Check for updates

OPEN ACCESS

EDITED BY Panagiota Anyfanti, Aristotle University of Thessaloniki, Greece

REVIEWED BY Philipp Douschan, Medical University of Graz, Austria Ayman Geddawy, Prince Sattam Bin Abdulaziz University, Saudi Arabia

*CORRESPONDENCE Ying Feng Image: fengying@ccmu.edu.cn Xianbo Wang Image: wangxb@ccmu.edu.cn

RECEIVED 22 October 2024 ACCEPTED 20 January 2025 PUBLISHED 31 January 2025

CITATION

Li Y, Zhu B, Shi K, Lu Y, Zeng X, Li Y, Zhang Q, Feng Y and Wang X (2025) Advances in intrahepatic and extrahepatic vascular dysregulations in cirrhotic portal hypertension. *Front. Med.* 12:1515400. doi: 10.3389/fmed.2025.1515400

COPYRIGHT

© 2025 Li, Zhu, Shi, Lu, Zeng, Li, Zhang, Feng and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Advances in intrahepatic and extrahepatic vascular dysregulations in cirrhotic portal hypertension

Yanqiu Li, Bingbing Zhu, Ke Shi, Yu Lu, Xuanwei Zeng, Yongqi Li, Qun Zhang, Ying Feng* and Xianbo Wang*

Center for Integrative Medicine, Beijing Ditan Hospital, Capital Medical University, Beijing, China

Cirrhotic portal hypertension, the most prevalent and clinically significant complication of liver cirrhosis, manifests as elevated portal venous pressure and is associated with severe complications. Although much research on the mechanisms of portal hypertension has focused on liver fibrosis, less attention has been given to the role of intrahepatic and extrahepatic vascular dysfunction, particularly with respect to extrahepatic vasculature. While the role of hepatic fibrosis in cirrhotic portal hypertension is undeniable, the underlying mechanisms involving intrahepatic and extrahepatic vasculature are highly complex. Sinusoidal capillarization and endothelial dysfunction contribute to increased intrahepatic vascular resistance. Hemodynamic changes in the extrahepatic circulation, including splanchnic vasodilation and hyperdynamic circulation, play a significant role in the development of portal hypertension. Additionally, therapeutic strategies targeting these vascular mechanisms are diverse, including improvement of sinusoidal microcirculation, therapies targeting hepatic stellate cells activation, and pharmacological modulation of systemic vascular tone. Therefore, in this review, we will discuss the vascularrelated mechanisms and treatment progress of portal hypertension in cirrhosis to provide a new theoretical basis and practical guidance for clinical treatment.

KEYWORDS

cirrhotic portal hypertension, sinusoidal capillarization, endothelial dysfunction, splanchnic vasodilation, hyperdynamic circulation, nitric oxide

1 Introduction

Liver cirrhosis is pathologically characterized by hepatocyte necrosis, fibrous tissue proliferation, and intrahepatic vascular remodeling (1). Portal hypertension is one of the most common and severe complications of cirrhosis, significantly impacting patients' quality of life and prognosis (2). Except for fibrosis, the mechanisms underlying portal hypertension are complex, involving both intrahepatic and extrahepatic factors. The development of portal hypertension is primarily attributed to increased intrahepatic vascular resistance and increased portal venous inflow. Increased intrahepatic vascular resistance arises from structural changes induced by hepatic fibrosis and regenerative nodule formation. Additionally, the activation of hepatic stellate cells (HSCs) and the excessive proliferation of myofibroblasts play crucial roles (3). The activation of these cells not only elevates intrahepatic vascular resistance but also promotes further fibrosis through the release of various cytokines and growth factors (4). Moreover, endothelial dysfunction and microvascular structural alterations significantly impact hepatic hemodynamics (5). The anticoagulant properties of liver sinusoidal endothelial cells are diminished, promoting thrombosis, which further exacerbates portal hypertension (6). Extrahepatically, vascular remodeling of the portal venous system and the formation of

portosystemic collaterals are important compensatory mechanisms (7). While these vascular changes can partially alleviate pressure within the portal system, they can also lead to serious complications such as esophageal and gastric varices and gastrointestinal bleeding.

However, the intrahepatic and extrahepatic vascular regulatory mechanisms governing portal hypertension are complex and a comprehensive understanding remains elusive. Current therapeutic options, while capable of reducing portal pressure and preventing bleeding to some extent, have limited efficacy and are associated with side effects. Therefore, this review aims to systematically summarize the intrahepatic and extrahepatic vascular mechanisms of portal hypertension in liver cirrhosis, analyze the progress of existing research and explore potential therapeutic strategies and future research directions. Through a comprehensive understanding of the pathophysiological mechanisms of portal hypertension, we expect to provide a theoretical basis and novel insights for clinical management, thereby improving the prognosis and quality of life for patients with cirrhosis.

2 Intrahepatic vascular changes in cirrhotic portal hypertension

During the process of cirrhosis, the liver's internal vasculature undergoes significant structural changes, mainly manifested as sinusoidal remodeling and capillarization. In terms of functional changes, endothelial cell damage leads to reduced nitric oxide (NO) synthesis, imbalance of systolic and diastolic vascular factors, and increased blood flow resistance, ultimately leading to the occurrence and development of portal hypertension (Figure 1).

2.1 Intrahepatic structural changes

During the development of liver cirrhosis, the intrahepatic vascular system undergoes significant structural changes. Major structural changes include hepatic sinus remodeling and capillarization. Hepatic sinuses are special capillary-like structures. Liver sinus endothelial cells (LSECs) in hepatic sinuses have fenestrae that allow direct contact between blood and hepatocyte, facilitating the exchange of substances (5). In liver cirrhosis, hepatic sinus remodeling and capillary vascularization are key pathophysiological changes and affect intrahepatic hemodynamics. Hepatic sinus remodeling involves LSECs dysfunction, capillarization, and activation of Kupffer cells and HSCs (Figure 2).

LSECs dysfunction and capillarization play a role in hepatic sinus remodeling. LSECs maintain important physiological functions in healthy liver, including regulation of liver blood flow, substance exchange, and immune surveillance (5, 8). In liver cirrhosis, LSECs lose



The intrahepatic and extrahepatic mechanisms of cirrhotic portal hypertension. LSECs, liver sinus endothelial cells; HSCs, hepatic stellate cells; HVR, hepatic vascular resistance; PBF, portal blood flow; VSMC, vascular smooth muscle cell.



their normal fenestrae structure and form a continuous basement membrane, a process known as hepatic sinusoidal capillarization (4, 5). Capillary vascularization leads to increased blood flow resistance and increased portal vein pressure. The formation of basement membrane is another key link in the hepatic sinusoidal capillarization. Dysfunctional LSECs secrete large amounts of basement membrane components, such as type IV collagen and laminin (9). These components are deposited beneath the LSECs, forming a continuous basement membrane (5). This basement membrane increases blood flow resistance and impedes the exchange of substances. Besides, dedifferentiation of LSECs means LSECs lost their characteristic fenestrated structure (10). This change causes the hepatic sinuses to lose their high permeability, obstructing material exchange and increasing blood flow resistance (11). The reasons for the loss of fenestration include endothelial cell damage, chronic inflammatory response and persistent cytokine stimulation.

Activation of HSCs play a central role in hepatic sinus remodeling and capillary vascularization. In liver cirrhosis, HSCs

change from a resting state to an active state, similar to myofibroblasts, producing large amounts of extracellular matrix (ECM) (12, 13). These ECM components further promote the formation of basement membrane and hepatic sinusoidal capillarization. HSCs also release a variety of pro-fibrotic factors, such as transforming growth factor- β (TGF- β) and platelet-derived growth factor (PDGF), which further promotes the fibrotic process and hepatic sinus remodeling (14).

Kupffer cells are resident macrophages in liver that play important immune and clearance functions in the hepatic sinuses (15). In liver cirrhosis, Kupffer cells are activated and release multiple inflammatory mediators and chemokines, such as tumor necrosis factor (TNF- α), interleukin-6 (IL-6), and reactive oxygen species (ROS) (16). These inflammatory mediators not only cause local inflammatory response, but also promote the activation of HSCs and the dysfunction of LSECs (17). It also causes hepatocytes apoptosis and necrosis by releasing ROS and cytokines, and exacerbating hepatic sinus remodeling and fibrosis.

2.2 Intrahepatic functional changes

2.2.1 Endothelial dysfunction

Endothelial cell damage occurs in the early stages of cirrhosis and portal hypertension. Chronic inflammation and oxidative stress are the main factors leading to endothelial cell damage in liver cirrhosis (18). Chronic hepatitis, alcoholic liver disease, non-alcoholic fatty liver disease (NAFLD) and other causes can cause persistent inflammation of liver, this chronic inflammation will lead to endothelial cell damage and dysfunction. Oxidative stress is caused by excessive production of ROS or insufficient antioxidant capacity, and this oxidative stress environment further damages endothelial cells (19).

In cirrhotic portal hypertension, there is a reduction in NO synthesis and release. Liver endothelial cells synthesize NO via endothelial nitric oxide synthase (eNOS) in healthy liver, a potent vasodilator that maintains vascular tone and normal blood flow (20). However, eNOS expression and activity are significantly reduced in cirrhosis, leading to reduced synthesis and release of NO (20, 21). The reduced NO production and decreased NO bioavailability directly leads to a decrease in the diastolic ability of intrahepatic blood vessels (21, 22). The blood vessels show continuous contraction, increasing portal system resistance. NO is not only a vasodilator, but also has the effect of inhibiting platelet adhesion and aggregation (23). The reduction of NO causes platelets to adhere to and aggregate on the damaged endothelial surface, forming tiny thrombi, further blocking blood vessels and aggravating microcirculatory disorders (6). The microthrombi formation not only increases the intrahepatic blood flow resistance, but may also lead to local ischemia and further tissue damage (24).

In contrast to the decreased NO, vasoconstrictors are significantly increased in liver cirrhosis. Endothelial dysfunction is also manifested by excessive release of vasoconstrictive factors such as endothelin-1 (ET-1). ET-1 is a potent vasoconstriction factor secreted by endothelial cells (13), inducing contraction of vascular smooth muscle cells (VSMCs) and increasing vascular resistance by binding to its receptor (13). In liver cirrhosis, due to endothelial cell damage and stimulation of inflammatory mediators, the expression and release of ET-1 are significantly increased (25), resulting in continuous vasoconstriction and further aggravating of portal hypertension (26).

2.2.2 Contraction of HSCs

Activated HSCs have contractile function similar to smooth muscle cells, which is mainly achieved through cytoskeletal remodeling and actin (α -SMA) expression (27). Contractile HSCs can directly increase hepatic sinusoidal resistance and impede blood flow in the portal system (28). α -SMA is a typical myofibroblast marker, and its expression level is closely related to the contractility of HSCs. Through the regulation of intracellular calcium concentration and the interaction of actin-myosin system (29, 30), contractile HSCs can significantly increase the intrahepatic vascular resistance and further lead to portal hypertension (31). Activated HSCs not only have contractile function, but also secrete a variety of vasoconstrictor factors, such as ET-1 and angiotensin II (AngII) (13). These factors act on intrahepatic blood vessels through autocrine and paracrine pathways, further promoting vasoconstriction and fibrosis (32, 33).

2.2.3 Vasoactive substance imbalance

There is an imbalance between NO and ET-1 in cirrhotic portal hypertension. The reduction of NO and the increase of ET-1 in

patients with cirrhosis are typical manifestations of vasoactive substance imbalance (34). The decrease of NO leads to weakened vasodilation, while the increase of ET-1 leads to enhanced vasoconstriction (32). This imbalance between vasodilation and contraction directly leads to increased intrahepatic vascular resistance and increased portal pressure (35).

