



Emerging Molecular Targets for the Treatment of Refractory Sarcoidosis

Gonçalo Boleto^{1,2}, Matheus Vieira^{1,2}, Anne Claire Desbois^{1,2,3,4,5}, David Saadoun^{1,2,3,4,5} and Patrice Cacoub^{1,2,3,4,5*}

¹ AP-HP, Groupe Hospitalier Pitié-Salpêtrière, Department of Internal Medicine and Clinical Immunology, Paris, France,

² Centre de Référence des Maladies Auto-Immunes et Systémiques Rares, Centre de Référence des Maladies

Auto-Inflammatoires et de l'Amylose, Bordeaux, France, ³ Sorbonne Université, UPMC Univ Paris 06, UMR 7211,

Inflammation-Immunopathology-Biotherapy Department (DHU i2B), Paris, France, ⁴ INSERM, UMR_S 959, Paris, France,

⁵ CNRS, FRE3632, Paris, France

Sarcoidosis is a multisystem granulomatous disease of unknown origin that has variable clinical course and can affect nearly any organ. It has a chronic course in about 25% of patients. Corticosteroids (CS) are the cornerstone of therapy but their long-term use is associated with cumulative toxicity. Commonly used CS-sparing agents include methotrexate, cyclophosphamide, azathioprine, and mycophenolate mofetil. Twenty to forty percentage of sarcoidosis patients are refractory to these therapies or develop severe adverse events. Therefore, additional and targeted CS-sparing agents are needed for chronic sarcoidosis. Macrophage activation, interferon response, and formation of the granuloma are mainly mediated by T helper-1 responses. Different pro-inflammatory cytokines such as interleukin (IL)-8, IL-12, IL-6, and tumor necrosis factor-alpha (TNF- α) have been shown to be highly expressed in sarcoidosis-affected tissues. As a result of increased production of these cytokines, Janus kinase-signal transducer and activator of transcription (JAK-STAT) signaling is constitutively active in sarcoidosis. Several studies of biological agents that target TNF- α have reported their efficacy and appear today as a second line option in refractory sarcoidosis. Some case series report a positive effect of tocilizumab an anti-IL-6 monoclonal antibody in this setting. More recently, JAK inhibition appears as a new promising strategy. This review highlights key advances on the management of chronic refractory sarcoidosis. Novel therapeutic strategies and treatment agents to manage the disease are described.

Keywords: sarcoidosis, therapy, JAK inhibitors, interleukin-1, interleukin-6, granuloma

INTRODUCTION

Sarcoidosis is a multisystem granulomatous disease that can involve virtually any organ though the lungs and the lymphatics are the most commonly affected sites (1). The disease may remit spontaneously or upon treatment usually within the first 2–3 years after diagnosis but can have a chronic course in about 25% of cases (2). The exact cause of sarcoidosis is still not known. However, genetic susceptibility and environmental factors have been suggested as contributors of disease development (3, 4). Several studies strongly suggest that sarcoidosis might be the result of an exaggerated granulomatous reaction to a microbial-induced host response and persistent presence of antigens causing sarcoid lesions (5). Systematic treatment

OPEN ACCESS

Edited by:

Mehdi Mirsaedi,
University of Miami, United States

Reviewed by:

Kunihiro Yamaoka,
Keio University, Japan
Luis Ferreira Moita,
Gulbenkian Institute of Science
(IGC), Portugal

*Correspondence:

Patrice Cacoub
patrice.cacoub@aphp.fr

Specialty section:

This article was submitted to
Rheumatology,
a section of the journal
Frontiers in Medicine

Received: 12 August 2020

Accepted: 02 November 2020

Published: 24 November 2020

Citation:

Boleto G, Vieira M, Desbois AC,
Saadoun D and Cacoub P (2020)
Emerging Molecular Targets for the
Treatment of Refractory Sarcoidosis.
Front. Med. 7:594133.
doi: 10.3389/fmed.2020.594133

is often reserved for life-threatening organ involvement [i.e., severe interstitial lung disease, central nervous system (CNS), kidneys, liver, heart] or severe disabling functional symptoms such as skin disease, arthritis and bone disease or posterior uveitis (1). Corticosteroids (CS) remain the mainstay of treatment; however, their long-term use is associated with cumulative toxicity. Alternative therapies commonly used as CS-sparing agents include hydroxychloroquine, methotrexate, azathioprine and mycophenolate mofetil (6). About 10% of patients are refractory to these first and second-line therapies or develop adverse events necessitating additional CS-sparing targeted agents (7). Despite limited data on extra pulmonary manifestations, cyclophosphamide (CYC) has been successfully used for refractory CNS and cardiac disease (8–10). However, the known toxic and carcinogenic profile of CYC limits its use especially in young patients. In this setting, biological agents that target the tumor necrosis factor (TNF) have been introduced as a third-line therapy and have proved to be effective in a proportion of patients with severe/refractory sarcoidosis (11). Despite their excellent safety profile in other rheumatic conditions, TNF- α antagonists use showed to be less well tolerated with severe infections and malignancies being more frequent during sarcoidosis treatment (12). Hence, there is currently an unmet therapeutic need for a proportion of refractory or intolerant patients to TNF antagonists. Identifying new molecular targets for the treatment of refractory sarcoidosis should be a priority. Here, we review new therapeutic approaches for the treatment of refractory sarcoidosis with targeted biologic and synthetic agents.

A summary of the current data on the potential therapeutic targets in refractory sarcoidosis is available in **Table 1**. A focus on the reported cases of targeted biologic and synthetic agents (other than TNF inhibitors) use in sarcoidosis is available in **Supplementary Table 1**. **Figure 1** illustrates the pathophysiology and potential therapeutic targets of refractory sarcoidosis.

TUMOR NECROSIS FACTOR-ALPHA INHIBITION

TNF- α plays a crucial role in the development of noncaseating granulomas in a variety of diseases. Previous studies showed that high levels of TNF- α from alveolar macrophages correlated with disease progression (13). Two randomized controlled trials have investigated infliximab, a chimeric monoclonal anti-TNF- α antibody, therapy in sarcoidosis and showed significant though modest improvement in lung function after 14 weeks of treatment (14, 15). Another randomized controlled trial showed significant improvement in extrapulmonary sarcoidosis as assessed by a the extrapulmonary physician organ severity tool (ePOST) in patients treated with infliximab (16). A large French nationwide multicentric retrospective study on 132 patients with refractory sarcoidosis showed TNF-antagonists to be efficient in about two-thirds of patients, despite higher rates of adverse events (11). Adalimumab, another monoclonal anti-TNF- α antibody, showed to be efficient in a randomized controlled clinical trial of 16 patients with skin sarcoidosis (17). These positive effects of TNF inhibition are supported by a high

relapse rate after discontinuation of infliximab therapy in 47 patients with severe sarcoidosis (18). Despite the lack of high quality evidence, TNF inhibition can currently be considered as standard-of-care in severe cases of refractory sarcoidosis.

