



OPEN ACCESS

EDITED BY

Kehinde Samuel Olaniyi,
University of KwaZulu-Natal, South Africa

REVIEWED BY

Sujata Kar,
Ravenshaw University, India
Nazan Yurtcu,
Sivas Cumhuriyet University Faculty of
Medicine, Türkiye

*CORRESPONDENCE

Lianwen Zheng
✉ davezheng@sohu.com

RECEIVED 25 March 2023

ACCEPTED 12 June 2023

PUBLISHED 28 June 2023

CITATION

Gao Y, Zou Y, Wu G and Zheng L (2023)
Oxidative stress and mitochondrial dysfunction
of granulosa cells in polycystic ovarian
syndrome.
Front. Med. 10:1193749.
doi: 10.3389/fmed.2023.1193749

COPYRIGHT

© 2023 Gao, Zou, Wu and Zheng. This is an
open-access article distributed under the terms
of the [Creative Commons Attribution License
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction
in other forums is permitted, provided the
original author(s) and the copyright owner(s)
are credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted which
does not comply with these terms.

Oxidative stress and mitochondrial dysfunction of granulosa cells in polycystic ovarian syndrome

Yiyin Gao, Yinggang Zou, Guijie Wu and Lianwen Zheng*

Reproductive Medical Center, Department of Obstetrics and Gynecology, The Second Hospital of Jilin University, Changchun, China

Polycystic ovarian syndrome (PCOS) is one of the leading causes of anovulatory infertility in women, affecting 5%–15% of women of reproductive age worldwide. The clinical manifestations of patients include ovulation disorders, amenorrhea, hirsutism, and obesity. Life-threatening diseases, such as endometrial cancer, type 2 diabetes, hyperlipidaemia, hypertension, and cardiovascular disease, can be distant complications of PCOS. PCOS has diverse etiologies and oxidative stress (OS) plays an important role. Mitochondria, as the core organelles of energy production, are the main source of reactive oxygen species (ROS). The process of follicular growth and development is extremely complex, and the granulosa cells (GCs) are inextricably linked to follicular development. The abnormal function of GCs may directly affect follicular development and alter many symptoms of PCOS. Significantly higher levels of OS markers and abnormal mitochondrial function in GCs have been found in patients with PCOS compared to healthy subjects, suggesting that increased OS is associated with PCOS progression. Therefore, the aim of this review was to summarize and discuss the findings suggesting that OS and mitochondrial dysfunction in GCs impair ovarian function and induce PCOS.

KEYWORDS

polycystic ovarian syndrome, oxidative stress, granulosa cells, mitochondrial function, reactive oxygen species, infertility

1. Introduction

Polycystic ovarian syndrome (PCOS) is a common reproductive endocrine disorder, in which patients often present with ovulatory disturbances, amenorrhoea, and androgen excess disorders (1). PCOS in adults is diagnosed when at least two of the following three criteria are met: impaired ovarian function, clinical and/or biochemical hyperandrogens, and polycystic ovaries (2). Despite a long history of PCOS research, its pathogenesis remains unclear. It is widely believed that dysregulation of androgen and gonadotropin secretion, oxidative stress (OS), abnormal mitochondrial function and glucose metabolism disorders lead to impaired energy and metabolic regulation, and insulin resistance (IR) and hyperandrogenism are core factors in PCOS etiology (3).

Molecular oxygen (O_2) is essential for all aerobic organisms. There are two unpaired electrons in different orbitals in the outer shell of O_2 , which facilitates radical formation (4). Superoxide (O_2^-) is the reduced product of O_2 and the precursor of other reactive oxygen species (ROS). Superoxide anions (O_2^-), hydroxyl radicals (OH^-), peroxy (ROO), alkoxy (RO), and hydroperoxy (HO_2) are the most biologically important ROS (5). Appropriate ROS levels play an important role in the oxidative modification of proteins, such as receptors, ion channels, and transcription factors (6, 7). However, excessive ROS levels cause an imbalance in the redox

system and lead to OS, which, in turn, has cytotoxic effects by initiating the peroxidation of membrane phospholipids and altering nucleic acids, lipids, and proteins. These processes lead to altered cell physiology, including apoptosis, increased membrane permeability, reduced enzyme activity, DNA damage, mitochondrial alterations, and ATP depletion (8, 9).

The process of follicular growth and development is complex and delicate, and granulosa cells (GCs) are inextricably linked to this process (10). Abnormalities in GC function may directly affect follicular development and alter several features of PCOS (11). OS may alter the function of GCs and oocytes and affect the follicular fluid (FF) microenvironment, as well as ovarian bioactivities, such as hormonal signaling, oocyte maturation, and ovarian steroidogenesis, and eventually have a negative impact on reproductive outcomes (12).

OS is considered a potential trigger for the development of PCOS, as numerous studies have shown that markers of oxidative damage are significantly elevated in the serum of patients with PCOS compared to those in normal subjects (13, 14). There are several mechanisms underlying OS in the development of PCOS. In this review, we focused on the OS of GCs in the pathogenesis of PCOS by reviewing *in vitro* and *in vivo* studies. Understanding the role of OS in PCOS may provide future directions for the development of treatment strategies.

2. GCs and their role in PCOS

Ovarian GCs are round or oval in shape and closely distributed around the oocyte, which comprises the cellular components of the follicle together with the oocyte; corpus luteum cells constitute the cellular components of the follicle (15, 16). As the largest cell population in the follicle, the proliferation and differentiation status of GCs directly affect the quality of oocytes, which consequently affects the functional activities of the ovary, such as the initiation and development of follicles, ovulation, and corpus luteum formation (17). Mitochondrial function and developmental potential of oocytes can be enhanced by surrounding GCs via mitophagy (18). In the oocyte-GC complex, low molecular weight signals or nutrient molecules are transferred between cells as a means of transporting nutrients to the oocyte and sharing metabolic intermediates and products. The transmission capacity increases as the ratio of oocyte volume to surface area increases (10). As the oocyte grows, the zona pellucida gradually separates GCs from the oocyte, and the GCs contact the oocyte by producing filamentous pseudopods that cross the zona pellucida (19). Thus, oocytes acquire glycolytic and cholesterol metabolites from GCs, which cannot be synthesized independently (20). The growth and development of oocytes depend on the production of such metabolites by GCs and their transfer across the zona pellucida to the oocyte, which is known as metabolic coupling. The greater the number of GCs that are metabolically coupled to the oocyte, the faster the oocyte grows (21). A distinctive sign of follicular development is the rapid growth and proliferation of GCs, whereas follicular atresia is mainly caused by the apoptosis of GCs. In addition, GCs work together with follicular membrane cells to complete the synthesis of ovarian hormones and maintain a microenvironment conducive to oocyte growth and maturation (22).

GCs play a supportive role in physiological folliculogenesis and greatly influence pathological folliculogenesis in PCOS, premature ovarian failure (POF), and some malignant diseases, such as ovarian

granulosa cell tumors (GCT) (11). Abnormalities in GC function may directly affect follicular development and alter several PCOS features. In PCOS patients, it is difficult for multiple small antral follicles to develop into larger dominant follicles. In addition, the function of GCs differs in patients with PCOS. A significant increase in follicle number and proliferation of GCs was observed in a PCOS mouse model, as well as in the ovaries of patients with PCOS (23, 24). Therefore, abnormal proliferation of ovarian GCs is likely to play a role in the pathogenesis of PCOS. Furthermore, previous studies have found that increased levels of apoptosis in GCs are one of the key factors contributing to follicular dysplasia and impaired oocyte maturation. Peng found that the level of apoptosis in GCs is significantly elevated in patients with PCOS, and that knocking down the expression of PATL2, a gene associated with oocyte development, using siRNA significantly prevented apoptosis of GCs (25). In addition, the anti-Müllerian hormone (AMH) secreted by GCs inhibits sinus follicle development, and decreased AMH levels are a prerequisite for ovulation. In contrast, androgens in the follicular fluid of patients with PCOS can block the decrease in AMH levels either directly or through the AMH-Smad signaling pathway. Moreover, GCs have a reduced ability to respond to persistently elevated AMH levels, resulting in follicular development arrest (26).