The expression of prostacyclin (PGI2) and cyclooxygenase-2 (COX-2) is also decreased in liver cirrhosis. PGI2 is a potent vasodilator that inhibits the VSMCs contraction by activating adenylate cyclase (AC) to produce cyclic adenosine phosphate (cAMP) (36). In cirrhosis, the synthesis and release of PGI2 are reduced, resulting in reduced vasodilation (37, 38). COX-2 is a key enzyme in the PGI2 synthesis, and its reduced expression directly affects PGI2 production, further weakening the vasodilation ability (39).

The role of AngII and Angiotensin-converting enzyme (ACE) in the pathophysiology of cirrhotic portal hypertension should not be overlooked. ACE is a key enzyme for AngII production, and its increased activity leads to increased AngII levels (40). AngII is a potent vasoconstriction that causes VSMCs to contract and increase vascular resistance by binding to AngII receptor (41). In addition, AngII also has a pro-fibrotic effect, further aggravating liver fibrosis and portal hypertension by stimulating the activation of HSCs and the production of ECM (42). Inhibiting AngII expression can decrease collagen synthesis (42, 43).

The compensation of vasodilators is obviously insufficient. In cirrhosis, although the levels of certain vasodilator factors such as adrenomedullin (AM) and brain natriuretic peptide (BNP) are elevated to certain extent in an attempt to counteract the overexpression of vasoconstrictors (44, 45), their compensatory effects are often insufficient to maintain normal vascular tone and balance. AM has a strong vasodilatory effect by increased cAMP generation (46). BNP inhibits VSMCs contraction by increasing the production of cyclic guanosine phosphate (cGMP) (47). However, in cirrhosis, the compensatory mechanisms of these vasodilator factors are unable to fully offset the overexpression of vasoconstrictor, resulting in vascular tone imbalance and increased portal pressure.

3 Extrahepatic vascular changes

Portal hypertension in cirrhosis leads to major changes in the extrahepatic vascular and systemic circulation. These changes include increased portal blood flow, splanchnic vasodilation, portal-systemic collateral formation, hyperdynamic circulation, and abnormalities in the intestinal and pulmonary microcirculation. Complex molecular mechanisms involve angiogenesis, vasodilation, and oxidative stress (Figure 1).

3.1 Extrahepatic portal vascular changes

Portal hypertension is one of the core pathological changes in liver cirrhosis. It is caused by many factors, among which changes of extrahepatic portal vein are particularly critical. These changes mainly include a significant increase in portal blood flow and the formation of portosystemic collateral circulation.

Extrahepatic vascular changes in liver cirrhosis include portal blood flow and systemic hemodynamics. Due to the dilation of visceral blood vessels, especially in the gastrointestinal and splenic region, splanchnic blood flow increases, eventually leading to increased blood flow to portal vein (48-50). In addition, changes in systemic hemodynamics are particularly critical. Patients with liver cirrhosis are usually accompanied by a hypovolemic state and reduced effective blood volume in the systemic circulation (51). This state activates the adrenal glands and the sympathetic nervous system, causing systemic vasoconstriction, especially in the renal and splenic vessels, thereby reducing blood flow elsewhere and increasing portal blood flow (52). At the same time, activation of the renin-angiotensin system by the kidney increases the release of vasoconstrictor factors, which further promotes an increase in portal blood flow. Alterations in splanchnic vasculature and systemic hemodynamics in cirrhosis interact to result in a significant increase in portal blood flow (49).

As portal vein pressure continues to rise, portosystemic collaterals are formed to relieve the pressure (53). Although this compensatory mechanism helps to reduce portal vein pressure in the short term, its long-term consequences can lead to a series of complications, including esophageal and gastric varices, hypersplenism, etc. (54). The vascular regulation mechanisms in this process involves are complex. Firstly, one of the core mechanisms of portosystemic collateral circulation is angiogenesis. In cirrhotic portal hypertension, multiple factors induce overexpression of angiogenic factors such as vascular endothelial growth factor (VEGF) and PDGF (55-57). These factors initiate the formation of new collateral circulation by promoting the proliferation and migration of vascular endothelial cells (56). Hypoxia-inducible factor (HIF-1 α) is another key regulator. HIF-1 α , stimulated by hypovolemia, not only induces VEGF expression, but also promotes other related angiogenic factors production, further accelerating the generation of collateral circulation (58, 59). Secondly, remodeling of existing blood vessels is also a mechanism that cannot be ignored during the formation of portosystemic collateral circulation. Especially in the esophagus and fundus of the stomach, the original microvascular network expands and remodels driven by portal pressure, forming functional varicose veins (60). Smooth muscle cells and collagen deposition in the blood vessel wall increase, which enhances the capacity of the blood vessel and allows greater blood flow to pass through (61). At the same time, this vascular remodeling process is accompanied by thinning of the vessel wall, increasing the risk of varicose vein rupture bleeding (57, 61). In addition, inflammatory factors not only promote angiogenesis, but also accelerate local blood vascular remodeling (57). At the same time, leukocyte infiltration will also accelerate blood vessels dilation and the formation of collateral circulation.

3.2 Changes in systemic circulation

Splanchnic vasodilation is one of the core features of systemic circulatory changes in patients with cirrhotic portal hypertension (62). Splanchnic vasodilation causes a series of adverse consequences, including hypovolemia, ascites and so on (7). In cirrhosis, visceral vascular endothelial cells are dysfunctional, and the levels of vasodilators such as NO, PGI2, and carbon monoxide (CO) are significantly increased (63–65). These factors are mainly produced by endothelial cells. Increased production of vasodilator synthase

promotes vascular smooth muscle relaxation and leads to vasodilation (63, 66). In addition, vasoconstrictor factors are relatively reduced. Although vasoconstrictor factors such as ET-1 also increase, their effects are offset by a large number of vasodilator factors. Moreover, the responsiveness of vascular smooth muscle to contractile stimuli decreases (67). Long-term exposure to high concentrations of vasodilator factors reduces the sensitivity of VSMCs to normal contractile stimulation (68). Inflammatory responses also participate in visceral vasodilation and increase vascular permeability, leading to ascites and tissue edema.

Hyperdynamic circulation is another important feature of cirrhotic patients, which is closely related to splanchnic vasodilation and the formation of portosystemic collateral circulation (49). The main manifestations of hyperdynamic circulation include increased cardiac output, decreased peripheral vascular resistance and decreased renal blood flow (69, 70). The occurrence of hyperdynamic circulation involves multiple mechanisms. The first is increased systemic NO production. NO not only causes splanchnic vasodilation, but also reduces arteriolar and capillary resistance through systemic vasodilation, thereby increasing cardiac output (71). This compensatory mechanism is to maintain the oxygen supply requirements of peripheral tissues. Secondly, the sympathetic nervous system and renin-angiotensin system activation are also involved in this process (72). In response to the decrease in systemic vascular resistance, the sympathetic nervous system and renin-angiotensin system are activated to increase blood volume by constricting blood vessels and retaining sodium and water (73). However, this compensatory mechanism has limited effect under the action of NO and other vasodilator factors, and instead aggravates vasodilation and reduce renal blood flow (74). In addition, the role of systemic inflammation in this process cannot be ignored (70). Inflammatory factors promote fluid exudation by increasing vascular permeability, leading to aggravation of ascites. Finally, the compensatory response of the heart will cause some adverse consequences. Long-term compensatory load increase may lead to cirrhotic cardiomyopathy, in which the heart is unable to maintain normal function under increased load (75, 76).

3.3 Microcirculation changes

The microcirculation, as the most subtle component of the vascular system, includes small arteries, capillaries and small veins, responsible for the transport of oxygen and nutrients and the discharge of waste. Cirrhotic portal hypertension has a profound impact on the extrahepatic microcirculation such as intestinal tract and lung through complex mechanisms.

The abnormalities of intestinal microcirculation in patients with cirrhosis are mainly manifested in intestinal ischemia, obvious vascular congestion in the intestinal wall, increased vascular permeability, and resulting in intestinal wall edema (77–79). It affects the absorption of nutrients and aggravates the malnutrition of patients (78). In addition, portal hypertension aggravates local intestinal inflammatory response, destroy intestinal barrier function, increase the risk of bacterial translocation, and thus induce systemic inflammatory response syndrome (SIRS) (80). It also promotes the entry of intestinal endotoxins into the portal vein system, aggravating liver inflammation and fibrosis (81). The mechanism of abnormal

intestinal microcirculation involves multiple aspects. Firstly, portal hypertension causes intestinal venous congestion, vasodilation, and slow blood flow, leading to insufficient microcirculatory perfusion (79). Secondly, there is an imbalance of vasoactive substances. Increased vasoconstrictor factors, such as ET-1, lead to intestinal microvascular spasm and further aggravate ischemia (82). The decreased bioavailability of NO leads to vascular endothelial dysfunction and further deteriorates intestinal microcirculation (83). In addition, the oxidative stress response is enhanced in cirrhosis, leading to endothelial damage, weakening the dilation ability of blood vessels, and ultimately affecting microcirculatory function (84).

Changes in pulmonary microcirculation are a common but easily overlooked complication in patients with cirrhotic portal hypertension. In terms of pulmonary microcirculation, the main manifestations are hepatopulmonary syndrome and hepatogenic pleural effusion. Hepatopulmonary syndrome is characterized by abnormal dilation of pulmonary blood vessels and redistribution of intrapulmonary blood flow (85), leading to oxygenation dysfunction (86); while hepatogenic pleural effusion is caused by obstruction of lymphatic drainage. In contrast to hepatogenic pleural effusion, hepatic hydrothorax develops when ascitic fluid moves from the peritoneal cavity into the pleural space through diaphragmatic defects (87), which is unrelated to pulmonary microcirculatory disorders. The mechanism of pulmonary microcirculation changes involves multiple aspects. The first is that pulmonary vasodilation occurs primarily at the level of alveolar capillaries. In liver cirrhosis, excessive synthesis of NO will cause abnormal expansion of pulmonary capillaries, increasing pulmonary blood flow, but decreasing oxygen diffusion efficiency, resulting in hypoxemia (88). In addition, the exchange time of oxygen between the alveoli and blood is insufficient, resulting in gas exchange disorder (89, 90). Increased arteriovenous shunting also leads to alveolar ventilation-blood flow imbalance (91). The chronic inflammatory response and oxidative stress will increase the permeability of pulmonary capillaries, leading to pulmonary edema and further aggravate the damage of lung function (90, 92, 93). Recent studies have revealed that increased pulmonary expression of placental growth factor (PIGF) and VEGF-A plays a central role in pathological angiogenesis (94). The von Willebrand factor-angiopoietin axis activation and altered circadian rhythm proteins, particularly BMAL1, significantly affect hypoxic responses and vascular remodeling (88, 95). Additionally, bacterial translocation and endotoxemia contribute to pulmonary inflammation through recruitment of intravascular monocytes that produce proangiogenic factors (96). These molecular mechanisms create extensive pulmonary microvascular alterations, including capillary dilatation, arteriovenous malformations, and altered vascular reactivity. In contrast, portopulmonary hypertension (POPH) represents a distinct entity characterized by pulmonary arterial hypertension in the setting of portal hypertension (97). The pathophysiology involves pulmonary vasoconstriction, vascular remodeling, and in situ thrombosis. Key molecular pathways include endothelial dysfunction with decreased NO and prostacyclin production, upregulation of ET-1 and serotonin pathways, and proliferation of pulmonary arterial smooth muscle cells (98). BMP9 is a sensitive and specific biomarker of POPH, which could predict transplant-free survival and the presence of pulmonary arterial hypertension in liver disease (99). The mechanical stress from increased pulmonary blood flow in the hyperdynamic circulatory state may trigger endothelial injury, initiating these pathological cascades.

