



## OPEN ACCESS

## EDITED BY

Yukihiro Shiga,  
University of Montreal Hospital Research  
Centre (CRCHUM), Canada

## REVIEWED BY

Zhengbo Shao,  
The Second Affiliated Hospital of Harbin  
Medical University, China

## \*CORRESPONDENCE

Rachel S. Chong  
✉ Rachel.chong.s.j@snec.com.sg

## SPECIALTY SECTION

This article was submitted to  
Ophthalmology,  
a section of the journal  
Frontiers in Medicine

RECEIVED 02 November 2022

ACCEPTED 04 January 2023

PUBLISHED 24 January 2023

## CITATION

Loo JH, Wang Z and Chong RS (2023)  
Caveolin-1 in vascular health and glaucoma: A  
critical vascular regulator and potential  
therapeutic target. *Front. Med.* 10:1087123.  
doi: 10.3389/fmed.2023.1087123

## COPYRIGHT

© 2023 Loo, Wang and Chong. This is an  
open-access article distributed under the terms  
of the [Creative Commons Attribution License  
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction  
in other forums is permitted, provided the  
original author(s) and the copyright owner(s)  
are credited and that the original publication in  
this journal is cited, in accordance with  
accepted academic practice. No use,  
distribution or reproduction is permitted which  
does not comply with these terms.

# Caveolin-1 in vascular health and glaucoma: A critical vascular regulator and potential therapeutic target

Jing Hong Loo<sup>1</sup>, Zhaoran Wang<sup>2</sup> and Rachel S. Chong<sup>3,4\*</sup>

<sup>1</sup>Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore, <sup>2</sup>Duke-NUS Medical School, Singapore, Singapore, <sup>3</sup>Glaucoma Department, Singapore National Eye Center, Singapore, Singapore, <sup>4</sup>Ocular Imaging Department, Singapore Eye Research Institute, Singapore, Singapore

Caveolin-1 (Cav-1) is an integral scaffolding membrane protein found in most cell types. Cav-1 has been found to contribute significantly to ocular function, with mutations of Cav-1 being associated with a genetic risk of glaucoma development. Raised intraocular pressure (IOP) is a major modifiable risk factor for glaucoma. Cav-1 may be involved in both IOP-dependent and independent mechanisms involving vascular dysregulation. Systemic vascular diseases including hypertension, diabetes and hyperlipidaemia, have been shown to be associated with glaucoma development. Cav-1 is closely interlinked with endothelial nitric oxide synthase pathways that mediate vascular function and prevent cardiovascular diseases. Endothelial nitric oxide synthase and endothelin-1 are key vasoactive molecules expressed in retinal blood vessels that function to autoregulate ocular blood flow (OBF). Disruptions in the homeostasis of OBF have led to a growing concept of impaired neurovascular coupling in glaucoma. The imbalance between perfusion and neuronal stimulation arising from Cav-1 depletion may result in relative ischemia of the optic nerve head and glaucomatous injury. OBF is also governed by circadian variation in IOP and systemic blood pressure (BP). Cav-1 has been shown to influence central BP variability and other circadian rhythms such as the diurnal phagolysosomal digestion of photoreceptor fragments and toxic substrates to maintain ocular health. Overall, the vast implications of Cav-1 on various ocular mechanisms leading to glaucoma suggest a potential for new therapeutics to enhance Cav-1 expression, which has seen success in other neurodegenerative diseases.

## KEYWORDS

glaucoma, Caveolin-1 (Cav-1), systemic vascular risk factors, ocular blood flow, neurovascular coupling (NVC), glaucoma therapy

## Caveolin-1 in vascular health

Caveolin-1 (Cav-1) is a major coat protein of caveolae, which are flask-shaped invaginations of the plasma membrane ubiquitously found in various cell types, particularly adipocytes, endothelial cells, epithelial cells and fibroblasts. Caveolae have been discovered to play roles in lipid transportation (1), membrane traffic (2), and signal transduction (3). Although there are three caveolin genes identified in mammals, namely Cav-1, -2, and -3, Cav-1 is notably essential for caveolae formation and function (4, 5). Through a multitude of signaling cascades, Cav-1 has been implicated in cardiovascular disease, atherosclerosis, diabetes, cancer, and a variety of degenerative muscular dystrophies (6). In the cardiovascular system, Cav-1 particularly contributes to the functions of endothelial cells *via* interacting with endothelial nitric oxide synthase (eNOS) and regulating the release of nitric oxide (NO) (7).

## Caveolin-1 involvement in glaucoma

While Cav-1 has been extensively studied in extra-ocular diseases, its role in ocular function and diseases has only recently received attention. Cav-1 is expressed abundantly in Muller glia, retinal and choroidal vasculature, and retinal pigment epithelium (RPE) (8). Mutations of Cav-1 gene are associated with an increased genetic risk of primary open-angle glaucoma development across various population cohorts (9–12).

Glaucoma is a neurodegenerative disease characterized by progressive loss of retinal ganglion cells (RGC) and optic nerve degeneration that results in irreversible visual field deficits. Raised intraocular pressure (IOP) is a major modifiable risk factor for glaucoma (13). The effect of Cav-1 deficiency on IOP homeostasis has been evaluated in various pre-clinical studies, with Cav-1 deficient mice displaying significantly higher IOP (14–16). A postulated mechanism underlying ocular hypertension in Cav-1 deficiency is the resultant overreactive eNOS signaling pathways. While NO is a potent vasodilator and has a crucial role in lowering IOP, chronic dysregulation of eNOS may lead to outflow tract dysfunction. Indeed, Cav-1 depletion has been independently associated with increased conventional outflow resistance leading to decreased drainage of aqueous humor from the anterior chamber (17). Additionally, new findings have shed light on Cav-1 potential role as mechanosensors in the Schlemm's canal and trabecular meshwork that protects against mechanical stress from IOP fluctuations (18). Cav-1 may contribute to increased IOP and increase the susceptibility of the optic nerve head (ONH) to cellular damage due to altered outflow tract mechanoprotection.

