#### Check for updates

#### OPEN ACCESS

EDITED BY Paris Vasileios Stefanoudis, University of Oxford, United Kingdom

#### REVIEWED BY

Alec Aitken, University of Saskatchewan, Canada Santiago E. A. Pineda Metz, Alfred Wegener Institute Helmholtz Centre for Polar and Marine Research (AWI), Germany

#### \*CORRESPONDENCE

Candace J. Grimes CGrimes1327@gmail.com Kenneth M. Halanych HalanychK@uncw.edu

#### SPECIALTY SECTION

This article was submitted to Deep-Sea Environments and Ecology, a section of the journal Frontiers in Marine Science

RECEIVED 09 November 2022 ACCEPTED 24 March 2023 PUBLISHED 14 April 2023

#### CITATION

Grimes CJ, Donnelly K, Ka C, Noor N, Mahon AR and Halanych KM (2023) Community structure along the Western Antarctic continental shelf and a latitudinal change in epibenthic faunal abundance assessed by photographic surveys. *Front. Mar. Sci.* 10:1094283. doi: 10.3389/fmars.2023.1094283

#### COPYRIGHT

© 2023 Grimes, Donnelly, Ka, Noor, Mahon and Halanych. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Community structure along the Western Antarctic continental shelf and a latitudinal change in epibenthic faunal abundance assessed by photographic surveys

Candace J. Grimes<sup>1\*</sup>, Kyle Donnelly<sup>1</sup>, Cheikhouna Ka<sup>2</sup>, Nusrat Noor<sup>2</sup>, Andrew R. Mahon<sup>3</sup> and Kenneth M. Halanych<sup>1\*</sup>

<sup>1</sup>Center for Marine Science, University of North Carolina Wilmington, Wilmington, NC, United States, <sup>2</sup>Department of Biological Sciences, Auburn University, Auburn, AL, United States, <sup>3</sup>Department of Biology, Central Michigan University, Mount Pleasant, MI, United States

The Southern Ocean's continental shelf communities harbor high benthic biodiversity. However, most census methods have relied on trawling or dredging rather than direct observation. Benthic photographic and videographic transect surveys serve a key role in characterizing marine communities' abundance and diversity, and they also provide information on the spatial arrangement of species within a community. To investigate diversity and abundance in Southern Ocean benthic communities, we employed photographic transects during cruises aboard the RVIB Nathanial B. Palmer (November 2012) and the ASRV Laurence M. Gould (February 2013). One kilometer long photographic transects were conducted at 8 sites along 6,000 km of Western Antarctica from the tip of the Antarctic Peninsula to the Ross Sea from which epifaunal echinoderms, tunicates, arthropods, cnidarians, poriferans, and annelids were identified and counted allowing estimations of biodiversity. Our results do not support a latitudinal trend in diversity, but rather a decrease in abundance of macrofaunal individuals at higher latitude sites. All communities sampled on the Western Antarctic shelf were primarily dominated by ophiuroids, pycnogonids, holothuroids, and demosponges. However, the most abundant taxon across all sites was Ophionotus victoriae, followed by the symbiotic partners lophon sp. (demosponge) and Ophioplinthus spp. (ophiuroid). Data also confirm that the Southern Ocean is composed of discretely unique benthic communities. These results provide critical understanding of the current community structure and diversity serving as a baseline as the Antarctic continental shelf changes due to rising ocean temperatures, climate change, and collapse of large ice sheets.

#### KEYWORDS

Antarctica, benthic biodiversity, photographic surveys, invertebrates, community structure

# **1** Introduction

Waters surrounding the Western Antarctic region house great faunal abundance, richness, and diversity. Although over 8,000 invertebrate species have been described from the Southern Ocean, an estimated 17,000-20,000 invertebrate species may exist there (Gutt et al., 2004) with the highest diversity reported in the Western Antarctic (Gutt and Starmans, 1998; Grange and Smith, 2013; Pineda-Metz et al., 2019). A similar situation exists on Arctic continental shelves, where species diversity is under described in large part due to inaccessibility caused by ice (Piepenburg et al., 2011). Although the Antarctic marine continental shelf biome is sometimes considered one ecoregion, there is considerable variation in the physical environment which can affect organismal distribution and abundance (Convey and Peck, 2019), ultimately impacting community structure. Through photo/video transects, bottom trawls, and box cores, previous research on the Antarctic shelf has revealed great diversity (Gutt et al., 2013) and abundance of epifaunal invertebrates (e.g. Dayton et al., 1974; Grebmeier and Barry, 1991; Gerdes et al., 1992; Gutt and Starmans, 1998; Sumida et al., 2008; Grange and Smith, 2013; Post et al., 2017; Pineda-Metz et al., 2019; Cummings et al., 2021; Zwerschke et al., 2022). These studies describe high numbers of ophiuroids, holothuroids, sponges, cnidarians, and annelids. Similar to destructive techniques, non-destructive imaging technologies (or SCUBA at shallow depths) can accurately characterize spatial relationships and communities of epifaunal species (Pineda-Metz and Gerdes, 2018; Cummings et al., 2021). Previous work in continental shelf regions of the Southern Ocean has revealed more sea cucumbers, urchins, and anthozoans near the Western Antarctic Peninsula and higher numbers of brittle stars and sponges in the Ross Sea (Barry et al., 2003; Sumida et al., 2008). Unlike the Arctic Ocean, where a latitudinal diversity hypothesis is supported, the Southern Ocean has thus far shown little support for a latitudinal decrease in diversity (Gray, 2001; Clarke et al., 2007; Aronson et al., 2009; Griffiths, 2010).

Around the continent, seawater temperature, salinity, and pH remain relatively constant. Antarctic waters are some of the most thermally stable environments on the planet varying on average roughly 1 °C (Clarke, 1988; Arntz et al., 1994; Pineda-Metz et al., 2020). As a result, many organisms in the region are stenothermic, being only able to live in a narrow temperature range (Clarke, 1988; Arntz et al., 1994; Aronson et al., 2009). Close to the edge of the continental shelf however, there are sources of environmental variation (Hellmer et al., 2012; Isla and Gerdes, 2019), such as regions where relatively warm water from the circumpolar deep water (CDW) currents are pushed onto the shelf (e.g., in the Bellingshausen Sea; see Ruan et al., 2021) resulting in different environmental zones (e.g., increased temperature, decreased oxygen) that might harbor distinct benthic communities. Variations in physical ocean characteristics offer insight to faunal diversity, distribution, and abundance in the Western Antarctic (Halanych and Mahon, 2018). Along with the direct influence of CDW influencing physical water parameters, CDW intrusion is currently understood to be an important driver in accelerated melting rates of sea ice (Jacobs and Comiso, 1997; Stern et al.,

