



Translational Research Studies Unraveling the Origins of Psoriatic Arthritis: Moving Beyond Skin and Joints

Janne W. Bolt^{1,2,3}, Chaja M. J. van Ansenwoude^{1,2,3}, Ihsan Hammoura^{1,2,3},
Marleen G. van de Sande^{1,2,3} and Lisa G. M. van Baarsen^{1,2,3*}

¹ Department of Rheumatology & Clinical Immunology, Amsterdam Institute for Infection & Immunity, Amsterdam UMC, University of Amsterdam, Amsterdam, Netherlands, ² Department of Experimental Immunology, Amsterdam Institute for Infection & Immunity, Amsterdam UMC, University of Amsterdam, Amsterdam, Netherlands, ³ Amsterdam Rheumatology & Immunology Center (ARC), Academic Medical Center, Amsterdam, Netherlands

OPEN ACCESS

Edited by:

David Simon,
University of Erlangen
Nuremberg, Germany

Reviewed by:

Qiang Xu,
Guangzhou University of Chinese
Medicine, China
Anabela Barcelos,
New University of Lisbon, Portugal
Meghna Jani,
The University of Manchester,
United Kingdom

*Correspondence:

Lisa G. M. van Baarsen
e.g.vanbaarsen@amsterdamumc.nl

Specialty section:

This article was submitted to
Rheumatology,
a section of the journal
Frontiers in Medicine

Received: 19 May 2021

Accepted: 12 July 2021

Published: 17 August 2021

Citation:

Bolt JW, van Ansenwoude CMJ,
Hammoura I, van de Sande MG and
van Baarsen LGM (2021) Translational
Research Studies Unraveling the
Origins of Psoriatic Arthritis: Moving
Beyond Skin and Joints.
Front. Med. 8:711823.
doi: 10.3389/fmed.2021.711823

Patients with psoriatic arthritis (PsA) are suffering from a decreased quality of life despite currently available treatments. In the latest years, novel therapies targeting the IL-17/IL-23 and TNF pathways improved clinical outcome. Despite this, remission of disease is not achieved in a considerable group of patients, continuous treatment is very often required to reach clinical remission, and prevention of PsA in patients with psoriasis (PsO) is currently impossible. A better understanding of PsA pathogenesis is required to develop novel treatment strategies that target inflammation and destruction more effectively and at an early stage of the disease, or even before clinically manifest disease. The skin is considered as one of the sites of onset of immune activation, triggering the inflammatory cascade in PsA. PsO develops into PsA in 30% of the PsO patients. Influenced by environmental and genetic factors, the inflammatory process in the skin, entheses, and/or gut may evolve into synovial tissue inflammation, characterized by influx of immune cells. The exact role of the innate and adaptive immune cells in disease pathogenesis is not completely known. The involvement of activated IL-17A+ T cells could implicate early immunomodulatory events generated in lymphoid organs thereby shaping the pathogenic inflammatory response leading to disease. In this perspective article, we provide the reader with an overview of the current literature regarding the immunological changes observed during the earliest stages of PsA. Moreover, we will postulate future areas of translational research aimed at increasing our knowledge on the molecular mechanisms driving disease development, which will aid the identification of novel potential therapeutic targets to limit the progression of PsA.

Keywords: early psoriatic arthritis, psoriasis, immunopathogenesis, translational, animal models

KEY MESSAGES

- Most translational research studies investigated blood, skin, and synovial tissues from PsA patients with established disease and showed that IL-23/IL-17 axis and TNF are of importance for the pathogenesis of PsA together with DAMPs, DCs, PMNs, keratinocytes and T cells.
- Consensus is needed to define the different phases of PsA development starting from the preclinical phase.

- Prospective studies that follow PsO patients toward the development of PsA are required to identify predictive biomarkers as well as the molecular and cellular processes associated with development of PsA.
- Translational research studies in tissues beyond skin and synovium, such as lymphoid organs, will shed new light on the immunological processes potentially initiating disease pathogenesis.
- To test potential targets for treatment, improved animal models are needed that better resemble the transition from PsO to PsA.

INTRODUCTION

Psoriatic arthritis (PsA) is a heterogeneous immune-mediated inflammatory disease with musculoskeletal symptoms including arthritis, enthesitis, dactylitis, and axial involvement. Psoriasis (PsO), affecting both skin and nails, is present in most PsA patients (1). Other non-musculoskeletal features linked to PsA are uveitis and colitis, which are observed in a smaller number of patients. Prevalence of PsA is equal in women and men, who suffer from subsequent structural damage and loss of quality of life (2, 3). In 80% of the patients PsA is preceded by PsO and 30% of the PsO patients develop PsA over an average time of 10 years (2–8). Recognition and characterization of this subgroup of PsO patients with an increased risk for (developing) PsA could facilitate early identification of PsA and for this reason screening tools for dermatologists to ease the recognition of PsA have been developed (5, 9–11). These screening tools are used with different levels of success (5), and more than one out of ten PsA diagnoses is missed in the PsO population (12–14). Diagnostic delays until 5 years are reported (15) and this hampers the start of treatment in the early phase of disease. Diagnosis in the first year after the onset of symptoms has been shown to decrease structural damage and improve clinical and patient-reported outcomes (16–19). These improvements are probably caused by the earlier initiation of treatment. Despite revolutionary improvements in treatment over the last years, disease remission is only achieved in up to 15% of the patients (20), and it is not possible to prevent the transition from PsO to PsA. Preventive treatment strategies are hampered by the absence of predictive biomarkers for PsA development that allow the identification of those patients with a very high risk to develop disease, and lack of knowledge on potential cellular or molecular treatment targets. Many efforts have been made to delineate the molecular pathways involved in PsA pathogenesis. However, knowledge on altered molecular pathways in individuals with an increased risk of developing PsA is lacking. Insight into such pathways may lead to the discovery of novel drug targets for preventive treatment in those at-risk individuals and may lead to the identification of biomarkers associated with PsA development. Here we will first give an overview of the current understanding of PsA from both a clinical and immunological perspective based on studies in animal models and humans focused on the earliest, even preclinical, phases of PsA. Secondly, we will describe future areas

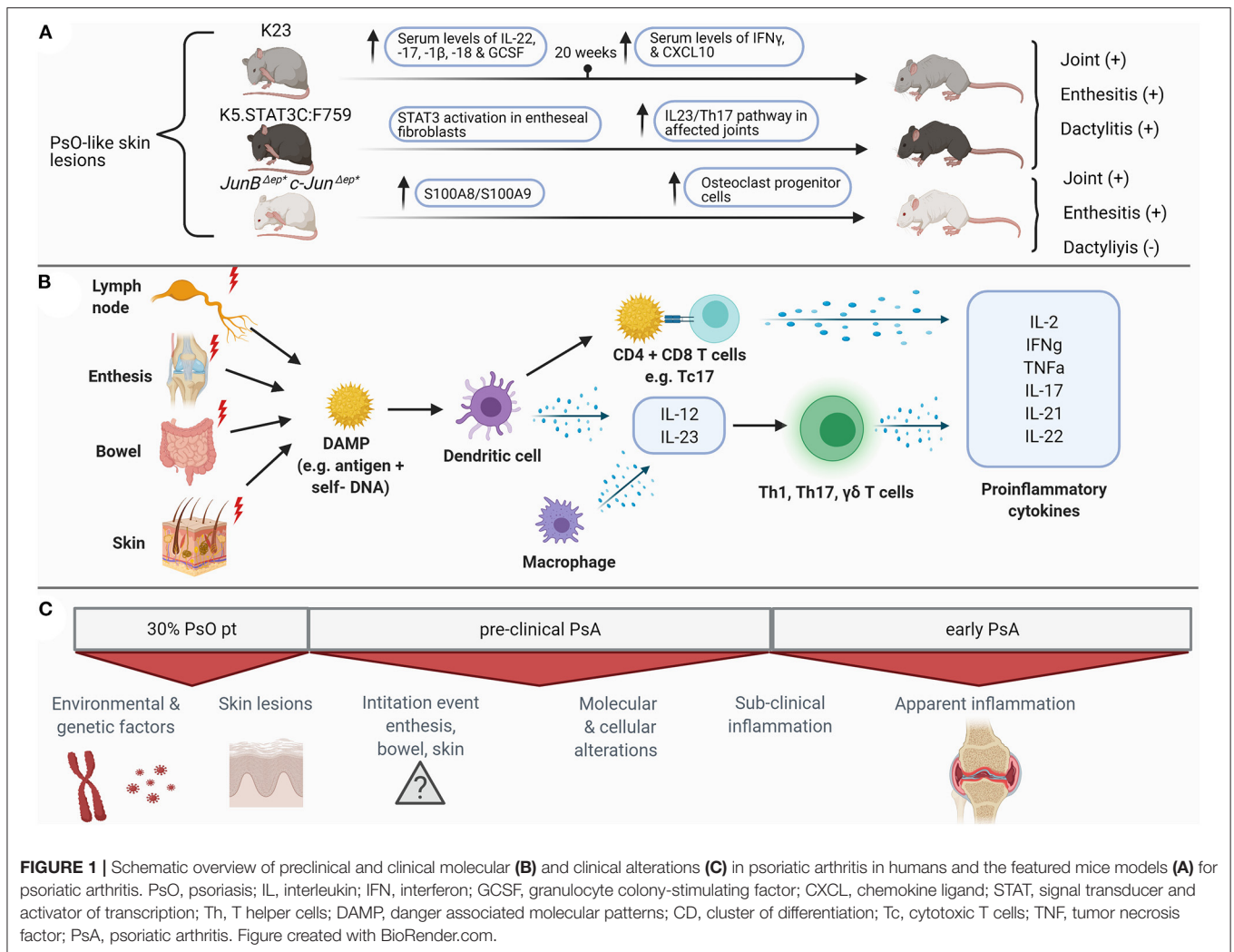
of translational research, which will increase our knowledge of the molecular mechanisms driving disease development.