Cav-1 has also been implicated in altering vascular function, both systemically and within the eye. These alterations in vascular profile may contribute to greater glaucoma risk, as described in this mini-review.

## The role of Cav-1 in mediating systemic vascular risk factors for glaucoma

Various systemic vascular risk factors including hypertension or hypotension, diabetes mellitus, hyperlipidaemia, atherosclerotic diseases and migraine have been associated with glaucoma development. Hypertension has a direct causative link to glaucoma risk by means of increased ciliary blood flow and aqueous humor production coupled with decrease outflow due to elevated episcleral venous pressure (19). Hypotension is particularly associated with normal-tension glaucoma since it lowers ocular perfusion pressure (OPP), resulting in optic nerve ischemia and glaucomatous degeneration (20). Circadian variation of blood pressure (BP) may have a role in glaucoma development too. A meta-analysis in 2015 pooled evidence from epidemiological studies and established nocturnal BP fall as a risk factor for progressive visual field losses in glaucoma (21). In presence of the other vascular risk factors, nocturnal dipping exacerbates poor optic nerve perfusion and glaucomatous optic neuropathy (22). The association between diabetes and glaucoma may be explained by a few key mechanisms. Hyperglycaemia and dysregulation in lipid metabolism results in oxidative stress, vascular dysregulation and eventual neuronal injury (23–25). Hyperglycaemia of the aqueous humor also leads to structural remodeling at the trabecular meshwork and impaired

aqueous humor outflow (26). Atherosclerotic diseases include a spectrum of disease conditions from coronary artery disease to peripheral vascular disease and stroke. The association between atherosclerotic diseases and glaucoma has been extensively studied (27–30), however current evidence is insufficient to support a direct causal relationship between the two due to potential confounding factors from the underlying pathophysiological processes involved. Finally, migraine is associated with systemic vasospasm causing relative ischemia, thereby increasing the risk of glaucoma, particularly normal-tension glaucoma (31–33).

Many of the aforementioned systemic vascular risk factors have been linked to Cav-1. From diabetes to lipid disorders and pulmonary fibrosis, Cav-1 plays an integral role in maintaining vascular homeostasis and controlling atherosclerosis formation through lipoprotein trafficking across the vascular endothelium (34–36). Central to this physiology is the regulatory setup of Cav-1/eNOS. eNOS is constitutively expressed in vascular endothelium and produces the vasodilatory gas NO which maintains endothelial function and health (37). eNOS bounded to caveolae is rendered inactive by its direct association with caveolin scaffolding domain of Cav-1 (38). Cav-1 directly competes with calmodulin (an activator of eNOS) for binding to the active site of eNOS (39). Furthermore, Cav-1 also regulates eNOS expression levels by inhibiting serine/threonine amino acid kinase Akt phosphorylation of eNOS, thus governing the basal level of NO in endothelial cells (40). While Cav-1 depletion is characterized by chronic hyperactivation of eNOS, a decoupling of the de-inhibited eNOS may occur, thus resulting in a decreased bioavailability of NO (41). Reduced NO production is associated with vascular dysfunction and cardiovascular mortality (42).

## Cav-1 mediated regulation of ocular blood flow *via* NO-dependent and independent pathways

Autoregulation of ocular blood flow (OBF) in the retinal vasculature enables a relatively stable supply of blood and metabolites despite fluctuation in OPP (43). OPP is calculated as derived from the subtraction of IOP from mean arterial pressure (44). Variations in mean arterial pressure and IOP results in corresponding variations in OPP. Within a range of OPP, OBF remains constant due to autoregulation of vascular tone in the retinal and ONH (45).

Two vasoactive factors, namely NO and ET-1, are crucial in the autoregulatory mechanism (46). NO is a potent vasodilator released by endothelial cells and acts on pericytes to cause vasodilation (47). ET-1 is a potent vasoconstrictor that exerts its effect *via* ETA, ETB1, and ETB2 receptors. ETA and ETB2 receptors are found on vascular smooth muscle cells and causes vasoconstriction while ETB1 is found on endothelial cells and cause vasodilation (48). The counterregulatory effects of NO and ET-1 maintains an appropriate vascular tone and constant blood flow to the ONH.

Impaired autoregulation is seen in glaucomatous optic neuropathy. This arises from cellular dysfunction leading to an imbalance of vasoactive factors and ischemia at the ONH (49, 50). Numerous studies have shown that elevated levels of ET-1 are associated with disease pathology (51–54). Blocking of ET-1 receptors in mice increased OBF and protects from glaucomatous

injury (55). Alterations in NO signaling pathways either through upregulation or downregulation are also implicated in glaucoma. High NO may increase ocular perfusion but cause oxidative stress and injury to neurons due to formation of reactive oxygen species (56). Decreased NO levels are found in the aqueous humor of glaucoma patients (57). Characteristic RGC loss and vascular dysfunction seen in glaucoma are more prominent with decreased NO production and impairment in its downstream NO-cGMP signaling pathways (58).

ET-1 and NO dysregulation is partly mediated by Cav-1. The effect of Cav-1 on NO homeostasis has been explained in the previous section. An intrinsic regulatory interaction also exists between Cav-1 and ET-1. Both the scaffolding domain and C-terminal domain of Cav-1 can bind to ET receptors and this localizes the complex to the caveolae membrane (59); it has been suggested that compartmentalization of ETB receptor/Cav-1 complexes within caveolae ensures signal transduction and prevents rapid endocytosis of the receptor (60). An early study demonstrated that disruption of caveolae structure significantly diminishes ET-1-induced phosphorylation of ERK 1/2 and subsequent signal propagation (60). This interplay between Cav-1 and mediators of vascular tone suggests the crucial role of Cav-1 in ocular vascular health. Cav-1 deficiency is associated with vascular dysregulation which may predispose to structural neuronal injury at the ONH in the presence of existing stressors like IOP, thus exacerbating disease progression (61).

