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Exosome prospects in the diagnosis and treatment of non-alcoholic fatty liver disease

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The growing prevalence of NAFLD and its global health burden have provoked considerable research on possible diagnostic and therapeutic options for NAFLD. Although various pathophysiological mechanisms and genetic factors have been identified to be associated with NAFLD, its treatment remains challenging. In recent years, exosomes have attracted widespread attention for their role in metabolic dysfunctions and their efficacy as pathological biomarkers. Exosomes have also shown tremendous potential in treating a variety of disorders. With increasing evidence supporting the significant role of exosomes in NAFLD pathogenesis, their theragnostic potential has become a point of interest in NAFLD. Expectedly, exosome-based treatment strategies have shown promise in the prevention and amelioration of NAFLD in preclinical studies. However, there are still serious challenges in preparing, standardizing, and applying exosome-based therapies as a routine clinical option that should be overcome. Due to the great potential of this novel theragnostic agent in NAFLD, further investigations on their safety, clinical efficacy, and application standardization are highly recommended.

KEYWORDS

NASH, non-alcoholic fatty liver, exosome, NAFLD, extracellular vesicle (EV), microRNA

Highlights

- Different exosomes participate in the development of non-alcoholic fatty liver disease.
- Exosomes are ideal biomarkers for the diagnosis of non-alcoholic fatty liver disease.
- Exosomal microRNAs can differentiate the different stages of non-alcoholic fatty liver disease.
- Stem cell-derived exosomes can ameliorate the course of non-alcoholic fatty liver disease.
- Exosomal blocking can prevent non-alcoholic fatty liver disease progression.

1 Introduction

Affecting one out of four adults globally, Non-alcoholic fatty liver disease (NAFLD) remains a growing worldwide health issue (1). It is highly associated with metabolic syndrome and commonly accompanies insulin resistance, obesity, and dyslipidemia (1). Moreover, it is increasing due to the Growing obesity and diabetes prevalence (2, 3).

NAFLD is a histological definition describing macrovesicular steatosis in more than 5% of hepatocytes in people with no or little alcohol consumption. It includes two major types: non-alcoholic fatty liver (NAFL) and non-alcoholic steatohepatitis (NASH).

Non-alcoholic fatty liver (NAFL), or simple steatosis, is a form of NAFLD. While NAFL is typically accompanied by a lower risk of liver-related mortality, it can progress to non-alcoholic steatohepatitis (NASH) (4). NASH is more commonly associated with cirrhosis and hepatocellular carcinoma (HCC), both of which can lead to liver-associated death (4). Histologically, NASH consists of ballooning degeneration and lobular inflammation along with steatosis, with or without perisinusoidal fibrosis (5). However, present theories propose a dynamic two-way cycling between NAFL and NASH, with fibrosis advancing slowly in most patients (4). The reality that the initiation of progressive fibrosis ultimately determines clinical results raises doubts about the importance of differentiating between NAFL and NASH (4).

Although there has been a lot of effort in developing non-invasive tools for NAFLD diagnosis, the diagnosis of NAFLD is mainly based on liver biopsy (6, 7). However, it is an invasive and costly procedure, and, considering the prevalence of the disease, impractical. Therefore, non-invasive diagnostic methods that could be accurate can be crucial in this area. Along with lifestyle modification and weight loss, several agents, including pioglitazone, semaglutide, and obeticholic acid, have shown promise in the therapy of NAFLD (7). However, with more than 50 treatment agents currently being studied in different clinical phases, the treatment of NAFLD remains challenging (7).

Exosomes are nanosized extracellular vesicles that have significant roles in the pathogenesis of different diseases (8). Recently, their theragnostic potentials have been broadly discussed in various diseases, including cardiac, renal, hepatic, and neurological diseases, as well as neoplasms (8). In the presenting paper, we aimed to review the potential and perspectives of exosomes as a diagnostic and therapeutic agent in NAFLD.

2 Exosomes

Exosomes are bi-lipid membrane extracellular vesicles sized from 50 to 140 nm (9–11). They are secreted by nearly all cells and can be found in all body fluids (12–15). Exosomes carry a wide variety of proteins, DNAs, RNAs, and lipids, with specific characterization based on the secreting cell (16–19). They have higher levels of specific proteins [such as heat shock protein 70 (HSP 70), tetraspanins (CD9, CD 63, CD 81, and CD 82), ALG-2-interacting protein X (ALIX), tumor susceptibility gene 101 (TSG101)] and lipids [such as flotillin and glycosylphosphatidylinositol-anchored protein (LBPA)] (20–22).

Exosome biogenesis involves complex intracellular pathways with distinct mechanisms for different EV subtypes (23). The endosomal system plays a central role in exosome biogenesis (24). Traditionally, the endosomal sorting complex required for transport (ESCRT) machinery has been considered essential for the biogenesis of exosomes (24). The biogenesis of exosomes involves four key steps: cargo sorting, the development and maturation of MVBs, the transportation of these MVBs, and finally, the fusion of MVBs with the cellular membrane (25). Each process is modulated through the competition or coordination of multiple mechanisms, whereby diverse repertoires of molecular cargos are sorted into distinct subpopulations of exosomes, resulting in the high heterogeneity of exosomes.

Recognition of Ubiquitinated Cargo is the first step in exosome biogenesis (24). The process begins with the recognition of ubiquitinated cargo by the ESCRT-0 complex (24). Ubiquitination is a post-translational modification where a small protein called ubiquitin is attached to a target protein (26). This serves as a recognition signal for ESCRT-dependent cargo sorting. Once the ubiquitinated cargo is recognized, it is sequestered into endosomal microdomains by the ESCRT-I and ESCRT-II complexes (25). These complexes also initiate the budding of the endosomal membrane into the lumen of the endosome, forming the nascent ILV. The ESCRT-III complex is responsible for driving the budding process to completion, leading to the formation of ILVs (24). It does this by assembling into a spiral-shaped structure on the endosomal membrane, which constricts the neck of the budding ILV. ILVs are formed within the lumen of an endosome through the inward budding of the endosomal membrane. When multiple ILVs accumulate within an endosome, it is then referred to as a multivesicular body (MVB). MVBs can either fuse with lysosomes for degradation or with the plasma membrane, releasing ILVs as exosomes.

Recent findings suggest the existence of an alternative pathway for sorting exosomal cargo into MVBs in an ESCRT-independent manner (24). Nonetheless, the pathways may not be completely distinct (27). This ESCRT-independent pathway seems to depend on raft-based microdomains for the lateral segregation of cargo within the endosomal membrane (28). For instance, the nSMase2-ceramide pathway is vital for ESCRT-independent exosome biogenesis (29). Besides, Tetraspanins are a family of proteins that play a crucial role in the biogenesis of exosomes, including the ESCRT-independent pathway (30). They are characterized by their four transmembrane domains, which allow them to interact with various other proteins, cholesterol, and gangliosides. These interactions lead to the formation of tetraspanin-enriched microdomains (TEMs) on the membrane (30). These TEMs can influence membrane bending and actin polymerization, which are essential steps in the formation of multivesicular bodies (MVBs) and, subsequently, exosomes (30). In addition to their role in exosome biogenesis, tetraspanins are also sorted into exosomes in an ALIX- and ESCRT-III-dependent manner (31). This sorting process occurs independently of other ESCRTs but requires lysobisphosphatidic acid (LBPA) *in vivo*.

Lately, exosomes have got much attention in diagnostics. They constitute a key intracellular communication system and act as a key factor in the pathogenesis of different diseases via intercellular signaling cascades (23, 32–35). Therefore, detecting exosomes, molecules carried by them such as nucleic acids, and their surface proteins may be used as an early, non-invasive, and potentially accurate diagnostic tool in different diseases (8). Their potential diagnostic value has been proven in cancers and diseases of the lungs, kidneys, liver, and central nervous system (36–42).

Exosome offers favorable features as a therapeutic agent. It has the same homing behavior as the secreting parent cell (43). Additionally, it benefits from a bilipid membrane, which allows it to carry considerable amounts of contents and protect its cargo from degradation by chemicals and enzymes. Considering that along with their targeted activity, exosomes are fruitful options in drug delivery applications. Moreover, they have significantly lower immunogenicity than virus-based drug delivery systems and

liposomes (44). Besides, exosomes derived from mesenchymal stem cells provide immunomodulatory and anti-inflammatory effects, promote cellular viability, and facilitate cellular proliferation and neoangiogenesis (45–50). In certain settings, exosomes can have regenerative and homeostatic, thus therapeutic effects on diseased tissue (51). On top of that, exosomes lack risks of carcinogenesis and immune response to cancers and infections, compared with cell therapies, offering high therapeutic potential with appreciable safety (9).

3 Pathophysiology of NAFLD

The complex pathophysiology of NAFLD has not yet been fully clarified. However, it is believed that it is related to the interaction among genetic, environmental, and individual factors (52).