INTERLEUKIN-6 BLOCKING

A subset of CD4⁺ effector T cell population called T helper 17 (Th17), which express a transcription factor known as retinoic acid-related orphan receptor (ROR) γ γ , has been described in sarcoid lesions (19). Active sarcoidosis patients have an increased Th17/T regulator (Treg) ratio in the peripheral blood and bronchoalveolar lavage fluid (BALF) that is reversed by immunosuppressive therapy (20). Interleukin-6 (IL-6) is a key pleiotropic cytokine. It induces the development of Th17 cells from naïve T cells together with transforming growth factor β (TGF- β) and it inhibits anti-inflammatory Treg cells (21). Previous reports showed increased IL-6 levels in the BALF of sarcoidosis patients. Significant correlations were found between IL-6 levels and CD4⁺/CD8⁺ ratio, IL-8 and BALF neutrophil percentage (22–24). Another study reported increased IL-6 levels in the cerebrospinal fluid (CSF) of patients with neurosarcoidosis as compared to patients with multiple sclerosis or other inflammatory disorders. CSF concentration of IL-6 > 50 pg/mL was associated with a higher risk of relapse or progression of neurosarcoidosis (25). Genetic variations in the genes encoding IL-6 were preferentially upregulated in patients with severe and progressive sarcoidosis thus giving further evidence of the pathogenic role of this proinflammatory cytokine (26, 27). IL-6 is a potent up regulator of serum amyloid A protein (SAA), an acute phase reactant which has demonstrated a potential key role in the pathogenesis of sarcoidosis (28).

Currently, there are two humanized IL-6 receptor monoclonal antibodies [tocilizumab (TCZ) and sarilumab (SAR)] that have been approved for the treatment of different inflammatory rheumatic conditions. Despite robust evidence on the role of IL-6 in the pathogenesis of sarcoidosis, data on the use of anti-IL-6 agents in sarcoidosis are scarce. Previous case reports showed clinical improvement under TCZ in patients with chronic sarcoidosis associated with Castleman's disease (29) and Still's disease (30). In a case series of four patients with refractory chronic sarcoidosis the authors reported dramatic responses to TCZ with improved symptoms and organ function allowing steroid tapering (31). In a model of experimental uveitis, Yoshimura et al. investigated the role of IL-6 in the formation of refractory ocular inflammation (32). The authors showed that IL-6-deficient mice had reduced Th17 responses and ameliorated ocular inflammation. Using the same model of experimental uveitis, systemic administration of an anti-IL-6 receptor antibody also ameliorated ocular inflammation by suppressing Th17 responses. TCZ has shown a positive signal in the management of patients with ocular involvement (refractory uveitis, cystoid macular oedema) refractory to conventional immunosuppressive drugs (33, 34). Of note, despite these encouraging data, cases of paradoxical new-onset sarcoidosis during TCZ therapy must also be acknowledge. To our knowledge, four reports [three patients

TABLE 1 | Current data on the potential therapeutic targets in refractory sarcoidosis.

Target	Agents	Evidence	Primary endpoint	Ongoing clinical trials
IL-6	Tocilizumab Sarilumab	Case-reports		Phase II (Sarilumab) (NCT04008069)
IL-1	Anakinra Canakinumab	Basic studies		Phase II (Canakinumab) for pulmonary sarcoidosis (NCT02888080) Phase II (Anakinra) for cardiac sarcoidosis (NCT04017936)
IL-17	Secukinumab Ixekizumab Brodalumab	Case-reports Phase II RCT (Secukinumab for non-infectious uveitis)	NA (only 4 patients)	None
IL-12/23	Ustekinumab (IL-12/23) Guselkumab (IL-23) Risankizumab (IL-23)	Phase II RCT (Ustekinumab for pulmonary and skin sarcoidosis)	Negative	None
CTLA4	Abatacept	Basic studies		STAR trial (NCT00739960) (the study has been terminated prematurely due to funding constraints)
B-cell	Rituximab Belimumab	Case-reports Open-label phase I/II trial (Rituximab for pulmonary sarcoidosis)	Negative	None
JAK	Ruxolitinib Tofacitinib Baracitinib Upadacitinib Filgotinib	Case-reports		Open-label trial (Tofacitinib) for pulmonary sarcoidosis (NCT03793439) Open-label trial (Tofacitinib) for cutaneous sarcoidosis (NCT03910543)
PDEA4	Apremilast	Open-label trial (cutaneous sarcoidosis)	Positive	Phase II/III open-label trial for cutaneous sarcoidosis (NCT00794274)

IL, interleukin; RCT, randomized-clinical trial; NA, not evaluated; CTLA4, cytotoxic T-lymphocyte antigen 4; JAK, janus kinase; PDEA4, phosphodiesterase type 4.

with rheumatoid arthritis (RA) (35–37) and one with giant cell arteritis (38)] have described cases of cutaneous and mediastinal sarcoidosis triggered by TCZ therapy. The exact mechanism for these paradoxical reactions is not known, but data on murine models of granulomatosis suggest that anti-IL-6 treatment might enhance TNF- α production contributing to granuloma formation (39, 40). Currently there is an ongoing clinical trial comparing the effectiveness and the safety of SAR in patients with glucocorticoid-dependent sarcoidosis (NCT04008069).

