Check for updates

OPEN ACCESS

EDITED BY Diogo Alpuim Costa, CUF Oncologia, Portugal

REVIEWED BY Zarema Albakova, Lomonosov Moscow State University, Russia Zizhao Yang, Zhejiang University, China Yusuf Tutar,

University of Health Sciences, Türkiye *CORRESPONDENCE

Bo Tang ⊠ tangbocq@vip.sina.com Hao Zhang ⊠ syzhanghao@zju.edu.cn

[†]These authors have contributed equally to this work

RECEIVED 11 October 2023 ACCEPTED 12 December 2023 PUBLISHED 09 January 2024

CITATION

Zhu J, Dai Y, Tang B and Zhang H (2024) The association between serum heat shock protein 72 and intestinal permeability with intestinal microbiota and clinical severity in patients with cerebral infarction. *Front. Med.* 10:1302460. doi: 10.3389/fmed.2023.1302460

COPYRIGHT

© 2024 Zhu, Dai, Tang and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The association between serum heat shock protein 72 and intestinal permeability with intestinal microbiota and clinical severity in patients with cerebral infarction

Jiahui Zhu[†], Yijie Dai[†], Bo Tang* and Hao Zhang*

Department of Neurology, Hangzhou First People's Hospital, Hangzhou, China

Objectives: We aimed to compare serum heat shock protein 72 (HSP72) and intestinal permeability in patients with cerebral infarction (CI) and healthy individuals to reveal their correlations and link to gut microbiota alterations and clinical severity of CI.

Methods and results: Stool samples of 50 patients with CI and 46 healthy volunteers were analyzed through 16S rRNA gene sequencing to characterize intestinal flora profiles. Serum HSP72 and zonulin were assayed using enzyme-linked immunoassay (ELISA). The obtained data were then subjected to comparative and correlative analysis. We found that the levels of zonulin and serum HSP72 were significantly higher in the CI group compared to the healthy group. Serum HSP72 and zonulin levels were positively correlated in the CI group and correlated positively with the clinical severity of CI. β diversity showed significant differences in intestinal microbiota composition between the two groups. In the CI patient group, the abundance of bacteria *Eubacterium_fissicatena_group*, *Eubacterium_eligens_group*, and *Romboutsia* manifested a remarkably positive correlation with serum HSP72. The abundance of bacteria *Eubacterium_fissicatena_group* and *Acetivibrio* had a significantly positive correlation with zonulin levels.

Conclusion: Our findings indicated that an increase in serum HSP72 and zonulin levels was manifested in patients with CI and was related to specific gut microbiota alterations and the clinical severity of CI.

KEYWORDS

intestinal flora, cerebral infarction, serum heat shock protein 72, intestinal permeability, zonulin, clinical severity of CI, ELISA, NIHSS

Introduction

Stroke, a devastating and lethal disease, is currently second among the leading causes of death globally and the third contributor to disability in the world (1). Besides the neurological defects caused by the infarct site, the major cause of death is the peripheral tissue damage induced by post-stroke leaky gut. Previous reports have shown that the abundance of opportunistic pathogens and the corresponding product of metabolism changes increased

significantly in patients with cerebral infarction (CI) compared to healthy controls (2). Incidentally, elevated inflammatory levels after CI efficiently facilitated the increase of intestinal mucosal permeability. Strict regulation of intestinal integrity is essentially related to host antimicrobial immune defense. Regrettably, previous studies on CI and specific alterations of intestinal flora have not effectively obtained explicit conclusions.

The level of the serum zonulin, a protein produced by the intestine epithelium, is proportional to gut permeability. *In vitro* studies demonstrated that endogenous human zonulin was responsible for increased permeability in the jejunum and ileum (3). An overgrowth of intraluminal microorganism and gluten contribute significantly to intestinal zonulin release (4). Zonulin secretion has been reported to be induced through the MyD88-dependent pathway, which is followed by dissociation of the protein ZO-1 from the tight junctional complex, thus generating leaky gut syndrome (5). Of note, zonulin is the biological door to a vast number of diseases such as autoimmune diseases, neuropsychiatric diseases, and cancer through its intestinal barrier regulation function (6, 7).

Heat shock protein 72 (HSP72), a critical component of inducible HSPs (iHSPs), is a ubiquitous molecule that exerts efficient effects on cellular survivability and tolerance to stressors. Correspondingly, animal experiments showed that the upregulation of HSP72 in ischemic cerebral tissue confirmed a crucial protective role in the outcome following stroke (8). Recent evidence indicates that bacterial components and metabolites specifically control the expression of HSPs (9, 10). HSP72 can be detected in quite a few body fluids, such as pleural fluid (11), cerebrospinal (12), synovia (12), bronchoalveolar lavage fluid (13), and serum (14). Recent evidence indicated that the concentration of the stress-inducible HSP72 homolog, HSP70, increased in patients with heart failure (15) and atherosclerosis (16). Strikingly, previous studies have documented that the serum expression level of HSP70 significantly increased following acute middle cerebral artery occlusion (MCAO) in rats (16). Nevertheless, whether there is a change in human serum HSP72 after CI has not been addressed. Thereafter, how serum HSP72 correlates with intestinal dysfunction and CI severity has not been accurately explored.

Accordingly, in this study, we deciphered gut microbiome profiles through 16S rRNA gene sequencing of stool samples, quantified serum HSP72 and zonulin via enzyme-linked immunoassay (ELISA), further compared and analyzed the levels of serum HSP72 and zonulin in patients with CI and healthy controls, and explored the link between the levels of serum HSP72 and zonulin to alterations in intestinal microbiota and clinical severity of CI.

