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Biomass-derived fiber materials for biomedical applications

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With the development of sustainable materials, biomass-derived fiber materials have attracted significant research interest because of their excellent performance, cost-effectiveness, and environmental friendliness. Following their extensive use in biomedical applications, biomass-derived fiber materials are now increasingly used in antibacterial skin-wound dressings, bone tissue engineering, drug delivery, etc. However, their practical medical applications are still in their infancy, requiring some reference and strategic directions for the future clinical applications of biomaterials in medicine and cross-discipline use, some of which are provided in this review. The sources and characteristics of the different biomass-derived fibers are introduced briefly, encompassing polysaccharide fiber and protein fiber's two categories. Then, we summarize the common applications of biomass and its derived fibers in the medical field in recent years. Finally, we discuss the application and advantages of three-dimensional printing technology combined with biomass-derived materials in modern medicine for a better understanding of the practical applications of biomass-derived fiber materials.

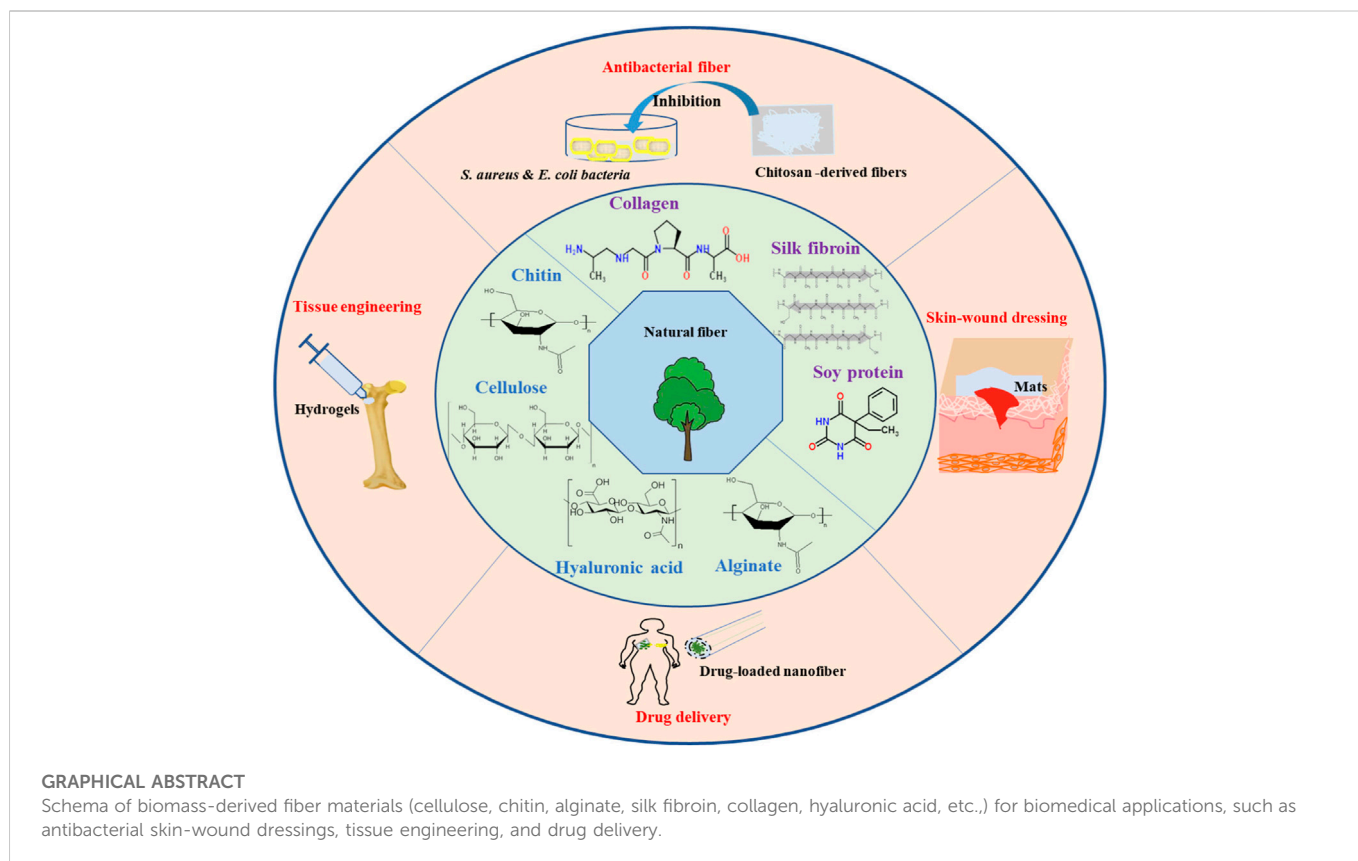
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1 Introduction

Polymers extracted from traditional petroleum materials have significantly promoted the development of modern life. However, petroleum-derived polymers cause environmental pollution and are non-renewable. Green chemistry makes chemical reactions and processes more environment-friendly (Pleissner and Kummerer, 2020). Over the past few years, renewable resources have provided an opportunity for the sustainable development of function by allowing us to use biomass materials in our daily lives (Srivastava et al., 2021). Biomass materials without chemical modification are mainly composed of carbon, hydrogen, and oxygen and are degraded easily by natural microorganisms into water, carbon dioxide, and other small molecules, which means their products can reenter the natural cycle. Therefore, advanced green biomass fibers, such as lignin-based carbon fibers and nanofibers, which are based on renewable resources, have great potential as sustainable composites (Venkata Mohan et al., 2016).

The sources of biomass materials are abundant and widely distributed. Biomass-derived materials mainly include polysaccharide fiber materials, such as cellulose, chitin, starch, sodium alginate, pectin, hyaluronic acid (HA), etc., and protein fiber materials, such as silk fibroin, collagen, corn protein, and soy protein. At present, biomass-derived fiber materials have been used in environmental protection (Hassan and Carr, 2021), biosensor sensors (Liedel, 2020), photocatalysts (Son et al., 2021), and others applications. Indeed, as long as specific quality measures are satisfied, biomass-derived fiber materials are promising candidates for the



biomedical field. The applications of biomass-derived fibers have, recently, been extended from antibacterial skin-wound dressings to bone tissue engineering, artificial blood vessels, drug-sustained release, and other areas, because of their excellent biodegradability, non-toxicity, renewability, and polymer chemical reaction. For example, Huang et al. prepared injectable hydrogels for the treatment of deep burn wounds by mixing modified cellulose nanofibers with carboxymethyl chitosan (CS) (Huang et al., 2018). Kawata et al. demonstrated that calcium phosphate/chitin nanofiber hydrogels could promote the formation of calcium phosphate crystals, which can improve the mechanical properties of scaffolds according to mineralization time (Kawata et al., 2016). Additionally, with the development of three-dimensional (3D) printing technology in biomass-derived materials, the development of multiscale, multilateral, and multifunctional structures for biomedical products will be a trend in the future. Huang et al. fabricated a lung tissue scaffold with 10 layers with high shape fidelity and fiber alignment via 3D printing technology (Huang et al., 2021); Tsukamoto et al. developed 3D cardiac scaffolds with an oriented structure, which possess better contractile properties and orientation-controlled structure (Tsukamoto et al., 2020); Sun et al. prepared a 3D printed collagen/chitosan scaffold to ameliorate axon regeneration and neurological recovery after spinal cord injury (Sun et al., 2019). With the development of 3D printing technology, biomass-derived materials will be more suitable for the treatment of fine structures and complex tissues in the future.

In this review, we attempt to provide readers with an overview of the sources and characteristics of different biomass-derived fibers and

a comprehensive study of the common applications of biomass and their derived fibers in the medical field in recent years. Moreover, we put special emphasis on the applications of biomass-derived fibers as bioinks for 3D printing and on the advantages that 3D printing technology combined with biomass-derived materials can bring to modern medicine, such as in tissue engineering, wound healing, and organoids. To reach the target property of 3D-printed biomass-derived materials, it is important to understand their composition and material characteristics. Overall, biomass-derived fiber materials are promising candidates for the commercialization of medical materials, and we expect the points shared in this review to give an impetus to further exploration of biomass-derived fiber materials for biomedical applications.

2 Source and composition of biomass-derived materials

2.1 Polysaccharide fiber materials

Polysaccharides originate from plants, animals, and microorganisms, and are abundant in the biological world with a wide range of functional and structural features. Polysaccharide fiber materials mainly include cellulose, chitin, starch, sodium alginate, pectin, and HA. They have been widely applied in various fields, including food, medicine, pharmaceuticals, energy components, cosmetics, packaging, dyes and pigments, antimicrobials, and water treatment (Akshay Kumar et al., 2021).

2.1.1 Cellulose

Cellulose, the main component of lignocellulosic biomass, is the most abundant natural polymer on earth. The annual global production of cellulose is approximately 75–100 billion tons (Du et al., 2019). Cellulose, a large molecular polysaccharide consisting of glucose, is synthesized in nature as a single molecule with linear chains of glucose-based residues, which self-assemble at biosynthetic sites. Approximately 30 individual cellulose molecules are assembled into fibrils, which are packaged into larger microfibrils, which in turn are assembled into cellulose fibers. The constituent molecules of individual microfibrils are packed tightly to prevent penetration by enzymes and small molecules such as water (Khalid et al., 2017). Therefore, cellulose is insoluble in water and organic solvents. The main sources of cellulose are wood pulp, dissolving wood pulp, cotton, cotton staples, and other fiber pulps. Cellulose derived from plants and bacteria exists in the form of cellulose nanofibers, nanoparticles, hydrogels, and aerogels. Cellulose has attracted increasing attention in the field of biomedical materials owing to its biocompatibility, biodegradability, renewability, environmental friendliness, and non-toxicity.

2.1.2 Chitin and chitosan

Chitin is the second most common polymer on earth after cellulose and is mainly derived from the exoskeletons of insects and marine crustaceans. In its native state, chitin occurs as ordered crystalline microfibrils that form structural components in the exoskeletons of arthropods or the cell walls of fungi and yeast. Until now, the main commercial sources of chitin have been crab and shrimp shells. In industrial processes, chitin is extracted by dissolving calcium carbonate with an acid treatment and then dissolving the protein in an alkaline solution. Hence, chitin exhibits a high degree of thermodynamic stability. The poor solubility of chitin in solvent extraction makes the extraction process difficult and laborious (Zhang et al., 2021). With advances in technology and a better understanding of its physiological and biological properties, chitin can be deacetylated and converted into chitosan (CS), a water-soluble polysaccharide (El Knidri et al., 2018). Nanomaterials based on chitin and its derivatives are frequently used in biomedical fields, such as in hemostasis, wound healing, antimicrobial agents, cell culture, tissue regeneration, and skin protection, owing to their unique biocompatibility, biodegradability, non-toxicity, metal ion chelation, and antimicrobial properties (Liu et al., 2019a; Ahmad et al., 2020).

