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Rapid physiological colouration change is a challenge - but not a hindrance - to successful photo identification of giant sunfish (*Mola alexandrini*, Molidae)

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Photo ID is a common tool in ecology, but has not previously been attempted for the ocean sunfishes (*Mola* spp., Molidae; 'molids'). The technique, based on body patterns, could potentially be informative for studying the seasonal occurrence of giant sunfish (*Mola alexandrini*) on the Bali reefs (Indonesia), where this species is an important drawcard for the local SCUBA diving tourism. However, molids are capable of rapid physiological colouration change, which may complicate the application of the method. Our study aimed to determine if photo ID is nevertheless achievable and informative. To test this, we created the citizen-science platform 'Match My Mola' for crowd-sourcing imagery (photos and video) of *M. alexandrini* in Bali, and undertook trial matching (n=1,098 submissions). The submitted imagery revealed a wide range of pattern clarity, from fish with no pattern to bold displays. Video confirmed physiological colouration change can occur in seconds in this species from low to high contrast, and cause individuals to look very different between moments. However, individual patterns appear to be stable although at least some parts can become inconspicuous during low contrast displays. Despite of this, photo ID is possible, including in some instances, where only partial patterns are visible on one image compared with another. However, true negatives (confirming two fish are not the same) can be challenging. Most identified matches were of fish photographed by different divers on the same day. Only a small number (n=9) were found with resighting durations ≥ 1 day (1 – 2,652 days). These matches demonstrate that at least some individuals return to the same reefs both within and between seasons, with the resighting duration of 7.2 years constituting the longest known example of molid site fidelity. Comparing body morphology between resightings of > 1 year (n=6) revealed limited indications of growth, contradicting the current understanding of rapid growth in captive molids (*Mola mola*), and highlighting the knowledge gap regarding growth in the wild. Continued photo ID in the Bali area could provide valuable complementary information to future growth studies using other methods as well as provide further insights into molid site fidelity.

KEYWORDS

citizen science, skin pattern stability, bold display, growth, Nusa Penida Marine Protected Area, Bali, Indonesia

1 Introduction

Photo identification of individual animals has become an established method in population ecology and is used across a wide range of taxa, including marine megafauna such as elasmobranchs, reptiles and mammals (Blount et al., 2022; Petso et al., 2022). The method has also been applied to teleosts (e.g., Giglio et al., 2014; Chaves et al., 2016; Couffer, 2017; Mucientes et al., 2019; Pedersen and Mohammed, 2021; Sèbe, 2021), but has to date not been applied to the ocean sunfishes ('molids'). The family comprises five species (Fricke et al., 2022), including the world's heaviest bony fish, giant sunfish *Mola alexandrini* (Ranzani, 1839) *sensu* (Sawai et al., 2018), capable of reaching at least 2.7 ton in body weight and at least 3.3 m in total length (TL) (Gomes-Pereira et al., 2022; Sawai and Nyegaard, 2022).

A main challenge to photo identification of individual molids (hereafter termed photo ID) lies in obtaining sufficient images. Encounter rates are generally low globally, and molid tourism is rare (Nyegaard, 2018; Thys et al., 2020). The reefs of the Nusa Penida Marine Protected Area (MPA) in Bali, Indonesia (Figure 1) provide an exception. Here, *M. alexandrini* appear seasonally to solicit parasite cleaning services from reef fish, and serve as an important draw card for local SCUBA diving tourism (Konow et al., 2006; Thys et al., 2016). While these molids occur in the area year round from the surface to well below the recreational SCUBA diving limit (40 m), SCUBA divers most commonly encounter individuals between c. July/August and November/December, locally known as the "sunfish season" (Nyegaard, 2018). Such encounters yield numerous opportunities for molid photography.

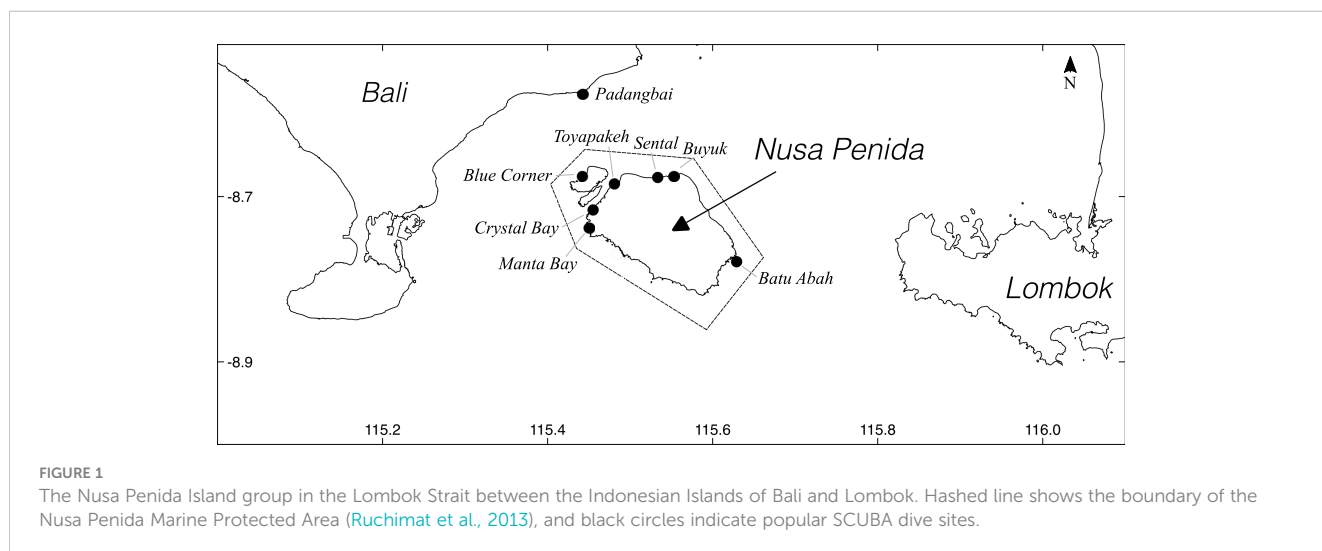
Despite the local popularity of the molids, their seasonal occurrence is not well understood. The fish are locally referred to as 'mola-mola', but have been morphologically and genetically identified as *M. alexandrini* (Thys et al., 2016; Nyegaard, 2018; see also Riawan et al., 2019), although we note not everyone agrees on the nomenclature (Britz, 2022). The size range of individuals has not been studied in the Nusa Penida MPA, but most fish appear comparable in length to the height of a human diver. Our approximation is c. 130 – 200 cm TL, based on personal *in situ*

observations and review of imagery [see Nyegaard (2018) and Supplemental Material Figure 1 for examples]. It is not known if individuals of this size are sexually mature, but for the close relative *Mola mola* (Linnaeus, 1758) maturation occurs around 150 – 160 cm TL (Forsgren et al., 2020). Further, it is not known if *M. alexandrini* are year-round residents in the area, or if they are seasonal visitors whose presence may be linked to seasonal cold water upwelling along the southern coastline of Bali and neighbouring islands (Thys et al., 2016; Nyegaard, 2018). While photo ID could be informative, not all taxa are suitable for this method, as it relies on clear, individual markings that are stable over time (Marshall and Pierce, 2012; Urian et al., 2015).

1.1 Individual markings

The elaborate body patterns on *M. alexandrini* consist of dusky-white spots, stripes and irregular shapes against a dark reddish-brown colour dorsally and laterally, merging into a dusky-white area ventrally (Sawai et al., 2018) (Figure 2A). The high degree of pattern intricacy and variation between individuals points to a well-suited taxa for photo ID, as also suggested by Kushimoto et al. (2022) for *M. mola* and sharptail sunfish *Masturus lanceolatus* (Liénard, 1840). However, the boldness of these skin patterns can change rapidly in all *Mola* species (Sawai, 2017; Nyegaard et al., 2018; Sawai et al., 2018; Thys et al., 2020; Kushimoto et al., 2022). Such 'physiological colouration change' is not uncommon among teleosts, and has been reported for numerous taxa (Kodric-Brown, 1998; Rosenthal and Marshall, 2011; Sköld et al., 2016). It is achieved through aggregation, dispersal and interaction of various light-absorbing and light reflectance pigment granules within dermal chromatophore units (Thurman, 1998; Sugimoto, 2002; Ligon and McCartney, 2016; Sköld et al., 2016).

Physiological colouration change has not been formally documented in wild molids, but has been reported at least as far back as Duhamel du Monceau (1777). In captivity, it is known that *M. mola* can produce striking shifts in body patterns during feeding and handling [M. Howard, pers comm in Thys et al. (2020)], and



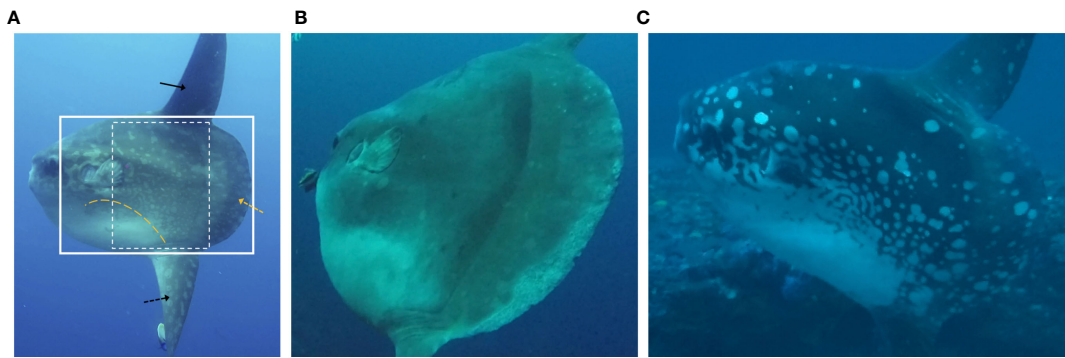


FIGURE 2

Three different *Mola alexandrini* individuals in the Nusa Penida Marine Protected Area. (A) Intricate patterns across the body and clavus (yellow hashed arrow); the pattern on both dorsal and anal fins (black solid and hashed arrows, respectively) are often obscured by shadow effects (e.g., dorsal fin here). White box: target area for photo identification; yellow hashed line: confluence of darker dorsal and lighter ventral colouration. (B) Inconspicuous and (C) highly conspicuous patterns during low contrast and bold displays, respectively. Cropped stills from unmanipulated Gopro video, natural light, orange filter. (C) Image by Richard Horner, printed with permission.

that such changes can occur in less than a minute (Kushimoto et al., 2022). These changes to pattern boldness can temporally impact individual ‘distinctiveness’ and thereby complicate photo ID (e.g., Urian et al., 2015). The challenge of photo ID in taxa capable of physiological colouration change has indeed been noted by other researchers, including for dusky grouper [*Epinephelus marginatus* (Lowe, 1834)] (Lelong, 1999), manta rays (*Mobula* spp.) (Ari, 2014), goliath grouper [*Epinephelus itajara* (Lichtenstein, 1822)] (Giglio et al., 2014) and small-spotted catshark [*Scyliorhinus canicula* (Linnaeus, 1758)] (Navarro et al., 2018).