Cav-1 depletion is also associated with disruptions in blood-retinal barrier integrity and venous morphology, that are independent of eNOS activity. Gu et al. have reported hyperpermeability of the large branch retinal veins of the superficial retina, and enlargement of retinal veins in Cav-1 knockout mice (62). These alterations were found to be independent of NOS-expression and activity. It is therefore possible that Cav-1 mediates vascular dysfunction in the eye through both NO-dependent and independent mechanisms, where the former regulates capillary dilation and the latter stabilizes vessel wall integrity in retinal veins.

## Neurovascular coupling in glaucoma

Neuronal activity is tightly matched to OBF in the eye in what is termed as neurovascular coupling (NVC) (45). An increase in neuronal stimulation is associated with a corresponding increase in blood flow to meet the metabolic requirements of the retinal tissue. This NVC response is mediated by the neurovascular unit which comprises vascular cells, glial cells and neurons (63, 64). Defective NVC has been described in primary open-angle glaucoma (65, 66). In response to flicker-light stimulation, the increase in OBF in glaucoma patients was found to be significantly lower than that of healthy subjects (67). Various mechanisms may explain the defective NVC response in glaucoma. Firstly, glaucoma is characterized by RGC apoptosis due to various possible causes involving raised IOP, oxidative stress and mitochondrial dysfunction—decreased neuronal signaling from RGCs may drive reduced NVC (68–70). Secondly, decreased gap junction expression in the retinal and ONH also affects communication between cells of the neurovascular unit (71, 72). Lastly, the integrity of retinal barrier is compromised due to the loss of tight junctions (73), leading to both compromised blood supply and transendothelial migration of inflammatory cells causing further neuronal injury (74).

On the back of increasing evidence of Cav-1 involvement in glaucoma, our own study in Cav-1 knockout mice showed defective NVC at the ONH as assessed by laser speckle flowgraphy (16). This is associated with changes in vessel morphology as well as a decrease in electrophysiological function of RGCs (16). While the temporal association has yet to be clearly-established, it is possible that vascular dysfunction contributes to defective NVC which is associated with early functional RGC injury before structural losses are seen. Apart from its role in mediating vascular tone as described in the previous sections, Cav-1 may influence microvascular structural characteristics by downregulating vascular endothelial growth factor (75). Defective Cav-1 promotes angiogenesis, but the excessive vascular branching pattern may lead to poorer perfusion instead (76). These findings support the theory that defective Cav-1 is associated with vascular dysfunction and impaired NVC. However, the precise involvement of glial cells or retinal microvasculature in regulating the NVC process remains to be seen—particularly in the context of glaucoma.

## Cav-1 and disruption of circadian rhythms

Numerous studies have shown that circadian variation in BP, IOP, and OPP are risk factors for the development of glaucoma. Progression of visual field loss in glaucoma patients has been associated with a larger range of diurnal IOP fluctuations and nocturnal pressure spikes (77–79). IOP tends to be higher at night due to decreased aqueous humor drainage *via* the trabecular meshwork and uveoscleral pathway (80). Similarly, diurnal variation in BP and nocturnal dipping may contribute to glaucoma pathogenesis as well (81, 82). Nocturnal BP reduction is attributed to a fall in sympathetic tone with reduced circulating levels of catecholamines (83). OPP is driven by a complex interplay between BP and IOP; fluctuations in either will translate to variations in OPP (84). Abrupt variations in OPP beyond the capacity of autoregulatory mechanisms may thus cause unstable OBF (85, 86), triggering a sequence of ischemic and reperfusion injury at the ONH.

Limited studies have described Cav-1 involvement in circadian rhythms disruptions causing glaucoma development. An experimental study by Desjardins et al. (87) showed that Cav-1 deficient mice exhibit decreased very low frequency BP variability. Administration of caveolin scaffolding domain reversed this drop in BP variability. The authors attributed this to the increased NO production *ex vivo* arising from reduced allosteric inhibition by Cav-1. While the bandwidth of spectral analysis cannot be directly applicable to human, the study does provide invaluable insights regarding the function of Cav-1 on NO production and control of central BP variability. Another circadian rhythm implicated in Cav-1 depletion is the diurnal pattern of renewal of photoreceptor outer segment (88). RPE supports photoreceptors neurons *via* the diurnal clearance of outer segment fragments (89). Cav-1 depletion impairs phagolysosome degradation by reversing the diurnal activity of enzymes in the RPE (88). Rod photoreceptor visual function is found to be decreased with Cav-1 knockout (8).

While the present evidence for cav-1 involvement in circadian regulation remains scant, the unique role of Cav-1 in mediating ocular perfusion *via* multiple pathways warrants further studies into how circadian disruptions may influence Cav-1 function.

## Potential therapeutic targets

Current treatment for glaucoma relies heavily on ocular hypotensive medications. Reduction in IOP has proven effective in preventing and slowing disease progression (85). In addition to its effectiveness, IOP-lowering medications exhibit minimal systemic adverse effects and high rates of patient tolerability (90). However, continued disease progression occurs in a small subset of patients despite adequate IOP lowering (91, 92). Furthermore, a modest proportion of patient experienced glaucomatous optic neuropathy despite having normal IOP, in what is termed as normal-tension glaucoma (93). This suggests that there are other IOP-independent mechanisms that may contribute to glaucoma development (94).