CHALLENGES IN EARLY RECOGNITION OF PSORIATIC ARTHRITIS

Clinical recognition of PsA in an early phase of the disease and consensus on its terminology is needed to identify and study early molecular changes in PsA patients. Recognition of PsA is however challenging due to the subtlety of the first symptoms and as a result, around 15% of all PsA patients are missed at the dermatology clinics (5, 12). Various clinical screening tools have been developed to accelerate early identification (9, 10, 21). These tools are based on studies that evaluated risk factors for having PsA when having PsO and contain various clinical features (22), such as nail (23) and scalp involvement, increased severity of PsO, family history of PsO (23), a high body mass index and late onset of PsO (6, 24–27), to be more frequently present in these patients. However, the downside of these questionnaire-based tools is the varying specificities and sensitivities, especially when applied in populations with a lower prevalence of PsA (9, 10, 21). The concept and terminology of early PsA have only recently gained popularity among researchers in the field of PsA. The challenges that occur in the recognition and terminology of early PsA hamper the availability of a clear overview of literature focusing on this topic. Currently, collaborating initiatives between dermatologists and rheumatologists are focusing on early PsA and these initiatives will hopefully accomplish more consensus on duration and terminology in early PsA (28). The importance of adopting such a concept has been proven in rheumatoid arthritis (RA), where consensus on early disease stages resulted in more uniform terminology for early disease and facilitated clinical trials in patients with early, or even at-risk, RA. There is no consensus on the exact terminology and duration of the early phase of PsA (28). In this perspective article, we will refer to early PsA as the first 2 years after diagnosis of PsA although others have suggested a shorter period after PsA diagnosis to be defined as early disease (22).

IMMUNOPATHOGENESIS OF PSORIATIC ARTHRITIS: LESSONS LEARNED FROM FUNDAMENTAL RESEARCH

The clear challenges in recognizing early PsA are reflected by the limited number of human studies that have focused on elucidating the immunological drivers of early PsA. For this reason, current knowledge regarding immunopathogenesis is mostly derived from human samples obtained from patients with established PsA (summarized in **Figure 1**). Here we will discuss the current knowledge on immunopathogenesis in established and early PsA and highlight some of the pivotal cytokines involved in PsA pathogenesis. Moreover, we will compare human studies with several animal models reflecting clinical and immunological aspects of human PsA. The current available animal models, shown in **Table 1**, include different



mechanisms for disease induction. These models reflect at least partly human PsA pathogenesis, but none of these models are completely representative.

Initiation of Psoriatic Arthritis

It has been postulated that the initiation of PsA occurs after triggering by environmental factors in gut, entheses, or skin in patients who are genetically more susceptible to development of this disease (37). The genetics of PsA have been reviewed in detail (38–41). In short, both HLA and non-HLA coding genes are associated with PsA. HLA coding genes linked to PsA have supported the role for peptide binding and T cell activation in PsA pathogenesis (40). In the non-HLA coding groups genes are related to innate immunity, cytokines (TNF, IFN, and IL-23/17) and antigen processing and presentation (38). However, genes that are specifically associated with development of PsA in PsO patients remain scarce (39). Two HLA-B27 haplotypes (C*01 and C*02, respectively) (42, 43) and IL13 gene polymorphism were suggested to be independent associated with PsA development in PsO patients (44, 45). The role of the gut microbiome in PsA pathogenesis has recently gained interest (46). Compared

to healthy individuals, PsO and PsA patients have different gut microbiome profiles (46). PsO and PsA patients have overlapping gut microbiome profiles, but differences between the two are also observed (46). Next to microbiome alterations, higher levels of fecal calprotectin and asymptomatic gastrointestinal inflammation are observed in PsA patients (47). It is suggested that intestinal permeability, alterations in immune homeostasis in the bowel and an imbalance of fatty-acid-producing bacteria could be involved in the immunopathogenesis of PsA, but these topics will need further investigation to understand their exact involvement in PsA (46). It is suggested that enthesitis can trigger arthritis via the so-called synovio-entheseal complex (SEC) which highlights the regional relationship between entheses and neighboring synovial membrane (48). Tissue studies investigating molecular alterations in human entheses that could drive PsA onset are highly challenging to perform and thus scarce (49). The few available studies in SpA have shown early vascularization and immune cell infiltration in entheses (50), and have implicated a possible disease-inducing role via enthesal mesenchymal stromal cells (51) and the production of pivotal pro-inflammatory cytokines by resident myeloid cells

(52). In contrast to the gut and the entheses, several studies have investigated the underlying molecular alterations in synovium of PsA patients and psoriatic skin lesions. Of interest is the comparison between skin of PsO and PsA patients, which is informative to understand the relation between the two diseases. These kinds of studies are unfortunately scarce (53, 54), but cross-sectional comparisons do point to molecular differences between skin of PsO and PsA patients (54).

From Psoriasis to Psoriatic Arthritis

In psoriatic skin, an imbalance in the composition of the microbiome (55) and colonization with pathogens (56) is observed, which is suggested to play a role in the external triggering of disease. These findings, as well as mechanical trauma, are thought to initiate the onset of psoriasis by activating keratinocytes (57). Activated and stressed keratinocytes and infiltrating neutrophils then release self-DNA and antimicrobial peptides (AMPs) (57) of which some are overlapping in PsO and PsA (58). The complex of self-DNA and AMP is protected from extracellular nuclease degradation and functions as a danger-associated molecular pattern (DAMP) in the psoriatic skin where it activates abundantly present plasmacytoid dendritic cells (pDCs) and myeloid dendritic cells (mDCs) (59–61). The disruption of these complexes via topical Imiquimod treatment, which is a toll like receptor ligand, was shown to result in the alleviation of the psoriatic skin lesions in the imiquimod-induced psoriasis-like mice model (62) and supports the importance of these complexes in PsA. After the activation by DAMPs, the dermal mDCs probably migrate to lymph nodes (LN) and stimulate differentiation and proliferation of T cells through the production of cytokines such as IL-12 and IL-23 resulting in Th1 and Th17 differentiation (63–65). The exact location and mechanisms of T cell activation have not yet been uncovered (66), but it is proposed that in skin and synovium this takes place through a common antigen as analyses revealed the presence of similar T cell clones in both tissues of PsA patients (67). This theory is supported by the observation of the antigen LL37 (a cationic AMP) being present in both synovial tissue and skin (60, 68). In one study, even antibodies targeting this antigen have been detected in synovial fluid and plasma of patients (60), but the exact role of these autoantibodies in PsA pathogenesis needs further investigation and confirmation. Upon activation, T cells migrate to peripheral tissues such as joints and skin, which is reflected by the abundant amount of Th17 cells in peripheral blood and synovial fluid already in the early stages of PsA (69). This finding suggests that the upregulation of Th1 and Th17 cells and their effector cytokines are essential for the start of a continuous inflammatory response (69). Recently, IL-17A+ CD8+ T cells gained interest, also due to their strong association with the major histocompatibility complex (MHC) class I, of which the genes predispose to PsA (70). CD8+ T cells are polyfunctional enabling the production of a wide range of cytokines both in skin and joints (66). In blood of PsA patients, more memory CD8+ T cells have been found compared to healthy controls and PsO patients (53), underlining a possible important role of memory CD8+ T cells in PsA. Also in synovial fluid of PsA patients, these CD8+ cells are abundantly present and are clonally expanded more extensively than CD4+ T cells

(66, 71). Transcriptome analysis showed that these expanded CD8+ T cells in the joint are memory cells expressing tissue-homing as well as tissue-resident markers (Trm cells) (71). The role of T cells in steering PsA development is supported by a study in JunB/c-Jun double-mutant mice, in which mice keratinocyte-specific JunB and c-Jun transcription factors are deleted in adult mice (by inducible knock out) causing psoriatic-like plaques with accumulation of neutrophils, macrophages, and T cells in the and joint inflammation (34). It was shown that when inducing absence of functional B and T cells by creating Rag2-deficient JunB/c-Jun double-mutant mice (34), these mice develop psoriatic skin lesions (34) though with a strong reduction in inflammation of the joints. It is not clear which exact mechanisms are responsible for the migration of T cells to different peripheral tissues. It is hypothesized that in PsA T cells migrate from the skin or peripheral blood to the other affected tissues (72). In 2 mice studies specific molecular alterations in the skin resulted in joint inflammation with a clear role for T cells in the JunB/C-Jun double mutant mice (29, 31, 34) (**Table 1**). However detailed migration of T cells was not investigated. The final inflammatory response in PsA is characterized by a self-perpetuating positive feedback loop with recruitment of various immune cells such as macrophages and polymorphonuclear leukocytes (PMNs) in all affected organs, keratinocyte proliferation and neutrophil accumulation in the skin, bone metabolism alterations in the joint and spine, and angiogenesis causing the clinical spectrum of PsA (63, 73, 74). Up until now, B cells are considered to have no profound role in this inflammatory response (75), but this is an ongoing debate in the field (71).

