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\*CORRESPONDENCE Mateusz Kamil Ożóg [mateusz.ozog@sum.edu.pl](mailto:mateusz.ozog@sum.edu.pl)

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# [Pathophysiology and clinical](https://www.frontiersin.org/articles/10.3389/fmed.2023.1121270/full)  [aspects of epiretinal membrane –](https://www.frontiersin.org/articles/10.3389/fmed.2023.1121270/full)  [review](https://www.frontiersin.org/articles/10.3389/fmed.2023.1121270/full)

Mateusz Kamil Ożóg<sup>1,2\*</sup>, Marta Nowak-Wąs<sup>1,3</sup> and Wojciech Rokicki<sup>3,4</sup>

1 Department of Histology and Cell Pathology, Faculty of Medical Sciences in Zabrze, Medical University of Silesia, Katowice, Poland, 2Department of Histology, Cytophysiology and Embryology, Faculty of Medicine, Academy of Silesia, Zabrze, Poland, <sup>3</sup>Department of Ophthalmology, Kornel Gibiński University Clinical Center, Medical University of Silesia, Katowice, Poland, 4Department of Ophthalmology, Faculty of Medical Sciences in Katowice, Medical University of Silesia, Katowice, Poland

The epiretinal membrane (ERM) is a pathological tissue formed at the vitreoretinal interface. The formation of this tissue is associated with numerous symptoms related to disturbances of vision. These types of lesions may arise idiopathically or be secondary to eye diseases, injuries and retinal surgeries. ERM tissue contains numerous cell types and numerous cytokines, which participate in its formation. The aim of this paper is to summarize information about the etiology, epidemiology, pathophysiology and treatment of ERM, with a brief description of the main cells that build the ERM – as well as the cytokines and molecules related to ERM pathogenesis – being provided in addition.

### KEYWORDS

epiretinal membrane, macular pucker, myofibroblasts, pre-macular fibrosis, retina, cellophane maculopathy

## 1. Introduction

The epiretinal membrane (ERM), commonly known as macular pucker or cellophane maculopathy, is a pathological tissue formed at the junction of the vitreous body and the retina – the vitreoretinal interface. The formation of this tissue is associated with symptoms related to visual disturbances. The lesions of this type may arise idiopathically or be secondary to eye diseases, injuries and retinal surgery. In the case of idiopathic changes, a number of factors may favor the development of these lesions [\(1](#page-7-0)).

ERM tissue contains many cell types deriving from different parts of eyeball: retinal pigment epithelium (RPE) cells, fibrocytes, fibrous astrocytes, myofibroblast-like cells, glial cells, endothelial cells (ECs), and macrophages. The cellular composition of this tissue varies individually and depends on the cause of the lesions and the participation of numerous cytokines such as growth factors, tumor necrosis factor (TGF) or chemokines [\(2](#page-7-1)).

The aim of this paper is to summarize information on the etiology, epidemiology, pathophysiology, and treatment of ERM.

## 2. Pathophysiology of ERM

The vitreous body, consisting of a colorless, highly hydrated gel matrix, fills the space called the vitreous chamber located posteriorly to the ciliary body. The vitreous body adheres loosely to the retina, connecting most strongly around the ora serrata and the optic nerve [\(3\)](#page-7-2). Its outer layer – vitreous cortex – is made of collagen, while the inside is filled with vitreous humor

containing mainly water (98–99%), fibrous protein called vitrosin, type II collagen fibers, glycosaminoglycans, hyaluronates, opticins and other proteins [\(4](#page-7-3)).

The vitreous body contains a small number of cells, mainly phagocytes, that remove cellular debris and hyalocytes the main function of which is to produce hyaluronans [\(5\)](#page-7-4). Vitreous body is involved in maintaining proper intraocular pressure, protecting the lens against oxidative stress and is one of the optical centers ([3](#page-7-2), [6](#page-7-5)).

With age, an increase in liquefaction and fiber aggregation occurs, one which may lead to many ophtalmic diseases ([7,](#page-7-6) [8](#page-7-7)). As a result of these processes, the volume of the vitreous body decreases, the said body collapses, and the collagen fibers reorganize. This leads to changes in the shape of the vitreous body and posterior vitreous detachment (PVD). If this process is not complete, and macula and vitreous body come into contact, a posterior vitreomacular adhesion (VMA) is formed at the point of this contact. This process may lead to the formation of vitreomacular traction (VMT), which involves foveal contour distortion and retinal layer disorders, with a possible elevation of the retina above the pigment epithelium (RPE), not interrupting, however, the retina's continuity. In this state most patients self-heal due to completion of PVD. When VMT is not self-healed and a hole in the macula occurs, the next step in the evolution of the pathology could be vitreoschisis, which occurs in the case of the half of PVD patients. During this process posterior vitreous cortex splits, leaving the outermost layer attached to the macula, while the remainder of the vitreous collapses forward. This can lead to the proliferation within the retinal vitreous residue, i.e., the formation of an epiretinal membrane (ERM) ([9,](#page-7-8) [10](#page-7-9)) [\(Figure 1\)](#page-1-0).