The multiple roles of Cav-1 in modulating ocular health and glaucoma risk suggest the potential for new therapeutic strategies that increase Cav-1 expression or augment its downstream signaling. While research on Cav-1 therapeutics remains in its infancy, success with Cav-1 gene therapy for chronic diseases have been described in few recent studies. Lin et al. demonstrated that electroporation-mediated transfer of the Cav-1 gene protects against bleomycin-induced pulmonary fibrosis in mouse lungs *via* downregulation of inflammasome activity and reduction in monocyte recruitment and circulating cytokines (95). The use of electroporation to deliver gene targets is currently being explored in clinical trials for cancer and vaccines (96). It thus remains to be seen if the promising outcomes of Cav-1 gene therapy for idiopathic pulmonary fibrosis can be replicated in humans as well. Cav-1 therapy has also been shown to preserve or delay neurodegeneration in a preclinical model of Alzheimer's disease. Wang et al. showed that synapsin-promoted Cav-1 gene therapy was able to maintain neuronal and synaptic morphology and preserve hippocampal function such as memory and learning in mice with Alzheimer's disease (97). Further translational or clinical research may focus on whether the therapeutic potential of Cav-1 can be exploited for neuroprotective effects in the human eye, possibly averting RGC loss and glaucoma development.

## Conclusions

Raised IOP has long been regarded as the only modifiable risk factor for glaucoma. However, adequate IOP lowering with anti-glaucoma medications may not always deter glaucoma progression. Hence, other factors independent of IOP may be involved in the complex pathogenesis of glaucoma. The “vascular theory” affecting neuronal function has gained attention recently with new evidence showing vascular dysregulation may precede RGC loss (98). Patients with cardiovascular risk factors are at an increased risk of glaucoma. Dysregulation of OBF due to altered levels of vasoactive substances may lead to disruption in blood supply of the ONH. Impaired

NVC can also cause a mismatch of neuronal stimulation and ocular perfusion. All these disturbances in vascular function may manifest as altered vessel morphology and vascular dropout seen in early glaucoma (99).

Cav-1 plays an important role in regulating various pathways involved in the “vascular theory” of glaucoma. There is consistent evidence describing the association between Cav-1 depletion and systemic cardiovascular disease, impaired autoregulation and defective NVC. While the underlying mechanism has not been fully elucidated, understanding this crucial association may pave the way for future therapeutics that focus on restoring vascular health to avert glaucomatous degeneration. Cav-1 therapeutics have shown promising outcomes for other disease, raising hopes that a similar approach can be applied to glaucoma prevention. Future research should focus on exploring the intricate interplay between Cav-1 and vascular dysregulation and exploiting the translative potential of Cav-1 therapy for alternative glaucoma treatment.

## Author contributions

JHL wrote the first draft of the manuscript. ZW formatted and proofread the manuscript. RC provided overall leadership and wrote sections of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

## Funding

Supported by NMRC Clinician-Scientist Individual Grant JRNMR177301 to investigate the role of Caveolin-1 in Inner Retina Neurovascular Coupling in Experimental Glaucoma.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## References

- Li WP, Liu P, Pilcher BK, Anderson R. Cell-specific targeting of caveolin-1 to caveolae, secretory vesicles, cytoplasm or mitochondria. *J Cell Sci.* (2001) 114:1397–408. doi: 10.1242/jcs.114.7.1397
- Anderson RG. The caveolae membrane system. *Annu Rev Biochem.* (1998) 67:199–225.
- Smart EJ, Graf GA, McNiven MA, Sessa WC, Engelman JA, Scherer PE, et al. Caveolins, liquid-ordered domains, and signal transduction. *Mol Cell Biol.* (1999) 19:289–304.
- Liu P, Rudick M, Anderson RG. Multiple functions of caveolin-1. *J Biol Chem.* (2002) 277:41295–8. doi: 10.1074/jbc.R200020200