There are different hypotheses about NAFLD pathophysiology. In the past, the “two-hit hypothesis” was used to explain the mechanism of NAFLD and NASH pathophysiology, but currently, the “multiple-hit model” is suggested (52). According to the two-hit model, the initial phase, often referred to as the “first hit,” involves the build-up of fat, specifically triglycerides, within the liver tissue and the development of insulin resistance. These elements are considered the key drivers of hepatic steatosis when the accumulation surpasses the threshold of 5% (53). The “second hit” is characterized by alterations in the levels of inflammatory cytokines and adipokines, mitochondrial dysfunction, and oxidative stress. These changes can trigger necroinflammation and fibrosis within the liver tissue (54, 55). However, recent investigations have shown that the “two-hit hypothesis” cannot fully explain the exact mechanism of human NAFLD. Therefore, the “multiple-hit model” is currently the most recognized theory. It suggests a broader metabolic dysfunction due to the interplay of genetic and environmental influences, as well as alterations in the crosstalk between the liver and various organs and tissues, such as the adipose tissue, pancreas, and gut (52, 54–57). Despite this, the initial stages, or “first hits,” are still believed to be the accumulation of fat in the liver triggered by obesity and insulin resistance (52).

NASH and NAFLD begin with the excessive accumulation of triglycerides (TG) in the liver (58). The fatty acids used for this hepatic TG accumulation are derived from three sources: (I) dietary fat taken up in the intestine, (II) *de novo* lipogenesis from glucose and fructose, and (III) dysregulated adipocytes’ triglyceride lipolysis leading to excessive fatty acid transport to the liver (58–60). When the liver’s capacity to utilize carbohydrates and fatty acids, as the primary metabolic energy substrates, is impaired, toxic lipid species are accumulated in hepatocytes. It, in turn, induces hepatocellular injury and death, which results in genomic instability and fibrogenesis that put the liver at risk of cirrhosis and hepatocellular carcinoma. However, the specific toxic lipid species that promote cell injury are not fully recognized (6). Lipotoxic lipids accumulation causes hepatocellular injury through endoplasmic reticulum (ER) stress (61), a dysregulated unfolded protein response (UPR) (62), activation of the inflammasome and apoptotic pathways (63), an increased hepatic Hedgehog (Hh) signaling, and inflammation (64). There are many external factors contributing to hepatocellular injury, including insulin resistance (65), adipokine

dysregulation (66), hepatic ATP depletion (67), uric acid-induced hepatocyte mitochondrial dysfunction (68), and the effects of intestinal microbiota products (69). Fibrogenesis results from extracellular signaling from injured hepatocytes, liver sinusoidal endothelial cells, activated Kupffer cells, T cells, B cells, and natural killer cells, promoting hepatic stellate cells (HSCs) activation. Activated HSCs transdifferentiate into fibrogenic myofibroblasts that can produce extracellular matrix proteins at a higher rate than their degradation (70). Finally, Progressive fibrosis leads to cirrhosis, which subsequently promotes portal hypertension and liver failure, which is the main reason for liver-related mortality in NAFLD (6).

4 Exosome in NAFLD

4.1 Role of exosomes in NAFLD pathophysiology

4.1.1 Exosomes in the pathogenesis of NAFLD

Several recent studies point to a significant role of Extracellular Vesicles (EVs) in the pathophysiology and progression of NAFLD and NASH. EVs play a central role in normal intercellular communication. Different types of liver cells, including human adult liver stem cells, cholangiocytes, hepatocytes, hepatic dendritic cells, and hepatic stellate cells, function as both exosome-secreting and exosome-targeted (42). Various liver conditions, including NAFLD, seem to increase the basal EV’s secretion (71). Povero et al. (72) have shown a considerable rise in the concentration of EVs in the liver and blood of diet-induced NAFLD animals. Normal hepatocytes produce exosomes that carry several cargos, such as proteins and miRNAs (73). However, exosomes secreted from damaged hepatocytes are essential in inducing hepatocellular inflammation and fibrosis during liver damage. These effects are through intercellular communication between different cell types (74). The roles of exosomes and exosomal miRNAs in NAFLD pathogenesis and the interactions between different organs are illustrated in Figures 1, 2.

4.1.2 Lipid accumulation

NAFLD is characterized by excessive hepatic lipid accumulation (75). The accumulation of lipids in the liver cells results in the release of stress signals, triggering the activation of inflammatory pathways that, when perpetuated, lead to chronic injury and fibrosis over time. Hepatocytes release EVs in response to lipotoxic fatty acids, which leads to HSCs fibrogenic activation and promotes macrophage chemotaxis (76, 77). Inflammation and fibrosis are essential for NAFLD progression (78).

Li et al. (79) demonstrated that the exosomal *miR-199a-5p* induces lipid build-up in the liver through a down-regulation of its target gene, hepatic Mammalian sterile 20-like kinase 1 (*MST1*). This leads to an alteration in Sterol regulatory element-binding protein 1 (*SREBP1c*), AMP-activated protein kinase (AMPK) signaling cascades and consequently suppresses Carnitine palmitoyltransferase I α (*CPT1 α*) lipolysis gene and induces Fatty acid synthase (*FASN*) lipogenesis gene expression.

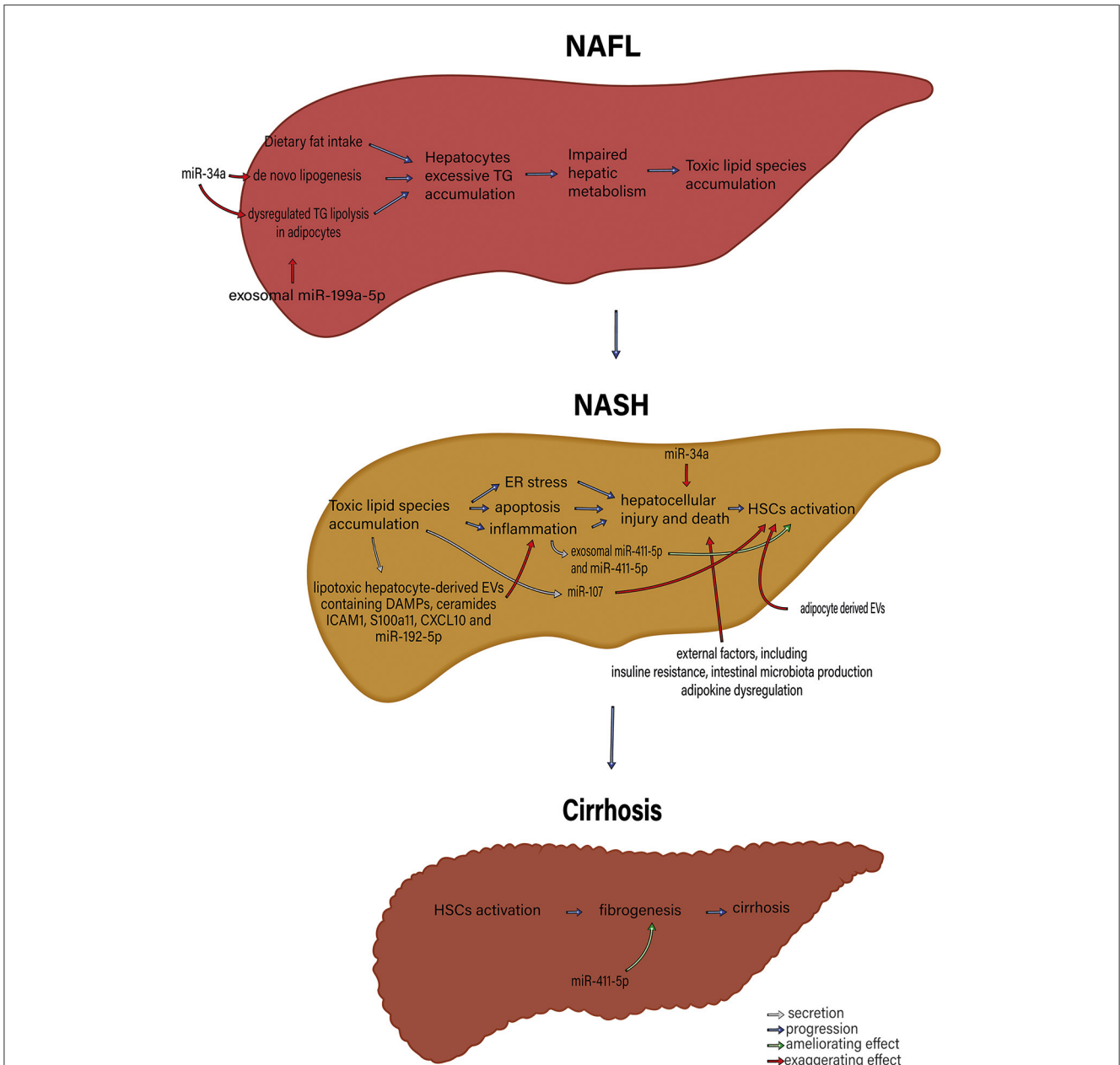
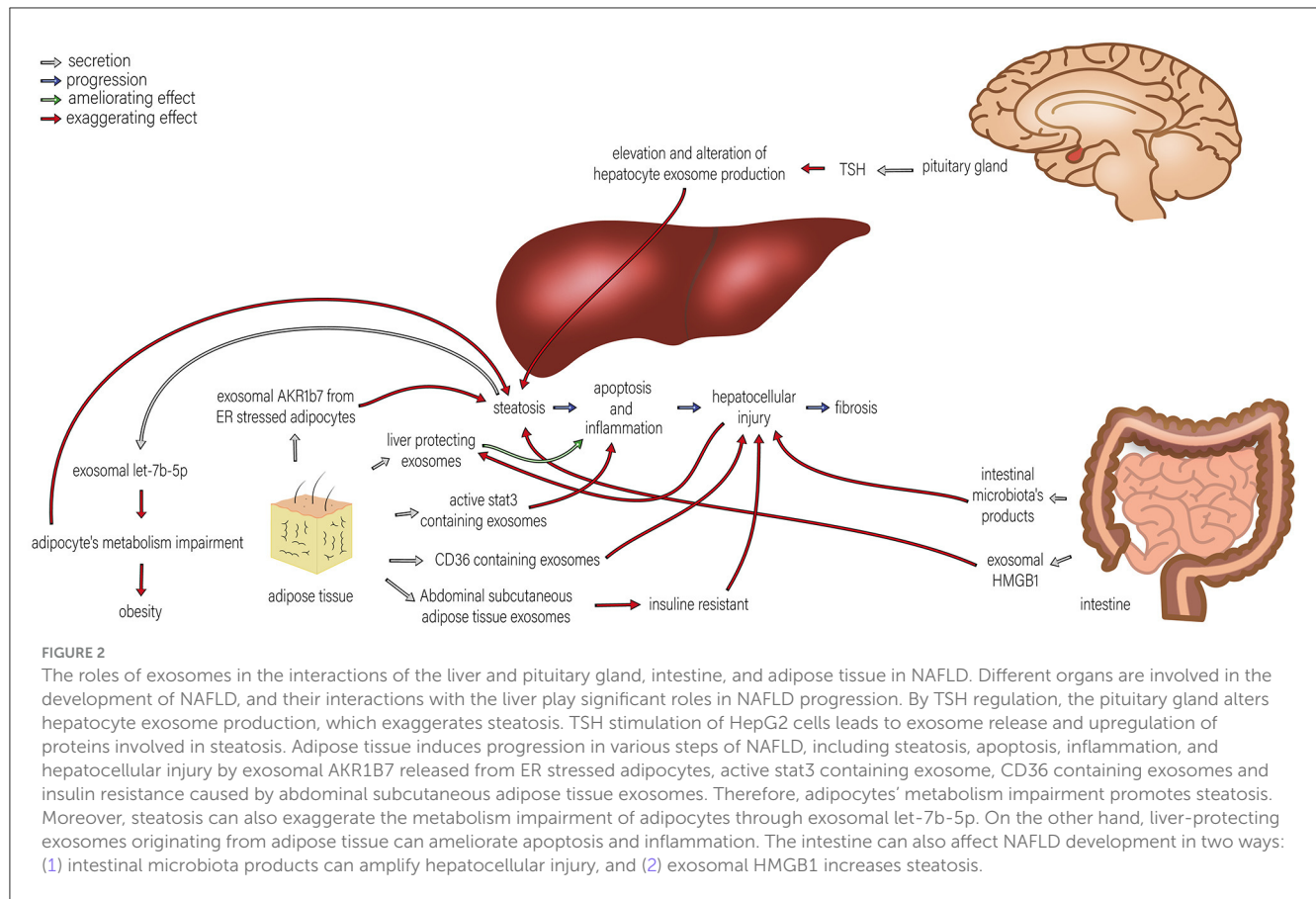