Since patients with PCOS may be affected by obesity and hyperinsulinemia, they often exhibit a chronic inflammatory state. Evidence suggests that OS and chronic inflammation caused by mitochondrial damage in GCs are associated with the pathogenesis of PCOS. A study investigating the metabolomics of FF in PCOS showed that mitochondrial dysfunction of GCs, redox potential imbalance, and increased cellular OS levels in PCOS patients could explain, to some extent, the metabolic disorders observed in PCOS (27).

3. Markers of OS in PCOS

OS refers to the excessive production of ROS *in vivo* or impaired production of antioxidants, resulting in an imbalance between oxidants and antioxidants, which may lead to DNA damage and apoptosis (28). ROS represent a class of molecules that originate from the metabolism of oxygen in aerobic organisms and include the one-electron reduction product of oxygen, superoxide anion (O_2^-), two-electron reduction product, hydrogen peroxide (H_2O_2), three-electron reduction product, hydroxyl radical (OH), and nitric oxide (NO) (28). ROS function as intracellular secondary messengers by regulating signal transduction pathways, gene expression, post-translational modification of proteins, and participation in cell proliferation, differentiation, and apoptosis (29). Highly complex antioxidant enzymes and nonenzymatic systems regulate ROS scavenging. Various antioxidant enzymes, such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione reductase (GR), glucose 6-phosphate dehydrogenase and isocitrate dehydrogenase can attenuate OS and tissue damage in cells by scavenging ROS (30). However, the main mechanism of resistance to excessive ROS damage is the nonenzymatic antioxidant system. It contains antioxidants, such as vitamins and polyphenols, and directly neutralizes ROS in the body (31). Trace elements, such as Cu, Zn, and Mn, are cofactors for antioxidant enzymes and are essential components of the enzymatic antioxidant system (30).

The detection of OS and antioxidant biomarkers has been suggested as a useful tool for assessing the risk of oxidative damage in various diseases (32). Table 1 shows the results of several OS markers detected in patients with PCOS in different studies.

Malondialdehyde (MDA) is a stable biomarker of lipid peroxidation, as reported in several studies of patients with PCOS. Circulating MDA concentrations increased by 47% in patients with PCOS compared to controls (41). In addition, Palacio et al. confirmed that MDA levels in erythrocytes were significantly higher in patients with PCOS than in BMI- and age-matched control populations (42). Another intracellular signaling molecule, NO, is involved in many biological processes and can be toxic in high concentrations. Although a meta-analysis revealed no significant difference in NO levels in patients with PCOS compared to controls (41), another study by Nacul et al. reported a significant negative correlation between NO levels and IR-related indices, including fasting insulin levels and homeostatic model assessments in patients with PCOS (35). AGEs are a class of stable end products formed by the non-enzymatic reaction of free amino groups of molecules, such as proteins, amino acids, and lipids with aldehyde groups of reducing sugars. AGEs have been reported as markers of OS in PCOS, and there is growing evidence that AGEs may alter steroid biosynthesis in PCOS by affecting enzyme function, inducing inflammatory changes and IR, and ultimately affecting androgen synthesis and follicular development (42). Tantalaki et al. investigated the effect of dietary intake of AGEs on hormone secretion. The results indicated that PCOS patients with a diet high in AGEs had higher levels of serum AGEs along with elevated levels of testosterone, free androgen index, and androstenedione, confirming the association between AGEs and excessive androgen levels (43). XO is an enzyme that participates in O_2^- generation, which is also involved in the metabolism of purines and catalyzes the production of ROS (36). Isik et al. found that serum C-reactive protein (CRP), platelet and platelet-specific volume (PCT) values, and XO activity were significantly higher in patients with PCOS, while SOD activity was decreased. XO activity was positively correlated with fasting plasma insulin and IR index and negatively correlated with quantitative insulin sensitivity check index levels. Therefore, XO may be used as a sensitive marker to assess the OS of patients with PCOS (44).

Generally, several cellular mechanisms defend against free radicals in the organelles, cell membranes, and extracellular environment. One mechanism involves enzymes, such as CAT, GPx, and SOD, which prevent the production or rapidly inactivate free radicals. Another mechanism interrupts the propagation of the lipid peroxidation chain by reacting with intermediate free radicals, which work as scavenging molecules, including albumin, bilirubin, ascorbic acid, fat-soluble vitamin E, and coenzyme Q10. Finally, the process of scavenging molecules damaged by oxidative attack is used to re-establish normal structure. Table 2 indicates several antioxidant markers in PCOS, but most of the findings are conflicting and further studies are required to elucidate the mechanisms underlying the changes in antioxidant levels in PCOS patients.

SOD catalyzes the production of H_2O_2 from O_2^- , which is further converted to H_2O by GPx, thereby eliminating ROS and performing its antioxidant function. Serum SOD activity in patients with PCOS has been reported to differ, with Sabuncu et al. claiming that serum SOD was significantly higher in patients with PCOS than in controls, whereas Zhang et al. reached the opposite conclusion (51, 52). Selem

TABLE 1 Oxidative stress markers in patients with PCOS.

Biomarkers	Origin	Changes in PCOS patients
MDA	Serum; Erythrocyte	↑ (33, 34)
NO	Serum	↑ (35) or (36, 37)
AGEs	Serum	↑ (38)
XO	Serum	↑ (36, 39)
TOS	Serum, FF	↑ (40)

MDA, malondialdehyde; NO, nitric oxide; AGEs, advanced glycation end products; XO, xanthine oxidase; TOS, total oxidant status.

TABLE 2 Anti-oxidative stress markers in patients with PCOS.

Biomarkers	Origin	Changes in PCOS patients
SOD	Serum; Erythrocyte; FF	↑ (39) or ↓ (39, 45, 46)
GPx	Serum	↓ (47) or (36)
CAT	Serum	↓ (45)
TAC	FF, Serum	↑ (45) or ↓ (44, 48) or (37)
GSH	Serum	↓ (45)
NAD+	GCs	↓ (49)
Vitamin E	Serum	↓ (50)
Vitamin C	Serum	↓ (50)

SOD, superoxide dismutase; GPx, glutathione peroxidase; TAC, total antioxidant capacity; GSH, glutathione; NAD+, nicotinamide adenine dinucleotide.

et al. reported lower SOD activity in both the serum and FF of patients with PCOS and concluded that serum SOD activity may be a clinical parameter for determining systemic OS in PCOS (44). Further studies are required to explore the antioxidant defense mechanisms of SOD in PCOS. However, neither study showed that GPx activity in patients with PCOS differed from that in controls (36, 52), although Savic-Radojevic et al. reported significantly lower GPx activity in patients with PCOS than in controls (53).

