



Rediscovering Beta-2 Microglobulin As a Biomarker across the Spectrum of Kidney Diseases

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There is currently an unmet need for better biomarkers across the spectrum of renal diseases. In this paper, we revisit the role of beta-2 microglobulin (β_2 M) as a biomarker in patients with chronic kidney disease and end-stage renal disease. Prior to reviewing the numerous clinical studies in the area, we describe the basic biology of β_2 M, focusing in particular on its role in maintaining the serum albumin levels and reclaiming the albumin in tubular fluid through the actions of the neonatal Fc receptor. Disorders of abnormal β_2 M function arise as a result of altered binding of β_2 M to its protein cofactors and the clinical manifestations are exemplified by rare human genetic conditions and mice knockouts. We highlight the utility of β_2 M as a predictor of renal function and clinical outcomes in recent large database studies against predictions made by recently developed whole body population kinetic models. Furthermore, we discuss recent animal data suggesting that contrary to textbook dogma urinary β_2 M may be a marker for glomerular rather than tubular pathology. We review the existing literature about β_2 M as a biomarker in patients receiving renal replacement therapy, with particular emphasis on large outcome trials. We note emerging proteomic data suggesting that β_2 M is a promising marker of chronic allograft nephropathy. Finally, we present data about the role of β_2 M as a biomarker in a number of non-renal diseases. The goal of this comprehensive review is to direct attention to the multifaceted role of β_2 M as a biomarker, and its exciting biology in order to propose the next steps required to bring this recently rediscovered biomarker into the twenty-first century.

Keywords: beta-2 microglobulin, chronic kidney disease, biomarkers, kidney transplantation, pediatric nephrology, acute kidney injury, multiple myeloma, glomerular filtration rate

INTRODUCTION

Chronic kidney disease (CKD) is a common public health issue associated with astonishingly high cardiovascular (CV) morbidity and mortality and high costs, particularly for patients with diabetic nephropathy. Patients with renal failure on maintenance dialysis have excess mortality, that is, eight times higher than that of the general population (1). Most patients die due to CV events related to both traditional and non-traditional risk factors (2) and this is true for both predialysis and dialysis patients. Attempts to modify cardiorenal risk in CKD by intensive glycemic (3) or blood pressure (4, 5) control, or combined RAAS inhibition (6–8) had modest efficacy and serious adverse events. In

light of these observations, it becomes imperative to acknowledge our lack of understanding of uremic toxicity and to reexamine assumptions about biological pathways that are potentially deranged in uremia. This understanding may then satisfy a significant unmet need for better biomarkers across the spectrum of CKD. Such markers may not only be used to risk stratify patients for future clinical studies, but may also suggest targets for future pharmacological interventions.

In this report, we aim to highlight the potential role of beta-2 microglobulin (β_2 M) as a marker and possibly a mediator of some of the complications of the uremic syndrome. The classical view of β_2 M has been that the molecule is relevant to the pathophysiology of dialysis-related amyloidosis (DRA) (9–19), a truly multifactorial syndrome. The molecule itself was considered to be a relatively non-toxic uremic retention solute, whose importance as a non-creatinine (Cr) renal filtration marker was overshadowed by cystatin, when the latter was chosen for investigation in the mid-1980s and 1990s (20–22). Nevertheless, there are compelling reasons to challenge this narrow view of β_2 M.

In this paper, we will first review the basic biology and rare genetic disorders (immunodeficiency 43, OMIM #241600) associated with dysfunction of β_2 M. This overview sets the stage for reconsidering the role of β_2 M by reviewing numerous studies published in the last 5 years. In particular, we will focus on recent reports examining the role of β_2 M as a marker of renal filtration and outcomes in renal diseases across the spectrum of CKD to end-stage renal disease (ESRD) and kidney transplantation. We will also review data from non-renal diseases, a field that is usually ignored in articles focusing on nephrologists. However, this rapidly expanding literature sheds some light into the potential pathogenic role of β_2 M in human disease. Due to space limitations, we will not cover the topic of β_2 M-related amyloidosis disorders, which extend all the way from rare familial non-neuropathic amyloidosis syndromes to DRA. This is a topic that has been recently reviewed both at the biochemical (23–26) and the clinical level (10, 11), with the early literature surveyed extensively more than 10 years ago (27).

In the concluding section of this review, we will attempt to synthesize the available data, informed by our analysis of the kinetics of β_2 M and the associations between concentrations of this biomarker with outcomes. We hope that our reflections will provoke the readers to critically rethink their own assumptions about the utility of β_2 M, this easily measured, forgotten, and rediscovered protein that accumulates in renal insufficiency.

β_2 M PHYSIOLOGY AND PATHOPHYSIOLOGY

Beta-2 microglobulin was first discovered in 1964 in the urine of subjects with Wilson's disease or cadmium poisoning (28). It is a 100-amino acid protein of relatively small molecular weight (11,800 Da, size 11 Å) and it is encoded by a gene in chromosome 15 in humans. The secondary structure of the molecule consists of two large beta sheets that are linked together by a single disulfide bond (29, 30). The tertiary structure of the molecule is thus similar to the constant domain of the immunoglobulins (**Figure 1**).

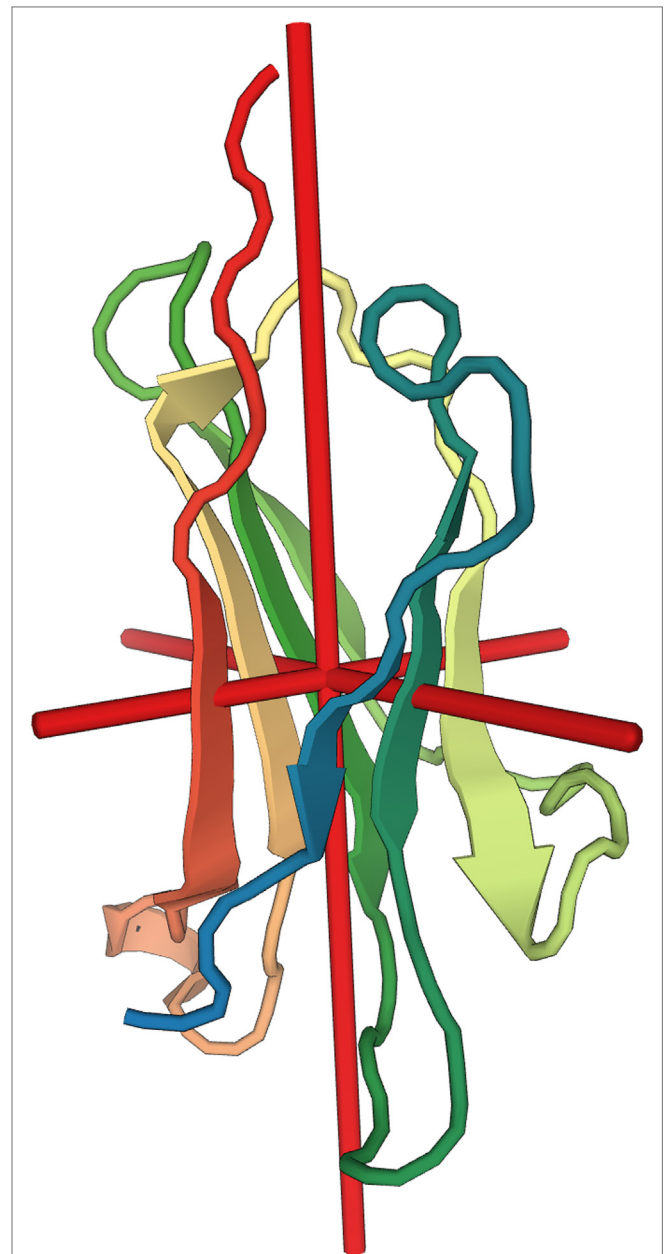
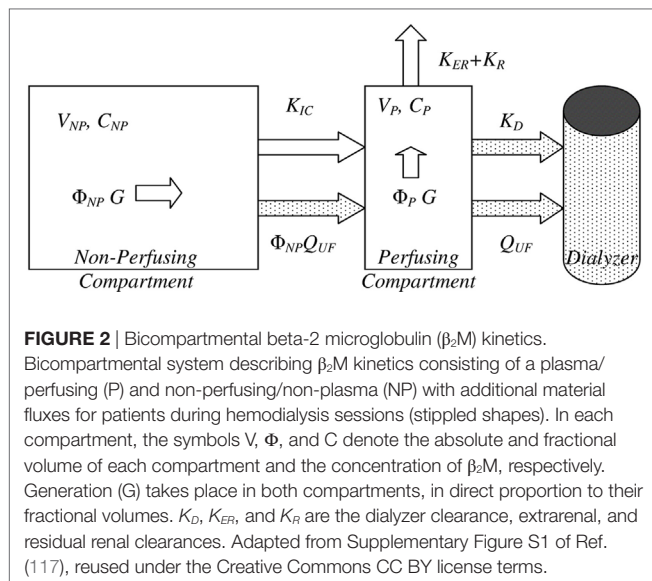


FIGURE 1 | Molecular structure of beta-2 microglobulin (β_2 M). Depiction of the secondary structure of β_2 M relative to the center of gravity of the molecule (red cross). X-ray diffraction at resolution of 1.13 Å (30). Image rendered from the Protein Data Bank entry 2YXF.

In contrast to the immunoglobulins, β_2 M does not form dimers but rather associates with the major histocompatibility complex I (MHC-I)/human leukocyte antigen I (HLA-I) on the surface of all nucleated cells. The interaction between β_2 M with the alpha chain of the HLA-I is essential for antigen presentation (31, 32). β_2 M also complexes with many non-classical MHC-I like molecules (MHC-Ib) such as CD1, MR1 (33), HLA-E, -F, -G (34, 35), neonatal Fc receptor (FcRn) (36–38), and HFE/HLA-H that are involved in mucosal immunity, tumor surveillance,



maternofetal immune tolerance, immunoglobulin and albumin homeostasis as well as iron metabolism. Disorders of β_2 M function thus arise from interruption of its interaction with classical and non-classical MHC-I molecules. Their consequences can be anticipated from the normal function of the β_2 M complexes. A thorough consideration of the entire spectrum of such disorders would by necessity encompass the entire complement of classical and non-classical MHC-I molecules and it is beyond the scope of this review (see (34, 39) and (40) for a β_2 M focused survey in the field of oncology). Nevertheless, the function of FcRn merits special mention as it provides a mechanistic link between β_2 M and another biomarker of special importance to nephrology, i.e., albumin.

The FcRn—A β_2 M-Dependent Non-Classical MHC I Molecule That Rescues Serum Proteins from Degradation

The discovery of the FcRn solved simultaneously two biological puzzles: the maternal transfer of antibodies to the offspring to protect from infections in early life and the persistence of serum albumin and immunoglobulins in the circulation (37, 41, 42). Studies in the early 1950s and 1960s demonstrated that maternal–fetal transfer of protective antibodies was dependent on the constant (Fc) part of the antibody. Furthermore, the same investigations showed that the same region underlines the long half-life of immunoglobulins vs. other proteins (~20 vs. ~5 days). The intestinal receptor responsible for transfer of antibodies from the mother’s milk to the placenta was cloned as the protein known today as FcRn (43, 44). It was subsequently shown that the same protein mediated the long half-life of immunoglobulin G (IgG) in the systemic circulation (45). Albumin also exhibits a long half-life similar to immunoglobulins, and the existence of a protection receptor had long been postulated. It was hypothesized

that a similar mechanism to that of the IgG rescue underlined the protection of albumin; this hypothesis-driven research led to the identification of the FcRn as the protein that also protects circulating albumin from degradation (38).

In contrast to the classical MHC-I molecules, the FcRn lacks sequence diversity and cannot present antigens. Nevertheless, it plays an important role in mucosal and systemic sites by rescuing immunoglobulins and albumin from degradation through the transepithelial pathway (transcytosis). β_2 M interacts with the heavy chain of the FcRn and is important for its proper function (44, 46), since mice deficient in β_2 M demonstrate abnormally short half-lives of IgG (47) and sequestration of the FcRn in the endoplasmic reticulum (48). Structural and biochemical data [reviewed extensively here (37, 41, 42)] suggest that the FcRn will bind simultaneously to albumin and IgG, but at different stoichiometries: a single albumin molecule per FcRn, whereas a single IgG will simultaneously bind to two FcRn molecules. The whole body kinetics of albumin and IgG rescue by the FcRn have been studied and a simplified model developed and fit to experimental data (49). This model makes several predictions: (1) almost one third of FcRn are available for albumin recycling, (2) the maximal capacity of albumin rescue is double that of IgG, (3) the molar recycling rate of albumin is three times that of IgG, and (4) two thirds of the plasma concentration of albumin are maintained by production rather than recycling.

More recent findings [reviewed in Ref. (37, 41)] suggest a more expanded role of the FcRn as an integral part of immune defense, bidirectionally transporting immunoglobulins and antigens to the mucosal immune system. Professional antigen-presenting cells take up antigen–IgG complexes through the classical Fc receptor (Fc γ R) at neutral pH, thus initiating receptor-mediated endocytosis. Acidification of phagolysosomes leads to “hand-off” between the Fc γ R and the FcRn and the delivery of the antigen to pathways that eventually load the antigen onto MHC-I and MHC-II molecules. The end result is a potent elicitation of CD4⁺ and CD8⁺ responses against bacterial and viral antigens (37, 41).

The Role of the FcRn in the Kidney

The FcRn is also expressed in the kidney, where it facilitates clearance of both immunoglobulins (50, 51) and albumin (52). There is some evidence for differential handling of albumin (reclamation back into the circulation) vs. immunoglobulins (elimination into the urine) (50, 53) by the intrarenal FcRn system. The role of the FcRn in renal physiology (albumin handling) and renal disease has been explored in numerous publications involving genetic knockouts and pharmacological interventions. This was shown in experiments in which wild type were transplanted to FcRn-knockout mice and *vice versa*. Transplantation of wild-type kidney to FcRn knockout mice resulted in amelioration of albuminuria and restoration of normal urinary IgG levels. This differential handling not only prevents accumulation of protein complexes that could potentially interfere with glomerular filtration, but also provides immune protection in the urinary tract. Importantly, impaired clearance of immunoglobulins by knocking out the FcRn did not result in accumulation of IgG in the glomerular basement membrane, but rendered such animals more susceptible to nephrotoxic insults (50). There is some evidence

that the podocyte FcRn system functions as an immune sensor that triggers the inflammatory response seen in certain glomerulonephritides. In particular, antithymocyte globulin treatment of podocytes increases the expression of FcRn from its high baseline state, leading to phosphorylation of the p38 mitogen-activated protein kinase, p38MAPK (54). In the same study, the percentage of glomeruli with at least two podocytes staining positive for the FcRn was characterized in human biopsies. The expression percentage was significantly higher in immune-mediated disease, including membranous nephropathy (46.7%), IgA nephropathy (66.7%), lupus nephritis (87.5%), and acute proliferative glomerulonephritis (100%), than in normal kidney samples (16.7%) ($P < 0.05$), whereas there was no significant difference between minimal-change disease and normal kidney. The relation between FcRn and p38MAPK signaling may be of pathogenetic significance since p38MAPK appears to be a major profibrotic pathway in diabetic (55), experimental nephrotic syndrome (56), and hypertensive kidney disease (57), whose inhibition leads to reduced blood pressure, sclerosis, podocyte injury, and apoptosis (58). In particular, one may postulate that activation of the β_2 M containing FcRn (e.g., by proteinuria) may trigger pathways of fibrosis inside the kidney through the p38MAPK pathway. This hypothesis, which needs to be verified experimentally, may underline the pathogenetic role of proteinuria in accelerating kidney disease progression to dialysis-dependent ESRD (59).

A major hypothesis in the nephrology literature is that proteinuria (albuminuria) underlines the progression of diverse forms of kidney disease (59, 60) and that the renoprotective effects of inhibitors of the renin-angiotensin system are partly mediated through their antiproteinuric effect (60–62). In this schema, increased oxidative stress through the NADPH oxidase system has been seen as a major contributor in promoting the progression of kidney disease (63–65), while antioxidant therapies have been proposed as a therapeutic intervention in CKD (66, 67). Interestingly, albumin overload itself has been shown to activate the renin-angiotensin system through oxidase stress and the NADPH pathway (68). These observations raise the possibility that FcRn-mediated albumin absorption may be a novel mechanism linking oxidative stress, activation of the renin-angiotensin system, and progression of kidney disease. This underexplored hypothesis has received some support in the literature. In particular, treatment with apocynin, an inhibitor of NADPH oxidase, reduced uptake of albumin by the FcRn and proteinuria in the puromycin model of nephrotic syndrome and proteinuric progressive kidney disease (69). Treatment with a monoclonal antibody against the FcRn reduced proteinuria in the same study. The possibility that the beneficial effects of renin-angiotensin inhibition are mediated to some degree through the FcRn has also been investigated in the literature. In particular, treatment of a mice anti-GBM model of glomerulonephritis with the direct renin inhibitor Aliskeren (70) reduced the glomerular deposition of IgG and reduced proteinuria in parallel with elevations in circulating IgG levels. In fact, animals that do not harbor the FcRn do not develop proteinuria and have reduced deposition of IgG compared to wild-type animals when anti-GBM nephritis is induced. The same data provided suggestive evidence that FcRn promotes the formation of subepithelial immune complex deposits (71). Finally, treatment

of podocytes with IgG derived from patients with lupus, entered the cytoplasm through the FcRn to upregulate the calcium/calmodulin-dependent protein kinase IV to activate genes linked to podocyte damage and T cell activation (72). Overall, these data suggest a role for the FcRn- β_2 M complex in both normal renal handling of albumin [along with the megalin/cubilin albumin receptor (73–75)] and IgG, as an initiating event in the podocyte injury observed in many immunologically mediated renal diseases, but also the oxidative stress that appears to underlie the progression of proteinuric forms of CKD.

Genetic Disorders of β_2 M Function

Specific mutations that interfere with the binding of β_2 M to its targets have been described in a number of conditions ranging from the rare *familial hypercatabolic* hypoproteinemia (immunodeficiency 43) (76–79) to the common *genetic hemochromatosis* (80, 81). The first two patients (siblings from a first cousin marriage) known to suffer from immunodeficiency 43 manifested a complex phenotype of hypoalbuminemia, hypogammaglobulinemia, skeletal abnormalities, and impaired delayed type hypersensitivity skin responses. These patients had circulating and total body pools of IgG less than 28% of the normal, despite having normal synthetic rates of immunoglobulins. The serum concentration for soluble HLA was less than 0.2% of normal, and iron indices were all within normal limits (79). The molecular defect was attributed to a single nucleotide transversion (G913C) in the first exon of β_2 M which impairs the function of FcRn, resulting in hypercatabolism of albumin and immunoglobulins. The immunological phenotype of β_2 M was investigated in a different consanguineous family, harboring a different homozygous splice site mutation in the first intron of the β_2 M gene (78). This mutation uncovered a cryptic splice site 4 nucleotides downstream of the canonical one, leading to a frameshift and premature termination of the β_2 M mRNA. The truncated protein had an extremely short half-life and patients had undetectable circulating and lymphocyte cell-surface β_2 M levels. HLA-I surface expression was undetectable, but there was intracellular accumulation of the HLA-A heavy chains. As anticipated, patients exhibited absence of all non-traditional MHC I molecules, i.e., CD1a, CD1b, CD1c, and FcRn from the surface of the monocytes. Similar to the first report, affected family members had severe hypoalbuminemia and hypogammaglobulinemia, with normal IgM and IgA levels. IgG responses to viral antigens were maintained, and the response to the anti-pneumococcal polysaccharide was only slightly reduced. The clinical phenotype was one of the recurrent respiratory tract infections with bronchiectasis, granulomatous dermatitis, and skin ulceration. None of the affected patients ever manifested proteinuria, possibly due to the extremely low levels of serum albumin. Circulating numbers of CD8⁺ cells were normal, but this T cell compartment consisted entirely of the $\gamma\delta$ cells. Skin lesions were infiltrated by these T cells, autoreactive NK cells, and perforin-producing CD27⁻CD28⁻CD4 cells similar to those seen in granulomatosis with polyangiitis. The NK compartment was functionally inactivated and this prevented the development of severe autoimmune phenomena against MHC-I-deficient “missing-self” cells (78, 82).

β_2 M knockout mice recapitulate many aspects of the human disease (83) and provide a model for the effects of a severe disruption in β_2 M binding. Such mice exhibit a wide variety of immunological aberrations including suboptimal IgG responses to antigenic stimulation (84), a higher catabolic rate of IgG (47) and albumin (85), hepatic and splenic iron overload (86–89), impaired interferon gamma (IFN- γ), and other cytokine responses (90–92), higher susceptibility to parasitic (93), mycobacterial (94, 95), certain viral (90, 96) and gram (–) infections (97, 98) as well as a higher susceptibility to virus induced tumors (99, 100). This animal model has also provided controlled evidence about the rescue role of β_2 M upon serum albumin, an effect that is mediated through the FcRn (38, 85). Interestingly enough these animals do not manifest albuminuria (101, 102), a feature that is attributed to the low circulating levels of albumin in these animals or possibly the “leaky” phenotype of β_2 M knockout mice. Furthermore, β_2 M-deficient mice are in general resistant to the development of proteinuria and renal disease (101, 102). When β_2 M is knocked out in the MRL-*fas^{lpr}* spontaneous lupus-like model, renal (but not skin) disease is inhibited (103, 104). Nevertheless, renal disease with the massive deposition of intrarenal immune complexes may be induced in such animals after specific and intense immunization protocols (105).

The association between β_2 M deficiency and iron overload is worthy of special mention, because it recapitulates some aspects of hereditary hemochromatosis. In the most common form of the latter disease, a C260Y mutation in the HFE molecule disrupts its association with β_2 M leading to systemic iron overload. The genetics and the clinical manifestations of hemochromatosis are very complex (106), but iron overload is seen irrespective of whether the genetic lesion refers to HFE or β_2 M. Nevertheless, there are important biochemical differences, since β_2 M-deficient mice have higher hepcidin levels which correlate inversely with the severity of hepatic iron overload (88). Furthermore, these animals fail to respond to iron overload by upregulating hepcidin levels. This may be due to abnormal cellular localization of hepcidin as seen in β_2 M silencing RNA knockdown experiments (107).