FIGURE 1
 The roles of exosome and exosomal miRNAs in NAFLD pathophysiology. microRNAs, including miR-34a and exosomal miR-199a-5p, that control *de novo* lipogenesis and dysregulated TG lipolysis in adipocytes significantly induce excessive hepatocyte TG accumulation. Subsequently, TG accumulation in hepatocytes gives rise to impaired hepatic metabolism and toxic lipid species accumulation. The accumulation of toxic lipid species leads to ER stress, apoptosis, and inflammation. These changes contribute to hepatocellular injury and death, followed by HSC activation. MiR-34a, exosomal miR-411-5p and miR-411-5p, miR-17, adipocyte-derived EV, lipotoxic hepatocyte-derived EVs containing DAMPs, ceramides, ICAM1, S100a11, CXCL10 and miR-195-5p and external factors including insulin resistance, intestinal microbiota production, and adipokine dysregulation participate in various parts of this process. HSC activation results in fibrosis and, eventually, cirrhosis. miR, microRNA; TG, triglyceride; ER, endoplasmic reticulum; DAMPs, Damage-associated molecular patterns; ICAM-1, Intercellular Adhesion Molecule 1; HSCs, Hepatic stellate cells; EVs, extracellular vesicles.

In another study, Xu et al. (80) reported that hepatocyte exosomal *miR-34a* plays an essential role in the development of NAFLD by increasing lipid absorption and synthesis and reducing fatty acid oxidation. It also promotes NAFLD's transition to NASH by regulating Kupffer cell activation, which promotes inflammatory responses, increases hepatic ROS levels, and induces hepatic apoptosis.

4.1.3 Hepatocellular inflammation

Lipotoxic hepatocyte-derived EVs cause hepatocellular inflammation. NAFLD is closely related to chronic inflammation associated with macrophages and neutrophils (78). Additionally, innate immune activation is fundamental in triggering hepatic inflammation in NASH (81). Hirsova et al. (82) have demonstrated that incubation of primary hepatocytes with



lysophosphatidylcholine (LPC) increased their EV secretion compared with control cells *in vitro*. EVs derived from hepatocytes, carrying TNF-related apoptosis-inducing ligand, activated *IL-1β* and *IL-6* messenger RNAs expression in macrophages derived from mice bone marrow, leading to inflammation and liver injury.

Regarding macrophage infiltration, Ibrahim et al. (83) have demonstrated that in hepatocyte lipotoxicity with LPC, Mixed lineage kinase 3 (MLK3) signaling induces the secretion of EVs containing elevated levels of CXCL10 from hepatocytes. These EVs induce bone marrow-derived macrophage chemotaxis. Moreover, Hepatocyte-derived ceramide-dependent EVs are enriched in several distinct damage-associated molecular patterns and cellular adhesion molecules, including ICAM1 and S100A11, that may influence immune cell responses, leading to hepatocellular inflammation (84). In addition, Kakazu et al. (77) have demonstrated that hepatocytes treated with palmitic acid (PA), a saturated fatty acid found in NAFLD hepatocytes, release significant amounts of C16:0 ceramide-enriched EVs in an inositol-requiring enzyme-1α (IRE1α)-dependent manner. IRE1A-stimulated hepatocyte-derived EVs, having ceramide-derived sphingosine 1-phosphate, enhance macrophage migration, which results in an inflammatory response in the liver (77, 85).

Moreover, in NAFLD, imbalanced macrophage polarization toward the pro-inflammatory M1 phenotype significantly contributes to disease progression (86). Conversely, M2 macrophages, particularly the M2a and M2c subtypes, exert

protective effects in NAFLD (87). M1 macrophages are involved in the immune response and immune monitoring through antigen presentation and the release of pro-inflammatory cytokines like *IL-1β* and *TNF-α* (87). On the other hand, M2 macrophages have limited antigen presentation capabilities and contribute to immune regulation by dampening the immune response with the secretion of inhibitory cytokines such as *IL-10*, *TGF-β*, and *Mrc* (88). M2 macrophages can contribute to tissue healing and renewal. The decision of whether macrophages exhibit a pro-inflammatory reaction that causes injury or an anti-inflammatory response that offers protection depends on the balance between M1 and M2 activation tendencies (89).

Liu et al. (90) found that exosomal *miR-192-5p* and hepatocyte-derived exosome levels are considerably higher in NASH patients' serum, similar to high-fat, high-cholesterol diet (HFHCD)-fed rat NASH models. Progression of NAFLD in HFHCD-fed rats is directly associated with serum *miR-192-5p* and proinflammatory M1 macrophage amounts along with proinflammatory cytokines expression (90). Another study (91) showed that exosomes originating from lipotoxic hepatocytes and containing *miR-192-5p* significantly activate macrophages. They polarize macrophages to proinflammatory M1 phenotype by regulating rapamycin-insensitive companion of mammalian target of rapamycin (Rictor)/Akt/Forkhead Box Transcription Factor O1 (FoxO1) signaling pathway. In addition, saturated fatty acids and Cholesterol, via inducing lysosomal dysfunction, promote exosomal *miR-122-5p* secretion from hepatocytes. It results in

pro-inflammatory M1 macrophage polarization and inflammatory activation (92).

The interaction between hepatic stellate cells (HSCs) and macrophages plays a crucial role in the pathogenesis of liver fibrosis (93, 94). Nevertheless, the specific molecular process that facilitates communication between hepatic stellate cells (HSCs) and liver macrophages, particularly the M2 subtype, remains incompletely understood. Exosomes are suggested to mediate the connection between mentioned cells, especially through the miRNAs (94). The polarization of M2 macrophages is strongly linked to the suppression of HSC activation and the reduction of liver fibrosis (95–97). Wan et al. (94) showed that HSC activity was significantly inhibited by exosomes derived from M2 macrophage. They found that exosomal *miRNA-411-5p* is reduced in the liver tissue and serum of HFHC diet-induced rat model of NASH. They also showed that exosomal *miR-411-5p* from M2 macrophages hinder the HSCs activity and lead to inactivation of stellate cells through downregulation of the expression of Calmodulin-Regulated Spectrin-Associated Protein 1 (CAMSAP1). The inhibition of HSCs occurred following the knockdown of CAMSAP1 as a direct target of *miRNA-411-5p*. Furthermore, the amount of *miR-411-5p* in exosomes derived from M2 macrophages was higher than that in M1 macrophages. Recent literature shows that *MiR-411-5p* may play a role in the regulation of hepatocellular carcinoma progression (98).

