Check for updates

OPEN ACCESS

EDITED BY Khan M. G. Mostofa, Tianjin University, China

REVIEWED BY Eurico D'Sa, Louisiana State University, United States Shuting Liu, Kean University, United States

*CORRESPONDENCE Bita Sabbaghzadeh Sabbagh@andrew.cmu.edu

RECEIVED 13 May 2024 ACCEPTED 19 June 2024 PUBLISHED 23 July 2024

CITATION

Sabbaghzadeh B, Uher G and Upstill-Goddard R (2024) "Dynamics of chromophoric dissolved organic matter in the Atlantic Ocean: unravelling provincedependent relationships, optical complexity, and environmental influences". *Front. Mar. Sci.* 11:1432133. doi: 10.3389/fmars.2024.1432133

COPYRIGHT

© 2024 Sabbaghzadeh, Uher and Upstill-Goddard. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

"Dynamics of chromophoric dissolved organic matter in the Atlantic Ocean: unravelling province-dependent relationships, optical complexity, and environmental influences"

Bita Sabbaghzadeh^{1,2*}, Guenther Uher² and Robert Upstill-Goddard²

¹Department of Marine Chemistry, Leibniz Institute for Baltic Sea Research, Rostock, Germany, ²School of Natural and Environmental Sciences, Newcastle University, Newcastle, United Kingdom

We report on the spatial distributions and optical characteristics of chromophoric dissolved organic matter (CDOM) in the sea surface microlayer (SML), subsurface seawater (SSW), and water column profiles down to 500 m across a range of Atlantic Ocean biogeochemical provinces during two cruises of the UK Atlantic Meridional Transect program (AMT24 and AMT25). We measured the CDOM absorption coefficient at 300 nm, a_{CDOM} (300), and determined CDOM spectral slopes across two UV wavelength ranges: S_1 (275-295 nm) and S_2 (350-400 nm). We used spectral slope ratios ($S_R: S_1/S_2$) to infer CDOM source characteristics and transformation history. During both cruises, SML $a_{CDOM}(300)$ was highest in the Northern Hemisphere, particularly in the North Atlantic Drift Region (NADR). CDOM was always enriched in the SML, with enrichment factors (SML a_{CDOM} (300) / SSW $a_{\text{CDOM}}(300)$) ranging from 1.03 to 2.00, reflecting preferential accumulation of CDOM in the SML. We also found a significant inverse correlation between $a_{\text{CDOM}}(300)$ and S_1 in both the SML (Spearman's rank correlation coefficient, $r^2 = -0.75$, p < 0.001, n = 114) and water column profiles ($r^2 = -0.74$, p < 0.001, n = 845). Biogeochemical province-dependent variations in the relationships between CDOM and chlorophyll a were also observed. In high-latitude regions, elevated $a_{CDOM}(300)$ and low S_R values indicated a dominance of terrestrially-derived CDOM, whereas oligotrophic subtropical areas showed lower $a_{CDOM}(300)$ and higher S_R values, suggestive of aged, refractory, and photodegraded biologically-derived CDOM. Taken together, these findings reveal a complexity of drivers affecting CDOM distributions and spectral properties, which may limit the use of CDOM in predictive relationships in the oceans. However, the potential use of chlorophyll a as a CDOM proxy may prove most successful in open ocean regions devoid of terrestrial inputs, where biological production predominates.

KEYWORDS

chromophoric dissolved organic matter (CDOM), Atlantic Ocean, biogeochemical provinces, optical complexity, environmental influences

1 Introduction

Chromophoric dissolved organic matter (CDOM) strongly influences the optical characteristics of the water column, impacting ocean color and contributing to photochemical and photobiological processes, particularly in the UV range. It affects photosynthesis and light availability for ocean color remote sensing. CDOM absorption is comparable to, or even exceeds, light absorption attributable to phytoplankton pigments. CDOM thus has significant implications for monitoring and understanding aquatic ecosystems (e.g., Siegel et al., 2002; Twardowski et al., 2004; Siegel et al., 2005). Marine CDOM arises from diverse sources and subsequently undergoes complex transformations. Two broad principal hypotheses regarding marine CDOM production have been posited. In the first, CDOM is predominantly produced in situ from marine organic matter, as supported by correlations with apparent oxygen utilization (AOU) (e.g., Coble, 2007; Nelson and Siegel, 2013). The second proposes significant contributions from terrestrial "humic-like" components, consistent with spectral and chemical analyses (Andrew et al., 2013; Nelson and Gauglitz, 2016). In coastal waters, CDOM is primarily allochthonous, deriving directly from river input, but autochthonous production by phytoplankton and bacteria also contributes (e.g., Blough and Del Vecchio, 2002; D'Sa and DiMarco, 2009). In open ocean waters away from significant terrestrial influence, CDOM is primarily autochthonous, deriving from primary production (e.g., Iuculano et al., 2019) and the microbial reworking of productivity-derived DOM. Bacterial degradation and photochemical bleaching are significant marine CDOM sinks. The modification of CDOM composition and light absorption characteristics via these processes gives rise to a "reactive" CDOM pool in the upper ocean, contrasting markedly with the "aged refractory CDOM" in deep waters where photochemical degradation is absent and microbial processing is comparatively very slow (Stedmon and Markager, 2001; Kitidis et al., 2006; Hansell and Carlson, 2014). Even so, much remains unknown about the origins, cycling, and precise chemical structure of oceanic CDOM, prompting a need for further enquiry. The interplay of CDOM production, cycling, and removal processes, both in surface and deep ocean waters, is heavily influenced by advection and vertical mixing (Nelson et al., 2007; Bracchini et al., 2010; Nelson and Siegel, 2013; Yamashita et al., 2013). Of particular note in this regard is the sea surface microlayer (SML), a highly dynamic environment (Carlson, 1983; Galgani and Engel, 2016) characterized by pronounced physical and biogeochemical gradients (Hardy, 1982; Cunliffe et al., 2013), and which maintains its integrity even under conditions of high surface turbulence (Wurl et al., 2011; Cunliffe et al., 2013).

Indeed, it is this high surface turbulence that actively maintains strong SML biogeochemical enrichments relative to underlying waters (Sabbaghzadeh et al., 2017). CDOM enrichment of the SML was first observed some time ago (Carlson and Mayer, 1980) and subsequently linked to phytoplankton production (Whitehead and Vernet, 2000). Hardy (1982) proposed that SML organic films promote the accumulation of biological exudates, and Whitehead and Vernet (2000) demonstrated a dinoflagellate response to sunlight exposure, manifested in the release of phenolics to the SML. Maximal bacterial and viral activity during phytoplankton blooms leads to further CDOM release (Wurl et al., 2011; Cunliffe et al., 2013). CDOM enrichment of the SML has also been shown to confer photoprotective properties for phytoplankton, which is particularly important in regions subject to high UV (Tilstone et al., 2010). A valuable means of investigating broad CDOM composition, and hence its likely origins and potential reactivity, is through variations in its optical properties expressed through changes in spectral slope (S). This is most commonly interrogated across two absorption wavelength intervals: S1 (S275-295) and S2 $(S_{350-400})$ (Helms et al., 2008). In regions dominated by biologicallyderived CDOM (e.g., the open ocean), S_1 exceeds S_2 , while in regions with predominantly photodegraded, terrestrially derived CDOM (e.g., coastal waters), S_2 exceeds S_1 (e.g., Helms et al., 2008; Yamashita and Tanoue, 2009). The ratio of these spectral slopes ($S_{\rm R}$: S_1/S_2) thus diagnoses the extent of biologically and photochemically-induced changes in CDOM molecular weight. Low S_R indicates high molecular weight CDOM, e.g., from terrestrial sources, while high S_R signifies low molecular weight CDOM, e.g., from photodegradation of autochthonous open ocean sources (e.g., Helms et al., 2008; Kowalczuk et al., 2013; Galgani and Engel, 2016). Prolonged light exposure leads to a loss of optical activity and the continued accumulation of refractory CDOM (e.g., Twardowski and Donaghay, 2002; Helms et al., 2008).