5. Drab M, Verkade P, Elger M, Kasper M, Lohn M, Lauterbach B, et al. Loss of caveolae, vascular dysfunction, and pulmonary defects in caveolin-1 gene-disrupted mice. *Science*. (2001) 293:2449–52. doi: 10.1126/science.1062688
6. Cohen AW, Hnasko R, Schubert W, Lisanti MP. Role of caveolae and caveolins in health and disease. *Physiol Rev*. (2004) 84:1341–79. doi: 10.1152/physrev.00046.2003
7. Goligorsky MS, Li H, Brodsky S, Chen J. Relationships between caveolae and eNOS: everything in proximity and the proximity of everything. *Am J Physiol Renal Physiol*. (2002) 283:F1–10. doi: 10.1152/ajprenal.00377.2001
8. Li X, McClellan ME, Tanito M, Garteiser P, Townner R, Bissig D, et al. Loss of caveolin-1 impairs retinal function due to disturbance of subretinal microenvironment. *J Biol Chem*. (2012) 287:16424–34. doi: 10.1074/jbc.M112.353763
9. Loomis SJ, Kang JH, Weinreb RN, Yaspan BL, Cooke Bailey JN, Gaasterland D, et al. Association of CAV1/CAV2 genomic variants with primary open-angle glaucoma overall and by gender and pattern of visual field loss. *Ophthalmology*. (2014) 121:508–16. doi: 10.1016/j.ophtha.2013.09.012
10. Thorleifsson G, Walters GB, Hewitt AW, Masson G, Helgason A, DeWan A, et al. Common variants near CAV1 and CAV2 are associated with primary open-angle glaucoma. *Nat Genet*. (2010) 42:906–9. doi: 10.1038/ng.661
11. Rong SS, Chen LJ, Leung CK, Matsushita K, Jia L, Miki A, et al. Ethnic specific association of the CAV1/CAV2 locus with primary open-angle glaucoma. *Sci Rep*. (2016) 6:27837. doi: 10.1038/srep27837
12. Kim S, Kim K, Heo DW, Kim JS, Park CK, Kim CS, et al. Expression-associated polymorphisms of CAV1-CAV2 affect intraocular pressure and high-tension glaucoma risk. *Mol Vis*. (2015) 21:548–54.
13. Weinreb RN, Aung T, Medeiros FA. The pathophysiology and treatment of glaucoma: a review. *JAMA*. (2014) 311:1901–11. doi: 10.1001/jama.2014.3192
14. Lei Y, Song M, Wu J, Xing C, Sun X. eNOS activity in CAV1 knockout mouse eyes. *Invest Ophthalmol Vis Sci*. (2016) 57:2805–13. doi: 10.1167/iovs.15-18841
15. Song M, Wu J, Lei Y, Sun X. Genetic deletion of the NOS3 gene in CAV1-/- mice restores aqueous humor outflow function. *Invest Ophthalmol Vis Sci*. (2017) 58:4976–87. doi: 10.1167/iovs.16-21072
16. Loo JH, Lee YS, Woon CY, Yong VHK, Tan B, Schmetterer L, et al. Loss of caveolin-1 impairs light flicker-induced neurovascular coupling at the optic nerve head. *Front Neurosci*. (2021) 15:764898. doi: 10.3389/fnins.2021.764898
17. Kizhatil K, Chlebowski A, Tolman NG, Freeburg NE, Ryan MM, Shaw NN, et al. An *in vitro* perfusion system to enhance outflow studies in mouse eyes. *Invest Ophthalmol Vis Sci*. (2016) 57:5207–15. doi: 10.1167/iovs.16-19481
18. Elliott MH, Ashpole NE, Gu X, Herrnberger L, McClellan ME, Griffith GL, et al. Caveolin-1 modulates intraocular pressure: implications for caveolae mechanoprotection in glaucoma. *Sci Rep*. (2016) 6:37127. doi: 10.1038/srep37127
19. Zhao D, Cho J, Kim MH, Guallar E. The association of blood pressure and primary open-angle glaucoma: a meta-analysis. *Am J Ophthalmol*. (2014) 158:615–27.e9. doi: 10.1016/j.ajo.2014.05.029
20. Binggeli T, Schoetzu A, Konieczka K. In glaucoma patients, low blood pressure is accompanied by vascular dysregulation. *EPMA J*. (2018) 9:387–91. doi: 10.1007/s13167-018-0155-5
21. Bowe A, Grünig M, Schubert J, Demir M, Hoffmann V, Kütting F, et al. Circadian variation in arterial blood pressure and glaucomatous optic neuropathy: a systematic review and meta-analysis. *Am J Hypertens*. (2015) 28:1077–82. doi: 10.1093/ajh/hpv016
22. Grzybowski A, Och M, Kanclerz P, Leffler C, Moraes CG. Primary open angle glaucoma and vascular risk factors: a review of population based studies from 1990 to 2019. *J Clin Med*. (2020) 9:761. doi: 10.3390/jcm9030761
23. Chopra V, Varma R, Francis BA, Wu J, Torres M, Azen SP, et al. Type 2 diabetes mellitus and the risk of open-angle glaucoma: the Los Angeles Latino Eye Study. *Ophthalmology*. (2008) 115:227–32.e1. doi: 10.1016/j.ophtha.2007.04.049
24. Su DH, Wong TY, Wong WL, Saw SM, Tan DT, Shen SY, et al. Diabetes, hyperglycemia, and central corneal thickness: the Singapore Malay Eye Study. *Ophthalmology*. (2008) 115:964–8.e1. doi: 10.1016/j.ophtha.2007.08.021
25. Nakamura M, Kanamori A, Negi A. Diabetes mellitus as a risk factor for glaucomatous optic neuropathy. *Ophthalmologica*. (2005) 219:1–10. doi: 10.1159/000081775
26. Sato T, Roy S. Effect of high glucose on fibronectin expression and cell proliferation in trabecular meshwork cells. *Invest Ophthalmol Vis Sci*. (2002) 43:170–5.
27. Jeganathan VS, Wong TY, Foster PJ, Crowston JG, Tay WT, Lim SC, et al. Peripheral artery disease and glaucoma: the singapore malay eye study. *Arch Ophthalmol*. (2009) 127:888–93. doi: 10.1001/archophth.2009.136
28. Chen YY, Hu HY, Chu D, Chen HH, Chang CK, Chou P, et al. Patients with primary open-angle glaucoma may develop ischemic heart disease more often than those without glaucoma: an 11-year population-based cohort study. *PLoS ONE*. (2016) 11:e0163210. doi: 10.1371/journal.pone.0163210
29. Marshall H, Mullany S, Qassim A, Siggs O, Hassall M, Ridge B, et al. Cardiovascular disease predicts structural and functional progression in early glaucoma. *Ophthalmology*. (2021) 128:58–69. doi: 10.1016/j.ophtha.2020.06.067
30. Song X, Li P, Li Y, Yan X, Yuan L, Zhao C, et al. Strong association of glaucoma with atherosclerosis. *Sci Rep*. (2021) 11:8792. doi: 10.1038/s41598-021-88322-4
