Check for updates

OPEN ACCESS

EDITED BY Savino Spadaro, University of Ferrara, Italy

REVIEWED BY Keita Ikeda, University of Virginia, United States Ali Golestani, University of Calgary, Canada

*CORRESPONDENCE Gustav Magnusson ⊠ gustav.magnusson@liu.se

RECEIVED 11 December 2023 ACCEPTED 16 April 2024 PUBLISHED 22 May 2024

CITATION

Magnusson G, Engström M, Georgiopoulos C, Cedersund G, Tobieson L and Tisell A (2024) High inspired CO₂ target accuracy in mechanical ventilation and spontaneous breathing using the Additional CO₂ method. *Front. Med.* 11:1352012. doi: 10.3389/fmed.2024.1352012

COPYRIGHT

© 2024 Magnusson, Engström, Georgiopoulos, Cedersund, Tobieson and Tisell. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

High inspired CO₂ target accuracy in mechanical ventilation and spontaneous breathing using the Additional CO₂ method

Gustav Magnusson^{1,2}*, Maria Engström^{1,2}, Charalampos Georgiopoulos^{2,3}, Gunnar Cedersund⁴, Lovisa Tobieson⁵ and Anders Tisell^{1,2}

¹Department of Health, Medicine and Caring Sciences, Linköping University, Linköping, Sweden, ²Center for Medical Image Science and Visualization (CMIV), Linköping University, Linköping, Sweden, ³Diagnostic Radiology, Department of Clinical Sciences, Medical Faculty, Lund University, Lund, Sweden, ⁴Department of Biomedical Engineering, Linköping University, Linköping, Sweden, ⁵Department of Neurosurgery in Linköping, and Department of Biomedical and Clinical Sciences, Linköping University, Linköping, Sweden

Introduction: Cerebrovascular reactivity imaging (CVR) is a diagnostic method for assessment of alterations in cerebral blood flow in response to a controlled vascular stimulus. The principal utility is the capacity to evaluate the cerebrovascular reserve, thereby elucidating autoregulatory functioning. In CVR, CO₂ gas challenge is the most prevalent method, which elicits a vascular response by alterations in inspired CO₂ concentrations. While several systems have been proposed in the literature, only a limited number have been devised to operate in tandem with mechanical ventilation, thus constraining the majority CVR investigations to spontaneously breathing individuals.

Methods: We have developed a new method, denoted Additional CO₂, designed to enable CO₂ challenge in ventilators. The central idea is the introduction of an additional flow of highly concentrated CO₂ into the respiratory circuit, as opposed to administration of the entire gas mixture from a reservoir. By monitoring the main respiratory gas flow emanating from the ventilator, the CO₂ concentration in the inspired gas can be manipulated by adjusting the proportion of additional CO₂. We evaluated the efficacy of this approach in (1) a ventilator coupled with a test lung and (2) in spontaneously breathing healthy subjects. The method was evaluated by assessment of the precision in attaining target inspired CO₂ levels and examination of its performance within a magnetic resonance imaging environment.

Results and discussion: Our investigations revealed that the Additional CO_2 method consistently achieved a high degree of accuracy in reaching target inspired CO_2 levels in both mechanical ventilation and spontaneous breathing. We anticipate that these findings will lay the groundwork for a broader implementation of CVR assessments in mechanically ventilated patients.

KEYWORDS

cerebrovascular reactivity, CO_2 gas challenge, ventilation, magnetic resonance imaging, carbon dioxide, vascular stimulus

1 Introduction

Cerebrovascular reactivity imaging (CVR) represents an innovative approach for the non-invasive exploration of cerebral hemodynamics. It involves the application of a vasoactive stimulus and simultaneous measurements of alterations in cerebral blood flow. The reactivity, quantified as the change in blood flow divided by the applied stimulus, serves as an indirect indicator of the local vasoregulatory reserve within the cerebral vasculature. Furthermore, this method enables the computation of the time delay in the blood flow response. Research has extensively examined the application of the CVR technique across various medical conditions, including arterial stenosis, moyamoya disease, brain tumors, dementia, small vessel disease, and subarachnoid hemorrhage (1). Despite the promising clinical potential of CVR in these diverse patient cohorts, it has not yet achieved widespread clinical adoption and remains predominantly a research tool. One key impediment to its broader utilization is the limited availability of commercial products for stimulus generation that can be applied across different clinical scenarios.

The established vascular stimulus in CVR measurement is a controlled alteration of arterial carbon dioxide partial pressure (P^a_{CO2}). The associated changes in blood flow are typically monitored using magnetic resonance imaging (MRI) in conjunction with the blood oxygenation level dependent (BOLD) signal (2). Various methods can be employed to manipulate $P^{a}_{CO_2}$, such as controlled breathing patterns, including deep breathing and breath-holding, or the administration of vasoactive drugs like Acetazolamide (3). However, the preferred approach, due to its reliability and reproducibility, is the administration of carbon dioxide within the inspired gas (4). Several methods described in the literature use reservoirs with a variable mixture of CO₂, O₂, and N₂, to target fractional CO₂ concentrations in the inspired gas $(F_{CO_2}^1)$ (5, 6). More sophisticated methods incorporate advanced controls, such as dynamic end-tidal forcing or prospective endtidal targeting, which enable precise targeting of subjects' end-tidal CO_2/O_2 levels, reflecting the gas concentrations in the alveoli (7, 8).

While the literature contains substantial information on methods for CO2 gas challenge in spontaneously breathing patients, there has been limited exploration in mechanically ventilated patients (9, 10). This gap in research may explain why CVR studies in mechanically ventilated patients have primarily focused on breathing pattern alterations or administration of vasoactive drugs (11-14). The aim of this study is to implement a method capable of administering CO₂ to both ventilated and non-ventilated patients. In contrast to other CO2 administration methods, our method does not generate the entire gas mixture but, instead, supplements the inspired gas with additional CO2 in proportion to the respiratory gas flow, as illustrated in Figure 1, drawing inspiration from nitric oxide systems (15). This approach does therefore not rely on the addition of external reservoirs or multiple gas sources, keeping the necessary modification of the breathing circuit minimal, which is desirable in an intensive care setting. We conducted tests of our method within mechanical ventilation, using a test lung, and in spontaneously breathing healthy subjects, comparing it to a conventional CO₂ gas challenge method, inspired by the approach detailed by Tancredi et al. (5).

2 Materials and methods

In this section, we outline the material and equipment employed in evaluating the Additional CO_2 method, directing readers to the Supplementary material for a comprehensive description of specific components used.

