

Fluorescence Quenching of Graphene Quantum Dots by Chloride Ions: A Potential Optical Biosensor for Cystic Fibrosis

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Ifrah Z, Shah Rukh A, Muhammad Nauman S, Maryam S and Rahat U (2022) Fluorescence Quenching of Graphene Quantum Dots by Chloride Ions: A Potential Optical Biosensor for Cystic Fibrosis. Front. Mater. 9:857432. doi: 10.3389/fmats.2022.857432 Cystic fibrosis is a genetic disorder caused by a mutation in the cystic fibrosis transmembrane conductance regulator (CFTR) gene that encodes an ABC transporterclass ion channel protein, resulting in sticky and thick mucus secretion which clogs the airways and leads to respiratory failure and other complications. It also results in higher chloride ions (CI7) in body fluids. Cystic fibrosis is generally detected using the sweat chloride test and ion exchange chromatography, which are lab restricted. Therefore, there is a dire need to develop portable assays to monitor circulatory changes (Cl⁻ ion detection) to detect CF at the point of care. In this work, fluorescence quenching of graphene quantum dots (GQDs) was used as a property of the optical sensor for chloride ion detection. GQDs were synthesized by varying the carbonizing temperature and time, and then their optical and fluorescence (FL) guenching was investigated upon exposure to chloride ions in comparison with different ionic species. GQDs synthesized at 160°C for 50 min were chosen as they displayed the highest fluorescence. The morphological and optical characterization confirmed the preparation of 12-15 nm GQDs, which were amorphous in nature with the peak emission observed at 462 nm when excited at 370 nm. The fluorescence quenching response of GQDs with Cl⁻ ions displayed linearity up to 100 mM with a correlation coefficient of 0.98 and the lowest detection limit of approximately 10 mM Cl⁻ ions.

Keywords: graphene quantum dot (GQD), cystic fbrosis, chloride ions (CI-), fluorescence quenching, citric acid

1 INTRODUCTION

Point-of-care diagnostics has attracted attention in the past few years because of the early and rapid detection of biomarkers for disease management and monitoring of diseases (Vashist 2017; Golubnitschaja 2009). Chloride is an essential electrolyte within the body fluids, and its elevated level in the serum, urine, cerebral spinal fluid, or sweat indicates cystic fibrosis (De Gregorio et al., 2014). Cystic fibrosis is a recessive genetic disorder, which induces defective ion transfer across the epithelial membranes. It has been reported that at least 70% of CF patients have the cystic fibrosis transmembrane conductance regulator (CFTR) gene with the major mutation delta F508 (Cui et al., 1995). The CFTR gene codes for an ATP-binding cassette transporter (ABC transporter) ion channel

protein that conducts chloride and thiocyanate ions across the epithelial cell membrane (Wang et al., 2011). Mutations of the CFTR gene elevate the concentration of chloride ions up to 60 mM and above in the epithelial fluids. The threshold value of chloride ions in these fluids is less than 20 mM (Quinton 1983). This results in the reduced chloride reabsorption from sweat that causes a sticky mucous secretion in multiple organs (Esteves et al., 2018). Most of the CF patients suffer from reduced lung function, change in overall physiological function, and increase in pancreaticobiliary and gastrointestinal tract malignancies (Simmonds et al., 2009; Horsley and Siddiqui 2015; Ooi and Durie 2016). Therefore, early detection of disease is of utmost importance to improve health and life expectancy of the patients.

The sweat chloride test is a conventional method used for the detection of CF, which is time consuming, requires skilled individuals, and prone to errors with very high false-negative rates at 12% (Farrell et al., 2001; LeGrys et al., 2009). Sweat analysis is non-invasive substitute to blood analysis which allows continuous monitoring of body parameters without making the patients uncomfortable. Moreover, high sensitivity and selectivity can be achieved using ion exchange chromatography technique, but it requires high maintenance cost and skilled operators. The other chloride analysis methods that include coulometry, colorimetry, and ion-selective electrodes (ISE) are not preferred because of low halide selectivity (McClatchey 2002). For example, iodide and bromide produce greater signals than that of chloride in ISE measurements (Bray et al., 1977). So, there is a dire need to develop a fully automated, low cost, sensitive and selective

rapid chloride sensing technology, which can be translated to clinical settings (Yager et al., 2008).