2.1.3 Sodium alginate

Alginate is a biopolymer commonly obtained from brown algae (Cyanobacteria, Alginidae) and bacteria (*Azotobacter vineyard* and *Pseudomonas* genus species) and it belongs to the polycationic copolymer family (Ling et al., 2019; Kothale et al., 2020). Alginate is a hydrophilic polysaccharide composed of linear copolymers containing (1,4) -linked β -D-mannouronic acid (M) and α -L-galonic acid (G) residues. The proportion of MM, GG, and MG blocks has a decisive influence on the physical properties of alginate (Sanchez-Ballester et al., 2021). Alginate has been used in the food and cosmetic industries as a thickener or viscosity increaser, owing to its viscosity, biodegradability, and non-toxicity (Kontominas, 2020). Ling et al. reported that alginate-CS microspheres could prevent coated drugs from damaging the reticulum endothelial system (Ling et al., 2019). Moreover, alginate has good

biocompatibility, low cost, ease of gelation, inertness, chemical compatibility, derivatives of natural sources, and easily available and feasible synthesis methods with several attractive properties, making it the biopolymer most widely applied in wound dressings, bone regeneration, new angiogenesis, protein delivery, cell delivery, therapeutic agents, oral drug delivery, and controlled release systems (Dhamecha et al., 2019).

2.1.4 Hyaluronic acid

HA, a non-sulfated anionic polysaccharide, is widely present in the human body and is essential for many cellular and tissue functions. HA is a non-sulfated glycosaminoglycan consisting of repeated D-glucuronic acid polymeric disaccharides linked by glycosidic bonds in an arrangement of alternating β -(1 \rightarrow 4) and β -(1 \rightarrow 3) bonds. The HA structure has a remarkable ability to retain/capture approximately 1000 times its weight in water (Bukhari et al., 2018). HA is a sulfate-free glycosaminoglycan found throughout the body, from the vitreous of the eye to the extracellular stroma of cartilage tissue. HA can be degraded by hyaluronidase in the body; therefore, its half-life is a few hours to days (Burdick and Prestwich, 2011). HA can be modified into HA-derivatives (thiol-modified HA, haloacetate-modified HA, hydrazide-modified HA, aldehyde-modified HA, and aldehyde-modified HA) according to the properties of the resulting material. HA and its derivatives have been used clinically for more than three decades (Peptu and Kowalczyk, 2018). Owing to its abundance in animals and humans, its biodegradability, non-toxicity, biocompatibility, non-immunogenicity, and non-inflammatory properties, it can be used for skin repair, cancer diagnosis, wound healing, tissue regeneration, anti-inflammatory purposes, and immune regulation. (Burdick and Prestwich, 2011; Tiwari and Bahadur, 2019).

2.2 Protein fiber materials

Natural protein fiber materials exhibit enhanced biocompatibility, bioactivity, and biodegradability. Because of their excellent biodegradability and biocompatibility, these proteins (collagen, silk, gelatin, soy protein, keratin, elastin, zein, and soy) are mostly used in the medical field in stents, sutures, wound healing, ligament replacement, and drug delivery. Compared with other natural biomaterials, they can support increased cell migration and proliferation.

2.2.1 Silk fibroin protein

Silk fibroin (SF) is present in the glands of filament-producing arthropods, including silkworms, spiders, scorpions, mites, and bees, and is spun into fibers during metamorphosis. Silk has a large molecular weight (200–350 kDa or higher) and a large repetitive modular hydrophobic domain, which is interrupted by small hydrophilic groups. The N- and C-termini of the SF are highly preserved. The SF protein consists of a heavy chain (H) and a light chain (L) linked by disulfide bonds. The H chain has a hydrophobic domain, and the L chain is hydrophilic and relatively elastic. The ratio of H to L determines the mechanical properties, bioactivity, and degradation behavior of SF (Kundu et al., 2013). The extraction of silk protein usually requires the addition of a variety of concentrated salt solutions, such as lithium bromide, calcium chloride/ethanol/

TABLE 1 Different bacteria reduction rates of CS-derived fibers.

Composition	Bacteria	Bacteria inhibition rate	References
CS	<i>S. aureus</i> and <i>E. coli</i> bacteria	99.2% and 95.6%	Li J et al. (2020)
CS/alginate fibers	<i>S. aureus</i> (MSSA and CA-MRSA)	83.47%–100%; and 76.26%–100%	Dumont et al. (2018)
CS/gallnut tannins fiber	<i>S. aureus</i> and <i>Candida albicans</i>	99.7% and 35%	Zhu et al. (2019)
Oxidized CS-grafted cashmere fibers	<i>S. aureus</i> and <i>E. coli</i> bacteria	90.4%–94.4% and 85.2%–91.8%	Li Z et al. (2020)
CS/tannic acid LBL Cellulose acetate fibers	<i>S. aureus</i> and <i>E. coli</i> bacteria	99% and 86%	Huang L et al. (2019)
CS/Oregano essential oil/PCL	<i>S. aureus</i> and <i>L. monocytogenes</i> and <i>S. enteritidis</i> and <i>E. coli</i> bacteria	52.72 ± 1.01% and 41.06 ± 3.12% and 49.37 ± 1.78% and 40.47 ± 3.36%	Augustine et al. (2020)
CS/PVA fibers	<i>S. aureus</i> and <i>E. coli</i> bacteria	67.4% and 58.9%	Wang et al. (2020)
HKUST-1/CS/PVA fibers	<i>S. aureus</i> and <i>E. coli</i> bacteria	99.0% and 99.0%	Wang et al. (2020)
SDS-nanoTiO ₂ /CS	<i>S. aureus</i> and <i>E. coli</i> bacteria	85.8% and 54.8%	Bao et al. (2019)

water, lithium chloride, or ionic liquids, under heated conditions. The low immunogenicity and antigenicity of SF rarely cause wound infections. Biodegradability and bio-absorbability materials are promising substitutes for many human protein (Kundu et al., 2013). Degummed filament silk fibers can form a variety of twisted structures, including membranes, hydrogels, 3D porous scaffolds, and particles, for use in vascular, skin, bone, cartilage, ligament, tendon, neural, and cardiac tissue regeneration (Yao et al., 2022).

2.2.2 Collagen and gelatin

Collagen is widely distributed in connective tissues throughout the body. The synthesis of natural collagen is complex and involves several biological processes *in vivo*. The processing property of collagen is an elegant structural motif in which three parallel polypeptide chains interact with each other in a left-handed polyproline type II (PPII) helical conformation coil, forming a single-residue right-handed triple helix. The abundant amide-amide hydrogen bonds in the triple helix structure give collagen strong thermal stability, mechanical strength, and the ability to interact with other biomolecules, making collagen an important structure for animal scaffolds (Shoulders and Raines, 2009). As a protein from mammals, collagen has advantages, such as an abundant source, rich biocompatibility, easy processing, hydrophilicity, low antigenicity, and being easily absorbable by the body (Sorushanova et al., 2019). Collagen has been used in leather, food, medicine, and photography. In the medical field, collagen scaffolds can serve as artificial skin, bones, tendons, and cartilage. Moreover, collagen can function as a carrier for drugs and growth factors. However, poor physicochemical properties of collagen, such as thermal stability, mechanical strength, and enzyme resistance, are often required to bind other synthetic polymers (polylactic acid) in bone regeneration (Liu X. et al., 2019).

Gelatin is a natural polymer with a molecular weight between 15 and 250 kDa that is hydrolyzed and degraded by collagen. Gelatin can be obtained from cow bones, fish, pig skin, and some insects through alkaline hydrolysis or acid hydrolysis (Campbell and Hotchkiss, 2017). Its unique amino acid structure gives it a variety of medical benefits: (1) It is highly soluble; (2) It contains important binding parts for cell attachment; (3) It is biodegradable and non-toxic to cells. However, for biological materials, gelatin has some drawbacks.

The main disadvantages of gelatin-based materials are poor mechanical properties, poor thermal stability, and a relatively short degradation rate (Bello et al., 2020). Gelatin is used in various areas, such as personal care products, pharmaceuticals, photography, nutraceuticals, food and liquid snacks (dessert courses, dairy products, liquid snacks, poultry and meat products, and confectionery) among others (Campbell and Hotchkiss, 2017).

2.2.3 Soy protein

SF, collagen, and gelatin are examples of animal protein used in biomedicine. Soy globulin, the main plant protein, is the main protein in soy. Soy protein has a variety of functional physicochemical properties for food applications, such as emulsification, foaming, gel, water, and fat absorption (Jahangirian et al., 2019). Soy protein isolate is used with other materials as a coating for protection or for physical or chemical surface modification (Hadzieva et al., 2017). In recent years, soy protein has been gradually applied to the medical field, as it can be prepared into hydrogels, micro and nanoparticles, fibers, and porous structures, suitable for different medical applications, such as tissue engineering, drug delivery, etc. Plant-derived proteins have a lower immunogenic potential than animal proteins, are easy to process, are certainly less likely to transmit disease and are relatively inexpensive. In addition, they have relatively low molecular weight and exhibit greater polarity (Mohammadinejad et al., 2016). With the emergence of new technologies and the improvement of the function of these protein materials, there will be more opportunities for effective disease treatment in the future.

3 Biomass-derived fiber materials for biomedical applications

3.1 Antibacterial fiber

CS can resist various microbes, including bacteria, molds, and yeasts. The main reason for this is that the negative surface charge of the bacteria interacts with the positively charged amino group of CS, resulting in a change in cell permeability and disruption of the cell membrane (Matica et al., 2019). Another mechanism is that CS

hydrolysis products can interact with microbial DNA, thereby disturbing mRNA, and protein synthesis (Romanazzi et al., 2018). Finally, CS can chelate metal ions (Ni^{2+} , Zn^{2+} , Co^{2+} , Fe^{2+} , and Cu^{2+}), which are essential nutrients for bacteria (Hosseinnejad and Jafari, 2016). CS of different molecular weights have different antibacterial effects. CS with a low molecular weight can pass through the cytomembrane and combine with the cell nucleus. However, CS with a high molecular weight can adhere to bacteria and disrupt nutrient transport (Yuan et al., 2020). Unfortunately, their poor mechanical characteristics and water sensitivity have severely restricted their application.

CS fibers are the most widely used form of CS material to improve their mechanical properties. Currently, CS fibers are mainly fabricated with various polymers, such as PVA, PEO, and PCL, to function as antibacterial agents in wound healing, tissue repair, and regeneration (Guo et al., 2020; Wang et al., 2020). Zhu et al. prepared CS/gallnut tannin fibers via blended solution spinning; the results showed that it significantly improved the lower hydrophilicity and higher dry and wet breaking strengths. In addition, the antimicrobial properties increased from 49% to 99.7% (Zhu et al., 2019). The different bacterial inhibition rates of CS-derived fibers in recent years are displayed in Table 1. Generally, the inhibition efficiency of CS-derived fibers against *S. aureus* was superior to that of *E. coli*.

Guo et al. prepared coaxial electrospun nanofibrous mats with CS/PEO as the shell and PCL as the core. After 48 h of incubation, the mats exhibited excellent antibacterial performance against *E. coli* and *S. aureus* (Guo et al., 2020). Wang et al. fabricated HKUST-1/CS/PVA fibers for application in wound healing processes against *E. coli* and *S. aureus* with 99% antibacterial efficiency (Wang et al., 2020). Antifungal activity against *C. albicans*, *C. glabrata*, and *S. cerevisiae* (Verlee et al., 2017) has been recently discovered. Antifungal properties are related to molecular weight (MW). Low MW (LMW) CS (17.4 kDa) is more likely to inhibit the mycelial growth of *R. stolonifera* (Hernandez-Lauzardo et al., 2008), whereas high MW (HMW) CS (350 kDa) is more effective for spore germination and mycelial growth of *A. kikuchiana* (Meng et al., 2010). Compared with the HMW CS, the LWH CS can penetrate the fungal cell wall more easily.