Liu et al. (91) found that *arginase 1* (Arg1), a marker of M2 macrophages, was reduced after an 8-week high-fat high-cholesterol diet (HFHCD) in a rat model, but other M2 markers, *chitinase 3-like protein 3* (Ym1) and *found in inflammatory zone 1* (Fizz1), remained unchanged throughout the 16-week diet. Interestingly, they discovered that hepatocyte-derived exosomal miR-192-5p did not induce M2 macrophage polarization, and neither did exosomes from PA-treated HepG2 cells affect M2 macrophage activation. They also observed a decrease in Ym1 expression in THP-1 macrophages after exposure to serum exosomes from NASH patients, an effect that was reversed with a miR-192-5p inhibitor. In another study, Zhao et al. (92) investigated the potential role of cholesterol-loaded hepatocytes in inducing macrophage polarization through exosome-related cross-talk. They found that exosomes from Huh7 cells loaded with ox-LDL and M β CD-cholesterol did not affect the percentage of M2 macrophages. In conclusion, these studies suggest that while certain exosomes and exosomal miRNAs released by cholesterol-loaded hepatocytes can influence M2 macrophage polarization, their effects are complex and may depend on the specific conditions and markers examined. Further research is needed to fully understand these mechanisms and their implications in NAFLD. This could potentially open new avenues for therapeutic interventions targeting macrophage polarization. Changing the macrophage polarization in hepatic tissue toward M2 macrophages may exert protective effects against NAFLD progression and liver fibrosis. This should be further studied as a potential therapeutic pathway in NAFLD.

Besides, hypoxia can induce NAFLD in obstructive sleep apnea (OSA) syndrome by exosomes. Hypoxia triggers the production of certain exosomes (99). Yang et al. (100) mentioned that OSA-induced exosomes promote hepatocyte steatosis and activate macrophages in liver tissue. These exosomes were observed to

enhance fat accumulation. They demonstrated that after the uptake of OSA-induced exosomes by macrophages, the polarization of macrophages toward the M1 type occurred. It led to the inhibition of sirtuin-3 (SIRT3)/AMP-activated protein kinase (AMPK) and autophagy. It also enhanced the activation of the nucleotide-binding domain, leucine-rich-containing family, and pyrin domain-containing-3 (NLRP3) inflammasomes. The use of 3-methyladenine (3-MA) to block autophagy prevented NLRP3 inflammasome activation and hindered M1 macrophage polarization. Moreover, they reported elevated levels of miR-421 in OSA-induced exosomes in OSA plus NAFLD mice and patients. In the liver tissues of OSA and OSA plus NAFLD mice, *miR-421* showed the same co-localization with macrophages. Hepatocytes exposed to intermittent hypoxia transferred *miR-421* to macrophages through exosomes to inhibit SIRT3 and contribute to macrophage M1 polarization. Accordingly, MiR-421 targeting reduced SIRT3 protein levels in macrophages. They also found that in *miR-421*^{-/-} mice subjected to OSA and NAFLD modeling, liver steatosis and M1 polarization were notably reduced. Knockout of *miR-421* alleviated the inhibitory effects of OSA-induced exosomes on SIRT3 and autophagy, leading to reduced liver steatosis and macrophage M1 polarization. On the other hand, they haven't found any change in the transcription of M2 macrophage genes Arg1 and Fizz1 Following the uptake of OSA exosomes by macrophages.

Other than the above mechanisms, Shen et al. (101) showed that decreased hepatocyte autophagy, found in non-alcoholic steatohepatitis, develops IL-1 β /TNF-induced hepatic injury and inflammation through the exosomal release of the damage-associated molecular pattern (DAMPs). In addition, liver inflammation could be caused by EVs secreted from other cells. For example, It has been reported that platelet-derived EVs may have a pro-inflammatory role in the liver (102). However, some studies have demonstrated the opposite (103).

4.1.4 Fibrogenesis

Hepatic stellate cells (HSCs) promote fibrogenesis through EVs intracellular communication. Literature (104) has shown that hepatocytes stimulated by palmitate (PA) secret significantly more exosomes with distinctive miRNA expression patterns, amplifying fibrotic gene expression in HSCs. In this regard, Wei Wang et al. (105) illustrated that hepatocytes treated with PA release more *miR-107*-enriched exosomes, leading to HSC proliferation and activation by two distinct pathways. These hepatocyte-derived exosomes transfer *miR-107* to HSCs, where *miR-107*, by directly inhibiting Dickkopf-1 (*DKK1*) expression, activates Wnt signaling. Additionally, these exosomes deliver *miR-107* to CD4⁺ T cells, where *miR-107* activates the Raf/MEK/ERK signaling pathway by upregulating *IL-9* expression via Forkhead box protein P1 (*Foxp1*) inhibition. These pathways lead to HSC activation, an essential component of NAFLD pathogenesis.

On the other hand, exosomes may exert protective effects on NAFLD. Studying on a diet-induced rat model of NASH, Qi et al. (106) proved that M2 macrophage-derived exosomes enriched with *miR-411-5p* directly reduce Calmodulin-Regulated Spectrin-Associated Protein 1 (CAMSAP1) expression. This

consequently inhibits hepatic stellate cell activation and fibrosis. In addition, Hou et al. (107) found that *miR-223*-enriched exosomes are released from myeloid cells in response to IL-6 signaling. They offer protective effects on NASH progression by inhibiting the expression of profibrotic genes that *miR-223* targets, including C-X-C motif chemokine (*Cxcl10*), NOD-, LRR—and pyrin domain-containing protein 3 (*Nlrp3*), transcriptional activator with PDZ-binding (*Taz*), and insulin-like Growth Factor 1 Receptor (*Igf1r*) in the liver.

4.1.5 Adipocyte-hepatocyte interaction

Adipocyte-hepatocyte interaction through EVs plays an essential role in NAFLD progression. Afrisham et al. (108) demonstrated that plasma exosomes isolated from obese women may play a role in NAFLD development by inducing insulin resistance, increasing hepatocellular TG levels, and decreasing hepatocellular FGF21 secretion. These exosomes promote a significant increase in the expression of *integrin $\alpha v \beta -5$* and tissue inhibitor of matrix metalloproteinase-1 (*TIMP-1*) along with a concomitant decrease in plasminogen activator inhibitor-1 (*PAI-1*) and matrix metalloproteinase-7 (*MMP-7*) expression in HepG2 cells. In hepatic stellate cells, these exosomes induce increased *integrin $\alpha v \beta -5$* and 8, *Smad-3*, *TIMP-1* and 4, and matrix metalloproteinase-9 (*MMP-9*) expression. Consequently, these processes dysregulate the tumor growth factor- β (TGF- β) signaling pathway, producing a profibrotic state in liver cells (109).

According to the tissue-cooperative homeostatic model of NAFLD, during the early phases of NAFLD progression, the falling levels of the hepatocellular production of *miR-122* are compensated by an increase in the adipose *miRNA*-containing exosome secretion (110). These molecules are eventually taken up by the hepatocytes and augment these cells' endogenous production of *miRNA*. Thus, metabolic damage to adipose tissue (due to liver disease progression) eventually decreases the external supply of liver-supporting *miRNAs*, leading to hepatic fibrosis and carcinogenesis.

Ogur et al. (111) determined that exosomes derived from Endoplasmic Reticulum (ER) stress-induced adipocytes transfer exosomal Aldo-keto-reductase 1b7 (*Akr1b7*) to hepatocytes. This elevates glycerol levels in liver cells, which gives rise to hepatic steatosis, inflammation, and eventually fibrosis. Additionally, Yan et al. (112) showed that PA-induced AMPK $\alpha 1$ inhibition in adipocytes promotes CD36-containing exosome secretion. These exosomes contribute to HepG2 cell damage and mediate the development of High-fat diet (HFD)-induced NAFL. Furthermore, exosomal *let-7b-5p* originated from hepatocytes is essential in the interaction between hepatocytes and adipocytes by TGF- β signaling. It impairs the energy balance of adipocytes by regulating *Adrb3* gene expression, resulting in hepatic steatosis and obesity (113). Besides, Fuchs et al. (114) found that subcutaneous abdominal adipose tissue-derived exosomes and raised levels of *PAI-1* take part in the pathophysiology of insulin resistance. Insulin resistance is a critical factor in the NAFLD pathobiology (115).

Exosomes of adipose tissue may also have beneficial effects on NAFLD. Zhao et al. (116) demonstrated that exosomes from adipose-derived stem cells (ADSCs) promote arginase-1 expression in macrophages by carrying active STAT3 and inducing

M2 phenotype macrophage polarization. This results in the inhibition of macrophage inflammatory responses. It suggests the potential efficacy of exosome and stem cell therapy in preventing NAFLD progression.