Total antioxidant capacity (TAC) refers to the ability of extracellular nonenzymatic antioxidants to scavenge free radicals, which can block the propagation chain of lipid peroxidation and prevent the expansion of free radical generation and subsequent biochemical damage (54). TAC is regarded as a better indicator of the total antioxidant status of a biological sample because it reflects the combined effects of various antioxidants (55). Several studies have reported TAC levels in patients with PCOS. A meta-analysis showed no significant difference in TAC between patients with PCOS and controls (53); however, Fenkci et al. showed significantly lower TAC levels in patients with PCOS (56). Verit et al. reported significantly elevated TAC levels in patients with PCOS compared to age- and BMI-matched controls (57). Although the complete mechanism underlying this elevation is unknown, they speculated that elevated TAC levels may be a responsive change in the body in answer to increased OS levels (57). As the most important non-enzymatic antioxidant, GSH is essential for regulating protein disulfide bonds and scavenging oxidants. Dinger et al. suggested that GSH depletion may be due to increased ROS production in patients with PCOS (58). NAD+ is involved in the regulation of the energy tricarboxylic acid cycle, oxidative phosphorylation, and intracellular glycolytic reactions,

and it is often considered as a central component of redox reactions with its reduced form, NADH (59). It has been shown that NAD⁺ levels in GCs of women with PCOS are lower than in controls, accompanied by elevated levels of inflammatory factors and abnormal mitochondrial dynamics (49). In addition, it has also been shown that concentrations of vitamin C and vitamin E, which have antioxidant capacity, are lower in PCOS patients than in controls (50).

4. Etiology related to OS in PCOS

Increased levels of ROS in ovarian GCs of women with PCOS may impair the function of mitochondrial oxidative metabolism, leading to abnormal morphology of GCs, resulting in inadequate energy supply and increased OS, while greatly inducing apoptosis of GCs. Elevated levels of ROS in FF are associated with poor oocyte and embryo quality (60). In turn, altered oocyte quality or capacity is thought to be a major influence on infertility or poor pregnancy outcomes in women with PCOS (61). Although more oocytes can be collected from women with PCOS, the rates of MII oocyte, 2PN fertilization, oogenesis, day 3 quality embryo and clinical pregnancy are lower than controls (12, 62).

OS plays an important role in the development of PCOS by influencing cellular functions in several ways. Various protein kinases are reportedly activated with increased OS damage, which in turn induces serine/threonine phosphorylation of insulin receptor substrates, leading to their degradation and reduced insulin sensitivity, thereby promoting the development of IR (63). However, with elevated ROS levels, large amounts of Ca²⁺ are released from the endoplasmic reticulum, leading to the dysregulation of intracellular calcium homeostasis and cellular necrosis. Calcium dysregulation eventually causes follicular arrest in patients with PCOS, which in turn leads to reproductive disorders, such as amenorrhoea and anovulation (64). The direct oxidation of proteins and free amino acid side chains leads to the formation of carbonyl products, resulting in protein denaturation and inactivation (65). Serum levels of advanced oxidation protein products (AOPPs) are significantly higher in PCOS patients than in healthy women, and therefore AOPPs may serve as a new marker of oxidant-mediated protein damage (66). Lipid peroxidation that occurs in the side chains of polyunsaturated fatty acids in the cell membrane is often a chain reaction, and the free radicals formed by the binding of fatty acid side chains to oxygen can, in turn, bind to other fatty acids to obtain hydrogen ions (67). Lipid peroxidation markers, such as thiobarbituric acid-reactive substances, oxidized low-density lipoproteins, and MDA, were detected at significantly higher levels in patients with PCOS than in normal controls. Serum lipid concentrations increase simultaneously with peroxidation (68).

5. Mitochondrial functions of GCs in PCOS

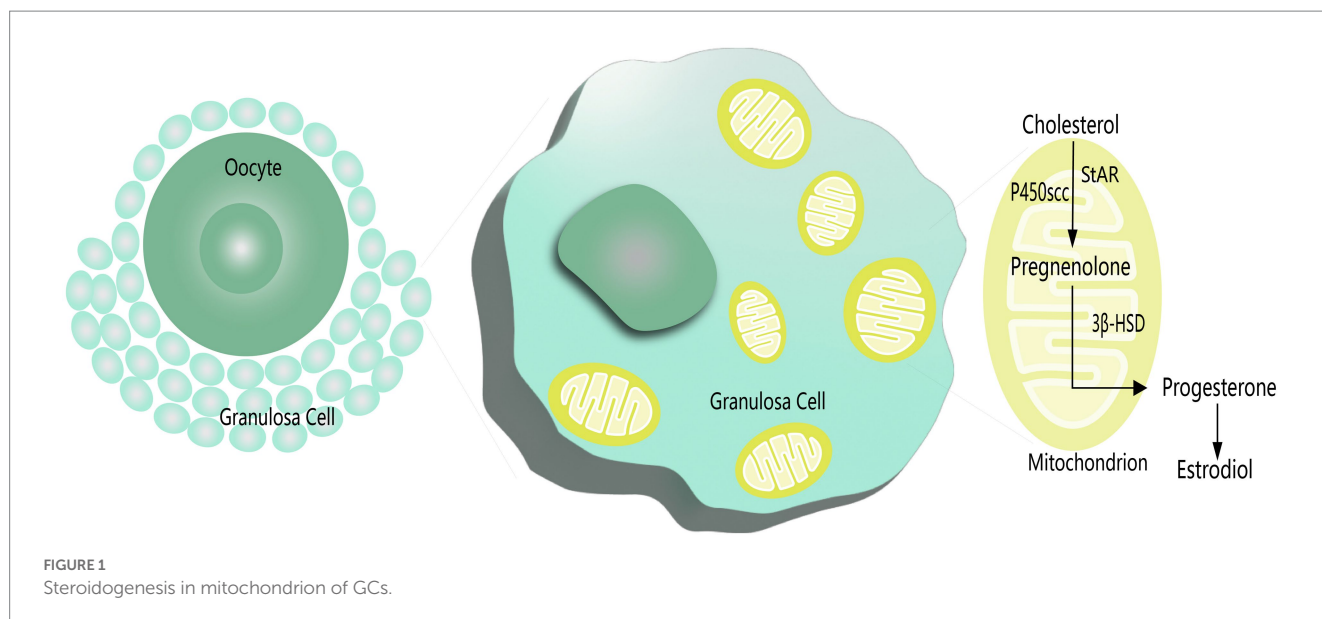
Mitochondria are important for energy production and insulin secretion and are essential regulators of apoptosis because they provide energy for normal cellular functions (69, 70). Mitochondrial abnormalities usually manifest as diseases of the entire organism and lead to varying degrees of metabolic disorders (71). The results of a

transcriptomic study showed altered expression of several genes involved in oxidative phosphorylation, mitochondrial function, and the sirtuin signaling pathway in the GCs of patients (72). However, little is known about the roles of mitochondrial regulatory factors in GCs.

As the powerhouse of the cell, mitochondria replenish ATP through oxidative phosphorylation (OxPhos) to maintain cellular energy homeostasis (73). Additionally, mitochondria produce ROS through the electron transport chain (ETC) complex, and excessive ROS production induces OS, leading to mitochondrial dysfunction and, consequently, an imbalance between the oxidant and antioxidant systems. The main ROS in the mitochondria is O^{2•-}, which is generated by electron leakage from the ETC and can react with O₂ (74). Mitochondria contain their own genetic material, mitochondrial DNA (mtDNA), which encodes seven genes in ETC complex I (75). Mutations in mtDNA can lead to mitochondrial dysfunction and induce IR by affecting OS homeostasis, energy deficiency, or hormone metabolism. Mutations in mtDNA mostly occur in mt-tRNA genes, which alter RNA processing by destabilizing tRNA tertiary structures, leading to defects in nucleotide modifications. Defects in mitochondrial protein synthesis reduce ATP production in GCs and pancreatic cells, resulting in the clinical phenotype of PCOS. The effects of mtDNA copy number and GC variation on the regulatory mechanisms of GC function have been the focus of scientific research. Currently, the role of mtDNA copy number in PCOS is contradictory, as different groups have reported positive, negative, or no associations between mtDNA copy number and PCOS (76–78). Au et al. demonstrated that the occurrence of mtDNA 4,977 bp deletion in GCs is associated with apoptosis (79). Overall, 33 mtDNA mutations were identified in patients with PCOS, and the majority of these mutations were found in the D-loop region (80).