1.2 Pattern stability

Skin pattern development and stability has not been investigated for *M. alexandrini*, but recent research documented stable and recognizable skin patterns in a captive *M. mola* individual over a 4.5 year period (Kushimoto et al., 2022). During this time the fish grew from c. 90 to c. 154 cm TL, with the images presented in Kushimoto et al. (2022) seemingly indicating that a slight ‘stretching’ may have occurred, but that the patterns were otherwise identical. However, given the large size spectrum of *Mola* spp., it is possible that new patterns could potentially develop with growth, akin to Turing’s reaction-diffusion model (Turing, 1952) of pattern formation in biological systems (for examples in teleosts see Kondo and Asai, 1995; Rosenthal and Marshall, 2011; Delcourt et al., 2018). This is particularly pertinent for *M. alexandrini* due to the large growth spectrum and changes in morphology with size (Sawai et al., 2017; Sawai et al., 2018; Gomes-Pereira et al., 2022; Sawai and Nyegaard, 2022). Most notably, *M. alexandrini* develops a large head bump, a large chin bump, and bulging lateral ridges with growth, and as for all *Mola* species, the dorsal and anal fins become shorter and broader relative to fish length (Watanabe and Sato, 2008; Nyegaard et al., 2018; Phillips et al., 2018; Sawai et al., 2018) (Figure 3).

The growth rate is unknown for *M. alexandrini*, however, rapid growth has been documented for *M. mola* in captivity, including an extreme case where an individual increased in mass from 57 to 880

kg in 15 months (Powell, 2001). Another study estimated that *M. mola* in captivity may reach c. 3 m in length in 20 – 23 years (Nakatsubo and Hirose, 2007), equating to a weight gain of c. 1,500 – 1,750 kg (Nakatsubo and Hirose, 2007; Phillips et al., 2018). If such rapid growth is representative for wild *M. alexandrini*, and if skin patterns change beyond recognition with growth, long-term photo identification could prove a significant challenge for this species.

This study aims to determine if individual photo ID based on skin patterns is possible and informative for *M. alexandrini*. To test this, we created the citizen-science platform ‘Match My Mola’ for crowd-sourcing imagery (photos and video) of *M. alexandrini* in Bali. We undertook trial Photo ID matching to explore if this is achievable despite the species’ capacity for rapid physiological colouration change. We further compared the body morphology

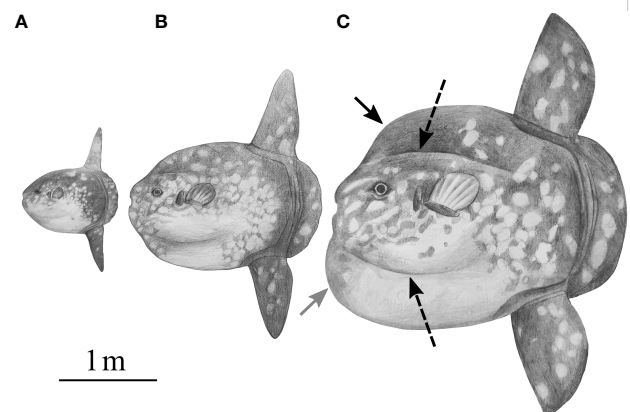


FIGURE 3

Generalised body morphology of giant sunfish (*Mola alexandrini*) of (A) 1 m (est. 48 kg), (B) 2 m (est. 484 kg), and (C) 3 m total length (est. 1,864 kg) (see Supplemental Material Figure 2 for examples). Head bump (black arrow), chin bump (grey arrow) and bulging lateral ridges (hashed arrows) develop with size. Estimated weights are based on the length-weight relationship for *M. alexandrini* in (Sawai and Nyegaard, 2022). Drawings by Robin Ljungfeldt Bryhni.

of matched individuals for indications of growth between resightings. Lastly, we discuss challenges, restrictions and possible applications of photo ID for this and other molid species.

2 Methods

2.1 Image submission

The ongoing citizen science project ‘Match My Mola’ was launched in 2013 to collect and curate new and existing *M. alexandri* imagery (photos and video) from the Bali area. Imagery can be submitted *via* an online platform, email, through social media or (at times) directly to a local project representative. The submitter is invited to name the fish. Imagery is also mined from internet sites Flickr, YouTube, and iNaturalist, as well as local tour operators’ social media accounts (Facebook and Instagram). In addition, images obtained during related molid research dives in the Nusa Penida MPA (Nyegaard, 2018) were added. The term ‘Photo Event’ here refers to one or more images of the same fish taken by the same photographer during the same dive and logged with ‘Match My Mola’.

2.2 Metadata processing

Photo Event metadata are collated in an off-line Photo Event Database built with Claris Filemaker Pro. Metadata includes date, time, location, temperature and depth of the Photo Event, date and platform of submission (or mining), photographer and submitter (or person posting), and all associated communication and links to online footprints. For mined Photo Events, the internet upload date is noted, and the authenticity of mined images and associated metadata are verified by contacting the photographer or person posting. All metadata (submitted and mined) are carefully scrutinised including comparisons of purported date of the Photo Event with image file date stamp (when available), date of submission/mining, and all digital and online footprints. Where information (provided or mined) is ambiguous or appears unreliable for any reason, the metadata resolution is reduced to the lowest verifiable level, such as general area in lieu of a dive site, or month and/or year in lieu of an exact date etc. Where the photographer or submitter cannot be contacted, the metadata is treated as ‘unverified’. For match pairs, all metadata are re-examined, rigorously scrutinised, the divers and background habitat in the images are compared and the photographers/submitters are contacted again if deemed necessary, to gain full confidence in the results.

2.3 Image processing

Photo Events are added to the database as follows: Up to three images (photos and/or video stills) are selected from each side of the fish to overall represent the lateral side of the body posterior of the eye (Figure 2A). As the dorsal and anal fins are used for propulsion

in Molidae (Watanabe and Sato, 2008), their patterns are often obscured by shadow effects, and the images are therefore cropped around the body of the fish (Figure 2A). Images showing faint body patterns are digitally enhanced using the application ‘Photos’ by Apple Inc. Following editing, images with indiscernible skin patterns are omitted (e.g., blurry, low resolution, sunfish in silhouette, grossly over- or under-exposed, distant sunfish, sunfish displaying no patterns). However, poor quality images, where skin patterns are nevertheless discernible, are included according to a subjective expectation of the possibility of matching. Each photo event, consisting of either images of the left, right, or both sides of the molid, is assigned a unique Photo Event number and filed in a left and right Photo Event Catalogue.

2.4 Matching

At the time of this study, 1,098 Photo Events from the Bali area were available from ‘Match My Mola’. Matching of Photo Events was undertaken by three observers for the left (n=606) and right (n=633) sides separately. Observer 1 matched all left and right side Photo Events, while observers 2 and 3 each matched a subset of these (Observer 2: n=358 left side, Observer 3: n=354 right side). The observers compared images side-by-side in a random, pairwise manner, using the skin patterns for matching. A particularly useful area for initiating a match check was found to be the confluence of the darker dorsal area and lighter ventral area (Figure 2A). All Match Events (i.e., two Photo Events deemed to depict the same fish) were verified by a fourth observer, who checked all visible patterns across the fish for similarity. Any physical marks such as clavus damage, skin irregularities, scars and injuries were used as secondary verification.

3 Results

3.1 Pattern clarity and physiological colouration change

The clarity of molid skin patterns varied markedly across Photo Events, ranging from fish with no or vague patterns (Figure 2B), to fish with intricate patterns in black and white across most of the body, clavus and fins (Figure 2C). Pattern clarity was clearly influenced by the physiological state of the patterns themselves (i.e., the result of physiological colouration change), as well as by external factors (e.g., light conditions, depth/light attenuation, water clarity, angle and distance between camera and fish, use of artificial light) combined with image quality (e.g., pixel resolution, focus, exposure, shutter speed). To distinguish between the true state of patterns versus the appearance on images, ‘boldness’ herein refers to the physiological state, while ‘conspicuity’ refers to the patterns as they appear on images, i.e., the result of the interplay between boldness, external factors and image quality. This distinction was made to acknowledge the inherent difficulty in determining true pattern boldness on arbitrary underwater images.

A total of n=363 Photo Events included video. Changes in pattern conspicuity was evident on more than a third of these, but

most occurred during movements of molid and/or camera. In other words, it was typically not possible to establish that physiological colouration change had indeed occurred, rather than a sudden change in light conditions causing the patterns to appear more conspicuous. Only in a small number of cases could the changes confidently be attributed to physiological colouration change. Combined, these indicated that 1) pattern boldening can happen rapidly in *M. alexandrini*, e.g., in <10 seconds (Figure 4), and can render the same fish looking very different between moments (Figure 5); 2) at least some parts of the body patterns can become inconspicuous during low contrast, but once ‘emerged’, the patterns remain stable during further physiological colouration change (Figure 6); 3) bold displays appear to be a result of ‘contrast enhancement’ seemingly due to both darkening dark areas, and lightening light areas of the underlying pattern. It is unclear, however, if *M. alexandrini* can conceal the skin patterns entirely, or if uniformly grey individuals are just ‘missing’ their patterns as an artefact of low quality images and/or suboptimal light conditions (e.g., Figures 4A, B) although high quality images of uniform grey individuals seemingly indicate the former (e.g., Figure 2B). Note here that ‘grey’ refers to the typical colour on underwater images, which may not reflect the true colour of the fish.