WHOLE BODY METABOLISM AND BIOMARKER KINETIC MODEL OF β_2 M

Beta-2 microglobulin is continuously generated by all nucleated cells of the body. The plasma level of β_2 M is thought to reflect release of molecules that are non-covalently bound to MHC-I into the circulation and once in the plasma β_2 M is freely filtered by the glomerulus (108). β_2 M is easily and accurately measurable with most of the commercial laboratories using the highly sensitive nephelometry method (109, 110). Serum β_2 M levels are not necessarily independent of sex, race, and ethnicity (111–113). However, in all studies to date it was found that elderly have higher serum β_2 M levels. As we will see later on in this review serum β_2 M levels also increase in solid organ malignancies, lymphoproliferative disorders such as myeloma and chronic lymphoblastic leukemia, and many autoimmune diseases such as Crohn's disease, Sjögren's syndrome, systemic lupus erythematosus, and

rheumatoid arthritis. All these are conditions, under which one would expect a higher number of cells bearing MHC molecules to be generated, or conditions in which higher shedding of β_2 M is observed (114–116).

The multiple influences affecting both generation and elimination of β_2 M raise the need for a quantitative understanding of the factors of generation, elimination, and body compartment distribution affecting the biomarker's concentrations. Our group produced such a *population-level* model by performing a kinetic-based meta-analysis of the existing studies in the field over the last 40 years (117). According to this model (Figure 2), β_2 M obeys bicompartamental kinetics and thus its behavior is a highly non-linear function of the relevant kinetic parameters. This is especially true in patients receiving hemodialysis (HD), who experience interdialytic (fluid ingestion) and intradialytic (ultrafiltration) compartment volume changes. The model may also be applied to study the kinetics of β_2 M in non-dialysis patients. In this case, considerable simplification is afforded by the lack of inter- and intradialytic volume changes and the discontinuous nature of dialytic clearance. A steady-state solution may in fact be recovered by solving the relevant bicompartamental system. However, this formula is too complex for practical use. This model, which is largely based on investigations in mostly Caucasian, young–middle-aged adult patients receiving HD, recapitulates many important clinical observations in both CKD and ESRD. The average serum β_2 M concentration in the “simulated” population was 1.59 ± 0.64 mg/l, while only 3.5% of simulated values were outside the upper reference range of 3 mg/l quoted in laboratory medicine references (118). This average compares favorably with the values previously reported to be: 1.53 mg/l (113), 1.62–1.86 (range of individuals with age compatible with the range in our kinetic meta-analysis) mg/l (112), and 1.9 ± 0.4 mg/l (119). Due to its derivation from first principles, this population kinetic biomarker model may also allow a more rigorous, quantitative evaluation of other factors (e.g., generation) affecting serum β_2 M concentration. This is a perspective that we explore in the conclusion of this review.

USING β_2 M TO ASSESS GLOMERULAR FUNCTION

There are various ways to assess renal function with changes in glomerular filtration rate (GFR) being the most widely used method. This is achieved by assessing the plasma or the urinary clearance of filtration markers with an ideal endogenous marker being the one that appears at a constant rate in plasma, is freely filtered by the glomerulus, is neither absorbed into the circulation nor secreted by the tubules, and it is not removed from extrarenal sites. Estimation of GFR by Cr-based equations lacks precision and accuracy due to non-renal determinants—such as non-renal removal, renal secretion, and variations in muscle mass—affecting serum Cr level. Researchers have been in constant search of an ideal filtration marker. In this section, we will review the evidence arguing for the adoption of β_2 M as an additional marker of glomerular filtration in CKD.

Role of β_2 M for the Assessment of GFR in Adults

Numerous studies to date have demonstrated large correlations between measures of renal function and suitably transformed serum levels of β_2 M (Table 1). These studies provide compelling reasons to suspect that one can estimate renal filtration with a β_2 M estimating equation. Recent research has explored the advance, if any, of such equations over the Cr-based estimated GFR (eGFR).

The Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) group developed a β_2 M-based GFR estimating equation in a cohort of 2,380 patients primarily comprised of Caucasians and African Americans with a mean measured GFR (mGFR), serum Cr, and serum β_2 M levels of 47.5 (\pm 21.7) ml/min/1.73 m², 1.9 (\pm 0.9) mg/dl, and 4.3 (\pm 2.4) mg/l, respectively (39). β_2 M was strongly positively correlated with serum cystatin C and Cr with Pearson coefficients of 0.9 and 0.78, respectively. Serum β_2 M was negatively correlated with GFR with a Pearson coefficient of -0.85 . The authors included the variables of age, sex, and race in the least error regression model for equation development—the coefficients for β_2 M were significant albeit small, similar to those of cystatin C, and smaller than those for Cr. Addition of these variables did not substantially improve equation performance in the whole cohort as well as various

subgroups, therefore, the final equation did not include these variables. This report also compared the precision and accuracy of equations using the metrics of interquartile range of error (difference between mGFR and eGFR of each subject), proportion of the patients in whom the eGFR was not within 30% ($1 - P_{30}$) and 20% ($1 - P_{20}$) of mGFR and root mean square error, respectively (Table 2). In that cohort, the CKD-EPI β_2 M equation achieved comparable accuracy to the CKD-EPI Cr, cystatin C, and the Cr–cystatin C equation. Nevertheless, the CKD-EPI β_2 M has an advantage over the other CKD-EPI equations in that it is independent of race, age, and sex.

Since the β_2 M estimating equation was not strongly correlated with age, sex, and race, the authors concluded that there are some other non-renal determinants of serum β_2 M and addition of those factors—if readily measurable—will lead to improvement in equation performance. One should note the apparent discrepancy between the lack of correction factors for age, gender, and race in the estimating equation for β_2 M and previously reported associations between these factors and the serum level of β_2 M (111, 112, 133). In multivariate adjusted models (133), only race (lower in blacks), smoking (higher in smokers), and proteinuria (higher in patients with proteinuria) retained inter-mediated associations with a higher serum β_2 M concentration. This discrepancy should be taken as evidence of the β_2 M to be a

TABLE 1 | Relationship between beta-2 microglobulin (β_2 M) and glomerular filtration rate (GFR) in adults.

Study	GFR measure	Correlation (1/ β_2 M)	Correlation (β_2 M)	Slope linear regression
Vincent et al. (120)	Inulin clearance	–	–	–0.87
Wibell et al. (121)	Inulin clearance	–	–0.94	–0.89
Swanson et al. (113)	lothalamate clearance	–	–	–0.82
Shea et al. (122)	lothalamate clearance	0.90	–	–
Inker et al. (123)	lothalamate clearance	–	–	–0.85
Aparicio et al. (124) ^b	⁵¹ Cr-EDTA	0.79	–	–0.75
Grubb et al. (20)	⁵¹ Cr-EDTA	0.59	–	–
Yun et al. (125) ^c	24-h creatinine clearance	–	–	–0.79
Jovanović et al. (126)	24-h creatinine clearance	0.80	–	–
Shea et al. (122)	24-h creatinine clearance	0.87	–	–
Aksun et al. (127) ^a	^{99m} Tc-DTPA GFR	–	–0.48	–
Bianchi et al. (128)	^{99m} Tc-DTPA GFR	0.76	–	–0.81
Donadio et al. (129)	^{99m} Tc-DTPA GFR	0.73	–	–
Donadio et al. (130)	^{99m} Tc-DTPA GFR	–	–	–0.81
Fry et al. (131) ^d	Timed urea collections	–	–0.63	–
Vilar et al. (132) ^d	Average of urea and creatinine collections	0.82	–0.72	–

The table reports the slope of the linear regression between log concentration and log clearance.

– Not reported.

^aStudied patients with type 2 diabetes.

^bStudied patients with sickle cell disease.

^cStudied patients with multiple myeloma.

^dStudied patients on maintenance dialysis.

TABLE 2 | Performance of beta-2 microglobulin (β_2 M), creatinine (Cr), and/or cystatin C-derived equations.

Equation	Interquartile range (95% CI)	$1 - P_{30}$ (%) (95% CI)	$1 - P_{20}$ (%) (95% CI)	Root mean square error (95% CI)
Chronic kidney disease (CKD)-EPI β_2 M	12.9 (12.2–13.8)	18.4 (16.2–20.8)	37.2 (34.6–40.1)	0.24 (0.231–0.257)
CKD-EPI Cr	11.6 (10.9–12.4)	16.4 (14.2–18.6)	34.5 (31.7–37.3)	0.224 (0.213–0.236)
CKD-EPI Cys	11.4 (10.6–12.4)	16.9 (14.9–18.6)	34.8 (32.1–37.6)	0.228 (0.217–0.239)
CKD-EPI Cr-Cys	9.3 (8.7–10.1)	11.3 (9.5–13.2)	25.5 (23.1–28.0)	0.189 (0.180–0.199)

somewhat superior marker of renal filtration that has higher correlations to the measured GFR and smaller correlations to these non-renal determinants than Cr *per se* (123, 133). Nevertheless, the influences of non-renal determinants on other factors affecting β_2 M kinetics (e.g., generation) nullify this putative advantage, so that the overall performance of CKD-EPI β_2 M equation is not different from that of other estimating equations.

To gain a better understanding of the performance of the CKD-EPI equation (134):

$$e\text{GFR}_{\beta_2\text{M}} = 133 \times \beta_2\text{M}^{-0.854}$$

we compared it to simulations based on our meta-analysis of the kinetic studies (117). These simulations, which were repeated for various levels of renal function, were then summarized with descriptive statistics (Figure 3), e.g., the mean (red), median (blue), and 95% quantile range (gray band). The relationship predicted by the CKD-EPI β_2 M equation (Figure 3, black) is essentially identical to the one predicted by the kinetic model, until about 40 ml/min. Below this level of GFR, the estimating equation predicts lower clearances for the same serum level of β_2 M. We can explain this divergence by considering that (a) a major underlying assumption of the kinetic model is that generation of β_2 M is not affected by renal impairment and (b) the non-renal determinants (e.g., variable generation) of serum β_2 M have been embedded into the coefficients of the estimating equation. It is worth remembering that the latter equation predicts an average relation that was estimated in cohorts with renal impairment and

an average mGFR of 47.7 ml/min/1.73m². If the generation rate of β_2 M varies at different levels of glomerular filtration, we would expect the statistical procedure used by the CKD-EPI investigators to balance out the influences of generation and elimination during model fitting. Furthermore, if generation is higher at lower levels of mGFR, then one would expect the CKD-EPI to provide a steeper curve between β_2 M and renal clearance than the true relation (as provided by the kinetic model) at both lower (Figure 3) and higher serum β_2 M levels. With respect to the latter point, the CKD-EPI investigators also reported that this equation underestimates mGFR at higher levels of renal function. Taken together, these observations reinforce the argument of the CKD-EPI group that poorly understood factors other than the age, sex, and race affect serum β_2 M levels. Even though the kinetic model does not allow us to pinpoint the nature of these factors, it can at least proportionate the influences of the generation and elimination processes. This feature may allow one to explore various “what-if-else” scenarios when designing clinical studies to further develop β_2 M as a biomarker.

Role of β_2 M for the Assessment of GFR in Pediatric Populations

Creatinine clearance is known to be an unreliable marker for the measurement of GFR in children due to the changing muscle mass with age. Hence other markers, including serum β_2 M, which is not influenced by muscle mass, have been investigated as potential markers that may more accurately estimate GFR. Studies published have shown mixed results. Some studies concluded that serum β_2 M may be a reliable marker to predict GFR (135–137), while others have not (138, 139). Furthermore, the recently developed CKD-Epi-Beta Trace- β_2 M formula (123) cannot be applied in children (140). In summary, the use of serum β_2 M as a measure of glomerular filtration function does not appear to be as useful in children relative to adults. However, the urinary β_2 M excretion has been used in the diagnosis of a wide variety of renal diseases in children as we discuss in the subsequent section.

URINARY β_2 M FOR THE ASSESSMENT OF TUBULAR FUNCTION

The removal of β_2 M from the serum is primarily by glomerular filtration but more than 99.9% of the filtered protein is reabsorbed and catabolized in the proximal convoluted tubule resulting in minimal urine concentration of β_2 M (usually less than 360 $\mu\text{g/l}$) (141, 142). The removal of β_2 M from the tubular fluid is postulated to be mediated through the megalin–cubilin complex (143, 144), on the basis of ligand blotting assays (145), megalin animal knockouts (146), and the human disease Donnai-Barrow/Facio-Ocular-Acoustico-Renal Syndrome (OMIM #222448) (147). This syndrome is associated with multisystemic abnormalities, developmental delay, and tubular proteinuria as a result of mutations in the *LRP2* megalin gene (148). This interaction may be mediated by the megalin component of the megalin–cubilin complex, since human cubilin mutations (megaloblastic anemia 1, OMIM #261100) manifest tubular proteinuria but with normal urinary β_2 M levels (149).

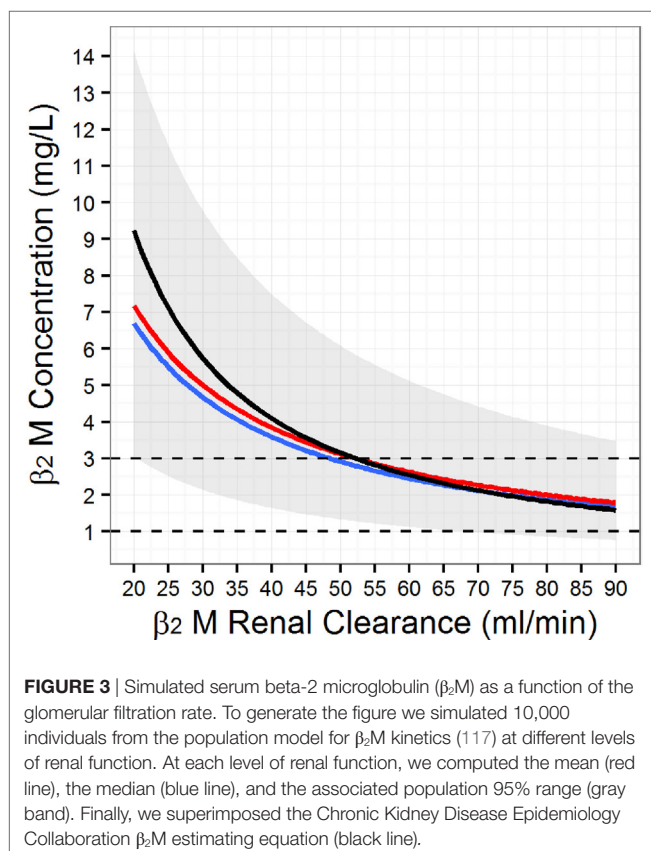


FIGURE 3 | Simulated serum beta-2 microglobulin (β_2 M) as a function of the glomerular filtration rate. To generate the figure we simulated 10,000 individuals from the population model for β_2 M kinetics (117) at different levels of renal function. At each level of renal function, we computed the mean (red line), the median (blue line), and the associated population 95% range (gray band). Finally, we superimposed the Chronic Kidney Disease Epidemiology Collaboration β_2 M estimating equation (black line).

Proteins endocytosed through the megalin/cubilin complex are targeted to the endosomes, where ligands are released from their receptors through acidification (143, 150). It is not known how much of β_2 M is degraded within the lysosomes, recycled with other MHC to the membrane surface, or transported to the basolateral surface (such as thyroglobulin or retinol binding protein). Data from experiments in rats provide evidence that β_2 M is targeted to the lysosomes (151), so that degradation appears to be the most likely fate for β_2 M. However, there is also conflicting evidence for convergent apical and basolateral endocytic systems in the proximal tubule (151, 152). As human proximal epithelial cells are capable of transcytosis of the FcRn- β_2 M-IgG (153), it is possible that some of the reabsorbed urinary β_2 M is transcytosed. However, to our knowledge no study has specifically looked for transcytosis of β_2 M absorbed through the megalin pathway. Experiments reported four decades ago provide some evidence for competitive inhibition for the absorptive tubular mechanism between β_2 M and other proteins in the tubular fluid (154–157). More recent experiments suggest similar transport kinetics arguing for a single mechanism mediating this process (158). Hence, one could anticipate variable urinary excretion levels of β_2 M in the presence of glomerular proteinuria. The tubular handling of β_2 M exhibits maturation during the neonatal period. The urinary β_2 M excretion peaks by the fifth day of life and gradually declines to adult level by 3 months of age (159). This feature suggests that urinary β_2 M not only may be a reliable biomarker of tubular toxicity but also it may even have an age dependent performance.

Role of β_2 M for the Assessment of Tubular Function in Adults

Since its initial discovery from the urine of humans with cadmium toxicity, β_2 M has been used to assess tubular function. More recently, the Nephrotoxicity Working Group of the Critical Path Institute Predictive Safety Testing Consortium assessed urinary β_2 M, along with other three biomarkers of nephrotoxicity (urinary clusterin, urinary cystatin C, and urinary total protein) in 10 mechanistic time-course studies involving 739 rats treated with eight nephrotoxins known to induce different types of renal lesions and two hepatotoxins as a means to assess specificity for kidney vs. other organ toxicity (160). Of note, β_2 M and cystatin C were specific for glomerular alternations, and with the exception of the gentamicin model, no systematic increase of either protein in urine or kidney tissue could be demonstrated when rats were exposed to other tubular toxins. Another recent animal toxicology study evaluated the performance of neutrophil gelatinase-associated lipocalin (NGAL) and four urinary biomarkers deemed acceptable by the regulatory authorities to detect acute drug-induced renal toxicity (161): β_2 M, cystatin C, kidney injury molecule-1 (KIM-1), and clusterin. In this particular study, urinary β_2 M and cystatin C increased early (prior to the detection of histological changes) and returned to the control range in the recovery phase. Furthermore, plasma β_2 M changes paralleled changes in urinary β_2 M, but correlations between the biomarker values varied according to the nephrotoxin. Nevertheless, a more

extensive evaluation of 12 markers for sensitivity (renal toxicity) and specificity (non-renal organ toxicity) in 22 rat studies, reveal that β_2 M (and cystatin C) had relatively poor area under the curve (AUC) for both tubular (AUC = 0.72) and glomerular (AUC = 0.85) toxicities (162). In the same study, urinary albumin had one of the best performance for both tubular (AUC = 0.90 vs. AUC = 0.96 of the best performing KIM-1) and glomerular (AUC = 0.99, best performing) toxicity. Collectively, the recent animal toxicology data raise important questions about both the specificity of urinary β_2 M for tubular lesions, i.e., this compound appears to detect glomerular injury better than tubular damage, and its overall utility relative to the more easily obtained assay for albuminuria. Interestingly, significant correlations between urinary β_2 M and other indices of renal damage (e.g., protein/Cr ratio) have been reported in IgA nephropathy (163) and systemic lupus erythematosus (164). Both these conditions are characterized by predominantly glomerular lesions. Consequently, controlled toxicological data and observational reports suggest some caution when interpreting urinary β_2 M elevations as indicative of a bona fide tubular process. In particular, one should always entertain the hypothesis of a glomerular process leading to proteinuria and competition of the filtered protein load for the reabsorption process in the proximal tubule (156) when interpreting a high urinary β_2 M level. Needless to say, there are virtually no human data about specific diagnostic cutoffs of the urinary β_2 M; even the three aforementioned toxicological rat studies provide different cutoff values.

A meta-analysis of various urinary biomarkers has confirmed that the value of urinary β_2 M may be limited in clinical acute kidney injury (AKI) due to sepsis (165). In this meta-analysis urinary β_2 M was found to be associated with changes in serum Cr and could differentiate between prerenal azotemia and tubular necrosis, but could not predict the clinically important outcome of need for renal replacement therapy. More recently, Zeng et al. conducted a study of diagnostic accuracy in 47 patients (166). The reference test was urinary β_2 M (normal urinary β_2 M range: 230–300 μ g/l). The sensitivity and specificity of urinary β_2 M in detecting tubular injury (assessed through KIM-1 staining in renal biopsies) was 86.6% and 64.7%, respectively. In summary, there is currently very limited evidence about the utility of urinary β_2 M in the diagnosis of AKI; most of the data come from the era of high-dose aminoglycoside therapy. Under these conditions, release of β_2 M in the urine may not even reflect actual toxicity (167). Important limitations of the literature to date are poor standardization of urine collection protocols for β_2 M and the poor stability of the analyte in acidic urine (168–171).

Various population-based studies have shown that urinary β_2 M levels can be used to detect tubular injury due to various toxins. β_2 M has been used as a marker of tubular dysfunction in subjects exposed to heavy metals such as cadmium with urinary β_2 M levels strongly correlating with serum cadmium levels (172–175). Rybakowski et al. showed that lithium-treated patients were more likely to have higher urinary β_2 M and lower eGFR than patients not treated with lithium (176). Beta-2 microglobulinuria was also seen in HIV patients on tenofovir (177–179).

Role of β_2 M for the Assessment of Tubular Function in Pediatric Populations Tubulo-Interstitial Diseases

Urinary excretion of β_2 M either in the form of fractional excretion of β_2 M (FE- β_2 M) or 24-h urine β_2 M excretion has been used in the diagnosis of tubulo-interstitial diseases (180). This study examined children with glomerular ($N = 114$), tubular ($N = 50$), or other ($N = 18$) renal diseases and showed that children with tubulo-interstitial disease had significantly higher FE- β_2 M (mean 4.27%) compared to children with glomerular disease alone (0.104%). This difference seen was not due to impairment in GFR alone as at any given eGFR, patients with tubulo-interstitial disease had higher FE- β_2 M compared to patients with glomerular disease alone (142). Children with glomerular disease and a high FE- β_2 M who underwent a renal biopsy ($N = 13$), were found to have focal areas of fibrosis, plasma cell or lymphocyte infiltration, or tubular atrophy. These patients were found to have poorer prognosis compared to patients who had pure glomerular disease. This finding was later refuted by a subsequent study (181), which showed that the urinary excretion of low-molecular weight protein (LMWP) in children with glomerular disease did not necessarily portend a poor prognosis. Urinary β_2 M excretion has also been used in the diagnosis of a variety of renal diseases that affect tubulo-interstitial function including tubule-interstitial nephritis with uveitis (182), hemoglobinopathies such as sickle cell disease (183, 184), as well as children who have received chemotherapy as part of their cancer treatment (185).

Localization of Urinary Tract Infection (UTI) and Detection of Urinary Obstruction

As urinary β_2 M level is an important reflection of tubular function of the kidneys, measuring the urine level has been used in the localization of UTI in children. Studies have shown that children with upper UTI tend to have higher urinary β_2 M excretion compared to children with lower UTI (186, 187) hence allowing for more accurate localization of infection and treatment strategies. Serum and urinary β_2 M have also been shown to be elevated in children with reflux nephropathy (188).

