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RECEIVED 23 May 2024 ACCEPTED 30 October 2024 PUBLISHED 19 November 2024

CITATION

Prosty C, Le M, Lu Y, Khoury L, Cormier M, Cheng MP, Fowler R, Murthy S, Tsang JL, Lejtenyi D, Ben-Shoshan M, Rahme E, Golchi S, Dendukuri N, Lee TC and Netchiporouk E (2024) Effect of omalizumab on inflammatory markers in COVID-19: an exploratory analysis of the COVID-19 immunologic antiviral therapy with omalizumab (CIAO) trial. *Front. Med.* 11:1437322. doi: 10.3389/fmed.2024.1437322

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Effect of omalizumab on inflammatory markers in COVID-19: an exploratory analysis of the COVID-19 immunologic antiviral therapy with omalizumab (CIAO) trial

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Background: The CIAO trial recently demonstrated a probable clinical benefit of omalizumab in the treatment of severe COVID-19; however, the mechanism underlying this benefit remains unclear. Therefore, we sought to longitudinally assess the impact of omalizumab on serum cytokines in CIAO trial patients to determine its mechanism of action.

Methods: Blood samples were collected on days 0, 2, 7, and 14 from patients recruited into the CIAO trial and who consented to this substudy. Blood samples were tested by a panel of 25 inflammatory cytokines, as well as for markers of mast cell activation. Levels of inflammatory biomarkers were compared over time between omalizumab- and placebo-treated patients by generalized linear mixed-effects model. Associations between biomarkers and clinical outcomes were investigated by mixed-effects logistic regression.

Results: Nineteen patients were recruited into this substudy; 10 were assigned to placebo and 9 to omalizumab. Monokine induced by gamma interferon was significantly positively associated with severe COVID-19 (Odds Ratio [OR]=1.06, 95%CI=1.00-1.11, p = 0.043). Further, omalizumab significantly reduced interleukin-15 (Coefficient=-0.95, p = 0.048) and macrophage inflammatory protein-1 (Coefficient=-1.31, p = 0.010) levels. However, neither was significant in analyses adjusting for multiple hypothesis testing.

Conclusion: Although limited by a small sample size, these results suggest that omalizumab's potential benefit in COVID-19 may be mediated independently of modulation of the measured serum biomarkers. Further studies are needed to investigate omalizumab's mechanism of action in COVID-19.

KEYWORDS omalizumab, coronavirus, COVID-19, clinical trial, cytokine

Introduction

Coronavirus disease 2019 (COVID-19) has had significant health and economic repercussions globally (1). The pathogenesis of COVID-19 is biphasic involving viral replication and dissemination followed by immune activation (2). Severe COVID-19 is postulated to involve pathological hyperinflammation (2, 3), which has been associated with impaired type I and III interferon (IFN) signaling (4-6). Type II inflammation has also been associated with severe disease (7). A small randomized controlled trial (RCT) studying dupilumab, a monoclonal antibody targeting IL4R- α , suggested a possible mortality benefit from inhibition of the type II inflammatory response in severe COVID-19 (8). Therefore, IFN and type II inflammation-targeting agents demonstrate potential as COVID-19 therapies.

Omalizumab is a monoclonal antibody that binds IgE and inhibits its interaction with the FceRI receptor on immune cells (9), used for refractory chronic spontaneous urticaria (10), asthma (11), and nasal polyps (12). Recent data suggest that omalizumab may have antiviral activity (13-15). In asthmatic patients, omalizumab significantly decreased the incidence of upper respiratory tract infections and the duration of viral shedding (15, 16). The postulated mechanism of action was inhibition of IgE-FceRI interaction augmenting type I IFN signaling (17, 18). Omalizumab has also been shown to attenuate type II inflammation/mast cell activation in asthmatic patients (13). In light of this evidence, the COVID-19 Immunologic Antiviral therapy with Omalizumab (CIAO) trial was conducted to assess the efficacy of omalizumab in moderate to severe COVID-19 (19). Despite early termination due to waning recruitment, among the 40 patients enrolled, omalizumab was associated with a 93% probability of reduction in death or mechanical ventilation on day 14 (19). The mechanism by which omalizumab conferred this benefit remains unclear. Thus, we sought to investigate omalizumab's mechanism of action in COVID-19 by analyzing its impact on longitudinal levels of various biomarkers.