Materials and methods

Study subjects

This cross-sectional study was conducted at Hangzhou First People's Hospital. We consecutively enrolled 81 patients with CI and 52 healthy individuals, respectively, from the Department of Neurology and Physical Examination Center. Herein, we rigorously screened out 50 patients with CI and 50 healthy controls who satisfied the inclusion criteria and ultimately participated in the study. Case groups were diagnosed within 2 weeks after sudden focal neurological deficits, with the acute infarct area in the corresponding brain region confirmed on the computed tomography (CT) and/or magnetic resonance imaging (MRI). Healthy controls reported never being diagnosed with any of the risk factors associated with CI, such as hypertension, diabetes, metabolic syndrome, and atrial fibrillation. Baseline characteristics such as demographic data including age, sex, dietary habits, medical histories (hypertension, diabetes, and atrial fibrillation), and laboratory data were collected. This study was approved by the Clinical Research Ethics Committee of Hangzhou First People's Hospital.

The inclusion criteria for the study subjects were determined as (1) aged between 40 and 80 years old; (2) grew up in southern China and maintained a healthy lifestyle, with healthy dietary and bowel habits; (3) body mass index range from 18 to 24; (4) had available blood and stool samples; and (5) signed informed consent before the experiments. Subjects were excluded if they (1) had a history of intracranial hemorrhage and other neuropsychiatric disorders; (2) were previously diagnosed with respiratory failure, heart failure, uremia, severe liver dysfunction, malignant tumors, and autoimmune disease; (3) had inflammatory bowel disease, gastrointestinal bleeding, and surgery, as well as other gastrointestinal dysfunction; and (4) had received treatment with antibiotics, probiotics, or hormone drugs within 2 months before the recruitment.

Sample collection

To avoid random error, fecal samples were taken with cotton swabs from the middle section of the stool samples provided and were kept in two 2 mL sterile frozen depository tubes with approximately 200 mg of samples in each tube. Fasting blood samples were collected in the coagulation vessels and centrifuged at 3000r for 10 min at -4° C. The serum was transferred to 1.5 mL frozen tubes. Preprocessed stool and serum samples were immediately transferred to the laboratory for storage at -80° C.

Microbial DNA extraction and sequence data analysis

The total DNA of fecal specimens was extracted using the E.Z.N.A.[®]Stool DNA Kit and samples were sequenced on the Illumina NovaSeq platform following the manufacturer's instructions. The complexity of the sample species diversity was characteristic of α diversity, calculated by the QIIME2. The β diversity was calculated by non-metric multidimensional scaling and plotted by the R package. Composition difference was analyzed by the Wilcoxon rank-sum test and Welch's t-test, based on which we constructed the heat map at the phyla level.

Quantitative assay of serum HSP72 and zonulin by ELISA

Serum HSP72 and zonulin were assayed through the Human HSP72 ELISA Kit and Zonulin ELISA Kit (Meimian Industrial, Jiangsu, China). A 50μ L proof sample was added to the standard well.

A 40 μ L sample diluent and a 10 μ L sample being tested were added to the enzymatic coating plate so that samples were eventually diluted to 5-fold. Samples were incubated with 100 μ L HRP-Conjugate reagent at 37°C for 60 min. Then, each well was filled with wash fluid, and the fluid was discarded after 30 s. This process was repeated five times, and each well was dried. Each well was incubated with chromogenic agent at 37°C for 15 min and then the reaction was terminated by adding 50 μ L of the stop solution to each well until the blue color turned yellow. Finally, the absorbance of each well was determined within the wavelength of 450 nm.

Statistical analysis

Data were analyzed using SPSS version 26.0 software and R version 3.6.1 statistical software. The continuous variable was denoted by mean ± SD, while categorical variables were denoted by numbers and percentages. Measurement data meeting normality was indicated by independent sample Student's t-test or analysis of variance and by Wilcoxon's rank sum test if the variable violated the assumption of normality. Categorical variable differences between groups were compared by a χ^2 test. We used binary logistic regression analysis to explore the relationship between serum HSP72 and zonulin levels and the occurrence of CI, after adjustment for age and sex. The area under the receiver operating characteristic (ROC) curve (AUC) was applied to evaluate the model's diagnostic performance for investigating the ability of serum HSP72 and zonulin levels to distinguish between patients with CI and controls. Multivariable linear regression models were conducted to examine the association between serum HSP72 and zonulin levels and clinical severity. The correlation between intestinal flora and clinical data was assessed by Spearman's rank correlation analysis. The data correlation conforming to the normal distribution was analyzed with the Pearson test. p-value <0.05 was defined as statistically significant.

Results

Study population and baseline characteristics

A total of 81 patients with CI and 52 healthy individuals were enrolled in this study. However, 31 patients and two control participants failed to defecate on time or provided unqualified fecal sampling, and were thus excluded from the subsequent analysis. Four healthy samples could not meet the 16S rRNA sequencing requirements due to low fecal volume, which led to unqualified amplification. Ultimately, only 96 subjects (50 patients with CI and 46 controls) remained in the next 16S rRNA analysis, and the final included samples were tested by ELISA. The National Institute of Health Stroke Scale (NIHSS) score was less than 16 in the CI group, for which the patients with severe CI had difficult fecal discharge the early next morning. All participants in the experiment were matched for age (CI group, 66.42 ± 6.92 ; healthy group, 65.15 ± 5.82 ; p = 0.233) and sex (M/F: case, 26/24; control, 25/21; p = 0.818). The demographic and baseline clinical characteristics of patients with CI and controls are shown in Table 1.