31. Wang JJ, Mitchell P, Smith W. Is there an association between migraine headache and open-angle glaucoma? Findings from the Blue Mountains Eye Study. *Ophthalmology*. (1997) 104:1714–9.
32. Cursiefen C, Wisse M, Cursiefen S, Jünemann A, Martus P, Korth M, et al. Migraine and tension headache in high-pressure and normal-pressure glaucoma. *Am J Ophthalmol*. (2000) 129:102–4. doi: 10.1016/S0002-9394(99)00289-5
33. McKendrick AM, Vingrys AJ, Badcock DR, Heywood JT. Visual field losses in subjects with migraine headaches. *Invest Ophthalmol Vis Sci*. (2000) 41:1239–47.
34. Frank PG, Pavlides S, Lisanti MP. Caveolae and transcytosis in endothelial cells: role in atherosclerosis. *Cell Tissue Res*. (2009) 335:41–7. doi: 10.1007/s00441-008-0659-8
35. Le Lay S, Blouin CM, Hajdouch E, Dugail I. Filling up adipocytes with lipids. Lessons from caveolin-1 deficiency. *Biochim Biophys Acta*. (2009) 1791:514–8. doi: 10.1016/j.bbali.2008.10.008
36. Bakhshi FR, Mao M, Shajahan AN, Piegeler T, Chen Z, Chernaya O, et al. Nitrosation-dependent caveolin 1 phosphorylation, ubiquitination, and degradation and its association with idiopathic pulmonary arterial hypertension. *Pulm Circ*. (2013) 3:816–30. doi: 10.1086/674753
37. Albrecht EW, Stegeman CA, Heeringa P, Henning RH, van Goor H. Protective role of endothelial nitric oxide synthase. *J Pathol*. (2003) 199:8–17. doi: 10.1002/path.1250
38. Bernatchez PN, Bauer PM, Yu J, Prendergast JS, He P, Sessa WC, et al. Dissecting the molecular control of endothelial NO synthase by caveolin-1 using cell-permeable peptides. *Proc Natl Acad Sci U S A*. (2005) 102:761–6. doi: 10.1073/pnas.0407224102
39. Michel JB, Feron O, Sacks D, Michel T. Reciprocal regulation of endothelial nitric-oxide synthase by Ca<sup>2+</sup>-calmodulin and caveolin. *J Biol Chem*. (1997) 272:15583–6.
40. Chen Z, Oliveira S, Zimnicka AM, Jiang Y, Sharma T, Chen S, et al. Reciprocal regulation of eNOS and caveolin-1 functions in endothelial cells. *Mol Biol Cell*. (2018) 29:1190–202. doi: 10.1091/mbc.E17-01-0049
41. Bendall JK, Alp NJ, Warrick N, Cai S, Adlam D, Rockett K, et al. Stoichiometric relationships between endothelial tetrahydrobiopterin, endothelial NO synthase (eNOS) activity, and eNOS coupling *in vivo*: insights from transgenic mice with endothelial-targeted GTP cyclohydrolase 1 and eNOS overexpression. *Circ Res*. (2005) 97:864–71. doi: 10.1161/01.RES.0000187447.03525.72
42. Qian J, Fulton D. Post-translational regulation of endothelial nitric oxide synthase in vascular endothelium. *Front Physiol*. (2013) 4:347. doi: 10.3389/fphys.2013.00347
43. Luo X, Shen YM, Jiang MN, Lou XF, Shen Y. Ocular blood flow autoregulation mechanisms and methods. *J Ophthalmol*. (2015) 2015:864871. doi: 10.1155/2015/864871
44. Prada D, Harris A, Guidoboni G, Siesky B, Huang AM, Arciero J, et al. Autoregulation and neurovascular coupling in the optic nerve head. *Surv Ophthalmol*. (2016) 61:164–86. doi: 10.1016/j.survophthal.2015.10.004
45. Wareham LK, Calkins DJ. The Neurovascular Unit in Glaucomatous Neurodegeneration. *Front Cell Dev Biol*. (2020) 8:452. doi: 10.3389/fcell.2020.00452
46. Nyborg NC, Nielsen PJ. The level of spontaneous myogenic tone in isolated human posterior ciliary arteries decreases with age. *Exp Eye Res*. (1990) 51:711–5.
47. Polak K, Luksch A, Berisha F, Fuchsjaeger-Mayrl G, Dallinger S, Schmetterer L, et al. Altered nitric oxide system in patients with open-angle glaucoma. *Arch Ophthalmol*. (2007) 125:494–8. doi: 10.1001/archophth.125.4.494
48. Schmidl D, Garhofer G, Schmetterer L. The complex interaction between ocular perfusion pressure and ocular blood flow: relevance for glaucoma. *Exp Eye Res*. (2011) 93:141–55. doi: 10.1016/j.exer.2010.09.002
49. Moore D, Harris A, Wudunn D, Kheradiya N, Siesky B. Dysfunctional regulation of ocular blood flow: a risk factor for glaucoma? *Clin Ophthalmol*. (2008) 2:849–61. doi: 10.2147/OPHTH.S2774
50. Feke GT, Pasquale LR. Retinal blood flow response to posture change in glaucoma patients compared with healthy subjects. *Ophthalmology*. (2008) 115:246–52. doi: 10.1016/j.ophtha.2007.04.055
51. Emre M, Örgül S, Haufschild T, Shaw SG, Flammer J. Increased plasma endothelin-1 levels in patients with progressive open angle glaucoma. *Br J Ophthalmol*. (2005) 89:60–3. doi: 10.1136/bjo.2004.046755
52. Kunimatsu S, Mayama C, Tomidokoro A, Araie M. Plasma endothelin-1 level in Japanese normal tension glaucoma patients. *Curr Eye Res*. (2006) 31:727–31. doi: 10.1080/02713680600837382
53. Cellini M, Strobbe E, Gizzi C, Balducci N, Toschi PG, Campos EC, et al. Endothelin-1 plasma levels and vascular endothelial dysfunction in primary open angle glaucoma. *Life Sci*. (2012) 91:699–702. doi: 10.1016/j.lfs.2012.02.013
54. Chen HY, Chang YC, Chen WC, Lane HY. Association between plasma endothelin-1 and severity of different types of glaucoma. *J Glaucoma*. (2013) 22:117–22. doi: 10.1097/IJG.0b013e31822e8c65
55. Howell GR, Macalinao DG, Sousa GL, Walden M, Soto I, Kneeland SC, et al. Molecular clustering identifies complement and endothelin induction as early events in a mouse model of glaucoma. *J Clin Invest*. (2011) 121:1429–44. doi: 10.1172/JCI44646
56. Tezel G. Oxidative stress in glaucomatous neurodegeneration: mechanisms and consequences. *Prog Retin Eye Res*. (2006) 25:490–513. doi: 10.1016/j.preteyeres.2006.07.003