Methods

Trial design

This substudy focused on the exploratory endpoints of the CIAO trial (19). Ethics approval was obtained from all participating sites and informed consent was acquired for each subject. Briefly, the CIAO trial was a multi-center randomized double-blind trial comparing a single dose of omalizumab 375 mg plus standard of care vs. placebo with standard of care in patients hospitalized for COVID-19 respiratory illness. The primary outcome was a composite of mechanical ventilation or all-cause mortality at 14 days with secondary endpoints including the time to clinical improvement of COVID-19 evaluated using the WHO 9-point ordinal scale (20). Data for this nested study were obtained

from the subset of patients recruited at the McGill University Health Centre (MUHC), who consented to have blood samples drawn during their hospitalization on days 0, 2, 7, and 14 $(\pm 2 \text{ days})$.

Sample processing

Samples were centrifuged and the serum was isolated and stored at -80° C for subsequent batched analysis. The total IgE (IgE) (day 0), tryptase (day 0), and C-reactive protein (CRP) (days 0, 2, 7, 14) assays were performed by the MUHC central laboratory. CD63 expression, a marker of basophil activation, was measured using the CD63 flocast flow cytometry assay (Buhlmann; Schonenbuch, Switzerland), as previously described, on days 0, 2, 7, and 14 (21). A panel of 25 different cytokines was measured for all samples (days 0, 2, 7, and 14) using the cytokine 25-plex human panel immunoassay according to manufacturer's instructions (Invitrogen; Vienna, Austria).

Statistical analyses

All statistical analyses were performed using R (4.3.2, Foundation for Statistical Computing, Vienna, Austria). Demographic and clinical data were summarized with descriptive statistics. Mixed-effects logistic regression was used to determine whether measured biomarkers were associated with severe COVID-19 (WHO ordinal COVID-19 score 5-7). In this model, patient identification and study day were random-effect and fixedeffect variables, respectively. Generalized linear mixed-effects models (GLMM) with a gamma distribution were used to model biomarkers over time using the *lme4* package (22). Fixed-effect terms were used for study day and omalizumab assignment and a randomeffect term for patient identification. A likelihood ratio test was performed to determine omalizumab's effect on biomarkers over time. p-values in each model were adjusted for multiple hypothesis testing separately using the Benjamini-Hochberg method (Q-value). Line graphs of the cytokine levels over time were plotted using the ggplot2 package (23).

Results

Patient demographics

Of the 40 patients recruited into the CIAO trial, 19 were recruited at the MUHC and consented to blood tests for this substudy. Nine patients were assigned to omalizumab and 10 to placebo. The patient demographics, comorbidities, vaccination status, receipt of concomitant interventions, and WHO COVID-19 ordinal scores are presented in Supplementary Table 1. The median age of the study population was 65.0 years (interquartile range 59.5–80.0) and 31.6% were female. Cardiovascular disease (63.2%), diabetes (52.6%), chronic kidney disease (26.3%), and cancer (21.1%) were the most common comorbidities. The majority (72.2%) of patients were vaccinated against COVID-19. At enrollment WHO ordinal severity scores for COVID-19 were most commonly 4 (36.8%), 3 (31.6%), 5 (21.1%), and 2 (10.5%). Concomitant COVID-19 therapies were received by all patients, including dexamethasone (100.0%), remdesivir (57.9%), tocilizumab (5.3%), and baricitinib (5.3%). The primary outcome, death or mechanical ventilation at day 14, occurred in 4 patients (21.1%).

Biomarker comparisons

Blood samples were obtained for 100% of participants on day 0, 73.7% on day 2, 52.6% on day 7, and 31.6% on day 14.

In unadjusted analyses, monokine induced by gamma interferon (MIG; OR=1.06, 95%CI=1.00–1.11, p = 0.043, Q=0.77) was significantly positively associated with severe COVID-19, but was not significant when adjusting for multiple hypothesis testing (Table 1). None of the remaining biomarkers were statistically significantly associated with the outcome.

Mean biomarker values over time are plotted in Figure 1 for omalizumab vs. placebo. In unadjusted analyses, omalizumab significantly decreased interleukin (IL)-15 (Coefficient = -0.95, p = 0.048, Q = 0.41) and macrophage inflammatory protein-1 (MIP-1; Coefficient = -1.31, p = 0.010, Q = 0.27); however, these were not statistically significant after adjustment. Receipt of omalizumab did not significantly affect any of the other 25 biomarkers Supplementary Table 2.