4.1.6 Other pathological pathways

EVs also contribute to the development of NAFLD through other pathophysiological mechanisms. In a study on *Asc^{-/-}* mice on a high-fat diet (HFD), Chen et al. (117) revealed a gut-liver axis mechanism in which exosomes play a significant role. In dysbiosis, exosomes act as the transporter of high mobility group box 1 (HMGB1) protein from the intestine to the liver, triggering hepatic steatosis. Besides, a preliminary study (118) on TSH-induced lipotoxicity in NAFLD revealed that TSH stimulation of HepG2 cells significantly increases their exosomal production and alters their exosomal proteomic profile. It leads to upregulation of proteins involved in different biological processes such as metabolism, inflammation, and apoptosis. Hence, it may be involved in NAFLD pathogenesis.

4.1.7 Exosomal micrnas

miRNAs are the most abundant cargo molecules transferred by exosomes (119). Specific *miRNAs* have been associated with NAFLD and may be effectual in its pathogenesis. They may also be used as non-invasive options in NAFLD diagnosis. Zhang and Pan (120) monitored serum exosomal *miRNAs* in children with NAFLD and identified 2,588 *miRNAs*. They revealed that in children with NAFLD, the expression of 80 *miRNAs*, importantly *miR-122-5p*, *miR-335-5p*, and *miR-27a*, differs from that of the control group. In another study, Zhou et al. (121) suggested that exosomal *miRNAs* might take part in the pathophysiology of NAFLD and revealed an upregulation of *miR-146b-3p*, *miR-155-5p*, *miR-122-5p*, and *miR-34a-5p* in NAFLD patients. *miR-122*, a highly liver-specific *miRNA*, takes part in lipid metabolism and is detected in the form of exosomes in the serum of NAFLD patients (122, 123). *miR-21* is another *miRNA* that is increased in the liver of NAFLD patients, as well as animal models of the disease. This *miRNA* regulates hepatocellular glucose and lipid metabolism. It works through a complex transcription network. At different stages, *miR-21* may be involved in NAFLD progression, including early steps of the initiation of hepatocellular steatosis and later steps of inflammation and fibrosis (124). A list of exosomal *miRNAs* that may be beneficial for NAFLD diagnosis is presented in Table 1.

4.2 Exosomes in NAFLD diagnosis

Studies have shown that exosomes, due to their contents, including *miRNAs*, may serve as potential diagnostic biomarkers for disease progression and severity in NAFLD (142). The content of exosomes can change in various diseases, and this is not only limited to exosomal *miRNAs*. As mentioned earlier, several exosomal *miRNAs* are considerably altered in NAFLD, and their potential role in NAFLD pathophysiology remains an important topic for a better understanding of NAFLD development and the mechanisms that help us ameliorate the

TABLE 1 Potential exosomal microRNAs in NAFLD diagnosis.

microRNA	Associated tissue	Associated condition	Alteration of microRNA levels	Target gene/protein	Pathogenesis	References
<i>miR-21</i>	Serum	NAFLD	Increased	<i>PPARα</i>	Dysregulating lipid metabolism in the liver	(125)
				<i>FABP7</i>	Dysregulating the transporting and metabolizing fatty acid	(126)
				<i>HMGCR</i>	Producing cholesterol and isoprenoids	(127)
				<i>HBP1</i>	Dysregulating the cell cycle, transcriptional repressor	(128)
		NASH	Increased	<i>SMAD7</i>	Suppressing the Inhibition of the TGF-β signaling	(129)
Liver fibrosis	Increased	<i>TIMP-3</i>	Inhibiting the matrix metalloproteinases	(130)		
<i>miR-122</i>	Serum	NAFLD	Increased	<i>SIRT1</i>	Suppressing the regulation of hepatic lipid metabolism Promoting hepatic oxidative stress Promoting hepatic inflammation	(131, 132)
<i>miR-27a</i>	Liver	NAFLD	Increased	<i>FASN</i> and <i>Scd1</i>	Attenuating hepatic <i>de novo</i> lipogenesis Alleviated obesity-initiated NAFLD	(133)
		MAFLD		<i>PINK1</i>	Inhibiting mitophagy Promoting MAFLD-related liver fibrosis	(134)
<i>miR-155-5p</i>	Liver	NAFLD	Increased	<i>STC1</i>	Inducing hepatic mitochondrial dysfunction, vascular insulin resistance Promoting endothelial cell activation and atherosclerosis	(135–137)
<i>miR-34a</i>	Serum	NASH NAFLD	Increased	<i>PPARα</i> and <i>SIRT1</i>	Hepatic lipid accumulation	(138)
<i>miR-192</i>	Serum	NAFLD	Decreased	<i>SREBF1</i>	Dysregulating lipid homeostasis in hepatocytes	(139)
<i>miR-181a</i>	Serum	NAFLD	Increased	<i>SIRT1</i>	Reducing insulin sensitivity Increasing gluconeogenesis and lipid synthesis	(140)
<i>miR-29a</i>	Serum	NAFLD	Decreased	<i>GSK3β</i>	Promoting of mitochondrial proteostatic stress	(141)
<i>miR-199a-5p</i>	Adipose tissue	NAFLD	Increased	<i>MST1</i>	Aggravating lipid accumulation in hepatocytes	(79)

PPARα, Peroxisome proliferator-activated receptor alpha; FABP7, Fatty Acid Binding Protein 7; HMGCR, 3-Hydroxy-3-Methylglutaryl-CoA Reductase; HBP1, HMG-Box Transcription Factor 1; SMAD7, SMAD Family Member 7; TIMP-3, Tissue inhibitors of metalloproteinases 3; SIRT1, Sirtuin 1; FASN, Fatty acid synthase; SCD1, Stearoyl-CoA Desaturase-1; PINK1, PTEN-induced kinase 1; STC1, Stanniocalcin 1; SREBF1, Sterol regulatory element-binding protein-1; GSK3β, Glycogen synthase kinase-3 beta; MST1, Macrophage Stimulating 1; MAFLD: Metabolic dysfunction-associated fatty liver disease.

condition. Meanwhile, the modification of the level of these miRNAs may provide a fruitful non-invasive diagnostic approach for NAFLD, considering the challenges in diagnosing NAFLD. A list of exosomal miRNAs potential for NAFLD diagnosis is presented in Table 1. Moreover, various studies have shown that changes in the level of exosome contents other than miRNAs, including protein FZD-7, can also act as diagnostic and prognostic biomarkers for NAFLD (143, 144). In addition to the content of exosomes, the level of certain EVs in mouse models and human subjects of NASH has been reported to be higher than that of normal individuals, suggesting specific exosomes' levels as potential factors for differentiating the stages of NAFLD (145). However, although exosomes offer favorable non-invasive methods

for the diagnosis and prognosis evaluation of NAFLD, there is a lack of comprehensive information regarding the sensitivity and specificity of exosome-based diagnostic methods. There is a need for further investigation to understand better the diagnostic capabilities of exosomes and their practical application in clinical settings for NAFLD assessment.

4.3 Exosome-related strategies for NAFLD treatment

Resmetirom has been recently approved by the Food and Drug Administration (FDA) and remains the only approved option

for NAFLD treatment along with lifestyle modifications, such as diet control and exercise. However, several drugs have shown potential therapeutic effects on NAFLD, such as elafibranor (by inhibiting lipid deposition), emricasan (via lowering cell death), IMMe124 (through regulating intestinal microenvironment and metabolism) (146). Moreover, Probiotics and prebiotics, symbiotic, fecal microbiome transplantation, and fasting-mimicking diet could also be beneficial in patients with NAFLD (147). Due to their unique characteristics, exosomes have gained attention as a new treatment option in different diseases. Exosomes have been studied for various clinical applications, including as a diagnostic/prognostic biomarker, cell-free therapeutic agent, drug delivery carrier, and cancer vaccines (148). Several exosome-based treatment approaches have shown efficacy in liver diseases, including NAFLD, which is described below. Figure 3 demonstrates potential exosome-based treatment approaches in NAFLD.

4.3.1 Cell-derived exosomes

In recent years, interest in therapeutic usage of cell derivatives such as exosomes over cell therapies has been increasing (149, 150). Exosomes are released by the majority of body cells and have been detected in nearly all fluids of the body, such as blood, cerebrospinal fluid, urine, amniotic fluid, breast milk, saliva, bile, malignant ascites, and lymph (151–158). Among all cell types, Mesenchymal stem cells (MSCs) are the most prolific producer of exosomes (159). MSCs can be derived from different body tissues, including peripheral blood, adipose tissue, bone marrow, umbilical cord, spleen, liver, pancreas, kidney, lung, thymus, and brain (160, 161). They are known as a subgroup of stromal stem cells. MSCs are adults' most researched type of stem cells in regenerative medicine owing to their extensive presence in body tissues and easy expansion procedure *in vitro*. They have shown the potential to prolong and save lives (162). Mesenchymal stem cell therapy has several disadvantages that must be considered, including the possibility of tracking in pulmonary capillaries, leading to pulmonary embolism, reduction in cell viability of MSCs in cryogenic storage during transport, and the risk of tumorigenesis after administration (163–166).

Exosomes mediate intracellular communication by transferring a variety of biomolecules like microRNAs (miRNAs), messenger RNAs (mRNAs), other non-coding RNAs, and lipids (167). Exosomes derived from stem cells have demonstrated the same therapeutic potential as their parental cells (162). It has been indicated that mesenchymal stem cell-derived exosome can alleviate liver fibrosis, decrease Alanine aminotransferase (ALT) and Aspartate transaminase (AST) levels, and mitigate liver inflammation (168). Thus, these exosomes, such as HUC-MSCs-derived exosomes, can be a promising therapeutic for NAFLD.