3.2 Matching

Observer 1 – 3 reported that matching was straight forward when pattern conspicuity was high (e.g., Figure 7), but was more challenging for fish with low pattern conspicuity, especially when image quality was low and/or differing angles between fish required mental geometric rotation. Not all observers found all matches; the subset of Photo Events matched by two observers yielded at total of 20 (left) and 22 (right) unique Match Events. Of these, Observer 1 found 80% (left) and 91% (right), Observer 2 found 100% (left) and Observer 3 found 59% (right).

Combined across all Photo Events (i.e., matched by one or two observers), the three observers found a total of 107 unique Match Events (60 left, 47 right). All Match Events were confirmed by the fourth observer, with no false matches identified. Approximately half of the Match Events (52% left, 54% right) were secondarily verified based on physical marks, or from matching based on the other side of the fish (e.g., Figure 7).

Most Match Events consisted of image pairs with similar skin pattern extents (e.g., Figure 7). A small number (n=17) consisted of images where only a subset of patterns was visible on one Photo Event compared with the other. All pairs were highly similar in body morphology (i.e. appeared to be the same fish), with n=6 pairs secondarily verified *via* physical marks. The differences in pattern extent appeared to be caused by a combination of low contrast display by the fish and poor image quality, which rendered parts of the patterns on one image indiscernible compared with the other (see Supplemental Material Figure 3 for an example).

The 107 unique Match Events mostly comprised same-day (n=32) or likely same-day Photo Events (incomplete/unverified data) (n=50), i.e., individual fish which had been photographed on the same day by different photographers and submitted

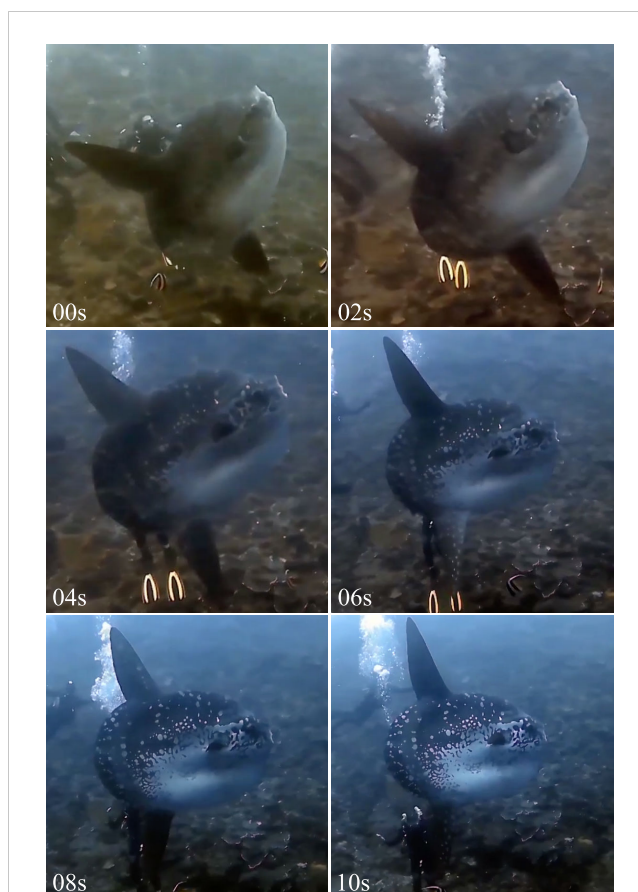


FIGURE 4

A giant sunfish (*Mola alexandrini*) during rapid physiological colouration change from low contrast to bold display. Unmanipulated stills in two second (s) intervals from digital video (natural light, red filter) by Amin Kancil, Nusa Penida Marine Protected Area, Indonesia (2019). Printed with permission. Video available at <https://www.facebook.com/amin.kancil/videos/2776134922397013/>.

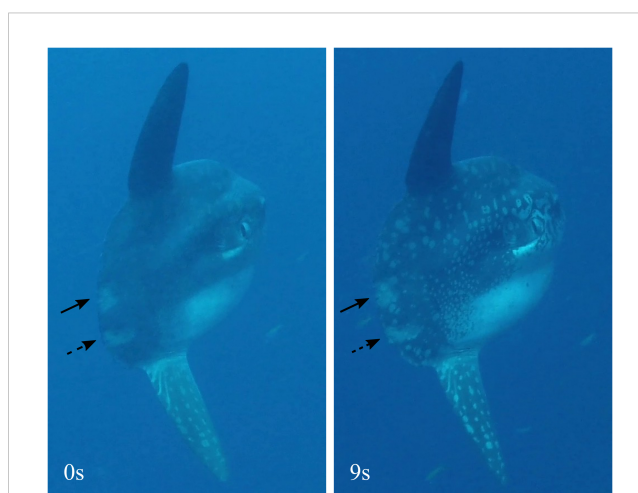


FIGURE 5

A *Mola alexandrini* individual in the Nusa Penida Marine Protected Area, Indonesia, revealing dots and patterns not evident seconds (s) prior. Arrows point to clavus damage, evident during both pattern states. Unmanipulated stills from video (natural light) by Sandra Clopp, printed with permission.

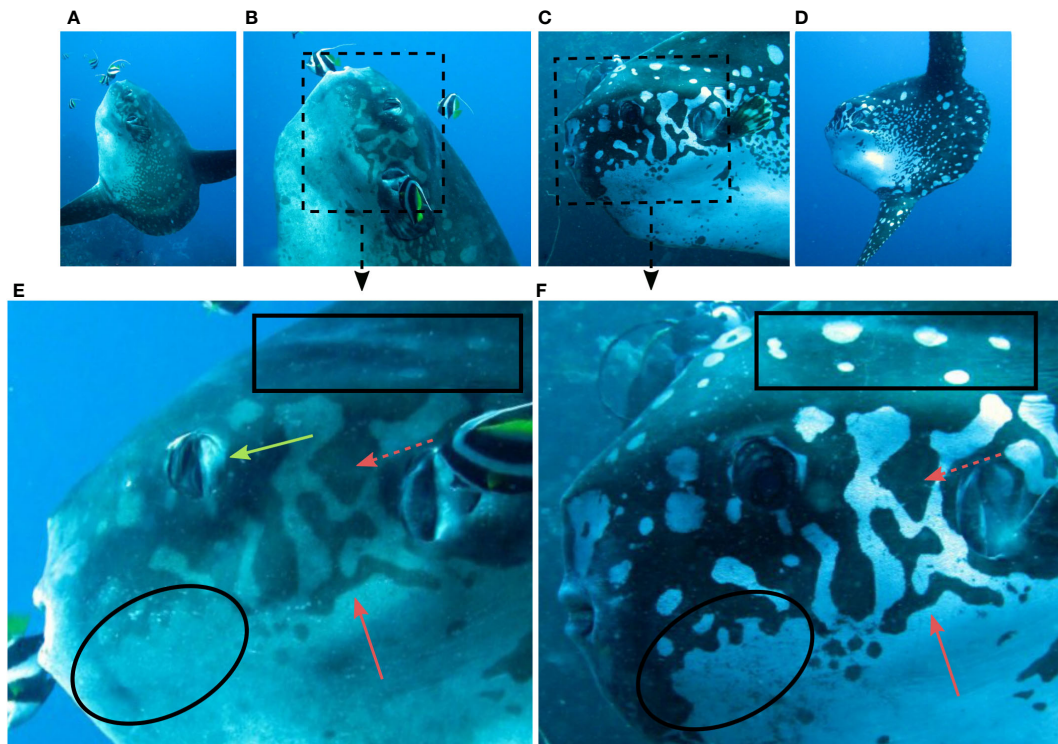


FIGURE 6

A *Mola alexandrini*, individual, Nusa Penida Marine Protected Area, Indonesia. (A) The fish was initially in cleaning position (head angled upwards, nictitating membranes covering eyes). As the photographer moved closer (B), the fish righted itself (C), withdrew the nictitating membranes and displayed bold skin patterns before (D) moving away. Inserts (E, F): The skin patterns were stable during the change (compare red arrows), but some patterns were inconspicuous or vague during low contrast (compare black squares and circles). Yellow arrow: nictitating membrane covering the eye. Unmanipulated photos (except (E, F), which were cropped). The images were taken within 60 seconds of each other by Roberto Piazza, 2012, printed with permission.