Urinary NGAL and β_2 M have been proposed as useful tests for the diagnosis of obstructive uropathy due to ureteropelvic junction obstruction as the levels were elevated in the pre- and peri-operative period and improved with the relief of obstruction (189). However, the control group in this study was comprised of healthy children with no renal impairment, hence it is unclear whether the elevated urinary NGAL and β_2 M level was a reflection of the impaired renal function or the obstruction itself.

Acute Kidney Injury (AKI)

Creatinine is a poor marker for AKI due to various factors including the influence of muscle mass, fluid status, and/or delayed increase in level after the occurrence of kidney injury making early intervention impossible. β_2 M has been investigated as a candidate biomarker for AKI as it is muscle mass-independent and the rise in serum β_2 M levels occurs earlier compared to the rise in serum Cr levels (190). A recent study (191) showed that

both serum cystatin C and β_2 M were better biomarkers compared to Cr in the detection of AKI in critically ill children. In a prospective study of 252 children who presented to the emergency department, urinary β_2 M, NGAL, and KIM-1 demonstrated good accuracy (AUC > 0.7–0.8) in predicting AKI (192). The caveat with using β_2 M as a biomarker for AKI is that the level varies with gestational age, hence caution will need to be exercised in using this serum marker in premature infants (193, 194).

In summary, measuring serum and urine β_2 M has been used in both predicting GFR and diagnosing renal diseases in children with variable success. Urinary β_2 M level has been helpful in the diagnosis and the monitoring of children afflicted by diseases that affect tubular function or in those who have or will be receiving medications that could affect the tubular function. Furthermore, urinary β_2 M may also be a useful marker for the early detection of AKI. Nevertheless, provocative animal and human data suggest that it may be a better marker of glomerular, rather than tubular injury.

β_2 M AS A BIOMARKER OF ADVERSE CLINICAL OUTCOMES AND MORTALITY IN CKD

There is limited research and evidence examining the role of serum β_2 M as a biomarker being able to predict adverse outcomes and mortality across the spectrum of predialysis CKD. We identified five recent studies examining the prognostic role of β_2 M in patients with CKD.

In the first study, the authors examined the relationship of plasma β_2 M levels to clinical and CV outcomes in 142 patients (mean age 67 years) at different stages of CKD. Plasma β_2 M levels increased with CKD stage and thus were highest in HD patients (195). Baseline plasma β_2 M levels were associated with vascular calcification but not with arterial stiffness or bone density. During a mean follow-up of 969 days, 44 patients died and 49 suffered a CV event. Higher plasma β_2 M levels were independently associated with overall and CV mortality and CV events in the whole cohort and with CV events in the predialysis cohort. Moreover, plasma β_2 M appeared to be a better predictor than well-established factors associated with outcomes in this population, such as eGFR (only for predialysis patients), inflammation biomarkers, and other factors included in a propensity score. Thus, they confirmed a strong relationship between plasma β_2 M levels and eGFR and the power of plasma β_2 M to predict overall and CV mortality and CV events in patients at different stages of CKD.

The association of serum β_2 M with hard clinical outcomes and its predictive ability was also examined in a prospective cohort study on behalf of the CKD Biomarker Consortium and the Chronic Renal Insufficiency Cohort (CRIC) Study Investigators (196). They examined the potential role of serum β_2 M as predictor of ESRD, mortality, and new-onset CV disease in 3,613 adults with CKD from the CRIC Study. During a 6-year median follow-up, 755 (21%) participants developed ESRD, 653 died, and 292 developed new-onset CV disease. After multivariable adjustment serum β_2 M was an independent predictor of ESRD, all-cause

mortality, and CV events. These associations were stronger than those observed for the Cr-based eGFR ($P \leq 0.02$).

Furthermore, in 2015 an interesting longitudinal cohort study came out by the same group on behalf of the CKD Biomarkers Consortium (197). They examined incident ESRD and mortality in 250 Pima Indians with type 2 diabetes (DM II) and whether serum β_2 M was associated with these outcomes. During a median follow-up of 14 years, 69 participants developed ESRD and 95 died. Serum β_2 M was associated with ESRD after adjustment for traditional risk factors and established filtration markers.

Another study examined the associations among serum β_2 M, malnutrition, inflammation, and atherosclerosis (MIA) in 312 patients with CKD between 2009 and 2015 (198). They found that serum β_2 M was more sensitive than serum Cr in predicting CV events and MIA syndrome. This study supports the hypothesis that CV events in patients with CKD should be understood as part of the MIA complex and that non-renal determinants of serum biomarkers provide prognostic information beyond that afforded by filtration biomarkers or their estimating equations.

Finally, an individual participant data meta-analysis was recently published by the CKD Biomarkers Consortium wherein they examined filtration markers, such as β_2 M, as predictors of ESRD and mortality (199). They included three general population/hazard ratio (GP/HR) studies ($n = 17,903$ participants) and three CKD studies ($n = 5,415$). They compared associations, risk prediction, and improvement in reclassification of eGFR using β -trace protein (BTP) (eGFR_{BTP}) and β_2 M (eGFR _{β_2 M}) alone and the average (eGFR_{avg}) of eGFR_{BTP}, (eGFR _{β_2 M}), Cr (eGFR_{Cr}), and cystatin C (eGFR_{cys}), to eGFR_{Cr}, eGFR_{cys}, and their combination (eGFR_{Cr-cys}) for ESRD (2,075 events) and death (7,275 events).

Mean (SD) follow-up times for ESRD and mortality for GP/HR and CKD studies were 13 (4), 6.2 (3.2), 14 (5), and 7.5 (3.9) years, respectively. Compared with eGFR_{Cr}, eGFR_{BTP} and (eGFR _{β_2 M}) improved risk associations and modestly improved prediction for ESRD and death even after adjustment for established risk factors. The authors concluded that these markers do not provide substantial additional prognostic information to eGFR_{Cr} and albuminuria, but may be appropriate in circumstances where eGFR_{Cr} is not accurate or albuminuria is not available.

In 2012, the Atherosclerosis Risk in Communities (ARIC) project investigated novel markers of kidney function as predictors of ESRD, CV disease, and mortality in the general population (200). They included 9,988 participants from population-based study in four US communities, followed for approximately 10 years. They utilized serum Cr-based eGFR calculated using the CKD-EPI equation and serum cystatin C, BTP, and β_2 M levels. The main outcomes were mortality, coronary heart disease, heart failure, and kidney failure. They found that higher serum cystatin C and β_2 M concentrations were associated more strongly with mortality ($n = 1,425$) than BTP level and all three biomarkers were associated more strongly with mortality than eGFR_{Cr} [adjusted HR for the upper 6, 7th percentile compared with the lowest quintile: 1.6 for eGFR_{Cr}, 2.9 (95% CI, 2.3–3.6) for serum cystatin C level, 1.9 (95% CI, 1.5–2.4) investigators for serum BTP level, and 3.0 (95% CI, 2.4–3.8) for serum β_2 M level]. Similar patterns were observed for coronary heart disease ($n = 1,279$), heart failure ($n = 803$), and kidney failure ($n = 130$).

The addition of serum cystatin C, BTP, and β_2 M levels to models including eGFR_{Cr} and all covariates, including urinary albumin-Cr ratio, significantly improved risk prediction for all outcomes ($P < 0.001$). They concluded that serum β_2 M and, to a lesser extent, serum BTP levels share cystatin C's advantage over eGFR_{Cr} in predicting hard clinical outcomes, including heart failure. These additional markers may be helpful in improving estimation of risk associated with decreased kidney function beyond current estimates based on eGFR_{Cr}. Subsequent investigations by the same group using data from the ARIC study have reported significant associations between serum β_2 M and sudden cardiac death (201) and fractures (119). Among the three biomarkers (Cr, BTP, and β_2 M)-based CKD-EPI estimating equations, β_2 M demonstrated the strongest association with sudden death [HR for fourth quartile vs. first quartile 3.48 (2.03–5.96) vs. ≤ 2.7 for the other kidney markers]. Renal filtration markers and albuminuria were shown to associate with fracture risk. Whereas the relationship between Cr-based CKD-EPI and risk of hospitalization for fracture was non-linear, there was a graded association between the inverse of serum β_2 M (HR per 1-SD decrease, 1.26, 95% CI, 1.15–1.37, $P < 0.001$). This risk was not attenuated and in fact increased when the investigators adjusted for the Cr-based eGFR_{Cr} to 1.37 (95% CI: 1.24–1.51, $P < 0.001$).

Therefore, when improved risk prediction (due to decreased GFR) is needed, serum β_2 M can be utilized as an alternative filtration marker beyond Cr. This finding was also independently reaffirmed in a subsequent prospective cohort study (202). In this study, the investigators sought to determine whether serum β_2 M levels have a stronger association with all-cause and CV mortality-like cystatin C compared to eGFR_{Cr} and to evaluate whether β_2 M improved risk classification beyond eGFR_{Cr}, in a nationally representative sample of adults ($n = 6,445$) in the US. Both studies mentioned above were performed on samples from the *general population*.

β_2 M IN ESRD

A non-traditional risk factor for CV mortality is the accumulation and high serum levels of β_2 M (195). The interpretation of the serum β_2 M in patients with ESRD is complicated by the non-linear, bi-compartmental kinetics, and large interindividual variability in kinetic parameters. This variability was recently quantified by our group in a patient-level meta-analysis of all studies reporting on kinetic parameters across the spectrum of CKD and ESRD (117). Using large-scale clinical trial simulations we showed that residual renal function is the major determinant of serum β_2 M concentrations even in patients receiving maintenance dialysis (195). Furthermore, enhanced dialytic removal of β_2 M will materially affect the biomarker's levels only when the residual renal clearance is less than 2 ml/min. These model-derived predictions are in substantial agreement with a large body of clinical data. They also support the further development of serum β_2 M as a measure of residual renal function in patients receiving renal replacement therapy. This topic has received some attention in the recent literature with some encouraging preliminary results (132, 203). In the following sections, we undertake an extensive review of the available literature

regarding serum β_2 M levels, delivered dialysis dose, method of clearance, and outcome measures. The key concepts behind the relevance of β_2 M in this field are the (a) middle molecule hypothesis [which in turn has directed the development of many of these dialysis techniques using β_2 M (204–209) as a proxy of other uremic toxins], (b) the lack of appreciation of the considerable effects of residual renal function in determining serum β_2 M levels even in patients receiving the most advanced forms of these therapies, and (c) the strong associations between β_2 M and outcomes (which rivals the magnitude of similar associations observed for other biomarkers, e.g., albumin) reported in these studies.

Conventional Hemodialysis

Observational studies originate strong message about the predictive power of serum β_2 M (210). The prognostic implication of serum β_2 M levels for the survival of HD patients was examined in 490 prevalent HD patients divided into two groups according to their serum β_2 M levels (lower and higher β_2 M group). During the follow-up period of 40 ± 15 months, there were 91 all-cause deaths (36 from CV causes). The results demonstrated that the serum β_2 M level is a significant predictor of mortality in HD patients, *independent of HD duration, diabetes, malnutrition, and chronic inflammation*. This observational study provides a modest argument about the *clinical importance of lowering serum β_2 M* in patients receiving maintenance HD. Counter to this argument is a report on the relationship between serum β_2 M and survival of chronic HD patients and of the association of serum β_2 M levels with mortality (211). Surprisingly, this study showed that higher serum β_2 M levels are associated with better survival in these patients. This paradoxical association may be a manifestation of “reverse epidemiology”, since nutritional status was an independent predictor of serum β_2 M concentration in the aforementioned study.

The association of inflammatory biomarkers and β_2 M has been the focus of many studies during the 1990s before the development of modern synthetic dialyzers (17, 212–215). This partially contradictory literature suggests an association between inflammation, triggered by membrane material, and serum β_2 M concentrations. To the extent that inflammation is a non-traditional factor for CV and overall mortality, as recently reviewed by Ref. (216), one would expect the association between serum β_2 M and mortality to be partly attributed to the confounding role of inflammation. Nevertheless, there is a paucity of more modern studies examining the association between serum β_2 M and risk factors for mortality in dialysis. A small study of 40 patients in high-flux (HF) HD for more than 6 months examined the association of serum β_2 M with inflammation and dyslipidemia as CV risk factors (217). There was no correlation of serum β_2 M with C-reactive protein (CRP) and IL-6 when HF membranes were used. During the follow-up period of 3 years, 6 out of 40 patients died from CV events. A significant relationship of β_2 M with dyslipidemia and mineral bone disorders, but not with inflammation was observed. Along the same lines, other groups have reported associations of serum β_2 M with suppressed interferon-gamma production, but not the traditional inflammatory marker of CRP when patients are switched from low flux (LF) to HF dialyzers (218, 219). These observations mirror similar findings in non-dialysis-dependent

CKD (198). Therefore, β_2 M might have an important role in the development of CV diseases, independent of other traditional and non-traditional risk factors even when patients are dialyzed with highly permeable HF membranes. There is a need for large, modern studies in this era of HF dialyzers and ultrapure dialyzate to better understand the magnitude and significance of β_2 M in patients receiving maintenance HD.

Observational studies and randomized clinical trials (RCTs) suggest that HF HD efficiently removes β_2 M from the blood and has positive effects on the survival and morbidity of uremic patients when compared with LF HD. The bulk of information [96% of all patients and events in the most recent meta-analysis by the Cochrane group (220)] is provided by two large multicenter RCTs. The hemodialysis (HEMO) study was a RCT designed to examine the impact of two treatment parameters (dialysis dosage based on urea Kt/V and membrane permeability) on clinical outcomes of maintenance HD patients (221). In the HEMO study, membrane flux was defined by the clearance of β_2 M (surrogate for the clearance of middle molecules). The primary analysis of the HEMO study did not show a statistically significant reduction in the rate of the primary outcome and all-cause mortality. In secondary analyses, however, a 20% decrease in cardiac death was observed for the HF group compared with the LF group. In the subgroup of patients who had been on dialysis for >3.7 years before enrollment in HEMO, HF was associated with lower all-cause mortality, cardiac deaths (221, 222), and cerebrovascular events (223). As expected, the cumulative mean predialysis serum β_2 M level during follow-up in the HF arm was statistically significantly lower than that in the LF arms. Furthermore, predialysis serum β_2 M levels predicted all-cause mortality even after adjustment for years on dialysis and residual kidney function (224). A subsequent, secondary analysis of HEMO examined the association of serum β_2 M levels and dialyzer β_2 M kinetics with the two most common causes of deaths in the HEMO study: cardiac and infectious diseases (225). In this report, the cumulative mean predialysis serum level of the middle molecule, β_2 M, correlated positively with the relative risk for infectious deaths in the HEMO study.

The Membrane Permeability Outcome (MPO) study is the second largest RCT to investigate the impact of membrane permeability on survival in incident HD patients. This study adopted a novel design, in that it specifically made a distinction between patients who had low albumin (≤ 4 g/dl) and normal albumin (> 4 g/dl) as separate randomization groups (226). The target patient population in MPO was different from the HEMO cohort, which only enrolled patients with no residual renal function who had been on dialysis for more than 3 months. In MPO, patients with serum albumin ≤ 4 g/dl had significantly better survival in the HF group compared with the LF group (227, 228). A *post hoc* secondary analysis showed that HF membranes may significantly improve survival in diabetic patients. No difference was found in patients with normal albumin levels. Our group reanalyzed the data from the HEMO and MPO studies to take into account dialyzer reuse in HEMO (reuse was not permitted in MPO). Our secondary analysis (229) reaffirmed the message from these two large, high-quality RCTs: HF dialysis with non-reused dialyzers was associated with an adjusted HR of 0.63 (95% CI: 0.51–0.78),

relative to their LF counterparts. Reductions of serum β_2 M explained only one-third of the mortality benefit of the non-reused dialyzers in this report, raising the possibility that there are other, non- β_2 M mediated, beneficial effects of HF dialysis.

Other investigations have attempted to shed a light into the non- β_2 M-related effects of HF dialysis. One recent study explored the effect of membrane flux on CV risk factors and on β_2 M plasma levels in patients treated with extended dialysis (between 5 and 8 h for all patients). In this trial, patients were randomly assigned to the treatment sequences LF/HF dialysis vs. HF/LF dialysis in a crossover design after a 3-month run-in period, with each phase lasting 9 months (230). This study did not find an influence of HF filters on several traditional CV risk factors, despite the significant reduction of plasma β_2 M levels at the end of the HF phase. At the time of this writing, the beneficial effects of HF dialysis on CV outcomes can only partly be attributed or explained to reductions in plasma β_2 M levels, or even to improvements in immune function (218). Even though we do have firm evidence from the HEMO and MPO that therapies associated with more efficient dialytic removal of plasma β_2 M will improve CV outcomes, reduction in the plasma levels of this marker only partly explain this effect.

Collectively, the bulk of available evidence highlights the potential of plasma β_2 M and its higher removal to serve as biomarker of outcomes, particularly CV mortality, in patients receiving conventional thrice weekly dialysis. Pitfalls of reverse epidemiology, the less than perfect association of reduced plasma β_2 M with survival, study limitations and finally the disparate effects of plasma β_2 M in patients with hypoalbuminemia suggest that additional biomarkers are needed to both understand the effects of HF dialysis on clinical outcomes and provide a causal explanation about the role of β_2 M in mediating these outcomes.

Hemodiafiltration

Online hemodiafiltration (OL-HDF), the most efficient renal replacement therapy, enables enhanced removal of small and large uremic toxins by combining diffusive and convective solute transport. Four meta-analyses of RCTs and narrative reviews in this area showed inconsistent results concerning the effect of convective treatments in improving general and CV survival. Nevertheless, these analyses suggest that OL-HDF may significantly reduce intradialytic symptomatic hypotension (231–236). Simulation studies anticipate that there should be a steep effect of convection volume (dose of OL-HDF) and achieved plasma β_2 M levels in patients receiving HDF (117). These simulation results originate from measurements in actual patients receiving convective therapies (237). By inference, one would expect OL-HDF to be associated with improved survival in prevalent dialysis patients receiving higher convection volume. This hypothesis is supported by observational studies and secondary analyses of RCTs. This evidence, reviewed further below, indicates that the observed reduction in mortality associated with OL-HDF correlates with the convection volumes delivered. The Dialysis Outcomes and Practice Patterns Study, an observational study involving 2,165 patients, was the first to identify the role of convection volume in patient outcome (238). This study showed that 15–25 l of substitution volume per session (not including weight loss for

extracellular fluid control) resulted in a 35% reduction in mortality with high-efficiency OL-HDF relative to LF HD.

The hypothesis of an effect of convective volume on outcomes was also explored in a *post hoc* fashion in the large HDF trials reported in the last 5 years. Although the CONTRAST Study, a RCT of OL-HDF vs. LF-HD involving 714 patients was not able to prove the superiority of OL-HDF over conventional LF HD in its primary end point of mortality, *post hoc* analysis identified that larger volumes of convection fluid were associated with a significant reduction in all-cause and CV mortality (239). The Turkish HDF Study (240) was a RCT involving 782 patients which compared survival rates for OL-HDF versus HF HD; again, no significant differences in primary end points were observed, but *post hoc* analysis indicated significantly reduced mortality in the subgroup of patients receiving the largest substitution volumes (>17.4 l/session). Finally, the ESHOL Study, a prospective RCT comparing postdilution OL-HDF with HF-HD involving 906 prevalent patients, reported a 30% reduction in all-cause mortality, 33% in CV mortality, and 61% risk reduction in mortality from stroke (241). Interestingly, in this study a mean delivered convection volume of 23.7 l/session was required to achieve this magnitude of reduction in mortality.

The convection volume threshold and the range associated with survival advantage were assessed in a large cohort of incident adult patients ($n = 2,293$) treated by postdilution OL-HDF over a 101-month period (237). The relative survival rate of OL-HDF patients, adjusted for age, gender, comorbidities, vascular access, albumin, CRP, and dialysis dose, was found to increase at about 55 l/week and to plateau at 70–75 l/week. Similar analysis of pre-dialysis plasma β_2 M concentrations found a nearly linear decrease as convection volume increased from 40 to 75 l/week. Thus, a convection dose target based on convection volume should be considered and needs to be confirmed by prospective trials as a new determinant of dialysis adequacy in patients receiving convective therapies.

An individual pooled participant analysis of the largest trials mentioned above is in line with these observations (242), suggesting a better survival when a convection volume of at least 23 l/session was delivered. Nevertheless, none of the large convective therapies trials has targeted these high volumes. Since patients were not randomized to these high targets, it is very likely that the results of these *post hoc* analyses are strongly confounded by other factors (234). In particular, high convection volumes can only be achieved if the dialysis access can support a high enough flow rate to keep the dialysis filtration fraction at a safe range (less than 30%). Participants with better functioning accesses and/or those receiving longer treatments, factors that are known to be linked to better patient outcomes, would thus have received higher convection volumes. Only well-designed RCTs with rigorous controlled convection volume targets can provide unambiguous evidence for the beneficial effects of higher convection volumes on outcomes.

Hemofiltration

Hemofiltration is a pure convective form of renal replacement therapy, which does not utilize a dialysis component. The effect of on-line high-flux hemofiltration (OL-HF hemofiltration) vs.

LF HD on mortality in CKD was studied in a small RCT (243). They compared OL-HF hemofiltration with ultrapure LF HD, assessing survival and morbidity in patients with ESRD. It was an investigator-driven, prospective, multicenter, 3-year-follow-up, centrally randomized study with no blinding and based on the intention-to-treat principle. Prevalent patients with ESRD (age, 16–80 years; vintage > 6 months) receiving renal replacement therapy at 20 Italian dialysis centers were included and centrally randomly assigned to HD ($n = 32$) or hemofiltration ($n = 32$). All-cause mortality, hospitalization rate for any cause, prevalence of dialysis hypotension, standard biochemical indexes, and nutritional status were monitored. There was significant improvement in survival with hemofiltration compared with HD (78%, hemofiltration vs. 57%, HD) at 3 years of follow-up after allowing for the effects of age ($P = 0.05$). β_2 M plasma levels remained constant in HD patients (33.90 ± 2.94 mg/dl at baseline and 36.90 ± 5.06 mg/dl at 3 years), but decreased significantly in hemofiltration patients (30.02 ± 3.54 mg/dl at baseline vs. 23.9 ± 1.77 mg/dl; $P < 0.05$). This was a small preliminary intervention study with a high dropout rate and problematic generalizability. They concluded that OL-HF hemofiltration may improve survival independent of Kt/V in patients with ESRD, with a significant decrease in plasma β_2 M levels and increased BMI. A larger study is required to confirm these results. Such a study could include an arm of higher volume OL-HDF in order to probe the differential effects (if any) of pure convection vs. mixed convection/diffusion in achieving lower plasma β_2 M levels and improving patient outcomes.