Pivotal Cytokines Involved in the Immunopathogenesis of Psoriatic Arthritis

In the inflammatory response of PsA, various cytokines are released. Many of these cytokines are found in both PsO and PsA, but not all are equally present which may indicate at least partially different mechanisms of disease pathogenesis. One of the key cytokines is type I interferon (IFN), which is important for the onset of both the innate and adaptive immune response. Via release of IFN α by activated pDC in the skin (61), IFN α stimulates activation of myeloid DCs (57, 61, 65, 76) resulting in T-cell differentiation and proliferation as discussed earlier. IFN α activity is increased in psoriatic skin compared to unaffected skin (77) and in synovial fluid of patients with PsA compared to patients with osteoarthritis (60). Accordingly, activation of IFN α pathway in synovium is similar to skin (60) and suggests an important role for this cytokine at an early stage of the immunopathogenesis of PsA. Type II IFN, IFN γ , also induces different processes in PsA by activating antigen-presenting cells early in the psoriatic cascade (64). This cytokine is present in both PsA synovium and lesional skin, and levels correlate with disease severity (78). Blocking of IFN γ was earlier proposed as a potential treatment for PsA. However, so far no IFN blocking treatments have been developed for PsA, possibly because at the same time IL-17 targeting treatments emerged (64). However, IFN could still be of interest as is exemplified by the stimulation of keratinocytes in psoriasis-like skin in mice caused by IFN γ (79). This increase resulted in increased levels of Th1 and Th17 cells

TABLE 1 | Characteristics of animals models for psoriatic arthritis and their contribution to understanding disease pathogenesis.

Animal model	Mice	Type of induction	Clinical or imaging disease manifestation	Histological disease manifestation	Suggested contribution to understanding pathogenesis PsA
R26STAT3Cstopfl/fl CD4Cre transgenic mice (29, 30)	R26STAT3Cstopfl/fl CD4Cre mice	Hyperactive expression of STAT3C in naïve T cells, resulting in expansion of Th17 cells.	Psoriasis-like skin inflammation (+) Arthritis (-) Enthesitis (-) Dactylitis (-) Bone erosion (+) Osteopenia (+)	Psoriasis-like skin inflammation (+) Synovitis (+) Enthesitis (+)	STAT3 signaling and Th17 cells are of importance for PsA-like disease development. RANKL expression and subsequent osteoclast differentiation is related to IL-17 from Th17 cells that act on mesenchymal cells.
K5.Stat3C:F759 transgenic mice (31)	K5.Stat3C:F759 mice	Hyperactive expression of STAT3C in keratinocytes and constitutive activation of IL-6 signaling that together lead to persistent STAT3 activation due to impaired SOCS3-negative feedback with induction of IL17A.	Psoriasis-like skin inflammation (+) Arthritis (+) Enthesitis (+) Dactylitis (+)	Psoriasis-like skin inflammation (+) Synovitis (+)	Facilitating role of skin inflammation for joint inflammation via crosstalk between keratinocytes and entheseal fibroblasts mediated by IL-6 and IL-23/Th17-associated STAT3 activation.
Mannan induced psoriatic disease in B10Q mice (32, 33)	B10Q mice	Intra peritoneal injection of Mannan, a ligand for mannose receptors at the host-fungus interactions and known as the major trigger of interleukin-17 (IL-17) production in B10Q mice.	Psoriasis-like skin inflammation (+) Arthritis* (+) Enthesitis (-) Dactylitis (-) New bone formation (+)	Psoriasis-like skin inflammation (+) Synovitis (-) Enthesitis (+) New bone formation (+)	Supporting role for macrophages in inducing PsA-like disease via triggering of IL-17A secretion by $\gamma\delta$ T cells and ILC3. These studies showed that reactive oxygen species (ROS) production by macrophages is protective for PsA-like disease whereas nitric oxygen species promote disease.
Mannan induced psoriatic disease in B10Q.Ncf1 ^{m1j/m1} mice (32, 33)	B10Q.Ncf1 ^{m1j/m1} mice	The Ncf1 mutation impairs superoxide anion production by the NOX2 complex and impaired ROS production by macrophages. This worsened the clinical phenotype.	Psoriasis-like skin inflammation (+) Arthritis (+) Enthesitis (+) New bone formation (+)	Psoriasis-like skin inflammation (+) Synovitis (+) Enthesitis (+) New bone formation (+) Erosions (+)	
JunB/c-Jun double-mutant mice (34)	JunBf/f c-Junf/f K5-Cre-ERT mice	Inducible epidermal deletion of JunB and c-Jun. In humans the JunB transcription factor is localized in psoriasis susceptibility locus 6 and regulates cell proliferation, differentiation, stress responses and cytokine expression.	Psoriasis-like skin inflammation (+) Arthritis (+) Dactylitis (-)	Psoriasis-like skin inflammation (+) Synovitis (+) Bone erosions (+)	Epidermal modulation of JunB leads to the secretion of chemokines and cytokines that recruit inflammatory cells in the skin resulting in PsO-like skin lesions and later development of arthritis. This underscores that epidermal changes can induce joint inflammation.
K23 mice (35)	R23/K14CreERT2 mice	Inducible-conditional IL-23 overexpression in keratinocytes.	Psoriasis-like skin inflammation (+) Arthritis (+) Enthesitis (-) Dactylitis (+)	Psoriasis-like skin inflammation (+) Synovitis (+) Enthesitis (+) Dactylitis (+) Bone erosions (+)	IL-23 plays a role in the initiation PsO, which later transits into PsA.
IL-17A minicircle gene transfer model in C57BL/6J mice (36)	C57BL/6J mice	Systemic overexpression of mouse IL-17A <i>in vivo</i> through injection of IL-17A minicircle construct in tail vein resulting in transduction of hepatocytes.	Psoriasis-like skin inflammation (+) Arthritis (-) Enthesitis (-) Dactylitis (-) Bone erosion (+)	Psoriasis-like skin inflammation (+) Bone erosions (+)	Link between IL-17, myelopoiesis and neutrophils. IL-17A induces PsO like skin lesions. IL-17 can induce bone erosions in absence of synovitis.

PsA, psoriatic arthritis; STAT, signal transducer and activator of transcription; Th, T helper cells; RANKL, receptor activator of nuclear factor kappa- β ligand; IL, interleukin; SOCS3, Suppressor of cytokine signaling 3; ILC, innate lymphoid cells; NOX, nicotinamide adenine dinucleotide phosphate oxidase; ROS, reactive oxygen species; NO, nitrogen oxide; PsO, psoriasis.

*Animals showed joint swelling without synovitis on histology.

in the skin of these animals. Additionally, IFN γ can stimulate the inhibition or induction of osteoclasts depending on the relative levels of RANKL and IFN γ (36). The influence of IFN γ on both keratinocytes and osteoclasts suggests regulation of fundamental mechanisms by this cytokine. For this reason, further research into IFN for new treatment targets could be promising. One of the cytokines produced by activated myeloid DCs present in the skin is IL-23, which drives Th17- activation and differentiation. IL-23 is detected in psoriatic skin lesions and inflamed PsA synovium, though levels are highly variable between patients (63, 80). Blocking of IL-23 is very efficient for disease reduction in humans (81–83) and the pivotal role of IL-23 in PsA is also confirmed in K23 mice (35). In these mice, the induction of conditional transgenic expression of IL-23 in keratinocytes resulted in PsA-like symptoms with PsO like skin inflammation followed by arthritis, dactylitis and enthesitis. Th17 cells activated by IL-23 produce several cytokines including IL-17A, IL-17F, and IL-22. IL-17A is another essential cytokine in the pathogenesis and treatment of PsA (84). IL-17A can be produced by T helper cells as well as by innate (like) cells such as innate lymphoid cells (ILCs) (85), iNKT, $\gamma\delta$ T cells, MAIT cells. IL-17A blockade is effective in PsA (84, 86, 87) and results in a decrease in synovial macrophages and neutrophils as well as synovial IL-17A mRNA expression (88). In R26Stat3Cstopfl/fl CD4Cre mice in which the conditional allele of the hyperactive STAT3 gene, STAT3C, is expressed selectively in T lymphocytes resulting in enrichment of Th17 cells (29). It was shown that induced IL-17A expression resulted in both cutaneous and musculoskeletal PsA-like symptoms (**Table 1**). In addition, IL-17A minicircle (mc) injection induced systemic overexpression of IL-17A in C5BL/6J mice resulted in systemic bone erosions and PsO like skin lesions. When in these mice arthritis is induced by collagen (CIA model) then time to arthritis development is reduced with an increased arthritis severity (36). These findings also directly link IL-17A to skin and musculoskeletal symptoms resembling PsA (29). A cytokine that works synergistically with IL-17 is TNF, which is produced by many immune cells and is a key regulator of pro-inflammatory gene transcription, cytokine secretion, cytotoxicity and differentiation of T-helper cells (89). TNF is important for the induction of inflammatory responses, granulopoiesis, psoriasis skin lesions (90), enthesitis (52), and the pathological formation (91) and destruction of bone (92) via induction of RANKL (93). Blockade of TNF is very effective in PsA (94–96) and results in a decrease of inflammation and angiogenesis in skin (97, 98) and synovium (98, 99) and thus clinical PsA symptoms. Several other cytokines such as the IL-10 (100, 101) family [of which IL-22 (102, 103) more specifically], IL-17F (104), IL-36 (105–107), and IL-9 (108) are also suggested to play a role in the immunopathogenesis of PsA, but will not be further discussed here.