In addition, mitochondrial swelling has been found in degenerated GCs and is associated with apoptosis and follicular atresia (81). Karuputhula et al. found reduced GC counts in patients with endometriosis and PCOS compared to controls, accompanied by decreased cell viability and mitochondrial abnormalities (82). Similar findings have been reported for mitochondrial dysfunction and mutations in GCs from patients with endometriosis or PCOS (83, 84). One of the representative functions of GCs is steroid hormone secretion. Mitochondria are the central organelles for steroidogenesis, with the initiating and rate-limiting steps of biosynthesis occurring in the mitochondria (Figure 1) (85). Studies have shown that estradiol (E₂) levels are significantly reduced in the serum and FF of patients with endometriosis, which is associated with reduced mitochondrial quality and mitochondrial membrane potential (MMP) in GCs accompanied by elevated apoptosis in GCs (86). Gong et al. also observed decreased MMP and elevated apoptosis in GCs with higher TOS in serum and FF in patients with PCOS, while treatment with growth factor may alleviate OS and improve GCs function (40).

GCs treated with methamphetamine (MA), a central nervous system stimulant, showed significant swelling and degeneration of the mitochondria with significant enlargement of the mitochondrial cristae septum accompanied by a decline in ovarian function (87). Similarly, most GCs exposed to ozone have swollen mitochondria and display vacuolisation and degeneration of the cristae and matrix. 4-Hydroxynonenal (4-HNE), produced during lipid oxidation, modifies mitochondrial proteins, leading to mitochondrial dysfunction. 4-HNE immunostaining was significantly increased in



the GCs of the ozone-exposed group, suggesting a significant impairment of mitochondrial function (88). Decreased mitochondrial function directly leads to decreased GC function, which in turn leads to decreased oocyte quality and ovarian reserve function and has a negative impact on reproductive outcomes.

6. Glucose metabolism of GCs in PCOS

Another downstream mechanism that plays an important role in PCOS is glucose metabolism, which also participates in the bidirectional signaling between GCs and oocytes. Since oocytes have low glycolytic activity, they preferentially utilize pyruvate and lactate produced by GCs via the mitochondrial OxPhos pathway to meet their energy requirements (89). In contrast, impaired glycolytic metabolism of GCs leads to insufficient energy substrates for oocytes and GCs, resulting in excessive ROS production due to dysfunctional mitochondrial OxPhos and ultimately impairing oocyte quality.

A recent multi-omics analysis of GCs in PCOS identified abnormalities in metabolism-related signals, such as glycolysis/glyoxylate production (90). Under hypoxic conditions, cells use the glycolytic pathway to produce ATP instead of the oxygen-independent metabolic pathway, which contributes to cell proliferation of GCs and oocyte development during hypoxia (91, 92). As a regulator of the cellular response to hypoxia, the expression of hypoxia-inducible factor 1 α (HIF- α) was increased under hypoxic conditions that induces a metabolic switch from OxPhos to glycolysis (93). The reduced mitochondrial activity of GCs in PCOS patients may inhibit the signaling of HIF-1 α , delaying the switch in energy metabolism during follicle growth and affecting normal follicle development (94).

Researchers have observed a significant decrease in glycolytic rate and reduced expression of key glycolytic enzymes in GCs of PCOS patients including hexokinase, muscular phosphofructokinase, platelet derived phosphofructokinase, and muscular pyruvate kinase (95). Besides, oocytes from women with PCOS exhibit increased mitochondrial ROS and decreased MMP with downregulation of

glucose-6-phosphate dehydrogenase (G6PD) (12). Mazloomi et al. postulated that increased ROS levels in GCs impair glycolysis, reduce energy levels, and ultimately disrupt GCs' function since ROS levels were inversely correlated with the expression of key glycolytic enzymes in PCOS. Zhang et al. found that these alterations were associated with a deficiency of Sirtuin 3 (SIRT3) in GCs, which resides primarily in mitochondria and regulates mitochondrial proteins via deacetylation (96). SIRT3 in GCs is involved in processes such as metabolic homeostasis and OS by targeting mitochondrial enzymes, such as glutamate dehydrogenase (97). In patients with PCOS, the deficiency of SIRT3 in GCs may lead to elevated mitochondrial ROS and altered cellular insulin signal pathways by altering the acetylation status of NDUFS1. The incapacity of mitochondria in GCs to maintain ATP synthesis promotes OS and impairs glucose metabolism, and eventually negatively influence the maturation of oocyte (12).

7. Treatment strategy of PCOS

Given the role of mitochondrial dysfunction in the development of PCOS, treatments targeting the mitochondria to improve PCOS symptoms have become a research hotspot. Vitamin D can directly act on mitochondria, which improves cell biosynthesis and anti-apoptosis levels, and upregulates antioxidant expression, thereby reducing ROS levels. Moreover, vitamin D stimulates the expression of mitochondrial transcription factor A, increases mtDNA copy number, and improves the integrity of the mitochondrial membrane (98). In DHEA-induced PCOS rat models, vitamin D supplementation improved ovarian and uterine morphology, and reduced body weight and obesity (99). Additionally, vitamin D regulates steroid production and reduces progesterone (P) and E₂ production by reducing the expression of steroidogenic enzymes. The application of the mitochondria-targeted antioxidant MitoQ10 can improve IR status, reduce the expression levels of apoptosis-related proteins, and improve damaged mitochondrial function in PCOS rats (100).

The mitochondria-targeted antioxidant melatonin enhances SIRT1 expression, thus regulating PINK1/Parkin-mediated

mitochondrial autophagy. The application of melatonin in PCOS patients significantly improves mitochondrial damage in GCs (101). Growth hormone can alleviate OS-induced apoptosis in GCs of patients with PCOS by activating the PI3K/Akt signaling pathway. The acute-phase protein PTX3 is locally expressed at the site of inflammation and can serve as a sensitive marker of endothelial dysfunction in patients (102). PTX3 is synthesized by GCs and plays an important role in normal oocyte development and fertilization. In contrast, PTX3 levels in the follicular fluid of PCOS patients are positively correlated with serum androgen concentrations, suggesting that the chronic inflammatory state of PCOS is associated with hyperandrogenemia (103). CRISPR/Cas9 technology has been used to screen genes involved in the regulation of mitochondrial metabolism in GCs, revealing several tumor suppressors with enhanced mitochondrial function, including HBP1, TP53, PTEN, CTNNA2, and WISP1 (104). Among them, HBP1 is involved in regulating mitochondrial production and transcription of the mitochondrial transcription factor (TFAM), a key gene associated with mitochondrial DNA production. Further experiments in mice showed that the knockdown of HBP1 expression enhanced mitochondrial function, reduced apoptosis-related signals in GCs, and promoted follicle development and oocyte maturation (104). Besides, inner mitochondrial membrane peptidase subunit 2 (Immp2l) deficiency leads to mitochondrial dysfunction in GCs, which further leads to increased OS, decreased E2 levels, and altered gene expression levels of Wnt/ β -catenin and steroid hormone synthesis pathways, such as CYP19A1, while melatonin is able to reverse these changes (105). The addition of estradiol-17 β to the culture medium significantly increased telomere length in GCs, which became more obvious with increasing E2 concentrations (47). Last but not least, as more than 10 enzymes from glycolytic and TCA cycle metabolic pathways were found to be differentially hypo- or hyperacetylated in PCOS, possible therapeutic strategies should be evaluated in the future by targeting miRNAs that specifically regulate the expression of these enzymes (106).