Peritoneal Dialysis (PD)

The association of β_2 M and patient survival in patients receiving PD is underexplored. In the largest observational study to date 771 PD patients were selected from the Clinical Research Center registry for ESRD cohort in Korea in order to examine the association of serum β_2 M levels with all-cause mortality (244). The patients were categorized into three groups by tertiles of serum β_2 M levels, and the median follow-up period was 39 months. The all-cause mortality rate was significantly different according to tertiles of serum β_2 M in PD patients ($P = 0.03$). Multivariate Cox proportional analysis showed that the HR for all-cause mortality was 1.02 (95% CI 1.01–1.04, $P = 0.006$) per 1 mg/l increase in serum β_2 M after adjustment for multiple confounding factors that relate to malnutrition and for inflammation markers. However, serum β_2 M was not associated with all-cause mortality after adjustment for residual renal clearance. Even though these results are supportive of the potential role of the serum β_2 M level as a predictor of mortality in PD, they suggest that this association is a reflection of the residual renal function, a powerful predictor of mortality in patients receiving PD (245–247).

The effects of higher peritoneal clearance of serum β_2 M on mortality in PD patients are much less certain. Relevant data come from a study which investigated whether baseline peritoneal loss and clearance of albumin and other proteins is a risk factor of death (248). Mass-transfer area coefficient of Cr and peritoneal clearances of albumin, β_2 M, α_2 -macroglobulin, and IgG were calculated during a standard peritoneal permeability analysis. The total amount of albumin loss in the dialysate was

also calculated. Overall mortality was studied with an intention-to-treat analysis. High baseline albumin clearance was associated with fast transport status, the presence of peripheral arterial disease, and a high comorbidity index, whereas CRP did not differ from the patients with low albumin clearance. Age, high comorbidity score, CRP > 10 mg/l, and a low serum albumin were associated with mortality. Peritoneal albumin clearances and albumin loss were not associated with death in crude and adjusted analysis. Similarly, peritoneal clearances of IgG, α_2 -macroglobulin, and β_2 M were not determinants of survival. They concluded that baseline peritoneal albumin and protein clearances are associated with signs of comorbidity, but this does not have a measurable effect on patient survival. However, these findings are tempered by the fact that higher clearances are associated with a fast-transport phenotype, which itself is a predictor of worse outcomes in PD (249, 250). Future studies should examine the effects of PD clearance irrespective of membrane transport status to better clarify the role of β_2 M and its clearance in PD.

β_2 M IN KIDNEY TRANSPLANTATION

Chronic allograft damage is still a leading cause of graft failure 1-year posttransplantation (251). The pathophysiology of this entity is still not clearly understood but both alloantigen-dependent and alloantigen-independent factors act together to initiate inflammatory reactions that eventually lead to loss of nephrons followed by interstitial fibrosis and tubular atrophy (IF/TA) in the graft (252). Alloantigen-dependent factors that can lead to chronic allograft damage include recurrent T-cell-mediated rejection, antibody-mediated rejection, and the presence of donor-specific antibodies (253). Alloantigen-independent factors that can lead to chronic allograft damage include ischemia/reperfusion injury, donor age, arterial hypertension of the donor, drug toxicity, infections, diabetes and hypertension in the recipient, recurrent and *de novo* glomerular disease, and the presence of proteinuria.

Non-invasive diagnostic studies that may help in determining whether chronic allograft damage is present include monitoring for proteinuria, monitoring for donor specific antibodies, and monitoring for changes in the serum creatinine (254–257). But none of these tests are specific for making a diagnosis of chronic allograft damage, and elevations in serum creatinine lag behind the histological changes observed in chronic allograft damage. Thus, identifying urinary biomarkers that can detect early tubular injury would be beneficial in helping to identify those patients who need an allograft biopsy earlier on so that further progression of chronic allograft damage is prevented. Similar to the available evidence from patients with non-transplant CKD, serum β_2 M-based eGFR (<30 vs. >60 ml/min) has been found to predict CV events [HR: 2.56 (95% CI: 1.35–4.88; $P = 0.004$)], overall mortality [HR: 4.09 (95% CI: 2.21–7.54; $P < 0.001$)], and dialysis dependent kidney failure [HR: 15.53 (95% CI: 6.99–34.51; $P < 0.001$)] in allograft recipients (258). The predictive ability of elevations in serum β_2 M for subsequent allograft loss has also been reported by other groups (259). Many *de novo* donor-specific antibodies recognize free serum β_2 M (260), but the significance of this association, i.e., whether it simply reflects false-positive reactions (more likely) or it is pathophysiologically significant (less likely) remains to be established.

Urinary β_2 M As a Biomarker in Chronic Allograft Damage

Proteomic analysis has been used in various studies in an attempt to identify a protein biomarker pattern that can help reveal chronic allograft damage. A very promising approach (261) used surfaced-enhanced laser-desorption/ionization time-of-flight mass spectrometry (SELDI-TOF-MS) to identify urinary proteins as biomarkers for chronic allograft damage. In this retrospective study, there were 34 renal transplant patients (disease group) with histologically proven chronic allograft damage, with an eGFR less than 45 ml/min who were more than 1-year posttransplantation. These patients were compared to a “control” group of 36 renal transplant patients with normal renal function (eGFR > 50 ml/min). Significantly higher concentrations of β_2 M were observed in the urine of the patients with chronic allograft damage compared with the controls (261). In another study (262), using the same population as that in Ref. (261), OrbiTrap mass spectrometry was utilized to analyze the urine further for identification of more biomarkers specific to chronic allograft damage. Again β_2 M was shown to be significantly increased in chronic allograft damage, with an approximately 50-fold increase of β_2 M expression in this cohort compared to the control group ($P < 0.0001$). Other proteins that were significantly increased in the chronic allograft damage cohort were clusterin and NGAL. Apolipoprotein A1 and uromodulin levels were significantly decreased in the same cohort compared to the control group (262).

Despite these encouraging observations, we currently lack a firm understanding of the pathophysiological processes underlying chronic urinary β_2 M elevations in kidney transplant recipients. One possibility is that they reflect chronic immunological injury. This is certainly possible, since urinary β_2 M is increased in patients with *acute rejection*. A previous study using unbiased proteomic analysis (SELDI-TOF-MS) identified many urinary fragments in the mass/charge (m/z) region 5,270–5,550 (region I; five peaks), 7,050–7,360 (region II; three peaks), and 10,530–11,100 (region III; five peaks) that always occurred together; the normal urine pattern had no peak clusters in these m/z regions (263). Interestingly, about 18% of patients with stable allograft function exhibited this pattern. A follow-up investigation by the same group used liquid chromatography–mass spectroscopy techniques to identify these peaks as cleaved β_2 M. The authors concluded that fragmented urinary β_2 M can serve as a potential biomarker for acute tubular injury due to rejection in renal allografts (264). The association of urinary β_2 M with acute rejection has been noted in two other unbiased proteomic studies utilizing matrix-associated laser desorption ionization time-of-flight mass spectroscopy (265, 266) by the same group. Whereas the first study suggested specificity for acute rejection, the second one did not, as β_2 M elevations were also seen in patients with non-transplant forms of CKD. Interestingly, another group reported that urinary β_2 M is elevated in renal transplant recipients even in the setting of good allograft function; this was different from patients with non-transplant-associated CKD who had high urinary β_2 M levels only when the Cr clearance was less than 30 ml/min/1.73 m² (267). One could postulate that elevations in urinary β_2 M in the absence of changes in serum Cr could be used to detect acute

rejection early. Furthermore, chronic elevations in urinary β_2 M could reflect ongoing low-grade immunological injury, leading to IF/TA and eventually to allograft loss. Larger studies are needed to obtain a better understanding of the factors affecting urinary β_2 M and its determinants in renal transplant recipients.

In summary, urinary β_2 M may be sensitive for this entity, but further research in this area is needed to identify whether it can be used as a reliable biomarker for identifying patients with early chronic allograft damage due to immunological factors who need an allograft biopsy for the effective management of this complex disease process.

β_2 M IN NON-RENAL DISEASES

Not only the pathology related to renal disease but also non-renal etiologies have influence on serum β_2 M level. Higher serum β_2 M level can be seen in patients smoking, of non-black race, and with a higher amount of protein excretion in the urine (133). As β_2 M is a light chain subunit of MHC class I antigens, it is present in all nucleated cells, especially on immunocompetent cells such as macrophages, active T and B lymphocytes. During normal cell turn over, it is released into the body fluids. Pathologies with high cell turnover, such as hemato-oncological conditions, and rheumatologic diseases are associated with higher serum β_2 M levels (268, 269). As many of these conditions may be associated with the subsequent development of CKD (270–272), we will briefly review some of these associations. Our aim is not to compile an exhaustive presentation of this large and rapidly expanding literature. Rather, we aim to draw attention to representative reports from other areas of Internal Medicine and highlight the relevant key messages for Nephrologists.

Hemato-Oncological Pathology

In hematological malignancies, such as leukemia, lymphoma, and multiple myeloma, serum β_2 M level is found to be elevated, despite preserved renal function. It has been reported that 60% of patients with mantle cell lymphoma have high pretreatment serum β_2 M level (273). This elevated value is independently associated with unfavorable prognosis of most of the hematological malignancies (269, 273–277). These associations persist despite adjustment for well-validated clinical prognostic scores and therapy indicators (274, 278, 279).

In multiple myeloma, serum β_2 M level is the main determinant of the International Staging System (ISS, Stage I: β_2 M < 3.5 mg/dl and albumin > 3.5 g/dl, Stage II: β_2 M > 3.5 but less than 5.5 mg/dl or β_2 M < 3.5 mg/l and albumin < 3.5 g/dl, Stage III β_2 M > 5.5 mg/l). β_2 M predicts not only the prognosis but also the progression of asymptomatic disease (HR 3.30; $P = 0.002$) (280) and even outcomes after stem cell transplantation (281, 282). The association between serum β_2 M and albumin upon patient prognosis in myeloma, not only is reminiscent of similar associations noted in the dialysis literature but also is very robust statistically. Even recent proposals for a revised ISS (283) based on emerging biomarkers (e.g., chromosomal abnormalities) or the levels of soluble free light chains (284) have highlighted the prognostic significance of high serum β_2 M levels. Although the

association between β_2 M and prognosis has been interpreted to reflect a higher tumor burden in myeloma (285) or a more aggressive (286, 287) myeloma subtype, it is important to realize that renal insufficiency may underline at least in part the higher levels of serum β_2 M seen in this disease (288). Despite the different cell origin, chronic leukocytic leukemia recapitulates the findings from multiple myeloma. β_2 M is a well-recognized adverse prognostic factor in this disease (289–291); even though one would expect the utility of this marker to be higher at the latter than the earlier stage of this disease, this hypothesis is not entirely borne out by observations (290). Even patients with early-stage disease have elevated serum level of β_2 M, which may reflect more aggressive behavior of the malignant process. This alternative hypothesis, i.e., that serum elevations of this biomarker may reflect the combination of higher tumor burden and more aggressive biology, is supported by observations that higher serum levels of β_2 M are associated with shorter time to therapy (291). Furthermore, failure to normalize serum β_2 M after 6 months of kinase inhibitor therapy (ibrutinib) was associated with inferior progression-free survival [HR 16.9 (95% CI: 1.3–220.0), $P = 0.031$] for ibrutinib-treated patients. This association persisted after multivariate adjustments (292).

Elevated serum β_2 M level can also be seen in patients with solid cancers, such as ovarian cancer (293), gall bladder cancer (294), prostate cancer (295), breast cancer (296), and renal cell carcinoma (297). Its higher value is closely related to the poor prognosis and aggressive characteristics of the tumor (293–297). Due to the high prevalence of high β_2 M in patients with ovarian cancer, β_2 M has been incorporated into the FDA approved OVA1 multianalytes assay for risk stratification of adnexal masses (298, 299). OVA1 measures the serum levels of five analytes, CA125, transthyretin, apolipoprotein A1, transferrin, and β_2 M. Results are reported as high or low risk for ovarian cancer and are used to determine whether referral to gynecologic oncology is required prior to surgical treatment of an adnexal mass.

More recently, evidence has emerged that implicates serum β_2 M level as a global biomarker of occult malignancy [HR: 1.25 (95% CI: 1.06–1.47; $P = 0.002$ for the trend of higher risk with increasing β_2 M quartile)], and more narrowly colorectal cancer risk [HR: 2.21 (95% CI: 1.32–3.70; $P = 0.001$ for the trend of higher risk with increasing β_2 M quartile)]. These associations, which were not attenuated after adjustment for an inflammatory biomarker, CRP, or even renal function (eGFR) of 12,300 patients, were noted in the prospective ARIC study. Significant associations were also observed for mortality from total, lung, and hematological cancers (300).

Autoimmune Disease

Serum β_2 M is elevated in autoimmune diseases as well. Higher serum levels are seen in patients with systemic lupus erythematosus and adult-onset Still disease, especially in those with active diseases and hemophagocytic syndrome (268, 301). After therapy, the serum β_2 M level decreased significantly (268). Urinary β_2 M has been shown to correlate with overall and renal disease activity scores and proteinuria (164). Patients with active primary Sjögren's syndrome, notably those with increased systemic

disease activity (302) and history of lymphoma (303, 304), and hemophagocytic lymphohistiocytosis (305) were also found to have elevated urinary β_2 M level. Interestingly, higher levels of urinary β_2 M are found in patients with active primary Sjögren's syndrome and impaired eGFR (306).

β_2 M: SYNTHESIS AND THE WAY FORWARD

In this review, we examined the recent literature linking elevated serum circulating and urinary β_2 M levels to outcomes across the spectrum of renal impairment and also its role as biomarker in non-renal diseases. This literature suggests that β_2 M may be a particularly strong (sensitive) biomarker for both morbidity and mortality across numerous clinical conditions. This lack of specificity for particular clinical states, necessitates the application of a suitable context that would allow the interpretation of alterations in serum and/or urinary β_2 M levels. Such a framework would by necessity be context specific given the ubiquity in expression of β_2 M. For future applications in non-dialysis-dependent CKD, such a model would most likely have to incorporate additional biomarkers to derive a complex, multivariate measurement of renal function. Existing approaches such as the combined β_2 M and BTP formula referenced previously (199) show one possible research thread that may yield fruitful results. Nevertheless, it is worth remembering that the concentrations of several LMWP's retained in CKD are poorly predicted by different eGFR formulas in a CKD population (stages 2–5 not on dialysis) (307). If this is indeed the case, then one may not improve much upon existing estimating equations through simple formulas based on biomarker level averaging, e.g., as was done when developing the cystatin-C/Cr eGFR (308) and β_2 M-BTP formulas (123). This raises the question of alternative approaches for the full subsequent development of β_2 M as a biomarker in CKD.

We believe that the way forward for β_2 M should be based on quantitative models for generation (non-renal determinants) and elimination (renal determinants) of this biomarker. For example, the population kinetic model we put forward (117) for the exploration of the “middle molecule” hypothesis for uremic toxicity maintains the separation between the processes of generation and elimination, while generating predictions that verify clinical observations in the dialysis population. When applied outside its intended application domain, i.e. in the field of CKD, it generates predictions that should be contrasted against the relationship between plasma β_2 M and the measured GFR estimated from extensive database analyses. Hence this model, developed on fewer than 150 patients, who were nonetheless extensively phenotyped, draws attention for further research on the generation (non-renal determinant) mechanisms that affect the serum levels of β_2 M.

Future research should expand this model to account for changes in the concentration of β_2 M in the urine. Such studies are urgently needed, because the urinary β_2 M appears to have a much larger utility than previously recognized. Animal toxicology experiments conducted under rigorously controlled

conditions (160, 162) and provocative observations in glomerular diseases (163, 164) suggest that urinary β_2 M may in fact be a better marker of glomerular than tubular damage. This finding, backed by controlled experiments in hundreds of animals and observations in a much smaller number of humans, seems to go against textbook dogma. The latter, however, is based on a handful of observations (20 normal, 15 with glomerular, and 15 with tubular pathology due to cadmium poisoning and hereditary syndromes) made 50 years ago at the dawn of the clinical chemistry and renal pathology and before the complex β_2 M-dependent mechanisms of protein transport in the kidney were deciphered (141). It is high time that studies acknowledging the biology of β_2 M and its complex compartmental kinetics in serum and urine are undertaken so that the role of β_2 M as a biomarker be clarified. The outcomes of this exercise are not merely academic and are not limited to the field of Nephrology, considering the importance of β_2 M in other fields (mainly hematological oncology). Furthermore, this re-examination has the potential to link β_2 M to another important biomarker, albumin whose kinetics and biofluid levels are controlled by β_2 M through the FcRn.

At this point, we would like to put forward a hypothesis that we think ties together many observations in both CKD and ESRD: the emergence of non-renal processes as determinants of serum β_2 M levels as renal function declines and the beneficial effects of HF-HD dialysis in patients with hypoalbuminemia as highlighted by reports from our group (229, 309). We hypothesize that this higher generation comes not from high cell turnover, e.g., as in oncological conditions, but from altered cell binding of β_2 M to the many proteins that it chaperones. According to the model, the level of renal function is the main determinant of plasma β_2 M concentrations by affecting both removal (glomerular filtration) and generation of free β_2 M. Interference with the binding of β_2 M to MHC and non-classical MHC molecules [possibly in the endosomes where these interactions are initiated (310, 311)] by other uremic retention solutes constitutes a major source for the heightened generation of β_2 M in uremia. Clinical manifestations of these alterations result both from the higher concentrations of β_2 M (e.g., DRA) as well as the altered MHC/non-classical MHC function, i.e., the phenotype of β_2 M deficiency highlighted in the β_2 M knockout mice. Observations in these animals parallel the clinical observations and laboratory associations in patients with renal dysfunction: dysregulated IFN- γ production (312, 313), tuberculosis (314–316), acute infections, and suboptimal antibody responses [all reviewed in Ref. (317)]. Patients with renal dysfunction or on dialysis have also a higher incidence of tumors that are considered to be of viral origin in registry studies (318–323). According to this model, the hypoalbuminemia seen in many dialysis patients is a reflection of a widespread abnormality in albumin rescue through the FcRn.

This hypothesis could provide an explanation for the beneficial effects of HF-HD in hypoalbuminemic patients. Although suggested by some *in vitro* studies (324, 325), other *ex vivo* (326, 327) and *in vivo* (328) investigations did not demonstrate an effect of flux on β_2 M gene transcription or protein expression. On the other hand, both *in vitro* (329) and *in vivo* (328) flow cytometric studies

have shown that dialysis with LF membranes is associated with a larger dissociation of β_2 M from the HLA-I complex compared to their HF-HD counterparts. To the extent that this model is true, it would provide a partial molecular explanation for the clinical associations between higher β_2 M concentrations (greater disruption of β_2 M binding) and infectious mortality (210, 224, 225). This model yields hypotheses about the binding behavior of β_2 M to its targets, and the resultant regulation of biological processes, e.g., IFN- γ and immunoglobulin levels in relation to higher clearance (renal or dialytic), which can be tested in small randomized crossover studies and laboratory experiments (218, 313, 329, 330). In that regard, a “candidate toxin” approach based on the EUTOX Uremic Solutes Database (331) could provide a way to test a number of known toxins *in vitro* experiments for a disruptive effect on the protein complexes of β_2 M. Given the potent effect of inflammatory stimuli on the function of the MHC/ β_2 M system, future work in this area should attempt to control for the confounding effects of microinflammation, which is prevalent in dialysis patients, as well as possible dialysis membrane-immune system interactions.

CONCLUSION

In conclusion, β_2 M is a promising marker to assess glomerular and tubular function in adults. It has similar performance to the Cr-based estimating equations as a measure of renal function, but may be more strongly associated with CV morbidity and mortality than Cr, or other small molecular renal filtration markers. β_2 M is also an important, emerging biomarker in numerous non-renal diseases. Plasma and urinary β_2 M levels can be reliably and cost effectively measured, which makes it an ideal screening tool. Plasma and urinary β_2 M levels can increase in certain conditions, which might limit its efficacy as a diagnostic marker in these populations. Future studies should be undertaken with the aim to link alterations in plasma and urinary β_2 M levels to its renal and non-renal determinants and also to the levels of albumin, which is regulated by the complex of the β_2 M–FcRn.

AUTHOR CONTRIBUTIONS

Each author was assigned a particular subtopic within this review. They researched the literature independently and provided their contribution to the corresponding author who integrated the manuscript. CA: conceived the paper idea, researched the biology and genetic disorders of β_2 M, animal toxicology studies, the kinetic model, and compiled the manuscript. SSC: researched the literature about non-renal diseases. Y-HN: researched the pediatric nephrology literature. M-ER: researched the literature of renal replacement therapies and the outcome studies in CKD. She also generated the first draft of the manuscript. KS: researched the literature on the use of the biomarker for glomerular and tubular disorders. PS: researched the literature on renal transplantation. AT critically reviewed the manuscript and drafted the response to author comments. All the authors were responsible for reviewing the final version of the article that was submitted to the journal.

