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EDITED BY

Ian James Martins,
University of Western Australia, Australia

REVIEWED BY

Xiaolei Li,
University of Pennsylvania, United States
Xianfang Rong,
IMiracle, China

*CORRESPONDENCE

Yan Tian
✉ 905790434@qq.com
Hua-you Luo
✉ km-lhy@qq.com

[†]These authors have contributed equally to this work and share first authorship

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The function of the inter-alpha-trypsin inhibitors in the development of disease

Xin-feng Zhang^{1†}, Xiao-li Zhang^{1†}, Li Guo¹, Yun-ping Bai², Yan Tian^{1*} and Hua-you Luo^{1*}

¹Department of Gastrointestinal and Hernia Surgery, The First Affiliated Hospital of Kunming Medical University, Kunming, China, ²Department of Otolaryngology, The First Affiliated Hospital of Kunming Medical University, Kunming, China

Through the formation of covalent connections with hyaluronic acid (HA), the inter- α -trypsin inhibitor ($I\alpha I$) family collaborates to preserve the stability of the extracellular matrix (ECM). The five distinct homologous heavy chains (ITI_H) and one type of light chain make up the $I\alpha I$ family. ITI_H alone or in combination with bikunin (BK) has been proven to have important impacts in a number of earlier investigations. This implies that BK and ITI_H might be crucial to both physiological and pathological processes. The functions of BK and ITI_H in various pathophysiological processes are discussed independently in this paper. In the meanwhile, this study offers suggestions for further research on the roles of BK and ITI_H in the course of disease and summarizes the plausible mechanisms of the previous studies.

KEYWORDS

inter- α -trypsin inhibitor ($I\alpha I$) family, bikunin (BK), ITI_Hs, hyaluronic acid (HA), extracellular matrix (ECM)

1 Introduction

Members of the ancient and distinctive inter- α -trypsin inhibitor ($I\alpha I$) family have developed throughout hundreds of millions of years of vertebrate history (1). Hepatocytes are the primary source of the 225 kDa $I\alpha I$ family protein complexes, which are found in the blood at high concentrations of 0.15 to 0.5 mg/mL (2). Human $I\alpha I$ family consists of three different polypeptide chains, namely bikunin (BK), heavy chain 1 and heavy chain 2 (3). $I\alpha I$ family, which makes up the majority of family members in human serum, is thought to be dormant until it enters the target tissue, where it is cleaved by TNF-stimulated gene 6 protein (TSG-6). After that, heavy chains (HCs) are transferred to hyaluronic acid (HA), a significant part of the extracellular matrix (ECM), through the formation of temporary covalent bonds with TSG-6 (4). TSG-6 is essential for the interaction with HA because it facilitates two following ester exchange reactions: it binds HC1 or HC2 of $I\alpha I$ family covalently and then moves them to the HA fraction in this complex, where the heavy chain conjugates and releases free TSG-6 (5, 6). HCs have been found to function as structural proteins that can directly cross-link HA that is secreted (7). $I\alpha I$ HC forms a strong bond with HA produced by fibroblasts in culture (7). In addition, the physiological correlation of HA with members of the $I\alpha I$ HC family has been linked to various cell types that show HA-containing outer membranes. For example, the maturation process of oocytes and experiments *in vitro* culture of mesothelial cells (8). In summary, the $I\alpha I$ family of proteins interacts with HA to maintain the stability of the ECM, which is a critical function in numerous illnesses. This work aims to provide an overview of

the mechanisms underlying the occurrence and development of I α I family proteins that have been linked to current disorders. Additionally, it offers suggestions for future research on I α I family proteins.

2 The structure and function of inter- α -trypsin inhibitor family members

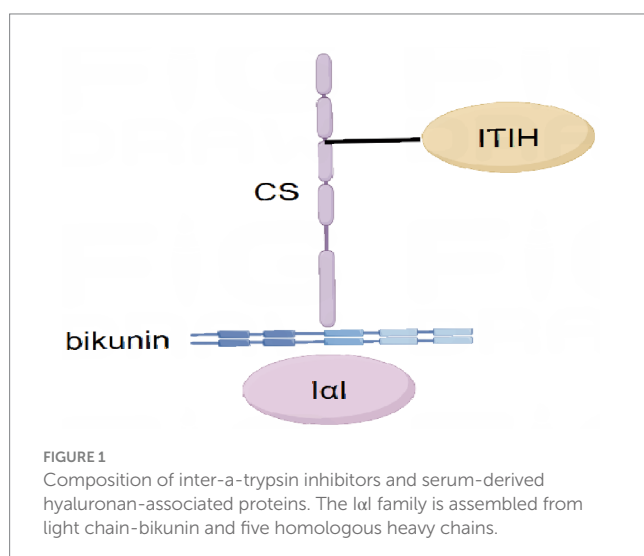
2.1 Bikunin

Bikunin (BK), the light chain, and five homologous heavy chains come together to form the I α I family (9) (Figure 1). There are various homologous heavy chains (ITIH), and the one that has been discovered to date has five members (ITIH1, ITIH2, ITIH3, ITIH4, and ITIH5), despite the fact that there is only one type of light chain. The α -1-microglobulin/BK precursor (AMBP) encodes the light chain and α -1 microglobulin, a member of the lipid transport protein superfamily independent of the ITI family either physically or functionally (10). The ITI light chain is referred to as “BK” because it has two tandem repeats of a Kunitz-type structural domain (11). Serum proteoglycans generated from liver are known as BK isoforms. They contain chondroitin sulfate (CS) chains, which are mainly esterified by one or two glycoproteins referred to as “heavy chains” (HC). There are three primary serum isoforms that have been extensively documented: pro-alpha-proteinase inhibitor and inter-alpha-trypsin inhibitor, which carry one and two HC, respectively, and urotrypsin inhibitor, which corresponds to BK and is connected to the CS chain (10). These complexes create the characteristic ITI proteoglycosaminoglycan-protein structure (12).

2.2 ITIH

Apart from BK, the ITI heavy chain also serves biological purposes (13). Numerous diseases, including inflammatory reactions in local tissues, acute inflammation, tumor growth, and problems connected to psychiatry, have been linked to ITIHs, according to

studies (13, 14). Subsequent investigations have also revealed that ITIHs, a main constituent of ITI, plays an important role, either by itself or in conjunction with BK (13). The von Willebrand A-type structural domains and the vault are two of the modules that make up the H chain, which can communicate with the ECM. The carboxyl group of the matching H-chain’s C-terminal aspartic acid residue and the C-6 hydroxyl group of the internal N-acetylamino galactose residue of BK’s chondroitin sulfate component are what link the H-chain to the protein. ITIHs can undergo an ester exchange reaction to covalently conjugate to locally generated HA. Serum-derived hyaluronan-associated protein is made up of D-glucuronic acid and N-acyl-D-glucosamine, which together make up the desialylated polymer known as HA (2). ITIHs bind covalently to HA to stabilize the ECM (15). Many extracellular proteins released by cells make up the ECM, which controls intercellular communication as well as biological activities (16). Many interacting proteins and proteoglycans that control cellular activity have an impact on the stiffness or rigidity of HA (17, 18). One important element of the ECM is HA (19). Together, these molecules of the ECM—HA, proliferating cells, migration, and tumor metastasis—maintain the stability of the ECM by forming a reticular structure (20, 21). A common ECM component, HA is a high molecular weight polymer that does not need to be modified further (21). Numerous biological processes, including wound healing, differentiation, and cell motility, are facilitated by HA-binding proteins, especially membrane-bound receptors like CD44 and RHAMM (22, 23). For instance, during the healing of skin injury, synthesis is elevated. The covalent transfer of heavy chains from I α I to HA is catalyzed by TsG-6, and the resulting HC-HA complex is engaged in remodeling and inflammatory processes in both healthy and pathological contexts (24). Furthermore, HA participates in angiogenic processes. The size of HA affects its angiogenic potential: short segments of HA produced in inflammation and tissue damage are strongly angiogenic (25), while high molecular weight HA possesses vasopressor qualities (26). After degradation, HA becomes a pro-angiogenic ligand (15). For instance, it has been discovered in earlier research that HA stimulates angiogenesis in lung damage and is even linked to abnormal angiogenesis (17). Increased ITIHs protein may prevent the HA-CD44 and ECM pathways from degrading, which would have anti-angiogenic effects. This could be one of the key ways that the protein ITIHs functions as a putative oncogene.