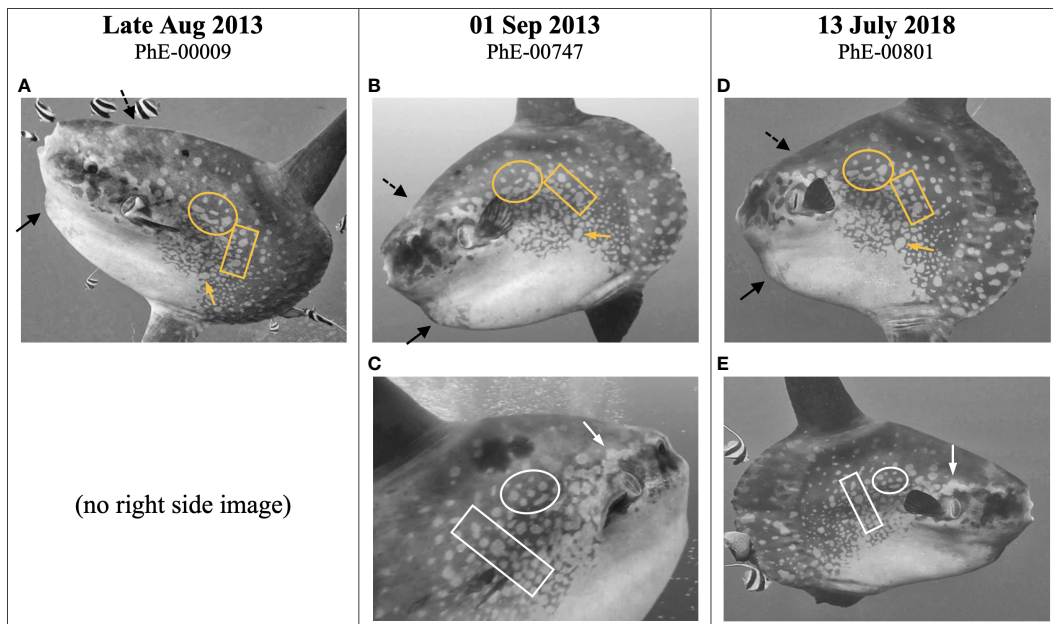


FIGURE 7

Three separate Photo Events of the same *Mola alexandrini* individual (“Concept”) at dive site Crystal Bay, Nusa Penida Marine Protected Area, Indonesia (Figure 1). See Supplemental Material Table 1 for full metadata details. Coloured arrows and boxes indicate examples of matching patterns. Black arrows point to chin bump (solid) and head bump (hashed). ‘PhE’ is the Photo Event number. (A) Photo by Sabrina Si Sadi; stills from video by (B, C) Michael Shark and (D, E) Niamh Lynch, printed with permission. All images were converted to black and white and the contrast was adjusted to enhance the patterns.

independently of each other. A further $n=9$ Match Events had unverified and/or insufficient metadata to determine the duration between resightings. The remaining Match Events ($n=15$) comprised nine individuals, which had been photographed with intervals of 1–2,652 days (Figure 8). The high quality of the extensively scrutinised metadata for the associated Photo Events boosted confidence in the results (see Supplemental Material Table 1 for details).

3.3 Fish size

Remarkably, all matched individuals had similar body morphologies between Photo Events despite some being separated by several years (Figures 7, 9A–E). For example, Fish “Fosforito” was photographed with 3.2 years interval with no indication of morphological changes (and thereby growth) during this time. Generally, no size estimates are available for Match My Mola submissions, however, an accurate size estimate was available for “Fosforito” (c. 160–180 cm TL) from the second Photo Event, where first author M. Nyegaard was able to compare her own height to the length of the fish during in-water biopsy sampling. Back-casting using the Von Bertalanffy growth curve for captive *M. mola* (Nakatsubo and Hirose, 2007), “Fosforito” would have been c. 63–95 cm TL at the first Photo Event 3.2 years earlier (PhE-00007; Figure 9C). Using the length-weight relationship for *M. alexandrini* (Sawai and Nyegaard, 2022) the difference in body mass would have been substantial; c. 10–41 kg (first Photo Event), and c. 231–341 kg (second Photo Event). However, such a small size during the first Photo Event is incompatible with both the body morphology and the in-water observations by the experienced dive guide who submitted the photograph (K. Belykh, pers com 2013). Instead, the comparable body morphologies indicate limited growth between the two Photo Events.

Similar theoretical calculations and hypothesized body sizes between Photo Events become increasingly unrealistic for other Match Events with longer re-sighting durations (Figure 7, Figures 9D, E). For example, the individual labelled “Phili” was

re-sighted after 7.2 years (Figure 8). This fish had remarkably similar body morphology during both Photo Events and was also similar in size relative to the surrounding cleaner fish (Figure 9E) indicating limited growth between re-sightings. For comparison, during 7.2 years in captivity, *M. mola* is estimated to grow from the ‘size at age 0’ (*Mola* spp. hatch at c. 2 mm; Martin and Drewry, 1978) to c. 209 cm TL, corresponding to an estimated weight gain of 493–572 kg (Nakatsubo and Hirose, 2007; Phillips et al., 2018).

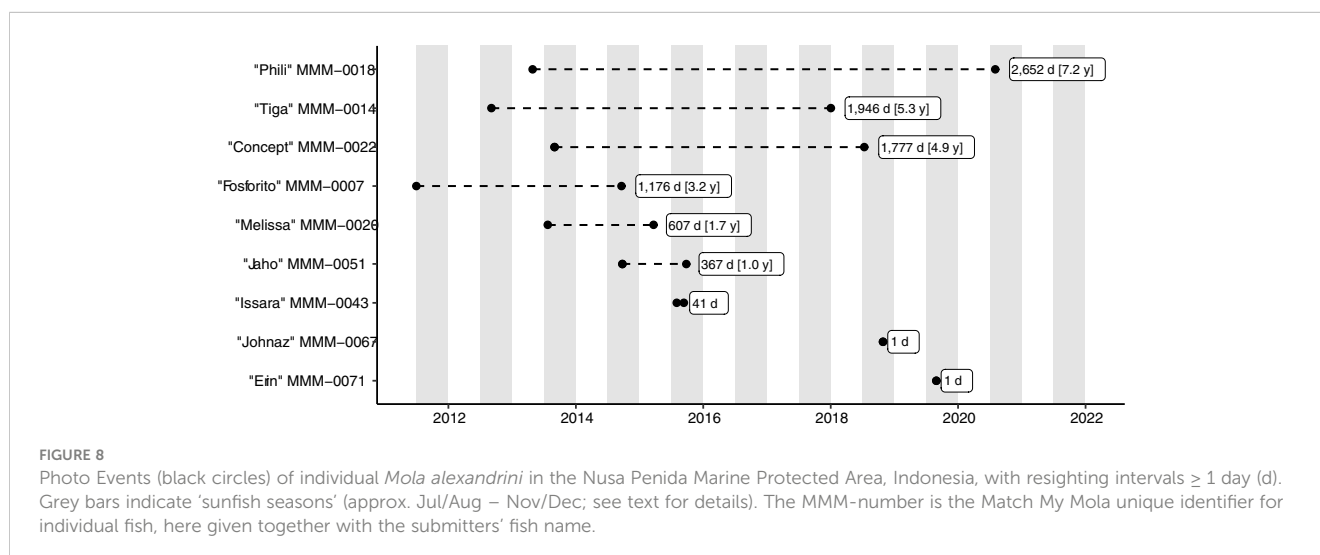
4 Discussion

We undertook trial Photo ID matching of giant sunfish (*M. alexandrini*) based on 1,098 citizen science imagery submissions from the Bali area. Matching was possible despite the capacity of *M. alexandrini* to rapidly (within seconds) change the boldness of their skin patterns, and consequently look very different between moments. Most matches were same-day photos by different divers, with a small number ($n=9$) with longer resighting durations (1–2,652 days). For these, the body morphology was virtually identical between matches, suggesting limited growth, starkly contradicting the current understanding of rapid growth in captive molids (*M. mola*).

4.1 Matching

Our study confirmed that physiological colouration change in *M. alexandrini* from low contrast to bold patterns can occur in seconds, and that the same individual can look very different between moments, corroborating observations of captive *M. mola* (Kushimoto et al., 2022). This ability clearly influences pattern conspicuity on images and thereby the ‘distinctiveness’ of individual between moments (e.g., Urian et al., 2015).

Our study confirmed that true positive matches (correctly determining that two individuals are the same) is nevertheless possible, as also suggested for *M. mola* (Kushimoto et al., 2022). Our study furthermore confirmed that it is possible to match



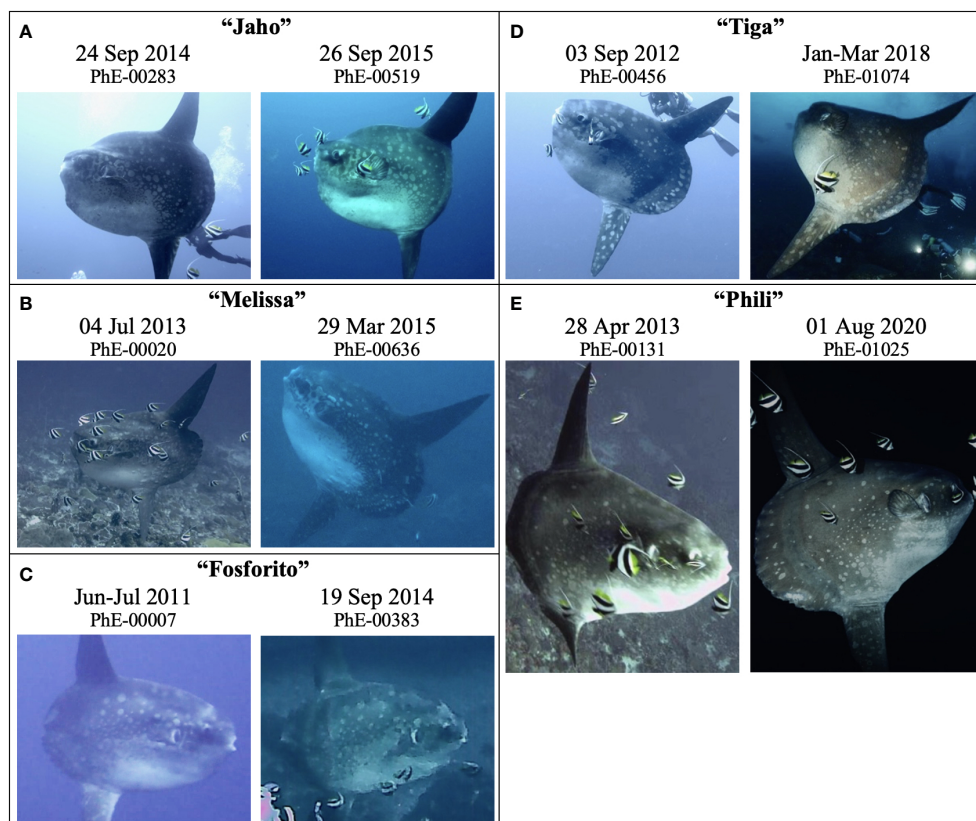


FIGURE 9

Five different individuals of *Mola alexandrini*, photographed in the Nusa Penida Marine Protected Area, with 1.0 – 7.2 years interval. 'PhE' is the Photo Event number. See [Supplemental Material Table 1](#) for full metadata details, and [Supplemental Material Figure 4](#) and [Supplemental Material Figure 5](#) for detailed comparison of skin patterns between Photo Events. Photos or stills from video by: (A) (left) Choocart Treetavekul; (B) (left) Emanuel Groult; (B) (right) Marianne Nyegaard; (B) (right) Ben Coffey; (C) (left) Lembongan Dive Adventure; (C) (right) Jonathan Andersen; (D) (left) Tomoki Ueda; (D) (right) Benny Gunawan; (E) (left) Aquatic Alliance; (E) (right) Keith Mash. All images printed with permission.

individuals where the skin pattern extent differs between images. However, determining true negative matches (correctly determining that two individuals are not the same) is challenging for fish in low contrast display, in particular where image quality is low. Aggressive image grading to exclude fish with little or vague patterns could solve this issue, but would require development of a 'cut-off' point (non-trivial), and would reduce the number of images available for positive matches – a significant issue due to the time consuming task of crowd sourcing images and reliable metadata.