Even though the immunopathogenesis of PsA is incompletely understood, the above-mentioned studies indicate a role for both innate and adaptive immune cells, as well as for keratinocytes. The molecular mechanisms steering activation of those cells are yet to be further unraveled. However, it is clear that the IL-23/IL-17 axis and TNF are essential in establishing and enhancing the inflammatory response, resulting in the positive inflammatory feedback loop, bone formation and destruction. A limiting factor

is that many of these human studies focused on patients with established PsA, while only a few studies have focused on early PsA patients. Furthermore, the PsA mice models do not always report the same clinical spectrum as human PsA. Therefore, more human studies and improved animal models are required focussing on the earliest or even preclinical phase of PsA.

CHALLENGES IN STUDYING THE EARLIEST PHASES OF PSORIATIC ARTHRITIS

Studies on early and preclinical disease are required as these studies will improve our knowledge on the pathogenesis of PsA and the transition of PsO to PsA which is essentially required if we ultimately want to come to preventive treatment or even a cure for PsA. Such studies will help to better understand the molecular pathways steering the transition from PsO to PsA and aid the identification of potential drug targets aimed at preventing the onset of PsA. Ultimately, this may lead to the development of risk stratification tools that identify those individuals at the highest risk of PsA who may benefit from preventative treatment. To study the earliest phases of PsA, consensus on the definition of early and preclinical PsA stages is needed as proposed in a recent review (22). A recent Delphi study could find consensus on some of the terminology, but especially the early phase needs further discussion (28). Such definitions will ease the use of preclinical terminology in studies that investigate clinical, imaging and molecular characteristics that determine the transition from PsO towards PsA in prospective cohort studies. These prospective studies will provide identification of novel risk factors associated with the development of PsA. So far, only a few prospective studies have been reported (109–111). An eight-year follow-up study in PsO patients confirmed nail involvement (RR 2.5, $p = 0.002$) associated with PsA development, additionally to a severe psoriasis phenotype (RR 5.4, $p = 0.006$), presence of low level of education (RR 0.30, $p = 0.049$), and uveitis (RR 31.5, $p = 0.0002$) (109). This study also showed that in PsO patients symptoms like arthralgia in women (HR 2.59, $p = 0.02$), fatigue (HR 2.36, $p = 0.007$), heel pain (HR 4.18, $p = 0.02$), and stiffness (HR 2.03, $p = 0.045$) at baseline were predictive for PsA development, as well as worsening of some of these symptoms (111). It is hypothesized that onset of clinically manifest PsA is preceded by a phase of subclinical enthesitis or arthritis (22). Subclinical inflammation could be visualized by various imaging techniques (22) and is not reflected by clinical symptoms. Subclinical enthesitis on ultrasound (US) was shown in the knees of around 83% of PsO patients compared to individuals without PsO that showed no signs of enthesitis on US (112). Another study using US in PsO patients showed subclinical enthesitis including grayscale and power Doppler in 49.3% in PsO patients compared to healthy controls (113). After 42 weeks of ustekinumab treatment in these PsO patients, their inflammation scores decreased by 47.5%. High-resolution peripheral quantitative CT in a prospective cohort of PsO patients with a mean follow-up of 28 months showed that subclinical enthesitis, characterized by structural enthesal lesions, was significantly associated with an increased

risk for PsA development (HR 5.10, $p = 0.008$) (114). High volumetric cortical bone mineral density was associated with a lower risk for PsA development (HR 0.64). An imaging study with MRI in PsO patients has shown that synovitis in the hands of PsO patients combined with arthralgia, is suggestive of subclinical inflammation with a risk of 60% for PsA development after 1 year of follow-up (110). In a cohort of PsO patients with sub-clinical inflammation, characterized by synovitis and enthesitis on MRI and CT, this inflammation improved after 24 weeks of secukinumab treatment (115). Bone erosion and osteoproliferation remained stable during treatment and scores representing synovitis and tenosynovitis significantly decreased ($p = 0.005$). In this cohort also arthralgia significantly decreased ($p = 0.003$). Whether subclinical inflammation reflects an increased risk of development of clinically manifest disease needs further validation in additional prospective longitudinal studies. Prospective studies on both clinical and imaging factors in relation to disease development over time will help to identify specific groups that have a high risk for PsA development. In-depth tissue studies in this high-risk group to further understand the immunological alterations preceding disease onset may aid the development of innovative and even preventive therapies limiting disease progression.

NEW HORIZONS FOR TRANSLATIONAL STUDIES

As said, studying target tissues of early PsA patients and PsO patients with a high risk of PsA development will provide novel insights into the immunopathogenesis of PsA. Therefore, detailed analyses of tissues involved in the initial triggering of PsA are crucial. As earlier discussed, the onset of PsA is thought to take place outside the joints at sites such as entheses, skin and gut. Even though activated immune cells can migrate from affected psoriatic tissues to draining lymph nodes (LN) where they trigger T helper cell differentiation and initiate an inflammatory response, human lymphoid organs have not yet been studied in PsA. As LNs are the epicenter for T-cell activation and differentiated and activated T cells play a key role in PsA, it will be of great relevance to study those cellular responses in LNs, especially during the onset of PsA. Our research group has recently successfully initiated LN tissue sampling (116–119) of patients with inflammatory arthritis, RA-risk individuals and healthy controls (116–119). We have found altered frequencies of immune cells in LN biopsies of patients with RA and RA-risk individuals (117, 119). We could also show B cell depletion in these inguinal LN biopsies of RA patients after rituximab treatment, while switched memory B cells were more resistant to therapy (120). Preliminary findings in LN biopsies of patients with PsA revealed an unexpected increase in innate cells when compared with healthy LN tissue, which is under current investigation (data not shown). We postulate that LN tissue sampling and analysis will be highly valuable in further elucidating the immune responses involved in the pathogenesis of PsA in addition to studying all other involved tissues in PsA collected during the earliest or preclinical phase of disease. There have already been efforts made to investigate molecular

differences between skin of PsO and PsA patients in cross-sectional studies. Differences have been reported (53, 54), but it is unclear if these observed molecular changes in skin precede or follow PsA development which stresses the importance to study patients during the transition from PsO to PsA using a prospective study design with serial tissue sampling. Although this is highly challenging, only this will show which molecular and cellular processes in the skin change during the transition from PsO to PsA. Additionally, this will show whether specific changes in the skin precede onset of musculoskeletal symptoms or whether the psoriatic skin changes after the onset of PsA as this is still under debate (121). Similar prospective follow-up studies could be applied to other target tissues, but tissues like bone and entheses are in general more challenging to collect in a prospective study. Studying synovial biopsies over time is challenging but possible, even in the absence of arthritis (122–128). When possible, it is preferred to study serial paired tissue samples of various tissues in the same patients. These studies are unfortunately sparse, which is not surprising, since the collection may be seen as too invasive for the patient. Due to the challenges of human in-depth tissue studies, animal models could support human studies. These animal models can give insights into immune alterations in various tissues which are difficult to study in humans, such as the entheses and bone. Moreover, by blocking specific pathways or molecules, causal relationships to development of disease can be investigated. But for this, improved animal models are needed that, contrary to the current models, better resemble the transition process from PsO to PsA in humans.

Concluding we have observed that many efforts have been made to unravel the immunopathogenesis of PsA, however prospective tissue studies in well-defined high-risk individuals are needed to further understand the disease pathogenesis and transition from PsO to PsA. Together with animal models that resemble this transition, these studies will provide further understanding of PsA development and ultimately may result in the development of treatments preventing the onset of PsA in PsO individuals at risk for this disease.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding authors.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

LB received funding from a ZonMw VIDI project (91718371). MS received funding from a ZonMw VENI project (09150161810112). LB and MS received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 847551 (ARCAID).

