Check for updates

OPEN ACCESS

EDITED BY Felix Carl Fabian Schmitt, Heidelberg University Hospital, Germany

REVIEWED BY Vivienne Theobald, Heidelberg University Hospital, Germany Chiara Middel, Heidelberg University Hospital, Germany

*CORRESPONDENCE Larissa Henze ⊠ larissa.henze@med.uni-rostock.de

RECEIVED 26 November 2023 ACCEPTED 15 December 2023 PUBLISHED 11 January 2024

CITATION

Henze L, Grunwald L, Felser S, Witte M, Grosse-Thie C, Roolf C, Murua Escobar H and Junghanss C (2024) Abdominal venous thromboses: detection of the *JAK2* p.V617F mutation by next-generation ultradeep sequencing—A prevalence study of patients in Mecklenburg-West Pomerania (2017–2021). *Front. Med.* 10:1344769. doi: 10.3389/fmed.2023.1344769

COPYRIGHT

© 2024 Henze, Grunwald, Felser, Witte, Grosse-Thie, Roolf, Murua Escobar and Junghanss. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms. Abdominal venous thromboses: detection of the *JAK2* p.V617F mutation by next-generation ultradeep sequencing—A prevalence study of patients in Mecklenburg-West Pomerania (2017–2021)

Larissa Henze^{1*}, Luise Grunwald¹, Sabine Felser¹, Maria Witte², Christina Grosse-Thie¹, Catrin Roolf¹, Hugo Murua Escobar¹ and Christian Junghanss¹

¹Department of Medicine, Clinic III—Hematology, Oncology, and Palliative Medicine, Rostock University Medical Center, Rostock, Germany, ²Department of General, Visceral, Thoracic, Vascular, and Transplant Surgery, Rostock University Medical Center, Rostock, Germany

Background: Abdominal venous thromboses are rare thrombotic events with heterogeneous etiologies. They are related to myeloproliferative neoplasms (MPNs) in some patients and can occur as first signs of the disease. MPNs are characterized by mutations in the genes of Janus kinase 2 (JAK2), myeloproliferative leukemia virus oncogene (MPL), and calreticulin (CALR).

Methods: Within the prospective trial "Prevalence of JAK2 mutations in patients with abdominal venous thromboses" (JAK2 MV study; German Clinical Trials Register: DRKS00026943), the peripheral blood of patients with abdominal venous thromboses in Mecklenburg-West Pomerania, a federal state located in north-east Germany, was analyzed by next-generation ultradeep sequencing for MPN-associated mutations. Clinical characteristics and blood cell counts were also of interest. The primary endpoint was the detection of the mutation *JAK2* p.V617F. Secondary endpoints were the detection of other acquired variants of JAK2, as well as MPL and CALR.

Results: A total of 68 patients with abdominal venous thromboses were included from February 2017 to January 2021, with splanchnic veins affected in 65 patients. The mutation *JAK2* p.V617F was present in 13 patients (19%), with four patients showing low variant allele frequencies (VAF 0.1% to 1.9%). The time interval from the thrombotic event to analysis was longer for patients with the mutation. The mutation *MPL* p.W515R was detected in three cases, all of them with low VAF. One patient among them had a concurrent mutation of *JAK2* p.V617F. The mutations *CALR* type I or type II were not found.

Discussion: By analyzing peripheral blood for the mutation *JAK2* p.V617F, an important cause of these rare thrombotic events can be identified. The development of a diagnostic workup with next-generation ultradeep sequencing for the analysis of the *JAK2* p.V617F mutation and further mutations has the potential to better understand the etiology of abdominal venous thromboses in

individual patients in regional clinical care, as abdominal venous thromboses are diagnosed by various medical disciplines.

KEYWORDS

abdominal venous thromboses, splanchnic vein thrombosis, JAK2 mutation, ultradeep sequencing, anticoagulation

Introduction

Abdominal venous thromboses are rare events (1). In the literature, hepatic vein thromboses (also known as Budd-Chiari syndrome), portal vein thrombosis, splenic vein thrombosis, and mesenteric vein thromboses are grouped as splanchnic vein thromboses (SVT) (1-4). Incidences range from 0.7 per million people per year for hepatic vein thromboses to 70 per million people for portal vein thromboses (5-7). Thromboses in the abdominal region may also involve the vena cava inferior and other abdominal veins such as renal or ovarian veins. Multiple causes and triggers of abdominal venous thromboses have been described: cirrhosis, inflammatory disease, particularly pancreatitis and inflammatory bowel disease (8), abdominal surgery (9), malignancies (8), myeloproliferative neoplasms (MPN) (1, 10), and congenital or acquired coagulation disorders (1, 11). Abdominal venous thromboses can be complicated by acute sequelae such as liver failure, gastrointestinal bleeding (1), intestinal infarction (12), perforation, peritonitis, and sepsis (11) and can lead to chronic complications, mostly the formation of varices with bleeding risk or re-thrombosis (1, 13). Therefore, in addition to the diagnosis of abdominal venous thrombosis, clarification of causality is fundamental for further treatment decisions and clinical outcomes (2).