GQDs are zero-dimensional (0D) material that exhibit extraordinary electronic and optical properties due to their edge effects and quantum confinement (Sun et al., 2013; Lin et al., 2014). Recently, graphene quantum dots have been widely used as fluorescent probes for the development of luminescent sensing assays (Li and Zhu 2013; Fan et al., 2015). Among other carbon allotropes graphene quantum dots (GQDs) display enthralling characteristics such as low-cost, non-toxicity, excellent biocompatibility, and photoluminescent stability (Li et al., 2013; Ganguly et al., 2019; Saravanan et al., 2020; Das et al., 2021; Bhattacharyya et al., 2020). The photoluminescent emission of graphene quantum dots can be extensively changed from the deep ultraviolet region to the near infrared region by regulating the shape, size, defects, hybridization of carbon lattice, functional groups, edge configuration, or by tuning the excitation wavelength or concentration of GQDs (Lai et al., 2020; Wang et al., 2016). A number of studies have confirmed the fluorescent quenching of GQDs (Saravanan et al., 2020; Das et al., 2018). The guenching mechanism between quantum dots and metal ions are mainly divided into Förster resonance energy transfer (FRET), the inner filter effect (IFE), and the dynamic and static quenching process (Qi et al., 2015). Usually, GQDs can be synthesized by two approaches, which are the top-down approach and the bottom-up approach. In the top-down approach, the zero-dimensional graphene quantum dots are fabricated from cutting of a particular carbon source material such as graphite, graphene oxide, carbon black, and carbon fiber (Ponomarenko et al., 2008). The disadvantages of



FIGURE 1 | Schematic diagram of the mechanism used for the synthesis of graphene quantum dots from citric acid.



synthesis *via* top-down approach includes critical synthesis conditions, difficulty in controlling the size distribution of products, which results in the formation of disordered and defective structures, and poorly crystalline nanomaterials (Dong et al., 2012a). In the bottom-up approach, condensation of smaller units of carbon derivatives results in the formation of bigger units of GQDs. The processes involve pyrolysis, oxidation, and hydrothermal heating. The most important advantage of the bottom-up approach is that it allows precise control over the morphology and the size distribution of the products (Wu et al., 2007).

In this study we explored the photoluminescence intensity of GQDs to monitor the chloride changes in cystic fibrosis. GQDs were prepared by optimizing the reaction parameters for controlled surface passivation, prepared *via* pyrolysis of citric acid, which is a bottom-up approach. Physiochemical and morphological characterization methods displayed the size of an average of 15 nm and showed the characteristic peaks *via* UV–Vis spectroscopy, XRD, FTIR, and SEM/EDX. GQDs were found to be disc-shaped, amorphous particles displaying blue phospholuminescence, with the maximum emission peak at 462 nm. The photoluminescent quenching effect of GQD was





observed by the addition of chloride ions which showed linearity in the range of 5–90 mM. The results of this study show that the fluorescent behavior of the GQDs makes possible the development of rapid, simple, and effective tests for the detection of chloride ions.

2 MATERIAL AND METHODS

2.1 Materials

Citric acid, sodium hydroxide (NaOH), sodium chloride (NaCl), and potassium chloride (KCl) were purchased from Daejung Chemicals and Metals. Potassium acetate, potassium nitrate, sodium carbonate, magnesium sulfate, and sodium phosphate were procured from Sigma-Aldrich. Hydrochloric acid (HCL) was obtained from BDH Laboratory Supplies, England. Deionized water was used throughout the experiment.