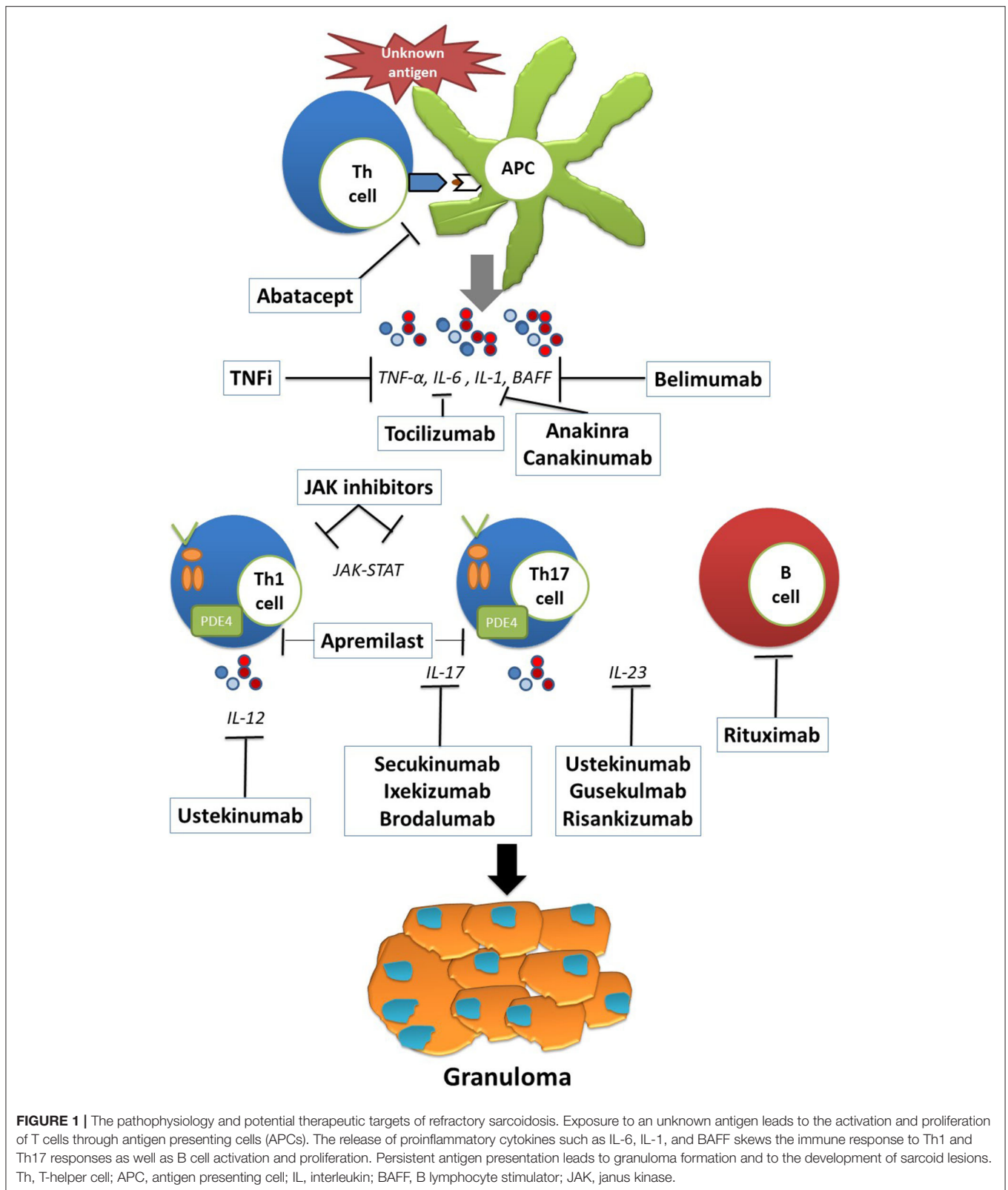
INTERLEUKIN-1 BLOCKING

Interleukin-1 (IL-1) is a cytokine with potent pro-inflammatory properties that has been shown to be implicated in the pathogenesis of sarcoidosis (41). An imbalance between the levels of IL-1 β and IL-1 receptor antagonist has been observed in the BALF of patients with pulmonary sarcoidosis (42, 43). Anakinra is a recombinant human IL-1 receptor antagonist that was firstly approved for the treatment of RA. It is currently used mostly for the management autoinflammatory conditions and difficult-to-treat gout (44, 45). We were unable to identify case reports or trials of anakinra use in the treatment of sarcoidosis. However, two cases of anakinra-induced sarcoidosis have been reported (46, 47). A phase 2 randomized controlled trial assessing the efficacy and safety of canakinumab, another IL-1 antagonist, in patients with pulmonary sarcoidosis (NCT02888080) has completed the recruitment process. The Interleukin-1 Blockade for Treatment of Cardiac Sarcoidosis (MAGiC-ART) trial (NCT04017936) is another ongoing phase

2 randomized-controlled trial evaluating anakinra for the treatment of cardiac sarcoidosis.

INTERLEUKIN-17 BLOCKING

Recent investigations indicate that Th17 cells are key players in all stages of granuloma formation and are upregulated in patients with active sarcoidosis (20, 48). The hallmark of the Th17 pathway is the production of IL-17. This proinflammatory cytokine with pleiotropic properties has been shown to be involved in the pathogenesis of various granulomatous diseases (49). Ten Berge et al. showed enhanced IL-17 expression in granulomas as well as increased numbers of IL-17 memory Th cells in the circulation and BALF of newly diagnosed sarcoidosis patients (50). Ostadkarampour et al. demonstrated the presence of T cells producing IL-17 in response to a mycobacterial antigen in patients with pulmonary sarcoidosis. The authors also observed higher levels of IL-17 and IL-17 producing cells in patients with Löfgren's syndrome suggesting a potential biomarker for the prognosis of sarcoidosis (51). Increased IL-17 responses might be related to aberrant metabolic pathways as shown by the abundant expression of hypoxia inducible factor (HIF) isoforms in granulomas and their association with Glut1 protein levels and enhanced IL-17 production. Downregulation of HIF in sarcoidosis peripheral blood mononuclear cells lead to a decrease in IL-17 production (52). An elevated expression of IL-17 receptor C on CD8⁺ T cells in peripheral blood was found in patients with ocular sarcoidosis (53).



Currently, three human monoclonal antibodies that target IL-17 have been approved for the treatment of psoriasis, psoriatic arthritis and axial spondyloarthritis (secukinumab, ixekizumab

and brodalumab). In Crohn's disease, which is another disorder characterized by granuloma formation, IL-17 inhibition can exacerbate the inflammatory disease. In sarcoidosis patients, the

role of IL-17 inhibition is less clear (54). Most of the available evidence on IL-17 blockade in patients with sarcoidosis comes from case reports. Previous reports suggest a favorable effect of secukinumab in two cases of sarcoidosis associated with TNF antagonists (55, 56). Conversely, our review of the literature identified two cases of sarcoidosis that either developed or worsened in patients taking secukinumab and ixekizumab (57, 58). However, a previous multicentre, randomized, double blind, phase II trial, assessing the efficacy and safety of secukinumab in non-infectious uveitis did not identify any safety concerns in the included patients with sarcoidosis ($n = 4$) (59). Further trials assessing the efficacy and safety of IL-17 blockade in sarcoidosis are warranted.

INTERLEUKIN-12 AND INTERLEUKIN-23 BLOCKING

Sarcoid granuloma formation is characterized by an influx of Th1 cells which spontaneously express IL-2 receptors and release interferon γ (IFN- γ), TNF- α , and IL-2 (5). IL-12 through its receptor (IL-12R) expressed on Th1 cells is one of the most important cytokines for inducing T cell response toward Th1 differentiation (60). Multiple studies have reported overexpression of IL-12 by activated alveolar macrophages in patients with active pulmonary sarcoidosis (61–65). IL-23 is a member of the IL-12 cytokine family which promotes Th17 responses through its receptor (IL-23R) (66). Transcriptomic analyses demonstrated upregulation of IL-23 in sarcoid lesions (67). IL-23R polymorphisms have also been shown to be implicated in the pathogenesis of sarcoidosis (68).

Ustekinumab is a monoclonal antibody that binds to the shared p40 unity of human IL-12 and IL-23, thus blocking Th1 and Th17 responses, respectively. It is approved for the treatment of psoriasis, psoriatic arthritis, Crohn's disease and ulcerative colitis (69). In a phase II, multicentre, randomized, double-blind, placebo-controlled trial (70), Judson et al. evaluated the safety and efficacy of ustekinumab in patients with chronic pulmonary and/or cutaneous sarcoidosis (70). Patients received ustekinumab 180 mg subcutaneously at week 0 and 90 mg at week 8, 16, and 24. Despite an excellent safety profile, ustekinumab failed to achieve the primary efficacy endpoint [change from baseline at week 16 in % predicted forced vital capacity (FVC%)] and there were no significant improvements in the major secondary endpoints. Guselkumab and risankizumab are specific anti-IL-23 monoclonal antibodies recently approved for the treatment of moderate-to-severe psoriasis. However, no data is currently available in patients with sarcoidosis. Similarly to IL-17 antagonists, several cases of induced or worsened sarcoidosis with ustekinumab or guselkumab have been reported in the literature (71–74).

CYTOTOXIC T-LYMPHOCYTE ANTIGEN 4 BLOCKADE

Cytotoxic T-lymphocyte antigen 4 (CTLA-4) is a costimulatory molecule that is an important regulator of T cell activation and proliferation. CTLA-4 polymorphisms were shown to

significantly influence phenotypes of sarcoidosis (75). Abatacept is a fusion protein of the extracellular domain of the CTLA-4 linked to a modified Fc of human immunoglobulin 1 (IgG1) inhibiting the activation of T cell responses (76). It is currently approved for the treatment of RA. To the best of our knowledge, there are no published studies or case reports concerning abatacept in sarcoidosis. We identified a prospective open-label trial (STAR trial NCT00739960) evaluating abatacept in refractory sarcoidosis. Unfortunately, this study has been terminated prematurely due to funding constraints.