In this work, we examined the variability of CDOM spectral characteristics through the uppermost 500 m of the Atlantic Ocean water column, including the deep chlorophyll maximum (DCM), and incorporating specialized sampling of the SML. Eleven "biogeographic provinces" representing three Longhurst biomes (Coastal, Westerly, and Trade wind) (Longhurst, 1995) were sampled during the boreal summer/autumn and austral spring. We used CDOM spectral characteristics to examine regionality in CDOM source characteristics and potential transformations, and we specifically contrasted the spectral characteristics of the SML with subsurface seawater (SSW).

Abbreviations: CDOM, chromophoric dissolved organic matter; SML, sea surface microlayer; SSW, subsurface seawater; AMT, Atlantic Meridional Transect; $a_{CDOM}(300)$, CDOM absorption coefficient at 300 nm; S, spectral slope; S₁, spectral slope across wavelength ranges of 275-295; S₂, spectral slope across wavelength ranges of 350-400; S_{R} , spectral slope ratio; NADR, North Atlantic Drift Region; EF, enrichment factor; r^2 , Spearman's rank correlation coefficient; p, probability; n, sample number; AOU, apparent oxygen utilization; ECSW, European Continental Shelf Water; NAST, North Atlantic Subtropical; NAST (E), North Atlantic Subtropical (Eastern); NAST (W) North Atlantic Subtropical (Western); NATR, North Atlantic Tropical Region; WTRA, Western Tropical Atlantic; ETRA, Eastern Tropical Atlantic; SATL, South Atlantic Gyre; SSTC, South Subtropical Convergence Zone; FKLD, Falkland Islands; CTD, Conductivity, temperature, and depth; BODC, British Oceanographic Data Centre; U_{10n}, wind speed at a reference height of 10 m corrected to neutral stability; MAAs, Mycosporine-like Amino Acids.

2 Material and methods

2.1 Study area

Samples were collected on two cruises of *RRS* James Clark Ross during the UK Atlantic Meridional Transect (AMT) program (https://www.pml.ac.uk/science/Projects/Atlantic-Meridional-Transect-(AMT)). Cruise AMT24 (21.09 – 06.11.2014) extended from Immingham (UK) to Punta Arenas (Chile), while cruise AMT25 (11.09 – 04.11.2015) extended from Immingham to the Falkland Islands (Figure 1). Sampling of the SML and SSW is described in detail in Sabbaghzadeh et al. (2017). In brief, we sampled the SML twice daily, during predawn and solar noon hydrocasts, using a Garrett Screen (mesh 16; wire diameter 0.36 mm; effective surface area 2025 cm²) (Garrett, 1965). SSW samples were collected simultaneously via a hull-mounted "nontoxic" seawater inlet at 7 m depth. To examine vertical CDOM profiles, we sampled using the ship's conductivity, temperature, and depth (CTD) rosette at a total of 27 stations during both cruises. Sampling always encompassed five specified light penetration depths (97%, 55%, 33%, 14%, and 1%), the base of the mixed layer, the DCM, and the depths corresponding to maximum and minimum dissolved oxygen (O_2) saturation. Samples were transferred into previously rinsed 250 ml borosilicate volumetric flasks (analytical-grade glassware, SCHOTT, GL 45, 250 ml, Fisher brand). Prior to use, all glassware used in sampling and subsequent analysis was precombusted at 450°C for 8 hours to eliminate any residual organic matter (Stubbins et al., 2006).

2.2 CDOM determination

Immediately following collection, 20 ml subsamples were transferred into a ground glass syringes (SAMCO) and gently filtered (10 ml min⁻¹) through 0.22 μ m surfactant-free single use Luer-Lok syringe filters (MILLEX GP, Millipore, PES membrane).



FIGURE 1

Cruise transects and sampling sites during AMT24 (white triangles) and AMT25 (black triangles). Oceanic provinces (Longhurst, 1995; Reygondeau et al., 2013) indicated are: European Continental Shelf Water (ECSW); North Atlantic Drift Region (NADR); Eastern and Western North Atlantic Subtropical (NAST (E), NAST (W)); North Atlantic Tropical (NATR); Western and Eastern Tropical Atlantic (WTRA, ETRA); South Atlantic Gyre (SATL); South Subtropical Convergence Zone (SSTC); and Falkland Islands (FKLD). The monthly climatology of sea surface chlorophyll *a* for October was obtained from NASA MODIS-Aqua (https://modis.gsfc.nasa.gov/data/dataprod/chlor_a.php).

To minimize contamination during the filtration process, syringes, filters, and glassware were initially rinsed with pure laboratory water (18.2 Ω Milli-Q, Millipore System Inc., USA, hereinafter referred to as Milli-Q) followed by a small aliquot of the sample (~ 3 ml).

CDOM spectra were captured using a single-beam UV-Visible high-performance liquid core waveguide (LCW) spectrophotometer (Ultrapath UV, World Precision Instruments, USA). On AMT25, the LCW had an effective optical pathlength of 204.01 ± 1.0 cm. On AMT24, a shorter effective optical pathlength of 33.70 ± 0.5 cm, determined according to Miller et al. (2002), was used due to previous instrument damage. Detection limits were 0.0092 $\mathrm{m}^{\text{-1}}$ and 0.0023 $\mathrm{m}^{\text{-1}}$ for AMT25 and AMT24, respectively. After the LCW was flushed by drawing < 10 ml of filtered seawater from its sample container using a peristaltic pump, absorbance spectra of the samples were recorded over the wavelength range 275-700 nm at 1nm resolution against 36 g L⁻¹ sodium chloride (NaCl) (hereinafter referred to as reference solution; Miller et al., 2002). This reference solution was prepared using granular NaCl (Fisher Scientific, CAS: 7647–14-5, \geq 99.0) precombusted at 400°C for 6 - 8 hours to eliminate any potential organic contamination (Miller et al., 2002; Helms et al., 2008).

To avoid artefacts arising from temperature differences between seawater samples and the reference solution, the samples were stored in the dark at laboratory temperature for 1 hour prior to analysis (e.g., Pegau and Zaneveld, 1993). To monitor for instrument drift and LCW contamination, absorbance spectra of Milli-Q were recorded before and after processing of a CTD cast. The baselines of Milli-Q recorded on the same day usually agreed to within \pm 0.0005 absorbance units at 275 nm. To avoid artefacts from sample residue, wetted Ultrapath components were cleaned daily with 1 ml of Ultrapath Waveguide cleaning solution (WPI, USA) followed by 20 ml Milli-Q, and cleaning was repeated until Milli-Q absorbance returned to the expected base level.