8. Conclusion

OS plays an important role in the development of PCOS. This paper reviews and summarizes information on the role of GCs in the pathogenesis of PCOS, the association between OS and PCOS, OS markers, antioxidant capacity in patients with PCOS, and the

participation of mitochondrial function. To date, most studies have been based on assays performed on serum or FF samples collected from patients with PCOS, and some data have produced conflicting results. More animal and *in vitro* studies are necessary to elucidate the mechanisms by which OS affects fertility in patients with PCOS, and randomized controlled clinical trials are needed to explore more effective targets for the treatment of PCOS.

Author contributions

LZ contributed to the conception and design of the study and revised the manuscript. YG wrote the first draft of the manuscript. YZ and GW wrote sections of the manuscript. All authors contributed to the article and approved the submitted version.

Funding

The work was supported by the Natural Science Foundation of Jilin Province, China (YDZJ202201ZYTS084).

Acknowledgments

The authors would like to thank Editage (www.editage.cn) for English language editing.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Swaroop A, Jaipuria AS, Gupta SK, Bagchi M, Kumar P, Preuss HG, et al. Efficacy of a novel fenugreek seed extract (*Trigonella foenum-graecum*, Furocyst) in polycystic ovary syndrome (PCOS). *Int J Med Sci.* (2015) 12:825–31. doi: 10.7150/ijms.13024
2. Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod.* (2004) 19:41–7. doi: 10.1093/humrep/deh098
3. Teede HJ, Hutchison SK, Zoungas S. The management of insulin resistance in polycystic ovary syndrome. *Trends Endocrinol Metab.* (2007) 18:273–9. doi: 10.1016/j.tem.2007.08.001
4. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol.* (2007) 39:44–84. doi: 10.1016/j.biocel.2006.07.001
5. Zorbakhsh S. Effect of antioxidants on preimplantation embryo development in vitro: a review. *Zygote.* (2021) 29:179–93. doi: 10.1017/S0967199420000660
6. Scialò F, Fernández-Ayala DJ, Sanz A. Role of mitochondrial reverse Electron transport in ROS signaling: potential roles in health and disease. *Front Physiol.* (2017) 8:428. doi: 10.3389/fphys.2017.00428
7. De Giusti VC, Caldiz CI, Ennis IL, Pérez NG, Cingolani HE, Aiello EA. Mitochondrial reactive oxygen species (ROS) as signaling molecules of intracellular pathways triggered by the cardiac renin-angiotensin II-aldosterone system (RAAS). *Front Physiol.* (2013) 4:126. doi: 10.3389/fphys.2013.00126
8. Schieber M, Chandel NS. ROS function in redox signaling and oxidative stress. *Curr Biol.* (2014) 24:R453–62. doi: 10.1016/j.cub.2014.03.034
9. Slimen IB, Najjar T, Ghram A, Dabbebi H, Ben Mrad M, Abdrabbah M. Reactive oxygen species, heat stress and oxidative-induced mitochondrial damage. A review. *Int J Hyperth.* (2014) 30:513–23. doi: 10.3109/02656736.2014.971446
10. Thomas FH, Vanderhyden BC. Oocyte-granulosa cell interactions during mouse follicular development: regulation of kit ligand expression and its role in oocyte growth. *Reprod Biol Endocrinol.* (2006) 4:19. doi: 10.1186/1477-7827-4-19