4 Pharmacological interventions based on intrahepatic vascular changes

Intrahepatic vascular changes play a role in liver cirrhosis. And therapeutic strategies targeting intrahepatic vessels mainly focus on anti-fibrotic treatment, improvement of hepatic sinusoidal microcirculation, and treatment targeting HSCs (Table 1).

4.1 Antifibrotic therapy

Antifibrotic therapy is one of the cornerstones of cirrhotic portal hypertension treatment. Liver fibrosis leads to structural remodeling and functional abnormalities of intrahepatic blood vessels, thereby causing increased portal pressure. Therefore, inhibiting and reversing the process of liver cirrhosis has become a key strategy to reduce portal pressure.

Firstly, inhibiting excessive deposition of ECM. The main pathological feature of liver cirrhosis is the massive ECM deposition (100). Antifibrotic treatments aim to reduce or reverse the accumulation of ECM (3). Many drugs and molecular targets can intervene in this process, including blocking liver fibrosis formation by inhibiting the TGF- β signaling pathway (14). TGF- β is an important profibrotic factor in liver fibrosis, and inhibiting its activity can significantly reduce the degree of fibrosis. Activators of matrix metalloproteinases (MMPs) can promote ECM degradation (101); while inhibitors of tissue inhibitors of metalloproteinases (TIMPs) can reduce ECM deposition (102). Therefore, MMP/TIMP balance is a potential therapeutic target for regulating the extracellular matrix (102, 103). Activation of HSCs can lead to imbalances in MMP2/ TIMP2 and MMP9/TIMP1, aggravating fibrosis (104, 105).

Secondly, applying antioxidant and anti-inflammatory treatment. Oxidative stress and inflammatory responses also play a key role in the process of liver cirrhosis. Therefore, antioxidants and anti-inflammatory drugs are used to alleviate oxidative stress damage to liver cells. Common antioxidants include vitamin E, lipoic acid, etc., which improve liver fibrosis by reducing the ROS production (106, 107). Wang Q, et al. found that glycyrrhizic acid inhibited oxidative stress injury through targeting AKR7A2 in HSCs, reduced the activated HSCs proliferation and reversed hepatic fibrosis (108). In addition, anti-inflammatory drugs such as glucocorticoids and certain immunomodulators can reduce the chronic inflammatory response and progression of liver cirrhosis. Qin BF, et al. found that specnuezhenide inflammatory response via SIRT6-P2X7R/NLRP3 pathway and improve fibrosis (109).

Targeting HSCs activation is important in pharmacological treatments for patients with cirrhotic portal hypertension. Activated HSCs are the main effector cells of liver cirrhosis. By inhibiting the activation of HSCs or promoting their apoptosis, ECM production can be effectively reduced (110). Some drugs, such as retinoic acid receptor gamma agonists (such as retinoic acid) and peroxisome proliferator-activated receptor gamma (PPAR γ) agonists, have shown the potential to inhibit HSCs activation (111). Benedicto AM, et al. have shown that interference with mitochondrial function could target HSCs to inhibit fibrosis (110). Tung HC, et al. demonstrated that inhibition of heme-thiolate monooxygenase CYP1B1 could decrease HSCs activation and fibrosis (12).

TABLE 1	Evidence for pharmacological	therapy targeting int	rahepatic and extrahepati	c vascular dysregulations i	n portal hypertension.
		the set of the getting in the		e raceatar ajeregatatierie.	

	Categories	Agent	Model	Research subjects	Direct target	Mechanism
Pun et al. (143)	Clinical drugs	Fructooligos- accharides	BDL rats	Intrahepatic vessels	ROS/eNOS	Ameliorate of dysbiosis and oxidative stress
Asada et al. (144)	_	Tofogliflozin	CCl ₄ rats	LSECs, HSCs	SGLT 2 inhibitors	Improving endothelial dysfunction
Fan et al. (57)		Cediranib	BDL rats	Intrahepatic and extrahepatic vessels	VEGFR-2	Improving vascular remodeling and contractility
Vairappan et al. (145)		Candesartan cilexetil	CCl ₄ mice	HUVECs	Nostrin-eNOS-NO	Improving endothelial dysfunction
Tai et al. (146)	-	Celecoxib	TAA rats	LSECs	eNOS	NO homeostasis
Noah et al. (147)		Empagliflozin	CCl4 rats	LSECs, HSCs	Gal-1/NRP-1	Suppression of angiogenesis
Zheng et al. (148)		Telmisartan	CCl4, BDL rats	Liver and mesenteric tissue	KLF-4, eNOS	Reducing angiogenesis and vascular remodeling
Zhu et al. (67)	Small molecular agents	8-OH-DPAT	TAA, BDL, PPVL rats	VSMCs	5-HT receptor 1A	Inducing the contraction of portal vein
Zhao et al. (61)		Imperatorin	CCl₄ rats	HSCs	TGF-β	Reducing hepatic fibrosis and vascular remodeling
Li et al. (149)		Urolithin A	CCl _{4,} BDL mice	HSCs	Glutaminas-e1	Inhibiting fibrogenesis and HSCs contraction
Gunarathne et al. (150)		MrgD	BDL, PPVL, CCl₄ rats	Splanchnic vessels	Mas receptor	Mesenteric Vasodilation
Wang et al. (151)		DPP4i	CCl ₄ rats	Mesenteric arterioles	Nox4	Normalizing arterial hypocontractility
Pun et al. (152)		Glycyrrhizin	BDL rats	Mesenteric vessels	VEGF	Attenuating portosystemic collateral shunting
Boyer-Diaz et al. (153)		Lanifibranor	TAA, BDL rats	HSCs, LSECs	Pan-PPAR agonist	Ameliorating hepatic microvascular function
Brusilovskaya et al. (154)		TADA	BDL rats	Intrahepatic vessels	PDE-5 inhibitor	Reducing sinusoidal vascular resistance
Tsai et al. (155)		Obeticholic acid	BDL rats	Intrahepatic vessels	Farnesoid X receptor agonist	Inhibiting vasoconstriction
Hu et al. (156)		AICAR	BDL, PPVL, CCl₄ rats	LSECs	AMPK/NO	Improving NO bioavailability
Castillo (157)		PHIN-156	BDL rats	Vasopressin receptor	V1a partial agonists	Reducing portal blood flow
Jones (158)		BI 685509	TAA rats	Portosystemic shunting	sGC, cGMP	NO-independent sGC activator
Zhao et al. (39)		PTUPB	CCl4 rats	Intrahepatic and extrahepatic vessels	sEH/COX-2/TGF-β	Inhibiting intra-or extrahepatic angiogenesis and vascular remodeling

BDL, bile duct ligation; eNOS, endothelial nitric oxide synthase; ROS, reactive oxygen species; CCl₄ Carbon tetrachloride; LSECs, liver sinus endothelial cells; HSCs, hepatic stellate cells; SGLT2, Sodium glucose transferase 2; VEGFR-2, vascular endothelial growth factor receptor 2; HUVECs, human umbilical vein endothelial cells; NO, Nitric oxide; TAA, thioacetamide; Gal-1, galactin-1; NRP-1, Neuropilin-1; KLF-4, Krüppel-like factor-4; PPVL, partial portal vein ligation; VSMCs, vascular smooth muscle cells; 5-HT, 5-hydroxytryptamine; TGF-β, Transforming growth factor-β; MrgD, Mas-related G protein-coupled receptor type D; DPP4i, Dipeptidyl peptidase-4 inhibitor; Nox4, NADPH oxidase 4; VEGF, vascular endothelial growth factor; Pan-PPAR, pan-peroxisome proliferator-activated receptor; TADA, Soluble guanylyl cyclase stimulation and phosphodiesterase-5; PDE-5, phosphodiesterase-5; AICAR, 5-aminoimidazole-4carboxyamide ribonucleoside; AMPK, Adenosine 5'-monophosphate-activated protein kinase; V1a, vasopressin 1a; sGC, soluble guanylyl cyclase; cGMP, cyclic guanosine monophosphate; PTUPB, 4-(5-phenyl-3-57-pyrazol-1-yl) -benzenesulfonamide; sEH, soluble epoxide hydrolase; COX-2, cyclooxygenase-2. Promoting hepatocyte regeneration can improve liver function and indirectly reduce fibrosis. Growth factors such as hepatocyte growth factor (HGF) and epidermal growth factor (EGF) have been shown to promote liver regeneration and reduce fibrosis. Wang P, et al. have identified hepatic Snai1 and Snai2 as key transcriptional regulators of liver regeneration and fibrosis (112). Novel stem cell therapies, such as mesenchymal stem cells (MSCs), possess immunomodulatory and anti-inflammatory capabilities that make them an attractive approach for promoting liver regeneration (113). The primary mechanism involves promoting apoptosis of HSCs and subsequently stimulating hepatocyte proliferation, thereby replacing damaged hepatocytes and reducing liver fibrosis (114).

4.2 Improvement of hepatic sinusoidal microcirculation

Hepatic sinusoidal microcirculation disorder is one of the important mechanisms for portal hypertension. It can directly reduce intrahepatic vascular resistance, thereby reducing portal pressure.

Statins, the most widely used lipid-lowering drugs, have been found in recent years to have the potential to improve liver cirrhosis and hepatic sinusoidal microcirculation (115, 116). Statins increase the expression and activity of eNOS, promoting the production of NO, and reducing hepatic sinusoidal resistance (116). In addition, statins can inhibit the production of inflammatory factors, such as TNF- α and IL-6, thereby reducing microcirculation disorders caused by inflammation (117). Statins can also protect LSECs and maintain their normal function by reducing the ROS production (117, 118). Statins can also reduce hepatic sinusoidal contraction and improve microcirculation by inhibiting Rho kinase activity (119). Some studies have shown that simvastatin and atorvastatin can significantly reduce portal pressure and improve the prognosis of patients with cirrhosis (120, 121). However, it should be noted that statins should be used with caution in patients with advanced cirrhosis to avoid potential hepatotoxicity (122). Statins represent the most clinically advanced antifibrotic therapy, with multiple Phase III trials demonstrating their potential in portal hypertension. Simvastatin and atorvastatin have shown particular promise, with data supporting their safety in compensated cirrhosis. However, their use in advanced cirrhosis requires careful monitoring.

In liver cirrhosis, NO production is reduced due to endothelial cell dysfunction, leading to increased vasoconstriction. Therefore, exogenous NO donors, such as nitrates, can improve the expansion ability of liver sinuses and reduce intrahepatic vascular resistance (32). In addition, NO donors can inhibit the contraction of HSCs and reduce their compression on hepatic sinusoids, thereby improving microcirculation (20). Villanueva C, et al. have found that isosorbide mononitrate (ISMN) can significantly reduce portal pressure and prevent variceal rebleeding, especially during acute application (123). However, long-term use may lead to the reduced tolerance and effectiveness of treatment. To overcome tolerance issues, researchers are exploring intermittent dosing regimens and novel NO donors. For example, NCX-1000 is a liver-targeted NO donor that can specifically release NO, promising to improve therapeutic efficacy and reduce systemic side effects (124). In

addition, combination treatment strategies combining statins and nitrates also show good promise. This combination can work synergistically through different mechanisms to improve hepatic sinusoidal microcirculation more effectively. Nicorandil and atorvastatin may alleviate hepatic sinusoidal microcirculatory disorders by improving liver function, anti-inflammation and antioxidation (125).