All spectra were corrected for dark current, recorded 3 times, and averaged for further processing. We used a reference solution of constant salinity because high sample numbers precluded the use of reference solutions with matching salinities for refractive index correction (Nelson et al., 2007). Sample salinities ranged from 34 to 37.6, with most samples falling between 35 and 37. To account for the effects of salinity variations, we compared the apparent absorbance spectra of our NaCl reference solution against Milli-Q to those of salinity 35 and 37 artificial seawater (Dickson, 1990; Grasshoff et al., 2009). This comparison revealed spectra similar to those reported in Nelson et al. (2007), with apparent absorbance increasing towards shorter wavelengths. Our artificial seawater solution showed slightly higher apparent absorbance units at 275 nm.

To address variations in sample salinity, we calculated salinitydependent spectral correction factors from the apparent absorbance spectra of the NaCl reference solution, $A(\lambda)_{NaCl,36}$, and artificial seawater, $A(\lambda)_{ref,SS}$, at sample salinity, SS (Equation 1):

$$F(\lambda) = A(\lambda)_{\text{NaCl},36} - A(\lambda)_{\text{ref},\text{SS}}.$$
 (1)

Following Nelson et al. (2007), $A(\lambda)_{ref,SS}$ was obtained from linear extrapolation of the apparent absorbance spectra of salinity 35 and 37 artificial seawater, $A(\lambda)_{ref,35}$ and $A(\lambda)_{ref,37}$, respectively (Equation 2):

$$A(\lambda)_{\rm ref,SS} = (A(\lambda)_{\rm ref,35} + (A(\lambda)_{\rm ref,37} - A(\lambda)_{\rm ref,35}))/37 - 35$$
$$\times (SS - 35).$$
(2)

Sample absorbance spectra were then corrected for refractive index effects by subtracting F(λ). Absorbance spectra exhibited minor yet perceptible offsets (< ± 0.01 AUs) at longer wavelengths, where CDOM absorbance is expected to be negligible. These small offsets could plausibly be explained by a combination of instrument drift and residual errors in our refractive index correction (e.g. Kitidis et al., 2006; Nelson et al., 2007; Matsuoka et al., 2012; Organelli et al., 2014). Offset-corrected Napierian CDOM absorption coefficients at wavelength λ , a_{CDOM} (λ), were calculated by subtracting the average absorbance between 675 and 695 nm from the absorbance spectrum (Equation 3):

$$a_{\rm CDOM}(\lambda) = 2.303 \times [A(\lambda) - A_{(675 - 695)}] \times l^{-1}$$
 (3)

where 2.303 is a conversion factor and l is the effective optical pathlength of the LCW in meters.

For this study, the absorption coefficient at a wavelength of 300 nm, $a_{\text{CDOM}}(300)$, was selected as a proxy for CDOM because CDOM absorbance dominates in the UV-B region and is notably above the detection limit (Kitidis et al., 2006; Stubbins et al., 2006). To further characterize CDOM, we determined spectral slopes in the shortwave (275–295 nm, S_1) and long-wave UV (350–400 nm, S_2) by linear regression of log-transformed absorption coefficient spectra (Helms et al., 2008).

The spectral slope ratio, $S_R = S_1/S_2$, was used to infer CDOM source characteristics and transformation history. Any accumulation or depletion patterns of compounds in the SML relative to the corresponding SSW were investigated by quantifying enrichment factors (EFs) (Equation 4). EF is calculated as follows:

$$EF = Compound_{SML}/Compound_{SSW}$$
(4)

where EF > 1 indicates compound enrichment in the SML, whereas EF < 1 represents compound depletion in the SML compared to the corresponding SSW. Additional ancillary data, including temperature, salinity, and oxygen, were obtained from the CTD rosette (Seabird Electronics (SBE) 9 plus underwater unit). Chlorophyll a depth profiles were calibrated against fluorometric chlorophyll a measurements of Niskin bottle samples (Welschmeyer, 1994). These data were obtained from the British Oceanographic Data Centre (BODC) (www.BODC.ac.uk). Wind speeds were measured from the ship's anemometer located on the top of the foremast at ~ 20.8 m height above sea level and logged on the ship's computer every 10 minutes. The true wind speed and direction were corrected for the ship's heading by the respective vector. Wind speeds were finally corrected to U_{10n}, the corresponding wind speed at a reference height of 10 m corrected to neutral stability (Nightingale et al., 2000).

3 Results

3.1 CDOM and its optical properties in the SML of the Atlantic Ocean: consistency of the enrichment pattern

Mean SML $a_{CDOM}(300)$ showed no significant difference (*t-test*; p = 0.128) between the two cruises (AMT 24; $\bar{x} \pm SD$: 0.47 \pm 0.28 m⁻¹, range: 0.16 – 1.52 m⁻¹; AMT 25; $\bar{x} \pm SD$: 0.41 \pm 0.13 m⁻¹, range: 0.24 – 0.82 m⁻¹) (Supplementary Table S1). During both cruises, we observed a significant decrease in SML $a_{CDOM}(300)$ from NADR towards NAST (*t-test*; p < 0.05), to around nearly half its initial value ($\bar{x} \pm SD$: 0.44 \pm 0.27 m⁻¹), reflecting a transition to low CDOM waters of the oligotrophic Northern Atlantic gyre. Conversely, higher S_1 in

NAST than in NADR (*t*-*test*; p < 0.05) was consistent with an inverse relationship between SML $a_{CDOM}(300)$ and S_1 .

We also observed an increase in SML $a_{CDOM}(300)$ across the productive and nutrient-enriched tropical regions (NATR) on both cruises, with a mean of $\bar{x} \pm SD$: 0.48 \pm 0.22 m⁻¹ (Supplementary Table S1). Further interhemispheric comparison showed mean SML a_{CDOM} (300) in the Northern Hemisphere (AMT24; $\bar{x} \pm SD$: 0.58 \pm 0.34 m⁻¹; AMT25; $\bar{x} \pm SD$: 0.43 \pm 0.13 m⁻¹) to be higher than in the Southern Hemisphere (AMT24; $\bar{x} \pm SD$: 0.35 \pm 0.11 m⁻¹; AMT25; $\bar{x} \pm SD$: 0.36 \pm 0.13 m⁻¹). However, this interhemispheric difference was statistically significant only during AMT24 (*t*-*test*, *p* < 0.05) (Figure 2).

The latitudinal data for both cruises revealed notable features. Distinct maxima in SML $a_{\rm CDOM}(300)$ were observed in NADR (AMT24; $\bar{x} \pm SD$: 0.96 \pm 0.30 m⁻¹; AMT25; $\bar{x} \pm SD$: 0.60 \pm 0.10 m⁻¹)



FIGURE 2

Latitudinal distribution of $a_{CDOM}(300)$ in the SML (top) and in the SSW at 7 m depth (middle), and the $a_{CDOM}(300)$ Enrichment Factor (EF $a_{CDOM}(300)$, bottom) during AMT24 (circles) and AMT25 (triangles). Error bars for $a_{CDOM}(300)$ are the standard deviation of the mean for each sample. The horizontal dashed line represents no SML enrichment in $a_{CDOM}(300)$ (EF=1).

and in NATR (AMT24; $0.65 \pm 0.27 \text{ m}^{-1}$; AMT25; $0.40 \pm 0.05 \text{ m}^{-1}$). The subtropical gyres (NAST and SATL) had the lowest SML a_{CDOM} (300), reflecting oligotrophic water masses in these less productive regions. SSW a_{CDOM} (300) distribution mirrored that in the SML, ranging from 0.18 to 1.5 m⁻¹, being maximal in NADR (Figure 2).