The study "Prevalence of JAK2 Mutations in Patients with Abdominal Venous Thrombosis" (JAK2 MV Study) prospectively investigated patients with abdominal venous thromboses in the federal state of Mecklenburg-West Pomerania (MV) in northeast Germany (see Supplementary Figure 1) for the presence of mutations in the gene of Janus kinase 2 (JAK2). The mutation JAK2 p.V617F represents one of the major diagnostic criteria of MPN (14, 15) and is frequently found in patients with abdominal venous thromboses. JAK2 is a non-receptor tyrosine kinase (16). Activated JAK2 generates the transcription factors known as "Signal transducers and activators of transcription." This JAK-STAT pathway is involved in the proliferation, differentiation, and self-renewal of the hematopoietic system. The mutation JAK2 p.V617F constitutively activates the tyrosine kinase (16), leading to increased erythropoiesis and thrombopoiesis and thus resulting in the phenotype MPN (17). Other known driver mutations of MPN, though less commonly detected, exist for myeloproliferative leukemia virus oncogene (MPL p.W515R) and calreticulin (CALR type I and CALR type II) (18). Both of these also result in the activation of the JAK-STAT pathway: mutations in the MPL gene that encodes the thrombopoietin receptor by affecting the intracellular domain of the protein followed by ligand-independent signaling through JAK2, mutations in the CALR gene that alter binding of CALR to MPL associated with JAK2, and thereby over-activating the JAK-STAT signaling pathway (19-22). Both mutations of MPL and CALR were included in the JAK2 MV study as secondary endpoints.

The implementation of the study in the federal state of Germany allows for the depiction of the reality of clinical care for patients with abdominal venous thromboses in a defined region with existing data about population and infrastructure. It also helps to better characterize one of the main causes of abdominal venous thromboses. Potentially, it can lay the groundwork for the development of a diagnostic workup in general clinical care in this area.

Methods

Study design

The investigator-initiated JAK2 MV trial was a prospective cohort study to analyze patients in MV with abdominal venous thromboses for the presence of acquired single nucleotide variants in the JAK2 gene. The primary endpoint was the detection of the *JAK2* p.V617F mutation. Secondary endpoints were the detection of other acquired variants of JAK2, as well as the mutations *MPL* p.W515R and *CALR* type I and type II, which are also part of the diagnostic criteria of MPN (14, 15).

Patients in MV with thromboses in hepatic veins, portal veins, splenic veins, mesenteric veins, and/or other abdominal veins were included in the study, regardless of the timepoint of the thrombotic event. For this study, all thrombotic events within 1 month before inclusion were classified as recent events, whereas the others were termed past events. Thrombosis had to be confirmed by imaging [ultrasound, computed tomography (CT), and magnetic resonance imaging (MRI)] or surgically for inclusion in the study. Written informed consent by the patient was required. Patients < 18 years were excluded.

A total of 5 ml of peripheral blood (anticoagulant EDTA) was collected for next-generation sequencing (NGS) and a blood smear for cytology. A standardized recording of demographic, clinical, and laboratory parameters using a questionnaire was within the scope of the study.

The organization of the study, data collection, and analysis, as well as laboratory work (NGS and cytology) were performed at Clinic III—Hematology, Oncology, and Palliative Medicine, Rostock University Medical Center.

The Ethics Committee of Rostock University Medical Center has approved the trial (approval date: 22 November 2016, A 2016– 0200), and the trial is registered in the German Clinical Trials Register (DRKS00026943).

Study implementation

At the beginning of the study, all departments of internal medicine (n = 27) and surgery (n = 28) at hospitals in MV with at least 80 beds, all specialists in hematology and oncology (n = 17), and ~ 7 months after the start of the study, all specialists in gastroenterology (n = 20) were informed about the study by post. In addition to a general brochure about the trial, a patient information sheet, an informed consent form, and a clinical questionnaire were included in the postal mailing. The trial was also promoted at regional educational and scientific meetings.

Patients were informed and included by their treating physicians, who also collected peripheral blood and answered the clinical questionnaire. Blood samples were sent by standard mail. Only upon receipt of the blood sample, together with a valid informed consent form and clinical questionnaire, the analysis of the blood sample was proceeded within the study. Sample processing and analysis were performed pseudonymously.

In cases where a mutation was identified, the respective physician was informed by phone and by written report.

Next-generation sequencing

A dedicated targeted sequencing gene panel for NGS was designed and validated for use in routine diagnostics. In addition to the total protein-coding sequences of JAK2, the panel also captures the total protein-coding sequences of MPL and CALR. Nucleic acid segments with known mutation hotspots were covered by at least two amplicons in the panel design. Nucleic acid isolation as well as the performance of NGS are outlined in the study by Grunwald et al. (23). Generally, in routine diagnostics for conventional applications, a sensitivity of NGS of approximately 2% is described (24). Here, validation of the above-described NGS panel detected variant allele frequencies (VAF) of 1% and below as the condition of high amplicon coverage (> 2,000) was fulfilled as ultradeep sequencing.

Statistical analysis

Descriptive participants' characteristics were calculated. Interval-scaled data were tested for normal distribution using the Shapiro-Wilk test. Mean differences were tested using Fisher's exact test or the Mann–Whitney *U*-test. The significance level was set at $p \leq 0.050$. SPSS 25.0 (SPSS Inc., Chicago, IL, United States) was used for the analysis.