REFERENCES

1. USRDS. *USRDS 2015 Annual Data Report: Atlas of Chronic Kidney Disease and End-Stage Renal Disease in the United States*. Bethesda, MD: National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases (2015).
2. Brosius FC, Hostetter TH, Kelepouris E, Mitsnefes MM, Moe SM, Moore MA, et al. Detection of chronic kidney disease in patients with or at increased risk of cardiovascular disease: a science advisory from the American Heart Association Kidney and Cardiovascular Disease Council; the Councils on High Blood Pressure Research, Cardiovascular Disease in the Young, and Epidemiology and Prevention; and the Quality of Care and Outcomes Research Interdisciplinary Working Group: developed in collaboration with the National Kidney Foundation. *Hypertension* (2006) 48:751–5. doi:10.1161/CIRCULATIONAHA.106.177321
3. Coca SG, Ismail-Beigi F, Haq N, Krumholz HM, Parikh CR. Role of intensive glucose control in development of renal end points in type 2 diabetes mellitus: systematic review and meta-analysis intensive glucose control in type 2 diabetes. *Arch Intern Med* (2012) 172:761–9. doi:10.1001/archinternmed.2011.2230
4. Estacio RO, Jeffers BW, Hiatt WR, Biggstaff SL, Gifford N, Schrier RW. The effect of nisoldipine as compared with enalapril on cardiovascular outcomes in patients with non-insulin-dependent diabetes and hypertension. *N Engl J Med* (1998) 338:645–52. doi:10.1056/NEJM199803053381003
5. ACCORD Study Group; Cushman WC, Evans GW, Byington RP, Goff DC, Grimm RH, et al. Effects of intensive blood-pressure control in type 2 diabetes mellitus. *N Engl J Med* (2010) 362:1575–85. doi:10.1056/NEJMoa1001286
6. Parving H-H, Brenner BM, McMurray JJV, de Zeeuw D, Haffner SM, Solomon SD, et al. Cardiorenal end points in a trial of aliskiren for type 2 diabetes. *N Engl J Med* (2012) 367:2204–13. doi:10.1056/NEJMoa1208799
7. ONTARGET Investigators, Yusuf S, Teo KK, Pogue J, Dyal L, Copland I, et al. Telmisartan, ramipril, or both in patients at high risk for vascular events. *N Engl J Med* (2008) 358:1547–59. doi:10.1056/NEJMoa0801317
8. Fried LF, Emanuele N, Zhang JH, Brophy M, Conner TA, Duckworth W, et al. Combined angiotensin inhibition for the treatment of diabetic nephropathy. *N Engl J Med* (2013) 369:1892–903. doi:10.1056/NEJMoa1303154
9. Jadoul M, Garbar C, van Ypersele de Strihou C. Pathological aspects of beta(2)-microglobulin amyloidosis. *Semin Dial* (2001) 14:86–9. doi:10.1046/j.1525-139x.2001.00037.x
10. Labriola L, Jadoul M. Dialysis-related amyloidosis: is it gone or should it be? *Semin Dial* (2017) 30(3):193–6. doi:10.1111/sdi.12590
11. Scarpioni R, Ricardi M, Albertazzi V, De Amicis S, Rastelli F, Zerbini L. Dialysis-related amyloidosis: challenges and solutions. *Int J Nephrol Renovasc Dis* (2016) 9:319–28. doi:10.2147/IJNRD.S84784
12. Copeland SD. Amyloidosis and its impact on patients with ESRD. *Nephrol Nurs J* (2006) 33:31–3.
13. Danesh F, Ho LT. Dialysis-related amyloidosis: history and clinical manifestations. *Semin Dial* (2001) 14:80–5. doi:10.1046/j.1525-139x.2001.00035.x
14. Gal R, Korzets A, Schwartz A, Rath-Wolfson L, Gafer U. Systemic distribution of beta 2-microglobulin-derived amyloidosis in patients who undergo long-term hemodialysis. Report of seven cases and review of the literature. *Arch Pathol Lab Med* (1994) 118:718–21.
15. Jadoul M, Garbar C, Noël H, Sennesael J, Vanholder R, Bernaert P, et al. Histological prevalence of β_2 -microglobulin amyloidosis in hemodialysis: a prospective post-mortem study. *Kidney Int* (1997) 51:1928–32. doi:10.1038/ki.1997.262
16. Jadoul M, Garbar C, Vanholder R, Sennesael J, Michel C, Robert A, et al. Prevalence of histological beta2-microglobulin amyloidosis in CAPD patients compared with hemodialysis patients. *Kidney Int* (1998) 54:956–9. doi:10.1046/j.1523-1755.1998.00064.x
17. Miyata T, Inagi R, Iida Y, Sato M, Yamada N, Oda O, et al. Involvement of beta 2-microglobulin modified with advanced glycation end products in the pathogenesis of hemodialysis-associated amyloidosis. Induction of human monocyte chemotaxis and macrophage secretion of tumor necrosis factor-alpha and interleukin-1. *J Clin Invest* (1994) 93:521–8. doi:10.1172/JCI117002
18. Yamamoto S, Gejyo F. Historical background and clinical treatment of dialysis-related amyloidosis. *Biochim Biophys Acta* (2005) 1753:4–10. doi:10.1016/j.bbapap.2005.09.006
19. Campistol JM. Dialysis-related amyloidosis after renal transplantation. *Semin Dial* (2001) 14:99–102. doi:10.1046/j.1525-139x.2001.00038.x
20. Grubb A, Simonsen O, Sturfelt G, Truedsson L, Thysel H. Serum concentration of cystatin C, factor D and beta 2-microglobulin as a measure of glomerular filtration rate. *Acta Med Scand* (1985) 218:499–503. doi:10.1111/j.0954-6820.1985.tb08880.x
21. Grubb A. Diagnostic value of analysis of cystatin C and protein HC in biological fluids. *Clin Nephrol* (1992) 38(Suppl 1):S20–7.
22. Norlund L, Fex G, Lanke J, Von Schenck H, Nilsson JE, Leksell H, et al. Reference intervals for the glomerular filtration rate and cell-proliferation markers: serum cystatin C and serum beta 2-microglobulin/cystatin C-ratio. *Scand J Clin Lab Invest* (1997) 57:463–70. doi:10.3109/00365519709084595
23. Corlin DB, Heegaard NHH. β_2 -microglobulin amyloidosis. *Subcell Biochem* (2012) 65:517–40. doi:10.1007/978-94-007-5416-4_19
24. Heegaard NHH. β_2 -microglobulin: from physiology to amyloidosis. *Amyloid* (2009) 16:151–73. doi:10.1080/13506120903151775
25. Stoppini M, Bellotti V. Systemic amyloidosis: lessons from β_2 -microglobulin. *J Biol Chem* (2015) 290:9951–8. doi:10.1074/jbc.R115.639799
26. Eichner T, Radford SE. Understanding the complex mechanisms of β_2 -microglobulin amyloid assembly. *FEBS J* (2011) 278:3868–83. doi:10.1111/j.1742-4658.2011.08186.x
27. Winchester JF, Salsberg JA, Levin NW. Beta-2 microglobulin in ESRD: an in-depth review. *Adv Ren Replace Ther* (2003) 10:279–309. doi:10.1053/j.arrt.2003.11.003
28. Berggård I, Bearn AG. Isolation and properties of a low molecular weight β_2 -globulin occurring in human biological fluids. *J Biol Chem* (1968) 243:4095–103.
29. Becker JW, Reeke GN. Three-dimensional structure of beta 2-microglobulin. *Proc Natl Acad Sci USA* (1985) 82:4225–9. doi:10.1073/pnas.82.12.4225
30. Iwata K, Matsuura T, Sakurai K, Nakagawa A, Goto Y. High-resolution crystal structure of beta2-microglobulin formed at pH 7.0. *J Biochem* (2007) 142:413–9. doi:10.1093/jb/mvm148
31. Cox J, Yewdell J, Eisenlohr L, Johnson P, Bennink J. Antigen presentation requires transport of MHC class I molecules from the endoplasmic reticulum. *Science* (1990) 247:715–8. doi:10.1126/science.2137259
32. Perarnau B, Siegrist C-A, Gillet A, Vincent C, Kimura S, Lemonnier FA. Beta 2-microglobulin restriction of antigen presentation. *Nature* (1990) 346:751–4. doi:10.1038/346751a0
33. Yamaguchi H. Association of MR1 protein, an MHC class I-related molecule, with β_2 -microglobulin. *Biochem Biophys Res Commun* (2002) 290:722–9. doi:10.1006/bbrc.2001.6277
34. Clements CS, Kjer-Nielsen L, McCluskey J, Rossjohn J. Structural studies on HLA-G: implications for ligand and receptor binding. *Hum Immunol* (2007) 68:220–6. doi:10.1016/j.humimm.2006.09.003
35. Gonen-Gross T, Achdout H, Arnon TI, Gazit R, Stern N, Horejsi V, et al. The CD85j/leukocyte inhibitory receptor-1 distinguishes between conformed and {beta}2-microglobulin-free HLA-G molecules. *J Immunol* (2005) 175:4866–74. doi:10.4049/jimmunol.175.8.4866
36. Roopenian DC, Akilesh S. FcRn: the neonatal Fc receptor comes of age. *Nat Rev Immunol* (2007) 7:715–25. doi:10.1038/nri2155
37. Pyzik M, Rath T, Lencer WI, Baker K, Blumberg RS. FcRn: the architect behind the immune and nonimmune functions of IgG and albumin. *J Immunol* (2015) 194:4595–603. doi:10.4049/jimmunol.1403014
38. Chaudhury C, Mehnaz S, Robinson JM, Hayton WL, Pearl DK, Roopenian DC, et al. The major histocompatibility complex-related Fc receptor for IgG (FcRn) binds albumin and prolongs its lifespan. *J Exp Med* (2003) 197:315–22. doi:10.1084/jem.20021829
39. Pratheek BM, Nayak TK, Sahoo SS, Mohanty PK, Chattopadhyay S, Chakraborty NG, et al. Mammalian non-classical major histocompatibility complex I and its receptors: important contexts of gene, evolution, and immunity. *Indian J Hum Genet* (2014) 20:129–41. doi:10.4103/0971-6866.142855
40. Nomura T, Huang W-C, Zhou HE, Josson S, Mimata H, Chung LWK. β_2 -Microglobulin-mediated signaling as a target for cancer therapy. *Anticancer Agents Med Chem* (2014) 14:343–52. doi:10.2174/18715206113139990092
41. Stapleton NM, Einarsdóttir HK, Stemerding AM, Vidarsson G. The multiple facets of FcRn in immunity. *Immunol Rev* (2015) 268:253–68. doi:10.1111/imr.12331
42. Martins JP, Kennedy PJ, Santos HA, Barrias C, Sarmiento B. A comprehensive review of the neonatal Fc receptor and its application in drug delivery. *Pharmacol Ther* (2016) 161:22–39. doi:10.1016/j.pharmthera.2016.03.007

43. Simister NE, Mostov KE. An Fc receptor structurally related to MHC class I antigens. *Nature* (1989) 337:184–7. doi:10.1038/337184a0
44. Israel EJ, Patel VK, Taylor SF, Marshak-Rothstein A, Simister NE. Requirement for a beta 2-microglobulin-associated Fc receptor for acquisition of maternal IgG by fetal and neonatal mice. *J Immunol* (1995) 154:6246–51.
45. Junghans RP, Anderson CL. The protection receptor for IgG catabolism is the beta2-microglobulin-containing neonatal intestinal transport receptor. *Proc Natl Acad Sci U S A* (1996) 93:5512–6. doi:10.1073/pnas.93.11.5512
46. Berryman M, Rodewald R. Beta 2-microglobulin co-distributes with the heavy chain of the intestinal IgG-Fc receptor throughout the transepithelial transport pathway of the neonatal rat. *J Cell Sci* (1995) 108:2347–60.
47. Ghetie V, Hubbard JG, Kim JK, Tsen MF, Lee Y, Ward ES. Abnormally short serum half-lives of IgG in beta 2-microglobulin-deficient mice. *Eur J Immunol* (1996) 26:690–6. doi:10.1002/eji.1830260327
48. Zhu X, Peng J, Raychowdhury R, Nakajima A, Lencer WI, Blumberg RS. The heavy chain of neonatal Fc receptor for IgG is sequestered in endoplasmic reticulum by forming oligomers in the absence of β_2 -microglobulin association. *Biochem J* (2002) 367:703–14. doi:10.1042/bj20020200
49. Kim J, Hayton WL, Robinson JM, Anderson CL. Kinetics of FcRn-mediated recycling of IgG and albumin in human: pathophysiology and therapeutic implications using a simplified mechanism-based model. *Clin Immunol* (2007) 122:146–55. doi:10.1016/j.clim.2006.09.001
50. Akilesh S, Huber TB, Wu H, Wang G, Hartleben B, Kopp JB, et al. Podocytes use FcRn to clear IgG from the glomerular basement membrane. *Proc Natl Acad Sci U S A* (2008) 105:967–72. doi:10.1073/pnas.0711515105
51. Dobrinskikh E, Okamura K, Kopp JB, Doctor RB, Blaine J. Human podocytes perform polarized, caveolae-dependent albumin endocytosis. *Am J Physiol Renal Physiol* (2014) 306:F941–51. doi:10.1152/ajprenal.00532.2013
52. Tenten V, Menzel S, Kunter U, Sicking E-M, van Roeyen CRC, Sanden SK, et al. Albumin is recycled from the primary urine by tubular transcytosis. *J Am Soc Nephrol* (2013) 24:1966–80. doi:10.1681/ASN.2013010018
53. Sarav M, Wang Y, Hack BK, Chang A, Jensen M, Bao L, et al. Renal FcRn reclaims albumin but facilitates elimination of IgG. *J Am Soc Nephrol* (2009) 20:1941–52. doi:10.1681/ASN.2008090976
54. Gan H, Feng S, Wu H, Sun Y, Hu R, Zhao Z, et al. Neonatal Fc receptor stimulation induces ubiquitin c-terminal hydrolase-1 overexpression in podocytes through activation of p38 mitogen-activated protein kinase. *Hum Pathol* (2012) 43:1482–90. doi:10.1016/j.humpath.2011.10.025
55. Kang S-W, Natarajan R, Shahed A, Nast CC, LaPage J, Mundel P, et al. Role of 12-lipoxygenase in the stimulation of p38 mitogen-activated protein kinase and collagen α_5 (IV) in experimental diabetic nephropathy and in glucose-stimulated podocytes. *J Am Soc Nephrol* (2003) 14:3178–87. doi:10.1097/01.ASN.0000099702.16315.DE
56. Koshikawa M, Mukoyama M, Mori K, Suganami T, Sawai K, Yoshioka T, et al. Role of p38 mitogen-activated protein kinase activation in podocyte injury and proteinuria in experimental nephrotic syndrome. *J Am Soc Nephrol* (2005) 16:2690–701. doi:10.1681/ASN.2004121084
57. Kavvadas P, Weis L, Abed AB, Feldman DL, Dussaule J-C, Chatziantoniou C. Renin inhibition reverses renal disease in transgenic mice by shifting the balance between profibrotic and antifibrotic agents novelty and significance. *Hypertension* (2013) 61:901–7. doi:10.1161/HYPERTENSIONAHA.111.00639
58. Kato Y, Mori K, Kasahara M, Osaki K, Ishii A, Mori KP, et al. Natriuretic peptide receptor guanylyl cyclase-A pathway counteracts glomerular injury evoked by aldosterone through p38 mitogen-activated protein kinase inhibition. *Sci Rep* (2017) 7:46624. doi:10.1038/srep46624
59. Abbate M, Zoja C, Remuzzi G. How does proteinuria cause progressive renal damage? *J Am Soc Nephrol* (2006) 17:2974–84. doi:10.1681/ASN.2006040377
60. Ruggenti P, Cravedi P, Remuzzi G. Mechanisms and treatment of CKD. *J Am Soc Nephrol* (2012) 23:1917–28. doi:10.1681/ASN.2012040390
61. Ruggenti P, Fassi A, Iliava AP, Bruno S, Iliev IP, Brusegan V, et al. Preventing microalbuminuria in type 2 diabetes. *N Engl J Med* (2004) 351:1941–51. doi:10.1056/NEJMoa042167
62. Okamura K, Dummer P, Kopp J, Qiu L, Levi M, Faubel S, et al. Endocytosis of albumin by podocytes elicits an inflammatory response and induces apoptotic cell death. *PLoS One* (2013) 8:e54817. doi:10.1371/journal.pone.0054817
63. Sedeek M, Nasrallah R, Touyz RM, Hébert RL. NADPH oxidases, reactive oxygen species, and the kidney: friend and foe. *J Am Soc Nephrol* (2013) 24:1512–8. doi:10.1681/ASN.2012111112
64. Wan C, Su H, Zhang C. Role of NADPH oxidase in metabolic disease-related renal injury: an update. *Oxid Med Cell Longev* (2016) 2016:e7813072. doi:10.1155/2016/7813072
65. Maekawa H, Inagi R. Stress signal network between hypoxia and ER stress in chronic kidney disease. *Front Physiol* (2017) 8:74. doi:10.3389/fphys.2017.00074
66. Small DM, Coombes JS, Bennett N, Johnson DW, Gobe GC. Oxidative stress, anti-oxidant therapies and chronic kidney disease. *Nephrology (Carlton)* (2012) 17:311–21. doi:10.1111/j.1440-1797.2012.01572.x
67. Jun M, Venkataraman V, Razavian M, Cooper B, Zoungas S, Ninomiya T, et al. Antioxidants for chronic kidney disease. *Cochrane Database Syst Rev* (2012) (10):CD008176. doi:10.1002/14651858.CD008176.pub2
68. Cao W, Zhou QG, Nie J, Wang GB, Liu Y, Zhou ZM, et al. Albumin overload activates intrarenal renin-angiotensin system through protein kinase C and NADPH oxidase-dependent pathway. *J Hypertens* (2011) 29:1411–21. doi:10.1097/HJH.0b013e32834786f0
69. Kinugasa S, Tojo A, Sakai T, Tsumura H, Takahashi M, Hirata Y, et al. Selective albuminuria via podocyte albumin transport in puromycin nephrotic rats is attenuated by an inhibitor of NADPH oxidase. *Kidney Int* (2011) 80:1328–38. doi:10.1038/ki.2011.282
70. Kang JH, Baik HW, Yoo S-M, Kim JH, Cheong HI, Park C-G, et al. Aliskiren regulates neonatal Fc receptor and IgG metabolism with attenuation of anti-GBM glomerulonephritis in mice. *Nephron* (2016) 134:272–82. doi:10.1159/000448789
71. Olaru F, Luo W, Suleiman H, John PLS, Ge L, Mezo AR, et al. Neonatal Fc receptor promotes immune complex-mediated glomerular disease. *J Am Soc Nephrol* (2014) 25:918–25. doi:10.1681/ASN.2013050498
72. Ichinose K, Ushigusa T, Nishino A, Nakashima Y, Suzuki T, Horai Y, et al. Lupus nephritis IgG induction of calcium/calmodulin-dependent protein kinase IV expression in podocytes and alteration of their function. *Arthritis Rheumatol* (2016) 68:944–52. doi:10.1002/art.39499
73. Amsellem S, Gburek J, Hamard G, Nielsen R, Willnow TE, Devuyst O, et al. Cubilin is essential for albumin reabsorption in the renal proximal tubule. *J Am Soc Nephrol* (2010) 21:1859–67. doi:10.1681/ASN.2010050492
74. Bern M, Sand KMK, Nilsen J, Sandlie I, Andersen JT. The role of albumin receptors in regulation of albumin homeostasis: implications for drug delivery. *J Control Release* (2015) 211:144–62. doi:10.1016/j.jconrel.2015.06.006
75. Dickson LE, Wagner MC, Sandoval RM, Molitoris BA. The proximal tubule and albuminuria: really! *J Am Soc Nephrol* (2014) 25:443–53. doi:10.1681/ASN.2013090950
76. Waldmann TA. Disorders of immunoglobulin metabolism. *N Engl J Med* (1969) 281:1170–7. doi:10.1056/NEJM196911202812107
77. Waldmann TA, Terry WD. Familial hypercatabolic hypoproteinemia. A disorder of endogenous catabolism of albumin and immunoglobulin. *J Clin Invest* (1990) 86:2093–8. doi:10.1172/JCI114947
78. Ardeniz Ö, Unger S, Onay H, Ammann S, Keck C, Cianga C, et al. β_2 -microglobulin deficiency causes a complex immunodeficiency of the innate and adaptive immune system. *J Allergy Clin Immunol* (2015) 136:392–401. doi:10.1016/j.jaci.2014.12.1937
79. Wani MA, Haynes LD, Kim J, Bronson CL, Chaudhury C, Mohanty S, et al. Familial hypercatabolic hypoproteinemia caused by deficiency of the neonatal Fc receptor, FcRn, due to a mutant beta2-microglobulin gene. *Proc Natl Acad Sci U S A* (2006) 103:5084–9. doi:10.1073/pnas.0600548103
80. de Sousa M, Reimão R, Lacerda R, Hugo P, Kaufmann SH, Porto G. Iron overload in beta 2-microglobulin-deficient mice. *Immunol Lett* (1994) 39:105–11. doi:10.1016/0165-2478(94)90094-9
81. Pietrangelo A. Hereditary hemochromatosis: pathogenesis, diagnosis, and treatment. *Gastroenterology* (2010) 139:e1–2. doi:10.1053/j.gastro.2010.06.013
82. Orr MT, Lanier LL. Natural killer cell education and tolerance. *Cell* (2010) 142:847–56. doi:10.1016/j.cell.2010.08.031
83. Santos M, Schilham MW, Rademakers LH, Marx JJ, de Sousa M, Clevers H. Defective iron homeostasis in beta 2-microglobulin knockout mice recapitulates hereditary hemochromatosis in man. *J Exp Med* (1996) 184:1975–85. doi:10.1084/jem.184.5.1975
84. Spriggs MK, Koller BH, Sato T, Morrissey PJ, Fanslow WC, Smithies O, et al. Beta 2-microglobulin-, CD8+ T-cell-deficient mice survive inoculation with high doses of vaccinia virus and exhibit altered IgG responses. *Proc Natl Acad Sci U S A* (1992) 89:6070–4. doi:10.1073/pnas.89.13.6070