Significantly higher SSW $a_{\rm CDOM}(300)$ occurred in the Northern Hemisphere than in the Southern Hemisphere during both cruises (*t*-*test*, *p* < 0.05). Potential accumulation or depletion of CDOM in the SML relative to SSW is evidenced by EFs for $a_{\rm CDOM}(300)$. We observed SML $a_{\rm CDOM}(300)$ enrichment at all locations sampled (i.e., EF $a_{\rm CDOM}(300) > 1$) (Figure 2). In the Atlantic Ocean, EF $a_{\rm CDOM}(300)$ vere identified around ~ 42°N on NADR, 10°N on the WTRA, and 15°S on the SATL during AMT24, aligning with findings from AMT25 (Figure 2). Despite a robust positive correlation observed between SML $a_{\rm CDOM}(300) = 1.12$ SSW $a_{\rm CDOM}(300) + 0.13$, $r^2 = 0.64$, n = 103), EF $a_{\rm CDOM}(300)$ was not found to be significantly associated with SML $a_{\rm CDOM}(300)$ (i.e., SML $a_{\rm CDOM}(300) = 0.57$ EF $a_{\rm CDOM}(300) + 1.43$, $r^2 = 0.05$, n = 103).

Additionally, we investigated the relationship between EF a_{CDOM} (300) and spectral absorption characteristics, revealing no significant linear regression between EF a_{CDOM} (300), S_1 , and S_R (p > 0.05). The latitudinal distribution of S_1 in the SML revealed a contrasting pattern, with elevated S_1 being observed at less productive northerly and southerly latitudes (Supplementary Figure S1). In the SML S_1 varied between 0.02 and 0.04 nm⁻¹ ($\bar{x} \pm SD$: 0.03 \pm 0.01 nm⁻¹). The oligotrophic gyres, centered around 30°N and 20°S, had the highest SML S_1 ($\bar{x} \pm SD$: 0.04 \pm 0.01 nm⁻¹). In contrast, the lowest SML S_1 (< 0.03 nm⁻¹) was consistently observed north of 40°N and south of 40°S on both cruises (Supplementary Figure S1).

Mean S_R was $\bar{x} \pm SD$: 1.65 \pm 0.54 (range: 0.15 – 2.70) on AMT24, and $\bar{x} \pm SD$: 2.11 \pm 0.56 (range: 1.03 – 3.60) on AMT25. Maximum S_R (i.e., $S_R > 2$) occurred in the oligotrophic regions of NAST and SATL. In SML samples around 15°N, a distinct CDOM absorption shoulder between 325 and 350 nm (Supplementary Figure S2) was consistent

with the findings of Tilstone et al. (2010), who reported a similar feature at short wavelengths in the dissolved optical fraction of organic matter off the Iberian Peninsula. Although Tilstone et al. (2010) suggested the presence of mycosporine-like amino acids (MAAs), our observations indicate that samples containing MAAs were primarily found in the SML of NATR during both cruises, but totaling only four samples. The presence of MAAs was less frequent in other oceanic provinces, where CDOM dominated the spectral absorbance within the SML at those locations. The $a_{\text{CDOM}}(300)$ was analyzed in relation to salinity, water temperature, and, wind speed across the SML. Although inverse correlations were observed between $a_{\text{CDOM}}(300)$ and both salinity and temperature in all studied samples, these correlations were not statistically significant (Supplementary Table S2). SML $a_{CDOM}(300)$ showed non-statistically significant increases at higher wind speeds (U_{10n}) (p > 0.001) (Supplementary Table S3). Furthermore, no significant correlation was identified between EF $a_{CDOM}(300)$ and wind speed along the transects (Pearson's correlation = -0.124, p = 0.213, n = 103).

3.2 Depth-cross sections of CDOM in the Atlantic Ocean

The depth-cross section of $a_{\text{CDOM}}(300)$ exhibited the average of $\bar{x} \pm SD$: 0.30 \pm 0.12 m⁻¹ (range: 0.10 – 0.87 m⁻¹) during AMT24 and the average of $\bar{x} \pm SD$: 0.27 \pm 0.10 m⁻¹ (range of 0.13 – 0.55 m⁻¹) during AMT25, with no statistically significant difference between the two cruises (p = 0.003). The latitudinal cross-section of chlorophyll *a* also indicates elevated concentrations in the Northern Hemisphere and equatorial zones compared to the Southern Hemisphere. The highest concentrations were observed north of 30°N, within the equatorial region, and south of 40°S. This trend is particularly pronounced in the data from AMT24 (Figure 3).

A peak in $a_{\text{CDOM}}(300)$ at the very end of transects in ECSW waters down to 150 m depth, observed during both AMT24 and AMT25,



FIGURE 3

Latitudinal cross sections of chlorophyll *a* (mg m⁻³) (left panel) and depth-cross sections of $a_{CDOM}(300)$ (m⁻¹) including the Atlantic SML (right panel) during AMT24 (top) and AMT25 (bottom). The chlorophyll data were derived from a fluorescence sensor fitted on the CTD rosette. Black dots represent latitude and depth of individual samples. Biogeochemical provinces are: European Continental Shelf Water (ECSW); North Atlantic Drift Region (NADR); North Atlantic Subtropical Region (NAST); North Atlantic Tropical Region (NATR); Western/Eastern Tropical Atlantic (W/E TRA); South Atlantic Gyre (SATL); South Subtropical convergence zone (SSTC); and Falkland Islands (FKLD), all bordered by solid black lines.



might be associated with regional chlorophyll a maxima. Additionally, the sustained higher levels of $a_{\text{CDOM}}(300)$ observed within the water column in ECSW and NADR could be attributed to the advection of terrestrially-derived material from the continental shelf, which was more pronounced during AMT25. The second set of regional/ latitudinal maxima of $a_{\rm CDOM}(300)$ was identified in the northern tropical region between NATR and W/E TRA, at depths greater than 20 m in NATR and around 50 m in W/E TRA, respectively (Figure 3). In the southern Hemisphere, $a_{\text{CDOM}}(300)$ was statistically lower than in the Northern Hemisphere (p < 0.001). At high Southern Hemisphere latitudes, specifically in SSTC and FKLD, a_{CDOM}(300) was slightly elevated in upper layers compared to the lower latitudes of SATL, correlating with a shallower DCM at approximately 50 m (Figure 3 and Supplementary Table S4). $a_{CDOM}(300)$ minima on the other hand, exhibited less variability, characterized by a deep DCM and low surface chlorophyll a, as documented by Aiken et al. (2000), in both the northern and southern Atlantic gyres (NAST and SATL) (Figure 3). Additionally, within the upper 100 m of the water column in the oligotrophic SATL, a_{CDOM}(300) showed more homogeneity and lower concentrations than in NAST (Supplementary Table S4). The S1 maxima coincided with DCM depths observed in NAST above ~ 50 m, above ~ 20 m in the equatorial region of NATR and W/E TRA, and in low latitudes in the SATL, consistently observed between the two expeditions. On the other hand, S_1 minima were determined at the northern and southern ends of transects (specifically ECSW, NADR, SSTC, and FKLD (Figure 4).