3.2 Skin-wound dressings

Globally, wound dressings, as skin tissue engineering materials, have drawn considerable attention from scientists and clinicians. However, traditional dressings, including gauze, lint, cotton wool, and bandages, are often easily dried and adhere to the skin, causing wound pain and discomfort (Khalid et al., 2017). In addition, conventional dressings often cannot resist bacterial infections and provide sufficient tensile strength. Moreover, their single function in wound healing cannot fulfill the requirements of complex wounds, such as infected wounds and diabetic chronic wounds (Mostafalu et al., 2017; Chen et al., 2018). An ideal wound dressing can: (1) provide a moist environment, (2) remove the exudate, (3) possess air permeability, and (4) possess antibacterial properties. Currently, natural biopolymeric fibers show considerable potential as wound dressing candidates because of their excellent moisture retention, high sorption capacity, outstanding biodegradability, and appropriate mechanical properties (Xi et al., 2018; Homaeigohar and Boccaccini, 2020; Xi et al., 2020). Biomass-derived fibers mainly

originate from naturally occurring polymers, such as cellulose, HA, chitin/CS, collagen, and SF.

Cellulose has been extensively investigated for tissue regeneration by mimicking the extracellular matrix (ECM) structure. Compared with commercial wound dressings, cellulose-fiber dressings are equipped with more advantages, including (1) maintaining a moist environment at the wound site to relieve pain; (2) promoting granulation tissues and re-epithelialization formation, and diminishing scar formation; (3) being equipped with good molding and mechanical properties to meet the requirements of complex wound healing (burns and chronic ulcers, damaged tissues, etc.); (4) protection of the wound against bacterial infection (Luo et al., 2021); and (5) excellent light transmittance and high porosity to speed up wound healing (Xia et al., 2020). Cellulose-fiber dressings can be multi-functionalized by incorporating other biopolymers, such as enzymes, collagen, CS, hormones, and gallic acid (Li et al., 2019; Wutticharoenmongkol et al., 2019). For instance, Xia et al. fabricated transparent wound dressings using porous cellulose fibers coated with CS, enabling visualization of wound healing (Xia et al., 2020). However, the special functions of cellulose fibers at different stages of wound healing, such as inflammation, proliferation, and remodeling, should be further investigated in subsequent studies. Hamid et al. prepared PVA/CS/silk sericin/tetracycline porous fibers possessing hygroscopic properties, antibacterial activity, and biocompatibility *in vivo* and *in vitro* (Bakhsheshi-Rad et al., 2020).

HA is a promising biopolymer for biomedical applications, especially wound dressings. The rich carboxyl and hydroxyl groups of the HA structure provide it with a highly hydrophilic character that facilitates cell adhesion and wound healing. Moreover, HA can encourage hemostasis, prompt collagen deposition and fibrosis, regulate inflammation balance, and promote re-epithelialization via the enzymatic reaction of hyaluronidase and chemical interaction (Zhu et al., 2017; Xia et al., 2020). The pivotal role of HA-fiber dressing in wound healing is in the inflammation phase (Graca et al., 2020). HA can facilitate the recruitment of neutrophils to eliminate debris and dead tissues and subsequently release cytokines, such as TNF- α , IL-1 β , and IL-8, which are involved in the inflammatory response (Tavianatou et al., 2019). HA with a high molecular weight (HMW-HA, $>1 \times 10^6$ Da) is fragmented to low molecular weight (LMW-HA, $1-25 \times 10^4$ Da), recruiting leukocytes and monocytes into the wound site. Finally, the cell surface receptors (TLR2 and TLR4) of these cells interact with LMW-HA to promote the expression of these cytokines, further cascading amplification effects (Zamboni et al., 2018; Tavianatou et al., 2019). However, the mechanical stability of HA-fiber dressings is a significant challenge. To date, the characteristics of HA-fiber dressings have been constantly developed through the incorporation of biomacromolecules and chemical modification to be widely applied in the clinic, for example, the HylaSponge[®] System (Mahedia et al., 2016), Hyalomatrix[®] (Longinotti, 2014), and Hyalosafe[®] (Longinotti, 2014). Moreover, HA-fibers advance into differently structured dressings (sponges, films, hydrogels, and electrospun membranes) with natural and synthetic biomolecules, such as PCL, PLGA, collagen, SF, and CS (Zhou et al., 2016; Graca et al., 2020). In the future, HA-fiber dressings with multiple functions will likely be developed.

Chitin is often insoluble in common solvents; thus, it is usually deacetylated to CS in an alkaline environment by dissolving it in organic acids, such as dilute aqueous formic, acetic, and lactic acids. In the early stages of wound healing, CS can induce polymorphonuclear

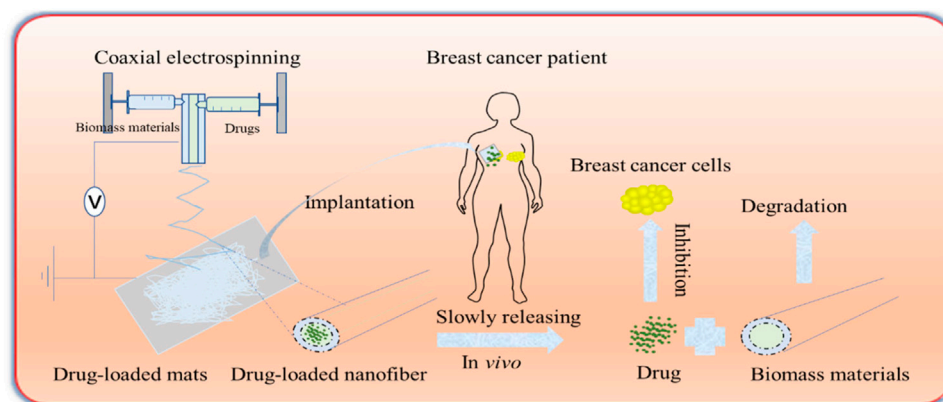


FIGURE 2

Schema of biomass-derived fibers for drug delivery. The drugs and biomass materials are made into porous nanofiber mats by coaxial electrospinning. The mats are transferred into the tumor areas of patients with breast cancer, etc. The drugs are released gradually, and the drugs inhibit tumor growth, while the biomass materials can be degraded naturally.

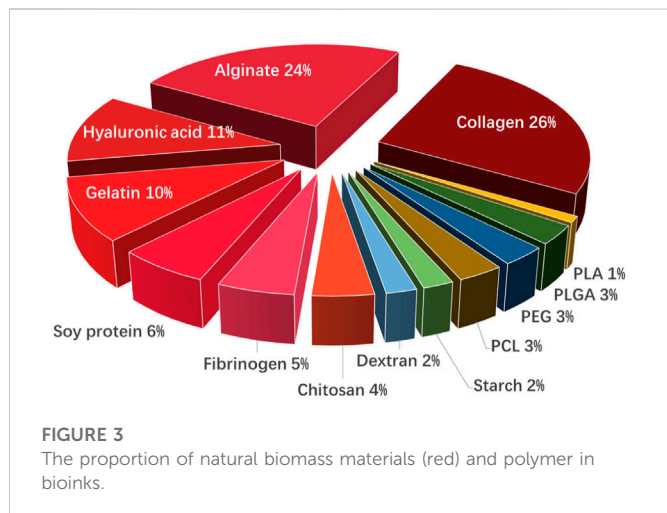
hydroxyapatite (Roseti et al., 2017). Biopolymeric nanofibers can mimic the bone ECM, where the cell can interact with the nanostructure surface of the material and rapidly grow. Chitin and CS, similar to the ECM, can be degraded into small molecular amino acids and polysaccharides *in vivo*, and have been processed into different structures, such as nanofibers, hydrogels, and aerogels for application in bone tissue repair and regeneration. Chitin/CS-fiber scaffolds and calcium phosphate/chitin hydrogels have been shown to facilitate the repair of bone defects without an inflammatory response *in vivo* (Gupta et al., 2015). Moreover, Chitin and CS can be dissolved in acidic solvents with other polymers, such as PCL, PLA, and nylon 6, to change their mechanical performance and biological properties. Research has shown that the function mechanism of chitin/CS is that it can activate osteogenesis-related signaling pathways, Runx2, BMP-2, BMP-4, and collagen-1, and facilitate the expression of alkaline phosphatase (ALP) (early biomineralization marker) and osteointegration relative protein, which contributes to mineralization and bone regeneration (Liu et al., 2013). Overall, chitin/CS-fiber scaffolds mainly affect bone regeneration engineering by promoting cell growth and osteogenic differentiation, accelerating angiogenesis of mineralized bone tissue, and promoting drug delivery. Apart from Chitin and CS, alginate has shown great potential in bone tissue engineering owing to its good biocompatibility, biodegradation, and size handling. In addition, alginate nanofiber blended with calcium phosphate cement can activate osteoblastic cell immobilization, proliferation, and differentiation (Jayachandran et al., 2022). Alginate scaffolds can promote human umbilical cord mesenchymal stem cell differentiation, and express ALP and osteosarcoma-associated proteins (Su et al., 2021). Alginate saline gel has strong gelling ability, low toxicity, high availability, and low cost, characteristics that make it the most common material in bone tissue engineering and bio-printing.

Proteins with non-toxicity, biodegradability, and biocompatibility make them an alternative to bone, ligaments, and cartilage. Owing to swelling *in vivo*, collagen must cross-link with other polymers (CaPs) to overcome its disadvantages and poor mechanical and dimensional stability (Bao et al., 2020).

The nature of the robust elastic property of SF protein corresponds with the robust mechanical strength of the bone. A group of SF surfaces, such as -OH, -COOH, and -N-H, can provide sites for hydroxyapatite (Kundu et al., 2012), and then the hydroxyapatite is mineralized to form bone under certain conditions. Typically, SF can guide calcium phosphate formation and promote the deposition of osteoblasts along nanofibers to produce a new matrix. More importantly, the degradation rate of SF is 6–12 months *in vivo*, which is helpful for the repair of bone defects, and the degradation products of SF (glycine and alanine) can be reused as templates for neo-protein synthesis. Therefore, SF does not often stimulate excessive inflammatory reactions. Moreover, SF-chitin hybrid nanofiber/TGF- β 1-functionalized scaffolds displayed a significant effect on chondrocyte proliferation, difference, and adhesion *in vivo* and *in vitro* (Cheng et al., 2021). Cheng et al developed core-shell SF/PCL/PVA mats by loading a controlled release system of BMP2 and CTGF using layer-by-layer techniques. Its positive effects on bone tissue repair and healing were confirmed *in vitro* and *in vivo* (Cheng et al., 2019). SF has been manufactured in a variety of forms, including films, artificial fibers, sponges, and hydrogels, which have been successfully deployed in a variety of tissue engineering applications (Sun et al., 2021). In the future, the fabrication by 3D bioprinting of nanoscale SF multi-level structures with high structural resolutions will make the application of SF more popular.