REFERENCES

- Ritchlin CT, Colbert RA, Gladman DD. Psoriatic arthritis. *N Engl J Med*. (2017) 376:957–70. doi: 10.1056/NEJMra1505557
- Wilson FC, Icen M, Crowson CS, McEvoy MT, Gabriel SE, Kremers HM. Incidence and clinical predictors of psoriatic arthritis in patients with psoriasis: a population-based study. *Arthritis Care Res*. (2009) 61:233–9. doi: 10.1002/art.24172
- Jamshidi F, Bouzari N, Seirafi H, Farnaghi F, Firooz A. The prevalence of psoriatic arthritis in psoriatic patients in Tehran, Iran. *Arch Iran Med*. (2008) 11:162–5.
- Reich K, Krüger K, Mössner R, Augustin M. Epidemiology and clinical pattern of psoriatic arthritis in Germany: a prospective interdisciplinary epidemiological study of 1511 patients with plaque-type psoriasis. *Br J Dermatol*. (2009) 160:1040–7. doi: 10.1111/j.1365-2133.2008.09023.x
- Mease PJ, Gladman DD, Helliwell P, Khraishi M, Fuiman J, Bananis E, et al. Comparative performance of psoriatic arthritis screening tools in patients with psoriasis in European/North American dermatology clinics. *J Am Acad Dermatol*. (2014) 71:649–55. doi: 10.1016/j.jaad.2014.05.010
- Alenius GM, Stenberg B, Stenlund H, Lundblad M, Dahlqvist SR. Inflammatory joint manifestations are prevalent in psoriasis: prevalence study of joint and axial involvement in psoriatic patients, and evaluation of a psoriatic and arthritic questionnaire. *J Rheumatol*. (2002) 29:2577–82.
- Ohara Y, Kishimoto M, Takizawa N, Yoshida K, Okada M, Eto H, et al. Prevalence and clinical characteristics of psoriatic arthritis in Japan. *J Rheumatol*. (2015) 42:1439–42. doi: 10.3899/jrheum.141598
- Christophers E, Barker J, Griffiths C, Daudén E, Milligan G, Molta C, et al. The risk of psoriatic arthritis remains constant following initial diagnosis of psoriasis among patients seen in European dermatology clinics. *J Eur Acad Dermatol Venereol*. (2010) 24:548–54. doi: 10.1111/j.1468-3083.2009.03463.x
- Gladman DD, Schentag CT, Tom BDM, Chandran V, Brockbank J, Rosen C, et al. Development and initial validation of a screening questionnaire for psoriatic arthritis: the toronto psoriatic arthritis screen (ToPAS). *Ann Rheum Dis*. (2009) 68:497–501. doi: 10.1136/ard.2008.089441
- Tinazzi I, Adami S, Zanolin EM, Caimmi C, Confente S, Girolomoni G, et al. The early psoriatic arthritis screening questionnaire: A simple and fast method for the identification of arthritis in patients with psoriasis. *Rheumatology*. (2012) 51:2058–63. doi: 10.1093/rheumatology/kes187
- De Groot M, Picavet DI, Van Kuijk AWR, Tak PP, Bos JD, De Rie MA, et al. A prospective, randomized, placebo-controlled study to identify biomarkers associated with active treatment in psoriatic arthritis: effects of adalimumab treatment on lesional and nonlesional skin. *Dermatology*. (2013) 225:298–303. doi: 10.1159/000343290
- Villani AP, Rouzaud M, Sevrain M, Barnetche T, Paul C, Richard MA, et al. Prevalence of undiagnosed psoriatic arthritis among psoriasis patients: systematic review and meta-analysis. *J Am Acad Dermatol*. (2015) 73:242–8. doi: 10.1016/j.jaad.2015.05.001
- Spelman L, Su JC, Fernandez-Peñas P, Varigos GA, Cooper AJ, Baker CS, et al. Frequency of undiagnosed psoriatic arthritis among psoriasis patients in Australian dermatology practice. *J Eur Acad Dermatol Venereol*. (2015) 29:2184–91. doi: 10.1111/jdv.13210
- Mease PJ, Palmer JB, Hur P, Strober BE, Lebwohl M, Karki C, et al. Utilization of the validated psoriasis epidemiology screening tool to identify signs and symptoms of psoriatic arthritis among those with psoriasis: a cross-sectional analysis from the US-based Corrona Psoriasis Registry. *J Eur Acad Dermatol Venereol*. (2019) 33:886–92. doi: 10.1111/jdv.15443
- Kavanaugh A, Helliwell P, Ritchlin CT. Psoriatic arthritis and burden of disease: patient perspectives from the population-based multinational assessment of psoriasis and psoriatic arthritis (MAPPS) survey. *Rheumatol Ther*. (2016) 3:91–102. doi: 10.1007/s40744-016-0029-z
- Van Mens LJJ, De Jong HM, Fluri I, Nurmohamed MT, Van de Sande MGH, Kok M, et al. Achieving remission in psoriatic arthritis by early initiation of TNF inhibition: a double-blind, randomised, placebo-controlled trial of golimumab plus methotrexate versus placebo plus methotrexate. *Ann Rheum Dis*. (2019) 78:610–6. doi: 10.1136/annrheumdis-2018-214746
- Coates LC, Moverley AR, McParland L, Brown S, Navarro-Coy N, O'Dwyer JL, et al. Effect of tight control of inflammation in early psoriatic arthritis (TICOPA): a UK multicentre, open-label, randomised controlled trial. *Lancet*. (2015) 386:2489–98. doi: 10.1016/S0140-6736(15)00347-5
- Tillett W, Jadon D, Shaddick G, Cavill C, Korendowych E, De Vries CS, et al. Smoking and delay to diagnosis are associated with poorer functional outcome in psoriatic arthritis. *Ann Rheum Dis*. (2013) 72:1358–61. doi: 10.1136/annrheumdis-2012-202608
- Haroon M, Gallagher P, FitzGerald O. Diagnostic delay of more than 6 months contributes to poor radiographic and functional outcome in psoriatic arthritis. *Ann Rheum Dis*. (2015) 74:1045–50. doi: 10.1136/annrheumdis-2013-204858
- Hagège B, Tan E, Gayraud M, Fautrel B, Gossec L, Mitrovic S. Remission and low disease activity in psoriatic arthritis publications: a systematic literature review with meta-analysis. *Rheumatology*. (2020) 59:1818–25. doi: 10.1093/rheumatology/keaa030
- Ibrahim G, Buch M, Lawson C, Waxman R. Evaluation of an existing tool for psoriatic arthritis: the psoriasis epidemiology screening tool (PEST) questionnaire. *Clin Exp Immunol*. (2009) 27:469–74.
- Scher JU, Ogdie A, Merola JF, Ritchlin C. Preventing psoriatic arthritis: focusing on patients with psoriasis at increased risk of transition. *Nat Rev Rheumatol*. (2019) 15:153–66. doi: 10.1038/s41584-019-0175-0
- Langenbruch A, Radtke MA, Krensel M, Jacobi A, Reich K, Augustin M. Nail involvement as a predictor of concomitant psoriatic arthritis in patients with psoriasis. *Br J Dermatol*. (2014) 171:1123–8. doi: 10.1111/bjd.13272
- Egeberg A, Skov L, Zachariae C, Gislason GH, Thyssen JP, Mallbris L. Duration of psoriatic skin disease as risk factor for subsequent onset of psoriatic arthritis. *Acta Derm Venereol*. (2018) 98:546–50. doi: 10.2340/00015555-2912
- Queiro R, Tejón P, Alonso S, Coto P. Age at disease onset: a key factor for understanding psoriatic disease. *Rheumatology*. (2014) 53:1178–85. doi: 10.1093/rheumatology/ket363
- Queiro R, Alperi M, Alonso S, Ballina J, Huergo-Zapico L, Fernandez-Guizan A, et al. Patients with psoriatic arthritis may show differences in their clinical and genetic profiles depending on their age at psoriasis onset. *Clin Exp Rheumatol*. (2012) 30:476–80.
- El Miedany Y, El Gaafary M, Youssef S, Ahmed I, Nasr A. Tailored approach to early psoriatic arthritis patients: clinical and ultrasonographic predictors for structural joint damage. *Clin Rheumatol*. (2015) 34:307–13. doi: 10.1007/s10067-014-2630-2
- Perez-Chada LM, Haberman RH, Chandran V, Rosen CF, Richlin C, et al. Consensus terminology for preclinical phases of psoriatic arthritis for use in research studies: results from a Delphi consensus study. *Nat Rev Rheumatol*. (2021) 17:238–43. doi: 10.1038/s41584-021-00578-2
- Yang L, Fanok MH, Mediero-Munoz A, Fogli LK, Corciulo C, Abdollahi S, et al. Augmented Th17 differentiation leads to cutaneous and synovio-enthesal inflammation in a novel model of psoriatic arthritis. *Arthritis Rheumatol*. (2018) 70:855–67. doi: 10.1002/art.40447
- Fogli LK, Sundrud MS, Goel S, Bajwa S, Jensen K, Derudder E, et al. T Cell-derived IL-17 mediates epithelial changes in the airway and drives pulmonary neutrophilia. *J Immunol*. (2013) 191:3100–11. doi: 10.4049/jimmunol.1301360
- Yamamoto M, Nakajima K, Takaishi M, Kitaba S, Magata Y, Kataoka S, et al. Psoriatic inflammation facilitates the onset of arthritis in a mouse model. *J Invest Dermatol*. (2015) 135:445–53. doi: 10.1038/jid.2014.426
- Zhong J, Scholz T, Yau ACY, Guerard S, Hüffmeier U, Burkhardt H, et al. Mannan-induced Nos2 in macrophages enhances IL-17-driven psoriatic arthritis by innate lymphocytes. *Sci Adv*. (2018) 4:2–11. doi: 10.1126/sciadv.aas9864
- Khmaladze I, Kelkka T, Guerard S, Wing K, Pizzolla A, Saxena A, et al. Mannan induces ROS-regulated, IL-17A-dependent psoriasis arthritis-like disease in mice. *Proc Natl Acad Sci USA*. (2014) 111:E3669–78. doi: 10.1073/pnas.1405798111
- Zenz R, Eferl R, Kenner L, Florin L, Hummerich L, Mehic D, et al. Psoriasis-like skin disease and arthritis caused by inducible epidermal deletion of Jun proteins. *Nature*. (2005) 437:369–75. doi: 10.1038/nature03963
- Chen L, Deshpande M, Grisotto M, Smaldini P, Garcia R, He Z, et al. Skin expression of IL-23 drives the development of psoriasis and psoriatic arthritis in mice. *Sci Rep*. (2020) 10:4–14. doi: 10.1038/s41598-020-65269-6