2013). In the spring time, CDW intrusion effects coupled with higher temperatures cause an annual melting of sea ice (De Jong et al., 2015). Upon sea ice melt in the spring and summer months, iron is released into the water column leading to an annual influx which is an important trigger for phytoplankton blooms that increases primary production, likely influencing the local benthic community (Jacobs and Comiso, 1997; De Jong et al., 2015). Due to the complex oceanography of the region however, mechanisms that cause variation in CDW intrusions annually and inter-annually are poorly understood (Morrison et al., 2020). In the Ross Sea, strong winds in the spring time can create a large area devoid of sea ice, also known as a polynya, which promotes phytoplankton growth (Smith and Gordon, 1997). Predictions of ocean warming driven by climate change threaten the thermal stability of this region, which may lead to detrimental effects on abundance and fitness of regional benthic fauna (Clarke et al., 2007). The warming and freshening of Southern Ocean waters may stress organisms both thermally and nutritionally. Warmer waters can directly affect organismal homeostasis, but these warmer waters also have the potential to alter the quantity and quality of detrital nutrients reaching the seafloor (Wu et al., 2021; Fernández-González et al., 2022). These potential effects may stress organisms and drastically impact the local diversity and abundance of organisms (Pineda-Metz et al., 2020). In addition, phytoplankton blooms stimulated by upwelling, ice-melt, or iron influx from warm water intrusions have been described in Western Antarctica and result in decreased oxygen levels as the phytoplankton decompose (Deppeler and Davidson, 2017; Tripathy and Jena, 2019). In many cases, these environmental factors are influenced by global climate change (Convey and Peck, 2019), further necessitating study and characterization of benthic communities as they experience such variation.

In this study, our goal was to describe epifaunal communities across 6,000km of the Western Antarctic continental shelf, beginning near the tip of the peninsula continuing west, and correlate potential environmental drivers to benthic macrofauna dominance through the use of benthic photographic transects (Figure 1). We expected to see a higher diversity and abundance of epifauna in sites with harder substrates (pebbles and cobbles) and phytodetritus because they provide a stable point of attachment and sustenance, respectively (Post et al., 2017; Gutt et al., 2019). Additionally, we hypothesized a greater relative abundance of brittle stars and sea cucumbers on softer sediments due to their feeding and burrowing behaviors. Understanding environmental influences on communities gives the scientific community valuable insight into how the Western Antarctic (WA) benthos, and the Antarctic ecosystem as a whole, could react to global climate change. Moreover, studies such as this help fill research gaps in global marine communities so latitudinal trends can be better understood.

# 2 Materials and methods

During cruise NBP 12-10 aboard the *RVIB Nathanial B. Palmer* (NBP) and cruise LMG 13-12 on the *ASRV Laurence M. Gould* (LMG) along Western Antarctica between 2012-2013, photographic



FIGURE 1

Examples of organismal diversity from each of the sites: (A) Montravel Rock, (B) Recovitza Island, (C) Seymour Island, (D) Bellingshausen, (E) Wright's Gulf, (F) Eastern Amundsen, (G) Near Ross Sea, (H) Mid Ross Sea. Macrofaunal identifications are: aa = Astrotoma agassizii, ab2 = Abatus sp. 2, bs = Bathyplotes sp. 1, cn19 = cnidarian 19, likely Umbellula sp., cn31 = cnidarian 31, likely Primnoella sp., echinoid 2 = echinoid 2, ho2 = holothurian 2, oi = Ophioplinthus spp. and lophon sp. symbiosis, op2 = ophiuroid 2, op13 = ophiuroid 13, op14 = ophiuroid 14, ov = Ophionotus victoriae, pk = Promachocrinus kerguluensis, po6 = porifera 6, po13 = porifera 13, py2 = pychnogonid 2, and tu8 = tunicate 8.

transects were conducted via a downward facing camera setup known as the 'Yoyo' camera (Smith et al., 2012; Grange and Smith, 2013). The Yoyo camera consists of a steel frame supporting a digital camera (10.2 megapixel, 20mm, Nikon D-80 Camera) and two parallel red lasers 10cm apart (Figure 2A). The unit is fitted with a transmissometer and audible contact that triggers a switch and takes the photo of the seafloor from ~2.5m above (Grange and Smith, 2013) imaging an approximately  $3m^2$  area. The camera unit is repeatedly raised and lowered, hence the yo-yo name, to take multiple images across a transect as the ship moves at approximately 1 knot. Photos were color corrected and white balanced, as well as cropped to the central 135cm x 135cm section, which included the two red laser dots (a scale-bar representing 10 cm on the benthos), to reduce edge effects. Flat and relatively drop stone-free sites were selected based on overall topography to facilitate subsequent organismal collections by Blake Trawl (speed = 1 knot with variable tow durations dependent on tension) to support taxonomic identifications. As a result, sites tended to lack relief and had soft sand and/or pebble bottoms. In this study, photographic data were collected for 1 km long transects between 301-731m depth at 8 sites from the tip of the Western Peninsula to the Ross Sea (Figure 1B). Site position and distance from the mainland were collected via Geographic Positioning System (GPS) to include latitude and longitude as potential factors for analysis.

For each transect, benthic environmental data was collected via a Sea-Bird CTD system, providing water column information from the sea surface to the seafloor once per site (Data available at https:// www.marine-geo.org: NBP 12-10 doi 10.7284/901177 and LMG 13-12 doi 10.7284/901981). We included maximum fluorescence from the CTD cast in the euphotic zone as a proxy for primary productivity. Sea ice cover and lack of fine-scale satellite coverage across the transect prevented obtaining long-term productivity estimates via satellite imagery. Maximum sea ice coverage for each site was gathered from the National Snow and Ice Data Center (NSIDC, 2023). For each cropped and color-corrected photo, the substrate was classified as sandy, sandy with pebbles (stones < 5cm), or sandy with pebbles and cobble (stones 5-50cm) and examined as a parameter to determine community differences between substrate types. If pebbles and cobbles covered more than 5cm<sup>2</sup>, then the photo was no longer classified as sandy, instead it was classified as 'sand and pebbles' or 'sand, pebbles, and cobbles', respectively. In addition to the substrate characterization, we also scored whether phytodetritus on the seafloor was present (i.e., visibly covered more than 5 cm<sup>2</sup>: Figures 1E, F). Highly collinear variables ( $|\mathbf{r}| \ge 0.80$ : distance from mainland) were removed from analysis, and trends in abiotic data collected were elucidated via principal component analysis (PCA).

During the 1 km transect at each site, 101-323 photos were taken roughly every 5-10 m. Subsequently, photos were filtered to remove



FIGURE 2



images with substantial turbidity, overlap between photos or poor photo quality. Of the remaining images, 50 photos were randomly selected and analyzed per site to represent the 1 km-long transects following the protocol of Grange and Smith (2013) to allow compatibility. Images were analyzed to identify organismal morphotypes and all morphotypes were assigned to the lowest taxonomic classification possible. For purposes of our analyses, species were also binned into higher taxonomic categories including Annelida, Ascidiacea, Malacostraca, Pycnogonida, Anthozoa, Staurozoa, Asteroidea, Crinoidea, Echinoidea, Holothuroidea, Ophiuroidea, Brachiopoda, Cephalopoda, Gastropoda, Demospongiae, and Hexactinellida. Benthic epifaunal invertebrates were assigned to the "Miscellaneous" group if identification was not possible (e.g., due to viewing angle or obstruction). Colonial fauna were included as individuals if substrate was visible between the zooids. Statistical analyses were run both with and without the miscellaneous group to account for all organisms distinguishable on the bottom, but our primary outcomes were not significantly impacted by the presence of the miscellaneous category (Supplementary Data). Species richness (Margalef, 1958), Pielou's evenness (Pielou, 1966), and diversity indices (Shannon, 1948; Simpson, 1949) were calculated and compared. Infauna, meiofauna, tube-dwelling annelids (such as chaetopterids) and other taxa (e.g., Bryozoa and Hydrozoa) were not included in the study due to the difficult nature of identifying species/morphotypes based on surface evidence and photo resolution.