4.2 Physiological colouration change

The biological function of rapidly changing displays in *Mola* spp. remains unclear. Physiological colouration change in fish is complex and can serve both as intra- and inter-specific communication, as well as crypsis and means of predator avoidance (Rosenthal and Marshall, 2011; Sköld et al., 2016; Figon and Casas, 2018). As the bold skin patterns in *M. alexandrini* render individuals striking and highly visible, it presumably serves as a signal rather than camouflage [although the visual ability of the receiver ultimately determines how a display is perceived (Stuart-Fox and Moussalli, 2009; Marshall et al., 2019)].

In some taxa, bold displays are associated with aposematism (signaling toxicity) (e.g., Figon and Casas, 2018), however this is doubtful for *M. alexandrini*. Despite previous beliefs that *Mola* spp. contain tetrodotoxin and is toxic to marine mammals (e.g., Halstead 1968 in Gladstone, 1988) this remains unsubstantiated (Baptista et al., 2020; Baptista et al., 2022) and seems unlikely as molids fall prey to both orca (Visser et al., 2023) and sealions (Powell, 2001; Nyegaard et al., 2019).

In other teleost taxa, boldening of patterns and/or darkening body colours is a fright response (Potts, 1974; Beeching, 1995), and in yet others is associated with intra-specific aggression and/or dominance (Hamilton and Peterman, 1971; Dawkins and Guilford, 1993; Delcourt et al., 2018). Although quantitative data is lacking, the boldening of skin patterns in *M. alexandrini* in the Nusa Penida MPA appears to be associated with disturbance during cleaner fish interactions (*M. Nyegaard pers obs*). While a fright response cannot be ruled out, the bold patterns may be a warning signal to intimidate intruding divers by accentuating the molid's large body size. Indeed, harassed *M. alexandrini* individuals sometimes 'glide' slowly past the intruding divers rather than swimming off, seemingly 'show-casing' both the large body size and vibrant lateral display (*M Nyegaard, pers obs*). In turn, *M. alexandrini* engaged in cleaning are typically displaying low contrast patterns

(M Nyegaard pers obs). This may be a neutral pattern state, or could be a component of cleaner-client signaling, perhaps serving to render skin parasites more visible to the cleaner fish, as seen in some teleost taxa (Hobson, 1965; Hobson, 1969; Hobson, 1971; Caves et al., 2018).

If boldening of skin patterns in *M. alexandrini* is indeed a response to diver disturbance then any behavioral differences between individual *M. alexandrini* (akin to the bold-shy trait continuum; Hulthén et al., 2014) could potentially introduce bias in photo identification towards individuals (or perhaps sex)? which more readily display bold patterns. Bias within the same individual could even arise over time if molids develop a tolerance to diver disturbances (e.g. Titus et al., 2015), or shy individuals become more bold (e.g., Hulthén et al., 2014), influencing their propensity to display bold patterns. Such challenges and biases must be considered in relation to individual research questions where photo ID is applied, including for creation of discovery curves and application of capture-recapture models (e.g. Germanov et al., 2019).

4.3 Within-season Match Events

The findings of several Match Events consisting of same-day Photo Events submitted by different photographers was not unexpected as molids in the Nusa Penida MPA are frequently surrounded by large numbers of divers (Nyegaard, 2018; M. Nyegaard, T. Thys, J. Karmy, M. Welly pers. obs.). In contrast, the low number of multi-day matches within the same sunfish season was unexpected, as telemetry studies have found that at least some individual *M. alexandrini* exhibit seasonal site fidelity to the area within sunfish seasons (Thys et al., 2016; Nyegaard, 2018). During the 2015 sunfish season, four individuals tagged with high-accuracy Fast-loc satellite tags appeared to utilise the Lombok Strait to forage, with occasional visits very close to shore, both off Nusa Penida and Bali (Nyegaard, 2018) – presumably to seek cleaner fish interactions. However, only four within-season Match Events were found in the current study, with two of these being on consecutive days. It is unclear if our results reflect low within-season re-visitation rates by individuals, or if re-visitations occur more frequently, but were not captured by the relatively low number of Photo Events per season in this study. Or, if once photographed, fish avoid re-entering areas replete with piscine paparazzi, akin to hook-shy fish avoiding hooks for some time after having been hooked once (e.g., Brown, 2015).

4.4 Inter-annual Match Events

The number of inter-annual Match Events (n=6), while low, provided unexpected results with regards to the limited indications of growth over the 1.0 – 7.2 year duration between Photo Events. One possible explanation is that the six re-sighted individuals were outliers, which had simply failed to grow, allowing them to be matched due to their unchanged body patterns. However, captive *M. mola* individuals retain stable body patterns which are

recognisable over at least 4.5 years and across a growth span of at least c. 90 – c. 156 cm (Kushimoto et al., 2022). Consequently, we would expect to find inter-annual matches of *M. alexandrini* exhibiting at least some evidence of growth, e.g., with increased size of the head bump and chin bump, increased swelling of the lateral ridges, and decreased aspect ratios of the dorsal and anal fins, however, no such matches were found in this study.

Another possibility is that all matched individuals were males at or near their maximum size, exhibiting slow growth. In *M. alexandrini* females appear to grow larger than males (Sawai and Nyegaard, 2022), similar to both *M. mola* (Sawai et al., 2011; Sawai et al., 2018) and *Ma. lanceolatus* (Liu et al., 2009). The maximum size of *M. alexandrini* males is not known, but is at least 197 cm TL (Vogelnest, 2003; see Supplemental Material Figure 6). An all-male Match Event scenario at Nusa Penida, however, still does not account for the lack of inter-seasonal matches of individuals exhibiting at least some evidence of growth (i.e., females) – unless only males frequent the Nusa Penida MPA. As external morphological sexual dimorphism has not been established for *M. alexandrini* (apart from size) the latter cannot currently be explored based on ‘Match My Mola’ imagery.

It is also possible our findings reflect markedly slower growth in wild *M. alexandrini* compared with captive *M. mola*. Captivity indeed appears to influence life history traits in *M. mola*. Nakatsubo and Hirose (2007) found that both body weight and gonadosomatic index were higher relative to TL in captive versus wild *M. mola*. Further, the diet of captive *M. mola* must be carefully controlled to avoid obesity (Howard et al., 2020). Currently, the only study on growth in wild Molidae is for *Ma. lanceolatus*. Based on vertebrae growth ring chronology, Liu et al. (2009) estimated longevity (i.e., years to reach 99% of the estimated maximum size) of 105 and 82 years for female and male *Ma. lanceolatus*, respectively. While this lends credibility to a slow-growth scenario for wild *M. alexandrini*, it is currently unknown if the findings of Liu et al. (2009) are representative of wild *Mola* spp. In summary, our findings highlight the current knowledge gaps in age and growth in wild *Mola* spp. It is essentially unclear whether these unusual fishes are short-lived and exhibit fast growth (akin to captive *M. mola*), or are long(er)-lived with slow(er) growth (as reported in wild *Ma. lanceolatus*).

Molid age and growth information is critical to gauge resilience to anthropogenic pressures such as interactions with commercial fisheries, and to establish effective conservation management plans (Liu et al., 2015; Forsgren et al., 2020; Nyegaard et al., 2020). *Mola* spp. otoliths are small and granular (Nolf and Tyler, 2006) and inadequate for age estimates, so other approaches are needed (Hays et al., 2020). Eye lens nuclei chronology (e.g., Nielsen et al., 2016), vertebrae chronology (e.g., Liu et al., 2009), or epigenetics (e.g., Mayne et al., 2021) may potentially be informative, and if growth is slow then bomb radiocarbon dating (e.g., Allen and Andrews, 2012) could be relevant for very large *M. alexandrini* individuals. Photo ID catalogues such as ‘Match My Mola’ could potentially provide important supportive information. Specifically, by establishing relative morphometrics (for example dorsal fin aspect ratio relative to fish length) body size and growth between Photo Events could potentially be estimated. However, further research is needed to establish such relative morphometrics for *M.*

alexandrini, and to explore the robustness of such an approach. Relative morphometrics could be obtained underwater from live molids using photogrammetry (Deakos, 2010) and from preserved museum specimens (Nyegaard and Sawai, 2018).

4.5 Population indicators

Very few multi-day matches were found in our study. While a direct comparison is not possible due to differing methods, a photo ID study of manta rays [*Mobula alfredi* (Krefft, 1868)] in the Nusa Penida MPA recently found 624 uniquely identified individuals (6,087 images; June 2004–April 2018), with 82% of these sighted more than once, 29% sighted more than 10 times, and 5% sighted 31–99 times (Germanov et al., 2019).