B-CELL INHIBITION

Innate and T cell immunity play major roles in the pathogenesis of sarcoidosis. Several studies suggest a potential involvement of B cell immune responses in this disease (5, 77). Hypergammaglobulinemia and B cell accumulation in sarcoid lesions are frequently observed (78, 79). Saussine et al. showed increased levels of B-cell-activating factor (BAFF), also called BlyS (B lymphocyte stimulator), which is a cytokine involved in the survival and maturation of B cells in patients with active sarcoidosis (80). Rituximab (RTX) is an anti-CD20 monoclonal chimeric antibody that selectively depletes CD20⁺ B cell population. RTX was first approved for the treatment of non-Hodgkin B-cell lymphoma and later for rheumatoid arthritis and anti-neutrophil cytoplasmic antibody (ANCA) associated vasculitis. Despite several case reports of the effectiveness of RTX in patients with refractory sarcoidosis (81–90) data from clinical trials are scarce. In a prospective, open-label phase I/II trial, the authors assessed the effect of RTX in 10 patients with symptomatic moderate-to-severe pulmonary sarcoidosis refractory to corticosteroids plus one or more corticosteroid-sparing agents, including methotrexate and azathioprine (91). The authors observed very modest and inconsistent improvements in FVC% and 6-min walk test (6MWD) with only two patients having >10% improvement of FCV% and three patients having >50 m improvement in 6MWD by week 52. The prednisone doses were not changed during the study. RTX was generally well tolerated with only one patient hospitalized for pneumonia. Two patients died of respiratory failure linked to sarcoidosis progression. Belimumab, a human monoclonal antibody that inhibits BAFF/BlyS, is approved for the treatment of systemic lupus erythematosus and may represent a potential drug candidate for the management of refractory sarcoidosis. However, our literature review did not identify reports or ongoing trials concerning belimumab in sarcoidosis.

JANUS KINASE INHIBITION

The activation of macrophages in sarcoid lesions is thought to be driven by Th1 immune responses and mediated by several cytokines including interferon- γ (IFN- γ) (92, 93). IFN- γ activates the Janus Kinase (JAK)-signal transducer and activator of transcription (STAT) signaling pathway which is involved in upregulating a set of genes involved in the inflammatory response (94). Transcriptomic analysis showed that JAK-STAT pathway

activation signatures, especially STAT1 pathway, is activated in granulomatous diseases (95–98).

There are currently four JAK inhibitors available for clinical use: ruxolitinib for the treatment of myeloproliferative disorders; tofacitinib for rheumatoid arthritis (RA), psoriatic arthritis and ulcerative colitis; and baricitinib, upadacitinib and filgotinib for RA. Our literature review identified 7 reports concerning JAK inhibitors use (ruxolitinib $n = 3$; tofacitinib $n = 3$; baricitinib $n = 1$) in cases of refractory sarcoidosis. Ruxolitinib, a JAK 1 and 2 inhibitor, improved multivisceral involvement in three patients with sarcoidosis among whom two were treated for concomitant polycythemia vera (99–101). Damsky et al. reported dramatic positive effects of tofacitinib, a JAK 1 and 3 inhibitor, in a patient with severe cutaneous sarcoidosis refractory to topical and systemic corticosteroids and multiple corticosteroid-sparing agents including minocycline, hydroxychloroquine, methotrexate, tacrolimus and adalimumab. During treatment, the authors observed a downregulation of JAK/STAT signature on skin samples (102). The positive effect of tofacitinib was also observed in four patients with refractory cutaneous sarcoidosis ($n = 3$) and granuloma annular ($n = 1$) (103). Tofacitinib resulted in a mean improvement in clinical activity and histologic resolution was documented in all patients. The same group reported another case of severe multiorgan sarcoidosis involving the lungs, lymph nodes, bones, and skin, not controlled with prednisone, mycophenolate sodium, methotrexate, infliximab, rituximab and intravenous immunoglobulins (104). In this patient, tofacitinib treatment resulted in clinical remission of cutaneous lesions as well as resolution of positron emission tomography hypermetabolized lesions in internal organs after 6 months. Scheinberg et al. reported the case of a 35-year-old female patient with fever, arthralgia and hilar and cervical adenopathy revealing sarcoidosis initially treated with prednisone (105). Due to persistent daily low grade fever and arthralgia she was started on baricitinib, a selective JAK 1 and 2 inhibitor, with complete resolution of clinical symptoms and lymph node disease after week 12. Despite these encouraging data, the safety profile of JAK inhibitors in sarcoidosis remains to be confirmed. There are currently two ongoing open-label trials evaluating tofacitinib 5 mg twice daily as a corticosteroid-sparing agent in pulmonary sarcoidosis (NCT03793439) and in the treatment of cutaneous sarcoidosis and granuloma annular (NCT03910543).

PHOSPHODIESTERASE TYPE 4 INHIBITION

Inhibition of PDEA4 prevents cyclic AMP (cAMP) being hydrolysed to AMP resulting in increased intracellular levels

of cAMP and inhibiting the expression of proinflammatory cytokines (106). Apremilast is an orally administered phosphodiesterase type 4 (PDEA4) inhibitor approved for the treatment of psoriasis and psoriatic arthritis. Baughman et al. reported the effects of apremilast in 15 patients with active cutaneous sarcoidosis (107). The authors observed significant improvements in skin scores in 14 patients treated with apremilast 20 mg twice daily. Three patients developed worsening of cutaneous sarcoid lesions within 3 months after discontinuation of apremilast. A phase 2 and 3 open-label trial evaluating the efficacy and safety of apremilast in chronic cutaneous sarcoidosis (NCT00794274) has been completed; however the results are not yet available.

CONCLUSION

Refractory sarcoidosis, broadly defined as failure to attain clinical remission after appropriate treatment with corticosteroids and conventional immunosuppressant, is associated with increased morbidity and mortality. At present, TNF antagonists can be considered standard-of-care therapy in severe cases of refractory sarcoidosis. However, treatment options in patients with multidrug-refractory sarcoidosis failing initial anti-TNF therapy is a challenging issue in clinical practice. In the clinical practice, assessing adherence, ruling out differential diagnosis, checking anti-TNF blood concentrations and testing for anti-drug antibodies, should be done before classifying patients as refractory and intensifying treatment. In this setting, new potential useful agents including IL-6, IL-17, and JAK inhibitors with evidence consisting largely of observation or uncontrolled studies are the most promising ones. Data from ongoing prospective clinical trials should give further information on clinical effects of these agents in refractory sarcoidosis.

AUTHOR CONTRIBUTIONS

All authors were involved in drafting the article or revising it critically for important intellectual content, and approved the final version to be submitted for publication. PC had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmed.2020.594133/full#supplementary-material>

REFERENCES

- Valeyre D, Prasse A, Nunes H, Uzunhan Y, Brillet P-Y, Müller-Quernheim J. Sarcoidosis. *Lancet Lond Engl*. (2014) 383:1155–67. doi: 10.1016/S0140-6736(13)60680-7
- Pereira CAC, Dornfeld MC, Baughman R, Judson MA. Clinical phenotypes in sarcoidosis. *Curr Opin Pulm Med*. (2014) 20:496–502. doi: 10.1097/MCP.0000000000000077
- Newman LS, Rose CS, Bresnitz EA, Rossman MD, Barnard J, Frederick M, et al. A case control etiologic study of sarcoidosis: environmental and