2.1 Rationale

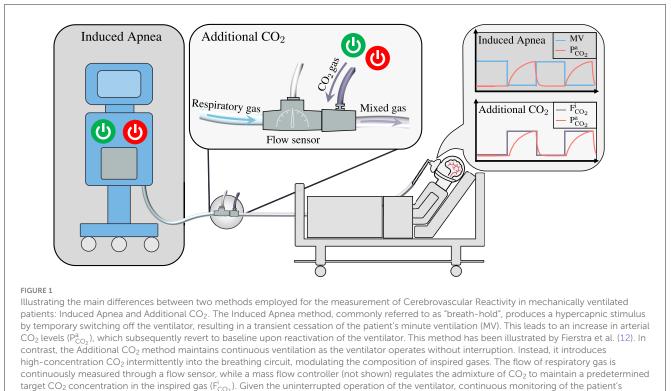
While conducting CVR experiments, the manipulation of gases arterial partial pressures is common practice, yet direct measurement of these pressures is infrequently performed. Instead, alveolar pressures, derived from end-tidal values, are typically used due to their strong correlation with arterial pressures in lung-healthy individuals (16). In the alveolar, we typically talk about gases fractional concentrations (F_{gas}^A) and partial pressures (P_{gas}^A) interchangeably since there is a direct translation between the two: $P_{gas}^A = F_{gas}^A \times (P_{atm} - P_{H_2O})$, where P_{atm} is the atmospheric pressure and P_{H_2O} is the partial pressure of water vapor at body temperature. The steady-state fractional alveolar concentrations of O₂ and CO₂ (F_{O_2/CO_2}^A) are determined by alveolar ventilation (\dot{V}_A), O₂ consumption/CO₂ production (\dot{V}_{O_2/CO_2}), and inspired gas concentrations (F_{O_2/CO_2}^i), as described by Equations (1, 2) (8).

$$F_{O_2}^{A} = F_{O_2}^{i} - \frac{V_{O_2}}{V_A}$$
(1)

$$F_{CO_2}^{A} = F_{CO_2}^{i} + \frac{\dot{V}_{CO_2}}{\dot{V}_{A}}$$
(2)

Equation (2) elucidates potential manipulation of $F^A_{CO_2}$, with controlled breathing patterns influencing alveolar ventilation \dot{V}_A , administration of vasoactive drugs altering \dot{V}_{CO_2} , or direct modification inspired CO₂. Among these techniques, direct manipulation of inspired CO₂ offers increased repeatability, as it targets the corresponding parameter in Equation (2) directly.

A common and straightforward method to modify inspired CO2 is to adjust the gas content in a reservoir from which the subject breathes. However, in mechanical ventilation, this presents challenges due to the presence of internal one-way valves in the ventilator, ensuring gas flow in only one direction. One workaround is illustrated by Winter et al. (9), involving the placement of two external reservoirs in a sealed compartment. Gas control occurs in one reservoir from which the subject breathes, while the ventilator ventilates the other. However, this approach necessitates a complex external breathing circuit and requires patient separation from the ventilator. Another option is manual ventilation, where gas control in a self-inflated bag occurs simultaneously with manual compression. Nonetheless, this method does not equate to mechanical ventilation (10). These limitations of reservoir-based alterations in mechanical ventilation may explain why most CVR studies on ventilated patients primarily focus on either breathing pattern alterations or administration of vasoactive drugs (11–14). Ideally, a simple method enabling $F_{CO_2}^i$ alterations while requiring minimal modification of the breathing circuit and no separation of the ventilator and patient would be preferable.



 O_2 and CO_2 levels ensues, offering an added layer of safety and control.

The proposed Additional CO₂ method addresses this need. By measuring respiratory gas flow and adding a proportional flow of CO₂, it does not rely on a reservoir to alter the gas content in inspired gas. The only modification to the breathing circuit is the incorporation of a flow sensor, gas inlet, and potentially a humidifier (if not already included) to facilitate mixing of the additional CO₂ and respiratory gas. Moreover, since the method relies on the measurement of respiratory gas flow, it has the potential to be further developed to enable direct targeting of $F_{CO_2}^A$ by adjusting the administered $F_{CO_2}^i$ on a breath-to-breath basis according to changes in \dot{V}_A (see Equation 2). Additionally, a source of oxygen could also be incorporated to allow simultaneous targeting of $F_{O_2}^A$.

2.2 Additional CO₂ system

A prototype system, Additional CO_2 System, was devised to assess the Additional CO_2 method, consisting of four primary components: gas source (100 % CO_2 , 5 L canister, AirLiquide), a control unit (including a microcontroller, Arduino Beetle, DFRobotic), gas control (flow sensor, SFM3200, Sensirion and mass flow controller, SFC5500, Sensirion) and graphical user interface (GUI, Python program running on a laptop, in-house developed). The flow sensor was read by the control unit, which also managed the mass flow controller. The proportional relationship between the setpoint of the mass flow controller and the flow of the flow sensor was computed at the GUI and transmitted to the control unit. The underlying calculation involved the solving of a mass balance equation for a target $F_{CO_2}^i$:

$$\begin{split} F^{i}_{CO_{2}} &= \frac{F^{res}_{CO_{2}} \times \dot{M}_{res} + F^{add}_{CO_{2}} \times \dot{M}_{add}}{\dot{M}_{res} + \dot{M}_{add}} \\ \Rightarrow \frac{\dot{M}_{add}}{\dot{M}_{res}} &= \frac{F^{i}_{CO_{2}} - F^{res}_{CO_{2}}}{F^{add}_{CO_{2}} - F^{i}_{CO_{2}}} \end{split}$$
(3)

where $\dot{M}_{res/add}$ and $F_{CO_2}^{res/add}$ are the mass flow and fractional CO₂ concentration of the respiratory (res) and additional (add) gas. A consequence of introducing additional CO₂ in this manner is the concurrent reduction of O₂ concentration in the inspired gas. The GUI also displayed this change in $F_{O_2}^i$ to the user. Moreover, to ensure safety, strict limits were imposed on the maximal and minimal $F_{CO_2}^i$ and $F_{O_2}^i$ concentrations, set at 5 and 19 %, respectively.

To specify the target $F_{CO_2}^i$ concentration, a user loaded a JSON protocol file containing target values and corresponding time durations via the GUI. For a more comprehensive description of the constructed system and the components employed, refer to Section 1.1 in the Supplementary material.

2.3 Reservoir CO₂ system

A reference system, modeled after the design by Tancredi et al. (5), was assembled to facilitate a comparative analysis with our Additional CO₂ System in spontaneous breathing. This system, from here on referred to as Reservoir CO₂ System, was established using three mass flow controllers (SLA5850, Brooks Instrument)

connected to sources of O₂, CO₂, and N₂. By altering the setpoints of each controller, a specific gas mixture was created and stored in a reservoir, from which a subject would draw breath. The same GUI mentioned earlier was employed to oversee the mass flow controllers. Users could specify target $F_{O_2}^i$ and total flow rate, in addition to $F_{CO_2}^i$ concentrations, using a protocol file similar to the one used for the Additional CO₂ System. The same constraints on maximal and minimal $F_{CO_2}^i$ and $F_{O_2}^i$ concentrations, as described above, remained in effect.

Note that this system was exclusively used during in spontaneous breathing and not during in mechanical ventilation (see Section 3.1 below). The reason being that the one-way valves within a mechanical ventilator only allow gas in the inspiration part of the circuit to flow toward the patient. Without more sophisticated modification to the breathing circuit, as described by Winter et al. (9) or Venkatraghavan et al. (10), control over the gas in a breathing reservoir is not achievable. For further details regarding the system and its components, please refer to Section 1.2 in the Supplementary material.