Results

Study participants

Between 22 February 2017 and 31 January 2021, 73 blood samples were submitted, of which 68 could be included and analyzed in the study (Figure 1). From Rostock originated 84% of samples, and 16% were from different locations in MV (n = 4 from Ribnitz-Damgarten, n = 2 from Greifswald, n = 2 from Stralsund,

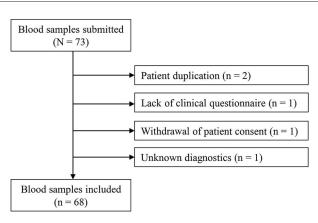
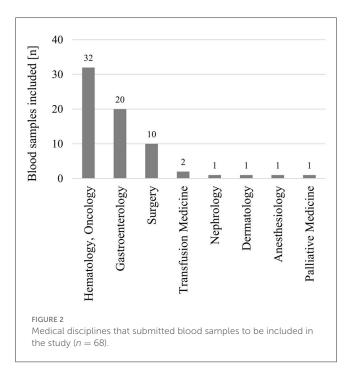


FIGURE 1

Flowchart of the study. Unknown diagnostics: the diagnostic method for the detection of the thrombosis was not known (Thrombosis had to be confirmed by imaging or surgically for inclusion in the study).



n = 1 from Neubrandenburg, n = 1 from Waren, and n = 1 from Wismar) (see Supplementary Figure 1). Figure 2 lists the medical disciplines that submitted blood samples.

Patients' characteristics are summarized in Table 1. Overall, 40 (59%) patients were male, 28 (41%) were female. The median age was 62.0 years (range 28–84 years) for male patients and 53.5 years (range 29–80 years) for female patients. In 65 of the 68 patients with abdominal venous thromboses (96%), splanchnic veins were involved. Thrombosis was most commonly localized in the portal vein (n = 52), followed by the splenic vein (n = 27) and mesenteric veins (n = 27). In 33 patients, thromboses were located in two or more abdominal veins. In total, 37 patients had a recent thrombotic event. For the other patients, the thrombotic diagnosis lasted from 32 days to 20 years (in the mean 12 months).

TABLE 1 Characteristics of study participants (n = 68).

	Total	JAK2 p.V617F negative	<i>JAK</i> 2 p.V61	p (negative v positive)	
	n = 68	n = 55	VAF 0.1–1.9 % n = 4	$VAF \ge 2 \%$ n = 9	
Sex, <i>n</i> (%)					0.123
Female	28 (41)	20 (36)	3 (75)	5 (56)	
Male	40 (59)	35 (64)	1 (25)	4 (44)	
Age (years)					0.167
Median (min-max)	61 (28-84)	60 (28-84)	69 (60–77)	64 (44–73)	
Thrombotic site, <i>n</i>					
Portal vein	52	40	3	9	0.636
Hepatic vein	5	4	0	1	0.22
Splenic vein	27	22	1	3	0.502
Mesenteric vein	27	22	1	3	0.553
SVT	65	53	3	9	0.353
Other veins	9	6	2	1	0.327
Single vs. multiple sites, n (%)					0.107
Single site	26 (38)	23 (42)	2 (50)	1 (11)	
Multiple sites	33 (49)	27 (49)	2 (50)	4 (44)	
Unknown	9 (13)	5 (9)	0	4 (44)	
Date of thrombotic event, <i>n</i> (%)					0.041*
= 1 month</td <td>37 (54)</td> <td>33 (60)</td> <td>2 (50)</td> <td>2 (22)</td> <td></td>	37 (54)	33 (60)	2 (50)	2 (22)	
>1 month	29 (43)	20 (36)	2 (50)	7 (78)	
Unknown	2 (3)	2 (4)	0	0	
Treatment, n					
Surgery	8	4	1	3	0.041*
Thrombolysis°	3	3	0	0	
Anticoagulation	54	45	4	5	0.13
Not specified	7	6	0	1	
Hepatomegaly, n (%)					0.142
Yes	10 (15)	9 (16)	0	1 (11)	
No	49 (72)	40 (73)	4 (100)	5 (56)	
Unknown	6 (9)	3 (6)	0	3 (33)	
Not specified	3 (4)	3 (6)	0	0	
Splenomegaly, n (%)					0.373
Yes	19 (28)	15 (27)	0	4 (44)	
No	41 (60)	34 (62)	4 (100)	3 (33)	
Unknown	5 (7)	3 (6)	0	2 (22)	
Not specified	3 (4)	3 (6)	0	0	
Previous thrombosis, n (%)					0.239
Yes	7 (10)	4 (7)	0	3 (33)	
No	44 (65)	37 (67)	3 (75)	4 (44)	
Unknown	14 (21)	12 (22)	1 (25)	1 (11)	
Not specified	3 (4)	2 (4)	0	1 (11)	

(Continued)

TABLE 1 (Continued)

	Total	<i>JAK</i> 2 p.V617F negative	JAK2 p.V617F positive		p (negative vs. positive)	
	n = 68	n = 55	VAF 0.1–1.9 % n = 4	VAF ≥ 2 % <i>n</i> = 9		
Thrombophilia, n (%)					1	
Yes	3 (4)	3 (6)	0	0		
No	37 (54)	30 (55)	2 (50)	5 (56)		
Unknown	27 (40)	22 (40)	2 (50)	2 (50) 3 (33)		
Not specified	1 (2)	0	0 1 (11)			
Malignancy, n (%)					0.291	
Yes°°	21 (31)	19 (35)	1 (25) 1 (11)			
No	34 (50)	25 (46)	2 (50)	7 (78)		
Unknown	12 (18)	10 (18)	1 (25)	1 (11)		
Not specified	1 (2)	1 (2)	0	0		
Liver cirrhosis, n (%)					0.193	
Yes	8 (12)	5 (9)	0	3 (33)		
No	49 (72)	42 (76)	2 (50)	5 (56)		
Unknown	11 (16)	8 (15)	2 (50)	1 (11)		

SVT, splanchnic vein thrombosis; VAF, variant allele frequency. $^{\circ}$ No systematic recording of lysis therapy in the JAK2-MV trial. $^{\circ\circ}$ Malignancies reported in patients without detection of the JAK2 p.V617F mutation: (suspected) pancreatic cancer (n = 6), lymphoma, carcinoma of unknown primary (suspected,) gastric cancer (suspected), lung cancer (suspected), cholangiocellular carcinoma, lymphoma (n = 2 each), rectal cancer, intraductal papillary mucinous neoplasia, liver tumor, chronic lymphocytic leukemia (n = 1 each). Malignancies reported in patients with the mutation JAK2 p.V617F are listed in Table 2. *Statistical significant.