Formed ERM tissue can cause macular edema or distortion of the vitreoretinal interface, which may lead to visual disturbances, including blurred vision and a reduction of visual acuity ([11](#page-7-10)) ([Figure 2\)](#page-2-0).

<span id="page-1-0"></span>

### 2.1. ERM tissue formation

Pathological cell proliferation on the internal limiting membrane (ILM) surface is the basis for the formation of pathological tissue at the interface between the vitreous body and the retina. As a result of PVD, ILM dehiscence occurs, which leads to the migration of microglial cells to the surface. Microglial cells then interact with the neighboring hyalocytes and laminocytes of the vitreous cellular membrane [\(12](#page-7-11)). These cells then differentiate into fibroblast-like cells, which are directly responsible for the formation of the collagen scaffolding of the ERM [\(13](#page-7-12), [14\)](#page-7-13) ([Figure 3\)](#page-2-1).

ERMs can arise idiopathically or secondary to certain disease states, and as a result of a trauma or surgery within the eyeball.

In the case of the idiopathic ERM retinal glial cells, hyalocytes, fibroblasts, and myofibroblasts generated by cell migration and differentiation of microglial cells, hyalocytes, and laminocytes predominate ([2,](#page-7-1) [14](#page-7-13)).

In secondary ERM, the presence of the retinal pigment epithelial cells, macrophages, T cells, and B cells is observed due to inflammation appearing in the etiology of this lesion [\(15](#page-7-14), [16\)](#page-7-15).

The leading theory presents the sequence of changes leading to the emergence of the ERM thus:

- 1) Microglial cells migrate to the surface of the retina as a consequence of PVD and the resulting formation of the cracks within the ILM.
- 2) The fragments of the vitreous membrane remaining on the surface of the ILM contain hyalocytes, which, due to contact with microglial cells, differentiate into myofibroblasts, while microglial cells may differentiate into fibroblasts.
- 3) PVD-induced ILM avulsion enhances the action of some ERM-promoting cytokines [\(17–](#page-7-16)[19](#page-7-17)).

### 2.2. Idiopathic ERM

The idiopathic form of ERM affects approximately 95% of patients and is directly related to cellular proliferation resulting from PVD.

It is possible to distinguish 2 types of iERM. Type I is formed due to collagen of the vitreous body coming into contact with the internal limiting membrane (ILM) of the retina, which results in the production of a collagen membrane separating the two structures. Type II, on the other hand, results from cell proliferation that takes place directly on the ILM surface with or without a small layer of collagen in between [\(20\)](#page-7-18).

## 2.3. Secondary ERM

The secondary form of ERM develops in the course of eye diseases or as a result of injuries of the eyeball or the surgeries performed on it.

As a result of these disorders, inflammation within the retina occurs, which leads to an inflammatory infiltration and, ultimately, a migration of cells from other layers of the retina to the ERM being in the process of formation  $(21)$  ([Table 1\)](#page-3-0).

<span id="page-2-0"></span>

### FIGURE 2

Transmission electron micrograph showing epiretinal membrane (ERM) and internal limiting membrane (ILM). a – original micrograph showing comparison of ERM and ILM structure (original magnification x4200), b – micrograph showing detailed structure of ERM and phagocytes: N – cell nuclei, asterix (\*) – vacuoles, arrowhead (^) – dense granules (magnificated and focused micrograph a).

<span id="page-2-1"></span>

### FIGURE 3

Transmission electron micrograph showing collagen fibers in epiretinal membrane. ERM – epiretinal membrane, ILM – internal limiting membrane, c – collagen fibers.

## 3. Histopatology of ERM

## 3.1. The cells that build ERM

ERM is usually made up of two layers placed on the ILM. The outer layer directly overlying the ILM consists of randomly oriented

<span id="page-3-0"></span>



<span id="page-3-1"></span>TABLE 2 Cells that built epiretinal membranes.

proteins, while the inner layer consists of one or more layers of cells from the retina [\(22](#page-7-20)). The cells observed are glial cells, hyalocytes, RPE cells, macrophages, fibroblasts, and myofibroblast-like cells. Apart from the latter type, the source of the cells within the ERM is uncertain. It is known, however, that myofibroblast-like cells are formed by differentiating from other cell types within ERM [\(22,](#page-7-20) [23\)](#page-7-21) ([Table 2\)](#page-3-1).

### 3.2. Cytokines and molecules related to ERM pathogenesis

A number of cytokines and molecules related to this process which directly or indirectly influence the development of this pathology have been identified. These cells are responsible for the production of proteins that constitute the ERM extracellular matrix ([31](#page-7-22)) [\(Table 3](#page-4-0)).

## 4. Epidemiology

The main risk factors for ERM are age and PVD. PVD is observed in the case of 70% of patients in the initial stages of the disease. Although PVD can appear early in life or childhood, ERM usually appears after the age of 50, and the risk of its appearance increases exponentially with age.

Gender was not observed to exert any influence as to the risk of the appearance of ERM, although some studies indicate a slight predominance of women among the afflicted [\(60\)](#page-8-0). Statistically, significant differences in risk of occurrence seem to emerge due to ethnicity – in studies concerning the American society, the highest number of ERM cases is identified in the population of Chinese origin, followed by the ethnic groups of, respectively, Latin American, Caucasian and African descent.