2.2 Preparation of Graphene Quantum Dots

Six different fluorescent GOD samples were synthesized using the carbonization method. Briefly, a small flask containing 2 g citric acid was heated from 160 to 200 C with a selected reaction time of 30 and 50 min for each temperature variation as shown in Supplementary Table S1. The citric acid liquidates after 5 min, and slowly turns from colorless to pale yellow to yellowish orange, which is the visual cue that the pyrolysis is occurring. After the initiation of the reaction, the dropwise addition of this yellowish orange solution into 100 ml of 10 mg/ml NaOH, at a constant temperature was carried out. The aqueous solution of GQD was obtained. It was further purified by centrifugation at 15,000 rpm for 30 min, and then GQDs were obtained in a pellet form. The schematic overview of the mechanism involved for synthesizing GQDs is shown in Figure 1. Different temperature (160, 180, and 200°C) variations were performed for the synthesis of GODs keeping in mind the thermogravimetric analysis (TGA) of citric acid. The samples were named according to their reaction conditions: the temperature followed by the reaction time in brackets, (i.e., temperature-time).

2.3 Morphological Characterization

A scanning electron microscope (SEM) and EDX analysis were performed to obtain information about the surface topography and elemental composition of graphene quantum dots. Similarly, X-ray diffraction (XRD) was performed using an STOE Powder Diffractometer θ - θ (STOE Inc. Germany) in order to ensure the morphological nature, that is, crystallinity or amorphous nature of GQDs. Moreover, Fourier-transform infrared spectroscopy (FTIR) was used to obtain an infrared spectrum of absorption or emission of the samples, which reflects the bonding interaction in materials.

2.4 Optical Characterization

The optical properties of the GQDs were initially determined under the excitation of a handheld 365 nm ultra-violet (UV) lamp (UVP) and a UV–Vis spectrophotometer (UVD-2950) was used to collect the absorption spectra in the range of 200–800 nm. Fluorescence properties of each graphene quantum dot sample were confirmed





using a fluorescence spectrometer (Horiba Fluoromax-4). As an excitation source, this system is equipped with a 150 W continuous wavelength xenon arc lamp, two Czerny–Turner monochromators for excitation and emission selection, adjustable slits in front of each monochromator, and a detector (R928P photomultiplier tube) whose response ranges from 180 to 850 nm. The slit width of excitation monochromator determines the bandpass of light incident on the sample whereas the emission monochromator's slits control the intensity of the fluorescence signal recorded by the signal detector. Throughout the experiment the slit widths on excitation and emission were adjusted at 2 nm.

Furthermore, the excitation monochromator was set at 370 nm and emissions were acquired within a range of 410-600 nm. The emission spectra were taken using the front-face fluorescence (FFF) sample geometry of the spectrometer setup. The cuvette was fixed in the sample holder such that the excitation angle was set at 57° as recommended in the setup manual. A total of 50 mM concentration of each ion was prepared, and 1ml of ion sample was mixed with 2 ml of (6 mg/ml) GQD sample in a cuvette and was incubated for 5 min. All the emission spectra were acquired with varying concentrations of ions ranging from 10 to 200 mM. The

quenching fluorescence patterns of graphene quantum dots with different concentrations of sodium chloride were analyzed. Every sample was analyzed as triplicates and the resultant graphs are an average of those spectra. To ensure that quenching effect observed was specific to only chloride ions, the quenching of GQDs were compared with another chloride-containing compound that is, potassium chloride (KCl).

3 RESULTS

3.1 Morphological Characterization Results

The surface morphology and the size of the GQD were analyzed using the scanning electron microscope (SEM) (Tescan Vega 3). Samples were prepared by diluting the as-prepared GQDs in the ratio 1:5, in deionized water and the diluent was dropped on a glass slide. Scanning was performed at 20 kV, with the maximum magnification of ×65000, at the working distance of 5.07 mm. Figures 2A-D shows the images of GQD surface zoomed at different resolutions (20 um, 5 um, 1 um, and 500 nm). The size of the GQD as seen in the (c) image is approximately measured as 15.78 nm and can be seen as being disc-like particles, as observed in Figure 2D. Similarly, EDX analysis further confirms that the fluorescence intensity is strictly due to the graphene quantum dots and not due to the presence of some impurities. However, a small percentage of sodium (Na) was also observed which can be attributed to NaOH, which was the dispersion solution of GQD. Most likely it could have been left behind from the centrifugation step, during the sample preparation stage. The observed atomic percentage of carbon was 40.72%, oxygen was 45.70%, and sodium was 14.08%.

