

Noctiluca Scintillans Distribution Largely Regulated by Phytoplankton Biomass in the East China Sea and Southern Yellow Sea

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OPEN ACCESS

Edited by:

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Reviewed by:

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Specialty section:

This article was submitted to Marine Biogeochemistry, a section of the journal Frontiers in Marine Science

Received: 18 March 2022 Accepted: 24 May 2022 Published: 23 June 2022

Citation:

Sheng L, Jiang Z, Sun Z, Zhu Y, Zhai H, Ding L, Tong M, Chen J, Chen Q and Zeng J (2022) Noctiluca Scintillans Distribution Largely Regulated by Phytoplankton Biomass in the East China Sea and Southern Yellow Sea. Front. Mar. Sci. 9:899334. doi: 10.3389/fmars.2022.899334 ¹ Ocean College, Zhejiang University, Zhoushan, China, ² Key Laboratory of Marine Ecosystem Dynamics, Second Institute of Oceanography, Ministry of Natural Resources, Hangzhou, China, ³ Key Laboratory of Marine Space Resource Management Technology, Ministry of Natural Resources, Hangzhou, China, ⁴ Marine Ecosystem Observation and Research Station on the Changjiang Estuary, Ministry of Natural Resources, Hangzhou, China, ⁵ Key Laboratory of Nearshore Engineering Environment and Ecological Security of Zhejiang Province, Hangzhou, China, ⁶ Key Laboratory of Satellite Ocean Environment Dynamics, Second Institute of Oceanography, Ministry of Natural Resources, Hangzhou, China, ⁷ School of Oceanography, Shanghai Jiao Tong University, Shanghai, China

Noctiluca scintillans feeds on a large number of phytoplankton, including diatoms and dinoflagellates, and frequently forms a red tide in the East China Sea (ECS) and southern Yellow Sea (SYS). However, the spatiotemporal distribution pattern, controlling factors, and long-term change of N. scintillans in the ECS and SYS remain unclear. In the present study, we collected N. scintillans samples from the ECS and SYS throughout the four seasons of 2011. We sampled phytoplankton and environmental parameters simultaneously. The depth-integrated abundance (DIA) of N. scintillans was the highest and lowest in summer and winter, respectively. N. scintillans is distributed abundantly in eutrophic coastal waters and the Changjiang Estuary, which are characterized by high concentrations of phytoplankton and chlorophyll-a. A Spearman correlation test demonstrated that its DIA in the upper 30-m water column was generally more significantly associated with phytoplankton abundance and chlorophyll-a concentration than with temperature and salinity. The results of the generalized additive models revealed that chlorophyll-a concentration explained more of the variation in N. scintillans abundance than temperature and salinity throughout the year, particularly in warm seasons. These findings indicate that the seasonal and spatial changes of N. scintillans are largely regulated by phytoplankton biomass. Compared with the historical data from 1959 and 2002, the abundance of N. scintillans in the Changjiang Estuary increased considerably in 2011 with increasing phytoplankton abundance resulting from accelerated eutrophication and warming. These results clarify the controlling factors, red-tide formation mechanism, and changing trends associated with the N. scintillans in the ECS and SYS.

Keywords: noctiluca scintillans, East China Sea, Southern Yellow Sea, controlling factors, phytoplankton

INTRODUCTION

Noctiluca scintillans is widely distributed in temperate, subtropical, and tropical coastal waters (Elbrächter and Qi, 1998; Harrison et al., 2011). It tends to be more abundant, even forming a red tide, in eutrophic inshore waters, estuaries, and upwellings because it requires a large supply of food based on phytoplankton (e.g., diatoms and dinoflagellates) to maintain its high growth rate (Harrison et al., 2011). The red tide formed by *N. scintillans* adheres to the gills of aquatic animals, causing them to suffocate from a lack of oxygen, and the ingestion of fish eggs causes a decline in fish stocks (Enomoto, 1956). N. scintillans contains a large amount of ammonium in its cells, which may affect the quality of seawater and the ecological environment (Ara et al., 2013; Zhou et al., 2020). N. scintillans influences the plankton food web and biogeochemical processes by feeding on microalgae, copepod eggs, fish eggs, and protozoa (Hattori, 1962; Zhang et al., 2016).

N. scintillans can be divided into two types according to the presence of the symbiotic algae Protoeuglena noctiluca: green N. scintillans with symbiotic algae and red N. scintillans without symbiotic algae (Harrison et al., 2011). N. scintillans reproduces asexually through the binary fission of vegetative cells and also sexually through the formation of gametocytes to produce gametes (Song et al., 2016; Sathish et al., 2020). Studies have demonstrated that the temporal and spatial distribution of N. scintillans is affected by various physical, chemical, and biological factors in different waters. Tada et al. (2004) revealed that the water stability from late spring to early summer promoted an increase in N. scintillans abundance. The proliferation of N. scintillans is generally regarded as an indirect result of eutrophication because the increase in nutrient input promotes a higher phytoplankton biomass, thereby providing more food for N. scintillans (Dela-Cruz et al., 2003; Smitha et al., 2022).