Rare taxa that were found at only one or two sites were removed from community analysis. Abundance data for remaining taxa in each photo were 4th-root transformed to reduce the impact of species dominance by a few select species. Then, faunal assemblages were visualized with a distance-based

redundancy analysis (dbRDA) based on calculations of a zeroadjusted Bray-Curtis similarity matrix in PRIMER version 7 (Bray and Curtis, 1957; Clarke et al., 2014). We performed a cluster analysis to estimate relatedness of communities and to group sites together based on community compositions and utilized a similarity profile (SIMPROF) to find statistically significant groupings. Then we completed a similarity percentage (SIMPER) analysis to identify taxa responsible for the similarities and dissimilarities between sites and regions. Environmental variables were visualized with a principal component analysis (PCA) then superimposed on the dbRDA to illustrate abiotic influences on community composition. To elucidate abiotic-biotic relationships, we conducted permutational analysis of variance (PERMANOVA) and subsequent PERMDISP to characterize dispersion. We used distance-based linear models with DistLM protocol to determine the environmental factors primarily responsible for community differences based on Akaike information criterion (AIC).

# **3** Results

A total of 400 photos from 8 transects were included in this study, resulting in a total count of 6,947 organisms belonging to 204 morphotypes (or "taxa") in the 16 taxonomic groups examined (Tables S1, S2). Benthic communities in the Peninsula region had higher organismal density than other regions (Table 1). Images across all 8 sites varied from having just 1 organism present to 89 individuals of multiple species and tended to have a sandy bottom (Figure 1B) as opposed to a sandy bottom containing pebbles (Figure 1D).

Site	Abundance (n/m <sup>2</sup> )	# of Species	Richness	Evenness	Shannon	Simpson	
Amundsen							
Bellingshausen	3.13	5.74	1.64	0.93	1.02	0.83	
Eastern Amundsen	8.99	16.46	2.60	0.90	1.80	0.85	
Wright's Gulf	4.92	12.08	2.35	0.93	1.67	0.89	
Peninsula							
Montravel Rock	28.14	50.5	2.40	0.80	1.83	0.80	
Seymour Island	14.35	26.16	2.17	0.75	1.49	0.69	
Ross and Recovitza							
Mid Ross Sea	6.68	12.88	1.98	0.89	1.51	0.80	
Near Ross Shelf	2.56	4.68	1.70	0.95	1.11	0.87	
Recovitza Island	5.77	10.52	1.43	0.82	1.14	0.66	

TABLE 1 Mean values for abundance, number of species, species richness, Pielou's Evenness, Shannon-Wiener Diversity Index, and Simpson Index for all sites.

### 3.1 Community assemblage characteristics

Dominant organisms in each site varied between ophiuroids, holothuroids, demosponges, and pycnogonids (Figure 2B and Table 2). Anthozoans, ophiuroids, and ascidians were ubiquitous at all sites, but ophiuroids dominated the Montravel Rock, Seymour Island, and the Bellingshausen Sea sites. Based on the cluster and SIMPROF analysis of benthic communities, we grouped the sites into three regions: the Peninsula region, the Amundsen region, and the Ross Sea region (Figure S1). There were 28 photos (16 Amundsen, 11 Ross Sea, and 1 Peninsula) that did not cluster within the same regions as the rest of the photos from their sites providing evidence of within group dissimilarity (Figure S2). There were 16 photos from the Amundsen in a deeply branching cluster with 15 photos from the Ross Sea indicating the lack of similarity within this particular group indicating patchiness along the transect. These Amundsen region photos were characterized by reduced numbers of cnidarians, ophiuroids, and echinoids compared to other photos of the region. The 11 Ross Sea photos that clustered within the Amundsen Region had less annelids, holothuroids, and pycnogonids than the rest of the photos in the Ross Sea and Recovitza Island region. Generally, sites cluster by relatives location according to epifaunal community assemblages (Figures 3A, S2): the Amundsen region includes the Eastern Amundsen, Bellingshausen, and Wright's Gulf sites, the Ross region includes the Mid Ross, Near Ross Shelf, and Recovitza Island sites, and the Peninsula region includes the Seymour Island and Montravel Rock sites (Figure 3A). Pycnogonid sp. 1, pycnogonid sp. 2, Ophionotus victoriae, Sterechinus sp. 1, Ophioplinthus spp., Iophon sp., and the yellow branching sponge (Figures 1A, C) were the primary species that helped differentiate the three regions we define here (Figures S3A, B). Based on the cluster and SIMPROF analysis (Figure S2), Recovitza Island communities were more similar to the Ross Region sites and have been included with them in the remainder of the analysis although we note some degree of within group dissimilarity (Figure S1). We saw a higher prevalence of echinoids (e.g., Sterechinus spp.) and holothuroids (e.g., Rhipidothuria racowitzai and sediment dwelling holothuroid 2) in the Amundsen Region than the other two regions. The Iophon sp. (demosponge), and Ophioplinthus spp. (ophiuroid) symbiont pair dominated the Peninsula Region and was significantly less abundant in all other regions (Table 2). The brittle star Ophionotus victoriae is abundant throughout the Southern Ocean, but it typically dominated softer sediments of the Peninsula region and Ross Sea region. The ophiuroid morphotype of Ophiuroid 4, characterized by small spiny pinkred arms with a darker central disc) was common in the Ross Sea region and is possibly Amphiura sp. We also saw an increase in Hexactinellida (glass sponges) in the Ross region compared to other regions (Figure 2B).

Our three regions had significant heterogenous dispersions with potentially significant differences between communities (Table 3). Our study regions illustrate distinct community compositions (Figure 2B) and a latitudinal trend in abundance (again with exception of Recovitza Island; Figures 4B, S4). Northernmost sites portrayed higher relative abundances of demosponges and ascidians on average (12.5% and 19%, respectively) than southernmost sites (4% and 3%, respectively) which contained higher percentages echinoids and holothuroids (9% and 18%, respectively). Macrofaunal abundances in the Ross region and Recovitza Island were significantly lower than that of the northernmost Peninsula region sites. Based on a SIMPER analysis between sites (Table 2), epifaunal communities in the Ross Sea and near Recovitza Island significantly differed from other communities near the peninsula as they were dominated by pycnogonids (~65%: pycnogonid 1 and pycnogonid 2 - possibly Nymphon spp.), malacostracan crustaceans (~5%: Mysid sp. 1), and anthozoan cnidarians (3%: possibly Hormosoma sp.). These taxa and those listed in Table 2 for each region are the crucial taxa that may be used to discriminate between