3.4 Drug delivery

Cancer, as one of the world's biggest challenges, causes enormous medical and financial burdens. Traditionally, cancer therapy has included surgery, radiation, and chemotherapy. However, these treatments can easily cause systemic adverse reactions while killing the tumors. Progress has been made through biomaterial-based implantable drug delivery in cancer therapy (Yadav et al., 2021). The ideal drug delivery includes the following features: (1) local drug delivery (the drug is released directly into the tumor, thus



four model drugs (naproxen, indomethacin, ibuprofen, and sulindac) (Tungprapa et al., 2007). However, this treatment has the risk of causing initial burst release; thus, different modified strategies have been developed, including the application of tri-axial electrospinning (Yang et al., 2019), increasing the surface hydrophobicity of drug carriers, and increasing porosity (Chen et al., 2020). Cellulose sulfate is an ester or cellulose sulfated from cellulose. Compared to natural cellulose, its antimicrobial properties and solubility are greatly improved. In addition, the excellent film-forming property allows it to effectively encapsulate drugs (rifampicin and risedronate) (Vehlow et al., 2016) and cytokines, such as BMP2 (Muller et al., 2018). Cellulose nitrate, acetate cellulose (Chen et al., 2020), carboxymethyl cellulose (Chen et al., 2019), ethyl cellulose, and methyl cellulose are effective carriers for drug delivery (Oprea and Voicu, 2020). Schematics of biomass-derived fibers for drug delivery are shown in Figure 2.

sharply reducing the drug dose and side effects of the drug); (2) sustained drug release (the drug should avoid abrupt drug release and maintain function concentration for a longer period); and (3) drug stability (the drug-delivery system should preserve the loaded drug to avoid degradation and removal by enzymes or other media *in vivo* before it is released) (Bastiancich et al., 2016; Conde et al., 2016; Talebian et al., 2018).

Cellulose is a natural and promising material in the biomedical field owing to its availability and good mechanical properties. Its notable mechanical characteristics, water solubility, hydrolytic stability, and natural biocompatibility facilitate its biomedical application. It has been produced on electrospun fibers coated with

4 3D printing of biomass-derived fiber materials

4.1 From material bionics to structure bionics

Ideal medical biomimicry materials model the complexity and heterogeneity of these tissues, including the required biological functions, sufficient strength, and stiffness to maintain structural integrity, and microscopic porous structures. But, common nanofibers *via* traditional process methods, such as electrostatic spinning, chemical vapor deposition, electrospinning, and solution blowing, can only still create biomaterials for tissues with relatively simple structures and compositions, such as skin and bone. However,

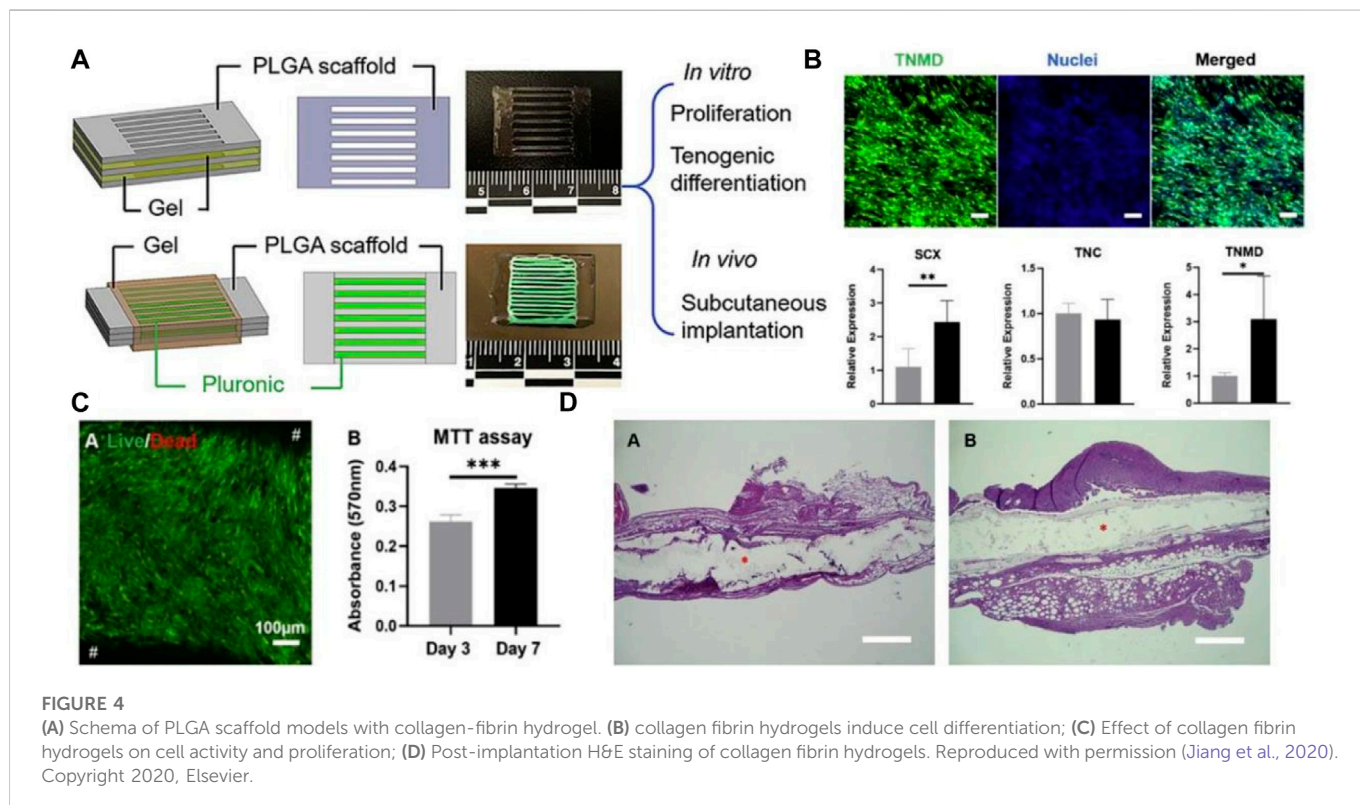


TABLE 2 The 3D printing of biomass polysaccharide fiber materials in medical application.

Types	Printed samples/structure	Function and potential applications	References
Cellulose- Poly (ethylene glycol) diacrylate nanocrystal hydrogels	Human ear construct scaffolds	High repeatability, fidelity, and mechanical integrity; reconstructive surgery of microtia and anotia	Palaganas et al. (2017)
SF-oxidized bacterial cellulose hydrogels	Lung tissue scaffolds with 10 layers	High shape fidelity and fiber alignment; Lung tissue engineering	Huang et al. (2021)
Nanocellulose and alginate hydrogels	Human ear and sheep meniscus scaffolds	High shape fidelity and printing resolution; Cartilage Tissue Engineering	Markstedt et al. (2015)
Bacterial cellulose nanofibers/SF/gelatin composite hydrogels	Human meniscus scaffolds with hierarchical pores	High shape fidelity and mechanical property; soft tissue reconstruction	Huang et al. (2019)
Nano-fibrillated cellulose, carboxymethyl cellulose, and citric acid	Honeycomb bone trabecular scaffolds with porous	High-controlled porosity, mechanical strength, and biocompatibility; Bone tissue engineering applications	Stiglic et al. (2022)
Chitosan hydrogels incorporated with cellulose nanocrystals	Knee meniscus scaffolds	High mechanical properties and biocompatibility; bone tissue engineering	Maturavongsadit et al. (2021)
Nano-fibrillated cellulose	Long-term stability scaffolds for cartilaginous tissues	High biosafety and mechanical stability; Cartilaginous tissues engineering	Apelgren et al. (2021)
Gelatin/chitosan hydrogels	Multi-furcated vessels and heterogeneous porous scaffolds	High physiological stability, mechanical strength, semi-permeability, hemocompatibility; Vascular network tissue engineering	Su et al. (2022)
Hydroxyapatite, poly (dopamine), and carboxymethyl CS	Porous scaffolds	High mechanical properties and water absorption, photothermal properties; Anti-tumor and bone repair engineering	Yao et al. (2021)
Fibronectin/gelatin	Cardiac scaffolds with an oriented structure	Better contractile properties and orientation-controlled structure; Cardiac tissue engineering	Tsukamoto et al. (2020)
Gelatin/carboxymethyl chitin/hydroxyapatite composite gel	Honeycomb bone trabeculae	High water uptake ratio, cell retention capability, infiltration, attachment, proliferation; Bone tissue engineering	Gupta et al. (2019)

the fine structure and complex composition of organisms are far beyond the capabilities of manufacturing technology, which limits biomimetic research on tissue regeneration. 3D printing has shown great promise in the design of biomaterials with multiscale, multilateral, and multifunctional structures (Ashammakhi et al., 2019; Ng et al., 2019). According to statistics, the proportion of biomass materials (~85%) in 3D printing is much higher than that of polymer materials (Khoeni et al., 2021) (Figure 3). Collagen, accounting for 26%, is the most common 3D-printed biomass material, which represents the major components of our natural ECM, providing overall organizational stiffness and integrity with distinct properties, and functionalities (Lee C et al., 2021). Alginate (24%) is the second most used biomass material in bioinks, which can improve the quality of the built (reduced spattering) or printed entity and the viscosity of the ink (Rastogi and Kandasubramanian, 2019). HA (11%) stands out for its excellent physical, chemical, and biological properties, which can provide printing suitability, improve its mechanical properties and printing with loaded cells, etc. (Ding et al., 2023). With this technology, biomass-derived materials have been successfully made into fibers, channels, sheets, coils, meshes, and porous 3D constructs that mimic tissue components. Rees A et al. discovered that, *via* 3D bioprinting, CS was formed into a 3D scaffold of grid structure with 9 layers, which has open porosity and the potential to carry and release antimicrobial components (Rees et al., 2015). Su H et al. fabricated free-standing multifurcated vessels and complicated vascular networks in heterogeneous porous scaffolds by 3D printing technology (Su et al., 2022). Gupta D et al. prepared a 3D printed gelatin/carboxymethyl chitin/hydroxyapatite composite bioactive gel scaffold with a controlled hierarchical structure (Gupta et al., 2019).

4.2 Advantages of 3D printing of biomass-derived composite fiber

Biomass-derived materials, including cellulose, lignin, starch, collagen, alginate, and CS, are the most used materials for 3D printing. They not only meet the needs of sustainability, but also reduce the side effects of some synthetic polymers in biomedical applications, e.g., degradability, recyclability, harmful decomposition of products, release of additives, and reduced cell attachment (Liu et al., 2019b). However, the inherent physicochemical properties of some biomass-derived materials, such as poor stability and adhesion, certainly pose difficult challenges to their 3D printing application. Hence, mixing different types of biopolymers with complementary advantages or combining with other ingredients are alternative methods or strategies to formulate composite inks with good processability, printability, mechanics, and bioactivity, and the performance and function of biomaterial products can also be improved and biomass-derived applications can be broadened. For instance, by combining agarose and type I collagen, Kopf, M. et al. developed a hydrogel mixture capable of coating human umbilic artery smooth muscle cells for a long time and that could be 3D printed on demand (Kopf et al., 2016). 3D structure-scaffolds can be prepared by adding methylcellulose into a 3% alginate solution, providing better printability, and maintaining the advantage for cell embedding (Schutz et al., 2017). 3D bioprinters can distribute materials while moving in the X, Y, and Z directions, which can prepare complex cartilage structures, such as a human ear and sheep meniscus, with high shape fidelity and printing resolution (Markstedt et al., 2015). Stiglic et al. developed an organic acid cross-linked 3D-printed cellulose nanocomposite biological scaffold with controllable porosity, mechanical strength, and biocompatibility (Stiglic et al., 2022).

TABLE 3 The 3D printing of biomass protein fiber materials in medical application.