The East China Sea (ECS) and southern Yellow Sea (SYS) are located off the west coast of the Pacific Ocean; they span latitudes from subtropical to temperate waters, and despite considerable seasonal changes, they are mainly controlled by the warm, salty, oligotrophic Kuroshio system (including the Taiwan Warm Current and Yellow Sea [YS] Warm Current) and the lowsalinity, eutrophic coastal current system (Changjiang Diluted Water, ECS Coastal Current, and YS Coastal Current; Figure 1). In summer, the Kuroshio mainstream flows toward the northeast along the ECS shelf break (200-1,000 m isobaths), and the Taiwan Warm Current enhances and flows northeastward along the 50-m isobath under the prevailing southwest monsoon. Notably, the low-salinity (≤31) Changjiang Diluted Water flows northeastward, and dominates the Changjiang Estuary, southern SYS, and northern ECS. Deeper water is controlled by YS Cold Water (≤10°C) in the SYS. In winter, the Taiwan Warm Current weakens and the high-temperature YS Warm Current intrudes into the SYS. The temperature, salinity, and nutrient concentrations vary greatly on spatial and seasonal scales because of the water mass movement of the Kuroshio and coastal current in addition to solar irradiance. Studies on N. scintillans in the ECS and SYS have been limited to coastal waters, including the Changjiang Estuary (Xu, 2009),

Jiaozhou Bay (Tian et al., 2017; Wang et al., 2018), and Yantai coast (Zhang et al., 2020c). No multidisciplinary investigations on the seasonal and spatial distribution and population dynamics of *N. scintillans* across the entire ECS and SYS have been conducted. The controlling factors and regulation mechanism of *N. scintillans* in the ECS and SYS remain unclear; therefore, a large-scale baseline survey is warranted.

N. scintillans is a eurythermal and euryhaline species; thus, it tolerates different temperatures and salinity in different regions. Xu (2009) revealed that the most suitable temperature range for N. scintillans in the Changjiang Estuary ranges from 15°C to 20° C. However, N. scintillans red tides were also observed in the Changjiang Estuary during summer when the water temperature was between 28°C and 30°C (Qi et al., 2019). This finding demonstrates that temperature is not the most crucial factor for inducing N. scintillans blooms in the Changjiang Estuary. Tseng et al. (2011) noted that N. scintillans was abundant in the eutrophic low-salinity waters of the southern and western parts of the ECS, especially in the Changjiang Estuary, and Zhang et al. (2020c) revealed that the rapid growth of N. scintillans in Yantai, China, was caused by an increase in phytoplankton abundance. Remote sensing data reveal that N. scintillans blooms frequently coincide with the fronts of the Changjiang Diluted Water, which are characterized by low salinity and abundant food (phytoplankton; Sriwoon et al., 2010; Qi et al., 2019). Furthermore, N. scintillans blooms were observed in the offshore waters of the ECS to at least 126°E, which have relatively high salinity (Qi et al., 2019). The eutrophic Changjiang Diluted Water and coastal current promote phytoplankton growth (Zhu et al., 2009; Jiang et al., 2015; Jiang et al., 2019; Liblik et al., 2020). The N. scintillans in the ECS and SYS is the red type (Harrison et al., 2011) that consumes large amounts of phytoplankton cells through heterotrophic feeding. Therefore, we assume that the spatial and temporal distribution of N. scintillans in the ECS and SYS is influenced more by phytoplankton biomass rather than by temperature and salinity.

In the present study, we conducted four cruises in the ECS and SYS for all four seasons of 2011. We collected *N. scintillans* using net-collection and water-collection methods. Our objectives were to (1) compare the temporal and spatial distribution of *N. scintillans* using different collection methods; (2) explore the adaptability of *N. scintillans* to temperature, salinity, phytoplankton abundance, and other factors; and (3) ascertain the long-term changes in *N. scintillans* in the Changjiang Estuary compared with historical data since 1959. The present study improves our understanding of the distribution pattern, controlling factors, red-tide formation mechanism, and changing trend of *N. scintillans* in the ECS and SYS.

MATERIALS AND METHODS

Study Area

Four multidisciplinary cruises were conducted in the ECS and SYS for the four seasons (i.e., spring [April–March 2011], summer



[July 2011], autumn [October–November 2011], and winter [December 2011–January 2012]). A total of 67, 62, 68, and 67 stations were investigated in spring, summer, autumn, and winter, respectively. **Figure 1** presents the study area and all sampling stations. Section D is the dividing line between the ECS and SYS.

Sample Collection

At each station, temperature, salinity, density (σ t), and water depth were measured *in situ* by using a conductivity– temperature–depth (CTD) recorder (SBE 917 Plus, Seabird, Sea Bird Corporation, USA). The water column stratification index (SI) was defined as $\Delta \sigma$ t = bottom density – surface density.

Water samples at different depths (2 or 3, 10, 30, 50, 75, 100, 150, and 200 m, as well as the bottom and deep chlorophyll-a [Chl-a] maximum) were collected in 12-L Niskin bottles. The collected seawater was used to determine nutrients and Chl-a concentrations and conduct the microscopic counting of *N*. *scintillans* abundance.

For the net-collected samples, phytoplankton and *N.* scintillans samples were collected vertically from the bottom (3-5 m above the seabed) to the surface using a plankton net with a 76-µm mesh (inner diameter of the net mouth: 37 cm; length: 280 cm) at each station. The amount of filtered water was recorded using a digital flow meter (Model 438115, Hydro Bios). All net-collected and water-collected samples were preserved using 4% formalin, and 2–3 mL of the net-collected samples were extracted for counting using a Leica DMI 3000B microscope. After 24–48 h of sedimentation, the water-collected samples (~1,000 mL) of *N. scintillans* were concentrated to 10–100 mL and then sieved using an 8-µm nylon sieve. The *N. scintillans* was counted in the sieve using a Leica DMI 3000B microscope.

After a specific amount of seawater was filtered through a 0.45- μ m polycarbonate membrane, a continuous flow analyzer (Skalar San++, Netherlands) was employed to determine the concentrations of dissolved inorganic nitrogen (DIN: NO₃⁻+NO₂⁻+NH₄⁺) and phosphorus (DIP) in a laboratory. The sample of Chl-*a* was obtained by filtering 100–250 mL of seawater (200- μ m sieve silk) through a 0.7- μ m Whatman GF/F filter membrane. After being extracted using 10 mL of 90% acetone at –20°C for 24 h, the concentration of Chl-*a* was measured using a Turner Design Fluorometer. The temperature, salinity, nutrient, and phytoplankton datasets used in this analysis have already been published (Quan, et al., 2013; Jiang et al., 2018; Jiang et al., 2020).