- occupational risk factors. *Am J Respir Crit Care Med.* (2004) 170:1324–30. doi: 10.1164/rccm.200402-249OC
4. Iannuzzi MC. Genetics of sarcoidosis. *Semin Respir Crit Care Med.* (2007) 28:15–21. doi: 10.1055/s-2007-970330
 5. Zissel G, Prasse A, Müller-Quernheim J. Immunologic response of sarcoidosis. *Semin Respir Crit Care Med.* (2010) 31:390–403. doi: 10.1055/s-0030-1262208
 6. Baughman RP, Nunes H, Sweiss NJ, Lower EE. Established and experimental medical therapy of pulmonary sarcoidosis. *Eur Respir J.* (2013) 41:1424–38. doi: 10.1183/09031936.00060612
 7. Baughman RP, Culver DA, Judson MA. A concise review of pulmonary sarcoidosis. *Am J Respir Crit Care Med.* (2011) 183:573–81. doi: 10.1164/rccm.201006-0865CI
 8. Doty JD, Mazur JE, Judson MA. Treatment of corticosteroid-resistant neurosarcoidosis with a short-course cyclophosphamide regimen. *Chest.* (2003) 124:2023–6. doi: 10.1378/chest.124.5.2023
 9. Chapelon-Abrie C, Sene D, Saadoun D, Cluzel P, Vignaux O, Costedoat-Chalumeau N, et al. Cardiac sarcoidosis: diagnosis, therapeutic management and prognostic factors. *Arch Cardiovasc Dis.* (2017) 110:456–65. doi: 10.1016/j.acvd.2016.12.014
 10. Joubert B, Chapelon-Abrie C, Biard L, Saadoun D, Demeret S, Dormont D, et al. Association of prognostic factors and immunosuppressive treatment with long-term outcomes in neurosarcoidosis. *JAMA Neurol.* (2017) 74:1336–44. doi: 10.1001/jamaneurol.2017.2492
 11. Jamilloux Y, Cohen-Aubart F, Chapelon-Abrie C, Maucourt-Boulch D, Marquet A, Pérard L, et al. Efficacy and safety of tumor necrosis factor antagonists in refractory sarcoidosis: a multicenter study of 132 patients. *Semin Arthritis Rheum.* (2017) 47:288–94. doi: 10.1016/j.semarthrit.2017.03.005
 12. Maneiro JR, Salgado E, Gomez-Reino JJ, Carmona L, BIOBADASER Study Group. Efficacy and safety of TNF antagonists in sarcoidosis: data from the Spanish registry of biologics BIOBADASER and a systematic review. *Semin Arthritis Rheum.* (2012) 42:89–103. doi: 10.1016/j.semarthrit.2011.12.006
 13. Marques LJ, Zheng L, Poulakis N, Guzman J, Costabel U. Pentoxifylline inhibits TNF-alpha production from human alveolar macrophages. *Am J Respir Crit Care Med.* (1999) 159:508–11. doi: 10.1164/ajrccm.159.2.9804085
 14. Baughman RP, Drent M, Kavuru M, Judson MA, Costabel U, du Bois R, et al. Infliximab therapy in patients with chronic sarcoidosis and pulmonary involvement. *Am J Respir Crit Care Med.* (2006) 174:795–802. doi: 10.1164/rccm.200603-402OC
 15. Rossman MD, Newman LS, Baughman RP, Teirstein A, Weinberger SE, Miller W, et al. A double-blinded, randomized, placebo-controlled trial of infliximab in subjects with active pulmonary sarcoidosis. *Sarcoidosis Vasc Diffuse Lung Dis.* (2006) 23:201–8. Available online at: <https://pubmed.ncbi.nlm.nih.gov/18038919/>
 16. Judson MA, Baughman RP, Costabel U, Flavin S, Lo KH, Kavuru MS, et al. Efficacy of infliximab in extrapulmonary sarcoidosis: results from a randomised trial. *Eur Respir J.* (2008) 31:1189–96. doi: 10.1183/09031936.00051907
 17. Pariser RJ, Paul J, Hirano S, Torosky C, Smith M. A double-blind, randomized, placebo-controlled trial of adalimumab in the treatment of cutaneous sarcoidosis. *J Am Acad Dermatol.* (2013) 68:765–73. doi: 10.1016/j.jaad.2012.10.056
 18. Vorselaars ADM, Verwoerd A, van Moorsel CHM, Keijsers RGM, Rijkers GT, Grutters JC. Prediction of relapse after discontinuation of infliximab therapy in severe sarcoidosis. *Eur Respir J.* (2013) 43:602–9. doi: 10.1183/09031936.00055213
 19. Facco M, Cabrelle A, Teramo A, Olivieri V, Gnoato M, Teolato S, et al. Sarcoidosis is a Th1/Th17 multisystem disorder. *Thorax.* (2011) 66:144–50. doi: 10.1136/thx.2010.140319
 20. Huang H, Lu Z, Jiang C, Liu J, Wang Y, Xu Z. Imbalance between Th17 and regulatory T-Cells in sarcoidosis. *Int J Mol Sci.* (2013) 14:21463–73. doi: 10.3390/ijms141121463
 21. Kimura A, Kishimoto T. IL-6: regulator of Treg/Th17 balance. *Eur J Immunol.* (2010) 40:1830–5. doi: 10.1002/eji.201040391
 22. Sahashi K, Ina Y, Takada K, Sato T, Yamamoto M, Morishita M. Significance of interleukin 6 in patients with sarcoidosis. *Chest.* (1994) 106:156–60. doi: 10.1378/chest.106.1.156
 23. Girgis RE, Basha MA, Malirik M, Popovich J, Iannuzzi MC. Cytokines in the bronchoalveolar lavage fluid of patients with active pulmonary sarcoidosis. *Am J Respir Crit Care Med.* (1995) 152:71–5. doi: 10.1164/ajrccm.152.1.7599865
 24. Takizawa H, Satoh M, Okazaki H, Matsuzaki G, Suzuki N, Ishii A, et al. Increased IL-6 and IL-8 in bronchoalveolar lavage fluids (BALF) from patients with sarcoidosis: correlation with the clinical parameters. *Clin Exp Immunol.* (1997) 107:175–81. doi: 10.1046/j.1365-2249.1997.d01-905.x
 25. Chazal T, Costopoulos M, Maillart E, Fleury C, Psimaras D, Legendre P, et al. The cerebrospinal fluid CD4/CD8 ratio and interleukin-6 and–10 levels in neurosarcoidosis: a multicenter, pragmatic, comparative study. *Eur J Neurol.* (2019) 26:1274–80. doi: 10.1111/ene.13975
 26. Bihl MP, Laule-Kilian K, Bubendorf L, Rutherford RM, Baty F, Kehren J, et al. Progressive pulmonary sarcoidosis—a fibroproliferative process potentially triggered by EGR-1 and IL-6. *Sarcoidosis Vasc Diffuse Lung Dis.* (2006) 23:38–50. doi: 10.1183/09031936.00051907
 27. Grutters JC, Sato H, Pantelidis P, Ruven HJT, McGrath DS, Wells AU, et al. Analysis of IL6 and IL1A gene polymorphisms in UK and Dutch patients with sarcoidosis. *Sarcoidosis Vasc Diffuse Lung Dis.* (2003) 20:20–7.
 28. Chen ES, Song Z, Willett MH, Heine S, Yung RC, Liu MC, et al. Serum amyloid A regulates granulomatous inflammation in sarcoidosis through toll-like receptor-2. *Am J Respir Crit Care Med.* (2010) 181:360–73. doi: 10.1164/rccm.200905-0696OC
 29. Awano N, Inomata M, Kondoh K, Satake K, Kamiya H, Moriya A, et al. Mixed-type multicentric Castleman's disease developing during a 17-year follow-up of sarcoidosis. *Intern Med Tokyo Jpn.* (2012) 51:3061–6. doi: 10.2169/internalmedicine.51.8120
 30. Semiz H, Kobak S. Coexistence of sarcoidosis and adult onset Still disease. *Rheumatol Clin.* (2019) 15:e18–20. doi: 10.1016/j.reuma.2017.04.004
 31. Sharp M, Donnelly SC, Moller DR. Tocilizumab in sarcoidosis patients failing steroid sparing therapies and anti-TNF agents. *Respir Med X.* (2019) 1:100004. doi: 10.1016/j.yrmex.2019.100004
 32. Yoshimura T, Sonoda K-H, Ohguro N, Ohsugi Y, Ishibashi T, Cua DJ, et al. Involvement of Th17 cells and the effect of anti-IL-6 therapy in autoimmune uveitis. *Rheumatol Oxf Engl.* (2009) 48:347–54. doi: 10.1093/rheumatology/ken489
 33. Silpa-Archa S, Oray M, Preble JM, Foster CS. Outcome of tocilizumab treatment in refractory ocular inflammatory diseases. *Acta Ophthalmol.* (2016) 94:e400–6. doi: 10.1111/aos.13015
 34. Vegas-Revenga N, Calvo-Río V, Mesquida M, Adán A, Hernández MV, Beltrán E, et al. Anti-IL6-receptor tocilizumab in refractory and noninfectious uveitic cystoid macular edema: multicenter study of 25 patients. *Am J Ophthalmol.* (2019) 200:85–94. doi: 10.1016/j.ajo.2018.12.019
 35. Nutz A, Pernet C, Combe B, Cohen JD. Sarcoidosis induced by tocilizumab: a paradoxical event? *J Rheumatol.* (2013) 40:1773–4. doi: 10.3899/jrheum.130278
 36. Bustamente L, Buscot M, Marquette CH, Roux C. Sarcoidosis and tocilizumab: is there a link? *Clin Exp Rheumatol.* (2017) 35:716.
 37. Shono Y, Kamata M, Takeoka S, Ikawa T, Tateishi M, Fukaya S, et al. Cutaneous sarcoidosis in a patient with rheumatoid arthritis receiving tocilizumab. *J Dermatol.* (2018) 45:e217–8. doi: 10.1111/1346-8138.14268
 38. Del Giorno R, Iodice A, Mangas C, Gabutti L. New-onset cutaneous sarcoidosis under tocilizumab treatment for giant cell arteritis: a quasi-paradoxical adverse drug reaction. Case report and literature review. *Ther Adv Musculoskelet Dis.* (2019) 11:1759720X19841796. doi: 10.1177/1759720X19841796
 39. Yimin null, Kohanawa M. A regulatory effect of the balance between TNF-alpha and IL-6 in the granulomatous and inflammatory response to *Rhodococcus aurantiacus* infection in mice. *J Immunol.* (2006) 177:642–50. doi: 10.4049/jimmunol.177.1.642
 40. Yimin null, Kohanawa M, Minagawa T. Up-regulation of granulomatous inflammation in interleukin-6 knockout mice infected with *Rhodococcus aurantiacus*. *Immunology.* (2003) 110:501–6. doi: 10.1111/j.1365-2567.2003.01762.x
 41. Rolfe MW, Standiford TJ, Kunkel SL, Burdick MD, Gilbert AR, Lynch JP, et al. Interleukin-1 receptor antagonist expression in sarcoidosis. *Am Rev Respir Dis.* (1993) 148:1378–84. doi: 10.1164/ajrccm/148.5.1378