36. Adamopoulos IE. IL-17A gene transfer induces bone loss and epidermal hyperplasia associated with psoriatic arthritis. *Ann Rheum Dis.* (2015) 74:1284–92. doi: 10.1136/annrheumdis-2013-204782
37. McGonagle D, Aydin SZ, Gül A, Mahr A, Direskeneli H. 'MHC-I-opathy'-unified concept for spondyloarthritis and Behçet disease. *Nat Rev Rheumatol.* (2015) 11:731–40. doi: 10.1038/nrrheum.2015.147
38. Rahmati S, Tsoi L, O'Rielly D, Chandran V, Rahman P. Complexities in genetics of psoriatic arthritis. *Curr Rheumatol Rep.* (2020) 22:7–9. doi: 10.1007/s11926-020-0886-x
39. Mulder MLM, van Hal TW, Wenink MH, Koenen H, Van den Hoogen F, De Jong E, et al. Clinical, laboratory, and genetic markers for the development or presence of psoriatic arthritis in psoriasis patients: a systematic review. *Arthritis Res Ther.* (2021) 23:168. doi: 10.1186/s13075-021-02545-4
40. Winchester R, Fitzgerald O. MHC class I associations beyond HLA-B27: The peptide binding hypothesis of psoriatic arthritis and its implications for disease pathogenesis. *Curr Opin Rheumatol.* (2020) 32:330–6. doi: 10.1097/BOR.0000000000000720
41. O'Rielly DD, Rahman P. Clinical and molecular significance of genetic loci associated with psoriatic arthritis. *Best Pract Res Clin Rheumatol.* (2021) 35:101691. doi: 10.1016/j.berh.2021.101691
42. Winchester R, Minevich G, Steshenko V, Kirby B, Kane D, Greenberg DA, et al. HLA associations reveal genetic heterogeneity in psoriatic arthritis and in the psoriasis phenotype. *Arthritis Rheum.* (2012) 64:1134–44. doi: 10.1002/art.33415
43. Eder L, Chandran V, Pellet F, Shanmugarajah S, Rosen CF, Bull SB, et al. Human leucocyte antigen risk alleles for psoriatic arthritis among patients with psoriasis. *Ann Rheum Dis.* (2012) 71:50–5. doi: 10.1136/ard.2011.155044
44. Eder L, Chandran V, Pellet F, Pollock R, Shanmugarajah S, Rosen CF, et al. IL13 gene polymorphism is a marker for psoriatic arthritis among psoriasis patients. *Ann Rheum Dis.* (2011) 70:1594–8. doi: 10.1136/ard.2010.147421
45. Bowes J, Eyre S, Flynn E, Ho P, Salah S, Warren RB, et al. Evidence to support IL-13 as a risk locus for psoriatic arthritis but not psoriasis vulgaris. *Ann Rheum Dis.* (2011) 70:1016–9. doi: 10.1136/ard.2010.143123
46. Myers B, Brownstone N, Reddy V, Chan S, Thidobeaux Q, Truong A, Bhutani T, et al. The gut microbiome in psoriasis and psoriatic arthritis. *Best Pract Res Clin Rheumatol.* (2019) 33:101494. doi: 10.1016/j.berh.2020.101494
47. Adarsh MB, Dogra S, Vaiphei K, Vaishnavi C, Sinha SK, Sharma A. Evaluation of subclinical gut inflammation using faecal calprotectin levels and colonic mucosal biopsy in patients with psoriasis and psoriatic arthritis. *Br J Dermatol.* (2019) 181:401–2. doi: 10.1111/bjd.17745
48. McGonagle D, Lories RJU, Tan AL, Benjamin M. The concept of a "synovio-entheseal complex" and its implications for understanding joint inflammation and damage in psoriatic arthritis and beyond. *Arthritis Rheum.* (2007) 56:2482–91. doi: 10.1002/art.22758
49. Perrin C. Focal synovial inflammation versus enthesitis theory in distal psoriatic arthritis. *Am J Dermatopathol.* (2021) Publish Ah:1–5. doi: 10.1097/DAD.0000000000001947
50. McGonagle D, Marzo-Ortega H, O'Connor P, Gibbon W, Hawkey P, Henshaw K, et al. Histological assessment of the early enthesitis lesion in spondyloarthropathy. *Ann Rheum Dis.* (2002) 61:534–7. doi: 10.1136/ard.61.6.534
51. Russell T, Watad A, Bridgewood C, Rowe H, Khan A, Rao A, et al. IL-17A and TNF modulate normal human spinal enthesal bone and soft tissue mesenchymal stem cell osteogenesis, adipogenesis, and stromal function. *Cells.* (2021) 10:341. doi: 10.3390/cells10020341
52. Bridgewood C, Watad A, Russell T, Palmer TM, Marzo-Ortega H, Khan A, et al. Identification of myeloid cells in the human enthesis as the main source of local IL-23 production. *Ann Rheum Dis.* (2019) 78:929–33. doi: 10.1136/annrheumdis-2018-214944
53. Leijten EFA, Id O, Kempen TSVAN, Pouw JN, Kleinrensink NJ, Vincken NL, et al. *Tissue-Resident Memory CD8+ T Cells From Skin Differentiate Psoriatic Arthritis From Psoriasis.* 0–3.
54. Cretu D, Liang K, Saraon P, Batruch I, Diamandis EP, Chandran V. Quantitative tandem mass-spectrometry of skin tissue reveals putative psoriatic arthritis biomarkers. *Clin Proteom.* (2015) 12:1–8. doi: 10.1186/1559-0275-12-1
55. Lewis DJ, Chan WH, Hinojosa T, Hsu S, Feldman SR. Mechanisms of microbial pathogenesis and the role of the skin microbiome in psoriasis: a review. *Clin Dermatol.* (2019) 37:160–6. doi: 10.1016/j.clindermatol.2019.01.011
56. Gudjonsson JE, Thorarinnsson AM, Sigurgeirsson B, Kristinnsson KG, Valdimarsson H. Streptococcal throat infections and exacerbation of chronic plaque psoriasis: a prospective study. *Br J Dermatol.* (2003) 149:530–4. doi: 10.1046/j.1365-2133.2003.05552.x
57. Barnas JL, Ritchlin CT. Etiology and pathogenesis of psoriatic arthritis. *Rheum Dis Clin North Am.* (2015) 41:643–63. doi: 10.1016/j.rdc.2015.07.006
58. Bierkarre H, Harder J, Cuthbert R, Emery P, Leuschner I, Mrowietz U, et al. Differential expression of antimicrobial peptides in psoriasis and psoriatic arthritis as a novel contributory mechanism for skin and joint disease heterogeneity. *Scand J Rheumatol.* (2016) 45:188–96. doi: 10.3109/03009742.2015.1091497
59. Gilliet M, Cao W, Liu YJ. Plasmacytoid dendritic cells: sensing nucleic acids in viral infection and autoimmune diseases. *Nat Rev Immunol.* (2008) 8:594–606. doi: 10.1038/nri2358
60. Frasca L, Palazzo R, Chimenti MS, Alivernini S, Tulosso B, Bui L, et al. Anti-LL37 antibodies are present in psoriatic arthritis (PsA) patients: new biomarkers in PsA. *Front Immunol.* (2018) 9:1–16. doi: 10.3389/fimmu.2018.01936
61. Lande R, Gregorio J, Facchinetti V, Chatterjee B, Wang YH, Homey B, et al. Plasmacytoid dendritic cells sense self-DNA coupled with antimicrobial peptide. *Nature.* (2007) 449:564–69. doi: 10.1038/nature06116
62. Cai Y, Shen X, Ding C, Qi C, Li K, Li X, et al. Pivotal role of dermal IL-17-producing $\gamma\delta$ T cells in skin inflammation. *Immunity.* (2011) 35:596–610. doi: 10.1016/j.immuni.2011.08.001
63. Nerviani A, Boutet MA, Tan WSG, Goldmann K, Prukayastha N, Atjos Lajtos T, et al. IL-23 skin and joint profiling in psoriatic arthritis: novel perspectives in understanding clinical responses to IL-23 inhibitors. *Ann Rheum Dis.* (2021) 80:591–7. doi: 10.1136/annrheumdis-2020-218186
64. Dai H, Adamopoulos IE. Psoriatic arthritis under the influence of IFN γ . *Clin Immunol.* (2020) 218:108513. doi: 10.1016/j.clim.2020.108513
65. Diani M, Altomare G, Reali E. T cell responses in psoriasis and psoriatic arthritis. *Autoimmun Rev.* (2015) 14:286–92. doi: 10.1016/j.autrev.2014.11.012
66. Steel KJA, Srenathan U, Ridley M, Durham LE, Wu SH, Ryan SE, et al. Polyfunctional, proinflammatory, tissue-resident memory phenotype and function of synovial interleukin-17A+CD8+ T cells in psoriatic arthritis. *Arthritis Rheumatol.* (2020) 72:435–47. doi: 10.1002/art.41156
67. Tassiulas I, Duncan SR, Centola M, Theofilopoulos AN, Boumpas DT. Clonal characteristics of T cell infiltrates in skin and synovium of patients with psoriatic arthritis. *Hum Immunol.* (1999) 60:479–91. doi: 10.1016/S0198-8859(99)00034-8
68. Paulsen F, Pufe T, Conradi L, Varoga D, Tsokos M, Papendieck J, et al. Antimicrobial peptides are expressed and produced in healthy and inflamed human synovial membranes. *J Pathol.* (2002) 198:369–77. doi: 10.1002/path.1224
69. Leipe J, Grunke M, Dechant C, Reindl C, Kerzendorf U, Schulze-Koops H, et al. Role of Th17 cells in human autoimmune arthritis. *Arthritis Rheum.* (2010) 62:2876–85. doi: 10.1002/art.27622
70. Bowes J, Budu-Aggrey A, Huffmeier U, Uebe S, Steel K, Hebert HL, et al. Dense genotyping of immune-related susceptibility loci reveals new insights into the genetics of psoriatic arthritis. *Nat Commun.* (2015) 6:1–10. doi: 10.1038/ncomms8741
71. Penkava F, Velasco-Herrera MDC, Young MD, Yager N, Nwosu LN, Pratt AG, et al. Single-cell sequencing reveals clonal expansions of pro-inflammatory synovial CD8 T cells expressing tissue-homing receptors in psoriatic arthritis. *Nat Commun.* (2020) 11. doi: 10.1038/s41467-020-18513-6
72. Diani M, Casciano F, Marongiu L, Longhi M, Altomare A, Pigatoo P, et al. Increased frequency of activated CD8+ T cell effectors in patients with psoriatic arthritis. *Sci Rep.* (2019) 9:10870. doi: 10.1038/s41598-019-47310-5
73. Candia L, Marquez J, Hernandez C, Zea AH, Espinoza LR. Toll-like receptor-2 expression is upregulated in antigen-presenting cells from patients with psoriatic arthritis: a pathogenic role for innate immunity? *J Rheumatol.* (2007) 34:374–9.