Discussion

After adjustment, we did not find a statistically significant association between the biomarkers studied and disease severity or any significant differences in biomarkers over time as a function of omalizumab treatment.

Previous studies have noted that SARS-CoV2-specific IgE are correlated with COVID-19 severity (24, 25). While these results, combined with omalizumab's possible efficacy in COVID-19 may suggest a specific role for IgE or type II immunity in the pathogenesis of COVID-19, they may also merely be indicative of global immune dysregulation (7). Indeed, we did not identify any significant associations between disease severity and type II inflammatory cytokines, IFN- α , and markers of mast cell activation, in this study with limited sample size. Other studies on the subject revealed conflicting results with one study suggesting a positive association between type II inflammation and COVID-19 mortality (26), whereas another failed to detect a significant association between type II/type I inflammatory cytokine imbalance and death (27).

Our findings suggest that omalizumab's possible clinical benefit in COVID-19 is mediated by mechanisms other than modulation of serum levels of IFN- α or type II inflammation. These results contrast those for dupilumab in COVID-19, which demonstrated a probable clinical benefit of dupilumab and an associated reduction in type II inflammatory cytokines (e.g., eotaxin-3 and YKL-40) compared to placebo (8). Notwithstanding our study's limitations, there are several alternative explanations for these findings. All patients received

concomitant corticosteroids which are known to broadly suppress inflammatory cytokines, including type II inflammation (28), and this may have obscured any omalizumab-induced differences and biased results toward the null. Alternatively, Djukanović et al. demonstrated that, compared to placebo, omalizumab treatment significantly reduced IL-4+ staining cells and eosinophils in bronchial biopsies in asthmatic patients (13). Therefore, omalizumab's effect may be mediated by immune modulation at the tissue rather than serum level. Additionally, the assay used for cytokine detection in this study does not analyze the levels of certain type II inflammatory cytokines (e.g., IL-9) and type I interferons (e.g., IFN- β), which have been shown in other studies to be modulated by omalizumab (18, 29). Consequently, we cannot exclude the possibility that omalizumab affected unmeasured cytokines.

TABLE 1 Mixed-effects logistic regression of biomarkers associated with severe COVID-19 (WHO ordinal COVID-19 score 5–7).

Biomarker	OR (95%CI)	<i>p</i> -value	Q-value
CD63+ (%)	1.59 (0.91–2.78)	0.10	0.77
CRP (mg/L)	1.03 (0.97–1.10)	0.28	0.95
Eotaxin (pg/ml)	0.72 (0.33-1.57)	0.40	0.95
GM-CSF (pg/ml)	NA	NA	NA
IFN-alpha (pg/ml)	0.98 (0.86-1.13)	0.82	0.95
IFN-gamma (pg/ml)	NA	NA	NA
IL-1 beta (pg/ml)	NA	NA	NA
IL-10 (pg/ml)	NA	NA	NA
IL-12/IL-23 (pg/ml)	1.05 (1.00-1.09)	0.072	0.77
IL-13 (pg/ml)	1.17 (0.70–1.93)	0.55	0.95
IL-15 (pg/ml)	0.97 (0.88-1.08)	0.58	0.95
IL-17A (pg/ml)	0.80 (0.49-1.31)	0.37	0.95
IL-1RA (pg/ml)	1.00 (0.99–1.00)	0.83	0.95
IL-2 (pg/ml)	0.99 (0.93-1.06)	0.82	0.95
IL-2R (pg/ml)	1.00 (1.00-1.00)	0.81	0.95
IL-4 (pg/ml)	1.00 (0.97-1.03)	0.94	0.95
IL-5 (pg/ml)	NA	NA	NA
IL-6 (pg/ml)	0.96 (0.90-1.03)	0.28	0.95
IL-7 (pg/ml)	0.89 (0.71–1.11)	0.29	0.95
IL-8 (pg/ml)	1.00 (0.98-1.02)	0.79	0.95
IP-10 (pg/ml)	1.00 (0.86-1.12)	0.95	0.95
MCP-1 (pg/ml)	1.00 (0.99–1.01)	0.54	0.95
MIG (pg/ml)	1.06 (1.00–1.11)	0.043	0.77
MIP-1 alpha (pg/ml)	1.00 (0.94–1.06)	0.87	0.95
MIP-1 beta (pg/ml)	1.00 (0.98-1.02)	0.72	0.95
RANTES (pg/ml)	1.00 (1.00-1.00)	0.93	0.95
TNF-alpha (pg/ml)	NA	NA	NA
Tryptase (ug/mL)*	0.90 (0.67-1.20)	0.47	0.95
IgE (ug/L)*	1.00 (1.00-1.01)	0.14	0.81