85. Kim J, Bronson CL, Wani MA, Oberyszyn TM, Mohanty S, Chaudhury C, et al. Beta 2-microglobulin deficient mice catabolize IgG more rapidly than FcRn- alpha-chain deficient mice. *Exp Biol Med (Maywood)* (2008) 233:603–9. doi:10.3181/0710-RM-270
86. Schaible UE, Collins HL, Priem F, Kaufmann SHE. Correction of the iron overload defect in beta-2-microglobulin knockout mice by lactoferrin abolishes their increased susceptibility to tuberculosis. *J Exp Med* (2002) 196:1507–13. doi:10.1084/jem.20020897
87. Rodrigues P, Lopes C, Mascarenhas C, Arosio P, Porto G, De Sousa M. Comparative study between Hfe^{-/-} and beta2m^{-/-} mice: progression with age of iron status and liver pathology. *Int J Exp Pathol* (2006) 87:317–24. doi:10.1111/j.1365-2613.2006.00491.x
88. Muckenthaler MU, Rodrigues P, Macedo MG, Minana B, Brennan K, Cardoso EM, et al. Molecular analysis of iron overload in beta2-microglobulin-deficient mice. *Blood Cells Mol Dis* (2004) 33:125–31. doi:10.1016/j.bcmd.2004.05.003
89. Miranda CJ, Makui H, Andrews NC, Santos MM. Contributions of β_2 -microglobulin-dependent molecules and lymphocytes to iron regulation: insights from HfeRag1^{-/-} and β_2 mRag1^{-/-} double knock-out mice. *Blood* (2004) 103:2847–9. doi:10.1182/blood-2003-09-3300
90. Klingel K, Schnorr J-J, Sauter M, Szalay G, Kandolf R. beta2-microglobulin-associated regulation of interferon-gamma and virus-specific immunoglobulin G confer resistance against the development of chronic coxsackievirus myocarditis. *Am J Pathol* (2003) 162:1709–20. doi:10.1016/S0002-9440(10)64305-2
91. Sherwood ER, Lin CY, Tao W, Hartmann CA, Dujon JE, French AJ, et al. {beta}2 microglobulin knockout mice are resistant to lethal intraabdominal sepsis. *Am J Respir Crit Care Med* (2003) 167:1641–9. doi:10.1164/rccm.200208-9500C
92. Vikingsson A, Pederson K, Muller D. Altered kinetics of CD4⁺ T cell proliferation and interferon-gamma production in the absence of CD8⁺ T lymphocytes in virus-infected beta2-microglobulin-deficient mice. *Cell Immunol* (1996) 173:261–8. doi:10.1006/cimm.1996.0276
93. Tarleton RL, Koller BH, Latour A, Postan M. Susceptibility of beta 2-microglobulin-deficient mice to *Trypanosoma cruzi* infection. *Nature* (1992) 356:338–40. doi:10.1038/356338a0
94. D'Souza CD, Cooper AM, Frank AA, Ehlers S, Turner J, Bendelac A, et al. A novel nonclassical beta2-microglobulin-restricted mechanism influencing early lymphocyte accumulation and subsequent resistance to tuberculosis in the lung. *Am J Respir Cell Mol Biol* (2000) 23:188–93. doi:10.1165/ajrcmb.23.2.4063
95. Flynn JL, Goldstein MM, Triebold KJ, Koller B, Bloom BR. Major histocompatibility complex class I-restricted T cells are required for resistance to *Mycobacterium tuberculosis* infection. *Proc Natl Acad Sci U S A* (1992) 89:12013–7. doi:10.1073/pnas.89.24.12013
96. Hildeman D, Salvato M, Whitton JL, Muller D. Vaccination protects beta 2 microglobulin deficient mice from immune mediated mortality but not from persisting viral infection. *Vaccine* (1996) 14:1223–9. doi:10.1016/S0264-410X(96)00028-X
97. Cogen AL, Moore TA. Beta2-microglobulin-dependent bacterial clearance and survival during murine *Klebsiella pneumoniae* bacteremia. *Infect Immun* (2009) 77:360–6. doi:10.1128/IAI.00909-08
98. Ladel CH, Flesch IE, Arnoldi J, Kaufmann SH. Studies with MHC-deficient knock-out mice reveal impact of both MHC I- and MHC II-dependent T cell responses on *Listeria monocytogenes* infection. *J Immunol* (1994) 153:3116–22.
99. Tarakanova VL, Suarez F, Tibbetts SA, Jacoby MA, Weck KE, Hess JL, et al. Murine gammaherpesvirus 68 infection is associated with lymphoproliferative disease and lymphoma in BALB beta2 microglobulin-deficient mice. *J Virol* (2005) 79:14668–79. doi:10.1128/JVI.79.23.14668-14679.2005
100. Drake DR, Lukacher AE. Beta 2-microglobulin knockout mice are highly susceptible to polyoma virus tumorigenesis. *Virology* (1998) 252:275–84. doi:10.1006/viro.1998.9455
101. Mozes E, Kohn LD, Hakim F, Singer DS. Resistance of MHC class I-deficient mice to experimental systemic lupus erythematosus. *Science* (1993) 261:91–3. doi:10.1126/science.8316860
102. Mixter PF, Russell JQ, Durie FH, Budd RC. Decreased CD4-CD8- TCR-alpha beta + cells in lpr/lpr mice lacking beta 2-microglobulin. *J Immunol* (1995) 154:2063–74.
103. Chan OT, Paliwal V, McNiff JM, Park SH, Bendelac A, Shlomchik MJ. Deficiency in beta(2)-microglobulin, but not CD1, accelerates spontaneous lupus skin disease while inhibiting nephritis in MRL-Fas(lpr) mice: an example of disease regulation at the organ level. *J Immunol* (2001) 167:2985–90. doi:10.4049/jimmunol.167.5.2985
104. Christianson GJ, Blankenburg RL, Duffy TM, Panka D, Roths JB, Marshak-Rothstein A, et al. beta2-microglobulin dependence of the lupus-like autoimmune syndrome of MRL-lpr mice. *J Immunol* (1996) 156:4932–9.
105. Tsumiyama K, Hashiramoto A, Takimoto M, Tsuji-Kawahara S, Miyazawa M, Shiozawa S. IFN- γ -producing effector CD8 T lymphocytes cause immune glomerular injury by recognizing antigen presented as immune complex on target tissue. *J Immunol* (2013) 191:91–6. doi:10.4049/jimmunol.1203217
106. Barton JC, Edwards CQ, Acton RT. HFE gene: structure, function, mutations, and associated iron abnormalities. *Gene* (2015) 574:179–92. doi:10.1016/j.gene.2015.10.009
107. Bhatt L, Horgan CP, McCaffrey MW. Knockdown of beta2-microglobulin perturbs the subcellular distribution of HFE and hepcidin. *Biochem Biophys Res Commun* (2009) 378:727–31. doi:10.1016/j.bbrc.2008.11.118
108. Forman DT. Beta-2 microglobulin – an immunogenetic marker of inflammatory and malignant origin. *Ann Clin Lab Sci* (1982) 12:447–52.
109. Mayo Medical Laboratories. *FB2MU – Overview: Beta-2-Microglobulin, Urine* (2017). Available from: <http://www.mayomedicallaboratories.com/test-catalog/Overview/75161>
110. Mayo Medical Laboratories. *B2M – Overview: Beta-2-Microglobulin (Beta-2-M), Serum* (2017). Available from: <http://www.mayomedicallaboratories.com/test-catalog/Overview/9234>
111. Diamondstone LS, Tollerud DJ, Fuchs D, Wachter H, Brown LM, Maloney E, et al. Factors influencing serum neopterin and beta 2-microglobulin levels in a healthy diverse population. *J Clin Immunol* (1994) 14:368–74. doi:10.1007/BF01546321
112. Juraschek SP, Coresh J, Inker LA, Levey AS, Köttgen A, Foster MC, et al. Comparison of serum concentrations of β -trace protein, β_2 -microglobulin, cystatin C, and creatinine in the US population. *Clin J Am Soc Nephrol* (2013) 8:584–92. doi:10.2215/CJN.08700812
113. Swanson RA, Tracy RP, Katzmann JA, Wilson DM, Young DS. Beta 2-microglobulin determined by radioimmunoassay with monoclonal antibody. *Clin Chem* (1982) 28:2033–9.
114. Aulitzky WE, Grosse-Wilde H, Westhoff U, Tilg H, Aulitzky W, Gastl G, et al. Enhanced serum levels of soluble HLA class I molecules are induced by treatment with recombinant interferon-gamma (IFN-gamma). *Clin Exp Immunol* (1991) 86:236–9. doi:10.1111/j.1365-2249.1991.tb05802.x
115. Nachbaur K, Troppmair J, Bieling P, Kotlan B, König P, Huber C. Cytokines in the control of beta-2 microglobulin release. I. In vitro studies on various haemopoietic cells. *Immunobiology* (1988) 177:55–65. doi:10.1016/S0171-2985(88)80092-5
116. Nachbaur K, Troppmair J, Kotlan B, König P, Aulitzky W, Bieling P, et al. Cytokines in the control of beta-2 microglobulin release. II. In vivo studies with recombinant interferons and antigens. *Immunobiology* (1988) 177:66–75. doi:10.1016/S0171-2985(88)80092-5
117. Roumelioti ME, Nolin T, Unruh ML, Argyropoulos C. Revisiting the middle molecule hypothesis of uremic toxicity: a systematic review of beta 2 microglobulin population kinetics and large scale modeling of hemodialysis trials in silico. *PLoS One* (2016) 11:e0153157. doi:10.1371/journal.pone.0153157
118. Gooptu M, Staros E. *Beta2-Microglobulin: Reference Range, Interpretation, Collection and Panels* (2016). Available from: <http://emedicine.medscape.com/article/2086864-overview?pa=WQUfwdeUR9XVfjtQ2OZY-GA496bSjaFuIjwC25mpHSzFLBWwCtyhZPKTpV4%2BNyPC8S1v8zJyV73GUYw5rsbWA%3D%3D>
119. Daya NR, Voskertchian A, Schneider ALC, Ballew S, McAdams DeMarco M, Coresh J, et al. Kidney function and fracture risk: the atherosclerosis risk in communities (ARIC) study. *Am J Kidney Dis* (2016) 67:218–26. doi:10.1053/j.ajkd.2015.06.020
120. Vincent C, Pozet N, Revillard JP. Plasma beta 2 microglobulin turnover in renal insufficiency. *Acta Clin Belg Suppl* (1980) 35:2–12. doi:10.1080/2295337.1980.11718773
121. Wibell L, Evrin PE, Berggård I. Serum 2 -microglobulin in renal disease. *Nephron* (1973) 10:320–31. doi:10.1159/000180203

122. Shea PH, Maher JF, Horak E. Prediction of glomerular filtration rate by serum creatinine and beta 2-microglobulin. *Nephron* (1981) 29:30–5. doi:10.1159/000182234
123. Inker LA, Tighiouart H, Coresh J, Foster MC, Anderson AH, Beck GJ, et al. GFR estimation using β -trace protein and β_2 -microglobulin in CKD. *Am J Kidney Dis* (2016) 67:40–8. doi:10.1053/j.ajkd.2015.07.025
124. Aparicio SA, Mojiminiyi S, Kay JD, Shepstone BJ, de Ceulaer K, Serjeant GR. Measurement of glomerular filtration rate in homozygous sickle cell disease: a comparison of 51Cr-EDTA clearance, creatinine clearance, serum creatinine and beta 2 microglobulin. *J Clin Pathol* (1990) 43:370–2. doi:10.1136/jcp.43.5.370
125. Yun J-P, Suh C, Lee E, Chang JW, Yang WS, Park JS, et al. Comparison of serum beta 2-microglobulin and 24 hour urinary creatinine clearance as a prognostic factor in multiple myeloma. *J Korean Med Sci* (2006) 21:639–44. doi:10.3346/jkms.2006.21.4.639
126. Jovanović D, Krstivojević P, Obradović I, Durdević V, Dukanović L. Serum cystatin C and beta2-microglobulin as markers of glomerular filtration rate. *Ren Fail* (2003) 25:123–33. doi:10.1081/JDI-120017475
127. Aksun SA, Ozmen D, Ozmen B, Parildar Z, Mutaf I, Turgan N, et al. Beta2-microglobulin and cystatin C in type 2 diabetes: assessment of diabetic nephropathy. *Exp Clin Endocrinol Diabetes* (2004) 112:195–200. doi:10.1055/s-2004-817933
128. Bianchi C, Donadio C, Tramonti G, Consani C, Lorusso P, Rossi G. Reappraisal of serum beta2-microglobulin as marker of GFR. *Ren Fail* (2001) 23:419–29. doi:10.1081/JDI-100104725
129. Donadio C, Lucchesi A, Ardini M, Giordani R. Cystatin C, beta 2-microglobulin, and retinol-binding protein as indicators of glomerular filtration rate: comparison with plasma creatinine. *J Pharm Biomed Anal* (2001) 24:835–42. doi:10.1016/S0731-7085(00)00550-1
130. Donadio C, Lucchesi A, Ardini M, Donadio E, Giordani R. Serum levels of beta-trace protein and glomerular filtration rate – preliminary results. *J Pharm Biomed Anal* (2003) 32:1099–104. doi:10.1016/S0731-7085(03)00215-2
131. Fry AC, Singh DK, Chandna SM, Farrington K. Relative importance of residual renal function and convection in determining beta-2-microglobulin levels in high-flux haemodialysis and on-line haemodiafiltration. *Blood Purif* (2007) 25:295–302. doi:10.1159/000104870
132. Vilar E, Boltiador C, Wong J, Viljoen A, Machado A, Uthayakumar A, et al. Plasma levels of middle molecules to estimate residual kidney function in haemodialysis without urine collection. *PLoS One* (2015) 10:e0143813. doi:10.1371/journal.pone.0143813
133. Liu X, Foster MC, Tighiouart H, Anderson AH, Beck GJ, Contreras G, et al. Non-GFR determinants of low-molecular-weight serum protein filtration markers in CKD. *Am J Kidney Dis* (2016) 68:892–900. doi:10.1053/j.ajkd.2016.07.021
134. Parikh CR, Koyner JL. Biomarkers in acute and chronic kidney diseases. *Brenner and Rector's The Kidney*. Elsevier (2015). p. 926–55.
135. Filler G, Witt I, Priem F, Ehrlich JHH, Jung K. Are cystatin C and β_2 -microglobulin better markers than serum creatinine for prediction of a normal glomerular filtration rate in pediatric subjects? *Clin Chem* (1997) 43:1077–8.
136. Ikezumi Y, Honda M, Matsuyama T, Ishikura K, Hataya H, Yata N, et al. Establishment of a normal reference value for serum β_2 microglobulin in Japanese children: reevaluation of its clinical usefulness. *Clin Exp Nephrol* (2013) 17:99–105. doi:10.1007/s10157-012-0658-7
137. Ikezumi Y, Uemura O, Nagai T, Ishikura K, Ito S, Hataya H, et al. Beta-2 microglobulin-based equation for estimating glomerular filtration rates in Japanese children and adolescents. *Clin Exp Nephrol* (2015) 19:450–7. doi:10.1007/s10157-014-1015-9
138. Filler G, Priem F, Lepage N, Sinha P, Vollmer I, Clark H, et al. Beta-trace protein, cystatin C, beta(2)-microglobulin, and creatinine compared for detecting impaired glomerular filtration rates in children. *Clin Chem* (2002) 48:729–36.
139. Van Acker KJ, Vlietinck RF, Steels PM. Estimation of glomerular filtration rate from beta 2-microglobulin serum levels in children. *Int J Pediatr Nephrol* (1984) 5:59–62.
140. Filler G, Alvarez-Elias AC, Westreich KD, Huang S-HS, Lindsay RM. Can the new CKD-EPI BTP-B2M formula be applied in children? *Pediatr Nephrol* (2016) 31:2175–7. doi:10.1007/s00467-016-3493-3
141. Peterson PA, Evrin P-E, Berggård I. Differentiation of glomerular, tubular, and normal proteinuria: determinations of urinary excretion of β_2 -microglobulin, albumin, and total protein. *J Clin Invest* (1969) 48:1189–98. doi:10.1172/JCI106083
142. Portman RJ, Kissane JM, Robson AM. Use of beta 2 microglobulin to diagnose tubulo-interstitial renal lesions in children. *Kidney Int* (1986) 30:91–8. doi:10.1038/ki.1986.156
143. Christensen EI, Birn H, Storm T, Weyer K, Nielsen R. Endocytic receptors in the renal proximal tubule. *Physiology (Bethesda)* (2012) 27:223–36. doi:10.1152/physiol.00022.2012
144. Nielsen R, Christensen EI, Birn H. Megalin and cubilin in proximal tubule protein reabsorption: from experimental models to human disease. *Kidney Int* (2016) 89:58–67. doi:10.1016/j.kint.2015.11.007
145. Orlando RA, Rader K, Authier F, Yamazaki H, Posner BI, Bergeron JJ, et al. Megalin is an endocytic receptor for insulin. *J Am Soc Nephrol* (1998) 9:1759–66.
146. Leheste J-R, Rolinski B, Vorum H, Hilpert J, Nykjaer A, Jacobsen C, et al. Megalin knockout mice as an animal model of low molecular weight proteinuria. *Am J Pathol* (1999) 155:1361–70. doi:10.1016/S0002-9440(10)65238-8
147. Storm T, Tranebjaerg L, Frykholm C, Birn H, Verroust PJ, Nevés T, et al. Renal phenotypic investigations of megalin-deficient patients: novel insights into tubular proteinuria and albumin filtration. *Nephrol Dial Transplant* (2013) 28:585–91. doi:10.1093/ndt/gfs462
148. Kantarci S, Al-Gazali L, Hill RS, Donnai D, Black GCM, Bieth E, et al. Mutations in LRP2, which encodes the multiligand receptor megalin, cause Donnai-Barrow and facio-oculo-acoustico-renal syndromes. *Nat Genet* (2007) 39:957–9. doi:10.1038/ng2063
149. Storm T, Emma F, Verroust PJ, Hertz JM, Nielsen R, Christensen EI. A patient with cubilin deficiency. *N Engl J Med* (2011) 364:89–91. doi:10.1056/NEJMc1009804
150. Christensen EI, Birn H. Megalin and cubilin: multifunctional endocytic receptors. *Nat Rev Mol Cell Biol* (2002) 3:258–68. doi:10.1038/nrm778
151. Sundin DP, Cohen M, Dahl R, Falk S, Molitoris BA. Characterization of the beta 2-microglobulin endocytic pathway in rat proximal tubule cells. *Am J Physiol* (1994) 267:F380–9.
152. Cohen M, Sundin DP, Dahl R, Molitoris BA. Convergence of apical and basolateral endocytic pathways for beta 2-microglobulin in LLC-PK1 cells. *Am J Physiol* (1995) 268:F829–38.
153. Kobayashi N, Suzuki Y, Tsuge T, Okumura K, Ra C, Tomino Y. FcRn-mediated transcytosis of immunoglobulin G in human renal proximal tubular epithelial cells. *Am J Physiol Renal Physiol* (2002) 282:F358–65. doi:10.1152/ajprenal.0164.2001
154. Gauthier C, Nguyen-Simonnet H, Vincent C, Revillard JP, Pellet MV. Renal tubular absorption of beta 2 microglobulin. *Kidney Int* (1984) 26:170–5. doi:10.1038/ki.1984.151
155. Bernard A, Amor AO, Viau C, Lauwerys R. The renal uptake of proteins: a nonselective process in conscious rats. *Kidney Int* (1988) 34:175–85. doi:10.1038/ki.1988.163
156. Thielemans N, Lauwerys R, Bernard A. Competition between albumin and low-molecular-weight proteins for renal tubular uptake in experimental nephropathies. *Nephron* (1994) 66:453–8. doi:10.1159/000187863
157. Bernard A, Viau C, Ouled A, Lauwerys R. Competition between low- and high-molecular-weight proteins for renal tubular uptake. *Nephron* (1987) 45:115–8. doi:10.1159/000184090
158. Thakkar H, Lowe PA, Price CP, Newman DJ. Measurement of the kinetics of protein uptake by proximal tubular cells using an optical biosensor. *Kidney Int* (1998) 54:1197–205. doi:10.1046/j.1523-1755.1998.00098.x
159. Karlsson FA, Hardell LI, Hellsing K. A prospective study of urinary proteins in early infancy. *Acta Paediatr Scand* (1979) 68:663–7. doi:10.1111/j.1651-2227.1979.tb18435.x
160. Dieterle F, Perentes E, Cordier A, Roth DR, Verdes P, Grenet O, et al. Urinary clusterin, cystatin C, β_2 -microglobulin and total protein as markers to detect drug-induced kidney injury. *Nat Biotechnol* (2010) 28:463–9. doi:10.1038/nbt.1622
161. Kuwata K, Nakamura I, Ide M, Sato H, Nishikawa S, Tanaka M. Comparison of changes in urinary and blood levels of biomarkers associated with proximal tubular injury in rat models. *J Toxicol Pathol* (2015) 28:151–64. doi:10.1293/tox.2014-0039
162. Vlasakova K, Erdos Z, Troth SP, McNulty K, Chapeau-Campredon V, Mokrzycki N, et al. Evaluation of the relative performance of 12 urinary