Types	Printed samples/structure	Function and potential applications	References
SF-Gelatin	Columnar and porous scaffolds	Excellent mechanical properties and degradation rate; Cartilage Tissue Engineering	Shi et al. (2017)
Collagen/CS	Porous scaffolds	High mechanical properties and biocompatibility; Tissue engineering of nerve repair	Sun et al. (2019)
Collagen/SF	Multi-layer porous structure scaffolds	Optimal biocompatibility, strength, and precise microarchitecture; Tissue engineering of nerve repair	Li et al. (2021)
Polycaprolactone (PCL)/silk fibroin (SF)	Meniscus	Extraordinary biomechanical properties and biocompatibility	Li J et al. (2020)
SF/collagen/hydroxyapatite	Irregular resembling mandible and porous scaffolds	Show the ideal water absorption and porosity; Reconstruction of Alveolar Bone Defects	Liu et al. (2022)
SF/collagen/hydroxyapatite biological	Round porous scaffolds	Show high biocompatibility and excellent mechanical properties; Cartilage Tissue Engineering	Lee J et al. (2021)
Hydroxyapatite nanocrystals and collagen fibers	Irregular resembling radius porous and major-size scaffolds	Enhance angiogenesis, osteogenesis, and osteointegration; Tissue Engineering for bone defect repair	Ma et al. (2018)
HA and collagen, or gelatin hydrogel	Brain tumor organoids	Provide viable and accurate organoids; drug development-based screening studies and precision medicine applications	Clark et al. (2022)
Collagen-based waveform microfibers	Waveform fiber-guiding scaffolds	Show high biocompatibility and excellent mechanical properties; Periodontal Ligament Reconstruction	Lin et al. (2021)

hyaluronic acid (HA), Chitosan (CS), silk fibroin.

4.3 3D printed polysaccharide/protein fiber materials in medicine application

Biomass-derived materials that are 3D printed can be applied in bone/articular cartilage defect repair, nerve tissue repair, soft tissue repair, and other types of repair. Ventola et al. incorporated human nasal septum chondrocytes into cellulose-alginate composite ink for 3D bioprinting to improve the shape fidelity and stability of biomedical materials (Ventola, 2014). Jiang et al. developed multilayer 3D printed scaffolds for rotator cuff tendon regeneration via combined collagen-fibrin hydrogels (Jiang et al., 2020) (Figure 4). Yao M et al. prepared HA, poly (dopamine), and carboxymethyl CS composite scaffolds using 3D printing technology, which had an anti-osteosarcoma effect and bone repair properties (Yao et al., 2021). Liu H et al. used 3D printing technology to develop SF/hydroxyapatite/collagen scaffolds with ideal water absorption and porosity, which could significantly promote the reconstruction of mandibular defects (Liu et al., 2022). Those 3D-printed polysaccharide/protein fiber materials possess high mechanical properties and biocompatibility, which can be widely applied in bone tissue engineering. Recent polysaccharide/protein fiber-based 3D printing studies with various applications regarding the different types of biomass fibers are summarized in Table 2; Table 3.

5 Conclusion and outlook

With the development of interdisciplinary research between medicine and bioengineering, emerging research has been devoted to the application of biomass materials in medicine. Combining the clinical needs and physical characteristics of biomass-derived fiber materials, the development of excellent and novel medical materials is

promising for application in a wider range of fields. In general, most biomass-derived fiber materials contain polysaccharide and protein fibers from two major sources, which offer excellent antibacterial properties and a significant recovery function of tissues that contribute to the development of antibacterial/antibiofilm materials, skin-wound dressings, tissue engineering, drug-sustained release, and other areas. Furthermore, the multiscale, multilateral, and multifunctional structures of biomass-derived fiber materials are available by 3D printing technology, which has been recognized as an effective and promising method for preparing biomass-derived fibers, owing to its advantages in speed, accuracy, and flexibility (Ji et al., 2020).

Meanwhile, 3D printing of biomass-derived fiber materials is also a promising prospect for drug detection. In past decades, drug screening and the development of new drugs have often relied on genetically modified animals as disease models. However, the use of animal models raises serious ethical issues and brings with it inevitable limitations in accurately representing human tissues in the pathophysiological context due to genetic differences (Vanderburgh et al., 2017). 3D printing of biomass-derived materials can provide platforms for drug delivery, screening, and development by accurately depositing biomaterials containing patient-derived cells and simulating the natural environment of the diseased body. Falcone G et al. prepared floating ribocendazole delivery systems, which can prolong gastric retention times (Falcone et al., 2022). Li Q et al. developed novel gastro-floating tablets with 3D extrusion-based printing, prolonging the gastric residence time of dapsin and increasing the drug release rate (Li et al., 2018). As for floating drug delivery systems, 3D printing of biomass-derived materials can highly prolong the drug action time, thereby improving bioavailability, patient compliance, and therapeutic efficacy. In addition, 3D printing of biomass-derived materials can also serve as the platform for drug screening and development. Yi et al. prepared

a radial oxygen gradient using a 3D gas permeation barrier containing ECM (glycosaminoglycan and HA) of brain cancer cells and endothelial cells to generate central hypoxia and simulate an anoxic environment for drug resistance research (Yi et al., 2019). Clark C et al. prepared hyaluronan and collagen bioink-supported 3D patient-derived brain tumor organoids, which can provide viable and accurate organoids for drug screening and development studies and precision medicine applications (Clark et al., 2022). Although 3D printing of biomass-derived materials has great advantages in drug delivery, screening, and development, its long-term effects and biosafety *in vivo* are still uncertain, and, thus, it needs more clinical studies in the future.

In conclusion, we summarized the common applications of biomass and its derived fibers in the medical field in recent years and discussed the application and advantages of 3D printing technology combined with biomass-derived materials in modern medicine. Our review may benefit from a better understanding of practical applications in biomass-derived fiber materials. Overall, although there have been several successes *in vitro* and *in vivo* trials of biomaterials, none have been processed to the human testing stage. Therefore, the translational benefits of biomass-based tissue engineering are still far from being applied to patients. This applies not only to tissue engineering, drug development, and organoids but also to other biomedicine fields. Therefore, translational research is a challenge to overcome in the future.

Author contributions

DL wrote the initial manuscript and created the figures and table. YW conceptualized and discussed the review. HG and WH reviewed

and edited the manuscript. All authors have contributed to the work and approved the final version of the manuscript.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Ahmad, S. I., Ahmad, R., Khan, M. S., Kant, R., Shahid, S., Gautam, L., et al. (2020). Chitin and its derivatives: Structural properties and biomedical applications. *Int. J. Biol. Macromol.* 164, 526–539. doi:10.1016/j.jbiomac.2020.07.098
- Akshay Kumar, K. P., Zare, E. N., Torres-Mendieta, R., Waclawek, S., Makvandi, P., Cernik, M., et al. (2021). Electrospun fibers based on botanical, seaweed, microbial, and animal sourced biomacromolecules and their multidimensional applications. *Int. J. Biol. Macromol.* 171, 130–149. doi:10.1016/j.jbiomac.2020.12.205
- Alven, S., and Aderibigbe, B. A. (2020). Chitosan and cellulose-based hydrogels for wound management. *Int. J. Mol. Sci.* 21 (24), 9656. doi:10.3390/ijms21249656
- Apelgren, P., Amoroso, M., Saljo, K., Lindahl, A., Brantsing, C., Stridh Orrhult, L., et al. (2021). Long-term *in vivo* integrity and safety of 3D-bioprinted cartilaginous constructs. *J. Biomed. Mater. Res. B Appl. Biomater.* 109 (1), 126–136. doi:10.1002/jbm.b.34687
- Ashammakhi, N., Ahadian, S., Xu, C., Montazerian, H., Ko, H., Nasiri, R., et al. (2019). Bioinks and bioprinting technologies to make heterogeneous and biomimetic tissue constructs. *Mater. Today Bio* 1, 100008. doi:10.1016/j.mtbio.2019.100008
- Augustine, R., Rehman, S. R. U., Ahmed, R., Zahid, A. A., Sharifi, M., Falahati, M., et al. (2020). Electrospun chitosan membranes containing bioactive and therapeutic agents for enhanced wound healing. *Int. J. Biol. Macromol.* 156, 153–170. doi:10.1016/j.jbiomac.2020.03.207
- Bakhsheshi-Rad, H. R., Ismail, A. F., Aziz, M., Akbari, M., Hadisi, Z., Omid, M., et al. (2020). Development of the PVA/CS nanofibers containing silk protein sericin as a wound dressing: *In vitro* and *in vivo* assessment. *Int. J. Biol. Macromol.* 149, 513–521. doi:10.1016/j.jbiomac.2020.01.139
- Bao, W., Li, M., Yang, Y., Wan, Y., Wang, X., Bi, N., et al. (2020). Advancements and Frontiers in the high performance of natural hydrogels for cartilage tissue engineering. *Front. Chem.* 8, 53. doi:10.3389/fchem.2020.00053
- Bao, Y., Zhou, Q., Zhang, M., Zhang, H., Luan, Q., Zhou, W., et al. (2019). Wet-spun nanoTiO₂/chitosan nanocomposite fibers as efficient and retrievable absorbent for the removal of free fatty acids from edible oil. *Carbohydr. Polym.* 210, 119–126. doi:10.1016/j.carbpol.2019.01.035
- Bastiancich, C., Danhier, P., Preat, V., and Danhier, F. (2016). Anticancer drug-loaded hydrogels as drug delivery systems for the local treatment of glioblastoma. *J. Control Release* 243, 29–42. doi:10.1016/j.jconrel.2016.09.034
- Bello, A. B., Kim, D., Kim, D., Park, H., and Lee, S. H. (2020). Engineering and functionalization of gelatin biomaterials: From cell culture to medical applications. *Tissue Eng. Part B Rev.* 26 (2), 164–180. doi:10.1089/ten.TEB.2019.0256
- Bhattacharjee, P., Kundu, B., Naskar, D., Kim, H. W., Maiti, T. K., Bhattacharya, D., et al. (2017). Silk scaffolds in bone tissue engineering: An overview. *Acta Biomater.* 63, 1–17. doi:10.1016/j.actbio.2017.09.027
- Bukhari, S. N. A., Roswandi, N. L., Waqas, M., Habib, H., Hussain, F., Khan, S., et al. (2018). Hyaluronic acid, a promising skin rejuvenating biomedicine: A review of recent updates and pre-clinical and clinical investigations on cosmetic and nutraceutical effects. *Int. J. Biol. Macromol.* 120, 1682–1695. doi:10.1016/j.jbiomac.2018.09.188
- Burdick, J. A., and Prestwich, G. D. (2011). Hyaluronic acid hydrogels for biomedical applications. *Adv. Mater* 23 (12), H41–H56. doi:10.1002/adma.201003963
- Campbell, R., and Hotchkiss, S. (2017). "Carrageenan industry market overview," in *Tropical seaweed farming trends, problems and opportunities* (Berlin, Germany: Springer), 193–205.
- Chen, G., Yu, Y., Wu, X., Wang, G., Ren, J., and Zhao, Y. (2018). Bioinspired multifunctional hybrid hydrogel promotes wound healing. *Adv. Funct. Mater.* 28 (33), 1801386. doi:10.1002/adfm.201801386
- Chen, J., Duan, H., Pan, H., Yang, X., and Pan, W. (2019). Two types of core/shell fibers based on carboxymethyl chitosan and Sodium carboxymethyl cellulose with self-assembled liposome for buccal delivery of carvedilol across TR146 cell culture and porcine buccal mucosa. *Int. J. Biol. Macromol.* 128, 700–709. doi:10.1016/j.jbiomac.2019.01.143
- Chen, Y., Qiu, Y., Chen, W., and Wei, Q. (2020). Electrospun thymol-loaded porous cellulose acetate fibers with potential biomedical applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* 109, 110536. doi:10.1016/j.msec.2019.110536