X-ray diffraction (XRD) was performed to ensure the morphological nature that is, crystallinity or amorphous nature of the graphene quantum dots. XRD of GQD was performed using the STOE Powder Diffractometer θ - θ (STOE Inc. Germany) with current 40 Ma and operating voltage 40Kv at scan rate 0.5°/min. The XRD pattern of GQDs shows a broad peak at around $2\theta \sim 32$ indicating the amorphous nature of the GQDs (Li et al., 2018; Kundu and Pillai 2020) as seen in **Figure 3**. As calculated by Bragg's equation the lattice size is 0.202 nm, suggesting that the carbonization of citric acid produced graphite structures, which has compact interlayer spacing. The small particle size is also evident from the broad peak. The absence of any peak characterized for graphene oxide and graphite, which is a sharp peak usually appears around $2\Theta = 19^{\circ}$ indicate that the prepared sample is indeed a graphene quantum dot. As reported, GQD exhibits absorption of carboxyl as well as hydroxyl



group showing that nanomaterial contains -COOH groups at its surfaces. In **Figure 4** is shown the FTIR spectra of GQDs which revealed the broadening of several peaks related to oxygen functional groups. The band appeared at 3,438 cm⁻¹ is assigned to the stretching vibration of the O–H group (Qi et al., 2015). Similarly, 1,612 cm⁻¹ absorption band is attributed to aromatic bending of C=O. Both these bands correspond to the complete dehydration of citric acid to GQD. An absorption FTIR band at 1,275 cm⁻¹ relates to the stretching vibration of hydroxyl groups. Furthermore, the absorption band shown at 1,082 cm⁻¹ is due to V_{c-o} and peak at 1,403 cm⁻¹ suggests the existence of C-H bond (Zhu et al., 2011). The band detected at 823 cm⁻¹ is attributed to CH₂ rocking vibration (Badatya et al., 2021; Tang et al., 2012). The stretching vibrations of C-H and C-O at 2,917 cm⁻¹ and 1,275 cm⁻¹ indicate that GQDs contain some incompletely carbonized citric acid.

3.2 Optical Results

The graphene quantum dots prepared at different carbonization temperature and time were subjected to irradiation by a UV-lamp to confirm their fluorescence. As evident from **Figure 5B**, the prepared samples displayed blue-colored luminescence under the 365 nm wavelength UV lamp. Similarly, the aqueous GQDs suspension is yellow to brown in color (**Figure 5A**) and transparent in the daylight. Furthermore, as can be seen in

Figure 5A, in visible light the prepared GQD display a slight yellow hue in their aqueous solutions such that their intensity varies according to the reaction conditions. Likewise, the intensity of blue fluorescence also depends upon the reaction condition. Based on this observation, it can be seen that the GQD sample (160°C-50 min) displays the highest amount of fluorescence.

Similarly, the absorption spectrum of GQDs in water was analyzed using a spectrophotometer (UVD-2950). Usually, two types of characteristic features are observed in the spectra of GQD, the first is a shoulder at ~350 nm, corresponding to an $n-\pi^*$ plasmon peak. Another characteristic feature of GO appears at ~ 230 nm and corresponds to a $\pi-\pi^*$ plasmon peak (Shen et al., 2012). As shown in **Figure 6**, the two peaks were observed at 236 and 358 nm. The intense peak present at 236 nm corresponds to GQDs, which is due to the free zigzag sites present in GQDs.

In addition to that, **Figure 7A** depicts the fluorescence emission spectra under varying excitation of each prepared GQDs solution. These bands are mostly peaked at 460 nm confirming the fluorescence emission of GQD. Studies have shown that the size of GQDs is of critically important because it determines the excitation/emission band (Dong et al., 2012b). Usually in GQDs, numerous electronic transitional states occurred due to the molecular orbitals of C=C sp² carbon domains and amino edges functional groups, and that is the



reason of showing different emission spectra. As observed, the 160°C -50 min sample displays the highest fluorescence at 370 nm, so we chose this sample for our further experiments. **Figure 7B** corresponds to the excitation/emission spectrum of GQDs (160°C -50 min). It shows excitation maximum at 370 nm, whereas emission band maximum at 462 nm corresponding to the blue luminescence displayed. To further check the PL properties of GQDs, the effect of changing the excitation wavelength on the emission spectrum was analyzed. The excitation wavelength was varied in increasing in the increments of 10 such as 350, 360, 370, and 380 nm (**Figure 7C**). Moreover, from **Figure 7C** it is clearly observed that the emission band of 460 nm of the GQD is independent from the excitation wavelengths.