*IgE and tryptase were only performed on day 0; therefore, only a conventional logistic regression was performed for these laboratories.

NA: Not applicable. Too few values above the threshold of detection to perform a logistic regression.



This study has multiple strengths including being conducted within the context of a randomized double-blind RCT, which minimized confounding by severity. Analyses were adjusted for multiple hypothesis testing to reduce the risk of type I error. Despite these strengths, this study is subject to limitations. First, the study population was small which may increase the risk of type II error. Second, the substudy population was further reduced by a loss to follow-up for blood sample collection on later study days, due to death or discharge. Third, many of the cytokine levels fell beneath the limit of detection of the assay, possibly due to concomitant receipt of corticosteroids.

Conclusion

This study suggests that the possible mortality benefit demonstrated by omalizumab in the CIAO trial could be mediated independently of modulation of serum IFN- α and type II inflammatory cytokines. Further studies are necessary to elucidate the mechanism of action of omalizumab in COVID-19 and other viral illnesses.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by McGill Research Ethics Board, McGill University, Montreal, Canada. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

CP: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources, Software, Validation, Visualization, Writing - original draft, Writing - review & editing. ML: Conceptualization, Data curation, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Writing - original draft, Writing - review & editing. YL: Data curation, Formal analysis, Methodology, Validation, Writing - original draft, Writing review & editing. LK: Data curation, Investigation, Project administration, Writing - original draft, Writing - review & editing. MC: Data curation, Investigation, Writing - original draft, Writing - review & editing. MPC: Investigation, Project administration, Writing - original draft, Writing review & editing. RF: Data curation, Investigation, Writing - original draft, Writing - review & editing. SM: Investigation, Writing - original draft, Writing - review & editing. JT: Investigation, Writing - original draft, Writing - review & editing. DL: Investigation, Validation, Writing - original draft, Writing - review & editing. MB-S: Investigation, Writing - original draft, Writing - review & editing. ER: Methodology, Validation, Writing - original draft, Writing - review & editing. SG: Formal analysis, Writing - original draft, Writing - review & editing. ND: Formal analysis, Writing - original draft, Writing - review & editing. TL: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Supervision, Validation, Writing - original draft, Writing - review & editing. EN: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Validation, Writing - original draft, Writing - review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. The study was

References

1. WHO. Coronavirus (COVID-19) Dashboard. (2023). Available at: https://covid19. who.int (Accessed August 27)

funded by the Ministre de l'Économie et Innovation/Fonds de Recherche Santé Québec and the MUHC Foundation. Novartis donated the study drug.

Conflict of interest

ML reports funding support from the Canadian Institutes of Health Research (CIHR) Vanier Doctoral Scholarship. MC has been an Advisory Board/Speaker/Consultant for GSK, AstraZeneca, Sanofi/Regeneron, Novartis, Boehringer-Ingelheim, Trudell, Valeo, and COVIS. MPC reports grants from the McGill Interdisciplinary Initiative in Infection and Immunity and personal fees from GEn1E Lifesciences (as a member of the scientific advisory board) and personal fees from nplex biosciences (as a member of the scientific advisory board). JT reports a Physicians' Services Incorporated Foundation grant to her institution, outside the submitted work. She is co-chair of the Canadian Community ICU Research Network (CCIRNet) and vice-chair of the Quest Community Health Centre board of directors. SM reports a grant from the Health Research Foundation, Innovative Medicines Canada. MB-S reports salary support from Fonds de recherche du Québec - Santé. MB-S is part of the advisory Board or equivalent: Bausch, Stallergenes, Novartis, and Sanofi. Participating or participated in a clinical trial: Novartis, Aimmune, & Sanofi. ER reports funds and consulting fees from Janssen in the course of an unrelated study. TL reports salary support from Fonds de recherche du Québec - Santé. EN has been an Advisory Board/ Speaker/Consultant and/or received Investigator Initiated Educational and/or Research funding from AbbVie Inc., Bausch Health, Beiersdorf, Boehringer Ingelheim International, Bristol Myers Squibb, Eli Lilly, Galderma SA., Janssen Inc., LEO Pharma, Medexus, Novartis Pharmaceuticals, Pfizer Inc., Sanofi Genzyme, Sun Pharmaceuticals, and UCB.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed.2024.1437322/ full#supplementary-material