57. Doganay S, Evreklioglu C, Turkoz Y, Er H. Decreased nitric oxide production in primary open-angle glaucoma. *Eur J Ophthalmol.* (2002) 12:44–8. doi: 10.1177/112067210201200109
58. Galassi F, Renieri G, Sodi A, Ucci F, Vannozzi L, Masini E, et al. Nitric oxide proxies and ocular perfusion pressure in primary open angle glaucoma. *Br J Ophthalmol.* (2004) 88:757–60. doi: 10.1136/bjo.2003.028357
59. Chun M, Liyanage UK, Lisanti MP, Lodish HF. Signal transduction of a G protein-coupled receptor in caveolae: colocalization of endothelin and its receptor with caveolin. *Proc Natl Acad Sci U S A.* (1994) 91:11728–32.
60. Yamaguchi T, Murata Y, Fujiyoshi Y, Doi T. Regulated interaction of endothelin B receptor with caveolin-1. *Eur J Biochem.* (2003) 270:1816–27. doi: 10.1046/j.1432-1033.2003.03544.x
61. Tribble JR, Costa VP, Sergott RC, Spaeth GL, Smith M, Wilson RP, et al. The influence of primary open-angle glaucoma upon the retrobulbar circulation: baseline, postoperative and reproducibility analysis. *Trans Am Ophthalmol Soc.* (1993) 91:245–61; discussion 61–5.
62. Gu X, Fliesler SJ, Zhao YY, Stallcup WB, Cohen AW, Elliott MH, et al. Loss of caveolin-1 causes blood-retinal barrier breakdown, venous enlargement, and mural cell alteration. *Am J Pathol.* (2014) 184:541–55. doi: 10.1016/j.ajpath.2013.10.022
63. Hamilton NB, Attwell D, Hall CN. Pericyte-mediated regulation of capillary diameter: a component of neurovascular coupling in health and disease. *Front Neuroenerg.* (2010) 2:5. doi: 10.3389/fnene.2010.00005
64. Attwell D, Buchan AM, Charpak S, Lauritzen M, Macvicar BA, Newman EA, et al. Glial and neuronal control of brain blood flow. *Nature.* (2010) 468:232–43. doi: 10.1038/nature09613
65. Garhöfer G, Zawinka C, Huemer KH, Schmetterer L, Dorner GT. Flicker light-induced vasodilatation in the human retina: effect of lactate and changes in mean arterial pressure. *Invest Ophthalmol Vis Sci.* (2003) 44:5309–14. doi: 10.1167/iovs.03-0587
66. Gugleta K, Kochkorov A, Waldmann N, Polunina A, Katamay R, Flammer J, et al. Dynamics of retinal vessel response to flicker light in glaucoma patients and ocular hypertensives. *Graefes Arch Clin Exp Ophthalmol.* (2012) 250:589–94. doi: 10.1007/s00417-011-1842-2
67. Gugleta K, Fuchsjaeger-Mayrl G, Orgül S. Is neurovascular coupling of relevance in glaucoma? *Surv Ophthalmol.* (2007) 52:S139–43. doi: 10.1016/j.survophthal.2007.08.009
68. Qu J, Wang D, Grosskreutz CL. Mechanisms of retinal ganglion cell injury and defense in glaucoma. *Exp Eye Res.* (2010) 91:48–53. doi: 10.1016/j.exer.2010.04.002
69. Liu B, Neufeld AH. Expression of nitric oxide synthase-2 (NOS-2) in reactive astrocytes of the human glaucomatous optic nerve head. *Glia.* (2000) 30:178–86. doi: 10.1002/(SICI)1098-1136(200004)30:2<178::AID-GLIA7>3.0.CO;2-C
70. Saccà SC, Izzotti A. Oxidative stress and glaucoma: injury in the anterior segment of the eye. *Prog Brain Res.* (2008) 173:385–407. doi: 10.1016/S0079-6123(08)01127-8
71. Shibata M, Oku H, Sugiyama T, Kobayashi T, Tsujimoto M, Okuno T, et al. Disruption of gap junctions may be involved in impairment of autoregulation in optic nerve head blood flow of diabetic rabbits. *Invest Ophthalmol Vis Sci.* (2011) 52:2153–9. doi: 10.1167/iovs.10-6605
72. Malone P, Miao H, Parker A, Juarez S, Hernandez MR. Pressure induces loss of gap junction communication and redistribution of connexin 43 in astrocytes. *Glia.* (2007) 55:1085–98. doi: 10.1002/glia.20527
73. Ivanova E, Kovacs-Oller T, Sagdullaev BT. Domain-specific distribution of gap junctions defines cellular coupling to establish a vascular relay in the retina. *J Comp Neurol.* (2019) 527:2675–93. doi: 10.1002/cne.24699
74. Soto I, Howell GR. The complex role of neuroinflammation in glaucoma. *Cold Spring Harb Perspect Med.* (2014) 4:8. doi: 10.1101/cshperspect.a017269
75. Liu J, Razani B, Tang S, Terman BI, Ware JA, Lisanti MP, et al. Angiogenesis activators and inhibitors differentially regulate caveolin-1 expression and caveolae formation in vascular endothelial cells. Angiogenesis inhibitors block vascular endothelial growth factor-induced down-regulation of caveolin-1. *J Biol Chem.* (1999) 274:15781–5.
76. Mirzapour-Shafiyi F, Kametani Y, Hikita T, Hasegawa Y, Nakayama M. Numerical evaluation reveals the effect of branching morphology on vessel transport properties during angiogenesis. *PLoS Comput Biol.* (2021) 17:e1008398. doi: 10.1371/journal.pcbi.1008398
77. Asrani S, Zeimer R, Wilensky J, Gieser D, Vitale S, Lindenmuth K, et al. Large diurnal fluctuations in intraocular pressure are an independent risk factor in patients with glaucoma. *J Glaucoma.* (2000) 9:134–42. doi: 10.1097/00061198-200004000-00002