Endothelin (ET) receptor antagonist is another choice of hepatic sinusoidal microcirculation improvement. There is an observed up-regulation of the ET-1 gene accompanied by a compensatory down-regulation of the ET A receptor (ETAR) gene in the human portal vein (25). Blocking the ET-1/ETAR pathway using selective ETAR antagonists (ERAs) represents a promising therapeutic strategy for liver cirrhosis treatment (26). Ten Hove M, et al. have demonstrated that engineered SPIONs functionalized with ETAR antagonist had improved liver fibrosis through the inhibition of HSCs activation (13). A selective ET-A antagonists, such as BQ 123 and Ambrisentan, decrease the portal pressure in cirrhotic patients (126).

4.3 Treatment targeting the contractile function of HSCs

HSCs play a central role in the development of portal hypertension. Activated HSCs are not only the main ECM producers that lead to liver fibrosis, but also have contractile properties and are directly involved in the regulation of hepatic sinusoidal resistance. Therefore, therapeutic strategies targeting HSCs have become a hot topic in recent years. HSCs have contractile properties and are directly involved in the regulation of liver sinusoidal resistance (127). ET-1 is a potent vasoconstrictor that can cause HSCs to contract. The use of ET-1 receptor antagonists can reduce HSCs contraction and reduce liver sinusoidal resistance (25). In addition, AngII can promote HSCs contraction and proliferation (128). AngII receptor antagonists, such as losartan, can reduce HSCs contraction and improve liver sinusoidal microcirculation (42). Moreover, Nanotechnology can be used to achieve targeted drug delivery, improve therapeutic effects and reduce side effects. Vitamin A-modified liposomes can specifically deliver drugs to HSCs because HSCs are the main vitamin A storage cells in liver. This strategy can be used to deliver anti-fibrotic drugs, siRNA or gene therapy vectors. Kaili Wang et al. constructed hyaluronic acid (HA) modified liposomes co-delivering all-trans retinoic acid (RA) and L-arginine (L-arg) to reverse hepatic fibrosis (129). Lingfeng Zhang et al. designed chondroitin sulfate-modified and vismodegib-loaded nanoparticles (CS-NPs/VDG) to efficiently normalize the fenestrae phenotype of LSECs and restore HSCs to quiescent state by inhibiting Hedgehog signaling pathway (130). Additionally, stem cell therapy is a treatment method targeting HSCs that has attracted much attention in recent years. MSCs have multidirectional differentiation potential and immunomodulatory functions, and can inhibit HSCs activation by secreting various anti-inflammatory and anti-fibrotic factors (131, 132). Preliminary clinical studies have shown that stem cell therapy is effective in reducing liver fibrosis and improving liver function (131). These have inspired new ways of thinking about treating liver fibrosis.

5 Pharmacological interventions based on changes in extrahepatic vessels

Extrahepatic vascular dilation, increased blood flow, and changes in peripheral vascular resistance exacerbate the portal pressure. Therefore, treatment strategies based on changes in extrahepatic blood vessels aim to reduce portal blood flow, regulate vascular tension, and improve systemic hemodynamic balance (Table 1). The clinical development of therapies targeting extrahepatic vascular changes shows a clear stratification in terms of evidence and approval status. Non-selective beta blockers (NSBBs) and vasoconstrictors represent the current standard of care, supported by Class I, Level A evidence. Propranolol, carvedilol, and nadolol are FDA-approved and widely used in clinical practice. For acute complications, terlipressin and octreotide have established roles in management protocols. Beyond these approved therapies, several novel approaches are in various stages of clinical development. Understanding this therapeutic hierarchy is essential for optimal clinical decision-making and future research directions.

5.1 NSBBs

NSBBs are classic drugs for cirrhotic portal hypertension (133). The main mechanisms include reducing cardiac output by blocking β1 receptors, and causing splanchnic vasoconstriction and reducing portal blood flow by blocking \u03b22 receptors. NSBBs can effectively reduce the burden on portal system, and are especially suitable for preventing esophageal and gastric variceal bleeding (134). Propranolol and Carvedilol are commonly used NSBBs. These drugs are widely used for the primary prevention of portal hypertension, which is to prevent bleeding from varicose veins that are not yet bleeding (135). For patients who have already suffered bleeding, NSBBs are also used for secondary prevention to reduce the risk of rebleeding (136). Although NSBBs are effective in reducing portal pressure and preventing variceal rupture, not all patients can tolerate these drugs, especially those with hypotension or severe cardiac dysfunction (137). In addition, NSBBs may interact with other medications, so they should be used with caution.

5.2 Vasoconstrictors

Vasoconstrictors reduce portal pressure by constricting the visceral arterial system and reducing blood flow to the portal vein. Their main target is the visceral vascular smooth muscle, directly or indirectly regulating its contractile function. They are usually used to treat acute complications of portal hypertension, such as gastric variceal. Terlipressin, one of the most commonly used vasoconstrictors clinically, reduces portal vein blood flow by selectively acting on V1 receptors in visceral blood vessels (138). Long-term continuous infusion of terlipressin can significantly increase cardiac reserve and attenuate a hyperdynamic state (139). Octreotide have similar effect for the management of variceal bleeding (140). The main side effects of vasoconstrictors include increased blood pressure, myocardial ischemia, and impaired renal function. Therefore, patients with cardiovascular disease or renal insufficiency should be used with extreme caution and closely monitored. In addition, long-term use of

these drugs may lead to decreased renal perfusion and increase the risk of AKI (141).

5.3 Angiogenesis inhibition therapy

An important feature of cirrhotic portal hypertension is abnormal angiogenesis in the visceral vascular system, especially in the spleen and intestinal areas. These abnormal neovascularization structures are unstable and permeable, leading to increased portal vein pressure. Therefore, targeted treatment strategies to inhibit abnormal angiogenesis have gradually become the focus of research. Cediranib may ameliorate extrahepatic hyperdynamic circulation by targeting angiogenesis. This is achieved through the inhibition of vascular endothelial growth factor receptor 2 (VEGFR-2) signaling, thereby reducing both portal collateral vessel formation and eNOS-mediated vasodilation and vascular remodeling (57). Hydroxysafflor yellow A is a multi-target tyrosine kinase inhibitor that inhibits the VEGF and PDGF signaling pathways, thereby inhibiting abnormal angiogenesis (142). Although anti-angiogenic therapy has great potential in theory, its clinical application is still being explored. These drugs may cause systemic side effects such as hypertension, bleeding, and delayed wound healing, so they should still be used with caution.

6 Future prospective

Future research will focus on further elucidating the intricate intrahepatic and extrahepatic vascular regulatory mechanisms underlying portal hypertension in cirrhosis.

· Nanotechnology for Targeted Therapies

Emerging nanotechnology offers new possibilities for the treatment of portal hypertension. Utilizing nanocarriers enables precise drug delivery to specific cells or tissues, such as HSCs or LSECs. This not only enhances drug concentration at the site of action but also minimizes off-target effects on healthy tissues. This strategy holds significant promise for improving both the efficacy and safety of therapeutic interventions. However, nanotechnology approaches are currently in preclinical development and require additional safety data.

• The Promise of Stem Cell Therapy

Stem cell therapy, a burgeoning therapeutic modality, has demonstrated potential in early clinical trials for reducing liver fibrosis. MSCs, through their immunomodulatory and antiinflammatory properties, can suppress HSC activation and reduce ECM production, thereby slowing the progression of fibrosis. Future research will further investigate the long-term efficacy of stem cell therapy in individuals with cirrhosis and explore strategies to enhance stem cell functionality through gene editing techniques. Stem cell therapies, while showing promise in animal studies, are still in early development phases. MSCs has progressed to Phase I trials, focusing primarily on safety assessments in cirrhotic patients.

Optimization of Existing Drugs and Development of Novel Agents

Future research will focus on optimizing the efficacy and safety of existing drugs. For example, while statins have shown promise in improving sinusoidal microcirculation and reducing portal pressure, their long-term safety requires further validation. The development of novel agents targeting pathological mechanisms like angiogenesis and vasodilation will also be a priority. Although anti-angiogenic drugs, such as VEGF inhibitors, are effective, they can cause systemic side effects like hypertension. Therefore, research will focus on improving drug targeting and minimizing adverse reactions. The safety and preliminary efficacy of novel NO donors are in the early clinical development stage. New small molecules targeting specific pathways require toxicology studies. VEGF inhibitors, obeticholic acid, and rifaximin combined with statins have entered the clinical research stage.

7 Conclusion

Cirrhotic portal hypertension involves complex intrahepatic and extrahepatic vascular mechanisms. Comprehensive treatments such as improving microcirculation and regulating vascular tension can effectively reduce portal pressure, alleviate complications, and improve patient prognosis. More research is needed in the future to validate drugs targeting intrahepatic and extrahepatic vascular disorders in order to improve treatment of portal hypertension.

Author contributions

YaL: Writing – original draft, Writing – review & editing. BZ: Writing – review & editing. KS: Writing – review & editing. YuL: Conceptualization, Writing – review & editing. XZ: Visualization, Writing – review & editing. YoL: Visualization, Writing – review & editing. QZ: Formal analysis, Writing – review & editing. YF: Supervision, Writing – review & editing. XW: Supervision, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This research was funded by the high-level Chinese Medicine Key Discipline Construction Project (No. zyyzdxk-2023005, to Xianbo Wang);

References

1. Dong Z, Wang Y, Jin W. Liver cirrhosis: molecular mechanisms and therapeutic interventions. *MedComm* (2020). (2024) 5:e721. doi: 10.1002/mco2.721

2. Guixé-Muntet S, Quesada-Vázquez S, Gracia-Sancho J. Pathophysiology and therapeutic options for cirrhotic portal hypertension. *Lancet Gastroenterol Hepatol.* (2024) 9:646–63. doi: 10.1016/S2468-1253(23)00438-7

3. Akkız H, Gieseler RK, Canbay A. Liver fibrosis: from basic science towards clinical Progress, focusing on the central role of hepatic stellate cells. *Int J Mol Sci.* (2024) 25:7873. doi: 10.3390/ijms25147873

4. Gao J, Lan T, Kostallari E, Guo Y, Lai E, Guillot A, et al. Angiocrine signaling in sinusoidal homeostasis and liver diseases. *J Hepatol.* (2024) 81:543–61. doi: 10.1016/j. jhep.2024.05.014

5. Qu J, Wang L, Li Y, Li X. Liver sinusoidal endothelial cell: an important yet often overlooked player in the liver fibrosis. *Clin Mol Hepatol.* (2024) 30:303–25. doi: 10.3350/ cmh.2024.0022

6. Hon WM, Lee KH, Khoo HE. Nitric oxide in liver diseases: friend, foe, or just passerby? *Ann N Y Acad Sci.* (2002) 962:275–95. doi: 10.1111/j.1749-6632.2002. tb04074.x

7. Mauro E, Gadano A. What's new in portal hypertension? *Liver Int*. (2020) 40:122–7. doi: 10.1111/liv.14366

8. Poisson J, Lemoinne S, Boulanger C, Durand F, Moreau R, Valla D, et al. Liver sinusoidal endothelial cells: physiology and role in liver diseases. *J Hepatol.* (2017) 66:212-27. doi: 10.1016/j.jhep.2016.07.009 Capital's Funds for Health improvement and Research (No. 2024-1-2173, to Xianbo Wang); National Natural Science Foundation of China (No. 82474419, to Xianbo Wang and 82474426, to Ying Feng); Beijing Municipal Natural Science Foundation (No. 7232272, to Ying Feng); and Beijing Traditional Chinese Medicine Technology Development Fund Project (No. BJZYZD-2023-12, to Xianbo Wang).