2. Lamers MM, Haagmans BL. SARS-CoV-2 pathogenesis. Nat Rev Microbiol. (2022) 20:270–84. doi: 10.1038/s41579-022-00713-0

3. del Valle DM, Kim-Schulze S, Huang HH, Beckmann ND, Nirenberg S, Wang B, et al. An inflammatory cytokine signature predicts COVID-19 severity and survival. *Nat Med.* (2020) 26:1636–43. doi: 10.1038/s41591-020-1051-9

4. Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann HH, Zhang Y, et al. Autoantibodies against type I IFNs in patients with life-threatening COVID-19. *Science*. (2020) 370. doi: 10.1126/science.abd4585

5. Zhang Q, Bastard P, Liu Z, le Pen J, Moncada-Velez M, Chen J, et al. Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. *Science*. (2020) 370. doi: 10.1126/science.abd4570

 Hadjadj J, Yatim N, Barnabei L, Corneau A, Boussier J, Smith N, et al. Impaired type I interferon activity and inflammatory responses in severe COVID-19 patients. *Science*. (2020) 369:718–24. doi: 10.1126/science.abc6027

7. Lucas C, Wong P, Klein J, Castro TBR, Silva J, Sundaram M, et al. Longitudinal analyses reveal immunological misfiring in severe COVID-19. *Nature*. (2020) 584:463–9. doi: 10.1038/s41586-020-2588-y

8. Sasson J, Donlan AN, Ma JZ, Haughey HM, Coleman R, Nayak U, et al. Safety and efficacy of Dupilumab for the treatment of hospitalized patients with moderate to severe coronavirus disease 2019: a phase 2a trial. *Open Forum Infect Dis.* (2022) 9. doi: 10.1093/ ofid/ofac343

9. Easthope S, Jarvis B. Omalizumab. Drugs. (2001) 61:253-60. doi: 10.2165/00003495-200161020-00008

10. Maurer M, Rosén K, Hsieh HJ, Saini S, Grattan C, Gimenéz-Arnau A, et al. Omalizumab for the treatment of chronic idiopathic or spontaneous Urticaria. *N Engl J Med.* (2013) 368:924–35. doi: 10.1056/NEJMoa1215372

11. Busse W, Corren J, Lanier BQ, McAlary M, Fowler-Taylor A, Cioppa GD, et al. Omalizumab, anti-IgE recombinant humanized monoclonal antibody, for the treatment of severe allergic asthma. *J Allergy Clin Immunol.* (2001) 108:184–90. doi: 10.1067/ mai.2001.117880

12. Gevaert P, Omachi TA, Corren J, Mullol J, Han J, Lee SE, et al. Efficacy and safety of omalizumab in nasal polyposis: 2 randomized phase 3 trials. *J Allergy Clin Immunol.* (2020) 146:595–605. doi: 10.1016/j.jaci.2020.05.032

13. Djukanović R, Wilson SJ, Kraft M, Jarjour NN, Steel M, Chung KF, et al. Effects of treatment with anti-immunoglobulin E antibody Omalizumab on airway inflammation in allergic asthma. *Am J Respir Crit Care Med.* (2004) 170:583–93. doi: 10.1164/rccm.200312-1651OC

14. Cardet JC, Casale TB. New insights into the utility of omalizumab. J Allergy Clin Immunol. (2019) 143:923–926.e1. doi: 10.1016/j.jaci.2019.01.016

15. Konstantinou GN, Podder I, Karapiperis D. Omalizumab prevents respiratory illnesses in non-atopic chronic spontaneous urticaria patients: a prospective, parallel-group, pilot pragmatic trial. *Clin Transl Allergy*. (2023) 13:e12279. doi: 10.1002/clt2.12279