The lower S_1 values were identified at depths exceeding 30 m in NATR and 80 m in W/E TRA, coinciding with $a_{\text{CDOM}}(300)$ maxima, while S_1 minima were observed in these regions at depths exceeding ~ 50 m (Figures 3, 4). Following the S_1 pattern, S_R maxima were predominantly observed in the upper layers of the oligotrophic NAST, primarily at depths shallower than ~ 60 m, and in the SATL. Additionally, there was a notable high S_R at depths below 200 m at the southern end of the transect, specifically FKLD, more prominently evident on AMT24 (Figure 4). Conversely, S_R minima were identified at the northern end of the transects, i.e., ECSW, and in the tropical and equatorial regions at depths below 20 m, aligning with the S_1 trend and in contrast to the $a_{\text{CDOM}}(300)$ distribution (Figures 3, 4).

Upon further investigation, we found the overall relationship between $a_{\text{CDOM}}(300)$ and S_1 in the Atlantic Ocean to exhibit a significant inverse correlation, both in the SML (r = -0.80, p < 0.001, n = 114) and in depth profiles ($r^2 = -0.60$, p < 0.001, n = 845) (Figure 5).

In general, S_1 consistently showed an asymptotic decrease with increasing $a_{\text{CDOM}}(300)$ on both cruises, with the highest S_1



FIGURE 5

 a_{CDOM} (300) vs S_1 in the SML (left) and in depth profiles (0 m \leq depth \leq 500 m) (right) during AMT24 and AMT25. The dashed line represents the best fit of the data to a linear model (left).



coinciding with the lowest $a_{\text{CDOM}}(300)$ in both the SML and in depth profiles (Figure 5). The highest $a_{\text{CDOM}}(300)$, associated with the lowest S_1 , was observed in ECSW and NADR depth profiles, while the highest S_1 associated with the lowest $a_{\text{CDOM}}(300)$ was observed in the oligotrophic waters of SATL during both AMT24 and AMT25. The overall trend displayed a less pronounced shift, and the regression curve exhibited a slight leveling off within the depth profiles in the tropical region (Figure 5).

3.3 CDOM-chlorophyll a association

Considering that CDOM in the open ocean is primarily derived from phytoplankton (e.g., Carder et al., 1989; Hansell and Carlson, 2014), we examined potential relationships between $a_{\text{CDOM}}(300)$ and chlorophyll *a*. For the depth profile data, we found no statistically significant correlations (Spearman rank order correlation coefficient; $r^2 = 0.45$, p > 0.001, n = 661) (Figure 6).

We also found no significant correlations between SML a_{CDOM} (300) and chlorophyll a (p > 0.001) for the complete dataset. Removing a group of high latitude samples (NADR circled in blue, Figure 6), improved the correlation ($r^2 = 0.74$, p < 0.001, n = 31).

4 Discussion

SML $a_{CDOM}(300)$ in the Atlantic Ocean was maximal at high latitudes in NADR. Conversely, a significant decrease in SML a_{CDOM} (300) was observed in the oligotrophic region of NAST during both transects. The elevated SML $a_{CDOM}(300)$ at higher latitudes has been previously attributed to increased regional productivity and biomass levels in continental shelf waters during sampling in boreal spring (Stubbins et al., 2006, unpublished data). However, further investigation during this study revealed no significant correlation between SML $a_{CDOM}(300)$ and chlorophyll *a* at high latitudes in the Northern Hemisphere, indicating the influence of other regional sources of CDOM such as terrestrial deposition. Additionally, the enrichment and removal of oceanic CDOM within the SML involves complex processes and multiple parameters, including photodegradation, bacterial production and consumption, and zooplankton grazing, which may camouflage any single relationship between chlorophyll a and CDOM (e.g., Carlson, 1983, 1993). Similarly, in our study, $a_{CDOM}(300)$ and chlorophyll a for water column samples down to a maximum depth of 500 m were not significantly correlated. This leads us to suggest that any association between CDOM and chlorophyll a might be biogeochemical province-dependent, with a stronger correlation observed towards lower latitudes where terrestrial CDOM sources are less diverse. On the other hand, instantaneous chlorophyll concentrations may not be suitable indicators of CDOM abundance, as CDOM production from phytoplankton-derived organic matter involves microbial processing over timescales of months (e.g., Nelson et al., 1998; Boss et al., 2001). In support of this, an earlier study conducted during the phytoplankton spring bloom in the Sargasso Sea reported no significant correlation between $a_{\text{CDOM}}(300)$ and chlorophyll a ($r^2 =$ 0.45, p > 0.05) (Nelson et al., 1998). In a recent study conducted during AMT26 (Dall'Olmo et al., 2017) a positive correlation was observed between chlorophyll and $a_{CDOM}(400)$ but with substantial variabilities in this relationship. These variabilities suggest that the dynamics of CDOM are influenced by processes distinct from those governing phytoplankton pigments. While acknowledging the correlation, the study implies that chlorophyll may not be an exclusive indicator of CDOM. The findings underscore the presence of additional factors beyond phytoplankton dynamics that contribute to shaping the distribution of CDOM in the Atlantic Ocean. Considering previous suggestions and the findings of our study, we propose that dynamics of CDOM optical properties in the SML may be better explained by a combination of both biological and non-biological factors. This supports the notion that CDOM is also produced as a degradation product of phytoplankton via zooplankton grazing and, primarily, bacterial degradation (e.g., Siegel and Michaels, 1996; Nelson et al., 1998). Consequently, we support the idea that chlorophyll is an inadequate proxy for estimating CDOM due to their temporal decoupling in the oceans.

The robust positive correlation between SML $a_{\text{CDOM}}(300)$ and SSW $a_{\text{CDOM}}(300)$, as identified in this study, aligns with previous research reporting statistically significant relationships between SML $a_{\text{CDOM}}(305)$, SML (DOC), and SML (DMS) concentrations and their corresponding subsurface waters in oceanic samples (e.g., Yang et al., 2006, 2009). The SML consistently exhibited enrichment in CDOM compared to the corresponding subsurface waters, with EF $a_{\text{CDOM}}(300)$ maxima observed in the Northern tropical region and EF $a_{\text{CDOM}}(300)$ minima in the Southern Hemisphere. These findings are consistent with those of Tilstone et al. (2010), who identified SML $a_{\text{CDOM}}(300)$ maxima with concentrations more than twice those of subsurface waters (2.0 m depth) in an active upwelling area off the Iberian Peninsula. In Jiazhou Bay, China, during autumn and winter, CDOM was also found to be enriched in the SML compared to the underlying waters, with an EF a_{CDOM} (300) of 1.84 (Zhang and Yang, 2013).

A negative correlation between $a_{\text{CDOM}}(300)$ and salinity and temperature in SML samples was reported previously by Kitidis et al. (2006), who observed a decrease in $a_{\text{CDOM}}(350)$ with increasing salinity, indicating CDOM removal. Similarly, we also found a negative relationship between $a_{\text{CDOM}}(300)$ and these variables; however, the correlation was not statistically significant.

The "removal" of CDOM with increasing salinity may coincide with surface waters, where the rise in salinity driven by high solar irradiance correlates with photochemical removal.

Additionally, the observed positive correlation between $a_{\rm CDOM}$ (300) and wind speed (U_{10n}) during this study may support our previous suggestion that the enrichment of organic compounds in the SML occurs through wind-driven bubble scavenging, which facilitates the transport of these compounds from the SSW to the SML (Sabbaghzadeh et al., 2017). However, this study did not reveal a strong correlation between instantaneous wind speeds and CDOM EFs. Therefore, we propose that under conditions of high wind speeds, turbulence, and generated bubbles, the enrichment of the SML is a complex process that obscures any simple relationships between wind speed and the enrichment of CDOM in the SML. This proposition aligns with the insights of Wurl et al. (2011), who also suggested an intricate enrichment process at high wind speeds, impacting the simple relationship between organic matter enrichment in the SML and wind speeds in both coastal and oceanic samples.