Acknowledgments

Authors are grateful to all members of Center for Integrative Medicine of Beijing Ditan Hospital for their contributions to the manuscript preparation. All authors approved the final version of the article, including YaL, BZ, KS, YuL, XZ, YoL, QZ, YF, XW.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The authors declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

9. Lehmann J, Praktiknjo M, Nielsen MJ, Schierwagen R, Meyer C, Thomas D, et al. Collagen type IV remodelling gender-specifically predicts mortality in decompensated cirrhosis. *Liver Int.* (2019) 39:885–93. doi: 10.1111/liv.14070

10. Huang R, Deng J, Zhu C-P, Liu SQ, Cui YL, Chen F, et al. Sulodexide attenuates liver fibrosis in mice by restoration of differentiated liver sinusoidal endothelial cell. *Biomed Pharmacother*. (2023) 160:114396. doi: 10.1016/j.biopha.2023.114396

11. DeLeve LD. Liver sinusoidal endothelial cells in hepatic fibrosis. *Hepatology*. (2015) 61:1740–6. doi: 10.1002/hep.27376

12. Tung H-C, Kim J-W, Zhu J, Li S, Yan J, Liu Q, et al. Inhibition of heme-thiolate monooxygenase CYP1B1 prevents hepatic stellate cell activation and liver fibrosis by accumulating trehalose. *Sci Transl Med.* (2024) 16:eadk8446. doi: 10.1126/ scitranslmed.adk8446

13. Ten Hove M, Smyris A, Booijink R, Wachsmuth L, Hansen U, Alic L, et al. Engineered SPIONs functionalized with endothelin a receptor antagonist ameliorate liver fibrosis by inhibiting hepatic stellate cell activation. *Bioact Mater.* (2024) 39:406–26. doi: 10.1016/j.bioactmat.2024.05.034

14. Ye L, Huang J, Liang X, Guo W, Sun X, Shao C, et al. Jiawei Taohe Chengqi decoction attenuates CCl4 induced hepatic fibrosis by inhibiting HSCs activation via TGF- β 1/CUGBP1 and IFN- γ /Smad 7 pathway. *Phytomedicine*. (2024) 133:155916. doi: 10.1016/j.phymed.2024.155916

15. Guillot A, Tacke F. Liver macrophages revisited: the expanding universe of versatile responses in a spatiotemporal context. *Hepatol Commun.* (2024) 8:e0491. doi: 10.1097/HC9.000000000000491

17. Sauer J, Steixner-Kumar AA, Gabler S, Motyka M, Rippmann JF, Brosa S, et al. Diverse potential of secretome from natural killer cells and monocyte-derived macrophages in activating stellate cells. *Front Immunol.* (2024) 15:1232070. doi: 10.3389/fimmu.2024.1232070

18. Kumar S, Duan Q, Wu R, Harris EN, Su Q. Pathophysiological communication between hepatocytes and non-parenchymal cells in liver injury from NAFLD to liver fibrosis. *Adv Drug Deliv Rev.* (2021) 176:113869. doi: 10.1016/j.addr.2021.113869

19. Pu Y, Zhang S, Zhou R, Huang N, Li H, Wei W, et al. IL-17A up-regulates expression of endothelial tissue factor in liver cirrhosis via the ROS/p 38 signal pathway. *Biochem Biophys Res Commun.* (2016) 470:41–7. doi: 10.1016/j.bbrc.2015.12.093

20. Perramón M, Navalón-López M, Fernández-Varo G, Moreno-Lanceta A, García-Pérez R, Faneca J, et al. Liver-targeted nanoparticles delivering nitric oxide reduce portal hypertension in cirrhotic rats. *Biomed Pharmacother*. (2024) 171:116143. doi: 10.1016/j. biopha.2024.116143

21. Jung E, Baek EB, Hong E-J, Kang JH, Park S, Park S, et al. TXNIP in liver sinusoidal endothelial cells ameliorates alcohol-associated liver disease via nitric oxide production. *Int J Biol Sci.* (2024) 20:606–20. doi: 10.7150/ijbs.90781

22. Vukotic R, Di Donato R, Roncarati G, Simoni P, Renzulli M, Gitto S, et al. 5-MTHF enhances the portal pressure reduction achieved with propranolol in patients with cirrhosis: A randomized placebo-controlled trial. *J Hepatol.* (2023) 79:977–88. doi: 10.1016/j.jhep.2023.06.017

23. Albornoz L, Bandi JC, Otaso JC, Laudanno O, Mastai R. Prolonged bleeding time in experimental cirrhosis: role of nitric oxide. *J Hepatol*. (1999) 30:456–60. doi: 10.1016/S0168-8278(99)80105-6

24. Ding J, Zhao F, Cui J, Zhu M, Zhao W. Platelet activation is upregulated in cirrhotic patients with portal vein thrombosis. *Thromb Haemost.* (2023) 124:739–52. doi: 10.1055/a-2190-8705

25. Owen NE, Williams TL, Maguire JJ, Kuc RE, Davenport EE, Davenport AP. Microarray analysis demonstrates up-regulation of the endothelin-1 gene with compensatory down-regulation of the ETA receptor gene in human portal vein. *Biosci Rep.* (2024) 44:BSR20240528. doi: 10.1042/BSR20240528

26. Owen T, Carpino G, Chen L, Kundu D, Wills P, Ekser B, et al. Endothelin receptor-A inhibition decreases Ductular reaction, liver fibrosis, and angiogenesis in a model of cholangitis. *Cell Mol Gastroenterol Hepatol.* (2023) 16:513–40. doi: 10.1016/j. jcmgh.2023.06.005

27. Ortiz C, Klein S, Reul WH, Magdaleno F, Gröschl S, Dietrich P, et al. Neprilysindependent neuropeptide Y cleavage in the liver promotes fibrosis by blocking NPYreceptor 1. *Cell Rep.* (2023) 42:112059. doi: 10.1016/j.celrep.2023.112059

28. Wu Y, Li Z, Wang S, Xiu A, Zhang C. Carvedilol inhibits angiotensin II-induced proliferation and contraction in hepatic stellate cells through the rho A/rho-kinase pathway. *Biomed Res Int.* (2019) 2019:7932046. doi: 10.1155/2019/7932046

29. van Dijk F, Teekamp N, Post E, Schuppan D, Kim YO, Zuidema J, et al. The antifibrotic potential of a sustained release formulation of a PDGF β -receptor targeted rho kinase inhibitor. *J Control Release*. (2019) 296:250–7. doi: 10.1016/j. iconrel.2018.12.039

30. Li Z, Ding Q, Ling L-P, Wu Y, Meng DX, Li X, et al. Metformin attenuates motility, contraction, and fibrogenic response of hepatic stellate cells in vivo and in vitro by activating AMP-activated protein kinase. *World J Gastroenterol.* (2018) 24:819–32. doi: 10.3748/wjg.v24.i7.819

31. Wang F, Jia Y, Li M, Wang L, Shao J, Guo Q, et al. Blockade of glycolysis-dependent contraction by oroxylin a via inhibition of lactate dehydrogenase-a in hepatic stellate cells. *Cell Commun Signal.* (2019) 17:11. doi: 10.1186/s12964-019-0324-8

32. Duong HTT, Dong Z, Su L, Boyer C, George J, Davis TP, et al. The use of nanoparticles to deliver nitric oxide to hepatic stellate cells for treating liver fibrosis and portal hypertension. *Small.* (2015) 11:2291–304. doi: 10.1002/smll.201402870

33. Tièche S, De Gottardi A, Kappeler A, Shaw S, Sägesser H, Zimmermann A, et al. Overexpression of endothelin-1 in bile duct ligated rats: correlation with activation of hepatic stellate cells and portal pressure. *J Hepatol.* (2001) 34:38–45. doi: 10.1016/S0168-8278(00)00031-3

34. Abdel-Razik A, Mousa N, Abdelsalam M, Abdelwahab A, Tawfik M, Tawfik AM, et al. Endothelin-1/nitric oxide ratio as a predictive factor of response to therapy with Terlipressin and albumin in patients with Type-1 Hepatorenal syndrome. *Front Pharmacol.* (2020) 11:9. doi: 10.3389/fphar.2020.00009

35. Tung H-C, Hsu S-J, Tsai M-H, Lin TY, Hsin IF, Huo TI, et al. Homocysteine deteriorates intrahepatic derangement and portal-systemic collaterals in cirrhotic rats. *Clin Sci (Lond)*. (2017) 131:69–86. doi: 10.1042/CS20160470

36. Sacerdoti D, Pesce P, Di Pascoli M, Brocco S, Cecchetto L, Bolognesi M. Arachidonic acid metabolites and endothelial dysfunction of portal hypertension. *Prostaglandins Other Lipid Mediat.* (2015) 120:80–90. doi: 10.1016/j. prostaglandins.2015.05.008

37. Skill NJ, Theodorakis NG, Wang YN, Wu JM, Redmond EM, Sitzmann JV. Role of cyclooxygenase isoforms in prostacyclin biosynthesis and murine prehepatic portal

hypertension. Am J Physiol Gastrointest Liver Physiol. (2008) 295:G953–64. doi: 10.1152/ ajpgi.00013.2008

38. Rosado E, Rodríguez-Vilarrupla A, Gracia-Sancho J, Monclús M, Bosch J, García-Pagán JC. Interaction between NO and COX pathways modulating hepatic endothelial cells from control and cirrhotic rats. *J Cell Mol Med.* (2012) 16:2461–70. doi: 10.1111/j. 1582-4934.2012.01563.x

39. Zhao Z, Zhang C, Lin J, Zheng L, Li H, Qi X, et al. COX-2/sEH dual inhibitor PTUPB alleviates CCI 4-induced liver fibrosis and portal hypertension. *Front Med (Lausanne)*. (2021) 8:761517. doi: 10.3389/fmed.2021.761517

40. Mastoor Z, Diz-Chaves Y, González-Matías LC, Mallo F. Renin-angiotensin system in liver metabolism: gender differences and role of incretins. *Meta*. (2022) 12:411. doi: 10.3390/metabo12050411

41. Grace JA, Herath CB, Mak KY, Burrell LM, Angus PW. Update on new aspects of the renin-angiotensin system in liver disease: clinical implications and new therapeutic options. *Clin Sci (Lond)*. (2012) 123:225–39. doi: 10.1042/CS20120030

42. Yu FJ, Dong PH, Fan XF, Lin Z, Chen YP, Li J. Down-regulation of angiotensin II by sh RNA reduces collagen synthesis in hepatic stellate cells. *Int J Mol Med.* (2010) 25:801–6. doi: 10.3892/ijmm_00000407

43. Abd El-Rahman SS, Fayed HM. Targeting Ang II/AT1R signaling pathway by perindopril inhibits ongoing liver fibrosis in rat. *J Tissue Eng Regen Med.* (2019) 13:2131–41. doi: 10.1002/term.2940

44. Araujo T, Vohra I, Palacios P, Katiyar V, Flores E, Randhawa T, et al. B-type natriuretic peptide (BNP) predicts 90-day mortality and need for paracentesis in cirrhotic patients without systolic heart failure. *Sci Rep.* (2021) 11:1697. doi: 10.1038/s41598-020-78946-3

45. Fábrega E, Casafont F, Crespo J, De la Peña J, San Miguel G, De las Heras G, et al. Plasma adrenomedullin levels in patients with hepatic cirrhosis. *Am J Gastroenterol.* (1997) 92:1901–4.