Geographic differences were also observed to play a part: ERM is more common in Europe and South America, and the



### <span id="page-4-0"></span>TABLE 3 Cytokines and molecules involved in epiretinal formation.



least common in Asia. This phenomenon is probably related to lifestyle and diet. Other factors contributing to the development of ERM are obesity, type II diabetes, hypertension and hypercholesterolaemia ([60](#page-8-0), [61\)](#page-8-1).

## 5. Symptoms and diagnosis

The symptoms reported by patients depend on the stage (phase of development) and type of ERM. In some cases, the presence of ERM does not produce clinical symptoms and is diagnosed accidentally. Patients usually complain of: visual disturbances – metamorphopsia, micropsia or macropsia, photopsia, decreased visual acuity, diplopia, and a loss of central vision. The diagnosis of ERM is based on a clinical examination and Optical Coherence Tomography (OCT). In the case of fundoscopy, a cellophane reflex or wrinkling on the retinal surface resulting from the contracture of the membrane can be observed. It usually affects the foveal and parafoveal area. Cystoid macular oedema (CMO), lamellar or full-thickness macular holes (MHs) and/or small retinal hemorrhages can be seen in association with ERM. The diagnosis of idiopathic ERM is based on the exclusion of other ophthalmic diseases like retinal vascular diseases including diabetic retinopathy, retinal vein occlusion, uveitis and other inflammatory diseases, trauma, intraocular tumors, and retinal tear or detachment [\(62\)](#page-8-2).

There are several ERM classification systems based on OCT findings. No classification, however, is currently suggested for general use in clinical practice ([63](#page-8-3)) [\(Figure 4](#page-5-0)).

<span id="page-5-0"></span>

Stages of ERM showed on optical coherence tomography (OCT) images. I – ERMs are mild and thin. Foveal depression is present. II – ERMs with a widening of the outer nuclear layer and loss of the foveal depression. III – ERMs with continuous ectopic inner foveal layers crossing the entire foveal area. IV – ERMs are thick with continuous ectopic inner foveal layers and disrupted retinal layers. Based on clasification by Govetto et al. ([64](#page-8-14)).

<span id="page-5-1"></span>



From the additional diagnostic tools, one worth mentioning is fluorescein angiography (FA), which is useful in the case of secondary ERM when it comes to identifying preoperatively the underlying cause of like intraocular tumors or retinal vascular diseases. Macular edema can also be confirmed with angiography ([62](#page-8-2)). The OCT examination allows not only to deepen the diagnosis, but also to distinguish different stages of the ERM development [\(64\)](#page-8-14) ([Table 4](#page-5-1) and [Figure 5\)](#page-6-0).

## 6. ERM treatment

The management options for ERM are limited and consist of observation or surgical intervention. Official guidelines for performing the surgical ERM removal have not been established. Before taking appropriate intervention measures, it is advisable to discuss with the patient all the possible benefits and complications related to the surgery in relation to the severity of the patient's symptoms and their lifestyle [\(65\)](#page-8-15).

During the surgical intervention called pars plana vitrectomy (PPV), the epiretinal membrane is removed and the retinal tractions are released. ILM is considered to be the scaffold for myofibroblast proliferation, thus it is commonly removed alongside with ERM, in order to minimize the risk of ERM recurrence. Sometimes PPV is combined with a simultaneous cataract surgery involving intraocular lens implantation (phacovitrectomy). During the surgery, special dyes, like triamcinolone acetonide, trypan blue, indocyanine green (ICG), and brilliant blue, are used to distinguish ERM from the retinal layers ([66](#page-8-16)).

It is possible to discern two types of PPV – complete vitrectomy, which involves the whole vitreous body being detached from the retina and removed, and limited vitrectomy, which involves removing only the central part of the vitreous body. Most of the times, the complete vitrectomy is performed, however, there is no difference when it comes to the results of it and the limited vitrectomy. Limited vitrectomy is usually faster and potentially produces fewer long-term side effects ([67](#page-8-17)).

<span id="page-6-0"></span>

The surgical treatment of ERM provides excellent postoperative visual outcomes and is a relatively safe procedure. Improvement in the vascularity of the choroid was also observed ([68\)](#page-8-18). Like any other surgical intervention, ERM surgery can cause complications, such as endophtalmitis, retinal detachment or ERM recurrence [\(69\)](#page-8-19).

## 7. Conclusion

It is estimated that the main risk factor that significantly preceding the development of ERM, i.e., PDV, occurs in about 2% of the population. The main epidemiological factor associated with PDV is old age ([60](#page-8-0), [61](#page-8-1), [70](#page-8-20)). An increased incidence of ERM is therefore to be expected due to the aging of the population. By understanding the successive processes leading to the formation of ERM, we better understand the causes of not only this pathology but also PDV. Currently, it is not possible to predict the development of this pathology based on biochemical tests, while the development of knowledge about the histological structure of ERM and the expression of cytokines and molecules related to ERM pathophysiology may in the future allow for the detection of a biochemical marker allowing for early detection of ERM development without the need for costly OCT, development guidelines for qualifying patients for surgical treatment and a potential conservative treatment regimen. Currently, there is also no prophylaxis for idiopathic ERM development, but by understanding the inflammatory factors involved in this process, it will be possible to develop some. In the case of secondary ERM, in

addition to reducing the risk by appropriate treatment of the cause, it is possible, thanks to potential biochemical markers, to include some of these patients in regular observation due to high risk of ERM development.