UV-resistant phytoplankton exudates such as MAAs, which share many optical characteristics with CDOM, may be found in the SML (Tilstone et al., 2010). Tilstone et al. (2010) identified the presence of MAAs in the NATR, which recorded one of the highest EF a_{CDOM} (300) in the Atlantic Northern Hemisphere. However, distinct CDOM absorption spectra related to the presence of MAAs were not frequently observed in our work. An alternative possibility is that photo-resistant CDOM derived from algae is transformed into an inactive form resistant to photodegradation (e.g., Nelson et al., 1998).

To further characterize CDOM, we determined CDOM spectral slope factors after Helms et al. (2008) and we found lower SML(S_R) compared to SSW(S_R). One possible explanation for the lower spectral slopes in the SML compared to underlying waters is the local microbial release of CDOM directly into the SML due to light exposure. Microbial processes, such as growth, exudation, and lysis in the euphotic zone, may lead to the production of CDOM with higher molecular weight, resulting in a lower spectral slope ratio (e.g., Helms et al., 2008; Kowalczuk et al., 2013; Galgani and Engel, 2016). Another possibility for the lower SML (S_R) compared to underlying waters could be the terrestrial source of high molecular weight CDOM in the coastal waters. The maximum SML(S_R) in NAST and SATL regions, associated with high S_1 compared to other provinces, may indicate CDOM photodegradation in these clear, less productive water masses (Helms et al., 2008).

Our $a_{\rm CDOM}(300)$ depth profiles concur with those previously reported for oceanic samples. A mean $a_{\rm CDOM}(300) < 0.50$ within the entire water column is close to values reported for the Mid-Atlantic Bight, the western tropical Atlantic, and the Sargasso Sea (e.g., Nelson et al., 1998; Del Vecchio and Blough, 2004). On AMT20, Kowalczuk et al. (2013) also measured $a_{\rm CDOM}(305)$ with an average of $\bar{x} = 0.23$ m⁻¹ to a depth of 300 m in the Atlantic Ocean.

The occurrence of $a_{CDOM}(300)$ maxima in ECSW and NADR in waters to about 150 m depth observed mostly on AMT24 was linked to regional chlorophyll *a* maxima. On AMT25, the high $a_{CDOM}(300)$ persisted throughout the water column to a depth of about 500 m. The observation that the DCM did not deepen further on AMT25 relative to AMT24 suggests that this elevated $a_{CDOM}(300)$ could be attributable to the advection of terrestrially-derived material from the continental shelf, warranting further investigation. Additionally, understanding the potential influence of increased mixing in coastal regions during AMT25 also requires further study. In regions such as NATR and W/E TRA, where high $a_{\text{CDOM}}(300)$ levels extended to deeper waters, we recognize the need for additional sampling to elucidate the reasons behind these observations. Given that our data predominantly cover the top 200 m, providing limited resolution in deeper waters, we recommend future studies to include sampling with better resolution to comprehensively investigate the potential causes of high-depth CDOM levels. Additionally, ECSW ($S_{\rm R}$) minima (i.e., ~ 1.5) concurred with the findings of Kowalczuk et al. (2013), who reported the lowest $S_{\rm R}$ values (i.e., $S_{\rm R} \sim 2.0$) in the uppermost 100 m of the western European continental shelf, suggesting the presence of high molecular weight terrestrially-derived CDOM. Low CDOM in oligotrophic regions may result from reduced productivity alone or in combination with accelerated CDOM depletion due to photodegradation, leading to CDOM transformation in optically clear waters. This transformation is associated with elevated S_1 (~ 0.05) and maximum S_R values (e.g., Helms et al., 2008; Kowalczuk et al., 2013; Zhang and Yang, 2013). The observed high S_R in NAST and SATL aligns with previous reports by Kowalczuk et al. (2013), indicating high $S_{\rm R}$ (~ 3.0) in the Atlantic oligotrophic gyres, and thus suggestive of CDOM photodegradation in these less productive waters. Our data consistently show that S_R is highest in the subtropical gyres, which receive the highest irradiance. This elevated $S_{\rm R}$ is maintained throughout the two studied seasons, as evidenced by similar high S_R values during both expeditions.

The earlier hypotheses proposed CDOM distribution to be primarily governed by CDOM production near the DCM and its subsequent upward transport through diffusion and biological transport, facilitated by the diurnal migration of plankton. Its removal has primarily been attributed to photooxidation, particularly during summer (e.g., Siegel and Michaels, 1996). However, our findings are suggestive of a province-dependent pattern of CDOM distributions in the upper 500 m of the Atlantic Ocean. In the low latitudes of NATR and W/E TRA, a distinct pattern of high $a_{\text{CDOM}}(300)$ and low S_{R} indicates discharge of terrestriallyderived CDOM. In contrast, the oligotrophic subtropical regions of NAST and SATL exhibit lower $a_{\text{CDOM}}(300)$ and higher S_{R} that presumably represent degraded and older 'background' (refractory) biologically-derived CDOM (e.g., Stedmon and Markager, 2001; Kitidis et al., 2006; Hansell and Carlson, 2014). While no strong correlation was identified between $a_{CDOM}(300)$ and chlorophyll *a*, either in the SML or in depth profiles, additional investigation revealed the potential for a linear regression model to predict CDOM absorption coefficient using chlorophyll *a* as a proxy with better than 70% accuracy in some regions (Figure 6). Notably, this predictive model exhibited province-dependent characteristics. We contend that predicting CDOM from chlorophyll *a* may potentially be of most value in open ocean waters where *in-situ* biological production is the predominant CDOM source, and where additional CDOM sources, such as terrestrially-derived material is absent.

5 Conclusion

Our broad findings regarding oceanic CDOM properties are consistent with those from previous studies. By combining these findings, we have identified a complex set of potential drivers of CDOM distribution and spectral properties, clearly reflected in the biogeochemical province-dependency we have observed. Of particular note is the strong province-dependency of CDOM vs chlorophyll a, which will likely confound any efforts to predict one from the other, especially in coastal waters where we consistently found the correlations to be weakest. An especially novel aspect of our work was the comparably complex behavior we observed for CDOM in the SML, which deviates from that characteristic of the underlying waters. The processes responsible for CDOM enrichment in the SML clearly challenge their adequate explanation via simple relationships, for example those dominated solely by wind-driven turbulence and bubble-mediated transport. To conclude, understanding the drivers of CDOM dynamics in the oceans remains a substantial challenge, which is further complicated by region-specific considerations. We believe that additional studies complementing the work we have described here will drive further advances in our understanding of the complex behavior of CDOM in the marine environment.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: British Oceanographic Data center (BODC) (www.BODC.ac.uk).