	Av. Abund	Av. Sim	Sim SD	Contrib%			
Peninsula region (33.41%)							
Ophionotus victoriae	11.58	17.76	1.24	53.16			
Ophiuroidea sp. 20	5.83	4.88	0.60	14.59			
Iophon sp. 1	5.83	4.88	0.60	14.59			
Porifera sp. 13 (yellow branching sponge)	2.87	1.22	0.31	3.66			
Tunicate sp. 9 (Synoicum sp)?	1.22	0.79	0.43	2.36			
Amundsen region (11.24%)							
Ophiuroidea sp. 4	1.39	4.14	0.48	36.83			
Elpidiidae sp. 1	0.79	1.45	0.33	12.87			
Ophiuroidea sp. 2	0.39	0.82	0.20	7.32			
Porifera sp. 4	0.89	0.69	0.24	6.17			
Ophiuroidea sp. 7	0.35	0.52	0.19	4.60			
Ross and Recovitza Island regio (17.06%)							
Pycnogonida sp. 1	2.37	10.15	0.75	59.46			
Ophionotus victoriae	1.63	2.88	0.36	16.90			
Pycnogonida sp. 2	0.43	0.95	0.26	5.57			
Mysid sp. 1	0.25	1.04	0.21	5.04			
Cnidarian sp. 26 (Hormosoma sp.)?	0.37	0.50	0.17	2.93			

TABLE 2 Dominant 5 macrofaunal invertebrates based on SIMPER analysis for each region with average abundance (Av. Abund), average similarity within site (Av. Sim) including standard deviation (Sim SD), and percent contribution to the community (Contrib%).

The percentage after the site name is the total percent similarity within the site.

the Ross (and Recovitza) region, the Peninsula region, and the Amundsen region (Table S3).

The Peninsula region portrayed lower evenness and Simpson diversity values than the Amundsen region as a result of dominance

by the yellow rope sponge, brittle stars (*Ophioplinthus* spp.), and their symbiotic sponge (*Iophon* sp. [Table 2: Figure 1A]). The Amundsen Region typically had higher Simpson diversity values than the other regions, but there was no difference in this diversity



#### FIGURE 3

(A) Distance-based redundancy analysis (dbRDA) plot based on the calculations of a zero-adjusted Bray-Curtis similarity matrix showing community composition relationships where proximity estimates community similarity with Principal Component Analysis (PCA) aligned and overlain on top to show environmental trends. Asterisk indicates the one image with a drop stone found in Seymour Island. (B) PCA of the environmental variables.

	Comparison	t	p (perm)
Substrate Type	Sand and Pebbles vs Sand, Pebbles, and 2.38 Cobbles		< 0.01
	Sand and Pebbles vs Sand	4.99	< 0.01*
	Sand, Pebbles, and Cobbles vs Sand	3.24	< 0.01
Phytodetritus	Present vs Absent	6.16	< 0.01
	Amundsen vs Ross	6.77	< 0.01*
Region	Amundsen vs Peninsula	8.32	< 0.01*
	Ross vs Peninsula	8.52	< 0.01*

TABLE 3 PERMANOVA results (no. permutations: 9,999) to compare benthic communities revealing significant differences between substrate type and phytodetritus denoted by bold p values.

\* indicates heterogenous dispersions between comparison groups which might explain the differences shown between region. T-values are the result of the pairwise comparisons.

index between the regions. Although the Peninsula region was dominated by a few species (Table 2: *Ophioplinthus* spp., *Iophon* sp., and *Ophionotus victoriae*), it contained the highest species richness and highest Shannon Diversity values, closely followed by the Amundsen region (Figures 4A, C). No significant latitudinal trends can be described here due to the inclusion of the Recovitza Island Site with the Ross region, but there is a discernable decreasing trend when looking at the sites individually (Figure S4). However, species richness, abundance, and Shannon diversity tend to decrease progressing southward (Peninsula region > Amundsen region > Ross Sea region: Figures 4B–D).

## 3.2 Environmental characteristics

Based on abiotic data (Table 4), sites generally group together based on location along our transect, similar to the assemblages

mentioned above. The PCA analysis indicates the strongest driver of differences in PC1 which accounted for 46.8% of the variation (Table S5) was DO content of bottom waters which was negatively correlated with temperature and maximum fluorescence, our proxy for chlorophyll (Table S6). Percent sea ice cover, water depth, latitude, and salinity were the most influential factors for PC2 which accounted for 22.6% of variation in the environmental characteristics (Figure 3B). Phytodetritus presence and pebble substrate were associated with higher chlorophyll, temperatures, and lower oxygen levels near the benthos in the Amundsen region.

Amundsen region sediments included pebbles and cobble stones whereas other regions were primarily comprised of finer sandy sediments, and Amundsen region benthos was also characterized by the presence of algal detritus. One drop stone (>50cm) was encountered at the Seymour Island Site (denoted with an asterisk in Figure 2A) and was classified as containing sand, pebble, and cobble. Salinity and temperature were positively



I otal number of species (A), Abundance (B), Shannon Diversity Index (C), and Simpson Diversity Index (D) for each region. \* indicates significan difference from at least 1 other region based with homogeneous dispersions (Table S4).

Site	Water Depth (m)	Oxygen (ml/l)	Temperature (°C)	Salinity (PSU)	Maximum Fluores- cence (mg/ml <sup>3</sup> )	Maximum Sea Ice Cover (%)	рН
			Amunds	en			
Bellingshausen	420	4.1	1.3	34.8	45	15	7.4
Eastern Amundsen	731	4.4	1.1	34.6	18.7	40	7.4
Wright's Gulf	302	4.3	0.9	34.6	29	30	7.2
Peninsula							
Montravel Rock	550	7	-1.5	34.0	0.8	15	7.1
Seymour Island	301	6.9	-1.8	34.6	0.9	80	7.1
Ross and Recovitza							
Mid Ross Sea	511	6.4	-1.6	34.2	21	15	7.3
Near Ross Shelf	606	6.3	-1.7	34.4	10	15	7.5
Recovitza Island	630	6.2	-1	34.4	0.6	15	7.2

#### TABLE 4 Environmental characteristics for each site in each region.

correlated and displayed an inverse relationship with oxygen. For the localities we sampled, Amundsen region sites were also the warmest (average = 1.13 °CC), but there was no difference in temperature between Peninsula region and Ross Sea region sites. Peninsula region sites had the highest DO of all sites (average = 6.95 ml l<sup>-1</sup>), while Amundsen region sites had the lowest (average = 4.26 ml l<sup>-1</sup>).

# 3.3 Community assemblage relationship to environmental characteristics

Macrofaunal communities were significantly different based on the substrate type and presence of phytodetritus (Table 3 and Figures S4A, B). Sites with phytodetritus, cobbles, and pebbles in the Amundsen region were characterized by a higher abundance of echinoids and holothuroids while the sandier sediments without phytodetritus of the Peninsula region contained more ophiuroids and sponges. Sea spiders were the most abundant group on the finer sediments of the Recovitza Island site. PCA vectors illustrate a significant correlation (Spearman Rank: rho = 0.443, p = 0.001; overlain in Figure 3A) between environmental parameters and epifaunal communities. The Amundsen region and Peninsula region are primarily differentiated based on PC1. Therefore, maximum fluorescence, DO, and temperature emerge as important factors driving community distinctness along the Western Antarctic coast (Table S6). In contrast, the Ross region and Recovitza Island were differentiated from the Peninsula and Amundsen regions based on PC2 whose core factors were pH, water depth, and percent sea ice cover. Distance based linear models show the importance DO, depth, temperature, and sea ice cover in predicting community compositions on the western Antarctic shelf at these sites (Tables S7).