46. Elnagdy M, Barve S, McClain C, Gobejishvili L. cAMP signaling in pathobiology of alcohol associated liver disease. *Biomol Ther.* (2020) 10:1433. doi: 10.3390/biom10101433

47. Holditch SJ, Schreiber CA, Harris PC, LaRusso NF, Ramirez-Alvarado M, Cataliotti A, et al. B-type natriuretic peptide overexpression ameliorates hepatorenal fibrocystic disease in a rat model of polycystic kidney disease. *Kidney Int.* (2017) 92:657–68. doi: 10.1016/j.kint.2017.02.017

48. Pun CK, Huang H-C, Chang C-C, Hsu SJ, Chuang CL, Huang YH, et al. Low-dose alcohol exacerbates hyperdynamic circulation and shunting in non-alcoholic cirrhotic rats. *Biosci Rep.* (2024) 44:BSR20240354. doi: 10.1042/BSR20240354

49. Kotani K, Kawada N. Recent advances in the pathogenesis and clinical evaluation of portal hypertension in chronic liver disease. *Gut Liver*. (2024) 18:27–39. doi: 10.5009/gnl230072

50. Simonetto DA, Singal AK, Garcia-Tsao G, Caldwell SH, Ahn J, Kamath PS. ACG clinical guideline: disorders of the hepatic and mesenteric circulation. *Am J Gastroenterol.* (2020) 115:18–40. doi: 10.14309/ajg.00000000000486

51. Téllez L, Albillos A. Non-selective beta-blockers in patients with ascites: the complex interplay among the liver, kidney and heart. *Liver Int.* (2022) 42:749–61. doi: 10.1111/liv.15166

52. McGrath MS, Wentworth BJ. The renin-angiotensin system in liver disease. Int J Mol Sci. (2024) 25:5807. doi: 10.3390/ijms25115807

53. Juncu S, Minea H, Girleanu I, Huiban L, Muzica C, Chiriac S, et al. Clinical implications and Management of Spontaneous Portosystemic Shunts in liver cirrhosis. *Diagnostics (Basel)*. (2024) 14:1372. doi: 10.3390/diagnostics14131372

54. Simón-Talero M, Roccarina D, Martínez J, Lampichler K, Baiges A, Low G, et al. Association between portosystemic shunts and increased complications and mortality in patients with cirrhosis. *Gastroenterology*. (2018) 154:1694–1705.e4. doi: 10.1053/j. gastro.2018.01.028

55. Mariotti V, Fiorotto R, Cadamuro M, Fabris L, Strazzabosco M. New insights on the role of vascular endothelial growth factor in biliary pathophysiology. *JHEP Rep.* (2021) 3:100251. doi: 10.1016/j.jhepr.2021.100251

56. Gana JC, Serrano CA, Ling SC. Angiogenesis and portal-systemic collaterals in portal hypertension. *Ann Hepatol.* (2016) 15:303–13. doi: 10.5604/16652681.1198799

57. Fan Q, Wu G, Chen M, Luo G, Wu Z, Huo H, et al. Cediranib ameliorates portal hypertensive syndrome via inhibition of VEGFR-2 signaling in cirrhotic rats. *Eur J Pharmacol.* (2024) 964:176278. doi: 10.1016/j.ejphar.2023.176278

58. Yang X, Wang Z, Kai J, Wang F, Jia Y, Wang S, et al. Curcumol attenuates liver sinusoidal endothelial cell angiogenesis via regulating Glis-PROX1-HIF-1 α in liver fibrosis. *Cell Prolif.* (2020) 53:e12762. doi: 10.1111/cpr.12762

59. Wang Q, Zhang F, Lei Y, Liu P, Liu C, Tao Y. micro RNA-322/424 promotes liver fibrosis by regulating angiogenesis through targeting CUL2/HIF-1 α pathway. *Life Sci.* (2021) 266:118819. doi: 10.1016/j.lfs.2020.118819

60. Maruyama H, Shiina S. Collaterals in portal hypertension: anatomy and clinical relevance. *Quant Imaging Med Surg.* (2021) 11:3867–81. doi: 10.21037/qims-20-1328

61. Zhao Z, Fan Q, Zhang C, Zheng L, Lin J, Chen M, et al. Imperatorin attenuates CCl4-induced cirrhosis and portal hypertension by improving vascular remodeling and

profibrogenic pathways. Eur J Pharmacol. (2024) 980:176833. doi: 10.1016/j. ejphar.2024.176833

62. Turco L, Reiberger T, Vitale G, la Mura V. Carvedilol as the new non-selective beta-blocker of choice in patients with cirrhosis and portal hypertension. *Liver Int.* (2023) 43:1183–94. doi: 10.1111/liv.15559

63. Lluch P, Segarra G, Medina P. Asymmetric dimethylarginine as a mediator of vascular dysfunction in cirrhosis. *World J Gastroenterol.* (2015) 21:9466–75. doi: 10.3748/wjg.v21.i32.9466

64. Bech JN, Aagaard NK, Pedersen RS, Sorensen TB, Vilstrup H, Pedersen EB. Renal effects of NO-inhibition in patients with cirrhosis vs. healthy controls: a randomized placebo-controlled crossover study. *Liver Int*. (2014) 34:211–9. doi: 10.1111/liv.12233

65. Serna E, Mauricio MD, Lluch P, Segarra G, Cortina B, Lluch S, et al. Basal release of nitric oxide in the mesenteric artery in portal hypertension and cirrhosis: role of dimethylarginine dimethylaminohydrolase. *J Gastroenterol Hepatol.* (2013) 28:880–6. doi: 10.1111/jgh.12119

66. Reiberger T, Payer BA, Schwabl P, Hayden H, Horvatits T, Jäger B, et al. Nebivolol treatment increases splanchnic blood flow and portal pressure in cirrhotic rats via modulation of nitric oxide signalling. *Liver Int.* (2013) 33:561–8. doi: 10.1111/liv.12101

67. Zhu C-P, Liu S-Q, Wang K-Q, Xiong HL, Aristu-Zabalza P, Boyer-Díaz Z, et al. Targeting 5-Hydroxytryptamine receptor 1A in the portal vein to decrease portal hypertension. *Gastroenterology*. (2024) 167:993–1007. doi: 10.1053/j.gastro.2024.06.007

68. Kondo R, Furukawa N, Deguchi A, Kawata N, Suzuki Y, Imaizumi Y, et al. Downregulation of Ca2+-activated cl-Channel TMEM16A mediated by angiotensin II in cirrhotic portal hypertensive mice. *Front Pharmacol.* (2022) 13:831311. doi: 10.3389/fphar.2022.831311

69. Miceli G, Calvaruso V, Casuccio A, Pennisi G, Licata M, Pintus C, et al. Heart rate variability is associated with disease severity and portal hypertension in cirrhosis. *Hepatol Commun.* (2023) 7:e0050. doi: 10.1097/HC9.00000000000000050

70. Ripoll C, Ibáñez-Samaniego L, Neumann B, Vaquero J, Greinert R, Bañares R, et al. Evaluation of the definition of hyperdynamic circulation in patients with cirrhosis and ascites. *Hepatol Commun.* (2022) 6:3528–38. doi: 10.1002/hep4.2102

71. Caracuel L, Sastre E, Llévenes P, Prieto I, Funes T, Aller MÁ, et al. Acute-onchronic liver disease enhances phenylephrine-induced endothelial nitric oxide release in rat mesenteric resistance arteries through enhanced PKA, PI3K/AKT and cGMP signalling pathways. *Sci Rep.* (2019) 9:6993. doi: 10.1038/s41598-019-43513-y

72. Møller S, Nørgaard A, Henriksen JH, Frandsen E, Bendtsen F. Effects of tilting on central hemodynamics and homeostatic mechanisms in cirrhosis. *Hepatology*. (2004) 40:811–9. doi: 10.1002/hep.1840400410

73. Gao Y, Liu X, Gao Y, Duan M, Hou B, Chen Y. Pharmacological interventions for cirrhotic ascites: from challenges to emerging therapeutic horizons. *Gut Liver*. (2024) 18:934–48. doi: 10.5009/gnl240038

74. Tandon P, Abraldes JG, Berzigotti A, Garcia-Pagan JC, Bosch J. Renin-angiotensinaldosterone inhibitors in the reduction of portal pressure: a systematic review and metaanalysis. J Hepatol. (2010) 53:273–82. doi: 10.1016/j.jhep.2010.03.013

75. Gananandan K, Wiese S, Møller S, Mookerjee RP. Cardiac dysfunction in patients with cirrhosis and acute decompensation. *Liver Int.* (2024) 44:1832–41. doi: 10.1111/liv.15896

76. Liu H, Naser JA, Lin G, Lee SS. Cardiomyopathy in cirrhosis: from pathophysiology to clinical care. JHEP Rep. (2024) 6:100911. doi: 10.1016/j.jhepr.2023.100911

77. Maksan S-M, Ryschich E, Ulger Z, Gebhard MM, Schmidt J. Disturbance of hepatic and intestinal microcirculation in experimental liver cirrhosis. *World J Gastroenterol.* (2005) 11:846–9. doi: 10.3748/wjg.v11.i6.846

78. Rochoń J, Kalinowski P, Szymanek-Majchrzak K, Grąt M. Role of gut-liver axis and glucagon-like peptide-1 receptor agonists in the treatment of metabolic dysfunction-associated fatty liver disease. *World J Gastroenterol.* (2024) 30:2964–80. doi: 10.3748/wjg. v30.i23.2964

79. Rusticeanu MA, Zimmer V. Alterations in intestinal mucosal barrier visualized by confocal laser Endomicroscopy in liver cirrhosis: A pilot trial (AMBIC). *Diagnostics* (*Basel*). (2024) 14:1606. doi: 10.3390/diagnostics14151606

80. Zhu X, Zhou Z, Pan X. Research reviews and prospects of gut microbiota in liver cirrhosis: a bibliometric analysis (2001-2023). *Front Microbiol.* (2024) 15:1342356. doi: 10.3389/fmicb.2024.1342356

81. Gioia S, Carnevale R, Tavano D, Overi D, Ridola L, Nardelli S, et al. Association between gut-derived endotoxins and Porto-sinusoidal vascular disorder with portal hypertension. *Aliment Pharmacol Ther.* (2023) 58:1205–16. doi: 10.1111/apt.17727

82. Nagasue N, Dhar DK, Yamanoi A, Emi Y, Udagawa J, Yamamoto A, et al. Production and release of endothelin-1 from the gut and spleen in portal hypertension due to cirrhosis. *Hepatology*. (2000) 31:1107–14. doi: 10.1053/he.2000.6596

83. Du Plessis J, Vanheel H, Janssen CEI, Roos L, Slavik T, Stivaktas PI, et al. Activated intestinal macrophages in patients with cirrhosis release NO and IL-6 that may disrupt intestinal barrier function. *J Hepatol*. (2013) 58:1125–32. doi: 10.1016/j.jhep.2013.01.038

84. Pijls KE, Jonkers DMAE, Elizalde M, Drittij-Reijnders MJ, Haenen GR, Bast A, et al. Is intestinal oxidative stress involved in patients with compensated liver cirrhosis? *Ann Hepatol.* (2016) 15:402–9. doi: 10.5604/16652681.1198816 85. Zhang J, Luo B, Tang L, Wang Y, Stockard CR, Kadish I, et al. Pulmonary angiogenesis in a rat model of hepatopulmonary syndrome. *Gastroenterology*. (2009) 136:1070–80. doi: 10.1053/j.gastro.2008.12.001

86. Rodríguez-Roisin R. Hepatopulmonary syndrome: A forgotten liver-induced lung vascular disorder. Arch Bronconeumol. (2023) 59:137–8. doi: 10.1016/j.arbres.2022.12.007