2.4 Ventilator and test lung

To evaluate the Additional CO_2 System in conjunction with mechanical ventilation, an anesthesia workstation (Primus Infinity Empowered, Dräger Medical) was used together with a test lung (AccuLung, Fluke Biomedical). The workstation was also used for sampling inspired and expired O_2 and CO_2 .

2.5 Breathing circuits

Two distinct breathing circuits were employed: one for mechanical ventilation of a test lung (Ventilator Circuit) and another for spontaneously breathing healthy subjects (Subject Circuit), as illustrated in Figure 2. In the Ventilator Circuit, which was only used together with the Additional CO_2 System, the flow sensor was connected to the ventilator's outlet, followed by a connector with a gas inlet to which the mass flow controller's outlet was attached. An empty humidifier was positioned immediately after the connector to serve as a small volume, ensuring a uniform gas mixture. A coaxial ventilator tube was affixed to the outlet of the humidifier, and an elbow connector with a sampling port connected the tube to the test lung.

In the Subject Circuit, the lower part of Figure 2 depicts the configurations used for the Additional CO_2 and Reservoir CO_2 Systems. The sole differences between the Additional CO_2 and Reservoir CO_2 configurations were the inclusion of the flow sensor (used solely in the Additional CO_2 System) and the length of the expandable tube. In the Reservoir CO_2 System, the expandable tube functioned as the gas reservoir, as elucidated by Tancredi et al. (5), and was extended to a length of 2 m, creating a reservoir with a size of about 700 ml (tube diameter ~22 mm). Given that normal tidal volumes in adults are ~500 ml, this size was deemed sufficient (17). Conversely, for the Additional CO_2 System, the expandable tube was minimized to 0.7 m. The reason for not completely removing the tube was the desire to maintain the flow sensor away from the center of the MRI scanner to avoid interference when using the system in a full BOLD-CVR setup (see Section 3.2 below). Apart from these variances, the breathing circuit remained uniform for the Additional CO_2 and Reservoir CO_2 Systems and comprised a connector with a gas inlet for the addition of pure CO_2 gas (in the Additional CO_2 System) or a gas mixture of O_2 , CO_2 , and N_2 (in the Reservoir CO_2 System). The direction of gas flow was regulated by two one-way valves, and a filter was added to eliminate particles from the inspired gas. A Y-piece separated the inspiration and expiration segments of the circuit, with an elbow connector featuring a sampling port connecting the Y-piece to the face mask (Mask 7450 V2, Vyaire). An in-house 3D printed adapter was used to accommodate the 22 mm elbow to the 30 mm port of the face mask. For a comprehensive inventory of components used, please refer to Supplementary Table S1.

3 Method

3.1 Assessment of inspired CO₂ target accuracy

The primary objective of this study was to assess the accuracy of the Additional CO_2 method in achieving the desired CO_2 target levels within the inspired gas. This assessment was conducted under two distinct scenarios: mechanical ventilation and spontaneous breathing.

The $F_{CO_2}^i$ target function employed in this evaluation encompassed a range of stimulus types, as illustrated in Figure 3. These stimuli included three box-stimulus at 1%, 3%, 5% CO₂, each lasting for 45 s, with an initial 60 s baseline period and a 45 s intermediate baseline. Subsequently, a ramp function was applied, increasing CO₂ concentration from 0 to 5% over 60 s, followed by the first half of a sinusoidal waveform with a peak concentration of 5% and a time period of 120 s. Finally, a 60 s baseline was appended, resulting in a total protocol duration of ~9 min.

3.1.1 Inspired CO₂ target accuracy in mechanical ventilation

The accuracy of inspired CO_2 levels during mechanical ventilation was evaluated using an anesthesia workstation (Primus Infinity Empowered, Dräger Medical) in conjunction with a test lung (AccuLung, Fluke Biomedical), and the Ventilator Circuit shown in the top part of Figure 2. To ensure a comprehensive assessment, a variety of ventilator conditions were considered, aligning with the specifications established by the European standard ISO 80601-2-12:2020 (18). Within this standard, two specific categories were explored: volume-control inflation (table 201.104) and pressure-control inflation (table 201.105). Due to limitations in the available settings of the AccuLung test lung, only the initial seven test cases from each table, totaling 14 test cases, were feasible. The complete list of these test cases is provided in Supplementary Table S4.

For each test case, randomized in order, the Additional CO_2 System administered CO_2 according to the $F_{CO_2}^i$ target function depicted in Figure 3 and the mass balance equation shown in Equation (3). The Primus workstation, functioning both

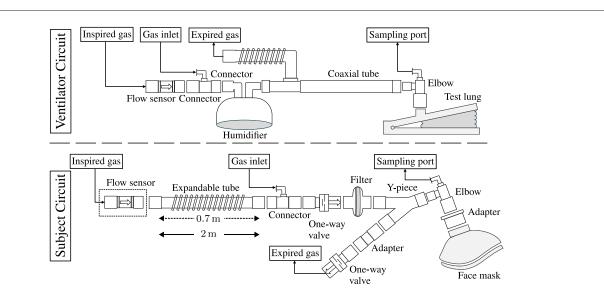
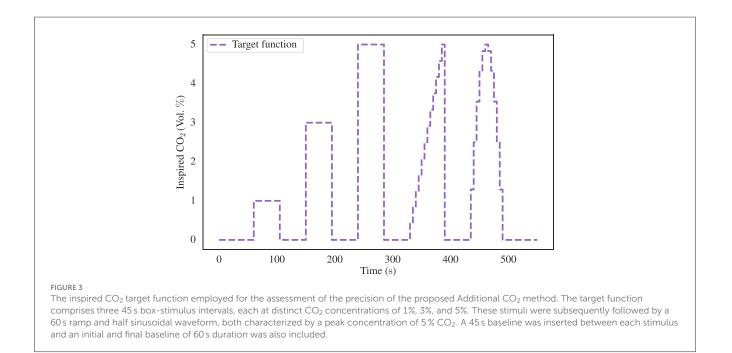


FIGURE 2

A schematic representation of the respiratory circuits employed in mechanical ventilation of a test lung (Ventilator Circuit) and spontaneously breathing healthy subjects (Subject Circuit). In the Ventilator Circuit, the respiratory circuit encompasses a flow sensor affixed to the outlet of the ventilator (not depicted), followed by a connector equipped with a luer port to facilitate the introduction of additional CO₂. To ensure the homogeneity of the gas mixture, an empty humidifier was incorporated to enable gas mixing. A coaxial tube connected to the humidifier and to the ventilator's inlet, with the distal end attached to the test lung via an elbow featuring a sampling port. In the Subject Circuit, the configuration of the respiratory circuit differed slightly for the Additional CO₂ and Reservoir CO₂ Systems. In the Reservoir CO₂ configuration, the deployment of a flow sensor was omitted, and the extendable tube was elongated from its minimal length of 0.7 m, as used in the Additional CO₂ configuration, to a length of 2 m, serving as a reservoir for the administered gas. The expandable tube was affixed to the gas inlet connector, followed by one-way valves and a Y-connector separating the inspiration and expiration part of the circuit A filter prevented particles from reaching the subjects who were breathing in the circuit through a face mask, which was fitted to the head with the help of an adjustable harness (not shown).



as a ventilator and gas analyzer, continuously sampled O_2 and CO_2 concentrations through the sampling port of the breathing circuit at an approximate frequency of 60 Hz. Using the sampled O_2 and CO_2 curves, the inspired O_2 and CO_2 levels were calculated by an automated Python script. The script identified the inspiratory phase and computed both the peak and baseline levels of O_2 and CO_2 to measure the variability within each inspiration. These

values were subsequently interpolated to ensure uniform sampling across all 14 ventilator test cases which enabled aggregation and computation of mean plus confidence intervals using the built-in functionalities of the Seaborn package in Python.