3 Advances in the study of BK and disease

In both physiological and pathological processes, BK has pleiotropic functions. BK appears to be involved in numerous activities, according to genetic studies of mice deficient the protein. Genes related to stress, apoptosis, proteases, aging, cytokines, HA metabolism, and female ovulation processes are dysregulated when BK is absent (27, 28). Subsequent research revealed that female mice deficient the BK gene exhibit considerably lower fertility (28). This results from a malfunction in the lateral protein precursors’ ability to form compounds with hyaluronic acid in the ovarian mound before ovulation. Therefore, hyaluronan, which is necessary for mammalian ovulation and fertilization, requires the delivery of serum-derived hyaluronan-related proteins by the chondroitin sulfate part of BK (29). Concurrently, research has produced intriguing findings on the

potential of the medication BK to lower the risk of preterm labor and enhance neonatal outcomes (30).

Furthermore, in pancreatitis, septic shock, and rheumatoid arthritis, it has been demonstrated that the BK core protein inhibits inflammation-associated proteases such as trypsin, elastase, and fibrinolytic enzymes (31, 32). As an anti-inflammatory, BK prevents the production of cytokines that are triggered by lipopolysaccharide (LPS). Through extracellular signal-regulated kinase signaling (ERK), calcium endocytosis, and endotoxin receptors, BK suppresses the generation of cytokines. Endotoxin stimulates calcium inward flow, generates phosphorylated ERK, and activates multiple transcription factors, including nuclear factor kappa B and early growth response-1, through endotoxin receptor signaling, all of which support the development of cytokines (33). It was discovered to prevent the generation of pro-inflammatory cytokines in a number of cell types in cellular tests (3, 34). Mice injected BK showed a decrease in multiple inflammatory markers in animal models of inflammatory disorders (35). Furthermore, BK knockout mice, *Bik* (−/−), were used for *in vitro* cytokine experiments and *in vivo* animal models. It was discovered that *Bik* (−/−) mice induced higher levels of endotoxin-induced death when compared to wild-type (Wt) mice; that application of BK significantly reduced LPS-induced lethality; and that endotoxin significantly increased endotoxin-induced lethality when compared to Wt mice. BK combined with application of BK inhibited the levels of these cytokines; additionally, BK inhibited endotoxin-induced up-regulation of cytokine expression by suppressing macrophage phosphorylation of ERK1/2, JNK, and p38; this implies that BK has a major anti-inflammatory function (36). In different acute and chronic inflammatory reactions, BK is a non-invasive circulating or urine biomarker (37).

BK has been found in cancer studies to inhibit tumor cell invasion through direct inhibition of fibrinolytic activity associated with tumor cells as well as urokinase-type fibrinogen activator (UPA) expression at the gene and protein levels, potentially through inhibition of MAP kinase signaling cascades and/or CD44 dimer. By interacting with different cell types' cartilage junction proteins and BK receptors, BK can be suppressed in order to prevent cell invasion (38). Furthermore, ovarian cancer cells' gene expression patterns are changed by BK, which prevents tumor cell invasion (39). In animal investigations, it was discovered that the exogenous injection of BK inhibited the growth of intraperitoneal ovarian tumors as well as peritoneal disseminated metastases (40). Treatment with BK in the adjuvant setting and/or in conjunction with cytotoxic medicines to enhance therapeutic efficacy may be helpful in postponing the beginning of metastases in patients with advanced ovarian cancer (41). Patients with ovarian cancer who had preoperative BK concentrations higher than 11.5 µg/mL were shown to have a significantly better prognosis than those with lower amounts. Patients with pretreatment values of 11.5 µg/mL had 2.2-fold higher Hazard Ratios (risk of mortality) than those with concentrations of more than 11.5 µg/mL of BK. Patients in both groups had a median survival of 26 months and more than 60 months, respectively (42). This implies that BK has a critical function and importance in determining the prognosis of ovarian cancer patients. Remarkably, there has been no report of BK overexpression being linked to human pathology (1).

The above cellular and animal experiments suggest that BK has potential medical value. It also plays a significant role in mammalian

ovulation and even directly affects fertility in knockout mice. Additionally, BK has important roles and functions as an anti-inflammatory protein and an anti-tumor invasive protein. These findings may lead to new ideas for diagnostic and therapeutic approaches in the future gene therapy of ovarian cancer, inflammation, and infertility. It might offer suggestions for later gene therapy treatments for ovarian cancer, inflammation, and infertility. Figure 2 presents a summary of the physiological and pathological processes that are mediated by BK.

4 Developments in the understanding of ITIH proteins and illnesses

ITIH has lately been found and thoroughly explained to be involved in several pathophysiological processes, such as inflammation and carcinogenesis (9, 43). Depending on the circumstances, these proteins may be positively or negatively regulated; nonetheless, there is compelling evidence that every member of the ITIH family is crucial to the development of tumors and cellular malignant processes (9, 44). ITIH proteins function as both pre- and anti-inflammatory acute phase proteins during inflammation, which leads to contradictory functions in the process. While the H3 chain is up-regulated and the related molecules behave as positive acute phase proteins, the H2 and BK chains are down-regulated and the associated molecules behave as negative acute phase proteins in acute inflammation. Inflammatory situations do not seem to have an impact on H1 chain (45). Strong evidence suggests that genes in the ITIH family may be tumor suppressors because these genes are highly down-regulated in a range of human solid tumors, including lung cancer, breast cancer and colon cancer (9). ITIH proteins stabilize the ECM in a way that inhibits tumor growth, which plays a significant role in carcinogenesis (46). ITIH's covalent binding to HA and its capacity to maintain the ECM are two potential pathways. ITIH2 expression and estrogen receptor expression are substantially associated ($p=0.001$) in breast cancer, according to a number of prior research. Cancer invasion and motility have been shown to be inhibited by estrogen (47). Because ITIH2 has an estrogen-binding structural domain that profoundly affects ECM integrity and may therefore be crucial for tumor growth and metastasis, there used to be a high correlation between ITIH2 expression levels and estrogen levels (48). ITIH2 is expressed in low-grade CNS cancers and normal brain tissues; however, it is not expressed in glioblastomas, especially glioblastoma multiforme, which is a highly invasive CNS tumor. This suggests that ITIH2 may have an anti-invasive function (49). The ITIH family gene has been linked in certain studies to shared genetic risk factors for schizophrenia and depression, in addition to its significant function in inflammation and malignancies. These findings imply that the ITIH family may potentially be implicated in the pathophysiology of psychiatric disorders (50). Furthermore, aberrant expression of ITIH family proteins has been found in neurodegenerative diseases (51, 52). The following table provides a summary of the roles and possible mechanisms of action of heavy chain participation (see Tables 1–5).

The ITIH family has five homologous variants that are encoded by various genes. Through an overview of current research pertaining to the ITIH family, we discovered that ITIH plays a role in numerous pathophysiological mechanisms. These comprise immunological responses, tumor growth, psychological issues, and