## Author contributions

MO and MN-W: conceptualization and writing—original draft preparation. WR: writing—review and editing, and supervision. All authors contributed to the article and approved the submitted version.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## References

<span id="page-7-0"></span>1. Kwok AK, Lai TY, Yuen KS. Epiretinal membrane surgery with or without internal limiting membrane peeling. *Clin Exp Ophthalmol* (2005) 33:379–85. doi: [10.1111/j.1442-9071.2005.01015.x](https://doi.org/10.1111/j.1442-9071.2005.01015.x)

<span id="page-7-1"></span>2. Oberstein SY, Byun J, Herrera D, Chapin EA, Fisher SK, Lewis GP. Cell proliferation in human epiretinal membranes: characterization of cell types and correlation with disease condition and duration. *Mol Vis* (2011) 17:1794–805.

<span id="page-7-2"></span>3. Rękas M., Rejdak M. (2020). *Choroby ciała szklistego i powierzchni szklistkowosiatkówkowej.Siatkówka i ciało szkliste.BCSC12.Seria Basic and clinical science course*. Edra Urban & Partner, Wrocław, Poland *2020*. pp. 371–374

<span id="page-7-3"></span>4. Donati S, Caprani SM, Airaghi G, Vinciguerra R, Bartalena L, Testa F, et al. Vitreous substitutes: the present and the future. *Biomed Res Int* (2014) 2014:351804. doi: [10.1155/2014/351804](https://doi.org/10.1155/2014/351804)

<span id="page-7-4"></span>5. Boneva SK, Wolf J, Wieghofer P, Sebag J, Lange AKC. Hyalocyte functions and nmunology. <br> *Expert* Rev Ophthalmol (2022) 17:249–62. doi: immunology. [10.1080/17469899.2022.2100763](https://doi.org/10.1080/17469899.2022.2100763)

<span id="page-7-5"></span>6. Ankamah E, Sebag J, Ng E, Nolan JM. Vitreous antioxidants, degeneration, and vitreo-retinopathy: exploring the links. *Antioxidants (Basel)* (2019, 2019) 9, 9:7. doi: [10.3390/antiox9010007](https://doi.org/10.3390/antiox9010007)

<span id="page-7-6"></span>7. Los LI, van der Worp RJ, van Luyn MJA, Hooymans JMM. Age-related liquefaction of the human vitreous body: LM and TEM evaluation of the role of proteoglycans and collagen. *Invest Opthalmol Vis Sci* (2003) 44:2828–33. doi: [10.1167/iovs.02-0588](https://doi.org/10.1167/iovs.02-0588)

<span id="page-7-7"></span>8. Tram NK, Swindle-Reilly KE. Rheological properties and age-related changes of the human vitreous humor. *Front Bioeng Biotechnol* (2018) 6:199. doi: [10.3389/](https://doi.org/10.3389/fbioe.2018.00199) [fbioe.2018.00199](https://doi.org/10.3389/fbioe.2018.00199)

<span id="page-7-8"></span>9. Ramovecchi P, Salati C, Zeppieri M. Spontaneous posterior vitreous detachment: a glance at the current literature. *World J Exp Med* (2021) 11:30–6. doi: [10.5493/wjem.v11.i3.30](https://doi.org/10.5493/wjem.v11.i3.30)

<span id="page-7-9"></span>10. Moon SY, Park SP, Kim Y-K. Evaluation of posterior vitreous detachment using ultrasonography and optical coherence tomography. *Acta Ophthalmol* (2020) 98:e29–35. doi: [10.1111/aos.14189](https://doi.org/10.1111/aos.14189)

<span id="page-7-10"></span>11. Frisina R, De Salvo G, Tozzi L, Gius I, Sahyoun JY, Parolini B, et al. Effects of physiological fluctuations on the estimation of vascular flow in eyes with idiopathic macular pucker. *Eye (Lond)* (2022) 37:1470–8. doi: [10.1038/s41433-022-02158-4](https://doi.org/10.1038/s41433-022-02158-4)

<span id="page-7-11"></span>12. Tsotridou E, Loukovitis E, Zapsalis K, Pentara I, Asteriadis S, Tranos P, et al. A review of last decade developments on epiretinal membrane pathogenesis. *Med Hypothesis Discov Innov Ophthalmol* (2020) 9:91–110.