- biomarkers for renal safety across 22 rat sensitivity and specificity studies. *Toxicol Sci* (2014) 138:3–20. doi:10.1093/toxsci/kft330
163. Shin JR, Kim SM, Yoo JS, Park JY, Kim SK, Cho JH, et al. Urinary excretion of β_2 -microglobulin as a prognostic marker in immunoglobulin A nephropathy. *Korean J Intern Med* (2014) 29:334–40. doi:10.3904/kjim.2014.29.3.334
 164. Choe J-Y, Park S-H, Kim S-K. Urine β_2 -microglobulin is associated with clinical disease activity and renal involvement in female patients with systemic lupus erythematosus. *Lupus* (2014) 23:1486–93. doi:10.1177/0961203314547797
 165. Bagshaw SM, Langenberg C, Haase M, Wan L, May CN, Bellomo R. Urinary biomarkers in septic acute kidney injury. *Intensive Care Med* (2007) 33:1285–96. doi:10.1007/s00134-007-0656-5
 166. Zeng X, Hossain D, Bostwick DG, Herrera GA, Zhang PL. Urinary beta2-microglobulin is a good indicator of proximal tubule injury: a correlative study with renal biopsies. *J Biomarkers* (2014) 2014:e492838. doi:10.1155/2014/492838
 167. Kaye WA, Griffiths WC, Camara PD, Trebbin WM, Solomon RJ, Diamond I. The significance of beta-2 microglobulinuria associated with gentamicin therapy. *Ann Clin Lab Sci* (1981) 11:530–7.
 168. Blumsohn A, Morris BW, Griffiths H, Ramsey CF. Stability of beta 2-microglobulin and retinol binding protein at different values of pH and temperature in normal and pathological urine. *Clin Chim Acta* (1991) 195:133–7. doi:10.1016/0009-8981(91)90133-W
 169. Donaldson MD, Chambers RE, Woolridge MW, Whicher JT. Stability of alpha 1-microglobulin, beta 2-microglobulin and retinol binding protein in urine. *Clin Chim Acta* (1989) 179:73–7. doi:10.1016/0009-8981(89)90024-7
 170. Bastable MD. Beta 2 microglobulin in urine: not suitable for assessing renal tubular function. *Clin Chem* (1983) 29:996–7.
 171. Bernard AM, Vyskocil AA, Mahieu P, Lauwerys RR. Assessment of urinary retinol-binding protein as an index of proximal tubular injury. *Clin Chem* (1987) 33:775–9.
 172. Hu J, Li M, Han T, Chen J, Ye L, Wang Q, et al. Benchmark dose estimation for cadmium-induced renal tubular damage among environmental cadmium-exposed women aged 35–54 years in two counties of China. *PLoS One* (2014) 9:e115794. doi:10.1371/journal.pone.0115794
 173. Ke S, Cheng X-Y, Zhang J-Y, Jia W-J, Li H, Luo H-F, et al. Estimation of the benchmark dose of urinary cadmium as the reference level for renal dysfunction: a large sample study in five cadmium polluted areas in China. *BMC Public Health* (2015) 15:656. doi:10.1186/s12889-015-2021-x
 174. Kim Y-D, Yim D-H, Eom S-Y, Moon S-I, Park C-H, Kim G-B, et al. Temporal changes in urinary levels of cadmium, N-acetyl- β -D-glucosaminidase and β_2 -microglobulin in individuals in a cadmium-contaminated area. *Environ Toxicol Pharmacol* (2015) 39:35–41. doi:10.1016/j.etap.2014.10.016
 175. Nishijo M, Suwazono Y, Ruangyuttikarn W, Nambunmee K, Swaddiwudhipong W, Nogawa K, et al. Risk assessment for Thai population: benchmark dose of urinary and blood cadmium levels for renal effects by hybrid approach of inhabitants living in polluted and non-polluted areas in Thailand. *BMC Public Health* (2014) 14:702. doi:10.1186/1471-2458-14-702
 176. Rybakowski JK, Abramowicz M, Chłopocka-Wozniak M, Czekalski S. Novel markers of kidney injury in bipolar patients on long-term lithium treatment. *Hum Psychopharmacol* (2013) 28:615–8. doi:10.1002/hup.2362
 177. Nishijima T, Gatanaga H, Komatsu H, Tsukada K, Shimbo T, Aoki T, et al. Renal function declines more in tenofovir- than abacavir-based antiretroviral therapy in low-body weight treatment-naive patients with HIV infection. *PLoS One* (2012) 7:e29977. doi:10.1371/journal.pone.0029977
 178. Oboho I, Abraham A, Benning L, Anastos K, Sharma A, Young M, et al. Tenofovir use and urinary biomarkers among HIV-infected women in the Women's Interagency HIV Study (WIHS). *J Acquir Immune Defic Syndr* (2013) 62:388–95. doi:10.1097/QAI.0b013e31828175c9
 179. Gatanaga H, Tachikawa N, Kikuchi Y, Teruya K, Genka I, Honda M, et al. Urinary beta2-microglobulin as a possible sensitive marker for renal injury caused by tenofovir disoproxil fumarate. *AIDS Res Hum Retroviruses* (2006) 22:744–8. doi:10.1089/aid.2006.22.744
 180. Joyce E, Glasner P, Ranganathan S, Swiatecka-Urban A. Tubulointerstitial nephritis: diagnosis, treatment, and monitoring. *Pediatr Nephrol* (2017) 32:577–87. doi:10.1007/s00467-016-3394-5
 181. Tomlinson PA, Dalton RN, Hartley B, Haycock GB, Chantler C. Low molecular weight protein excretion in glomerular disease: a comparative analysis. *Pediatr Nephrol* (1997) 11:285–90. doi:10.1007/s004670050278
 182. Hettinga YM, Scheerlinck LME, Lilien MR, Rothova A, de Boer JH. The value of measuring urinary β_2 -microglobulin and serum creatinine for detecting tubulointerstitial nephritis and uveitis syndrome in young patients with uveitis. *JAMA Ophthalmol* (2015) 133:140–5. doi:10.1001/jamaophthol.2014.4301
 183. Badr M, El Koumi MA, Ali YF, El-Morshedy S, Almonem NA, Hassan T, et al. Renal tubular dysfunction in children with sickle cell haemoglobinopathy. *Nephrology (Carlton)* (2013) 18:299–303. doi:10.1111/nep.12040
 184. Unal S, Kotan C, Delibas A, Oztas Y, Cystatin C. Beta2 microglobulin, N-acetyl-beta-D-glucosaminidase, retinol-binding protein, and endothelin 1 levels in the evaluation of sickle cell disease nephropathy. *Pediatr Hematol Oncol* (2015) 32:250–7. doi:10.3109/08880018.2013.810317
 185. Stefanowicz J, Owczuk R, Aleksandrowicz E, Owczarzak A, Kurylak A, Adamkiewicz-Drożyńska E, et al. Renal function and low-molecular-weight proteins (cystatin C, β_2 -microglobulin, neutrophil gelatinase-associated lipocalin) in child and young adult cancer survivors. *J Pediatr Hematol Oncol* (2012) 34:461–6. doi:10.1097/MPH.0b013e318257fd89
 186. Fang J, Luan J, Zhu G, Qi C, Wang D. Detection of PCT and urinary β_2 -MG enhances the accuracy for localization diagnosing pediatric urinary tract infection. *J Clin Lab Anal* (2016). doi:10.1002/jcla.22088
 187. Korpala-Szczyrska N, Uszycia-Karcz M, Mierzewski P. Urinary excretion of beta-2-microglobulin in children with urinary tract infections. *Pediatr Res* (1989) 26:505–505. doi:10.1203/00006450-198911000-00035
 188. Salvaggio E, Menonna NM, Ricci R, Ferrara P, Nardini F. [Beta 2 microglobulin in the diagnosis of reflux nephropathy in childhood]. *Pediatr Med Chir* (1988) 10:83–8.
 189. Madsen MG, Nørregaard R, Palmfeldt J, Olsen LH, Frøkiaer J, Jørgensen TM. Urinary NGAL, cystatin C, β_2 -microglobulin, and osteopontin significance in hydronephrotic children. *Pediatr Nephrol* (2012) 27:2099–106. doi:10.1007/s00467-012-2217-6
 190. El-Frargy MS, El-Refaey AM, Eid R, Hussien MA. Serum cystatin-C and BETA 2-microglobulin as accurate markers in the early diagnosis of kidney injury in neonates: a single center study. *Saudi J Kidney Dis Transpl* (2015) 26:712–7. doi:10.4103/1319-2442.160151
 191. Herrero-Morin JD, Málaga S, Fernández N, Rey C, Diéguez MA, Solís G, et al. Cystatin C and beta2-microglobulin: markers of glomerular filtration in critically ill children. *Crit Care* (2007) 11:R59. doi:10.1186/cc5923
 192. Du Y, Zappitelli M, Mian A, Bennett M, Ma Q, Devarajan P, et al. Urinary biomarkers to detect acute kidney injury in the pediatric emergency center. *Pediatr Nephrol* (2011) 26:267–74. doi:10.1007/s00467-010-1673-0
 193. Askenazi DJ, Koralkar R, Levitan EB, Goldstein SL, Devarajan P, Khandrika S, et al. Baseline values of candidate urine acute kidney injury biomarkers vary by gestational age in premature infants. *Pediatr Res* (2011) 70:302–6. doi:10.1203/PDR.0b013e3182275164
 194. DeFreitas MJ, Seeherunvong W, Katsoufis CP, RamachandraRao S, Duara S, Yasin S, et al. Longitudinal patterns of urine biomarkers in infants across gestational ages. *Pediatr Nephrol* (2016) 31:1179–88. doi:10.1007/s00467-016-3327-3
 195. Liabeuf S, Lenglet A, Desjardins L, Neiryck N, Glorieux G, Lemke HD, et al. Plasma beta-2 microglobulin is associated with cardiovascular disease in uremic patients. *Kidney Int* (2012) 82:1297–303. doi:10.1038/ki.2012.301
 196. Foster MC, Coresh J, Hsu C-Y, Xie D, Levey AS, Nelson RG, et al. Serum β -trace protein and β_2 -microglobulin as predictors of ESRD, mortality, and cardiovascular disease in adults with CKD in the chronic renal insufficiency cohort (CRIC) study. *Am J Kidney Dis* (2016) 68(1):68–76. doi:10.1053/j.ajkd.2016.01.015
 197. Foster MC, Inker LA, Hsu C-Y, Eckfeldt JH, Levey AS, Pavkov ME, et al. Filtration markers as predictors of ESRD and mortality in Southwestern American Indians with type 2 diabetes. *Am J Kidney Dis* (2015) 66:75–83. doi:10.1053/j.ajkd.2015.01.013
 198. Wu H-C, Lee L-C, Wang W-J. Associations among serum beta 2 microglobulin, malnutrition, inflammation, and advanced cardiovascular event in patients with chronic kidney disease. *J Clin Lab Anal* (2017) 31(3):e22056. doi:10.1002/jcla.22056
 199. Inker LA, Coresh J, Sang Y, Hsu C-Y, Foster MC, Eckfeldt JH, et al. Filtration markers as predictors of ESRD and mortality: individual participant data meta-analysis. *Clin J Am Soc Nephrol* (2017) 12:69–78. doi:10.2215/CJN.03660316

200. Astor BC, Shafi T, Hoogeveen RC, Matsushita K, Ballantyne CM, Inker LA, et al. Novel markers of kidney function as predictors of ESRD, cardiovascular disease, and mortality in the general population. *Am J Kidney Dis* (2012) 59:653–62. doi:10.1053/j.ajkd.2011.11.042
201. Suzuki T, Agarwal SK, Deo R, Sotoodehnia N, Grams ME, Selvin E, et al. Kidney function and sudden cardiac death in the community: the Atherosclerosis Risk in Communities (ARIC) Study. *Am Heart J* (2016) 180:46–53. doi:10.1016/j.ahj.2016.07.004
202. Foster MC, Inker LA, Levey AS, Selvin E, Eckfeldt J, Juraschek SP, et al. Novel filtration markers as predictors of all-cause and cardiovascular mortality in US adults. *Am J Kidney Dis* (2013) 62(1):42–51. doi:10.1053/j.ajkd.2013.01.016
203. Wong J, Sridharan S, Berdeprado J, Vilar E, Viljoen A, Wellsted D, et al. Predicting residual kidney function in hemodialysis patients using serum β -trace protein and β_2 -microglobulin. *Kidney Int* (2016) 89:1090–8. doi:10.1016/j.kint.2015.12.042
204. Scribner BH, Babb AL. Evidence for toxins of “middle” molecular weight. *Kidney Int Suppl* (1975) 3:349–51.
205. Babb AL, Ahmad S, Bergström J, Scribner BH. The middle molecule hypothesis in perspective. *Am J Kidney Dis* (1981) 1:46–50. doi:10.1016/S0272-6386(81)80011-X
206. Leypoldt JK, Holmes CJ, Rutherford P. Clearance of middle molecules during haemodialysis and haemodiafiltration: new insights. *Nephrol Dial Transplant* (2012) 27:4245–7. doi:10.1093/ndt/gfs475
207. Thomas G, Jaber BL. Innovation in the treatment of uremia: proceedings from the Cleveland clinic workshop: convective therapies for removal of middle molecular weight uremic toxins in end-stage renal disease: a review of the evidence. *Semin Dial* (2009) 22:610–4. doi:10.1111/j.1525-139X.2009.00665.x
208. Panichi V, De Ferrari G, Saffiotti S, Sidoti A, Biagioli M, Bianchi S, et al. Comparison of on-line HDF modes automated TMP control vs. volume control on achieved convective volume and middle molecule clearance. *Nephrol Dial Transplant* (2012) 27:ii205–6. doi:10.1093/ndt/gfs224
209. Tattersall J. Clearance of beta-2-microglobulin and middle molecules in haemodiafiltration. *Contrib Nephrol* (2007) 158:201–9. doi:10.1159/000107251
210. Okuno S, Ishimura E, Kohno K, Fujino-Katoh Y, Maeno Y, Yamakawa T, et al. Serum beta2-microglobulin level is a significant predictor of mortality in maintenance haemodialysis patients. *Nephrol Dial Transplant* (2009) 24:571–7. doi:10.1093/ndt/gfn521
211. Kim KM, Kim S-S, Kim H, Koo T, Im EY, Kim SB. Higher serum beta2-microglobulin levels are associated with better survival in chronic hemodialysis patients: a reverse epidemiology. *Clin Nephrol* (2011) 75:458–65. doi:10.5414/CNP75458
212. Carreno MP, Rousseau Y, Poignet JL, Jahns G, Cholley B, Kazatchkine MD, et al. Dissociation between beta-2 microglobulin and IL-1 production in hemodialyzed patients. *Nephrol Dial Transplant* (1997) 12:2365–74. doi:10.1093/ndt/12.11.2365
213. Caruana RJ, Lobel SA, Leffell MS, Campbell H, Cheek PL. Tumor necrosis factor, interleukin-1 and beta 2-microglobulin levels in chronic hemodialysis patients. *Int J Artif Organs* (1990) 13:794–8.
214. Memoli B, Libetta C, Rampino T, Canton AD, Conte G, Scala G, et al. Hemodialysis related induction of interleukin-6 production by peripheral blood mononuclear cells. *Kidney Int* (1992) 42:320–6. doi:10.1038/ki.1992.292
215. Pertosa G, Marfella C, Tarantino EA, Di Cillo M, Manno C, Russo R, et al. Involvement of peripheral blood monocytes in haemodialysis: in vivo induction of tumour necrosis factor alpha, interleukin 6 and beta 2-microglobulin. *Nephrol Dial Transplant* (1991) 6(Suppl 2):18–23.
216. Akchurin OM, Kaskel F. Update on inflammation in chronic kidney disease. *Blood Purif* (2015) 39:84–92. doi:10.1159/000368940
217. Topçiu-Shufta V, Miftari R, Haxhibeqiri V, Haxhibeqiri S. Association of beta-2 microglobulin with inflammation and dyslipidemia in high-flux membrane hemodialysis patients. *Med Arch* (2016) 70:348–50. doi:10.5455/medarh.2016.70.348-350
218. Lonnemann G, Novick D, Rubinstein M, Passlick-Deetjen J, Lang D, Dinarello CA. A switch to high-flux helixone membranes reverses suppressed interferon-gamma production in patients on low-flux dialysis. *Blood Purif* (2003) 21:225–31. doi:10.1159/000070694
219. Lonnemann G, Bahlmann FH, Freise J, Hertel B, Dinarello CA. Low-flux hemodialysis suppresses interferon-gamma production: the possible role of beta2-microglobulin. *Clin Nephrol* (2009) 72:170–6. doi:10.5414/CNP72170
220. Palmer SC, Rabindranath KS, Craig JC, Roderick PJ, Locatelli F, Strippoli GFM. High-flux versus low-flux membranes for end-stage kidney disease. *Cochrane Database Syst Rev* (2012) (9):CD005016. doi:10.1002/14651858.CD005016.pub2
221. Eknoyan G, Beck GJ, Cheung AK, Daugirdas JT, Greene T, Kusek JW, et al. Effect of dialysis dose and membrane flux in maintenance hemodialysis. *N Engl J Med* (2002) 347:2010–9. doi:10.1056/NEJMoa021583
222. Cheung AK, Levin NW, Greene T, Agodoa L, Bailey J, Beck G, et al. Effects of high-flux hemodialysis on clinical outcomes: results of the HEMO study. *J Am Soc Nephrol* (2003) 14:3251–63. doi:10.1097/01.ASN.0000096373.13406.94
223. Delmez JA, Yan G, Bailey J, Beck GJ, Beddhu S, Cheung AK, et al. Cerebrovascular disease in maintenance hemodialysis patients: results of the HEMO Study. *Am J Kidney Dis* (2006) 47:131–8. doi:10.1053/j.ajkd.2005.09.031
224. Cheung AK, Rocco MV, Yan G, Leypoldt JK, Levin NW, Greene T, et al. Serum β -2 microglobulin levels predict mortality in dialysis patients: results of the HEMO study. *J Am Soc Nephrol* (2006) 17:546–55. doi:10.1681/ASN.2005020132
225. Cheung AK, Greene T, Leypoldt JK, Yan G, Allon M, Delmez J, et al. Association between serum β_2 -microglobulin level and infectious mortality in hemodialysis patients. *Clin J Am Soc Nephrol* (2008) 3:69–77. doi:10.2215/CJN.02340607
226. Locatelli F, Gaulty A, Czekalski S, Hannedouche T, Jacobson SH, Loureiro A, et al. The MPO Study: just a European HEMO Study or something very different? *Blood Purif* (2008) 26:100–4. doi:10.1159/000110574
227. Locatelli F, Martin-Malo A, Hannedouche T, Loureiro A, Papadimitriou M, Wizemann V, et al. Effect of membrane permeability on survival of hemodialysis patients. *J Am Soc Nephrol* (2009) 20:645–54. doi:10.1681/ASN.2008060590
228. Locatelli F, Cavalli A, Manzoni C, Pontoriero G. The membrane permeability outcome study. *Contrib Nephrol* (2011) 175:81–92. doi:10.1159/000333816
229. Argyropoulos C, Roumelioti M-E, Sattar A, Kellum JA, Weissfeld L, Unruh ML. Dialyzer reuse and outcomes of high flux dialysis. *PLoS One* (2015) 10:e0129575. doi:10.1371/journal.pone.0129575
230. Chazot C, Kirchgessner J, Pham J, Vo-Van C, Lorriaux C, Hurot J-M, et al. Effect of membrane permeability on cardiovascular risk factors and β_2 m plasma levels in patients on long-term haemodialysis: a randomised crossover trial. *Nephron* (2015) 129:269–75. doi:10.1159/000380767
231. Locatelli F, Vecchio LD, Milia VL. Haemodiafiltration at higher volumes and patient survival. *Contrib Nephrol* (2016) 189:1–8. doi:10.1159/000450632
232. Mostovaya IM, Blankestijn PJ, Bots ML, Covic A, Davenport A, Grooteman MPC, et al. Clinical evidence on hemodiafiltration: a systematic review and a meta-analysis. *Semin Dial* (2014) 27:119–27. doi:10.1111/sdi.12200
233. Nistor I, Palmer SC, Craig JC, Saglimbene V, Vecchio M, Covic A, et al. Convective versus diffusive dialysis therapies for chronic kidney failure: an updated systematic review of randomized controlled trials. *Am J Kidney Dis* (2014) 63:954–67. doi:10.1053/j.ajkd.2013.12.004
234. Nistor I, Palmer SC, Craig JC, Saglimbene V, Vecchio M, Covic A, et al. Haemodiafiltration, haemofiltration and haemodialysis for end-stage kidney disease. *Cochrane Database Syst Rev* (2015) (5):CD006258. doi:10.1002/14651858.CD006258.pub2
235. Susantitaphong P, Siribamrungwong M, Jaber BL. Convective therapies versus low-flux hemodialysis for chronic kidney failure: a meta-analysis of randomized controlled trials. *Nephrol Dial Transplant* (2013) 28:2859–74. doi:10.1093/ndt/gft396
236. Wang AY, Ninomiya T, Al-Kahwa A, Perkovic V, Gallagher MP, Hawley C, et al. Effect of hemodiafiltration or hemofiltration compared with hemodialysis on mortality and cardiovascular disease in chronic kidney failure: a systematic review and meta-analysis of randomized trials. *Am J Kidney Dis* (2014) 63:968–78. doi:10.1053/j.ajkd.2014.01.435
237. Canaud B, Barbieri C, Marcelli D, Bellocchio F, Bowry S, Mari F, et al. Optimal convection volume for improving patient outcomes in an international incident dialysis cohort treated with online hemodiafiltration. *Kidney Int* (2015) 88:1108–16. doi:10.1038/ki.2015.139