# 4 Discussion

A trend toward decreasing abundances of macrofauna approaching the South Pole was observed (Figure S5), however, a significant latitudinal trend, due to the Recovitza Island outlier, was lacking. Further, Antarctic communities in close proximity are distinct in nature (Montravel Rock vs Recovitza Island). Faunal abundances from the Western Antarctic shelf presented here  $(2.44 - 28.14 \text{ individuals/m}^2)$  are similar to previous studies in the Western Antarctic fjords, and other Antarctic shelf sites  $(2.6 - 46 \text{ individuals/m}^2)$ , but higher than Western Antarctic shelf community estimates (0.38 - 0.5 individuals/m<sup>2</sup>; (Barry et al., 2003; Sumida et al., 2008; Grange and Smith, 2013). Cummings et al., (2021) reports evidence of comparable megafaunal abundances on the Ross Sea continental shelf through video transects (0.448 - 3.0 individuals/m<sup>2</sup>), however, they did not find pycnogonids whereas we found them abundant in this region. All 8 sites had ophiuroids (e.g. Ophionotous victoriae and Ophioplinthus spp.) located at the base of anthozoan cnidarians which supports hypotheses of their shade-seeking behavior (Hendler et al., 1995; Johnsen and Kier, 1999). Similar ophiuroid-anemone associations have been described in the Arctic near faunal islands (Syvitski et al., 1989). Although many O. victoriae were observed in the open, this shade-seeking behavior could bias diversity and abundance estimates as not all of those present in the area may be recorded. Throughout these Western Antarctic sites, crinoids (mainly Promachocrinus kerguelensis), and the filter-feeding ophiuroid Astrotoma agassizii were typically seen atop cobble, sponges, cnidarians, and tunicates (Figure 5B) to collect food from water column (Meyer, 1983), but not always (Figure 1 at Recovitza Island).



#### FIGURE 5

Example of sampling sites (A) Montravel Rock with yellow rope sponge, (B) Mid Ross Sea site, (C) Eastern Amundsen with abundant holothuroids, and (D) the drop stone found at the Seymour Island site.

## 4.1 Epifaunal communities along the Western Antarctic coast

Echinoderms are one of the most dominant fauna of the Antarctic continental shelf (Dayton et al., 1974; Clarke and Johnston, 2003; Chiantore et al., 2006; Moles et al., 2015). However, cnidarians, sponges, and pycnogonids were abundant enough to influence assemblages along the Western Antarctic Peninsula sites sampled for this investigation. Similar to data presented here from the Peninsula region (Figure 5A), a recent study in the Antarctic peninsula has shown high abundances of ophiuroids, ascidians, and demosponges (Gutt et al., 2019). Within the Bransfield Straight, Gutt et al. (2019) found dominance of varying phyla (e.g. Echinodermata, Hydrozoa, and Bryozoa) which further shows the patchy nature of this region on a relatively small spatial scale (100km). Previous studies on the continental shelf in the Ross Sea denoted high abundances of suspension feeding bryozoans, anthozoans, crinoids, demosponges, and ascidians (Cummings et al., 2021), which is similar to our results except that we also recorded an abundance of pycnogonids which were abundant in the Ross region and dominated Recovitza Island. Further, comparing our Amundsen sites to previous studies is difficult due to lack of data, but the abundance of holothuroids and echinoids at these sites is notable (Figure 5C).

Although we did not see a latitudinal trend in overall diversity across the Western Antarctic benthic communities, we did see a shift in these communities from dominance by a single taxon to a more diverse representation. Previously, higher Shannon diversity was observed on the open continental shelf compared with fjord sites (Grange and Smith, 2013). Communities at similar depths showed higher diversity and evenness on the shelf, but higher species number and abundance in the fjords. Given the proximity of these fjord and shelf sites and the lack of environmental data included, drawing comparisons between these studies is difficult. However, based on our results, predicted changes with climate warming on salinity and temperature in fjords will drastically impact the communities present there (Grange and Smith, 2013).

## 4.2 Distinct environments driving community composition

Our results indicate Antarctic benthic community variability resulting from abiotic differences on a local scale. Between the Amundsen region and other two regions, differences in maximum fluorescence (>2.3 mg/ml<sup>3</sup>), temperature (> 1.9  $^{\circ}$ C) and DO (> 1.8 ml/l), compared to salinity (0 PSU at minimum) lead us to emphasize the maximum fluorescence, temperature and DO

impacts. The relationships between temperature, salinity, depth, pressure, and DO in marine environments make disentangling singular effects of one factor on a community difficult. Amundsen region sites with higher maximum fluorescence, lower DO, and higher temperature than other regions might have had more echinoids and holothuroids than other sites due to less competition in these zones from organisms that require more oxygen as a result of a higher metabolic rate, such as ophiuroids and crinoids (Woods et al., 2009; Hughes et al., 2011). These environmental characteristics are influenced by decomposition of algal particulates on the seafloor (Iken et al., 2010; Menezes et al., 2017) likely resulting in the community differences observed. These three factors in the Amundsen Sea are typically heavily impacted by algal blooms which was likely occurring during the data collection for this project (Summer 2013). Further, the abundance of holothuroids was higher in the Amundsen region than the Ross or Peninsula region where the maximum fluorescence was highest, potentially indicating a shallower calcium carbonate saturation horizon in this region (Griffiths, 2010). Phytodetritus, higher chlorophyll concentrations in the water column, and a shallower saturation horizon in the Amundsen Sea and Wright's Gulf would help explain the greater abundance of filter-feeding holothuroids over those organisms more reliant on calcium carbonate exoskeletons, such as ophiuroids or crinoids (Griffiths, 2010). Another important factor in distinguishing communities presented here is the maximum sea ice coverage as it facilitated the differentiation between Peninsula region and Ross Sea regions. Within the Peninsula region, sea ice coverage was increasingly variable as the Montravel Rock site had less than 60% of the sea ice cover than the Seymour Island site, however, these communities present with high abundances of ophiuroids and other filter-feeders (e.g. demosponges, crinoids, and ascidians) as previously described (Sumida et al., 2008; Gutt et al., 2019). Finally, the sea ice coverage for the summer in the Southern Ocean has reached a new historic low (Charctic, 2023) which will likely impact benthic communities.