42. Mikuniya T, Nagai S, Shimoji T, Takeuchi M, Morita K, Mio T, et al. Quantitative evaluation of the IL-1 beta and IL-1 receptor antagonist obtained from BALF macrophages in patients with interstitial lung diseases. *Sarcoidosis Vasc Diffuse Lung Dis.* (1997) 14:39–45.
43. Mikuniya T, Nagai S, Takeuchi M, Mio T, Hoshino Y, Miki H, et al. Significance of the interleukin-1 receptor antagonist/interleukin-1 beta ratio as a prognostic factor in patients with pulmonary sarcoidosis. *Respir Int Rev Thorac Dis.* (2000) 67:389–96. doi: 10.1159/000029536
44. Cvetkovic RS, Keating G, Anakinra. *BioDrugs.* (2002) 16:303–11. doi: 10.2165/00063030-200216040-00005
45. Kalliolias GD, Liossis S-NC. The future of the IL-1 receptor antagonist anakinra: from rheumatoid arthritis to adult-onset Still's disease and systemic-onset juvenile idiopathic arthritis. *Expert Opin Investig Drugs.* (2008) 17:349–59. doi: 10.1517/13543784.17.3.349
46. Sacre K, Pasqualoni E, Descamps V, Choudat L, Debray M-P, Papo T. Sarcoid-like granulomatosis in a patient treated by interleukin-1 receptor antagonist for TNF-receptor-associated periodic syndrome. *Rheumatol Oxf Engl.* (2013) 52:1338–40. doi: 10.1093/rheumatology/kes377
47. Friedman BE, English JC. Drug-induced sarcoidosis in a patient treated with an interleukin-1 receptor antagonist for hidradenitis suppurativa. *JAAD Case Rep.* (2018) 4:543–5. doi: 10.1016/j.jidcr.2018.03.007
48. Crouser ED. Role of imbalance between Th17 and regulatory T-cells in sarcoidosis. *Curr Opin Pulm Med.* (2018) 24:521–6. doi: 10.1097/MCP.0000000000000498
49. Curtis MM, Way SS. Interleukin-17 in host defence against bacterial, mycobacterial and fungal pathogens. *Immunology.* (2009) 126:177–85. doi: 10.1111/j.1365-2567.2008.03017.x
50. Ten Berge B, Paats MS, Bergen IM, van den Blink B, Hoogsteden HC, Lambrecht BN, et al. Increased IL-17A expression in granulomas and in circulating memory T cells in sarcoidosis. *Rheumatol Oxf Engl.* (2012) 51:37–46. doi: 10.1093/rheumatology/ker316
51. Ostadkarampour M, Eklund A, Moller D, Glader P, Olgart Höglund C, Lindén A, et al. Higher levels of interleukin IL-17 and antigen-specific IL-17 responses in pulmonary sarcoidosis patients with Löfgren's syndrome. *Clin Exp Immunol.* (2014) 178:342–52. doi: 10.1111/cei.12403
52. Talreja J, Talwar H, Bauerfeld C, Grossman LI, Zhang K, Tranchida P, et al. HIF-1 α regulates IL-1 β and IL-17 in sarcoidosis. *eLife.* (2019) 8:e44519. doi: 10.7554/eLife.44519
53. Wu W, Jin M, Wang Y, Liu B, Shen D, Chen P, et al. Overexpression of IL-17RC associated with ocular sarcoidosis. *J Transl Med.* (2014) 12:152. doi: 10.1186/1479-5876-12-152
54. Orrell KA, Murphrey M, Kelm RC, Lee HH, Pease DR, Laumann AE, et al. Inflammatory bowel disease events after exposure to interleukin 17 inhibitors secukinumab and ixekizumab: postmarketing analysis from the RADAR (“research on adverse drug events and reports”) program. *J Am Acad Dermatol.* (2018) 79:777–8. doi: 10.1016/j.jaad.2018.06.024
55. Toussirot E, Bernard C, Bossert M. Safety of the use of anti-IL17A treatment in a patient with certolizumab-induced sarcoidosis. *Clin Exp Rheumatol.* (2019) 37:344–5.
56. Eichhoff G. Management with secukinumab of tumour necrosis factor inhibitor-induced pulmonary sarcoidosis-like reaction in a patient with psoriasis. *Clin Exp Dermatol.* (2020) 45:455–6. doi: 10.1111/ced.14101
57. Nyckowski T, Ceilleux R, Wilson J. Sarcoidosis developing during secukinumab therapy: case report. *SKIN J Cutan Med.* (2017) 1:95–9. doi: 10.25251/skin.1.2.7
58. Sambharia M, Magge T, Ramakrishna S. Worsening of pulmonary sarcoidosis induced by ixekizumab: a rare paradox. *Chest.* (2018) 154:898A. doi: 10.1016/j.chest.2018.08.808
59. Letko E, Yeh S, Foster CS, Pleyer U, Brigell M, Grosskreutz CL, et al. Efficacy and safety of intravenous secukinumab in noninfectious uveitis requiring steroid-sparing immunosuppressive therapy. *Ophthalmology.* (2015) 122:939–48. doi: 10.1016/j.ophtha.2014.12.033
60. Trinchieri G. Interleukin-12 and its role in the generation of TH1 cells. *Immunol Today.* (1993) 14:335–8. doi: 10.1016/0167-5699(93)90230-I
61. Mroz RM, Korniluk M, Stasiak-Barmuta A, Chyczewska E. Increased levels of interleukin-12 and interleukin-18 in bronchoalveolar lavage fluid of patients with pulmonary sarcoidosis. *J Physiol Pharmacol.* (2008) 59 (Suppl. 6):507–13.