<span id="page-7-12"></span>13. Bianchi L, Altera A, Barone V, Bonente D, Bacci T, De Benedetto E, et al. Untangling the extracellular matrix of idiopathic epiretinal membrane: a path winding among structure, interactomics and translational medicine. *Cells* (2022) 11:2531. doi: [10.3390/cells11162531](https://doi.org/10.3390/cells11162531)

<span id="page-7-13"></span>14. Myojin S, Yoshimura T, Yoshida S, Takeda A, Murakami Y, Kawano Y, et al. Gene expression analysis of the irrigation solution samples collected during vitrectomy for idiopathic epiretinal membrane. *PLoS One* (2016) 11:e0164355. doi: [10.1371/journal.](https://doi.org/10.1371/journal.pone.0164355) [pone.0164355](https://doi.org/10.1371/journal.pone.0164355)

<span id="page-7-14"></span>15. Sheybani A, Harocopos GJ, Rao PK. Immunohistochemical study of epiretinal membranes in patients with uveitis. *J Ophthalmic Inflamm Infect* (2012) 2:243–8. doi: [10.1007/s12348-012-0074-x](https://doi.org/10.1007/s12348-012-0074-x)

<span id="page-7-15"></span>16. Ueki M, Morishita S, Kohmoto R, Fukumoto M, Suzuki H, Sato T, et al. Comparison of histopathological findings between idiopathic and secondary epiretinal membranes. *Int Ophthalmol* (2016) 36:713–8. doi: [10.1007/s10792-016-0194-7](https://doi.org/10.1007/s10792-016-0194-7)

<span id="page-7-16"></span>17. Yamashita T, Uemura A, Sakamoto T. Intraoperative characteristics of the posterior vitreous cortex in patients with epiretinal membrane. *Graefes Arch Clin Exp Ophthalmol* (2008) 246:333–7. doi: [10.1007/s00417-007-0745-8](https://doi.org/10.1007/s00417-007-0745-8)

18. Hamoudi H. Epiretinal membrane surgery: an analysis of sequential or combined surgery on refraction, macular anatomy and corneal endothelium. *Acta Ophthalmol* (2018) 96:1–24. doi: [10.1111/aos.13690](https://doi.org/10.1111/aos.13690)

<span id="page-7-17"></span>19. Foos RY. Vitreoretinal juncture; epiretinal membranes and vitreous. *Invest Ophthalmol Vis Sci* (1977) 16:416–22.

<span id="page-7-18"></span>20. Wiznia RA. Posterior vitreous detachment and idiopathic preretinal macular gliosis. *Am J Ophthalmol* (1986) 102:196–8. doi: [10.1016/0002-9394\(86\)90144-3](https://doi.org/10.1016/0002-9394(86)90144-3)

<span id="page-7-19"></span>21. Lee GW, Lee SE, Han SH, Kim SJ, Kang SW. Characteristics of secondary epiretinal membrane due to peripheral break. *Sci Rep* (2020) 10:20881. doi: [10.1038/](https://doi.org/10.1038/s41598-020-78093-9) [s41598-020-78093-9](https://doi.org/10.1038/s41598-020-78093-9)

<span id="page-7-20"></span>22. da Silva RA, Roda VM, Matsuda M, Siqueira PV, Lustoza-Costa GJ, Wu DC. Cellular components of the idiopathic epiretinal membrane. *Graefes Arch Clin Exp Ophthalmol* (2022) 260:1435–44. doi: [10.1007/s00417-021-05492-7](https://doi.org/10.1007/s00417-021-05492-7)

<span id="page-7-21"></span>23. Shibuya M. (2011). Vascular endothelial growth factor (VEGF) and its receptor (VEGFR) signaling in angiogenesis: a crucial target for anti-and pro-Angiogenic therapies. *Genes Cancer* 2:1097–1105. doi: [10.1177/1947601911423031](https://doi.org/10.1177/1947601911423031)

<span id="page-7-23"></span>24.Vishwakarma S, Gupta RK, Jakati S, Tyagi M, Pappuru RR, Reddig K, et al. Molecular assessment of Epiretinal membrane: activated microglia, oxidative stress and inflammation. *Antioxidants (Basel)* (2020) 9:654. doi: [10.3390/](https://doi.org/10.3390/antiox9080654) [antiox9080654](https://doi.org/10.3390/antiox9080654)

<span id="page-7-24"></span>25. Bringmann A, Wiedemann P. Involvement of Müller glial cells in epiretinal membrane formation. *Graefes Arch Clin Exp Ophthalmol* (2009) 247:865–83. doi: [10.1007/s00417-009-1082-x](https://doi.org/10.1007/s00417-009-1082-x)

<span id="page-7-25"></span>26. Colakoglu A, Balci AS. Potential role of Müller cells in the pathogenesis of macropsia associated with epiretinal membrane: a hypothesis revisited. *Int J Ophthalmol* (2017) 10:1759–67. doi: [10.18240/ijo.2017.11.19](https://doi.org/10.18240/ijo.2017.11.19)

<span id="page-7-26"></span>27. Schumann RG, Gandorfer A, Ziada J, Scheler R, Schaumberger MM, Wolf A, et al. Hyalocytes in idiopathic epiretinal membranes: a correlative light and electron microscopic study. *Graefes Arch Clin Exp Ophthalmol* (2014) 252:1887–94. doi: [10.1007/](https://doi.org/10.1007/s00417-014-2841-x) [s00417-014-2841-x](https://doi.org/10.1007/s00417-014-2841-x)

<span id="page-7-27"></span>28. Kohno RI, Hata Y, Kawahara S, Kita T, Arita R, Mochizuki Y, et al. Possible contribution of hyalocytes to idiopathic epiretinal membrane formation and its contraction. *Br J Ophthalmol* (2009) 93:1020–6. doi: [10.1136/bjo.2008.155069](https://doi.org/10.1136/bjo.2008.155069)

<span id="page-7-28"></span>29. Tang S, Gao R, Wu DZ. Macrophages in human epiretinal and vitreal membranes in patients with proliferative intraocular disorders. *Yan Ke Xue Bao* (1996) 12:28–32.