11. Tu J, Cheung AH, Chan CL, Chan WY. The role of microRNAs in ovarian granulosa cells in health and disease. *Front Endocrinol.* (2019) 10:174. doi: 10.3389/fendo.2019.00174
12. Zhang Q, Ren J, Wang F, Pan M, Cui L, Li M, et al. Mitochondrial and glucose metabolic dysfunctions in granulosa cells induce impaired oocytes of polycystic ovary syndrome through Sirtuin 3. *Free Radic Biol Med.* (2022) 187:1–16. doi: 10.1016/j.freeradbiomed.2022.05.010
13. Özer A, Bakacak M, Kiran H, Ercan Ö, Köstü B, Kanat-Pektaş M, et al. Increased oxidative stress is associated with insulin resistance and infertility in polycystic ovary syndrome. *Ginekol Pol.* (2016) 87:733–8. doi: 10.5603/GP.2016.0079
14. Mancini A, Bruno C, Vergani E, d'Abate C, Giacchi E, Silvestrini A. Oxidative stress and low-grade inflammation in polycystic ovary syndrome: controversies and new insights. *Int J Mol Sci.* (2021) 22:1667. doi: 10.3390/ijms22041667
15. Weakley BS. Electron microscopy of the oocyte and granulosa cells in the developing ovarian follicles of the golden hamster (*Mesocricetus auratus*). *J Anat.* (1966) 100:503–34.
16. Hummitchsch K, Anderson RA, Wilhelm D, Wu J, Telfer EE, Russell DL, et al. Stem cells, progenitor cells, and lineage decisions in the ovary. *Endocr Rev.* (2015) 36:65–91. doi: 10.1210/er.2014-1079
17. Turathum B, Gao EM, Chian RC. The function of cumulus cells in oocyte growth and maturation and in subsequent ovulation and fertilization. *Cells.* (2021) 10:2292. doi: 10.3390/cells10092292
18. Shen Q, Liu Y, Li H, Zhang L. Effect of mitophagy in oocytes and granulosa cells on oocyte quality. *Biol Reprod.* (2021) 104:294–304. doi: 10.1093/biolre/iaoa194
19. Anderson E, Wilkinson RF, Lee G, Meller S. A correlative microscopical analysis of differentiating ovarian follicles of mammals. *J Morphol.* (1978) 156:339–66. doi: 10.1002/jmor.1051560303
20. Su YQ, Sugiura K, Eppig JJ. Mouse oocyte control of granulosa cell development and function: paracrine regulation of cumulus cell metabolism. *Semin Reprod Med.* (2009) 27:32–42. doi: 10.1055/s-0028-1108008
21. Schultz RM. Roles of cell-to-cell communication in development. *Biol Reprod.* (1985) 32:27–42. doi: 10.1095/biolreprod32.1.27
22. Havelock JC, Rainey WE, Carr BR. Ovarian granulosa cell lines. *Mol Cell Endocrinol.* (2004) 228:67–78. doi: 10.1016/j.mce.2004.04.018
23. Färkkilä A, Haltia UM, Tapper J, McConechy MK, Huntsman DG, Heikinheimo M. Pathogenesis and treatment of adult-type granulosa cell tumor of the ovary. *Ann Med.* (2017) 49:435–47. doi: 10.1080/07853890.2017.1294760
24. Yildiz BO, Bozdogan G, Yapici Z, Esinler I, Yarali H. Prevalence, phenotype and cardiometabolic risk of polycystic ovary syndrome under different diagnostic criteria. *Hum Reprod.* (2012) 27:3067–73. doi: 10.1093/humrep/des232
25. Peng SL, Wu QF, Xie Q, Tan J, Shu KY. *PATL2* regulated the apoptosis of ovarian granulosa cells in patients with PCOS. *Gynecol Endocrinol.* (2021) 37:629–34. doi: 10.1080/09513590.2021.1928066
26. Dilaver N, Pellatt L, Jameson E, Ogunjimi M, Bano G, Homburg R, et al. The regulation and signalling of anti-Müllerian hormone in human granulosa cells: relevance to polycystic ovary syndrome. *Hum Reprod.* (2019) 34:2467–79. doi: 10.1093/humrep/dez214
27. Zhao H, Zhao Y, Li T, Li M, Li J, Li R, et al. Metabolism alteration in follicular niche: the nexus among intermediary metabolism, mitochondrial function, and classic polycystic ovary syndrome. *Free Radic Biol Med.* (2015) 86:295–307. doi: 10.1016/j.freeradbiomed.2015.05.013
28. Halliwell B. Antioxidants in human health and disease. *Annu Rev Nutr.* (1996) 16:33–50. doi: 10.1146/annurev.nu.16.070196.000341
29. Allen RG, Tresini M. Oxidative stress and gene regulation. *Free Radic Biol Med.* (2000) 28:463–99. doi: 10.1016/S0891-5849(99)00242-7
30. Al-Gubory KH, Fowler PA, Garrel C. The roles of cellular reactive oxygen species, oxidative stress and antioxidants in pregnancy outcomes. *Int J Biochem Cell Biol.* (2010) 42:1634–50. doi: 10.1016/j.biocel.2010.06.001
31. Johnson LJ, Meacham SL, Kruskall LJ. The antioxidants--vitamin C, vitamin E, selenium, and carotenoids. *J Agromedicine.* (2003) 9:65–82. doi: 10.1300/J096v09n01_07
32. Mohammadi M. Oxidative stress and polycystic ovary syndrome: a brief review. *Int J Prev Med.* (2019) 10:86. doi: 10.4103/ijpvm.IJPVM_576_17
33. Deepika ML, Nalini S, Maruthi G, Ramchander V, Ranjith K, Latha KP, et al. Analysis of oxidative stress status through MN test and serum MDA levels in PCOS women. *Pak J Biol Sci.* (2014) 17:574–7. doi: 10.3923/pjbs.2014.574.577
34. Palacio JR, Iborra A, Ulcova-Gallova Z, Badia R, Martinez P. The presence of antibodies to oxidative modified proteins in serum from polycystic ovary syndrome patients. *Clin Exp Immunol.* (2006) 144:217–22. doi: 10.1111/j.1365-2249.2006.03061.x
35. Năcul AP, Andrade CD, Schwarz P, de Bittencourt PI Jr, Spritzer PM. Nitric oxide and fibrinogen in polycystic ovary syndrome: associations with insulin resistance and obesity. *Eur J Obstet Gynecol Reprod Biol.* (2007) 133:191–6. doi: 10.1016/j.ejogrb.2006.09.009
36. BASKOL G, AYGEN E, ERDEM F, CANIKLIOĞLU A, NARIN F, ŞAHİN Y, et al. Assessment of paraoxonase 1, xanthine oxidase and glutathione peroxidase activities, nitric oxide and thiol levels in women with polycystic ovary syndrome. *Acta Obstet Gynecol Scand.* (2012) 91:326–30. doi: 10.1111/j.1600-0412.2011.01337.x
37. Kohzadi M, Khazaei MR, Choobaz F, Khazaei M. Relationship between serum levels of anti-Müllerian hormone, adiponectin and oxidative stress markers in patients with polycystic ovary syndrome. *Int J Fertil Steril.* (2020) 14:27–33. doi: 10.22074/ijfs.2020.5809
38. Christakou C, Economou F, Livadas S, Piperi C, Adamopoulos C, Marinakis E, et al. Strong and positive association of endothelin-1 with AGEs in PCOS: a causal relationship or a bystander. *Hormones.* (2011) 10:292–7. doi: 10.14310/horm.2002.1320
39. Isik H, Aymoglu O, Timur H, Sahbaz A, Harma M, Can M, et al. Is xanthine oxidase activity in polycystic ovary syndrome associated with inflammatory and cardiovascular risk factors. *J Reprod Immunol.* (2016) 116:98–103. doi: 10.1016/j.jri.2016.06.002
40. Gong Y, Luo S, Fan P, Jin S, Zhu H, Deng T, et al. Growth hormone alleviates oxidative stress and improves oocyte quality in Chinese women with polycystic ovary syndrome: a randomized controlled trial. *Sci Rep.* (2020) 10:18769. doi: 10.1038/s41598-020-75107-4
41. Murri M, Luque-Ramírez M, Insenser M, Ojeda-Ojeda M, Escobar-Morreale HF. Circulating markers of oxidative stress and polycystic ovary syndrome (PCOS): a systematic review and meta-analysis. *Hum Reprod Update.* (2013) 19:268–88. doi: 10.1093/humupd/dms059
42. Garg D, Merhi Z. Relationship between advanced glycation end products and steroidogenesis in PCOS. *Reprod Biol Endocrinol.* (2016) 14:71. doi: 10.1186/s12958-016-0205-6
43. Tantalaki E, Piperi C, Livadas S, Kollias A, Adamopoulos C, Koulouri A, et al. Impact of dietary modification of advanced glycation end products (AGEs) on the hormonal and metabolic profile of women with polycystic ovary syndrome (PCOS). *Hormones.* (2014) 13:65–73. doi: 10.1007/BF03401321
44. Seleem AK, El Refaey AA, Shaalan D, Sherbiny Y, Badawy A. Superoxide dismutase in polycystic ovary syndrome patients undergoing intracytoplasmic sperm injection. *J Assist Reprod Genet.* (2014) 31:499–504. doi: 10.1007/s10815-014-0190-7
45. Uçkan K, Demir H, Turan K, Sarikaya E, Demir C. Role of oxidative stress in obese and nonobese PCOS patients. *Int J Clin Pract.* (2022) 2022:4579831. doi: 10.1155/2022/4579831
46. Fortune JE. Ovarian follicular growth and development in mammals. *Biol Reprod.* (1994) 50:225–32. doi: 10.1095/biolreprod50.2.225
47. Endo M, Kimura K, Kuwayama T, Monji Y, Iwata H. Effect of estradiol during culture of bovine oocyte-granulosa cell complexes on the mitochondrial DNA copies of oocytes and telomere length of granulosa cells. *Zygote.* (2014) 22:431–9. doi: 10.1017/S0967199412000603
48. Kanafchian M, Esmaeilzadeh S, Mahjoub S, Rahsepar M, Ghasemi M. Status of serum copper, magnesium, and Total antioxidant capacity in patients with polycystic ovary syndrome. *Biol Trace Elem Res.* (2020) 193:111–7. doi: 10.1007/s12011-019-01705-7
49. Wang Y, Yang Q, Wang H, Zhu J, Cong L, Li H, et al. NAD⁺ deficiency and mitochondrial dysfunction in granulosa cells of women with polycystic ovary syndrome. *Biol Reprod.* (2021) 105:371–80. doi: 10.1093/biolre/iaob078
50. Kurdoglu Z, Ozkol H, Tuluce Y, Koyuncu I. Oxidative status and its relation with insulin resistance in young non-obese women with polycystic ovary syndrome. *J Endocrinol Investig.* (2012) 35:317–21. doi: 10.3275/7682
51. Zhang D, Luo WY, Liao H, Wang CF, Sun Y. The effects of oxidative stress to PCOS. *Sichuan Da Xue Xue Bao Yi Xue Ban.* (2008) 39:421–3. doi: 10.3969/j.issn.1672-173X.2008.03.019
52. Sabuncu T, Vural H, Harma M, Harma M. Oxidative stress in polycystic ovary syndrome and its contribution to the risk of cardiovascular disease. *Clin Biochem.* (2001) 34:407–13. doi: 10.1016/S0009-9120(01)00245-4
53. Savic-Radojevic A, Bozic Antic I, Coric V, Bjekic-Macut J, Radic T, Zarkovic M, et al. Effect of hyperglycemia and hyperinsulinemia on glutathione peroxidase activity in non-obese women with polycystic ovary syndrome. *Hormones.* (2014) 14:101–8. doi: 10.14310/horm.2002.1525
54. Gupta S, Finelli R, Agarwal A, Henkel R. Total antioxidant capacity-relevance, methods and clinical implications. *Andrologia.* (2021) 53:e13624. doi: 10.1111/and.13624
55. Mancini A, Festa R, Di Donna V, Leone E, Littarru GP, Silvestrini A, et al. Hormones and antioxidant systems: role of pituitary and pituitary-dependent axes. *J Endocrinol Investig.* (2010) 33:422–33. doi: 10.1007/BF03346615
56. Fencki V, Fencki S, Yilmazer M, Serteser M. Decreased total antioxidant status and increased oxidative stress in women with polycystic ovary syndrome may contribute to the risk of cardiovascular disease. *Fertil Steril.* (2003) 80:123–7. doi: 10.1016/S0015-0282(03)00571-5
57. Verit FF, Erel O. Oxidative stress in nonobese women with polycystic ovary syndrome: correlations with endocrine and screening parameters. *Gynecol Obstet Investig.* (2008) 65:233–9. doi: 10.1159/000113046
58. Dinger Y, Akcay T, Erdem T, Ilker Saygili E, Gundogdu S. DNA damage, DNA susceptibility to oxidation and glutathione level in women with polycystic ovary syndrome. *Scand J Clin Lab Invest.* (2005) 65:721–8. doi: 10.1080/00365510500375263