Comparison of serum HSP72 and zonulin levels between the two groups

The quantification and comparison of serum HSP72 and zonulin between the two groups are shown in Table 1 and Figure 1. The mean serum HSP72 level was $307.46 \pm 42.59 \text{ pg./g}$ in patients with CI, while the mean value was $176.61 \pm 44.75 \text{ pg./g}$ in controls. In light of this, patients with CI had significantly higher serum HSP72 than controls (p < 0.01). Accordingly, the mean zonulin level was $135.70 \pm 30.74 \text{ ng/mL}$ in patients with CI, while the mean value was $105.83 \pm 24.04 \text{ ng/mL}$

Clinical parameter	Cl group (<i>n</i> = 50)	Healthy group (n = 46)	<i>p</i> -value
Sex, male	26 (52%)	25 (54%)	0.818
Age, y	66.42 ± 6.92	65.15 ± 5.82	0.233
NIHSS score	10.88 ± 2.67	/	/
Hypertension (n, %)	36 (72%)	0	< 0.01**
Diabetes (n, %)	25 (50%)	0	< 0.01**
Atrial fibrillation (<i>n</i> , %)	3 (6%)	0	< 0.01**
Smoking history (<i>n</i> , %)	9 (18%)	6 (13%)	0.504
TG (mmol/L)	1.53 ± 0.76	1.21 ± 0.60	0.013*
TC (mmol/L)	4.21 ± 0.97	2.64 ± 0.68	< 0.01**
LDL (mmol/L)	2.32 ± 0.84	2.43 ± 0.54	0.203
HCY (umol/L)	16.55 ± 10.90	8.81±3.03	< 0.01**
UA (umol/L)	314.28±89.72	350.30±42.75	0.016*
Serum HSP72 (pg/g)	307.46±42.59	176.61 ± 44.75	< 0.01**
Zonulin (ng/ml)	135.70 ± 30.74	105.83 ± 24.04	< 0.01**

TG, triglyceride; TC, total cholesterol; LDL, low-density lipoprotein; HCY, homocysteine; UA, uric acid; HSP72, heat shock protein 72. Variables are expressed as mean \pm SD or n (%). *p < 0.05. **p < 0.01.



Violin plot representing data density distribution and median with interquartile range of serum HSP72 (A) and zonulin (B) between the two groups. **, p < 0.01.

mL in controls. Moreover, serum zonulin was significantly higher in the CI group than in healthy controls (p < 0.01).

We then investigated whether serum HSP72 or zonulin distinguished between patients with CI and healthy individuals. Binary logistic regression analysis indicated that higher levels of serum HSP72 (OR = 1.10, 95% CI = 1.03-1.11; p < 0.01) and zonulin (OR = 1.04, 95% CI = 1.02-1.07; p < 0.01) were more likely to associated with CI patients after adjustment for sex and age.

According to binary logistic analysis, ROC curve analysis was performed to predict CI occurrence. Strikingly, the AUC, which represented the prediction precision, raised from 0.57 in the basic model (age + sex, p = 0.22) to 0.98 in a way that added serum HSP72 levels, which discriminated clearly between the CI and healthy groups in the logistic regression analysis (p < 0.01). The AUC was 0.79 with the addition of the serum level of zonulin (p < 0.01), which also discriminated markedly between the CI and healthy groups in the logistic regression analysis. Furthermore, when bringing age, sex, serum HSP72, and zonulin into the full model, the AUC reached 0.98 (p < 0.01) (Figure 2A; Table 2).

Association between serum HSP72 and zonulin levels and the link to CI severity

Based on Pearson correlation analysis, we further observed that serum HSP72 ($\rho = 0.93$, p < 0.01) and zonulin ($\rho = 0.97$, p < 0.01) positively correlated with NIHSS scores (Figures 2B,C). Of note, we found a positive correlation between the levels of serum HSP72 and zonulin within the CI group ($\rho = 0.87, p < 0.01$) (Figure 2D). Similarly, after adjustment for age, sex, smoking history, previous related diseases, and routine blood test results in bias correlation analysis, serum HSP72 ($\rho = 0.93$, p < 0.01) and zonulin ($\rho = 0.97$, p < 0.01) levels continuously correlated positively with NIHSS scores in patients with CI. With the multivariate linear regression model, we found that serum HSP72 (b = 0.06, t = 14.37, p < 0.01) and zonulin (b = 0.02, t = 7.16, t)p < 0.01) were persistently positively correlated with NIHSS scores. These data demonstrated that higher levels of serum HSP72 and zonulin were associated with a more severe degree of CI.



FIGURE 2

(A) ROC curves for estimating the accuracy of cerebral infarction (CI) using basic model (age + sex, AUC = 0.57, p = 0.22) raised with the addition of serum HSP72 and zonulin (AUC = 0.98, p < 0.01; AUC = 0.79, p < 0.01). In a full model comprising age, sex, serum HSP72, and zonulin, the accuracy of evaluating the occurrence of CI showed significant differences between CI and healthy individuals (AUC = 0.98, p < 0.01). (B,C) Scatter diagram manifested positive correlations between NIHSS scores and serum HSP72 (B) and zonulin (C). (D) Scatter diagram showed correlations between serum HSP72 and zonulin. ROC = receiving operating characteristic. AUC = area under the curve.

Intestinal flora correlated with serum HSP72 and zonulin

 α diversity was used to describe the species diversity of individual samples. The Wilcoxon rank-sum test, depicted through Chao 1, Good's coverage, Simpson, and Shannon indexes, showed no statistical

TABLE 2 Receiver operating characteristic association statistics for forecasting cerebral infarction.