Data Analysis

The depth-integrated abundance (DIA) of *N. scintillans* in the whole water column at each station was calculated.

On the basis of the abundance of *N. scintillans* at each water layer, the water-collected DIA C_t of the *N. scintillans* at each station was calculated using the equation as follows:

$$C_t = \sum_{i=1}^{n-1} \frac{(C_i + C_i + 1)}{2} \times (D_i + 1 - D_i)(1 \le i \le n - 1),$$

where C_i is the *N. scintillans* abundance of the *i*th layer (cells/L), C_{i+1} is the *N. scintillans* abundance of the *i* + 1st layer (cells/L), D_i is the sampling depth of the *i*th layer (m), D_{i+1} is the sampling depth of the *i* + 1st layer (m), and *n* is the number of sampling layers.

According to the net-collected abundance of *N. scintillans* at each station, the DIA C_d of *N. scintillans* at each station was calculated as follows:

$$C_d = C \times Z_{\max}$$

Where *C* is the abundance of net-collected *N*. *scintillans* and Z_{max} is the water depth (m) at each station.

On the basis of the environmental variables (temperature, salinity, and nutrients) at each station, the depth-average value (N_t) of each variable in the upper 30-m water column at each station was calculated as follows:

$$N_t = \sum_{i=1}^{n-1} \frac{(N_i + N_i + 1)}{2} (1 \le i \le n - 1)$$

Where N_i is the variables of the *i*th layer, N_{i+1} is the variable of the *i* + 1st layer, and *n* is the number of sampling layers.

The high density of *N. scintillans* was generally observed in the upper 30-m water column. Therefore, a Spearman correlation analysis and generalized additive models (GAMs) were applied to determine the relationship between the DIA of *N. scintillans* and the depth-average values of the environmental variables (except SI) in the upper 30-m water column. A Spearman correlation analysis and GAMs were applied using SPSS 20.0 and R, respectively. The establishment of GAMs was completed using the mgcViz package (version 4.0.2). The expression of the GAMs was expressed was as follows:

$$Y = \alpha + f_1(X_1) + f_2(X_2) + f_3(X_3) + f_4(X_4) + f_5(X_5)$$

In the aforementioned equation, the abundance at several stations was 0; thus, *Y* is the log (C_d +1)-transformed value of *N*. *scintillans* abundance; α is the intercept of the fit function; f_1, f_2, f_3, f_4 , and f_5 are the smooth functions of the five variables of *X*; X_1 and X_2 are the depth-average temperature and salinity, respectively, in the upper 30-m water column, respectively; X_3 is the SI; X_4 is the depth-integrated Chl-*a* in the upper 30-m water column; and X_5 is the log-transformed C_d of phytoplankton. The *N*. *scintillans* in the ECS and SYS is a heterotrophic dinoflagellate and cannot directly utilize nutrients (Harrison et al., 2011). Thus, DIN and DIP were not included in the formula. Model building followed a stepwise forward approach with the largest cumulative deviation explained value. The model with the lowest value of Akaike information criterion (AIC) was selected as the optimal model.

The relationship between C_d and C_t was determined using linear regression. A Kruskal–Wallis one-way analysis of variance by ranks was performed to detect significant differences in the C_d and C_t of *N. scintillans* and environmental variables among different seasons. These statistical tests were applied using SPSS 20.0. We used the net-collection method to describe the spatial distribution pattern in the abundance of *N. scintillans*. Because C_d ranged from 0 to 10⁸ cells/m² across the entire ECS and SYS, a log (C_d +1) transformation was performed to present the spatial distribution pattern of *N. scintillans*. The distribution of environmental factors and *N. scintillans* abundance was drawn using Ocean Data View (ODV) 4.5.

RESULTS

Environment Conditions

Our results revealed significant (p < 0.01) seasonal differences in temperature and salinity both on the surface and in the upper 30-m

water column (Table 1). The SI and depth-average nutrients (DIN and DIP) results also varied significantly among the seasons (p <0.01). The depth-average and surface temperature were higher in summer (21.5°C and 25.0°C) and autumn (21.7°C and 21.7°C) than in winter (15.3°C and 15.3°C) and spring (9.9°C and 10.0°C). Because of the presence of the low-salinity Changjiang Diluted Water extension off the Changjiang Estuary, the depth-average salinity of the upper 30-m water column and surface salinity were lower in summer (31.8 and 30.6) and autumn (32.8 and 32.8) than in spring (33.1 and 33.0) and winter (33.3 and 33.3). Furthermore, because of the enhancement of the Changjiang Diluted Water and YS Cold Water Mass, the SI was higher in summer (4.69) and autumn (1.42) than in spring (0.22) and winter (0.27). The depthaverage concentrations of the DIN and DIP were markedly lower in summer and autumn than in winter and spring because the water bodies mixed to a greater degree in winter and spring than in summer and autumn.

In winter and spring, temperature and salinity were high in the southern and eastern parts of the ECS and in the southeastern part of the SYS because of the intrusion of the Kuroshio, Taiwan Warm Current, and YS Warm Current (**Figure 2**). In summer and autumn, the temperature and salinity were higher in the southern part of the ECS than in the northern part of the ECS and in the SYS influenced by the Changjiang Diluted Water and YS Coastal Current. The southern part of the SYS and the northern ECS were controlled by the Changjiang Diluted Water, which had a strong halocline, and the deeper SYS was dominated by the YS Cold Water Mass, which had a strong thermocline; consequently, a high SI was detected off the Changjiang Estuary and in the central SYS.