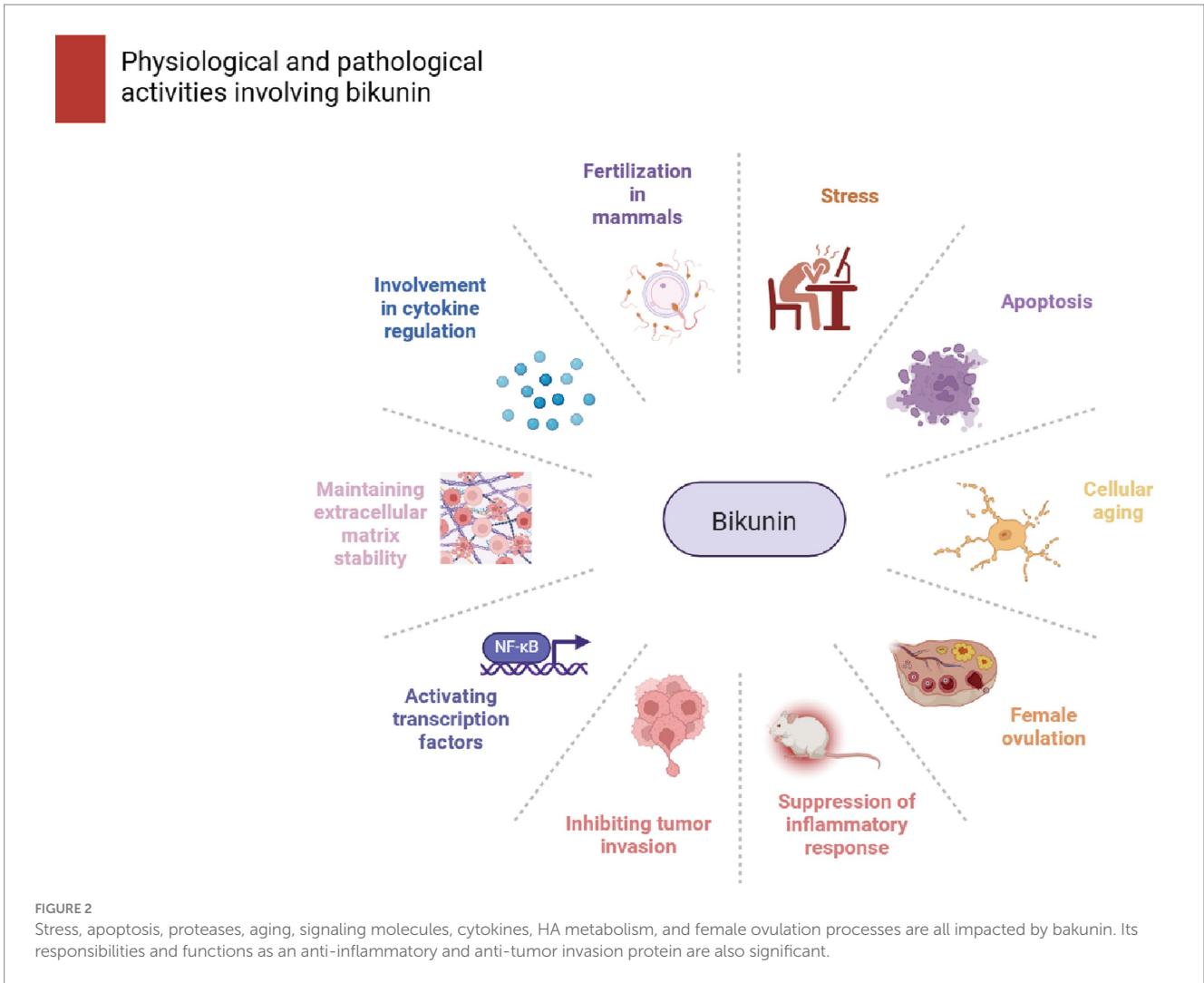


TABLE 1 Functions and potential mechanisms of ITIH1.

	Functions	Mechanisms	References
ITIH1	Insulin sensitivity	① ITIH1 binding directly to HA on skeletal muscle and adipose tissue surface causes ECM stability and subsequent insulin resistance ② Increased ITIH1 may impact β-cell activity	(16)
ITIH1	Indicators for ankylosing spondylitis diagnosis	Markedly elevated in ankylosing spondylitis patients, $p = 0.035$ ROC distinguished between ankylosing spondylitis patients (AUC = 0.98)	(53)
ITIH1	Evaluation of hepatic fibrosis	Liver fibrosis development and impaired ECM stability are linked to lower ITIH1 levels	(54)
ITIH1	Control of adherence of leukocytes	Adhesion of the inflammatory HA is regulated by thrombin cleavage of ITIH1	(55)
ITIH1	Modulator of immunity	Blocks the C3 complement pathway	(56)
ITIH1	Potential biomarker for hepatocellular carcinoma that is both diagnostic and predictive	Hepatocellular carcinoma patients have much lower levels of ITIH1 expression, and this downregulation is detrimental to the prognosis of these patients	(57)
ITIH1	Involved in the development of hepatocellular carcinoma	By stimulating the PI3K/AKT signaling pathway and epigenetically suppressing ITIH1 transcription, KDM5C can encourage the malignant progression of hepatocellular carcinoma	(58)
ITIH1	Markers for radiographic knee osteoarthritis development prediction	Shows promise for enhancing clinical practice radiographic knee osteoarthritis incidence prediction	(59)
ITIH1	Potential biomarkers for ovarian cancer	Variations in expression in the serum of patients with ovarian cancer	(60)
ITIH1	Biomarker for coronary heart disease risk stratification	When paired with additional proteins, ITIH1 exhibits a Lasso-logistic score that is highly effective in categorization (cross-validated area under the curve = 0.74)	(61)

TABLE 2 Functions and potential mechanisms of ITIH2.

	Functions	Mechanisms	References
ITIH2	Increases intercellular adhesion, suppresses cell proliferation, and prevents glioblastoma invasion	Reduces the activity of the PI3K/AKT signaling pathway	(49)
ITIH2	Biomarker for multiple sclerosis in children	Patients with pediatric multiple sclerosis had considerably reduced serum levels of ITIH2 protein	(62)
ITIH2	Pancreatic cancer diagnostic biomarker	Area under the curve = 0.947; ROC separates pancreatic cancer from chronic pancreatitis	(63)
ITIH2	Potential candidates associated with diabetic retinopathy	Comparing the protein profiles of vitreous humor in patients with idiopathic macular lentiginos who are not diabetics, it was discovered that they had significantly decreased expression of ITIH2 protein	(64)
ITIH2	Indicators of serum proteins for osteoarticular. Tuberculosis detection	For the diagnosis of osteoarticular tuberculosis, the AUC of ITIH2 was 0.7167 (95% CI: 0.5846–0.8487)	(65)
ITIH2	Hinders the growth of mother cells in gliomas	Tumor cell contact is inhibited by gonadotropin-releasing hormone interaction	(66)

TABLE 3 Functions and potential mechanisms of ITIH3.

	Functions	Mechanisms	References
ITIH3	Play in the development of the human brain	Differential spatiotemporal expression of ITIH3 in the developing human brain is shown by analysis of spatiotemporal histology data, which indicates that the SNP locus rs25352629 of ITIH3 is a susceptibility variable for autism	(67)
ITIH3	Intrahepatic cholestasis in pregnancy: diagnostic indicators	For the diagnosis of intrahepatic cholestasis in pregnancy, the AUC of ITIH3 was 0.8163	(68)
ITIH3	Strongly correlated with a history of attempted suicide in bipolar illness and schizophrenia cases	After multiple testing was taken into account, there was a substantial correlation found between the risk allele for the SNP locus rs2239547 of ITIH3 and a history of suicide attempts	(14)
ITIH3	Strongly correlated with the likelihood of getting schizophrenia	The ITIH3 SNP locus rs3617 may impact neurodevelopment and protein function, raising the risk of schizophrenia	(69)
ITIH3	Potentially forecast ovarian cancer's susceptibility to cisplatin	Platinum resistance and a poor prognosis are strongly correlated with low expression of ITIH3 protein in ovarian cancer tissues	(70)
ITIH3	Unknown hereditary susceptibility to myocardial infarction	In human atherosclerotic lesions, macrophages and vascular smooth muscle cells express the ITIH3 protein	(71)
ITIH3	It is a helpful biomarker for gastric cancer early detection Assesses the likelihood of gastric cancer	According to ROC, ITIH3 has a maximum sensitivity of 96% and a maximum specificity of 66% in the identification of gastric cancer. Furthermore, patients with early gastric cancer had considerably greater plasma levels of ITIH3 than non-cancerous patients ($p < 0.001$)	(72, 73)
ITIH3	Might not be adequate as a potent biomarker for early gastric cancer detection	Between early and advanced gastric cancer, there was no difference in expression; the AUC value was 0.65 (95% CI: 0.55–0.75)	(74)
ITIH3	Enhances acute and chronic heart failure and hypertrophic cardiomyopathy. Treatment with biomarker candidate	Proteomic analysis was used to identify putative core biomarkers	(75)
ITIH3	A biomarker for the detection of pancreatic cancer	In comparison to controls, expression in pancreatic cancer was 1.80 times greater	(76)
ITIH3	It is a biomarker for the early detection of rheumatoid arthritis	Utilizing innovative diagnostic techniques, rheumatoid arthritis diagnosis approaches 100% sensitivity and specificity	(77)
ITIH3	Contributes to the development of synovial joint disease	Significant elevation of ITIH3 in synovial joint diseases	(78)

inflammation. These protein families play a role in the modification of extracellular structures necessary for cell migration and the growth of malignant tumors. ITIH family has intricate functions. Various disease processes may involve distinct heavy chain proteins.

Different diseases may involve the same heavy chain protein. This implies that ITIH family may not be as accurate a diagnostic or prognostic marker for some disorders. Large-scale multicenter clinical trials are required in the future to assess if ITIH family may

TABLE 4 Functions and potential mechanisms of ITIH4.