	AUC (95% CI)	<i>p</i> -value
Basic (age, sex)	0.57 (0.46–0.69)	0.22
Basic + serum HSP72	0.98 (0.96–1.00)	< 0.01**
Basic + zonulin	0.79 (0.70-0.88)	< 0.01**
Full model (age, sex, serum HSP72 and zonulin)	0.98 (0.95–1.00)	< 0.01**

AUC= area under the receiver operating characteristic curve; CI = confidence interval. **p < 0.01.

difference in species richness and evenness between the two groups (Figure 3A). β diversity is an index reflecting the difference in composition and distribution of bacteria between groups. NMDS plots showed significant differences in intestinal microbiota composition between the two groups (unweighted Unifrac stress=0.17 and weighted UniFrac stress=0.15) (Figure 3B). We then analyzed the differences in colony abundance at the 67 genus level (Figure 3C). Accordingly, the CI group was more abundant with 27 features, while the healthy group was enriched with 40 features. These data indicated that the microbial abundance in the CI group was much lower than that in the healthy group.

We further performed the Spearman correlation analysis on the differentially abundant bacteria genus with serum HSP72 and zonulin. We found significant correlations between the bacteria genus with both serum HSP72 and zonulin. Among the correlation heatmap for CI participants, the abundance of bacteria *Eubacterium_fissicatena_* group, Eubacterium_eligens_group, and Romboutsia manifested a remarkably positive correlation with serum HSP72 (ρ =0.28, p=0.04; ρ =0.30, p=0.03; ρ =0.28, p=0.04). The abundance of bacteria *Eubacterium_fissicatena_group* and *Acetivibrio* had a significantly





positive correlation with zonulin levels ($\rho = 0.34$, p = 0.02; $\rho = 0.28$, p = 0.04) (Figure 4). Of note, the CI group was characterized by a decreased abundance of *Eubacterium_fissicatena_group*, *Eubacterium_eligens_group*, and *Romboutsia* and an increased abundance of *Acetivibrio* (p < 0.05).

Discussion

The present study is the first to elucidate how both serum HSP72 and zonulin levels interrelate to alterations in intestinal microbiota and CI symptom severity in patients compared with healthy controls. After adjustment for confounders, we subsequently demonstrated that an increase in serum HSP72 and zonulin was associated with the symptom severity of CI and gut microbiota alterations.

Accordingly, we demonstrated higher levels of serum HSP72 in patients with CI than in healthy controls. Notably, the levels of serum HSP72 were positively associated with CI clinical severity. Intracellular HSP72 distributes in nearly every cell of the body and provides multiple cell survival functions such as restricting protein aggregation, promoting protein refolding, and acting as protein chaperoning. Current research pays attention to confirming intra-cellular HSP72 may confer an immunostimulatory effect that, on the one hand, facilitates innate immune responses against acute pathogenic substances, whereas, on the other hand, serum HSP72 accelerates the

inflammatory process in people with various diseases such as hypertension and atherosclerosis (17). As a danger signal, circulation HSP72 is capable of specifically stimulating NO, TNF- α , IL-10, and IL-6 secretion from macrophages and neutrophils (18). It is noteworthy that HSP70 interacts with microglia and macrophages through toll-like receptors (TLRs) (17). The combination of HSP70 and TLRs induces the expression of genes encoding inflammationassociated molecules and cytokines through the activation of transcription factors activator protein 1 (AP-1) and nuclear factor kB (NF-KB) (19). In this connection, theoretically, HSP72 levels are related to the severity of inflammation and stress in organisms, which explains the higher levels of serum HSP72 in patients with severe CI. Previous research confirmed that overexpressing HSP72 facilitated a satisfactory prognosis of cerebral ischemia-reperfusion injury, which may be attributed to c-Jun N-terminal kinase 3 signaling pathway inhibition and Akt1 activation (20). George et al. found an association between HSP72 overexpression and reduced reactive astrocytes after stroke, which may contribute to neuroprotection (21). Moreover, Shailaja et al. suggested that HSP72 knockdown significantly upregulates apoptosis-inducing factors and ROS levels in both anoxia/reoxygenation (22). Recent work by Xu et al. (8) demonstrated that the upregulation of HSP72 in ischemic cerebral tissue has a crucial protective role in the outcome of stroke. In addition, it is noteworthy that endogenous HSP72 is released through necrotic or lytic cell death and activation of the α-adrenergic receptor pathway in response to environmental stressors in ways that can be detected in blood and excreta (23). We speculated that the elevation of the serum HSP72 level after CI mainly resulted from two leading causes. The first is that intracranial ischemic death cells rupture and release HSP72 to peripheral blood circulation. The other is that the sympathetic adrenal axis activated after CI triggers a large amount of HSP72 released from tissue cells and intracranial inflammatory cells to alleviate brain inflammation and oxidative damage. Recent evidence revealed that high levels of serum HSP72 were usually associated with enhanced oxidative profiles and ascending rates of mortality among septic patients (24). To this point, our results suggested that HSP72 may play a role in the deterioration of CI.