4 DISCUSSION

4.1 Graphene Quantum Dots as a Fluorescence Biosensor

A 50 mM mixture of anions and cations (sodium phosphate, potassium acetate, potassium nitrate, magnesium sulfate, sodium carbonate, and sodium hydroxide) were taken in order to draw comparison of the quenching exclusivity of chloride ions. As **Figure 8** demonstrates, sodium chloride shows considerable quenching compared to the other ionic compounds which proves that it exhibit quenching effect even in the presence of the other ions. Human specimens such as sweat are composed of a combination of multiple ions, it is important for the study to mimic that somehow. The carbonate ions present in the sodium carbonate adds certain compounds on the surface of the GQDs which causes an observable red shift in the spectra while the other compounds do not display any kind of shift in the emission spectra of graphene quantum dots.

Furthermore, the quenching fluorescence patterns of NaCl with GQDs were monitored with various concentrations of NaCl ranging from 10 to 200 mM as shown in **Figure 9**. GQDs displayed a linear quenching curve until 90 mM of chloride

ion concentration. It can be argued that at this concentration all the interacting sites are full and further addition causes the sites to clutter up showing an increased fluorescence. Moreover, the quenching capability of GQDs was also studied with potassium chloride, to ensure that the quenching effect is being observed due to chloride ions and not due to the cations. It can be seen in **Figure 10**, almost identical spectra were observed, hence confirming our initial observation that chloride ions are responsible for the quenching effect observed.

The attenuation of fluorescence in the quenching mechanism results from the presence of chloride ions, which lead to non-radiative relaxation of the excited graphene quantum dot. In accordance, dynamic quenching is characterized by linear dependence of fluorescence quenching; and non-linear behavior suggests that both the static and dynamic processes are taking place. In this study, the NaCl acts as a quencher; we plotted the peak fluorescence intensity (I_0/I) vs. chloride ion concentration.

The calibration equation is as follows:

$$Y\left(\frac{I_{\circ}}{I}\right) = 0.00459X(conc.) + 1.01459.$$

The results demonstrated a linear dependence the between Io/ I and Cl-concentration, between the concentration range of 10–90 mM as shown in **Figure 11**. By increasing the chloride concentration beyond this concentration non-linearity is observed, which suggests that both static and dynamic quenching processes are occurring (Lakowicz, 1983).

5 CONCLUSION

In this study, the optical properties of the GQDs are investigated to monitor the changes in the chloride ions for the detection of cystic fibrosis. The GQDs are prepared by pyrolysis of citric acid showing disc-shaped having an average size of 15 nm. Additionally, they are amorphous particles displaying blue phospholuminescence (PL) with



the maximum emission peak at 462 nm. The PL quenching effect of GQD was observed by the addition of chloride ions, which showed linearity in the range of 10–90 mM. Within this range, the GQD display a linear response to Cl^- with a correlation coefficient of 0.98. The sensitivity and specificity to chloride ions suggest the potential application of this easily synthesized and the green GQD-based

sensing system as an alternative route for the time-consuming method of laboratory sweat testing for the diagnosis of cystic fibrosis. It also provides an alternative to the expensive and toxic organic dyes that are rarely used in laboratory settings. This also paves the way for point-of-care testing, as the tests may be calibrated for hand-held fluorescence spectrophotometers.





DATA AVAILABILITY STATEMENT

Publicly available datasets were analyzed in this study. This data can be found here: The original contribution made in form of data generated is mentioned in the article. Any further information can be provided by corresponding author upon request.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmats.2022.857432/full#supplementary-material.

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