Data on the clinical situation were presented by the referring physician and recorded as text on the questionnaire. The presenting symptoms were variable. Pain, gastrointestinal complaints, and liver-associated symptoms were most often mentioned. Of note, the diagnosis was incidental (no symptoms) in 13 patients.

CT was used most frequently (n = 54), followed by ultrasound (n = 21) and MRI (n = 5) as diagnostic procedures. Multiple methodologies were applied to 13 patients. In three patients, diagnosis was established during surgery.

As therapeutic procedure, surgery was performed on eight patients (12%) (Table 1). The use of anticoagulation was reported for 54 (79%) patients in total (Table 1) with the following distribution: low-molecular-weight heparins (LMWH) (n = 24), heparin (n = 3), phenprocoumon (n = 7), direct oral anticoagulants (n = 10), and unspecified (n = 6). Furthermore, enoxaparin in prophylactic dosage was mentioned for two patients, as was clopidogrel \pm acetylsalicylic acid for two patients. For three patients, information about thrombolysis was listed. It has to be noted, though, that this treatment option was not systematically queried in the standardized reporting form (Table 1).

Prevalence of the mutation JAK2 p.V617F

The mutation *JAK2* p.V617F was detected in 13 of 68 study participants (VAF 0.2–43%), corresponding to 19%. Four of the thirteen patients had VAF between 0.1% and 1.9%,

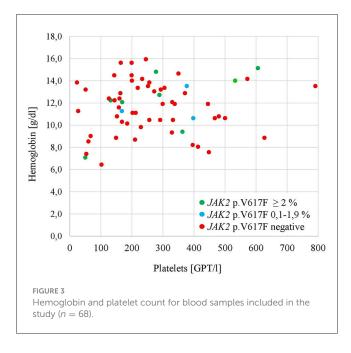
which is below the conventional cut-off used for routine diagnostics by NGS. The characteristics of study participants with the mutation *JAK2* p.V617F are summarized in Table 2. Strikingly, all but one study participant with the mutation had portal vein thrombosis. Five patients also had thromboses in other abdominal veins, and two patients had experienced a previous thrombotic event. Tumor diseases were reported in these patients as "suspected leukemia" (MPN38; VAF 43.0%), possibly MPN, and carcinoma of the esophagogastric junction (MPN36; VAF 1.4%).

Table 1 presents the characteristics of the total study population for patients without mutation of JAK2 p.V617F and for patients with JAK2 p.V617F mutation (VAF 0.1–1.9% and VAF \geq 2%, respectively). Patients with a mutation of JAK2 p.V617F, regardless of VAF, were summarized for statistical analysis. Individuals with the mutation were more likely to be female and older. Patients with the mutation also tended more often to have suffered from a previous thrombotic event or to have liver cirrhosis, whereas malignancy was less commonly suspected. Hepatomegaly and splenomegaly were comparable between patients with and without the mutation of JAK2 p.V617F. Figure 3 visualizes the findings of the hemoglobin and platelet counts of the three groups, showing a scattering of the values without classification of the groups. Patients with the mutation JAK2 p.V617F had a significantly longer history of abdominal venous thromboses. They were significantly more often treated by surgery (4 of 13 patients). Nine of the thirteen patients with the mutation JAK2 p.V617F received anticoagulants.

Code	<i>JAK2</i> p.617F VAF (%)	Portal vein	Hepatic vein	Splenic vein	Mesenteric vein	Other veins	Previous thrombosis	Suspected malignancy	Liver cirrhosis
MPN38	43.0	+	Unknown	Unknown	Unknown	Unknown	+	+ 4	+
MPN18	31.4	+	Unknown	Unknown	Unknown	Unknown	Not specified	-	-
MPN43	28.4 ⁶	+	Not specified	Not specified	Not specified	Not specified	-	-	+
MPN30	24.6	+	Unknown	Unknown	Unknown	Unknown	-	Unknown	Unknown
MPN42	17.3	+	+	+	Not specified	-	-	-	-
MPN11	13.2	+	Unknown	+	+	+ 1	Unknown	-	-
MPN44	13.2	+	-	-	-	-	+	-	-
MPN67	12.3	+	-	-	+	-	+	-	+
MPN72	4.5	+	-	+	+	-	-	-	-
MPN36	1.4	-	-	-	-	+ 2	-	+ 5	-
MPN34	0.6	+	-	+	+	Not specified	Unknown	Unknown	Unknown
MPN45	0.3	+	-	-	-	+ 3	-	-	-
MPN28	0.2	+	Not specified	Not specified	Not specified	Not specified	-	-	Unknown

TABLE 2 Patients with the mutation JAK2 p.V617F: site of thrombosis, previous thrombotic event, malignancy, and liver cirrhosis.

VAF, variant allele frequency; +, yes; -, no. ¹ venous confluence; ² left renal veins; ³ plexus venosus uterinus. ⁴ suspected leukemia; ⁵ carcinoma of the esophagogastric junction. ⁶ in addition MPL p.W515R with VAF 0.2%.