74. Boutet MA, Nerviani A, Gallo Afflitto G, Pitzalis C. Role of the IL-23/IL-17 axis in psoriasis and psoriatic arthritis: the clinical importance of its divergence in skin and joints. *Int J Mol Sci.* (2018) 19:530. doi: 10.3390/ijms19020530
75. Armas-González E, Díaz-Martín A, Domínguez-Luis MJ, Arce-Franco MT, Herrera-García A, Hernández-Hernández MV, et al. Differential antigen-presenting B cell phenotypes from synovial microenvironment of patients with rheumatoid and psoriatic arthritis. *J Rheumatol.* (2015) 42:1825–34. doi: 10.3899/jrheum.141577
76. Mackern-Oberti JP, Llanos C, Vega F, Salazar-Onfray F, Riedel CA, Bueno SM, et al. Role of dendritic cells in the initiation, progress and modulation of systemic autoimmune diseases. *Autoimmun Rev.* (2015) 14:127–39. doi: 10.1016/j.autrev.2014.10.010
77. Nestle FO, Conrad C, Tun-Kyi A, Homey B, Gombert M, Boyman O, et al. Plasmacytoid dendritic cells initiate psoriasis through interferon- α production. *J Exp Med.* (2005) 202:135–43. doi: 10.1084/jem.20050500
78. Wade SM, Canavan M, McGarry T, Low C, Wade SC, Mullan RH, et al. Association of synovial tissue polyfunctional T-cells with DAPSA in psoriatic arthritis. *Ann Rheum Dis.* (2018) 47:350–4. doi: 10.1136/annrheumdis-2018-214138
79. Gunderson AJ, Mohammed J, Horvath FJ, Podolsky MA, Anderson CR, Glick AB. CD8+ T cells mediate RAS-induced psoriasis-like skin inflammation through IFN- γ . *J Invest Dermatol.* (2013) 133:955–63. doi: 10.1038/jid.2012.390
80. Celis R, Planell N, Fernández-Sueiro JL, Sanmarti R, Ramirez J, Gonzalez-Alvaro I, et al. Synovial cytokine expression in psoriatic arthritis and associations with lymphoid neogenesis and clinical features. *Arthritis Res Ther.* (2012) 14:R93. doi: 10.1186/ar3817
81. Kavanaugh A, Ritchlin C, Rahman P, Puig L, Gottlieb AB, Li S, et al. Ustekinumab, an anti-IL-12/23 p40 monoclonal antibody, inhibits radiographic progression in patients with active psoriatic arthritis: Results of an integrated analysis of radiographic data from the phase 3, multicentre, randomised, double-blind, placebo-c. *Ann Rheum Dis.* (2014) 73:1000–6. doi: 10.1136/annrheumdis-2013-204741
82. Mease PJ, Rahman P, Gottlieb AB, Kollmeier AP, Hsia EC, Lu XL, et al. Guselkumab in biologic-naïve patients with active psoriatic arthritis (DISCOVER-2): a double-blind, randomised, placebo-controlled phase 3 trial. *Lancet.* (2020) 395:1126–36. doi: 10.1016/S0140-6736(20)30263-4
83. Deodhar A, Helliwell PS, Boehncke WH, Kollmeier AP, Hsia EC, Subramanian RA, et al. Guselkumab in patients with active psoriatic arthritis who were biologic-naïve or had previously received TNF α inhibitor treatment (DISCOVER-1): a double-blind, randomised, placebo-controlled phase 3 trial. *Lancet.* (2020) 395:1115–25. doi: 10.1016/S0140-6736(20)30265-8
84. McInnes IB, Mease PJ, Kirkham B, Kavanaugh A, Ritchlin CT, Rahman P, et al. Secukinumab, a human anti-interleukin-17A monoclonal antibody, in patients with psoriatic arthritis (FUTURE 2): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet.* (2015) 386:1137–46. doi: 10.1016/S0140-6736(15)61134-5
85. Leijten EFA, van Kempen TS, Boes M, Michels-van Amelsfort JMR, Hijnen D, Hartgring SAY, et al. Brief report: enrichment of activated group 3 innate lymphoid cells in psoriatic arthritis synovial fluid. *Arthritis Rheumatol.* (2015) 67:2673–78. doi: 10.1002/art.39261
86. Mease PJ, McInnes IB, Kirkham B, Kavanaugh A, Rahman P, Van der Heijde D, et al. Secukinumab inhibition of interleukin-17A in patients with psoriatic arthritis. *N Engl J Med.* (2015) 373:1329–39. doi: 10.1056/NEJMoa1412679
87. Nash P, Kirkham B, Okada M, Rahman P, Combe B, Burmester GR, et al. Ixekizumab for the treatment of patients with active psoriatic arthritis and an inadequate response to tumour necrosis factor inhibitors: results from the 24-week randomised, double-blind, placebo-controlled period of the SPIRIT-P2 phase 3 trial. *Lancet.* (2017) 389:2317–27. doi: 10.1136/annrheumdis-2017-eular.1576
88. van Mens LJJ, van de Sande MGH, Menegatti S, Chen S, Blijdroop ICJ, De Jong HM, et al. Brief report: interleukin-17 blockade with secukinumab in peripheral spondyloarthritis impacts synovial immunopathology without compromising systemic immune responses. *Arthritis Rheumatol.* (2018) 70:1994–2002. doi: 10.1002/art.40581
89. Pfeffer K. Biological functions of tumor necrosis factor cytokines and their receptors. *Cytokine Growth Factor Rev.* (2003) 14:185–91. doi: 10.1016/S1359-6101(03)00022-4
90. Ettehadhi P, Greaves MW, Wallach D, Aderka D, Camp RDR. Elevated tumour necrosis factor- α biological activity in psoriatic skin lesions. *Clin Exp Immunol.* (1994) 96:146–51. doi: 10.1111/j.1365-2249.1994.tb06244.x
91. Mori G, Cantatore FP, Brunetti G, Oranger A, Colaiani G, Quarta L, et al. Synovial fluid fibroblasts and lymphocytes support the osteoclastogenesis in human psoriatic arthritis. *Ann N Y Acad Sci.* (2007) 1117:159–64. doi: 10.1196/annals.1402.013
92. Zhao B, Grimes SN, Li S, Hu X, Ivashkiv LB. TNF-induced osteoclastogenesis and inflammatory bone resorption are inhibited by transcription factor RBP-J. *J Exp Med.* (2012) 209:319–34. doi: 10.1084/jem.20111566
93. Taams LS, Steel KJA, Srenathan U, Burns LA, Kirkham BW. IL-17 in the immunopathogenesis of spondyloarthritis. *Nat Rev Rheumatol.* (2018) 14:453–66. doi: 10.1038/s41584-018-0044-2
94. Mease PJ, Fleischmann R, Deodhar AA, Wollenhaupt J, Khraishi M, Kielar D, et al. Effect of certolizumab pegol on signs and symptoms in patients with psoriatic arthritis: 24-week results of a Phase 3 double-blind randomised placebo-controlled study (RAPID-PsA). *Ann Rheum Dis.* (2014) 73:48–55. doi: 10.1136/annrheumdis-2013-203696
95. Gladman DD, Mease PJ, Ritchlin CT, Choy EHS, Sharp JT, Ory PA, et al. Adalimumab for long-term treatment of psoriatic arthritis: forty-eight week data from the Adalimumab effectiveness in psoriatic arthritis trial. *Arthritis Rheum.* (2007) 56:476–88. doi: 10.1002/art.22379
96. Mease PJ, Kivitz AJ, Burch FX, Siegel EL, Cohen SB, Ory P, et al. Etanercept treatment of psoriatic arthritis: Safety, efficacy, and effect on disease progression. *Arthritis Rheum.* (2004) 50:2264–72. doi: 10.1002/art.20335
97. de Groot M, Teunissen MBM, Picavet DI, de Rie MA, Bos JD. Reduction of different inflammatory cell types of the innate immune system in psoriatic skin during etanercept treatment. *Exp Dermatol.* (2010) 19:754–6. doi: 10.1111/j.1600-0625.2010.01089.x
98. Goedkoop AY, Kraan MC, Teunissen MBM, Picavet DI, De Rie MA, Bos JD, et al. Early effects of tumour necrosis factor α blockade on skin and synovial tissue in patients with active psoriasis and psoriatic arthritis. *Ann Rheum Dis.* (2004) 63:769–73. doi: 10.1136/ard.2003.018085
99. Van Kuijk AWR, Gerlag DM, Vos K, Wolbink G, De Groot M, De Rie MA, et al. A prospective, randomised, placebo-controlled study to identify biomarkers associated with active treatment in psoriatic arthritis: effects of adalimumab treatment on synovial tissue. *Ann Rheum Dis.* (2009) 68:1303–9. doi: 10.1136/ard.2008.091389
100. McInnes IB, Illei GG, Danning CL, Yarboro CH, Crane M, Kuroiwa T, et al. IL-10 improves skin disease and modulates endothelial activation and leukocyte effector function in patients with psoriatic arthritis. *J Immunol.* (2001) 167:4075–82. doi: 10.4049/jimmunol.167.7.4075
101. Danning CL, Illei GG, Hitchon C, Greer MR, Boumpas DT, McInnes IB. Macrophage-derived cytokine and nuclear factor κ B p65 expression in synovial membrane and skin of patients with psoriatic arthritis. *Arthritis Rheum.* (2000) 43:1244–56. doi: 10.1002/1529-0131(200006)43:6<1244::AID-ANR7>3.0.CO;2-2
102. Benham H, Norris P, Goodall J, Wechalekar MD, FitzGerald O, Szentpetery A, et al. Th17 and Th22 cells in psoriatic arthritis and psoriasis. *Arthritis Res Ther.* (2013) 15:1. doi: 10.1186/ar4317
103. Mitra A, Raychaudhuri SK, Raychaudhuri SP. Functional role of IL-22 in psoriatic arthritis. *Arthritis Res Ther.* (2012) 14:R65. doi: 10.1186/ar3781
104. Burns LA, Maroof A, Marshall D, Steel K, Lalnunhlimi S, Cole S, et al. Presence, function, and regulation of IL-17F-expressing human CD4+ T cells. *Eur J Immunol.* (2020) 50:568–80. doi: 10.1002/eji.2019.48138
105. Boutet MA, Nerviani A, Lliso-Ribera G, Lucchesi D, Prediletto E, Ghiardi GM, et al. Interleukin-36 family dysregulation drives joint inflammation and therapy response in psoriatic arthritis. *Rheumatology.* (2020) 59:828–38. doi: 10.1093/rheumatology/kez358
106. Van Kuijk AWR, Reinders-Blankert P, Smeets TJM, Dijkmans BAC, Tak PP. Detailed analysis of the cell infiltrate and the expression of mediators of synovial inflammation and joint destruction in the synovium of patients