238. Canaud B, Bragg-Gresham JL, Marshall MR, Desmeules S, Gillespie BW, Depner T, et al. Mortality risk for patients receiving hemodiafiltration versus hemodialysis: European results from the DOPPS. *Kidney Int* (2006) 69:2087–93. doi:10.1038/sj.ki.5000447
239. Grooteman MPC, van den Dorpel MA, Bots ML, Penne EL, van der Weerd NC, Mazairac AHA, et al. Effect of online hemodiafiltration on all-cause mortality and cardiovascular outcomes. *J Am Soc Nephrol* (2012) 23:1087–96. doi:10.1681/ASN.2011121140
240. Ok E, Asci G, Toz H, Ok ES, Kircelli F, Yilmaz M, et al. Mortality and cardiovascular events in online haemodiafiltration (OL-HDF) compared with high-flux dialysis: results from the Turkish OL-HDF Study. *Nephrol Dial Transplant* (2013) 28:192–202. doi:10.1093/ndt/gfs407
241. Maduell F, Moreso F, Pons M, Ramos R, Mora-Macia J, Carreras J, et al. High-efficiency postdilution online hemodiafiltration reduces all-cause mortality in hemodialysis patients. *J Am Soc Nephrol* (2013) 24:487–97. doi:10.1681/ASN.2012080875
242. Peters SAE, Bots ML, Canaud B, Davenport A, Grooteman MPC, Kircelli F, et al. Haemodiafiltration and mortality in end-stage kidney disease patients: a pooled individual participant data analysis from four randomized controlled trials. *Nephrol Dial Transplant* (2016) 31:978–84. doi:10.1093/ndt/gfv349
243. Santoro A, Mancini E, Bolzani R, Boggi R, Cagnoli L, Francioso A, et al. The effect of on-line high-flux hemofiltration versus low-flux hemodialysis on mortality in chronic kidney failure: a small randomized controlled trial. *Am J Kidney Dis* (2008) 52:507–18. doi:10.1053/j.ajkd.2008.05.011
244. Koh ES, Lee K, Kim SH, Kim YO, Jin DC, Song HC, et al. Serum β₂-microglobulin predicts mortality in peritoneal dialysis patients: a prospective cohort study. *Am J Nephrol* (2015) 42:91–8. doi:10.1159/000439060
245. Liu X, Dai C. Advances in understanding and management of residual renal function in patients with chronic kidney disease. *Kidney Dis (Basel)* (2017) 2:187–96. doi:10.1159/000449029
246. Mathew AT, Fishbane S, Obi Y, Kalantar-Zadeh K. Preservation of residual kidney function in hemodialysis patients: reviving an old concept. *Kidney Int* (2016) 90:262–71. doi:10.1016/j.kint.2016.02.037
247. Termorshuizen F, Korevaar JC, Dekker FW, van Manen JG, Boeschoten EW, Krediet RT, et al. The relative importance of residual renal function compared with peritoneal clearance for patient survival and quality of life: an analysis of the Netherlands Cooperative Study on the Adequacy of Dialysis (NECOSAD)-2. *Am J Kidney Dis* (2003) 41:1293–302. doi:10.1016/S0272-6386(03)00362-7
248. Balafa O, Halbesma N, Struijk DG, Dekker FW, Krediet RT. Peritoneal albumin and protein losses do not predict outcome in peritoneal dialysis patients. *Clin J Am Soc Nephrol* (2011) 6:561–6. doi:10.2215/CJN.05540610
249. Brimble KS, Walker M, Margetts PJ, Kundhal KK, Rabbat CG. Meta-analysis: peritoneal membrane transport, mortality, and technique failure in peritoneal dialysis. *J Am Soc Nephrol* (2006) 17:2591–8. doi:10.1681/ASN.2006030194
250. Rajakaruna G, Caplin B, Davenport A. Peritoneal protein clearance rather than faster transport status determines outcomes in peritoneal dialysis patients. *Perit Dial Int* (2015) 35:216–21. doi:10.3747/pdi.2013.00217
251. Pascual M, Theruvath T, Kawai T, Tolkoff-Rubin N, Cosimi AB. Strategies to improve long-term outcomes after renal transplantation. *N Engl J Med* (2002) 346:580–90. doi:10.1056/NEJMra011295
252. Heemann U, Lutz J. Pathophysiology and treatment options of chronic renal allograft damage. *Nephrol Dial Transplant* (2013) 28:2438–46. doi:10.1093/ndt/gft087
253. Boor P, Floege J. Renal allograft fibrosis: biology and therapeutic targets. *Am J Transplant* (2015) 15:863–86. doi:10.1111/ajt.13180
254. Bargnoux AS, Servel AC, Piéroni L, Dupuy AM, Badiou S, Garrigue V, et al. Accuracy of GFR predictive equations in renal transplantation: validation of a new turbidimetric cystatin C assay on Architect c8000. *Clin Biochem* (2012) 45:151–3. doi:10.1016/j.clinbiochem.2011.10.018
255. Buron F, Hadj-Aissa A, Dubourg L, Morelon E, Steghens J-P, Ducher M, et al. Estimating glomerular filtration rate in kidney transplant recipients: performance over time of four creatinine-based formulas. *Transplantation* (2011) 92:1005–11. doi:10.1097/TP.0b013e3182301602
256. Gourishankar S, Leduc R, Connett J, Cecka JM, Cosio F, Fieberg A, et al. Pathological and clinical characterization of the ‘troubled transplant’: data from the DeKAF Study. *Am J Transplant* (2010) 10:324–30. doi:10.1111/j.1600-6143.2009.02954.x
257. Mannon RB, Kirk AD. Beyond histology: novel tools to diagnose allograft dysfunction. *Clin J Am Soc Nephrol* (2006) 1:358–66. doi:10.2215/CJN.01681105
258. Foster MC, Weiner DE, Bostom AG, Carpenter MA, Inker LA, Jarolim P, et al. Filtration markers, cardiovascular disease, mortality, and kidney outcomes in stable kidney transplant recipients: the FAVORIT trial. *Am J Transplant* (2017). doi:10.1111/ajt.14258
259. Trailin AV, Pleten MV, Ostapenko TI, Iefimenko NF, Nikonenko OS. High serum level of β₂-microglobulin in late posttransplant period predicts subsequent decline in kidney allograft function: a preliminary study. *Dis Markers* (2015) 2015:562580. doi:10.1155/2015/562580
260. Michel K, Santella R, Steers J, Sahajpal A, Downey FX, Thohan V, et al. Many de novo donor-specific antibodies recognize β₂-microglobulin-free, but not intact HLA heterodimers. *HLA* (2016) 87:356–66. doi:10.1111/tan.12775
261. Johnston O, Cassidy H, O’Connell S, O’Riordan A, Gallagher W, Maguire PB, et al. Identification of β₂-microglobulin as a urinary biomarker for chronic allograft nephropathy using proteomic methods. *Proteomics Clin Appl* (2011) 5:422–31. doi:10.1002/prca.201000160
262. Cassidy H, Slyne J, O’Kelly P, Traynor C, Conlon PJ, Johnston O, et al. Urinary biomarkers of chronic allograft nephropathy. *Proteomics Clin Appl* (2015) 9:574–85. doi:10.1002/prca.201400200
263. Schaub S, Rush D, Wilkins J, Gibson IW, Weiler T, Sangster K, et al. Proteomic-based detection of urine proteins associated with acute renal allograft rejection. *J Am Soc Nephrol* (2004) 15:219–27. doi:10.1097/01.ASN.0000101031.52826.BE
264. Schaub S, Wilkins JA, Antonovici M, Krokkin O, Weiler T, Rush D, et al. Proteomic-based identification of cleaved urinary beta2-microglobulin as a potential marker for acute tubular injury in renal allografts. *Am J Transplant* (2005) 5:729–38. doi:10.1111/j.1600-6143.2005.00766.x
265. Oetting WS, Rogers TB, Krick TP, Matas AJ, Ibrahim HN. Urinary beta2-microglobulin is associated with acute renal allograft rejection. *Am J Kidney Dis* (2006) 47:898–904. doi:10.1053/j.ajkd.2006.01.034
266. Zhang Y, Oetting WS, Harvey SB, Stone MD, Monkkonen T, Matas AJ, et al. Urinary peptide patterns in native kidneys and kidney allografts. *Transplantation* (2009) 87:1807–13. doi:10.1097/TP.0b013e3181a66595
267. Donadio C, Puccini R, Lucchesi A, Giordani R, Rizzo G. Urinary excretion of proteins and tubular enzymes in renal transplant patients. *Ren Fail* (1998) 20:707–15. doi:10.3109/08860229809045167
268. Wakabayashi K, Inokuma S, Matsubara E, Onishi K, Asashima H, Nakachi S, et al. Serum β₂-microglobulin level is a useful indicator of disease activity and hemophagocytic syndrome complication in systemic lupus erythematosus and adult-onset Still’s disease. *Clin Rheumatol* (2013) 32:999–1005. doi:10.1007/s10067-013-2220-8
269. Yoo C, Yoon DH, Suh C. Serum beta-2 microglobulin in malignant lymphomas: an old but powerful prognostic factor. *Blood Res* (2014) 49:148–53. doi:10.5045/br.2014.49.3.148
270. Anders H-J, Vielhauer V. Renal co-morbidity in patients with rheumatic diseases. *Arthritis Res Ther* (2011) 13:222. doi:10.1186/ar3256
271. Chiu H-Y, Huang H-L, Li C-H, Chen H-A, Yeh C-L, Chiu S-H, et al. Increased risk of chronic kidney disease in rheumatoid arthritis associated with cardiovascular complications – a national population-based cohort study. *PLoS One* (2015) 10:e0136508. doi:10.1371/journal.pone.0136508
272. Kronbichler A, Mayer G. Renal involvement in autoimmune connective tissue diseases. *BMC Med* (2013) 11:95. doi:10.1186/1741-7015-11-95
273. Yoo C, Yoon DH, Kim S, Huh J, Park C-S, Park C-J, et al. Serum beta-2 microglobulin as a prognostic biomarker in patients with mantle cell lymphoma. *Hematol Oncol* (2016) 34:22–7. doi:10.1002/hon.2188
274. Seo S, Hong JY, Yoon S, Yoo C, Park JH, Lee JB, et al. Prognostic significance of serum beta-2 microglobulin in patients with diffuse large B-cell lymphoma in the rituximab era. *Oncotarget* (2016) 7:76934–43. doi:10.18632/oncotarget.12734
275. Tsimberidou A-M, Kantarjian HM, Wen S, O’Brien S, Cortes J, Wierda WG, et al. The prognostic significance of serum β₂ microglobulin levels in acute myeloid leukemia and prognostic scores predicting survival: analysis of 1,180 patients. *Clin Cancer Res* (2008) 14:721–30. doi:10.1158/1078-0432.CCR-07-2063
276. von Hohenstaufen KA, Conconi A, de Campos CP, Franceschetti S, Bertoni F, Margiotta Casaluci G, et al. Prognostic impact of monocyte count at

- presentation in mantle cell lymphoma. *Br J Haematol* (2013) 162:465–73. doi:10.1111/bjh.12409
277. Wang Q, Qin Y, Zhou S, He X, Yang J, Kang S, et al. Prognostic value of pre-treatment serum beta-2 microglobulin level in advanced classical Hodgkin lymphoma treated in the modern era. *Oncotarget* (2016) 7:72219–28. doi:10.18632/oncotarget.12663
278. Coutinho R, Lobato J, Esteves S, Carvalho S, Pierdomenico F, Bernardo P, et al. The NCCN-IPI helps to identify very high-risk patients with DLBCL and can be improved by other independent clinical factors including bulky disease and beta2-microglobulin. *Blood* (2015) 126:3888–3888.
279. Kanemasa Y, Shimoyama T, Sasaki Y, Tamura M, Sawada T, Omuro Y, et al. Beta-2 microglobulin as a significant prognostic factor and a new risk model for patients with diffuse large B-cell lymphoma. *Hematol Oncol* (2016). doi:10.1002/hon.2312
280. Rossi D, Fangazio M, De Paoli L, Puma A, Riccomagno P, Pinto V, et al. Beta-2-microglobulin is an independent predictor of progression in asymptomatic multiple myeloma. *Cancer* (2010) 116:2188–200. doi:10.1002/cncr.24959
281. Rajkumar SV, Fonseca R, Lacy MQ, Witzig TE, Lust JA, Greipp PR, et al. Beta2-microglobulin and bone marrow plasma cell involvement predict complete responders among patients undergoing blood cell transplantation for myeloma. *Bone Marrow Transplant* (1999) 23:1261–6. doi:10.1038/sj.bmt.1701787
282. Stella-Holowiecka B, Czerw T, Holowiecka-Goral A, Giebel S, Wojnar J, Holowiecki J. Beta-2-microglobulin level predicts outcome following autologous hematopoietic stem cell transplantation in patients with multiple myeloma. *Transplant Proc* (2007) 39:2893–7. doi:10.1016/j.transproceed.2007.08.052
283. Palumbo A, Avet-Loiseau H, Oliva S, Lokhorst HM, Goldschmidt H, Rosinol L, et al. Revised international staging system for multiple myeloma: a report from International Myeloma Working Group. *J Clin Oncol* (2015) 33:2863. doi:10.1200/JCO.2015.61.2267
284. García de Veas Silva JL, Bermudo Guitarte C, Menéndez Valladares P, Rojas Noboa JC, Kestler K, Duro Millán R. Prognostic value of serum free light chains measurements in multiple myeloma patients. *PLoS One* (2016) 11:e0166841. doi:10.1371/journal.pone.0166841
285. Rajkumar SV. Myeloma today: disease definitions and treatment advances. *Am J Hematol* (2016) 91:90–100. doi:10.1002/ajh.24236
286. Papanikolaou X, Alapat D, Rosenthal A, Stein C, Epstein J, Owens R, et al. The flow cytometry-defined light chain cytoplasmic immunoglobulin index and an associated 12-gene expression signature are independent prognostic factors in multiple myeloma. *Leukemia* (2015) 29:1713–20. doi:10.1038/leu.2015.65
287. van Rhee F, Bolejack V, Hollmig K, Pineda-Roman M, Anaissie E, Epstein J, et al. High serum-free light chain levels and their rapid reduction in response to therapy define an aggressive multiple myeloma subtype with poor prognosis. *Blood* (2007) 110:827–32. doi:10.1182/blood-2007-01-067728
288. Park S, Han B, Kim K, Kim SJ, Jang JH, Kim WS, et al. Renal Insufficiency in newly-diagnosed multiple myeloma: analysis according to International Myeloma Working Group consensus statement. *Anticancer Res* (2014) 34:4299–306.
289. Chen C, Puvvada S. Prognostic factors for chronic lymphocytic leukemia. *Curr Hematol Malig Rep* (2016) 11:37–42. doi:10.1007/s11899-015-0294-x
290. Gentile M, Cutrona G, Neri A, Molica S, Ferrarini M, Morabito F. Predictive value of β_2 -microglobulin (β_2 -m) levels in chronic lymphocytic leukemia since Binet A stages. *Haematologica* (2009) 94:887–8. doi:10.3324/haematol.2009.005561
291. Rossi D, Zucchetto A, Rossi FM, Capello D, Cerri M, Deambrogi C, et al. CD49d expression is an independent risk factor of progressive disease in early stage chronic lymphocytic leukemia. *Haematologica* (2008) 93:1575–9. doi:10.3324/haematol.13103
292. Thompson PA, O'Brien SM, Xiao L, Wang X, Burger JA, Jain N, et al. β_2 -microglobulin normalization within 6 months of ibrutinib-based treatment is associated with superior PFS in CLL. *Cancer* (2016) 122:565–73. doi:10.1002/cncr.29794
293. Høgdaal E, Fung ET, Christensen IJ, Yip C, Nedergaard L, Engelholm SA, et al. Proteomic biomarkers for overall and progression-free survival in ovarian cancer patients. *Proteomics Clin Appl* (2010) 4:940–52. doi:10.1002/prca.200900171
294. Sun J, Yang Z, Miao X, Zou Q, Li J, Liang L, et al. ATP5b and β_2 -microglobulin are predictive markers for the prognosis of patients with gallbladder cancer. *J Mol Histol* (2015) 46:57–65. doi:10.1007/s10735-014-9597-9
295. Zhang Y, Wang L, Ji P, Zhao G, Zhong G, Wang Z. Correlation of serum β_2 -microglobulin levels with prostate-specific antigen, Gleason score, clinical stage, tumor metastasis and therapy efficacy in prostate cancer. *Arch Med Res* (2013) 44:259–65. doi:10.1016/j.arcmed.2013.03.008
296. Klein T, Levin I, Niska A, Koren R, Gal R, Schachter J, et al. Correlation between tumour and serum beta 2m expression in patients with breast cancer. *Eur J Immunogenet* (1996) 23:417–23. doi:10.1111/j.1744-313X.1996.tb00132.x
297. Lucarelli G, Dittono P, Bettocchi C, Vavalo A, Rutigliano M, Galleggiante V, et al. Diagnostic and prognostic role of preoperative circulating CA 15-3, CA 125, and beta-2 microglobulin in renal cell carcinoma. *Dis Markers* (2014) 2014:e689795. doi:10.1155/2014/689795
298. Ueland FR. A perspective on ovarian cancer biomarkers: past, present and yet-to-come. *Diagnostics (Basel)* (2017) 7:14. doi:10.3390/diagnostics7010014
299. Ueland FR, Desimone CP, Seamon LG, Miller RA, Goodrich S, Podzielinski I, et al. Effectiveness of a multivariate index assay in the preoperative assessment of ovarian tumors. *Obstet Gynecol* (2011) 117:1289–97. doi:10.1097/AOG.0b013e31821b5118
300. Prizant AE, Linabery AM, Lutsey PL, Selvin E, Nelson HH, Folsom AR, et al. Circulating beta-2 microglobulin and risk of cancer: the atherosclerosis risk in communities study (ARIC). *Cancer Epidemiol Biomarkers Prev* (2016) 25:657–64. doi:10.1158/1055-9965.EPI-15-0849
301. Hermansen M-LF, Hummelshøj L, Lundsgaard D, Hornum L, Keller P, Fleckner J, et al. Increased serum β_2 -microglobulin is associated with clinical and immunological markers of disease activity in systemic lupus erythematosus patients. *Lupus* (2012) 21:1098–104. doi:10.1177/0961203312447668
302. Candon S, Gottenberg JE, Bengoufa D, Chatenoud L, Mariette X. Quantitative assessment of antibodies to ribonucleoproteins in primary Sjögren syndrome: correlation with B-cell biomarkers and disease activity. *Ann Rheum Dis* (2009) 68:1208–12. doi:10.1136/ard.2008.095257
303. Gottenberg J, Busson M, Cohen-Solal J, Lavie F, Abbed K, Kimberly R, et al. Correlation of serum B lymphocyte stimulator and β_2 microglobulin with autoantibody secretion and systemic involvement in primary Sjögren's syndrome. *Ann Rheum Dis* (2005) 64:1050–5. doi:10.1136/ard.2004.030643
304. Gottenberg J-E, Seror R, Miceli-Richard C, Benessiano J, Devauchelle-Pensec V, Dieude P, et al. Serum levels of beta2-microglobulin and free light chains of immunoglobulins are associated with systemic disease activity in Primary Sjögren's Syndrome. Data at enrollment in the prospective ASSESS cohort. *PLoS One* (2013) 8:e59868. doi:10.1371/journal.pone.0059868
305. Jiang T, Ding X, Lu W. The prognostic significance of beta2 microglobulin in patients with hemophagocytic lymphohistiocytosis. *Dis Markers* (2016) 2016:e1523959. doi:10.1155/2016/1523959
306. Amarante GBD, Zotin MC, Rocha E, Delgado AG Jr, Leite M Jr, Gomes CP. Renal tubular dysfunction in patients with primary Sjögren syndrome. *Clin Nephrol* (2014) 81:185–91. doi:10.5414/CN108142
307. Neiryneck N, Eloit S, Glorieux G, Barreto DV, Barreto FC, Liabeuf S, et al. Estimated glomerular filtration rate is a poor predictor of the concentration of middle molecular weight uremic solutes in chronic kidney disease. *PLoS One* (2012) 7:e44201. doi:10.1371/journal.pone.0044201
308. Stevens LA, Coresh J, Schmid CH, Feldman HI, Froissart M, Kusek J, et al. Estimating GFR using serum cystatin C alone and in combination with serum creatinine: a pooled analysis of 3418 individuals with CKD. *Am J Kidney Dis* (2008) 51:395–406. doi:10.1053/j.ajkd.2007.11.018
309. Argyropoulos C, Unruh ML. Analysis of time to event outcomes in randomized controlled trials by generalized additive models. *PLoS One* (2015) 10:e0123784. doi:10.1371/journal.pone.0123784
310. Bhatt L, Horgan CP, Walsh M, McCaffrey MW. The hereditary hemochromatosis protein HFE and its chaperone beta2-microglobulin localise predominantly to the endosomal-recycling compartment. *Biochem Biophys Res Commun* (2007) 359:277–84. doi:10.1016/j.bbrc.2007.05.100
311. Ye L, Liu X, Rout SN, Li Z, Yan Y, Lu L, et al. The MHC class II-associated invariant chain interacts with the neonatal Fc gamma receptor and modulates its trafficking to endosomal/lysosomal compartments. *J Immunol* (2008) 181:2572–85. doi:10.4049/jimmunol.181.4.2572

312. Ando M, Shibuya A, Tsuchiya K, Akiba T, Nitta K. Reduced capacity of mononuclear cells to synthesize cytokines against an inflammatory stimulus in uremic patients. *Nephron Clin Pract* (2006) 104:c113–9. doi:10.1159/000094446
313. Lonnemann G, Novick D, Rubinstein M, Dinarello CA. Interleukin-18, interleukin-18 binding protein and impaired production of interferon-gamma in chronic renal failure. *Clin Nephrol* (2003) 60:327–34. doi:10.5414/CNP60327
314. Ahmed AT, Karter AJ. Tuberculosis in California dialysis patients. *Int J Tuberc Lung Dis* (2004) 8:341–5.
315. Chia S, Karim M, Elwood RK, FitzGerald JM. Risk of tuberculosis in dialysis patients: a population-based study. *Int J Tuberc Lung Dis* (1998) 2:989–91.
316. Chou KJ, Fang HC, Bai KJ, Hwang SJ, Yang WC, Chung HM. Tuberculosis in maintenance dialysis patients. *Nephron* (2001) 88:138–43. doi:10.1159/000045974
317. Dalrymple LS, Go AS. Epidemiology of acute infections among patients with chronic kidney disease. *Clin J Am Soc Nephrol* (2008) 3:1487–93. doi:10.2215/CJN.01290308
318. Maisonneuve P, Agodoa L, Gellert R, Stewart J, Buccianti G, Lowenfels A, et al. Cancer in patients on dialysis for end-stage renal disease: an international collaborative study. *Lancet* (1999) 354:93–9. doi:10.1016/S0140-6736(99)06154-1
319. Stengel B. Chronic kidney disease and cancer: a troubling connection. *J Nephrol* (2010) 23:253–62.
320. Vajdic CM, McDonald SP, McCredie MRE, van Leeuwen MT, Stewart JH, Law M, et al. Cancer incidence before and after kidney transplantation. *JAMA* (2006) 296:2823–31. doi:10.1001/jama.296.23.2823
321. Yanik EL, Clarke CA, Snyder JJ, Pfeiffer RM, Engels EA. Variation in cancer incidence among patients with ESRD during kidney function and nonfunction intervals. *J Am Soc Nephrol* (2016) 27:1495–504. doi:10.1681/ASN.2015040373
322. Butler AM, Olshan AF, Kshirsagar AV, Edwards JK, Nielsen ME, Wheeler SB, et al. Cancer incidence among US Medicare ESRD patients receiving hemodialysis, 1996–2009. *Am J Kidney Dis* (2015) 65:763–72. doi:10.1053/j.ajkd.2014.12.013
323. Shang W, Huang L, Li L, Li X, Zeng R, Ge S, et al. Cancer risk in patients receiving renal replacement therapy: a meta-analysis of cohort studies. *Mol Clin Oncol* (2016) 5:315–25. doi:10.3892/mco.2016.952
324. Jahn B, Betz M, Deppisch R, Janssen O, Hänsch GM, Ritz E. Stimulation of beta 2-microglobulin synthesis in lymphocytes after exposure to Cuprophane dialyzer membranes. *Kidney Int* (1991) 40:285–90. doi:10.1038/ki.1991.212
325. Schoels M, Jahn B, Hug F, Deppisch R, Ritz E, Hänsch GM. Stimulation of mononuclear cells by contact with cuprophane membranes: further increase of beta 2-microglobulin synthesis by activated late complement components. *Am J Kidney Dis* (1993) 21:394–9. doi:10.1016/S0272-6386(12)80267-8
326. Campistol JM, Molina R, Bernard DB, Rodriguez R, Mirapeix E, Munoz-Gomez JM, et al. Synthesis of beta 2-microglobulin in lymphocyte culture: role of hemodialysis, dialysis membranes, dialysis-amyloidosis, and lymphokines. *Am J Kidney Dis* (1993) 22:691–9. doi:10.1016/S0272-6386(12)80432-X
327. Zingraff J, Beyne P, Ureña P, Uzan M, Man NK, Descamps-Latscha B, et al. Influence of haemodialysis membranes on beta 2-microglobulin kinetics: in vivo and in vitro studies. *Nephrol Dial Transplant* (1988) 3:284–90.
328. Haufe CC, Eismann U, Deppisch RM, Stein G. Expression of beta2-microglobulin and c-fos mRNA: is there an influence of high- or low-flux dialyzer membranes? *Kidney Int Suppl* (2001) 78:S177–81. doi:10.1046/j.1523-1755.2001.07816.x
329. Zaoui PM, Stone WJ, Hakim RM. Effects of dialysis membranes on beta 2-microglobulin production and cellular expression. *Kidney Int* (1990) 38:962–8. doi:10.1038/ki.1990.298
330. Traut M, Haufe CC, Eismann U, Deppisch RM, Stein G, Wolf G. Increased binding of beta-2-microglobulin to blood cells in dialysis patients treated with high-flux dialyzers compared with low-flux membranes contributed to reduced beta-2-microglobulin concentrations. Results of a cross-over study. *Blood Purif* (2007) 25:432–40. doi:10.1159/000110069
331. Vanholder R, Abou-Deif O, Argiles A, Baurmeister U, Beige J, Brouckaert P, et al. The role of EUTox in uremic toxin research. *Semin Dial* (2009) 22:323–8. doi:10.1111/j.1525-139X.2009.00574.x

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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