We observed that patients with CI had higher serum concentrations of zonulin than controls. Zonulin levels correlated with the degree of neurological deficit after CI. Our findings support previous studies that indicated that blood levels of zonulin are elevated in neuroinflammatory diseases, such as stroke, severe traumatic brain injury, neurodegenerative diseases, and psychiatric disorders (25). Produced by small intestine epithelium, the blood zonulin level becomes a reliable indicator reflecting intestinal permeability and inflammatory response level in vivo. Intriguingly, it is worth mentioning that intestinal epithelium plays an essential role in triggering the pathogenesis of numerous inflammatory diseases, and zonulin levels are provoked under acute stress states. Apart from the direct neural pathway connecting the intestine and the brain, it has long been appreciated that post-stroke dysbiosis gives rise to a downmodulated biosynthesis of $\gamma\delta T$ lymphocytes that directly impairs the immune system stability. It is worth mentioning that gut microbiota dysbiosis not only induces intestinal homeostasis damage but also stimulates the migration of $\gamma\delta T$ lymphocytes from the intestinal tract to the brain (26). Bacterial products and cellular components derived from the intestinal flora have an impact on the prognosis of patients with CI. Lipopolysaccharide (LPS) released by gut microbes after CI binds to TLRs and then activates the TLR4/P13K/Akt/MAPKs pathway, which sets the stage for matrix metalloproteinase 9 (MMP-9) expression in astrocytes and astrocytes migration and leads to intestinal leakage (27). Trimethylamine oxide (TMAO), a component of microbiota metabolite, is positively correlated with infarct size and severity of CI (28). Inflammatory signaling pathways mediated by TMAO involved NF-κB, pyrin dome-containing protein 3 (NLRP3) inflammasome, and the MAPK/JNK pathway in the peripheral and central nervous systems (29). Alternatively, the release of zonulin triggered by intestinal flora imbalance induces antigen influx from the intestinal lumen to the lamina propria and further exacerbates immune response, causing IFN- γ and TNF- α release (4, 30). Additionally, zonulin secretion was regarded as MyD88-dependent followed by protein ZO-1 dissociation from the tight junctional complex, which was responsible for both intestinal and extraintestinal inflammation, autoimmunity, and cancer (5, 6). Accordingly, the alterations of zonulin may be attributed to post-stroke dysbiosis and neuro-humoral mechanisms.

Moreover, we further observed the positive correlation between the levels of serum HSP72 and zonulin. A compromise of the intestinal mucosa is a result of increased severity and duration of stress and insufficient endogenous protective factors. Previous research has given insight into the endogenous protective mechanism of HSP72 within enterocytes. In 1999, an *in vitro* cell study conducted by Musch et al. (31) supported that HSP72 played a pivotal role in the integrity of the actin cytoskeleton and maintenance of epithelial barrier function under oxidant-induced stress. It remains a mystery whether serum HSP72 could be a reliable indicator reflecting intestinal permeability. We speculated that serum HSP72 may act on the brain-gut axis accelerating intestinal barrier destruction.

In the present study, the 16s rRNA sequencing results demonstrated that CI was associated with certain transformations in fecal bacteria. Consistent with the findings of previous publications, post-stroke dysbiosis was characterized by reduced diversity, decreased abundance of protective bacteria, and harmful bacterial overgrowth. Intestinal dysbiosis is effectively linked to several risk factors for stroke, such as diabetes, hypertension, and atherosclerosis, and also to stroke outcomes. However, previous studies on CI and specific alterations of intestinal flora failed to obtain unanimous and definite conclusions, which may result from different patient races, different DNA detection methods used by researchers, and different patient dietary habits.

Furthermore, we discovered that the levels of serum HSP72 and zonulin in patients with CI were correlated with the relative abundance of specific differential microbial genera. The abundance of bacteria Eubacterium_fissicatena_group, Eubacterium_eligens_group, and Romboutsia manifested a remarkably positive correlation with serum HSP72. The abundance of bacteria Eubacterium_fissicatena_group and Acetivibrio had a significantly positive correlation with zonulin levels. Indeed, we noted the CI group was associated with a significant decrease in the abundance of E. fissicatena group and E. eligens group. The genus Eubacterium, belonging to the bacterial phylum Firmicutes, has been identified to contribute to massive aspects of human health, for the majority of the family produce short-chain fatty acids (SCFAs), especially butyric acid. It is acknowledged and accepted that SCFAs act as a special nutrient and energy component of the intestinal epithelium, protect the intestinal mucosal barrier, and reduce inflammation levels in the body. Of note, Eubacterium has been shown

to detoxify toxic compounds into more benign forms in the intestine. Understandably, it has been reported recently that the reduction or absence of Eubacterium is associated with many diseases, such as depression, obesity, inflammatory bowel disease, type 2 diabetes, cardiovascular disease, and autism (32). However, the functional annotation of E. fissicatena group and E. eligens group remains poorly understood, partly because both of the bacterial species are rarely detected in feces in previous studies. Despite the protective nature of most members of the genus Eubacterium, recent evidence revealed that E. fissicatena group belongs to potentially disease-related bacteria that add to the risk of intestinal inflammation and metabolic disorders (33). Jing et al. reported that the increased abundance of E. fissicatena group had a positive correlation with serum TMAO levels, which was one of the independent risk factors of acute coronary syndrome (34). Nevertheless, E. fissicatena group was also regarded as butyrateproducing bacteria and beneficial bacteria suppressing intestinal inflammation (35). Another associated genus E. eligens group has been widely acknowledged to exhibit its probiotic effects. Using metagenomic analysis to estimate the gut microbiome profile in atherosclerosis patients, Sheng et al. revealed that the abundance of E. eligens group was positively correlated with propionate and butyrate production but was negatively correlated with inflammatory marker high-sensitivity C-reactive protein and visceral fat area. Similarly, E. eligens group played vital roles in the pathway CDP-diacylglycerol biosynthesis and was also significantly correlated with higher highdensity lipoprotein-cholesterol levels, which significantly modulate the lipid metabolism (36). Afterward, in vitro cell-based assays found that E. eligens group efficiently promoted the production of the antiinflammatory cytokine IL-10, suggesting the potential to be a therapeutic target for inflammatory diseases (37). In general, it seems reasonable in our results that E. fissicatena and E. eligens group, combined with serum HSP72 and zonulin, have the potential to be involved in the post-stroke systemic inflammatory response.