87. Chin A, Bastaich DR, Dahman B, Kaplan DE, Taddei TH, John BV. Refractory hepatic hydrothorax is associated with increased mortality with death occurring at lower MELD-Na compared to cirrhosis and refractory ascites. *Hepatology.* (2024) 79:844–56. doi: 10.1097/HEP.00000000000577

88. Dandavate V, Bolshette N, Van Drunen R, Manella G, Bueno-Levy H, Zerbib M, et al. Hepatic BMAL1 and HIF1 α regulate a time-dependent hypoxic response and prevent hepatopulmonary-like syndrome. *Cell Metab.* (2024) 36:2038–2053.e5. doi: 10.1016/j.cmet.2024.07.003

89. Douschan P, Kovacs G, Sassmann T, Stadlbauer V, Avian A, Foris V, et al. Pulmonary vascular disease and exercise hemodynamics in chronic liver disease. *Respir Med.* (2022) 202:106987. doi: 10.1016/j.rmed.2022.106987

90. Zhu J, Qiu J, Chen K, Wang W, Zheng S. Tea polyphenols and levofloxacin alleviate the lung injury of hepatopulmonary syndrome in common bile duct ligation rats through endotoxin-TNF signaling. *Biomed Pharmacother*. (2021) 137:111263. doi: 10.1016/j.biopha.2021.111263

91. Raevens S, Boret M, Fallon MB. Hepatopulmonary syndrome. JHEP Rep. (2022) 4:100527. doi: 10.1016/j.jhepr.2022.100527

92. Nascimento ESP, Nunes WMC, Guerra EM, da Roza MR, Silva-Costa S, Machado-Silva W, et al. Combined exercise training improved exercise capacity and lung inflammation in rats with hepatopulmonary syndrome. *Life Sci.* (2021) 287:120112. doi: 10.1016/j.lfs.2021.120112

93. Mangoura SA, Ahmed MA, Hamad N, Zaka AZ, Khalaf KA. Hepatoprotective effects of vildagliptin mitigates lung biochemical and histopathological changes in experimental hepatopulmonary syndrome model in rat. *Int Immunopharmacol.* (2024) 143:113254. doi: 10.1016/j.intimp.2024.113254

94. Li Y-J, Wu X-F, Wang D-D, Li P, Liang H, Hu XY, et al. Serum soluble vascular endothelial growth factor receptor 1 as a potential biomarker of Hepatopulmonary syndrome. *J Clin Transl Hepatol.* (2023) 000–1160. doi: 10.14218/JCTH.2022.00421

95. Horvatits T, Drolz A, Roedl K, Herkner H, Ferlitsch A, Perkmann T, et al. Von Willebrand factor antigen for detection of hepatopulmonary syndrome in patients with cirrhosis. *J Hepatol.* (2014) 61:544–9. doi: 10.1016/j.jhep.2014.04.025

96. Zhang Z-J, Yang C-Q. Progress in investigating the pathogenesis of hepatopulmonary syndrome. *Hepatobiliary Pancreat Dis Int.* (2010) 9:355–60.

97. Krowka MJ, DuBrock HM, Raevens S, Fallon MB. Unmet needs in diagnosis and management of portopulmonary hypertension: how far have we reached? *Hepatology*. (2024). doi: 10.1097/HEP.00000000001218

98. Khoramzadeh M, Dehghanian A, Ketabchi F. Roles of endothelin B receptors and endothelial nitric oxide synthase in the regulation of pulmonary hemodynamic in cirrhotic rats. *J Cardiovasc Pharmacol.* (2019) 73:178–85. doi: 10.1097/ FIC.0000000000000650

99. Nikolic I, Yung L-M, Yang P, Malhotra R, Paskin-Flerlage SD, Dinter T, et al. Bone morphogenetic protein 9 is a mechanistic biomarker of Portopulmonary hypertension. *Am J Respir Crit Care Med.* (2019) 199:891–902. doi: 10.1164/rccm.201807-1236OC

100. Luangmonkong T, Parichatikanond W, Olinga P. Targeting collagen homeostasis for the treatment of liver fibrosis: opportunities and challenges. *Biochem Pharmacol.* (2023) 215:115740. doi: 10.1016/j.bcp.2023.115740

101. Geervliet E, Karkdijk E, Bansal R. Inhibition of intrahepatic monocyte recruitment by Cenicriviroc and extracellular matrix degradation by MMP1 synergistically attenuate liver inflammation and fibrogenesis in vivo. *Sci Rep.* (2024) 14:16897. doi: 10.1038/s41598-024-67926-6

102. Shan L, Wang F, Zhai D, Meng X, Liu J, Lv X. Matrix metalloproteinases induce extracellular matrix degradation through various pathways to alleviate hepatic fibrosis. *Biomed Pharmacother*. (2023) 161:114472. doi: 10.1016/j.biopha.2023.114472

103. Xie H, Su D, Zhang J, Ji D, Mao J, Hao M, et al. Raw and vinegar processed Curcuma wenyujin regulates hepatic fibrosis via bloking TGF- β /Smad signaling pathways and up-regulation of MMP-2/TIMP-1 ratio. *J Ethnopharmacol.* (2020) 246:111768. doi: 10.1016/j.jep.2019.01.045

104. Robert S, Gicquel T, Bodin A, Fautrel A, Barreto E, Victoni T, et al. Influence of inflammasome pathway activation in macrophages on the matrix metalloproteinase expression of human hepatic stellate cells. *Int Immunopharmacol.* (2019) 72:12–20. doi: 10.1016/j.intimp.2019.03.060

105. Ren J-J, Huang T-J, Zhang Q-Q, Zhang HY, Guo XH, Fan HQ, et al. Insulin-like growth factor binding protein related protein 1 knockdown attenuates hepatic fibrosis via the regulation of MMPs/TIMPs in mice. *Hepatobiliary Pancreat Dis Int.* (2019) 18:38–47. doi: 10.1016/j.hbpd.2018.08.008

106. Panera N, Braghini MR, Crudele A, Smeriglio A, Bianchi M, Condorelli AG, et al. Combination treatment with Hydroxytyrosol and vitamin E improves NAFLD-related fibrosis. *Nutrients*. (2022) 14:3791. doi: 10.3390/nu14183791

107. Yaqub F, Latief N, Butt H, Naseer N, Riazuddin S. Alpha lipoic acid priming enhances the hepatoprotective effect of adipose derived stem cells in CCl4 induced

hepatic injury in-vitro. Eur J Pharmacol. (2021) 906:174201. doi: 10.1016/j. ejphar.2021.174201

108. Wang Q, Lu T, Song P, Dong Y, Dai C, Zhang W, et al. Glycyrrhizic acid ameliorates hepatic fibrosis by inhibiting oxidative stress via AKR7A2. *Phytomedicine*. (2024) 133:155878. doi: 10.1016/j.phymed.2024.155878

109. Qin B-F, Zhang J-J, Feng Q-Y, Guo X, Sun HM, Song J. Specnuezhenide ameliorates hepatic fibrosis via regulating SIRT6-mediated inflammatory signaling cascades. *J Ethnopharmacol.* (2024) 335:118646. doi: 10.1016/j.jep.2024.118646

110. Benedicto AM, Lucantoni F, Fuster-Martínez I, Diaz-Pozo P, Dorcaratto D, Muñoz-Forner E, et al. Interference with mitochondrial function as part of the antifibrogenic effect of Rilpivirine: A step towards novel targets in hepatic stellate cell activation. *Biomed Pharmacother*. (2024) 178:117206. doi: 10.1016/j.biopha.2024.117206

111. Lv M, Chen S, Shan M, Si Y, Huang C, Chen J, et al. Arctigenin induces activated HSCs quiescence via AMPK-PPARγ pathway to ameliorate liver fibrosis in mice. *Eur J Pharmacol.* (2024) 974:176629. doi: 10.1016/j.ejphar.2024.176629

112. Wang P, Kang Q, Wu W-S, Rui L. Hepatic Snai 1 and Snai 2 promote liver regeneration and suppress liver fibrosis in mice. *Cell Rep.* (2024) 43:113875. doi: 10.1016/j.celrep.2024.113875

113. Yao L, Hu X, Yuan M, Liu P, Zhang Q, Wang Z, et al. Human umbilical cordderived mesenchymal stromal cells alleviate liver cirrhosis through the hippo/YAP/Id1 pathway and macrophage-dependent mechanism. *Int Immunopharmacol.* (2023) 123:110456. doi: 10.1016/j.intimp.2023.110456

114. Yadav P, Singh SK, Rajput S, Allawadhi P, Khurana A, Weiskirchen R, et al. Therapeutic potential of stem cells in regeneration of liver in chronic liver diseases: current perspectives and future challenges. *Pharmacol Ther.* (2024) 253:108563. doi: 10.1016/j.pharmthera.2023.108563

115. Tapper EB, Zhao Z, Henderson J. Statins for the prevention of cirrhosis complications: an American emulation of the stat liver trial. *Hepatol Commun.* (2024) 8:e0530. doi: 10.1097/HC9.00000000000530

116. Gil M, Khouri L, Raurell I, Rafael D, Andrade F, Abasolo I, et al. Optimization of statin-loaded delivery nanoparticles for treating chronic liver diseases by targeting liver sinusoidal endothelial cells. *Pharmaceutics*. (2023) 15:2463. doi: 10.3390/pharmaceutics15102463

117. Kezer CA, Schmidt KA, Shah VH. Statin the course: navigating unchartered territory in cirrhosis. *Hepatol Commun.* (2024) 8:e0456. doi: 10.1097/HC9.000000000000456

118. Abdelhamid AM, Saber S, Hamad RS, Abdel-Reheim MA, Ellethy AT, Amer MM, et al. STA-9090 in combination with a statin exerts enhanced protective effects in rats fed a high-fat diet and exposed to diethylnitrosamine and thioacetamide. *Front Pharmacol.* (2024) 15:1454829. doi: 10.3389/fphar.2024.1454829

119. Trebicka J, Schierwagen R. Statins, rho GTPases and KLF2: new mechanistic insight into liver fibrosis and portal hypertension. *Gut.* (2015) 64:1349–50. doi: 10.1136/ gutjnl-2014-308800

120. Pose E, Solà E, Lozano JJ, Juanola A, Sidorova J, Zaccherini G, et al. Treatment with simvastatin and Rifaximin restores the plasma Metabolomic profile in patients with decompensated cirrhosis. *Hepatol Commun.* (2022) 6:1100–12. doi: 10.1002/hep4.1881

121. Kronborg TM, Schierwagen R, Trošt K, Gao Q, Moritz T, Bendtsen F, et al. Atorvastatin for patients with cirrhosis. A randomized, placebo-controlled trial. *Hepatol Commun.* (2023) 7:e0332. doi: 10.1097/HC9.00000000000332

122. Muñoz AE, Pollarsky FD, Marino M, Cartier M, Vázquez H, Salgado P, et al. Addition of statins to the standard treatment in patients with cirrhosis: safety and efficacy. *World J Gastroenterol.* (2021) 27:4639–52. doi: 10.3748/wjg.v27.i28.4639

123. Villanueva C, Aracil C, Colomo A, Lopez-Balaguer JM, Piqueras M, Gonzalez B, et al. Clinical trial: a randomized controlled study on prevention of variceal rebleeding comparing nadolol + ligation vs. hepatic venous pressure gradient-guided pharmacological therapy. *Aliment Pharmacol Ther.* (2009) 29:397–408. doi: 10.1111/j.1365-2036.2008.03880.x