78. Liu JH, Zhang X, Kripke DF, Weinreb RN. Twenty-four-hour intraocular pressure pattern associated with early glaucomatous changes. *Invest Ophthalmol Vis Sci.* (2003) 44:1586–90. doi: 10.1167/iovs.02-0666
79. Wax MB, Camras CB, Fiscella RG, Girkin C, Singh K, Weinreb RN, et al. Emerging perspectives in glaucoma: optimizing 24-h control of intraocular pressure. *Am J Ophthalmol.* (2002) 133:S1–10. doi: 10.1016/S0002-9394(02)01459-9
80. Liu H, Fan S, Gulati V, Camras LJ, Zhan G, Gbate D, et al. Aqueous humor dynamics during the day and night in healthy mature volunteers. *Arch Ophthalmol.* (2011) 129:269–75. doi: 10.1001/archophthalmol.2011.4
81. Melgarejo JD, Maestre GE, Mena LJ, Lee JH, Petitto M, Chávez CA, et al. Normal-tension glaucomatous optic neuropathy is related to blood pressure variability in the Maracaibo Aging Study. *Hypertens Res.* (2021) 44:1105–12. doi: 10.1038/s41440-021-00687-1
82. Shin JW, Jo YH, Song MK, Won HJ, Kook MS. Nocturnal blood pressure dip and parapapillary choroidal microvasculature dropout in normal-tension glaucoma. *Sci Rep.* (2021) 11:206. doi: 10.1038/s41598-020-80705-3
83. Sherwood A, Steffen PR, Blumenthal JA, Kuhn C, Hinderliter AL. Nighttime blood pressure dipping: the role of the sympathetic nervous system. *Am J Hypertens.* (2002) 15:111–8. doi: 10.1016/S0895-7061(01)02251-8
84. Choi J, Jeong J, Cho HS, Kook MS. Effect of nocturnal blood pressure reduction on circadian fluctuation of mean ocular perfusion pressure: a risk factor for normal tension glaucoma. *Invest Ophthalmol Vis Sci.* (2006) 47:831–6. doi: 10.1167/iovs.05-1053
85. Heijl A, Leske MC, Bengtsson B, Hyman L, Bengtsson B, Hussein M, et al. Reduction of intraocular pressure and glaucoma progression: results from the Early Manifest Glaucoma Trial. *Arch Ophthalmol.* (2002) 120:1268–79. doi: 10.1001/archophth.120.10.1268
86. Delaney Y, Walshe TE, O'Brien C. Vasospasm in glaucoma: clinical and laboratory aspects. *Optom Vis Sci.* (2006) 83:406–14. doi: 10.1097/01.opx.0000225877.13217.01
87. Desjardins F, Lobysheva I, Pelat M, Gallez B, Feron O, Dessy C, et al. Control of blood pressure variability in caveolin-1-deficient mice: role of nitric oxide identified *in vivo* through spectral analysis. *Cardiovasc Res.* (2008) 79:527–36. doi: 10.1093/cvr/cvn080
88. Sethna S, Chamakkala T, Gu X, Thompson TC, Cao G, Elliott MH, et al. Regulation of phagolysosomal digestion by caveolin-1 of the retinal pigment epithelium is essential for vision. *J Biol Chem.* (2016) 291:6494–506. doi: 10.1074/jbc.M115.687004
89. Baba K, Goyal V, Tosini G. Circadian regulation of retinal pigment epithelium function. *Int J Mol Sci.* (2022) 23:5. doi: 10.3390/ijms23052699
90. Beckers HJ, Schouten JS, Webers CA, van der Valk R, Hendrikse FS. A effects of commonly used glaucoma medications: comparison of tolerability, chance of discontinuation, and patient satisfaction. *Graefes Arch Clin Exp Ophthalmol.* (2008) 246:1485–90. doi: 10.1007/s00417-008-0875-7
91. Anderson DR. Collaborative normal tension glaucoma study. *Curr Opin Ophthalmol.* (2003) 14:86–90. doi: 10.1097/00055735-200304000-00006
92. Cockburn DM. Does reduction of intraocular pressure (IOP) prevent visual field loss in glaucoma? *Am J Optom Physiol Opt.* (1983) 60:705–11.
93. Chen MJ. Normal tension glaucoma in Asia: epidemiology, pathogenesis, diagnosis, and management. *Taiwan J Ophthalmol.* (2020) 10:250–4. doi: 10.4103/tjo.tjo\_30\_20
94. Killer HE, Pircher A. Normal tension glaucoma: review of current understanding and mechanisms of the pathogenesis. *Eye.* (2018) 32:924–30. doi: 10.1038/s41433-018-0042-2
95. Lin X, Barravecchia M, Matthew Kottmann R, Sime P, Dean DA. Caveolin-1 gene therapy inhibits inflammasome activation to protect from bleomycin-induced pulmonary fibrosis. *Sci Rep.* (2019) 9:19643. doi: 10.1038/s41598-019-55819-y
96. Heller R, Heller LC. Gene electrotransfer clinical trials. *Adv Genet.* (2015) 89:235–62. doi: 10.1016/bs.adgen.2014.10.006
97. Wang S, Leem JS, Podvin S, Hook V, Kleschevnikov N, Savchenko P, et al. Synapsin-caveolin-1 gene therapy preserves neuronal and synaptic morphology and prevents neurodegeneration in a mouse model of AD. *Mol Ther Methods Clin Dev.* (2021) 21:434–50. doi: 10.1016/j.omtm.2021.03.021
98. Jia Y, Wei E, Wang X, Zhang X, Morrison JC, Parikh M, et al. Optical coherence tomography angiography of optic disc perfusion in glaucoma. *Ophthalmology.* (2014) 121:1322–32. doi: 10.1016/j.ophtha.2014.01.021
99. Liu L, Jia Y, Takusagawa HL, Pechauer AD, Edmunds B, Lombardi L, et al. Optical coherence tomography angiography of the peripapillary retina in glaucoma. *JAMA Ophthalmol.* (2015) 133:1045–52. doi: 10.1001/jamaophthalmol.2015.2225