	Functions	Mechanisms	References
ITIH4	Pro-inflammatory reaction	Activation of the PGK1-ITIH4 axis causes a pro-inflammatory response	(79)
ITIH4	Cholestatic liver disease biomarkers	Cholestatic liver dysfunction was associated with considerably higher plasma ITIH4 values	(80)
ITIH4	Biomarkers that show how NAFLD progresses and how hepatocellular carcinoma develops as a result	Compared to patients with simple steatosis and virus-associated hepatocellular carcinoma, patients with hepatocellular carcinoma-NAFLD had considerably higher serum ITIH4 levels Following hepatectomy, patients with hepatocellular carcinoma-NAFLD who had greater serum ITIH4 levels had a worse prognosis	(81)
ITIH4	A possible therapeutic target to prevent the spread of tumors	HuH7 cell movement was markedly reduced by ITIH4 overexpression and increased by ITIH4 knockdown, respectively. Patients with hepatocellular carcinoma who had a good prognosis had tumor tissues with higher levels of ITIH4 expression than those who had a bad prognosis	(82)
ITIH4	Distinguishes multisystemic and unisystemic histiocytosis of Langerhans cells	Significant variations in ITIH4 expression were found between the two illness groups by peptideomics analysis	(83)
ITIH4	Participated in repeated abortions	The activation of the IL-6 signaling pathway by ITIH4 ^(ΔN688) promotes inflammatory responses The long isoforms of ITIH4 have different functions in controlling cell invasion, migration, proliferation, and inflammatory responses	(84)
ITIH4	Beneficial for anticipating and monitoring sepsis	The expression of ITIH4 varies with the stage of sepsis	(85)
ITIH4	Possible prognostic biomarker for psoriasis patients' liver fibrosis brought on by methotrexate	Patients with psoriasis who had liver fibrosis brought on by methotrexate had abnormally high levels of it in their urine	(86)
ITIH4	Early gastric cancer biomarkers	When compared to patients receiving will-care controls, patients with early gastric cancer had noticeably higher serum levels of ITIH ITIH4 expression increased during <i>Helicobacter pylori</i> infection	(87, 88)
ITIH4	Biomarkers for ovarian cancer	Patients with ovarian cancer had far lower ITIH4 expression levels in their urine than did controls	(89)
ITIH4	Evaluation of the diagnostic and prognostic importance in individuals with hepatitis B-associated hepatocellular carcinoma and liver cirrhosis caused by the virus	Patients diagnosed with hepatocellular carcinoma had lower serum ITIH4 levels, and those with hepatitis B-associated hepatocellular carcinoma also had shorter survival times	(90)
ITIH4	Associated with autoimmune disease	Prevention of neutrophil recruitment in order to reduce inflammatory autoimmunity	(91)
ITIH4	Prognostic markers for aneurysmal subarachnoid hemorrhage in humans (ASAH)	① Prevent inflammation ② Early on ASAH, a substantial drop in serum ITIH4 concentrations was highly correlated with a poor prognosis and the severity of the illness	(92)
ITIH4	Prostate cancer and prostatic hyperplasia identification	Patients with prostate cancer had considerably higher ITIH4 protein levels	(93)
ITIH4	Ischemic acute stroke biomarkers	As the acute ischemic stroke condition progressed, ITIH4 eventually returned to normal	(94)
ITIH4	Preventing the growth of melanoma	A finding not further explained in the text	(95)
ITIH4	Monitoring of patients with postoperative breast cancer	ITIH4 was substantially lower in postoperative breast cancer patients than in controls	(96)
ITIH4	Accurately forecasting hyperlipidemia	C/T polymorphism of single nucleotides at IVS17 + 8 of ITIH4. Ninety percent of those missing the T allele went on to develop hypercholesterolemia. Of those with the T allele, only 10% experienced hypercholesterolemia ($p < 0.0001$)	(97)
ITIH4	Potential Alzheimer's disease serum biomarkers	Increased intact size of ITIH4 protein and decreased fragmentation of ITIH4; increased ITIH4 expression	(51, 52)
ITIH4	Interstitial cystitis biomarkers	ITIH4 plasma concentrations were greater in the patients ($p = 0.019$)	(98)

TABLE 5 Functions and potential mechanisms of ITIH5.

	Functions	Mechanisms	References
ITIH5	Indicator biomarkers for bile duct cancer diagnostics	In comparison to the control group, which included people with hepatocellular carcinoma, benign illness, chronic hepatitis B, and healthy persons, the cholangiocarcinoma group had greater serum ITIH5 levels. The AUC for ITIH5 varied between 0.839 and 0.851, indicating a significant difference between bile duct cancer and the control group	(99)
ITIH5	Prevention of the development of pancreatic cancer	Prevention of the spread of pancreatic cancer	(100)
ITIH5	Prevention of the development of bladder cancer	Overexpression of ITIH5 in bladder cancer cells prevented colony spreading and cell migration	(101)
ITIH5	Prevention of the spread of breast cancer	ITIH5-overexpressing breast cancer cells nearly totally failed to develop lung metastasis in a mouse metastasis model	(102)
ITIH5	Stops the spread of cervical cancer	Reduced the growth and invasiveness of tumor spheres to a large degree, which increased the rate of death in cervical cancer cells	(103)
ITIH5	Adipokine released by adipocytes	By comparing gene expression in subcutaneous and visceral fat microarray investigations in obese and lean subjects, ITIH5 was discovered as a novel adipokine	(104)
ITIH5	Involved in wound healing	The transformation of fibroblasts into myofibroblasts, which is dependent on growth factor β 1, requires the interaction of ITIH5 with cell surface HA	(105)
ITIH5	Involved in breast cancer development	Breast cancers exhibit a marked downregulation of ITIH5 expression. While ITIH5 expression is either consistently missing or significantly downregulated in invasive ductal carcinomas, normal breast epithelial cells express ITIH5 substantially. Both benign breast cell lines and breast cancer cell lines did not exhibit ITIH5 gene expression. Breast cancer development may be linked to the lack of ITIH5 expression	(106)
ITIH5	Prevention of the development of cervical cancer	ITIH5 overexpression resulted in a marked decrease in cell migration and clone formation as well as a considerable inhibition of cervical cancer cell proliferation	(107)
ITIH5	Prevents the growth of colon cancer cell	Colon cancer cells were unable to proliferate when ITIH5 was overexpressed	(108)
ITIH5	Prevention of the development of pancreatic cancer	ITIH5 may obstruct many oncogenic signaling pathways, including as the PI3K/AKT pathway. Changes in cell migration and the creation of local adhesions may result from this. These alterations in the cells could be connected to ITIH5's ability to prevent metastasis in pancreatic cancer	(109)
ITIH5	Prevention of the development of non-small cell lung cancer	Lower expression of ITIH5 is linked to a worse prognosis in non-small cell lung cancer	(110)
ITIH5	Responsible for the emergence of clinical metabolic abnormalities and obesity	Obese people express ITIH5 more than lean people do, and dieting-induced weight loss reduces ITIH5 expression	(111)
ITIH5	Contribute to the pathophysiology of congenital megacolon	Elevated ITIH5 expression prevents cell migration and proliferation	(112)

be used as a prognostic or diagnostic biomarker for specific illnesses. Despite the fact that numerous studies have demonstrated that the ITIH family prevents the growth of different types of solid tumors. The ITIH family is a putative oncogene, but further investigation and analysis are required, and it's still unclear how it works.

5 Summary

To jointly preserve the stability of the ECM, members of the $\text{I}\alpha\text{I}$ family bind covalently to HA. Hence the characteristics play a role in several physiological and pathological processes. The $\text{I}\alpha\text{I}$

family contains the protein BK, which is important for mammalian ovulation and human cancer. It is also an anti-invasive protein. The ITIH family is important for inflammation, immunity, psychiatric disorders, tumorigenesis, and development. Although BK has been extensively researched, its exact mechanism of action in the pathophysiological processes involving the ITIH family is still unclear. In order to examine the possible mechanisms and offer a theoretical foundation for targeted therapy of the associated disorders, a significant amount of research is required going forward. In addition, existing studies have shown that BK and ITIH family have been found to play a role in both inflammation and tumors, but whether there is an interaction between the two remains unclear.

Author contributions

X-fZ: Investigation, Writing – original draft, Data curation. X-lZ: Writing – original draft, Data curation, Investigation. LG: Writing – original draft. Y-pB: Writing – original draft. YT: Supervision, Writing – review & editing. H-yL: Supervision, Writing – review & editing.