Prevalence of the mutations *MPL* p.W515R and *CALR* type I and type II

Mutations of *MPL* p.W515R were detected in three cases with VAFs of 1.2%, 0.4%, and 0.2%, thus showing low VAF. The latter patient had a concurrent mutation of *JAK2* p.V617F (MPN43; VAF 28.4%) (Table 2). CALR mutations were not found.

Discussion

In the literature, in 50%-75% of patients with abdominal venous thromboses, the etiology could be identified (8, 11, 22). In addition to thrombophilia, acquired local and systemic pathophysiological factors are responsible; among the latter, MPNs are of particular importance (3, 22). MPNs associated with abdominal venous thromboses predominantly show the mutation JAK2 p.V617F (2). In the prospective study "Prevalence of JAK2 mutations in patients with abdominal venous thrombosis" (JAK2 MV study) described here, a prevalence of 19% was found for the mutation JAK2 p.V617F, whereas the mutation MPL p.W515R was only detected in two additional patients (3%) with low VAF. This is in line with other reports about the distribution of the driver mutations in the genes of JAK2, MPL, and CALR in patients with SVT with and without MPN (25-27). While all driver mutations can cause hyperproliferation of myeloid progenitor cells, the mutation JAK2 p.V617F has been described to contribute to a prothrombotic state by different mechanisms, like enhanced vascular and intercellular adhesion of neutrophils and increased formation of neutrophil extracellular traps (18, 28-30), all of which have not yet been fully elucidated (31). However, as shown in this study and others, the association between the mutation JAK2 p.V617F and abdominal venous thromboses is clinically evident. Ageno et al. (1) reported a frequency of JAK2 p.V617F of 20.1

% in a multicentric cohort of patients with SVT with thrombotic events ranging back < 6 months. In the prospective multicenter pilot study for rivaroxaban as a treatment for SVT, JAK2 p.V617F was detected in 13 of 50 patients tested (26%); patients with Budd-Chiari syndrome or liver cirrhosis were excluded (32). In a monocentric study from 1994 to 2021, published by Colaizzo et al. (13), JAK2 p.V617F was found in 29.8% by systematic screening of 152 patients at the time of diagnosis of SVT without concomitant cirrhosis or carcinoma. In this study, patients were followed up (median for 64 months, range 1-214 months) for both detection of JAK2 p.V617F and diagnosis of MPN: 9 of 13 patients with the initial finding of JAK2 p.V617F and 6 of 8 patients with detection of JAK2 p.V617F during follow-up were diagnosed with MPN during the course of the study. Analysis of JAK2 p.V617F was performed using real-time quantitative PCR; the detection limit was defined at 1.5%, but information on VAF was not given.

In the JAK2 MV study, ultradeep sequencing was used to detect and quantify the mutation JAK2 p.V617F. Given the sensitivity of ultradeep sequencing, VAF from 0.1% to 1.9% could also be described. In the study cohort, four patients showed JAK2 p.V617F with VAF in this range, and nine patients showed VAF >/= 2% (range 4.5%-43.0%). Patients with evidence of the mutation were diagnosed earlier with abdominal venous thromboses, in line with the observation of Colaizzo et al. (13). How et al. (2, 33) postulate that abdominal venous thromboses are early events, preceding the diagnosis of MPN in some patients. In a Danish population study with nearly 20,000 participants, Cordua et al. (17) showed an increased odds ratio for venous and arterial thromboses in case JAK2 p.V617F was present with low VAF without a diagnosis of MPN. In light of this, the high sensitivity of the sequencing method is particularly important. In parallel, within the JAK2 MV study, the presence of JAK2 p.V617F in patients was not associated with hepatomegaly, splenomegaly, or blood count abnormalities (clinical or laboratory signs of MPN). As depicted in Figure 1, hemoglobin levels and platelet counts varied. Particularly hemoglobin levels > 16.5 g/dl or > 16.0 g/dl (referring to men or women), or platelet counts > 450 GPT/l as major diagnostic criteria for MPN (polycythemia vera and essential thrombocythemia, respectively), according to the International Consensus Classification of 2022 (15), were only seen in a few cases with the presence of JAK2 p.V617F. This implies that high blood cell counts are not the main trigger factors for thrombotic events. The abdominal venous thromboses may influence the development of blood count abnormalities through portal hypertension, inflammation, or bleeding episodes (27). More importantly, as described above, abdominal venous thromboses can be found early before evidence of MPN by diagnostic criteria (14, 15).

Consistent with the literature (2, 13, 33), individuals with abdominal venous thromboses and *JAK2* p.V617F were more likely to be female. Individuals with *JAK2* p.V617F tended to be older. In the literature, comparative data about age refer to individuals with SVT compared to patients with other thromboses (2) or patients with MPN (4, 33). In this regard, patients with SVT tend to be younger.

Population aspects

The JAK2 MV study had no defined exclusion criteria except for age < 18 years. The study was open to all patients with current or precedent evidence of abdominal venous thromboses in MV. As a consequence, the study cohort was heterogeneous, with competing causes of the thrombotic event, but it represented general clinical care. This highlights the study because, in contrast to other reports, it was not confined to patients of referral centers (1, 7, 12, 13, 34) or MPN or SVT registries (4). Compared to the literature (1, 13), the prevalence of *JAK2* p.V617F in total was lower, but the mutation was still found in about one in five patients. Patients with SVT were represented within the study cohort at 96%.