- Cheng, G., Dai, J., Dai, J., Wang, H., Li, Z., Liu, Y., et al. (2021). Extracellular matrix imitation utilizing nanofibers-embedded biomimetic scaffolds for facilitating cartilage regeneration. *Chem. Eng. J.* 410 (2), 128379. doi:10.1016/j.cej.2020.128379
- Cheng, G., Yin, C., Tu, H., Jiang, S., Wang, Q., Zhou, X., et al. (2019). Controlled Co-delivery of growth factors through layer-by-layer assembly of core-shell nanofibers for improving bone regeneration. *ACS Nano* 13 (6), 6372–6382. doi:10.1021/acsnano.8b06032
- Clark, C., Yoo, K. M., Sivakumar, H., Strumpf, K., Laxton, A., Tatter, S. B., et al. (2022). Immersion bioprinting of hyaluronan and collagen bioink-supported 3D patient-derived brain tumor organoids. *Biomed. Mater.* 18, 015014. doi:10.1088/1748-605X/aca05d
- Conde, J., Shomron, N., and Artzi, N. (2016). Biomaterials for abrogating metastasis: Bridging the gap between basic and translational research. *Adv. Healthc. Mater.* 5 (18), 2312–2319. doi:10.1002/adhm.201600414
- Dhamecha, D., Movsas, R., Sano, U., and Menon, J. U. (2019). Applications of alginate microspheres in therapeutics delivery and cell culture: Past, present and future. *Int. J. Pharm.* 569, 118627. doi:10.1016/j.ijpharm.2019.118627
- Ding, Y.-W., Zhang, X.-W., Mi, C.-H., Qi, X.-Y., Zhou, J., and Wei, D.-X. (2023). Recent advances in hyaluronic acid-based hydrogels for 3D bioprinting in tissue engineering applications. *Smart Mater. Med.* 4, 59–68. doi:10.1016/j.smaim.2022.07.003
- Du, H., Liu, W., Zhang, M., Si, C., Zhang, X., and Li, B. (2019). Cellulose nanocrystals and cellulose nanofibrils based hydrogels for biomedical applications. *Carbohydr. Polym.* 209, 130–144. doi:10.1016/j.carbpol.2019.01.020
- Dumont, M., Villet, R., Guirand, M., Montebault, A., Delair, T., Lack, S., et al. (2018). Processing and antibacterial properties of chitosan-coated alginate fibers. *Carbohydr. Polym.* 190, 31–42. doi:10.1016/j.carbpol.2017.11.088
- El Knidri, H., Belaabed, R., Addaou, A., Laajeb, A., and Lahsini, A. (2018). Extraction, chemical modification and characterization of chitin and chitosan. *Int. J. Biol. Macromol.* 120, 1181–1189. doi:10.1016/j.ijbiomac.2018.08.139
- Falcone, G., Real, J. P., Palma, S. D., Aquino, R. P., Del Gaudio, P., Garofalo, E., et al. (2022). Floating ricobendazole delivery systems: A 3D printing method by Co-extrusion of sodium alginate and calcium chloride. *Int. J. Mol. Sci.* 23 (3), 1280. doi:10.3390/ijms23031280
- Farokhi, M., Mottaghtalab, F., Fatahi, Y., Khademhosseini, A., and Kaplan, D. L. (2018). Overview of silk fibroin use in wound dressings. *Trends Biotechnol.* 36 (9), 907–922. doi:10.1016/j.tibtech.2018.04.004
- Graca, M. F. P., Miguel, S. P., Cabral, C. S. D., and Correia, I. J. (2020). Hyaluronic acid-based wound dressings: A review. *Carbohydr. Polym.* 241, 116364. doi:10.1016/j.carbpol.2020.116364
- Guo, H., Tan, S., Gao, J., and Wang, L. (2020). Sequential release of drugs from a dual-delivery system based on pH-responsive nanofibrous mats towards wound care. *J. Mater. Chem. B* 8 (8), 1759–1770. doi:10.1039/c9tb02522g
- Gupta, A. K., Mita, K., Arunkumar, K. P., and Nagaraju, J. (2015). Molecular architecture of silk fibroin of Indian golden silkworm, *Antheraea assama*. *Sci. Rep.* 5, 12706. doi:10.1038/srep12706
- Gupta, D., Singh, A. K., Dravid, A., and Bellare, J. (2019). Multiscale porosity in compressible cryogenically 3D printed gels for bone tissue engineering. *ACS Appl. Mater. Interfaces* 11 (22), 20437–20452. doi:10.1021/acsmi.9b05460
- Hadzieva, J., Mladenovska, K., Crcarevska, M. S., Dodov, M. G., Dimchevska, S., Geskovski, N., et al. (2017). Lactobacillus casei encapsulated in soy protein isolate and alginate microparticles prepared by spray drying. *Food Technol. Biotechnol.* 55 (2), 173–186. doi:10.17113/ftb.55.02.17.4991
- Hassan, M. M., and Carr, C. M. (2021). Biomass-derived porous carbonaceous materials and their composites as adsorbents for cationic and anionic dyes: A review. *Chemosphere* 265, 129087. doi:10.1016/j.chemosphere.2020.129087
- Hernandez-Lauzardo, A. N., Bautista-Banos, S., Velazquez-Del Valle, M. G., Mendez-Montealvo, M. G., Sanchez-Rivera, M. M., and Bello-Perez, L. A. (2008). Antifungal effects of chitosan with different molecular weights on *in vitro* development of *Rhizopus stolonifer* (Ehrenb.:Fr.) Vuill. *Carbohydr. Polym.* 73 (4), 541–547. doi:10.1016/j.carbpol.2007.12.020
- Homaeigohar, S., and Boccaccini, A. R. (2020). Antibacterial biohybrid nanofibers for wound dressings. *Acta Biomater.* 107, 25–49. doi:10.1016/j.actbio.2020.02.022
- Hosseinnejad, M., and Jafari, S. M. (2016). Evaluation of different factors affecting antimicrobial properties of chitosan. *Int. J. Biol. Macromol.* 85, 467–475. doi:10.1016/j.ijbiomac.2016.01.022
- Huang, J., Cheng, Y., Wu, Y., Shi, X., Du, Y., and Deng, H. (2019). Chitosan/tannic acid bilayers layer-by-layer deposited cellulose nanofibrous mats for antibacterial application. *Int. J. Biol. Macromol.* 139, 191–198. doi:10.1016/j.ijbiomac.2019.07.185
- Huang, L., Du, X., Fan, S., Yang, G., Shao, H., Li, D., et al. (2019). Bacterial cellulose nanofibers promote stress and fidelity of 3D-printed silk based hydrogel scaffold with hierarchical pores. *Carbohydr. Polym.* 221, 146–156. doi:10.1016/j.carbpol.2019.05.080
- Huang, L., Yuan, W., Hong, Y., Fan, S., Yao, X., Ren, T., et al. (2021). 3D printed hydrogels with oxidized cellulose nanofibers and silk fibroin for the proliferation of lung epithelial stem cells. *Cellul. (Lond)* 28 (1), 241–257. doi:10.1007/s10570-020-03526-7
- Huang, W., Wang, Y., Huang, Z., Wang, X., Chen, L., Zhang, Y., et al. (2018). On-demand dissolvable self-healing hydrogel based on carboxymethyl chitosan and cellulose nanocrystal for deep partial thickness burn wound healing. *ACS Appl. Mater. Interfaces* 10 (48), 41076–41088. doi:10.1021/acsmi.8b14526
- Jahangirian, H., Azizi, S., Rafiee-Moghaddam, R., Baratvand, B., and Webster, T. J. (2019). Status of plant protein-based green scaffolds for regenerative medicine applications. *Biomolecules* 9 (10), 619. doi:10.3390/biom9100619
- Jayachandran, V., Murugan, S. S., Dalavi, P. A., Gurusanthappa Vishalakshi, Y. D., and Seong, G. H. (2022). Alginate-based composite microspheres: Preparations and applications for bone tissue engineering. *Curr. Pharm. Des.* 28 (13), 1067–1081. doi:10.2174/1381612828666220518142911
- Ji, A., Zhang, S., Bhagia, S., Yoo, C. G., and Ragauskas, A. J. (2020). 3D printing of biomass-derived composites: Application and characterization approaches. *RSC Adv.* 10 (37), 21698–21723. doi:10.1039/d0ra03620j
- Jiang, X., Wu, S., Kuss, M., Kong, Y., Shi, W., Streubel, P. N., et al. (2020). 3D printing of multilayered scaffolds for rotator cuff tendon regeneration. *Bioact. Mater.* 5 (3), 636–643. doi:10.1016/j.bioactmat.2020.04.017
- Kawata, M., Azuma, K., Izawa, H., Morimoto, M., Saimoto, H., and Ifuku, S. (2016). Biomimetic mineralization of calcium phosphate crystals on chitin nanofiber hydrogel for bone regeneration material. *Carbohydr. Polym.* 136, 964–969. doi:10.1016/j.carbpol.2015.10.009
- Khalid, A., Khan, R., Ul-Islam, M., Khan, T., and Wahid, F. (2017). Bacterial cellulose-zinc oxide nanocomposites as a novel dressing system for burn wounds. *Carbohydr. Polym.* 164, 214–221. doi:10.1016/j.carbpol.2017.01.061
- Khoeni, R., Nosrati, H., Akbarzadeh, A., Eftekhari, A., Kavetsky, T., Khalilov, R., et al. (2021). Natural and synthetic bioinks for 3D bioprinting. *Nat. Synthetic Bioprinting* 1 (8), 2000097. doi:10.1002/anbr.202000097
- Kontominas, M. G. (2020). Use of alginates as food packaging materials. *Foods* 9 (10), 1440. doi:10.3390/foods9101440
- Kopf, M., Campos, D. F., Blaeser, A., Sen, K. S., and Fischer, H. (2016). A tailored three-dimensionally printable agarose-collagen blend allows encapsulation, spreading, and attachment of human umbilical artery smooth muscle cells. *Biofabrication* 8 (2), 025011. doi:10.1088/1758-5090/8/2/025011
- Kothale, D., Verma, U., Dewangan, N., Jana, P., Jain, A., and Jain, D. (2020). Alginate as promising natural polymer for pharmaceutical, food, and biomedical applications. *Curr. Drug Deliv.* 17 (9), 755–775. doi:10.2174/1567201817666200810110226
- Kundu, B., Rajkhowa, R., Kundu, S. C., and Wang, X. (2013). Silk fibroin biomaterials for tissue regenerations. *Adv. Drug Deliv. Rev.* 65 (4), 457–470. doi:10.1016/j.addr.2012.09.043
- Kundu, S. C., Kundu, B., Talukdar, S., Bano, S., Nayak, S., Kundu, J., et al. (2012). Nonmulberry silk biopolymers. *Biopolymers* 97 (6), 455–467. doi:10.1002/bip.22024
- Lee, C. F., Hsu, Y. H., Lin, Y. C., Nguyen, T. T., Chen, H. W., Nabilla, S. C., et al. (2021). 3D printing of collagen/oligomeric proanthocyanidin/oxidized hyaluronic acid composite scaffolds for articular cartilage repair. *Polym. (Basel)* 13 (18), 3123. doi:10.3390/polym13183123
- Lee, J. M., Suen, S. K. Q., Ng, W. L., Ma, W. C., and Yeong, W. Y. (2021). Bioprinting of collagen: Considerations, potentials, and applications. *Macromol. Biosci.* 21 (1), e2000280. doi:10.1002/mabi.202000280
- Leppiniemi, J., Lahtinen, P., Paajanen, A., Mahlberg, R., Metsa-Kortelainen, S., Pinomaa, T., et al. (2017). 3D-Printable bioactivated nanocellulose-alginate hydrogels. *ACS Appl. Mater. Interfaces* 9 (26), 21959–21970. doi:10.1021/acsmi.7b02756
- Li, D., Cheng, Y., Shahzadi, I., Jiang, G., Yi, Y., Shi, X., et al. (2019). Egg source natural proteins LBL modified cellulose nanofibrous mats and their cellular compatibility. *Carbohydr. Polym.* 213, 329–337. doi:10.1016/j.carbpol.2019.02.096
- Li, J., Fang, T., Yan, W., Zhang, F., Xu, Y., and Du, Z. (2020). Structure and properties of oxidized chitosan grafted cashmere fiber by amide covalent modification. *Molecules* 25 (17), 3812. doi:10.3390/molecules25173812
- Li, Q., Guan, X., Cui, M., Zhu, Z., Chen, K., Wen, H., et al. (2018). Preparation and investigation of novel gastro-floating tablets with 3D extrusion-based printing. *Int. J. Pharm.* 535 (1–2), 325–332. doi:10.1016/j.ijpharm.2017.10.037
- Li, X. H., Zhu, X., Liu, X. Y., Xu, H. H., Jiang, W., Wang, J. J., et al. (2021). The corticospinal tract structure of collagen/silk fibroin scaffold implants using 3D printing promotes functional recovery after complete spinal cord transection in rats. *J. Mater. Sci. Mater. Med.* 32 (4), 31. doi:10.1007/s10856-021-06500-2
- Li, Z., Wu, N., Cheng, J., Sun, M., Yang, P., Zhao, F., et al. (2020). Biomechanically, structurally and functionally meticulously tailored polycaprolactone/silk fibroin scaffold for meniscus regeneration. *Theranostics* 10 (11), 5090–5106. doi:10.7150/thno.44270
- Liedel, C. (2020). Sustainable battery materials from biomass. *ChemSusChem* 13 (9), 2110–2141. doi:10.1002/cssc.201903577
- Lin, H. H., Chao, P. G., Tai, W. C., and Chang, P. C. (2021). 3D-Printed collagen-based waveform microfibrous scaffold for periodontal ligament reconstruction. *Int. J. Mol. Sci.* 22 (14), 7725. doi:10.3390/ijms22147725
- Ling, K., Wu, H., Neish, A. S., and Champion, J. A. (2019). Alginate/chitosan microparticles for gastric passage and intestinal release of therapeutic protein nanoparticles. *J. Control Release* 295, 174–186. doi:10.1016/j.jconrel.2018.12.017
- Liu, H., Peng, H., Wu, Y., Zhang, C., Cai, Y., Xu, G., et al. (2013). The promotion of bone regeneration by nanofibrous hydroxyapatite/chitosan scaffolds by effects on integrin-BMP/Smad signaling pathway in BMSCs. *Biomaterials* 34 (18), 4404–4417. doi:10.1016/j.biomaterials.2013.02.048