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References

- Lord MS, Melrose J, Day AJ, Whitelock JM. The inter-alpha-trypsin inhibitor family: versatile molecules in biology and pathology. *J Histochem Cytochem.* (2020) 68:907–27. doi: 10.1369/0022155420940067
- Zhuo L, Hascall VC, Kimata K. Inter-alpha-trypsin inhibitor, a covalent protein-glycosaminoglycan-protein complex. *J Biol Chem.* (2004) 279:38079–82. doi: 10.1074/jbc.R300039200
- Fries E, Blom AM. Bikunin—not just a plasma proteinase inhibitor. *Int J Biochem Cell Biol.* (2000) 32:125–37. doi: 10.1016/S1357-2725(99)00125-9
- Jessen TE, Odum L. Role of tumour necrosis factor stimulated gene 6 (TSG-6) in the coupling of inter-alpha-trypsin inhibitor to hyaluronan in human follicular fluid. *Reproduction.* (2003) 125:27–31. doi: 10.1530/rep.0.1250027
- Sanggaard KW, Scavenius C, Rasmussen AJ, Wisniewski HG, Thogersen IB, Enghild JJ. The TSG-6/HC2-mediated transfer is a dynamic process shuffling heavy chains between glycosaminoglycans. *J Biol Chem.* (2010) 285:21988–93. doi: 10.1074/jbc.M109.041046
- Colon E, Shyuhina A, Cowman MK, Band PA, Sanggaard KW, Enghild JJ, et al. Transfer of inter-alpha-inhibitor heavy chains to hyaluronan by surface-linked hyaluronan-TSG-6 complexes. *J Biol Chem.* (2009) 284:2320–31. doi: 10.1074/jbc.M807183200
- Huang L, Yoneda M, Kimata K. A serum-derived hyaluronan-associated protein (SHAP) is the heavy chain of the inter alpha-trypsin inhibitor. *J Biol Chem.* (1993) 268:26725–30. doi: 10.1016/S0021-9258(19)74373-7
- Chen L, Mao SJ, Larsen WJ. Identification of a factor in fetal bovine serum that stabilizes the cumulus extracellular matrix. A role for a member of the inter-alpha-trypsin inhibitor family. *J Biol Chem.* (1992) 267:12380–6.
- Hamm A, Veeck J, Bektas N, Wild PJ, Hartmann A, Heindrichs U, et al. Frequent expression loss of inter-alpha-trypsin inhibitor heavy chain (ITIH) genes in multiple human solid tumors: a systematic expression analysis. *BMC Cancer.* (2008) 8:25. doi: 10.1186/1471-2407-8-25
- Salier JP, Rouet P, Raguenez G, Daveau M. The inter-alpha-inhibitor family: from structure to regulation. *Biochem J.* (1996) 315:1–9. doi: 10.1042/bj3150001
- Gebhard W, Schreitmuller T, Hochstrasser K, Wachter E. Two out of the three kinds of subunits of inter-alpha-trypsin inhibitor are structurally related. *Eur J Biochem.* (1989) 181:571–6. doi: 10.1111/j.1432-1033.1989.tb14762.x
- Enghild JJ, Salvesen G, Thogersen IB, Valnickova Z, Pizzo SV, Hefta SA. Presence of the protein-glycosaminoglycan-protein covalent cross-link in the inter-alpha-inhibitor-related proteinase inhibitor heavy chain 2/bikunin. *J Biol Chem.* (1993) 268:8711–6. doi: 10.1016/S0021-9258(18)52933-1
- Zhuo L, Kimata K. Structure and function of inter-alpha-trypsin inhibitor heavy chains. *Connect Tissue Res.* (2008) 49:311–20. doi: 10.1080/03008200802325458
- Finseth PI, Sonderby IE, Djurovic S, Agartz I, Malt UF, Melle I, et al. Association analysis between suicidal behaviour and candidate genes of bipolar disorder and schizophrenia. *J Affect Disord.* (2014) 163:110–4. doi: 10.1016/j.jad.2013.12.018
- Pardue EL, Ibrahim S, Ramamurthi A. Role of hyaluronan in angiogenesis and its utility to angiogenic tissue engineering. *Organogenesis.* (2008) 4:203–14. doi: 10.4161/org.4.4.6926
- Kim TH, Koo JH, Heo MJ, Han CY, Kim YI, Park SY, et al. Overproduction of inter-alpha-trypsin inhibitor heavy chain 1 after loss of G α_{13} in liver exacerbates systemic insulin resistance in mice. *Sci Transl Med.* (2019) 11:eaan4735. doi: 10.1126/scitranslmed.aan4735
- Garantzotis S, Zudaire E, Trempus CS, Hollingsworth JW, Jiang D, Lancaster LH, et al. Serum inter-alpha-trypsin inhibitor and matrix hyaluronan promote angiogenesis

Conflict of interest

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- in fibrotic lung injury. *Am J Respir Crit Care Med.* (2008) 178:939–47. doi: 10.1164/rccm.200803-386OC
- Zhuo L, Yoneda M, Zhao M, Yingsung W, Yoshida N, Kitagawa Y, et al. Defect in SHAP-hyaluronan complex causes severe female infertility. A study by inactivation of the bikunin gene in mice. *J Biol Chem.* (2001) 276:7693–6. doi: 10.1074/jbc.C000899200
- Chanmee T, Ontong P, Itano N. Hyaluronan: a modulator of the tumor microenvironment. *Cancer Lett.* (2016) 375:20–30. doi: 10.1016/j.canlet.2016.02.031
- Zhao M, Yoneda M, Ohashi Y, Kurono S, Iwata H, Ohnuki Y, et al. Evidence for the covalent binding of SHAP, heavy chains of inter-alpha-trypsin inhibitor, to hyaluronan. *J Biol Chem.* (1995) 270:26657–63. doi: 10.1074/jbc.270.44.26657
- Fraser JR, Laurent TC, Laurent UB. Hyaluronan: its nature, distribution, functions and turnover. *J Intern Med.* (1997) 242:27–33. doi: 10.1046/j.1365-2796.1997.00170.x
- Slevin M, Krupinski J, Gaffney J, Matou S, West D, Delisser H, et al. Hyaluronan-mediated angiogenesis in vascular disease: uncovering RHAMM and CD44 receptor signaling pathways. *Matrix Biol.* (2007) 26:58–68. doi: 10.1016/j.matbio.2006.08.261
- Bourguignon LY. Matrix hyaluronan-activated CD44 signaling promotes keratinocyte activities and improves abnormal epidermal functions. *Am J Pathol.* (2014) 184:1912–9. doi: 10.1016/j.ajpath.2014.03.010
- Tan KT, McGrouther DA, Day AJ, Milner CM, Bayat A. Characterization of hyaluronan and TSG-6 in skin scarring: differential distribution in keloid scars, normal scars and unscarred skin. *J Eur Acad Dermatol Venereol.* (2011) 25:317–27. doi: 10.1111/j.1468-3083.2010.03792.x
- Deed R, Rooney P, Kumar P, Norton JD, Smith J, Freemont AJ, et al. Early-response gene signalling is induced by angiogenic oligosaccharides of hyaluronan in endothelial cells. Inhibition by non-angiogenic, high-molecular-weight hyaluronan. *Int J Cancer.* (1997) 71:251–6. doi: 10.1002/(sici)1097-0215(19970410)71:2<251::aid-ijc21>3.0.co;2-j
- West DC, Hampson IN, Arnold F, Kumar S. Angiogenesis induced by degradation products of hyaluronic acid. *Science.* (1985) 228:1324–6. doi: 10.1126/science.2408340
- Suzuki M, Kobayashi H, Tanaka Y, Kanayama N, Terao T. Reproductive failure in mice lacking inter-alpha-trypsin inhibitor (ITI)—ITI target genes in mouse ovary identified by microarray analysis. *J Endocrinol.* (2004) 183:29–38. doi: 10.1677/joe.1.05803
- Sato H, Kajikawa S, Kuroda S, Horisawa Y, Nakamura N, Kaga N, et al. Impaired fertility in female mice lacking urinary trypsin inhibitor. *Biochem Biophys Res Commun.* (2001) 281:1154–60. doi: 10.1006/bbrc.2001.4475
- Zhuo L, Salustri A, Kimata K. A physiological function of serum proteoglycan bikunin: the chondroitin sulfate moiety plays a central role. *Glycoconj J.* (2002) 19:241–7. doi: 10.1023/A:1025331929373
- Lepedda AJ, De Muro P, Capobianco G, Formato M. Role of the small proteoglycan bikunin in human reproduction. *Hormones.* (2020) 19:123–33. doi: 10.1007/s42000-019-00149-x
- Pratt CW, Pizzo SV. Mechanism of action of inter-alpha-trypsin inhibitor. *Biochemistry.* (1987) 26:2855–63. doi: 10.1021/bi00384a029
- Yingsung W, Zhuo L, Morgelin M, Yoneda M, Kida D, Watanabe H, et al. Molecular heterogeneity of the SHAP-hyaluronan complex. Isolation and characterization of the complex in synovial fluid from patients with rheumatoid arthritis. *J Biol Chem.* (2003) 278:32710–8. doi: 10.1074/jbc.M303658200
- Kobayashi H. Endogenous anti-inflammatory substances, inter-alpha-inhibitor and bikunin. *Biol Chem.* (2006) 387:1545–9. doi: 10.1515/BC.2006.192
- Nakamura H, Abe S, Shibata Y, Sata M, Kato S, Saito H, et al. Inhibition of neutrophil elastase-induced interleukin-8 gene expression by urinary trypsin inhibitor in human bronchial epithelial cells. *Int Arch Allergy Immunol.* (1997) 112:157–62. doi: 10.1159/000237448