62. Taha RA, Minshall EM, Olivenstein R, Ihaku D, Wallaert B, Tscopoulos A, et al. Increased expression of IL-12 receptor mRNA in active pulmonary tuberculosis and sarcoidosis. *Am J Respir Crit Care Med.* (1999) 160:1119–23. doi: 10.1164/ajrccm.160.4.9807120
63. Shigehara K, Shijubo N, Ohmichi M, Kon S, Shibuya Y, Takahashi R, et al. Enhanced mRNA expression of Th1 cytokines and IL-12 in active pulmonary sarcoidosis. *Sarcoidosis Vasc Diffuse Lung Dis.* (2000) 17:151–7.
64. Shigehara K, Shijubo N, Ohmichi M, Takahashi R, Kon S, Okamura H, et al. IL-12 and IL-18 are increased and stimulate IFN-gamma production in sarcoid lungs. *J Immunol.* (2001) 166:642–9. doi: 10.4049/jimmunol.166.1.642
65. Shigehara K, Shijubo N, Ohmichi M, Kamiguchi K, Takahashi R, Morita-Ichimura S, et al. Increased circulating interleukin-12 (IL-12) p40 in pulmonary sarcoidosis. *Clin Exp Immunol.* (2003) 132:152–7. doi: 10.1046/j.1365-2249.2003.02105.x
66. Parham C, Chirica M, Timans J, Vaisberg E, Travis M, Cheung J, et al. A receptor for the heterodimeric cytokine IL-23 is composed of IL-12Rbeta1 and a novel cytokine receptor subunit, IL-23R. *J Immunol.* (2002) 168:5699–708. doi: 10.4049/jimmunol.168.11.5699
67. Judson MA, Marchell RM, Mascelli M, Piantone A, Barnathan ES, Petty KJ, et al. Molecular profiling and gene expression analysis in cutaneous sarcoidosis: the role of interleukin-12, interleukin-23, and the T-helper 17 pathway. *J Am Acad Dermatol.* (2012) 66:901–10.e1–2. doi: 10.1016/j.jaad.2011.06.017
68. Kim HS, Choi D, Lim LL, Allada G, Smith JR, Austin CR, et al. Association of interleukin 23 receptor gene with sarcoidosis. *Dis Markers.* (2011) 31:17–24. doi: 10.1155/2011/185106
69. Sands BE, Sandborn WJ, Panaccione R, O'Brien CD, Zhang H, Johanns J, et al. Ustekinumab as induction and maintenance therapy for ulcerative colitis. *N Engl J Med.* (2019) 382:91. doi: 10.1056/NEJMoa1900750
70. Judson MA, Baughman RP, Costabel U, Drent M, Gibson KF, Raghu G, et al. Safety and efficacy of ustekinumab or golimumab in patients with chronic sarcoidosis. *Eur Respir J.* (2014) 44:1296–307. doi: 10.1183/09031936.00000914
71. Powell JB, Matthews P, Rattehalli R, Woodhead F, Perkins P, Powell G, et al. Acute systemic sarcoidosis complicating ustekinumab therapy for chronic plaque psoriasis. *Br J Dermatol.* (2015) 172:834–6. doi: 10.1111/bjd.13365
72. Gad MM, Bazarbashi N, Kaur M, Gupta A. Sarcoid-like phenomenon - ustekinumab induced granulomatous reaction mimicking diffuse metastatic disease: a case report and review of the literature. *J Med Case Rep.* (2019) 13:257. doi: 10.1186/s13256-019-2137-1
73. Kobak S, Semiz H. Ustekinumab-induced sarcoidosis in a patient with psoriatic arthritis. *Curr Drug Saf.* (2020) 15:163–6. doi: 10.2174/1574886315666200316113312
74. Thomas AS, Rosenbaum JT. Poor control of sarcoidosis-related panuveitis with an antibody to IL-23. *Ocul Immunol Inflamm.* (2020) 28:491–3. doi: 10.1080/09273948.2019.1569245
75. Hattori N, Niimi T, Sato S, Achiwa H, Maeda H, Oguri T, et al. Cytotoxic T-lymphocyte antigen 4 gene polymorphisms in sarcoidosis patients. *Sarcoidosis Vasc Diffuse Lung Dis.* (2005) 22:27–32.
76. Dubois EA, Cohen AF. Abatacept. *Br J Clin Pharmacol.* (2009) 68:480–1. doi: 10.1111/j.1365-2125.2009.03502.x
77. Iannuzzi MC, Rybicki BA, Teirstein AS. Sarcoidosis. *N Engl J Med.* (2007) 357:2153–65. doi: 10.1056/NEJMra071714
78. Lee N-S, Barber L, Akula SM, Sigounas G, Kataria YP, Arce S. Disturbed homeostasis and multiple signaling defects in the peripheral blood B-cell compartment of patients with severe chronic sarcoidosis. *Clin Vaccine Immunol CVI.* (2011) 18:1306–16. doi: 10.1128/01118-11
79. Fazel SB, Howie SE, Krajewski AS, Lamb D. B lymphocyte accumulations in human pulmonary sarcoidosis. *Thorax.* (1992) 47:964–7. doi: 10.1136/thx.47.11.964
80. Saussine A, Tazi A, Feuillet S, Rybojad M, Juillard C, Bergeron A, et al. Active chronic sarcoidosis is characterized by increased transitional blood B cells, increased IL-10-producing regulatory B cells and high BAFF levels. *PLoS ONE.* (2012) 7:e43588. doi: 10.1371/journal.pone.0043588
81. Beccastrini E, Vannozzi L, Bacherini D, Squatrito D, Emmi L. Successful treatment of ocular sarcoidosis with rituximab. *Ocul Immunol Inflamm.* (2013) 21:244–6. doi: 10.3109/09273948.2012.762982