Phytoplankton Biomass

The phytoplankton biomass in the ECS and SYS varied significantly (p < 0.01) among the seasons. The C_d values of Chl-*a* concentration and phytoplankton abundance were the highest in summer and lowest in winter. In spring, phytoplankton biomass was high in the coastal waters (**Figure 3**). In summer and autumn, the phytoplankton biomass was high in the Zhejiang coastal waters and Changjiang Estuary controlled by the ECS Coastal Current, upwelling, and Changjiang Diluted Water. In winter, the phytoplankton biomass was high in the southeast part of the ECS controlled by the Kuroshio where nutrients and light were not restricted.

N. Scintillans Abundance

The average C_d of the *N. scintillans* was the highest in summer, followed by autumn, spring, and winter in both the ECS and SYS (**Table 2**). C_d was higher in the SYS than in the ECS for all seasons except summer. High *N. scintillans* C_d (**Figure 4**) and C_t (**Figure S1**) were observed in coastal waters and the Changjiang Estuary during all seasons, which were controlled by the coastal current and Changjiang Diluted Water. Low *N. scintillans* C_d and C_t were observed in the offshore waters controlled by the Kuroshio and YS Cold Water Mass. Our data also revealed a significant correlation (p < 0.05) between the C_d and C_t of the *N*.

Environmental parameters	Spring(n=67)	Summer(n=62)	Autumn(n=68)	Winter(n=67)	н
SST	10.0 ± 3.0 ^d	25.0 ± 2.3ª	21.7 ± 2.2 ^b	15.3 ± 4.2°	196.9**
SSS	33.0 ± 1.0^{a}	30.6 ± 2.8^{b}	32.8 ± 1.3^{a}	33.3 ± 1.2ª	57.7**
Depth-average temperature	$9.9 \pm 3.1^{\circ}$	21.5 ± 3.8^{a}	21.7 ± 2.2^{a}	15.3 ± 4.1 ^b	162.4**
Depth-average salinity	33.1 ± 0.9^{a}	31.8 ± 1.6^{b}	32.8 ± 1.2^{a}	33.3 ± 1.1 ^a	38.7**
SI	$0.22 \pm 0.44^{\circ}$	4.69 ± 1.67^{a}	1.42 ± 1.16^{b}	0.27 ± 0.46^{d}	156.5**
Depth-average DIN	9.23 ± 4.29^{a}	6.80 ± 7.11^{b}	4.77 ± 3.74^{b}	5.99 ± 2.63 ^{ac}	45.9**
Depth-average DIP	0.44 ± 0.22^{a}	0.38 ± 0.55^{b}	0.40 ± 0.30^{b}	0.43 ± 0.24^{a}	16.8**
Phytoplankton	760.2 ± 4491.9 ^b	1316.7 ± 6930.8 ^{ac}	1177.3 ± 4088.7 ^a	56.4 ± 218.7 ^b	19.9**
Chl-a	65.33 ± 56.05 ^a	54.97 ± 61.55 ^{ab}	45.05 ± 22.92 ^{ab}	39.39 ± 13.86 ^b	25.0**
Net-collected N. scintillans	1.06 ± 5.06^{b}	359.89 ± 860.06^{a}	26.14 ± 146.31^{ab}	0.38 ± 1.48 ^{ab}	15.1**
Water-collected N. scintillans	3.90 ± 8.92^{ac}	173.06 ± 995.81^{a}	7.81 ± 23.48^{a}	1.35 ± 4.16^{bc}	196.9**

TABLE 1 | Comparison of depth-integrated abundance of *N. scintillans* and environmental variables (mean ± standard deviation) in the East China Sea and southern

 Yellow Sea in different seasons by using Kruskal–Wallis (*H*) test.

SST, sea surface temperature (°C); SSS, sea surface salinity; SI, stratification index (kg m⁻³), which was defined as $\Delta \sigma t =$ bottom density – surface density. Depth-average DIN, DIP, Depth-average dissolved inorganic nitrogen (µmol/L), phosphorus (µmol/L). Depth-average temperature (°C), salinity, and nutrients show the values in the upper 30-m water column. Chl-a (mg/m²), phytoplankton (× 10⁶ cells/m²) and N. scintillans (× 10⁴ cells/m²) show the depth-integrated abundance. Different lowercase letters indicate a significant difference (p < 0.05) in the same row. **p < 0.01.

scintillans in the ECS and SYS, and the corresponding slope (**Figure S2**) was higher in autumn (0.78) and winter (0.75) than in spring (0.62) and summer (0.64).

We observed an obvious difference between the density of *N.* scintillans at different depths along section P in all seasons (**Figure 5**). However, a high density of *N. scintillans* was generally observed in the upper 30-m water column. Notably, the highest abundance (5,314 cells/L) was observed in summer at a depth of 10 m at station T07 (upwelling water), which was characterized by abundant phytoplankton, high nutrient concentrations and salinity, and low temperatures (**Figure S3**). The highest C_t of *N. scintillans* in summer was also detected at this station (**Figure S1**).

Relationship Between *N. Scintillans* and Environmental Factors

The Spearman correlation demonstrated that the annual DIA of the *N. scintillans* in the upper 30-m water column was significantly (p < 0.05) and positively correlated with phytoplankton abundance, Chl-*a* concentration, temperature, and SI but significantly (p < 0.01) and negatively correlated with salinity (**Table 3**). Particularly in warm seasons (summer and autumn), the DIA of the *N. scintillans* was more significantly associated with phytoplankton abundance and Chl-*a* concentration than with temperature, salinity, and SI. **Figure 6** reveals that *N. scintillans* was abundant in the coastal waters controlled by the low-salinity Changjiang Diluted Water and coastal current, particularly in the fronts. A low density of *N. scintillans* was observed in the Kuroshio, YS Cold Water Mass, and YS Warm Current.