To effectively compare the aggregated values with the target $F^i_{\rm O_2}$ and $F^i_{\rm CO_2}$ levels, the aggregated data was time shifted 8 s to compensate for the sampling delay of 3 s and the presence

of dead space within the ventilator tubing. This dead space necessitated multiple breaths before any alteration in inspired CO_2 concentrations would manifest at the sampling port. While it is acknowledged that individual test runs would have experienced distinct time delays, accounting for variations in tidal volume and respiratory rate, it was determined that the uniform application of the same delay to all runs introduced a relatively minor error when compared with other sources of variation, such as the temporal misalignment between the onset of the stimulus and the start of the subsequent breath.

3.1.2 Inspired CO₂ target accuracy in spontaneous breathing

To conduct a comprehensive evaluation of the Additional CO_2 method, six healthy subjects (aged between 25 and 42, three males and three females) were recruited to assess accuracy of inspired CO_2 in spontaneous breathing. Additionally, we made a comparative analysis between our proposed system: Section 2.2, and the previously described system outlined by Tancredi et al. (5): Section 2.3.

The recruitment process strictly adhered to the principles outlined in the Helsinki Declaration, and ethical approval was obtained from the Swedish Ethical Review Authority (reference number: 2021-04825). Prior to their participation, the selected subjects underwent a screening process to ascertain the absence of pulmonary diseases or other chronic health conditions.

The Subject Circuit, lower part of Figure 2, was used in its different configurations for the Additional CO_2 and Reservoir CO_2 Systems. The acquisition of O_2 and CO_2 concentrations was made by the Primus workstation, now operating in surveillance mode. Notably, the workstation was not connected to the inspiration and expiration portions of the Subject Circuit, as these remained open to the surrounding room environment.

The same $F^1_{CO_2}$ target function employed in the mechanical ventilation assessment was used (see Figure 3). Subjects were instructed to maintain calm and normal breathing while the target stimulus was administered. The experiment was repeated for both the Additional CO2 and Reservoir CO2 configurations for each subject, resulting in a total of 12 experimental runs. The sequence in which these two methods were employed was randomized in blocks to mitigate any order effects. Furthermore, it is relevant to mention that the Reservoir CO₂ method allowed for the specification of the total flow of fresh gas and inspired O₂ levels, which is not actively controlled in the Additional CO2 method. To facilitate comparison between the two methods, the target $F_{O_2}^1$ level in the Reservoir CO₂ System was set to 21 %, approximately corresponding to the ambient room concentration. Achieving the 15 L/min flow rate of fresh gas, as proposed by Tancredi et al. (5), was unattainable due to constraints imposed by the maximum flow capacity of the mass flow controllers in the Reservoir CO₂ System, which was limited to 10 L/min. Instead, we chose to use 8 L/min of fresh gas flow, which is approximately the upper limit of common minute ventilation in healthy adults which range between 6 and 8 L/min (17, 19). This also means minimum excessive usage of fresh gas, making the method comparable with the Additional CO₂ method, where gas is never delivered in excess but always in proportion to the measured respiratory gas flow.

A semi-automated Python script was used to compute inspired and end-tidal O2 and CO2 values from the sampled concentrations. Manual intervention was primarily required because of the interplay between inspired CO2 and subject production of CO2 during spontaneous breathing. Consequently, the sampled CO₂ trace exhibited increased variability, multiple peaks, and valleys compared to the more stable curves observed in passive ventilated test lung scenarios. The semi-automated script initially sought to identify expiration segments within the O_2/CO_2 traces by leveraging the characteristic exponential decay pattern typically exhibited during expiration. Subsequently, these expiration phases were used to pinpoint inspiration phases, situated between two expiration phases. To fully capture the range of values within each inspiration, 1-3 data points were tracked. Inspection of the script's initial estimations, including manual refinements when needed, was then performed. The script achieved a success rate of \sim 95% in accurately identifying expiration/inspiration phases. In the case of the Reservoir CO₂ System, the non-excessive flow of fresh gas (as discussed above) meant that primarily the initial inspired gas was being targeted well, with noticeable reduction in CO₂ levels during the late inspiration phase. However, since the late inspired gas typically does not reach the alveolar space but remains within the physiological dead-space, it was deemed motivated to disregard these late reductions in inspired CO₂ levels to avoid potentially biasing the accuracy of the Reservoir CO2 System negatively. While this approach may introduce a positive bias, it was considered preferable over a negative bias, particularly since the Reservoir CO₂ System served as the reference system for the Additional CO₂ System.

Data aggregation across subjects was facilitated by binning the identified inspired and end-tidal O_2 and CO_2 values into 5 s bins, a process that also introduced some degree of smoothing. Consequently, the need for time-shifting to compensate for the sampling delay of 3 s was eliminated. Inspired values from individual runs could then be aggregated across all subjects and directly compared to the target $F_{O_2}^i$ and $F_{CO_2}^i$ levels, again with the help of Seaborn package in Python. For the end-tidal O_2 and CO_2 values, baseline subject variations were first removed by subtracting the mean end-tidal value during the initial 60 s of each run before aggregating across subjects.

3.2 BOLD-CVR examination

In the context of assessing the Additional CO_2 method as a technique for CVR measurement, it is essential to note that the conventional approach to conducting CVR examinations relies on using the BOLD signal as a surrogate measure of blood flow. We therefore evaluated the Additional CO_2 method in an MRI environment in a subset of two subjects. They underwent BOLD-CVR examinations using both the Additional CO_2 and Reservoir CO_2 Systems, which were repeated twice in a test-retest experimental design, yielding a total of four runs per subject. The same Subject Circuit from the target accuracy assessment (lower part of Figure 2) was again used. Detailed information regarding

the experiment, including MRI and CO_2 protocols, as well as the generation of CVR maps, can be found in Section 2 of the Supplementary material.

Note that the Additional CO₂ System is also safe to use together with the Ventilator Circuit (upper part of Figure 2) inside an MRI environment, of course an MR Conditional ventilator will have to be used.