82. Cinetto F, Compagno N, Scarpa R, Malipiero G, Agostini C. Rituximab in refractory sarcoidosis: a single centre experience. *Clin Mol Allergy*. (2015) 13:19. doi: 10.1186/s12948-015-0025-9
83. Gottenberg JE, Guillevin L, Lambotte O, Combe B, Allanore Y, Cantagrel A, et al. Tolerance and short term efficacy of rituximab in 43 patients with systemic autoimmune diseases. *Ann Rheum Dis*. (2005) 64:913–20. doi: 10.1136/ard.2004.029694
84. Zella S, Kneiphof J, Haghikia A, Gold R, Woitalla D, Thöne J. Successful therapy with rituximab in three patients with probable neurosarcoidosis. *Ther Adv Neurol Disord*. (2018) 11:1756286418805732. doi: 10.1177/1756286418805732
85. Bompreszi R, Pati S, Chansakul C, Vollmer T. A case of neurosarcoidosis successfully treated with rituximab. *Neurology*. (2010) 75:568–70. doi: 10.1212/WNL.0b013e3181ec7ff9
86. Sawaya R, Radwan W. Sarcoidosis associated with neuromyelitis optica. *J Clin Neurosci*. (2013) 20:1156–8. doi: 10.1016/j.jocn.2012.09.030
87. Lower EE, Baughman RP, Kaufman AH. Rituximab for refractory granulomatous eye disease. *Clin Ophthalmol*. (2012) 6:1613–8. doi: 10.2147/OPTH.S35521
88. Dalia T, Liu D, Fraga GR, Springer J. A rare case of sarcoidosis presenting with cutaneous medium-vessel granulomatous vasculitis treated with rituximab. *J Clin Rheumatol*. (2018) 5.
89. Earle B, Wolf DS, Ramsay ES. Novel use of rituximab in treatment of refractory neurosarcoidosis in an 11-year-old girl. *J Clin Rheumatol*. (2019) 25:e101–3. doi: 10.1097/RHU.0000000000000900
90. Krause ML, Cooper LT, Chareonthaitawee P, Amin S. Successful use of rituximab in refractory cardiac sarcoidosis. *Rheumatol Oxf Engl*. (2016) 55:189–91. doi: 10.1093/rheumatology/kev309
91. Sweiss NJ, Lower EE, Mirsaeidi M, Dudek S, Garcia JGN, Perkins D, et al. Rituximab in the treatment of refractory pulmonary sarcoidosis. *Eur Respir J*. (2014) 43:1525–8. doi: 10.1183/09031936.00224513
92. Broos CE, Hendriks RW, Kool M. T-cell immunology in sarcoidosis: disruption of a delicate balance between helper and regulatory T-cells. *Curr Opin Pulm Med*. (2016) 22:476–83. doi: 10.1097/MCP.0000000000000303
93. Ramstein J, Broos CE, Simpson LJ, Ansel KM, Sun SA, Ho ME, et al. IFN- γ -Producing T-Helper 17.1 cells are increased in sarcoidosis and are more prevalent than T-helper type 1 cells. *Am J Respir Crit Care Med*. (2016) 193:1281–91. doi: 10.1164/rccm.201507-1499OC
94. Rosenbaum JT, Pasadhika S, Crouser ED, Choi D, Harrington CA, Lewis JA, et al. Hypothesis: sarcoidosis is a STAT1-mediated disease. *Clin Immunol*. (2009) 132:174–83. doi: 10.1016/j.clim.2009.04.010
95. Zhou T, Casanova N, Pouladi N, Wang T, Lussier Y, Knox KS, et al. Identification of Jak-STAT signaling involvement in sarcoidosis severity via a novel microRNA-regulated peripheral blood mononuclear cell gene signature. *Sci Rep*. (2017) 7:4237. doi: 10.1038/s41598-017-04109-6
96. Zhou T, Zhang W, Sweiss NJ, Chen ES, Moller DR, Knox KS, et al. Peripheral blood gene expression as a novel genomic biomarker in complicated sarcoidosis. *PLoS ONE*. (2012) 7:e44818. doi: 10.1371/journal.pone.0044818
97. Li H, Zhao X, Wang J, Zong M, Yang H. Bioinformatics analysis of gene expression profile data to screen key genes involved in pulmonary sarcoidosis. *Gene*. (2017) 596:98–104. doi: 10.1016/j.gene.2016.09.037
98. Rosenbaum JT, Hesselund A, Phan I, Planck SR, Wilson DJ. The expression of STAT-1 and phosphorylated STAT-1 in conjunctival granulomas. *Ocul Immunol Inflamm*. (2010) 18:261–4. doi: 10.3109/09273941003797934
99. Rotenberg C, Besnard V, Brillet PY, Giraudier S, Nunes H, Valeyre D. Dramatic response of refractory sarcoidosis under ruxolitinib in a patient with associated JAK2-mutated polycythemia. *Eur Respir J*. (2018) 52:1801482. doi: 10.1183/13993003.01482-2018
100. Levraut M, Martis N, Viau P, Suarez F, Queyrel V. Refractory sarcoidosis-like systemic granulomatosis responding to ruxolitinib. *Ann Rheum Dis*. (2019) 78:1606–7. doi: 10.1136/annrheumdis-2019-215387
101. Wei JJ, Kallenbach LR, Kreider M, Leung TH, Rosenbach M. Resolution of cutaneous sarcoidosis after Janus kinase inhibitor therapy for concomitant polycythemia vera. *JAAD Case Rep*. (2019) 5:360–1. doi: 10.1016/j.jidcr.2019.02.006
102. Damsky W, Thakral D, Emeagwali N, Galan A, King B. Tofacitinib treatment and molecular analysis of cutaneous sarcoidosis. *N Engl J Med*. (2018) 379:2540–6. doi: 10.1056/NEJMoa1805958
103. Damsky W, Thakral D, McGeary MK, Leventhal J, Galan A, King B. Janus kinase inhibition induces disease remission in cutaneous sarcoidosis and granuloma annulare. *J Am Acad Dermatol*. (2020) 82:612–21. doi: 10.1016/j.jaad.2019.05.098
104. Damsky W, Young BD, Sloan B, Miller EJ, Obando JA, King B. Treatment of multiorgan sarcoidosis with tofacitinib. *ACR Open Rheumatol*. (2020) 2:106–9. doi: 10.1002/acr2.11112
105. Scheinberg M, Maluf F, Wagner J. Steroid-resistant sarcoidosis treated with baricitinib. *Ann Rheum Dis*. (2020) 79:1259–60. doi: 10.1136/annrheumdis-2020-217271
106. Keating GM. Apremilast: a review in psoriasis and psoriatic arthritis. *Drugs*. (2017) 77:459–72. doi: 10.1007/s40265-017-0709-1
107. Baughman RP, Judson MA, Ingledue R, Craft NL, Lower EE. Efficacy and safety of apremilast in chronic cutaneous sarcoidosis. *Arch Dermatol*. (2012) 148:262–4. doi: 10.1001/archdermatol.2011.301

Conflict of Interest: PC has received consulting and lecturing fees from Abbvie, Alnylam, Bristol-Myers Squibb, Gilead, Glaxo Smith Kline, Innotech, Janssen, Servier and Vifor. DS has received consulting and lecturing fees from from Medimmune, Abbvie, Bristol Meyer Squibb, Celgene, Sanofi, Roche, Servier, Gilead, AstraZeneca and Glaxo Smith Kline.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Boleto, Vieira, Desbois, Saadoun and Cacoub. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.