- Liu, H., Wang, C., Sun, X., Zhan, C., Li, Z., Qiu, L., et al. (2022). Silk fibroin/collagen/hydroxyapatite scaffolds obtained by 3D printing technology and loaded with recombinant human erythropoietin in the reconstruction of alveolar bone defects. *ACS Biomater. Sci. Eng.* 8, 5245–5256. doi:10.1021/acsbomaterials.2c00690
- Liu, J., Song, P., Zhang, J., Nangong, Z., Liu, X., Gao, Y., et al. (2019a). Characteristics and function of the chitin binding protein from xenorhabdus nematophila. *Protein Pept. Lett.* 26 (6), 414–422. doi:10.2174/0929866526666190327143335
- Liu, J., Sun, L., Xu, W., Wang, Q., Yu, S., and Sun, J. (2019b). Current advances and future perspectives of 3D printing natural-derived biopolymers. *Carbohydr. Polym.* 207, 297–316. doi:10.1016/j.carbpol.2018.11.077
- Liu, X., Zheng, C., Luo, X., Wang, X., and Jiang, H. (2019c). Recent advances of collagen-based biomaterials: Multi-hierarchical structure, modification and biomedical applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* 99, 1509–1522. doi:10.1016/j.msec.2019.02.070
- Longinotti, C. (2014). The use of hyaluronic acid based dressings to treat burns: A review. *Burns Trauma* 2 (4), 162–168. doi:10.4103/2321-3868.142398
- Luo, H., Lan, H., Cha, R., Yu, X., Gao, P., Zhang, P., et al. (2021). Dialdehyde nanocrystalline cellulose as antibiotic substitutes against multidrug-resistant bacteria. *ACS Appl. Mater. Interfaces* 13 (29), 33802–33811. doi:10.1021/acscami.1c06308
- Ma, L., Wang, X., Zhao, N., Zhu, Y., Qiu, Z., Li, Q., et al. (2018). Integrating 3D printing and biomimetic mineralization for personalized enhanced osteogenesis, angiogenesis, and osteointegration. *ACS Appl. Mater. Interfaces* 10 (49), 42146–42154. doi:10.1021/acscami.8b17495
- Mahedia, M., Shah, N., and Amirlak, B. (2016). Clinical evaluation of hyaluronic acid sponge with zinc versus placebo for scar reduction after breast surgery. *Plast. Reconstr. Surg. Glob. Open* 4 (7), e791. doi:10.1097/GOX.0000000000000747
- Markstedt, K., Mantas, A., Tournier, I., Martinez Avila, H., Hagg, D., and Gatenholm, P. (2015). 3D bioprinting human chondrocytes with nanocellulose-alginate bioink for cartilage tissue engineering applications. *Biomacromolecules* 16 (5), 1489–1496. doi:10.1021/acs.biomac.5b00188
- Matica, M. A., Aachmann, F. L., Tondervik, A., Sletta, H., and Ostafe, V. (2019). Chitosan as a wound dressing starting material: Antimicrobial properties and mode of action. *Int. J. Mol. Sci.* 20 (23), 5889. doi:10.3390/ijms20235889
- Maturavongsadit, P., Narayanan, L. K., Chansoria, P., Shirwaiker, R., and Benhabbour, S. R. (2021). Cell-Laden nanocellulose/chitosan-based bioinks for 3D bioprinting and enhanced osteogenic cell differentiation. *ACS Appl. Bio Mater* 4 (3), 2342–2353. doi:10.1021/acscbam.0c01108
- Mehrotra, S., Chouhan, D., Konwarh, R., Kumar, M., Jadi, P. K., and Mandal, B. B. (2019). Comprehensive review on silk at nanoscale for regenerative medicine and allied applications. *ACS Biomater. Sci. Eng.* 5 (5), 2054–2078. doi:10.1021/acsbomaterials.8b01560
- Meng, X. H., Yang, L. Y., Kennedy, J. F., and Tian, S. P. (2010). Effects of chitosan and oligochitosan on growth of two fungal pathogens and physiological properties in pear fruit. *Carbohydr. Polym.* 81 (1), 70–75. doi:10.1016/j.carbpol.2010.01.057
- Mohammadinejad, R., Karimi, S., Iravani, S., and Varma, R. S. J. G. C. (2016). Plant-derived nanostructures: Types and applications. *Plant-derived nanostructures types Appl.* 18 (1), 20–52. doi:10.1039/c5cg01403d
- Mostafalu, P., Kiaee, G., Giatsidis, G., Khalilpour, A., Nabavinia, M., Dokmeci, M. R., et al. (2017). A textile dressing for temporal and dosage controlled drug delivery. *Adv. Funct. Mater.* 27 (41), 1702399. doi:10.1002/adfm.201702399
- Muller, M., Urban, B., Reis, B., Yu, X., Grab, A. L., Cavalcanti-Adam, E. A., et al. (2018). Switchable release of bone morphogenetic protein from thermoresponsive poly(NIPAM-co-dmaema)/cellulose sulfate particle coatings. *Polym. (Basel)* 10 (12), 1314. doi:10.3390/polym10121314
- Ng, W. L., Chua, C. K., and Shen, Y.-F. (2019). Print me an organ! Why we are not there yet. *Prog. Polym. Sci.* 97, 101145. doi:10.1016/j.progpolymsci.2019.101145
- Oprea, M., and Voicu, S. I. (2020). Recent advances in composites based on cellulose derivatives for biomedical applications. *Carbohydr. Polym.* 247, 116683. doi:10.1016/j.carbpol.2020.116683
- Palaganas, N. B., Mangadla, J. D., De Leon, A. C. C., Palaganas, J. O., Pangilinan, K. D., Lee, Y. J., et al. (2017). 3D printing of photocurable cellulose nanocrystal composite for fabrication of complex architectures via stereolithography. *ACS Appl. Mater. Interfaces* 9 (39), 34314–34324. doi:10.1021/acscami.7b09223
- Peptu, C., and Kowalczyk, M. (2018). “8 - biomass-derived polyhydroxyalkanoates: Biomedical applications,” in *Biomass as renewable raw material to obtain bioproducts of high-tech value*. Editors V. Popa and I. Volf (Amsterdam, Netherlands: Elsevier), 271–313.
- Pleissner, D., and Kummerer, K. (2020). Green chemistry and its contribution to industrial biotechnology. *Adv. Biochem. Eng. Biotechnol.* 173, 281–298. doi:10.1007/10_2018_73
- Przekora, A. (2019). The summary of the most important cell-biomaterial interactions that need to be considered during *in vitro* biocompatibility testing of bone scaffolds for tissue engineering applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* 97, 1036–1051. doi:10.1016/j.msec.2019.01.061
- Rastogi, P., and Kandasubramanian, B. (2019). Review of alginate-based hydrogel bioprinting for application in tissue engineering. *Biofabrication* 11 (4), 042001. doi:10.1088/1758-5090/ab331e
- Romanazzi, G., Feliziani, E., and Sivakumar, D. (2018). Chitosan, a biopolymer with triple action on postharvest decay of fruit and vegetables: Eliciting, antimicrobial and film-forming properties. *Front. Microbiol.* 9, 2745. doi:10.3389/fmicb.2018.02745
- Roseti, L., Parisi, V., Petretta, M., Cavallo, C., Desando, G., Bartolotti, I., et al. (2017). Scaffolds for bone tissue engineering: State of the art and new perspectives. *Mater. Sci. Eng. C Mater. Biol. Appl.* 78, 1246–1262. doi:10.1016/j.msec.2017.05.017
- Sanchez-Ballester, N. M., Bataille, B., and Soulaïrol, I. (2021). Sodium alginate and alginate acid as pharmaceutical excipients for tablet formulation: Structure-function relationship. *Carbohydr. Polym.* 270, 118399. doi:10.1016/j.carbpol.2021.118399
- Schutz, K., Placht, A. M., Paul, B., Bruggemeier, S., Gelsinsky, M., and Lode, A. (2017). Three-dimensional plotting of a cell-laden alginate/methylcellulose blend: Towards biofabrication of tissue engineering constructs with clinically relevant dimensions. *J. Tissue Eng. Regen. Med.* 11 (5), 1574–1587. doi:10.1002/term.2058
- Shi, W., Sun, M., Hu, X., Ren, B., Cheng, J., Li, C., et al. (2017). Structurally and functionally optimized silk-fibroin-gelatin scaffold using 3D printing to repair cartilage injury *in vitro* and *in vivo*. *Adv. Mater* 29 (29), 1701089. doi:10.1002/adma.201701089
- Shoulders, M. D., and Raines, R. T. (2009). Collagen structure and stability. *Annu. Rev. Biochem.* 78, 929–958. doi:10.1146/annurev.biochem.77.032207.120833
- Simoes, D., Miguel, S. P., Ribeiro, M. P., Coutinho, P., Mendonca, A. G., and Correia, I. J. (2018). Recent advances on antimicrobial wound dressing: A review. *Eur. J. Pharm. Biopharm.* 127, 130–141. doi:10.1016/j.ejpb.2018.02.022
- Son, B. T., Long, N. V., and Nhat Hang, N. T. (2021). The development of biomass-derived carbon-based photocatalysts for the visible-light-driven photodegradation of pollutants: A comprehensive review. *RSC Adv.* 11 (49), 30574–30596. doi:10.1039/d1ra05079f
- Sorushanova, A., Delgado, L. M., Wu, Z., Shologu, N., Kshirsagar, A., Raghunath, R., et al. (2019). The collagen suprafamily: From biosynthesis to advanced biomaterial development. *Adv. Mater* 31 (1), e1801651. doi:10.1002/adma.201801651
- Srivastava, R. K., Shetti, N. P., Reddy, K. R., Kwon, E. E., Nadagouda, M. N., and Aminabhavi, T. M. (2021). Biomass utilization and production of biofuels from carbon neutral materials. *Environ. Pollut.* 276, 116731. doi:10.1016/j.envpol.2021.116731
- Stiglic, A. D., Gurer, F., Lackner, F., Bracic, D., Winter, A., Gradisnik, L., et al. (2022). Organic acid cross-linked 3D printed cellulose nanocomposite bioscaffolds with controlled porosity, mechanical strength, and biocompatibility. *iScience* 25 (5), 104263. doi:10.1016/j.isci.2022.104263
- Su, H., Li, Q., Li, D., Li, H., Feng, Q., Cao, X., et al. (2022). A versatile strategy to construct free-standing multi-furcated vessels and a complicated vascular network in heterogeneous porous scaffolds via combination of 3D printing and stimuli-responsive hydrogels. *Mater. Horiz.* 9 (9), 2393–2407. doi:10.1039/d2mh00314g
- Su, X., Wang, T., and Guo, S. (2021). Applications of 3D printed bone tissue engineering scaffolds in the stem cell field. *Regen. Ther.* 16, 63–72. doi:10.1016/j.reth.2021.01.007
- Sun, W., Gregory, D. A., Tomeh, M. A., and Zhao, X. (2021). Silk fibroin as a functional biomaterial for tissue engineering. *Int. J. Mol. Sci.* 22 (3), 1499. doi:10.3390/ijms22031499
- Sun, Y., Yang, C., Zhu, X., Wang, J. J., Liu, X. Y., Yang, X. P., et al. (2019). 3D printing collagen/chitosan scaffold ameliorated axon regeneration and neurological recovery after spinal cord injury. *J. Biomed. Mater. Res. A* 107 (9), 1898–1908. doi:10.1002/jbm.a.36675
- Talebian, S., Foroughi, J., Wade, S. J., Vine, K. L., Dolatshahi-Pirouz, A., Mehrali, M., et al. (2018). Biopolymers for antitumor implantable drug delivery systems: Recent advances and future outlook. *Adv. Mater* 30 (31), e1706665. doi:10.1002/adma.201706665
- Tavianatou, A. G., Caon, I., Franchi, M., Piperigkou, Z., Galesso, D., and Karamanos, N. K. (2019). Hyaluronan: Molecular size-dependent signaling and biological functions in inflammation and cancer. *FEBS J.* 286 (15), 2883–2908. doi:10.1111/febs.14777
- Tiwari, S., and Bahadur, P. (2019). Modified hyaluronic acid based materials for biomedical applications. *Int. J. Biol. Macromol.* 121, 556–571. doi:10.1016/j.ijbiomac.2018.10.049
- Tsukamoto, Y., Akagi, T., and Akashi, M. (2020). Vascularized cardiac tissue construction with orientation by layer-by-layer method and 3D printer. *Sci. Rep.* 10 (1), 5484. doi:10.1038/s41598-020-59371-y
- Tu, H., Wu, G., Yi, Y., Huang, M., Liu, R., Shi, X., et al. (2019). Layer-by-layer immobilization of amphoteric carboxymethyl chitosan onto biocompatible silk fibroin nanofibrous mats. *Carbohydr. Polym.* 210, 9–16. doi:10.1016/j.carbpol.2019.01.047
- Tungprapa, S., Jangchud, I., and Supaphol, P. (2007). Release characteristics of four model drugs from drug-loaded electrospun cellulose acetate fiber mats. *Polymer* 48 (17), 5030–5041. doi:10.1016/j.polymer.2007.06.061
- Vanderburgh, J., Sterling, J. A., and Guelcher, S. A. (2017). 3D printing of tissue engineered constructs for *in vitro* modeling of disease progression and drug screening. *Ann. Biomed. Eng.* 45 (1), 164–179. doi:10.1007/s10439-016-1640-4
- Vehlow, D., Schmidt, R., Gebert, A., Siebert, M., Lips, K. S., and Muller, M. (2016). Polyelectrolyte complex based interfacial drug delivery system with controlled loading and improved release performance for bone therapeutics. *Nanomater. (Basel)* 6 (3), 53. doi:10.3390/nano6030053
- Venkata Mohan, S., Nikhil, G. N., Chiranjeevi, P., Nagendranatha Reddy, C., Rohit, M. V., Kumar, A. N., et al. (2016). Waste biorefinery models towards sustainable circular bioeconomy: Critical review and future perspectives. *Bioresour. Technol.* 215, 2–12. doi:10.1016/j.biortech.2016.03.130