4 Results

4.1 Target accuracy of inspired gases

The present section delves into the outcomes of the experiment aimed at evaluating the target accuracy of inspired CO2. To illustrate the analysis, Figure 4 shows example datasets. In the uppermost section of Figure 4, the Additional CO₂ System with the Ventilator Circuit is depicted, along with a randomly selected test case. The middle section showcases the Additional CO₂ System with the Subject Circuit in conjunction with a random subject, while the lowermost part illustrates the Reservoir CO2 System with the same subject. In all instances, the same $F_{CO_2}^1$ target function, as detailed in Figure 3, was used. Focusing on the magnified windows, the dynamic fluctuations in CO₂ concentration throughout the 5 % box-stimulus is shown. We see that the rise time of CO_2 in the Ventilator Circuit is longer than the other two configurations. Also, the Reservoir CO₂ System has a greater degree of variability in inspired CO₂ levels when contrasted with the Additional CO₂ System. This is due to the non-excessive usage of fresh gas in the Reservoir CO2 System, resulting in a sudden reduction of inspired CO₂ concentration at the end of the inspiration phase. To avoid negatively biasing the inspired CO2 target accuracy for the Reservoir CO₂ System, only the initial phase of the inspiration is being tracked, as discussed in Section 3.1.2.

To ascertain the performance of the experimental configurations the data from all runs were aggregated over all test cases/subjects as outlined in the Section 2. This process has enabled the computation of the mean and a 95% confidence interval for the inspired/end-tidal CO₂, as displayed in Figure 5. Significantly, it is apparent that the Additional CO₂ System achieves a similar inspired CO₂ target accuracy as the Reservoir CO₂ System, albeit with a consistent undershoot noted in the Ventilator Circuit. Also, similar end-tidal CO₂ changes are seen in both systems. Note that end-tidal values have been converted from volume percentages to partial pressures, assuming an atmospheric pressure of 760 mmHg and water vapor partial pressure of 47 mmHg.

The accuracy and precision of each setup, assessed by the mean deviation between measured and target $F^i_{CO_2}$, was quantified for each type of stimulus. The deviations, after eliminating transition periods for the box-stimuli (initial 10 s and final 5 s) and ramp-stimulus (final 5 s), are summarized in Figure 6. We see that the Additional CO₂ System is consistently targeting the inspired CO₂ to an accuracy better than 0.4 percentage points and performs similar well as the Reservoir CO₂ System across all targets.

Next, we redirect our attention toward the aggregated values of inspired and end-tidal oxygen, as depicted in Figure 7, with the uppermost graph showing the inspired O_2 levels, and the

lower graph showcasing the end-tidal O_2 values. We restrict us to presentation of data from the Additional CO₂ System + Subject Circuit and Reservoir CO₂ System + Subject Circuit configurations. Even though both systems target the same baseline O_2 concentration, 21 %, the Additional CO₂ System did so passively by the usage of room air, which is not exactly 21 %. To facilitate direct comparison between the two configurations, the measured and target $F_{O_2}^i$ levels have been normalized (divided) by their baseline value in Figure 7. Notably, even though the inspired O_2 varies drastically between the two methods, the end-tidal O_2 demonstrate similar patterns marked by a successive rise over time.

4.2 BOLD-CVR measurements

In Figure 8, we present illustrative BOLD-CVR maps obtained through the application of the Additional CO_2 and Reservoir CO_2 Systems within a single subject. It is imperative to emphasize that our objective is not to derive quantitative inferences, however, Figure 8 does unveil a qualitative congruence in the CVR maps yielded by both methods.

5 Discussion

5.1 Inspired CO₂ target attainment

When examining the illustrative data presented in Figure 4, noticeable disparities among the three distinct configurations (Additional CO₂ System + Ventilator Circuit, Additional CO₂ System + Subject Circuit, and Reservoir CO₂ System + Subject Circuit) become evident. First, the ventilator configuration displays slower CO₂ response compared to the two subject configurations. The distinct behavior arises primarily from the gas inlet's placement within these setups. In the Ventilator Circuit, the inlet is located proximal to the flow sensor, whereas, in the Subject Circuit, the inlet is proximal to the sampling port (as depicted in Figure 2). This discrepancy dictates the rate of CO₂ level alteration due to the volume within the tubes, as air is propelled forward in fixed tidal increments. The rationale for not placing the gas inlet proximal to the sampling port in the Ventilator Circuit is the need to minimize the distance between the flow sensor and the gas inlet due to the compression of air within the ventilator tube, characteristic of ventilator operation. Otherwise, a substantial disparity arises between the flow sensor's measured flow and the gas delivered by the mass flow controller. Secondarily, the Additional CO₂ configuration for the selected subject displays considerably less variance in inspired CO₂ values in comparison to the Reservoir CO2 configuration. A closer examination of the data reveals that in the Reservoir CO₂ configuration, the initial inspired CO₂ closely approximates the target value but then suddenly declines toward zero. This behavior is expected given our non-excessive usage of fresh gas [8 L/min instead of 15 L/min as proposed by Tancredi et al. (5)]. To avoid negatively biasing the inspired CO₂ target accuracy of the Reservoir CO2 System, we have only tracked the inspired CO₂ concentrations during the initial phase of inspiration and ignored late declines toward zero. The rationale is that only

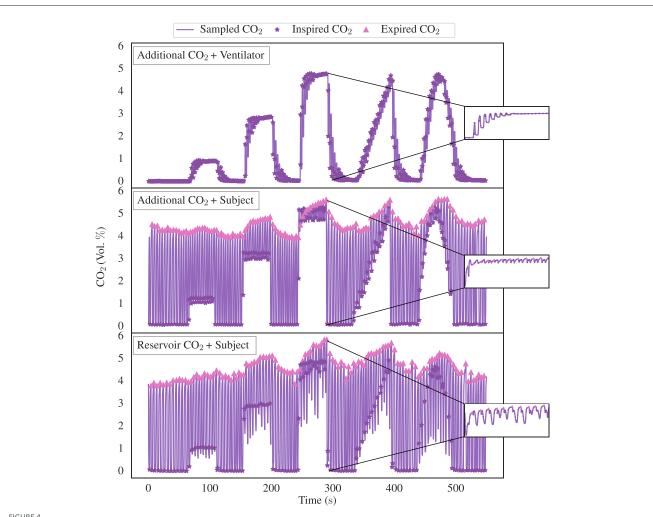


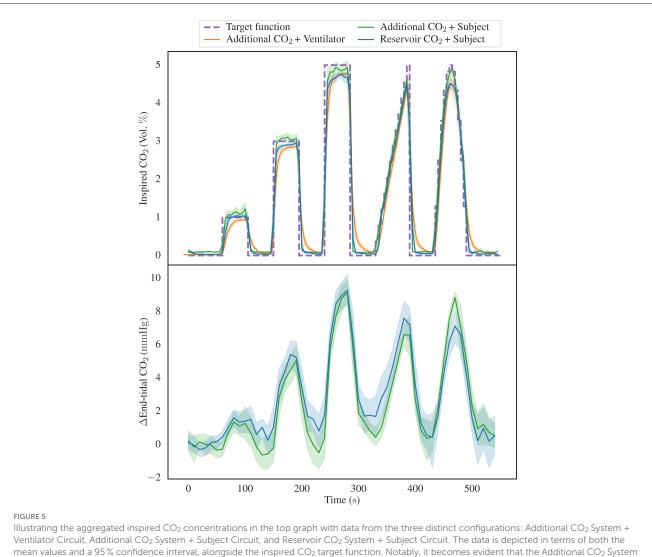
FIGURE 4

Illustrative data pertaining to inspired CO₂ target accuracy assessment, delineating the three distinct experimental configurations: the Additional CO2 System + Ventilator Circuit (depicted in the top graph), the Additional CO2 System + Subject Circuit (displayed in the middle graph), and the Reservoir CO₂ System + Subject Circuit (depicted in the bottom graph). All three instances use the identical target function as showcased in Figure 3. Upon closer examination within the magnified windows, we see the dynamic fluctuations in CO₂ concentration throughout the 5% box-stimulus. It becomes evident that the rise time of CO2 in the Ventilator Circuit exhibits a substantially slower response in comparison to the other two configurations. Furthermore, note the greater degree of variability in inspired CO2 levels for the Reservoir CO2 System when compared with the Additional CO₂ System. This is due to the non-excessive usage of fresh gas in our Reservoir CO₂ System, leading to a reduced inspired concentration at the end of the inspiration phase. However, for the sake of inspired CO₂ target accuracy of the Reservoir CO₂ System, only the concentration during the initial inspired phase is being tracked.