- with psoriatic arthritis: implications for treatment. *Ann Rheum Dis.* (2006) 65:1551–7. doi: 10.1136/ard.2005.050963
107. Frey S, Derer A, Messbacher ME, Baeten DLP, Bugatti S, Montecucco C, et al. The novel cytokine interleukin-36 α is expressed in psoriatic and rheumatoid arthritis synovium. *Ann Rheum Dis.* (2013) 72:1569–74. doi: 10.1136/annrheumdis-2012-202264
 108. Ciccia F, Guggino G, Ferrante A, Raimondo S, Bignone R, Rodolico V, et al. Interleukin-9 overexpression and Th9 polarization characterize the inflamed gut, the synovial tissue, and the peripheral blood of patients with psoriatic arthritis. *Arthritis Rheumatol.* (2016) 68:1922–31. doi: 10.1002/art.39649
 109. Eder L, Haddad A, Rosen CF, Lee KA, Chandran V, Cook R, et al. The incidence and risk factors for psoriatic arthritis in patients with psoriasis: a prospective cohort study. *Arthritis Rheumatol.* (2016) 68:915–23. doi: 10.1002/art.39494
 110. Faustini F, Simon D, Oliveira I, Kleyer A, Haschka J, Englbrecht M, et al. Subclinical joint inflammation in patients with psoriasis without concomitant psoriatic arthritis: a cross-sectional and longitudinal analysis. *Ann Rheum Dis.* (2016) 75:2068–74. doi: 10.1136/annrheumdis-2015-208821
 111. Eder L, Polachek A, Rosen CF, Chandran V, Cook R, Gladman DD. The development of psoriatic arthritis in patients with psoriasis is preceded by a period of nonspecific musculoskeletal symptoms: a prospective cohort study. *Arthritis Rheumatol.* (2017) 69:622–9. doi: 10.1002/art.39973
 112. Emad Y, Ragab Y, Bassyouni I, Moawayh O, Fawzy M, Saad A, et al. Enthesitis and related changes in the knees in seronegative spondyloarthropathies and skin psoriasis: magnetic resonance imaging case-control study. *J Rheumatol.* (2010) 37:1709–17. doi: 10.3899/jrheum.100068
 113. Savage L, Goodfield M, Horton L, Watad A, Hensor E, Emery P, et al. Regression of peripheral subclinical enthesopathy in therapy-naïve patients treated with ustekinumab for moderate-to-severe chronic plaque psoriasis: a fifty-two-week, prospective, open-label feasibility study. *Arthritis Rheumatol.* (2019) 71:626–31. doi: 10.1002/art.40778
 114. Simon D, Tascilar K, Kleyer A, Bayat S, Kampylafka E, Sokolova M, et al. Structural enthesal lesions in patients with psoriasis are associated with an increased risk of progression to psoriatic arthritis. *Arthritis Rheumatol.* (2020). doi: 10.1002/art.41239
 115. Kampylafka E, Simon D, D'Oliveira I, Linz C, Lerchen V, Englbrecht M, et al. Disease interception with interleukin-17 inhibition in high-risk psoriasis patients with subclinical joint inflammation - Data from the prospective IVEPSA study. *Arthritis Res Ther.* (2019) 21:1–9. doi: 10.1186/s13075-019-1957-0
 116. Karouzakis E, Hähnlein J, Grasso C, Semmelink JF, Tak PP, Gerlag DM, et al. Molecular characterization of human lymph node stromal cells during the earliest phases of rheumatoid arthritis. *Front Immunol.* (2019) 10:1–11. doi: 10.3389/fimmu.2019.01863
 117. Ramwadhoebe TH, Hähnlein J, Maijer KI, Van Boven LJ, Gerlag DM, Tak PP, et al. Lymph node biopsy analysis reveals an altered immunoregulatory balance already during the at-risk phase of autoantibody positive rheumatoid arthritis. *Eur J Immunol.* (2016) 46:2812–21. doi: 10.1002/eji.201646393
 118. Ramwadhoebe TH, Hähnlein J, Van Kuijk BJ, Choi IY, Van Boven LJ, Gerlag DM, et al. Human lymph-node CD8+ T cells display an altered phenotype during systemic autoimmunity. *Clin Transl Immunol.* (2016) 5:e67. doi: 10.1038/cti.2016.8
 119. Van Baarsen LGM, De Hair MJH, Ramwadhoebe TH, Zijlstra IJAJ, Maas M, Gerlag DM, et al. The cellular composition of lymph nodes in the earliest phase of inflammatory arthritis. *Ann Rheum Dis.* (2013) 72:1420–4. doi: 10.1136/annrheumdis-2012-202990
 120. Ramwadhoebe TH, Van Baarsen LGM, Boumans MJH, Bruijnen STG, Safy M, Berger FH, et al. Effect of rituximab treatment on T and B cell subsets in lymph node biopsies of patients with rheumatoid arthritis. *Rheumatology.* (2019) 58:1075–85. doi: 10.1093/rheumatology/key428
 121. Sakkas LI, Bogdanos DP. Are psoriasis and psoriatic arthritis the same disease? The IL-23/IL-17 axis data. *Autoimmun Rev.* (2017) 16:10–5. doi: 10.1016/j.autrev.2016.09.015
 122. de Hair MJH, de Sande MGH van, Maas M, Gerlag DM, Tak PP. Experience of patients undergoing mini-arthroscopy compared to MRI in the earliest phases of arthritis. *Int J Clin Med.* (2013) 4:1–5. doi: 10.4236/ijcm.2013.412A2001
 123. Van De Sande MGH, De Launay D, De Hair MJH, Garcia S, Van de Sande GPM, Wijbrandts CA, et al. Local synovial engagement of angiogenic TIE-2 is associated with the development of persistent erosive rheumatoid arthritis in patients with early arthritis. *Arthritis Rheum.* (2013) 65:3073–83. doi: 10.1002/art.38128
 124. Van De Sande MGH, Thurlings RM, Boumans MJH, Wijbrandts CA, Grazia Modesti M, Gerlag DM, et al. Presence of lymphocyte aggregates in the synovium of patients with early arthritis in relationship to diagnosis and outcome: is it a constant feature over time? *Ann Rheum Dis.* (2011) 70:700–3. doi: 10.1136/ard.2010.139287
 125. Lindberg J, Wijbrandts CA, van Baarsen LG, Nader G, Klareskog L, Catrina A, et al. The gene expression profile in the synovium as a predictor of the clinical response to infliximab treatment in rheumatoid arthritis. *PLoS ONE.* (2010) 5:11310. doi: 10.1371/journal.pone.0011310
 126. Van Baarsen EGM, De Jong TA, De Hair MJH, Semmelink JF, Choi IY, Gerlag DM, et al. Synovial gene signatures associated with the development of rheumatoid arthritis in at risk individuals: a prospective study. *bioRxiv Prepr.* doi: 10.1101/2021.04.27.440770
 127. De Hair MJH, Van De Sande MGH, Ramwadhoebe TH, Hansson M, Landewe R, Van Der Leij C, et al. Features of the synovium of individuals at risk of developing rheumatoid arthritis : Implications for understanding preclinical rheumatoid arthritis. *Arthritis Rheumatol.* (2014) 66:513–22. doi: 10.1002/art.38273
 128. Van De Sande MGH, De Hair MJH, Van Der Leij C, Klarenbeek PL, Bos WH, SMith MD, et al. Different stages of rheumatoid arthritis: features of the synovium in the preclinical phase. *Ann Rheum Dis.* (2011) 70:772–7. doi: 10.1136/ard.2010.139527

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Bolt, van Ansenwoude, Hammoura, van de Sande and van Baarsen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.