The study was restricted to MV. The federal state in northeast Germany is an area of low population density with 69 inhabitants per km², consisting of 1.6 million inhabitants (35). In contrast to urban areas, the distribution of medical specialists is limited, and specialized diagnostic centers are scarce. Patients with rare medical problems might be underdiagnosed. Initiating the prospective trial "Prevalence of JAK2 mutations in patients with abdominal venous thromboses" was a step toward distributing highly sophisticated diagnostic tools to potentially all patients in MV.

Diagnostic aspects

A unique feature of this study was the application of a highly sensitive and quantitative diagnostic method for the detection of the mutation JAK2 p.V617F: ultradeep sequencing. This allowed the detection of VAF below the sensitivity limitation of NGS or conventional applications such as quantitative polymerase chain reaction (36, 37). Baumeister et al. (37) mention that VAFs of 1% lead to subclinical manifestations of MPN. The JAK2 MV study was not designed to confirm the diagnosis of MPN by further investigation or to follow-up patients. Nevertheless, this aspect is of particular interest for patients with VAF of *JAK2* p.V617F between 0.1% and 1.9%. It should be emphasized that with the detection of *JAK2* p.V617F, further hematological diagnostics regarding MPN are required. This was communicated to the submitting colleagues. Due to the study design, no information about the results was available.

Limitations

The following limitations of the study have to be acknowledged. First, the study is restricted to MV and might therefore be influenced by the geographic or genetic characteristics of its population. Second, due to the population of MV and the rarity of the thrombotic manifestations, the cohort is small. Still, the results are comparable to data from the literature, taking into account the differences in the inclusion criteria. Third, because the study period ranged from 22 February 2017 to 31 January 2021, the data might be influenced by the COVID-19 pandemic in the last 12 months. During this time, 12 patients were included. Finally, the study was designed as a prevalence study, and a follow-up was not within the scope of the study protocol. Nevertheless, receiving results of further hematological diagnostics would be of great interest for patients with mutations in *JAK2* V617 with VAF \geq 2% to differentiate MPN from clonal hematopoiesis of indeterminate potential (CHIP) and in those with VAF between 1.0% and 1.9%.

Conclusion

The JAK2 MV study presents relevant aspects of general clinical care in patients with abdominal venous thromboses: Patients were diagnosed by various medical disciplines. Clinical criteria and blood counts were not conclusive for etiology. By ultradeep nextgeneration sequencing of peripheral blood samples, the mutation JAK2 p.V617F was detected in 19% of patients included in the JAK2 MV study. Looking ahead, longitudinal examination of VAF is of interest, as is patients' follow-up with a focus on additional hematological results, especially in the light of evolving data about the pathophysiological influence of CHIP (38). The JAK2 MV study suggests, along with others (38), that "CHIP-associated SVT" (38) or "SVT-predominant MPN" may become a novel clinical entity consisting of patients with abdominal venous thrombosis with the mutation JAK2 p.V617F. It has therapeutic and prognostic implications before fulfilling the diagnostic criteria of MPN, for example, continuation of anticoagulation, regular surveillance of blood cell counts, and clinical follow-up. Corresponding to the rapid development and distribution of highly sensitive diagnostic procedures like next-generation ultradeep sequencing patients with VAF < 2% will perhaps be included in the future (39). Meanwhile, a larger cohort of patients can contribute to better characterizing different patient groups with abdominal venous thromboses.

Implications

Patients with abdominal venous thromboses are seen by various medical disciplines.

In a prospective prevalence study in MV, a federal state in Germany, the mutation *JAK2* p.V617F was detected in 19% of patients with abdominal venous thromboses. Clinical or laboratory criteria did not provide evidence for the mutation *JAK2* p.V617F.

Establishing a diagnostic workout for peripheral blood samples in a region can help clarify the etiology of abdominal venous thrombotic events.

Peripheral blood samples are suitable for mutation analysis of *JAK2*. Ultradeep sequencing, as a highly sensitive and quantitative detection method, is advantageous. By using next-generation sequencing, other mutations of interest can be investigated at the same time.

If mutations are detected, further hematological diagnostics should be initiated, and regular follow-up is warranted.

Data availability statement

The data presented in this study are deposited in the sequence read archive (SRA) by the NIH, accession number SUB10682917.

Ethics statement

This study involving humans was approved by the Ethikkommission an der Medizinischen Fakultät der Universitaet Rostock, Rostock, Germany. The study was conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study.

Author contributions

LH: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Project administration, Supervision, Validation, Writing – original draft. LG: Data curation, Formal analysis, Investigation, Methodology, Writing – original draft. SF: Formal analysis, Visualization, Writing – review & editing. MW: Investigation, Writing – review & editing. CGT: Conceptualization, Funding acquisition, Writing – review & editing. CR: Conceptualization, Funding acquisition, Writing – review & editing. HME: Conceptualization, Formal analysis, Funding acquisition, Investigation, Methodology, Supervision, Validation, Writing – original draft. CJ: Resources, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. We thank the society of internal medicine in Mecklenburg West-Pomerania (Gesellschaft der Internisten Mecklenburg-Vorpommern e.V.) for funding the JAK2-MV study by the research grant of the year 2016 (received by LH, CGT, CR, and HME).