The low re-sightings rate in our study may be linked to the limited overlap between molids and SCUBA divers in the Bali area. Here, *M. alexandrini* forage in deep water of several hundred meters, spend little time near the surface (Nyegaard, 2018), and generally only overlap with SCUBA divers when seeking cleaner fish interactions on reefs of <40 m water depth (with a frequency currently unknown). In contrast, manta rays can be seen off Nusa Penida during a wider spectrum of behaviours, including cleaner fish interactions, foraging, courtship and cruising (Germanov et al., 2019). Individual manta ray observations and interactions furthermore occur over much longer periods, in much shallower waters and in much less challenging current conditions compared with *M. alexandrini* (Nyegaard pers obs). Overall, this provides more frequent and better photo opportunities for manta rays by both SCUBA divers and snorkelers, and allows for faster acquisition of high quality images for photo ID, which overall increases the chances of resighting the same animal when it is in the area.

Our low re-sightings rate may also reflect differences in site fidelity to the Nusa Penida area between manta rays and molids. *Mobula alfredi* individuals are generally sighted multiple times over several years (Germanov et al., 2019). In turn, at least some *M. alexandrini* individuals move widely (at least hundreds of km) both within and outside the sunfish season (Thys et al., 2016; Nyegaard, 2018), with unknown visitation rates to the Nusa Penida reefs both within, outside and between seasons.

In comparison, both small-scale site fidelity and wide movements have been recorded elsewhere for *M. alexandrini*. Acoustic telemetry in the Galapagos Islands found a high degree of site fidelity to a “sunfish hotspot” (Punta Vicente Roca), whereby three of five tagged *M. alexandrini* were detected nearly year-round over a duration of 202–733 days (Thys et al., 2017). However, another individual tagged with a satellite tag first remained local for c. one month, then moved 2,700 km westward in c. two weeks (Thys et al., 2017). Similarly, some individuals tagged in the Nusa Penida MPA have been tracked moving away hundreds of km, both within and outside the sunfish season (Thys et al., 2016; Nyegaard, 2018), and most recently, four *M. alexandrini* tagged in Taiwan moved between 552–6,952 km; two travelled north (one reached Japan after 178 days), while two travelled south and crossed the equator,

with one reaching New Caledonia after 240 days (Chang et al., 2021).

It is not clear if *M. alexandrini* in the Bali area belong to a large and/or open population, to what extent individuals remain in the area outside the sunfish season, nor if some/all undertake large scale movements of hundreds or thousands of kilometers. However, our study demonstrates that among a relatively limited number of Photo Events from the small area of Nusa Penida, we nevertheless resighted a small number of individuals with several years interval (longest of 7.2 years). This is to the best of our knowledge the longest example of site fidelity in any molid species, and the first confirmation that the same individual may return to the Nusa Penida MPA reefs between years. This site fidelity may be further explored through continued photo ID, in particular if combined with acoustic telemetry within the Nusa Penida MPA. Satellite tracking and population genetics could shed light on wider movements and connectivity with *M. alexandrini* elsewhere, including in the waters of Taiwan and Japan, where molids are fished commercially for human consumption (Nyegaard et al., 2020).

4.6 Other applications

Common uses of curated Photo-ID image databases include determining population parameters such as sex ratios and maturity based on external morphology, however, neither is currently possible for *M. alexandrini* (or other molids) as external morphological sexual dimorphism and size at maturity is unknown (Hays et al., 2020; Nyegaard et al., 2020). However, if relative morphometrics were established, population size distributions within and between seasons could be estimated and compared between seasons. Other potential applications include assessing injury rates from boat strikes and entanglement in fishing gear, which would be informative in understanding impacts from human marine activities within the Nusa Penida MPA.

To date, ‘Match My Mola’ has confirmed that molids seen by SCUBA divers on Nusa Penida and Bali reefs are almost exclusively *M. alexandrini* (Nyegaard, 2018). Recently, SLKT et al. (2021) also reported *M. mola* in the Nusa Penida MPA, however, examination of images provided to us by one of the authors points to *M. alexandrini* (Nyegaard pers obs). To date, the only confirmed records known to us of other species in the area are three videos of *Ma. lanceolatus* lodged with ‘Match My Mola’. These were recorded by: 1) snorkelers near the surface at Toyapakeh, Nusa Penida, on 06 July 2017; 2) SUBA divers at 40 m off Nusa Penida on 31 October 2018; and 3) SCUBA divers off Padangbai, east coast of Bali, on 03 October 2019 (Supplemental Material Figure 7). The videos appear to be of three different individuals, all swimming, and none interacting with cleaner fish. *Masturus lanceolatus* is known to occur in the Bali area from stranding events (M Nyegaard unpublished data) and is also listed as a species occurring in commercial and/or artisanal fisheries and/or at fish markets in southern Indonesia (White et al., 2013).

4.7 Automated recognition

As for other taxa, undertaking manual matching of *M. alexandrini* images is extremely time consuming. Computerised recognition based on algorithms and application of machine learning has been developed for numerous taxa (e.g., <https://www.wildme.org>; Cheema and Anand, 2017; Miele et al., 2020; Blount et al., 2022) and can significantly ease the matching task. However, these approaches typically require sizeable image datasets to train and test the matching algorithms. Obtaining sufficiently large datasets for elusive species such as molid can be challenging. An alternative approach that shows promising preliminary results for *M. alexandrini* combines traditional use of algorithms and machine learning, without the need for prior training (Pedersen et al., 2022, Pedersen et al., 2023). Further development of this method could increase match capacity, and be applied to other molid hotspots such as the Galapagos Islands (www.oceansunfish.org), and perhaps aid in photo identification of other elusive taxa.

Data availability statement

By default, the citizen science images submitted to Match My Mola are not shared or published unless explicit permission is provided by the photographer or submitter. The generated match data can be shared upon request. Requests to access the datasets should be directed to mnyegaard@oceansunfishresearch.org.

Ethics statement

The animal study was reviewed and approved by Animal Ethics Committee, Murdoch University, Western Australia (permit R2542/12).

Author contributions

MN founded the photo ID catalogue in 2013 and manages the database, JK and LM assisted in obtaining images, curating and verifying metadata and undertook image matching; MN undertook match checks, did summary statistics and wrote the manuscript, TT critically reviewed the manuscript, RD and MW provided field support to MN and scientific input in the initial stages of the initiative, provided assistance with obtaining Indonesian research permits, shared local knowledge and networks, and critically reviewed the manuscript. All authors contributed to the article and approved the submitted version.

References

- Allen, L. G., and Andrews, A. H. (2012). Bomb radiocarbon dating and estimated longevity of giant Sea bass (*Stereolepis gigas*). *Bull. South. Calif. Acad. Sci.* 111, 1–14. doi: 10.3160/0038-3872-111.1.1
- Ari, C. (2014). Rapid coloration changes of manta rays (Mobulidae). *Biol. J. Linn. Soc.* 113, 180–193. doi: 10.1111/bij.12321

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmars.2023.1179467/full#supplementary-material>