4.3.1.1 HUC-MSC exosomes

Many reports have illustrated that human umbilical cord mesenchymal stem cells (HUC-MSCs) exert therapeutic effects on liver diseases, including chemical-induced liver injury, decompensated liver cirrhosis, liver failure, and autoimmune liver diseases, including primary biliary cholangitis (169–174).

4.3.1.1.1 Lipid and glucose metabolism

HUC-MSCs improve NAFLD and metabolic syndrome by means of regulating lipid metabolism by promoting the expression of genes that are related to fatty acid oxidation and suppressing adipogenesis-related genes' expression in db/db mice (175). MSCs exert their therapeutic effects through secretory factors, including exosomes. Cheng et al. (176) showed that the treatment of palmitic acid (PA)-treated human normal liver cell line (L-O2 cells), human fetal hepatocyte line, with HUC-MSCs-exosomes improves cell viability and inhibits apoptosis. They showed that the level of *miR-627-5p* was higher in HUC-MSCs-exosomes compared with HUC-MSCs. Moreover, they found that the expression of G6Pc and PEPCK, gluconeogenesis-related proteins, along with FAS and SREBP-1c, lipid metabolism-related proteins, were repressed in these cells, while the expression of PPAR α , another lipid metabolism-related protein, was notably downregulated. Additionally, the study exhibited that the plasma levels of ALT, AST, total cholesterol (TC), triglyceride (TG), and blood glucose in the NAFLD rat model were repressed by exosome treatment. They also reported that *MiR-627-5p* improves lipid and glucose metabolism, key pathological components of NAFLD, in L-O2 cells by targeting fat mass and obesity-related gene (*FTO*). The *FTO* gene facilitates NAFLD development via increasing insulin resistance, oxidative stress, and lipid accumulation in liver cells (177, 178). Furthermore, exosome treatment has also been found to alleviate insulin resistance and liver damage and reduce fat accumulation in NAFLD rat models (176). Another study (179) also confirmed that HUC-MSC-exosome therapy can attenuate liver steatosis and regulate abnormal expression of *Fabp5*, *ACOX*, *PPAR- α* , *FAS*, *SREBP-1c*, and *CPT1 α* as lipid metabolism-related genes.

4.3.1.1.2 Oxidative stress

HUC-MSC-Exosomes have also been shown to have beneficial effects against oxidative stress and inflammation. Kang et al. (179) demonstrated that in rat models of NASH, HUC-MSCs exosomes enhanced the Nrf2 (Nuclear factor erythroid 2-related factor 2), a protective factor against oxidative stress and 1 [NAD (P) H quinone dehydrogenase 1], a part of cellular adaptive response to stress, which seems to play a significant part in treating NASH (180). Besides, HUC-MSC-derived exosomes reduce oxidative stress through lowering Malondialdehyde (MDA), CYP2E1, and reactive oxygen species (ROS) levels and elevating Superoxide dismutase (SOD) and GSH function as well (179). Furthermore, HUC-MSC-exosome therapy reduces inflammatory response through decreasing F4/80⁺ and CD11c⁺ macrophages and the levels of tumor necrosis factor-alpha (TNF- α) and Interleukins-6 (IL-6) (179).

4.3.1.2 BM-MSC exosomes

MSCs are classically separated from bone marrow (181). Bone marrow mesenchymal stem cells (BM-MSCs), also known as BM mesenchymal stromal cells, are multipotent mesenchymal precursor cells that have many favorable characteristics for regenerative therapy, including anti-inflammatory and immunomodulatory properties (182, 183). Studies have revealed that BM-MSCs have therapeutic potential in different diseases, including cardiovascular, lung, neural, hematopoietic, and

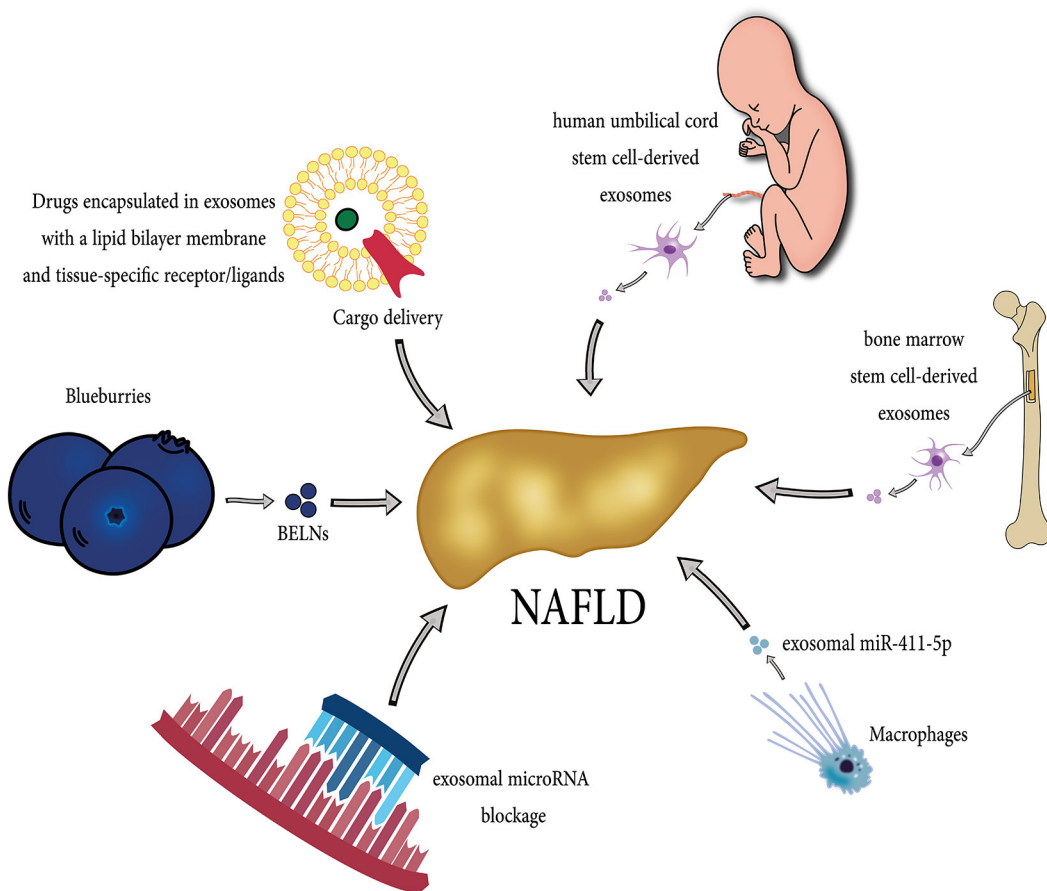


FIGURE 3
 Diverse potential exosome-based therapeutic strategies in NAFLD. Exosomes originating from different sources, including stem cells, bone marrow, macrophages, and Blueberries, can exert beneficial effects in the treatment of NAFLD. Each source contributes unique therapeutic properties, such as anti-inflammatory and regenerative capabilities, immune modulation, and fibrosis inhibition, making them a promising tool in NAFLD management. They can be administered safely in high doses due to their cell-free nature, exhibit low toxicity, remain stable during circulation, and have minimal immunogenicity. Moreover, researchers are actively exploring exosome-based therapies for NAFLD, including drug delivery (serving exosomes as drug carriers, delivering therapeutic cargo directly to liver cells), targeted approaches (blocking exosomal contents such as microRNAs), and modulation of inflammatory processes.

liver diseases, along with graft-vs.-host disease and cutaneous, tendon ligament, and musculoskeletal tissue repairing (184–186). BM-MSCs release paracrine factors which influence the surrounding microenvironment (187). In contrast with their favorable therapeutic characteristics, a few downsides can restrain their clinical utilization, primarily due to their potential for tumorigenicity and immunogenicity (188). Thus, using BM-MSCs paracrine factors maintains BM-MSCs properties without most disadvantages. Therefore, exosomes can potentially replace BM-MSCs as a safer therapeutic approach for tissue repair (162, 189).

BM-MSCs and BM-MSC-derived exosomes both can provide anti-steatotic effects through downregulating sterol regulatory element binding protein 1 (*SREB-1*), sterol regulatory element binding protein 2 (*SREB-2*) and cetyl coenzyme A carboxylase (*ACC*), suppressing lipid uptake and upregulating peroxisome proliferator-activated receptor alpha (*PPAR-α*) and carnitine palmitoyltransferase 1 (*CPT1*) fatty acid oxidation genes. BM-MSC-derived exosomes at 15 μg/kg, 30 μg/kg, and 120 μg/kg

have anti-steatotic effects in the HFD-induced NASH model. BM-MSCs or BM-MSCs-exosome co-treatment caused anti-apoptotic impacts via a meaningful reduction in *Bax/Bcl2* ratio and a rise in the expression of mitochondrial mitophagy genes, including Parkin RBR E3 Ubiquitin Protein Ligase (*Parkin*), phosphatase and tensin homolog (PTEN)-induced putative kinase 1 (*PINK1*), unc-51 like autophagy activating kinase 1 (*ULK1*), B-cell lymphoma 2/adenovirus E1B 19 kDa protein-interacting protein 3 (*BNIP3L*), autophagy related gene (*ATG5*), *ATG7* and *ATG12*. Furthermore, a notable depletion in the levels of AST and ALT has been observed in BM-MSCs or BM-MSCs-exosome co-treatment (190).