Because the evidence for algae (maximum fluorescence and phytodetritus absence) was lacking, Recovitza Island's significantly different community from the Peninsula region sites could be a result of the slight increase in temperature (0.45-0.8 °C) or decreased oxygen (0.6-0.7 ml l<sup>-1</sup>). Although this site lies within the Bransfield Strait, it is located at the convergence of the transitional waters from the Bellingshausen Sea and water currents from the Weddell Sea (Smith et al., 2012; Deppeler and Davidson, 2017; Tripathy and Jena, 2019). Complexity of the Bransfield water currents leads to environmental fluctuations likely resulting in the community differences presented in our study (e.g., dominance of pycnogonids at Recovitza Island and ophiuroids at Montravel Rock). This distinction in communities on either side of the Bransfield Strait is supported by the clear variation between communities on the northern (mixed assemblage) compared to the southern (more suspension feeders) side of the Bransfield Strait (Gutt et al., 2013). These hydrographic features aid in forming ecoregions in the Southern Ocean which result in these trends in community composition and diversity as shown in the present study (Deppeler and Davidson, 2017). Given impacts of these environmental fluctuations and water currents on the benthic community fluctuations at the local scale, Western Antarctica should be a priority for future climate change studies, especially the Amundsen Region which is the least studied region of all the Southern Ocean (Griffiths et al., 2011).

Although most sites presented in this study were sandy or muddy sediments by design, those that had pebble and cobble substrates in the Amundsen and Peninsula regions, portrayed comparable abundances and Shannon diversity values (Table 1) to those found in other sections of the shelf (Grange and Smith, 2013), the Weddell Sea (e.g. Gutt and Starmans, 1998; Pineda-Metz et al., 2019), and Eastern Antarctica (Post et al., 2017). However, the diversity of benthic communities in the southeastern Weddell Sea were higher than the western Weddell Sea and most of the regions presented in this study (Gutt and Starmans, 1998). Further, in the southeastern Weddell Sea, a greater abundance of holothuroids, bryozoans, and ascidians were found on substrates with less pebbles and a greater abundance of sponges, anthozoans, and other taxa found on finer sandy substrates (Gutt and Starmans, 1998), similar to our Amundsen region and Peninsula region sites, respectively. An increase in arthropods (pycnogonids and others) at the finer sediment sites (Ross Sea and Recovitza region) was found in the Weddell Sea (Gutt and Starmans, 1998) and the Sabrina Coast in Eastern Antarctica (Post et al., 2017). In the Amundsen region and along the Sabrina Coast, on pebble and cobble substrates, there was an increase in brachiopods. Additionally, drop stones and iceberg scour which affect substrate were not accounted for in this study but there was notable abundance and diversity of macrofauna on our one encountered drop stone at Seymour Island (Figure 5D). Drop stones can act as faunal islands in Antarctic and Arctic waters and facilitate dispersal of benthic fauna by serving as steppingstones (Boucot and Carney, 1981; Dale et al., 1989), and in the Arctic, they have been shown to house communities dominated by sponges, ascidians, bryozoans, and crinoids which were found on our one drop stone (Syvitski et al., 1989).

High levels of endemism (Hempel, 1985; Griffiths, 2010; Gutt et al., 2013; Miranda et al., 2021) in the western shelf suggest research is needed to understand how diversity, community composition, and faunal richness will change with climate. Given the higher richness, abundance, and diversity of Peninsula region communities compared to the rest of Western Antarctica, further studies should be employed to better understand changes along the continental shelf and assess regions where benthic community diversity may be of interest (e.g., highly stable or highly variable regions). Diversity trends observed here are similar to those seen in meiofaunal reports from Western Antarctica (Brannock et al., 2018) which show lower richness and diversity in the center of the Bransfield Straight and higher in the Amundsen region. Curiously though, the Eastern Peninsula sites (near our Seymour Island) presented the lowest richness, abundance, and diversity for meiofaunal communities (Brannock et al., 2018) which we believe supports our assertion for localized habitats formed by water currents as seen in the Bransfield Straight. Further microbial diversity was shown to be higher in further west sites (e.g., Bellingshausen Sea and Amundsen Sea) compared to Antarctic Peninsula sites (Learman et al., 2016), however, this might be an artifact of grouping all peninsula sites together.

With predicted decreased sea ice, increased temperature, decreased DO, and algal blooms, polar regions with a dearth of data such as the Amundsen region provide target areas for future research. Studies such as this provide necessary information on benthic communities to aid in characterizing the Southern Ocean's diverse ecoregions in this remote and isolated habitat without the need for potentially harmful trawling (Konecki and Targett, 1989). The Southern Ocean is a large and diverse habitat with varying benthic communities along the continental shelf. In order to identify economically and ecologically important ecoregions, we must continue to gather more data on biodiversity and spatial relationships in this area.

## Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/ Supplementary Material.

## Author contributions

KH and AM collected the data. CJG, AM, and KH designed the study. Photo processing and organismal identifications were conducted by CG, KD, CK, and NN. CG, KD, and KH wrote the manuscript. All authors contributed to the article and approved the submitted version.

## Funding

Funding for this project was provided by the National Science Foundation's Antarctic Program (ANT-1043745 to KH) and the Office of Polar Programs (OPP-1916661 to KH and OPP 1916665 to AM).

# References

Arntz, W., Brey, T., and Gallardo, V. (1994). Antarctic Zoobenthos. Oceanogr Mar. Biol. 32, 241–304.

Aronson, R. B., Moody, R. M., Ivany, L. C., Blake, D. B., Werner, J. E., and Glass, A. (2009). Climate change and trophic response of the Antarctic bottom fauna. *PloS One* 4, e4385. doi: 10.1371/journal.pone.0004385

Barry, J. P., Grebmeier, J. M., Smith, J., and Dunbar, R. B. (2003). Oceanographic versus seafloor-habitat control of benthic megafaunal communities in the SW Ross Sea, Antarctica. *Biogeochem Ross Sea* 78, 327-353. doi: 10.1029/078ARS21

Boucot, A. J., and Carney, R. S. (1981). "Principles of benthic marine paleoecology," (New York, USA: Academic Press).

Brannock, P. M., Learman, D. R., Mahon, A. R., Santos, S. R., and Halanych, K. M. (2018). Meiobenthic community composition and biodiversity along a 5500 km transect of Western Antarctica: a metabarcoding analysis. *Mar. Ecol. Prog. Ser.* 603, 47–60. doi: 10.3354/ meps12717

Bray, J. R., and Curtis, J. T. (1957). An ordination of the upland forest communities of southern Wisconsin. *Ecol. Monogr.* 27, 326–349. doi: 10.2307/1942268

Charctic, National Snow and Ice Data Center (Accessed March 18, 2023).

# **Acknowledgments**

We thank the National Science Foundation for funds to collect and analyze these data. We also thank the ships' crews, scientific support staff, and scientific crew of the *ASRV Laurence M. Gould* (LMG 13-12) and *RVIB Nathaniel B. Palmer* (NBP 12-10). We acknowledge and thank David Branson for initial photo editing steps. Further, we thank the reviewers for their constructive feedback on our manuscript. We especially thank the electronic technicians on the NBP 12-10 and LMG 13-12 cruises who assisted with the deployment of the Yoyo camera.

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

## Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmars.2023.1094283/full#supplementary-material

Chiantore, M., Guidetti, M., Cavallero, M., De Domenico, F., Albertelli, G., and Cattaneo-Vietti, R. (2006). Sea Urchins, sea stars and brittle stars from Terra Nova bay (Ross Sea, Antarctica). *Polar Biol.* 29, 467–475. doi: 10.1007/s00300-005-0077-2

Clarke, A. (1988). Seasonality in the Antarctic marine environment. Comp. Biochem. Physiol. Part B: Comp. Biochem. 90, 461–473.