Our research also added to previous reports that the CI group has an increased abundance of genus Acetivibrio, which manifested a positive correlation with zonulin levels. The genus Acetivibrio was equipped with efficient biological machinery transferring lignocellulose into ethanol and has been known to ferment carbohydrates to produce acetic acid (38). Yuan et al. (39) provided evidence that A. ethanolgignens group played a pivotal role in facilitating inflammation and lipid metabolism abnormalities as well as interfering with the energy supply process of the tricarboxylic acid cycle. Normal peristalsis, digestive, and absorption functions of the gut require a series of coordinated operations of intestinal cells. Intestinal flora disorders and systemic inflammation will allow for intestinal barrier disruption and the invasion of harmful substances into circulation. We speculated that Acetivibrio accelerated increased intestinal permeability through the induction of metabolic disturbance and energy intake difficulty of intestinal cells.

Genus *Romboutsia* are SCFA producers and immunomodulators in the gut, which act in the maintenance of intestinal barrier integrity. Our results revealed that a significantly increased abundance of *Romboutsia* genus was observed in the healthy group as compared to the CI group, and the higher the abundance of *Romboutsia* genus, the higher the levels of serum HSP72. In earlier studies, Gerritsen et al. (40) showed that, as a dominant taxon in the small intestine of rats, *Romboutsia* displayed a restricted capacity to synthesize amino acids and vitamins, whereas it was adept at the utilization of different relatively simple carbohydrates (40). Intriguingly, *Romboutsia* has the potential to engage in obesity-related metabolic abnormalities. Previous studies conducted by Zeng et al. (41) indicated that *Romboutsia* was positively associated with body weight, serum lipids, and UA. We, therefore, speculated that the decreased abundance of *Romboutsia* could be an indicator of post-stroke dysbiosis and that the genus *Romboutsia* may have something to do with serum HSP72 levels and post-stroke immunomodulatory effects.

Taken together, our findings proposed that an increase in serum HSP72 and zonulin was observed in patients with CI. It has to be emphasized that the levels of serum HSP72 and zonulin were related to the clinical severity of CI and specific gut microbiota alterations. Our present study has some limitations. First, it was a single-center cross-sectional study with inevitable time and place biases. Second, considering timely bowel movements, the NIHSS scores of CI patients enrolled in this study did not reach more than 15, which made it impossible to assess the relationship between extremely severe CI and the levels of serum HSP72 and zonulin. Finally, a considerable part of patients in the CI group were accompanied by different coexisting diseases that may have affected the results. In general, our results provided promising research prospects that the levels of serum HSP72 and zonulin have the potential to serve as prospective markers for distinguishing patients with CI from controls and mirroring disease severity. Further investigation is required to explore the definitive mechanisms of how serum HSP72 and zonulin act on the process of post-stroke systemic inflammation and intestinal dysbiosis.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving humans were approved by the studies involving human participants were reviewed and approved by the Affiliated Hangzhou First People's Hospital, Zhejiang University School of Medicine. The patients/participants provided their written informed consent to participate in this study. The studies were conducted in accordance with the local legislation and institutional requirements. The human samples used in this study were acquired from a by-product of routine care or industry. Written informed consent for participation was not required from the participants or the participants' legal guardians/next of kin in accordance with the national legislation and institutional requirements.

Author contributions

JZ: Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration,

Resources, Writing – original draft. YD: Writing – original draft. BT: Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – review & editing. HZ: Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. The work was supported by Zhejiang Provincial Medical and Health Technology Project for Young Backbone Talents (grant no. 2019RC234).

References

1. GBD 2019 Stroke Collaborators. Global, regional, and national burden of stroke and its risk factors, 1990-2019: a systematic analysis for the global burden of disease study 2019. *Lancet Neurol.* (2021) 20:795–820. doi: 10.1016/S1474-4422(21)00252-0

2. Tan C, Wu Q, Wang H, Gao X, Xu R, Cui Z, et al. Dysbiosis of gut microbiota and short-chain fatty acids in acute ischemic stroke and the subsequent risk for poor functional outcomes. *JPEN J Parenter Enteral Nutr.* (2021) 45:518–29. doi: 10.1002/ jpen.1861

3. Wang W, Uzzau S, Goldblum SE, Fasano A. Human zonulin, a potential modulator of intestinal tight junctions. J Cell Sci. (2000) 113:4435–40. doi: 10.1242/jcs.113.24.4435

4. Sturgeon C, Fasano A. Zonulin, a regulator of epithelial and endothelial barrier functions, and its involvement in chronic inflammatory diseases. *Tissue Barriers*. (2016) 4:e1251384. doi: 10.1080/21688370.2016.1251384

5. Fasano A. All disease begins in the (leaky) gut: role of zonulin-mediated gut permeability in the pathogenesis of some chronic inflammatory diseases. *F1000Res*. (2020) 9:9. doi: 10.12688/f1000research.20510.1

6. Fasano A. Zonulin and its regulation of intestinal barrier function: the biological door to inflammation, autoimmunity, and cancer. *Physiol Rev.* (2011) 91:151–75. doi: 10.1152/physrev.00003.2008