References

1. Ageno W, Riva N, Schulman S, Beyer-Westendorf J, Bang SM, Senzolo M et al. Long-term clinical outcomes of splanchnic vein thrombosis: results of an international registry. *JAMA Intern Med.* (2015) 175:1474–80. doi: 10.1001/jamainternmed.2015.3184

2. How J, Zhou A, Oh T. Splanchnic vein thrombosis in myeloproliferative neoplasms: pathophysiology and molecular mechanisms of disease. *Ther Adv Hematol.* (2017) 8:107–18. doi: 10.1177/2040620716680333

3. Mathew C, Zumberg M. Clots in unusual places: lots of stress, limited data, critical decisions. *Hematology Am Soc Hematol Educ Program.* (2021) 2021:92–9. doi: 10.1182/hematology.2021000237

4. Alvarez-Larrán A, Pereira A, Magaz M, Hernández-Boluda JC, Garrote M, Cuevas B et al. Natural history of polycythemia vera and essential thrombocythemia presenting with splanchnic vein thrombosis. *Ann Hematol.* (2020) 99:791–8. doi: 10.1007/s00277-020-03965-z

5. Rajani R, Melin T, Björnsson E, Broomé U, Sangfelt P, Danielsson A et al. Budd-Chiari syndrome in Sweden: epidemiology, clinical characteristics and survival - an 18-year experience. *Liver Int.* (2009) 29:253–9. doi: 10.1111/j.1478-3231.2008.01838.x

6. Acosta S, Alhadad A, Svensson P, Ekberg O. Epidemiology, risk and prognostic factors in mesenteric venous thrombosis. *Br J Surg.* (2008) 95:1245–51. doi: 10.1002/bjs.6319

7. Stefano V de, Vannucchi AM, Ruggeri M, Cervantes F, Alvarez-Larrán A, Iurlo A et al. Splanchnic vein thrombosis in myeloproliferative neoplasms: risk factors for recurrences in a cohort of 181 patients. *Blood Cancer J.* (2016) 6:e493. doi: 10.1038/bcj.2016.103

Acknowledgments

We thank all the patients who participated in the study. We thank the colleagues in Mecklenburg-West Pomerania for their contribution to the JAK2 MV study by including their patients.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The author(s) declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed.2023. 1344769/full#supplementary-material

SUPPLEMENTARY FIGURE 1 Regional distribution of blood samples included in the study (n = 68).

8. Riva N, Beyer-Westendorf J, Contino L, Bucherini E, Sartori MT, Grandone E et al. Treatment of portal, mesenteric, and splenic vein thrombosis with rivaroxaban: a pilot, prospective cohort study. *Blood.* (2021) 138:671. doi: 10.1182/blood-2021-153813

9. Valeriani E, Di Nisio M, Riva N, Cohen O, Garcia-Pagan J-C, Magaz M et al. Anticoagulant therapy for splanchnic vein thrombosis: a systematic review and metaanalysis. *Blood.* (2021) 137:1233–40. doi: 10.1182/blood.2020006827

10. Felser S, Rogahn J, Le Coutre P, Al-Ali HK, Schulze S, Muegge L-O et al. Anxieties, age and motivation influence physical activity in patients with myeloproliferative neoplasms - a multicenter survey from the East German study group for hematology and oncology (OSHO #97). Front Oncol. (2022) 12:1056786. doi: 10.3389/fonc.2022.1056786

11. Praktiknjo M, Meyer C, Strassburg CP, Trebicka J. Frische venöse Thrombose splanchnischer Gefäße: Zwei Fallberichte zur kathetergestützten lokalen Thrombolyse und -aspiration. *Internist.* (2017) 58:82–9. doi: 10.1007/s00108-016-0120-0

12. Abbattista M, Capecchi M, Martinelli I. Treatment of unusual thrombotic manifestations. *Blood.* (2020) 135:326–34. doi: 10.1182/blood.2019000918

13. Colaizzo D, Amitrano L, Guardascione MA, Tiscia GL, D'Andrea G, Longo VAC et al. Outcome of patients with splanchnic venous thrombosis presenting without overt MPN: a role for the JAK2 V617F mutation re-evaluation. *Thromb Res.* (2013) 132:e99–e104. doi: 10.1016/j.thromres.2013.07.014

14. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood.* (2016) 127:2391–405. doi: 10.1182/blood-2016-03-643544

15. Arber DA, Orazi A, Hasserjian RP, Borowitz MJ, Calvo KR, Kvasnicka H-M et al. International consensus classification of myeloid neoplasms and acute leukemias: integrating morphologic, clinical, and genomic data. *Blood.* (2022) 140:1200–28. doi: 10.1182/blood.2022015850

16. Demiriz IS, Kazanci MH, Menfaatli E, Jafari-Gharabaghlou D, Zarghami N. Allelic burden of Janus kinase 2 in a 6-month course of therapy for myeloproliferative neoplasms. *Mol Biol Rep.* (2023) 50:5687–95. doi: 10.1007/s11033-023-08511-4

17. Etheridge SL, Roh ME, Cosgrove ME, Sangkhae V, Fox NE, Chen J et al. JAK2V617F-positive endothelial cells contribute to clotting abnormalities in myeloproliferative neoplasms. *Proc Natl Acad Sci U S A*. (2014) 111:2295–300. doi: 10.1073/pnas.1312148111

18. Torres DG, Paes J, da Costa AG, Malheiro A, Silva GV, Mourão LPdS et al. JAK2 variant signaling: genetic, hematologic and immune implication in chronic myeloproliferative neoplasms. *Biomolecules.* (2022) 12:291. doi: 10.3390/biom12020291

19. Vainchenker W, Kralovics R. Genetic basis and molecular pathophysiology of classical myeloproliferative neoplasms. *Blood.* (2017) 129:667-79. doi: 10.1182/blood-2016-10-695940

20. Falchi L, Kantarjian HM, Verstovsek S. Assessing the thrombotic risk of patients with essential thrombocythemia in the genomic era. *Leukemia.* (2017) 31:1845–54. doi: 10.1038/leu.2017.150