the initial portion of inspired air is significant, as late inspired air resides in the physiological dead-space. However, since it is not possible from our data to distinguish which portion of the inspired CO₂ trace belongs to air remaining in the dead-space, there is a potential positive bias of the CO₂ target accuracy by ignoring the late inspiration phase. Nonetheless, as we are using the Reservoir CO₂ System as a reference system for the Additional CO2 System, any positive bias will not undermine the conclusions drawn regarding the accuracy for the Additional CO₂ System.

Directing our focus to the aggregated inspired CO₂ levels (upper portion of Figure 5), it becomes apparent that the Additional CO2 method consistently adheres to the target value within the subject dataset. Notably, it performs equally well as the Reservoir CO₂ method, a comparison further strengthened in Figure 6, which elucidates the mean divergence between measured and target $F_{CO_2}^1$. We also see that the end-tidal CO₂ levels in the lower portion of Figure 5, exhibit small discrepancy between the two methods, motivating our decision to ignore the late inspired CO2 level reductions in the Reservoir CO2 System. It also highlights that the Reservoir CO2 method seems to work well at non-excessive fresh gas flows, as long as the flow is sufficient to cover alveolar ventilation.

In revisiting the upper portion of Figure 5, it is noteworthy that the Additional CO2 System consistently undershoots the target value in the Ventilator Circuit. While the offset is relatively small, amounting to <0.4 percentage points (see Figure 6), understanding the rationale behind this deviation holds intrinsic value. One plausible explanation pertains to the sensitivity of the SFM3200 flow sensor to flow profile. Preliminary assessments suggest that turbulent flow yields higher readings than laminar flow. Consequently, if the ventilator (Primus workstation) produces a higher degree of laminar gas flow relative to the gas used during the

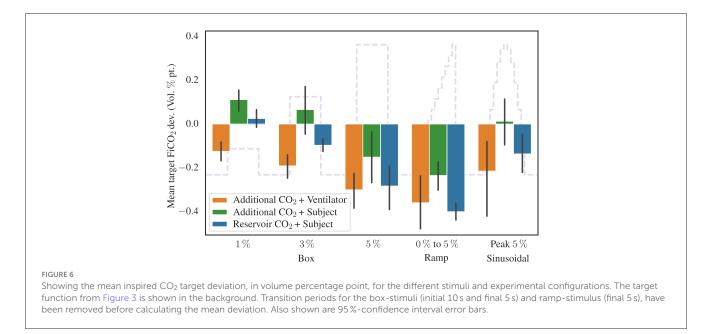


Illustrating the aggregated inspired CO₂ concentrations in the top graph with data from the three distinct configurations: Additional CO₂ System + Ventilator Circuit, Additional CO₂ System + Subject Circuit, and Reservoir CO₂ System + Subject Circuit. The data is depicted in terms of both the mean values and a 95 % confidence interval, alongside the inspired CO₂ target function. Notably, it becomes evident that the Additional CO₂ System network of the aggregated in the ventilator configuration. In the lower graph, the aggregated end-tidal CO₂ levels, although a consistent undershoot is observed in the ventilator configuration. In the lower graph, the aggregated end-tidal CO₂ baseline deviations from the two sets of subject data are presented. We here again notice the similarity between the two methods.

calibration of the Additional CO_2 System (incorporating the flow sensor and mass flow controller), this might elucidate the observed persistent undershoot evident in Figure 5. However, further investigations are requisite to explain this apparent discrepancy in mechanical ventilation. Although any offset is undesirable from a standpoint of repeatability, a consistent target undershoot arguably fares better than a consistent target overshoot concerning subject safety and tolerance.

5.2 Resulting oxygen concentrations

Figure 7 shows the inspired and end-tidal O_2 concentrations for the two subject configurations, using the Additional CO_2 and Reservoir CO_2 Systems. As delineated in the Section 2, the Additional CO_2 method does not actively regulate inspired O_2 concentration, rather, it manifests as a direct consequence of adding CO_2 to the inspired gas. Hence, it is unsurprising that the target $F^i_{\mathrm{O}_2}$ level for the Additional CO_2 method, depicted by the dashed red line in the upper portion of Figure 7, inversely mirrors the target $F^i_{CO_2}$ level (see Figure 3). In the Reservoir CO₂ method, O2 levels are actively controlled by the system and have been maintained at a constant 21 %, as indicated by the dotted red line in Figure 7. To facilitate a comparison between the two methods, the measured and target $F_{O_2}^i$ values have been scaled by their baseline value. Given the conspicuous dissimilarities in measured $F_{O_2}^1$ levels, one might reasonably anticipate notable discrepancies in end-tidal O2 levels. However, a close examination of the lower segment of Figure 7 reveals a lack of pronounced differentiation between the two methods. This phenomenon arises from the recognition that inspired concentration is not the sole determinant of end-tidal values. Variations in alveolar ventilation, by increased or decreased breath frequency and depth, will typically affect end-tidal O2 [(and CO_2), see Equations 1, 2]. Inspecting the end-tidal O_2 curve for the Reservoir CO₂ dataset reveals a progressive elevation over time, even though the inspired O2 concentration stays fixed,



signifying an increasing alveolar ventilation as the experiment unfolds. This seems reasonable given the automatic triggering of reflexes to stimulate deeper and more frequent breaths when CO_2 is inspired (20). Similarly, in the Additional CO_2 configuration, endtidal O_2 levels appear to rise as the experiment progresses, even though inspired O_2 transiently decreases. Hence, the variations in alveolar ventilation obscures the disparities between the Additional CO_2 and Reservoir CO_2 methods. Advanced control systems, such as prospective end-tidal targeting, account for these changes in alveolar ventilation to provide a more precise and reproducible stimulus (8).

It is worthy of note that during the mechanical ventilation of the test lung, tidal volumes, and consequently alveolar ventilation, remained constant when the test lung was ventilated using pressure-control inflation, but not when volume-control was employed. This discrepancy is understandable since, in volumecontrol ventilation, the ventilator administers a predefined tidal volume, with any additional CO₂ gas adding to this volume. Conversely, pressure-control ventilation involves the establishment of a fixed inspiration pressure (Pinsp) at the outset of each breath, maintained for a predetermined duration (Tinsp). In such scenarios, tidal volume becomes dependent solely upon Pinsp, Tinsp and the compliance of the test lung (or patient), why the addition of CO₂ gas does not alter the administered volume. Hence, in the practical application of mechanical ventilation in patients using pressure-control, a reduction in end-tidal O2 levels is to be anticipated when employing the Additional CO₂ method to administer CO₂.