7. Aho V, Houser MC, Pereira P, Chang J, Rudi K, Paulin L, et al. Relationships of gut microbiota, short-chain fatty acids, inflammation, and the gut barrier in Parkinson's disease. *Mol Neurodegener*. (2021) 16:6. doi: 10.1186/s13024-021-00427-6

8. Xu L, Xiong X, Ouyang Y, Barreto G, Giffard R. Heat shock protein 72 (Hsp 72) improves long term recovery after focal cerebral ischemia in mice. *Neurosci Lett.* (2011) 488:279–82. doi: 10.1016/j.neulet.2010.11.047

9. Liu TS, Musch MW, Sugi K, Walsh-Reitz MM, Ropeleski MJ, Hendrickson BA, et al. Protective role of HSP72 against *Clostridium difficile* toxin A-induced intestinal epithelial cell dysfunction. *Am J Physiol Cell Physiol.* (2003) 284:C1073–82. doi: 10.1152/ ajpcell.00134.2002

10. Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cells.* (2004) 118:229–41. doi: 10.1016/j.cell.2004.07.002

11. Varano DVJ, Lansley SM, Porcel JM, Bielsa S, Brown JS, Creaney J, et al. Bacterial infection elicits heat shock protein 72 release from pleural mesothelial cells. *PLoS One.* (2013) 8:e63873. doi: 10.1371/journal.pone.0063873

12. Standage SW, Lahni PM, Ma W, Kernie SG, Wong HR, Wheeler DS. Cerebrospinal fluid levels of extracellular heat shock protein 72: a potential biomarker for bacterial meningitis in children. *J Pediatr Intensive Care*. (2014) 3:023–8. doi: 10.3233/PIC-14080

13. Wood KL, Nunley DR, Moffatt-Bruce S, Pope-Harman A, Huang Q, Shamo EN, et al. The role of heat shock protein 27 in bronchiolitis obliterans syndrome after lung transplantation. *J Heart Lung Transplant.* (2010) 29:786–91. doi: 10.1016/j. healun.2010.03.004

14. Pockley AG, Shepherd J, Corton JM. Detection of heat shock protein 70 (Hsp 70) and anti-Hsp 70 antibodies in the serum of normal individuals. *Immunol Investig.* (1998) 27:367–77. doi: 10.3109/08820139809022710

15. Zerikiotis S, Angelidis C, Dhima I, Naka KK, Kasioumi P, Kalfakakou V, et al. The increased expression of the inducible HSP 70 (HSP70A1A) in serum of patients with heart failure and its protective effect against the cardiotoxic agent doxorubicin. *Mol Cell Biochem.* (2019) 455:41–59. doi: 10.1007/s11010-018-3469-7

16. Krepuska M, Szeberin Z, Sotonyi P, Sarkadi H, Fehervari M, Apor A, et al. Serum level of soluble Hsp 70 is associated with vascular calcification. *Cell Stress Chaperones.* (2011) 16:257–65. doi: 10.1007/s12192-010-0237-3

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

17. Kim JY, Kim JW, Yenari MA. Heat shock protein signaling in brain ischemia and injury. *Neurosci Lett.* (2020) 715:134642. doi: 10.1016/j.neulet.2019.134642

18. Fleshner M, Johnson JD. Endogenous extra-cellular heat shock protein 72: releasing signal (s) and function. *Int J Hyperth.* (2005) 21:457–71. doi: 10.1080/02656730500088211

19. Arumugam TV, Okun E, Tang SC, Thundyil J, Taylor SM, Woodruff TM. Toll-like receptors in ischemia-reperfusion injury. *Shock.* (2009) 32:4–16. doi: 10.1097/SHK.0b013e318193e333

20. Qi D, Liu H, Niu J, Fan X, Wen X, Du Y, et al. Heat shock protein 72 inhibits c-Jun N-terminal kinase 3 signaling pathway via Akt 1 during cerebral ischemia. *J Neurol Sci.* (2012) 317:123–9. doi: 10.1016/j.jns.2012.02.011

21. Barreto GE, White RE, Xu L, Palm CJ, Giffard RG. Effects of heat shock protein 72 (Hsp 72) on evolution of astrocyte activation following stroke in the mouse. *Exp Neurol.* (2012) 238:284–96. doi: 10.1016/j.expneurol.2012.08.015

22. Kesaraju S, Nayak G, Prentice HM, Milton SL. Upregulation of Hsp 72 mediates anoxia/reoxygenation neuroprotection in the freshwater turtle via modulation of ROS. *Brain Res.* (2014) 1582:247–56. doi: 10.1016/j.brainres.2014.07.044

23. Johnson JD, Fleshner M. Releasing signals, secretory pathways, and immune function of endogenous extracellular heat shock protein 72. *J Leukoc Biol.* (2006) 79:425–34. doi: 10.1189/jlb.0905523

24. Sulzbacher MM, Sulzbacher LM, Passos FR, Bilibio B, Althaus WF, Weizenmann L, et al. A single dose of eHSP72 attenuates sepsis severity in mice. *Sci Rep.* (2020) 10:9198. doi: 10.1038/s41598-020-66011-y

25. Rahman MT, Ghosh C, Hossain M, Linfield D, Rezaee F, Janigro D, et al. IFNgamma, IL-17A, or zonulin rapidly increase the permeability of the blood-brain and small intestinal epithelial barriers: relevance for neuro-inflammatory diseases. *Biochem Biophys Res Commun.* (2018) 507:274–9. doi: 10.1016/j.bbrc.2018.11.021