Clarke, K. R., Gorley, R., Somerfield, P., and Warwick, R. (2014). *Change in marine communities: an approach to statistical analysis and interpretation* (Plymouth Marine Laboratory, Plymouth, UK: Primer-E Ltd).

Clarke, A., and Johnston, N. M. (2003). "Antarctic Marine benthic diversity," in *Oceanography and marine biology, an annual review, volume 41* (Boca Raton, Florida, United States: CRC Press), 55–57.

Clarke, A., Murphy, E. J., Meredith, M. P., King, J. C., Peck, L. S., Barnes, D. K., et al. (2007). Climate change and the marine ecosystem of the western Antarctic peninsula. *Philos. Trans. R. Soc. B: Biol. Sci.* 362, 149–166. doi: 10.1098/rstb.2006.1958

Convey, P., and Peck, L. S. (2019). Antarctic Environmental change and biological responses. *Sci. Adv.* 5, eaaz0888. doi: 10.1126/sciadv.aaz0888

Cummings, V. J., Bowden, D. A., Pinkerton, M. H., Halliday, N. J., and Hewitt, J. E. (2021). Ross Sea Benthic ecosystems: Macro-and mega-faunal community patterns

from a multi-environment survey. Front. Mar. Sci. 8, 629787. doi: 10.3389/ fmars.2021.629787

Dale, J. E., Aitken, A. E., Gilbert, R., and Risk, M. J. (1989). Macrofauna of Canadian arctic fjords. *Mar. Geology* 85 (2-4), 331–358.

Dayton, P. K., Robilliard, G. A., Paine, R. T., and Dayton, L. B. (1974). Biological accommodation in the benthic community at McMurdo sound, Antarctica. *Ecol. Monogr.* 44, 105–128. doi: 10.2307/1942321

De Jong, J., Stammerjohn, S., Ackley, S., Tison, J.-L., Mattielli, N., and Schoemann, V. (2015). Sources and fluxes of dissolved iron in the Bellingshausen Sea (West antarctica): The importance of sea ice, icebergs and the continental margin. *Mar. Chem.* 177, 518–535. doi: 10.1016/j.marchem.2015.08.004

Deppeler, S. L., and Davidson, A. T. (2017). Southern ocean phytoplankton in a changing climate. *Front. Mar. Sci.* 4, 40. doi: 10.3389/fmars.2017.00040

Fernández-González, C., Tarran, G. A., Schuback, N., Woodward, E. M. S., Arístegui, J., and Marañón, E. (2022). Phytoplankton responses to changing temperature and nutrient availability are consistent across the tropical and subtropical Atlantic. *Commun. Biol.* 5, 1035. doi: 10.1038/s42003-022-03971-z

Gerdes, D., Klages, M., Arntz, W., Herman, R., Galéron, J., and Hain, S. (1992). Quantitative investigations on macrobenthos communities of the southeastern Weddell Sea shelf based on multibox corer samples. *Polar Biol.* 12, 291–301. doi: 10.1007/ BF00238272

Grange, L. J., and Smith, C. R. (2013). Megafaunal communities in rapidly warming fjords along the West Antarctic peninsula: hotspots of abundance and beta diversity. *PloS One* 8, e77917. doi: 10.1371/journal.pone.0077917

Gray, J. S. (2001). Antarctic Marine benthic biodiversity in a world-wide latitudinal context. *Polar Biol.* 24, 633–641. doi: 10.1007/s003000100244

Grebmeier, J. M., and Barry, J. P. (1991). The influence of oceanographic processes on pelagic-benthic coupling in polar regions: a benthic perspective. *J. Mar. Syst.* 2, 495–518. doi: 10.1016/0924-7963(91)90049-Z

Griffiths, H. J. (2010). Antarctic Marine biodiversity-what do we know about the distribution of life in the southern ocean? *PloS One* 5, e11683. doi: 10.1371/journal.pone.0011683

Griffiths, H. J., Arango, C. P., Munilla, T., and Mcinnes, S. J. (2011). Biodiversity and biogeography of southern ocean pycnogonids. *Ecography* 34, 616–627. doi: 10.1111/j.1600-0587.2010.06612.x

Gutt, J., Arndt, J., Kraan, C., Dorschel, B., Schröder, M., Bracher, A., et al. (2019). Benthic communities and their drivers: A spatial analysis off the Antarctic peninsula. *Limnol Oceanogr* 64, 2341–2357. doi: 10.1002/lno.11187

Gutt, J., Griffiths, H. J., and Jones, C. D. (2013). Circumpolar overview and spatial heterogeneity of Antarctic macrobenthic communities. *Mar. Biodivers.* 43, 481–487. doi: 10.1007/s12526-013-0152-9

Gutt, J., Sirenko, B. I., Smirnov, I. S., and Arntz, W. E. (2004). How many macrozoobenthic species might inhabit the Antarctic shelf? *Antarctic Sci.* 16, 11–16. doi: 10.1017/S0954102004001750

Gutt, J., and Starmans, A. (1998). Structure and biodiversity of megabenthos in the Weddell and lazarev seas (Antarctica): ecological role of physical parameters and biological interactions. *Polar Biol.* 20, 229–247. doi: 10.1007/ s003000050300

Halanych, K. M., and Mahon, A. R. (2018). Challenging dogma concerning biogeographic patterns of Antarctica and the southern ocean. *Annu. Rev. Ecol Evolution Systematics* 49, 355–378. doi: 10.1146/annurev-ecolsys-121415-032139

Hellmer, H. H., Kauker, F., Timmermann, R., Determann, J., and Rae, J. (2012). Twenty-first-century warming of a large Antarctic ice-shelf cavity by a redirected coastal current. *Nature* 485 (7397), 225–228.

Hempel, G. (1985). arine biology of polar regions and effects of stress on marine organisms J. S. Gray and M. E. Christiansen eds. Wiley, Chichester, 3-33.

Hendler, G., Miller, J. E., Pawson, D. L., and Kier, P. M. (1995). Sea Stars, sea urchins, and allies: echinoderms of Florida and the Caribbean (Washington D.C: Smithsonian Institution).

Hughes, S. J. M., Ruhl, H. A., Hawkins, L. E., Hauton, C., Boorman, B., and Billett, D. S. (2011). Deep-sea echinoderm oxygen consumption rates and an interclass comparison of metabolic rates in asteroidea, crinoidea, echinoidea, holothuroidea and ophiuroidea. *J. Exp. Biol.* 214, 2512–2521. doi: 10.1242/ jeb.055954

Iken, K., Konar, B., Benedetti-Cecchi, L., Cruz-Motta, J. J., Knowlton, A., Pohle, G., et al. (2010). Large-Scale spatial distribution patterns of echinoderms in nearshore rocky habitats. *PloS One* 5, e13845. doi: 10.1371/journal.pone.0013845

Isla, E., and Gerdes, D. (2019). Ongoing ocean warming threatens the rich and diverse macrobenthic communities of the Antarctic continental shelf. *Prog. Oceanography* 178, 102180.