35. Futamura Y, Kajikawa S, Kaga N, Shibutani Y. Protection against preterm delivery in mice by urinary trypsin inhibitor. *Obstet Gynecol.* (1999) 93:100–8. doi: 10.1016/s0029-7844(98)00396-2
36. Wakahara K, Kobayashi H, Yagyu T, Matsuzaki H, Kondo T, Kurita N, et al. Bikunin suppresses lipopolysaccharide-induced lethality through down-regulation of tumor necrosis factor- α and interleukin-1 β in macrophages. *J Infect Dis.* (2005) 191:930–8. doi: 10.1086/428134
37. Lepedda AJ, Nieddu G, Cannas C, Formato M. Molecular and pathobiological insights of bikunin/UTII in cancer. *Mol Biol Rep.* (2023) 50:1701–11. doi: 10.1007/s11033-022-08117-2
38. Hirashima Y, Kobayashi H, Suzuki M, Tanaka Y, Kanayama N, Fujie M, et al. Characterization of binding properties of urinary trypsin inhibitor to cell-associated binding sites on human chondrosarcoma cell line HCS-2/8. *J Biol Chem.* (2001) 276:13650–6. doi: 10.1074/jbc.M009906200
39. Suzuki M, Kobayashi H, Tanaka Y, Hirashima Y, Kanayama N, Takei Y, et al. Bikunin target genes in ovarian cancer cells identified by microarray analysis. *J Biol Chem.* (2003) 278:14640–6. doi: 10.1074/jbc.M300239200
40. Suzuki M, Kobayashi H, Tanaka Y, Hirashima Y, Kanayama N, Takei Y, et al. Suppression of invasion and peritoneal carcinomatosis of ovarian cancer cell line by overexpression of bikunin. *Int J Cancer.* (2003) 104:289–302. doi: 10.1002/ijc.10950
41. Kobayashi H, Suzuki M, Hirashima Y, Terao T. The protease inhibitor bikunin, a novel anti-metastatic agent. *Biol Chem.* (2003) 384:749–54. doi: 10.1515/BC.2003.083
42. Matsuzaki H, Kobayashi H, Yagyu T, Wakahara K, Kondo T, Kurita N, et al. Plasma bikunin as a favorable prognostic factor in ovarian cancer. *J Clin Oncol.* (2005) 23:1463–72. doi: 10.1200/JCO.2005.03.010
43. Singh K, Zhang LX, Bendelja K, Heath R, Murphy S, Sharma S, et al. Inter-alpha inhibitor protein administration improves survival from neonatal sepsis in mice. *Pediatr Res.* (2010) 68:242–7. doi: 10.1203/PDR.0b013e3181e9fd0
44. Ohya K, Yoshimi H, Aibara N, Nakamura Y, Miyata Y, Sakai H, et al. Immune complexome analysis reveals the specific and frequent presence of immune complex antigens in lung cancer patients: a pilot study. *Int J Cancer.* (2017) 140:370–80. doi: 10.1002/ijc.30455
45. Daveau M, Rouet P, Scotte M, Faye L, Hiron M, Lebreton JP, et al. Human inter-alpha-inhibitor family in inflammation: simultaneous synthesis of positive and negative acute-phase proteins. *Biochem J.* (1993) 292:485–92. doi: 10.1042/bj2920485
46. Kopylov AT, Stepanov AA, Malsagova KA, Soni D, Kushlinsky NE, Enikeev DV, et al. Revelation of proteomic indicators for colorectal cancer in initial stages of development. *Molecules.* (2020) 25:619. doi: 10.3390/molecules25030619
47. Rochefort H, Platet N, Hayashido Y, Derocq D, Lucas A, Cunat S, et al. Estrogen receptor mediated inhibition of cancer cell invasion and motility: an overview. *J Steroid Biochem Mol Biol.* (1998) 65:163–8. doi: 10.1016/S0960-0760(98)00010-7
48. Zhang Q, Huang R, Tang Q, Yu Y, Huang Q, Chen Y, et al. Leucine-rich alpha-2-glycoprotein-1 is up-regulated in colorectal cancer and is a tumor promoter. *Oncotargets Ther.* (2018) 11:2745–52. doi: 10.2147/OTT.S153375
49. Werbowetski-Ogilvie TE, Agar NY, Waldkircher de Oliveira RM, Faury D, Antel JB, Jabado N, et al. Isolation of a natural inhibitor of human malignant glial cell invasion: inter alpha-trypsin inhibitor heavy chain 2. *Cancer Res.* (2006) 66:1464–72. doi: 10.1158/0008-5472.CAN-05-1913
50. He K, Wang Q, Chen J, Li T, Li Z, Li W, et al. ITIH family genes confer risk to schizophrenia and major depressive disorder in the Han Chinese population. *Prog Neuro-Psychopharmacol Biol Psychiatry.* (2014) 51:34–8. doi: 10.1016/j.pnpbp.2013.12.004
51. Zhang X, Yu W, Cao X, Wang Y, Zhu C, Guan J. Identification of serum biomarkers in patients with Alzheimer's disease by 2D-DIGE proteomics. *Gerontology.* (2022) 68:686–98. doi: 10.1159/000520961
52. Shi X, Ohta Y, Liu X, Shang J, Morihara R, Nakano Y, et al. Acute anti-inflammatory markers ITIH4 and AHSG in mice brain of a novel Alzheimer's disease model. *J Alzheimers Dis.* (2019) 68:1667–75. doi: 10.3233/JAD-181218
53. Fischer R, Trudgian DC, Wright C, Thomas G, Bradbury LA, Brown MA, et al. Discovery of candidate serum proteomic and metabolomic biomarkers in ankylosing spondylitis. *Mol Cell Proteomics.* (2012) 11:M1111.013904. doi: 10.1074/mcp.M1111.013904
54. Caillot F, Hiron M, Gorla O, Gueudin M, Francois A, Scotte M, et al. Novel serum markers of fibrosis progression for the follow-up of hepatitis C virus-infected patients. *Am J Pathol.* (2009) 175:46–53. doi: 10.2353/ajpath.2009.080850
55. Petrey AC, de la Motte CA. Thrombin cleavage of inter-alpha-inhibitor heavy chain I regulates leukocyte binding to an inflammatory hyaluronan matrix. *J Biol Chem.* (2016) 291:24324–34. doi: 10.1074/jbc.M116.755660
56. Briggs DC, Langford-Smith AWW, Birchenough HL, Jowitz TA, Kiely CM, Engchild JJ, et al. Inter-alpha-inhibitor heavy chain-1 has an integrin-like 3D structure mediating immune regulatory activities and matrix stabilization during ovulation. *J Biol Chem.* (2020) 295:5278–91. doi: 10.1074/jbc.RA119.011916
57. Chang QH, Mao T, Tao Y, Dong T, Tang XX, Ge GH, et al. Pan-cancer analysis identifies ITIH1 as a novel prognostic indicator for hepatocellular carcinoma. *Aging.* (2021) 13:11096–119. doi: 10.18632/aging.202765
58. Qian X, Bao ZM, Yao D, Shi Y. Lysine demethylase 5C epigenetically reduces transcription of ITIH1 that results in augmented progression of liver hepatocellular carcinoma. *Kaohsiung J Med Sci.* (2022) 38:437–46. doi: 10.1002/kjm2.12501