26. Yang Z, Wei F, Zhang B, Luo Y, Xing X, Wang M, et al. Cellular immune signal exchange from ischemic stroke to intestinal lesions through brain-gut axis. *Front Immunol.* (2022) 13:688619. doi: 10.3389/fimmu.2022.688619

27. Xian M, Cai J, Zheng K, Liu Q, Liu Y, Lin H, et al. Aloe-emodin prevents nerve injury and neuroinflammation caused by ischemic stroke via the PI3K/AKT/mTOR and NF-kappa B pathway. *Food Funct*. (2021) 12:8056–67. doi: 10.1039/d1fo01144h

28. Haghikia A, Li XS, Liman TG, Bledau N, Schmidt D, Zimmermann F, et al. Gut microbiota-dependent trimethylamine N-oxide predicts risk of cardiovascular events in patients with stroke and is related to proinflammatory monocytes. *Arterioscler Thromb Vasc Biol.* (2018) 38:2225–35. doi: 10.1161/ATVBAHA.118.311023

29. Zhou SY, Guo ZN, Yang Y, Qu Y, Jin H. Gut-brain axis: mechanisms and potential therapeutic strategies for ischemic stroke through immune functions. *Front Neurosci.* (2023) 17:1081347. doi: 10.3389/fnins.2023.1081347

30. Linninge C, Jonsson P, Bolinsson H, Onning G, Eriksson J, Johansson G, et al. Effects of acute stress provocation on cortisol levels, zonulin and inflammatory markers in low- and high-stressed men. *Biol Psychol.* (2018) 138:48–55. doi: 10.1016/j. biopsycho.2018.08.013

31. Musch MW, Sugi K, Straus D, Chang EB. Heat-shock protein 72 protects against oxidant-induced injury of barrier function of human colonic epithelial Caco 2/bbe cells. *Gastroenterology.* (1999) 117:115–22. doi: 10.1016/s0016-5085(99)70557-3

32. Mukherjee A, Lordan C, Ross RP, Cotter PD. Gut microbes from the phylogenetically diverse genus Eubacterium and their various contributions to gut health. *Gut Microbes.* (2020) 12:1802866. doi: 10.1080/19490976.2020.1802866

33. Zhuge A, Li S, Lou P, Wu W, Wang K, Yuan Y, et al. Longitudinal 16S rRNA sequencing reveals relationships among alterations of gut microbiota and nonalcoholic

fatty liver disease progression in mice. *Microbiol Spectr.* (2022) 10:e0004722. doi: 10.1128/spectrum.00047-22

34. Gao J, Yan KT, Wang JX, Dou J, Wang J, Ren M, et al. Gut microbial taxa as potential predictive biomarkers for acute coronary syndrome and post-STEMI cardiovascular events. *Sci Rep.* (2020) 10:2639. doi: 10.1038/ s41598-020-59235-5

35. Li L, Bao J, Chang Y, Wang M, Chen B, Yan F. Gut microbiota may mediate the influence of periodontitis on prediabetes. *J Dent Res.* (2021) 100:1387–96. doi: 10.1177/00220345211009449

36. Liu S, Zhao W, Liu X, Cheng L. Metagenomic analysis of the gut microbiome in atherosclerosis patients identify cross-cohort microbial signatures and potential therapeutic target. *FASEB J.* (2020) 34:14166–81. doi: 10.1096/fj.202000622R

37. Chung W, Meijerink M, Zeuner B, Holck J, Louis P, Meyer AS, et al. Prebiotic potential of pectin and pectic oligosaccharides to promote anti-inflammatory

commensal bacteria in the human colon. *FEMS Microbiol Ecol.* (2017) 93:93. doi: 10.1093/femsec/fix127

38. Wang N, Yan Z, Liu N, Zhang X, Xu C. Synergy of Cellulase systems between Acetivibrio thermocellus and Thermoclostridium stercorarium in consolidated-bioprocessing for cellulosic ethanol. *Microorganisms*. (2022) 10:10. doi: 10.3390/microorganisms10030502

39. Yuan GH, Zhang Z, Gao XS, Zhu J, Guo WH, Wang L, et al. Gut microbiotamediated tributyltin-induced metabolic disorder in rats. *RSC Adv.* (2020) 10:43619–28. doi: 10.1039/d0ra07502g

40. Gerritsen J, Hornung B, Renckens B, van Hijum S, Martins DSV, Rijkers GT, et al. Genomic and functional analysis of Romboutsia ilealis CRIB (T) reveals adaptation to the small intestine. *Peerj.* (2017) 5:e3698. doi: 10.7717/peerj.3698

41. Zeng Q, Li D, He Y, Li Y, Yang Z, Zhao X, et al. Discrepant gut microbiota markers for the classification of obesity-related metabolic abnormalities. *Sci Rep.* (2019) 9:13424. doi: 10.1038/s41598-019-49462-w

Glossary

CI	Cerebral infarction
ELISA	enzyme-linked immunoassay
NIHSS	National Institute of Health Stroke Scale
EGF	epidermal growth factor
ТЈ	tight junction
HSP72	heat shock protein 72
СТ	computed tomography
MRI	magnetic resonance imaging
ROC	receiver operating characteristic
AUC	area under the curve
SCFAs	short-chain fatty acids
ТМАО	trimethylamine N-oxide
TLRs	toll-like receptors
AP-1	activator protein 1
NF-ĸB	nuclear factor κΒ
LPS	lipopolysaccharide
MMP-9	metalloproteinase 9
МСАО	middle cerebral artery occlusion