El-Derany et al. (190) found that BM-MSC-derived exosome can be an ideal treatment option for NAFLD through a mechanism in which the upregulation of *miRNA-96-5p* leads to the inhibition of *caspase-2*. *MiRNA-96-5p* is found both in peripheral blood and bone marrow (191). Studies have demonstrated that HFD is associated with a downregulation of *miRNA-96-5p* in NAFLD models (190, 192, 193). Moreover, it has been proven that the suppression of *caspase-2* lowers lipo-apoptosis and inhibits

fibrogenic Hedgehog ligands production, leading to the transition of NAFL to Nash (190, 194, 195). Furthermore, evidence has revealed that the inhibition of *caspase-2* prevents hyperlipidemia and reduces hepatic steatosis, liver apoptosis, and mitochondrial mitophagy (190, 194).

4.3.1.3 Macrophage-derived exosomes

Severe stages of NAFLD are related to hepatic stellate cell (HCS) activation and their interactions with macrophages. Hepatic macrophages have critical roles in the initiation and progression of fibrosis (93, 196, 197). It has been found that the level of *miRNA-411-5p* is lower in NASH patients' plasma exosomes and liver samples of NAFLD patients (94). Moreover, M2 macrophage-derived exosomes contain a higher amount of *miRNA-411-5p* than M1 macrophages. Wan et al. (94) revealed that M2-derived exosomes suppress HCS activation by inhibiting the direct target of *miRNA-411-5p*, Calmodulin-Regulated Spectrin-Associated Protein 1 (*CAMSAP1*), during the transition of NASH to NAFL.

In macrophages and neutrophils, *miR-223* is the amplest miRNA that is functionally active and can be transferred from myeloid cells to other cells by exosomes (198–200). The serum level of *miR-223* was reported to be remarkably higher in NAFLD. Moreover, a positive correlation has been seen between the levels of serum *miR-223* and IL-6 (107). IL-6 signaling exerts anti-fibrotic effects in NAFLD-associated liver fibrosis by inducing myeloid cells to enhance *miR-223*-enriched exosome release. IL-6 increases the biogenesis of exosomes by upregulating gene expression without changing the *pre-miRNA-223* expression (107). Myeloid cell-derived *miRNA-223* enriched exosomes are delivered to the liver and inhibit the expression of *miR-223* target pro-fibrotic genes like *Igf1r*, *Cxcl10*, *Taz*, and *Nlrp3*. Subsequently, they help control Nash's progression. Therefore, macrophage-derived exosomes could be used as a therapeutic option that alters the progression of NASH to NAFL (107).

4.3.2 Exosome blocking

Exosomes are the communication bridge between cells, thus playing critical roles in the pathological development and progression of numerous diseases (201). Accordingly, changing the level of exosome secretion, inhibiting their activity, or changing their contents could be a potential approach for treating diseases.

4.3.2.1 Exosomal mirna blocking

Targeting exosomal miRNAs may also be an effective treatment approach in NAFLD. miRNAs are reported to have therapeutic roles in NAFLD by regulating lipid metabolism, inflammation, and fibrosis (90). As mentioned in the pathophysiology part, exosomal *miR-192-5p* is higher in exosomes derived from NASH hepatocytes and plays a role in NAFLD progression by triggering M1 macrophage polarization (90). It has been found that this mechanism can be reversed using a *miR-192-5p* inhibitor (91). Hence, targeting serum exosomal *miR-192-5p* can be a beneficial option for inhibiting the progression of NAFLD (91).

Moreover, *MiR-122-5p* is another prospective option in exosome blocking. Zhao et al. (92) demonstrated that *miR-122-5p* level was higher in exosomes originating from Huh7 cells,

a hepatic cell line loaded with cholesterol (including ox-LDL and M β CD-cholesterol). They used *anti-miR-122-5p* to block this exosomal miRNA. *Anti-miR-122-5p*-treated Huh7 cells-derived exosomes exerted lower effects in inducing TNF- α , IL-6, and IL-1 β expression. Additionally, exosome-mediated inflammation of macrophages was blocked.

4.3.2.2 Blocking liver-adipose tissue crosstalk

Adipose tissue exosomes may also be favorable therapeutic targets for NASH. Several studies have supported the role of pro-inflammatory mediators released from visceral adipose tissue (VAT) in the progression of NAFLD (202, 203). A cross-talk between adipose tissue and the liver is significant in NAFLD development and progression. This connection seems to be facilitated through exosomes (204). Stressed AT induces abnormal lipid accumulation in the liver, which may be the major contributor to NASH initiation and progression.

Studies have indicated that NASH development can be a consequence of Endoplasmic Reticulum (ER)-stress, which possibly changes miRNAs and metabolites of secretory exosomes (77, 205). Exosomes derived from ER-stress induced-AT can induce and aggravate NASH by delivering exosomal Aldo-keto reductase family 1 B7 (AKr1b7) (111). They can be taken up by hepatocytes and cause hepatic steatosis, inflammation, and fibrosis. Akr1b7 deficiency protects murine liver from NASH in HFD and methionine-choline-deficient diet (MCD)-fed mice. In addition, suppressing hepatic AKr1b7 reduces TG level and glycerol concentration and alleviates hepatic inflammation, ER stress, and lipid synthesis. Accordingly, Epalrestat, an Akr1b7 inhibitor, can suppress the effects of AT exosomes on ER stress and hepatic inflammation (68). Moreover, Treatment with GW4869, an exosome production inhibitor (by inhibiting sphingomyelinase), has been shown to alleviate ER stress and reduce lipid synthesis and hepatic inflammation (111).

4.3.2.3 TGF-B signaling blockage

TGF-B signaling plays a major part in various biological processes, such as cellular proliferation, differentiation, migration, and cell death. It is also an essential pathway in the regulation of liver homeostasis. TGF-B signaling helps NAFLD progression from initial lipid accumulation to fibrosis (206). Steatosis, weight gain, and impaired insulin sensitivity have been found to be associated with TGF-B signaling in a mouse model of NASH (207). Moreover, TGF-B signaling can modulate microRNA biogenesis at transcriptional and post-transcriptional levels (208). TGF-B induces hepatic secretion of exosomes carrying *let-7b-5p*, a miRNA. TGF- β -*let-7b-5p* pathway contributes to HFD-induced steatosis and obesity through lowering oxidative phosphorylation in mitochondria and inhibiting white adipose tissue (WAT) to brown fat conversion (113). Accordingly, *Tgfb2* loss in hepatocytes ameliorates HFD-induced NAFLD by improving mitochondrial biogenesis. It alleviates lipid accumulation and induces "browning" of WAT via inhibiting miRNA *let-7b-5p*-containing exosomes release from hepatocytes. In addition, *let-7b-5p* inhibitor can downregulate the expression of *Cd36*, *Fatp1*, and *Fabp1* (which are fatty acid transporter genes), upregulate mitochondrial genes' expression, like *Cox5b* and *Atp5a*, and reduce lipid accumulation

in CL-316,243 (CL)/cold-exposed mice and HFD-induced obese mice (113).

4.3.2.4 Drugs blocking exosomes

Some drugs are also effective in exosome blockage and provide benefits through exosome modulation. GW4869, a non-competitive neutral sphingomyelinase inhibitor, suppresses exosome secretion. It has been shown to mitigate HFD-induced NAFLD in mice models (112). Moreover, McCommis et al. (209) evaluated the efficacy of next-generation thiazolidinediones (MSDC-0602) in a mouse model of NASH. They have weak binding to peroxisome proliferator-activated receptor γ (PPAR γ) and still directly suppress and interact with mitochondrial pyruvate carrier (MPC). The effect of MSDC-0602 was through MPC2. They revealed that MSDC-0602 treatment, directly and indirectly, prevents and reverses stellate cell activation, liver fibrosis, and lipid accumulation in HTF-C mice. It is through the indirect modulation of exosomes derived from hepatocytes in an MPC-dependent manner. Furthermore, Yan et al. (112) found that metformin treatment inhibits HFD-induced NAFLD in mice, thorough activating AMP-activated protein kinase (AMPK α 1) in WAT, which leads to the inhibition of exosomes shedding into serum and WAT.

Ezetimibe is a lipid-lowering drug that selectively inhibits the absorption of cholesterol through the blockage of NBC1LI-dependent cholesterol transport in the small intestine. NBC1LI is also present in the liver (210–212). Many studies have demonstrated that ezetimibe treatment can significantly improve hepatic steatosis, ballooning score, and NAFLD histological features, even after its fibrotic changes (213, 214). It has been reported that ezetimibe treatment activates autophagy in human hepatocyte cells (215). In addition, Kim et al. (216) observed other beneficial effects of ezetimibe in NAFLD. They reported changes in Hepatocyte-macrophage interaction following ezetimibe treatment. They observed decreased Interleukin 1 Beta (IL-1B) mRNA and protein levels in macrophages cultured with EVs released from ezetimibe/PA-cotreated hepatocytes.