59. Hassa PO, Haenni SS, Elser M, Hottiger MO. Nuclear ADP-ribosylation reactions in mammalian cells: where are we today and where are we going. *Microbiol Mol Biol Rev.* (2006) 70:789–829. doi: 10.1128/MMBR.00040-05
60. Rajani S, Chattopadhyay R, Goswami SK, Ghosh S, Sharma S, Chakravarty B. Assessment of oocyte quality in polycystic ovarian syndrome and endometriosis by spindle imaging and reactive oxygen species levels in follicular fluid and its relationship with IVF-ET outcome. *J Hum Reproduct Sci.* (2012) 5:187–93. doi: 10.4103/0974-1208.101020
61. Palomba S, Daolio J, La Sala GB. Oocyte competence in women with polycystic ovary syndrome. *Trends Endocrinol Metab.* (2017) 28:186–98. doi: 10.1016/j.tem.2016.11.008
62. Nikbakht R, Mohammadjafari R, Rajabalipour M, Moghadam MT. Evaluation of oocyte quality in polycystic ovary syndrome patients undergoing ART cycles. *Fertil Res Pract.* (2021) 7:2. doi: 10.1186/s40738-020-00094-z
63. Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocr Rev.* (2012) 33:981–1030. doi: 10.1210/er.2011-1034
64. Rashidi B, Haghollahi F, Shariat M, Zayerii F. The effects of calcium-vitamin D and metformin on polycystic ovary syndrome: a pilot study. *Taiwan J Obstet Gynecol.* (2009) 48:142–7. doi: 10.1016/S1028-4559(09)60275-8
65. Dalle-Donne I, Aldini G, Carini M, Colombo R, Rossi R, Milzani A. Protein carbonylation, cellular dysfunction, and disease progression. *J Cell Mol Med.* (2006) 10:389–406. doi: 10.1111/j.1582-4934.2006.tb00407.x
66. Kaya C, Erkan AF, Cengiz SD, Dunder I, Demirel ÖE, Bilgihan A. Advanced oxidation protein products are increased in women with polycystic ovary syndrome: relationship with traditional and nontraditional cardiovascular risk factors in patients with polycystic ovary syndrome. *Fertil Steril.* (2009) 92:1372–7. doi: 10.1016/j.fertnstert.2008.08.016
67. Abuja PM, Albertini R. Methods for monitoring oxidative stress, lipid peroxidation and oxidation resistance of lipoproteins. *Clin Chim Acta.* (2001) 306:1–17. doi: 10.1016/S0009-8981(01)00393-X
68. González F, Rote NS, Minium J, Kirwan JP. Reactive oxygen species-induced oxidative stress in the development of insulin resistance and hyperandrogenism in polycystic ovary syndrome. *J Clin Endocrinol Metab.* (2006) 91:336–40. doi: 10.1210/jc.2005-1696
69. Andrieux P, Chevillard C, Cunha-Neto E, Nunes J. Mitochondria as a cellular hub in infection and inflammation. *Int J Mol Sci.* (2021):22. doi: 10.3390/ijms222111338
70. Bhatti JS, Bhatti GK, Reddy PH. Mitochondrial dysfunction and oxidative stress in metabolic disorders—a step towards mitochondria based therapeutic strategies. *Biochim Biophys Acta Mol Basis Dis.* (2017) 1863:1066–77. doi: 10.1016/j.bbdis.2016.11.010
71. Cozzolino M, Herraiz S, Titus S, Roberts L, Romeu M, Peinado I, et al. Transcriptomic landscape of granulosa cells and peripheral blood mononuclear cells in women with PCOS compared to young poor responders and women with normal response. *Hum Reprod.* (2022) 37:1274–86. doi: 10.1093/humrep/deac069
72. Graier WF, Trenker M, Malli R. Mitochondrial Ca²⁺, the secret behind the function of uncoupling proteins 2 and 3. *Cell Calcium.* (2008) 44:36–50. doi: 10.1016/j.ceca.2008.01.001
73. Tang JX, Thompson K, Taylor RW, Oláhová M. Mitochondrial OXPHOS biogenesis: co-regulation of protein synthesis, import, and assembly pathways. *Int J Mol Sci.* (2020) 21:3820. doi: 10.3390/ijms21113820
74. Guo R, Gu J, Zong S, Wu M, Yang M. Structure and mechanism of mitochondrial electron transport chain. *Biom J.* (2018) 41:9–20. doi: 10.1016/j.bj.2017.12.001
75. Johnston IG, Burgstaller JP. Evolving mtDNA populations within cells. *Biochem Soc Trans.* (2019) 47:1367–82. doi: 10.1042/BST20190238
76. Eriksen MB, Minet AD, Glinborg D, Gaster M. Intact primary mitochondrial function in myotubes established from women with PCOS. *J Clin Endocrinol Metab.* (2011) 96:E1298–302. doi: 10.1210/jc.2011-0278
77. Lee SH, Chung DJ, Lee HS, Kim TJ, Kim MH, Jeong HJ, et al. Mitochondrial DNA copy number in peripheral blood in polycystic ovary syndrome. *Metabolism.* (2011) 60:1677–82. doi: 10.1016/j.metabol.2011.04.010
78. Rabøl R, Svendsen PF, Skovbro M, Boushel R, Schjerling P, Nilas L, et al. Skeletal muscle mitochondrial function in polycystic ovarian syndrome. *Eur J Endocrinol.* (2011) 165:631–7. doi: 10.1530/EJE-11-0419
79. Au HK, Lin SH, Huang SY, Yeh TS, Tzeng CR, Hsieh RH. Deleted mitochondrial DNA in human luteinized granulosa cells. *Ann N Y Acad Sci.* (2005) 1042:136–41. doi: 10.1196/annals.1338.014
80. Dabrowski SA, Nikiforov NG, Eid AH, Nedosugova LV, Starodubova AV, Popkova TV, et al. Mitochondrial dysfunction and chronic inflammation in polycystic ovary syndrome. *Int J Mol Sci.* (2021) 22:3923. doi: 10.3390/ijms22083923
81. von Mengden L, Klamt F, Smitz J. Redox biology of human cumulus cells: basic concepts, impact on oocyte quality, and potential clinical use. *Antioxid Redox Signal.* (2020) 32:522–35. doi: 10.1089/ars.2019.7984
82. Karuputhala NB, Chattopadhyay R, Chakravarty B, Chaudhury K. Oxidative status in granulosa cells of infertile women undergoing IVF. *Syst Biol Reprod Med.* (2013) 59:91–8. doi: 10.3109/19396368.2012.743197