<span id="page-7-29"></span>30. Baek J, Park HY, Lee JH, Choi M, Lee JH, Ha M, et al. Elevated M2 macrophage markers in epiretinal membranes with ectopic inner foveal layers. *Invest Ophthalmol Vis Sci* (2020) 61:19. doi: [10.1167/iovs.61.2.19](https://doi.org/10.1167/iovs.61.2.19)

<span id="page-7-22"></span>31. Joshi M, Agrawal S, Christoforidis JB. Inflammatory mechanisms of idiopathic epiretinal membrane formation. *Mediat Inflamm* (2013) 2013:192582. doi: [10.1155/2013/192582](https://doi.org/10.1155/2013/192582)

<span id="page-7-30"></span>32. Bianchi E, Ripandelli G, Feher J, Plateroti AM, Plateroti R, Kovacs I, et al. Occlusion of retinal capillaries caused by glial cell proliferation in chronic ocular inflammation. *Folia Morphol (Warsz)* (2015) 74:33–41. doi: [10.5603/FM.2015.0006](https://doi.org/10.5603/FM.2015.0006)

<span id="page-7-31"></span>33. Giachos I, Chalkiadaki E, Andreanos K, Symeonidis C, Charonis A, Georgalas I, et al. Epiretinal membrane-induced intraretinal neovascularization. *Am J Ophthalmol Case Rep* (2021) 23:101180. doi: [10.1016/j.ajoc.2021.101180](https://doi.org/10.1016/j.ajoc.2021.101180)

<span id="page-7-32"></span>34. Stafiej J, Kaźmierczak K, Linkowska K, Żuchowski P, Grzybowski T, Malukiewicz G. Evaluation of TGF-Beta 2 and VEGF*α* gene expression levels in Epiretinal membranes and internal limiting membranes in the course of retinal detachments, proliferative diabetic retinopathy, macular holes, and idiopathic Epiretinal membranes. *J Ophthalmol* (2018) 2018:8293452. doi: [10.1155/2018/8293452](https://doi.org/10.1155/2018/8293452)

<span id="page-7-33"></span>35. Rezzola S, Guerra J, Krishna Chandran AM, Loda A, Cancarini A, Sacristani P, et al. VEGF-independent activation of Müller cells by the vitreous from proliferative diabetic retinopathy patients. *Int J Mol Sci* (2021) 22:2179. doi: [10.3390/ijms22042179](https://doi.org/10.3390/ijms22042179)

<span id="page-7-34"></span>36. Lennikov A, Mukwaya A, Fan L, Saddala MS, De Falco S, Huang H. Synergistic interactions of PlGF and VEGF contribute to blood-retinal barrier breakdown through canonical NFκB activation. *Exp Cell Res* (2020) 397:112347. doi: [10.1016/j.](https://doi.org/10.1016/j.yexcr.2020.112347) [yexcr.2020.112347](https://doi.org/10.1016/j.yexcr.2020.112347)

<span id="page-7-35"></span>37. Jang DI, Lee A-H, Shin H-Y, Song H-R, Park J-H, Kang T-B, et al. The role of tumor necrosis factor alpha (TNF-α) in autoimmune disease and current TNF-α inhibitors in therapeutics. *Int J Mol Sci* (2021) 22:2719. doi: [10.3390/ijms22052719](https://doi.org/10.3390/ijms22052719)

<span id="page-7-36"></span>38. Mirshahi A, Hoehn R, Lorenz K, Kramann C, Baatz H. Anti-tumor necrosis factor alpha for retinal diseases: current knowledge and future concepts. *J Ophthalmic Vis Res*  $(2012)$  7:39–44.

<span id="page-7-37"></span>39. Andrae J, Gallini R, Betsholtz C. Role of platelet-derived growth factors in physiology and medicine. *Genes Dev* (2008) 22:1276–312. doi: [10.1101/gad.1653708](https://doi.org/10.1101/gad.1653708)

<span id="page-7-38"></span>40. Edqvist PHD, Niklasson M, Vidal-Sanz M, Hallböök F, Forsberg-Nilsson K. Platelet-derived growth factor over-expression in retinal progenitors results in abnormal retinal vessel formation. *PLoS One* (2012) 7:e42488. doi: [10.1371/journal.pone.0042488](https://doi.org/10.1371/journal.pone.0042488)

<span id="page-7-39"></span>41. Vinores SA, Henderer JD, Mahlow J, Chiu C, Derevjanik NL, Larochelle W, et al. Isoforms of platelet-derived growth factor and its receptors in epiretinal membranes: immunolocalization to retinal pigmented epithelial cells. *Exp Eye Res* (1995) 60:607–19. doi: [10.1016/s0014-4835\(05\)80003-x](https://doi.org/10.1016/s0014-4835(05)80003-x)