All the established models are shown in **Table S1**, and the summary of the final optimal models is shown in **Table 4**. In summer, winter, and year-round models, the AIC value decreased with the increase of environmental factors. Therefore, the best model for summer had 5 factors, and the best model for the winter and full year had 4 factors (SI was not included in winter and full year because cumulative explained deviation was decreased after its adding). There were the two-factor (Temperature + SI) and one-factor (Chl-a) models in

spring and autumn, respectively, with the smallest AIC values and the best fit. The results of the GAMs demonstrated that relative to temperature and salinity, Chl-*a* concentration explained more of the variation in the abundance of *N*. *scintillans* throughout the year. **Table 4** and **Figure 7** reveal that Chl-a concentration, salinity, temperature, and phytoplankton abundance explained 33.76% of the variation in *N. scintillans* abundance with the smallest AIC value. Among these environmental parameters, Chl-*a* concentration explained 25.07% (p < 0.01) of the aforementioned variation. The optimal GAM for the full year was expressed using the equation as follows:

 $Y = \alpha + f_1(\text{Chl-}a) + f_2(\text{Salinity}) + f_3(\text{Temperature})$

$+ f_4(\log(\text{Phytoplankton}))$

In spring, temperature (42.24%) explained more of the variation in the abundance of *N. scintillans* relative to the other variables (9.97%). A multi-peak nonlinear relationship existed between temperature and *N. scintillans* abundance (**Figure 7**). The abundance of *N. scintillans* peaked at between approximately 6°C and 13°C. However, for the other seasons, Chl-*a* concentration explained more variation in the abundance of *N. scintillans* relative to the other variables (**Table 4**).

DISCUSSION

Comparison of *N. Scintillans* Sampled Using Net-Collection and Water-Collection Methods

N. scintillans samples are usually collected using plankton nets (i.e., net-collection method) and bottles equipped with a CTD recorder (i.e., water-collection method) (Murray and Suthers, 1999; Zhang et al., 2016). The net-collection method collects *N. scintillans* from the entire water column through vertical towing from the bottom to the surface. Because the single cell of *N. scintillans* is large (diameter of 200–600 μ m), the species of *N. scintillans* is not easy to be lost in net-collected samples. The water-collection method collects *N.*



FIGURE 2 | Distribution of depth-average (A–D) temperature (°C) and (E–H) salinity in the upper 30-m water column, and (I–L) stratification index (kg m⁻³) in the East China Sea and southern Yellow Sea.

scintillans from discrete depths. We observed an obvious difference between the density of *N. scintillans* at different depths along section P in all seasons (**Figure 5**). The density of *N. scintillans* varied greatly in the water column (particularly in summer and autumn; **Figure 5**), the C_t was higher than C_d in winter and spring and vice versa in summer and autumn (**Table 1**). And the high density of *N. scintillans* was generally observed in the upper 30-m water column. Notably, the highest abundance (5,314 cells/L) was observed in summer at a depth of 10 m at station T07 (**Figure S3**). Once a water layer with very high or very low abundance is not collected, the depth-integrated abundance will be greatly affected. Therefore the net-collected method can more accurately reflect its abundance in

the entire water column compared with the water-collected method. Jiang et al. (2020) found that the species number of phytoplankton in water samples was slightly less than that in net samples. However, this method can obtain its vertical distribution information. Studies have used both methods (Murray and Suthers, 1999; Jiang et al., 2018; Jiang et al., 2020); thus, we collected *N. scintillans* samples using these two methods.

Adaptability of *N. Scintillans* to Environmental Factors

Phytoplankton biomass seemed to be the most crucial factor in the regulation of the seasonal and spatial distribution of *N. scintillans* in



FIGURE 3 | Distribution of (A–D) chlorophyll-a concentration (mg/m²) and (E–H) phytoplankton abundance (× 10⁸ cells/m²) in the East China Sea and southern Yellow Sea.

the ECS and SYS. Chl-*a* had the highest correlation coefficient among all the environmental factors in all seasons except winter (**Table 3**). Moreover, the correlations between Chl-*a* concentration and *N. scintillans* abundance were much weaker in cold seasons (winter and spring) than in warm seasons. The phytoplankton abundance and Chl-*a* concentration were too low and evenly distributed across the entire ECS and SYS in winter (**Figure 3**), probably resulting in a low correlation coefficient between *N. scintillans* abundance and Chl-*a* concentration. GAMs confirmed that Chl-*a* concentration explained more variation in the abundance of *N. scintillans* than that of temperature and salinity throughout the year and in all seasons except spring (**Table 4**). The abundance of *N. scintillans* was relatively lower in spring than in warm seasons, which required less phytoplankton biomass and was largely influenced by temperature. In spring, diatom blooms (high Chl-*a* concentration and phytoplankton abundance) occurred in the western SYS (Jiang et al., 2019) and central Zhejiang coastal waters (**Figure 3**), being inconsistent with the high-abundance regions (southern waters of Shandong Peninsula and southern Zhejiang coastal waters) of *N. scintillans* (**Figure 4**). Under an increasingly eutrophic Changjiang Diluted Water, the density of phytoplankton increased and the C_d of *N. scintillans* was revealed to be significantly higher in summer and autumn than in winter and spring (**Table 1**). From spring to summer, the average C_d of phytoplankton increased from 760.21 × 10⁶ to 1,316.66 × 10⁶ cells/m², whereas the average C_d of *N. scintillans* increased from

TABLE 2 | The depth-integrated abundance (× 10⁴ cells/m²) of N. scintillans using net-collection method in the East China Sea and southern Yellow Sea.