16. Esquivel A, Busse WW, Calatroni A, Togias AG, Grindle KG, Bochkov YA, et al. Effects of Omalizumab on rhinovirus infections, illnesses, and exacerbations of

asthma. Am J Respir Crit Care Med. (2017) 196:985–92. doi: 10.1164/ rccm.201701-0120OC

17. Gill MA, Bajwa G, George TA, Dong CC, Dougherty II, Jiang N, et al. Counterregulation between the FcepsilonRI pathway and antiviral responses in human plasmacytoid dendritic cells. *J Immunol Baltim Md 1950*. (2010) 184:5999–6006. doi: 10.4049/jimmunol.0901194

18. Gill MA, Liu AH, Calatroni A, Krouse RZ, Shao B, Schiltz A, et al. Enhanced plasmacytoid dendritic cell antiviral responses after omalizumab. *J Allergy Clin Immunol.* (2018) 141:1735–1743.e9. doi: 10.1016/j.jaci.2017.07.035

19. Le M, Khoury L, Lu Y. COVID-19 immunologic antiviral therapy with Omalizumab (CIAO) – a randomized-controlled clinical trial. *Open Forum Infect Dis.* (2024). 11. doi: 10.1093/ofid/ofae102

20. Marshall JC, Murthy S, Diaz J, Adhikari NK, Angus DC, Arabi YM, et al. A minimal common outcome measure set for COVID-19 clinical research. *Lancet Infect Dis.* (2020) 20:e192–7. doi: 10.1016/S1473-3099(20)30483-7

21. Netchiporouk E, Moreau L, Rahme E, Maurer M, Lejtenyi D, Ben-Shoshan M. Positive CD63 basophil activation tests are common in children with chronic spontaneous Urticaria and linked to high disease activity. *Int Arch Allergy Immunol.* (2016) 171:81–8. doi: 10.1159/000451084

22. Bates D, Mächler M, Bolker B, Walker S. Fitting linear mixed-effects models using lme4. J Stat Softw. (2015) 67:1-48. doi: 10.18637/jss.v067.i01

23. Wickham H. ggplot2: Elegant Graphics for Data Analysis | SpringerLink. (2009). Available at: https://link.springer.com/book/10.1007/978-3-319-24277-4 (Accessed January 18, 2024).

24. Plūme J, Galvanovskis A, Šmite S, Romanchikova N, Zayakin P, Linē A. Early and strong antibody responses to SARS-CoV-2 predict disease severity in COVID-19 patients. *J Transl Med.* (2022) 20:176. doi: 10.1186/s12967-022-03382-y

25. Tan C, Zheng X, Sun F, He J, Shi H, Chen M, et al. Hypersensitivity may be involved in severe COVID-19. *Clin Exp Allergy*. (2022) 52:324–33. doi: 10.1111/cea.14023

26. Baker JR, Mahdi M, Nicolau DV, Ramakrishnan S, Barnes PJ, Simpson JL, et al. Early Th2 inflammation in the upper respiratory mucosa as a predictor of severe COVID-19 and modulation by early treatment with inhaled corticosteroids: a mechanistic analysis. *Lancet Respir Med.* (2022) 10:545–56. doi: 10.1016/S2213-2600(22)00002-9

27. Pavel AB, Glickman JW, Michels JR, Kim-Schulze S, Miller RL, Guttman-Yassky E. Th2/Th1 cytokine imbalance is associated with higher COVID-19 risk mortality. *Front Genet.* (2021) 12:12. doi: 10.3389/fgene.2021.706902

28. Braun CM, Huang SK, Bashian GG, Kagey-Sobotka A, Lichtenstein LM, Essayan DM. Corticosteroid modulation of human, antigen-specific Th1 and Th2 responses. J Allergy Clin Immunol. (1997) 100:400–7. doi: 10.1016/S0091-6749(97)70255-0

29. Takaku Y, Soma T, Nishihara F, Nakagome K, Kobayashi T, Hagiwara K, et al. Omalizumab attenuates airway inflammation and interleukin-5 production by mononuclear cells in patients with severe allergic asthma. *Int Arch Allergy Immunol.* (2013) 161:107–17. doi: 10.1159/000350852