4.3.3 Cargo delivery

Recently, a variety of different nano-based drug carriers have been used in order to improve the therapeutic efficacy of chemical and biomolecular drugs and ingredients (217). Exosomes have favorable characteristics for being used as a drug delivery agent, including small size, which facilitates its penetration into deep tissues (218), having a slightly negative zeta potential (219), biocompatibility, low immunogenicity, being able to cross the biological barriers such as the BBB (220) and escape immune clearance (221), low systemic toxicity, and long-term existence in the target tissue (221, 222). Moreover, compared to many synthetic drug delivery agents, using exosomes facilitates cellular uptake due to their specific surface proteins (218, 223).

The exosomal-based delivery system has been loaded with a variety of biomolecules for drug delivery like paclitaxel, doxorubicin, curcumin as a peptide or protein-based therapeutics containing STAT3 inhibitors, catalase, and also genetic material including siRNA (222), an effective carrier of miRNA that also stabilizes the miRNA in exosome (224, 225).

4.3.4 miRNA-based therapeutics

MiRNAs are non-coding RNAs that take part in the regulation of gene expression. Recently, some studies have revealed that miRNA-based therapies are feasible and effective in various diseases, especially cancers (226–228). For instance, *miR-16*-loaded minicell is in clinical investigation for the treatment of malignant pleural mesothelioma (229). MiRNAs may be potential treatment options for NAFLD as well. In the early stages of NAFLD, decreased intrahepatic miRNA levels are compensated by the elevated production of miRNA in adipose tissue. As NAFLD progresses, the external supply of liver-supporting miRNA falls gradually, which leads to the deterioration of liver function and an increase in hepatic carcinogenesis (110). Therefore, supplying damaged liver with external miRNA may be efficient in hindering the progression of NAFLD. Moreover, Baranova et al. (110) revealed that purified *miR-122* exosomes may be potential cell-free therapeutics for preventing HCC in NAFLD. Further research on the efficiency of miRNA therapy in NAFLD is needed.

Blocking pathogenic exosomal miRNAs is also a potential treatment approach for NAFLD. The level of *miR-199a-5P* is found to be considerably higher in NAFLD. Exosomal *miR-199a-5P* (Exo-miR-199a-5P) interferes with the metabolism of lipids in the liver by suppressing the expression of hepatic MST1 (79). Additionally, administration of Exo-miR-199a-5P induces hepatic lipid deposition in normo-caloric diet (NCD) and HFD mice (which is possibly via regulating AMPK and SREBP-1c signaling pathways) (79). It also elevates liver weights and plasma TG levels (79). Expectedly, exosomal *anti-miR-199* can significantly reduce lipid accumulation *in vivo* and *in vitro*. It also downregulates lipogenic genes and lowers serum and hepatic TG and cholesterol in HFD mice (79). Thus, treatment with exosomal *anti-miR-199a* can offer fruitful effects in the prevention and treatment of liver steatosis and dyslipidemia.

4.3.5 Exosomes-like nanoparticles

Plant-derived exosome-like nanoparticles (PDNPs) have shown potential in drug delivery and as therapeutic applications in different disease (230, 231). PDNPs are nano-sized vesicles released from edible plants, including lemon, apple, carrot, grapefruit, coconut, broccoli, grape, and ginger (232–241). Increasing evidence indicates that similar to mammalian cell-secreted exosomes, PDNPs have an important in intercellular communication (242, 243). In addition, they have certain superiorities compared to mammalian cell-secreted exosomes or artificial nanoparticles; they do not cause any detectable Immunogenicity or toxicity, they have high potential in delivering biomolecules, and using them is highly economical (95, 107, 110). They have been used in treating alcoholic fatty liver as well as IBD and colon cancer (234, 237, 244–246).

Blueberry nanoparticles have shown promise in the treatment of NAFLD. Blueberry is one of the most consumed berries that contains a wide diversity of bioactive compounds, such as flavonoids, anthocyanins, and polyphenols. Blueberry has antioxidative, anti-inflammatory, and anti-tumor activity and can have protective effects against cancers as well as Alzheimer's cardiovascular diseases, and depression (247–251). Zhao et al.

(252) investigated the anti-oxidative impacts of Blueberry-derived exosomes-like nanoparticles (BELNs) in hepatocytes. They demonstrated that BELNs could attenuate oxidative stress by modulating nuclear factor erythroid 2-related factor 2 (Nrf2) distribution, which results in the upregulation of the antioxidative proteins and enzymes expression such as SOD, NADPH/quinone oxidoreductase I (NQO1), Heme Oxygenase-1 (HO-1), glutathione peroxidase (GPx), and catalase (CAT). It also regulates Bcl-2-associated X protein (BAX), Bcl-2, and HO-1 apoptosis-related proteins' expression in the liver of HFD-fed mice and rotenone-induced HepG2 cells. BELN supplementation also improves liver dysfunction and insulin resistance in NAFLD, which is accompanied by a reduction in AST and ALT levels and fat deposition as well as a downregulation in acetyl-CoA carboxylase 1 (ACC1) and fatty acid synthase (FAS) expression (252).

4.3.6 Exosome-melatonin cotreatment

Melatonin can potentiate the therapeutic effects of exosomes; thus, it may be a favorable agent to be administered alongside exosomes. Melatonin is a methoxyindole that is secreted by the pineal gland and gastrointestinal tract during night and daytime, respectively (253). Melatonin has a critical role in the amelioration of different metabolic disorders in various tissues, including adipose and hepatic tissues (254–256). Melatonin exerts its beneficial effects in NAFLD exosomal therapy by activating Bmal1 expression in adipocytes (257). Eventually, Exosome-Melatonin co-treatment alleviates ER-stress-induced hepatic steatosis through a significant downregulation in the expression of adipocyte-derived exosomal resistin (257). Resistin is an adipocytokine that may aggravate hepatic steatosis via stimulating hepatic ER stress. Furthermore, exosome-melatonin supplementation can lower hepatic inflammation, fibrosis, and cell apoptosis (257).

4.4 Challenges and opportunities

The growing prevalence of NAFLD and its global health burden have provoked considerable research on possible diagnostic and therapeutic options for NAFLD. NAFLD is a multifactorial disease which includes a broad spectrum of liver damage. Although various pathophysiological mechanisms and genetic factors have been proven to be related to NAFLD, its treatment remains challenging. In recent years, exosomes have attracted widespread attention for their role in metabolic dysfunctions and their efficacy as pathological biomarkers. Exosomes have also shown tremendous potential in treating various disorders, such as wound healing and neurological and cardiovascular dysfunctions. With increasing evidence supporting the significant role of exosomes in NAFLD pathogenesis, their theragnostic potential has become a point of interest in NAFLD. Considering the low cost and high feasibility of EV detection and its non-invasiveness, using exosomes as a diagnostic biomarker can be a revolution in NAFLD diagnosis. Exosome-based therapies can also bring favorable results in preventing and treating NAFLD. However, there are still certain

limitations that need to be overcome, and many questions need to be answered before exosomes can be utilized as a routine treatment for liver diseases.

Contrary to the satisfactory results of experimental exosome therapy studies, exosomal therapy methods and safety are not entirely clarified (258). Exosome-based therapy methods face both pharmaceutical (production, isolation, and drug loading) and pharmacokinetic (biodistribution and cellular uptake) challenges in becoming clinical. One of the main limitations of exosome therapy is the preparation and purification of a large volume of desired exosomes. Moreover, designing standard methods to isolate and identify the types and sources of exosomes also needs to be investigated more since the purity and physicochemical properties of exosomes are strongly affected by their isolation method, and no globally accepted standard is currently available in this era. Besides, considering exosomes' degradation during isolation and freeze-thaw processes, standardization of isolation and storage conditions is also crucial (148, 259–261). Another emerging challenge in exosomal therapy is that natural exosomes, also known as cell-derived exosomes, contain a multitude of biomolecules with unknown effects. Cells in different environmental conditions release exosomes with diverse cargo, complicating the predictability of cell-derived exosome therapies (262). It also makes finding the effective agent challenging. Therefore, a comprehensive review of the content of exosomes and their effect on pathologic and healthy tissues is also necessary. Additionally, although exosomes are capable of cell targeting, concerns about their intrahepatic and intrasplenic accumulation remain to be solved, as well as their relatively short half-life. These problems may be manageable if different tissue targeting strategies are used in conjunction (168). Finally, suppressing exosome secretion or function as a therapeutic strategy is far from utilization, as it requires a further in-depth understanding of the exosome's role in normal physiology (259). Accordingly, there are still serious challenges in using exosome-based therapies as a routine clinical option that should be overcome.

5 Conclusion

Recent findings on exosomes have provided promising prospects of using them as feasible, costly, and reliable diagnostic options in NAFLD. In addition, exosomal-based therapies, having low immunogenicity and high biocompatibility, for drug delivery as an effective treatment option or by inhibiting their actions as a treatment strategy have been associated with favorable results in NAFLD. However, despite several pre-clinical studies on exosomes in liver disorders, the passage to the clinical setting remains unsure. There are still certain challenges to using exosomal-based therapies in NAFLD. Besides, the feasibility of exosome-related treatment strategies in NAFLD will await further evaluation due to the critical role of exosomes in numerous physiological conditions. Due to the great potential of this novel theragnostic agent in NAFLD, further investigations on their safety, clinical efficacy, and application standardization are highly recommended.

Author contributions

AT: Conceptualization, Investigation, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. MJ: Investigation, Writing – original draft. NS-P: Visualization, Writing – review & editing. AM: Investigation, Writing – original draft.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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