Author contributions

BS: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Resources, Validation, Visualization, Writing – original draft, Writing – review & editing. GU:

References

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the UK Natural Environment Research Council (NERC: grant NE/K00252X/1 to R.C.U.G.) and is a component of RAGNARoCC (Radiatively active gases from the North Atlantic Region and Climate Change), which contributes to NERC's Greenhouse Gas Emissions and Feedbacks program (http://www. nerc.ac.uk/research/funded/programmes/greenhouse/). AMT contributes to the international IMBER initiative and is supported by NERC National Capability funding to PML and the National Oceanography Centre, Southampton. This is AMT contribution number 295. The author confirms that the publication fee was supported by funding from the Leibniz Open Access Publishing Fund (https://www.leibniz-gemeinschaft.de/en/research/ open-science-and-digitalisation/open-access/journal-publishingfund.html), Germany. Additionally, part of the publication fee was covered by the editorial board, with Professor Mostofa agreeing to cover the remaining portion of the fee for the editorial points.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmars.2024.1432133/ full#supplementary-material

Aiken, J., Rees, N., Hooker, S., Holligan, P., Bale, A., Robins, D., et al. (2000). The Atlantic Meridional Transect: overview and synthesis of data. *Prog. Oceanogr.* 45, 257–312. doi: 10.1016/S0079-6611(00)00005-7

Andrew, A. A., Walker, S. L., Murray, J. W., and Codispoti, L. A. (2013). Chromophoric dissolved organic matter in the eastern Atlantic Ocean. *Mar. Chem.* 148, 15–24. doi: 10.1016/j.marchem.2012.11.001

Blough, N. V., and Del Vecchio, R. (2002). "Chromophoric DOM in the coastal environment," in *Biogeochemistry of marine dissolved organic matter*. Eds. C. A. Carlson and D. A. Hansell (Academic Press), 509–546. doi: 10.1016/B978-012323841-2/50012-9

Boss, E., Pegau, W. S., Zaneveld, J. R. V., and Barnard, A. H. (2001). Spatial and temporal variability of absorption by dissolved material at a continental shelf. *JGR: Oceans* 106, 9499–9507. doi: 10.1029/2000JC900008

Bracchini, L., Tognazzi, A., Dattilo, A. M., Decembrini, F., Rossi, C., and Loiselle, S. A. (2010). Sensitivity analysis of CDOM spectral slope in artificial and natural samples: an application in the central eastern mediterranean basin. *Aquat. Sci.* 72, 485–498. doi: 10.1007/s00027-0100150-y

Carder, K. L., Steward, R. G., Harvey, G. R., and Ortner, P. B. (1989). Marine humic and fulvic acids: Their effects on remote sensing of ocean chlorophyll. *Limnol. oceanogr.* 34, 68–81. doi: 10.4319/lo.1989.34.1.0068

Carlson, D. J. (1983). Dissolved organic materials in surface microlayers: Temporal and spatial variability and relation to sea state. *Limnol. Oceanogr.* 28, 415–431. doi: 10.4319/lo.1983.28.3.0415

Carlson, D. J. (1993). "The early diagenesis of organic matter: reaction at the air-sea interface," in *The Organic geochemistry: principles and applications*. Ed. M. H. Engel (Springer), 255–268.

Carlson, D. J., and Mayer, D. A. (1980). Organic films on ocean water. Science 208, 389-391.

Coble, P. G. (2007). Marine optical biogeochemistry: the chemistry of ocean color. *Chem. Rev.* 107, 402-418. doi: 10.1021/cr050350+

Cunliffe, M., Engel, A., Frka, S., Gašparović, B., Guitart, C., Murrell, J. C., et al. (2013). Sea surface microlayers: A unified physicochemical and biological perspective of the air–ocean interface. *Prog. Oceanogr.* 109, 104–116. doi: 10.1016/j.pocean.2012.08.004

Dall'Olmo, G., Brewin, R. J., Nencioli, F., Organelli, E., Lefering, I., McKee, D., et al. (2017). Determination of the absorption coefficient of chromophoric dissolved organic matter from underway spectrophotometry. *Optics express* 25, A1079–A1095. doi: 10.1364/OE.25.0A1079

Del Vecchio, R., and Blough, N. V. (2004). Spatial and seasonal distribution of chromophoric dissolved organic matter in the Middle Atlantic Bight. *Mar. Chem.* 89, 169–187. doi: 10.1016/j.marchem.2004.02.027

Dickson, A. G. (1990). Thermodynamics of the dissociation of boric acid in synthetic seawater from 273.15 to 318.15 K. *Deep-Sea Res. Part I: Oceanographic Res. Papers* 37, 755–766. doi: 10.1016/0198-0149(90)90004-F

D'Sa, E., and DiMarco, S. (2009). Seasonal variability and controls on chromophoric dissolved organic matter in a large river-dominated coastal margin. *Limnol. Oceanogr.* 54, 2233–2242. doi: 10.4319/lo.2009.54.6.2233

Galgani, L., and Engel, A. (2016). Changes in optical characteristics of surface microlayers hint to photochemically and microbially mediated DOM turnover in the upwelling region off the coast of Peru. *Biogeosci* 13, 2453–2473. doi: 10.5194/bg-13-2453-2016

Garrett, W. D. (1965). Collection of slick-forming materials from the sea surface. *Limnol. Oceanogr.* 10, 602–605. doi: 10.4319/lo.1965.10.4.0602

Grasshoff, K., Kremling, K., and Ehrhardt, M. (2009). *Methods of seawater analysis* (John Wiley & Sons).

Hansell, D. A., and Carlson, C. A. (2014). *Biogeochemistry of marine dissolved organic matter* (Academic Press).

Hardy, J. T. (1982). The sea surface microlayer: Biology, chemistry and anthropogenic enrichment. *Prog. Oceanogr.* 11, 307–328. doi: 10.1016/0079-6611(82)90001-5

Helms, J. R., Stubbins, A., Ritchie, J. D., Minor, E. C., Kieber, D. J., and Mopper, K. (2008). Absorption spectral slopes and slope ratios as indicators of molecular weight, source, and photobleaching of chromophoric dissolved organic matter. *Limnol. Oceanogr.* 53, 955–969. doi: 10.4319/lo.2008.53.3.0955

Iuculano, F., Álverez-Salgado, X. A., Otero., J., Catalá., T. S., Sobrino., C., Duarte, C. M., et al. (2019). Patterns and drivers of UV absorbing chromophoric dissolved organic matter in the euphotic layer of the open ocean. *Front. Mar. Sci.* 6. doi: 10.3389/fmars.2019.00320

Kitidis, V., Stubbins, A. P., Uher, G., Goddard, R. C. U., Law, C. S., and Woodward, E. M. S. (2006). Variability of chromophoric organic matter in surface waters of the Atlantic Ocean. *Deep Sea Res. Part II: Topical Stud. Oceanogr.* 53, 1666–1684. doi: 10.1016/j.dsr2.2006.05.009

Kowalczuk, P., Tilstone, G. H., Zabłocka, M., Röttgers, R., and Thomas, R. (2013). Composition of dissolved organic matter along an Atlantic Meridional Transect from fluorescence spectroscopy and Parallel Factor Analysis. *Mar. Chem.* 157, 170–184. doi: 10.1016/j.marchem.2013.10.004

Longhurst, A. (1995). Seasonal cycles of pelagic production and consumption. *Prog. Oceanogr.* 36, 77–167. doi: 10.1016/0079-6611(95)00015-1

Matsuoka, A., Bricaud, A., Benner, R., Para, J., Sempéré, R., Prieur, L., et al. (2012). Tracing the transport of colored dissolved organic matter in water masses of the Southern Beaufort Sea: relationship with hydrographic characteristics. *Biogeosci* 9, 925– 940. doi: 10.5194/bg-9-925-2012

Miller, R. L., Belz, M., Del Castillo, C., and Trzaska, R. (2002). Determining CDOM absorption spectra in diverse coastal environments using a multiple pathlength, liquid core waveguide system. *Continental Shelf Res.* 22, 1301–1310. doi: 10.1016/S0278-4343 (02)00009-2