<span id="page-7-40"></span>42. Hachana S, Larrivée B. TGF-β superfamily signaling in the eye: implications for ocular pathologies. *Cells* (2022) 11:2336. doi: [10.3390/cells11152336](https://doi.org/10.3390/cells11152336)

<span id="page-7-41"></span>43. Kanda A, Noda K, Hirose I, Ishida S. TGF-β-SNAIL axis induces Müller glialmesenchymal transition in the pathogenesis of idiopathic epiretinal membrane. *Sci Rep* (2019) 9:673. doi: [10.1038/s41598-018-36917-9](https://doi.org/10.1038/s41598-018-36917-9)

<span id="page-7-42"></span>44. Pafumi I, Favia A, Gambara G, Papacci F, Ziparo E, Palombi F, et al. Regulation of Angiogenic functions by angiopoietins through calcium-dependent signaling pathways. *Biomed Res Int* (2015) 2015:965271:1–14. doi: [10.1155/2015/965271](https://doi.org/10.1155/2015/965271)

<span id="page-7-43"></span>45. Joussen AM, Ricci F, Paris LP, Korn C, Quezada-Ruiz C, Zarbin M. Angiopoietin/ Tie2 signalling and its role in retinal and choroidal vascular diseases: a review of preclinical data. *Eye* (2021) 35:1305–16. doi: [10.1038/s41433-020-01377-x](https://doi.org/10.1038/s41433-020-01377-x)

<span id="page-7-44"></span>46. Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. *Cold Spring Harb Perspect Biol* (2014) 6:a016295. doi: [10.1101/cshperspect.a016295](https://doi.org/10.1101/cshperspect.a016295)

<span id="page-7-45"></span>47. Ghasemi H. Roles of IL-6 in ocular inflammation: a review. *Ocul Immunol Inflamm* (2018) 26:37–50. doi: [10.1080/09273948.2016.1277247](https://doi.org/10.1080/09273948.2016.1277247)

<span id="page-7-46"></span>48. Limb GA, Kapur S, Woon H, Franks WA, Jones SE, Chignell AH. Expression of mRNA for interleukin 6 by cells infiltrating epiretinal membranes in proliferative vitreoretinopathy. *Agents Actions* (1993) 38:C73–6. doi: [10.1007/BF01991142](https://doi.org/10.1007/BF01991142)

<span id="page-8-4"></span>49. Harjunpää H, Llort AM, Guenther C, Fagerholm SC. Cell adhesion molecules and their roles and regulation in the immune and tumor microenvironment. *Front Immunol* (2019) 10:1078. doi: [10.3389/fimmu.2019.01078](https://doi.org/10.3389/fimmu.2019.01078)

<span id="page-8-5"></span>50. Tang S, Le-Ruppert KC, Gabel VP. Expression of intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1) on proliferating vascular endothelial cells in diabetic epiretinal membranes. *Br J Ophthalmol* (1994) 78:370–6. doi: [10.1136/bjo.78.5.370](https://doi.org/10.1136/bjo.78.5.370)

<span id="page-8-6"></span>51. Chiquet M. Tenascin-C: from discovery to structure-function relationships. *Front Immunol* (2020) 11:611789. doi: [10.3389/fimmu.2020.611789](https://doi.org/10.3389/fimmu.2020.611789)

<span id="page-8-7"></span>52. Kobayashi Y, Yoshida S, Zhou Y, Nakama T, Ishikawa K, Arita R, et al. The role of tenascin-C in fibrovascular membrane formation in diabetic retinopathy. *Invest Ophthalmol Vis Sci* (2015) 56:224.

<span id="page-8-8"></span>53. Yun YR, Won JE, Jeon E, Lee S, Kang W, Jo H, et al. Fibroblast growth factors: biology, function, and application for tissue regeneration. *J Tissue Eng* (2010) 2010:218142. doi: [10.4061/2010/218142](https://doi.org/10.4061/2010/218142)

<span id="page-8-9"></span>54. Cassidy L, Barry P, Shaw C, Duffy J, Kennedy S. Platelet derived growth factor and fibroblast growth factor basic levels in the vitreous of patients with vitreoretinal disorders. *Br J Ophthalmol* (1998) 82:181–5. doi: [10.1136/bjo.82.2.181](https://doi.org/10.1136/bjo.82.2.181)

55. Frank RN, Amin RH, Eliott D, Puklin JE, Abrams GW. Basic fibroblast growth factor and vascular endothelial growth factor are present in epiretinal and choroidal neovascular membranes. *Am J Ophthalmol* (1996) 122:393–403. doi: [10.1016/s0002-9394\(14\)72066-5](https://doi.org/10.1016/s0002-9394(14)72066-5)

<span id="page-8-10"></span>56. Hueber A, Wiedemann P, Esser P, Heimann K. Basic fibroblast growth factor mRNA, bFGF peptide and FGF receptor in epiretinal membranes of intraocular proliferative disorders (PVR and PDR). *Int Ophthalmol* (1996) 20:345–50. doi: [10.1007/](https://doi.org/10.1007/BF00176889) [BF00176889](https://doi.org/10.1007/BF00176889)

<span id="page-8-11"></span>57. Ding X, Zhang R, Zhang S, Zhuang H, Xu G. Differential expression of connective tissue growth factor and hepatocyte growth factor in the vitreous of patients with high myopia versus vitreomacular interface disease. *BMC Ophthalmol* (2019) 19:25. doi: [10.1186/s12886-019-1041-1](https://doi.org/10.1186/s12886-019-1041-1)

<span id="page-8-12"></span>58. Shpak AA, Guekht AB, Druzhkova TA, Troshina AA, Gulyaeva N. Glial cell linederived neurotrophic factor in patients with age-related cataract. *Invest Ophthalmol Vis Sci* (2021) 62:718.