- Ventola, C. L. (2014). Medical applications for 3D printing: Current and projected uses. *P T 39* (10), 704–711.
- Verlee, A., Mincke, S., and Stevens, C. V. (2017). Recent developments in antibacterial and antifungal chitosan and its derivatives. *Carbohydr. Polym.* 164, 268–283. doi:10.1016/j.carbpol.2017.02.001
- Wang, S., Yan, F., Ren, P., Li, Y., Wu, Q., Fang, X., et al. (2020). Incorporation of metal-organic frameworks into electrospun chitosan/poly (vinyl alcohol) nanofibrous membrane with enhanced antibacterial activity for wound dressing application. *Int. J. Biol. Macromol.* 158, 9–17. doi:10.1016/j.ijbiomac.2020.04.116
- Wutticharoenmongkol, P., Hannirojram, P., and Nuthong, P. (2019). Gallic acid-loaded electrospun cellulose acetate nanofibers as potential wound dressing materials. *Polym. Adv. Technol.* 30, 1135–1147. doi:10.1002/pat.4547
- Xi, Y., Ge, J., Guo, Y., Lei, B., and Ma, P. X. (2018). Biomimetic elastomeric polypeptide-based nanofibrous matrix for overcoming multidrug-resistant bacteria and enhancing full-thickness wound healing/skin regeneration. *ACS Nano* 12 (11), 10772–10784. doi:10.1021/acsnano.8b01152
- Xi, Y., Ge, J., Wang, M., Chen, M., Niu, W., Cheng, W., et al. (2020). Bioactive anti-inflammatory, antibacterial, antioxidative silicon-based nanofibrous dressing enables cutaneous tumor photothermo-chemo therapy and infection-induced wound healing. *ACS Nano* 14 (3), 2904–2916. doi:10.1021/acsnano.9b07173
- Xia, J., Zhang, H., Yu, F., Pei, Y., and Luo, X. (2020). Superclear, porous cellulose membranes with chitosan-coated nanofibers for visualized cutaneous wound healing dressing. *ACS Appl. Mater. Interfaces* 12 (21), 24370–24379. doi:10.1021/acscami.0c05604
- Yadav, M., Goel, G., Hatton, F. L., Bhagat, M., Mehta, S. K., Mishra, R. K., et al. (2021). A review on biomass-derived materials and their applications as corrosion inhibitors, catalysts, food and drug delivery agents. *Curr. Res. Green Sustain. Chem.* 4, 100153. doi:10.1016/j.crgsc.2021.100153
- Yang, Y., Li, W., Yu, D. G., Wang, G., Williams, G. R., and Zhang, Z. (2019). Tunable drug release from nanofibers coated with blank cellulose acetate layers fabricated using tri-axial electrospinning. *Carbohydr. Polym.* 203, 228–237. doi:10.1016/j.carbpol.2018.09.061
- Yao, M., Zou, Q., Zou, W., Xie, Z., Li, Z., Zhao, X., et al. (2021). Bifunctional scaffolds of hydroxyapatite/poly(dopamine)/carboxymethyl chitosan with osteogenesis and anti-osteosarcoma effect. *Biomater. Sci.* 9 (9), 3319–3333. doi:10.1039/d0bm01785j
- Yao, X., Zou, S., Fan, S., Niu, Q., and Zhang, Y. (2022). Bioinspired silk fibroin materials: From silk building blocks extraction and reconstruction to advanced biomedical applications. *Mater Today Bio* 16, 100381. doi:10.1016/j.mtbio.2022.100381
- Yi, H. G., Jeong, Y. H., Kim, Y., Choi, Y. J., Moon, H. E., Park, S. H., et al. (2019). A bioprinted human-glioblastoma-on-a-chip for the identification of patient-specific responses to chemoradiotherapy. *Nat. Biomed. Eng.* 3 (7), 509–519. doi:10.1038/s41551-019-0363-x
- Yuan, M., Dai, F., Li, D., Fan, Y., Xiang, W., Tao, F., et al. (2020). Lysozyme/collagen multilayers layer-by-layer deposited nanofibers with enhanced biocompatibility and antibacterial activity. *Mater Sci. Eng. C Mater Biol. Appl.* 112, 110868. doi:10.1016/j.msec.2020.110868
- Zamboni, F., Vieira, S., Reis, R. L., Oliveira, J. M., and Collins, M. N. (2018). The potential of hyaluronic acid in immunoprotection and immunomodulation: Chemistry, processing and function. *Prog. Mater. Sci.* 97, 97–122. doi:10.1016/j.pmatsci.2018.04.003
- Zhang, X., Yuan, J., Li, F., and Xiang, J. (2021). Chitin synthesis and degradation in Crustaceans: A genomic view and application. *Mar. Drugs* 19 (3), 153. doi:10.3390/md19030153
- Zhou, J., Zhang, B., Liu, X., Shi, L., Zhu, J., Wei, D., et al. (2016). Facile method to prepare silk fibroin/hyaluronic acid films for vascular endothelial growth factor release. *Carbohydr. Polym.* 143, 301–309. doi:10.1016/j.carbpol.2016.01.023
- Zhu, X., Hou, X., Ma, B., Xu, H., and Yang, Y. (2019). Chitosan/gallnut tannins composite fiber with improved tensile, antibacterial and fluorescence properties. *Carbohydr. Polym.* 226, 115311. doi:10.1016/j.carbpol.2019.115311
- Zhu, Z., Wang, Y. M., Yang, J., and Luo, X. S. (2017). Hyaluronic acid: A versatile biomaterial in tissue engineering. *Plastic Aesthetic Res.* 4, 219–227. doi:10.20517/2347-9264.2017.71