124. Berzigotti A, Bellot P, De Gottardi A, Garcia-Pagan JC, Gagnon C, Spénard J, et al. NCX-1000, a nitric oxide-releasing derivative of UDCA, does not decrease portal pressure in patients with cirrhosis: results of a randomized, double-blind, dose-escalating study. *Am J Gastroenterol.* (2010) 105:1094–101. doi: 10.1038/ajg.2009.661

125. Abdel-Sattar AR, Abo-Saif AA, Aboyoussef AM. Nicorandil and atorvastatin attenuate carbon tetrachloride – induced liver fibrosis in rats. *Immunopharmacol Immunotoxicol*. (2020) 42:582–93. doi: 10.1080/08923973.2020.1830104

126. Zipprich A, Gittinger F, Winkler M, Dollinger MM, Ripoll C. Effect of ET-A blockade on portal pressure and hepatic arterial perfusion in patients with cirrhosis: A proof of concept study. *Liver Int.* (2021) 41:554–61. doi: 10.1111/liv.14757

127. Zhang F, Wang F, He J, Lian N, Wang Z, Shao J, et al. Regulation of hepatic stellate cell contraction and cirrhotic portal hypertension by Wnt/ β -catenin signalling via interaction with Gli 1. *Br J Pharmacol.* (2021) 178:2246–65. doi: 10.1111/bph.15289

128. Zhang Y, Xing M, Meng F, Zhu L, Huang Q, Ma T, et al. The mechanical mechanism of angiotensin II induced activation of hepatic stellate cells promoting portal hypertension. *Eur J Cell Biol.* (2024) 103:151427. doi: 10.1016/j.ejcb.2024.151427

129. Wang K, Chen H, Zheng J, Chen J, Chen Y, Yuan Y. Engineered liposomes targeting hepatic stellate cells overcome pathological barriers and reverse liver fibrosis. *J Control Release*. (2024) 368:219–32. doi: 10.1016/j.jconrel.2024.02.022

130. Zhang L-F, Deng W-Q, Huang Q-W, Zhang JJ, Wang Y, Zhou TJ, et al. Vicious cycle-breaking lipid nanoparticles remodeling multicellular crosstalk to reverse liver fibrosis. *Adv Mater*. (2024) 36:e2311474. doi: 10.1002/adma.202311474

131. Xu Y, Wang X-S, Zhou X-L, Lu WM, Tang XK, Jin Y, et al. Mesenchymal stem cell therapy for liver fibrosis need "partner": results based on a meta-analysis of preclinical studies. *World J Gastroenterol.* (2024) 30:3766–82. doi: 10.3748/wjg.v30. i32.3766

132. Ji G, Zhang Z, Wang X, Guo Q, Zhang E, Li C. Comprehensive evaluation of the mechanism of human adipose mesenchymal stem cells ameliorating liver fibrosis by transcriptomics and metabolomics analysis. *Sci Rep.* (2024) 14:20035. doi: 10.1038/ s41598-024-70281-1

133. Calès P, Bertrais S, Boursier J, Fouchard I, Oberti FSNIFF 16 group. Non-selective beta-blockers increase overall and liver mortality in alcoholic cirrhosis with MELD \geq 12 over 5 years of follow-up. *Liver Int*. (2021) 41:168–79. doi: 10.1111/liv.14674

134. Ranjan A, Jindal A, Maiwall R, Vashishtha C, Vijayaraghavan R, Arora V, et al. Midodrine plus propranolol versus propranolol alone in preventing first bleed in patients with cirrhosis and severe ascites: a randomized controlled trial. *Hepatol Int.* (2024) 18:1261–70. doi: 10.1007/s12072-024-10687-1

135. Villanueva C, Sapena V, Lo G-H, Seo YS, Shah HA, Singh V, et al. Improving primary prophylaxis of variceal bleeding by adapting therapy to the clinical stage of cirrhosis. A competing-risk meta-analysis of individual participant data. *Aliment Pharmacol Ther.* (2024) 59:306–21. doi: 10.1111/apt.17824

136. Balcar L, Mandorfer M, Hernández-Gea V, Procopet B, Meyer EL, Giráldez Á, et al. Predicting survival in patients with "non-high-risk" acute variceal bleeding receiving β -blockers+ligation to prevent re-bleeding. *J Hepatol.* (2024) 80:73–81. doi: 10.1016/j.jhep.2023.10.007

137. Gillespie S-L, Hanrahan TP, Rockey DC, Majumdar A, Hayes PC. Review article: controversies surrounding the use of carvedilol and other beta blockers in the management of portal hypertension and cirrhosis. *Aliment Pharmacol Ther.* (2023) 57:454–63. doi: 10.1111/apt.17380

138. Terbah R, Testro AG, Hoermann R, Majumdar A, Chapman B, Gow PJ, et al. Continuous home terlipressin infusion increases handgrip strength and reduces ascites-A prospective randomized crossover study. *Hepatology*. (2024) 80:605–20. doi: 10.1097/HEP.00000000000820

139. Terbah R, Koshy AN, Majumdar A, Vaz K, Testro A, Sinclair M. Long-term continuous Terlipressin infusion improves cardiac Reserve in Patients with Decompensated Cirrhosis. *Clin Gastroenterol Hepatol.* (2024). doi: 10.1016/j. cgh.2024.08.010

140. Rehman H, Rehman ST, Zulfiqar S, Awan S, Abid S. Real-world comparison of terlipressin vs. octreotide as an adjuvant treatment in the management of variceal bleeding. *Sci Rep.* (2024) 14:6692. doi: 10.1038/s41598-024-56873-x

141. Garcia-Tsao G, Abraldes JG, Rich NE, Wong VWS. AGA clinical practice update on the use of vasoactive drugs and intravenous albumin in cirrhosis: expert review. *Gastroenterology*. (2024) 166:202–10. doi: 10.1053/j.gastro.2023.10.016

142. Xue X, Li Y, Zhang S, Yao Y, Peng C, Li Y. Hydroxysafflor yellow A exerts antifibrotic and anti-angiogenic effects through mi R-29a-3p/PDGFRB axis in liver fibrosis. *Phytomedicine*. (2024) 132:155830. doi: 10.1016/j.phymed.2024.155830

143. Pun CK, Huang H-C, Chang C-C, Chuang CL, Hsu SJ, Hou MC, et al. Fructooligosaccharides reverses hepatic vascular dysfunction and dysbiosis in rats with liver cirrhosis and portal hypertension. *Eur J Clin Investig.* (2024) 54:e14287. doi: 10.1111/eci.14287

144. Asada S, Kaji K, Nishimura N, Koizumi A, Matsuda T, Tanaka M, et al. Tofogliflozin delays portal hypertension and hepatic fibrosis by inhibiting sinusoidal Capillarization in cirrhotic rats. *Cells.* (2024) 13:538. doi: 10.3390/ cells13060538

145. Vairappan B, Wright G, Sundhar M, Ravikumar TS. Candesartan cilexetil ameliorates NOSTRIN-NO dependent portal hypertension in cirrhosis and ACLF. *Eur J Pharmacol.* (2023) 958:176010. doi: 10.1016/j.ejphar.2023.176010

146. Tai Y, Zhao C, Zhang L, Tang S, Jia X, Tong H, et al. Celecoxib reduces hepatic vascular resistance in portal hypertension by amelioration of endothelial oxidative stress. *J Cell Mol Med.* (2021) 25:10389–402. doi: 10.1111/jcmm.16968

147. Noah AA, El-Mezayen NS, El-Ganainy SO, Darwish IE, Afify EA, et al. Reversal of fibrosis and portal hypertension by Empagliflozin treatment of CCl4-induced liver fibrosis: emphasis on gal-1/NRP-1/TGF-β and gal-1/NRP-1/VEGFR2 pathways. *Eur J Pharmacol.* (2023) 959:176066. doi: 10.1016/j.ejphar.2023.176066

148. Zheng L, Zhao Z, Lin J, Li H, Wu G, Qi X, et al. Telmisartan relieves liver fibrosis and portal hypertension by improving vascular remodeling and sinusoidal dysfunction. *Eur J Pharmacol.* (2022) 915:174713. doi: 10.1016/j.ejphar.2021.174713

149. Li R, Liu Z, Huang W, Guo Y, Xie C, Wu H, et al. Microbial-derived Urolithin A targets GLS1 to inhibit Glutaminolysis and attenuate cirrhotic portal hypertension. *Cell Mol Gastroenterol Hepatol.* (2024) 18:101379. doi: 10.1016/j. jcmgh.2024.101379

150. Gunarathne LS, Rajapaksha IG, Casey S, Qaradakhi T, Zulli A, Rajapaksha H, et al. Mas-related G protein-coupled receptor type D antagonism improves portal hypertension in cirrhotic rats. *Hepatol Commun.* (2022) 6:2523–37. doi: 10.1002/hep4.1987

151. Wang X, Gu H, Li K, Lin J, Zhu Y, Deng W. DPP4 inhibitor reduces portal hypertension in cirrhotic rats by normalizing arterial hypocontractility. *Life Sci.* (2021) 284:119895. doi: 10.1016/j.lfs.2021.119895

152. Pun CK, Huang H-C, Chang C-C, Chuang CL, Yen CH, Hsu SJ, et al. Glycyrrhizin attenuates portal hypertension and collateral shunting via inhibition of extrahepatic angiogenesis in cirrhotic rats. *Int J Mol Sci.* (2021) 22:7662. doi: 10.3390/ijms22147662

153. Boyer-Diaz Z, Aristu-Zabalza P, Andrés-Rozas M, Robert C, Ortega-Ribera M, Fernández-Iglesias A, et al. Pan-PPAR agonist lanifibranor improves portal hypertension and hepatic fibrosis in experimental advanced chronic liver disease. *J Hepatol.* (2021) 74:1188–99. doi: 10.1016/j.jhep.2020.11.045

154. Brusilovskaya K, Königshofer P, Lampach D, Szodl A, Supper P, Bauer D, et al. Soluble guanylyl cyclase stimulation and phosphodiesterase-5 inhibition improve portal hypertension and reduce liver fibrosis in bile duct-ligated rats. *United European Gastroenterol J.* (2020) 8:1174–85. doi: 10.1177/2050640620944140 155. Tsai Y-L, Liu C-W, Hsu C-F, Huang CC, Lin MW, Huang SF, et al. Obeticholic acid ameliorates hepatorenal syndrome in ascitic cirrhotic rats by down-regulating the renal 8-iso-PGF2α-activated COX-TXA2 pathway. *Clin Sci (Lond)*. (2020) 134:2055–73. doi: 10.1042/CS20200452

156. Hu L, Su L, Dong Z, Wu Y, Lv Y, George J, et al. AMPK agonist AICAR ameliorates portal hypertension and liver cirrhosis via NO pathway in the BDL rat model. *J Mol Med (Berl*). (2019) 97:423–34. doi: 10.1007/s00109-01746-4

157. Castillo GM, Yao Y, Guerra RE, Jiang H, Nishimoto-Ashfield A, Lyubimov AV, et al. Subcutaneous therapy for portal hypertension: PHIN-214, a partial vasopressin receptor 1A agonist. *Biomed Pharmacother*. (2024) 171:116068. doi: 10.1016/j.biopha.2023.116068

158. Jones AK, Chen H, Ng KJ, Villalona J, McHugh M, Zeveleva S, et al. Soluble guanylyl cyclase activator BI 685509 reduces portal hypertension and portosystemic shunting in a rat Thioacetamide-induced cirrhosis model. *J Pharmacol Exp Ther*. (2023) 386:70–9. doi: 10.1124/jpet.122.001532