Season	East China	Sea	Southern Yellow Sea		
	Mean ± S.D.	Maximum	Mean ± S.D.	Maximum	
Spring	0.53 ± 1.42	6.87	1.41 ± 6.90	37.25	
Summer	556.59 ± 1037.32	4733.18	462.84 ± 1138.88	4733.18	
Autumn	2.30 ± 6.47	41.78	57.39 ± 218.27	1072.34	
Winter	0.11± 0.17	0.65	0.75 ± 2.25	9.27	





 1.06×10^4 to 354.17×10^4 cells/m². This seasonal variation is consistent with other observations. For example, Sun et al. (2011) noted that the bimodal distribution of *N. scintillans* abundance throughout the year was consistent with the bimodal distribution of phytoplankton in Jiaozhou Bay. In addition, a correlation analysis demonstrated a significant relationship between Chl-*a*

concentration and *N. scintillans* abundance in the northern Indian Ocean (Koochaknejad and Hamzei, 2022).

N. scintillans was distributed abundantly in coastal waters and the Changjiang Estuary, controlled by the eutrophic low-salinity Changjiang Diluted Water and coastal current (**Figures 4** and **6**), which were characterized by high levels of phytoplankton and

TABLE 3 | Correlation coefficients (Spearman's *r*) between the depth-integrated abundance of *N.scintillans* and environmental factors in the East China Sea and southern Yellow Sea.

Time	Temperature	Salinity	SI	Chl-a	Phytoplankton
Spring	0.207	0.002	0.226	0.258	0.148
Summer	0.306*	-0.268*	0.331**	0.647**	0.559**
Autumn	-0.064	-0.167	-0.361**	0.537**	0.387**
Winter	-0.254*	-0.357**	-0.117	0.200	-0.080
Annual	0.158*	-0.251**	0.157*	0.492**	0.357**

*p < 0.05, **p < 0.01.



indicates a bloom of *N. scintillans* with abundance of 5314 cells/L at station T07. CW: including Changjiang Diluted Water, the ECS Coastal Water, and the SYS Coastal Water; YSWCW, Yellow Sea Warm Current Water; TWCW, Taiwan Warm Current Water; YSCWM, the deeper Yellow Sea Cold Water Mass; KSW, Kuroshio surface water; KSSW, Kuroshio subsurface water; SMW, shelf mixed water. (A) spring; (B) summer; (C) autumn; (D) winter.

Chl-a (Figure 3) and associated nutrients (data not shown). Figure 5 illustrates the higher abundance of N. scintillans in the upper 30-m water column, characterized by abundant phytoplankton. Table 3 verifies that the C_tof N. scintillans in the upper 30-m water column was significantly and positively correlated with phytoplankton abundance and Chl-a concentration in all seasons except winter, demonstrating that the density of N. scintillans in the ECS and SYS was highly associated with phytoplankton biomass. Hallegraeff et al. (2019) observed a higher abundance (even bloom) of N. scintillans in the coastal waters of Australia (because of abundant food sources) than in offshore waters. A long-term study in the German Gulf revealed that the spatial distribution of N. scintillans had the highest abundance near the coast, with a decreasing trend from the coast to the North Sea with decreasing food sources (Uhlig and Sahling, 1990).

N. scintillans in the ECS and SYS is a heterotrophic dinoflagellate (Harrison et al., 2011). Thus, food abundance (particularly phytoplankton) strongly influences its abundance. Zhang et al. (2020a) demonstrated that the community structure

and nutrient conditions of food might influence the metabolism and bloom of the species of N. scintillans, and an experimental study verified that well-fed N. scintillans undergo rapid cell division, whereas starved cells stop dividing (Elbrächter and Qi, 1998). The proportion of individuals in binary fission of N. scintillans was significantly and positively correlated with Chl-a concentration, indicating that multiplication requires sufficient food (Tian et al., 2017). Therefore, food supply is a key factor shaping proliferation. N. scintillans can feed on microalgae, copepod eggs, fish eggs, and protozoa in a natural environment (Zhang et al., 2016). However, phytoplankton is considered to be its main food (Harrison et al., 2011), and its bloom is attributed to the increase in phytoplankton abundance. As shown in Figures 3 and 4, the high-abundance regions of N. scintillans were generally consistent with high-value waters of phytoplankton abundance and Chl-a concentration.

Temperature is generally considered to limit the distribution and bloom of *N. scintillans*. Red *N. scintillans* is distributed from temperate to tropical waters with temperature between 10° C and 30° C (Zhu et al., 2009; Kirchner et al., 2012). The optimal

Time	Environmental parameters	R ² adjusted	Cumulative explained deviation (%)	AIC	Optimal model
Spring	Temperature	0.33	42.24	237.21	
	Temperature + SI	0.39	48.87	232.83	✓
	Temperature + SI + Salinity	0.39	50.15	233.37	
	Temperature + SI + Salinity + Chl-a	0.39	52.05	234.16	
	Temperature + SI + Salinity + Chl-a + Phytoplankton	0.38	52.21	235.54	
Summer	Chl-a	0.46	50.22	267.24	
	Chl-a + Temperature	0.64	68.22	243.91	
	Chl-a + Temperature + Phytoplankton	0.67	71.15	241.28	
	Chl-a + Temperature + Phytoplankton + Sl	0.71	76.52	235.55	
	Chl-a + Temperature + Phytoplankton + SI + Salinity	0.72	78.17	235.31	1
Autumn	Chl-a	0.42	48.01	275.82	✓
	Chl-a + Salinity	0.46	55.47	276.04	
	Chl-a + Salinity + Temperature	0.46	56.35	276.68	
	Chl-a + Salinity + Temperature + Phytoplankton	0.52	61.85	261.22	
Winter	Chl-a	0.09	12.60	273.99	
	Chl-a + Salinity	0.14	18.19	270.81	
	Chl-a + Salinity + Temperature	0.24	34.48	267.56	
	Chl-a + Salinity + Temperature + Phytoplankton	0.25	37.63	267.45	\checkmark
Annual	Chl-a	0.23	25.07	1091.83	
	Chl-a + Salinity	0.28	31.09	1077.74	
	Chl-a + Salinity + Temperature	0.30	33.60	1071.04	
	Chl-a + Salinity + Temperature + Phytoplankton	0.30	33.76	1064.43	1

