane lipid composition in a “window” between gel and non-lamellar structures. *J. Biol. Chem.* 271, 6801–6809. doi: 10.1074/jbc.271.12.6801
- Provencher, S. W. (1982). A constrained regularization method for inverting data represented by linear algebraic or integral equations. *Comput. Phys. Commun.* 27, 213–227. doi: 10.1016/0010-4655(82)90173-4
- Rivera, J., Cordero, R. J., Nakouzi, A. S., Frases, S., Nicola, A., and Casadevall, A. (2010). *Bacillus anthracis* produces membrane-derived vesicles containing biologically active toxins. *Proc. Natl. Acad. Sci. U.S.A.* 107, 19002–19007. doi: 10.1073/pnas.1008843107
- Romancino, D. P., Buffa, V., Caruso, S., Ferrara, I., Raccosta, S., Notaro, A., et al. (2018). Palmitoylation is a post-translational modification of Alix regulating the membrane organization of exosome-like small extracellular vesicles. *Biochim. Biophys. Acta Gen. Subj.* 1862, 2879–2887. doi: 10.1016/j.bbagen.2018.09.004
- Schrempf, H., Koebsch, I., Walter, S., Engelhardt, H., and Meschke, H. (2011). Extracellular streptomyces vesicles: amphorae for survival and defence. *Microb. Biotechnol.* 4, 286–299. doi: 10.1111/j.1751-7915.2011.00251.x
- Simpson, R. J., Lim, J. W., Moritz, R. L., and Mathivanan, S. (2009). Exosomes: proteomic insights and diagnostic potential. *Expert Rev. Proteomics* 6, 267–283. doi: 10.1586/epr.09.17
- Spinuzzi, F., Ferrero, C., Ortore, M. G., De Maria Antolinos, A., and Mariani, P. (2014). GENFIT: software for the analysis of small-angle X-ray and neutron scattering data of macro-molecules in solution. *J. Appl. Crystallogr.* 47, 1132–1139. doi: 10.1107/s1600576714005147
- Taylor, D. D., and Gercel-Taylor, C. (2008). MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol. Oncol.* 110, 13–21. doi: 10.1016/j.ygyno.2008.04.033
- Trauble, H., and Overath, P. (1973). The structure of *Escherichia coli* membranes studied by fluorescence measurements of lipid phase transitions. *Biochim. Biophys. Acta* 307, 491–512. doi: 10.1016/0005-2736(73)90296-4
- Valadi, H., Ekstrom, K., Bossios, A., Sjostrand, M., Lee, J. J., and Lotvall, J. O. (2007). Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* 9, 654–659. doi: 10.1038/ncb1596
- van der Pol, L., Stork, M., and van der Ley, P. (2015). Outer membrane vesicles as platform vaccine technology. *Biotechnol. J.* 10, 1689–1706. doi: 10.1002/biot.201400395
- Wang, X., Thompson, C. D., Weidenmaier, C., and Lee, J. C. (2018). Release of *Staphylococcus aureus* extracellular vesicles and their application as a vaccine platform. *Nat. Commun.* 9:379. doi: 10.1038/s41467-018-03847-z
- White, G. F., Racher, K. I., Lipski, A., Hallett, F. R., and Wood, J. M. (2000). Physical properties of liposomes and proteoliposomes prepared from *Escherichia coli* polar lipids. *Biochim. Biophys. Acta* 1468, 175–186. doi: 10.1016/s0005-2736(00)00255-8
- Wolters, D., Antoine, J. M., Myllyluoma, E., Schrezenmeier, J., Szajewska, H., and Rijkers, G. T. (2010). Guidance for substantiating the evidence for beneficial effects of probiotics: prevention and management of infections by probiotics. *J. Nutr.* 140, 698S–712S. doi: 10.3945/jn.109.113753

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Sarra, Celluzzi, Bruno, Ricci, Sennato, Ortore, Casciardi, Del Chierico, Postorino, Bordi and Masotti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.