- Baptista, M., Braga, A. C., Rosa, R., and Costa, P. R. (2022). Does ocean sunfish *Mola* spp. (Tetraodontiformes: Molidae) represent a risk for tetrodotoxin poisoning in the Portuguese coast? *Mar. Drugs* 20, 594. doi: 10.3390/md20100594
- Baptista, M., Figueiredo, C., Lopes, C., Costa, P. R., Dutton, J., Adams, D. H., et al. (2020). "Biotoxins, trace elements, and microplastics in the ocean sunfishes (Molidae)," in *The ocean sunfishes: evolution, biology and conservation*. Eds. T. M. Thys, G. C. Hays and J. D. R. Houghton (Boca Raton: CRC Press), 186–215. doi: 10.1201/9780429343360-11
- Beeching, S. C. (1995). Colour pattern and inhibition of aggression in the cichlid fish *Astronotus ocellatus*. *J. Fish Biol.* 47, 50–58. doi: 10.1111/j.1095-8649.1995.tb01872.x
- Blount, D., Gero, S., Van Oast, J., Parham, J., Kingen, C., Scheiner, B., et al. (2022). Flukebook: an open-source AI platform for cetacean photo identification. *Mamm. Biol. (Special Issue)* 102, 1005–1023. doi: 10.1007/s42991-021-00221-3
- Britz, R. (2022). Comments on the holotype of *Orthragoriscus alexandrinii*, Ranzani 1839 (Teleostei: Molidae). *Zootaxa* 5195, 391–392. doi: 10.11646/zootaxa.5195.4.6
- Brown, C. (2015). Fish intelligence, sentience and ethics. *Anim. Cogn.* 18, 1–17. doi: 10.1007/s10071-014-0761-0
- Caves, E. M., Green, P. A., and Johnsen, S. (2018). Mutual visual signaling between the cleaner shrimp *Ancylomenes pedersoni* and its client fish. *Proc. R. Soc. B Biol. Sci.* 285, 20180800. doi: 10.1098/rspb.2018.0800
- Chang, C. T., Chiang, W. C., Musyl, M. K., Popp, B. N., Lam, C. H., Lin, S. J., et al. (2021). Water column structure influences long-distance latitudinal migration patterns and habitat use of bumphead sunfish *Mola alexandrinii* in the Pacific Ocean. *Sci. Rep.* 11, 21934. doi: 10.1038/s41598-021-01110-y
- Chaves, L. C. T., Hall, J., Feitosa, J. L. L., and Côté, I. M. (2016). Photo-identification as a simple tool for studying invasive lionfish *Pterois volitans* populations. *J. Fish Biol.* 88, 800–804. doi: 10.1111/jfb.12857
- Cheema, G. S., and Anand, S. (2017). Automatic detection and recognition of individuals in patterned species. *Lect. Notes Comput. Sci. (including Subser. Lect. Notes Artif. Intell. Lect. Notes Bioinformatics)* 10536, 27–38. doi: 10.1007/978-3-319-71273-4_3
- Couffer, M. C. (2017). Individually-unique spot patterns of young-of-the-year giant sea bass (*Stereolepis gigas*) in captive-raised fish. *Bull. South. Calif. Acad. Sci.* 116, 98–109. doi: 10.3160/soca-116-02-98-109.1
- Dawkins, M. S., and Guilford, T. (1993). Colour and pattern in relation to sexual and aggressive behaviour in the bluehead wrasse *Thalassoma bifasciatum*. *Behav. Processes* 30, 245–251. doi: 10.1016/0376-6357(93)90136-F
- Deakos, M. H. (2010). Paired-laser photogrammetry as a simple and accurate system for measuring the body size of free-ranging manta rays *Manta alfredi*. *Aquat. Biol.* 10, 1–10. doi: 10.3354/ab00258
- Delcourt, J., Ovidio, M., Denoël, M., Muller, M., Pendeville, H., Deneubourg, J. L., et al. (2018). Individual identification and marking techniques for zebrafish. *Rev. Fish Biol. Fish.* 28, 839–864. doi: 10.1007/s11160-018-9537-y
- Duhamel du Monceau, H. (1777). *Traité général, des pesches, et histoire des poissons qu'elles fournissent, tant pour la subsistance des hommes, que pour plusieurs autres usages qui ont rapport aux arts et au commerce* (Paris: Saillant & Nyon; Plueve Desaint).
- Figon, F., and Casas, J. (2018). Morphological and physiological colour changes in the animal kingdom. *eLS* 1–11. doi: 10.1002/9780470015902.a0028065
- Forsgren, K., McBride, R. S., Nakatsubo, T., Thys, T. M., Carson, C. D., Tholke, E. K., et al. (2020). "Reproductive biology of the ocean sunfishes," in *The ocean sunfishes: evolution, biology and conservation*. Eds. T. M. Thys, G. C. Hays and J. D. R. Houghton (Boca Raton: CRC Press), 87–104. doi: 10.1201/9780429343360-6
- Fricke, R., Eschmeyer, W. N., and van der Laan, R. (2022) *Eschmeyer's catalog of fishes - version of 4 Oct 2022*. Available at: <https://www.calacademy.org/scientists/projects/eschmeyers-catalog-of-fishes> (Accessed October 23, 2022).
- Germanov, E. S., Bejder, L., Chabanne, D. B., Dharmadi, D., Hendrawan, I. G., Marshall, A. D., et al. (2019). Contrasting habitat use and population dynamics of reef manta rays within the Nusa Penida Marine Protected Area, Indonesia. *Front. Mar. Sci.* 6. doi: 10.3389/fmars.2019.00215
- Giglio, V. J., Adeli-Alves, J., and Bertoncini, A. A. (2014). Using scars to photo-identify the goliath grouper, *Epinephelus itajara*. *Mar. Biodivers. Rec.* 7. doi: 10.1017/S1755267214001080
- Gladstone, W. (1988). Killer whale feeding observed underwater. *J. Mammal.* 69, 629–630. doi: 10.2307/1381360
- Gomes-Pereira, J., Pham, C., Catarino, D., Miodonsky, J., Santos, M. A. R., Dionisio, G., et al. (2022). The heaviest bony fish in the world: a 2744 kg giant sunfish *Mola alexandrinii* (Ranzani, 1839) from the North Atlantic. *J. Fish Biol.* 102, 1–4. doi: 10.1111/jfb.15244
- Hamilton, W. J., and Peterman, R. M. (1971). Countershading in the colourful reef fish *Chaetodon lunula*: concealment, communication or both? *Anim. Behav.* 19, 357–364. doi: 10.1016/S0003-3472(71)80017-9
- Hays, G. C., Houghton, J. D. R., Thys, T. M., Adams, D. H., Ahuir-Baraja, A. E., Alvarez, J., et al. (2020). "Unresolved questions about ocean sunfishes, Molidae: a family comprising some of the world's largest teleosts," in *The ocean sunfishes: evolution, biology and conservation*. Eds. T. M. Thys, G. C. Hays and J. D. R. Houghton (Boca Raton: CRC Press), 280–296. doi: 10.1201/9780429343360-15
- Hobson, E. S. (1965). A visit with el barbero. *Underw. Nat.* 3, 5–10.
- Hobson, E. S. (1969). Comments on certain recent generalizations regarding cleaning symbiosis in fishes. *Pacific Sci.* 23, 35–39.
- Hobson, E. S. (1971). Cleaning symbiosis among California inshore fishes. *Fish. Bull.* 63, 491–523.
- Howard, M. J., Nakatsubo, T., Correia, J. P., Batista, H., Baylina, N., Taura, C., et al. (2020). "Sunfish on display. Husbandry of the ocean sunfish *Mola Mola*," in *The ocean sunfishes: evolution, biology and conservation*. Eds. T. M. Thys, G. C. Hays and J. R. Houghton (Boca Raton: CRC Press), 242–261. doi: 10.1201/9780429343360-11
- Hulthén, K., Chapman, B. B., Nilsson, P. A., Hollander, J., and Brönmark, C. (2014). Express yourself: bold individuals induce enhanced Morphological defences. *Proc. R. Soc. B Biol. Sci.* 281, 20132703. doi: 10.1098/rspb.2013.2703
- Kodric-Brown, A. (1998). Sexual dichromatism and temporary color changes in the reproduction of fishes. *Amer. Zool.* 38, 70–81. doi: 10.1093/icb/38.1.70
- Kondo, S., and Asai, R. (1995). A reaction-diffusion wave on the skin of the marine angelfish *Pomacanthus*. *Nature* 376, 765–768. doi: 10.1038/376765a0
- Konow, N., Fitzpatrick, R., and Barnett, A. (2006). Adult emperor angelfish (*Pomacanthus imperator*) clean giant sunfishes (*Mola Mola*) at Nusa Lembongan, Indonesia. *Coral Reefs* 25, 208. doi: 10.1007/s00338-006-0086-9
- Kushimoto, T., Kakino, A., and Shimomura, N. (2022). Possible individual identifications by the body surface marking patterns in the ocean sunfish *Mola Mola* and the sharptail sunfish *Masturus lanceolatus* (Molidae). *Ichthyol. Nat. Hist. Fishes Japan* 19, 1–7. doi: 10.34583/ichthy.19.0_1
- Lelong, P. (1999). Individual identification of dusky grouper, *Epinephelus marginatus* (Lowe 1834) by cephalic blotches. *Mar. Life* 9, 29–35.
- Ligon, R. A., and McCartney, K. L. (2016). Biochemical regulation of pigment motility in vertebrate chromatophores: a review of physiological color change mechanisms. *Curr. Zool.* 62, 237–252. doi: 10.1093/cz/zow051
- Liu, K. M., Lee, M. L., Joung, S. J., and Chang, Y. C. (2009). Age and growth estimates of the sharptail *Mola, Masturus lanceolatus*, in waters of eastern Taiwan. *Fish. Res.* 95, 154–160. doi: 10.1016/j.fishres.2008.08.013
- Liu, J., Zapfe, G., Shao, K. T., Leis, J., Matsuura, K., Hardy, G., et al. (2015) *Mola Mola* (errata version published in 2016). *The IUCN red list of threatened species 2015: e.T190422A97667070*. Available at: <https://www.iucnredlist.org/search?query=MolaMola&searchType=species>.
- Marshall, N. J., Cortesi, F., de Busserolles, F., Siebeck, U. E., and Cheney, K. L. (2019). Colours and colour vision in reef fishes: past, present and future research directions. *J. Fish Biol.* 95, 5–38. doi: 10.1111/jfb.13849
- Marshall, A. D., and Pierce, S. J. (2012). The use and abuse of photographic identification in sharks and rays. *J. Fish Biol.* 80, 1361–1379. doi: 10.1111/j.1095-8649.2012.03244.x
- Martin, F. D., and Drewry, G. E. (1978). Development of fishes of the mid-Atlantic bight: an atlas of egg, larval, and juvenile stages, vol VI - stromateidae through ogocephalidae. *Fort Collins*.
- Mayne, B., Espinoza, T., Roberts, D., Butler, G. L., Brooks, S., Korbie, D., et al. (2021). Nonlethal age estimation of three threatened fish species using DNA methylation: Australian lungfish, Murray cod and Mary river cod. *Mol. Ecol. Resour.* 21, 2324–2332. doi: 10.1111/1755-0998.13440
- Miele, V., Dussert, G., Spataro, B., Chamailé-Jammes, S., Allainé, D., and Bonenfant, C. (2020). Revisiting animal photo-identification using deep metric learning and network analysis. *Methods Ecol. Evol.* 12, 863–873. doi: 10.1111/2041-210X.13577
- Mucientes, G., Irisarri, J., and Villegas-Ríos, D. (2019). Interannual fine-scale site fidelity of male ballan wrasse *Labrus bergylta* revealed by photo-identification and tagging. *J. Fish Biol.* 95, 1151–1155. doi: 10.1111/jfb.14111
- Nakatsubo, T., and Hirose, H. (2007). Growth of captive ocean sunfish, *Mola Mola*. *Aquac. Sci.* 55, 403–407. doi: 10.1123/aquaculturesci.1953.55.403
- Navarro, J., Perezgrueso, A., Barria, C., and Coll, M. (2018). Photo-identification as a tool to study small-spotted catshark *Scyliorhinus canicula*. *J. Fish Biol.* 92, 1657–1662. doi: 10.1111/jfb.13609
- Nielsen, J., Hedeholm, R. B., Heinemeier, J., Bushnell, P. G., Christiansen, J. S., Olsen, J., et al. (2016). Eye lens radiocarbon reveals centuries of longevity in the Greenland shark (*Somniosus microcephalus*). *Sci* 353, 702–704. doi: 10.1126/science.aaf1703
- Nolf, D., and Tyler, J. C. (2006). Otolith evidence concerning interrelationships of caproid, zeiform and tetraodontiform fishes. *Bull. l'Institut R. Des. Sci. Nat. Belgique* 76, 147–189.
- Nyegaard, M. (2018) *There be giants! The importance of taxonomic clarity of the large ocean sunfishes (genus Mola, family Molidae) for assessing sunfish vulnerability to anthropogenic pressures*. Available at: <https://researchrepository.murdoch.edu.au/id/eprint/41666/> (Accessed 01 March 2023).
- Nyegaard, M., Andrzejczek, S., Jenner, C. S., and Jenner, M. N. M. (2019). Tiger shark predation on large ocean sunfishes (Family Molidae) – two Australian observations. *Environ. Biol. Fishes* 102, 1559–1567. doi: 10.1007/s10641-019-00926-y
- Nyegaard, M., Garcia-Barcelona, S., Phillips, N. D., and Sawai, E. (2020). "Fisheries interactions, distribution modelling and conservation issues of the ocean sunfishes," in *The ocean sunfishes: evolution, biology and conservation*. Eds. T. M. Thys, G. C. Hays and J. D. R. Houghton (Boca Raton: CRC Press), 216–242. doi: 10.1201/9780429343360
- Nyegaard, M., and Sawai, E. (2018). Species identification of sunfish specimens (Genera *Mola* and *Masturus*, family Molidae) from Australian and New Zealand natural history museum collections and other local sources. *Data Br.* 19, 2404–2415. doi: 10.1016/j.dib.2018.07.015