AIC, Akaike information criterion.



temperature for its growth ranges from 10°C; to 25°C; (Turkoglu, 2013; Gomes et al., 2014). Our results indicated that the C_d (359.89 × 10⁴ cells/m²) of *N. scintillans* in the ECS and SYS was substantially higher in summer (25.0°C; in average) than in other seasons (**Table 1**). However, *N. scintillans* bloomed with the highest density of up to 7.86 × 10⁷ cells/m² in the Changjiang Estuary and Zhejiang coastal waters (**Figures 2** and **4**) with a temperature (> 25°C) above the optimum growth temperature,

indicating that high temperature was not the limiting factors for the growth of *N. scintillans*. Field investigation and remote sensing data have revealed frequent blooms of *N. scintillans* in the Changjiang Estuary and adjacent waters in summer (Xu, 2009; Qi et al., 2019), and a Spearman correlation demonstrated that its density was positively correlated with temperature in spring and summer (**Table 3**). Notably, extremely high C_d of phytoplankton (11.36 × 10⁶ cells/m²) and Chl-*a* concentration (68.47 mg/m²) was observed in the Changjiang Estuary. We inferred that the promoting effect of phytoplankton on N. scintillans exceeded the limiting effect of high temperature. Although N. scintillans was negatively correlated with temperature in autumn and winter (Table 3), the explanation of Chl-a to variation in abundance of N. scintillans exceeded that of temperature (Table 4). Table 2 shows a higher abundance in the low-temperature SYS than in the high-temperature ECS in all seasons except summer (the hottest season). These results also supported our inference. Moreover, the annual abundance of N. scintillans was significantly and positively correlated with temperature. The low temperature (~4°C) in cold seasons did not limit distribution of N. scintillans in the ECS and SYS (Figure 7). Particularly in spring, temperature explained more variation in the abundance of N. scintillans than that of Chl-a concentration (Table 4). Our findings indicated a wide range of suitable temperatures (~4°C-30°C) and the high thermal adaptation of the species of N. scintillans. Therefore, temperature did not limit the distribution and bloom of N. scintillans in the ECS and SYS.

Notably, an extremely high N. scintillans abundance (particularly blooming at station T07) was observed in the Zhejiang coastal upwelling in summer caused by the strong intrusion of the deeper Taiwan Warm Current (Figures S1 and S3). Similarly, upwelling along the coast of China promoted the growth of phytoplankton, triggering a N. scintillans bloom (Zhang et al., 2020b). Dela-Cruz et al. (2008) also observed that N. scintillans was abundant in the mature upwelled waters in Australia. Figure S1 depicts a higher abundance of N. scintillans in the upper 20-m water column, characterized by relatively high Chl-a concentration. Moreover, our results revealed that the temperature (> 25°C;) was substantially higher in the upper 20 m than at other discrete depths beyond the optimal temperature for its growth. These findings therefore confirm that the abundance of N. scintillans in the ECS and SYS in warm seasons is largely regulated by phytoplankton biomass rather than temperature.

In addition, our results demonstrated that salinity did not limit the distribution of *N. scintillans* across the entire ECS and SYS. It was abundant in all seasons in the Changjiang Estuary and southern waters of the Shandong Peninsula, controlled by the low-salinity eutrophic Changjiang Diluted Water and coastal current (**Figures 2** and **4**). Because coastal low salinity is generally associated with high nutrient concentrations and the consequent abundant phytoplankton, the C_d of *N. scintillans* was negatively correlated with salinity in all seasons except spring. **Figures 4** and **6** verify that the species of *N. scintillans* was occasionally abundant in the high-salinity offshore waters controlled by the Taiwan Warm Current and Kuroshio. This observation is consistent with the remote sensing data revealing that *N. scintillans* blooms in the offshore saline waters of the ECS to at least 126° E (Qi et al., 2019). Several studies have also noted no marked relationship between *N. scintillans* abundance and salinity (Tada et al., 2004; Nasution et al., 2021). Our findings indicated that the abundance of *N. scintillans* in the ECS and SYS was largely shaped by phytoplankton rather than salinity.

Long-Term Changes in *N. Scintillans* Abundance

We collected historical data of N. scintillans abundance in the Changjiang Estuary using the net-collection method (Table 5). The abundance of N. scintillans in 2011 was considerably higher than that in earlier studies in all seasons except spring in 1959 and 2002. However, the annual average abundance increased sharply from 1.42 cells/m³ in 1959 to 1,466.60 cells/m³ in 2002, and then to 38,365.41 cells/m³ in 2011. Similarly, Wang et al. (2012) noted that N. scintillans abundance in the northern South China Sea in 2006–2008 was 27 times higher than that recorded in 1959. In the Adriatic Sea, N. scintillans bloomed as a result of eutrophication in the 1970s, but it did not bloom in the 1990s because of reduced eutrophication (Fonda-Umani et al., 2004). This increasing abundance of N. scintillans feeds on a larger number of phytoplankton and small protozoa (Hattori, 1962; Zhang et al., 2016), which may profoundly influence the planktonic food web and biogeochemical processes (e.g., primary production and carbon transfer) in the Changjiang Estuary, ECS, and SYS.