21. Cazzola M, Kralovics R. From Janus kinase 2 to calreticulin: the clinically relevant genomic landscape of myeloproliferative neoplasms. *Blood.* (2014) 123:3714–9. doi: 10.1182/blood-2014-03-530865

22. Colaizzo D, Amitrano L, Tiscia GL, Scenna G, Grandone E, Guardascione MA, et al. The JAK2 V617F mutation frequently occurs in patients with portal and mesenteric venous thrombosis. *J Thromb Haemost.* (2007) 5:55-61. doi: 10.1111/j.1538-7836.2006.02277.x

23. Grunwald L, Grosse-Thie C, Sender S, Knuebel G, Krohn S, Roolf C et al. Ultradeep targeted sequencing reveals low allele frequencies of somatic JAK2 and MPL variants in patients with abdominal vein thromboses: results of an ongoing prospective prevalence study in Mecklenburg-West Pomerania. *Biomark Res.* (2020) 8:73. doi: 10.1186/s40364-020-00254-9

24. Maslah N, Verger E, Schlageter M-H, Miclea J-M, Kiladjian J-J, Giraudier S et al. Next-generation sequencing for JAK2 mutation testing: advantages and pitfalls. *Ann Hematol.* (2019) 98:111–8. doi: 10.1007/s00277-018-3499-y

25. Cattaneo D, Bucelli C, Marchetti A, Lionetti M, Fermo E, Bellani V et al. Pathological and genomic features of myeloproliferative neoplasms associated with splanchnic vein thrombosis in a single-center cohort. *Ann Hematol.* (2023) 102:1409–20. doi: 10.1007/s00277-023-05217-2

26. Colaizzo D, Amitrano L, Guardascione MA, Favuzzi G, Tiscia GL, D'Andrea G et al. Clinical utility of screening for CALR gene exon 9 mutations in patients with splanchnic venous thrombosis. *Thromb Haemost.* (2015) 113:1381–2. doi: 10.1160/TH14-12-1055

27. Debureaux P-E, Cassinat B, Soret-Dulphy J, Mora B, Verger E, Maslah N et al. Molecular profiling and risk classification of patients with myeloproliferative neoplasms and splanchnic vein thromboses. *Blood Advances.* (2020) 4:3708–15. doi: 10.1182/bloodadvances.2020002414

28. Wolach O, Sellar RS, Martinod K, Cherpokova D, McConkey M, Chappell RJ et al. Increased neutrophil extracellular trap formation promotes thrombosis in myeloproliferative neoplasms. *Sci Transl Med.* (2018) 10:8292. doi: 10.1126/scitranslmed.aan8292

29. DeZern AE, Malcovati L, Ebert BL. CHIP, CCUS, and other acronyms: definition, implications, and impact on practice. *Am Soc Clin Oncol Educ Book*. (2019) 39:400–10. doi: 10.1200/EDBK_239083

30. Edelmann B, Gupta N, Schnoeder TM, Oelschlegel AM, Shahzad K, Goldschmidt J et al. JAK2-V617F promotes venous thrombosis through β 1/ β 2 integrin activation. *Journal of Clin Inv.* (2018) 128:4359–71. doi: 10.1172/JCI 90312

31. Eaton N, Subramaniam S, Schulte ML, Drew C, Jakab D, Haberichter SL et al. Bleeding diathesis in mice lacking JAK2 in platelets. *Blood Advances.* (2021) 5:2969–81. doi: 10.1182/bloodadvances.2020003032

32. Ageno W, Beyer Westendorf J, Contino L, Bucherini E, Sartori MT, Senzolo M et al. Rivaroxaban for the treatment of noncirrhotic splanchnic vein thrombosis: an interventional prospective cohort study. *Blood Adv.* (2022) 6:3569-78. doi: 10.1182/bloodadvances.2022007397

33. How J, Trinkaus KM, Oh ST. Distinct clinical, laboratory and molecular features of myeloproliferative neoplasm patients with splanchnic vein thrombosis. *Br J Haematol.* (2018) 183:310–3. doi: 10.1111/bjh.14958

34. Klute K, DeFilippis EM, Shillingford K, Chapin J, DeSancho MT. Clinical presentations, risk factors, treatment and outcomes in patients with splanchnic vein thrombosis: a single-center experience. *J Thromb Thrombolysis*. (2016) 42:267–71. doi: 10.1007/s11239-016-1337-4

35. Statistiken zu Mecklenburg-Vorpommern. (2023) Available online at: https://de. statista.com/themen/2450/mecklenburg-vorpommern/#editorsPicks (accessed August 5, 2023).

36. Gondek LP. CHIP is clonal hematopoiesis a surrogate for aging and other disease? *Hematology Am Soc Hematol Educ Program.* (2021) 2021:384–9. doi: 10.1182/hematology.2021000270

37. Baumeister J, Chatain N, Sofias AM, Lammers T, Koschmieder S. Progression of myeloproliferative neoplasms (MPN): diagnostic and therapeutic perspectives. *Cells.* (2021) 10:551. doi: 10.3390/cells10123551

38. Camerlo S, Ligato J, Rosati G, Carrà G, Russo I, Gobbi M de et al. Shedding light on the pathogenesis of splanchnic vein thrombosis. *Int J Mol Sci.* (2023) 24:262. doi: 10.3390/ijms24032262

39. Luque Paz D, Kralovics R, Skoda RC. Genetic basis and molecular profiling in myeloproliferative neoplasms. *Blood.* (2023) 141:1909–21. doi: 10.1182/blood.2022017578