59. Lourido L, Balboa-Barreiro V, Ruiz-Romero C, Rego-Perez I, Camacho-Encina M, Paz-Gonzalez R, et al. A clinical model including protein biomarkers predicts radiographic knee osteoarthritis: a prospective study using data from the osteoarthritis initiative. *Osteoarthr Cartil.* (2021) 29:1147–54. doi: 10.1016/j.joca.2021.04.011
60. Lin B, White JT, Wu J, Lele S, Old LJ, Hood L, et al. Deep depletion of abundant serum proteins reveals low-abundant proteins as potential biomarkers for human ovarian cancer. *Proteomics Clin Appl.* (2009) 3:853–61. doi: 10.1002/prca.200800141
61. Anwar MA, Dai DL, Wilson-McManus J, Smith D, Francis GA, Borchers CH, et al. Multiplexed LC-ESI-MRM-MS-based assay for identification of coronary artery disease biomarkers in human plasma. *Proteomics Clin Appl.* (2019) 13:e1700111. doi: 10.1002/prca.201700111
62. Solmaz I, Kocak E, Kaplan O, Celebier M, Anlar B. Analysis of plasma protein biomarkers in childhood onset multiple sclerosis. *J Neuroimmunol.* (2020) 348:577359. doi: 10.1016/j.jneuroim.2020.577359
63. Saraswat M, Joenavaara S, Seppanen H, Mustonen H, Haglund C, Renkonen R. Comparative proteomic profiling of the serum differentiates pancreatic cancer from chronic pancreatitis. *Cancer Med.* (2017) 6:1748–51. doi: 10.1002/prca.41107
64. Garcia-Ramirez M, Canals F, Hernandez C, Colome N, Ferrer C, Carrasco E, et al. Proteomic analysis of human vitreous fluid by fluorescence-based difference gel electrophoresis (DIGE): a new strategy for identifying potential candidates in the pathogenesis of proliferative diabetic retinopathy. *Diabetologia.* (2007) 50:1294–303. doi: 10.1007/s00125-007-0627-y
65. Chen X, Wang J, Wang J, Ye J, Di P, Dong C, et al. Several potential serum proteomic biomarkers for diagnosis of osteoarthral tuberculosis based on mass spectrometry. *Clin Chim Acta.* (2023) 547:117447. doi: 10.1016/j.cca.2023.117447
66. Tripathi PH, Akhtar J, Arora J, Saran RK, Mishra N, Polisetty RV, et al. Quantitative proteomic analysis of GnRH agonist treated GBM cell line LN229 revealed regulatory proteins inhibiting cancer cell proliferation. *BMC Cancer.* (2022) 22:133. doi: 10.1186/s12885-022-09218-8
67. Xie X, Meng H, Wu H, Hou F, Chen Y, Zhou Y, et al. Integrative analyses indicate an association between ITIH3 polymorphisms with autism spectrum disorder. *Sci Rep.* (2020) 10:5223. doi: 10.1038/s41598-020-62189-3
68. Chen L, Li J, You Y, Qian Z, Liu J, Jiang Y, et al. Secreted proteins in plasma and placenta as novel non-invasive biomarkers for intrahepatic cholestasis of pregnancy: a case-control study. *Heliyon.* (2023) 9:e21616. doi: 10.1016/j.heliyon.2023.e21616
69. Li K, Li Y, Wang J, Huo Y, Huang D, Li S, et al. A functional missense variant in ITIH3 affects protein expression and neurodevelopment and confers schizophrenia risk in the Han Chinese population. *J Genet Genomics.* (2020) 47:233–48. doi: 10.1016/j.jgg.2020.04.001
70. Liu Y, Shi L, Yuan C, Feng Y, Li M, Liu H, et al. Downregulation of ITIH3 contributes to cisplatin-based chemotherapy resistance in ovarian carcinoma via the Bcl-2 mediated anti-apoptosis signaling pathway. *Oncol Lett.* (2023) 25:61. doi: 10.3892/ol.2022.13646
71. Ebana Y, Ozaki K, Inoue K, Sato H, Iida A, Lwin H, et al. A functional SNP in ITIH3 is associated with susceptibility to myocardial infarction. *J Hum Genet.* (2007) 52:220–9. doi: 10.1007/s10038-006-0102-5
72. Chong PK, Lee H, Zhou J, Liu SC, Loh MC, Wang TT, et al. ITIH3 is a potential biomarker for early detection of gastric cancer. *J Proteome Res.* (2010) 9:3671–9. doi: 10.1021/pr100192h
73. Shang F, Wang Y, Shi Z, Deng Z, Ma J. Development of a signature based on eight metastatic-related genes for prognosis of GC patients. *Mol Biotechnol.* (2023) 65:1796–808. doi: 10.1007/s12033-023-00671-9
74. Uen YH, Lin KY, Sun DP, Liao CC, Hsieh MS, Huang YK, et al. Comparative proteomics, network analysis and post-translational modification identification reveal differential profiles of plasma Con A-bound glycoprotein biomarkers in gastric cancer. *J Proteome.* (2013) 83:197–213. doi: 10.1016/j.jpro.2013.03.007
75. Chen H, Tesic M, Nikolic VN, Pavlovic M, Vucic RM, Spasic A, et al. Systemic biomarkers and unique pathways in different phenotypes of heart failure with preserved ejection fraction. *Biomol Ther.* (2022) 12:1419. doi: 10.3390/biom12101419
76. Tonack S, Jenkinson C, Cox T, Elliott V, Jenkins RE, Kitteringham NR, et al. iTRAQ reveals candidate pancreatic cancer serum biomarkers: influence of obstructive jaundice on their performance. *Br J Cancer.* (2013) 108:1846–53. doi: 10.1038/bjc.2013.150
77. Chen HM, Tsai YH, Hsu CY, Wang YY, Hsieh CE, Chen JH, et al. Peptide-coated bacteriorhodopsin-based photoelectric biosensor for detecting rheumatoid arthritis. *Biosensors.* (2023) 13:929. doi: 10.3390/bios13100929
78. Zhang C, Gawri R, Lau YK, Spruce LA, Fazelinia H, Jiang Z, et al. Proteomics identifies novel biomarkers of synovial joint disease in a canine model of mucopolysaccharidosis I. *Mol Genet Metab.* (2023) 138:107371. doi: 10.1016/j.ymgme.2023.107371
79. Park HB, Choi BC, Baek KH. PGK1 modulates balance between pro- and anti-inflammatory cytokines by interacting with ITI-H4. *Biomed Pharmacother.* (2023) 161:114437. doi: 10.1016/j.biopha.2023.114437
80. Laursen TL, Bossen L, Pihl R, Trolborg A, Sandahl TD, Hansen AG, et al. Highly increased levels of inter-alpha-inhibitor heavy chain 4 (ITIH4) in autoimmune cholestatic liver diseases. *J Clin Transl Hepatol.* (2022) 10:796–802. doi: 10.14218/JCTH.2021.00515