83. Hsu AL, Townsend PM, Oehninger S, Castora FJ. Endometriosis may be associated with mitochondrial dysfunction in cumulus cells from subjects undergoing in vitro fertilization-intracytoplasmic sperm injection, as reflected by decreased adenosine triphosphate production. *Fertil Steril.* (2015) 103:347–52.e1. doi: 10.1016/j.fertnstert.2014.11.002
84. Zhang J, Bao Y, Zhou X, Zheng L. Polycystic ovary syndrome and mitochondrial dysfunction. *Reprod Biol Endocrinol.* (2019) 17:67. doi: 10.1186/s12958-019-0509-4
85. Miller WL. Disorders in the initial steps of steroid hormone synthesis. *J Steroid Biochem Mol Biol.* (2017) 165:18–37. doi: 10.1016/j.jsbmb.2016.03.009
86. Sreerangaraja Urs DB, Wu WH, Komrskova K, Postlerova P, Lin YE, Tzeng CR, et al. Mitochondrial function in modulating human granulosa cell steroidogenesis and female fertility. *Int J Mol Sci.* (2020) 21:3592. doi: 10.3390/ijms21103592
87. Wang L, Qu G, Dong X, Huang K, Kumar M, Ji L, et al. Long-term effects of methamphetamine exposure in adolescent mice on the future ovarian reserve in adulthood. *Toxicol Lett.* (2016) 242:1–8. doi: 10.1016/j.toxlet.2015.11.029
88. Shi L, Zhang J, Lai Z, Tian Y, Fang L, Wu M, et al. Long-term moderate oxidative stress decreased ovarian reproductive function by reducing follicle quality and progesterone production. *PLoS One.* (2016) 11:e0162194. doi: 10.1371/journal.pone.0162194
89. Sutton-McDowall ML, Gilchrist RB, Thompson JG. The pivotal role of glucose metabolism in determining oocyte developmental competence. *Reproduction.* (2010) 139:685–95. doi: 10.1530/REP-09-0345
90. Zhao R, Jiang Y, Zhao S, Zhao H. Multiomics analysis reveals molecular abnormalities in granulosa cells of women with polycystic ovary syndrome. *Front Genet.* (2021) 12:648701. doi: 10.3389/fgene.2021.648701
91. Stefano GB, Kream RM. Glycolytic coupling to mitochondrial energy production ensures survival in an oxygen rich environment. *Med Sci Monit.* (2016) 22:2571–5. doi: 10.12659/MSM.899610
92. Laganà AS, Rossetti P, Buscema M, La Vignera S, Condorelli RA, Gullo G, et al. Metabolism and ovarian function in PCOS women: a therapeutic approach with Inositols. *Int J Endocrinol.* (2016) 2016:6306410. doi: 10.1155/2016/6306410
93. Taylor CT, Scholz CC. The effect of HIF on metabolism and immunity. *Nat Rev Nephrol.* (2022) 18:573–87. doi: 10.1038/s41581-022-00587-8
94. Wang J, Wu X. The effects of mitochondrial dysfunction on energy metabolism switch by HIF-1 α signalling in granulosa cells of polycystic ovary syndrome. *Endokrynol Pol.* (2020) 71:134–45. doi: 10.5603/EPa2020.0002
95. Mazloomi S, Farimani MS, Tavilani H, Karimi J, Amiri I, Abbasi E, et al. Granulosa cells from immature follicles exhibit restricted glycolysis and reduced energy production: a dominant problem in polycystic ovary syndrome. *J Assist Reprod Genet.* (2023) 40:343–59. doi: 10.1007/s10815-022-02676-w
96. Tatone C, Di Emidio G, Barbonetti A, Carta G, Luciano AM, Falone S, et al. Sirtuins in gamete biology and reproductive physiology: emerging roles and therapeutic potential in female and male infertility. *Hum Reprod Update.* (2018) 24:267–89. doi: 10.1093/humupd/dmy003
97. Pacella-Ince L, Zander-Fox DL, Lan M. Mitochondrial SIRT3 and its target glutamate dehydrogenase are altered in follicular cells of women with reduced ovarian reserve or advanced maternal age. *Hum Reprod.* (2014) 29:1490–9. doi: 10.1093/humrep/deu071
98. Safaei Z, Bakhshalizadeh S, Nasr-Esfahani MH, Akbari Sene A, Najafzadeh V, Soleimani M, et al. Vitamin D3 affects mitochondrial biogenesis through mitogen-activated protein kinase in polycystic ovary syndrome mouse model. *J Cell Physiol.* (2020) 235:6113–26. doi: 10.1002/jcp.29540
99. Azhar A, Haider G, Naseem Z, Farooqui N, Farooqui MU, Rehman R. Morphological changes in the experimental model of polycystic ovary syndrome and effects of vitamin D treatment. *J Obstet Gynaecol Res.* (2021) 47:1164–71. doi: 10.1111/jog.14671
100. Ding Y, Jiang Z, Xia B, Zhang L, Zhang C, Leng J. Mitochondria-targeted antioxidant therapy for an animal model of PCOS-IR. *Int J Mol Med.* (2019) 43:316–24. doi: 10.3892/ijmm.2018.3977
101. Yi S, Zheng B, Zhu Y, Cai Y, Sun H, Zhou J. Melatonin ameliorates excessive PINK1/Parkin-mediated mitophagy by enhancing SIRT1 expression in granulosa cells of PCOS. *Am J Physiol Endocrinol Metab.* (2020) 319:E91–E101. doi: 10.1152/ajpendo.00006.2020
102. Sari U, Kaygusuz I, Kafali H. Is pentraxin 3 a new cardiovascular risk marker in polycystic ovary syndrome. *Gynecol Obstet Investig.* (2014) 78:173–8. doi: 10.1159/000363744
103. Pan J, Zhou C, Zhou Z, Yang Z, Dai T, Huang H, et al. Elevated ovarian pentraxin 3 in polycystic ovary syndrome. *J Assist Reprod Genet.* (2021) 38:1231–7. doi: 10.1007/s10815-021-02105-4
104. Dong Z, Huang M, Liu Z, Xie P, Dong Y, Wu X, et al. Focused screening of mitochondrial metabolism reveals a crucial role for a tumor suppressor Hbp1 in ovarian reserve. *Cell Death Differ.* (2016) 23:1602–14. doi: 10.1038/cdd.2016.47
105. He Q, Gu L, Lin Q, Ma Y, Liu C, Pei X, et al. The Imp21 mutation causes ovarian aging through ROS-Wnt/ β -catenin-estrogen pathway: preventive effect of melatonin. *Endocrinology.* (2020) 161:bqaa119. doi: 10.1210/endo/bqaa119
106. Gupta S, Ghulmiyyah J, Sharma R, Halabi J, Agarwal A. Power of proteomics in linking oxidative stress and female infertility. *Bio Med Res Int.* (2014) 2014:916212. doi: 10.1155/2014/916212