- Nyegaard, M., Sawai, E., Gemmill, N., Gillum, J., Loneragan, N. R., Yamanoue, Y., et al. (2018). Hiding in broad daylight: molecular and Morphological data reveal a new ocean sunfish species (Tetraodontiformes: Molidae) that has eluded recognition. *Zool. J. Linn. Soc.* 182, 631–658. doi: 10.1093/zoolinnean/zlx040
- Pedersen, M., Nyegaard, M., and Moeslund, T. B. (2023). Finding Nemo's Giant Cousin: Keypoint Matching for Robust Re-Identification of Giant Sunfish. *J. Mar. Sci. Eng.* 11, 889. doi: 10.3390/jmse11050889
- Pedersen, M., Haurum, J. B., Moeslund, T. B., and Nyegaard, M. (2022). Re-identification of giant sunfish using keypoint matching. *Proc. Northern Lights Deep Learn. Workshop 2022* 3. doi: 10.7557/18.6234
- Pedersen, M., and Mohammed, A. (2021). Photo identification of individual *Salmo trutta* based on deep learning. *Appl. Sci.* 11, 9039. doi: 10.3390/app11199039
- Petso, T., Jamisola, R. S., and Mpoeleng, D. (2022). Review on methods used for wildlife species and individual identification. *Eur. J. Wildl. Res.* 68. doi: 10.1007/s10344-021-01549-4
- Pedersen, M., Nyegaard, M., and Moeslund, T. B. (2022). Finding nemo's giant cousin: Keypoint matching for robust re-identification of giant sunfish. *J. Mar. Sci. Eng.* 11, 889. doi: 10.3390/jmse11050889
- Phillips, N. D., Kubicek, L., Payne, N. L., Harrod, C., Eagling, L. E., Carson, C. D., et al. (2018). Isometric growth in the world's largest bony fishes (genus *Mola*)? Morphological insights from fisheries bycatch data. *J. Morphol.* 279, 1312–1320. doi: 10.1002/jmor.20872
- Potts, G. W. (1974). The colouration and its behavioural significance in the corkwing wrasse, *Crenilabrus melops*. *J. Mar. Biol. Assoc. United Kingdom* 54, 925–938. doi: 10.1017/S0025315400057659
- Powell, D. C. (2001). *A fascination for fish: adventures of an underwater pioneer* (Berkeley and Los Angeles: University of California Press).
- Riawan, I. M. O., Setiabudi, G. I., Merdana, I. M., Mariasa, I. P. M., and Wirasastra, K. T. (2019). First molecular identification of sunfish in North Bali water. *Adv. Trop. Biodivers. Environ. Sci.* 3, 12. doi: 10.24843/atbes.2019.v03.i01.p04
- Rosenthal, G. G., and Marshall, N. J. (2011). Communication behavior: visual signals. *Encycl. Fish Physiol.* 1, 692–698. doi: 10.1016/B978-0-12-374553-8.00080-0
- Ruchimat, T., Basuki, R., and Welly, M. (2013). Nusa Penida Marine Protected Area (MPA) Bali - Indonesia: why need to be protected? *Transylvanian Rev. Syst. Ecol. Res.* 15, 193–202. doi: 10.2478/trser-2013-0016
- Sawai, E. (2017). *The mystery of ocean sunfishes [Manbou no himitsu]* (Tokyo: Iwanami Shoten Publishers).
- Sawai, E., and Nyegaard, M. (2022). A review of giants: examining the species identities of the world's heaviest extant bony fishes (ocean sunfishes, family Molidae). *J. Fish Biol.* 100, 1345–1364. doi: 10.1111/jfb.15039
- Sawai, E., Yamanoue, Y., Jawad, L., Al-Mamry, J., and Sakai, Y. (2017). Molecular and Morphological identification of *Mola* sunfish specimens (Actinopterygii: Tetraodontiformes: Molidae) from the Indian Ocean. *Species Divers.* 22, 99–104. doi: 10.12782/sd.22_99
- Sawai, E., Yamanoue, Y., Nyegaard, M., and Sakai, Y. (2018). Redescription of the bump-head sunfish *Mola alexandrini* (Ranzani 1839), senior synonym of *Mola ramsayi* (Giglioli 1883), with designation of a neotype for *Mola Mola* (Linnaeus 1758) (Tetraodontiformes: Molidae). *Ichthyol. Res.* 65, 142–160. doi: 10.1007/s10228-017-0603-6
- Sawai, E., Yamanoue, Y., Yoshita, Y., Sakai, Y., and Hashimoto, H. (2011). Seasonal occurrence patterns of *Mola* sunfishes (*Mola* spp. A and B; *Molidae*) in waters off the Sanriku region, eastern Japan. *Japanese J. Ichthyol.* 58, 181–187. doi: 10.11369/jji.58.181
- Sèbe, M. (2021). The use of photo-identification for moray eel monitoring. *Cah. Biol. Mar.* 62, 11–16. doi: 10.21411/CBM.A.F5F29BE0
- Sköld, H. N., Aspengren, S., Cheney, K. L., and Wallin, M. (2016). Fish chromatophores - from molecular motors to animal behavior. *Int. Rev. Cell Mol. Biol.* 321, 171–219. doi: 10.1016/bs.ircmb.2015.09.005
- SLKT, D., Kamal, M. M., Tarsidin, T., and Yulianto, G. (2021). Sunfish (*Mola* spp.) habitat characteristics on their appearance at dive tourism depths in Nusa Penida waters, Bali. *J. Biol. Trop.* 21, 149–156. doi: 10.29303/jbt.v21i1.2442
- Stuart-Fox, D., and Moussalli, A. (2009). Camouflage, communication and thermoregulation: lessons from colour changing organisms. *Philos. Trans. R. Soc B Biol. Sci.* 364, 463–470. doi: 10.1098/rstb.2008.0254
- Sugimoto, M. (2002). Morphological color changes in fish: regulation of pigment cell density and morphology. *Microsc. Res. Tech.* 58, 496–503. doi: 10.1002/jemt.10168
- Thurman, C. L. (1998). Rhythmic physiological colour change in Crustacea—a review. *Comp. Biochem. Physiol. Part C Comp. Pharmacol.* 91, 171–185. doi: 10.1016/0742-8413(88)90184-3
- Thys, T. M., Hearn, A. R., Weng, K. C., Ryan, J. P., and Peñaherrera-Palma, C. (2017). Satellite tracking and site fidelity of short ocean sunfish, *Mola ramsayi*, in the Galapagos Islands. *J. Mar. Biol.* 2017, Article ID 7097965. doi: 10.1155/2017/7097965
- Thys, T. M., Nyegaard, M., and Kubicek, L. (2020). "Ocean sunfishes and society," in *The ocean sunfishes: evolution, biology and conservation*. Eds. T. M. Thys, G. C. Hays and J. D. R. Houghton (Boca Raton: CRC Press), 263–279. doi: 10.1201/9780429343360-14
- Titus, B. M., Ryan, J. P., Weng, K. C., Erdmann, M., and Tresnati, J. (2016). Tracking a marine ecotourism star: movements of the short ocean sunfish *Mola ramsayi* in Nusa Penida, Bali, Indonesia. *J. Mar. Biol.* 2016, Article ID 8750193. doi: 10.1155/2016/8750193
- Titus, B. M., Daly, M., and Exton, D. A. (2015). Do reef fish habituate to diver presence? Evidence from two reef sites with contrasting historical levels of SCUBA intensity in the Bay Islands, Honduras. *PLoS One* 10, e0119645. doi: 10.1371/journal.pone.0119645
- Turing, A. M. (1952). The chemical basis for morphogenesis. *Phil. Trans. R. Soc Lond. B* 237, 37–72. doi: 10.1098/rstb.1952.0012
- Urian, K., Gorgone, A., Read, A., Balmer, B., Wells, R. S., Berggren, P., et al. (2015). Recommendations for photo-identification methods used in capture-recapture models with cetaceans. *Mar. Mammal Sci.* 31, 298–321. doi: 10.1111/mms.12141
- Visser, I., Nyegaard, M., and Fletcher, L. (2023). Orca, *Orcinus orca* (Linnaeus 1758) (Mammalia cetacea), interactions with ocean sunfishes (Family Molidae, genus *Mola* köleuter 1766 and *Masturus* gill 1884): a global review. *Biodivers. J.* 14, 61–164. doi: 10.31396/Biodiv.Jour.2023.14.1.61.164
- Vogelnest, L. (2003). "The tale of two ocean sunfishes (*Mola Mola*) - an unusual mortality in an unusual species," in *Wildlife Disease Association Australasian Section 2003 Annual Conference*, Healesville, Victoria: Wildlife Disease Association. 30–32.
- Watanabe, Y., and Sato, K. (2008). Functional dorsoventral symmetry in relation to lift-based swimming in the ocean sunfish *Mola Mola*. *PLoS One* 3, e3446. doi: 10.1371/journal.pone.0003446
- White, W. T., Last, P. R., Dharmadi, F. R., Chodrijah, U., Prisantoso, B. I., Pogonoski, J. J., et al. (2013). *Market fishes of Indonesia (jenis-jenis ikan di indonesia)*. ACIAR monograph no. 155 (Canberra: Australian Center for international Agriculture Research).