5.3 Compatibility with magnetic resonance imaging

We examined BOLD-CVR in two research subjects to evaluate the compatibility of the Additional CO₂ method with simultaneous MRI measurements. The dataset depicted in Figure 8 presents initial findings, serving as an illustrative demonstration of the feasibility of our proposed Additional CO_2 method. It is crucial to underscore, nonetheless, that a more extensive, in-depth inquiry is imperative to assess the applicability of the Additional CO_2 method in an MRI context. For a more detailed exploration of the BOLD-CVR experiment, we direct interested readers to Section 2 of the Supplementary material.

5.4 Limitations

Currently, the Additional CO_2 System only allows targeting of inspired CO_2 . However, in CVR experiments, it is the alveolar CO_2 concentration which is the parameter of interest. Thus, direct targeting of alveolar CO_2 would be preferable (4). In the last paragraph of Section 2.1, we have outlined potential enhancements to the Additional CO_2 method to enable targeting of alveolar O_2 and CO_2 .

Our Reservoir CO₂ System had some inherent limitations, particularly the flow of fresh gas which was limited due to technical factors. We used 8 L/min of fresh gas, arguing that this leads to a minimal usage of gas which is comparable to the Additional CO₂ System. However, as highlighted by Tancredi et al. (5), the Reservoir CO₂ method performs better with increased gas flow. To partially address this limitation, we adjusted how we computed inspired CO₂ levels for the Reservoir CO₂ System, disregarding late CO₂ level reductions, motivated by the fact that only the initial inspired gas reaches the alveolar space.

In comparing the Additional CO_2 and Reservoir CO_2 Systems, we only considered spontaneous breathing. A more comprehensive analysis would also include comparisons in mechanical ventilation. However, this requires more complicated breathing circuits for the reservoir approach, such as those detailed in Winter et al. (9) or Venkatraghavan et al. (10), which were not available to us.

When evaluating the Additional CO_2 System in mechanical ventilation, we only used a test lung, with no human

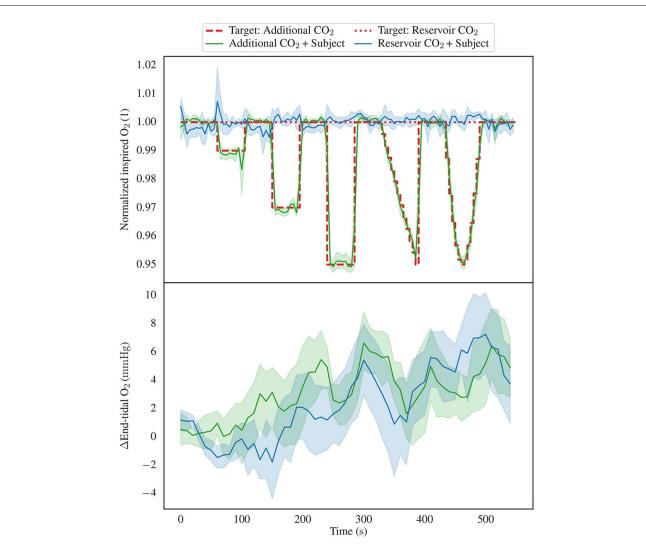


FIGURE 7

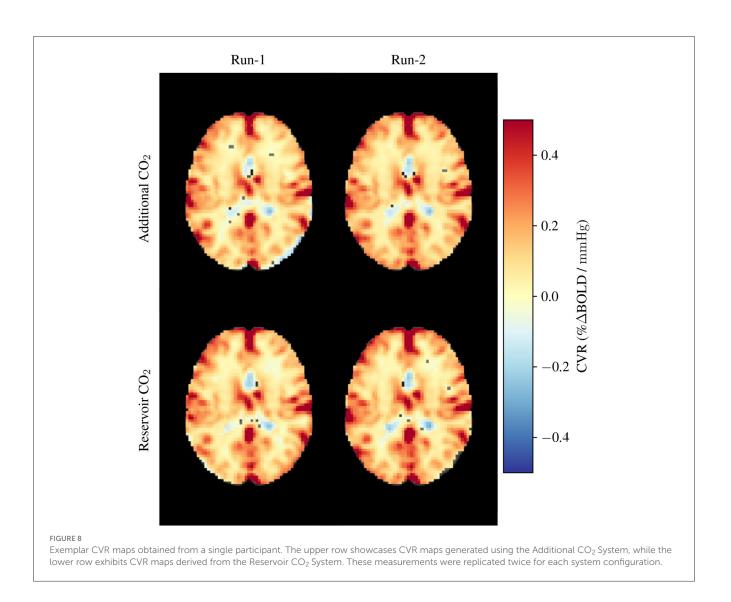
Showing the inspired and end-tidal O_2 levels, focusing exclusively on the Additional CO_2 System + Subject Circuit and Reservoir CO_2 System + Subject Circuit configurations. The top graph displays the inspired O_2 , including both the mean values and 95% confidence intervals. It is noteworthy that two distinct target function are depicted, in the Reservoir CO_2 configuration, the target inspired O_2 concentration remains constant, while in the Additional CO_2 configuration, it varies due to the introduction of additional CO_2 . Further, the measured and target inspired CO_2 concentrations have been normalized by their baseline value to allow for direct comparison between the two methods. The lower graph presents the aggregated end-tidal O_2 baseline deviations. Notably, both graphs exhibit analogous trends characterized by an increase in end-tidal O_2 levels over time, despite the notable disparity in inspired O_2 concentrations between the two experimental configurations.

subject being ventilated. Furthermore, our investigation only considered two types of ventilation mode: volume-control and pressure-control.

6 Conclusion

The contemporary landscape of CVR research predominantly features investigations conducted in spontaneously breathing subjects, with limited attention directed toward patients undergoing mechanical ventilation. A notable constraint contributing to this disparity resides in the lack of suitable apparatus for executing CO₂ gas challenges within a ventilator-dependent setting. Consequently, CVR assessments in ventilated patients have conventionally resorted to alternative stimuli, such as induced apnea, entailing cyclic activation and deactivation of the ventilator, or administration of vasoactive drugs, such as Acetazolamide.

In the present work, we propose a new method, which collaboratively interfaces with mechanical ventilation to administer a variable amount of CO_2 , referred to as Additional CO_2 . We systematically assess the precision of our proposed method in regulating the inspired CO_2 levels and compare its performance against an established method that relies on a gas reservoir containing a mixture of CO_2 at varying concentrations. Furthermore, we evaluate the compatibility of our devised system within an MRI environment, conducting



a BOLD-CVR study. Our findings support the efficacy of our method in maintaining precise inspired CO_2 levels in both mechanical ventilation and in spontaneous breathing. Moreover, it can integrate with an MRI scanner to generate BOLD-CVR maps.