Jacobs, S. S., and Comiso, J. C. (1997). Climate variability in the amundsen and Bellingshausen seas. J. Climate 10, 697–709. doi: 10.1175/1520-0442(1997)010<0697: CVITAA>2.0.CO;2

Johnsen, S., and Kier, W. M. (1999). Shade-seeking behaviour under polarized light by the brittlestar *Ophioderma brevispinum* (Echinodermata: Ophiuroidea). J. Mar. Biol. Assoc. United Kingdom 79, 761–763. doi: 10.1017/S0025315498000940 Konecki, J. T., and Targett, T. E. (1989). Eggs and larvae of *Nototheniops larseni* from the spongocoel of a hexactinellid sponge near Hugo island, Antarctic peninsula. *Polar Biol.* 10, 197–198. doi: 10.1007/BF00238495

Learman, D. R., Henson, M. W., Thrash, J. C., Temperton, B., Brannock, P. M., Santos, S. R., et al. (2016). Biogeochemical and microbial variation across 5500 km of Antarctic surface sediment implicates organic matter as a driver of benthic community structure. *Front. Microbiol.* 7, 284. doi: 10.3389/fmicb.2016.00284

Margalef, R. (1958). Information theory in biology. Gen. Syst. Yearbook 3, 36-71.

Menezes, V. V., Macdonald, A. M., and Schatzman, C. (2017). Accelerated freshening of Antarctic bottom water over the last decade in the southern Indian ocean. *Sci. Adv.* 3, e1601426. doi: 10.1126/sciadv.1601426

Meyer, D. M. J. D. (1983). Sea Lilies and feather stars. Am. Scientist 71, 354-365.

Miranda, T. P., Fernandez, M. O., Genzano, G. N., Cantero, Á.L.P., Collins, A. G., and Marques, A. C. (2021). Biodiversity and biogeography of hydroids across marine ecoregions and provinces of southern south America and Antarctica. *Polar Biol.* 44, 1669–1689. doi: 10.1007/s00300-021-02909-1

Moles, J., Figuerola, B., Campanya-Llovet, N., Monleon-Getino, T., Taboada, S., and Avila, C. (2015). Distribution patterns in Antarctic and subantarctic echinoderms. *Polar Biol.* 38, 799–813. doi: 10.1007/s00300-014-1640-5

Morrison, A., Hogg, A. M., England, M. H., and Spence, P. (2020). Warm circumpolar deep water transport toward Antarctica driven by local dense water export in canyons. *Sci. Adv.* 6, eaav2516. doi: 10.1126/sciadv.aav2516

Pielou, E. C. (1966). The measurement of diversity in different types of biological collections. J. Theor. Biol. 13, 131-144. doi: 10.1016/0022-5193(66)90013-0

Piepenburg, D., Archambault, P., Ambrose, W. G., Blanchard, A. L., Bluhm, B. A., Carroll, M. L., et al. (2011). Towards a pan-Arctic inventory of the species diversity of the macro-and megabenthic fauna of the Arctic shelf seas. *Mar. Biodivers.* 41, 51–70. doi: 10.1007/s12526-010-0059-7

Pineda-Metz, S. E., and Gerdes, D. (2018). Seabed images versus corer sampling: a comparison of two quantitative approaches for the analysis of marine benthic communities in the southern Weddell Sea (Southern ocean). *Polar Biol.* 41, 515–526. doi: 10.1007/s00300-017-2211-3

Pineda-Metz, S. E., Gerdes, D., and Richter, C. (2020). Benthic fauna declined on a whitening Antarctic continental shelf. *Nat. Commun.* 11, 1–7. doi: 10.1038/s41467-020-16093-z

Pineda-Metz, S. E., Isla, E., and Gerdes, D. (2019). Benthic communities of the filchner region (Weddell Sea, Antarctica). *Mar. Ecol. Prog. Ser.* 628, 37–54. doi: 10.3354/meps13093

Post, A. L., Lavoie, C., Domack, E. W., Leventer, A., Shevenell, A., and Fraser, A. D. (2017). Environmental drivers of benthic communities and habitat heterogeneity on an East Antarctic shelf. *Antarctic Sci.* 29, 17–32. doi: 10.1017/S0954102016000468

Ruan, X., Speer, K. G., Thompson, A. F., Schulze Chretien, L. M., and Shoosmith, D. R. (2021). Ice-shelf meltwater overturning in the Bellingshausen Sea. J. Geophys Research: Oceans 126, e2020JC016957. doi: 10.1029/2020JC016957

Shannon, C. E. (1948). A mathematical theory of communication. *Bell system Tech. J.* 27, 379–423. doi: 10.1002/j.1538-7305.1948.tb01338.x

Simpson, E. H. (1949). Measurement of diversity. *Nature* 163, 688–688. doi: 10.1038/ 163688a0

Smith, C. R., Demaster, D. J., Thomas, C., Sršen, P., Grange, L., Evrard, V., et al. (2012). Pelagic-benthic coupling, food banks, and climate change on the West Antarctic peninsula shelf. *Oceanography* 25, 188–201. doi: 10.5670/oceanog.2012.94

Smith, W. O.Jr., and Gordon, L. I. (1997). Hyperproductivity of the Ross Sea (Antarctica) polynya during austral spring. *Geophys Res. Lett.* 24, 233–236. doi: 10.1029/96GL03926

Stern, A., Dinniman, M., Zagorodnov, V., Tyler, S., and Holland, D. (2013). Intrusion of warm surface water beneath the McMurdo ice shelf, Antarctica. *J. Geophys Research: Oceans* 118, 7036–7048. doi: 10.1002/2013JC008842

Sumida, P. Y., Bernardino, A. F., Stedall, V. P., Glover, A. G., and Smith, C. R. (2008). Temporal changes in benthic megafaunal abundance and composition across the West Antarctic peninsula shelf: results from video surveys. *Deep Sea Res. Part II: Topical Stud. Oceanogr* 55, 2465–2477.

Syvitski, J. P., Farrow, G. E., Atkinson, R., Moore, P., and Andrews, J. (1989). Baffin Island fjord macrobenthos: bottom communities and environmental significance. *Arctic* 42, 232–247. doi: 10.14430/arctic1662

Tripathy, S., and Jena, B. (2019). Iron-stimulated phytoplankton blooms in the southern ocean: a brief review. *Remote Sens. Earth Syst. Sci.* 2, 64–77. doi: 10.1007/s41976-019-00012-y

Woods, H. A., Moran, A. L., Arango, C. P., Mullen, L., and Shields, C. (2009). Oxygen hypothesis of polar gigantism not supported by performance of Antarctic pycnogonids in hypoxia. *Proc. R. Soc. B: Biol. Sci.* 276, 1069–1075. doi: 10.1098/rspb.2008.1489

Wu, X., Niklas, K. J., and Sun, S. (2021). Climate change affects detritus decomposition rates by modifying arthropod performance and species interactions. *Curr. Opin. Insect Sci.* 47, 62–66. doi: 10.1016/j.cois.2021.05.002

Zwerschke, N., Sands, C. J., Roman-Gonzalez, A., Barnes, D. K., Guzzi, A., Jenkins, S., et al. (2022). Quantification of blue carbon pathways contributing to negative feedback on climate change following glacier retreat in West Antarctic fjords. *Global Change Biol.* 28, 8–20. doi: 10.1111/gcb.15898