We inferred that the increasing trend in N. scintillans abundance in the Changjiang Estuary was largely attributed to the increase in phytoplankton abundance. Severe eutrophication fuels the growth of phytoplankton and frequent algal blooms in the Changjiang Estuary (Jiang et al., 2014; Wei et al., 2021). The extensive use of chemical fertilizers and discharge of industrial wastewater and domestic sewage increases N and P loads from the Changjiang (Cui et al., 2020). During the past 50 years, phytoplankton abundance and Chl-a concentration in the Changjiang Estuary have increased significantly with increasing eutrophication (Jiang et al., 2014; Chen et al., 2020). For example, the summer net-collected phytoplankton abundance in the Changjiang Estuary (28.0-32.0°N, 122.0-123.5°E) increased from 12.0 cells/mL in 1959 to 39.4 cells/mL in 2002 and then to 65.7 cells/mL in 2011. Furthermore, light penetration has enhanced and consequently promoted primary production in the Changjiang Estuary since the interception of a large amount of sediment by the Three Gorges Dam in 2003. Yang et al. (2007) calculated that the construction of the Three Gorges Dam decreased the sediment input into the estuary by 31% between

TABLE 5 | Comparison of the net-collected abundance of N. scintillans (cells/m³) in different years in the Changjiang Estuary (28°00'–32°00'N, 122°00'–123°30'E).

Time	Spring	Summer	Autumn	Winter	Annual
1959	0.08	5.34	0.14	0.11	1.42
2002	5864.19	2.19	0	0.02	1466.60
2011	179.93	152795.13	433.40	53.18	38365.41

2003 and 2005. Therefore, the severe light limitation in the Changjiang Estuary was reduced, which fueled the growth and bloom of phytoplankton as a result of deteriorated eutrophication (Chen, 2003; Chen et al., 2020).

Warming is probably another reason for the increase in the abundance of *N. scintillans*. Because of accelerated warming and the strengthening of the Taiwan Warm Current transport (Tang et al., 2009), the summer sea temperature of the Changjiang Estuary increased significantly (by 1.31° C; with a rate of 0.49° C;/ decade) from 1982 to 2009 (Jiang et al., 2014). The increase in temperature also resulted in an enhanced stratification, which facilitated the growth of dinoflagellates with a motional ability (Guo et al., 2014; Xiang et al., 2019). Our results verify that the annual abundance of *N. scintillans* was significantly and positively correlated with the SI in the ECS and SYS (**Table 3**). Tada et al. (2004) revealed that from late spring to early summer, the stability of water bodies leads to an increase in *N. scintillans* populations on the surface. *N. scintillans* can obtain more food sources through vertical migration (Xiang et al., 2019).

CONCLUSIONS

We first compared the abundance of N. scintillans using watercollection and net-collection methods during large-scale surveys in China's marginal seas. Our results suggest that the DIA of the species of N. scintillans varied between net-collected and watercollected samples, but a significant correlation between the C_d and C_t of N. scintillans was revealed in the ECS and SYS. The present study provides high spatial resolution data sets of N. scintillans in the ECS and SYS and verifies our hypothesis that its seasonal and spatial distribution would be largely regulated by phytoplankton biomass rather than temperature and salinity throughout the year. Particularly in warm seasons when the abundance of N. scintillans increased, its abundance was highly associated with Chl-a concentration and phytoplankton abundance. However, temperature explained more variation in the abundance of N. scintillans than other Chl-a concentration in winter. N. scintillans was abundant in the Changjiang Estuary and southern waters of the Shandong Peninsula, controlled by the eutrophic Changjiang Diluted Water and coastal current with high phytoplankton abundance and Chl-a concentration. In addition, N. scintillans is prone to bloom in Zhejiang coastal upwelling waters, characterized by abundant nutrients and phytoplankton. Since 1959, the abundance of N. scintillans in the Changjiang Estuary has increased sharply with the increasing phytoplankton density resulting from accelerated eutrophication and warming. These findings identified the distribution pattern,

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Ara, K., Nakamura, S., Takahashi, R., Shiomoto, A., and Hiromi, J. (2013). Seasonal Variability of the Red Tide-Forming Heterotrophic Dinoflagellate *Noctiluca Scintillans* in the Neritic Area of Sagami Bay, Japan: Its Role in the Nutrient-Environment and Aquatic Ecosystem. *Plankton. Benthos. Res.* 8 (1), 9–30. doi: 10.3800/pbr.8.9 controlling factors, red-tide formation mechanism, and longterm change of *N. scintillans* in the ECS and SYS, which improves our understanding of its regional population succession and ecological function.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**. Further inquiries can be directed to the corresponding authors.

AUTHOR CONTRIBUTIONS

LS: Writing-original draft preparation and data curation. ZJ: Conceptualization, reviewing and editing. ZS: Visualization. HZ: Investigation. LD: Data processing. MT: Data curation. JC: Data curation and funding acquisition. QC: Funding acquisition. JZ: Funding acquisition, reviewing and editing. All authors contributed to the article and approved the submitted version.

ACKNOWLEDGMENTS

We are grateful to Professor Wensheng Jiang for providing the physical data. We also thank Wallace for editing. This work was funded by the National Key Research and Development Program of China (2021YFC3101702), Key R&D Program of Zhejiang (2022C03044), NSFC-Zhejiang Joint Fund for the Integration of Industrialization and Informatization (U1709201), National Basic Research Program of China (2010CB428903), National Programme on Global Change and Air-Sea Interaction (Phase II) —Hypoxia and Acidification Monitoring and Warning Project in the Changjiang Estuary, and Longterm Observation and Research Plan in the Changjiang Estuary and Adjacent East China Sea Project (LORCE) (SZ2001).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmars.2022. 899334/full#supplementary-material

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