<span id="page-8-13"></span>59. Nishikiori N, Tashimo A, Mitamura Y, Ohtsuka K. Glial cell line–derived neurotrophic factor in the vitreous of patients with proliferative diabetic retinopathy. *Invest Ophthalmol Vis Sci* (2005) 46:3103.

<span id="page-8-0"></span>60. Xiao W, Chen X, Yan W, Zhu Z, He M. Prevalence and risk factors of epiretinal membranes: a systematic review and meta-analysis of population-based studies. *BMJ Open* (2017) 7:e014644. doi: [10.1136/bmjopen-2016-014644](https://doi.org/10.1136/bmjopen-2016-014644)

<span id="page-8-1"></span>61.Ng CH, Cheung N, Wang JJ, Islam AF, Kawasaki R, Meuer SM, et al. Prevalence and risk factors for epiretinal membranes in a multi-ethnic United States population. *Ophthalmology* (2010) 118:694–9. doi: [10.1016/j.](https://doi.org/10.1016/j.ophtha.2010.08.009) [ophtha.2010.08.009](https://doi.org/10.1016/j.ophtha.2010.08.009)

<span id="page-8-2"></span>62. Fung AT, Galvin J, Tran T. Epiretinal membrane: a review. *Clin Exp Ophthalmol* (2021) 49:289–308. doi: [10.1111/ceo.13914](https://doi.org/10.1111/ceo.13914)

<span id="page-8-3"></span>63. Stevenson W, Prospero Ponce CM, Agarwal DR, Gelman R, Christoforidis JB. Epiretinal membrane: optical coherence tomography-based diagnosis and classification. *Clin Ophthalmol* (2016) 10:527–34. doi: [10.2147/OPTH.S97722](https://doi.org/10.2147/OPTH.S97722)

<span id="page-8-14"></span>64. Govetto A, Lalane RA 3rd, Sarraf D, Figueroa MS, Hubschman JP. Insights into Epiretinal membranes: presence of ectopic inner foveal layers and a new optical coherence tomography staging scheme. *Am J Ophthalmol* (2017) 175:99–113. doi: [10.1016/j.ajo.2016.12.006](https://doi.org/10.1016/j.ajo.2016.12.006)

<span id="page-8-15"></span>65. Folk JC, Adelman RA, Flaxel CJ, Hyman L, Pulido JS, Olsen TW. Idiopathic epiretinal membrane and vitreomacular traction preferred [practice](https://doi.org/10.1016/j.ophtha.2015.10.048)  pattern(®) guidelines. *Ophthalmology* (2016) 123:P152–81. doi: 10.1016/j. [ophtha.2015.10.048](https://doi.org/10.1016/j.ophtha.2015.10.048)

<span id="page-8-16"></span>66. Matoba R, Morizane Y. Surgical treatment of epiretinal membrane. *Acta Med Okayama* (2021) 75:403–13. doi: [10.18926/AMO/62378](https://doi.org/10.18926/AMO/62378)

<span id="page-8-17"></span>67. Forlini M, Date P, D'Eliseo D, Rossini P, Bratu A, Volinia A, et al. Limited vitrectomy versus complete vitrectomy for epiretinal membranes: a comparative multicenter trial. *J Ophthalmol* (2020) 2020:1. doi: [10.1155/2020/6871207](https://doi.org/10.1155/2020/6871207)

<span id="page-8-18"></span>68. Meduri A, Oliverio GW, Trombetta L, Giordano M, Inferrera L, Trombetta CJ. Optical coherence tomography predictors of favorable functional response in Naïve diabetic macular edema eyes treated with dexamethasone implants as a first-line agent. *J Ophthalmol* (2021) 2021:1. doi: [10.1155/2021/6639418](https://doi.org/10.1155/2021/6639418)

<span id="page-8-19"></span>69. Dawson SR, Shunmugam M, Williamson TH. Visual acuity outcomes following surgery for idiopathic epiretinal membrane: an analysis of data from 2001 to 2011. *Eye (Lond)* (2014) 28:219–24. doi: [10.1038/eye.2013.253](https://doi.org/10.1038/eye.2013.253)

<span id="page-8-20"></span>70. Shen Z, Duan X, Wang F, Wang N, Peng Y, Liu DTL, et al. Prevalence and risk factors of posterior vitreous detachment in a Chinese adult population: the Handan eye study. *BMC Ophthalmol* (2013) 13:33. doi: [10.1186/1471-2415-13-33](https://doi.org/10.1186/1471-2415-13-33)