While traditional methods using gas reservoirs remain beneficial for populations predominantly composed of spontaneously breathing patients due to their simplicity, our Additional CO_2 method presents a viable alternative in cases involving both mechanically ventilated and spontaneously breathing patients. This new method avoids the complexities and modifications required by reservoir-based approaches in ventilator circuits. We anticipate that our findings will encourage further CVR research in mechanically ventilated patients in the near future.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by Swedish Ethical Review Authority (reference number: 2021-04825). The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

GM: Conceptualization, Data curation, Investigation, Methodology, Software, Validation, Visualization, Writing – original draft, Writing – review & editing. ME: Funding acquisition, Project administration, Resources, Supervision, Writing – review & editing. CG: Supervision, Writing – review & editing. GC: Funding acquisition, Supervision, Writing – review & editing. LT: Supervision, Writing – review & editing. AT: Funding acquisition, Project administration, Resources, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was made possible through funding from the Swedish Research Council (Grant 2022-02886), the Swedish Brain Foundation (Grant FO2022-0109), and Region Östergötland (ALF grant). GC has used funding from the Swedish Research Council (Grant 2018-05418, 2018-03319, 2023-03186, and 2023-05460), EU project STRATIF-AI (Grant 101080875), and EU project PRECISE4Q (Grant 777107).

Acknowledgments

The authors gratefully acknowledge the Center for Magnetic Resonance Research (CMRR), University of Minnesota, USA, for generously providing the Multi-Band Multi-Echo BOLD sequence used in this study. Special appreciation was extended to Mats Johansson, Region Östergötland, Sweden, and Bengt Rangemalm, Linköping University, Sweden, for their invaluable technical assistance.

References

1. Sleight E, Stringer MS, Marshall I, Wardlaw JM, Thrippleton MJ. Cerebrovascular reactivity measurement using magnetic resonance imaging: a systematic review. *Front Physiol.* (2021) 12:643468. doi: 10.3389/fphys.2021.643468

2. Liu P, De Vis JB, Lu H. Cerebrovascular reactivity (CVR) MRI with CO_2 challenge: a technical review. Neuroimage. (2019) 187:104–15. doi: 10.1016/j.neuroimage.2018.03.047

3. Moreton FC, Dani KA, Goutcher C, O'Hare K, Muir KW. Respiratory challenge MRI: practical aspects. *NeuroImage Clin.* (2016) 11:667–77. doi: 10.1016/j.nicl.2016.05.003

4. Fierstra J, Sobczyk O, Battisti-Charbonney A, Mandell DM, Poublanc J, Crawley AP, et al. Measuring cerebrovascular reactivity: what stimulus to use? *J Physiol.* (2013) 591:5809–21. doi: 10.1113/jphysiol.2013.259150

5. Tancredi FB, Lajoie I, Hoge RD. A simple breathing circuit allowing precise control of inspiratory gases for experimental respiratory manipulations. *BMC Res Notes.* (2014) 7:1–8. doi: 10.1186/1756-0500-7-235

6. Lu H, Liu P, Yezhuvath U, Cheng Y, Marshall O, Ge Y. MRI mapping of cerebrovascular reactivity via gas inhalation challenges. *J Vis Exp.* (2014) 94:52306. doi: 10.3791/52306-v

7. Wise RG, Pattinson KT, Bulte DP, Chiarelli PA, Mayhew SD, Balanos GM, et al. Dynamic forcing of end-tidal carbon dioxide and oxygen applied to functional magnetic resonance imaging. *J Cereb Blood Flow Metab.* (2007) 27:1521–32. doi: 10.1038/sj.jcbfm.9600465

8. Slessarev M, Han J, Mardimae A, Prisman E, Preiss D, Volgyesi G, et al. Prospective targeting and control of end-tidal CO_2 and O_2 concentrations. *J Physiol.* (2007) 581:1207–19. doi: 10.1113/jphysiol.2007.129395

9. Winter JD, Fierstra J, Dorner S, Fisher JA, St Lawrence KS, Kassner A. Feasibility and precision of cerebral blood flow and cerebrovascular reactivity MRI measurements using a computer-controlled gas delivery system in an anesthetised juvenile animal model. *J Magn Reson Imaging*. (2010) 32:1068–75. doi: 10.1002/jmri.22230

10. Venkatraghavan L, Poublanc J, Han JS, Sobczyk O, Rozen C, Sam K, et al. Measurement of cerebrovascular reactivity as blood oxygen level-dependent magnetic resonance imaging signal response to a hypercapnic stimulus in

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed.2024. 1352012/full#supplementary-material

mechanically ventilated patients. J Stroke Cerebrovasc Dis. (2018) 27:301–8. doi: 10.1016/j.jstrokecerebrovasdis.2017.08.035

11. Brauer P, Kochs E, Werner C, Bloom M, Policare R, Pentheny S, et al. Correlation of transcranial doppler sonography mean flow velocity with cerebral blood flow in patients with intracranial pathology. *J Neurosurg Anesthesiol.* (1998) 10:80–5. doi: 10.1097/00008506-199804000-00003

12. Fierstra J, Burkhardt JK, van Niftrik CHB, Piccirelli M, Pangalu A, Kocian R, et al. Blood oxygen-level dependent functional assessment of cerebrovascular reactivity: feasibility for intraoperative 3 tesla MRI. *Magn Reson Med.* (2017) 77:806–13. doi: 10.1002/mrm.26135

13. Sari A, Yamashita S, Ohosita S, Ogasahara H, Yamada K, Yonei A, et al. Cerebrovascular reactivity to $\rm CO_2$ in patients with hepatic or septic encephalopathy. *Resuscitation.* (1990) 19:125–34. doi: 10.1016/0300-9572(90)90035-D

14. Deckers PT, Siero JCW, Mensink MO, Kronenburg A, Braun KPJ, van der Zwan A, et al. Anesthesia depresses cerebrovascular reactivity to acetazolamide in pediatric moyamoya vasculopathy. *J Clin Med.* (2023) 12:4393. doi: 10.3390/jcm12134393

15. Branson RD, Griebel J, Rodriquez D. A bench study of inhaled nitric oxide delivery during high frequency percussive ventilation. *Pediatr Pulmonol.* (2018) 53:337–41. doi: 10.1002/ppul.23934

16. McSwain SD, Hamel DS. End-tidal and arterial carbon dioxide measurements correlate across all levels of physiologic dead space. *Respir Care*. (2010) 55:288–93.

17. Hallett S, Toro F, Ashurst JV. *Physiology, Tidal Volume.* Treasure Island, FL: StatPearls Publishing (2023).

18. ISO 80601-2-12:2020. Medical Electrical Equipment – Part 2-12: Particular Requirements for Basic Safety and Essential Performance of Critical Care Ventilators. Brussels: European Committee for Standardization (2020).

19. Sapra A, Malik A, Bhandari P. Vital Sign Assessment. Treasure Island, FL: StatPearls Publishing (2023).

20. Carr JMJR, Caldwell HG, Ainslie PN. Cerebral blood flow, cerebrovascular reactivity and their influence on ventilatory sensitivity. *Exp Physiol.* (2021) 106:1425-48. doi: 10.1113/EP089446