81. Nakamura N, Hatano E, Iguchi K, Sato M, Kawaguchi H, Ohtsu I, et al. Elevated levels of circulating ITIH4 are associated with hepatocellular carcinoma with nonalcoholic fatty liver disease: from pig model to human study. *BMC Cancer*. (2019) 19:621. doi: 10.1186/s12885-019-5825-8
82. Lee EJ, Yang SH, Kim KJ, Cha H, Lee SJ, Kim JH, et al. Inter-alpha inhibitor H4 as a potential biomarker predicting the treatment outcomes in patients with hepatocellular carcinoma. *Cancer Res Treat*. (2018) 50:646–57. doi: 10.4143/crt.2016.550
83. Murakami I, Oh Y, Morimoto A, Sano H, Kanzaki S, Matsushita M, et al. Acute-phase ITIH4 levels distinguish multi-system from single-system Langerhans cell histiocytosis via plasma peptidomics. *Clin Proteomics*. (2015) 12:16. doi: 10.1186/s12014-015-9089-2
84. Li L, Choi BC, Ryo J, Song SJ, Pei CZ, Lee KY, et al. Opposing roles of inter-alpha-trypsin inhibitor heavy chain 4 in recurrent pregnancy loss. *EBioMedicine*. (2018) 37:535–46. doi: 10.1016/j.ebiom.2018.10.029
85. Larsen JB, Pihl R, Aggerbeck MA, Larsen KM, Hvas CL, Johnsen N, et al. Inter-alpha-inhibitor heavy chain H4 and sepsis-related coagulation disturbances: another link between innate immunity and coagulation. *Res Pract Thromb Haemost*. (2023) 7:100078. doi: 10.1016/j.rpth.2023.100078
86. van Swelm RP, Laarakkers CM, Kooijmans-Otero M, de Jong EM, Masereeuw R, Russel FG. Biomarkers for methotrexate-induced liver injury: urinary protein profiling of psoriasis patients. *Toxicol Lett*. (2013) 221:219–24. doi: 10.1016/j.toxlet.2013.06.234
87. Sun Y, Jin J, Jing H, Lu Y, Zhu Q, Shu C, et al. ITIH4 is a novel serum biomarker for early gastric cancer diagnosis. *Clin Chim Acta*. (2021) 523:365–73. doi: 10.1016/j.cca.2021.10.022
88. Jing D, Jin J, Mei Z, Zhu Q, Lu Y, Wang X. Effects of *Helicobacter pylori* infection and interleukin 6 on the expression of ITIH4 in human gastric cancer cells. *Transl Cancer Res*. (2020) 9:4656–65. doi: 10.21037/tcr-20-1766
89. Abdullah-Soheimi SS, Lim BK, Hashim OH, Shuib AS. Patients with ovarian carcinoma excrete different altered levels of urine CD59, kininogen-1 and fragments of inter-alpha-trypsin inhibitor heavy chain H4 and albumin. *Proteome Sci*. (2010) 8:58. doi: 10.1186/1477-5956-8-58
90. Noh CK, Kim SS, Kim DK, Lee HY, Cho HJ, Yoon SY, et al. Inter-alpha-trypsin inhibitor heavy chain H4 as a diagnostic and prognostic indicator in patients with hepatitis B virus-associated hepatocellular carcinoma. *Clin Biochem*. (2014) 47:1257–61. doi: 10.1016/j.clinbiochem.2014.05.002
91. Iwai T, Ohyama A, Osada A, Nishiyama T, Shimizu M, Miki H, et al. Role of inter-alpha-trypsin inhibitor heavy chain 4 and its citrullinated form in experimental arthritis murine models. *Clin Exp Immunol*. (2024) 215:302–12. doi: 10.1093/cei/uxae001
92. Tian H, Wang G, Zhong Q, Zhou H. Usability of serum inter-alpha-trypsin inhibitor heavy chain 4 as a biomarker for assessing severity and predicting functional outcome after human aneurysmal subarachnoid hemorrhage: a prospective observational cohort study at a single institution. *Clin Chim Acta*. (2024) 552:117679. doi: 10.1016/j.cca.2023.117679
93. Jayapalan JJ, Ng KL, Shuib AS, Razack AH, Hashim OH. Urine of patients with early prostate cancer contains lower levels of light chain fragments of inter-alpha-trypsin inhibitor and saposin B but increased expression of an inter-alpha-trypsin inhibitor heavy chain 4 fragment. *Electrophoresis*. (2013) 34:1663–9. doi: 10.1002/elps.201200583
94. Kashyap RS, Nayak AR, Deshpande PS, Kabra D, Purohit HJ, Taori GM, et al. Inter-alpha-trypsin inhibitor heavy chain 4 is a novel marker of acute ischemic stroke. *Clin Chim Acta*. (2009) 402:160–3. doi: 10.1016/j.cca.2009.01.009
95. Kormosh NG, Davidova TV, Kopyltsov VN, Serebryakova MV, Kabieva AO, Voyushin KE, et al. Conformational changes in inter-alpha-trypsin inhibitor heavy chain 4 activate its tumor-specific activity in mice with B16 melanoma. *Mol Med Rep*. (2015) 12:4483–93. doi: 10.3892/mmr.2015.3961
96. van den Broek I, Sparidans RW, van Winden AW, Gast MC, van Dulken EJ, Schellens JH, et al. The absolute quantification of eight inter-alpha-trypsin inhibitor heavy chain 4 (ITIH4)-derived peptides in serum from breast cancer patients. *Proteomics Clin Appl*. (2010) 4:931–9. doi: 10.1002/prca.201000035
97. Fujita Y, Ezura Y, Emi M, Sato K, Takada D, Iino Y, et al. Hypercholesterolemia associated with splice-junction variation of inter-alpha-trypsin inhibitor heavy chain 4 (ITIH4) gene. *J Hum Genet*. (2004) 49:24–8. doi: 10.1007/s10038-003-0101-8
98. Canter MP, Graham CA, Heit MH, Blackwell LS, Wilkey DW, Klein JB, et al. Proteomic techniques identify urine proteins that differentiate patients with interstitial cystitis from asymptomatic control subjects. *Am J Obstet Gynecol*. (2008) 198:e551–6. doi: 10.1016/j.ajog.2008.01.052
99. Chen M, Ma J, Xie X, Su M, Zhao D. Serum ITIH5 as a novel diagnostic biomarker in cholangiocarcinoma. *Cancer Sci*. (2024) 115:1665–79. doi: 10.1111/cas.16143
100. Sasaki K, Kurahara H, Young ED, Natsugoe S, Ijichi A, Iwakuma T, et al. Genome-wide in vivo RNAi screen identifies ITIH5 as a metastasis suppressor in pancreatic cancer. *Clin Exp Metastasis*. (2017) 34:229–39. doi: 10.1007/s10585-017-9840-3
101. Rose M, Gaisa NT, Antony P, Fiedler D, Heidenreich A, Otto W, et al. Epigenetic inactivation of ITIH5 promotes bladder cancer progression and predicts early relapse of pT1 high-grade urothelial tumours. *Carcinogenesis*. (2014) 35:727–36. doi: 10.1093/carcin/bgt375
102. Rose M, Klotten V, Noetzel E, Gola L, Ehling J, Heide T, et al. ITIH5 mediates epigenetic reprogramming of breast cancer cells. *Mol Cancer*. (2017) 16:44. doi: 10.1186/s12943-017-0610-2
103. Daum AK, Dittmann J, Jansen L, Peters S, Dahmen U, Heger JI, et al. ITIH5 shows tumor suppressive properties in cervical cancer cells grown as multicellular tumor spheroids. *Am J Transl Res*. (2021) 13:10298–314.
104. Dahlman I, Elsen M, Tennagels N, Korn M, Brockmann B, Sell H, et al. Functional annotation of the human fat cell secretome. *Arch Physiol Biochem*. (2012) 118:84–91. doi: 10.3109/13813455.2012.685745
105. Martin J, Midgley A, Meran S, Woods E, Bowen T, Phillips AO, et al. Tumor necrosis factor-stimulated gene 6 (TSG-6)-mediated interactions with the inter-alpha-inhibitor heavy chain 5 facilitate tumor growth factor beta1 (TGFbeta1)-dependent fibroblast to myofibroblast differentiation. *J Biol Chem*. (2016) 291:13789–801. doi: 10.1074/jbc.M115.670521
106. Himmelfarb M, Klopocki E, Grube S, Staub E, Klamann I, Hinzmann B, et al. ITIH5, a novel member of the inter-alpha-trypsin inhibitor heavy chain family is downregulated in breast cancer. *Cancer Lett*. (2004) 204:69–77. doi: 10.1016/j.canlet.2003.09.011
107. Dittmann J, Ziegfeld A, Jansen L, Gajda M, Klotten V, Dahl E, et al. Gene expression analysis combined with functional genomics approach identifies ITIH5 as tumor suppressor gene in cervical carcinogenesis. *Mol Carcinog*. (2017) 56:1578–89. doi: 10.1002/mc.22613
108. Klotten V, Rose M, Kaspar S, von Stillfried S, Knuchel R, Dahl E. Epigenetic inactivation of the novel candidate tumor suppressor gene ITIH5 in colon cancer predicts unfavorable overall survival in the CpG island methylator phenotype. *Epigenetics*. (2014) 9:1290–301. doi: 10.4161/epi.32089
109. Kosinski J, Sechi A, Hain J, Villwock S, Ha SA, Hauschulz M, et al. ITIH5 as a multifaceted player in pancreatic cancer suppression, impairing tyrosine kinase signaling, cell adhesion and migration. *Mol Oncol*. (2024) 18:1486–509. doi: 10.1002/1878-0261.13609
110. Dotsch MM, Klotten V, Schlenz M, Heide T, Braunschweig T, Veeck J, et al. Low expression of ITIH5 in adenocarcinoma of the lung is associated with unfavorable patients' outcome. *Epigenetics*. (2015) 10:903–12. doi: 10.1080/15592294.2015.1078049
111. Ruhl T, Sessler TM, Keimes JM, Beier JP, Villwock S, Rose M, et al. ITIH5 inhibits proliferation, adipogenic differentiation, and secretion of inflammatory cytokines of human adipose stem cells—a new key in treating obesity? *FASEB J*. (2024) 38:e23352. doi: 10.1096/fj.202301366R
112. Cai P, Li H, Huo W, Zhu H, Xu C, Zang R, et al. Aberrant expression of LncRNA-MIR31HG regulates cell migration and proliferation by affecting miR-31 and miR-31* in Hirschsprung's disease. *J Cell Biochem*. (2018) 119:8195–203. doi: 10.1002/jcb.26830