Nelson, N. B., and Gauglitz, J. M. (2016). Optical signatures of dissolved organic matter transformation in the global ocean. *Front. Mar. Sci.* 2, 118. doi: 10.3389/fmars.2015.00118

Nelson, N. B., and Siegel, D. A. (2013). The global distribution and dynamics of chromophoric dissolved organic matter. *Ann. Rev. Mar. Sci.* 5, 447–476. doi: 10.1146/ annurev-marine-120710-100751

Nelson, N. B., Siegel, D. A., Carlson, C. A., Swan, C., Smethie C.OMMAJr., W. M., and Khatiwala, S. (2007). Hydrography of chromophoric dissolved organic matter in the North Atlantic. *Deep Sea Res. Part I: Oceanographic Res. Pap.* 54, 710–731. doi: 10.1016/j.dsr.2007.02.006

Nelson, N. B., Siegel, D. A., and Michaels, A. F. (1998). Seasonal dynamics of colored dissolved material in the Sargasso Sea. *Deep Sea Res. Part I: Oceanographic Res. Pap.* 45, 931–957. doi: 10.1016/S0967-0637(97)00106-4

Nightingale, P. D., Liss, P. S., and Schlosser, P. (2000). Measurements of air-sea gas transfer during an open ocean algal bloom. *Geophysical Res. Lett.* 27, 2117–2120. doi: 10.1029/2000GL011541

Organelli, E., Bricaud, A., Antoine, D., and Matsuoka, A. (2014). Seasonal dynamics of light absorption by chromophoric dissolved organic matter (CDOM) in the NW Mediterranean Sea (BOUSSOLE site). *Deep Sea Res. Part I: Oceanographic Res. Papers* 91, 72–85. doi: 10.1016/j.dsr.2014.05.003

Pegau, W. S., and Zaneveld, J. R. V. (1993). Temperature-dependent absorption of water in the red and near-infrared portions of the spectrum. *Limnol. Oceanogr.* 38, 188–192. doi: 10.4319/lo.1993.38.1.0188

Reygondeau, G., Longhurst, A., Martinez, E., Beaugrand, G., Antoine, D., and Maury, O. (2013). Dynamic biogeochemical provinces in the global ocean. *Global Biogeochemical Cycles* 27, 1046–1058. doi: 10.1002/gbc.20089

Sabbaghzadeh, B., Upstill-Goddard, R. C., Beale, R., Pereira, R., and Nightingale, P. D. (2017). The Atlantic Ocean surface microlayer from 50° N to 50° S is ubiquitously enriched in surfactants at wind speeds up to 13 m s- 1. *Geophy. Res. Lett.* 44, 2852–2858. doi: 10.1002/2017GL072988

Siegel, D. A., Maritorena, S., Nelson, N. B., Behrenfeld, M. J., and McClain, C. R. (2005). Colored dissolved organic matter and its influence on the satellite-based characterization of the ocean biosphere. *Geophysical Res. Lett.* 32. doi: 10.1029/2005GL024310

Siegel, D. A., and Michaels, A. F. (1996). Quantification of non-algal light attenuation in the Sargasso Sea: Implications for biogeochemistry and remote sensing. *Deep Sea Res. Part II: Topical Stud. Oceanogr.* 43, 321–345. doi: 10.1016/0967-0645(96)00088-4

Siegel, D. A., Maritorena, S., Nelson, N. B., Hansell, D. A., and Lorenzi-Kayser, M. (2002). Global distribution and dynamics of colored dissolved and detrital organic materials. *J. Geophys. Res.* 107, 3228. doi: 10.1029/2001JC000965

Stedmon, C. A., and Markager, S. (2001). The optics of chromophoric dissolved organic matter (CDOM) in the Greenland Sea: An algorithm for differentiation between marine and terrestrially derived organic matter. *Limnol. Oceanogr.* 46, 2087–2093. doi: 10.4319/lo.2001.46.8.2087

Stubbins, A., Uher, G., Law, C. S., Mopper, K., Robinson, C., and Upstill-Goddard, R. C. (2006). Open-ocean carbon monoxide photoproduction. *Deep Sea Res. Part II: Topical Stud. Oceanogr.* 53, 1695–1705. doi: 10.1016/j.dsr2.2006.05.011

Tilstone, G. H., Airs, R. L., Vicente, V. M., Widdicombe, C., and Llewellyn, C. (2010). High concentrations of mycosporine-like amino acids and colored dissolved organic matter in the sea surface microlayer off the Iberian Peninsula. *Limnol. Oceanogr.* 55, 1835–1850. doi: 10.4319/lo.2010.55.5.1835

Twardowski, M. S., Boss, E., Sullivan, J. M., and Donaghay, P. L. (2004). Modeling the spectral shape of absorption by chromophoric dissolved organic matter. *Mar. Chem.* 89, 69–88. doi: 10.1016/j.marchem.2004.02.008

Twardowski, M. S., and Donaghay, P. L. (2002). Photobleaching of aquatic dissolved materials: Absorption removal, spectral alteration, and their interrelationship. *JGR: Oceans* 107, 6–1. doi: 10.1029/1999JC000281

Welschmeyer, N. A. (1994). Fluorometric analysis of chlorophyll-a in the presence of chlorophyll-b and phaeopigments. *Limnol. Oceanogr.* 39, 1985–1992. doi: 10.4319/ lo.1994.39.8.1985

Whitehead, K., and Vernet, M. (2000). Influence of mycosporine-like amino acids (MAAs) on UV absorption by particulate and dissolved organic matter in La Jolla Bay. *Limno. Oceanog.* 45, 1788–1796. doi: 10.4319/lo.2000.45.8.1788

Wurl, O., Wurl, E., Miller, L., Johnson, K., and Vagle, S. (2011). Formation and global distribution of sea-surface microlayers. *Biogeosci* 8, 121–135. doi: 10.5194/bg-8-121-2011

Yamashita, Y., Nosaka, Y., Suzuki, K., Ogawa, H., Takahashi, K., and Saito, H. (2013). Photobleaching as a factor controlling spectral characteristics of chromophoric dissolved organic matterino penocean. *Biogeosci* 10, 7207-7217. doi: 10.5194/bg-10-7207-2013

Yamashita, Y., and Tanoue, E. (2009). Basin scale distribution of chromophoric dissolved organic matter in the Pacific Ocean. *Limnol. Oceanogr.* 54, 598. doi: 10.4319/ lo.2009.54.2.0598

Yang, G. P., Jing, W. W., Li, L., Kang, Z. Q., and Song, G. S. (2006). Distribution of dimethylsulfide and dimethylsulfoniopropionate in the surface microlayer and subsurface water of the Yellow Sea, China during spring. *J. Mar. Systems* 62, 22–34. doi: 10.1016/j.jmarsys.2006.04.003

Yang, G. P., Levasseur, M., Michaud, S., Merzouk, A., Lizotte, M., and Scarratt, M. (2009). Distribution of dimethylsulfide and dimethylsulfoniopropionate and its relation with phytoneuston in the surface microlayer of the western North Atlantic during summer. *Biogeochem* 94, 243–254. doi: 10.1007/s10533-009-9323-y

Zhang, J., and Yang, G. (2013). Chemical properties of colored dissolved organic matter in the sea-surface microlayer and subsurface water of Jiaozhou Bay, China in autumn and winter. *Acta Oceanologica Sin.* 32, 26–39. doi: 10.1007/s13131-013-0306-4